

Supporting Information

Chromium Catalyzed Sustainable C-C and C-N Bond Formation: C-Alkylation and Friedländer Quinoline Synthesis Using Alcohols

Vaishnavi Atreya^a, Sachin Jalwal^a, and Subrata Chakraborty*

Department of Chemistry, Indian Institute of Technology Jodhpur, Karwar, Jodhpur, 342037,
Rajasthan

* Corresponding author: Email: subrata@iitj.ac.in

Table of Contents

| S.No. | Title | Page no. |
|-------|--|----------|
| 1 | General information | S2 |
| | Experimental section | |
| 2 | a. Synthesis of Ligand precursors and PONN ^H b. Synthesis of complex Cr-1 c. Determination of Magnetic Susceptibility by Evans method. | S2 |
| 3 | Spectra of various studies of Cr-1 and catalytic reactions | S8 |
| 4 | General procedure for the catalytic reactions of Table S1 | S11 |
| 5 | Table S1: Optimization table for the α -alkylation of acetophenone using p-methyl benzyl alcohol | S14 |
| 6 | Analytical data of isolated products and NMR spectra | S15 |
| 7 | NMR spectra for the isolated compounds | S25 |

1. General information

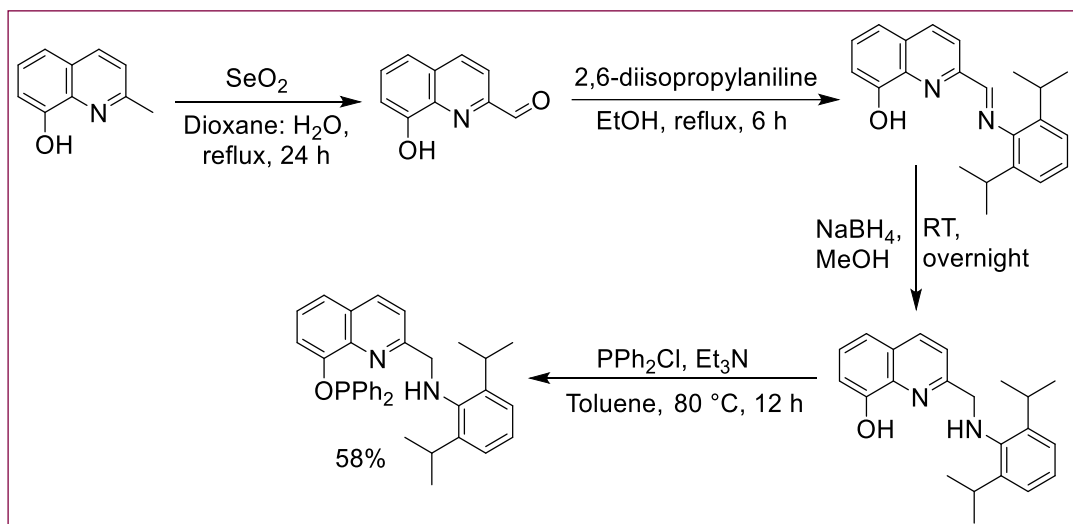
All manipulations were performed under inert nitrogen atmosphere by using standard Schlenk techniques and inside glove box, unless stated otherwise. Standard solvents were heated under reflux over suitable drying agent (sodium/benzophenone for THF, Et₂O, hexane; calcium hydride for CHCl₃ and kept under freshly dried molecular sieves), distilled, and stored over activated 4Å molecular sieves in the glovebox under nitrogen atmosphere. Deuterated solvents were purged with nitrogen and kept in glovebox. Unless otherwise stated, commercial reagents were used without purification, only benzyl alcohol and acetophenone were purified via vacuum distillation. Chromium chloride (CrCl₂) was purchased from Sigma- Aldrich, calcium hydride (CaH₂) and all other materials were purchased from Avra, BLDpharma, TCI and Sigma- Aldrich, stored under nitrogen and used as received. Ligand precursors were prepared according to literature procedure.^[1, 2] Reaction temperature reported is the temperature of the oil bath. All the reaction for ligand synthesis and substrate scope was monitored by TLC and isolated by column chromatography or flash chromatography (silica gel 60-120). Then product was condensed with the help of rotary evaporator. Isolated product was characterized by using ¹H NMR, ¹³C NMR from Bruker 500 MHz spectrometer using CDCl₃ or C₆D₆. All spectra were recorded at room temperature, unless otherwise noted. ATR-IR spectra was recorded on Perkin-Elmer FT-IR spectrometer. Solution-state UV–visible absorption spectrum was recorded on Cary 4000 UV–vis spectrophotometer using dichloromethane (DCM) solvent. Mass spectra were recorded on Agilent spectrometer. EPR analysis was performed on a JES-FA200 ESR Spectrometer in the X-band region (8.75–9.65 GHz). Elemental analysis was carried out on a Flash smart V CHNS/O.

2. Experimental Section:

a) Synthesis of ligand precursors and N-(((8-(((diphenylphosphaneyl)oxy)quinolin-2-yl)methyl)-2,6-diisopropylaniline (PONN^H)

(2-(((2,6-diisopropylphenyl)amino)methyl)quinolin-8-ol) was prepared according to the procedure in literature.^[1] 8-hydroxy-2-quinolinecarbaldehyde (1g, 5.83 mmol) which was obtained by oxidation of 8-methyl quinolinol by SeO₂, was heated in ethanol (30 mL) to 80 °C, followed by dropwise addition of ethanol solution of phenylamine (540 mg, 5.83 mmol in 30 mL ethanol). This reaction mixture was allowed to reflux for 6 h and then cooled to room temperature. The precursor was obtained in pure form by column chromatography using 1/3 dichloromethane/ petroleum ether (1% triethylamine) in the form of yellow needles (yield = 58%).

(2-(((2,6-diisopropylphenyl)amino)methyl)quinolin-8-ol) (1004 mg, 3 mmol) was dissolved in 20 ml toluene in a 100 mL Schlenk tube under nitrogen atmosphere, and an equimolar amount of triethyl amine (418 μL, 3 mmol) was added to it dropwise, while stirring vigorously. After 5 min., 3 mmol (540 μL) of chlorodiphenylphosphine was also added dropwise, while stirring the solution vigorously at room temp. Now the Schlenk tube was closed tightly and kept for heating at 80 °C overnight. The reaction mixture was cooled to room temperature and the solvent was evaporated under vacuum. The residue was extracted with 20 mL of toluene and filtered through celite. Again the solvent was evaporated under vacuum. This product was again washed with dried hexane, and finally obtained as colourless oily liquid in 58% yield after drying in vacuum. The purity of the ligand was checked with ³¹P NMR data. ³¹P NMR (203 MHz, Chloroform-*d*) δ 117.3. ¹H NMR (500 MHz, Chloroform-*d*) δ 7.93 (d, *J* = 8.4 Hz, 1H), 7.70 (dd, *J* = 11.5, 7.6 Hz, 1H), 7.63 (dd, *J* = 13.7, 7.5 Hz, 1H), 7.51 (t, *J* = 7.9 Hz, 6H), 7.29 (dt, *J* = 8.3, 4.1 Hz, 1H), 7.22 (d, *J* = 7.3 Hz, 1H), 7.16 (t, *J* = 7.3 Hz, 6H), 7.10 (d, *J* = 7.7 Hz, 1H), 7.07 (d, *J* = 8.2 Hz, 1H), 7.00 (d, *J* = 7.7 Hz, 1H), 6.84 (s, 1H), 4.78 (s, 1H), 3.60 (p, *J* = 6.6 Hz, 2H), 1.06 (d, *J* = 6.6 Hz, 12H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 152.78, 143.44, 135.40 (d, *J* = 7.1 Hz), 132.59 (d, *J* = 2.8 Hz), 131.85, 131.66, 131.51, 131.42 – 131.17 (m), 130.79, 130.70, 130.38, 129.61, 128.97, 128.87, 128.64 (d, *J* = 7.1 Hz), 128.45 (dd, *J* = 14.3, 9.7 Hz), 119.95, 117.53, 113.03, 45.82, 28.34, 24.49. IR frequencies (ATR): 3380 (N-H), 2962 (C=C-H), 2861 (C-C-H), 1589 (aryl C-H).



Scheme S1: Synthetic access to ligand N-((8-((diphenylphosphaneyl)oxy)quinolin-2-yl)methyl)-2,6-diisopropylaniline (PONNH)

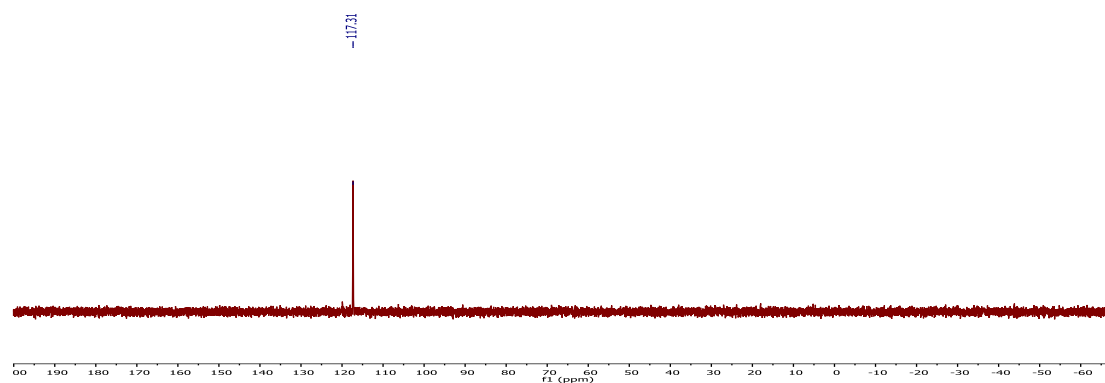


Figure S1. ^{31}P NMR spectrum of PONNH in CDCl_3 at room temperature.

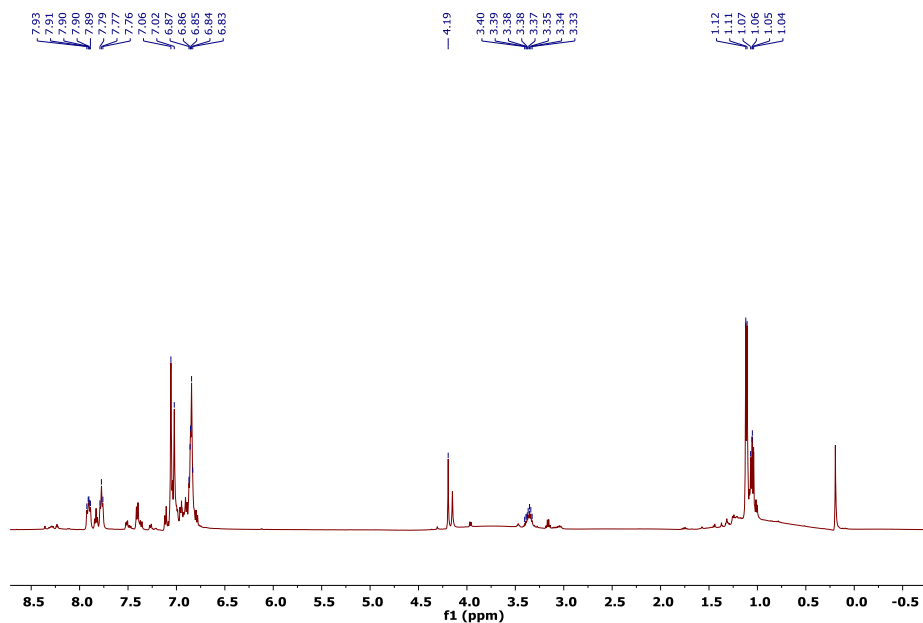


Figure S2: ^1H NMR spectrum of the PONNH^H ligand in C_6D_6 at room temperature (contains precursor impurities).

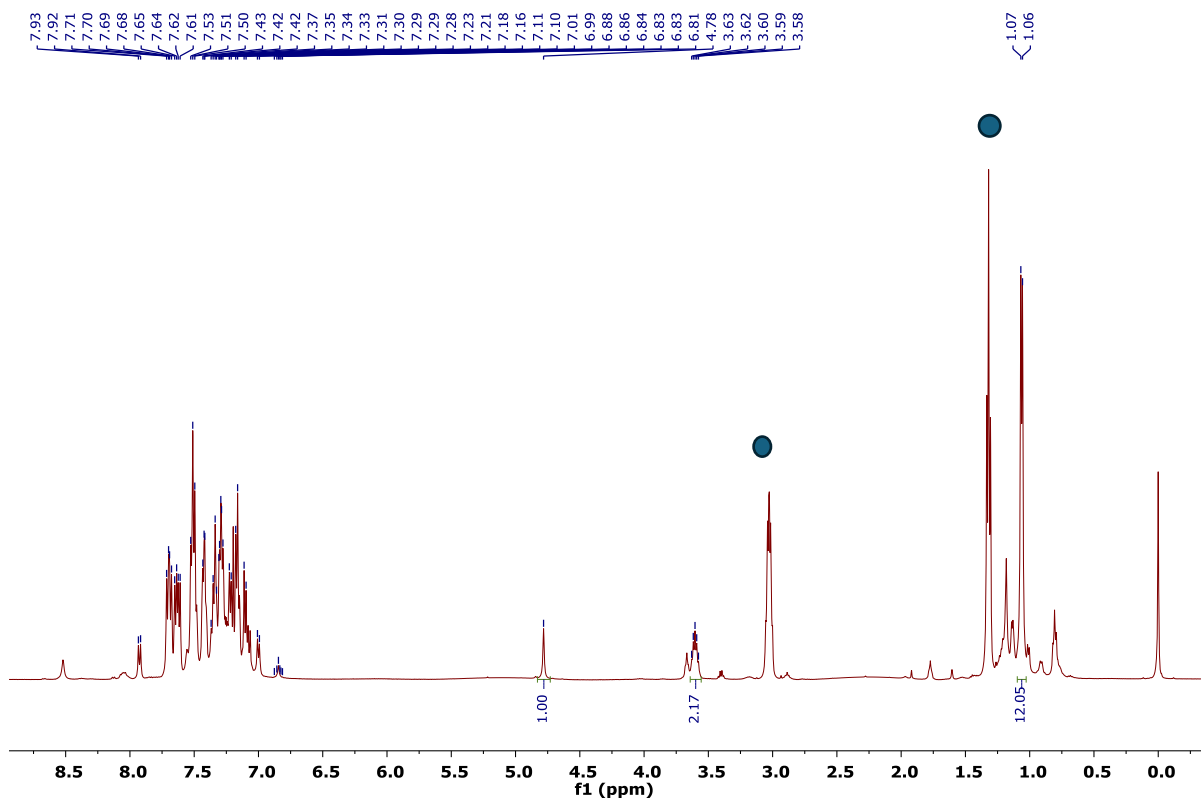


Figure S3: ^1H NMR spectrum of the crude reaction mixture for the synthesis of PONNH^H ligand in C_6D_6 at room temperature containing Et_3NH^+ salt ●.

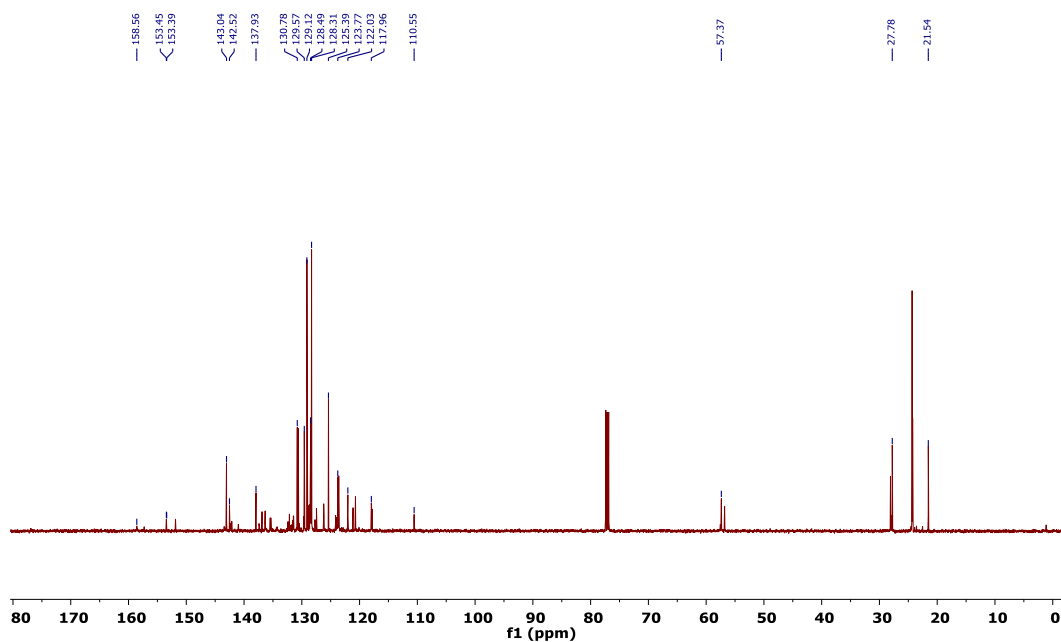


Figure S4: ^{13}C NMR spectrum of PONNH^H in CDCl_3 at room temperature.

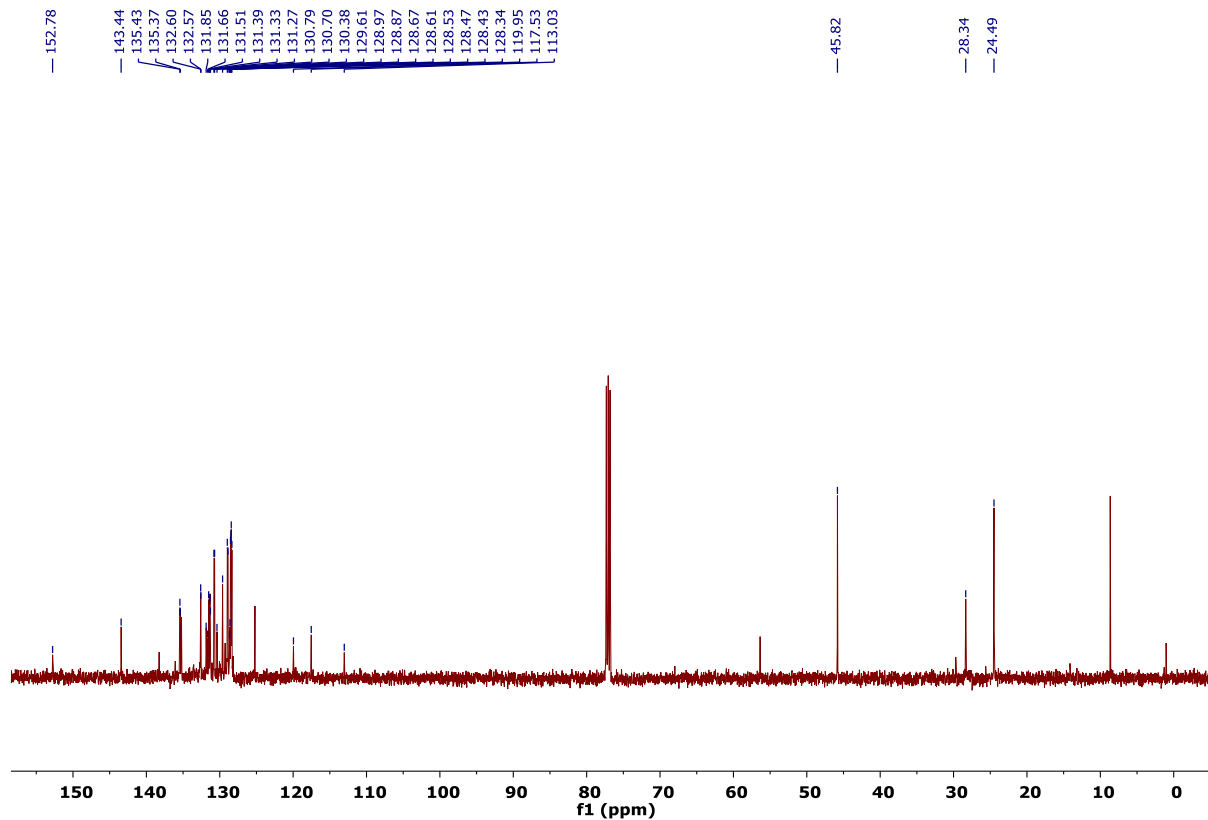


Figure S5: ^{13}C NMR spectrum of the crude reaction mixture of PONNH^H in CDCl_3 at room temperature.

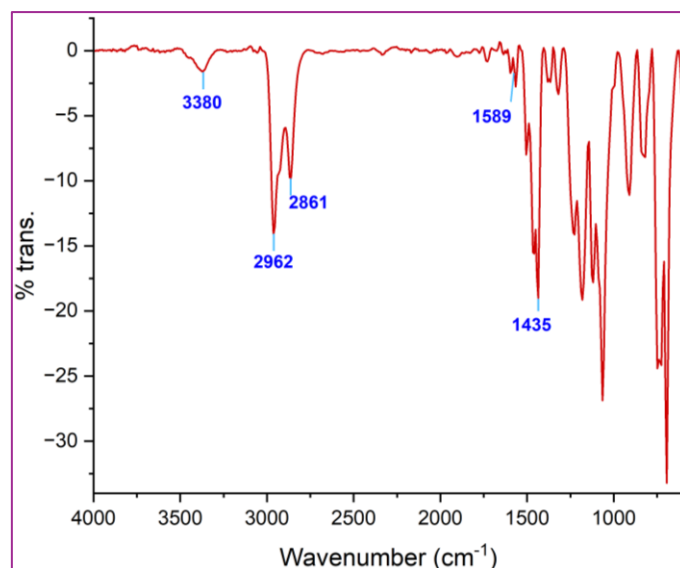
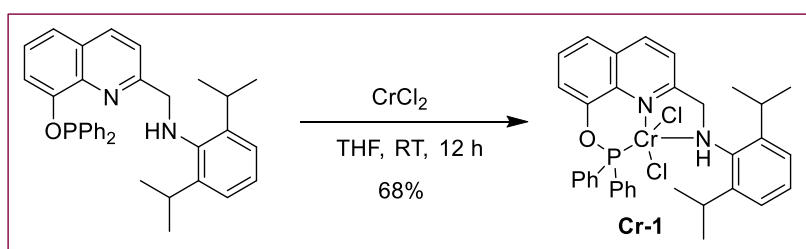


Figure S6: IR data for **PONNH** ligand (ATR)

b) Synthesis of $\text{CrCl}_2(\text{PONNH})$ (Cr-1)

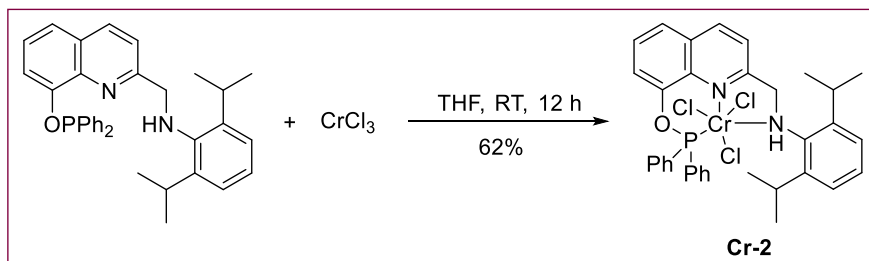
To a suspension of 5 mmol CrCl_2 in THF, 1 equiv. of PONNH^H ligand was added dropwise and stirred at room temperature overnight. Dark green coloured suspension formed was washed twice with hexane and dried in vacuo to get green crystalline powder. $^1\text{H NMR}$ (500 MHz, Benzene- d_6) δ 7.32 (s, 2H), 7.08 – 7.02 (m, 44H), 7.00 (s, 3H), 4.79 (s, 0H), 4.27 (s, 1H), 2.25 (s, 1H), 2.11 (s, 1H), 1.55 (s, 2H). **Selected IR data (ATR, cm^{-1}):** 3054 (N-H), 2959 (C-H) 1564(N-H). **UV-vis (DCM) λ/nm :** 274nm (sharp), 374 (broad). **HRMS calcd. for $\text{C}_{34}\text{H}_{35}\text{Cl}_2\text{CrN}_2\text{OP} + \text{Na}^+$:** 663.1161, observed: 663.4545. **Room temperature X-band EPR, g value:** 2.002. **Chemical Formula:** $\text{C}_{34}\text{H}_{35}\text{Cl}_2\text{CrN}_2\text{OP}$. **Elemental Analysis (calculated) :** C, 63.66; H, 5.50; Cl, 11.05; Cr, 8.10; N, 4.37; O, 2.49; P, 4.83. **Observed:** C, 59.205; H, 5.888; N, 3.736.



Scheme S2: Preparation of **Cr-1**.

c) Synthesis of $\text{CrCl}_3(\text{PONNH})$:

To a suspension of 80 mg (0.5 mmol) CrCl_3 in THF, 1 equiv. of THF solution of PONNH^H ligand (260 mg) was added dropwise and stirred at room temperature overnight. The brown coloured suspension so formed was dried in vacuo and washed twice with hexane to get golden brown powder in 62% yield. **HRMS calcd. for $\text{C}_{34}\text{H}_{35}\text{Cl}_3\text{CrN}_2\text{OP} + 2\text{Na}^+$:** 721.0958, observed: 721.0955, for $\text{C}_{34}\text{H}_{35}\text{Cl}_3\text{CrN}_2\text{OP} + 2\text{Na}^+ + 2\text{Li}^+$: 735.0958, observed: 735.1824; for $\text{C}_{34}\text{H}_{35}\text{Cl}_3\text{CrN}_2\text{OP} - \text{Cl}^- + \text{Na}^+$ 663.0998: , observed: 663.1467 .



Scheme S3: Preparation of **Cr-2**.

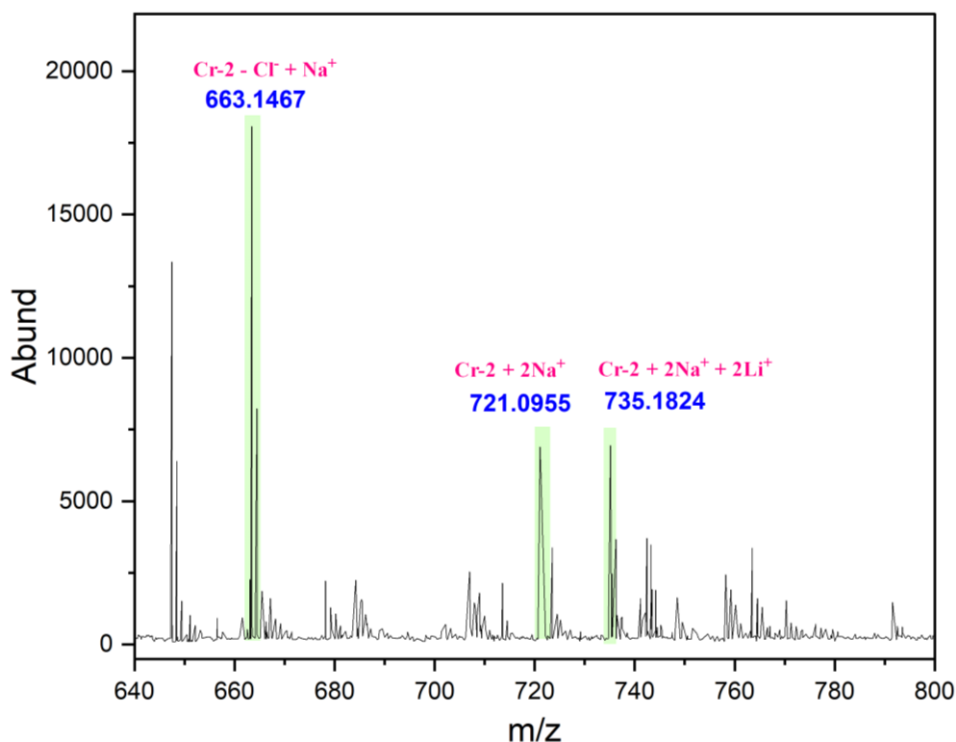


Figure S7: HRMS spectrum of **Cr-2**.

d) **Determination of Magnetic Susceptibility by Evans Method.**^[3, 4]

Weight of catalyst taken for NMR measurement = 2.5 mg

Molecular mass of Cr-1 = 641.5369

Shift in ¹H NMR peak of CDCl₃ (400 MHz) ($\Delta\nu$) = 0.1156 ppm = 46.24 Hz

By using the following formula for calculating χ_m :

$$\chi_m = \frac{\Delta\nu}{S_f \nu_0} \times \frac{1000}{c}$$

Where, $S_f = \frac{4\pi}{3}$, the value of χ_m was calculated to be equal to 0.00354 cm³mol⁻¹.

Putting this value in following equation, (taking the value of $\chi_m = \chi^{para}$), we get:

$$\mu_{eff} = \sqrt{8\chi_m^{para} T}$$

Where T = 298K, $\mu_{eff} = \sqrt{8 \cdot 225} = 2.86$.

3. Spectra of various studies of Cr-1 and catalytic reactions

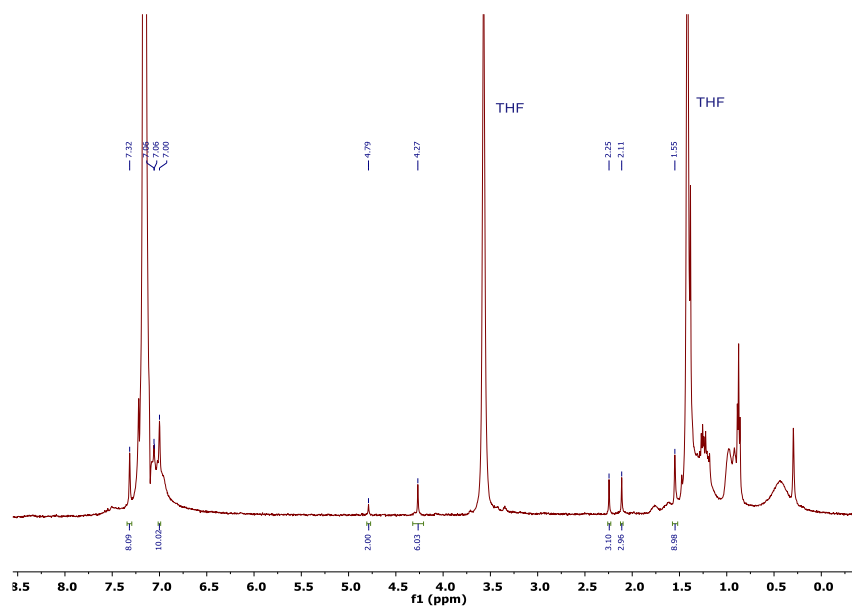


Figure S8: ^1H NMR for Cr-1.

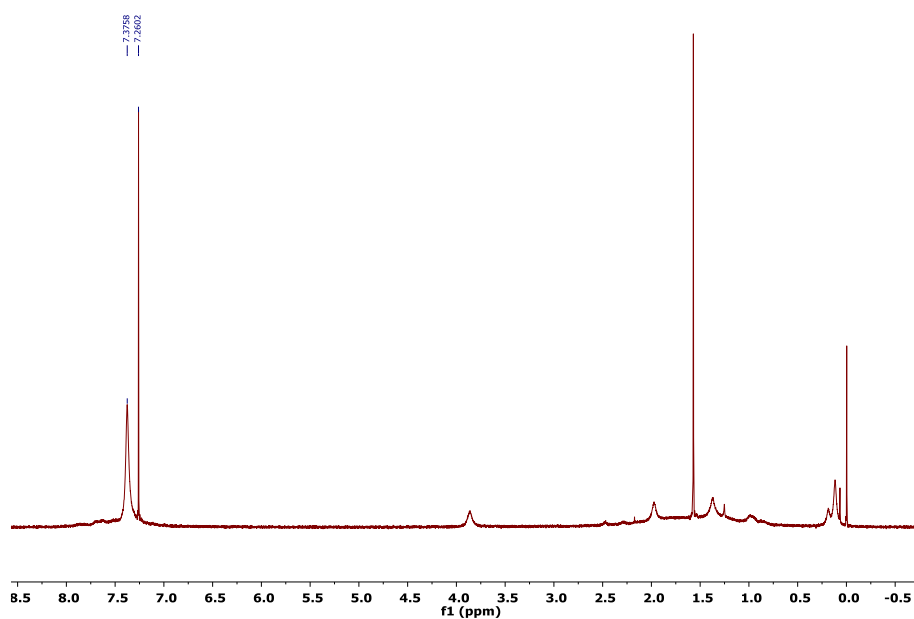


Figure S9: ^1H NMR data of Cr-1 for the Evans measurement.

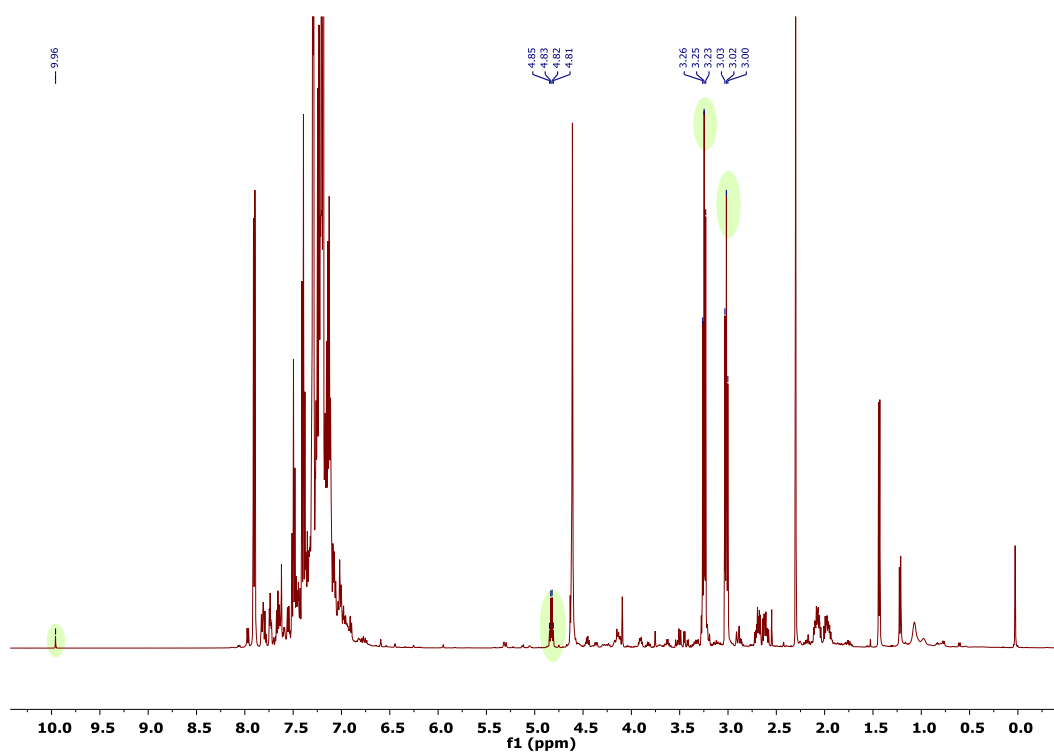


Figure S10: ^1H NMR of the catalytic reaction mixture of benzyl alcohol and acetophenone after 3h at 135 °C.

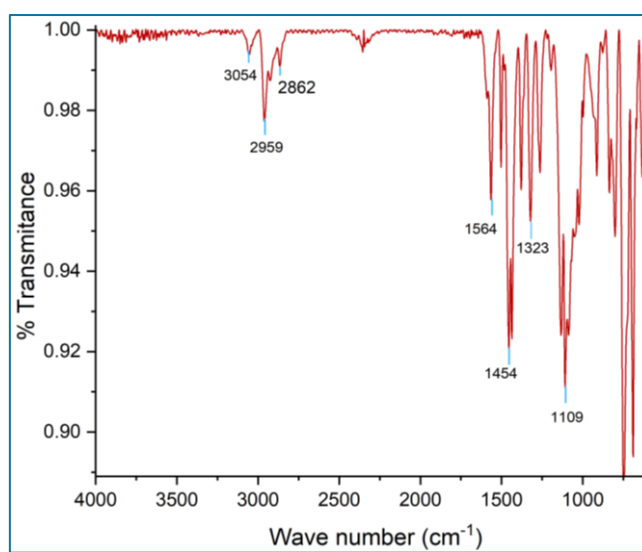


Figure S11: IR data for Cr-1.

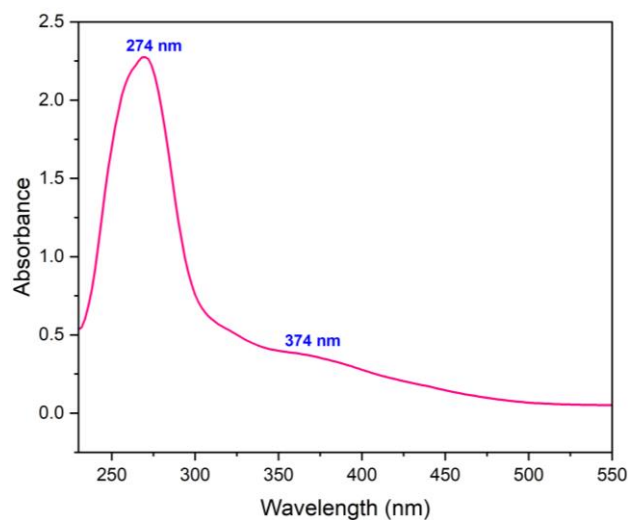


Figure S12: UV-Vis. spectrum of **Cr-1** in DCM.

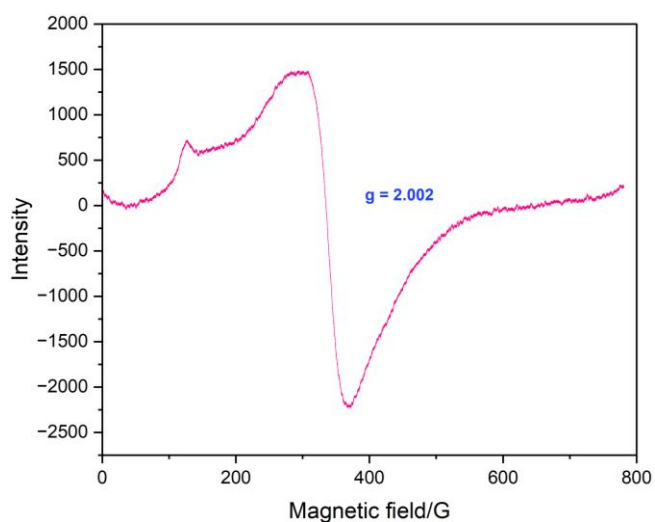


Figure S13: Room temperature X-band EPR for **Cr-1**.

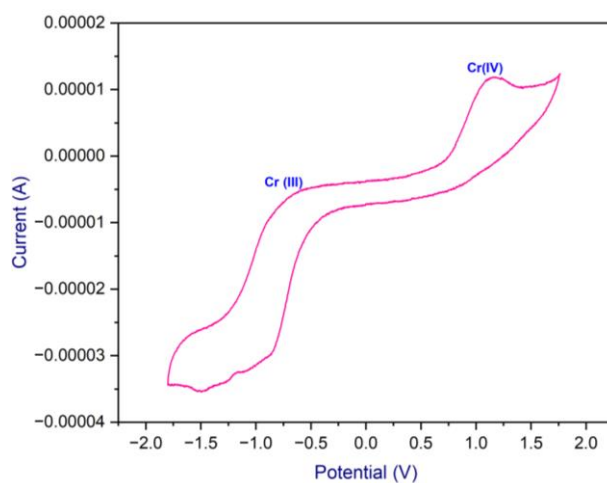


Figure S14: CV curve of **Cr-1**. Conditions: 1.0 mM analyte in 0.1 M TBAPF₆/MeCN, under dry argon atmosphere, glassy carbon working electrode, platinum wire counter electrode, Ag/AgCl reference electrode, 0.02 V/s scan rate.

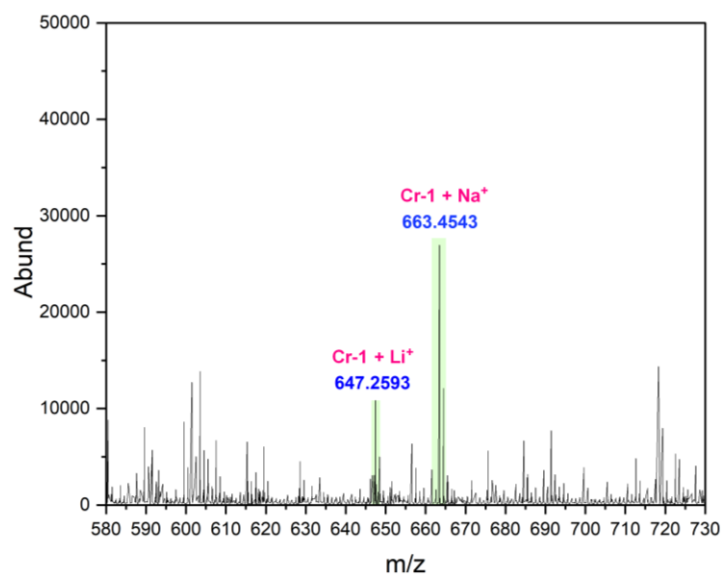


Figure S15: HRMS data for $\text{CrCl}_2(\text{PONNH})$ complex ($m/z + \text{Na}^+$ and $m/z + \text{Li}^+$)

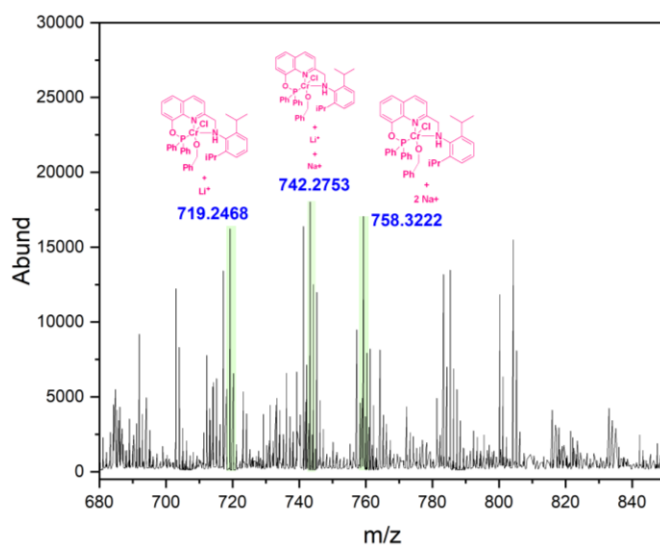


Figure S16: HRMS data for the formation of Alkoxy-complex **Cr-1b**.

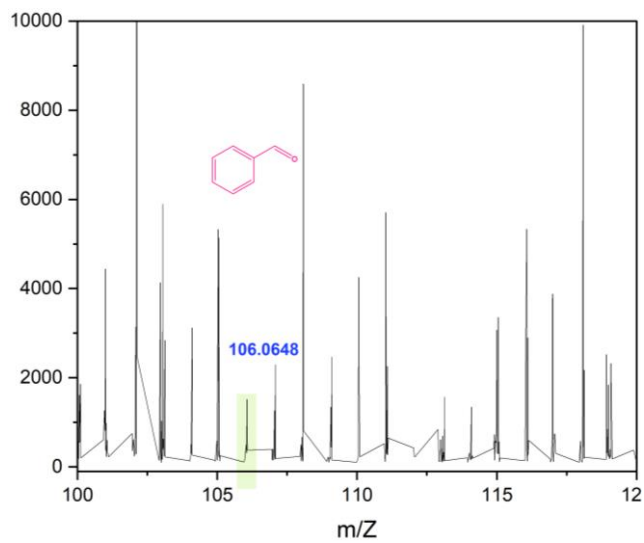


Figure S17: HRMS data for aldehyde formation during catalytic reaction via dehydrogenation of benzyl alcohol.

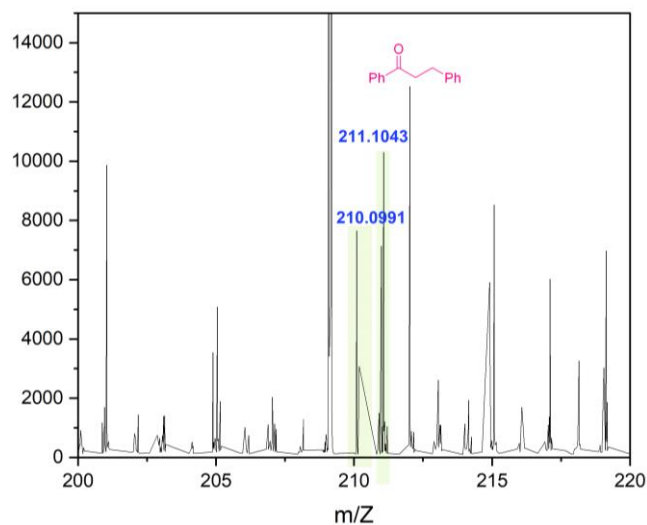


Figure S18: HRMS data for product formation 1,3-diphenylpropan-1-one (expected HRMS: $m/z = 212.1201$, observed = 212.0914)

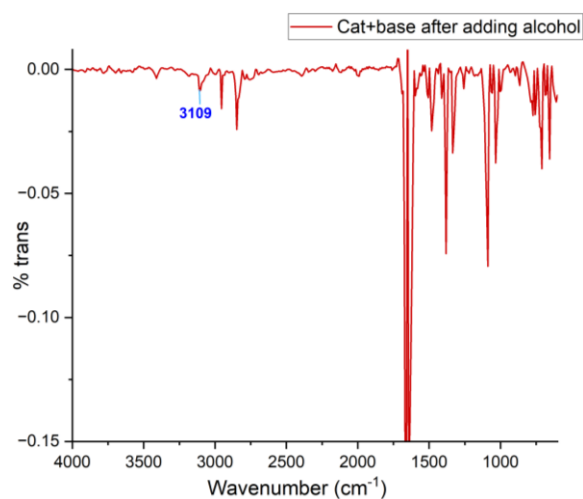


Figure S19: IR spectrum recorded after adding benzyl alcohol to the reaction mixture of complex **Cr-1** and *t*BuOK (ATR-IR).

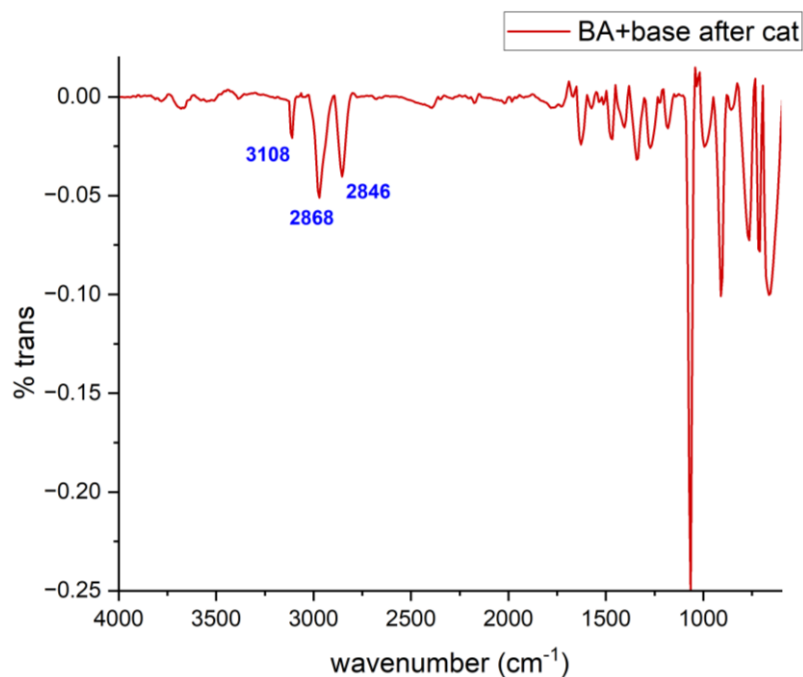
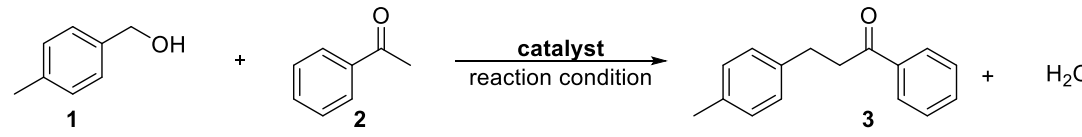
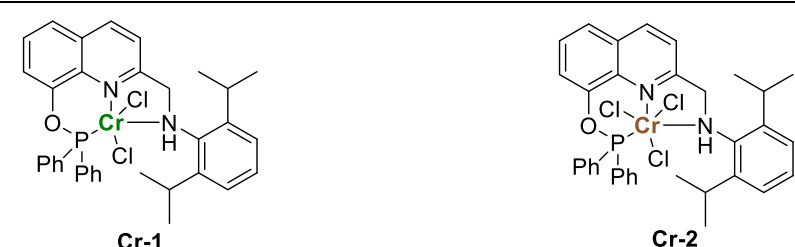


Figure S20: IR spectrum recorded after adding complex **Cr-1** to potassium benzyloxy species (formed by the reaction of benzyl alcohol with *t*BuOK) (ATR-IR).

5. General procedure for the catalytic reactions of Table S1:

0.5 mmol of *p*-methyl benzyl alcohol and 0.5 mmol of acetophenone were added to 0.015 mmol of catalyst **Cr-1** and 20 mol% of ^tBuOK (or other bases, as mentioned in Table 1) and the mixture was dissolved in 1.5 mL of toluene (or *t*-Amyl alcohol or 1,4-dioxane or THF; Table 1, entries 11, 12, 13, and 14 respectively) and placed in a 25 mL Schlenk tube under N₂. The tube was heated at 120-135 °C with stirring for the mentioned time. The reaction mixture was then cooled down and the products formation was monitored using TLC. Products were analyzed by NMR spectroscopy using 1,1,2,2-tetrachloroethane as internal standard.

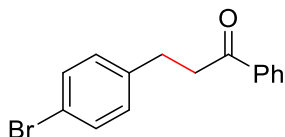
6. Table S1: Optimization table for the α -alkylation of acetophenone using *p*-methylbenzyl alcohol.^a

|  | | | | | | |
|--|---------------------------------------|-----------------------|---------------------------------|------------------|----------|----------|
|  | | | | | | |
| entry | base (mol %) | catalyst (mol %) | solvent (mL) | temperature (°C) | time (h) | yield(%) |
| 1 | KO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 120 | 24 | 62 |
| 2 ^b | KO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 76 |
| 3 | KO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 88 |
| 4 | KO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 36 | 92 |
| 5 | NaOH (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 84 |
| 6 | KOH (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 80 |
| 7 | NaO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 78 |
| 8 | Cs ₂ CO ₃ (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 86 |
| 9 | LiO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 24 | 84 |
| 10 | - | Cr-1 (3) | Toluene (1.5) | 135 | 24 | <5 |
| 11 | KO ^t Bu (20%) | Cr-1 (3) | <i>tert</i> -amyl alcohol (1.5) | 135 | 24 | 50 |
| 12 | KO ^t Bu (20%) | Cr-1 (3) | Dioxane (1.5) | 135 | 24 | trace |
| 13 | Cs ₂ CO ₃ (20%) | Cr-1 (3) | Dioxane (1.5) | 135 | 24 | trace |
| 14 | KO ^t Bu (20%) | Cr-1 (3) | THF(1.5) | 135 | 24 | trace |
| 15 ^c | KO ^t Bu (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 36 | 98 |
| 16 ^c | KO ^t Bu (20%) | Cr-1 (3) | chlorobenzene (1.5) | 135 | 36 | 45 |
| 17 ^c | KO ^t Bu (20%) | Cr-1 (3) | mesitylene (1.5) | 135 | 36 | 71 |
| 18 ^c | KO ^t Bu (20%) | - | Toluene (1.5) | 135 | 36 | trace |
| 19 ^c | KO ^t Bu (20%) | CrCl ₂ (3) | Toluene (1.5) | 135 | 36 | 20 |
| 20 ^c | LiOH (20%) | Cr-1 (3) | Toluene (1.5) | 135 | 36 | 87 |
| 21 ^{b,c} | KO ^t Bu (20%) | Cr-2 (3) | Toluene (1.5) | 135 | 24 | 59 |
| 22 ^d | NaO ^t Bu (5%) | Cr-2 (0.005) | - | 140 | 3 | 6 |
| 23 ^d | NaO ^t Bu (20%) | Cr-2 (3) | Toluene (1) | 135 | 24 | 54 |

^aReaction condition: *p*-methyl benzyl alcohol (0.5 mmol) and acetophenone (0.5 mmol). ^bUsing Benzyl alcohol as substrate. ^cYields were determined by ¹H NMR analysis based on substrate consumption or 1,1,2,2-tetrachloroethane as internal standard. ^dBenzyl alcohol (2 mmol) and 1-phenylethanol (2 mmol). Yields were determined by ¹H NMR analysis based on substrate consumption or 1,1,2,2-tetrachloroethane as internal standard.

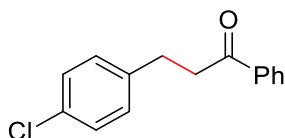
7. Analytical data of isolated products and NMR spectra

3a) 3-(4-bromophenyl)-1-phenylpropan-1-one⁵



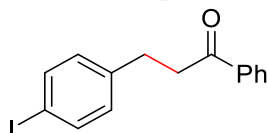
¹H NMR (500 MHz, Chloroform-*d*) δ 7.98 (d, J = 7.0 Hz, 2H), 7.62 – 7.56 (m, 1H), 7.48 (t, J = 7.7 Hz, 2H), 7.44 (d, J = 8.3 Hz, 2H), 7.16 (d, J = 8.4 Hz, 2H), 3.31 (t, J = 7.5 Hz, 2H), 3.06 (t, J = 7.5 Hz, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.27, 140.73, 137.20, 133.65, 132.01, 130.71, 129.11, 128.47, 120.35, 40.53, 29.89.

3b) 3-(4-chlorophenyl)-1-phenylpropan-1-one⁵



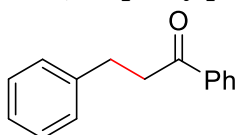
¹H NMR (500 MHz, Chloroform-*d*) δ 7.98 – 7.93 (m, 2H), 7.56 (d, J = 7.2 Hz, 1H), 7.46 (t, J = 7.7 Hz, 2H), 7.26 (d, J = 8.3 Hz, 2H), 7.19 (d, J = 8.3 Hz, 2H), 3.28 (t, J = 7.5 Hz, 2H), 3.05 (t, J = 7.5 Hz, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.31, 140.19, 137.18, 133.64, 132.31, 130.29, 129.07 (d, J = 6.8 Hz), 128.46, 40.59, 29.82.

3c) 3-(4-iodophenyl)-1-phenylpropan-1-one⁶



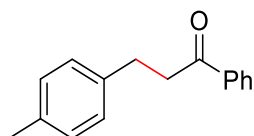
¹H NMR (500 MHz, Chloroform-*d*) δ 7.95 (d, J = 7.0 Hz, 2H), 7.61 (d, J = 8.3 Hz, 2H), 7.56 (t, J = 7.4 Hz, 1H), 7.46 (t, J = 7.7 Hz, 2H), 7.01 (d, J = 8.3 Hz, 2H), 3.28 (t, J = 7.6 Hz, 2H), 3.02 (t, J = 7.5 Hz, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.25, 141.43, 138.01, 137.21, 133.66, 131.06, 129.12, 128.49, 91.67, 40.51, 30.00.

3d) 1,3-diphenylpropan-1-one⁵



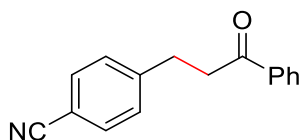
¹H NMR (500 MHz, Chloroform-*d*) δ 7.88 (d, J = 6.9 Hz, 2H), 7.47 (t, J = 7.4 Hz, 1H), 7.37 (t, J = 7.7 Hz, 2H), 7.21 (d, J = 7.3 Hz, 2H), 7.18 (d, J = 6.5 Hz, 2H), 7.13 (t, J = 7.1 Hz, 1H), 3.23 (t, 2H), 2.99 (t, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.70, 141.76, 137.33, 133.53, 129.07, 129.00, 128.89, 128.51, 126.60, 40.92, 30.60.

3e) 1-phenyl-3-(p-tolyl)propan-1-one⁵



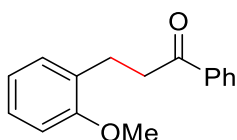
¹H NMR (500 MHz, Chloroform-*d*) δ 7.96 (d, J = 7.6 Hz, 2H), 7.56 (t, J = 7.5 Hz, 1H), 7.46 (t, J = 7.7 Hz, 2H), 7.16 (d, J = 8.0 Hz, 2H), 7.12 (d, J = 7.8 Hz, 2H), 3.31 – 3.27 (m, 2H), 3.07 – 3.01 (m, 2H), 2.33 (s, 3H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.37, 138.21, 136.91, 135.65, 133.05, 129.23, 128.62, 128.32, 128.07, 40.64, 29.74, 21.03.

3f) 4-(3-Oxo-3-phenylpropyl)benzotrile⁷



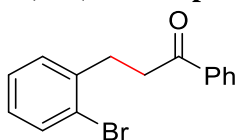
^1H NMR (500 MHz, Chloroform-*d*) δ 7.94 (d, $J = 7.0$ Hz, 2H), 7.57 (t, $J = 7.6$ Hz, 3H), 7.46 (t, 2H), 7.37 (d, $J = 8.0$ Hz, 2H), 3.33 (t, $J = 7.4$ Hz, 2H), 3.14 (t, $J = 7.4$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 198.72, 147.48, 137.03, 133.83, 132.80, 129.82, 129.19, 128.47, 119.46, 110.56, 39.92, 30.47.

3g) 3-(2-methoxyphenyl)-1-phenylpropan-1-one⁵



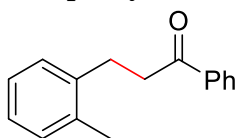
^1H NMR (500 MHz, Chloroform-*d*) δ 8.00 (d, $J = 8.1$ Hz, 2H), 7.56 (t, $J = 7.4$ Hz, 1H), 7.46 (t, $J = 7.6$ Hz, 2H), 7.26 – 7.19 (m, 2H), 6.92 (d, $J = 7.4$ Hz, 1H), 6.88 (d, $J = 8.6$ Hz, 1H), 3.84 (s, 3H), 3.28 (t, $J = 8.7$, 6.8 Hz, 2H), 3.07 (t, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 200.41, 157.95, 137.41, 133.32, 130.59, 129.95, 128.95, 128.54, 127.94, 120.96, 110.68, 55.62, 39.37, 26.16.

3h) 3-(2-bromophenyl)-1-phenylpropan-1-one⁸



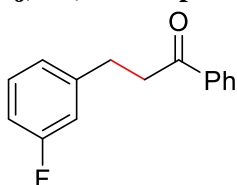
^1H NMR (500 MHz, Chloroform-*d*) δ 7.90 (d, $J = 7.0$ Hz, 2H), 7.48 (t, $J = 7.6$ Hz, 2H), 7.38 (t, $J = 7.7$ Hz, 2H), 7.24 (dd, $J = 7.6$, 1.8 Hz, 1H), 7.16 (dd, $J = 7.5$, 1.3 Hz, 1H), 7.00 (td, $J = 7.7$, 1.8 Hz, 1H), 3.24 (t, $J = 7.9$ Hz, 2H), 3.11 (t, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 199.41, 141.05, 137.23, 133.60, 133.37, 131.29, 129.09, 128.56, 128.46, 128.11, 124.84, 39.09, 31.28.

3i) 1-phenyl-3-(*o*-tolyl)propan-1-one⁸



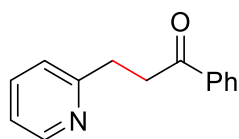
^1H NMR (500 MHz, Chloroform-*d*) δ 7.99 (d, $J = 8.3$ Hz, 2H), 7.58 (t, $J = 7.3$ Hz, 1H), 7.48 (t, $J = 7.7$ Hz, 2H), 7.21 (t, $J = 5.4$ Hz, 1H), 7.19 – 7.14 (m, 3H), 3.30 – 3.25 (m, 2H), 3.11 – 3.06 (m, 2H), 2.38 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 199.41, 139.42, 136.88, 136.03, 133.27, 133.00, 130.39, 128.79, 128.55, 128.20, 126.37, 126.23, 39.15, 27.55, 19.48.

3j) 3-(3-fluorophenyl)-1-phenylpropan-1-one⁸



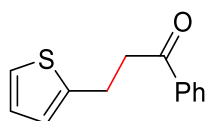
^1H NMR (500 MHz, Chloroform-*d*) δ 7.95 (d, $J = 7.9$ Hz, 2H), 7.55 (t, 1H), 7.45 (t, $J = 7.8$ Hz, 2H), 7.23 (t, 1H), 7.02 (d, $J = 7.6$ Hz, 1H), 6.95 (d, $J = 9.9$ Hz, 1H), 6.88 (td, $J = 8.6$, 2.6 Hz, 1H), 3.29 (t, $J = 8.2$, 7.0 Hz, 2H), 3.06 (t, $J = 7.6$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 199.24, 163.41 (d, $J = 245.5$ Hz), 144.32 (d, $J = 7.1$ Hz), 137.22, 133.64, 130.40 (d, $J = 8.6$ Hz), 128.49, 124.57 (d, $J = 2.7$ Hz), 115.79 (d, $J = 20.8$ Hz), 113.49 (d, $J = 20.9$ Hz), 40.44, 30.21 (d, $J = 1.7$ Hz). ^{19}F NMR (376 MHz, Chloroform-*d*) δ -113.44.

3l) 1-phenyl-3-(pyridin-2-yl)propan-1-one⁹



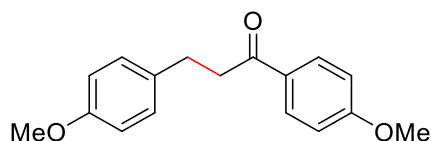
¹H NMR (500 MHz, Chloroform-*d*) δ 8.51 (d, *J* = 4.6 Hz, 1H), 7.98 (d, *J* = 6.9 Hz, 2H), 7.61 (td, *J* = 7.7, 1.9 Hz, 1H), 7.54 (t, *J* = 7.4 Hz, 1H), 7.44 (t, *J* = 7.7 Hz, 2H), 7.28 (d, *J* = 7.8 Hz, 1H), 7.12 (td, 1H), 3.52 (t, *J* = 7.2 Hz, 2H), 3.25 (t, *J* = 7.2 Hz, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.68, 161.01, 149.29, 137.29, 137.22, 133.49, 129.01, 128.55, 124.08, 121.83, 38.29, 32.32.

3m) 1-phenyl-3-(thiophen-2-yl)propan-1-one⁹



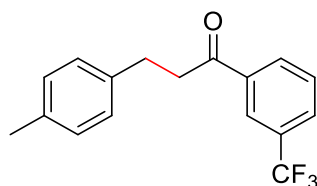
¹H NMR (500 MHz, Chloroform-*d*) δ 7.97 (d, *J* = 7.0 Hz, 2H), 7.57 (t, *J* = 7.3 Hz, 1H), 7.47 (t, *J* = 7.7 Hz, 2H), 7.13 (d, *J* = 3.9 Hz, 1H), 6.93 (t, 1H), 6.87 (d, *J* = 2.3 Hz, 1H), 3.37 (t, *J* = 6.6 Hz, 2H), 3.31 (t, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 199.07, 144.37, 137.23, 133.66, 129.12, 128.52, 127.34, 125.16, 123.86, 41.03, 24.70.

3p) 1,3-bis(4-methoxyphenyl)propan-1-one⁸



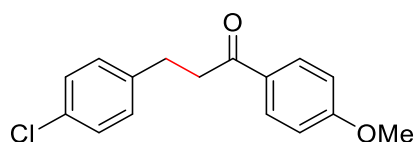
¹H NMR (500 MHz, Chloroform-*d*) δ 7.94 (d, *J* = 8.9 Hz, 2H), 7.17 (d, *J* = 8.6 Hz, 2H), 6.92 (d, *J* = 8.8 Hz, 2H), 6.85 (d, *J* = 8.6 Hz, 2H), 3.86 (s, 3H), 3.79 (s, 3H), 3.22 (t, *J* = 7.7 Hz, 2H), 3.00 (dd, *J* = 8.4, 7.0 Hz, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 198.40, 163.85, 158.39, 133.92, 130.73, 130.44, 129.77, 114.35, 114.14, 55.88, 55.69, 40.79, 29.91.

3q) 3-(p-tolyl)-1-(3-(trifluoromethyl)phenyl)propan-1-one



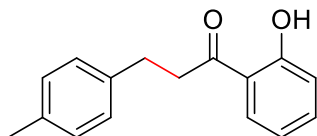
¹H NMR (500 MHz, Chloroform-*d*) δ 8.19 (s, 0H), 8.13 (d, *J* = 7.8 Hz, 0H), 7.81 (d, *J* = 7.8 Hz, 0H), 7.60 (t, *J* = 7.8 Hz, 1H), 7.14 (q, *J* = 8.0 Hz, 2H), 3.31 (t, *J* = 8.3, 6.9 Hz, 1H), 3.05 (t, *J* = 7.6 Hz, 1H), 2.33 (s, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 198.38, 138.26, 137.85, 136.32, 131.86, 131.62, 129.92, 129.77, 128.78, 125.39, 125.25, 41.24, 30.00, 21.47. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -62.80. HRMS: calculated: 292.1075; observed: 292.1298.

3s) 3-(4-chlorophenyl)-1-(4-methoxyphenyl)propan-1-one⁸



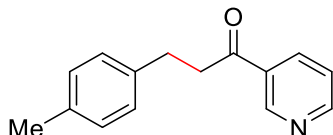
^1H NMR (500 MHz, Chloroform-*d*) δ 7.85 (d, $J = 8.9$ Hz, 1H), 7.17 (d, $J = 8.2$ Hz, 1H), 7.09 (d, $J = 8.4$ Hz, 1H), 6.84 (d, $J = 8.9$ Hz, 1H), 3.78 (s, 2H), 3.14 (t, $J = 7.6$ Hz, 1H), 2.94 (t, $J = 7.5$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 197.87, 163.95, 140.35, 132.21, 130.73, 130.28, 129.00, 114.19, 55.92, 40.23, 29.98.

3v) 2-(4-methylbenzyl)-3,4-dihydronaphthalen-1(2H)-one¹⁰



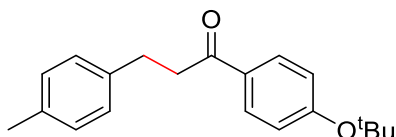
^1H NMR (500 MHz, Chloroform-*d*) δ 12.31 (s, 1H), 7.75 (d, $J = 8.1$ Hz, 1H), 7.46 (t, $J = 7.8$ Hz, 1H), 7.18 – 7.10 (m, 4H), 6.99 (d, $J = 8.4$ Hz, 1H), 6.91 – 6.85 (m, 1H), 3.31 (t, $J = 7.7$ Hz, 2H), 3.03 (t, $J = 7.7$ Hz, 2H), 2.33 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 205.91, 162.86, 138.00, 136.70, 136.24, 130.22, 129.67, 128.64, 119.68, 119.30, 118.95, 40.59, 30.02, 21.40.

3w) 1-(pyridin-3-yl)-3-(p-tolyl)propan-1-one



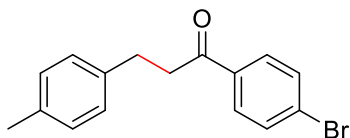
^1H NMR (500 MHz, Chloroform-*d*) δ 9.16 (s, 1H), 8.77 (d, $J = 3.1$ Hz, 1H), 8.23 (d, $J = 7.9$ Hz, 1H), 7.42 (dd, $J = 8.0, 4.8$ Hz, 1H), 7.13 (q, $J = 8.1$ Hz, 4H), 3.33 – 3.26 (m, 2H), 3.05 (t, $J = 7.6$ Hz, 2H), 2.32 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 198.28, 153.50, 149.66, 137.85, 136.08, 135.74, 132.37, 129.51, 128.50, 123.94, 41.09, 29.61, 21.23. HRMS: calculated = 225.1154. Observed m/z = 225.1627.

3x) 1-(4-(tert-butoxy)phenyl)-3-(p-tolyl)propan-1-one



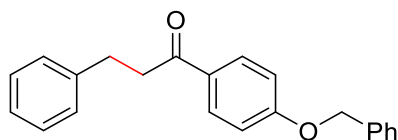
^1H NMR (500 MHz, Chloroform-*d*) δ 7.92 (d, $J = 8.4$ Hz, 2H), 7.48 (d, $J = 8.4$ Hz, 2H), 7.17 (d, $J = 7.9$ Hz, 2H), 7.12 (d, $J = 7.8$ Hz, 2H), 7.04 (s, 1H), 3.31 – 3.24 (m, 2H), 3.04 (t, $J = 7.8$ Hz, 2H), 2.34 (s, 3H), 1.35 (s, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 199.05, 156.77, 138.36, 135.60, 134.35, 129.23, 128.33, 128.06, 125.56, 40.57, 35.13, 31.13, 29.84, 21.05. HRMS: calculated = 296.1776. Observed m/z = 296.2017.

3y) 1-(4-bromophenyl)-3-(p-tolyl)propan-1-one¹⁰



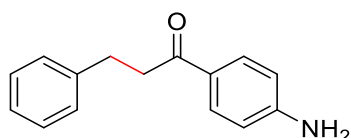
^1H NMR (500 MHz, Chloroform-*d*) δ 7.81 (d, $J = 8.5$ Hz, 2H), 7.59 (d, $J = 8.6$ Hz, 2H), 7.16 – 7.09 (m, 4H), 3.24 (t, $J = 8.4, 7.0$ Hz, 2H), 3.02 (t, $J = 7.7$ Hz, 2H), 2.33 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 198.50, 138.15, 135.96, 135.81, 132.13, 129.79, 129.46, 128.49, 128.41, 40.79, 29.84, 21.23.

3aa) 1-(4-benzylphenyl)-3-phenylpropan-1-one¹¹



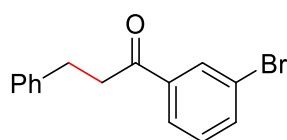
^1H NMR (500 MHz, Chloroform-*d*) δ 7.83 (d, J = 8.8 Hz, 1H), 7.32 (d, J = 1.8 Hz, 0H), 7.28 (d, J = 7.2 Hz, 1H), 7.24 (d, J = 7.2 Hz, 0H), 7.18 (d, J = 7.0 Hz, 1H), 7.14 (d, J = 7.3 Hz, 1H), 7.10 (td, J = 7.8, 7.3, 2.9 Hz, 1H), 7.03 (d, J = 7.0 Hz, 0H), 6.89 (d, J = 8.8 Hz, 1H), 5.00 (s, 1H), 3.13 (t, J = 7.7 Hz, 1H), 2.95 (t, J = 7.9 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 197.86, 162.64, 141.52, 136.23, 130.39, 129.08, 128.77, 128.58, 128.51, 128.45, 128.33, 127.56, 126.16, 114.64, 70.17, 40.19, 30.36.

3ab) 1-(4-aminophenyl)-3-phenylpropan-1-one⁸



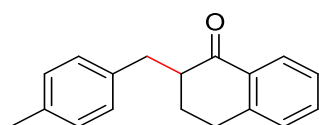
^1H NMR (500 MHz, Chloroform-*d*) δ 7.82 (d, J = 8.6 Hz, 2H), 7.30 (t, J = 7.5 Hz, 3H), 7.25 – 7.23 (m, 1H), 7.22 – 7.15 (m, 2H), 6.63 (d, J = 8.7 Hz, 2H), 3.20 (dd, J = 8.8, 6.7 Hz, 2H), 3.08 – 3.02 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 148.56, 147.66, 135.72, 134.26, 129.39, 128.50, 123.85, 71.50, 40.79, 31.63, 21.20.

3ac) 1-(3-bromophenyl)-3-phenylpropan-1-one¹²



^1H NMR (500 MHz, Chloroform-*d*) δ 7.99 (s, 1H), 7.78 (dt, J = 7.8, 1.3 Hz, 1H), 7.59 (ddd, J = 7.9, 2.0, 1.0 Hz, 1H), 7.25 – 7.23 (m, 2H), 7.21 (d, J = 7.4 Hz, 1H), 7.17 (d, J = 2.0 Hz, 2H), 7.15 (d, 1H), 7.12 (d, 1H), 3.19 (t, 2H), 2.98 (t, J = 7.6 Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 198.01, 141.19, 138.80, 136.14, 131.38, 130.44, 129.20, 128.64, 126.76, 126.48, 123.22, 40.76, 30.18.

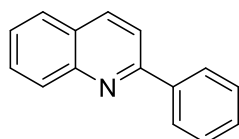
3ad) 8-hydroxy-2-(4-methylbenzyl)-3,4-dihydronaphthalen-1(2H)-one¹⁰



^1H NMR (500 MHz, Chloroform-*d*) δ 8.07 (dd, J = 7.9, 1.6 Hz, 1H), 7.46 (t, J = 7.4 Hz, 1H), 7.31 (t, J = 7.6 Hz, 1H), 7.22 (d, J = 7.8 Hz, 1H), 7.12 (s, 4H), 3.45 (dd, J = 13.8, 4.1 Hz, 1H), 3.00 – 2.87 (m, 3H), 2.73 (ddt, J = 11.6, 9.6, 4.3 Hz, 1H), 2.61 (dd, J = 13.8, 9.6 Hz, 1H), 2.33 (s, 1H), 2.12 (dq, J = 13.5, 4.5 Hz, 1H), 1.79 (dtd, J = 13.4, 11.6, 5.1 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 199.75, 144.28, 137.12, 135.85, 133.47, 132.73, 129.37, 129.32, 128.93, 127.77, 126.83, 49.74, 35.43, 28.82, 27.85, 21.26.

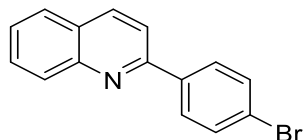
Friedländer Quinoline Synthesis

6a) 2-phenylquinoline¹³



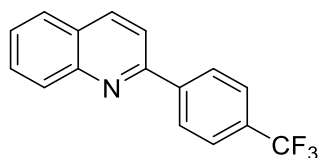
^1H NMR (500 MHz, Chloroform-*d*) δ 8.22 (t, $J = 9.2$ Hz, 2H), 8.18 (d, $J = 7.1$ Hz, 2H), 7.88 (d, $J = 8.5$ Hz, 1H), 7.83 (d, $J = 8.1, 1.4$ Hz, 1H), 7.74 (t, $J = 8.4, 6.9, 1.5$ Hz, 1H), 7.54 (t, 3H), 7.48 (t, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 157.38, 148.22, 139.61, 136.90, 129.75, 129.70, 129.40, 128.88, 127.64, 127.49, 127.22, 126.35, 119.06.

6b) 2-(4-bromophenyl)quinoline¹³



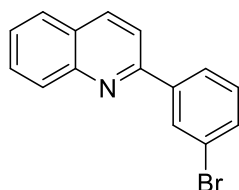
^1H NMR (500 MHz, Chloroform-*d*) δ 8.21 (d, $J = 8.6$ Hz, 1H), 8.17 (d, $J = 8.5$ Hz, 1H), 8.06 (d, $J = 8.5$ Hz, 2H), 7.82 (d, $J = 8.6$ Hz, 2H), 7.74 (t, $J = 7.7$ Hz, 1H), 7.65 (d, $J = 8.5$ Hz, 2H), 7.54 (t, $J = 7.5$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 156.48, 148.67, 138.92, 137.47, 132.44, 130.34, 130.15, 129.56, 127.96, 127.71, 127.00, 124.41, 118.96.

6c) 2-(4-(trifluoromethyl)phenyl)quinoline¹³



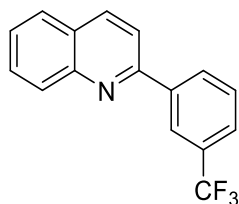
^1H NMR (500 MHz, Chloroform-*d*) δ 8.28 (d, $J = 8.1$ Hz, 2H), 8.24 (d, $J = 8.6$ Hz, 1H), 8.20 (d, $J = 8.6$ Hz, 1H), 7.89 – 7.82 (m, 2H), 7.80 – 7.73 (m, 3H), 7.56 (ddd, $J = 8.1, 6.8, 1.2$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 156.09, 148.71, 143.38, 137.59, 131.54, 130.46, 130.31, 128.30, 127.99, 127.90, 127.32, 126.20, 125.77, 119.21. ^{19}F NMR (376 MHz, Chloroform-*d*) δ -62.41.

6d) 2-(3-bromophenyl)quinoline¹³



^1H NMR (500 MHz, Chloroform-*d*) δ 8.37 (s, 1H), 8.23 (d, $J = 8.6$ Hz, 1H), 8.19 (d, $J = 8.5$ Hz, 1H), 8.08 (d, $J = 8.0$ Hz, 1H), 7.84 (d, $J = 8.6$ Hz, 2H), 7.77 – 7.74 (m, 1H), 7.60 (d, $J = 6.9$ Hz, 1H), 7.55 (d, $J = 7.3$ Hz, 1H), 7.40 (t, $J = 7.9$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 155.82, 148.38, 141.84, 135.28, 132.44, 130.84, 130.53, 130.12, 129.97, 127.70, 127.55, 126.88, 126.27, 123.36, 118.89.

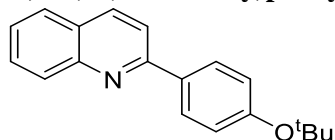
6e) 2-(3-(trifluoromethyl)phenyl)quinoline¹⁴



^1H NMR (500 MHz, Chloroform-*d*) δ 8.48 (s, 1H), 8.36 (d, $J = 7.8$ Hz, 1H), 8.26 (d, $J = 8.6$ Hz, 1H), 8.21 (d, $J = 8.5$ Hz, 1H), 7.89 (d, $J = 8.6$ Hz, 1H), 7.85 (d, $J = 8.2$ Hz, 1H), 7.81 – 7.70 (m, 3H), 7.65 (t, $J = 7.8$ Hz, 1H), 7.57 (t, $J = 7.5$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 155.78, 148.43, 140.54, 137.42, 131.68 –

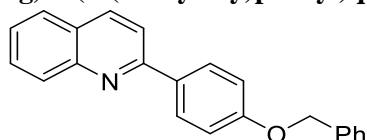
131.08 (m), 130.94, 130.23, 130.01, 129.53, 127.74, 127.02, 126.12, 124.64, 118.80. ^{19}F NMR (376 MHz, Chloroform-*d*) δ -62.52.

6f) 2-(4-(tert-butoxy)phenyl)quinoline



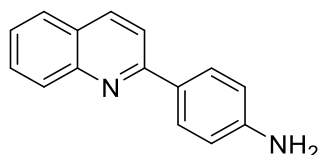
^1H NMR (500 MHz, Chloroform-*d*) δ 8.21 (d, J = 8.5 Hz, 2H), 8.11 (d, J = 8.5 Hz, 2H), 7.87 (d, J = 8.6 Hz, 1H), 7.82 (d, J = 8.0 Hz, 1H), 7.73 (ddd, J = 8.4, 6.8, 1.5 Hz, 1H), 7.56 (d, J = 8.4 Hz, 2H), 7.52 (t, J = 6.9 Hz, 1H), 1.39 (s, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 157.58, 152.86, 148.40, 129.87, 129.82, 127.67, 127.56, 127.32, 126.37, 126.07, 119.20, 34.98, 31.52. HRMS: calculated = 277.1467, observed = 277.1758, 278.1505.

6g) 2-(4-(benzyloxy)phenyl)quinoline¹⁵



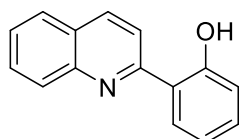
^1H NMR (500 MHz, Chloroform-*d*) δ 8.17 (dd, J = 11.2, 8.6 Hz, 4H), 7.82 (dd, J = 13.3, 8.4 Hz, 2H), 7.72 (t, J = 7.6 Hz, 1H), 7.50 (dd, J = 17.6, 7.1 Hz, 3H), 7.42 (t, J = 7.5 Hz, 2H), 7.35 (t, J = 7.3 Hz, 1H), 7.14 (d, J = 8.8 Hz, 2H), 5.16 (s, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (500 MHz, Chloroform-*d*) δ 160.24, 157.02, 136.99, 136.96, 129.86, 129.63, 129.16, 128.84, 128.25, 127.70, 127.64, 127.12, 126.19, 118.77, 115.40, 70.27, 28.20, 24.38.

6i) 4-(quinolin-2-yl)aniline¹⁶



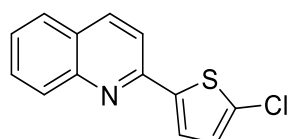
^1H NMR (500 MHz, Chloroform-*d*) δ 8.15 (d, J = 8.6 Hz, 2H), 8.04 (d, J = 8.4 Hz, 2H), 7.80 (dd, J = 12.2, 8.0 Hz, 2H), 7.69 (t, J = 7.7 Hz, 1H), 7.48 (t, J = 7.4 Hz, 1H), 6.81 (d, J = 8.5 Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 157.38, 148.16, 136.88, 129.79, 129.40, 129.13, 127.62, 127.00, 125.90, 118.62, 115.35.

6j) 2-(quinoline-2-yl)phenol¹⁷



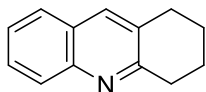
^1H NMR (500 MHz, Chloroform-*d*) δ 8.27 (d, J = 8.9 Hz, 1H), 8.04 (d, J = 9.0 Hz, 2H), 7.95 (dd, J = 8.1, 1.6 Hz, 1H), 7.83 (d, J = 1.4 Hz, 1H), 7.74 (ddd, J = 8.4, 6.9, 1.5 Hz, 1H), 7.55 (ddd, J = 8.1, 6.9, 1.1 Hz, 1H), 7.37 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.11 (dd, J = 8.2, 1.3 Hz, 1H), 6.97 (t, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 161.46, 158.38, 145.06, 138.21, 132.59, 131.02, 128.02, 127.93, 127.45, 127.20, 127.00, 119.39, 119.25, 119.16, 117.76.

6k) 2-(5-chlorothiophen-2-yl)quinoline¹⁸



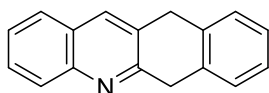
^1H NMR (500 MHz, Chloroform-*d*) δ 8.15 (d, J = 8.6 Hz, 2H), 7.77 (d, J = 8.1 Hz, 1H), 7.71 (d, J = 8.5 Hz, 2H), 7.50 (t, J = 7.7 Hz, 2H), 6.97 (d, J = 3.8 Hz, 1H).

6l) 1,2,3,4-tetrahydroacridine¹⁹



¹H NMR (500 MHz, Chloroform-*d*) δ 8.01 (d, *J* = 8.5 Hz, 1H), 7.83 (s, 1H), 7.73 – 7.68 (m, 1H), 7.61 (ddd, *J* = 8.4, 6.7, 1.5 Hz, 1H), 7.44 (ddd, *J* = 8.0, 6.7, 1.2 Hz, 1H), 3.15 (t, *J* = 6.5 Hz, 2H), 2.98 (t, *J* = 6.1 Hz, 2H), 2.03 – 1.97 (m, 2H), 1.93 – 1.87 (m, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 159.47, 146.47, 135.53, 131.25, 128.91, 128.22, 127.43, 127.12, 125.89, 33.58, 29.45, 23.37, 23.08.

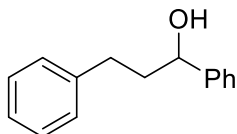
6m) 6,11-dihydrobenzo[*b*]acridine²⁰



¹H NMR (500 MHz, Chloroform-*d*) δ 8.60 (d, *J* = 7.8 Hz, 1H), 8.16 (d, *J* = 8.5 Hz, 1H), 7.92 (s, 1H), 7.75 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.66 (ddd, *J* = 8.4, 6.9, 1.5 Hz, 1H), 7.51 – 7.42 (m, 2H), 7.38 (td, *J* = 7.4, 1.5 Hz, 1H), 7.29 (d, *J* = 1.4 Hz, 1H), 3.15 – 3.11 (m, 2H), 3.02 (dd, *J* = 8.4, 5.5 Hz, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 153.57, 147.76, 139.64, 134.85, 133.98, 130.80, 129.92, 129.57, 128.89, 128.16, 128.07, 127.55, 127.13, 126.29, 29.03, 28.61.

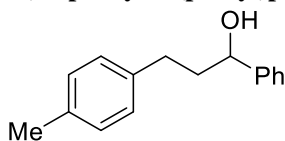
NMR data for β-alkylation of secondary alcohols:

8a) 1,3-diphenylpropan-1-ol²¹



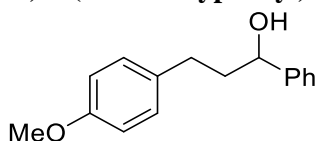
¹H NMR (500 MHz, Chloroform-*d*) δ 7.27 (d, *J* = 4.4 Hz, 4H), 7.19 (t, 3H), 7.11 (d, *J* = 7.6 Hz, 3H), 4.60 (dd, *J* = 7.8, 5.3 Hz, 1H), 2.75 – 2.46 (m, 2H), 2.10 – 1.92 (m, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 144.59, 141.80, 128.55, 128.47, 128.42, 127.68, 125.96, 125.89, 73.92, 40.49, 32.09.

8b) 1-phenyl-3-(*p*-tolyl)propan-1-ol²¹



¹H NMR (500 MHz, Chloroform-*d*) 7.27 (s, 4H), 7.21 (q, 1H), 7.02 (s, 4H), 4.60 (dd, *J* = 7.8, 5.4 Hz, 1H), 2.60 (dddd, *J* = 39.3, 14.1, 9.5, 6.1 Hz, 2H), 2.25 (s, 3H), 1.98 (dddd, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 145.07, 139.11, 135.73, 129.53, 128.95, 128.77, 128.05, 126.40, 74.33, 41.01, 32.04, 21.45.

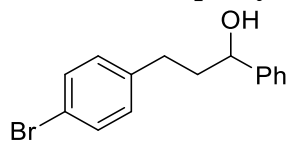
8c) 3-(4-methoxyphenyl)-1-phenylpropan-1-ol²²



¹H NMR (500 MHz, Chloroform-*d*) δ 7.25 (s, 4H), 7.19 (q, 1H), 7.02 (d, *J* = 8.7 Hz, 2H), 6.74 (d, *J* = 8.8 Hz, 2H), 4.57 (dd, *J* = 7.9, 5.3 Hz, 1H), 3.69 (s, 3H), 2.68 – 2.42 (m, 2H), 2.07 – 1.86 (m, 2H).

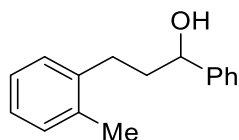
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 157.81, 144.68, 133.86, 129.36, 128.53, 127.63, 125.98, 113.86, 73.86, 55.29, 40.73, 31.16.

8d) 3-(4-bromophenyl)-1-phenylpropan-1-ol²²



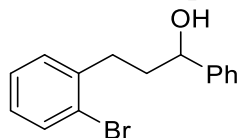
^1H NMR (500 MHz, Chloroform-*d*) δ 7.33 (d, $J = 8.1$ Hz, 2H), 7.28 (d, $J = 15.0$ Hz, 1H), 7.25 – 7.21 (m, 1H), 7.00 (d, $J = 8.0$ Hz, 1H), 4.60 (dd, $J = 7.9, 5.3$ Hz, 1H), 2.60 (dddd, $J = 30.1, 13.7, 9.5, 6.0$ Hz, 2H), 2.09 – 1.81 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 144.41, 140.77, 131.44, 130.24, 128.61, 127.78, 125.90, 119.60, 73.72, 40.26, 31.46.

8e) 1-phenyl-3-(o-tolyl)propan-1-ol²²



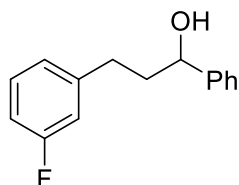
^1H NMR (500 MHz, Chloroform-*d*) δ 7.43 – 7.35 (m, 4H), 7.33 (q, 1H), 7.19 – 7.10 (m, 4H), 4.76 (dd, $J = 7.8, 5.3$ Hz, 1H), 2.73 (dddd, $J = 67.8, 14.1, 10.3, 5.7$ Hz, 2H), 2.30 (s, 3H), 2.16 – 1.94 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 144.60, 140.03, 136.00, 130.24, 128.77, 128.57, 127.70, 126.03, 126.00, 125.95, 74.25, 39.25, 29.46, 19.26.

8f) 3-(2-bromophenyl)-1-phenylpropan-1-ol²³

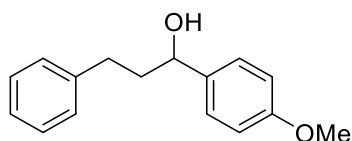


^1H NMR (500 MHz, Chloroform-*d*) δ 7.55 (d, $J = 8.0$ Hz, 1H), 7.39 (q, $J = 7.0$ Hz, 4H), 7.34 – 7.29 (m, 1H), 7.25 (d, $J = 3.4$ Hz, 1H), 7.08 (ddd, $J = 8.8, 5.7, 3.5$ Hz, 1H), 4.75 (dd, $J = 7.9, 5.4$ Hz, 1H), 2.87 (dddd, $J = 62.9, 13.7, 9.8, 6.0$ Hz, 2H), 2.15 – 2.01 (m, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, Chloroform-*d*) δ 144.40, 141.18, 132.86, 130.44, 128.56, 127.71, 127.67, 127.49, 125.95, 124.48, 73.98, 38.92, 32.62.

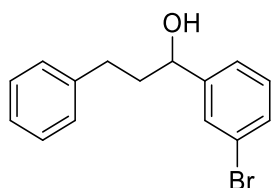
8h) 3-(3-fluorophenyl)-1-phenylpropan-1-ol²¹



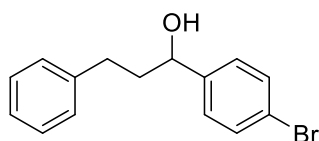
^1H NMR (500 MHz, Chloroform-*d*) δ 7.28 (d, $J = 3.2$ Hz, 3H), 7.22 (dq, $J = 5.8, 3.1$ Hz, 1H), 7.15 (q, $J = 8.0, 6.2$ Hz, 2H), 6.89 (d, $J = 7.7$ Hz, 1H), 6.81 (ddt, $J = 13.3, 8.3, 4.0$ Hz, 2H), 4.61 (dd, $J = 7.9, 5.3$ Hz, 1H), 2.71 – 2.56 (m, 2H), 2.11 – 1.89 (m, 2H). ^{19}F NMR (376 MHz, Chloroform-*d*) δ -113.69.

8k) 1-(4-methoxyphenyl)-3-phenylpropan-1-ol²¹

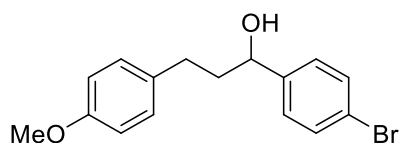
¹H NMR (500 MHz, Chloroform-*d*) δ 7.21 (q, 4H), 7.13 (dd, 3H), 6.83 (d, *J* = 8.7 Hz, 1H), 4.57 (dd, *J* = 7.8, 5.7 Hz, 0H), 3.74 (s, 1H), 2.72 – 2.53 (m, 1H), 2.13 – 1.92 (m, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 159.14, 141.88, 136.74, 128.47, 128.41, 127.25, 125.86, 113.92, 73.52, 55.32, 40.37, 32.16.

8l) 1-(3-bromophenyl)-3-phenylpropan-1-ol²¹

¹H NMR (500 MHz, Chloroform-*d*) δ 7.44 (d, *J* = 1.9 Hz, 1H), 7.34 (dd, *J* = 7.8, 1.7 Hz, 1H), 7.21 (ddd, *J* = 13.1, 8.0, 4.1 Hz, 3H), 7.17 – 7.10 (m, 4H), 4.58 (ddd, *J* = 7.7, 5.1, 2.2 Hz, 1H), 2.74 – 2.55 (m, 2H), 2.08 – 1.88 (m, 2H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 146.99, 141.46, 130.67, 130.12, 129.07, 128.50, 128.46, 126.02, 124.55, 122.67, 73.16, 40.52, 31.94.

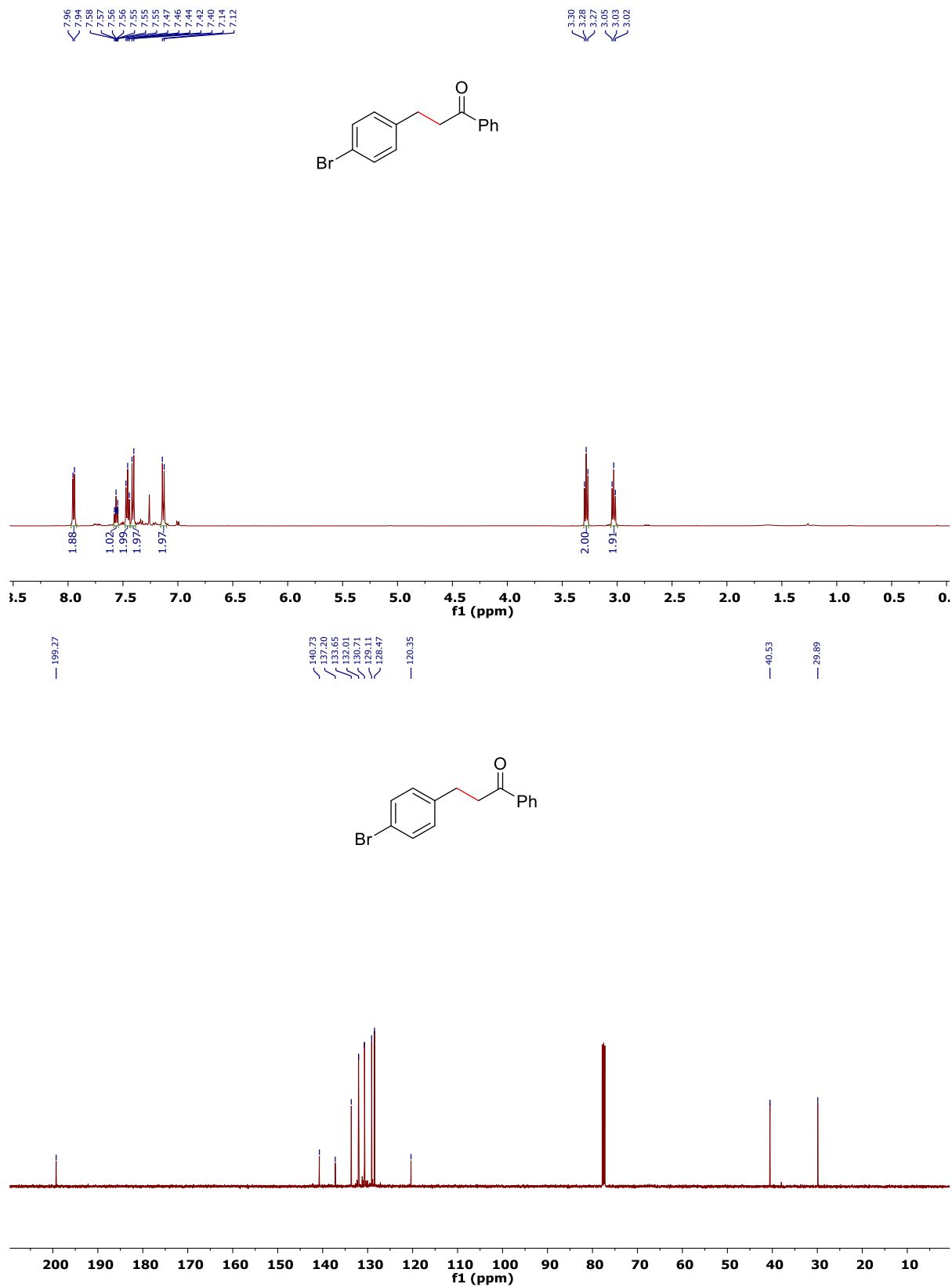
8m) 1-(4-bromophenyl)-3-phenylpropan-1-ol²¹

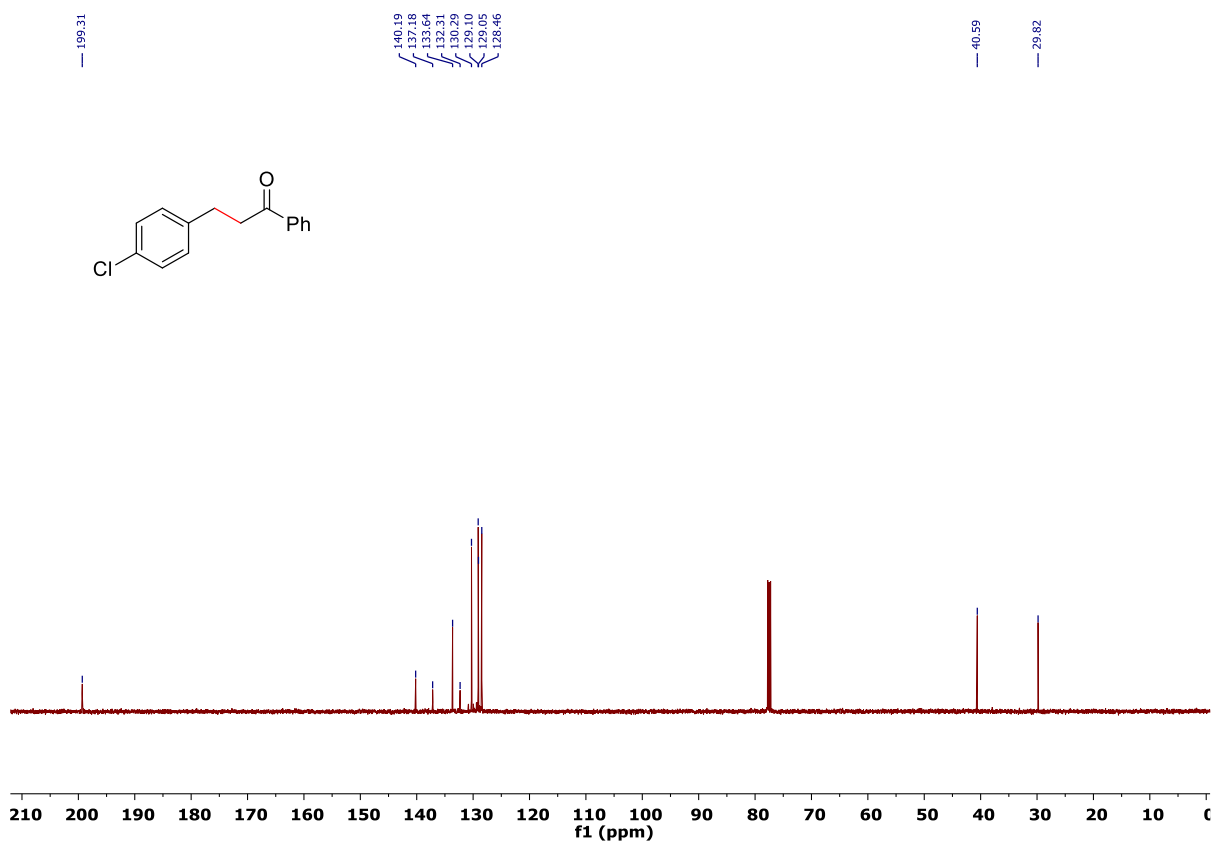
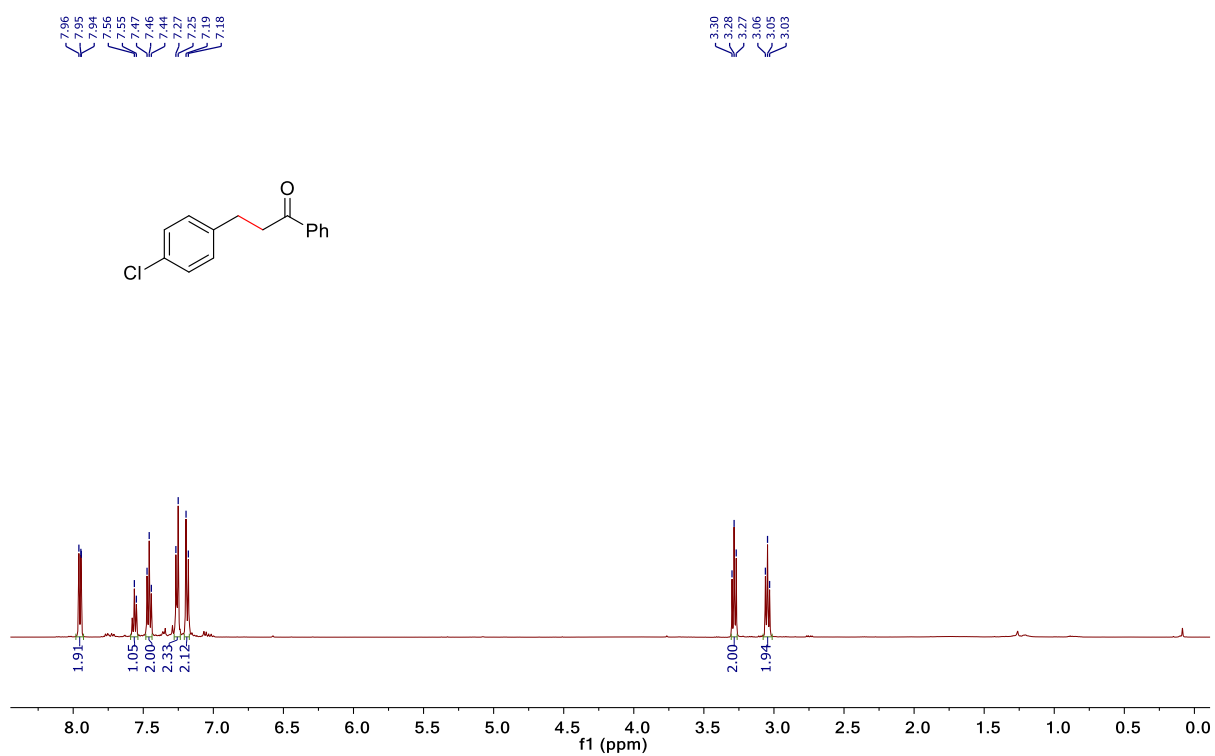
¹H NMR (500 MHz, Chloroform-*d*) δ 7.50 (d, *J* = 8.2 Hz, 1H), 7.38 (d, *J* = 4.3 Hz, 0H), 7.31 (q, *J* = 7.2, 6.6 Hz, 1H), 7.25 – 7.19 (m, 2H), 4.68 (dd, *J* = 7.9, 5.2 Hz, 0H), 2.72 (dddd, *J* = 29.8, 13.9, 9.4, 6.2 Hz, 1H), 2.18 – 1.96 (m, 1H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 143.57, 141.49, 131.61, 128.48, 128.44, 127.68, 126.00, 121.38, 73.20, 40.48, 31.92.

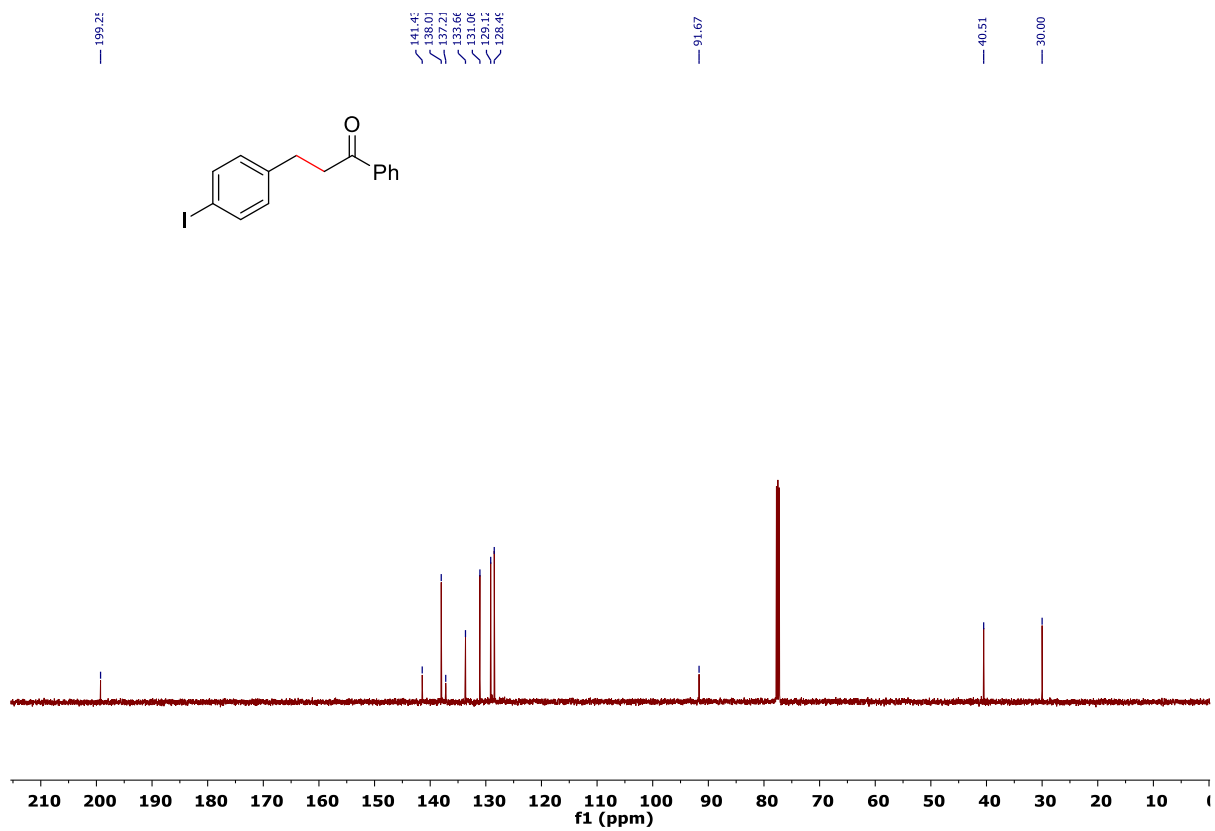
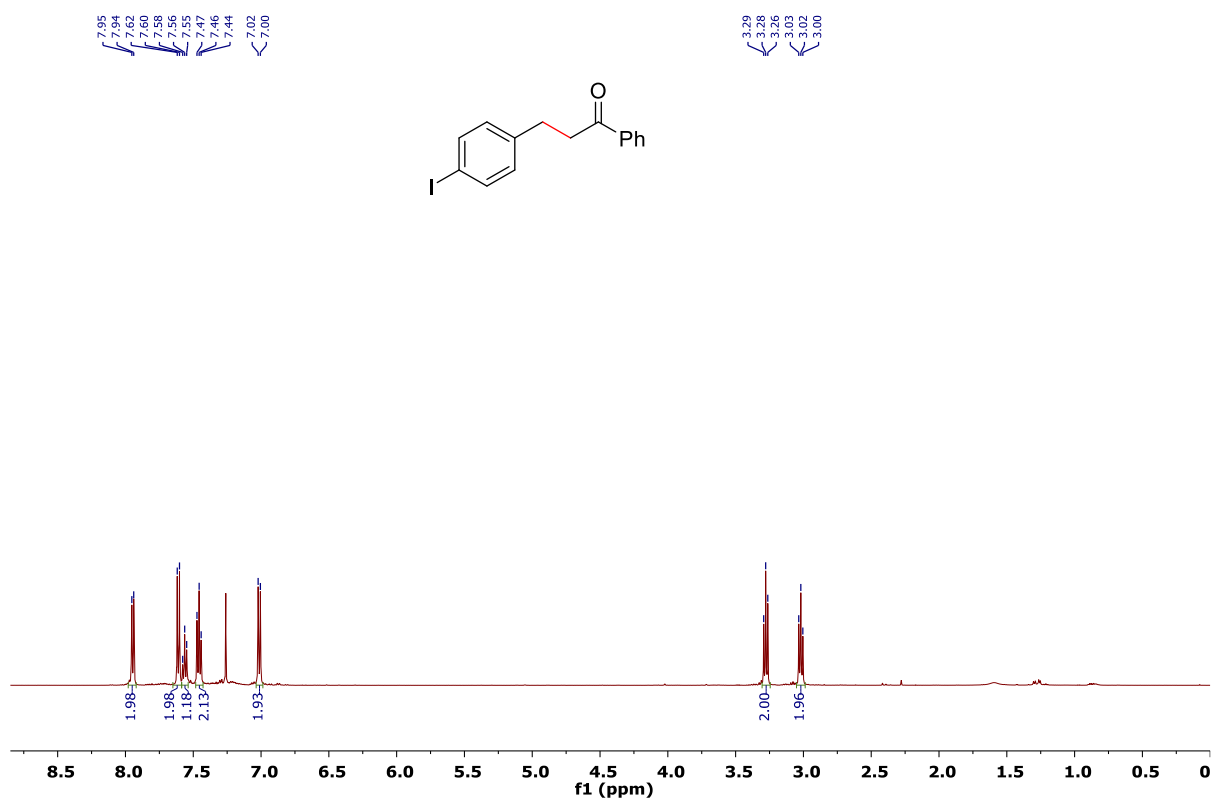
8n) 1-(4-bromophenyl)-3-(4-methoxyphenyl)propan-1-ol²⁴

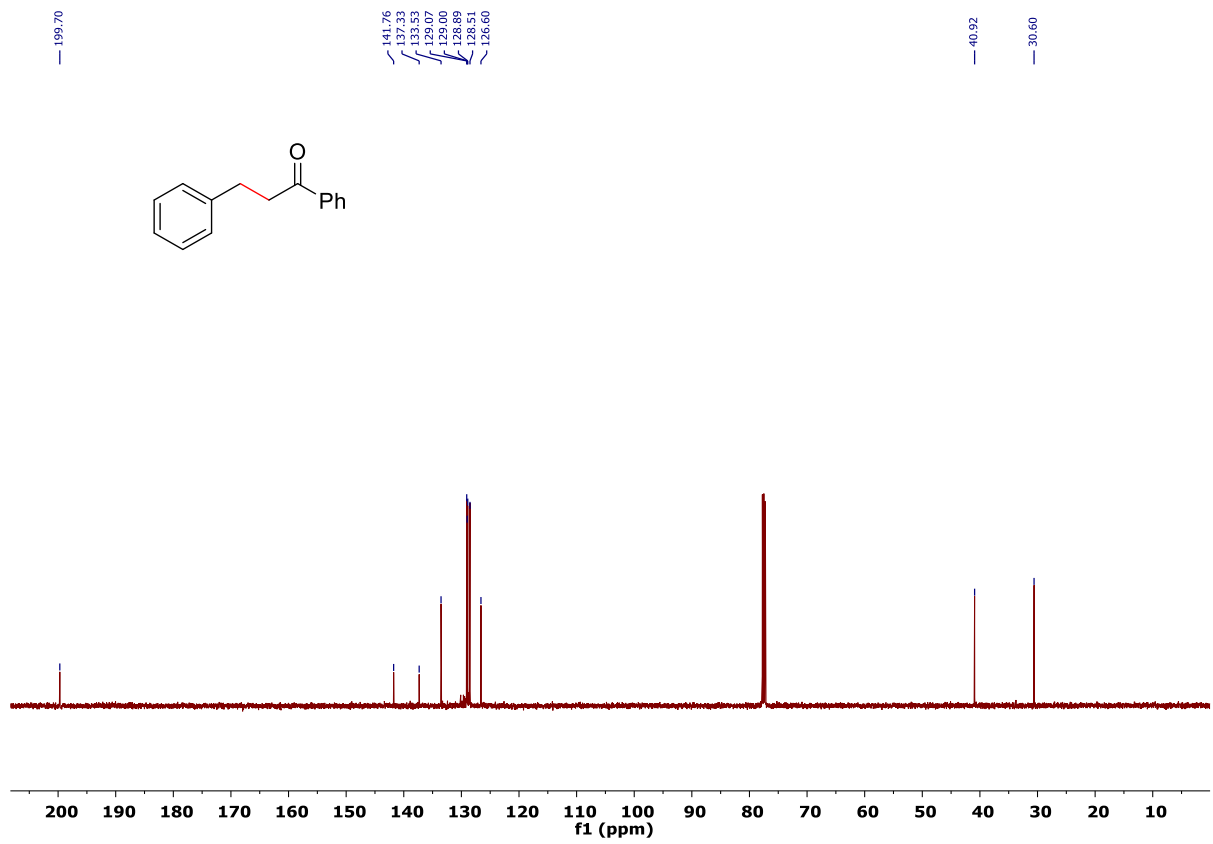
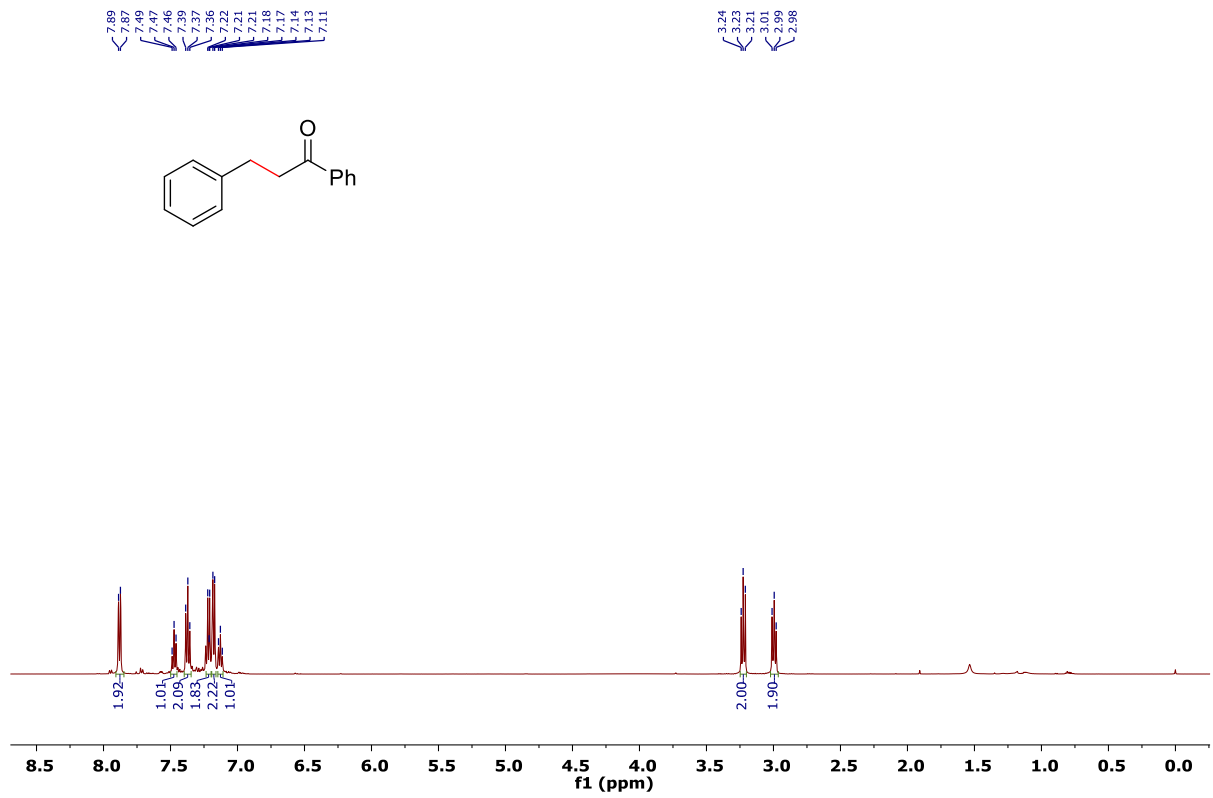
¹H NMR (500 MHz, Chloroform-*d*) δ 7.49 (d, *J* = 8.3 Hz, 0H), 7.23 (d, *J* = 6.7 Hz, 1H), 7.11 (d, *J* = 8.7 Hz, 1H), 6.85 (d, *J* = 8.9 Hz, 1H), 4.65 (ddd, *J* = 7.8, 5.2, 2.1 Hz, 0H), 3.81 (s, 1H), 2.66 (qdd, *J* = 14.3, 9.0, 6.2 Hz, 1H), 2.14 – 1.89 (m, 1H). ¹³C{¹H} NMR (126 MHz, Chloroform-*d*) δ 157.88, 143.64, 133.50, 131.59, 129.33, 127.69, 121.32, 113.90, 73.16, 55.29, 40.71, 31.00.

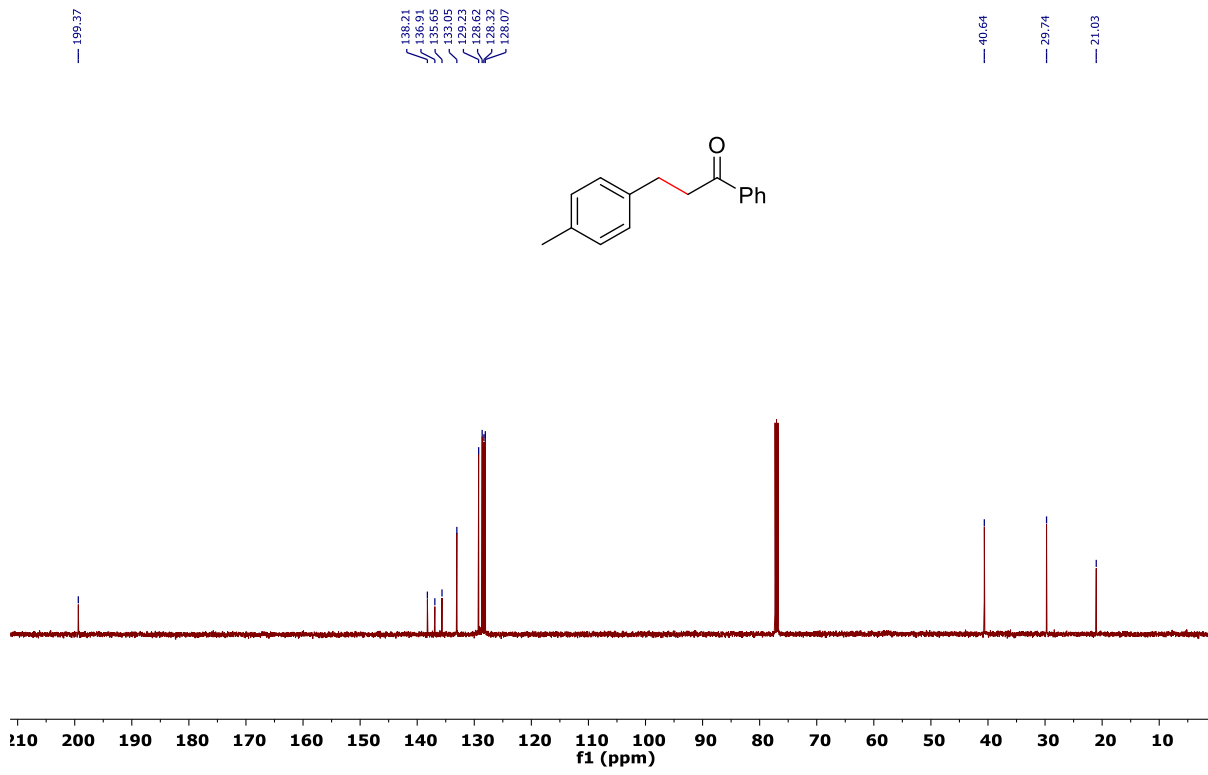
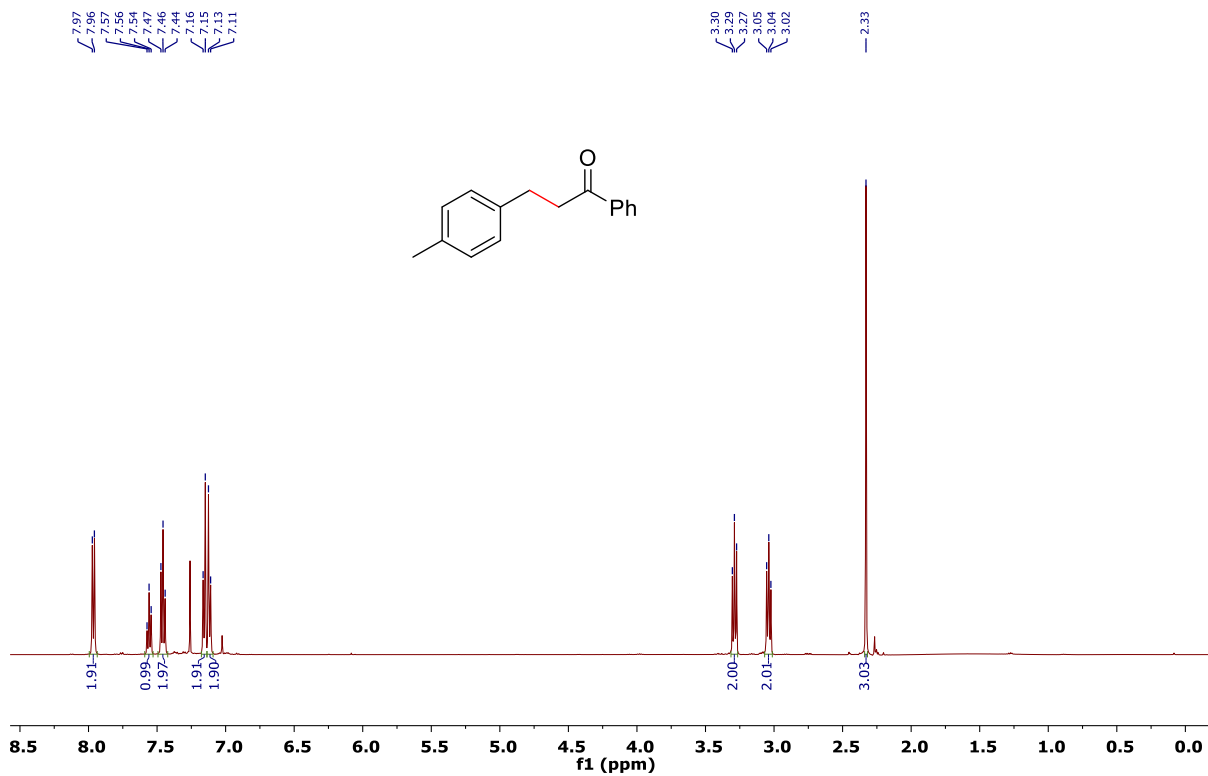
8. ^1H (500 MHz, CDCl_3) and ^{13}C NMR (126 MHz, CDCl_3) spectra for the isolated compounds:

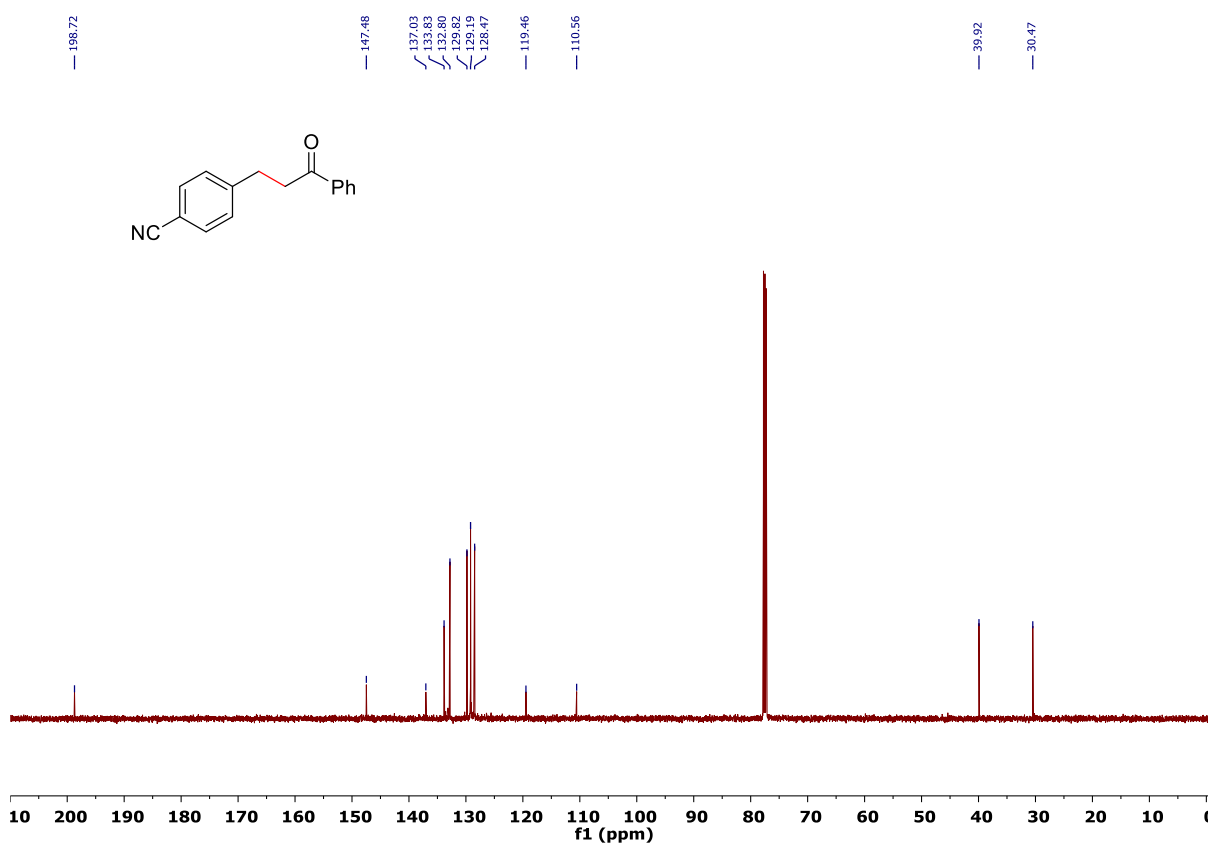
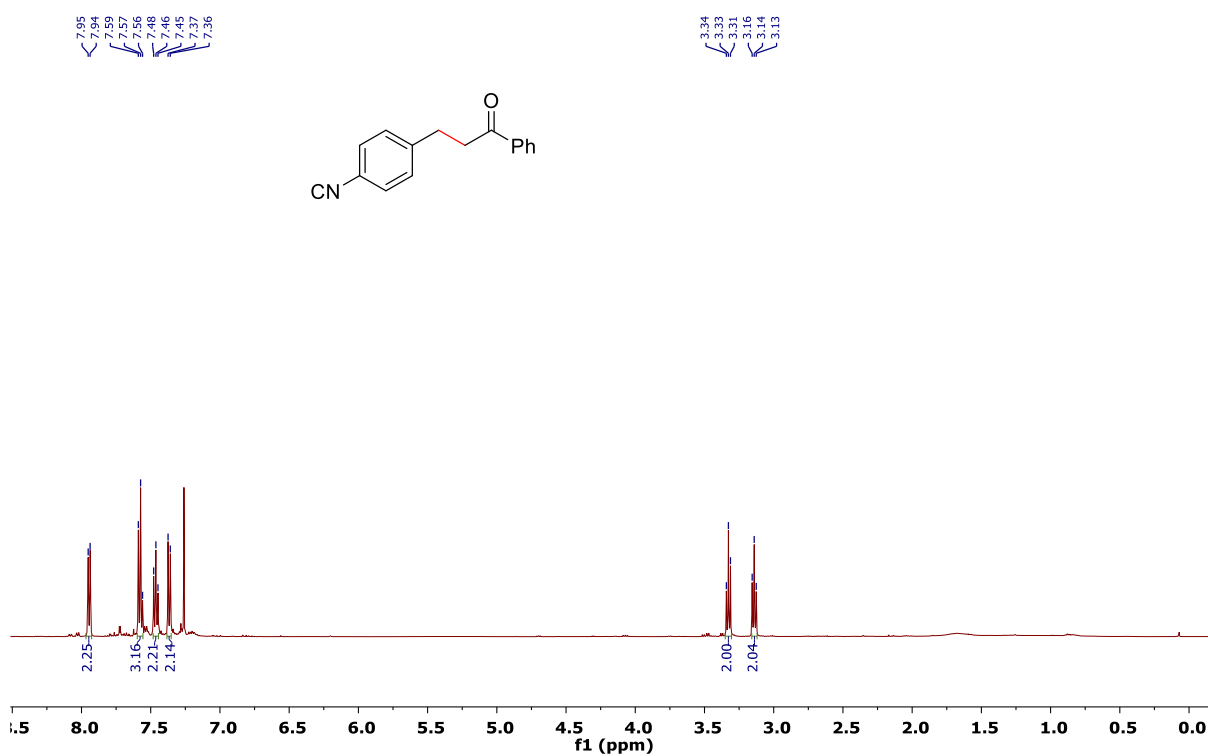


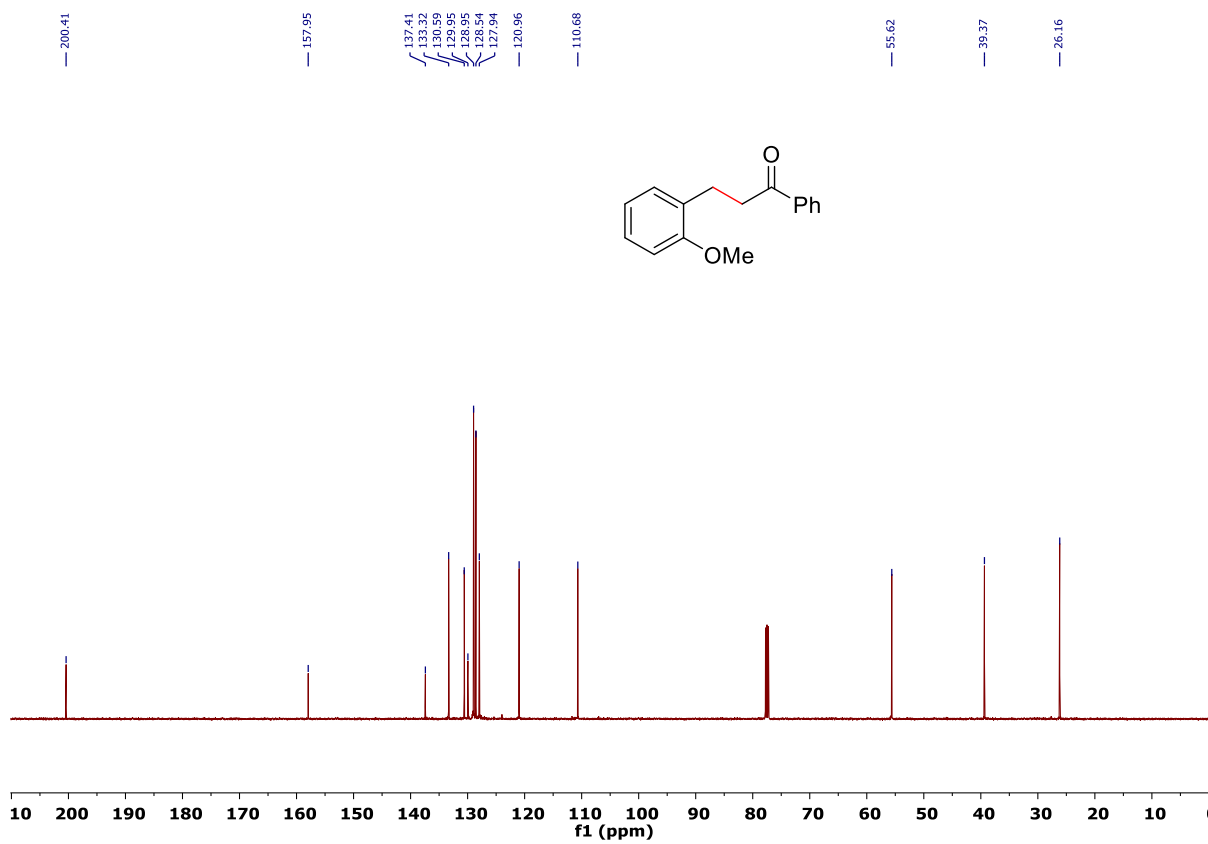
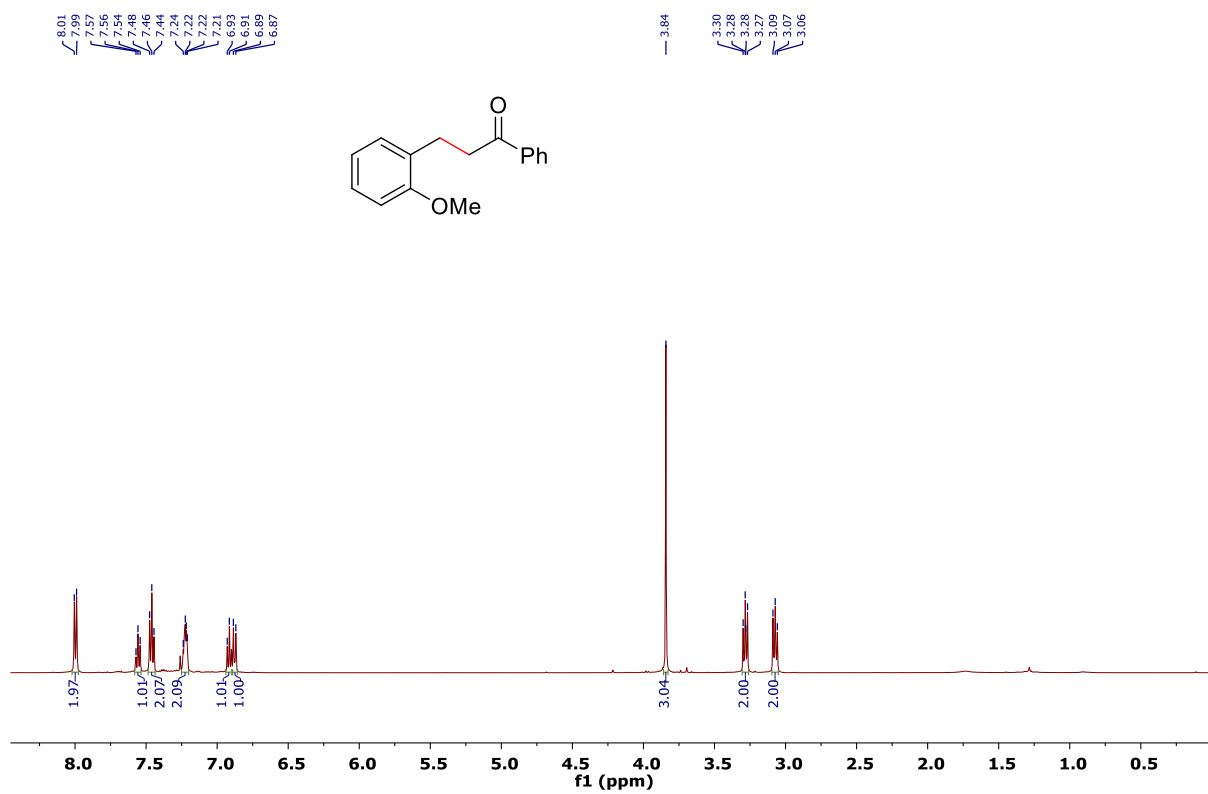


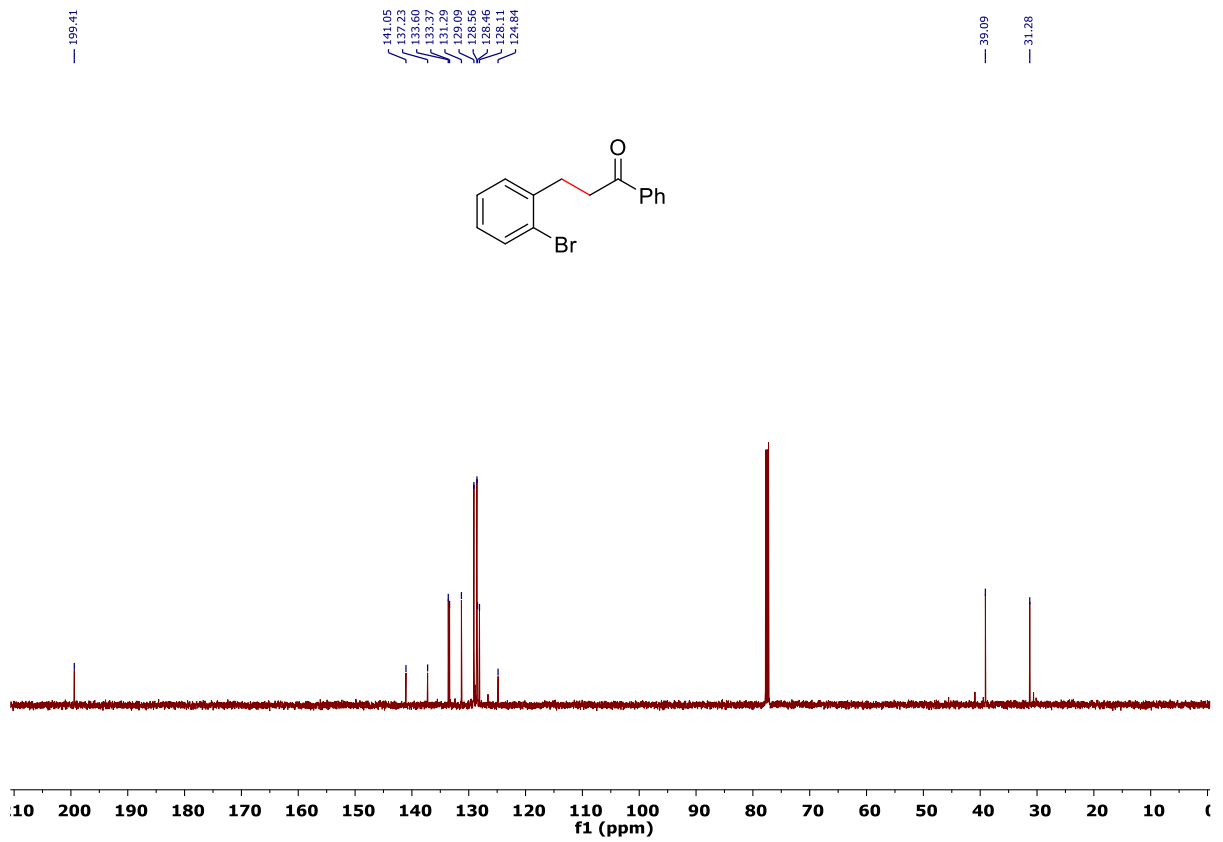
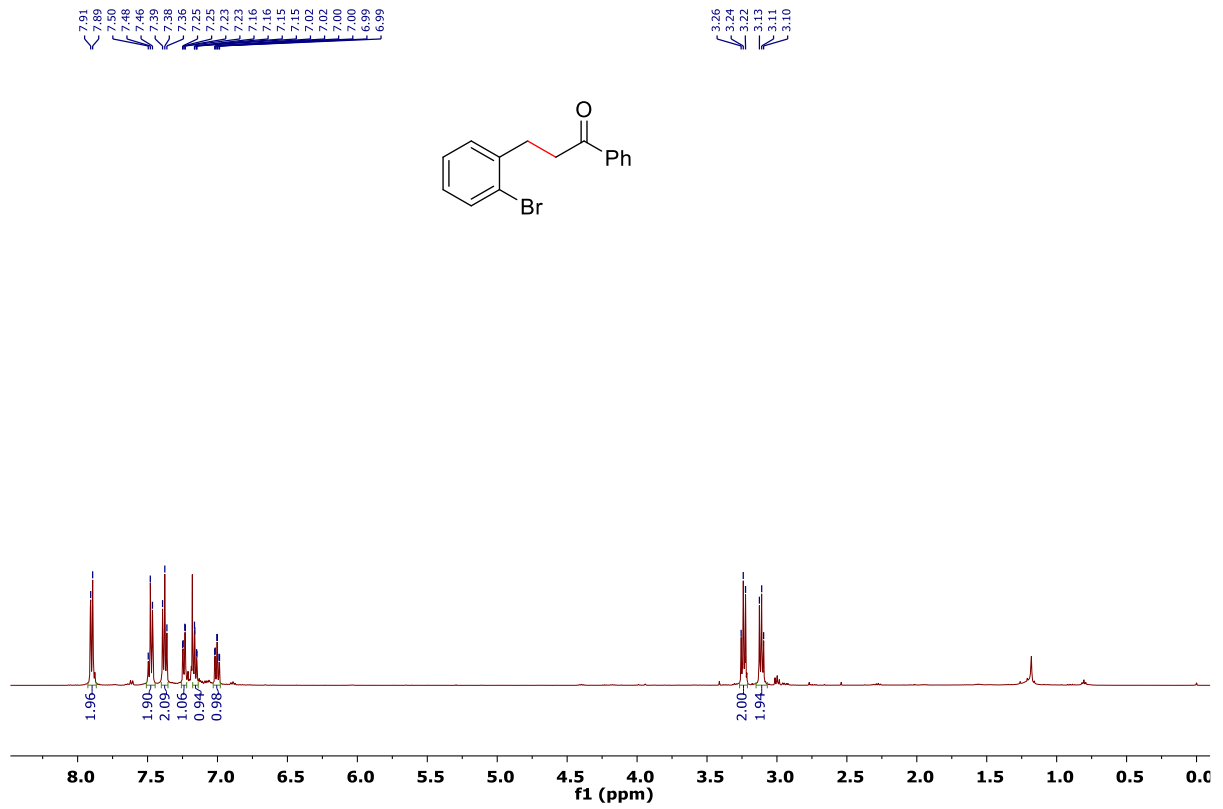


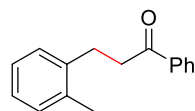
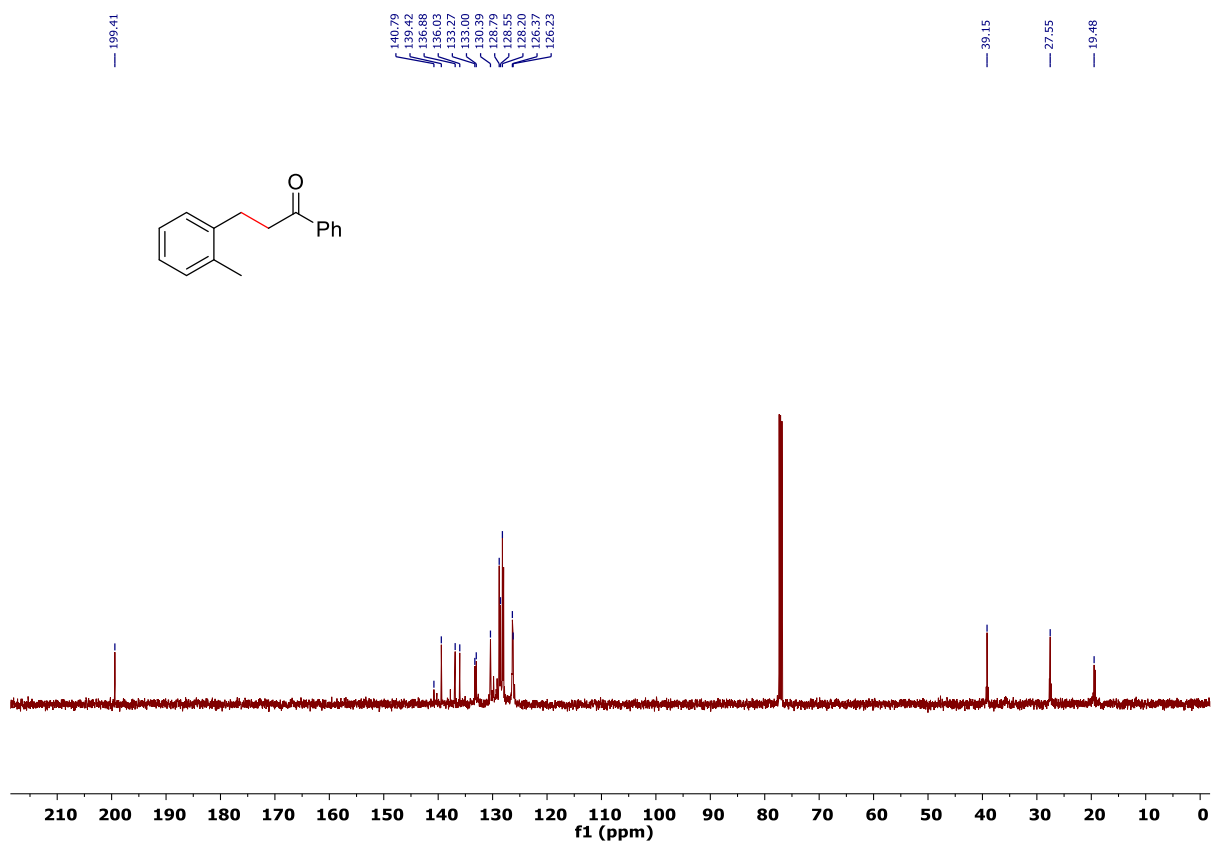
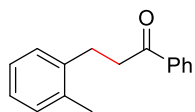
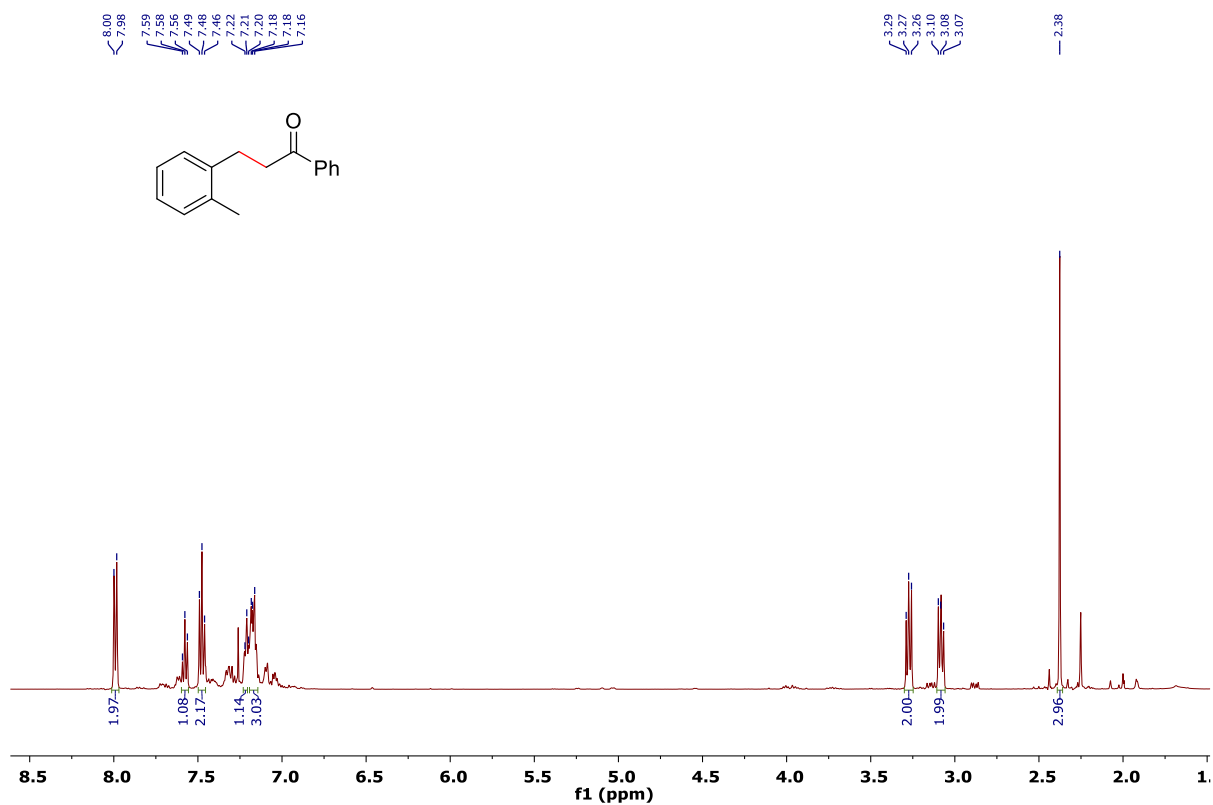


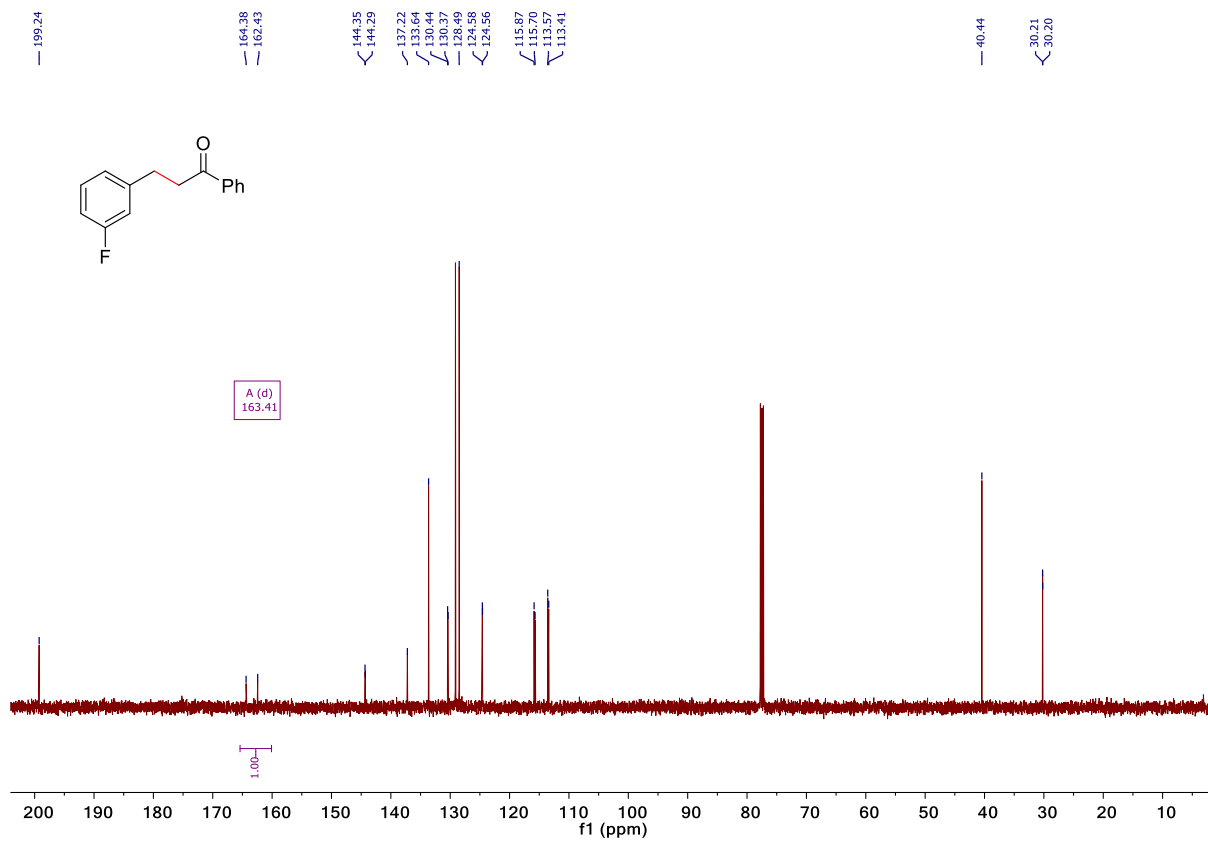
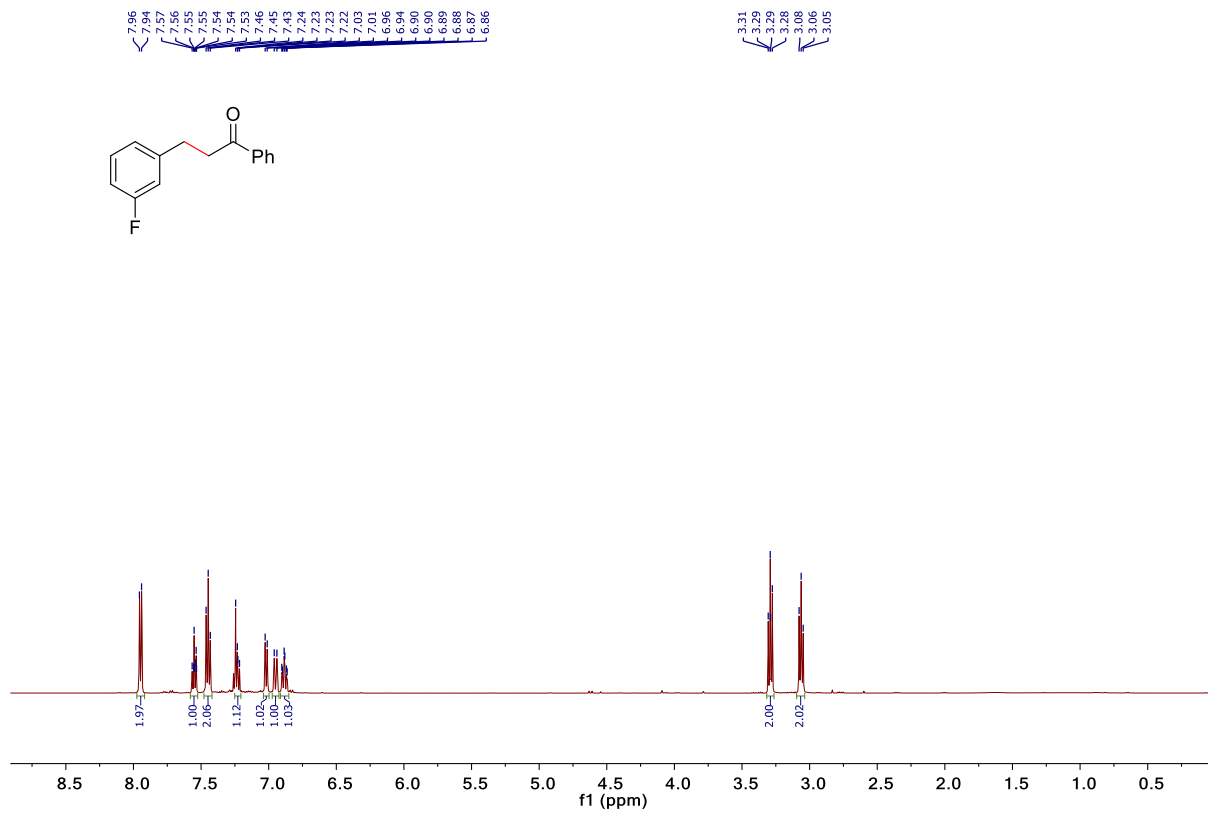




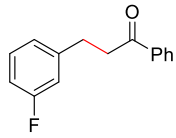




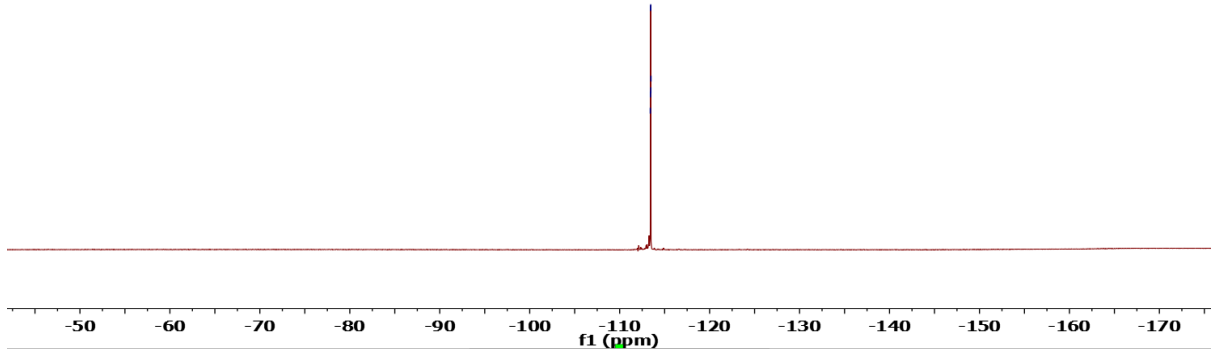




¹⁹F NMR



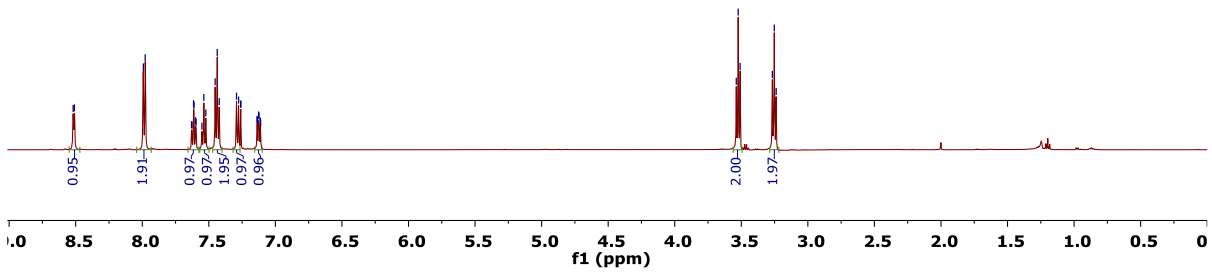
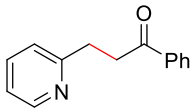
-113.41
-113.43
-113.44
-113.45
-113.46

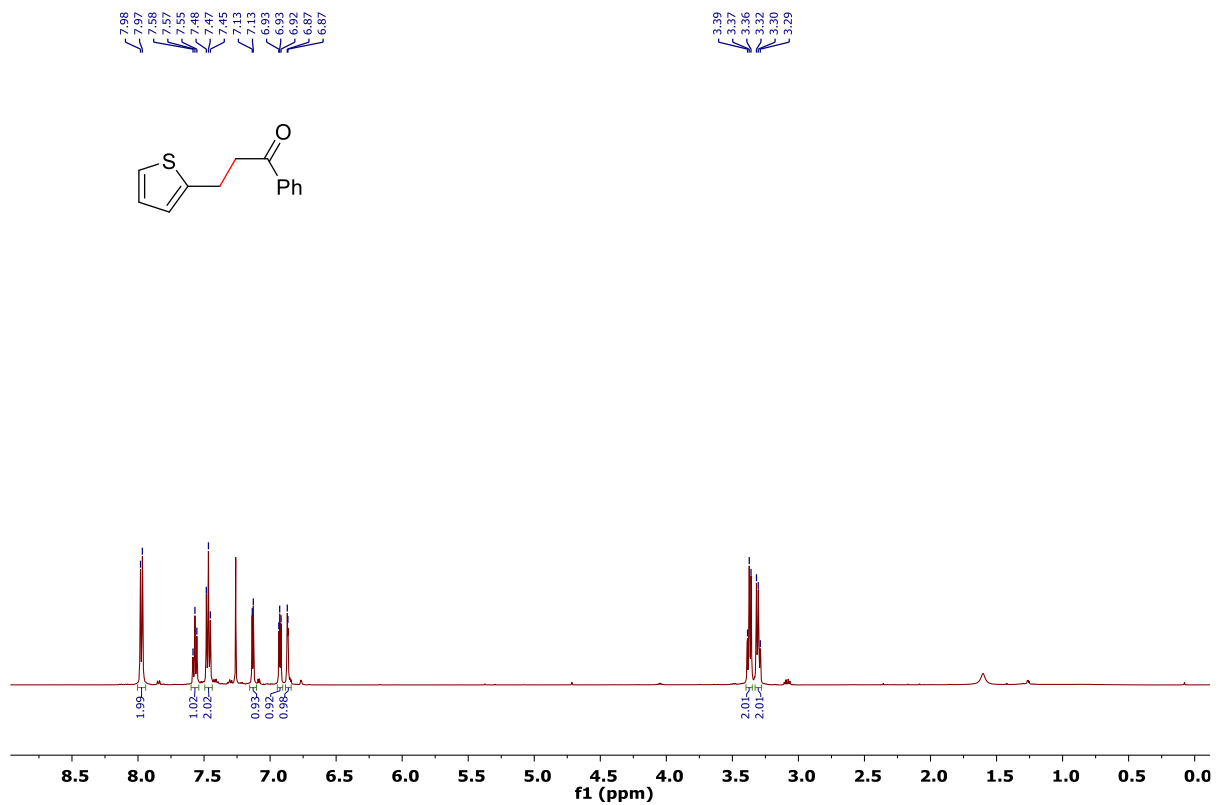
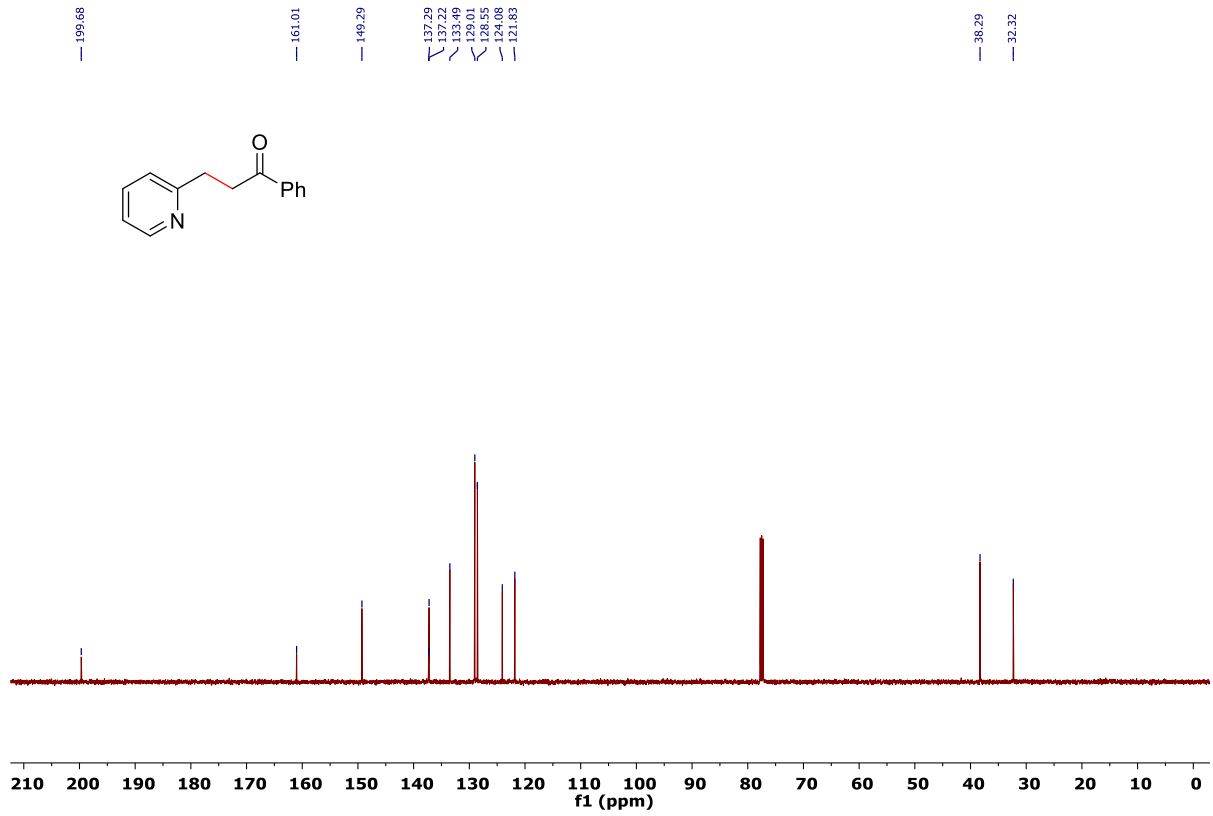


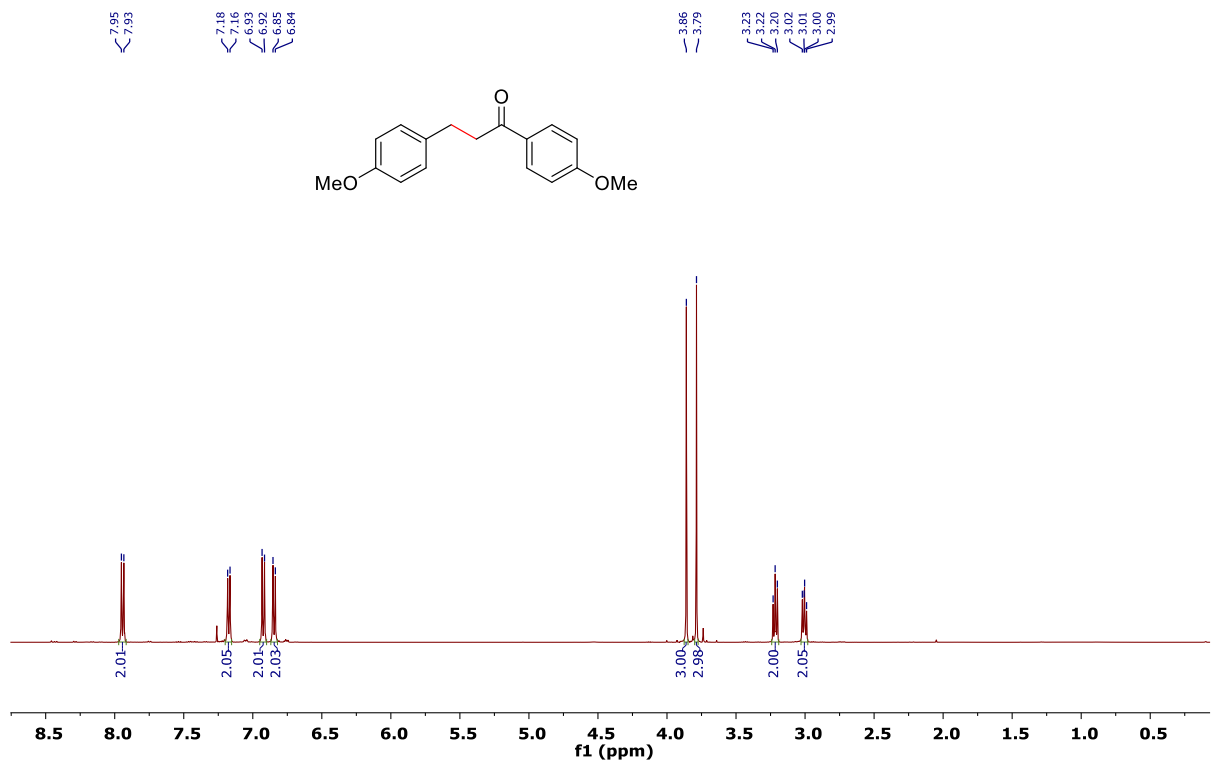
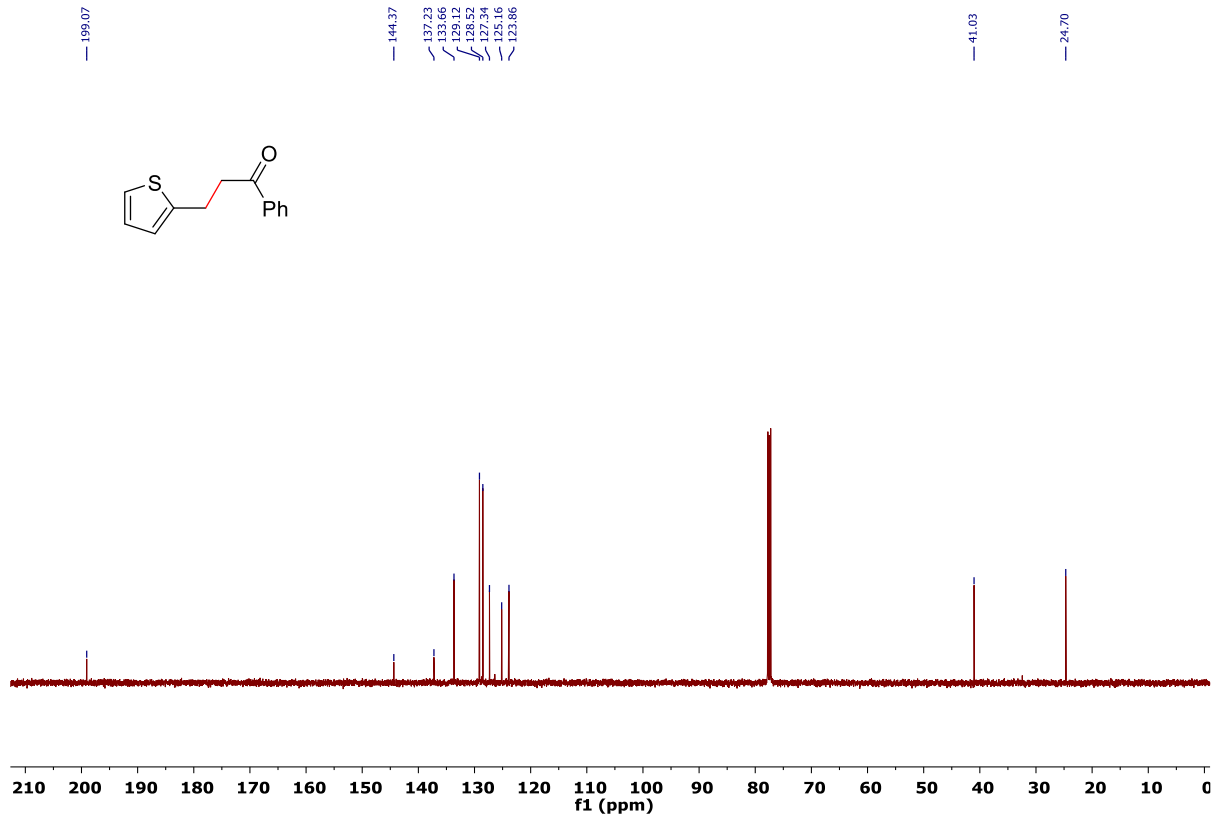
8.52
8.51

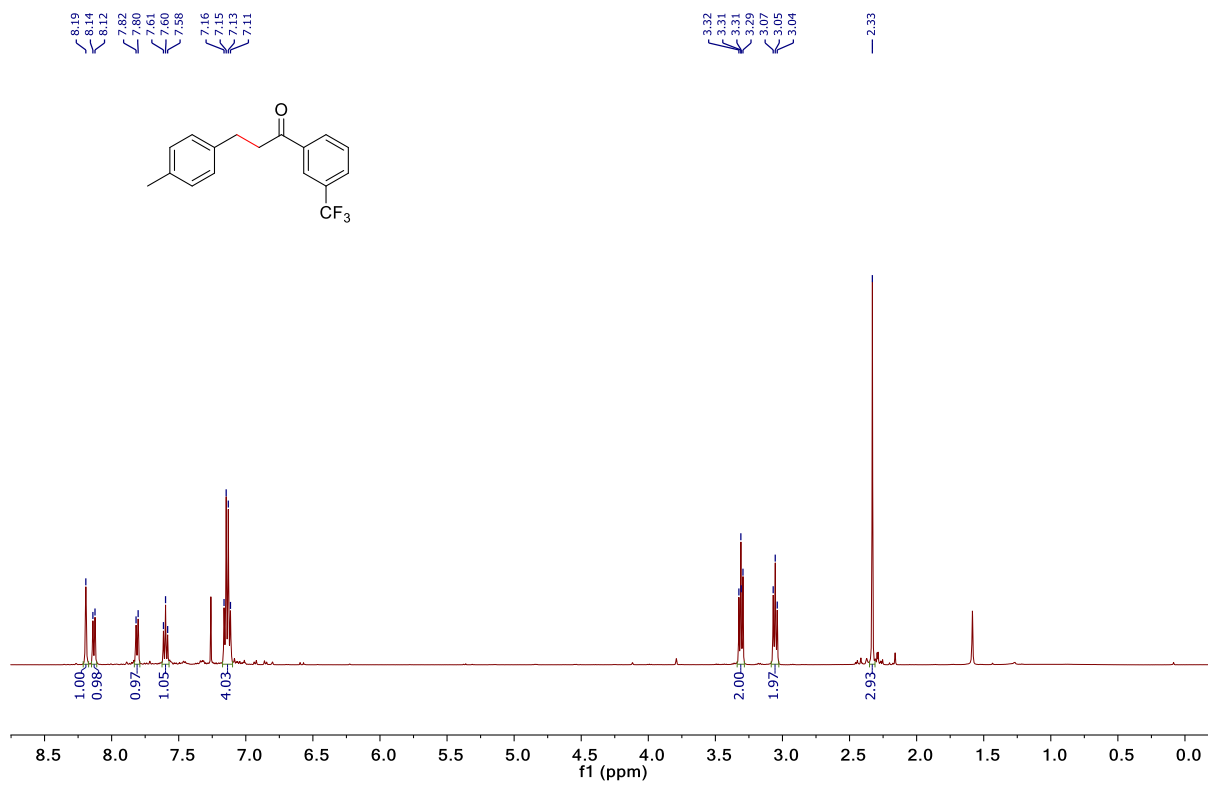
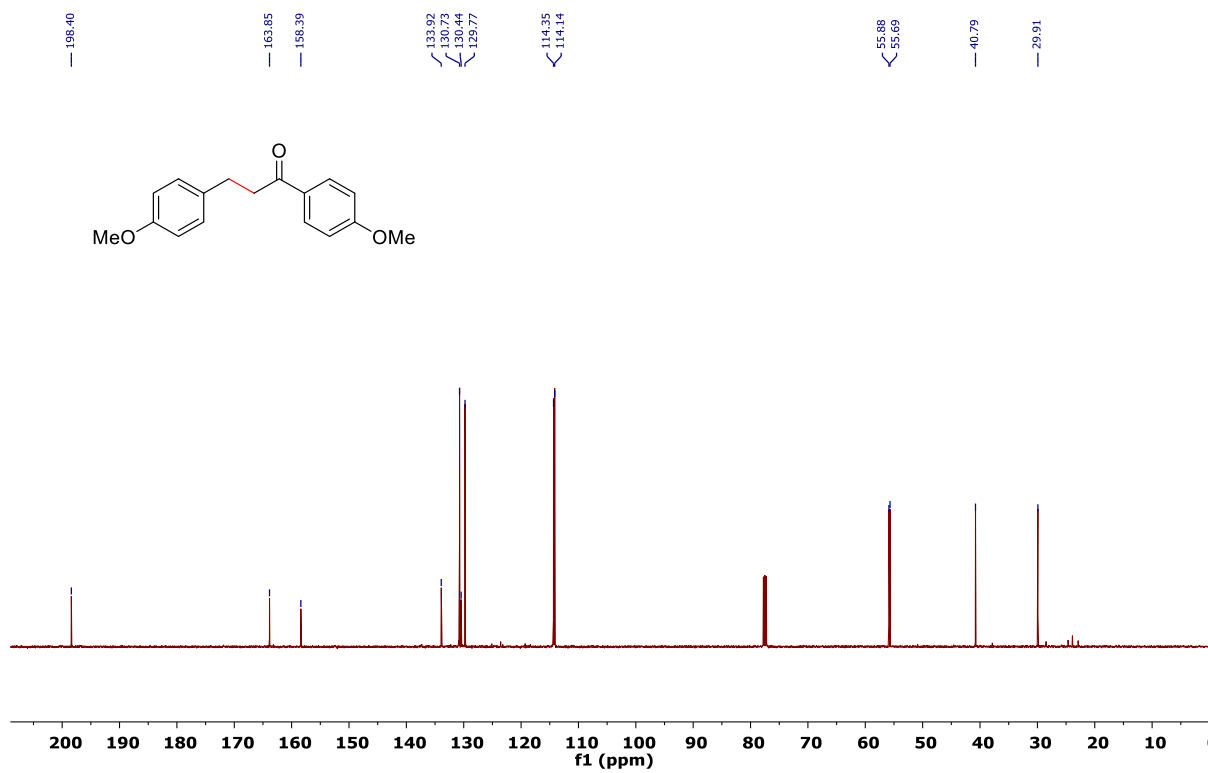
7.99
7.88
7.85
7.63
7.61
7.61
7.60
7.60
7.55
7.54
7.52
7.44
7.44
7.42
7.29
7.28
7.26
7.26
7.14
7.14
7.12
7.12
7.11
7.11

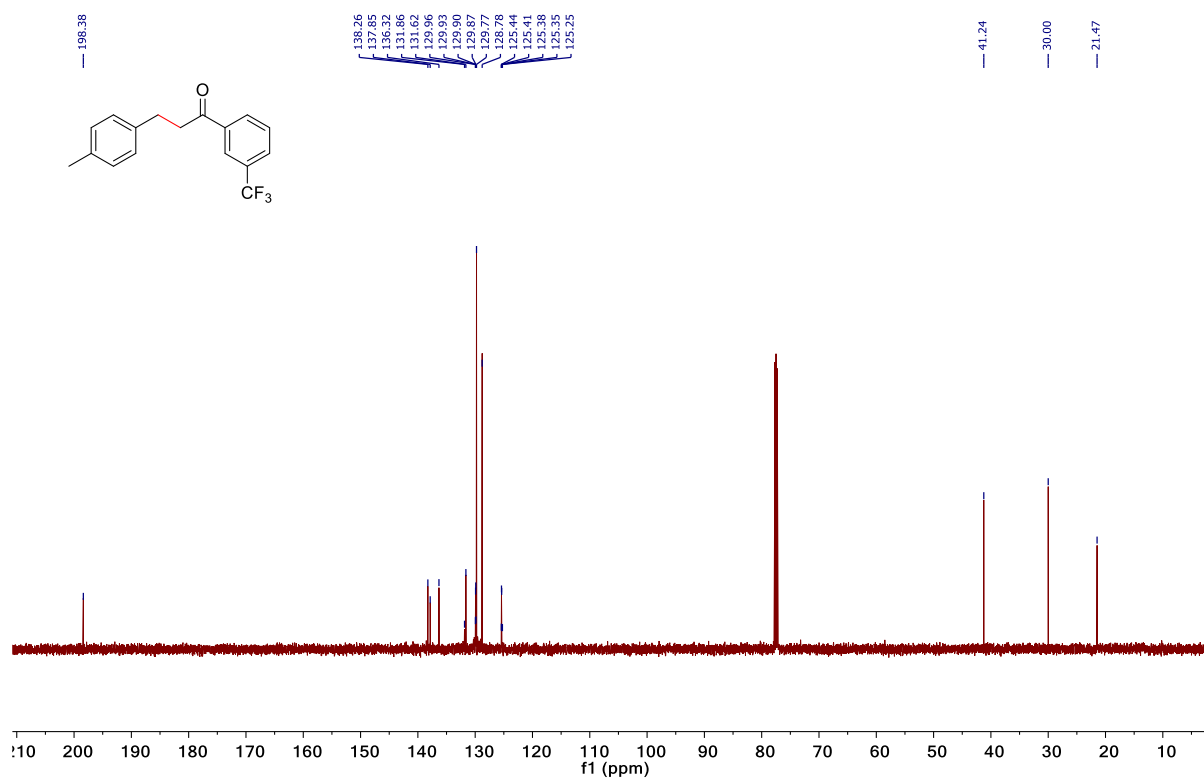
3.54
3.52
3.51
3.27
3.24



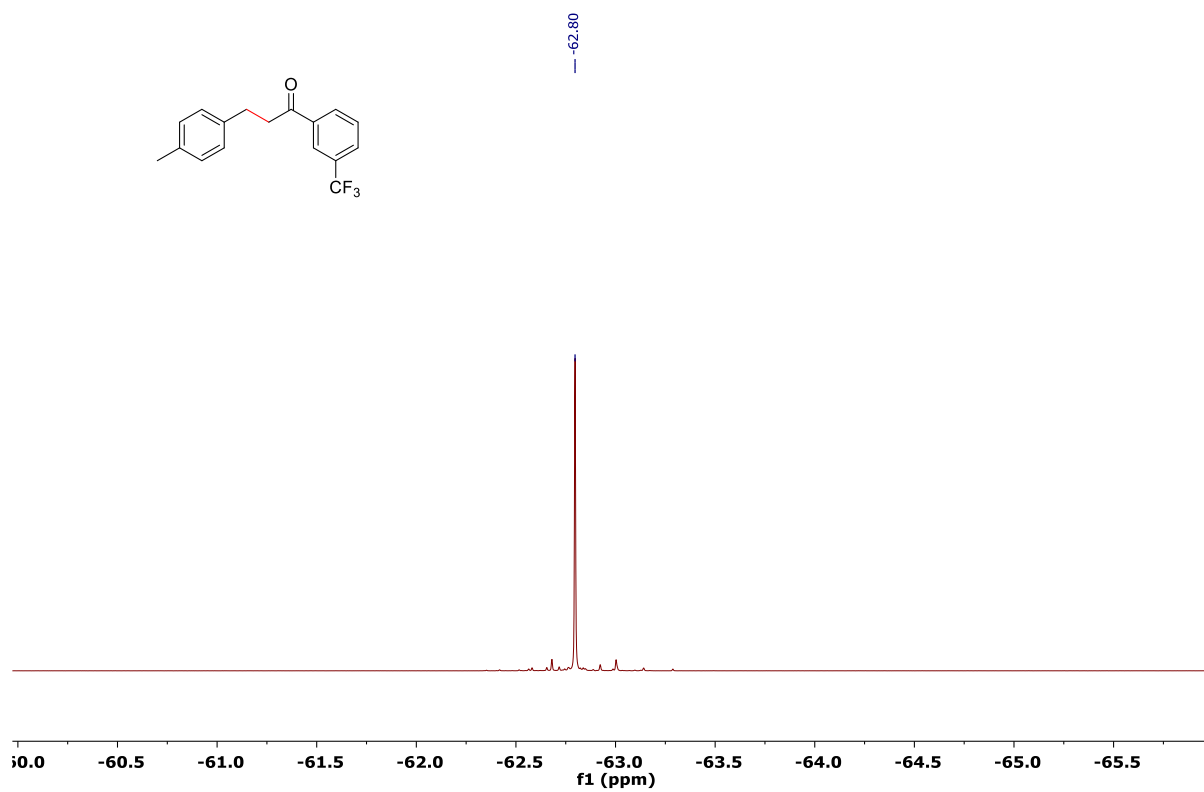


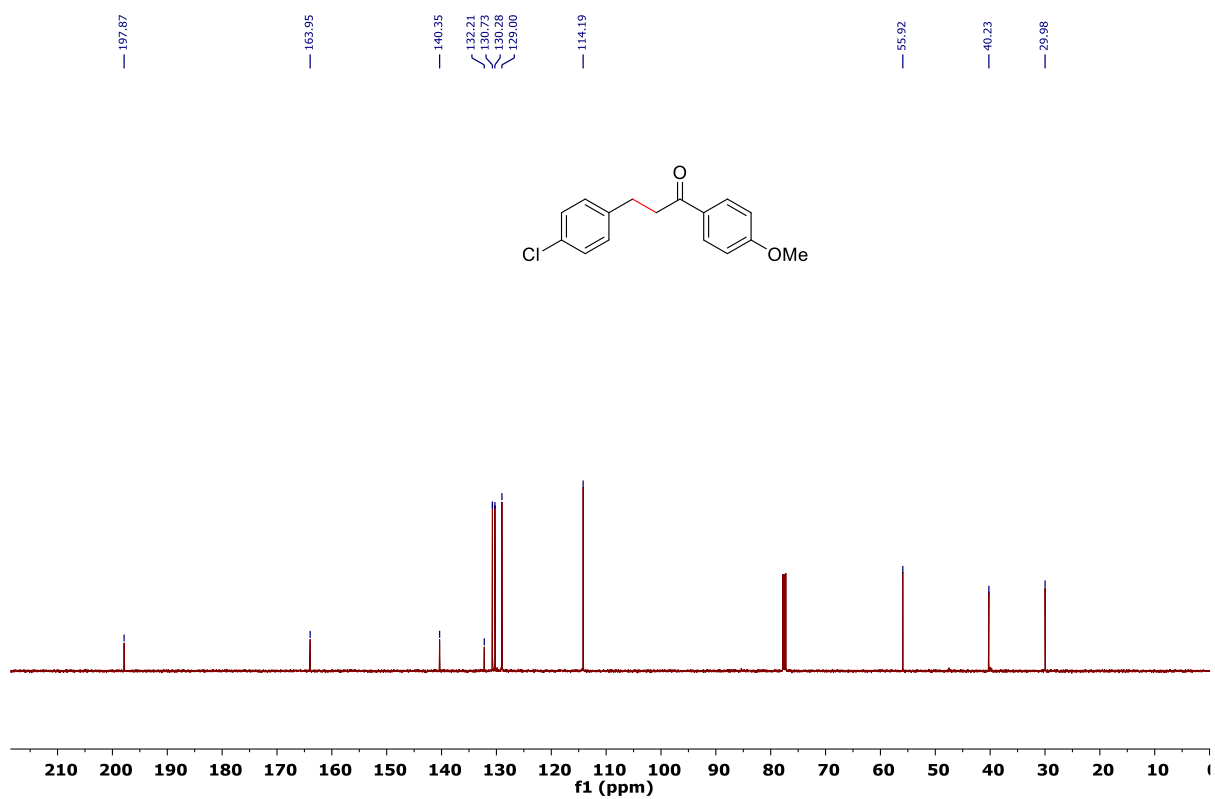
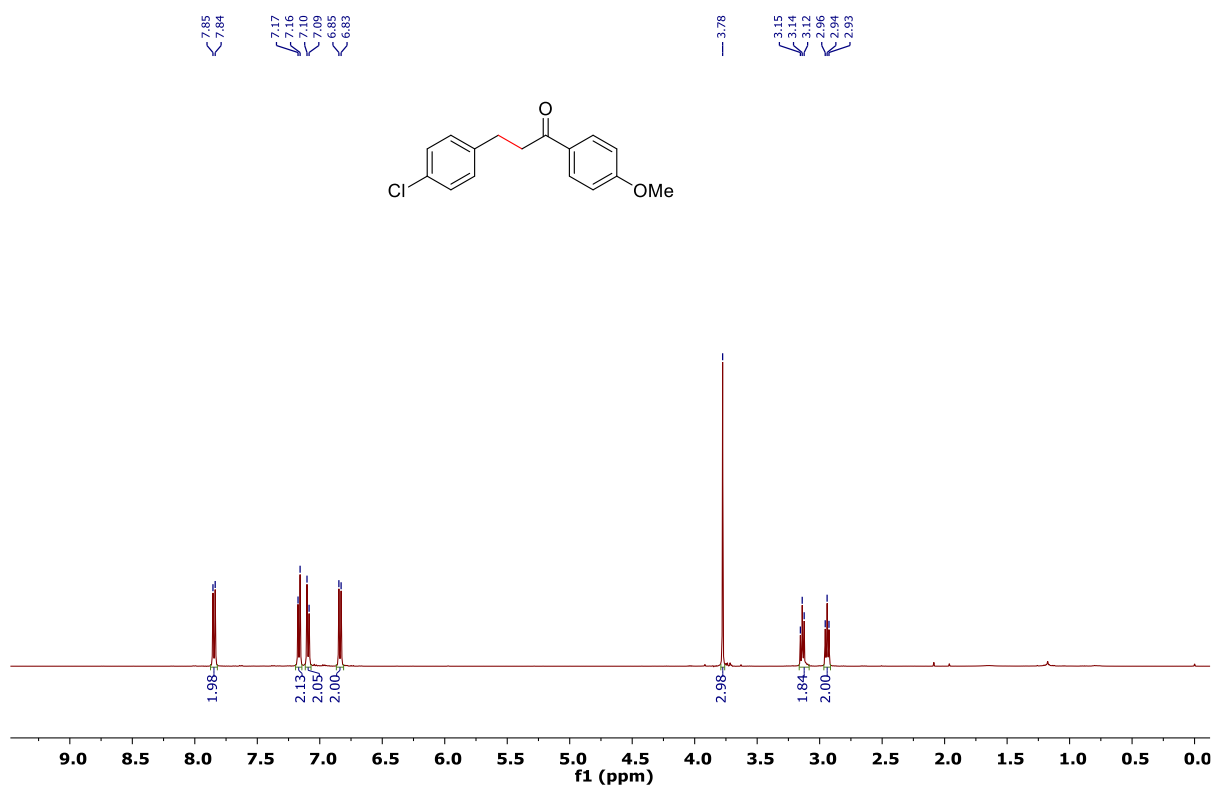


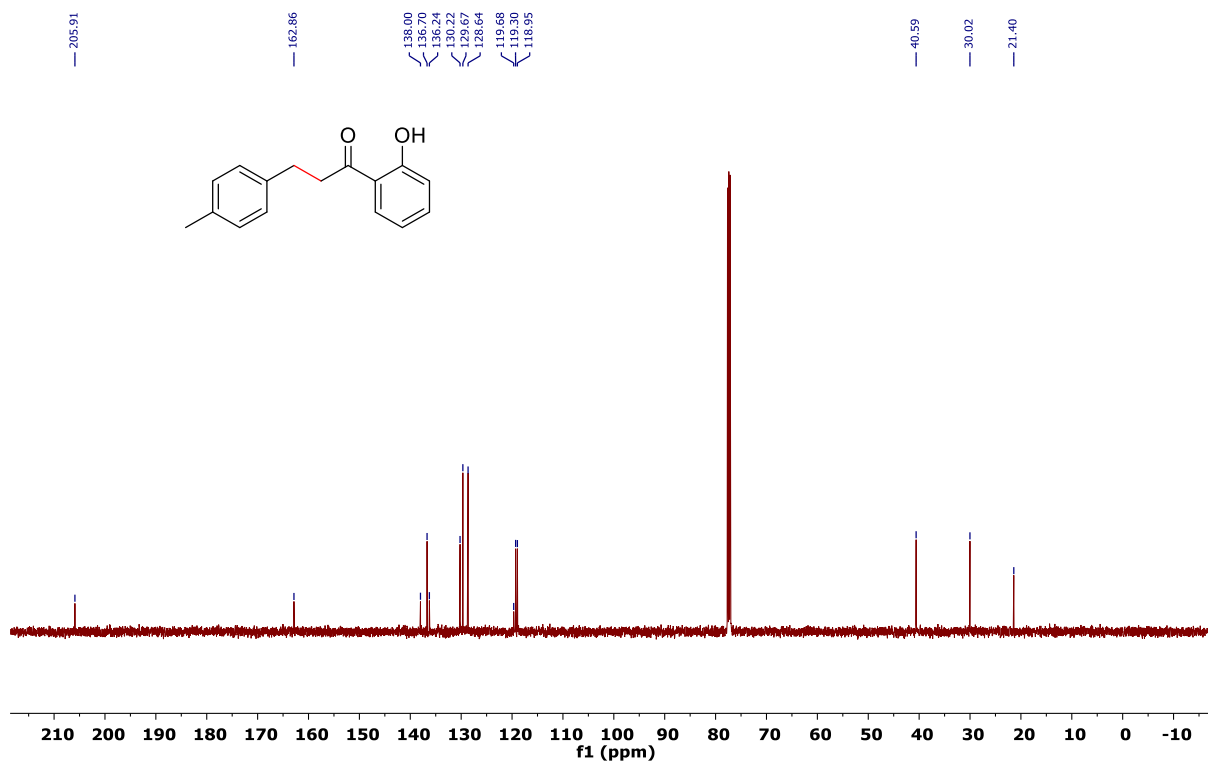
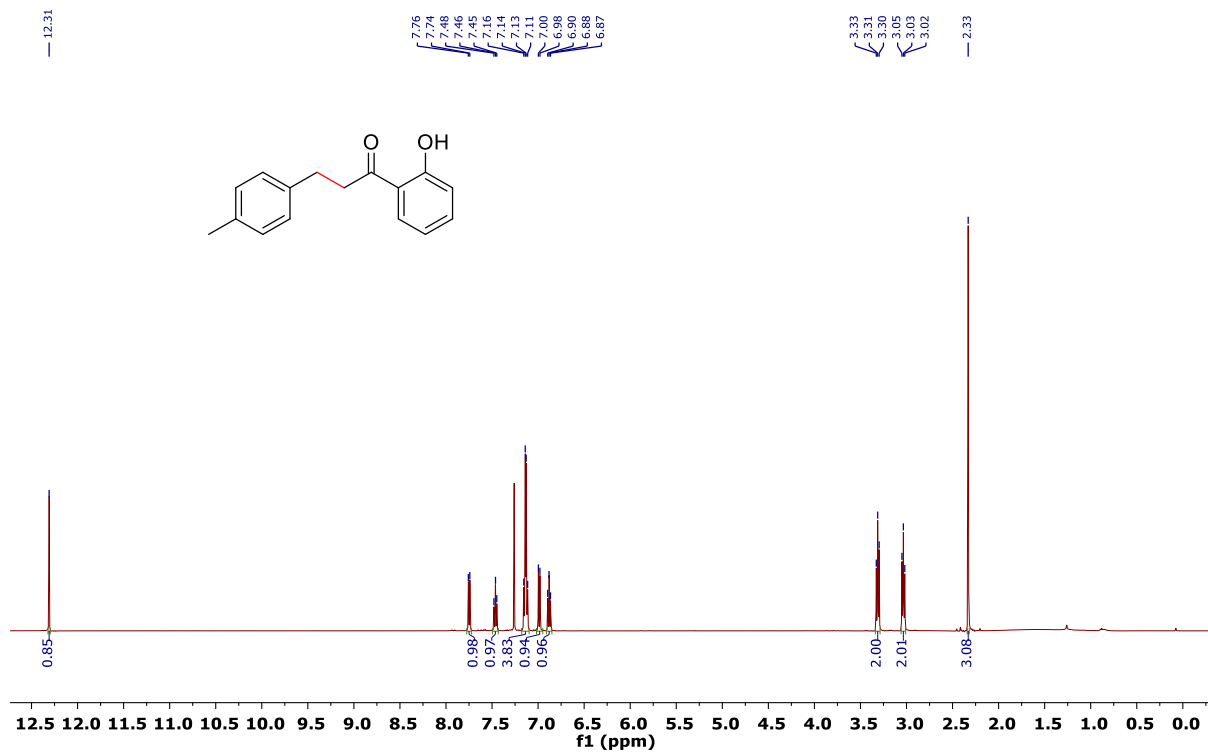


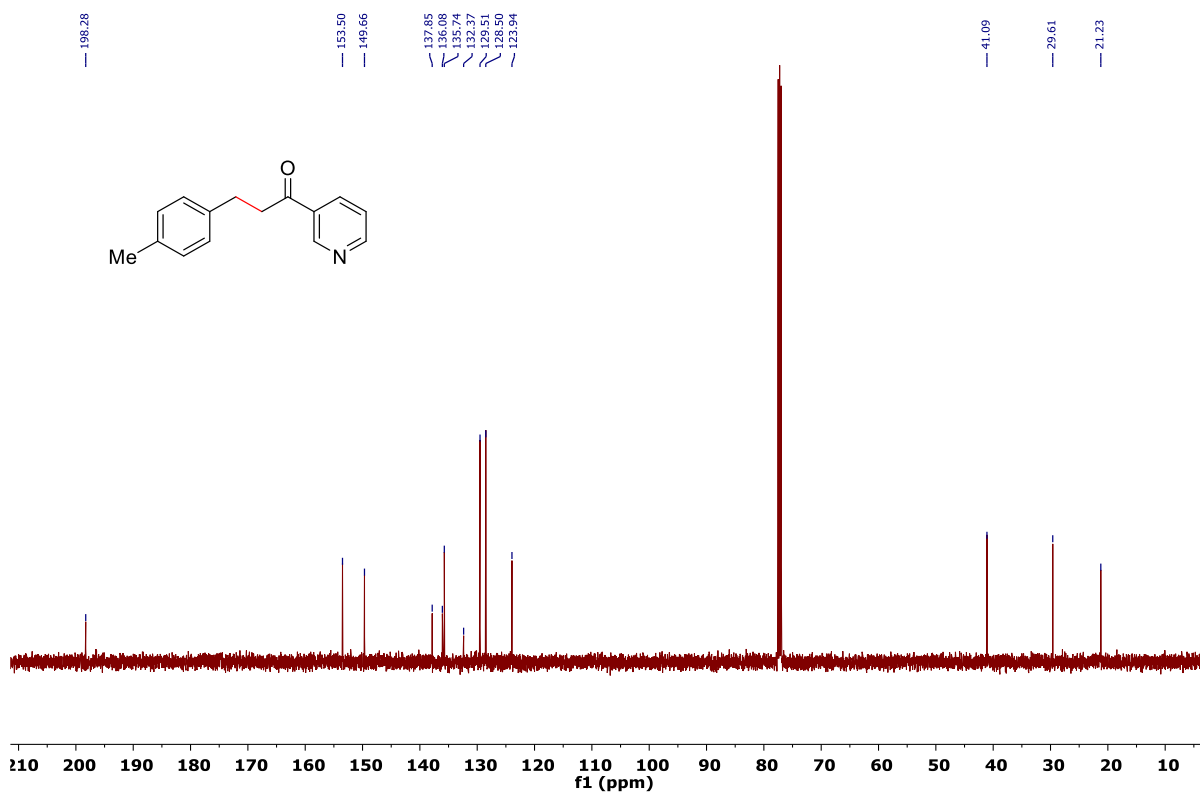
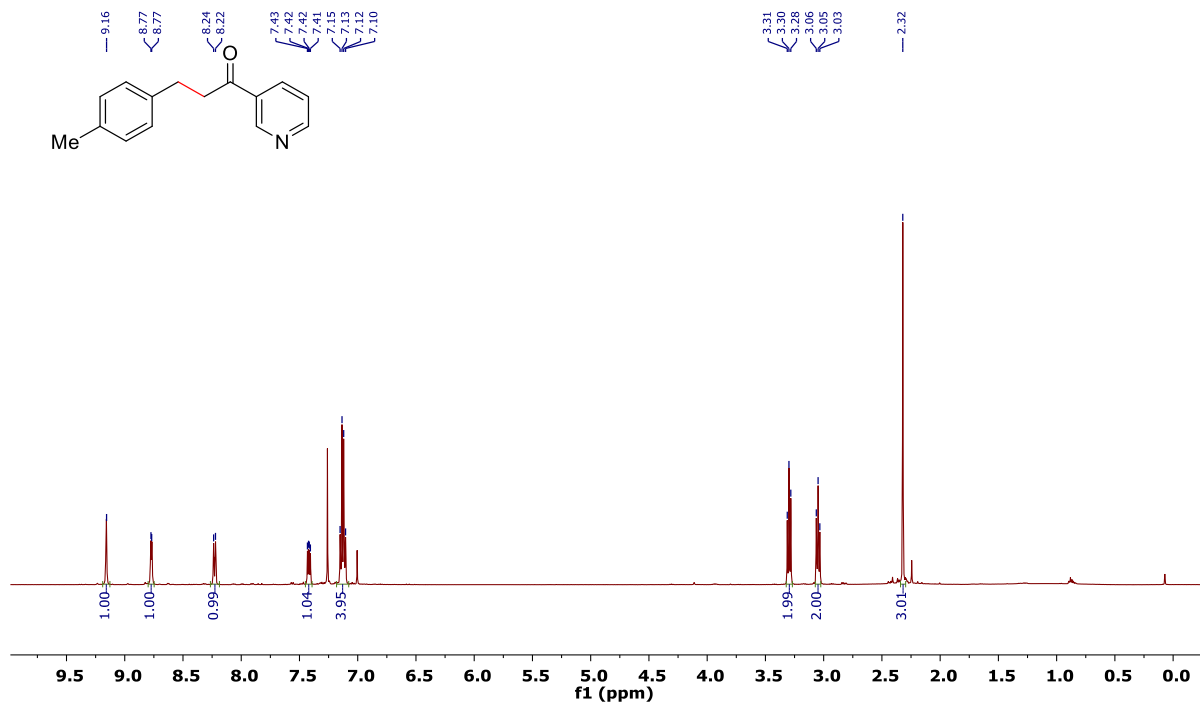


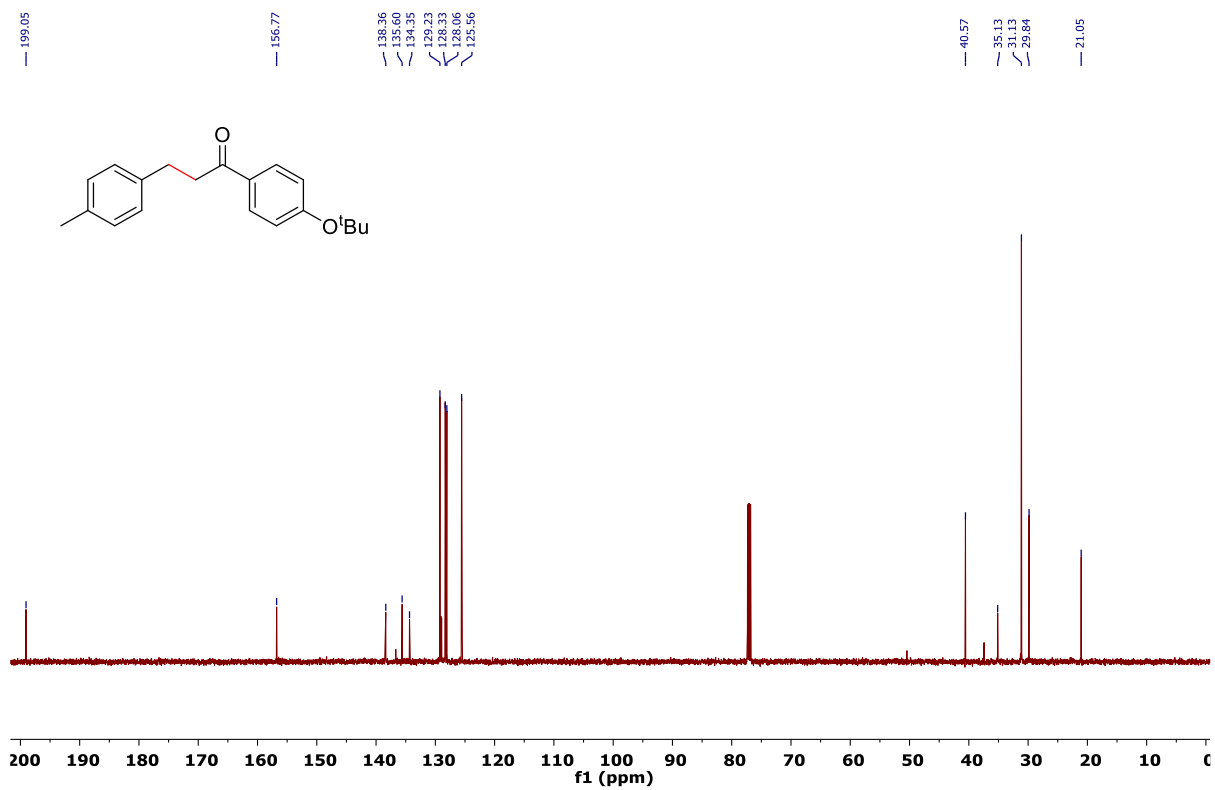
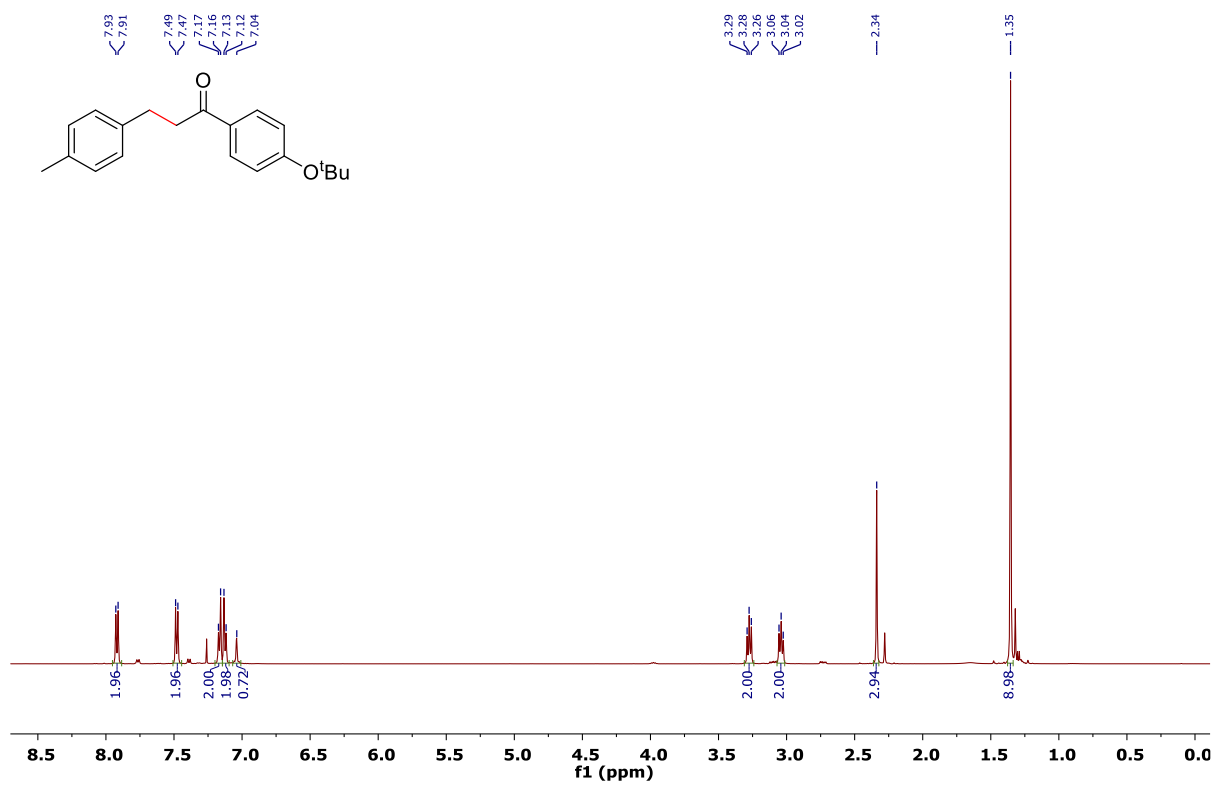
^{19}F NMR

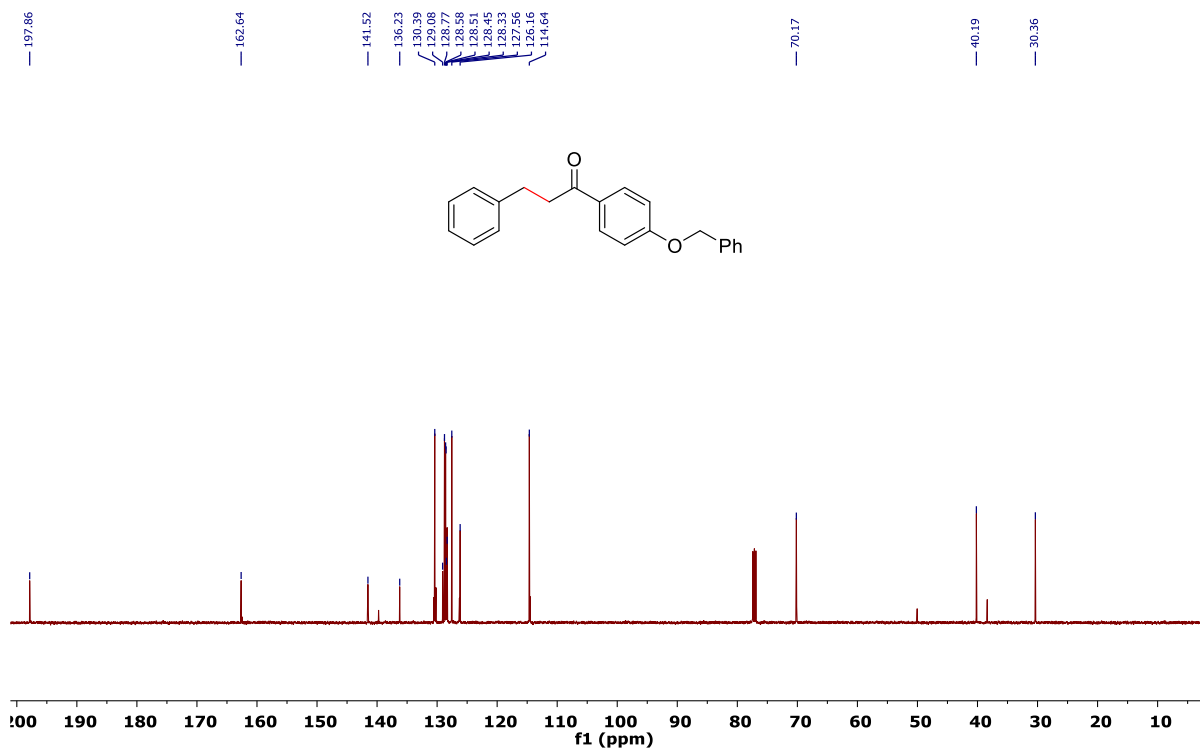
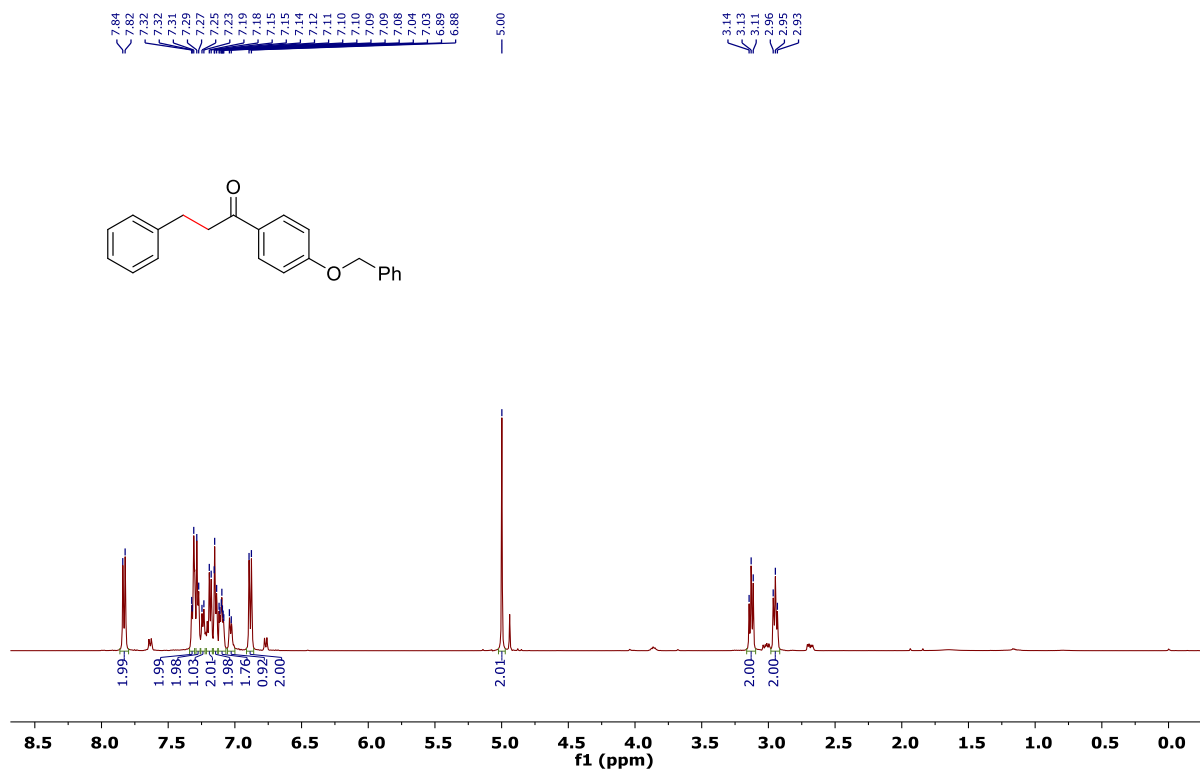


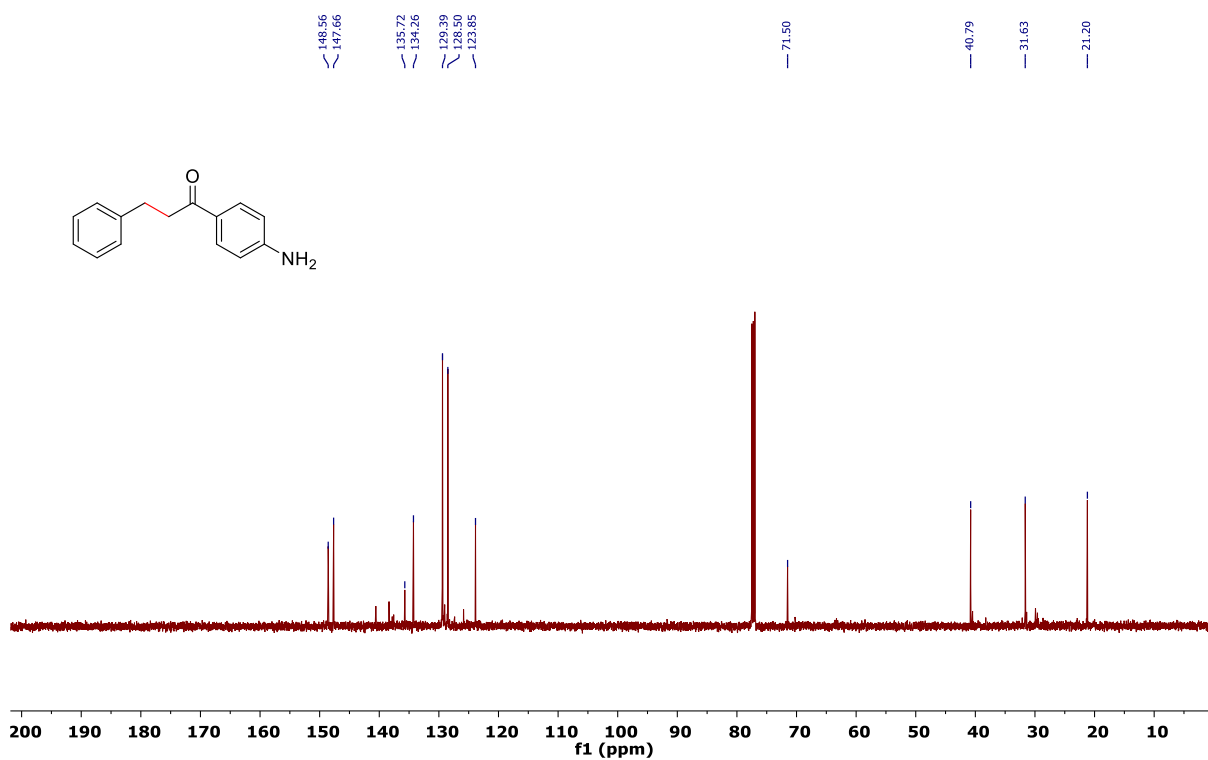
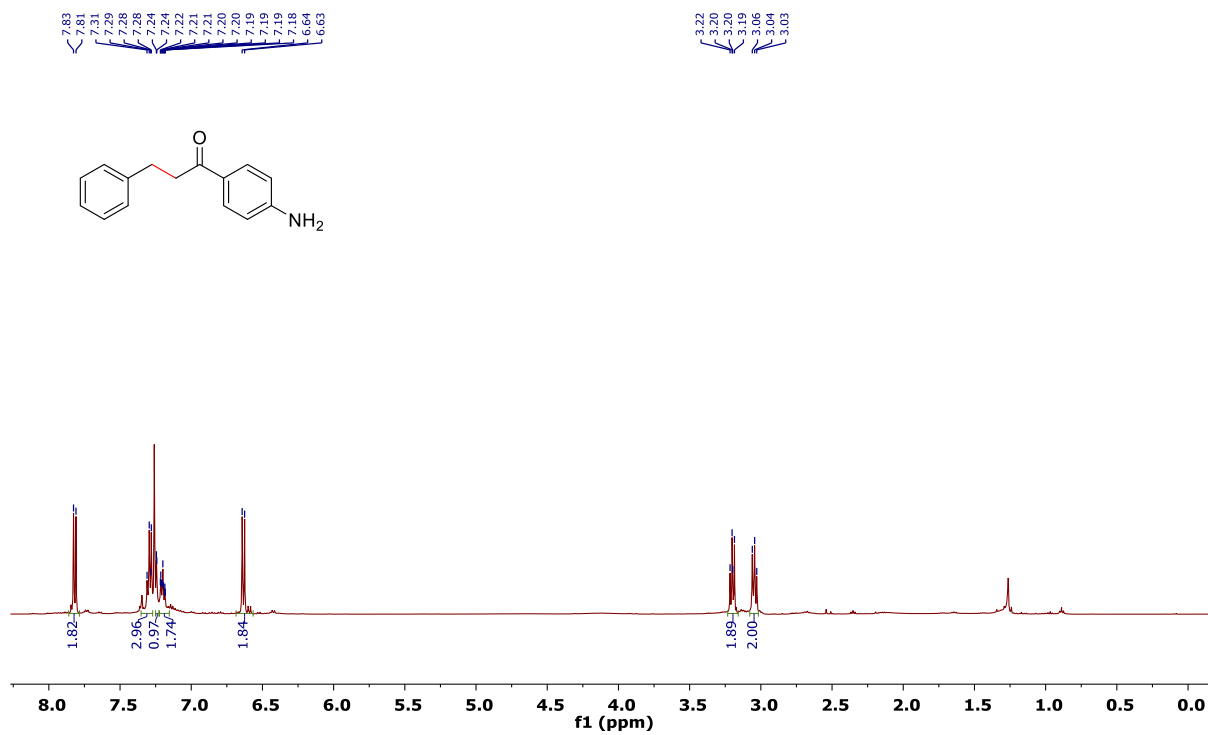


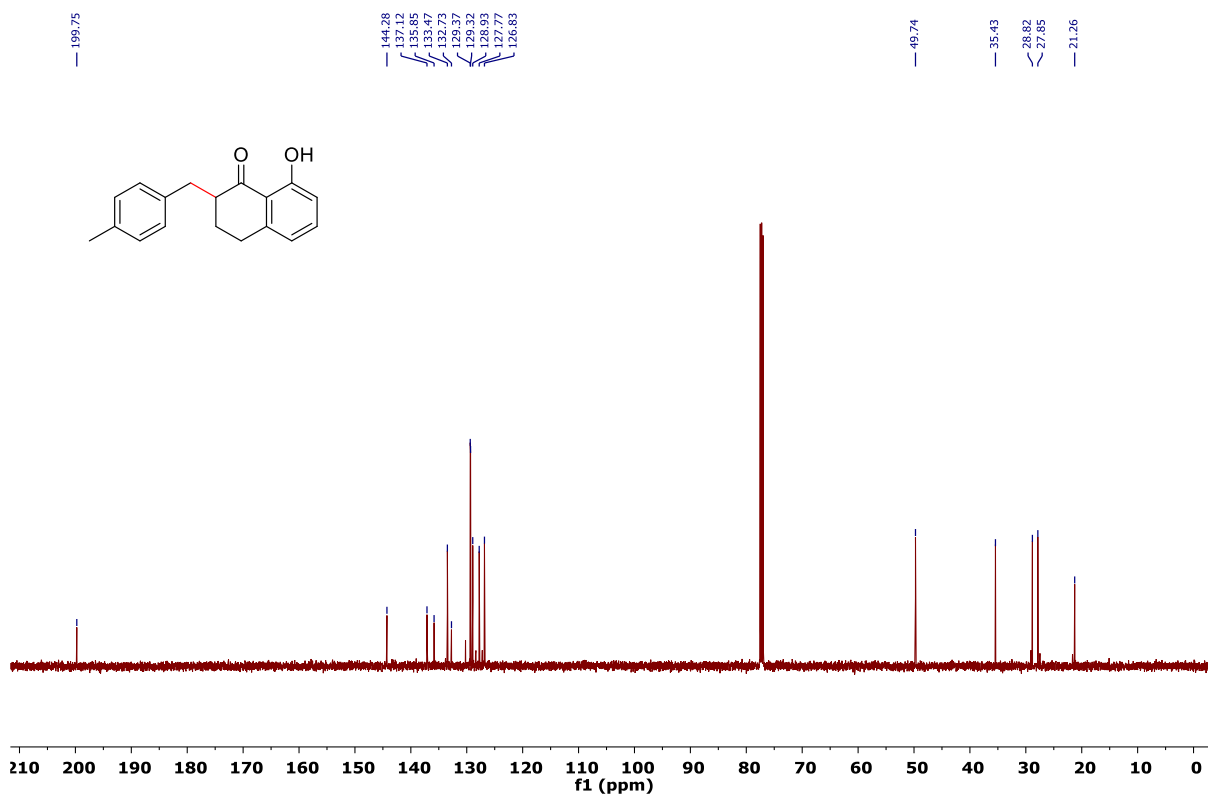
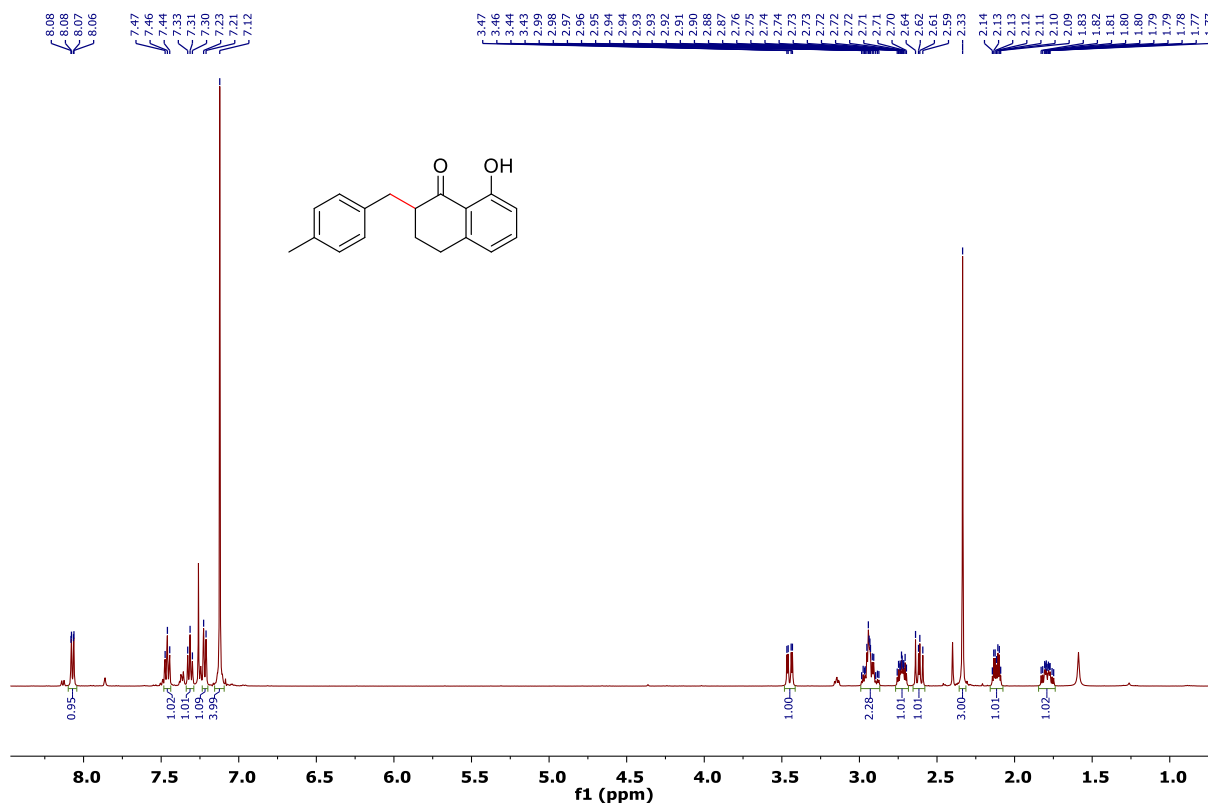


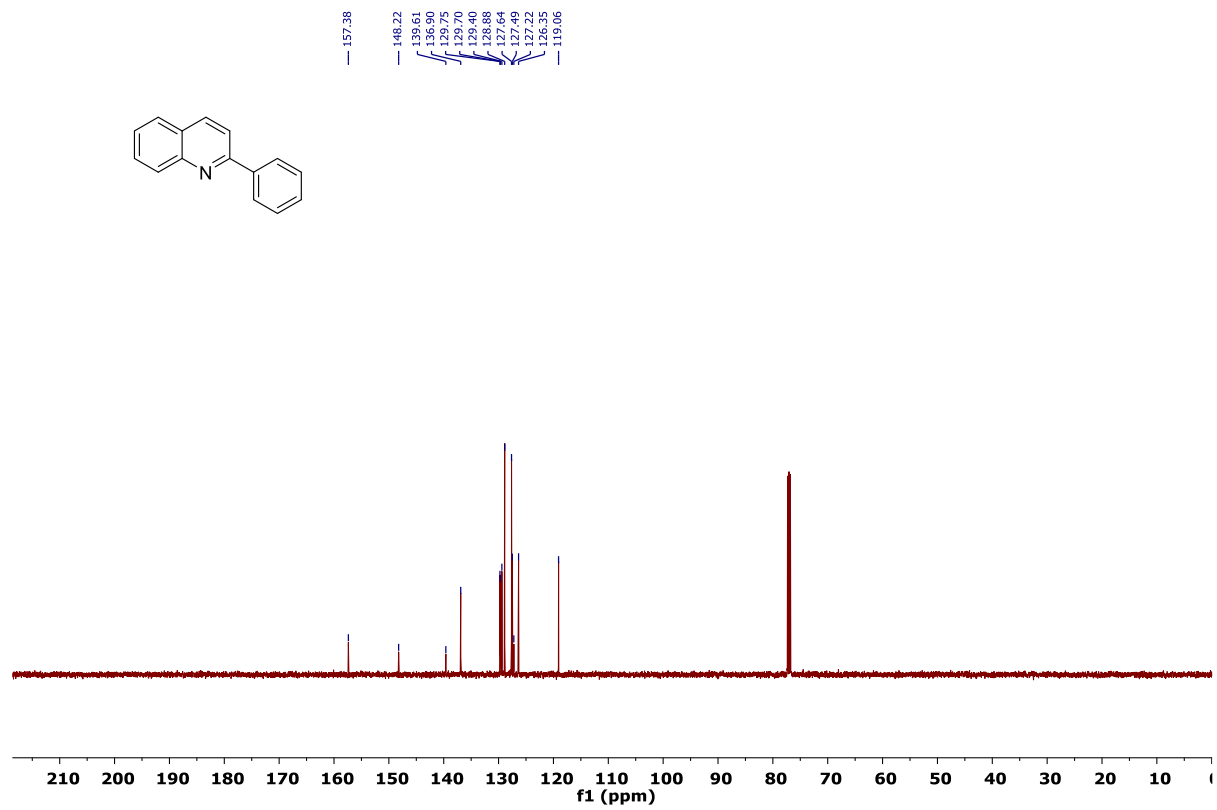
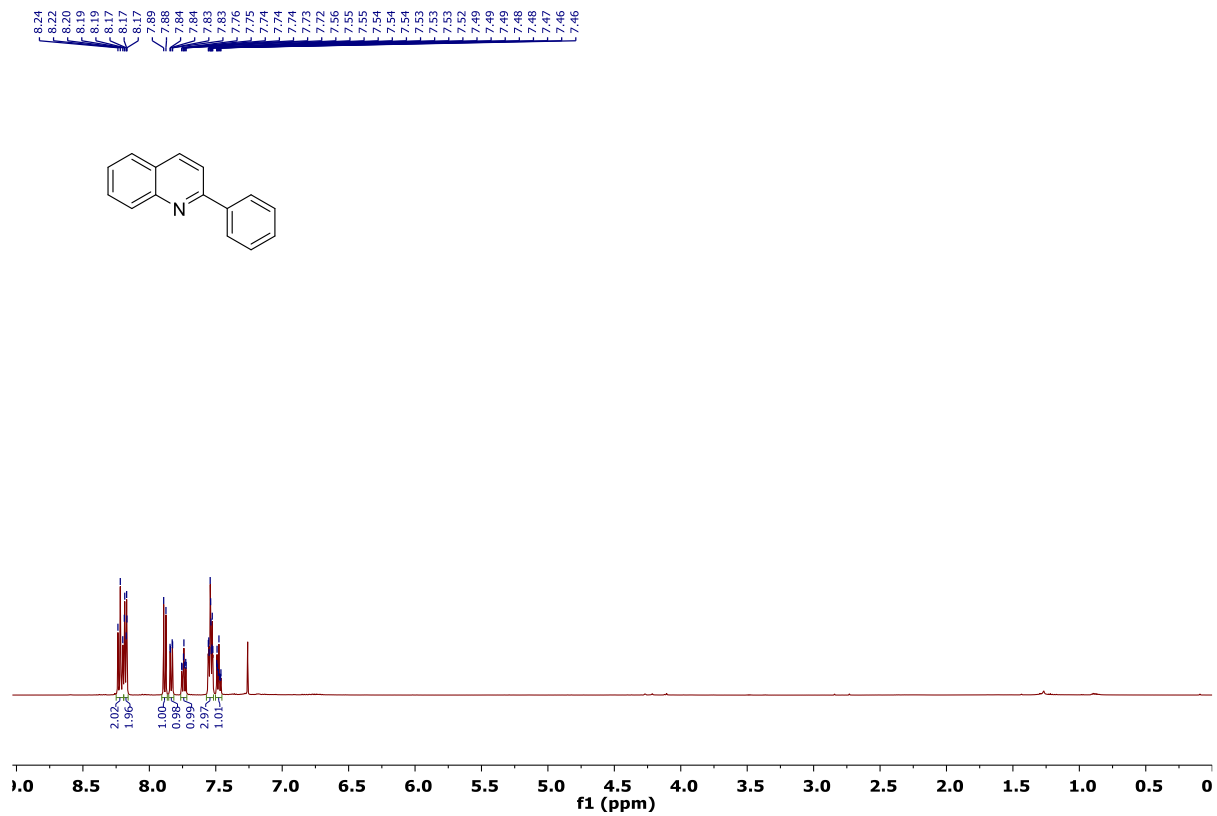


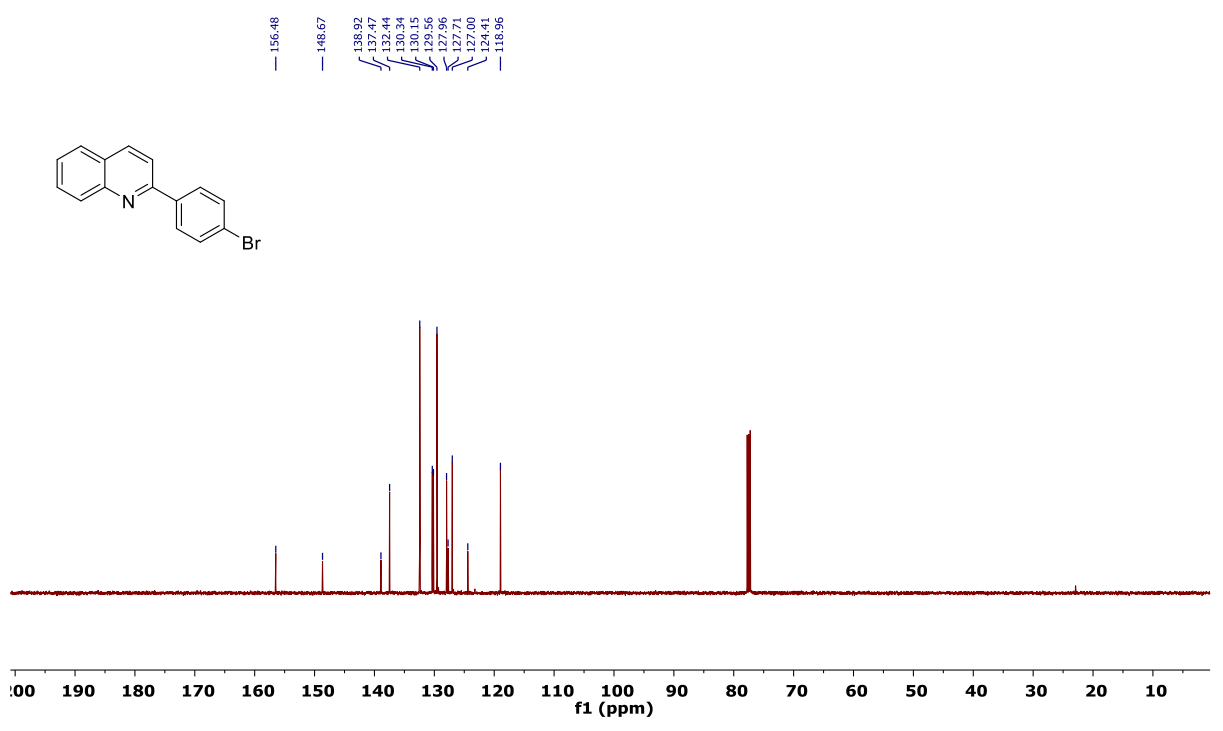
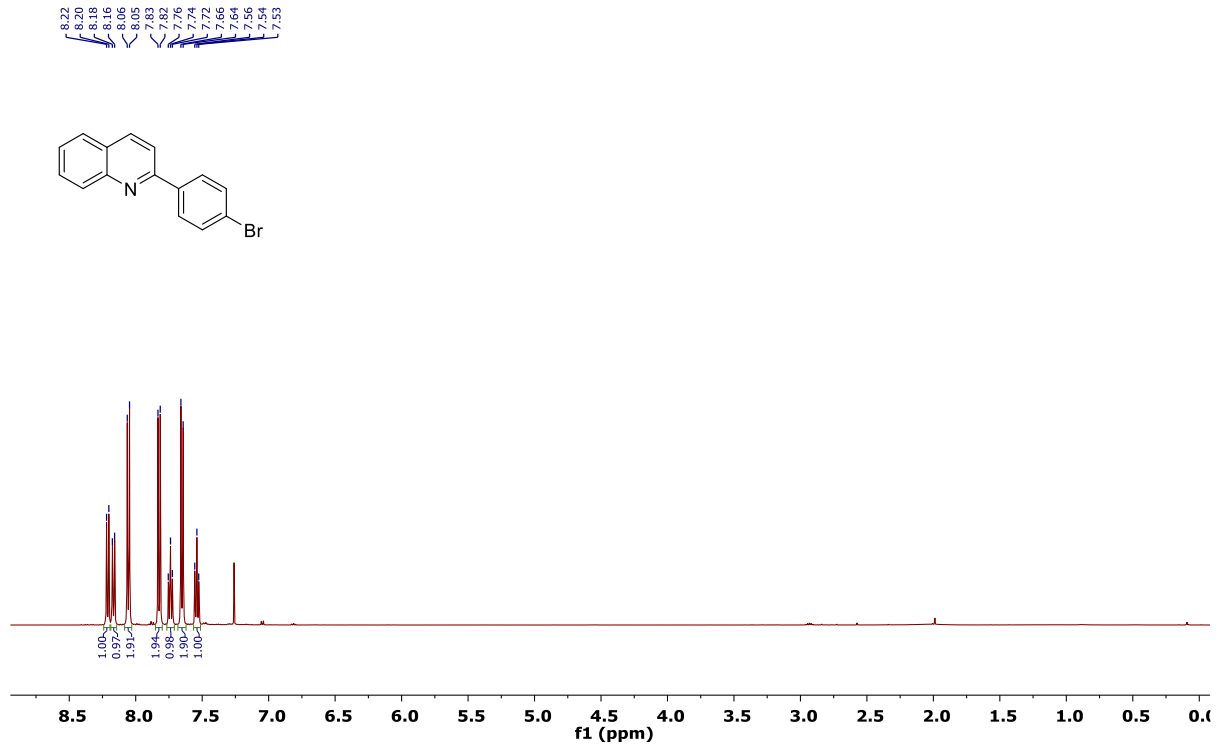




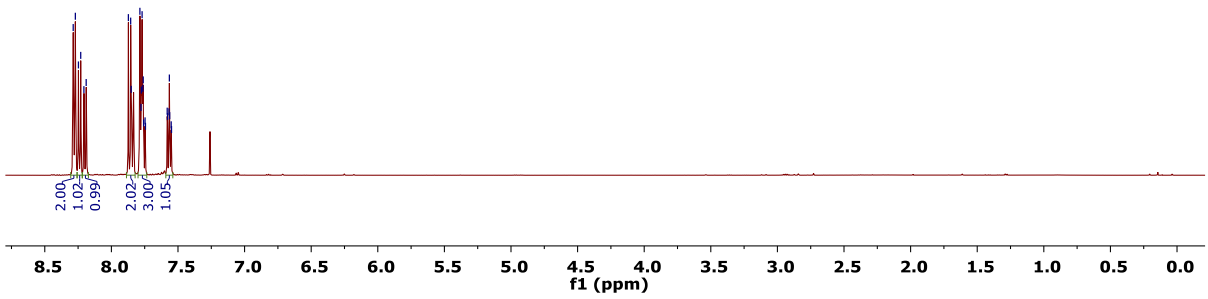
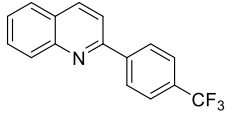




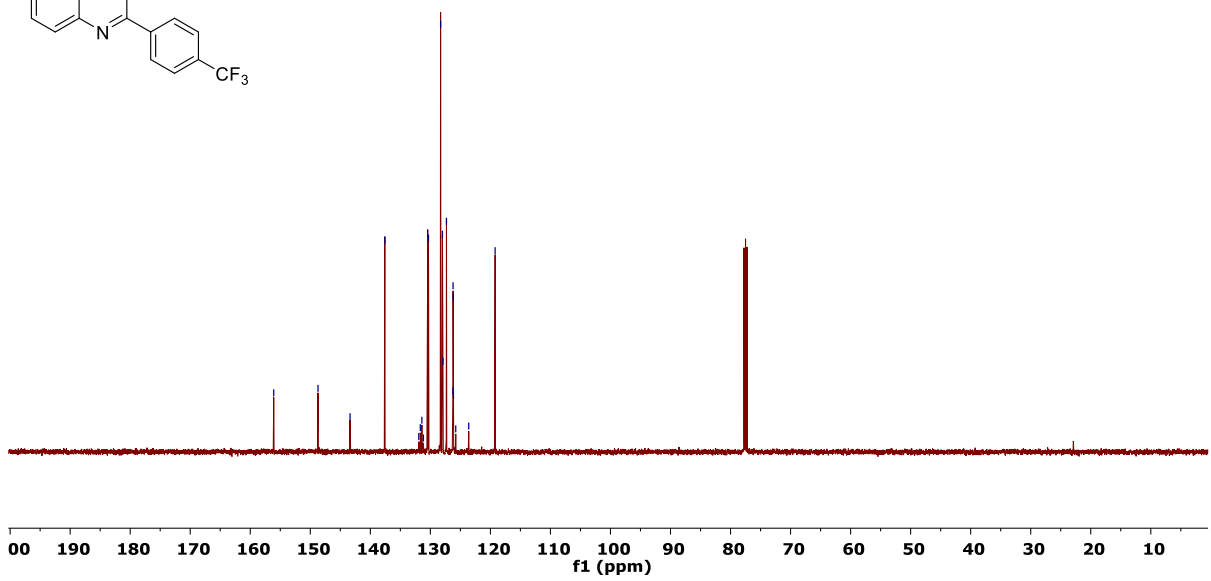
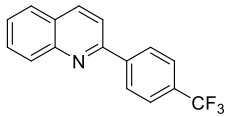




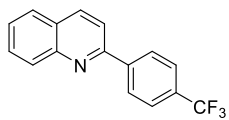
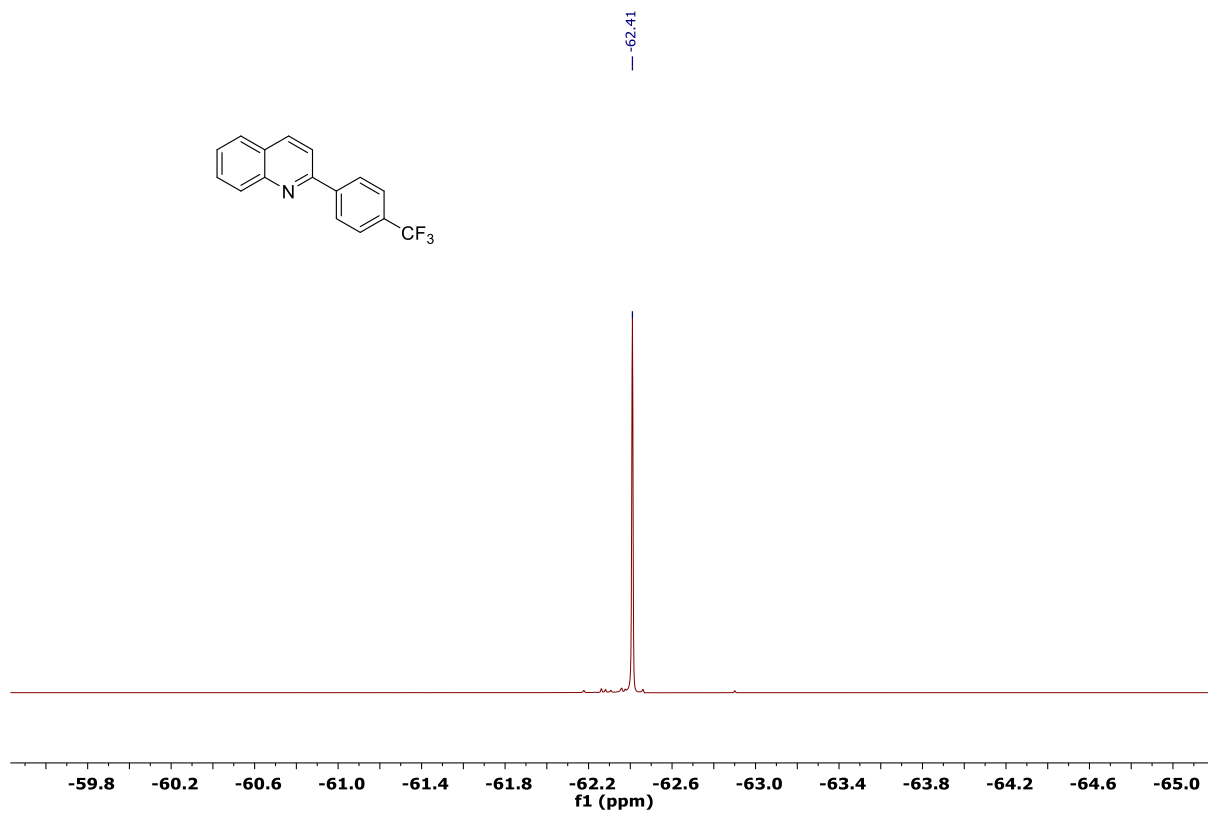
8.29
8.27
8.25
8.23
8.21
8.19
7.87
7.86
7.85
7.79
7.78
7.77
7.76
7.75
7.74
7.58
7.57
7.56
7.55



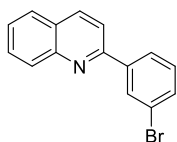
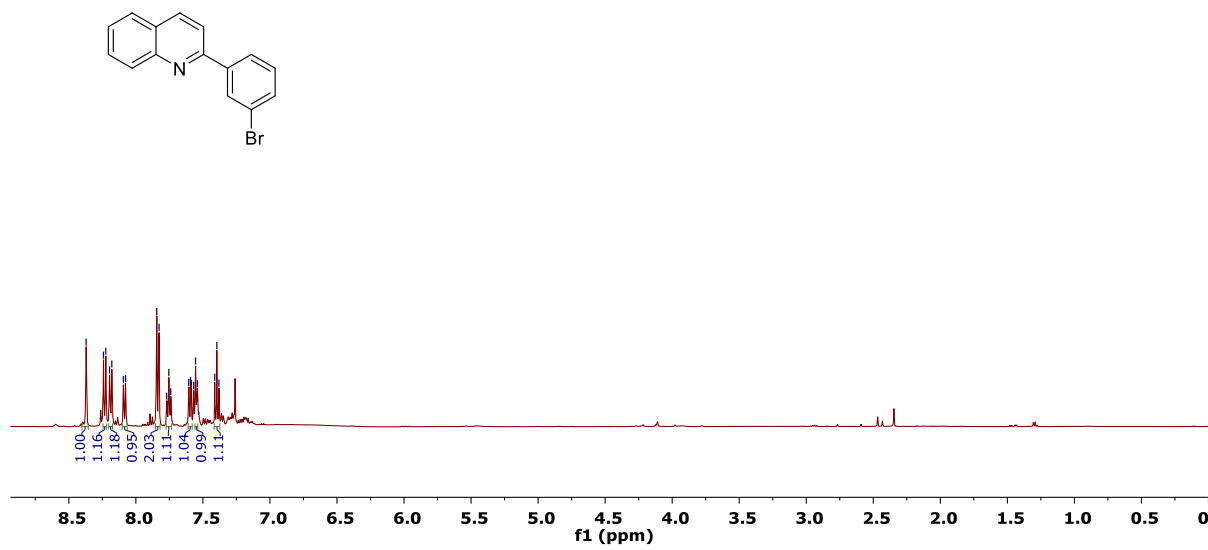
156.09
148.71
143.38
137.59
131.93
131.67
131.41
131.16
130.46
129.31
128.31
127.99
127.90
127.32
126.25
126.22
126.18
126.16
125.77
123.67
119.21



¹⁹F NMR



8.37
8.24
8.22
8.20
8.18
8.08
8.08
7.84
7.83
7.77
7.75
7.74
7.61
7.59
7.57
7.55
7.54
7.41
7.40
7.38



1.00

1.16

1.18

0.95

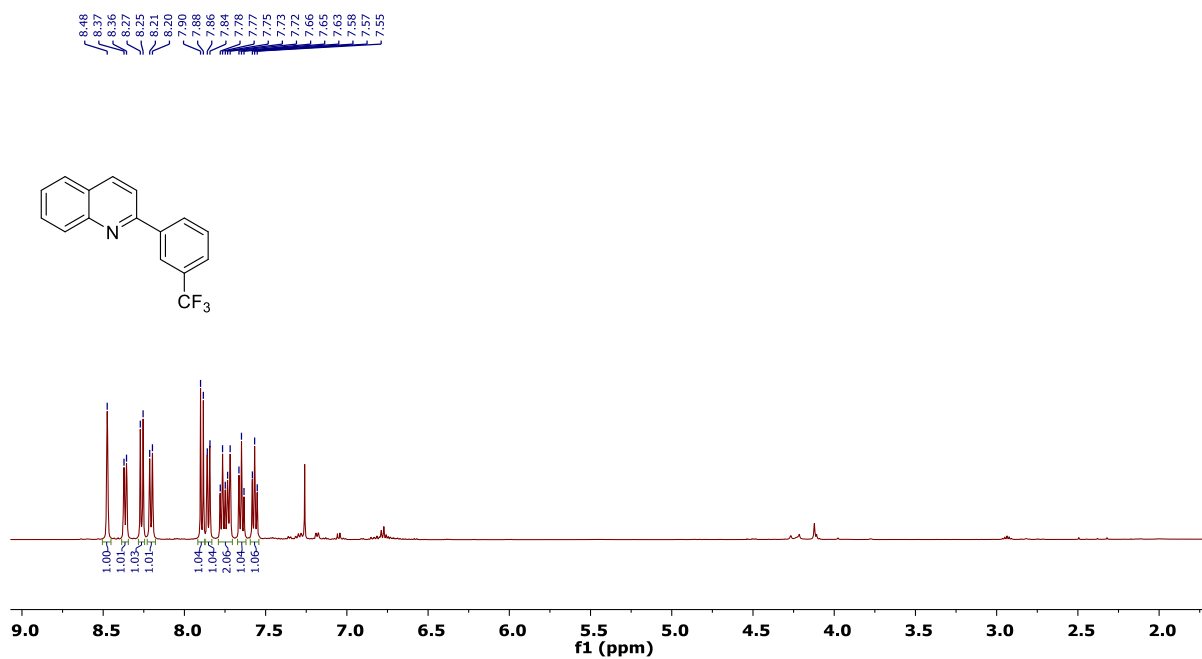
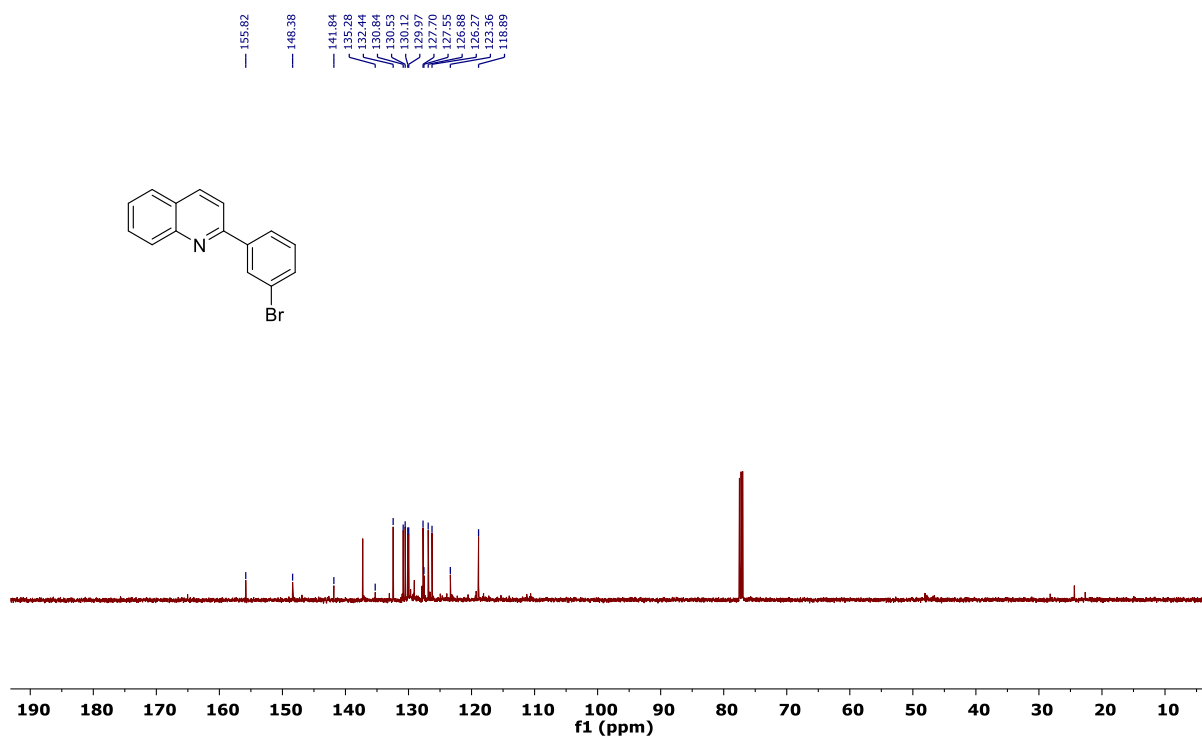
2.03

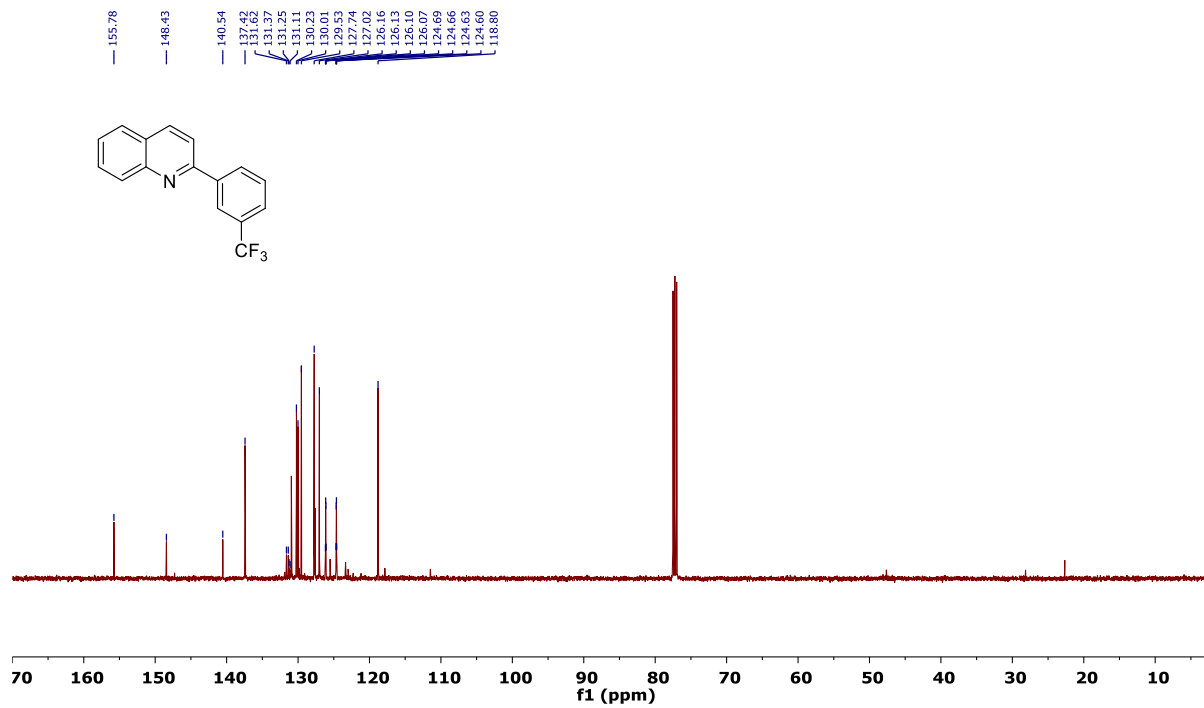
1.11

1.04

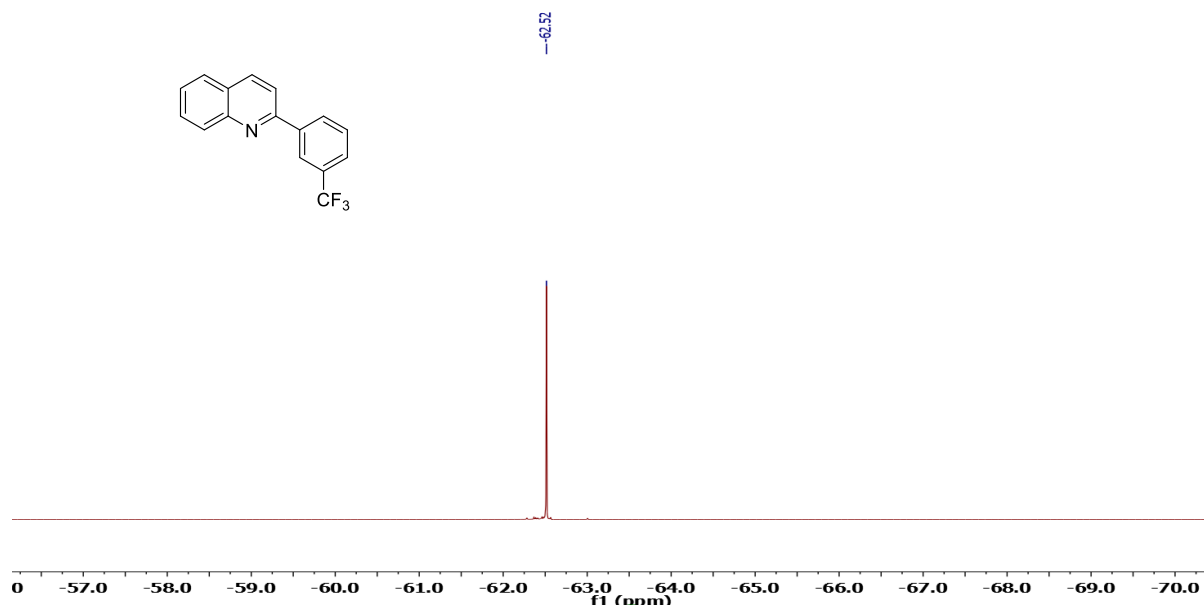
0.99

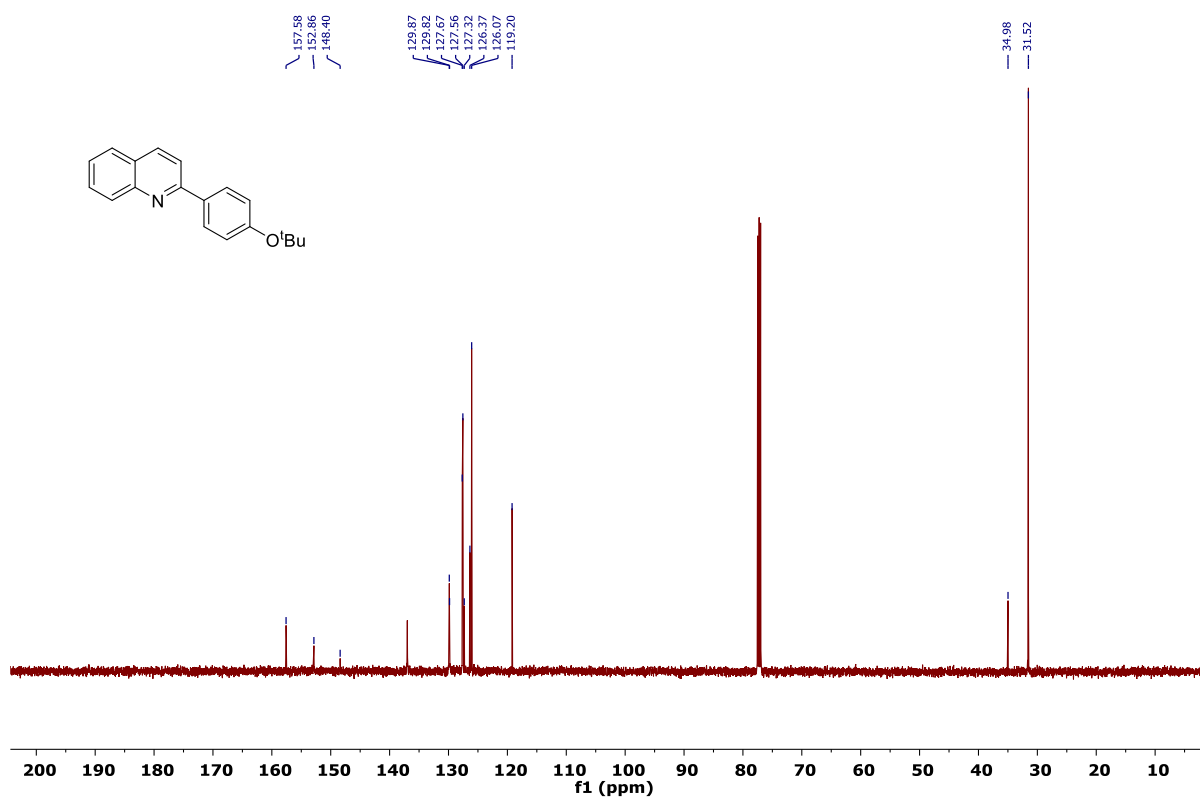
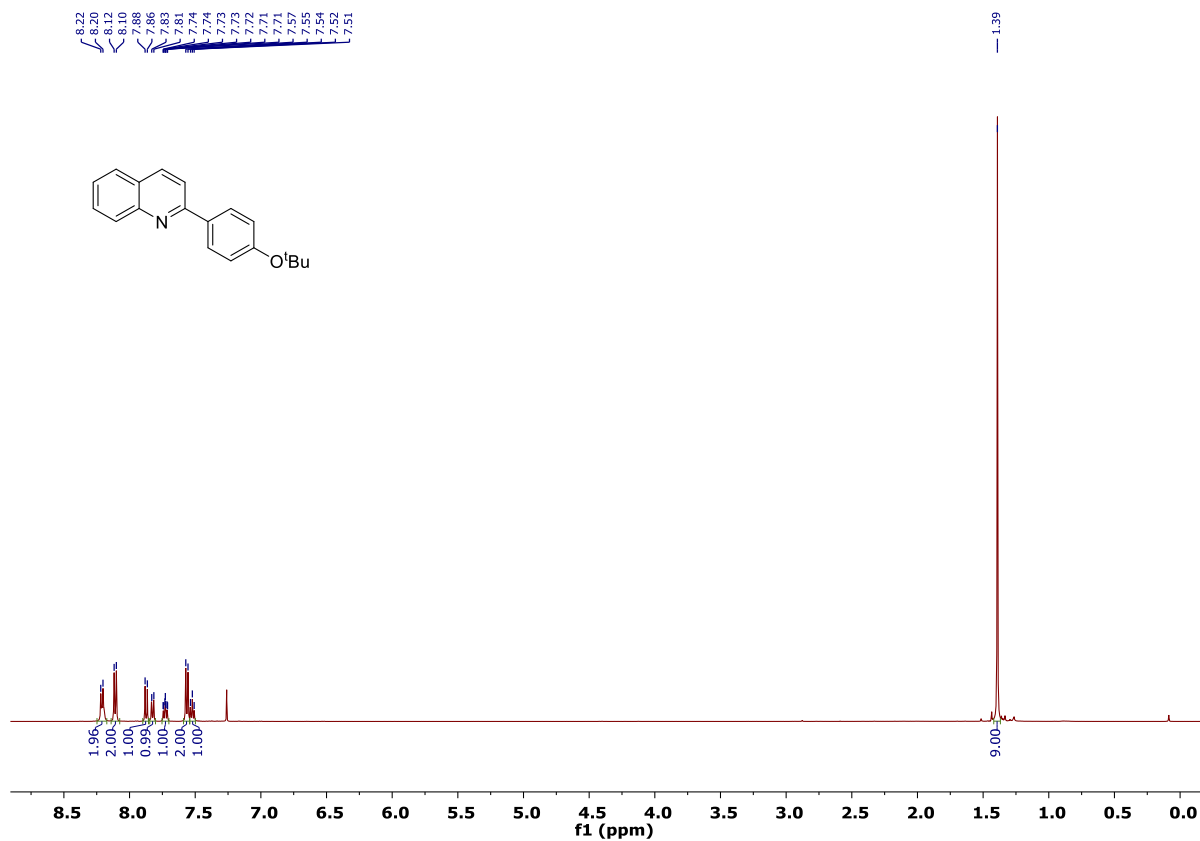
1.11

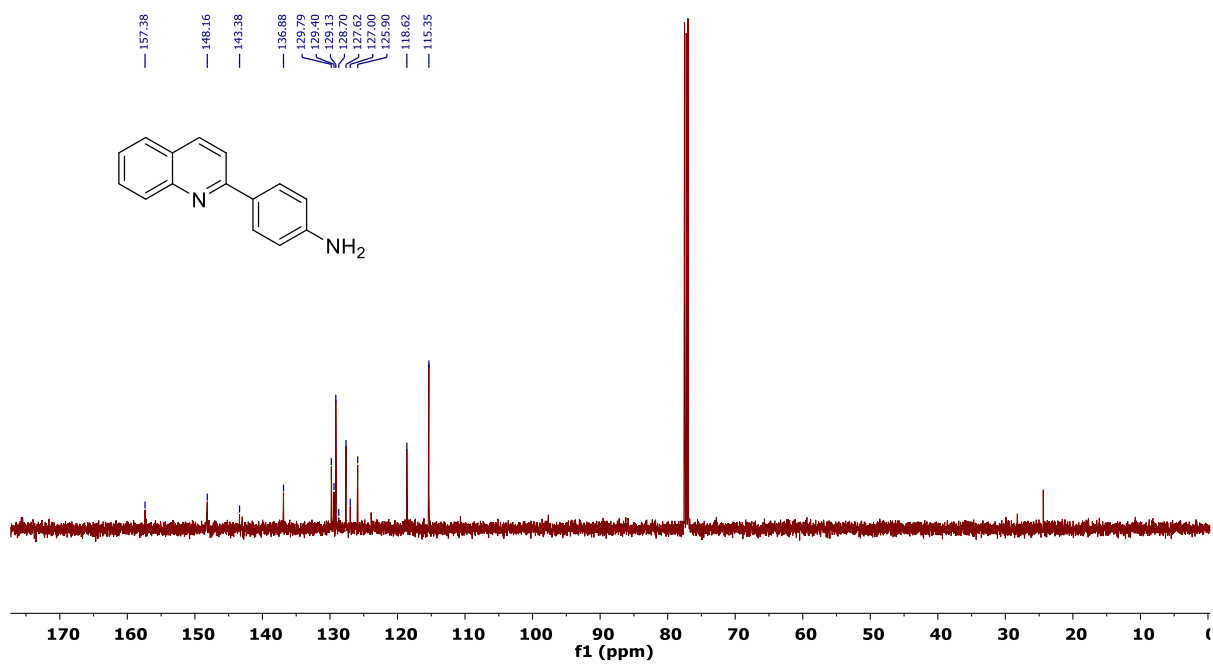
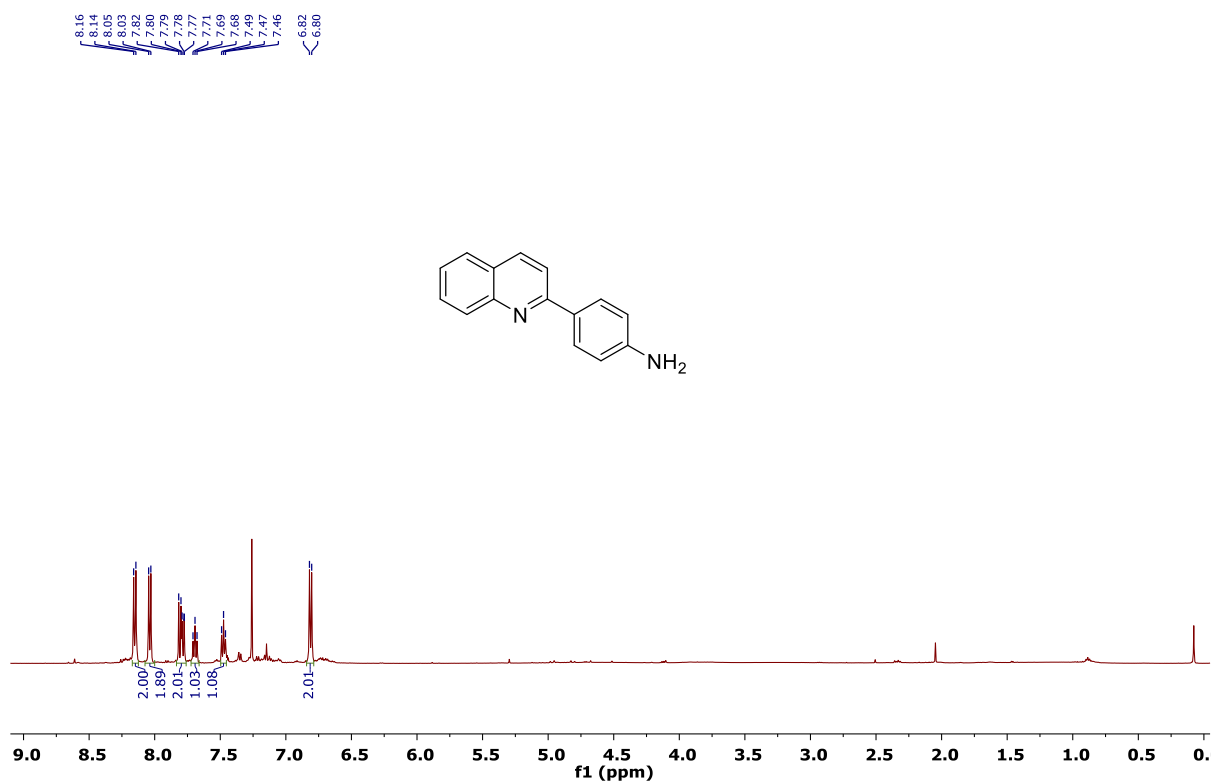




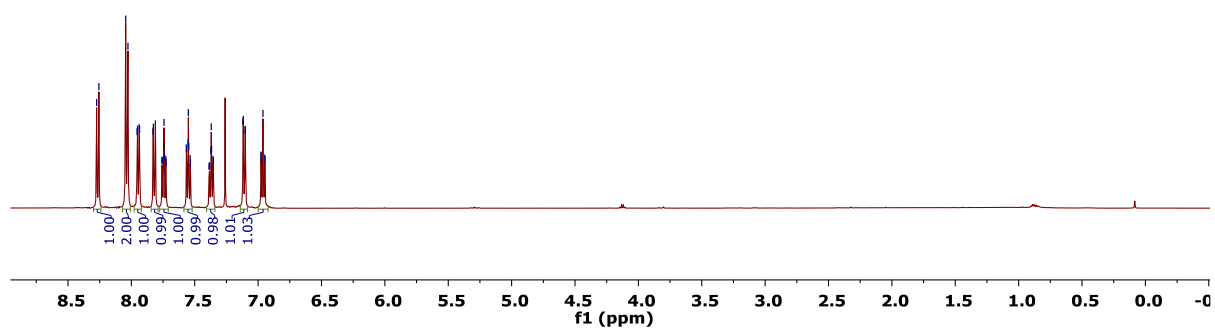
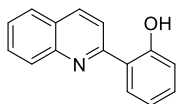
¹⁹F NMR



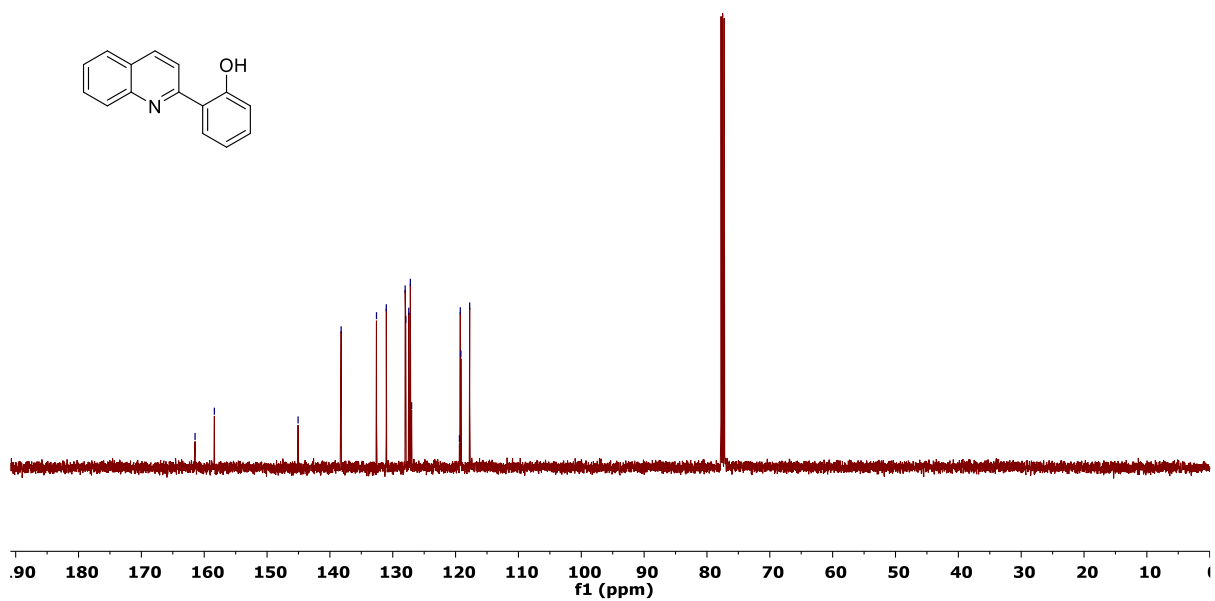
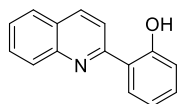


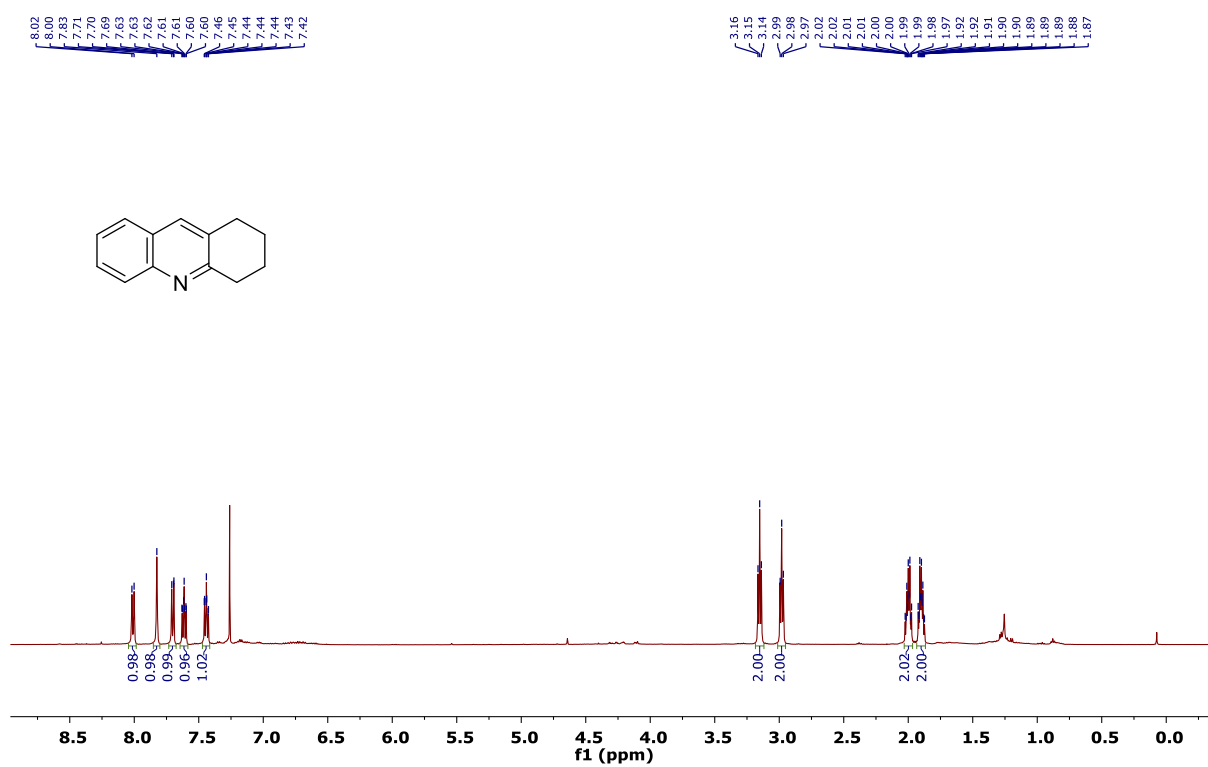
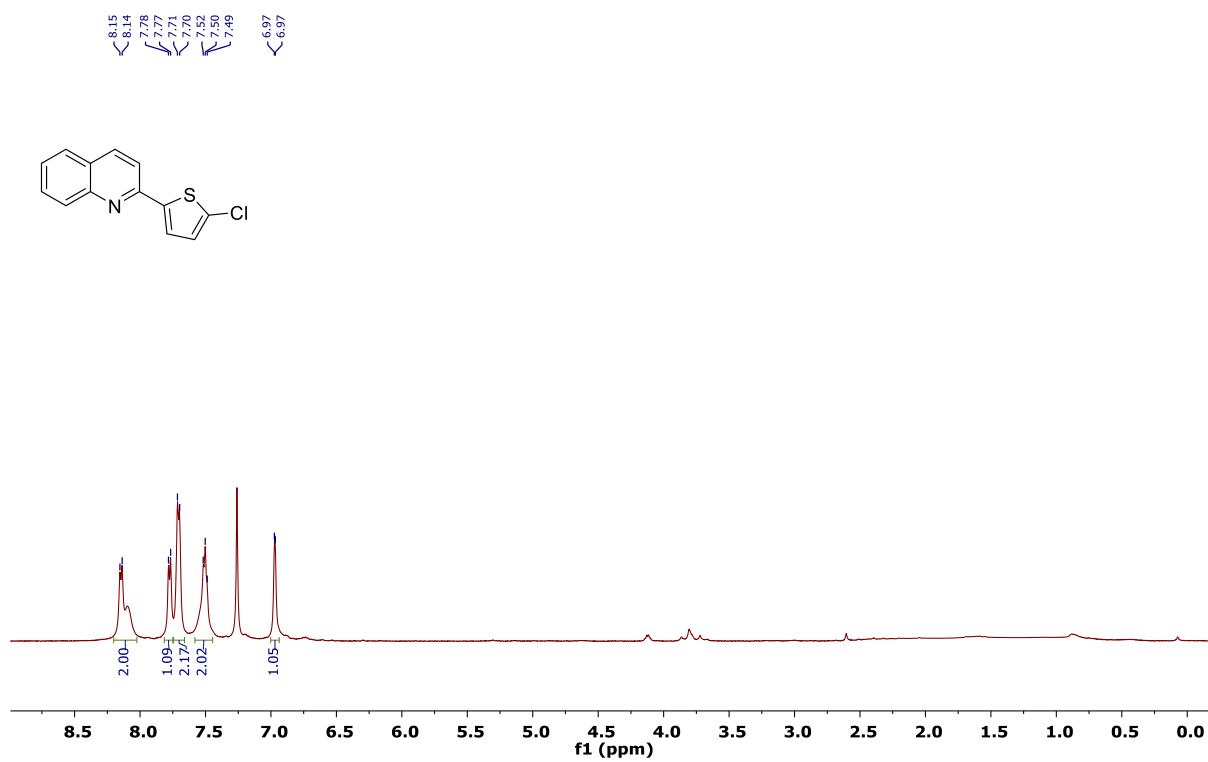


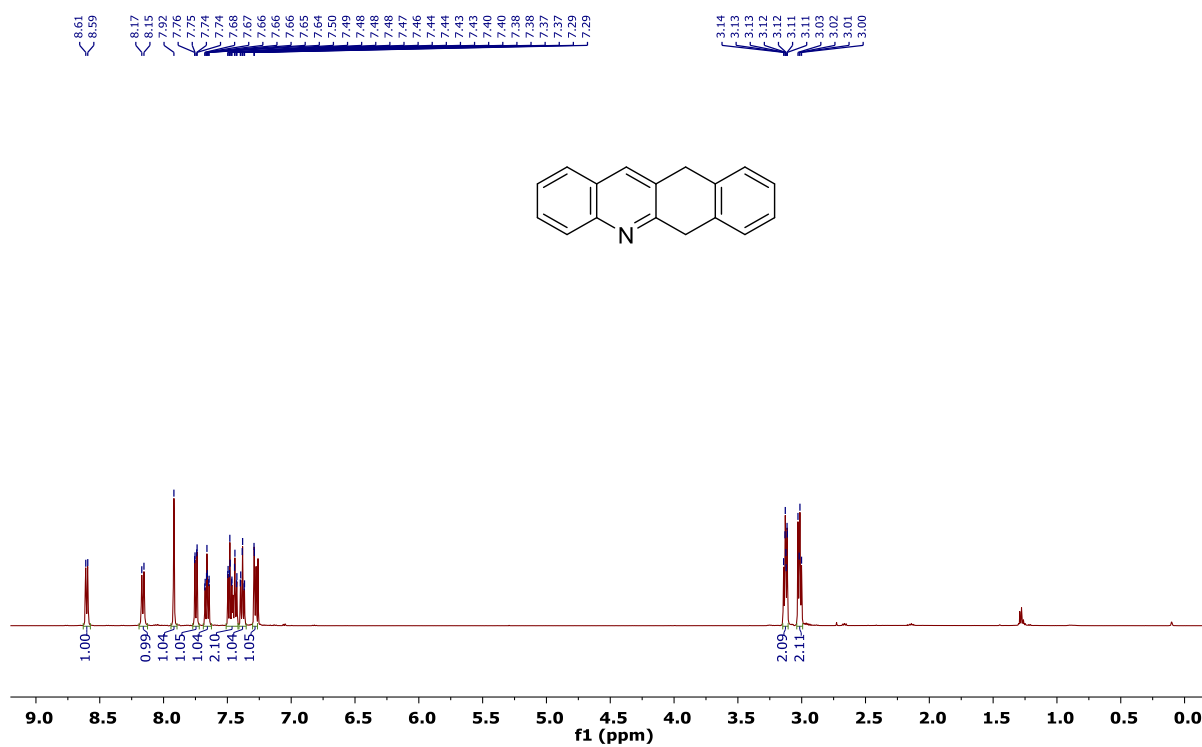
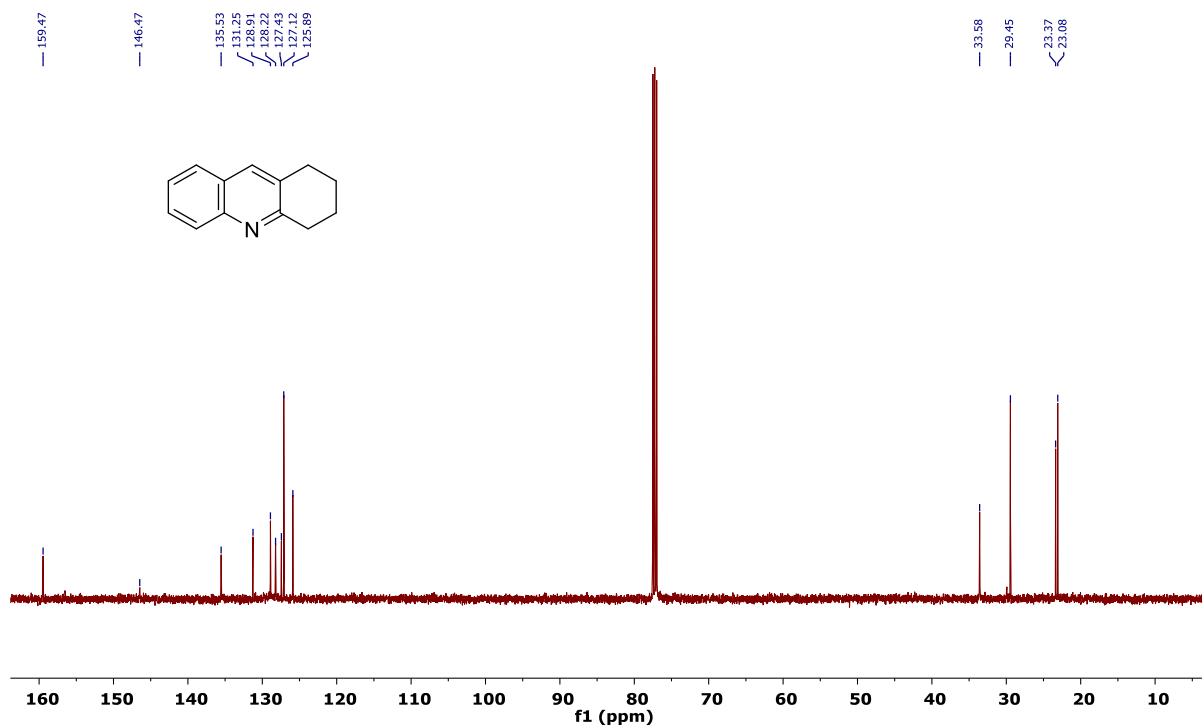
8.27
8.26
8.04
8.03
7.95
7.94
7.94
7.83
7.76
7.76
7.75
7.74
7.74
7.73
7.57
7.56
7.55
7.55
7.55
7.54
7.53
7.39
7.38
7.37
7.37
7.36
7.35
7.35
7.12
7.10
7.10
6.98
6.98
6.95
6.95

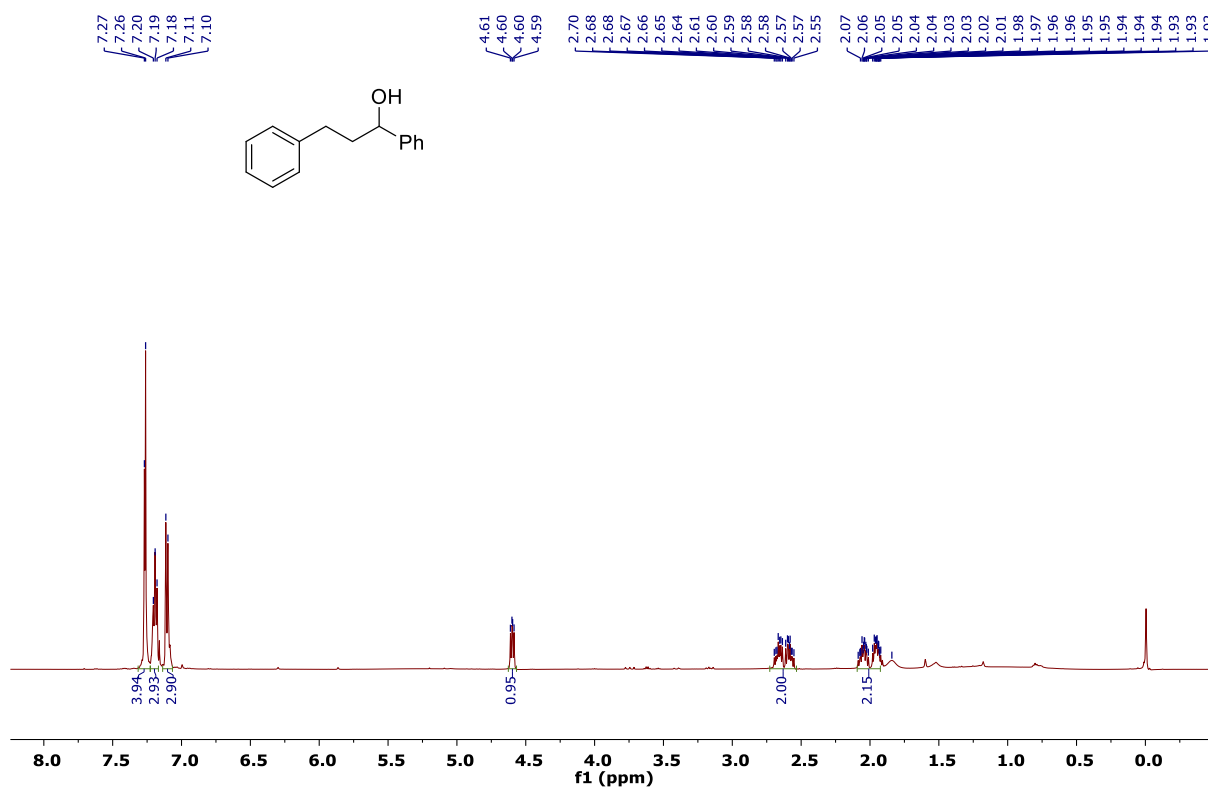
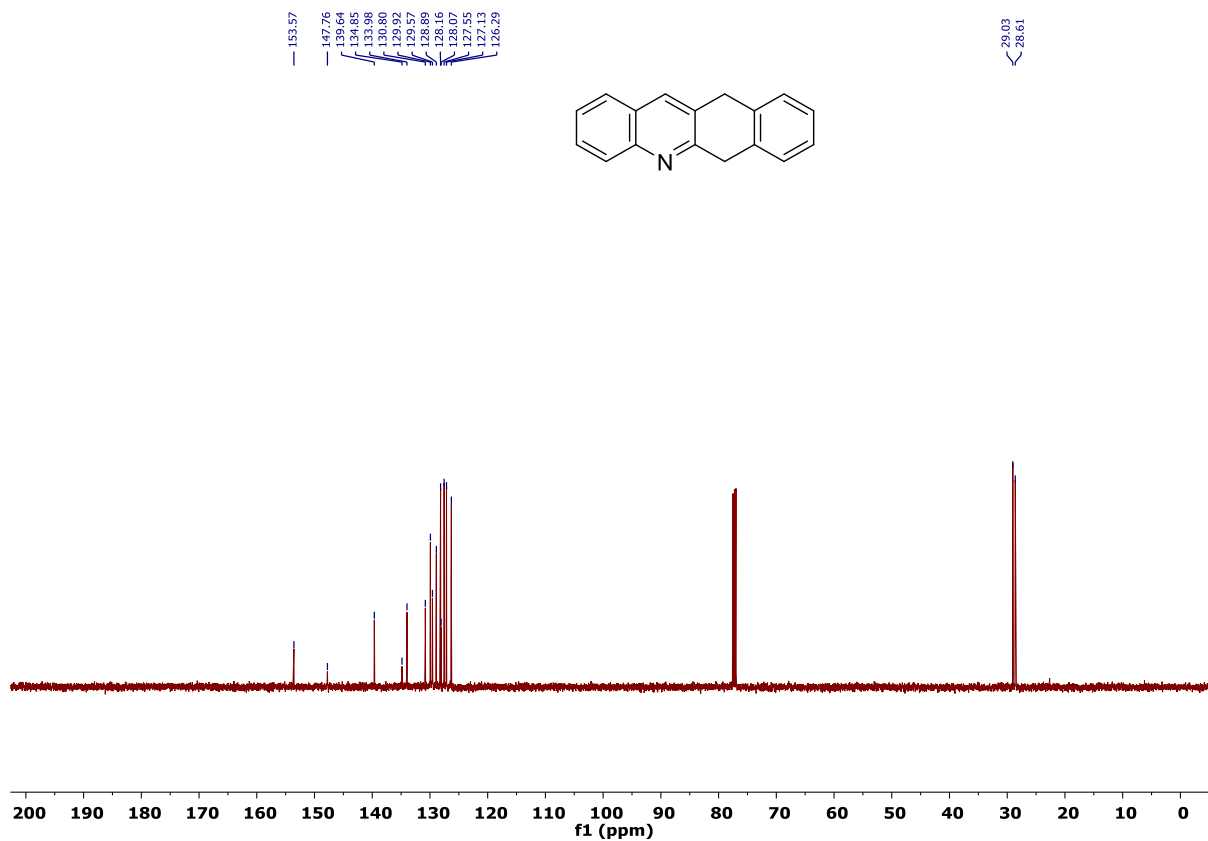


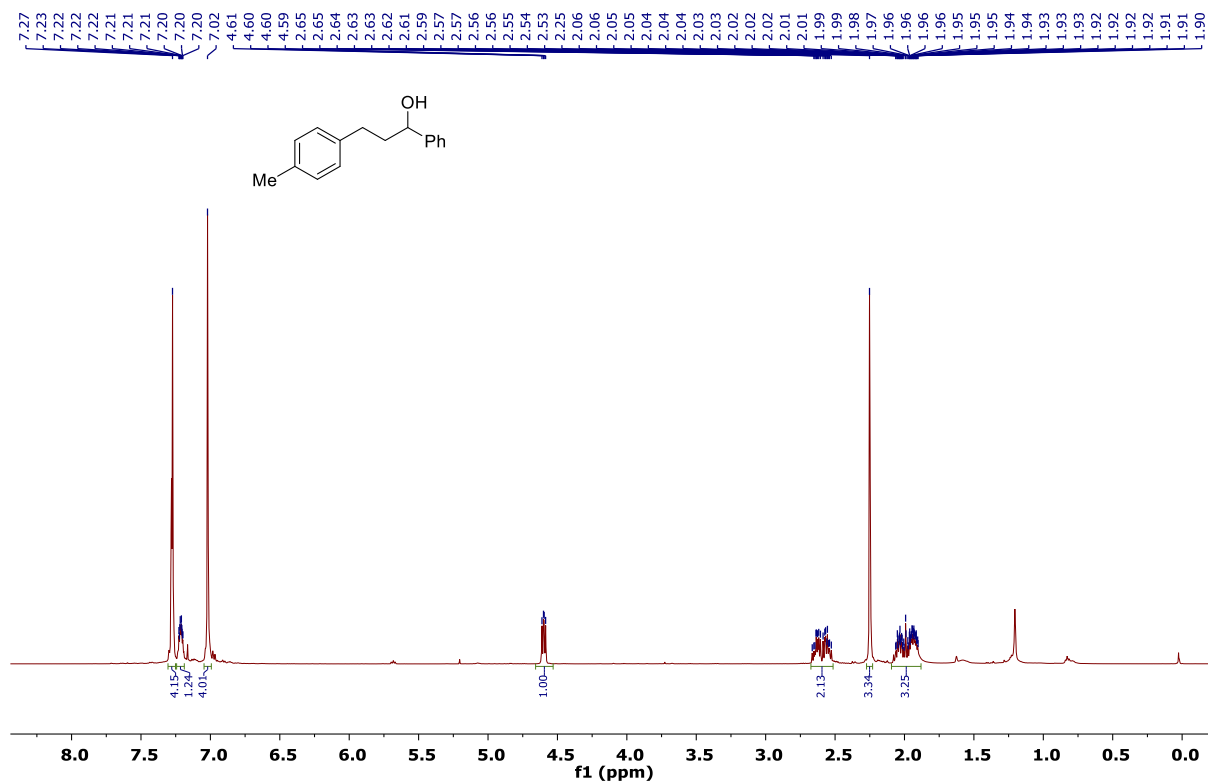
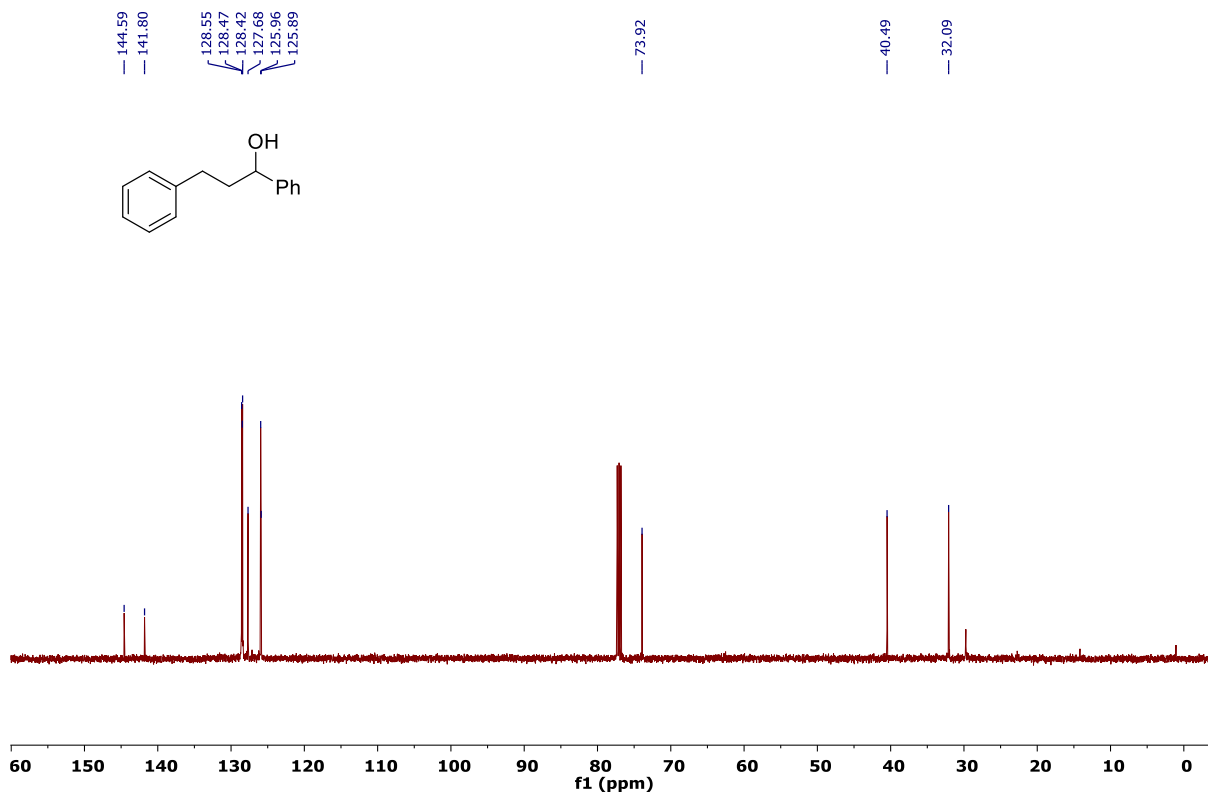
161.46
158.38
145.06
138.21
132.59
131.02
128.02
127.93
127.45
127.20
127.00
119.39
119.25
119.16
117.76

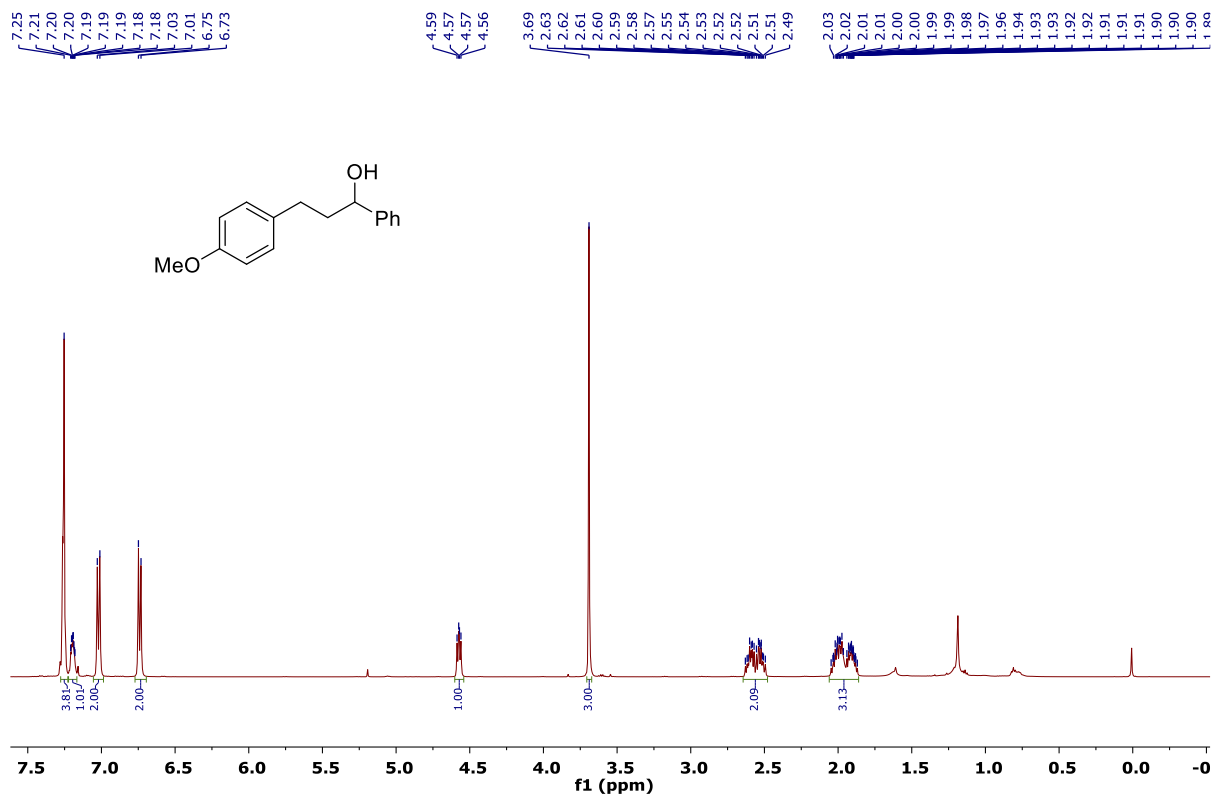
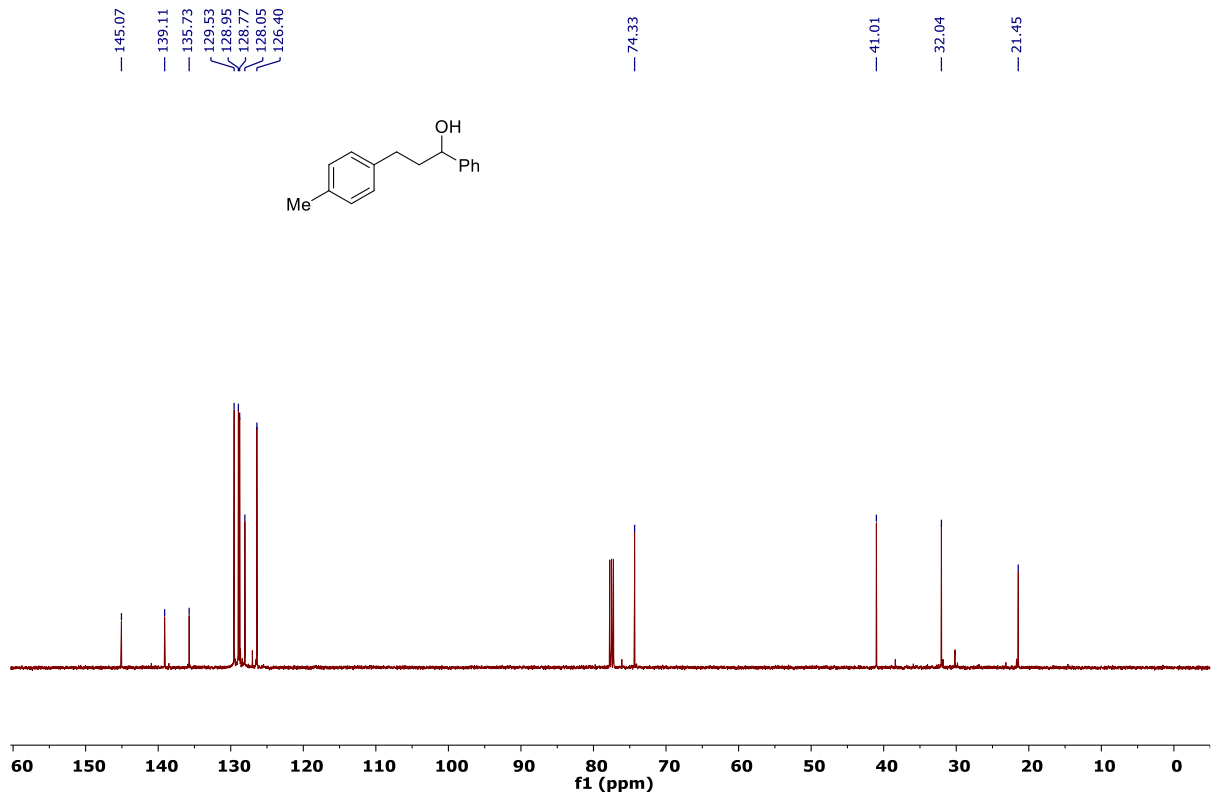


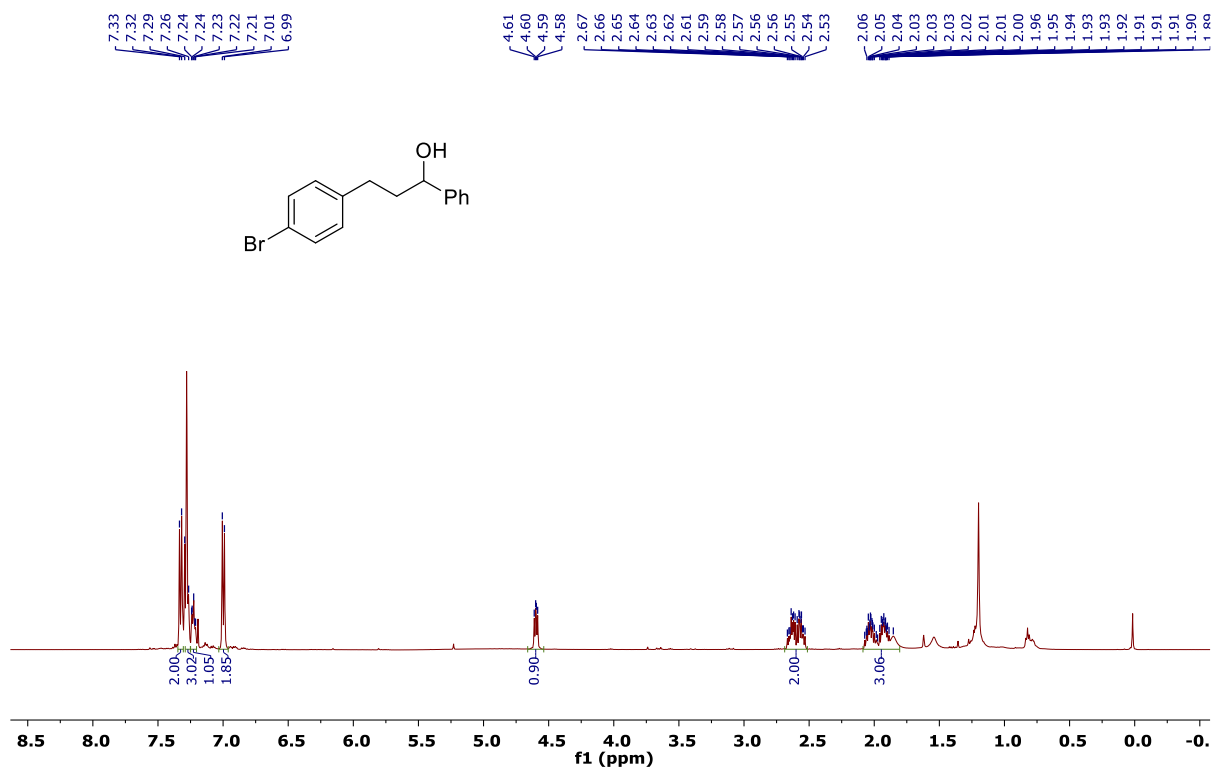
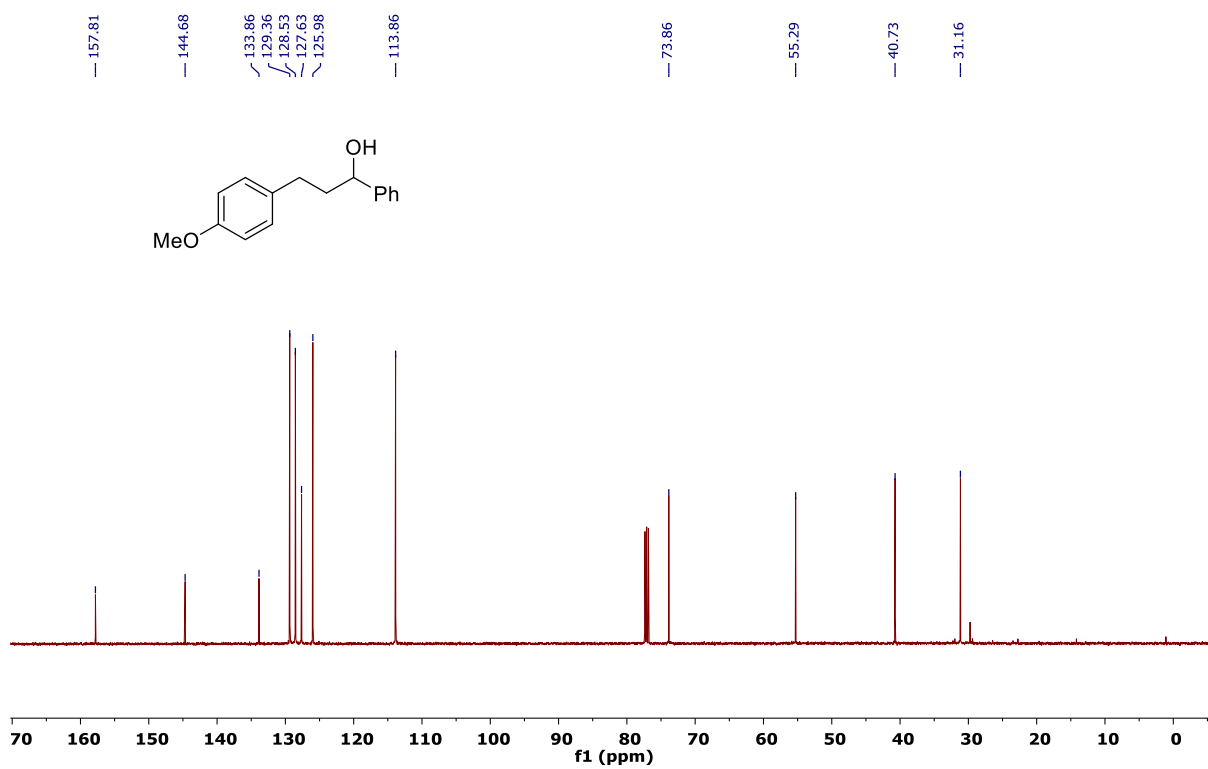


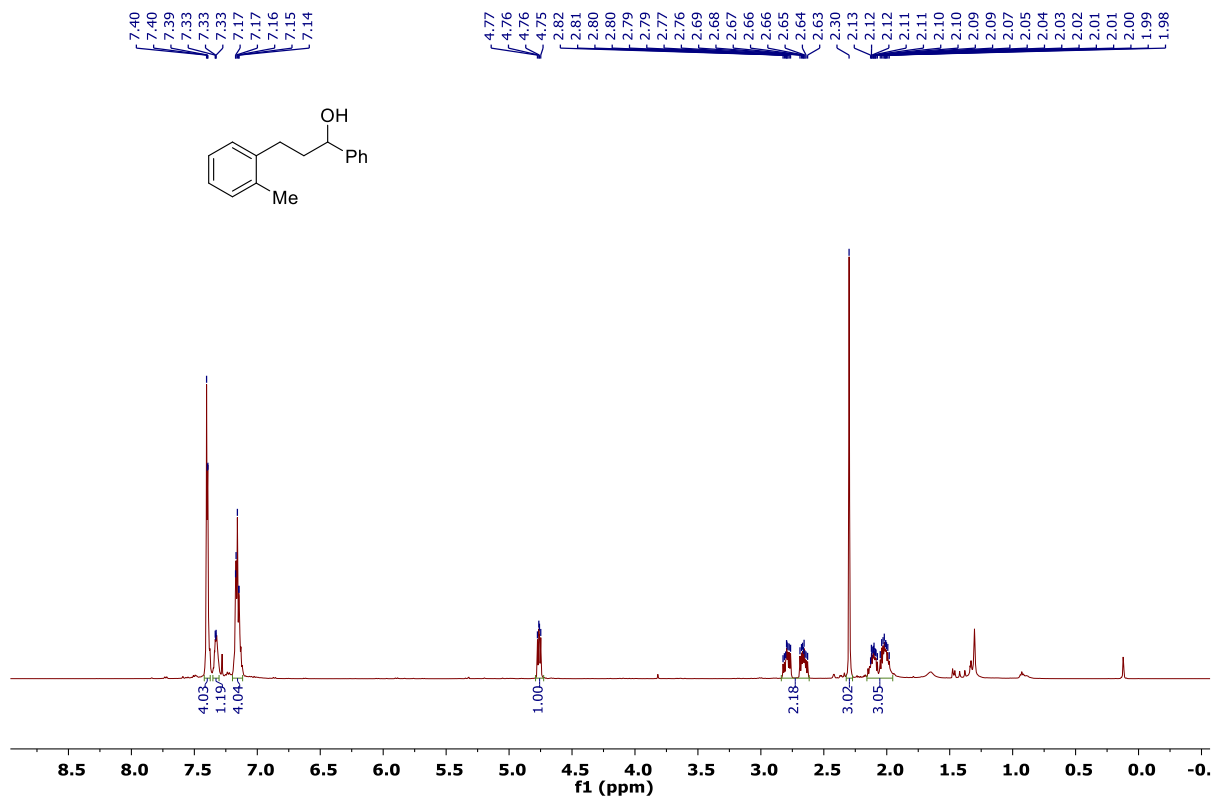
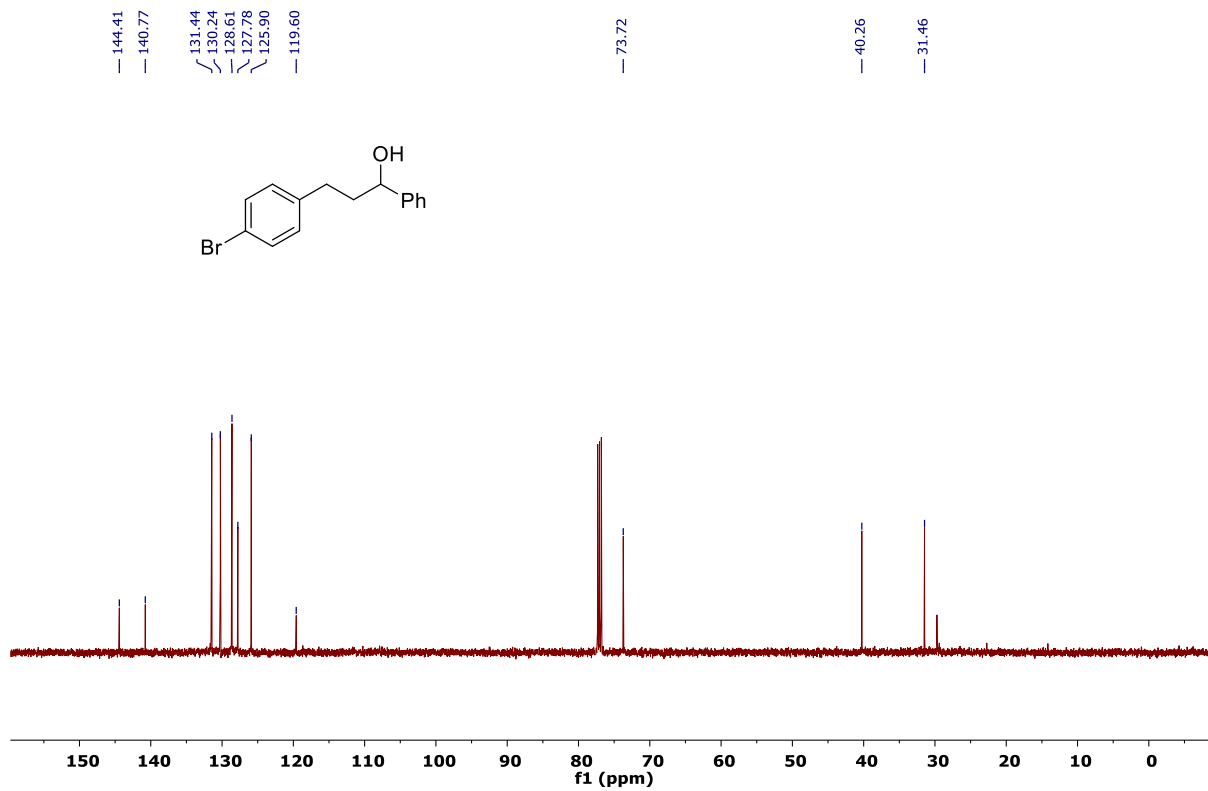


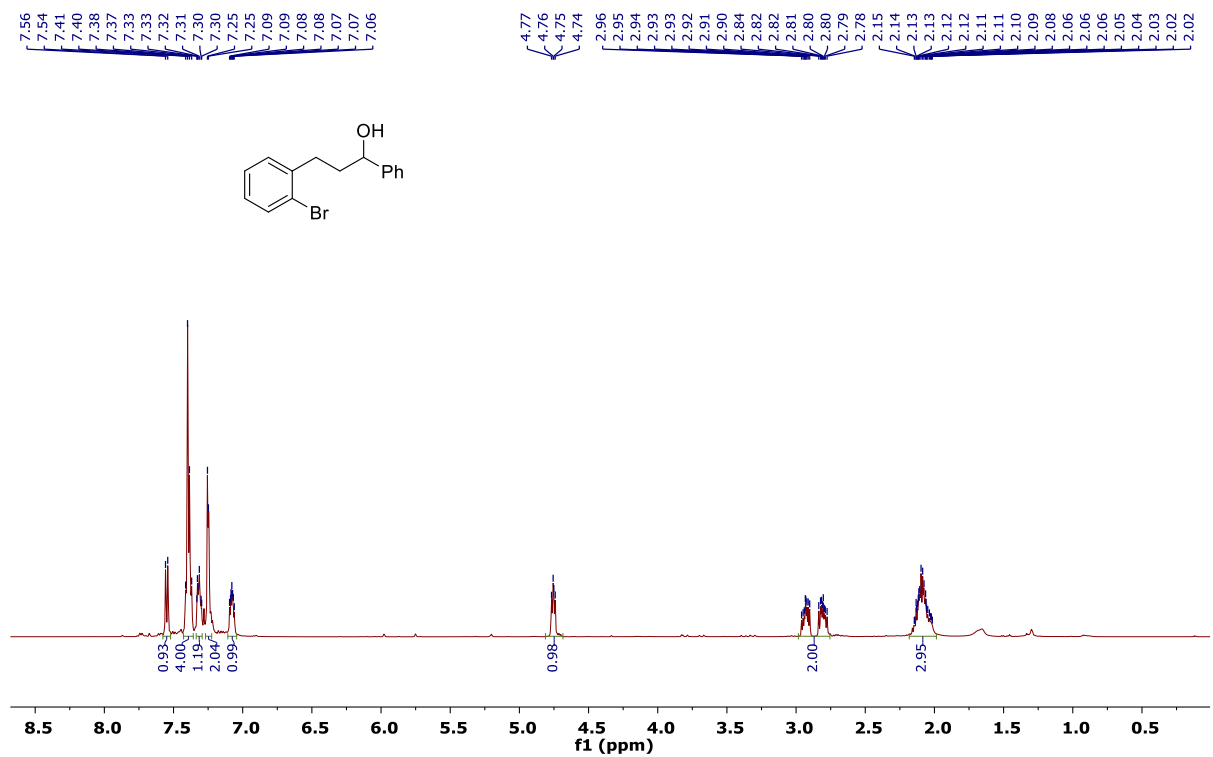
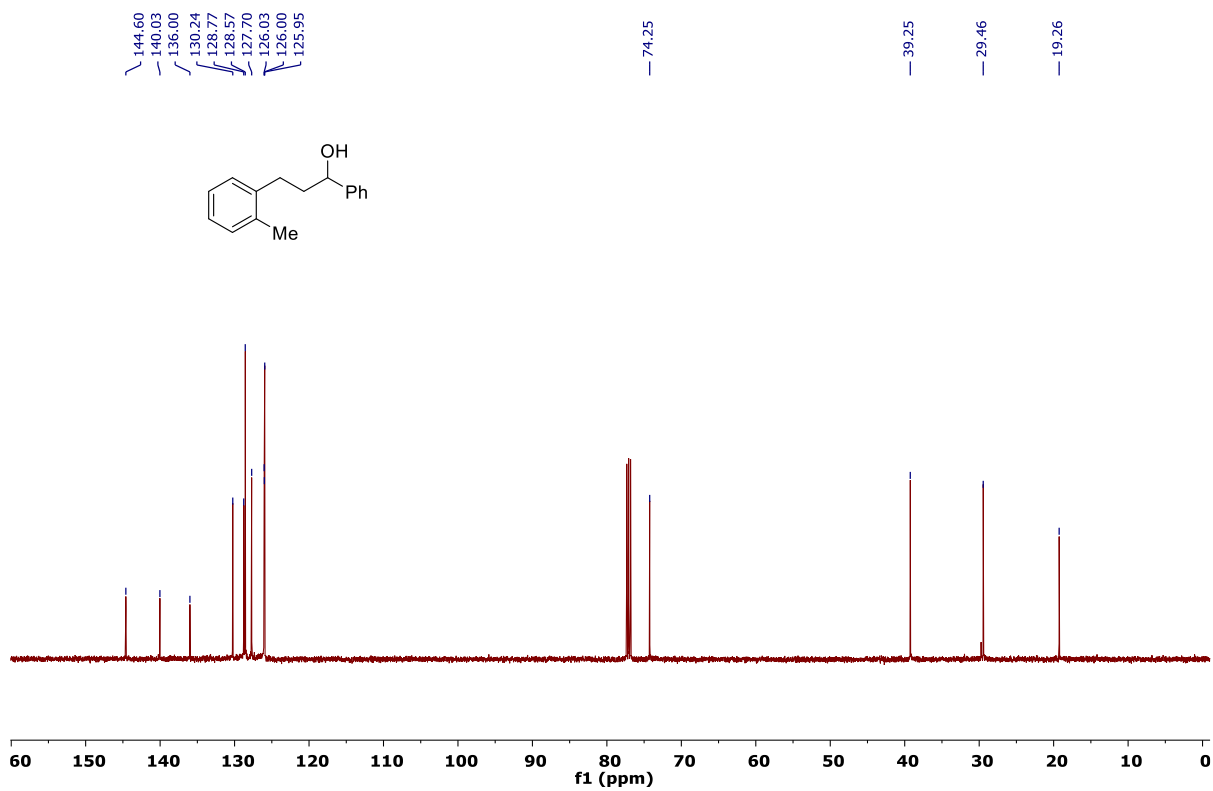


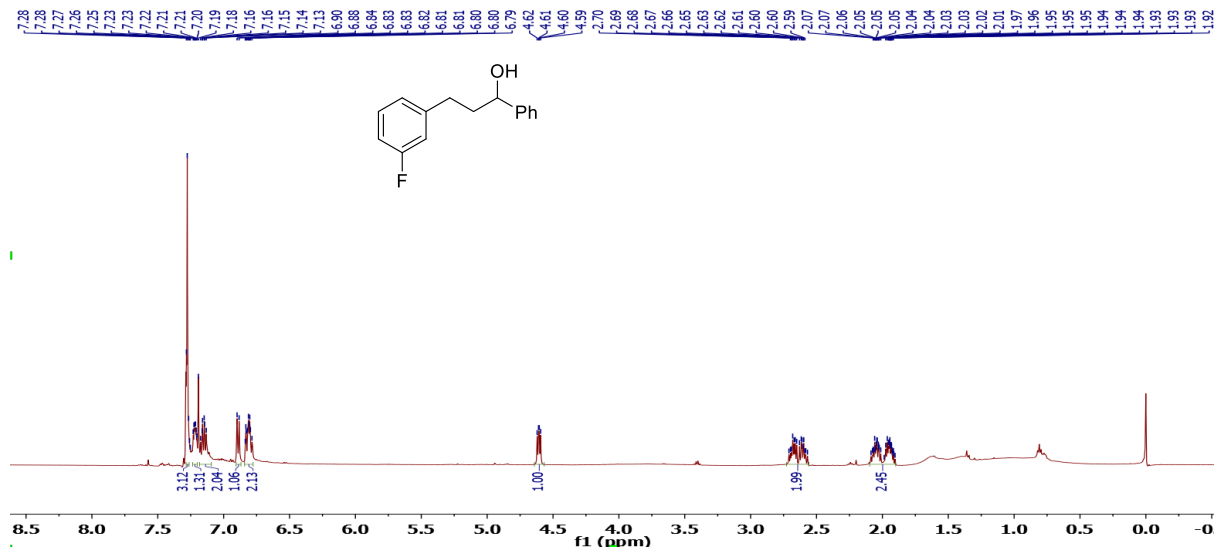
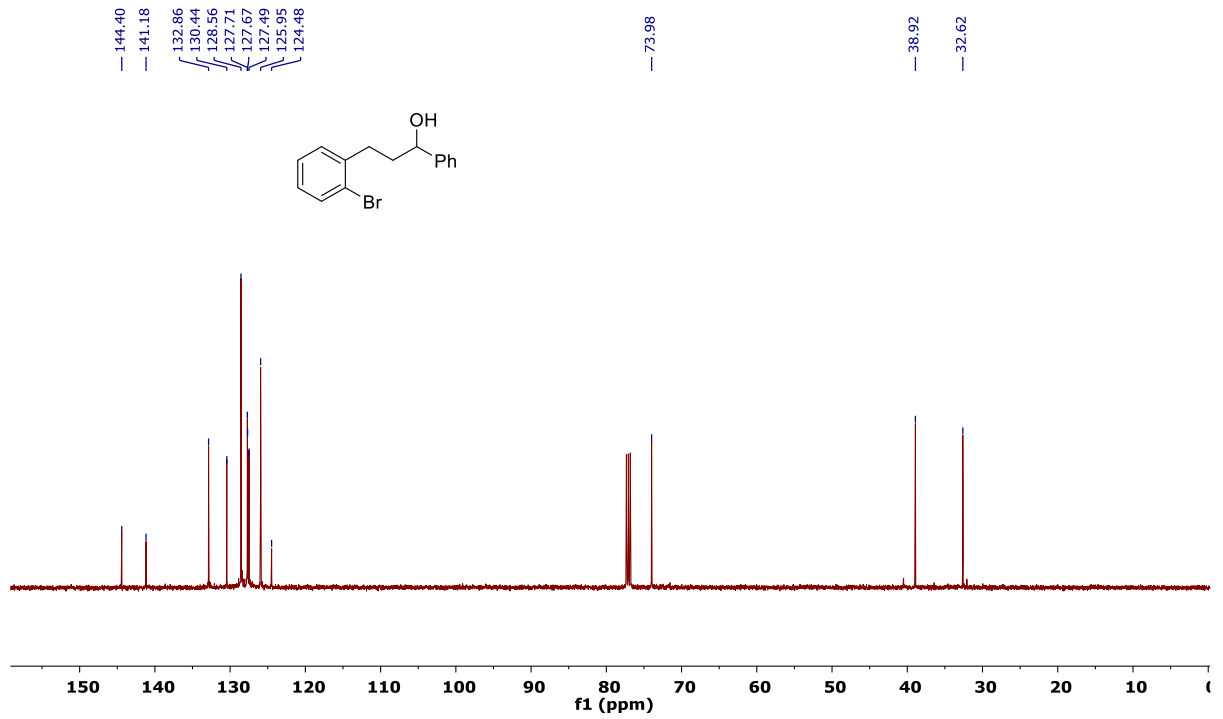




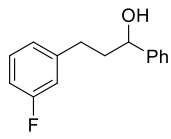




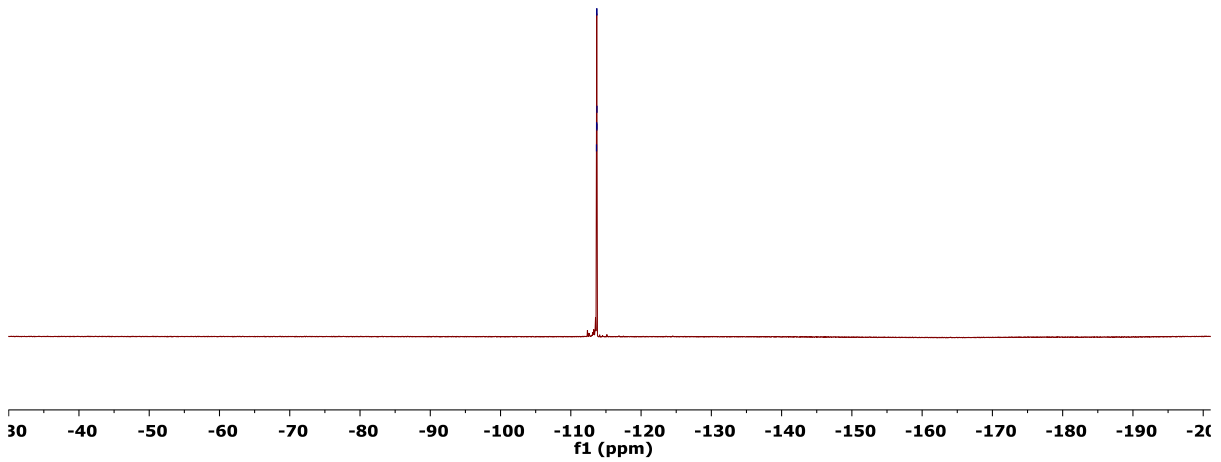




¹⁹F NMR



-113.66
-113.67
-113.68
-113.70
-113.71
-113.72

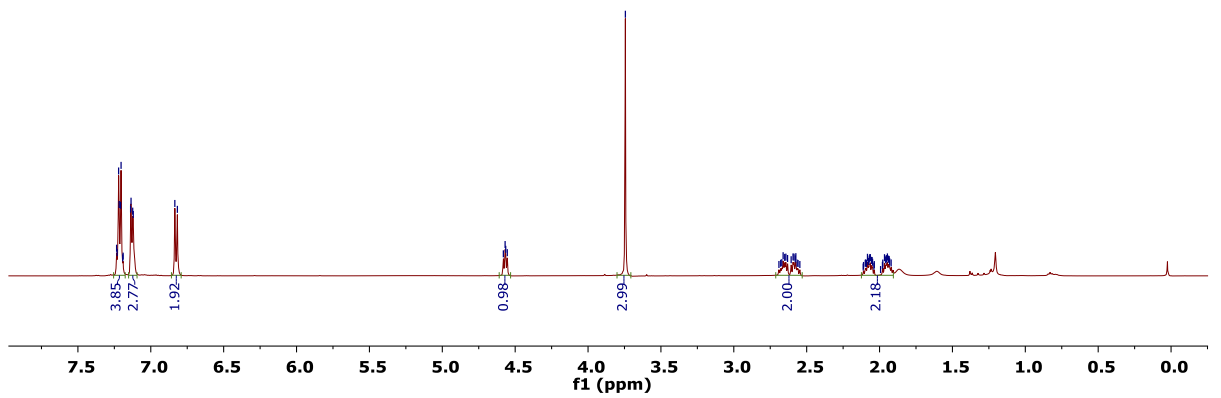
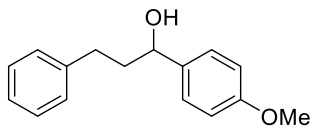


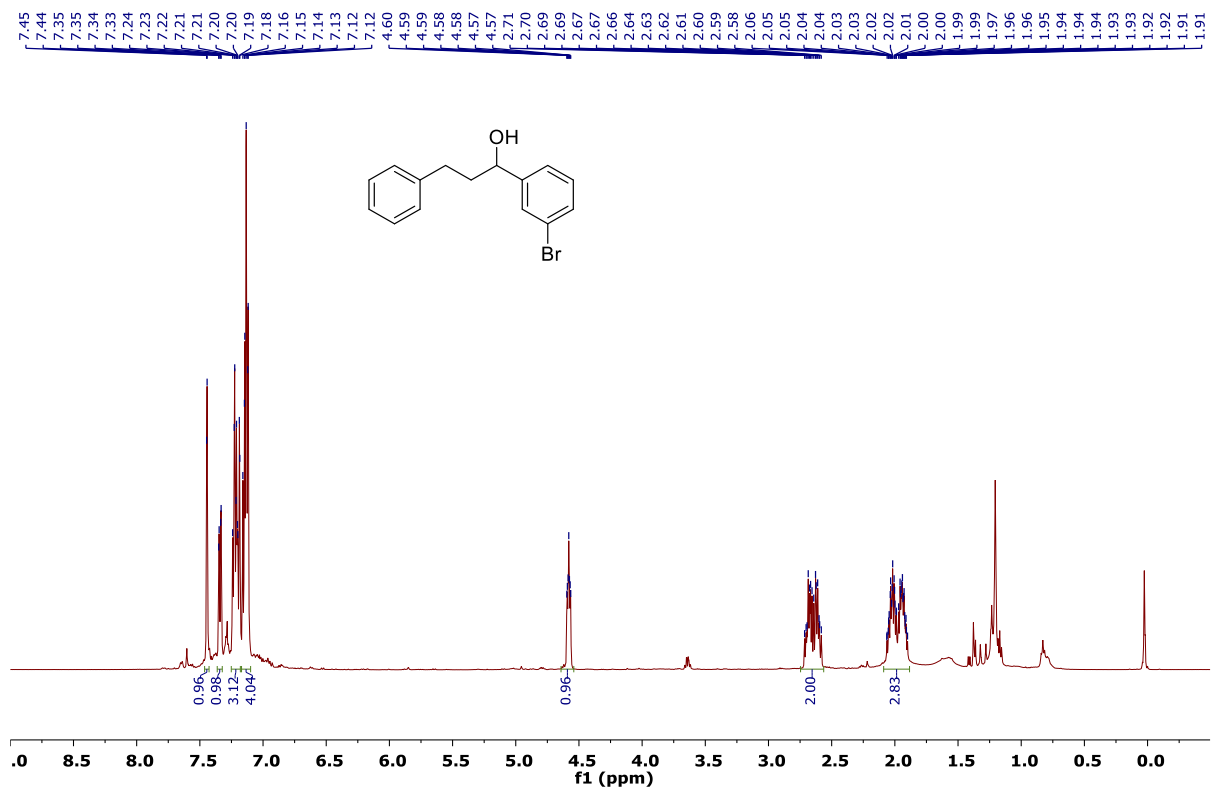
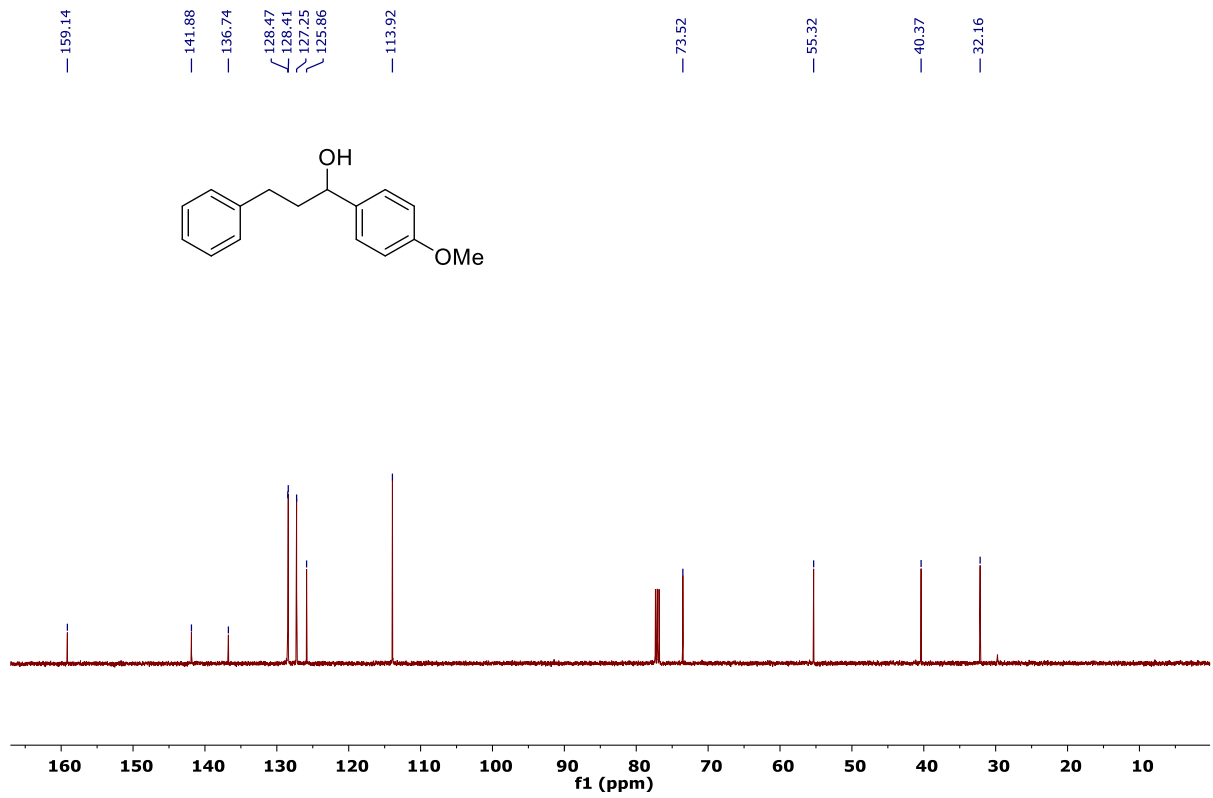
7.22
7.22
7.21
7.20
7.14
7.14
7.12
6.84
6.82

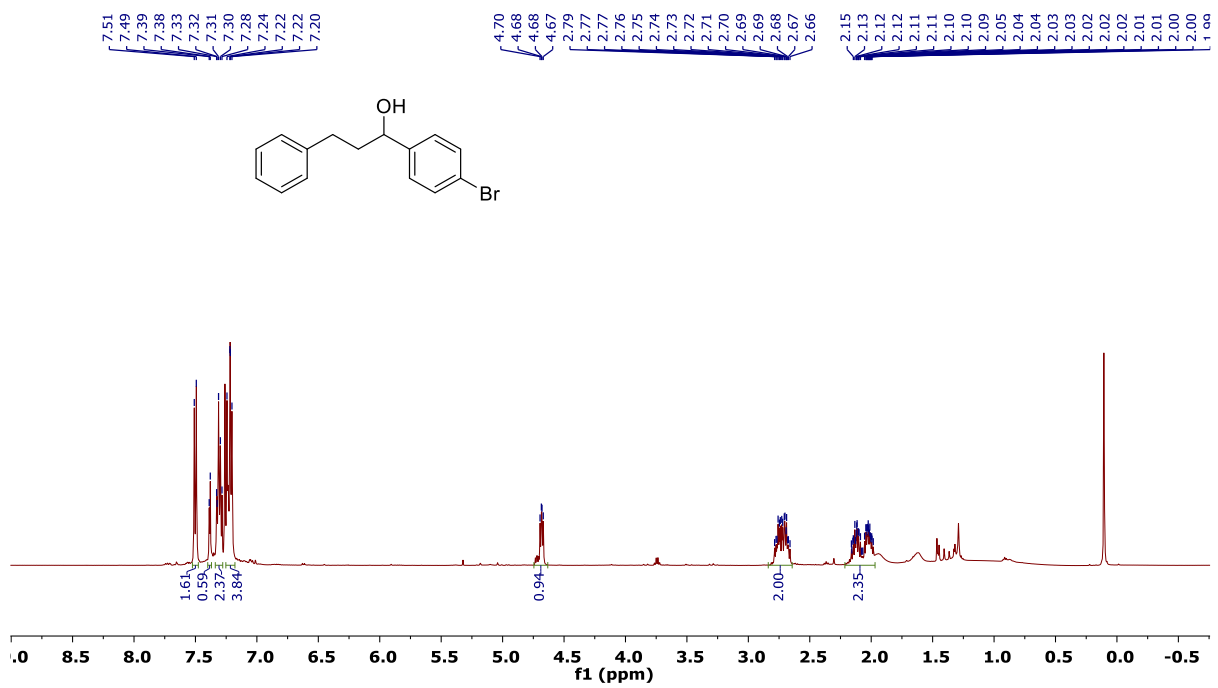
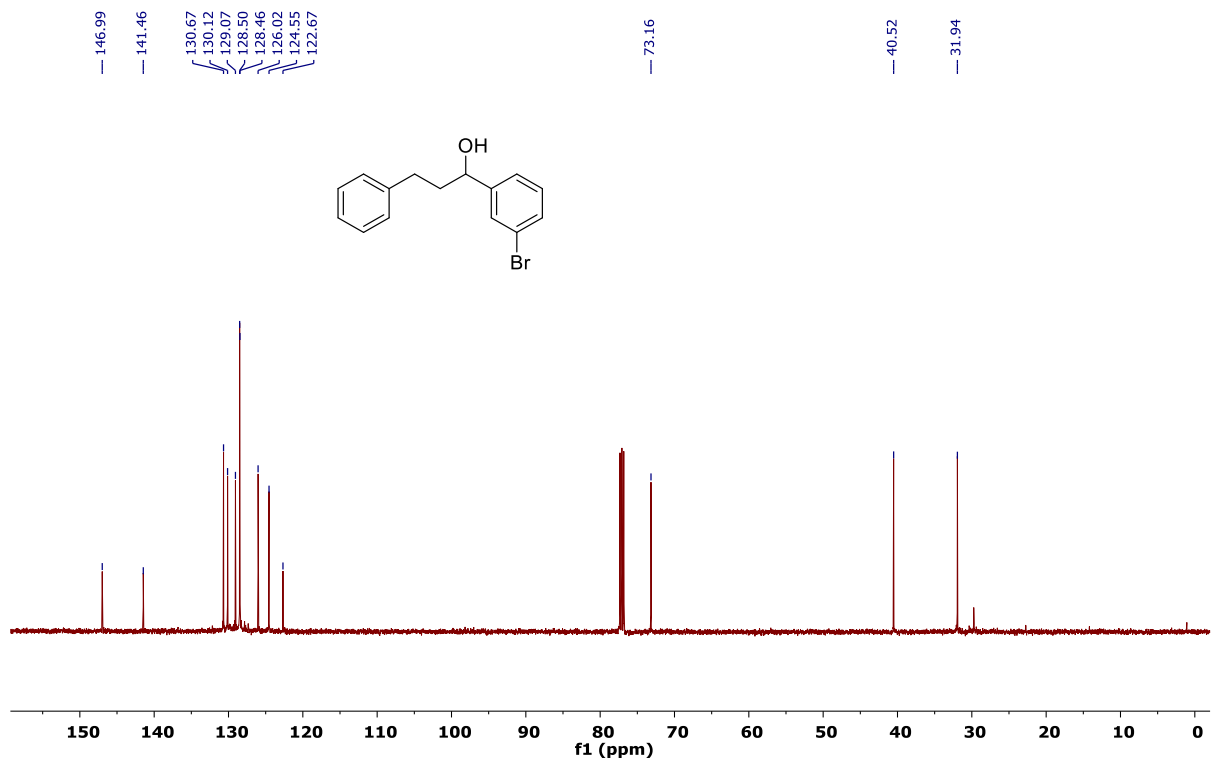
4.58
4.57
4.56
4.55

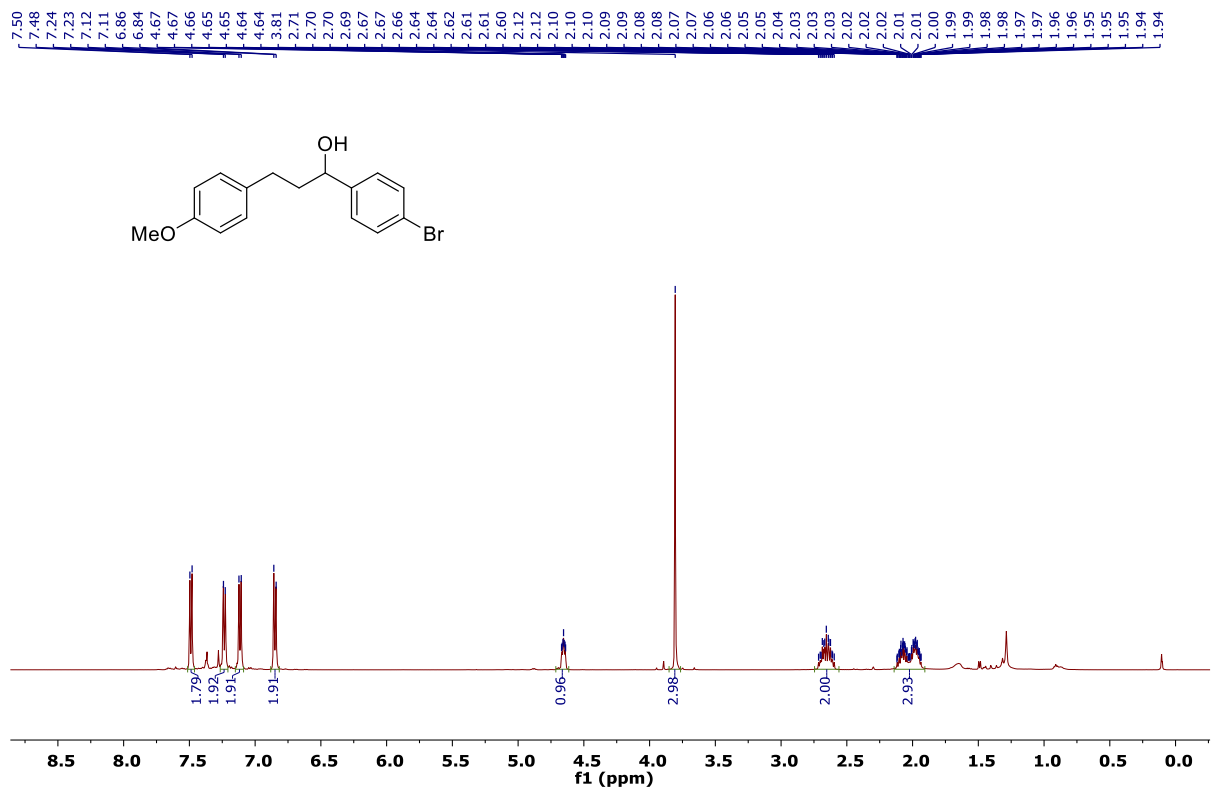
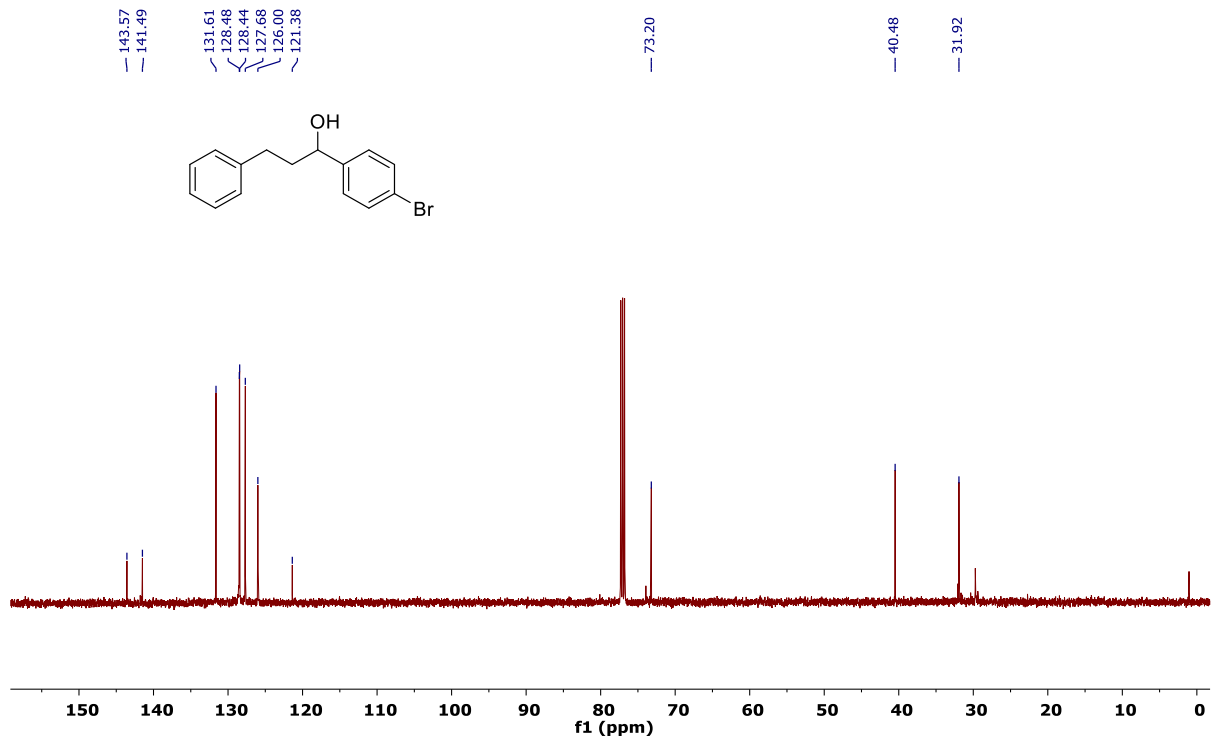
3.74
2.69
2.68
2.67
2.66
2.65
2.64
2.63
2.61
2.59
2.59
2.58
2.57
2.57
2.56
2.55

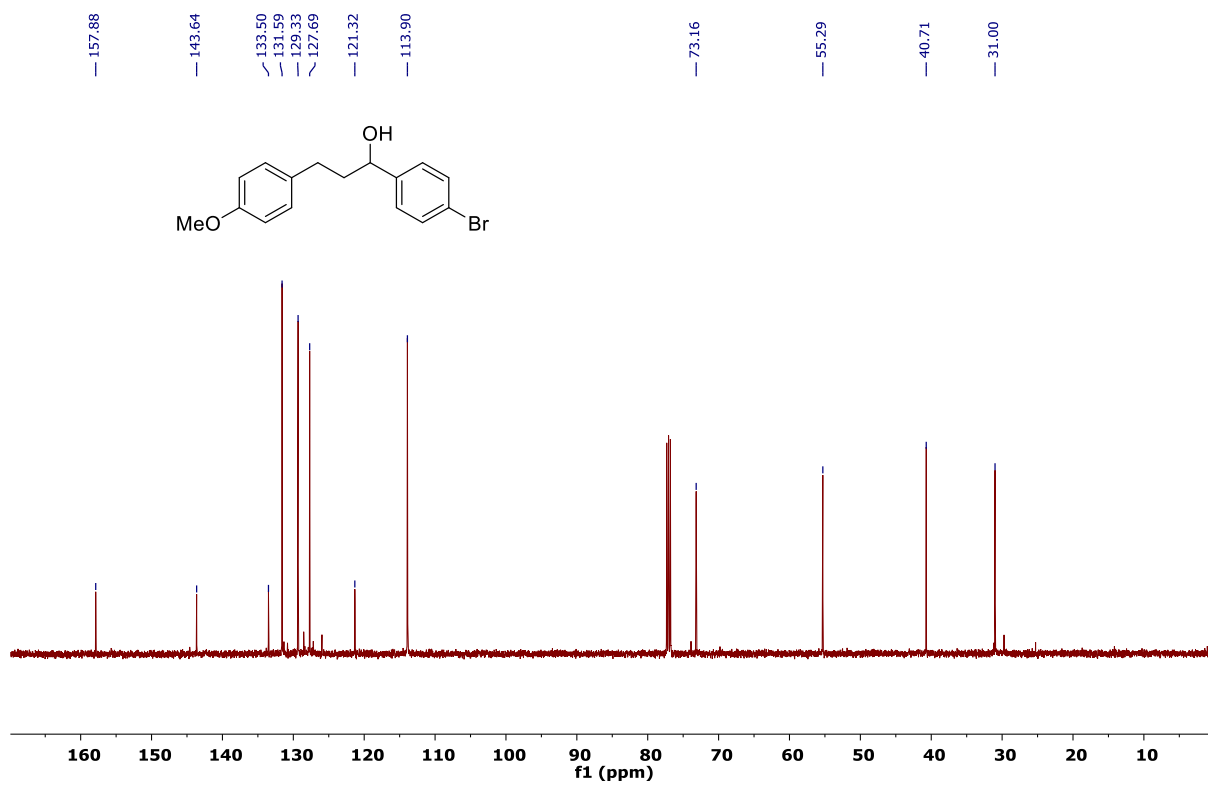
2.10
2.09
2.08
2.08
2.07
2.07
2.06
2.06
2.05
2.05
1.98
1.97
1.96
1.95
1.95
1.94
1.94
1.93
1.93
1.92











References:

- [1] P. Hu, Y.-L. Qiao, J.-Q. Wang and G.-X. Jin, *Organometallics*, **2012**, *31*, 3241–3247.
- [2] R. Favela-Mendoza, E. Rufino-Felipe, H. Valdés, R. A. Toscano, S. Hernandez-Ortega and D. Morales-Morales, *Inorganica Chim. Acta*, **2020**, *512*, 119920.
- [3] M. Uchida, C. H. Cortney, K. Bustos, E. Manzo, E. Sauls, J. Bouchard, R. Fukazawa and V. V. Krishnan, *J. Chem. Educ.*, **2023**, *100*, 4822–4827.
- [4] G. A. Bain and J. F. Berry, *J. Chem. Educ.*, **2008**, *85*, 532.
- [5] D.-W. Tan, H.-X. Li, D.-L. Zhu, H.-Y. Li, D. J. Young, J.-L. Yao and J.-P. Lang, *Org. Lett.*, 2018, **20**, 608–611.
- [6] G. Zhang, J. Wu, H. Zeng, S. Zhang, Z. Yin and S. Zheng, *Org. Lett.*, 2017, **19**, 1080–1083.
- [7] S. Chakraborty, P. Daw, Y. Ben David and D. Milstein, *ACS catalysis*, 2018, **8**, 10300–10305.
- [8] S. Chakraborty, S. Jalwal, A. Regina, V. Atreya and M. Paranjothy, *Dalton Trans.*.
- [9] R. Sharma, A. Mondal, A. Samanta, N. Biswas, B. Das and D. Srimani, *Adv. Synth. Catal.*, 2022, **364**, 2429–2437.
- [10] D. Wei, B. Feng, Q. Chen, W. Yue, Y. Wang and Z. Peng, *Org. Chem. Front.*, 2024, 10.1039/D3QO01862H.
- [11] D. Bhattacharyya, B. K. Sarmah, S. Nandi, H. K. Srivastava and A. Das, *Org. Lett.*, 2021, **23**, 869–875.
- [12] P. Liu, R. Liang, L. Lu, Z. Yu and F. Li, *J. Org. Chem.*, 2017, **82**, 1943–1950.
- [13] X. Xu, Y. Ai, R. Wang, L. Liu, J. Yang and F. Li, *J. Catal.*, 2021, **395**, 340–349.
- [14] C. S. Cho, B. T. Kim, T.-J. Kim and S. C. Shim, *Chem. Commun.*, 2001, 2576–2577.
- [15] D. Bhattacharyya, P. Adhikari, K. Deori and A. Das, *Catal. Sci. Technol.*, 2022, **12**, 5695–5702.
- [16] S. Parua, R. Sikari, S. Sinha, S. Das, G. Chakraborty and N. D. Paul, *Org. Biomol. Chem.*, 2018, **16**, 274–284.
- [17] S. W. Maurya, H. Sagir, M. D. Ansari and I. R. Siddiqui, *ChemistrySelect*, 2021, **6**, 13601–13608.
- [18] G. Joshi, A. A. Wani, S. Sharma, P. Bhutani, P. V. Bharatam, A. T. Paul and R. Kumar, *ACS Omega*, 2018, **3**, 18783–18790.
- [19] H. Vander Mierde, P. Van Der Voort, D. De Vos and F. Verpoort, *Eur J Org Chem*, 2008, **2008**, 1625–1631.
- [20] A. Banik, P. Datta and S. K. Mandal, *Org. Lett.*, 2023, **25**, 1305–1309.
- [21] J. Liu, W. Li, Y. Li, Y. Liu and Z. Ke, *Chem. Asian J.*, 2021, **16**, 3124–3128.
- [22] T. W. Ng, G. Liao, K. K. Lau, H. Pan and Y. Zhao, *Angew. Chem.*, 2020, **132**, 11480–11485.
- [23] H. Narjinari, N. Tanwar, L. Kathuria, R. V. Jasra and A. Kumar, *Catal. Sci. Technol.*, 2022, **12**, 4753–4762.
- [24] R. Babu, M. Subaramanian, S. P. Midya and E. Balaraman, *Org. Lett.*, 2021, **23**, 3320–3325.