Supporting Information

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1. Hydroxyl radical formation measured by 2-hydroxyterephthalic acid fluorescence

Fluorescence spectra were recorded every 5-10 min, during 40-60 minutes of irradiation. The TAOH formation curves for the experiments are shown in Figure S16-19. By applying the TAOH calibration curves it was possible to follow the formation of TAOH with time. Linear regressions were performed on these curves to obtain the rate of TAOH formation derived from the slope. Charbouillot et al. $(2011)^1$ presented a yield of OH formation from TAOH dependent on the pH, which allowed the calculation of OH formation as seen in Equation S1.

$$
\Gamma_{TAOH} = (0.0248 \pm 0.0059) \, pH + (0.046 \pm 0.035) \tag{S1}
$$

The pH in the solutions 5.7 and 6.7 in the VL experiments. In nitrate solutions, the pH was between 6.4 and 7.0, while for hydrogen peroxide the range was from 6.2 to 6.4. These differences in pH was considered by means of Equation S1, that was used to calculate the yield TAOH from each individual experiment. The slope of the linear fit of [TAOH] vs time of the different OH source concentrations was divided by the yield of OH formation (Eq. S1) , leading to figures S9- 11. These plot shows the OH formation rate as a function of the compounds. From the linear regression of this curve, the slope (and its variability) is found, resulting in a value for the OH formation rate depending on the species concentration (in units of M s⁻¹ M(comp)⁻¹). This method for estimating the OH formation rates was applied to vanillin and 4HB, as well as two other sources of OH in the aqueous phase, NO_3^- and H_2O_2 .

2. Aerosol samples

The sampling dates of the samples were $29/11/2021$ to $02/12/2021$ for the sample referred to as 021221, and 13/12/2021 to 14/12/2021 for the sample referred to as 141221. The dissolved organic carbon of the samples was analyzed as described in the main text and the resulting values were

73.42 and 64.01 mgC L-1 for the 021221 and 141221 samples, respectively. Blank filter extractions are regularly performed to track the extraction performance. Eventually, some simple carboxylic acids are reaching mg levels, while most complex organics were below the detection limit at tens of µg levels. This contamination represents a very small fraction of the organics present during our experiments.

3. Degradation of the triplet state probe, TMP

The triplet state steady-state concentration was estimated by using TMP with UPLC/UV detection. The degradation of TMP gave a first-order rate constant $(k_{obs, TMP})$, that similarly to singlet oxygen can be applied in calculation the steady-state concentration. Although TMP is common probe for triplet states, it can also be degraded by singlet oxygen and OH radicals. Therefore, $k_{obs,TMP}$ should be corrected for the influence of singlet oxygen, OH radicals, and also direct photolysis ($k_{\text{blank,TMP}}$). The steady-state concentrations of singlet oxygen and OH radicals in the two samples were estimated from previous experiments with the same sample extracts but slightly varying conditions (20 \degree C, 20 mL reactor). The singlet oxygen concentrations were determined to be $(5.17 \pm 0.23) \times 10^{-13}$ and $(5.59 \pm 0.26) \times 10^{-13}$ M for samples 02122021B and 141221B, respectively. The steady-state OH radical concentrations were calculated as described in section S4.

An average second-order rate constant between TMP and triplet states $(k_{3C^*,TMP})$ from a variety of sources (listed in the Table S1) was calculated to be $(1.46 \pm 1.17) \cdot 10^9$ M⁻¹ s⁻¹. Using this second-order rate constant of TMP with triplets, along with the second-order rate constant for TMP with singlet oxygen ($k_{1O2,TMP}$) and with OH radicals ($k_{OH,TMP}$), the steady-state triplet state concentrations are calculated as:

$$
\left[\begin{array}{c} {}^{3}C^{*} \end{array}\right]_{ss} = \frac{k_{obs, TMP} - (k_{blank, TMP} + k_{1OZ, TMP} [{}^{1}O_{2}]_{ss} + k_{OH, TMP} [OH]_{ss})}{k_{3C*, TMP}} \tag{S2}
$$

The value for $k_{OH,TMP}$ was an average value of the reaction of OH radicals with 2-methylphenol and 4-methylphenol², because no value for the rate constant of TMP and OH radicals was found.

When the steady-state triplet state concentrations of the photosensitizers were estimated, the concentrations were calculated as a simplified version of Equation S2, where the correction for singlet oxygen and OH radical interference with the probe was omitted.

Triplet state source	$k_{3C^*,TMP}(10^9 M^{-1} s^{-1})$	Refence
2-acetonaphthone	0.617 ± 0.016	Wenk 2013 ³
2-acetonaphthone	0.72 ± 0.01	Canonica 2000 ⁴
3-methoxyacetophenone	1.88 ± 0.21	Wenk 2013 ³
3-methoxyacetophenone	2.6 ± 0.3	Canonica 2000 ⁴
4-carboxybenzophenone	3.3 ± 2.4	McCabe & Arnold 2017 ⁵
Benzophenone	5.1 ± 0.9	Canonica 2000 ⁴
CDOM (Canal)	3.0 ± 1.1	al Housari 2009 ⁶
CDOM (Vaccarès)	4.8 ± 4.4	al Housari 2009 ⁶
DOM (Great Dismal Swamp)	0.77 ± 0.01	Erickson 2018 ⁷
DOM (Lake Bradford)	0.84 ± 0.03	Erickson 2018 ⁷
Fulvic acid (Pony Lake)	1.26 ± 0.02	Erickson 2018 ⁷
Fulvic acid (Suwannee River)	0.54 ± 0.01	Erickson 2018 ⁷
Humic acid (Suwannee River)	0.75 ± 0.03	Erickson 2018 ⁷
NOM (Mississippi River)	0.90 ± 0.02	Erickson 2018 ⁷
NOM (Suwannee River)	0.63 ± 0.02	Erickson 2018 ⁷
Watershed (CMH07)	1.4 ± 0.4	McCabe & Arnold 2017 ⁵

Table S1. Second-order rate constants between triplet states and TMP. CDOM: chromophoric dissolved organic matter.

3.1 Probe control experiments

Various probe molecules, including TMP, sorbic acid, but also other dienes have been used to indirectly assess and quantify different triplet states. It is important to note that these probes, such as TMP and sorbic acids, quantify distinct triplet pools, differing in quantity and reactive nature8. TMP is directly oxidized by organic matter triples states while dienes primarily engage through energy transfer, targeting high-energy triplets. When energy transfers to a diene occur, they generate triplet excited dienes, then probably secondary reactivities in water. We opted for TMP in our study due to its stability in aqueous solution and simple and rapid detectability through

High-Performance Liquid Chromatography (HPLC) analysis in complex media.

Figure S1. TMP degradation (in pct) by photolysis (black), nitrate (red), humic acid (blue), and humic acid and isopropanol (green).

In order to assess the direct oxidation of TMP by hydroxyl radicals, we conducted additional experiments where the concentration of TMP was monitored in the presence of nitrate ions $(NO₃)$) under photolysis (serving as a source of hydroxyl radicals) and humic acids (HA), known to generate excited states with and without isopropanol (ISO), a recognized hydroxyl radical scavenger (Figure S1).

Figure S1 illustrates that, after 3 hours of irradiation, there was no significant enhancement in TMP degradation, with approximately 20% of TMP undergoing degradation via direct photolysis. However, in the presence of HA, there was a marked increase in TMP degradation (which is expected due to the formation and then reactivity with excited states). Notably, the use of isopropanol did not alter this degradation, suggesting that isopropanol had no direct effect on the reactivity of the triplet states with TMP.

To test the effect of singlet molecular oxygen, an experiment with TMP+VL was performed with nitrogen to remove the molecular oxygen from the solution. N_2 was bubbled through the solution for 40 min (in the dark) before the start of the irradiation. Hereafter, the irradiation was started while nitrogen continued to be bubbled through the solution. An experiment with a standard TMP+VL solution was performed on the same day. Both experiments were performed in the glass reactor at 15 °C, similar to the experiments containing TMP described in the main text. The concentration of both compounds was 100 µM. The TMP was detected by UHPLC/UV detection (described in section S3.2). The results are shown in Figure S2.

Figure S2. Degradation of TMP (100 µM) with VL (100 µM) in a standard solution (containing O2, black) and in a solution that had been bubbled with nitrogen (without O2, orange).

The results of these experiments showed that the degradation of TMP was faster in the N_2 (without O_2) solution than in the standard solution (with O_2). This indicates that singlet oxygen does not have a significant effect on the degradation of TMP. It is possible that the faster degradation of TMP in an N_2 solution is due to less quenching of the triplet states by O_2 (i.e. because most O_2 is removed from the solution).

In this study, aerosol samples were collected and extracted as described in the main text. These samples were extracted for brown carbon, and hereby represent an aerosol constituent responsible for a large amount of triplet state formation. The degradation of TMP by the BrC extracts was compared to that of VL. This gives an indication of whether VL can be regarded as a proxy for aerosol components.

3.2 Experimental set-up

In this section, a sketch of the experimental set-up for the degradation of TMP is shown. This setup was also applied for analyzing the TAOH formation from the aerosol samples as described in the following section (S3.2).

3.3 Method for U(H)PLC/UV and UPLC/FLR analysis

The samples for the experiments regarding the degradation of TMP (described in the main text) were analyzed in the UPLC with UV detection. The method consisted of a flow of 0.6 ml/min with 95 % water (with 1% H₃PO₄) and 5 % methanol (ultrapure, UPLC grade) for the initial 0-4.5 min. Hereafter, the solvents were changed to have 5 % water (with 1% H₃PO₄) and 95 % methanol until 5.1 min, where after the method returned to 95 % water (with 1% H₃PO₄) and 5 % methanol (ending at 6 min).

UHPLC/UV detection for the TMP control experiments (with VL in O_2 or N_2) was performed with an ACQUITY UPLC HSS T3 by WATERS column (100 mm x 2.1 mm, 1.8µm particle size). The flow through the UHPLC was 0.3 mL/min and consisted of solvent A: H₂O with 0.1% formic acid, and/or solvent B: ACN with 0.1% formic acid. The gradient was as follows; 0-2 min: 1% solvent B and 99% solvent A, 2-13 min: the solvent changed gradually from 1 to 100% B (with solvent A as the remaining), 13-15 min: the solvent remained 100% B, 15-15.1 min: the solvent gradually changed from 100 to 1% solvent B (with solvent A as the remaining), 15.1-22 min: the solvent remained 1% B and 99% solvent A.

The OH steady-state concentration was determined for the two aerosol samples by applying terephthalic acid as a probe and analyzing the TAOH formation by UPLC with fluorescence detection. In the experiments the sample concentration was 10 mgC/L, and the TA concentration was 500 µM. A 50 ml reactor was used and the samples were irradiated for 60 min, using the same set-up as for the TMP experiments. The TA concentration was higher in these experiments compared to those performed with the other OH sources (ie. VL , H_2O_2 , and NO_3^-) because we needed a TAOH concentration that was sufficient for detection by UPLC/FLR. The method for this analysis had a flow of 0.6 ml/min and began with 95 % water (with 1% H₃PO₄) and 5 %

methanol (ultrapure, UPLC grade) from 0 min to 4.5 min. From 4.5 to 5 min, the solvent changed to 40 % water (with 1% H₃PO₄) and 60 % methanol and remained at this ratio until 5.1 min, where after it was changed back to 95 % water (with 1% H₃PO₄) and 5 % methanol. The method had a duration of 6 min.

4. OH steady-state concentrations

The calculations of the OH steady-state concentrations of in the various experiments were performed by the following equation, put forth by Lallement et al.⁹;

$$
[OH]_{ss} = \frac{k_{obs}}{\gamma_{TAOH} \cdot k_{TA,OH} \cdot [TA]} (S3)
$$

Where k_{obs} is the first-order rate constant for the formation of TAOH, γ_{TAOH} is the yield of formation of TAOH, *kTA,OH* is the second-order rate constant between TA and OH radicals, and [TA] is the concentration of TA.

The steady-state concentrations of OH for the applied sources of OH radicals can be found in Figures S3-6. The TAOH formation for all species is shown in Figures S16-20.

The [OH]ss was also determined in blank experiments with TA as described in section 3.2 (SI).

Table S2. OH steady-state concentrations. For nitrate anions, hydrogen peroxide, and VL, a range of concentrations are giving, indicating the minimum and the maximum concentration.

Compound	$\left[OH\right]_{ss}(M)$		
NO ₃	$(1.10 \pm 0.62) \cdot 10^{-16}$ to $(4.81 \pm 1.6) \cdot 10^{-15}$		
H_2O_2	$(5.15 \pm 0.97) \cdot 10^{-16}$ to $(7.31 \pm 1.5) \cdot 10^{-15}$		
VL	$(8.67 \pm 1.9) \cdot 10^{-17}$ to $(6.20 \pm 2.6) \cdot 10^{-16}$		
4HB	$(8.34 \pm 0.27) \cdot 10^{-17}$ to $(1.48 \pm 0.064) \cdot 10^{-16}$		

 \mathbb{L}

5. Figures

F**igure S3.** OH formation rate (black squares) and OH steady-state concentrations (red circles) for nitrate anions.

Figure S4. OH formation rate (black squares) and OH steady-state concentrations (red circles) for hydrogen peroxide.

Figure S5. OH formation rate (black squares) and OH steady-state concentrations (red circles) for VL.

Figure S6. OH formation rate (black squares) and OH steady-state concentrations (red circles) for 4HB.

Figure S7. Emission of the Xenon lamp (dotted black line, right y-axis), and molar absorption coefficients (left y-axis) of VL (blue, full line), 4HB (pink, full line), and TA (brown, full line).

Figure S8. Excitation emission matrix (EEM) spectra of the samples 021221 (a) and 141221 (b), and the three photosensitizers, 2-IC (c), 4-BBA (d), and VL (e). The color bars denote the fluorescence intensity (a.u.). All spectra were performed with slits of 5 nm for both excitation and emission. Note the difference in intensity scale of the fluorescence; the top row (a-c) share a scale with a maximum of 100, while the bottom row (d-e) share a scale with a maximum of 8.

Figure S9. OH formation rates from nitrate anions. A linear fit is applied and the resulting slope is $(4.16 \pm 0.51) \times 10^{-7}$ M s⁻¹ M(NO₃⁻)⁻¹. The points are results of duplicate experiments.

Figure S10. OH formation rate of H_2O_2 . A linear fit is applied and the resulting slope (1.64 \pm 0.06)

 \times 10⁻⁶M s⁻¹ M(H₂O₂)⁻¹. The points are results of duplicate experiments.

Figure S11. OH formation rate of VL. The left figure shows the whole plot (each point is the result

of triplicate experiments), while the right figure shows the linear fit that is applied to points from 5×10^{-5} to 3.75×10^{-4} M. The slope of the linear regression is 9.00×10^{-6} M s⁻¹ M(VL)⁻¹.

Figure S12. OH formation rate of 4HB. The left figure shows the whole plot, while the right figure shows the linear fit that is applied to points from 5×10^{-5} to 3.75×10^{-4} M. The slope of the linear regression is $(9.21 \pm 0.28) \times 10^{-7}$ M s⁻¹ M(4HB)⁻¹. The points applied in the linear regression is the results of triplicate experiments, while the three points at higher concentrations are from duplicate experiments.

6. OH formation rates in literature

In Table S3, literature values for OH formation rates are shown in various environments.

Table S3. OH formation rates from literature studies on aerosols or cloud/fog.

Reference	Type	Location	OH formation rate $(M s^{-1})$
Anastasio et Jordan 2004 Arctic 10	particles		Nunavut, Canada $2.8 \cdot 10^{-12}$ to $3.0 \cdot 10^{-9}$
<i>Anastasio et McGregor</i> Fog 2001 ¹¹		California, USA	2.4.10 ⁻¹⁰ to 1.9.10 ⁻⁹

7. Literature study of aerosol and cloud/fog concentrations of NO_3 , H_2O_2 , HULIS, and **phenols/nitrophenols**

In Table 1 in the main text, concentrations of the relevant species in aerosols and cloud/fog conditions are shown. These values originate from a literature study, in which concentrations of nitrate, hydrogen peroxide, and HULIS/(N)Ph concentrations were extracted from studies regarding measurements of aerosols or cloud/fog composition.

7.1 Method for " μ g m⁻³ to mol L⁻¹" conversion

Here, a method for converting the concentrations of considered species from μ g m⁻³ to mol L⁻¹ is shown.

[comp]
$$
\left(\frac{mol}{L}\right) = \frac{\rho \cdot [comp] \cdot 10^{-9}}{[particles] \cdot M}
$$
 (S4)

Where ρ is the density of particles in units of μg (particles) m^{-3} (particles), and is estimated to be 1.49 g cm-3 (average from eight articles reporting density of particles 20–27). *[particles]* is the average concentration of particulate matter (unit: μ g (particles) $m^{-3}(air)$) reported in correlation with the concentration of the given compound, *[comp]* (unit: $\mu g m^{-3}(air)$). *M* is the molar mass of the compound in units of g mol⁻¹.

7.2 Example of conversion

An example of the conversion of concentration is shown in the equation below.

*Peng et al.*²⁸ reports an annual nitrate concentration of 5.6 μ g m⁻³ and an annual particle concentration of 56.2 μ g m⁻³. Here, the conversion into mol L⁻¹ is shown.

$$
[NO3-] = \frac{1.49 \cdot 10^{12} \mu g m^{-3} \cdot 5.6 \mu g m^{-3} \cdot 10^{-9}}{56.2 \mu g m^{-3} \cdot 62.0 g mol^{-1}} = 2.39 mol L^{-1}
$$

7.3 Reported concentrations in literature

In the following tables, the result of the literature study is shown. The tables provide the references, the size and type of particles or cloud/fog, the location, the reported concentration of the species and particulate matter, as well as the species concentration converted into mol L^{-1} . This study was mainly focused on $PM_{2.5}$, though a few entries of other particle sizes were also included to have enough data.

7.3.1 Concentrations in aerosols

Table S4. Concentrations of nitrate anions in aerosols.

Table S5. Concentrations of hydrogen peroxide in aerosols.

Table S6. Concentrations of HULIS in aerosols. ^aThe values were reported in HULIS µg m⁻³ and was converted to μ gC m⁻³ by a mass-to-carbon ratio of HULIS of 1.9, that the authors (Lin et al., 2010) estimated from literature.

7.3.2 Concentrations in cloud/fog

Table S9. Concentrations of phenols and nitrophenols in cloud/fog conditions.

8. Additional figures

Figure S13. Boxplot OH of formation rates in rural clouds using (N)Ph concentrations for triplet state approximations.

Figure S14. Boxplot of OH formation rates in biomass burning aerosols HULIS concentrations were used for triplet state approximations. ^aIt was not possible to find H_2O_2 concentrations in biomass burning aerosols, and therefore an average of all urban concentrations was applied.

Figure S15. Zoom of the aerosol HULIS/NO₃ region of Figure 4 in the main text.

Figure S16. TAOH formation from H₂O₂ at various concentrations.

Figure S17. TAOH formation from NO₃ with various concentrations.

Figure S18. TAOH formation from 4HB with various concentrations.

Figure S19. The TAOH formation from various concentrations of VL, and from a TA control experiment.

Figure S20. TAOH formation from the two aerosol samples (021221 and 141221), as well as a blank TA experiment. All three experiments had a TA concentration of 500 µM.

9. Decay of triplet state source

Figure S21. UV-Vis spectra that show the decay of VL (left) and 4HB (right) during the experiments with concentrations of 0.037 mM. Both had approximately linear decays with firstorder rate constants of 7.59×10^{-5} s⁻¹ and 1.80×10^{-4} s⁻¹ for VL and 4HB, respectively.

Figure S22. UV-vis spectra showing the formation of light absorbing species in the visible range from both the VL (left) and 4HB (right) experiments with concentrations of 0.037 mM.

10. Quantum yields of OH formation

Quantum yields were calculated as previously proposed by Bianco et al. ¹⁵, according to Equation (S5).

$$
\Phi_{280 - 600nm} = \frac{R_{OH}}{R_{abs}} (S5)
$$

Here R_{OH} is the rate of OH formation from the compounds, and R_{abs} is the rate of light absorption by the solution and is calculated by the following (Equation S6) 89 ,

$$
R_{abs} = 2.303 \times 10^3 \times \Sigma (\alpha_{\lambda} \times I_{\lambda} \times \Delta \lambda) \text{ (S6)}
$$

where α_{λ} is the absorbance of the sample at the given wavelength (cm⁻¹), I_{λ} is the irradiance of the lamp at the given wavelength (mol_{photons} s⁻¹ cm⁻² nm⁻¹), and $\Delta\lambda$ is the wavelength interval (nm) . 2.3 is used for base conversion, and $10³$ is applied for unit conversion.

The screening factors are calculated by 90 ,

$$
sf = \frac{\Sigma(1 - 10^{-(\ell \times \alpha_{\lambda})} \times I_{\lambda})}{\Sigma(2.303 \times \ell \times \alpha_{\lambda} \times I_{\lambda})}
$$
 (S7)

where *l* is the path length of the light through the sample.

Table S10. Quantum yields, rate of light absorption, and screening factors for VL and 4HB $([VL]_0 = 37 \mu M$, $[4HB]_0 = 37 \mu M$, at the beginning (t = 0 min) and end (t = 40 min) of the experiments and from the aerosol samples 021221 and 141221 (t = 0 min). Values from VL and 4HB are from duplicate experiments. The errors are from one standard deviation.

	$\Phi_{(280-600nm)}$	R_{abs} (mol _{photons} L^{-1} s ⁻¹)	screening factor
VL $(t = 0$ min)	$(1.04 \pm 0.020) \times 10^{-4}$	$(2.27 \pm 0.097) \times 10^{-6}$	0.80 ± 0.003
$4HB(t=0 min)$	$(1.12 \pm 0.29) \times 10^{-4}$	$(6.89 \pm 1.53) \times 10^{-7}$	0.77 ± 0.05
Sample	3.26×10^{-5}	2.98×10^{-6}	0.75
02122021			
Sample	4.39×10^{-5}	1.87×10^{-6}	0.81
14122021			

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