Singlet oxygen is not the main source of electrolyte degradation in lithium-oxygen batteries

Supplementary Information

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General information

Reagents and solvents. Tetraethylene glycol dimethyl ether (tetraglyme, \geq 99%) and dimethyl sulfoxide (DMSO) were purchased from Sigma-Aldrich and dried over 4 Å molecular sieves for at least three days before use (H₂O <10 ppm determined by Karl Fischer titration). Lithium bis(trifluoromethanesulfonyl)imide (LiTFSI, 99.99%, Sigma-Aldrich) was dried at 85 °C under vacuum overnight (>12 h). 9,10-Dimethylanthracene (DMA, Sigma-Aldrich, 99%) was purified by recrystallisation in ethanol, melted under constant argon flow followed by recrystallisation in ethanol. The purified powder was dried at 85 °C under vacuum overnight (>12 h). 5,10,15,20-Tetraphenyl-21*H*,23*H*-porphine (TPP), 4,5,6,7-tetrachloro-2',4',5',7'-tetraiodofluorescein disodium salt (rose bengal, RB) and 1,4-dicyanobenzene were purchased (Sigma-Aldrich) and used without further purification.

Nuclear magnetic resonance (NMR) spectroscopy details. The ¹H NMR spectroscopy details for samples presented in **Figure S2** are as follows: Deuterated benzene (C_6D_6) was used as the lock solvent in a 1:9 solvent: sample ratio. NMR experiments were performed on a Bruker AVIII 400 (u_0 (¹H) = 400.17 MHz) spectrometer equipped with 5 mm z-gradient broadband multinuclear probe. The T_1 was calculated using an inversion recovery sequence, and a 10-second recycle delay (>5 T_1) was used throughout.

Samples for quantitative NMR studies (**Figure S3**) were prepared using CD₃CN as the lock solvent with a 9:1 solvent: sample ratio. The 1D and 2D NMR experiments were performed on a Bruker AVIII 600 (u_0 (¹H) = 600.13 MHz, with a proton-optimised TCI HCN cryoprobe spectrometer). The T_1 was determined using an inversion recovery sequence, and a 30-second recycle delay (>5 T_1) was used throughout.

¹H and ¹⁹F NMR spectra for **Figures 3, S5 - S8** were acquired on a Bruker AV(III)400 HD or a Bruker AV(III)500 HD fitted with a 5 mm BBFO and 5 mm prodigy BBO probes, respectively. Deuterated chloroform (CDCl₃) was used as the lock solvent.

The chemical shifts in ¹H spectra were referenced to the residual (partially) non-deuterated solvent according to Fulmer et al.¹ ¹⁹F NMR spectra were referenced through the solvent lock (²H) signal according to the IUPAC-recommended secondary referencing method following Bruker protocols.² Processing of all NMR data was conducted with Bruker TopSpin or Mestrelab Research Mnova software suites.

Experimental

On-line Mass-Spectrometry Analysis

On-line MS (Thermofisher Prima BT process mass spectrometer) was used to monitor the reactivity of ${}^{1}O_{2}$ with the selected solvents and to quantify the CO₂ evolved during the analysis of carbonate and carboxylate formed due to degradation. For monitoring the reactivity of ${}^{1}O_{2}$, the gas outlet of the ${}^{3}O_{2}$ purged samples was connected to an on-line MS, and once a stable ${}^{3}O_{2}$ baseline was achieved, samples were irradiated to begin ${}^{1}O_{2}$ production. When irreversible reactions occur with ${}^{1}O_{2}$, the measured ${}^{3}O_{2}$ concentration decreases as O₂ has been removed from flow. It is assumed all ${}^{1}O_{2}$ returns to the ground state before reaching the MS due to its short half-life (10-100 µs).^{3,4}

To study the amount of carbonate and carboxylate groups generated on WEs, acid and Fenton's reagent treatments were used as previously described.⁵ For ¹³C samples, the electrodes were rinsed with DME and dried under vacuum. Samples were connected to the on-line MS and Ar was used as the carrier gas. Once m/z 44 (CO₂) and m/z 45 (¹³CO₂) signals stabilised, phosphoric acid was added and after the baseline stabilised for the second time, Fenton's reagent was added to decompose all the carbonate and carboxylate species, accordingly.

Carbonate analysis - Acid treatment

2 M H₃PO₄ is added to the electrodes to decompose all present inorganic carbonates to CO₂ *i.e.* Li₂CO₃ to CO₂.⁵

Carboxylate analysis – Fenton's reagent treatment

Fenton's reagent⁶ was used to decompose carboxylates into CO₂. The reaction for Fenton's reagent generates radicals such as OH[•] and HO₂[•] which can attack electron deficient carbonyl sites such as in acetate and formate, releasing CO₂. 0.5 M FeSO₄ in 2 M H₃PO₃ and H₂O₂ (30%) was added to the electrodes after all CO₂ evolution from carbonate analysis had ceased.

Singlet Oxygen (¹O₂) Generation for On-line MS Experiments

Photocatalytic generation of ${}^{1}O_{2}$ was achieved using RB or TPP (<10 µM) with 530 nm or 660 nm light sources, respectively. The solutions were saturated with dry ${}^{3}O_{2}$ by continued sparging throughout the experiments. The experiments were either conducted in an Ar-filled glovebox or a cell connected to an on-line mass spectrometer. For experiments connected to the on-line mass-spectrometer, 20% O₂:80% Ar gas (dried over activated molecular sieves in the gas line) was bubbled into the samples whilst constantly stirring. 100% O₂ (N5.0 grade) was used for the experiments carried out in a glovebox. The chemical stability of the photosensitiser with the electrolyte and its electrochemical stability at 3.8 V are important selection criteria. Hence, RB was only used for chemical experiments, while TPP which is stable at higher oxidising potentials (**Figure S1**) was used in the electrochemical experiments.



Figure S1. Cyclic voltammogram of 7 μ M TPP in 0.1 M LiTFSI and tetraglyme. The CV was recorded with a planar Au working and counter electrodes and a fritted delithiated LiFePO₄ reference electrode at a scan rate of 100 mV s⁻¹.

¹O₂ oxidation of DMSO

DMSO was treated with ¹O₂ and O₂ consumption was monitored via online-mass spectrometry using the procedures described above (**Scheme S1**). Dimethyl sulfone (DMSO₂) was determined to be the major product, and the spectroscopic data (**Figure S2**) is consistent with those reported previously.⁷



Scheme S1. Photo-oxidation of DMSO via singlet oxygen.

FTIR spectra of the resulting mixture were recorded in an N₂-filled glovebox using a Thermo Fischer Nicolet iS50 FTIR Spectrometer. The spectra showed new peaks consistent with DMSO₂ (**Figure S2**). **FTIR** (cm⁻¹) 1142 (v_{as}SO₂), 763 (v_sCSC), 499 ($_{\delta}$ OSO), 465 ($_{\gamma \text{ or } \rho}$ SO₂).



Figure S2. **a**) ¹H NMR (400 MHz, C₆D₆, 298.1 K) spectra of the DMSO reaction solution before and after exposure to ¹O₂. **b**) FTIR spectra for DMSO (purple) and ¹O₂ treated DMSO (pink, DMSO₂ peaks highlighted.

Determination of ¹O₂ formation rate and comparison to the cell

The amount of ¹O₂ generation was confirmed by NMR spectroscopic experiments. DMA (75 mM) and TPP (7 µM) were dissolved in 0.1 M LiTFSI tetraglyme electrolyte and 100% dry O₂ was bubbled into the solution. The light was turned on after 2 h to ensure that the solution was initially saturated with ³O₂ before ¹O₂ generation. The duration of light exposure was recorded, and samples were analvsed by quantitative NMR to guantify DMA and 9,10-dimethyl-9,10-dihydro-9,10epidioxyanthracene (DMAO₂) (Figure S3). The amount of DMAO₂ was calculated by integration of the NMR data, and the rate of ¹O₂ generation under these photochemical conditions was estimated to be 1.56 x 10⁻⁵ mol min⁻¹ assuming 100% capture of ${}^{1}O_{2}$ by DMA. The efficiency of the ${}^{1}O_{2}$ generation was tested under an applied potential of 3.8 V and the amount of DMAO₂ is not affected by the voltage. Correcting for volume, 750 μ L of electrolyte gives a rate per volume of 2.08 × 10⁻² mol ¹O₂ L⁻¹min⁻¹. The rate of ¹O₂ formation in a cell containing 100 µL electrolyte and 1 mg carbon electrode discharging at 100 mA g^{-1}_{carbon} , and assuming 2% $^{1}O_{2}$ generation, is 6.12 × 10⁻⁶ mol $^{1}O_{2}$ L^{-1} min⁻¹ which is ~1000 times less than that found in the online-MS experiments. Assuming the steady-state concentration of ¹O₂ is directly proportional to the rate of formation, then the ¹O₂ concentration is also ~1000 greater in our degradation analysis compared to the cell and can be considered a harsher test of stability.



Figure S3. The ¹H NMR (600 MHz, CD₃CN, 298.1 K) spectra of a solution containing 75 mM DMA (*bottom*) and a ¹O₂ exposed solution (*top*) show resonances for DMA and DMAO₂. The aromatic peaks for DMAO₂ are marked in the dashed box.

Electrode Preparation

For ¹³C-PTFE composite working electrodes (WEs), amorphous carbon-13 powder (¹³C, Sigma-Aldrich) was treated at 900 °C under Ar:H₂ (95:5, v/v) flow for 5 h. The carbon powder was transferred to an Ar-filled glovebox. The WEs were prepared in an Ar-filled glovebox by mixing ¹³C and PTFE powder in an agate mortar. 10 mg of the composite material was pressed into a 7 mm diameter. The pellet was then pressed onto a stainless-steel mesh (100 mesh, Advent-RM) current collector.

Li₂O₂-preloaded working electrodes (¹³C-Li₂O₂ WEs) were prepared by grinding carbon-13 powder and in-house synthesised Li₂O₂ (80:20 w/w), and adding 15% PTFE powder to this mixture. Similar to ¹³C WE, ¹³C-Li₂O₂ composite material was pressed into 7 mm diameter pellets with 10 mg material loading.

LiFePO₄ was used as the counter and reference electrode material. To make LiFePO₄ electrodes, LiFePO₄, Super P, and PTFE (in water) were mixed in an 80:10:10 w/w ratio with isopropanol into a homogenous paste. This paste was cast on stainless steel mesh. The LiFePO₄ electrodes were chemically delithiated to ~20 % total capacity (Li_xFePO₄, x<1). For this, the electrodes were soaked in 1.5 mL acetic acid (Aldrich), 3.6 ml 30% H₂O₂ (Aldrich) and 250 ml MilliQ water for 20 minutes.⁹ The electrodes were washed with MilliQ water (18.2 MΩ cm) and isopropanol until all the acetic acid was removed, dried under vacuum at 85 °C overnight, and transferred into an Ar-filled glovebox.

Electrochemical Experiments

The effect of ¹O₂ under applied potential was investigated by applying a constant potential of 3.8 V Li|Li⁺ for 12 hours whilst photochemically generating ¹O₂ in-situ. The same electrochemical experiments were repeated with ³O₂ (without turning on the light source). The electrodes were carbon-13 (WE), LiFePO₄ (CE), and delithiated LiFePO₄ (REF). The electrolyte was 0.1 M in LiTFSI in tetraglyme containing 7 μ M TPP.

¹³C-Li₂O₂ WEs were used to investigate the effect of Li₂O₂ on degradation and for these experiments the cell was charged under Ar atmosphere at a constant potential of 3.8 V.

Cyclic voltammetry was performed in a heart-shaped flask with a planar Au working and counter electrodes and a fritted delithiated LiFePO₄ reference electrode at a scan rate of 100 mV s⁻¹.

Singlet Oxygen Induced Degradation Studies (Batch)

Photochemical Set-up

Photochemical reactions were conducted using green LED bulbs (5 W). Lamps were positioned with a bulb-to-vial distance of approximately 10-15 mm and fan cooling was used to maintain an external temperature of ~25-30 °C (**Figure S4**). Internal temperatures were measured to not exceed 27 °C. All reactions were magnetically stirred at 500 rpm using a PTFE coated stirrer bar.

Warning: Oxygenated ethers. Extended storage of oxygenated ether solvents, particularly in the absence of a stabiliser, is hazardous due to the accumulation of organic peroxides which can lead to unexpected explosions.¹⁰ Ensure temperatures do not exceed 30 °C due to the risk of detonation.





Tetraglyme & LiTFSI Experiments

Photosensitisation of triplet oxygen with RB or TPP has been reported as an efficient method for ${}^{1}O_{2}$ formation and subsequent ${}^{1}O_{2}$ mediated C-H hydroperoxidation of cyclic ethers (such as THF) and activated linear ethers.¹¹ Having successfully reproduced these results with THF (not shown), the method was deemed suitable for the evaluation of tetraglyme and 1 M LiTFSI in tetraglyme stability to ${}^{1}O_{2}$ by both flow and batch. A modified general procedure deviating from Su, M and co-authors' method is described below. Results for tetraglyme and 1 M LiTFSI in tetraglyme are summarised in **Tables S1** and **S2**, respectively. In both cases, no change to tetraglyme or the TFSI anion following prolonged exposure to ${}^{3}O_{2}$ with and without photocatalyst, nor when irradiated with green light in the absence of photocatalyst was detected by 1 H and 19 F NMR spectroscopies (**Figure S5-7**). Under reaction conditions producing ${}^{1}O_{2}$ (*Entry 4*, **Tables S1 & S2**) new resonances were observed in the 1 H NMR spectra at 11.0 – 8.5 ppm and 5.5 – 4.5 ppm suggesting tetraglyme degradation (**Figure S5 & S6, Scheme S2**). Conversely, the TFSI anion showed no change by 19 F NMR spectroscopy following prolonged exposure to ${}^{1}O_{2}$ (**Figure S7**), even with organic hydroperoxide species present, suggesting stability to both ${}^{1}O_{2}$ and tetraglyme degradation products.



Scheme S2. Photo-oxidation of tetraglyme via singlet oxygen.

General procedure: An oven-dried microwave vial equipped with a stirrer bar was taken inside a glovebox (nitrogen atmosphere) before the addition of tetraglyme solution (with or without 1 M LiTFSI) and rose bengal stock solution (if using) to a total volume of 4 mL. The vial was sealed with a septum-lined cap, removed from the glovebox and sparged with oxygen (5-10 min) before irradiating according to the described photochemical setup for 18 h. An internal standard, 1,4-dicyanobenzene (5.1 mg, 0.04 mmol, 0.01 M), was added to the resulting solution, a 50 μ L aliquot was taken, added to CDCl₃ (0.75 mL) and NMR spectroscopies measured.

Table S1. Stability test of tetraglyme under the photocatalytic generation of singlet oxygen.^a

Entry	Photocatalyst	Light source	Degradation detected	
1	None	None	NPD	
2	None	Green	NPD	
3	RB	None	NPD	
4	RB	Green	Traces	

^aReaction conducted according to the *general procedure*: Photocatalyst $(1 \times 10^{-5} \text{ M})$, tetragylme (4 mL), O₂, irradiated for 18 h. NPD = No products detected. Trace is denoted when the concentration of products is <1 mM.

Table S2. Stability test of 1 M LiTFSI tetraglyme solution under the photocatalytic generation of singlet oxygen.^a

Entry	Photocatalyst	Light source	Degradation detected	
			Tetraglyme	Litfsi
1	None	None	NPD	NPD
2	None	Green	NPD	NPD
3	RB	None	NPD	NPD
4	RB	Green	Trace	NPD

^aReaction conducted according to the *general procedure*: Photocatalyst $(1 \times 10^{-5} \text{ M})$, 1 M LiTFSI tetragylme (4 mL), O₂, irradiated for 18 h. NPD = No products detected. Trace is denoted when the concentration of products is <1 mM.

Supplementary Note 1: Tetraglyme degradation calculations

As indicated by ¹H NMR spectroscopy, tetraglyme underwent degradation in the presence ¹O₂ with observable signals in the regions of 11.0 – 8.5 ppm and 5.5 – 4.5 ppm (Figure 3 & S5, S6 & S8). These resonances are in the expected regions for organic hydroperoxide proton (OOH) and associated tertiary proton (OCH) environments, respectively. Given tetraglyme contains multiple sites vulnerable to oxidation (eight α -CH₂ positions), a series of mono, bis, or greater hydroperoxide species are the likely contributors to the observed signals. Under the assumption that each new resonance in the region of 11.0 - 8.5 ppm corresponds to a reacted ${}^{1}O_{2}$, we note seven significant peaks with the concentration for each of these new species to be approximately 0.2 - 0.8 mM as determined by ¹H NMR spectroscopic analysis. Thus, the estimated maximum concentration of accumulated ¹O₂ products is ca. 5.6 mM, with moles of ¹O₂ consumed approximated to be 2.2 $\times 10^{-5}$ mol.

- s in region, each 0.2 0.8 mM
- responds to a reacted ¹O₂

Scheme S3. Photo-oxidation sites at tetraglyme by singlet oxygen and highlighting the new ¹H NMR resonance region of 11.0 – 8.5 ppm

To understand the impact of tetraglyme degradation by ¹O₂ on the cell, we can compare the amount of degradation products formed per ¹O₂ in model experiments and relate this to known ¹O₂ formation in the cell. The NMR spectroscopic analysis above suggests that the rate of ¹O₂ formation is ca. 2.08 \times 10⁻² mol ¹O₂ L⁻¹min⁻¹ for photochemical reactions, considering the variance between our ¹O₂ quantification and degradation (Batch) studies, this value is expected to be a minimum rate and thus an overestimate of the impact of ¹O₂. Assuming the photocatalyst was active for only 4 hours before bleaching, the total amount of ¹O₂ formed is ca. 5.0 moles ¹O₂ L⁻¹. Taking the concentration of degradation products above and normalising this to the total ¹O₂ formed per unit volume, the moles of degradation products per mole of ${}^{1}O_{2}$ is 1.1 x 10⁻³ mol mol⁻¹. Assuming 2 % of all moles of oxygen form ¹O₂ in the cell during charge, then the percentage of degradation due to singlet oxygen in the cell is ca. 2.2 x 10⁻³%. Assuming a worst-case scenario where 10 % of oxygen used within the cell forms ¹O₂, then the estimated amount of degradation is still only 0.01 % of the discharge product which is not consistent with the 10 % found in practical cells. In summary, ¹O₂ degradation pathways of the solvent and salt are likely minor compared to other challenges existing in state-of-the-art systems.

Spectra



Figure S5. Highlighted baseline of ¹H NMR (500 MHz, CDCl₃, 298 K) spectra of tetraglyme experiments from **Table S1**, notable degradation products detected only under singlet oxygen forming reaction conditions (*entry 4*). PC = photocatalyst.



Figure S6. The highlighted baseline of ¹H NMR (400 MHz, CDCl₃, 298 K) spectra of 1 M LiTFSI tetraglyme experiments from **Table S2**, notable degradation products detected only under singlet oxygen forming reaction conditions (*entry 4*). PC = photocatalyst



Figure S7. ¹⁹F NMR (377 MHz, CDCl₃, 298 K) spectra of 1 M LiTFSI tetraglyme experiments from **Table S2** indicating complete stability of LiTFSI to ¹O₂ under photochemical conditions. *Left*) Full spectra window shown, the broad resonance at ~ 170 ppm is the PTFE probe background. *Right*) Highlight of LiTFSI singlet region indicating no change in multiplicity.



Figure S8. ¹H NMR (500 MHz, CDCl₃, 298 K) spectroscopy highlighting degraded tetraglyme attributed to singlet oxygen (**Table S1**, *entry 4*). Internal standards in the tetraglyme solution (0.01 M) and tetraglyme ¹³C carbon satellites are shown to indicate the trace quantities of degradation products.

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