

## Supplemental information

*Dynamics of residential indoor gas- and particle-phase water-soluble organic carbon: measurements during the CASA Experiment*

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## S1. Perturbations

### *Cooking without vinegar*

Green peppers, bacon and tater tots were prepared in the National Institute of Standards and Technology (NIST) Net-Zero Energy Residential Test Facility (NZERTF) kitchen for cooking perturbations. These meals were prepared during 3 consecutive days of the campaign (March 13<sup>th</sup> to 15<sup>th</sup>). The food items were sequentially pan-fried in heated canola oil for about 10 min on an electric hot plate (pan temperature was approximately 150 °C to 200 °C), removing each food item from pan before starting the next. Cooking day 1 consisted of 3 rounds of cooking under high relative humidity (RH) conditions (74 % RH). Cooking on days 2 and 3 provided a comparison of pan-fry and air-fry cooking emissions where the same food items were sequentially pan-fried in the morning and sequentially air-fried (about 10 min with air fryer temperature set to 190 °C) in the afternoon under normal RH conditions (30 % to 33 % RH). All cooking perturbations were performed by University of Colorado Boulder researchers.

### *Cooking with vinegar*

Two perturbations with vinegar were performed to specifically observe acetic acid dynamics under normal house conditions. First, 30 ml (2 Tbsp) of balsamic vinegar were boiled until fully evaporated in a heated pan (burner set to high heat). Then later on the same day, 30 ml (2 Tbsp) of balsamic vinegar were incorporated into a balsamic glaze for cooking with vinegar perturbations. Chopped onions, green peppers, zucchinis, and sliced mushrooms were sautéed in heated pan soiled with bacon grease (under high heat burner setting) until onions were caramelized and other vegetables were browned. Then, balsamic vinegar was added to the pan until visibly turning into a glaze. All cooking with vinegar perturbations were performed by University of Colorado Boulder researchers.

### *Product addition (i.e., pesticide)*

Approximately 30 g of commercial insecticide was sprayed on the first floor of the NZERTF. A box fan was operated to promote mixing of the sprayed pesticide which was 0.15% permethrin, 0.15% tetramethrin, and 99.70% other ingredients (i.e., organic solvents such as isobutane and propane and petroleum distillates). The Henry's Law constants ( $K_H$ ) of isobutane and propane are on the order of  $10^{-4}$  M atm<sup>-1</sup> and  $10^{-3}$  M atm<sup>-1</sup>, respectively.<sup>7</sup> The Henry's law constants for aromatic naphtha from petroleum can be on the order of  $10^0$  M atm<sup>-1</sup> and pyrethroids are typically on the order of  $10^2$  M atm<sup>-1</sup>.<sup>7</sup> Product additions were performed by NIST researchers.

### *Acid/base addition*

Mass-flow controlled additions of 500 mg min<sup>-1</sup> and 40 mg min<sup>-1</sup> of carbon dioxide (CO<sub>2</sub>) and ammonia (NH<sub>3</sub>), respectively, were injected into the supply flow of the Heat Recovery Ventilator (HRV) system and delivered peak mixing ratios of approximately 5000 ppm<sub>v</sub> CO<sub>2</sub> and

500 ppb<sub>v</sub> NH<sub>3</sub>. Note: we define mixing ratio as the ratio of the number density of a gaseous compound to the dry air number density (approximately  $2.43 \times 10^{19}$  molecules cm<sup>-3</sup> at standard ambient temperature and pressure) expressed as parts-per-million by volume (ppm<sub>v</sub>), parts-per-billion by volume (ppb<sub>v</sub>) or parts-per-trillion by volume (ppt<sub>v</sub>). Mixing ratios of CO<sub>2</sub> and NH<sub>3</sub> were allowed to decay for at least 3 h before repeated injections. In all, 2 to 3 injections of each species were performed under elevated RH conditions on a single day, then under lower, background RH conditions the following day.

#### *CASA cocktail injection*

A volatile organic compound (VOC) cocktail (i.e., acetone, toluene-d8, o-xylene, chlorobenzene, 2-pentanone, 2-heptanone, alpha-pinene, 1-hexene, isoprene, 1-octene and furfural) was injected into the NZERTF for a VOC addition perturbation (aka CASA cocktail injection) under normal house conditions. A volume of 0.5 mL of each compound was combined in a flask that was submerged in an 89 °C water bath until the solution was evaporated (approximately 10 min). During heating and evaporation, a 10.2 L min<sup>-1</sup> zero air carrier gas flow was passed over the flask headspace and into 1.14 m long Teflon tube that protruded through the porch door into the dining room of the NZERTF. The CASA cocktail perturbations were performed by University of Toronto researchers.

#### *Wood smoke injection*

A portable cocktail smoker was used to generate small amounts of smoke from combusting Ponderosa pine woodchips. There were 23 wood smoke injections between March 21<sup>st</sup> and April 6<sup>th</sup> performed by Colorado State University researchers.<sup>1</sup> Approximately 0.2 g to 0.5 g of woodchips were either burned in a cocktail smoker directly inside the test house or burned outside the house where the smoke was collected in a ~1 m<sup>3</sup> Teflon chamber and indirectly injected into the house; Teflon chamber-contained smoke was then injected into the house using a zero-air flow that diluted/pushed smoke in the chamber through a copper tube outlet that extended into the first floor of the house through a side door. For some experiments, smoke was injected under high (74 % RH; March 29<sup>th</sup> and March 30<sup>th</sup>) and compared to repeat experiments under low RH conditions (32 % to 36 % RH; April 5<sup>th</sup> and April 6<sup>th</sup>). Furthermore, for some experiments, wood smoke (typically obtained burning a total of ~6 g of woodchips in the cocktail smoke) was aged in the Teflon bag with 10 ppm<sub>v</sub> to 20 ppm<sub>v</sub> of ozone (O<sub>3</sub>) before injection. Also, O<sub>3</sub> injections occurred in both the smoke-free and smoke-exposed house.

#### *Ozone addition*

An O<sub>3</sub> generator was used to deliver a 0.001 L min<sup>-1</sup> flow of approximately 100 ppm<sub>v</sub> O<sub>3</sub> into the HRV (245 m<sup>3</sup> h<sup>-1</sup>) that supplied air to the main house; O<sub>3</sub> injections were typically 30 min to 90 min in duration and were performed by NIST personnel throughout the campaign. A total of 17 ozone experiments (1 to 4 O<sub>3</sub> injections each experiment) was performed by NIST personnel throughout the campaign.

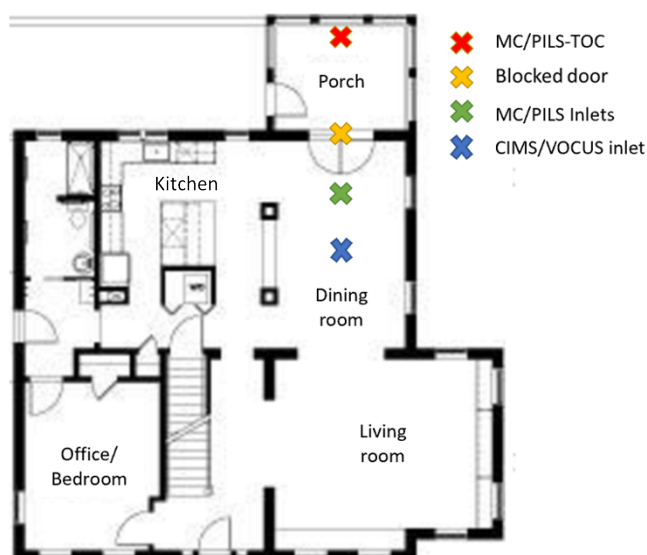
## Cleaning

Surface cleaning (dusting, vacuuming, and mopping) and air cleaning (i.e., portable commercial air cleaners and custom-built Corsi-Rosenthal boxes constructed of air conditioning filters and a fan) were performed. Two surface cleaning days were implemented during the campaign under normal temperature and RH conditions aimed at cleaning the NZERTF before smoke additions began (March 21<sup>st</sup>) and then again in the smoke-exposed house after smoke injections were completed (April 7<sup>th</sup>). Main house floors were vacuumed, and other surface were dusted (approximately 1 h), followed by floor mopping and surface wiping (approximately 30 min); the total floor area of the house 1<sup>st</sup> and 2<sup>nd</sup> floor in the main house was about 252 m<sup>2</sup>. Floors were mopped and surfaces were wiped with 110 ml (7.5 Tbsp) tri-sodium phosphate (TSP) and 90 ml (3/8 cups) of a commercial multiple purpose cleaner diluted in 5.7 L (1.5 gallons) of water, which was a recommended cleaning solutions for removal of smoke odor by the Red Cross.<sup>2</sup> Air cleaners were located on the first floor and operated after some smoke additions and represented a broad spectrum of cleaning mechanisms (e.g., HEPA filters, UV, photocatalytic oxidation, activated carbon, and electrostatic charge). The air and surface cleaning experiment was led by University of California, San Diego researchers.

## Window opening

All windows on the first and second floors and the front door of the NZERTF were opened during a high occupancy day (April 7<sup>th</sup>; 10 occupants inside the house for most of the afternoon) for about 5 h in which visitors were given tours of the house. At the end of the day, all windows were closed. Water soluble organic carbon (WSOC) measurements during window opening and closing were captured.

## S2. Experimental set-up details



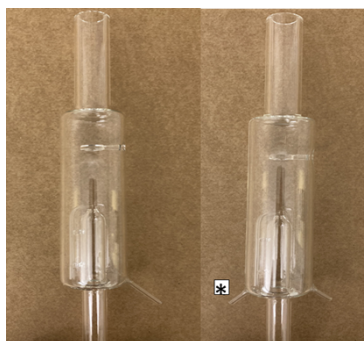
**Figure S1.** NZERTF 1<sup>st</sup> floor plan with sampling system (red) and sampling inlet position (green and blue) marked (x). The blocked porch door is also marked (gold).



**Figure S2.** Instruments for  $WSOC_g$  and  $WSOC_p$  on NZERTF porch. **1.** Total organic carbon analyzer (TOC) **2.** Particle-into-liquid sampler (PILS) **3.** Mist Chamber (MC).



**Figure S3.**  $WSOC$  sampling inlet locations. Room air was drawn to the MC ( $WSOC_g$ ) and PILS ( $WSOC_p$ ) through the Teflon tube and copper tube, respectively.



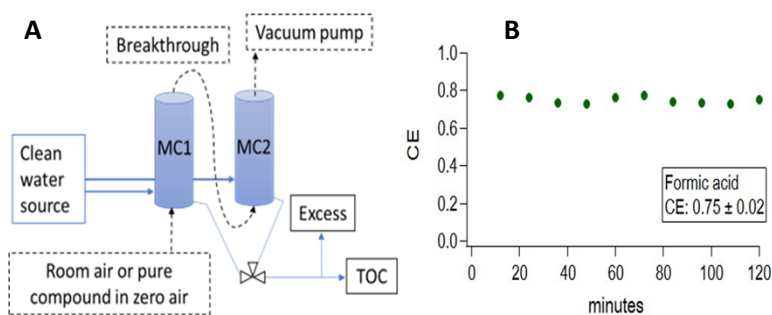
**Figure S4.** Traditional mist chamber (left) and real-time mist chamber (right). The new inlet that enabled continuous bulk water flow and real-time analysis is denoted (\*). The new inlet nozzle was added to the same side of MC as the capillary tube that draws water for mist generation. The real-time MC was operated with a water reservoir and droplet residence time of 8.3 min (10 mL of water/ $1.2 \text{ mL min}^{-1}$  of water flow) and 0.43 sec ( $(0.190 \text{ L MC volume} - 0.010 \text{ L bulk water volume})/0.42 \text{ L sec}^{-1}$  air flow), respectively.

### S3. Quality control

#### Determination of MC collection efficiency

Traditionally, MCs have been used as integrated samplers to measure individual compounds<sup>3</sup> and characterization of MC performance in collecting total gas phase WSOC (WSOC<sub>g</sub>) mixtures is limited but improving.<sup>4-6</sup> Collection efficiency was measured using two real-time MCs operated in series (MC1 = upstream MC, MC2 = downstream MC) and MC water from each was analyzed for WSOC in real-time, alternating between MC1 and MC2 every 6 min for total collection. A dynamic blank was performed before sampling air and MC water was analyzed with the vacuum pump off (no air flow through MC1 or MC2) and subtracted from sample TOC concentrations. Here, for MCs operated as a continuous real-time sampler, the collection efficiency was constant and a steady-state WSOC concentration was maintained (**Figure S5B**). Collection efficiency (CE) was calculated using the **Equation S1**.

$$CE = 1 - \frac{\text{Concentration in MC2}}{\text{Concentration in MC1}} \quad \text{Eqn S1}$$



**Figure S5.** Collection efficiency (CE) experimental set-up (A) and a representative CE vs

#### Summary and discussion of MC collection efficiency results

Mean ( $n = 10$ ) measured collection efficiencies and  $K_H$  values for glyoxal, formic acid, acetic acid, acetone and WSOC<sub>g</sub> in room air are listed in **Table S1** and compared to reported values for integrated MCs in literature. The collection efficiency for compounds with  $K_H > 10^3$  (formic acid, acetic acid, and glyoxal) were in the range of 63 % to 75 % whereas collection efficiency for acetone ( $K_H < 10^2 \text{ M atm}^{-1}$ ) was 41 %. Reversing the order of the MCs in series yielded the same result. The minimum collection efficiency observed for acetone was roughly 4 times higher than observed by Hennigan et al.<sup>7</sup>

**Table S1.** Measured mist chamber collection efficiencies for individual water-soluble organic gases and mixtures from this work and reported in literature.

Compound	Henry's law constant (mol L <sup>-1</sup> atm <sup>-1</sup> ) <sup>8</sup>	MC collection efficiency (%) (Henningan et al. 2018) <sup>5*</sup>	MC collection efficiency (%) (Duncan et al. 2019) <sup>9‡</sup>	MC collection efficiency (%) (this work) ‡
Glyoxal	4,000,000	109 ± 4		63 ± 2
Formic acid	8,000	104 ± 5	76 ± 5	75 ± 2
Acetic acid	4,000	94 ± 4	55 ± 4	74 ± 8
Acetone	31	11 ± 8		41 ± 3
WSOC <sub>g</sub> in room air	10 <sup>1</sup> to 10 <sup>6**</sup>		43 ± 10	57 ± 4

\*Mist chamber parameters: 10 mL water (initial), approximately 8 mL water (final), 28 L min<sup>-1</sup> air flow, room temperature (21 °C), 5 min collection time, 0.6 sec droplet residence time. Collection efficiency > 100 % were reported when the downstream MC was measured to have TOC levels below the dynamic blank and were reported as such to demonstrate method uncertainty.

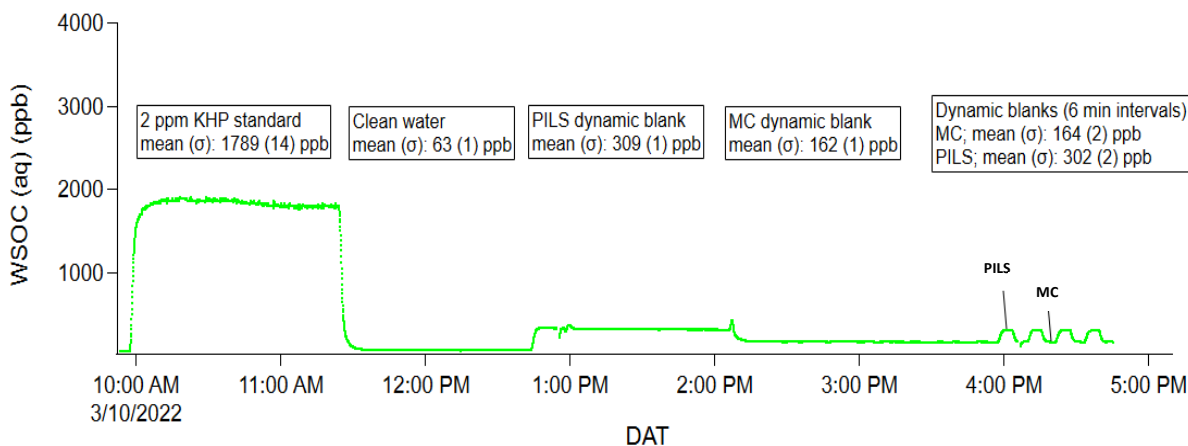
‡Mist chamber parameters: 25 mL water topped off as needed (due to evaporation), 25 L min<sup>-1</sup> air flow, room temperature (20 °C to 22 °C), 2 h collection time, 0.4 sec droplet residence time. Measured in a NC home.

‡Mist chamber parameters: 10 mL water (initial), approximately 10 mL water (final; constant water flow in and out while maintaining the meniscus height), 25 L min<sup>-1</sup> air flow, room temperature (20 °C to 22 °C), 2 h collection time, 0.43 sec droplet residence time and 8.3 min bulk water residence time. WSOC<sub>g</sub> in room air was measured in a kitchen used as a teaching lab in the Department of Nutrition at UNC.

\*\*General range of K<sub>H</sub> for water-soluble organic gases in indoor air.<sup>4,10</sup>

Note: MCs experience evaporative loss of bulk water due to air flow

### Calibrant check, clean water check, and dynamic blanks



**Figure S6.** TOC data (aqueous concentrations; ppb<sub>m</sub>) from measurements of a potassium hydrogen phthalate (KHP) calibrant check, clean water, PILS dynamic blank, MC dynamic

MC and PILS dynamic blank concentrations were 2.6 and 4.9 times higher than the TOC concentration measured in the clean water supply, respectively. After correcting for the clean water background (63 ppb<sub>m</sub>), the PILS dynamic blank WSOC signal was 11 % of the house background WSOC<sub>g</sub> concentration measured by MC-TOC. This indicates that there was 11 % breakthrough of WSOC<sub>g</sub> through the PILS upstream carbon denuder. Reported particulate WSOC (WSOC<sub>p</sub>) concentrations are corrected for WSOC<sub>g</sub> breakthrough. Note: ppb<sub>m</sub> is defined as the grams of compound (or in this case the grams of carbon) per billion grams of solution.

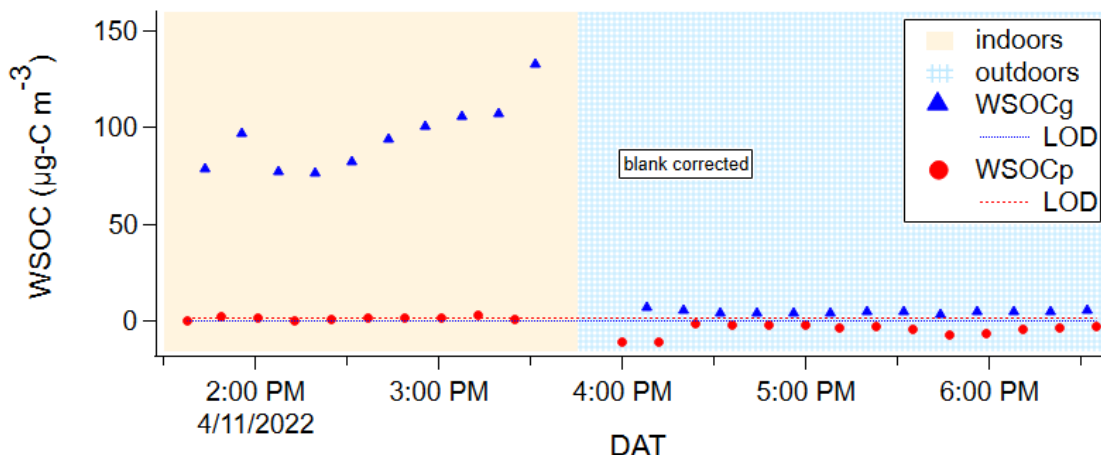
### S4. Indoor vs. outdoor WSOC measurements

#### Indoor-outdoor (I/O) ratios

**Figure S7** shows blank corrected indoor and outdoor WSOC concentrations on April 11. Indoor WSOC<sub>g</sub> was at least factor of 19 greater indoors ( $95 \mu\text{g-C m}^{-3} \pm 18 \mu\text{g-C m}^{-3}$ ) than outdoors ( $5 \mu\text{g-C m}^{-3} \pm 1 \mu\text{g-C m}^{-3}$ ), suggesting that there are substantial indoor WSOC<sub>g</sub> sources. Indoor and outdoor WSOC<sub>p</sub> concentrations were below the limit of detection (LOD) on average. On this day, people were freely entering and exiting the house and the front door was sometimes propped open which should explain the larger indoor WSOC<sub>g</sub> variability and lower indoor WSOC<sub>g</sub> concentrations compared to unperturbed time periods on other days. This also suggests that the WSOC<sub>g</sub> indoor to outdoor ratio may be even larger when the house's doors and windows are



closed. The  $WSOC_g$  outdoor measurements were near, but still above the LOD.  $WSOC_p$  measurements were near LOD indoors and below LOD outdoors. The I/O ratio was approximately 20 for the  $WSOC_g$  and greater than or equal to 1 for  $WSOC_p$  (using the LOD as the upper bound outdoor  $WSOC_p$  concentration).



**Figure S7.** Indoor and outdoor  $WSOC_g$  and  $WSOC_p$  concentrations (blank corrected). The LOD for the  $WSOC_g$  (LOD:  $0.5 \mu\text{g-C m}^{-3}$ ) and  $WSOC_p$  (LOD:  $2.0 \mu\text{g-C m}^{-3}$ ) are also shown.

## S5. Additional measurements

### *Aldehydes, ketones, and alcohols*

Gas-phase mixing ratios of C2 to C4 water-soluble aldehydes, ketones, and alcohols were measured using a Vocus proton-transfer-reaction (PTR-MS) mass spectrometer<sup>11</sup> by University of Toronto researchers. The instrument was in the garage of the NZERTF. The sample inlet was in the dining room and located approximately 1.6 m from the floor. The instrument exhibited an average mass resolution of approximately 10,000 ( $m/\Delta m$ ) to 11,000 ( $m/\Delta m$ ). Gases were sampled through a 30 m long, 12.7 mm OD, 9.5 mm ID Teflon line that was heated to 50 °C, through which 4 L  $\text{min}^{-1}$  of room air was flowing. A PTFE particulate filter in the inlet line removed particles so that only the gas phase was measured. The PTR-MS detects protonated molecular ions. Instrument backgrounds were performed frequently using a heated catalyst system. Calibrations for ethanol and acetaldehyde were conducted onsite every 3 h to 6 h for 5 min using flow from a cylinder containing 99.4 and 101.9 ppb<sub>v</sub> of these species, respectively. Acetone, butanone, and hydroxyacetone were all calibrated post-campaign. Acetone was calibrated using a standard cylinder with 992 ppb<sub>v</sub> of acetone. Butanone and hydroxyacetone were calibrated from liquid standards by injecting a set flow rate of each liquid standard solution from a syringe pump into a pre-determined zero air carrier gas flow (a detailed procedure has been published previously).<sup>12</sup> Since these three calibrations were performed after the measurement period, their signals were ratioed to toluene's post-campaign signal from toluene calibrations with the same cylinder mixture (1000 ppb<sub>v</sub> toluene in cylinder VOC mix). These ratios were compared to the toluene calibrations during the campaign (103.1 ppb<sub>v</sub> toluene in field

cylinder) to infer acetone, butanone, and hydroxyacetone mixing ratios during the measurement period.

In addition to structural isomers, PTR ionization can lead to ion fragmentation so that multiple species may contribute intensity to any specific ion signal. To assess interferences at the protonated molecular ions for each species, a fast-scanning gas chromatograph<sup>15</sup> was periodically interfaced to the inlet of the PTR-MS which provided chromatographic separation for each sampled ion. Using 19 chromatograms measured throughout the campaign, it was determined that a large portion of the signal of the protonated VOC ions was due to the targeted analyte, with 11 % interference for ethanol, 17 % for acetone, 11 % for acetaldehyde, 48 % for butanone, and 55 % for hydroxyacetone. The reported mixing ratios account for these interferences. Given the onsite nature of the calibrations for acetaldehyde and ethanol, we estimate the uncertainties in the mixing ratios for these compounds to be on the order of 20 %. For the species calibrated post-campaign, the potential uncertainties are harder to estimate but are likely larger, perhaps up to a factor of two. Data processing, including mass calibration and high-resolution peak fitting, was performed using Tofware software (v 3.2.5).

### *Organic Acids*

The gas-phase mixing ratios of C<sub>1</sub> to C<sub>5</sub> water-soluble carboxylic acids (i.e., formic, acetic, propanoic, butanoic, and pentanoic acids) were measured using a Time-of-Flight Chemical Ionization Mass Spectrometer (ToF-CIMS) equipped with iodide (I<sup>-</sup>) reagent ions. The ToF-CIMS was operated by Colorado State University researchers. The instrument exhibited an average mass resolution of approximately 4000 ( $m/\Delta m$ ). Detailed information regarding the ion chemistry and operational procedures can be found elsewhere.<sup>13,14</sup> Air samples from the dining room were drawn into the instrument, which was located in the garage of the NZERTF, via a 30.5 m heated 6.1 mm PFA (perfluoroalkoxy alkane) line, maintaining a flow rate of 5 L min<sup>-1</sup> to 12 L min<sup>-1</sup>. Hourly zero measurements and calibrations were conducted using ultra-high purity (UHP) nitrogen (N<sub>2</sub>) and certified standards of the C<sub>1</sub> to C<sub>5</sub> carboxylic acids. To maintain equivalent RH levels as ambient air, the zeroing N<sub>2</sub> flow consisted of a combination of dry N<sub>2</sub> and wet N<sub>2</sub>. Throughout each sampling day, the sensitivities of these acids exhibited variations of less than 20 %. All data processing, including mass calibration and high-resolution peak fitting, was performed using Tofware software (v 2.5.7).

### *Ozone*

O<sub>3</sub> mixing ratios in the main house were measured using an O<sub>3</sub> analyzer (LOD: 0.5 ppb<sub>v</sub>, precision: ± 1 ppb<sub>v</sub>) by NIST personnel on the first floor and outside the NZERTF using separate, but identical, sampling lines. The sample line was 9.1 m long PFA tube with a 6.3 mm OD and 0.45 μm in-line particle filter. The O<sub>3</sub> sampling system was designed to switch between the first floor and a sampling location outside the NZERTF. The combined flow rate through the sampling lines was 8 L min<sup>-1</sup>. The instrument itself was located in the attic of the NZERTF.

## *Formaldehyde*

Formaldehyde and formic acid mixing ratios in the main house were measured by NIST personnel using a quantum cascade tunable infrared direct absorption spectrometer (QC-TILDAS) which sampled at  $1 \text{ L min}^{-1}$  with an LOD of  $0.1 \text{ ppb}_v$  and a precision of  $20 \text{ ppt}_v$  to  $100 \text{ ppt}_v$ . The spectrometer measured formaldehyde on the first floor and outside the NZERTF; the instrument itself was located in the attic of the NZERTF and formaldehyde was measured using the same sampling system as  $\text{O}_3$ .

## S6. WSOC in house background

**Table S2.**  $WSOC_g$  and  $WSOC_p$  (mean  $\pm$   $\sigma$ ) of 4 s measurements ( $n = 30$ ) during 6 min sampling intervals) during unperturbed conditions (house background).  $WSOC_g$  concentrations were corrected using the measured effective collection efficiency of 57 %. Also shown are the fraction of airborne WSOC in the particle phase ( $F_p = WSOC_p/[WSOC_g + WSOC_p]$ ), temperature (T), relative humidity (RH) and air change rate with outdoor air (ACR;  $h^{-1}$ ). “House background periods” refer to time periods immediately before conditions were perturbed by scripted activities (typically 1 h to 2 h before the perturbation), 12 h to 16 h after the last perturbation, and sometimes shortly (1 h to 2 h) after perturbations that did not induce an observable change in WSOC in our system.  $\sigma$ : standard deviation

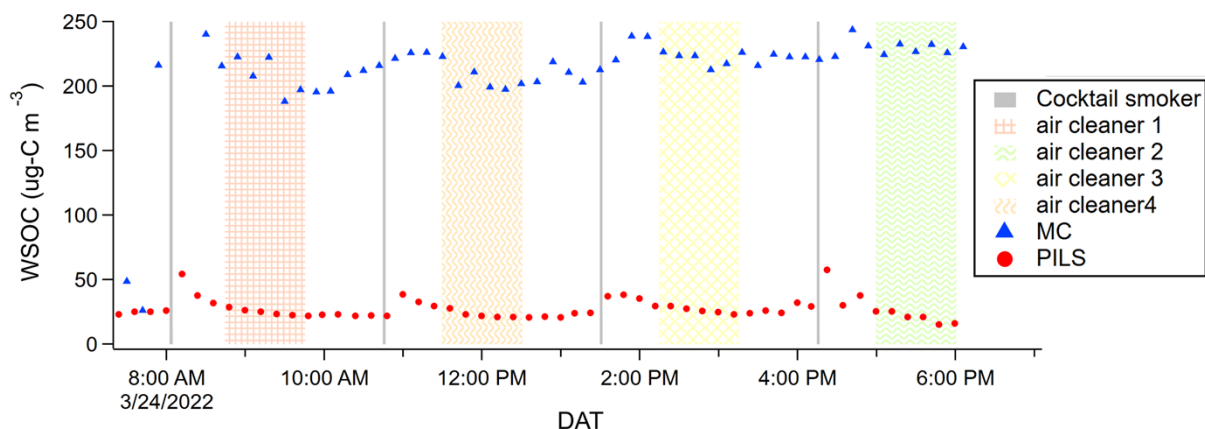
Date	Time	$WSOC_g \pm \sigma$ ( $\mu\text{g-C m}^{-3}$ )	$WSOC_p \pm \sigma$ ( $\mu\text{g-C m}^{-3}$ )	$F_p$	T ( $^{\circ}\text{C}$ )	RH (%)	ACR
8-Mar	11 AM- 12 PM	213 $\pm$ 5	10 $\pm$ 4	0.05	24	27	0.24
11-Mar	10 AM- 11 AM	228 $\pm$ 14	4 $\pm$ 2	0.02	24	28	
16-Mar	10:30 AM- 11:30 AM	258 $\pm$ 9	3 $\pm$ 1	0.01	24	32	
21-Mar	11 AM- 12 PM	196 $\pm$ 6	3 $\pm$ 1	0.02	24	37	
1-Apr	10:30 AM- 11:30 AM	182 $\pm$ 6	8 $\pm$ 1	0.04	23	34	
<b>Avg:</b>		215 $\pm$ 29	6 $\pm$ 3	0.03 $\pm$ 0.02	24.0 $\pm$ 0.3	31.5 $\pm$ 0.0	

**Table S3.** Ranges of compound class contributions to the WSOC<sub>g</sub> and concentrations of selected individual water soluble organic gases in the house background across March 11, March 21, and April 1.

Compound class	Contribution (%)	Species	Concentration (µg-C m <sup>-3</sup> )
Alcohol	33 - 45	Ethanol	46 - 82
Organic Acid	12 - 16	Acetic	12 - 24
		Formic	7 - 9
		Propanoic	3
		Butanoic	0.5 - 1
		Pentanoic	1 - 1.5
Aldehyde	4 - 7	Acetaldehyde	7 - 9
		Formaldehyde	5 - 6
Ketone	8 - 9	Acetone	11 - 16
		Butanone	1 - 3
		Hydroxyacetone	0.8 - 1.2
Sum of Known Compounds	51 - 72	Ethanol, acetic acid, formic acid, propanoic acid, butanoic acid, pentanoic acid, acetaldehyde, formaldehyde, acetone, butanone, and hydroxyacetone	101 - 144
Sum of Unknown Compounds	28 - 49	--	51 - 95

## S7. WSOC response to air cleaner operation

There was no clear change in WSOC induced by use of the air cleaners after wood smoke additions in our system (**Figure S8**).  $WSOC_g$  concentration may have decreased during operation of air cleaners 1 and 2 but is difficult to differentiate from the wood smoke peak decay due to ventilation and normal surface removal. More experiments are needed to verify these effects.

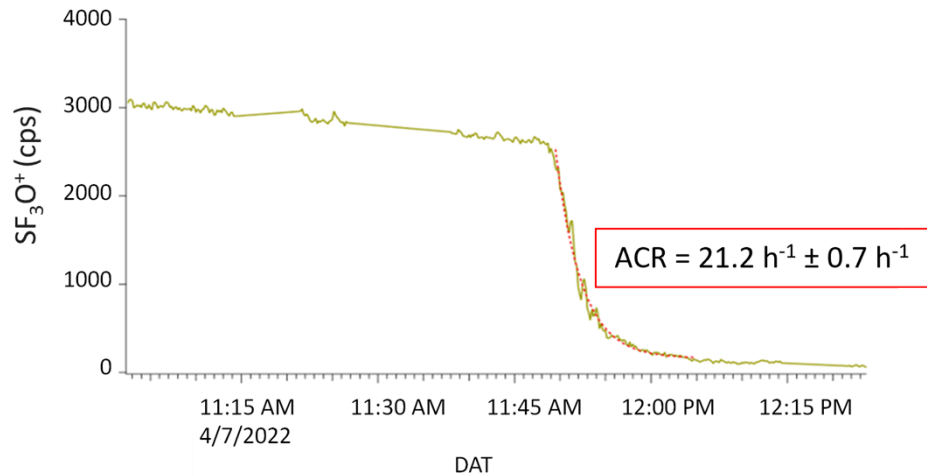


**Figure S8.**  $WSOC_g$  and  $WSOC_p$  concentrations during direct wood smoke injections (i.e., woodchips burned inside house) and operation of 4 different air cleaners—electrostatic (1), carbon filter (2), Corsi-Rosenthal box for gases (3; built with activated carbon filters), and Corsi-Rosenthal box for particles (4; built with traditional air filters for particle filtration). Error bars represent measurement uncertainties ( $\pm 1 \sigma$ ) and are mostly smaller than the data point markers. (DAT = date and time).

## S8. Air change rate with open windows

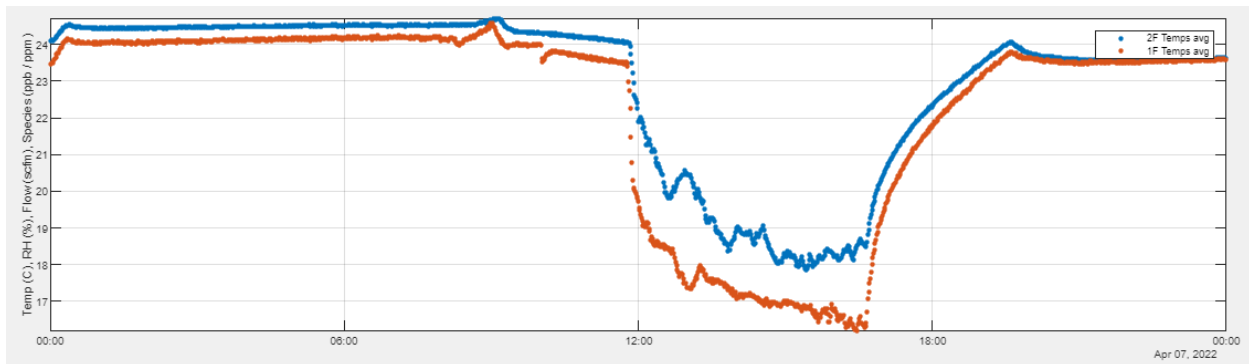
### *Determination of the elevated ventilation rate with windows open*

There was a second co-located time-of-flight mass spectrometer equipped with hydronium ion proton-transfer ionization (PTR-MS) at the Chemical Assessment of Surfaces and Air (CASA) experiment operated by NIST personnel. A  $5 \text{ L min}^{-1}$  sample flow was sampled through 30.5 m, 6.3 mm OD PFA lines. Sample lines were Teflon filtered and heated to  $50 \text{ }^\circ\text{C}$ . The instrument was zeroed and calibrated on an hourly basis.  $\text{SF}_6$  was measured using the PTR-MS via product ions  $\text{SF}_3\text{O}^+$  ion (mass-to-charge ratio: 104.962) with 10 s measurement resolution. The sensitivity, LOD, and signal-to-noise ratio of  $\text{SF}_3\text{O}^+$  was  $8 \text{ cps ppb}_v^{-1}$ ,  $350 \text{ ppt}_v$ , and 3, respectively. Additional product ions were  $\text{SF}_3^+$ ,  $\text{SF}_2^+$ , and  $\text{SF}^+$ , which were measured with less sensitivity. Tracer gas decay was used to determine the outdoor ventilation rate, or air change rate with outdoor air (ACR) (**Figure S9**).



**Figure S9.** Tracer gas decay (sulfur hexafluoride, SF<sub>6</sub>) during a window opening experiment on April 8<sup>th</sup>. The outdoor air change rate (ACR) during the natural ventilation event (at about 9:45 AM) was determined by fitting an exponential function to the decay of the measured SF<sub>6</sub> signal. Date and time (DAT) is in eastern daylight time (EDT).

The extremely high ACR with windows open (21 h<sup>-1</sup>) occurred because all windows and doors were opened nearly simultaneously at a time when that resulted in substantial thermal and wind pressure gradients. The outside temperature was 9 °C and the initial inside temperature was 24 °C. Figure XX demonstrates the impact on house temperatures when the house was opened (just prior to 12:00). This strong thermal gradient contributed to the high air change rate. In addition, thunderstorms were moving through the area resulting in local wind gusts.

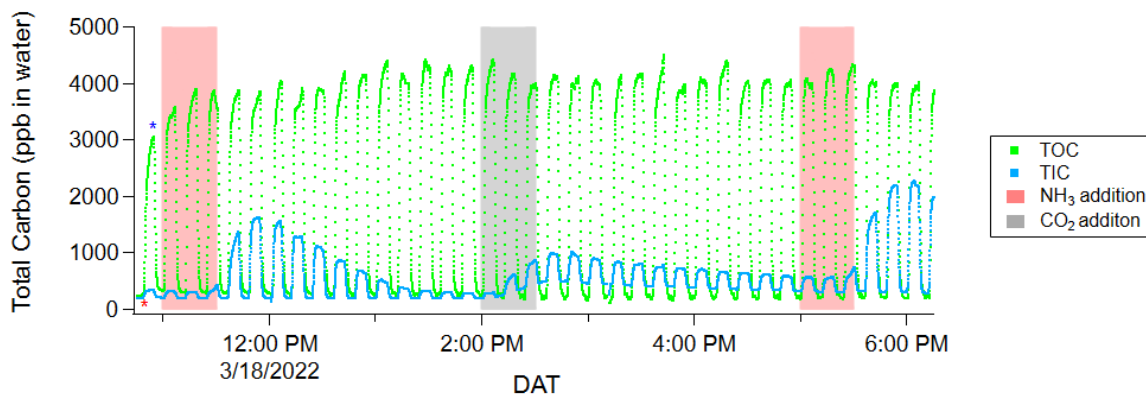


**Figure S10.** NZERTF average 1<sup>st</sup> and 2<sup>nd</sup> floor temperatures during open house event 04-07-22.

### S9. WSOC response to acid/base addition

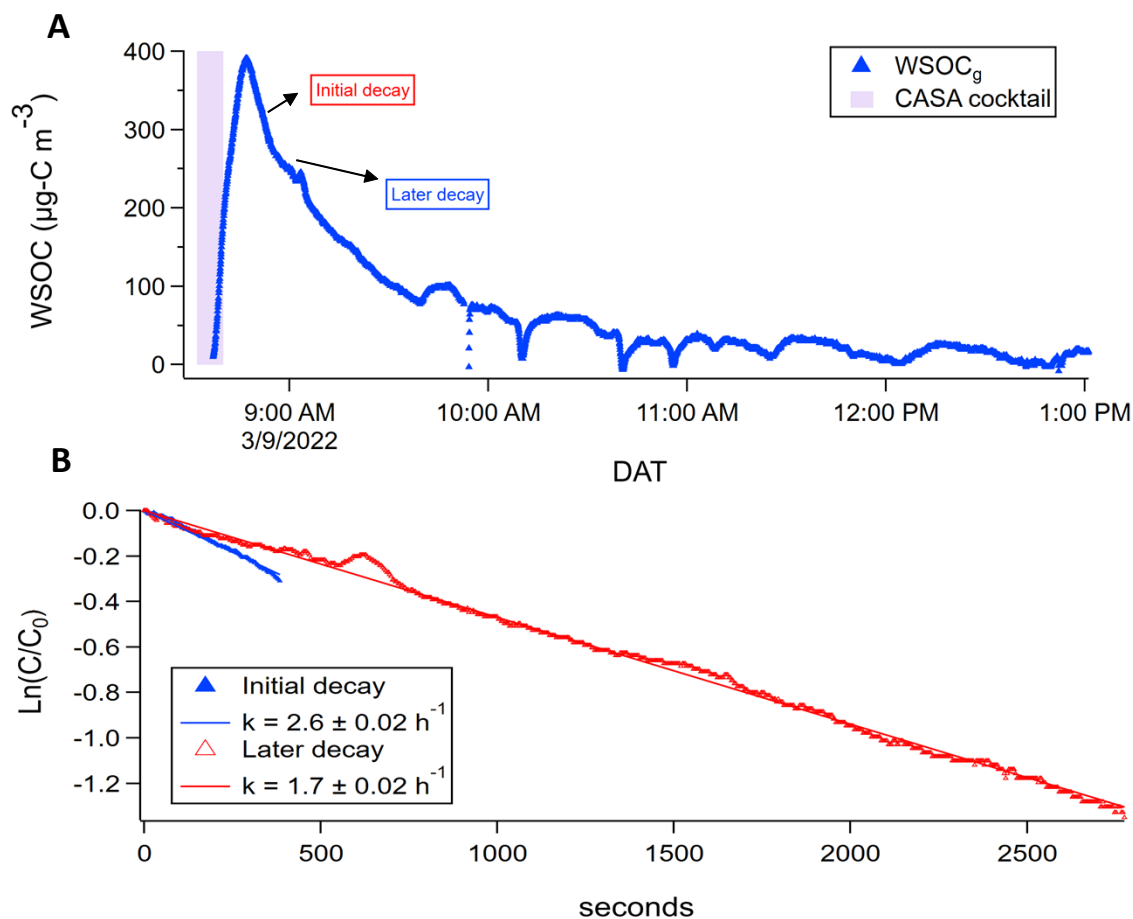
No observable change occurred in TOC but did for total inorganic carbon (TIC) upon addition of  $\text{NH}_3$  and  $\text{CO}_2$  (**Figure S10**). TIC increased upon  $\text{NH}_3$  injection for only the MC likely due to increase in MC water pH resulting in increased collection efficiency for acidic  $\text{CO}_2$ . During  $\text{CO}_2$  injections, TIC increased due to  $\text{CO}_2$  uptake into MC water and PILS aqueous sample flow (i.e., the carbon denuders upstream of PILS inlet does not remove  $\text{CO}_2$ ). Incorporation of  $\text{NH}_3$  and  $\text{CO}_2$  into MC water suggests Henry's law driven uptake of  $\text{NH}_3$  and  $\text{CO}_2$  to aqueous surface reservoirs inside the NZERTF likely also occurred and may have influenced surface pH conditions.

### S10. Change in decay rates with time



**Figure S11.** TOC (green markers) and TIC (blue markers) in  $\text{ppb}_m$  (grams carbon per billion grams of water) during semi-continuously monitoring of total carbon, alternating between the MC (blue asterisk; peaks) and PILS (red asterisk; valleys), during  $\text{NH}_3$  and  $\text{CO}_2$  injections.





**Figure S12.** Room background corrected WSOC<sub>g</sub> from volatile organic compound (VOC) “CASA cocktail” injection. **A.** WSOC<sub>g</sub> peak from VOC injection. *Initial* and *later* decay periods denoted. **B.** Log decay of WSOC<sub>g</sub> from VOC injection. *Initial* ( $k = 2.6 \pm 0.02 \text{ h}^{-1}$ ) and *later* ( $k = 1.7 \pm 0.02 \text{ h}^{-1}$ ) decay.

### S11. Disclaimer

Any equipment, instruments, software, or materials mentioned in this paper are solely for the purpose of accurately describing the experimental procedure. Mentioning these items does not constitute an endorsement or recommendation by NIST, nor does it imply that they are the best available options.

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