## **Supplementary Information**

Manganese-based nanozyme enables efficient mitigation of Huanglongbinginduced oxidative damage in *Citrus* 

Shuojun Li<sup>ac<sup>†</sup></sup>, Yuying Long<sup>b†</sup>, Guiyun Deng<sup>a</sup>, Yinghui Men<sup>d</sup>, Feifan Lu<sup>b</sup>, Zihan Wang<sup>c</sup>, Jiaying Li<sup>a</sup>, and Heyou Han<sup>ab</sup>\*

<sup>a</sup> National Key Laboratory of Agricultural Microbiology, College of Life Science and Technology, Huazhong Agricultural University, Wuhan, 430070, China.
<sup>b</sup>National Key Laboratory of Agricultural Microbiology, College of Chemistry, Huazhong Agricultural University, Wuhan, 430070, China.
<sup>c</sup> National Key Laboratory of Molecular Biology, CAS Center for Excellence in Molecular Cell Science, Shanghai Institute of Biochemistry and Cell Biology, 200031, China.
<sup>d</sup> Med-X Research Institute and School of Biomedical Engineering, Shanghai

<sup>a</sup> Med-X Research Institute and School of Biomedical Engineering, Shanghai JiaoTong University, Shanghai, 200030, China.

Correspondence: Heyou Han (<u>hyhan@mail.hzau.edu.cn</u>)

Table S1. Primers used in this study.

Name	Sequence
Cs6226SOD-F	GCTATAGGCTTGGGTAAATC
Cs6226SOD-R	CCGCCTTGTTGTAGTTAGTA
Cs6642SOD-F	GGCTCACGTTCTCCTCGTAG
Cs6642SOD-R	ATTTGAGGGAGACGCCGTG
Cs2588SOD-F	CAGTTTGGATATGTGGGAGC
Cs2588SOD-R	GGGTTCGCCTAAATTCACGA
Cs6866CAT-F	AATTCCCTGACATGGTCCAT
Cs6866CAT-R	AACACCAGATCCTTCCATGT
Cs5999CAT-F	ACAGCCACGCTACTCAGGAT
Cs5999CAT-R	TGTTCAAGACCAAGCGTCCA
Cs0069RBOH-F	AGCAACTCGTTGGTTCAGAA
Cs0069RBOH-R	GTAATATGCCGTGGACGATG
Actin-F	TCCAAGCAGCATGAAGATCAA
Actin-R	CAGACTCATCATACTCGCCCT
HLB-F	ACCAGTTCTTATTCCATTCGCCC
HLB-R	TCACAATCATGTTTTTCAAGCCAA



**Figure S1.** The stability of MONPs for long-term storage. The MONPs supernatant and DLS did not change after 1 month storage.



**Figure S2.** Characterization of the MONPs nano-enzyme. (A) Zeta potential analysis of the MONPs. (B) XRD (X-ray Powder diffractometer) pattern of the MONPs. (C) Raman pattern of the MONPs. (D&E) The fluorescence intensity of DCFH-DA after the reaction with X and XO in the absence and presence of the MONPs. X for xanthine and XO for xanthine oxidase, respectively. (Data are means, the statistic of peak points  $\pm$  SD, n=3).



**Figure S3.** Comfirm of the Clas-postive infection in HLB-*Citrus*. (A) PCR analysis of the genomic DNA of *C*las in *Citrus* tissues. H1, H2, H3 means the healthy Citrus; S1, S2, S3 means the HLB-*Citrus*. (B) Representative TEM images of the *C*las colonization in HLB-*Citrus*. Scale bar: 500 nm (right) and full size of 5  $\mu$ m (left).



**Figure S4**. Screen reactive oxygen species fluorescence probe for trace ROS localization in HLB-Citrus leaf. Scale bar: 20 µm.



Figure S5. Effects of MONPs on the Chlorophyll content of HLB-*Citrus* (Data are means  $\pm$  SD, n=3).



**Figure S6.** Assay of the *in vitro* antibacterial activity of MONPs. *Sinorhizobium meliloti Rm1021* (*S. meliloti Rm1021*) was co-incubated with MONPs at different concentrations (0, 20, 40, 60, 80, 100  $\mu$ g/mL). (A) Representative images of the diluted coating plate; (B) Quantitative analysis of antibacterial activity of the MONPs against *S. meliloti Rm1021* (Data are means ± SD, n=3).



**Figure S7.** Representative images of the *Citrus* leaf tissues staining with histochemical safranin O-fast green solution to observe the morphology.



**Figure S8.** GO enrichment histogram of top 30 GO function terms of HLB symptom related differentially expressed genes (DEGs).