### **Supplementary Information**

# Tuning La-O adsorption sites dispersion via hydrogen bond-capping organic-inorganic copolymerization strategy for enhanced phosphate removal

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#### **Section S1. Materials Preparation**

### Preparation of PVA-LHO copolymerized hydrogel (LaCPVA), pure PVA hydrogel and La-NP modified PVA hydrogel (La-PVA)

PvaLaC was prepared through the following repeated freezing-and-melting method. Initially, 10 mL of LHO at a concentration of 50 mg/mL was centrifuged at 8000 rpm for 5 min, and then the supernatant was removed. Aqueous solutions containing 10 wt. % PVA were formed by dissolving PVA in deionized water at 90°C for 6 h. The PVA aqueous solution was then uniformly mixed with the LHO precipitate to obtain a homogeneous emulsion. Next, the PVA solution was frozen using an ultralow temperature upright freezer maintained at -74±2°C. After 12 h, the frozen sample was thawed at room temperature for 6 h. This freeze and melt cycle were repeated three times. The pure PVA hydrogel was synthesized using the same procedure as mentioned above without the use of LHO. La-PVA was synthesized by substituting LHO with a La-NP solution.

## Preparation of PAM-LHO copolymerized hydrogel (LaCPAM), pure PAM hydrogel and La-NP modified PAM hydrogel (La-PAM)

Initially, 10 mL of the LHO solution (50 mg/mL) was centrifugated for 5 min at 8000 rpm, and the supernatant was discarded. Next, an aqueous AM solution (1.0 g of AM dissolved in 4 mL of  $H_2O$ ) was mixed uniformly with the LHO precipitate to generate a homogeneous emulsion. Subsequently, 4 mL of an MBAA alcoholic solution (5 mg/L) was added to the emulsion under vigorous stirring. Finally, 0.4 mL of a saturated KPS ethanol solution and 20 µL of TEMED were added to the mixture, which

had been pre-deoxygenated with  $N_2$  gas for 30 min. After a copolymerization reaction at 25°C for 24 h, the resulting white gel was washed with ethanol and deionized water to eliminate any residues, then dried under vacuum at -40°C for 24 h, resulting in the formation of bulk PamLaC. A pure PAM hydrogel was synthesized by following the procedure outlined above, excluding the inclusion of LHO. La-PAM was prepared by substituting LHO with a solution of La-NP.

#### Section S2. Adsorption Models

The isothermal adsorption curves were analyzed using the Langmuir and Freundlich equations. The Langmuir equation is formulated as follows:

$$q_e = \frac{q_m b C_e}{1 + b q_m}$$

where  $q_e$  represents the equilibrium adsorption concentration (mg/L),  $C_e$  is the equilibrium liquid-phase concentration (mg/L),  $q_m$  is the theoretical saturation sorption capacity (mg/g), b is a constant related to the adsorption heat <sup>1</sup>. The reliability of Langmuir equation could be assessed using the value of  $R_L$  as below:

$$R_L = \frac{1}{1 + bC_0}$$

where  $C_0$  is the initial adsorbent concentration. When  $0 < R_L \le 1$ , the experiment data could fit the Langmuir model; when  $R_L = 0$  or  $R_L > 1$ , the experiment data could not fit the model.

The Freundlich equation was utilized to describe non-ideal and multi-layer adsorption on heterogeneous adsorbents' surfaces <sup>2</sup>. The Freundlich equation is expressed as:

$$q_e = K_F C_e^{1/n}$$

Alternatively, this equation can be linearized as:

$$\lg q_e = \frac{1}{n} \lg C_e + \lg K_F$$

where  $K_F(mg^{1+n} L^n/g)$  and n are Freundlich constants.

The adsorption kinetics were performed in 100 mL of a 100.0 mg P/L solution at a dosage of 0.3 g/L at pH 7.0±0.2 agitated for 1.5 h at 300 rpm/min under 25°C. The pseudo-first-order (PFO) kinetic equation and pseudo-second-order (PSO) kinetic equation was applied to analyze adsorption kinetics.

The Liu isotherm model represents a synergistic amalgamation of the Langmuir and Freundlich isotherm models, with a departure from the Langmuir model's monolayer adsorption hypothesis and the Freundlich model's assumption of an unlimited adsorption capacity <sup>3</sup>. This approach posits that the active sites on the adsorbent surface exhibit a heterogeneous distribution of energies. The Liu equation is as follows:

$$q_e = \frac{Q_{max}(K_g C_e)}{1 + (K_g C_e)^{n_L}}$$

where  $K_g$  is the Liu equilibrium constant (L/mg);  $n_L$  is dimensionless exponent of the Liu equation, and  $Q_{max}$  is the maximum adsorption capacity of the adsorbent (mg/g).

The usage of La  $(g_{La})$  was calculated as follows:

$$g_{La} = \frac{Q_{LaCCH} - Q_{CH}}{W_{La}}$$

where  $Q_{LaCCH}$  (mg/g) is the maximum adsorption capacity of LaCCH,  $Q_{CH}$  is the

adsorption capacity of CH at the same equilibrium adsorption concentration calculated by the isotherm model,  $W_{La}$  is La element content (wt%) in LaCCH.

The PFO rate expression based on capacity is generally expressed as follows:

$$\frac{dq_t}{dt} = k_1(q_e - q_t)$$

Where  $K_1 \text{ (min}^{-1)}$  is the PFO rate constant;  $q_e \text{ (mg/g)}$  and  $q_t \text{ (mg/g)}$  are the amount of P adsorbed per unit weight of the adsorbent at equilibrium and time t.

The PSO equation is also based on the sorption capacity, which is expressed as follows:

$$\frac{dq_t}{dt} = k_2(q_e - q_t)^2$$

this equation also could be expressed by a linear model as follows:

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{1}{q_e} t$$

Where  $k_2 (g/(mg \cdot min))$  is the rate constant of second-order adsorption.

### Section S2. Supplemental Figures and Tables



Fig. S1 Ultrathin slices TEM images of CH.



Fig. S2 SEM-EDS mapping (a-c), TEM (d) and SAED (e) images of LaCCH.



Fig. S3 SEM-EDS mapping (a-d), TEM (e) and SAED (f) images of La-CH.



Fig. S4 TGA curves of LaCCH, La-CH and CH.



Fig. S5 Raman spectra of LaCCH, La-CH and CH.



**Fig. S6** Reconstructed 2D ToF-SIMS images of La<sup>+</sup> after 60 s sputtering, and Dynamic ToF-SIMS depth profiling of LaCCH (a) and La-CH (b). A darker color represents a stronger signal intensity.



**Fig. S7** Breakthrough curve of P adsorption by LaCCH with the feeding of a simulated effluent (TP=1.0 mg/L; COD=500 mg/L; [NaCl]=500 mg/L; [NaNO<sub>3</sub>-N]=50 mg/L; [NH<sub>4</sub>-N]=20 mg/L; pH=6.90-7.15; EBCT=6.25 BV/h, adsorbent dosage: 0.18 g/cm).



Fig. S8 Raman spectroscopy spectra of the LaCCH after P adsorption at pH=5.0-9.0.



Fig. S9 TEM (a), SAED (b) and EDS-mapping (c-d) images of LaCCH after P saturated.



# **Fig. S10** Adsorption isotherms of PVA/La-PVA/LaCPVA (a) and PAM/La-PAM/LaCPAM (b).



Fig. S11 TEM images of LaCCH (a), La-CH (b), LaCPVA (c), La-PVA (d), LaCPAM (e) and La-PAM (f).



**Fig. S12** The distribution of free path spacing corresponding to TEM images: (a) LaCPVA and PVA-La; (b) LaCCH and La-CH; (c) LaCPAM and La-PAM.



Fig.S13 Dependence of lg perimeter P on lg area A obtained from the binary TEM

images (Fig. S11) of La-PVA (a), LaCPVA (b), La-CH (c), LaCCH (d), La-PAM (e) and LaCPAM (f).

Langmuir Freundlich Liu  $q_e = \frac{k_L q_e c}{1 + k_L c}$  $q_e = k_F c^{\frac{1}{n}}$  $q_e = \frac{Q(kx)^n}{1 + (kx)^n}$ Equations  $R_{\rm L}{}^2$  $R_{\rm F}^2$ 1/n Q k  $R_l^2$  $k_{L} \\$  $k_{\rm F}$  $q_m$ п 0.92 1.82 75.28 0.25 1.74 0.96 LaCCH 108.27 0.124 16.14 0.87 La-CH 87.74 0.09 0.91 10.42 1.72 0.88 57.59 0.20 1.77 0.93 CH 49.08 0.12 0.96 2.00 0.91 0.98 8.10 65.64 0.21 1.71

Table S1 Parameters of isotherms fitting of LaCCH, La-CH and CH.

Table S2 Comparison of phosphorus adsorption capacity for different dephosphorization adsorbents

Adsorbent	рН	Initial concentration (mg/L)	$q_{ m m}$ (mg/g)	Ref.
La-MOFs	2.0-11.0	-	87.48-51.50	4
OA-La(OH) <sub>3</sub>	3.0-11.0	1-100	168	5
LaAl-BTC	2.0-12.0	5-1000	72.27-39.71	6
La@Fe	1.0-13.0	5-600	130–160	7
SCBC-La	3.0-9.0	50-120	48	8
LCM	2.0-10.0	10-100	77.49	9
HKL-LaOH	4.0-10.0	1-200	26.15	10
CCH@La	3.0-9.0	1-50	92.54	11
LDHs-Modified Biochar	3.0-11.0	10-60	10.64	12
FeCa-LDH	4.0-10.0	50-600	-	13
UiO-66- NH <sub>2</sub> @Mg(OH) <sub>2</sub>	4.0-11.0	10-150	130.39	14
FMBO-S	3.0-10.0	5-120	61.24	15
MOF-76(Ce)	3.0-11.0	5-25	72.97	16
Ca <sub>x</sub> La <sub>1</sub> - <i>x</i> MnO <sub>3</sub>	3.0-11.0	1-10	37.8	17
CaFe <sub>1:2</sub> -700	3.0-11.0	25-300	62–75	18
MgBC600	3.0-11.0	0.5-160	109.35	19
MMBC-200/600	3.0-11.0	75	83.06	20
Ce-BC	3.0-11.0	1-50	16.7	21

OH/NH <sub>2</sub> @MBC	5.0-9.0	-	52.53	22
La-MBC	4.0-8.0	0.5-15	27.49	23
OV-MgO	3.0-11.0	50-100	379.7	24
EFG	4.0-10.0	0-100	49.92	25
LC@AER	6.0-10.0	-	49.89	26
FeCaMg-ALE	2.0-12.0	0-100	20.1	27
Fe/Mn-BMBCs	3.0-11.0	5-150	44.0–53.8	28
Fe <sup>2+</sup> +HFO	2.0-10.0	5-200	51.7	29
La@PAN	3.0-6.0	10-500	83.33	30
MC-hal-2	3.0-11.0	-	136.7	31
Mg-La LDH	3.0-11.0	100-500	87.23	32
CSPGs-La	2.0-10.0	50-350	159.5	33
La-loaded geopolymer	4.0-12.0	5-60	33.65	34
biochar	2.0-10.0	0-400	98.5	35
LDH-biochar	7.0	0-200	160.8	36
CeAC-A	3.0-11.0	5-120	95.47	37
Mg <sub>60</sub> Al <sub>40</sub> -LDH	-	1-50	108.8	38
LZ	3.0-10.0	10-150	122.7	39
MGPA DN hydrogel	3.0-12.0	10-150	38.75	40
ZIF-L@GO	3.0-10.0	1-200	$116.3\pm2.1$	41
MBC	3.0-11.0	25-200	70.26	42
NH <sub>2</sub> -CNS-La	2.0-12.0	5-100	103.01	43
Fe-CLCAB	1.0-9.0	10-200	73.13	44
Ca/(Al-DWTAS)- LDO	2.0-11.0	10-100	110.14	45
CaMgAl-LDH	3.0-11.0	0-150	21.47	46
nano-CaO <sub>2</sub> /BFS	4.0-10.0	5-20	67.48	47
MAC@Zr	3.0-9.0	0-50	14.3	48
LaCCH	3.0-11.0	1-50	70.0	This study

Table S3 Kinetics con	stants for phosphate ads	orption on th	e LaCC	CH.
Kinetic n	$q_e (\mathrm{mg/g})$	$k_l$	R <sup>2</sup>	
like-pseudo-first-order	$q_t = q_e \left(1 - e^{-k_1 t}\right)$	68.4	9.30	0.95
like-pseudo-second-order	$q_t = \frac{q_e^2 k_2 t}{1 + q_e k_2 t}$	71.37	0.21	0.99

Samples	Species	B.E. <sup><i>a</i></sup> (eV)	FWHM <sup>b</sup> (eV)	G:L <sup>c</sup> ratio	Percent <sup>d</sup> (%)		
LaCCH	La-PO <sub>4</sub> <sup>3-</sup>	133.1	1.16	0:100	28.8		
	La-HPO <sub>4</sub> <sup>2-</sup>	132.1	1.27	14:86	71.2		

Table S4 Deconvolution of XPS P 2p spectra for LaCCH after P adsorption.

<sup>*a*</sup>Binding energy (B.E.); <sup>*b*</sup>The full width at half maximum (FWHM); <sup>*c*</sup>Gaussian: Lorentzian ratio; <sup>*d*</sup>The percentage represents the contribution of each peak to the total number of counts under the P 2p peak.

adsorption.						
Samples	Pe	ak	B.E. <sup><i>a</i></sup> (eV)	FWHM <sup><i>b</i></sup> (eV)	G:L <sup>c</sup> ratio	Percent <sup><math>d</math></sup> (%)
LaCCH	La	main	835.2	2.63	0:100	31.5
	La 3d <sub>5/2</sub>	satel lite	839.0	2.4	0:100	29.2
	La 3d <sub>3/2</sub>	main	851.9	2.57	2:98	21.8
		satel lite	855.8	2.3	0:100	17.6
LaCCH+P	La 3d <sub>5/2</sub>	main	835.5	2.1	0:100	23.3
		satel lite	837.8	2.9	93:7	40.1
	m	main	851.1	1.4	0:100	11.2
	La 3d <sub>3/2</sub>	satel lite	854.3	4.5	9:91	25.1

 Table S5 Deconvolution of XPS La 3d spectra for LaCCH sample before and after P

 adsorption

<sup>*a*</sup>Binding energy (B.E.); <sup>*b*</sup>The full width at half maximum (FWHM); <sup>*c*</sup>Gaussian: Lorentzian ratio; <sup>*d*</sup>The percentage represents the contribution of each peak to the total number of counts under the P 2p peak.

 Table S6 Deconvolution of XPS O 1s spectra for LaCCH sample before and after P

 adsorption

adsorption.						
Samples	Spacios	<b>B.E.</b> <i><sup><i>a</i></sup></i>	$FWHM^b$	G.I & ratio	Percent <sup>d</sup>	
	species	(eV)	(eV)	G.L. Tatio	(%)	
LaCCH	La–O	531.7	2.6	0:100	85.0	
	-OH	529.5	1.1	0:100	15.0	
LaCCH +P	La–O	532.0	1.8	0:100	27.9	

-OH	530.5	1.5	18:82	45.4
	531.6	1.1	19:81	26.7

<sup>*a*</sup>Binding energy (B.E.); <sup>*b*</sup>The full width at half maximum (FWHM); <sup>*c*</sup>Gaussian: Lorentzian ratio; <sup>*d*</sup>The percentage represents the contribution of each peak to the total number of counts under the P 2p peak.

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