Efficient simultaneously quantitative and qualitative detection of multiple phenols using highly water-stable Co2+‐doped Cu-BTC as electrocatalyst

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Chemicals

2Copper(II) nitrate trihydrate $(Cu(NO_3)_2.3H_2O, 99.99\%)$, polyvinylpyrrolidone (PVP, $M_w=8,000$), 2-AP, 2-CP and 2-NP compounds were purchased from Aladdin (Shanghai, China). Cobalt(II) nitrate hexahydrate $(Co(NO_3)_2.6H_2O, 99%)$ was acquired from Adamas (Shanghai, China). Ethanol $(C_2H_6O, 99.7%)$ and trimesic acid (BTC, 98%) were secured from Macklin (shanghai, China). Sodium hydroxide (NaOH, 99.7%) was obtained from Chongqing Chuandong Chemical (Chongqing, China). All chemicals were used as received without further purification. Configuration of 0.1 M phosphate-buffered saline (PBS) is that 17.907 g $Na₂HPO₄·12H₂O$, 7.8005 g NaH₂PO₄·2H₂O, 3.7275 g KCl were dissolved in 500 mL of ultrapure water after that the resulting solution was adjusted pH and fixed volume. The 5mM $[Fe(CN)_6]^{3-/4}$ solution is prepared as follows: 0.4116g K₃ $[Fe(CN)_6]$, 0.5280g $K_4[Fe(CN)_6]$ 3H₂O and 1.8638g KCl were dissolved in 250mL PBS solution with pH 7.0 and make up to volume in a volumetric flask.

Instrumentation

X-ray diffraction (XRD) was performed using a Shimadzu XRD-6100 X-ray diffractometer (40 kV/30 mA) with Cu Kα radiation source $(\lambda = 1.540 \text{ Å})$ at a scan rate of 5.0° min⁻¹ and 2θ values in the range of 3-50°. Field emission scanning electron microscopy (FESEM) was conducted using a Hitachi S-4800 electron microscope with an accelerating voltage of 5 kV. Transmission electron microscopy (TEM, JEOL JEM 2100) imaging was performed at an accelerating voltage of 200 kV. Fourier transform infrared (FTIR) spectra of the MOF samples were collected using a Shimadzu IRPrestige-21 spectrometer in the wavenumber region of 500 to 4000 cm-1 . X-ray photoelectron spectroscopy (XPS) of the sample was conducted using the Thermo escalab 250XI instrument with monochromic Al Ka (hv=1486.6eV) and at a power of 150 W. The high-resolution XPS peaks were calibrated concerning the C 1s peak at 284.8 eV and further processed using the Advantage software. Four-point probe resistivity test was conducted using the FM100GH highprecision resistivity test instrument. All the electrochemical measurements were performed at the CHI 660E electrochemical workstation. During the measurement, a standard three-electrode system including a glassy carbon working electrode (GCE, $\phi=3$ mm), platinum wire auxiliary electrode and the saturated calomel reference electrode was adopted.

Preparation of sodium-BTC (Na-BTC) linker

The Na-BTC linker was prepared by adding 120 mg of NaOH and 210 mg of trimesic acid into 150 mL of ethanol. After fully dissolving, the obtained solution was heated to 60 ℃ for 2 h in a water bath. Then, the Na-BTC product underwent centrifugation and ethanol washing three times. Finally, the product was dried at 60 ℃ overnight.

Fig. S1 (A), (B) The structure of Cu-BTC is viewed from different directions.

Fig. S2 (A), (B) SEM images of Cu-BTC.

Fig. S3 (A) SEM of Cu-BTC@Co. (B)-(E) Mapping images of Cu-BTC@Co. Energy-dispersive

X-ray spectroscopy (EDS) spectrum of Cu-BTC@Co(F).

Fig. S4 The XPS survey spectra of Cu-BTC@Co.

Table S1 Relative atomic percentages of Cu, Co, O and C in Cu-BTC@Co samples based on XPS

measurements.

Table S2 Four-point probe resistivity test results for Cu-BTC and Cu-BTC@Co.

The electrochemical active surface area of Cu-BTC@Co/GCE

Fig. S5A illustrated the influence of scan rate on the redox behavior of Cu-BTC@Co/GCE in 2.5 mM $[Fe(CN)_6]^{3-4}$ solution. With the scan rate increased from 0.02 V/s to 0.1 V/s, the redox peak currents also increased with almost no shift in the redox potentials. The plot of the square root of the scan rate against the peak current for the redox process (Fig. S5B) correlation coefficient value coefficients of 0.9989 and 0.9985 for the anodic and cathodic peaks with the peak current, respectively with the regression equations of Ipa (μ A) = 21.01911 v^{1/2} - 0.14104 and Ipc (μ A) = 28.18982 $v^{1/2}$ -1.33768. The electrochemical active surface area of Cu-BTC@Co/GCE is determined to be 1.457 cm² by calculating with the Randles-Sevcik equation.

Fig. S5 (A) CVs of Cu-BTC@Co/GCE as a function of scan rate (20-100 mV/s) and the plot of

the square root of scan rate vs. current (B).

Fig. S6 (A) CV comparison grapes of Cu-BTC@Co/GCE and Cu-BTC/GCE individually detected 2-AP, 2-CP (B) and 2-NP (C). (D) DPVs were obtained for the mixture of 2-AP, 2-CP, and 2-NP in PBS (pH 5.0) at Cu-BTC and Cu-BTC@Co-modified GCE.

Optimization of Experimental Conditions

The pH is one of the important factors affecting the electrochemical performance of electrochemical sensors. The response current of Cu-BTC@Co in 0.1 M PBS with different pH ranges (4.0 - 8.0) at the scan rate of 0.1 Vs⁻¹ was tested by DPVs, and corresponding to the concentration of 2-AP, 2-CP, 2-NP was 4.8 μ M, 0.06 μ M, 4.8 μ M, respectively. The results are shown in Fig. S7A-C, the peak currents at pH 5 of 2-AP and 2-CP are greater than those at pH 4. However, the peak current of the target pollutants sharply declines at pH 6, due to decreased electrochemical activity of the phenols with increased alkalinity of the solution. In addition, when

comparing the response current values of 2-NP at different pH, the value at pH 5.0 showed a small difference from that at pH 6.0. In summary, considering the response currents of these three phenols at different pH values in Cu-BTC@Co/GCE, the subsequent experiment was conducted at pH 5.0.

Owing to the quantity of Cu-BTC@Co on the electrode affecting its response current values, the concentration and the volume of Cu-BTC@Co added during electrode preparation were optimized. Firstly, Cu-BTC@Co concentrations of 0.5 mg/mL, 1mg/mL, 5 mg/mL, and 8 mg/mL were prepared. 5 µL Cu-BTC@Co with varying concentrations was added to the bare electro containing 12 µM 2-AP, 0.15 µM 2-CP and 24 µM 2-NP. The result is shown in Fig. S7D. When the electrochemical sensor prepared was used with Cu-BTC@Co at a concentration of 1 mg/mL, the maximum oxidation peaks were observed at 2-AP, 2-CP and 2-NP. Therefore 1mg/mL Cu-BTC@Co was the optimal concentration. To further investigate the detection performance of different volumes of 1 mg/ml Cu-BTC@Co, 2 μ L, 5 μ L and 10 μ L of 1 mg/mL Cu-BTC@Co were dropwise added onto the electrode as shown in Fig. S7E. The result showed that the sensor prepared by 5 µL of 1mg/mL Cu-BTC@Co exhibited excellent detection performance. To sum up, $5 \mu L$ of 1mg/mL Cu-BTC@Co was selected for preparing the electrochemical sensor.

Fig. S7 (A)-(C) Peak current column-line chart of Cu-BTC@Co/GCE containing 4.8 µM 2-AP, 0.06 µM 2-CP and 4.8 µM 2-NP in 0.1 M PBS (at different pH values: 4.0, 5.0, 6.0, 7.0, 8.0). (D) DPV plots of different concentrations (0.5 mg/ml, 1 mg/ml, 5 mg/ml, 8 mg/ml) and different volumes (E) of Cu-BTC@Co for the detection of the same concentration of 2-AP, 2-CP and 2-NP.

Fig. S8 (A) DPV comparison graphs of tap water and tap water addition phenols. (B) DPVs of 2- AP, 2-CP and 2-NP in tap water samples $(n = 3)$.

Fig. S9 (A) DPVs response of effluent samples with and without the addition of phenolic compounds. (B) DPVs of 2-AP、2-CP and 2-NP in effluent samples.

sample 2-AP			$2-CP$			$2-NP$			
	Added Found					RecoveryAdded Found RecoveryAdded Found			Recovery
	(μM)	(μM)	(%)	(μM)	(μM)	(%)	(μM)	(μM)	$(\%)$
1	9	9.05	100.5% 0.05		0.051	102%	5	5.04	100.8%
$\overline{2}$	11	11.48	104.3% 0.09		0.088	97.8%	8	8.05	100.6%
3	15	15.35	102.3% 0.11		0.1096	99.6%	11	10.76	97.8%

Table S3 Recovery results of 2-AP, 2-CP and 2-NP in tap water.

Scheme 1. Mechanism of oxidation of phenolic compounds at Cu-BTC@Co/GCE