Supplementary Information (SI) for Environmental Science: Nano. This journal is © The Royal Society of Chemistry 2024

## **Supporting Information for**

## Solid Phase Silver Sulfide Nanoparticles Contribute Significantly to Biotic Silver in Agricultural Systems

Yingnan Huang<sup>a#</sup>, Huijun Yan<sup>a,b#</sup>, Fei Dang<sup>a\*</sup>, Zhenyu Wang<sup>c,d</sup>, Jason C. White<sup>e</sup>,

Yujun Wang<sup>a</sup>

<sup>a</sup> State Key Laboratory of Soil and Sustainable Agriculture, Institute of Soil Science,

Chinese Academy of Sciences, Nanjing 211135, China

<sup>b</sup> College of Geography and Environment, Shandong Normal University, Jinan

250358, China

<sup>c</sup> Institute of Environmental Processes and Pollution Control, and School of

Environment and Civil Engineering, Jiangnan University, Wuxi 214122, China

<sup>d</sup> Jiangsu Engineering Laboratory for Biomass Energy and Carbon Reduction

Technology, Jiangnan University, Wuxi 214122, China

<sup>e</sup> Connecticut Agricultural Experiment Station, New Haven, CT 06504, USA

<sup>#</sup> Y.N.H. and H.J.Y. contributed equally to this work.

\*Corresponding author: Fei Dang

Email: fdang@issas.ac.cn



Figure S1. Rainfall data are based on historical re-analysis datasets from the European Centre for Medium-Range Weather Forecasts (ECMWF)/National Aeronautics and Space Administration (NASA), provided by www.xihe-energy.com. The vertical pink line indicates the time when the mesocosms were covered by a temporary roof.



Figure S2. The fresh weight of radish roots from NP<sub>red</sub> and NP<sub>black</sub> mesocosms. NP<sub>red</sub>: mesocosms with NPs in red soil; NP<sub>black</sub>: mesocosms with NPs in black soil. Data shown are the averages of two replicates  $\pm$  SD. All individuals were pooled together for each replicate. No significant difference was observed between the treatments at p > 0.05.



Figure S3. Bioaccumulation of Ag in (a) rice shoots, (b) rice roots, and (c) radish leaves after exposure to NPs. NP<sub>red</sub>: mesocosms with NPs in red soil; NP<sub>black</sub>: mesocosms with NPs in black soil. Data shown are the averages of four replicates  $\pm$  SD for rice shoots and radish leaves, and the averages of two replicates  $\pm$  SD for rice roots.



Figure S4. The available data on the levels of soil-borne NPs. The data are available in the literature.<sup>1-3</sup>

## Table S1. Physical and chemical properties of the sampled soils.

		Organic matter		CEC <sup>c</sup>	Total nitrogen <sup>d</sup>	Total Ag	Total Fe
Sample	Texture	content (g kg <sup>-1</sup> ) <sup>a *</sup>	pH <sup>b</sup> *	(mol kg <sup>-1</sup> )	(%)	$(mg kg^{-1})*$	$(g kg^{-1})^*$
Red soil	Loamy clay	$16.9\pm0.03$	$5.0\pm0.04$	12.3	0.16	$0.4\pm0.01$	37.1 ± 1.7
Black soil	Loamy clay	$27.6\pm0.1$	$6.7\pm0.1$	29.0	0.15	$0.2\pm0.01$	$28.6\pm0.9$

<sup>a</sup> Determination of organic matter by potassium dichromate oxidation and external heating. Data shown are the averages of two replicates  $\pm$  SD (n=2).

<sup>b</sup> pH was determined by potentiometric method with soil-water ratio 1:5. Data shown are the averages of two replicates  $\pm$  SD (n=2).

<sup>c</sup> Determination of cation exchange capacity (CEC) by EDTA-ammonium salt.

<sup>d</sup> Determination of total nitrogen by semi-trace Kjeldahl method.

\*A significant difference was observed between the soils (p < 0.05).

Table S2. Concentrations of major cations in the water column before NPs addition. The water was collected from a local Reservoir. The cations were measured by Inductively Coupled Plasma Optical Emission Spectrometry (iCAP7400, Thermo Fisher, USA). Data shown are the averages of four replicates  $\pm$  SD.

Cations (mg L <sup>-1</sup> )	NP <sub>Red</sub> mesocosm	$NP_{Black}$ mesocosm
$\mathrm{K}^{+}$ *	$2.5\pm0.7$	$0.8 \pm 0.4$
Ca <sup>2+ *</sup>	$8.3\pm0.7$	$76.1 \pm 8.2$
$Mg^{2+}*$	$5.2 \pm 1.0$	$13.8 \pm 1.0$

\* A significant difference was observed between the soils (p < 0.05).

Instance	Abundance			
Isotope	Ag <sub>2</sub> S-NPs <sup>a</sup>	$^{109}Ag_2S-NPs^b$		
<sup>107</sup> Ag	0.5180	0.0042		
<sup>109</sup> Ag	0.4820	0.9958		

Table S3. The isotope abundances of  $^{109}Ag_2S$ -NPs and  $Ag_2S$ -NPs.

<sup>a</sup> The abundance of <sup>107</sup>Ag and <sup>109</sup>Ag in Ag<sub>2</sub>S-NPs are the natural abundances of Ag, respectively.<sup>4</sup>

<sup>b</sup> The abundances of <sup>107</sup>Ag and <sup>109</sup>Ag in <sup>109</sup>Ag<sub>2</sub>S-NPs are calculated based on the determined concentrations of <sup>107</sup>Ag and <sup>109</sup>Ag by ICP-MS. Data shown are the averages of ten replicates.

Soil type	Plant	Time	Relative bioavailability of water-borne NPs to soil-borne NPs
	Duorra nico	1 <sup>st</sup> rice season	$2.4 \pm 1.5^{\mathrm{a}}$
	Brown rice	2 <sup>nd</sup> rice season	$0.2\pm0.1^{\mathrm{a}}$
NP <sub>red</sub> mesocosms			
	Duckweed	2 <sup>nd</sup> rice season	$8.1\pm0.01^{\text{b}}$
	Rice borer	2 <sup>nd</sup> rice season	$1.3 \pm 1.8^{b}$
	Brown rice	1 <sup>st</sup> rice season	$3.1\pm0.6^{\mathrm{a}}$
	Diowii nee	2 <sup>nd</sup> rice season	$1.8\pm0.6^{\mathrm{a}}$
NP <sub>black</sub> mesocosms			
	Duckweed	2 <sup>nd</sup> rice season	$6.8\pm2.8^{b}$
	Rice borer	2 <sup>nd</sup> rice season	$2.8 \pm 1.0^{\mathrm{b}}$
NP <sub>red</sub> mesocosms	Radish roots	-	$2.6\pm0.5^{\mathrm{a}}$
NP <sub>black</sub> mesocosms	Radish roots	-	$4.8\pm0.7^{\mathrm{a}}$

Table S4. The relative bioavailability of water-borne to soil-borne NPs. This is calculated as the percentage of biotic Ag from initially water-borne NPs (i.e., normalized by their exposure concentrations) divided by the percentage of biotic Ag from the soil-borne NPs.

<sup>a</sup> Data shown are the averages of four replicates  $\pm$  SD.

<sup>b</sup> Data shown are the averages of two replicates  $\pm$  SD.

Table S5. The soil-to-plant transfer factor for soil-borne NPs (TF, the ratio of Ag concentrations in edible tissues derived from soil-borne NPs to their soil-borne NPs levels).

Soil type	Plant	Time	TF value
ND magaaaama	Drown rice	1 <sup>st</sup> rice season	2.1×10-2
NP <sub>red</sub> mesocosms	DIOWIIIICE	2 <sup>nd</sup> rice season	1.4×10 <sup>-2</sup>
ND masaaasma	Proven rico	1 <sup>st</sup> rice season	1.8×10 <sup>-2</sup>
INF black Incsocosins	DIOWITICE	2 <sup>nd</sup> rice season	4.6×10 <sup>-3</sup>
NP <sub>red</sub> mesocosms	Radish roots	-	7.4×10 <sup>-2</sup>
NP <sub>black</sub> mesocosms	Radish roots	-	1.5×10 <sup>-2</sup>

## REFERENCES

1. A. Hong, Q. Tang, A. U. Khan, M. Miao, Z. Xu, and F. Dang, *et al.*, Identification and speciation of nanoscale silver in complex solid matrices by sequential extraction coupled with inductively coupled plasma optical emission spectrometry. *Anal. Chem.*, 2021, **93**, 1962-1968.

2. L. Li, Q. Wang, Y. Yang, L. Luo, R. Ding, and Z.G. Yang *et al.*, Extraction method development for quantitative detection of silver nanoparticles in environmental soils and sediments by single particle inductively coupled plasma mass spectrometry. *Anal. Chem.*, 2019, **91**, 9442-9450.

3. P. Wang, N. W. Menzies, P. G. Dennis, J. Guo, C. Forstner, and R. Sekine, *et al.*, Silver nanoparticles entering soils via the wastewater–sludge–soil pathway pose low risk to plants but elevated Cl concentrations increase Ag bioavailability. *Environ. Sci. Technol*, 2016, **50**, 8274-8281.

4. F. Dang, Y. Z. Chen, Y. N. Huang, H. Hintelmann, Y. B. Si and D. M. Zhou, Discerning the sources of silver nanoparticle in a terrestrial food chain by stable isotope tracer technique. *Environ. Sci. Technol*, 2019, **53**, 3802-3810.