## From defence to damage: The impact of seawater passivation on microbially influenced corrosion in CuNi 70/30 alloy

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## **Supplementary Data**

**Table S1.** Chemical composition of the natural seawater used for passivation and corrosion testing in this study.

Test	Units	Seawater	
Chloride	mg 1-1	20000	
Calcium	mg l <sup>-1</sup>	540	
Magnesium	mg l <sup>-1</sup>	1600	
Potassium	mg l <sup>-1</sup>	530	
Sodium	mg l-1	10000	
Iron	mg l <sup>-1</sup>	< 0.05	
Phosphate	mg l <sup>-1</sup>	0.08	
Sulphate	mg l <sup>-1</sup>	2600	
Nitrate	mg l <sup>-1</sup>	< 0.02	
Total alkalinity	mg l <sup>-1</sup>	170	
Total organic carbon	mg l <sup>-1</sup>	< 5	
рН	pH units	8.0	
Conductivity	μS cm <sup>-1</sup>	63000	

**Table S2.** EDS elemental composition of non-passivated and seawater passivated samples.

Sample types	Elemental composition (at %)						
	Cu	Ni	Fe	Mn	Cl	0	
Non-passivated	65.7 ± 0.1	$32.7 \pm 0.3$	0.6 ± 0.1	$1.0 \pm 0.1$	-	-	
Seawater passivated	$44.7\pm0.4$	$23.6 \pm 0.2$	$0.6 \pm 0.0$	$0.7\pm0.0$	0.3 ± 0.1	30.1 ± 0.4	



**Fig. S1.** SEM images after removal of surface film formed on seawater passivated samples for 35 days. The insets provide higher magnification images of the corresponding coupon surface.

## Non-passivated sample



**Fig. S2.** 3D optical profilometer images showing biofilm thickness on the non-passivated and seawater passivated samples following immersion in SRB inoculated MB medium for 28 days. The regions on the right-hand side of the images represent areas where the biofilm was removed, revealing the CuNi alloy surfaces.



**Fig. S3.** EDS spectra of biofilm formed on the non-passivated and seawater passivated samples following immersion in SRB inoculated MB medium for 28 days.