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SUPPLEMENTAL INFORMATION

for

Effect of intracellular algal organic matter and nitrate on disinfection byproduct formation in chlorinated water after UV/H₂O₂ and UV/Cl₂ advanced oxidation processes

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Table S1. Preliminary Data for Selecting the Filter				
Type of Filter	TOC (mg/L)	TN (mg/L)		
Ultrapure water	0.082	Not detectable		
Mixed cellulose ester 0.45 reagent water	0.049	Not detectable		
Nylon 0.45 reagent water	1.001	0.235		

Table S2. THM4 in Standard Solution

Compound	Compound abv	CAS	Concentration	RT (min)
Bromodichloromethane	BDCM	(75-27-4)	200 µg/mL	11.4
Bromoform	BF	(75-25-2)	200 µg/mL	28.6
Chloroform	CF	(67-66-3)	200 µg/mL	8.43
Dibromochloromethane	DBCM	(124-48-1)	200 µg/mL	22.3

THM and HAA extraction protocol:

For THMs liquid-liquid extraction consisted of adding 3.0 mL of methyl tert-butyl ether (MTBE) and 4 g Na₂SO₄ to 50 mL of the sample, shaking vigorously for 11 min, and then inverting for five minutes to allow water and MTBE phases to separate. After that, 1 mL of the MTBE phase was transferred to an autosampler vial, 10 μ L of 4-bromofluorobenzenewas added as an internal standard, and vials were stored at -20°C until analysis.

The liquid-liquid extraction method for HAAs consisted of transferring 40 mL of the water sample to a precleaned 60-mL glass vial with a PTFE-lined screw cap using a clean graduated cylinder for each sample, adding 2 mL concentrated sulfuric acid (97% ACS grade) and 16 g of Na₂SO₄, and shaking vigorously by hand until all Na₂SO₄ was dissolved. Next, 3.0 mL of MTBE with internal standard (120 µg/L of 1,2-dibromopropane) was added to sample and the mixture was shaken vigorously for 14 min, after which the phases were allowed to separate for 5 mins. Then 2 mL of the upper MTBE layer was transferred to a 15 mL centrifuge tube, mixed with 1 mL of 15% sulfuric acid in methanol, sealed and heated in a water bath at 40°C for 160 min. After the tubes cooled to room temperature, 8.5 mL of a 129 g/L Na₂SO₄ solution was added to each centrifuge tube, and upon separation the lower layer was discarded. Finally, 1 mL of

saturated NaHCO₃ solution was added, and the upper layer was transferred to an autosampler vial and stored at -20° C until analysis.



Figure S1. GC-ECD Standard curves of THMs: (a) chloroform, (b) bromodichloromethane, (c) dibromochloromethane, and (d) bromoform

Compound	Compound abv	CAS	Concentration
Bromochloroacetic acid	BCAA	(5589-96-8)	400 μg/mL
Bromodichloroacetic acid	BDCAA	(71133-14-7)	400 µg/mL
Chlorodibromoacetic acid	CDBAA	(5278-95-5)	1000 μg/mL
Dibromoacetic acid	DBAA	(631-64-1)	200 µg/mL
Dichloroacetic acid	DCAA	(79-43-6)	600 μg/mL
Monobromoacetic acid	MBAA	(79-08-3)	400 µg/mL
Monochloroacetic acid	MCAA	(79-11-8)	600 μg/mL
Tribromoacetic acid	TBAA	(75-96-7)	2000 μg/mL
Trichloroacetic acid	TCAA	(76-03-9)	200 μg/mL

Table S3. HAA9 in standard solution



Figure S2. GC-ECD standard curves for HAAs (continued on next page)



Figure S2. (continued) GC-ECD standard curves for HAAs

υ	Set S4. Optimized LC-MS/MS conditions for NDWA detection						
	Compounds	Parent Ion	Product Ion	Collision	Cell Accelerator		
		(m/z) [M+H] ⁺	(m/z) [M+H] ⁺	Energy (eV)	Voltage		
	NDMA	75	43	15	4		
	NDMA-d6	81	46	25	4		
	NDPA-d14	145	97	25	4		

Table S4. Optimized LC-MS/MS conditions for NDMA detection



Figure S3. LC-MS/MS Calibration Curve for NDMA