

Supplementary Materials

Text S1 Characterization and chemical analyses

The Brunauer–Emmett–Teller (BET) surface area, pore size distribution, and pore volume of biochar were measured according to the N₂ adsorption–desorption isotherm at 77 K (Quantachrome, Autosorb EVO, USA). The surface morphologies and crystalline phases of the adsorbents were characterized by scanning electron microscopy (SEM) (S-4800, Hitachi, Japan) and X-ray diffraction (XRD) (Smartlab 9, Rigaku Corporation, Japan), respectively.

Rifampicin wastewater solution was filtered through Whatman GF/F filters with a nominal pore size 0.45- μ m to obtain the DOM solution. Dissolved organic carbon (DOC) was measured by a TOC-L analyzer (Shimadzu Inc., Japan) as DOM content. Chemical oxygen demand (COD) was measured using a fast confined catalytic digestion-spectrophotometer method. Potassium dichromate was used as oxidant and mercury sulfate as a screening agent was used to remove main disturbing ion, chlorine ion.

UV–Vis spectra of DOM was collected using a Shimadzu UV-2600 spectrophotometer at 25 °C. The wavelength range was set from 200 to 600 nm with a 1 nm intervals. Ultrapure water (Milli-Q) was used for the blank reference. Absorbance values at 355 nm (a_{355}) and 254 nm (UV_{254}) were used to quantify the relative content of CDOM and aromatic structures in rifampicin wastewater. SFS of CDOM were collected using a molecular fluorescence spectrometer (F-7000, Hitachi, Japan) in the range of 250–600 nm with a constant offset of 40 nm. Raman scattering was eliminated by deducting ultrapure water (Milli-Q) as a blank subtraction. Solid powders were collected by freeze-drying the CDOM solutions for FTIR analysis using a Nicolet Nexus FTIR spectrometer.

Figures

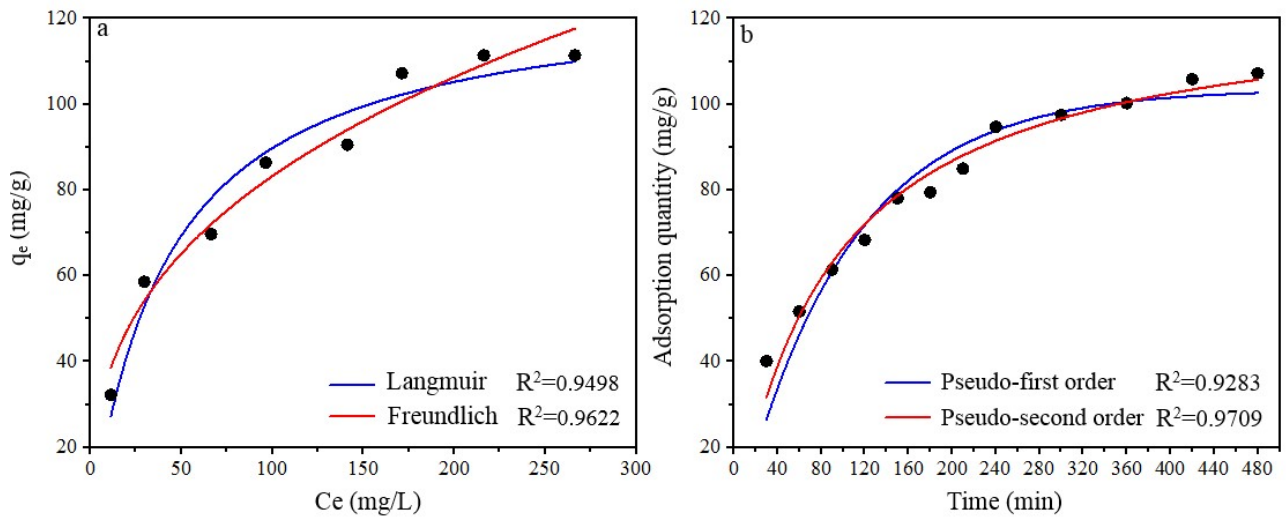


Fig. S1 The adsorption isotherm model (a) and kinetics model (b)

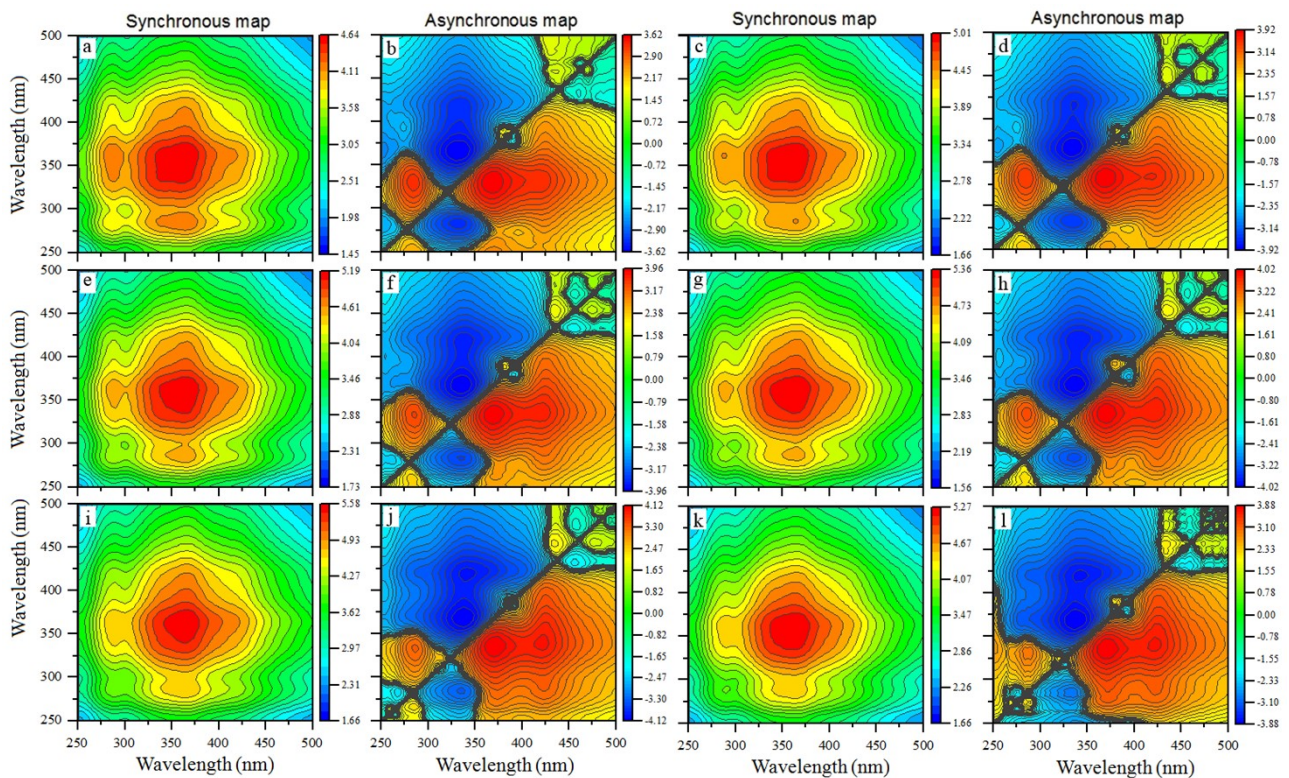


Fig. S2 2D-COS maps of rifampicin wastewater adsorbed by CSB in different COD concentrations: (a, b) 150 mg/L; (c, d) 200 mg/L; (e, f) 250 mg/L; (g, h) 300 mg/L; (i, j) 350 mg/L; and (k, l) 400 mg/L;

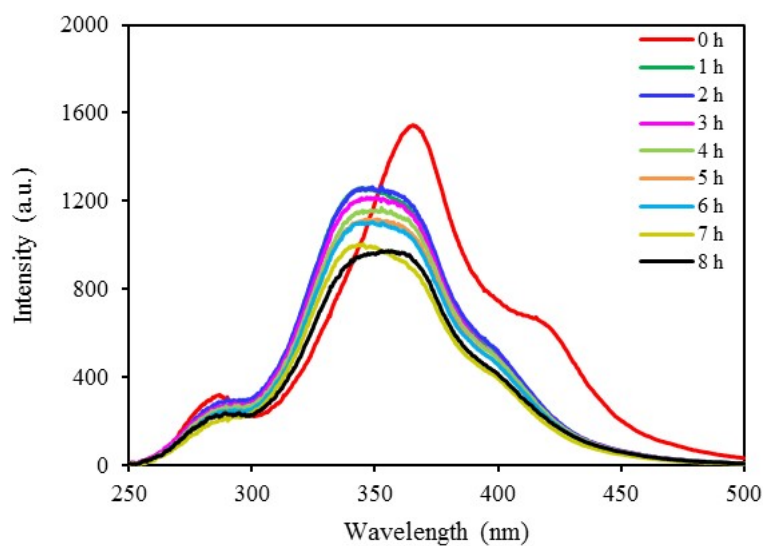


Fig. S3 The changes of fluorescent CDOM in adsorption dynamics process (COD=300 mg/L)

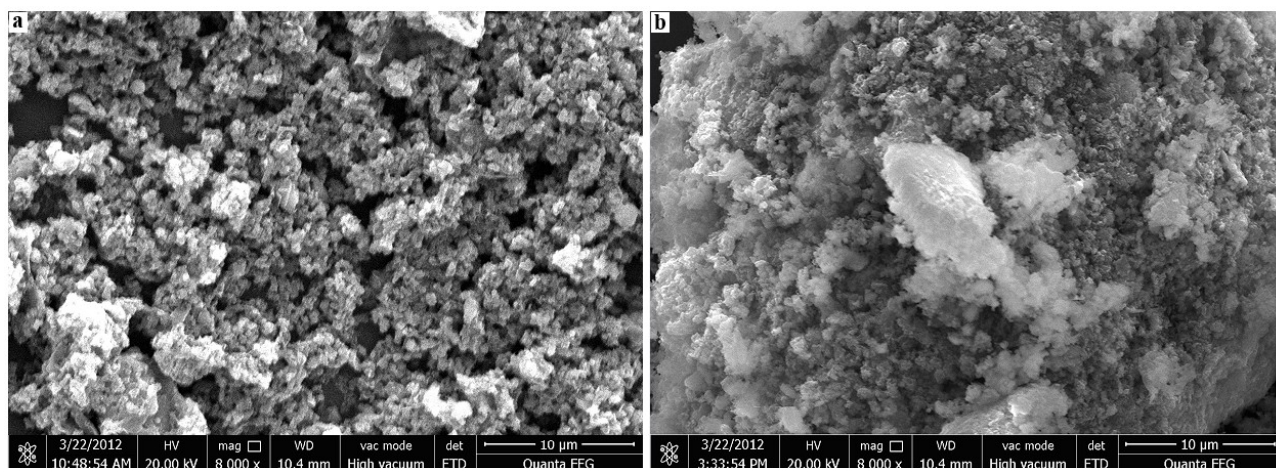


Fig. S4 SEM of CSB (a) before adsorption; (b) after adsorption

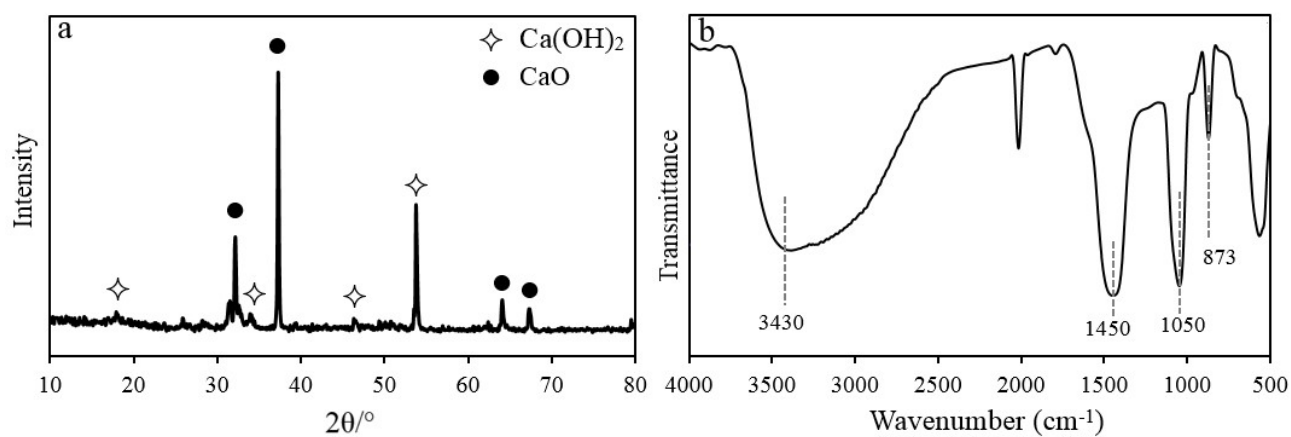


Fig S5 XRD and FTIR of CSB

Tables

Table S1 The positive and negative correlation coefficient of cross-peaks identified by the 2D-SFS-COS

$\begin{matrix} x1 \\ \backslash \\ x2 \end{matrix}$	334	367	375	390	402	422	465
284	-	+	+	+	+	+	+
334		+	+	+	+	+	+
367			+	-	+	+	-
375				-	-	+	-
390					+	+	-
402						+	-
422							-

1450

cm⁻¹

(carboxyl $\delta_{\text{O-H}}$, alkane δ_{CH_3} or skeleton vibration $\nu_{\text{C=C}}$), 1050 cm⁻¹ (alcohol or ethers $\nu_{\text{C-O}}$), 873 cm⁻¹ (the rocking vibration of CaO, nitro-compounds $\nu_{\text{C-N}}$), 920 cm⁻¹ (carboxyl $\gamma_{\text{C-OH}}$), 1225 cm⁻¹ (aryl group $\nu_{\text{C-N}}$, phenolic $\nu_{\text{C-O}}$), 722 cm⁻¹ (amide $\gamma_{\text{N-H}}$), and 1700-2000 cm⁻¹ (carbonyl group $\nu_{\text{C=O}}$).

Table S2 2D-FTIR-COS results on the assignment of cross- and auto- peaks in synchronous and asynchronous maps.

Wavenumber (cm ⁻¹)	Assignment
1700-2000	carbonyl group $\nu_{\text{C=O}}$
1450	carboxyl $\delta_{\text{O-H}}$, alkane δ_{CH_3} or skeleton vibration $\nu_{\text{C=C}}$
1225	aryl group $\nu_{\text{C-N}}$, phenolic $\nu_{\text{C-O}}$
1050	alcohol or ethers $\nu_{\text{C-O}}$
920	carboxyl $\gamma_{\text{C-OH}}$
873	the rocking vibration of CaO, nitro-compounds $\nu_{\text{C-N}}$
722	amide $\gamma_{\text{N-H}}$

Table S3 The positive and negative correlation coefficient of cross-peaks identified by the 2D-FTIR-COS

$\begin{matrix} x1 \\ \backslash \\ x2 \end{matrix}$	873	914	1050	1225	1450	1700-2000
722	+	+	+	+	+	-
873		-	+	-	+	-
914			+	-	+	-
1050				-	+	-
1225					+	-
1450						-

Table S4 BET parameters of CSB before and after adsorption

Sample	BET (m ² /g)	Pore volume (cm ³ /g)	Pore size (Å)
Before	111.86	0.614	219.31
After	61.41	0.476	167.88