

High-throughput selection of (new) enzymes: phage display-mediated isolation of alkylhalide hydrolases from a library of active-site mutated epoxide hydrolases

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Supporting Tables

Table S1 Oligonucleotides used in the construction of mutant cDNA library

Name	Sequence 5'→3'	Ratios in PCR	Mutation(s)	Note
Fp_PUC	CCCAGTCACGACGTTGTAAAACG		-	Fragment 1, forward primer
Rp_HYX1	TATAGCCTTTAGCCCCTCAAC		-	Fragment 1, reverse primer
Fp_HYN1A	GCTAAAGGCTATATATGGGGAAGATNDTNDTATATCGAGATTCAGGTACCGG	144	H153X, Y154X	Fragment A, forward primer
	GCTAAAGGCTATATATGGGGAAGATNDTVMAATATCGAGATTCAGGTACCGG	72		
	GCTAAAGGCTATATATGGGGAAGATNDTATGATATCGAGATTCAGGTACCGG	12		
	GCTAAAGGCTATATATGGGGAAGATNDTTGGATATCGAGATTCAGGTACCGG	12		
Fp_HYV1B	GCTAAAGGCTATATATGGGGAAGATVMAVMAATATCGAGATTCAGGTACCGG	36		
	GCTAAAGGCTATATATGGGGAAGATVMANDTATATCGAGATTCAGGTACCGG	72		
	GCTAAAGGCTATATATGGGGAAGATVMAATGATATCGAGATTCAGGTACCGG	6		
	GCTAAAGGCTATATATGGGGAAGATVMATGGATATCGAGATTCAGGTACCGG	6		
Fp_HYA1C	GCTAAAGGCTATATATGGGGAAGATATGATGATATCGAGATTCAGGTACCGG	1		
	GCTAAAGGCTATATATGGGGAAGATATGNDTATATCGAGATTCAGGTACCGG	12		
	GCTAAAGGCTATATATGGGGAAGATATGVMAATATCGAGATTCAGGTACCGG	6		
	GCTAAAGGCTATATATGGGGAAGATATGTGGATATCGAGATTCAGGTACCGG	1		
Fp_HYT1D	GCTAAAGGCTATATATGGGGAAGATTGGTGGATATCGAGATTCAGGTACCGG	1		
	GCTAAAGGCTATATATGGGGAAGATTGGNDTATATCGAGATTCAGGTACCGG	12		
	GCTAAAGGCTATATATGGGGAAGATTGGVMAATATCGAGATTCAGGTACCGG	6		
	GCTAAAGGCTATATATGGGGAAGATTGGATGATATCGAGATTCAGGTACCGG	1		
Rp_PF1	CTGTATGGCGCTAGGAC			Fragment A, reverse primer
Fp_PFN1A	ACATACCGCGATCCTGCANDTNDTATTTTCCTAAAGGCAAAGGCC	144	P188X, F189X	Fragment B, forward primer
	ACATACCGCGATCCTGCANDTVMATTTTCCTAAAGGCAAAGGCC	72		
	ACATACCGCGATCCTGCANDTATGTATTTTCCTAAAGGCAAAGGCC	12		
	ACATACCGCGATCCTGCANDTTGGTATTTTCCTAAAGGCAAAGGCC	12		
Fp_PFV1B	ACATACCGCGATCCTGCAMAVMAATTTTCCTAAAGGCAAAGGCC	36		

	ACATACCGCGATCCTGCAV MANDT TATTTTCCTAAAGGCAAAGGCC	72		
	ACATACCGCGATCCTGCAV MAATGT TATTTTCCTAAAGGCAAAGGCC	6		
	ACATACCGCGATCCTGCAV MATGGT TATTTTCCTAAAGGCAAAGGCC	6		
Fp_PFA1C	ACATACCGCGATCCTGCA ATGATG TATTTTCCTAAAGGCAAAGGCC	1		
	ACATACCGCGATCCTGCA ATGTGGT TATTTTCCTAAAGGCAAAGGCC	12		
	ACATACCGCGATCCTGCA ATGVMA TATTTTCCTAAAGGCAAAGGCC	6		
	ACATACCGCGATCCTGCA ATGTGGT TATTTTCCTAAAGGCAAAGGCC	1		
Fp_PFT1D	ACATACCGCGATCCTGCA TGGTGGT TATTTTCCTAAAGGCAAAGGCC	1		
	ACATACCGCGATCCTGCA TGGNDT TATTTTCCTAAAGGCAAAGGCC	12		
	ACATACCGCGATCCTGCA TGGVMA TATTTTCCTAAAGGCAAAGGCC	6		
	ACATACCGCGATCCTGCA TGGATG TATTTTCCTAAAGGCAAAGGCC	1		
Rp_TYR1	GTAACTGCACCAAGTAAACC			Fragment B, reverse primer
Fp_TYRN1A	GTTTCACTGGTGCAGTAACTAT NDT CGTGCTTTACCCATAAACTGG	6	Y235X	Fragment C, forward primer
Fp_TYRV1B	GTTTCACTGGTGCAGTAACTAT VMAC GTGCTTTACCCATAAACTGG	3		
Fp_TYRA1C	GTTTCACTGGTGCAGTAACTAT ATG CGTGCTTTACCCATAAACTGG	0.5		
Fp_TYRT1D	GTTTCACTGGTGCAGTAACTAT TGG CGTGCTTTACCCATAAACTGG	0.5		
Rp_PUC	AGCGGATAACAATTTACACAGG			Fragment C, reverse primer
Fp_PUC_EH	CTGAAATGAGCTGTTGACAATTAATCATCGGCTCGTATAATG			Fragment 1A, forward primer
Rp-BamHI	CAGGATCCCGGTATGTCAATATTTTCTTAAGAACAGACTTAGCACC			Fragment 1A, reverse primer
Fp_BamHI	GGTGCTAAGTCTGTTCTTAAGAAAATATTGACATACCGGGATCCTG		R184 silent mutation,	Fragment BC, forward primer, introducing <i>Bam</i> HI site
Rp_PUC_EH	GACCATGATTACGCCAAGCTTGCAATGC			Fragment BC, reverse primer
Fp_PCEH	CGGATAACAATTTACACAGG			Forward primer, StEH
Rp_BCEH	GGAAACAGCTATGACCATG			Reverse primer, StEH
NcoI_fp	AATCCTCTTCATCCACGGCTCCCTGAACT		H31H, silent mutation	Forward primer, removing one <i>Nco</i> I site
Paul_fp	CGACTGGGGCGCGCTAATTGCTTGCCATT		L109L, silent mutation	Forward primer, introducing <i>Paul</i> site
fp_D105A	GTGTTTGTCTGTTGCGCACGCCTGGGGCGCGTTAATTGCTTGG			Forward primer for D105A mutagenesis
rp_D105A	CCAAGCAATTAACGCGCCCCAGGCGTGCGCAACGACAAACAC			Reverse primer for D105A mutagenesis

Table S2 Deduced amino acid sequences of selected protein variants

Clone #	Amino acid position					Comments
	H153	Y154	P188	F189	Y235	
HYPFV-2_1, -2_2, -3_1, -3_2, -3_3	○ ^a	○	○	○	V (gtt)	
HYPFI-2, -3_1, -3_2, -3_3	○	○	○	○	I (att)	
HYPFR-1_1	○	○	○	○	R (cgt)	
HYPFR-1_2	○	○	○	○	R (cgt)	E198D (gag→gac)
HYPSE-2, -3	○	○	○	S (agt)	E (gaa)	
HYPFC-1, -3	○	○	○	○	C (tgt)	
HYPFD-1	○	○	○	○	D (gat)	
HYPFG-1	○	○	○	○	G (ggt)	
HYPFH-2	○	○	○	○	H (cat)	
VEPFR-1	V (gtt)	E (gaa)	○	○	R (cgt)	S311N (agc→aac)
GYFPV-1	G (ggt)	○	F (ttt)	P (cca)	V (gtt)	
HYFAV-3	○	○	F (ttt)	A (gca)	V (gtt)	
YNPFH-2	Y (tat)	N (aat)	○	○	H (cat)	
HYACK-1	○	○	A (gca)	C (tgt)	K (aaa)	
HYFKY-2	○	○	F (ttt)	K (aaa)	○	
HWPFT-2	○	W (tgg)	○	○	T (aca)	
ISTMK-3	I (att)	S (agt)	T (aca)	M (atg)	K (aaa)	
QMLGY-3	Q (caa)	M (atg)	L (ctt)	G (ggt)	○	
HYMMY-3	○	○	M (atg)	M (atg)		
CEYFC-3	C (tgt)	E (gaa)	Y (tat)	○	C (tgt)	
VKPFY-3	V (gtt)	K (aaa)	○	○	○	
HYFHY-3	○	○	F (ttt)	H (cat)	○	
HYAKM-3	○	○	A (gca)	K (aaa)	M (atg)	
HYEWW-3	○	○	E (gaa)	W (tgg)	W(tgg)	
WAPFH-3	W (tgg)	A (gca)	○	○	H (cat)	
HYYAW-3	○	○	Y (tat)	A (gca)	W (tgg)	
HYQQL-3	○	○	Q (caa)	Q (caa)	L (ctt)	
EAPFY-3	E (gaa)	A (gca)	○	○	○	
EQPQL-3	E (gaa)	Q (caa)	○	Q (caa)	V (gtt)	
KKMAW-3	K (aaa)	K (aaa)	M (atg)	A (gca)	W (tgg)	

^a ○, no substitution. Naming is according to amino acid substitution in the linear sequence. The number following the one-letter sequence refers to the round of phage display selection from which the clone was isolated.

Supporting Figures

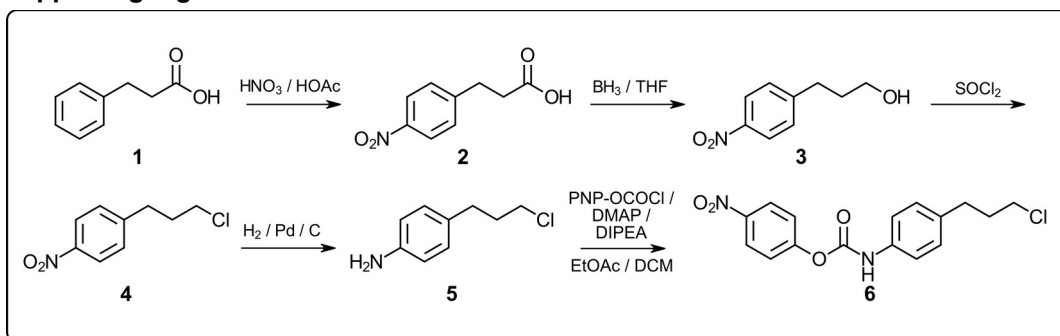


Fig. S1 Synthesis of building block 6 (see **Figs. S4–S7** for NMR spectra).

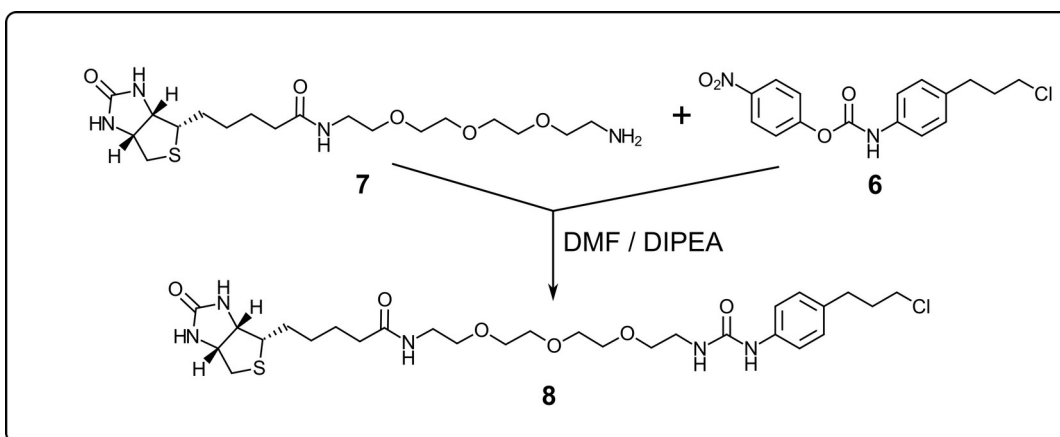


Fig. S2 Block synthesis of bait substrate (8) used in phage display selection (see **Fig. S8** for NMR spectra).

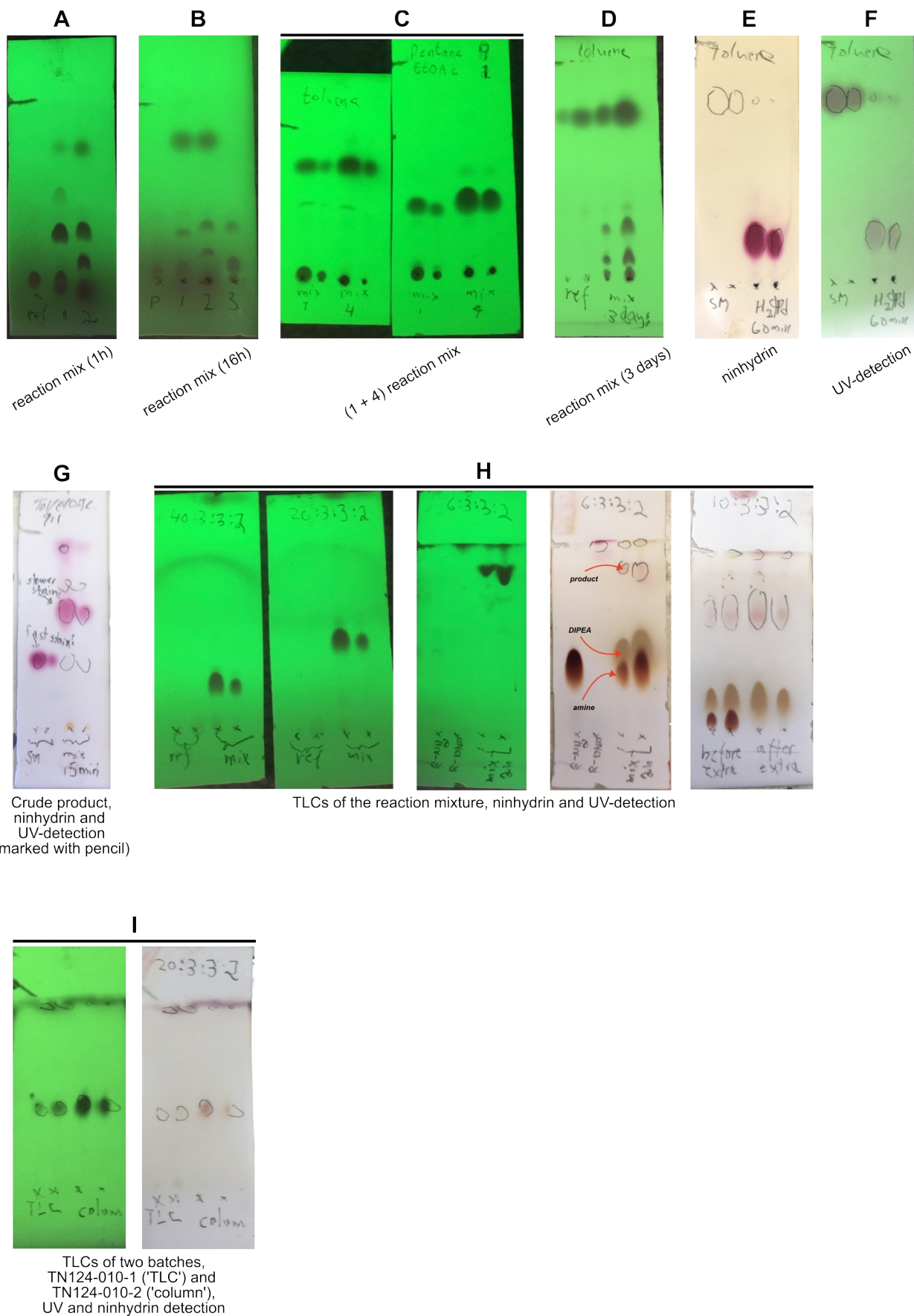


Fig. S3 TLC chromatograms of synthesis intermediates leading to compound **8**.

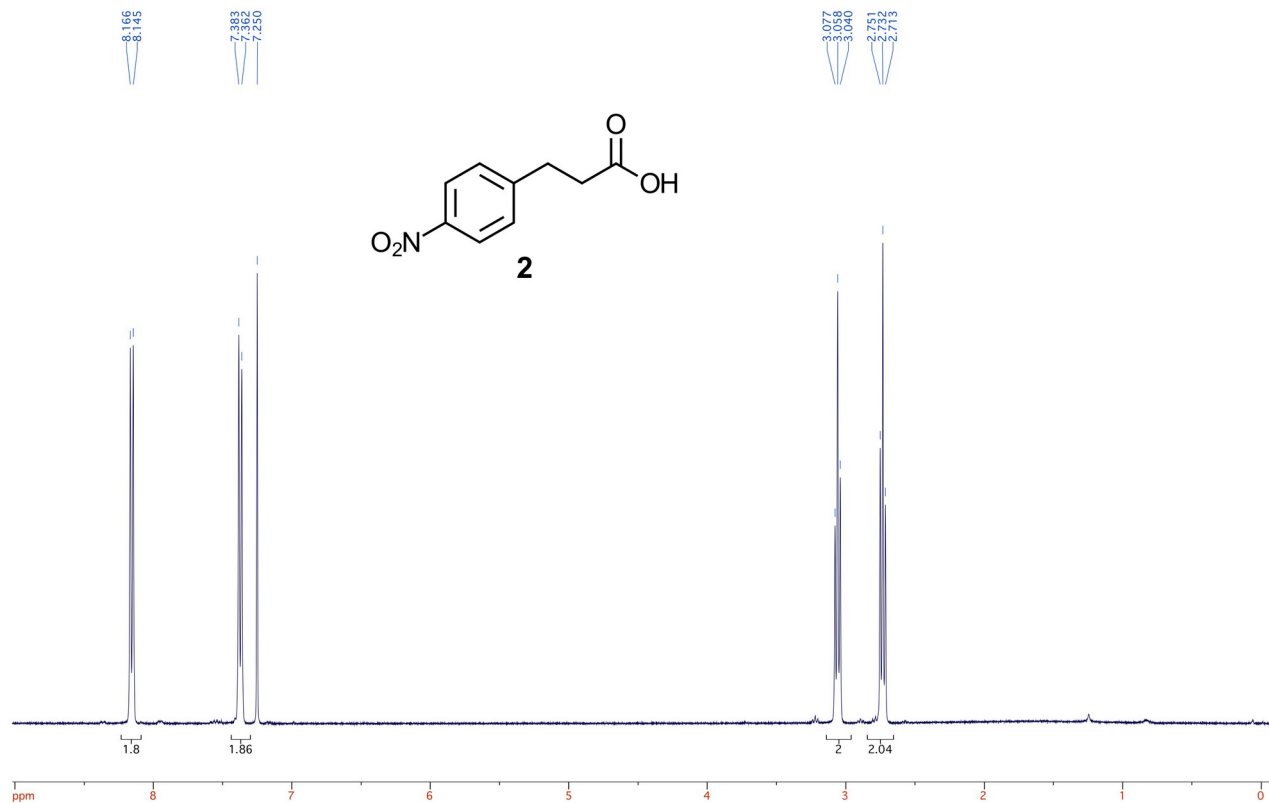


Fig. S4 ¹H-NMR spectrum of compound **2**.

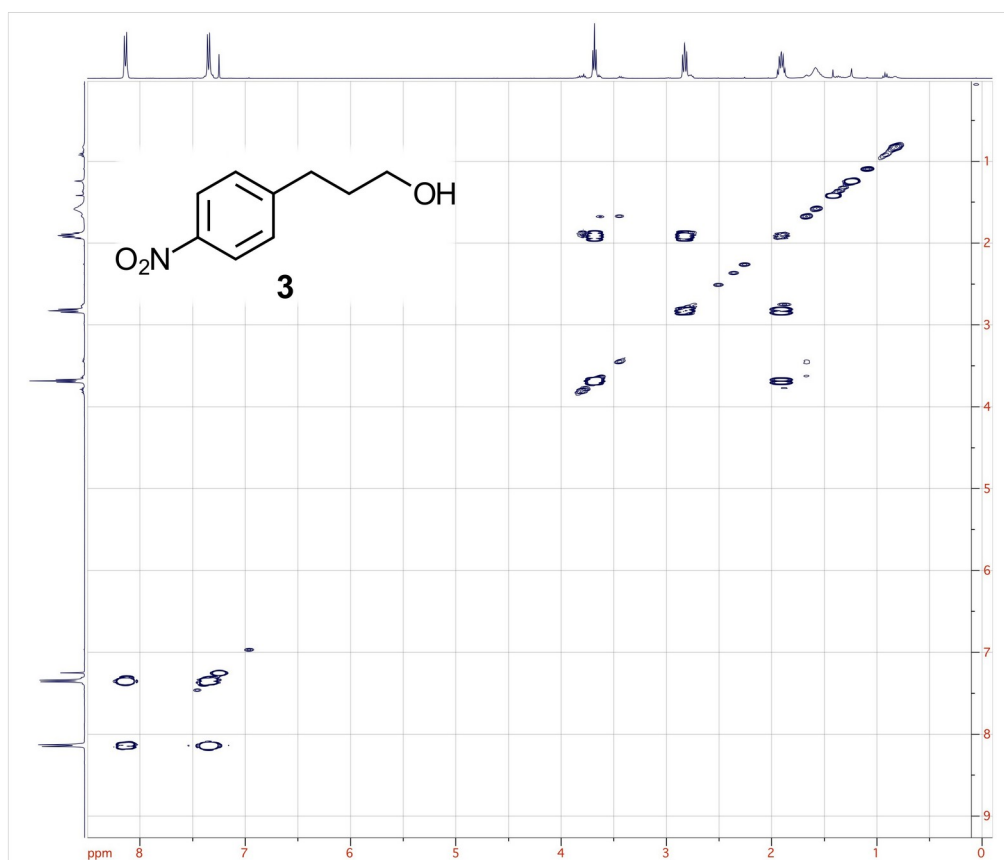
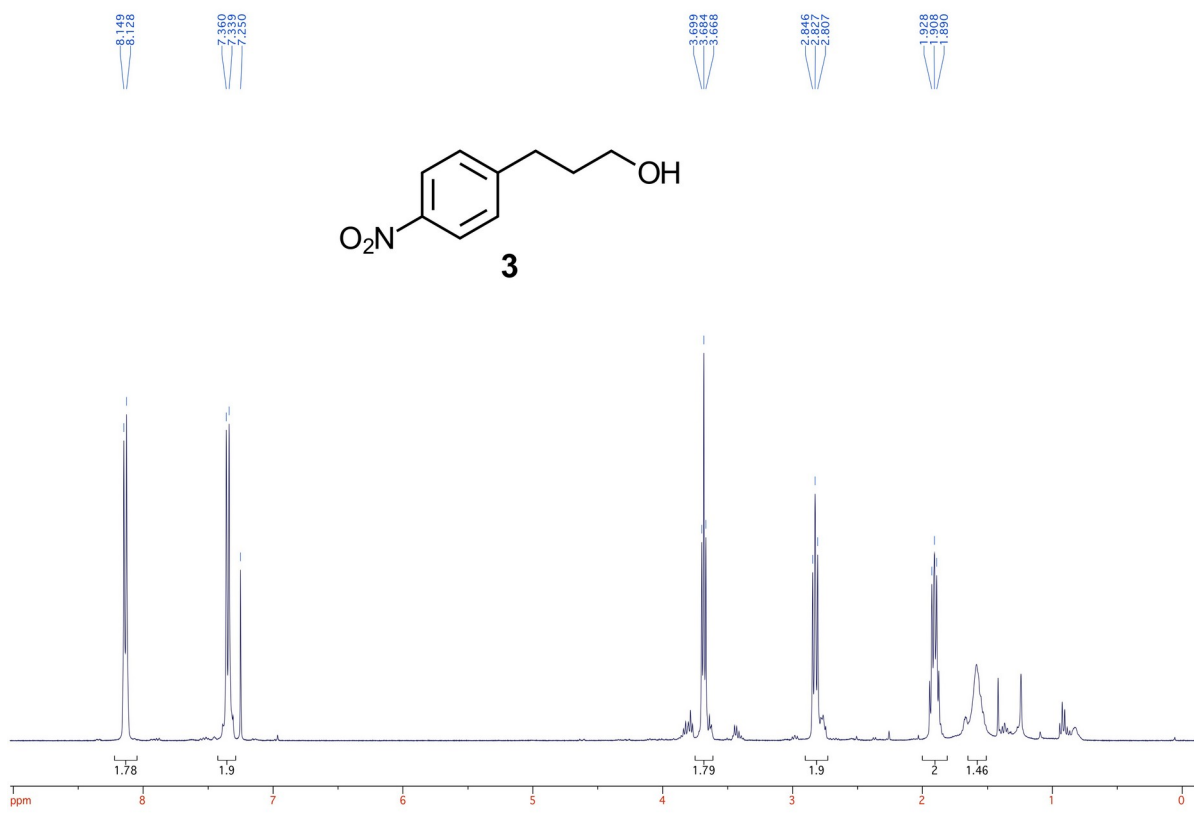


Fig. S5 ¹H-NMR (top) and COESY (bottom) spectra of compound 3.

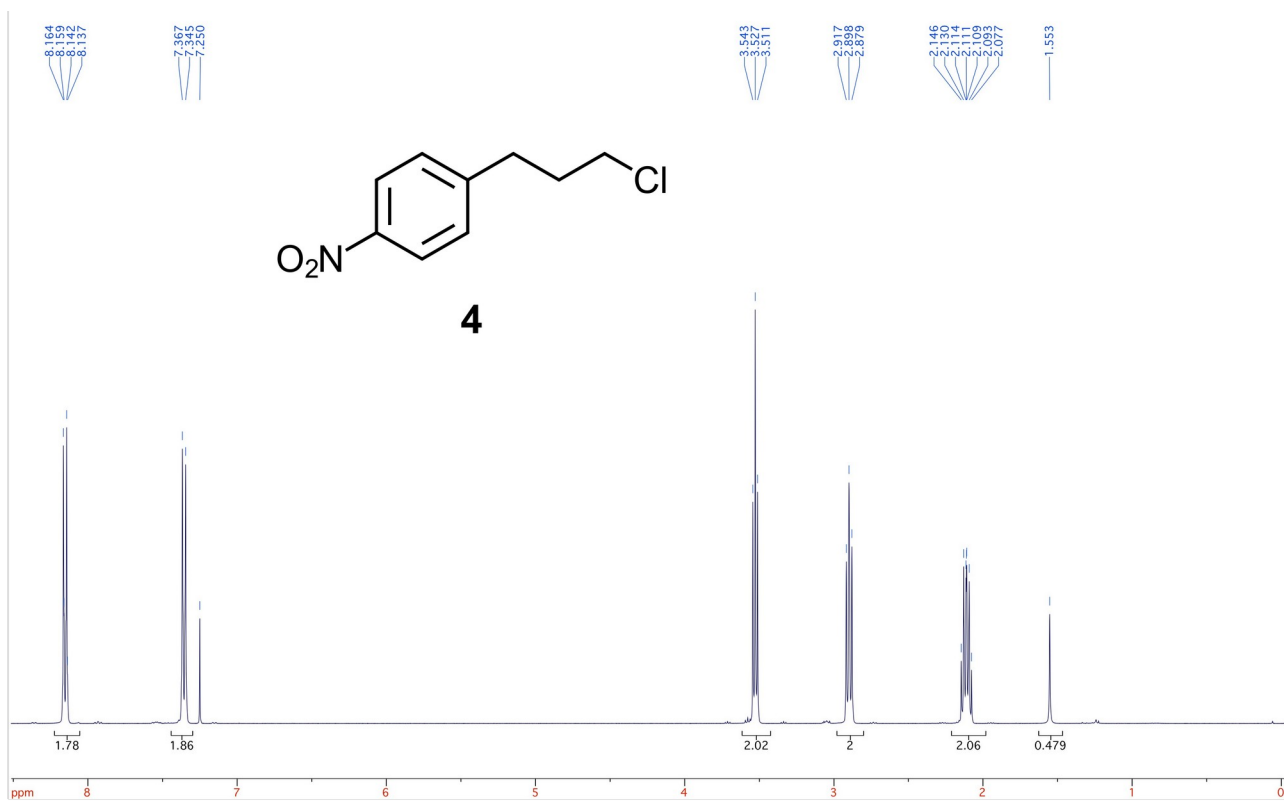


Fig. S6 ^1H - (top) and ^{13}C -NMR (bottom) spectra of compound **4**.

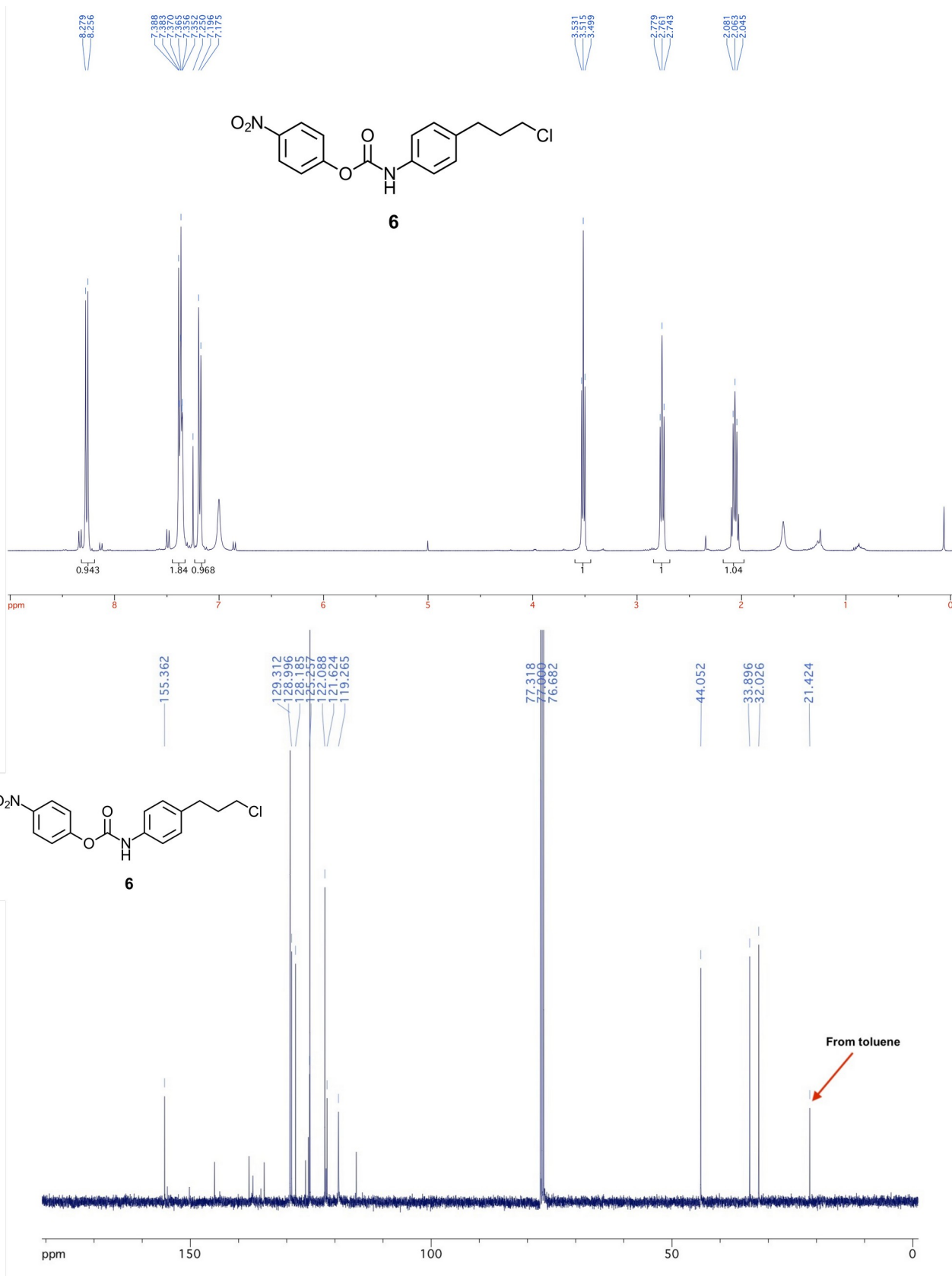


Fig. S7 ¹H- (top) and ¹³C-NMR (bottom) spectra of compound 6.

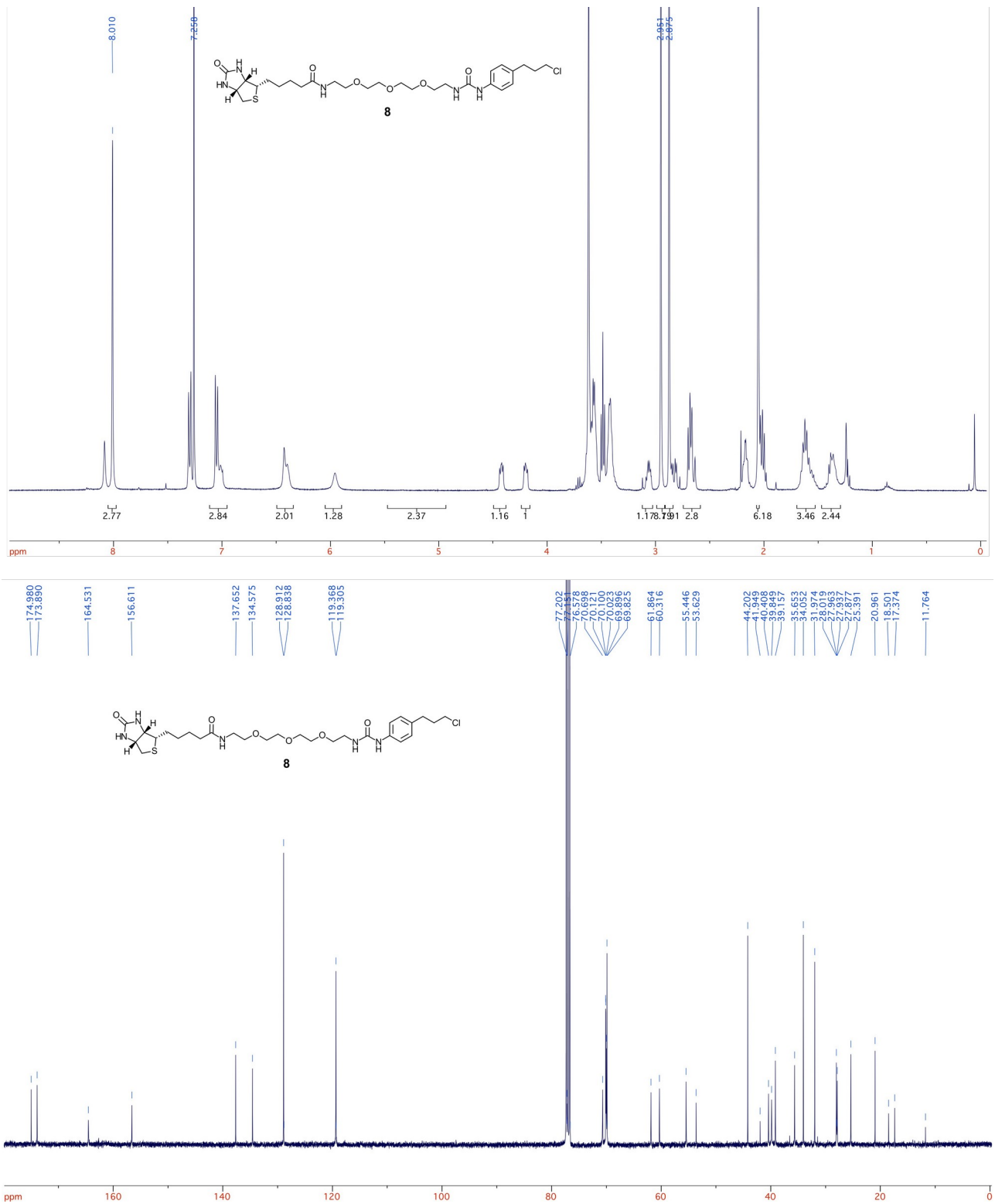


Fig. S8 ^1H - (top) and ^{13}C -NMR (bottom) spectra of compound **8**.

```

    30      32      107      109
... ATC CAC GGC ... // ... GGC GCG CTA ...
   I  H  G      G  A  L
   NcoI      PauI
  
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Fig. S9 Silent mutations (in yellow) in the StEH1 cDNA that removes an *NcoI* site at codons 30 – 32 and introduces a unique *Paul* site at codons 107 – 109.

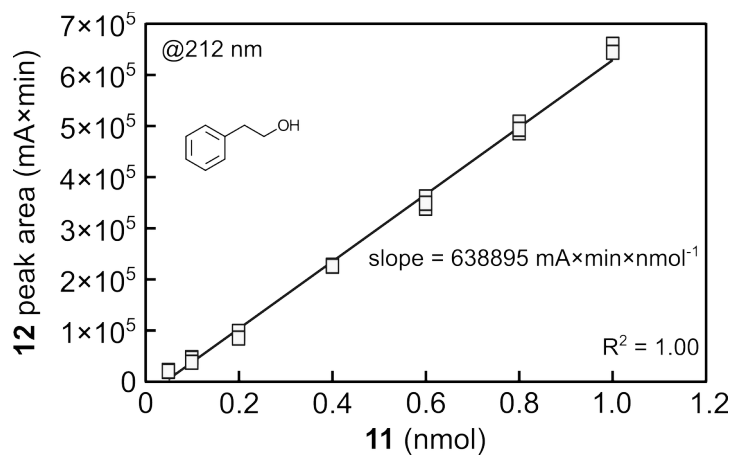


Fig. S10 Standard curve of peak area as a function of amount of alcohol **11**.