

Fig S1. Survival capacity of WF2020 in gastrointestinal conditions (A) and effect of WC2020 on egg laying (B) and body bends (C) in *C. elegans* N2, respectively. For the tolerance to gastrointestinal conditions, 3 mL of overnight bacterial suspensions were centrifuged and the corresponding pellet was diluted 1:1 in a sterile electrolyte solution simulating salivary juice (Simulated Salivary Juice, SSJ), composed of 6.2 g/L NaCl, 2.2 g/L KCl, 0.22 g/L CaCl₂, 1.2 g/L NaHCO₃ pH 6.9 and lysozyme with a final concentration of 100 mg/L. The mixed suspension was incubated for 5 min at 37 °C. Subsequently, the sample was diluted 3:5 with Simulated Gastric Juice (SGJ) containing 6.2 g/L NaCl, 2.2 g/L KCl, 0.22 g/L CaCl₂, 1.2 g/L NaHCO₃ pH 2.5 and 3 g/L pepsin, and incubated for 1 h at 37 °C. After incubation, 1 mL aliquot of the sample was serially diluted and plated, in triplicate, onto LB agar. The remaining sample was diluted 1:4 in Simulated Pancreatic Juice (SPJ) consisting of 6.4 g/L NaHCO₃, 0.239 g/L KCl, 1.28 g/L NaCl, 0.5% bile extract, 0.1% pancreatin at pH 7.2, and incubated for 3 h at 37 °C. At 2 and 3 h incubation times, 1 mL aliquots were withdrawn, serially diluted and plated on LB agar. In parallel, control samples were treated with Phosphate Buffer Saline (PBS) and subjected to the same procedure. For

egg laying and body bends, L1-stage worms incubated on the NGM plates supplemented with a suspension of 100 μL of 4.4×10^{10} CFU/mL WF2020 or OP50 cells. 20 worms on day 3 and 12 were randomly selected and used for the detection of egg laying (B) and body bends (C). The data of egg laying from three independent experiments were shown. Error bar: SD from replicates. Asterisked bars in each bar group differ significantly from those unmarked ($p < 0.05$).

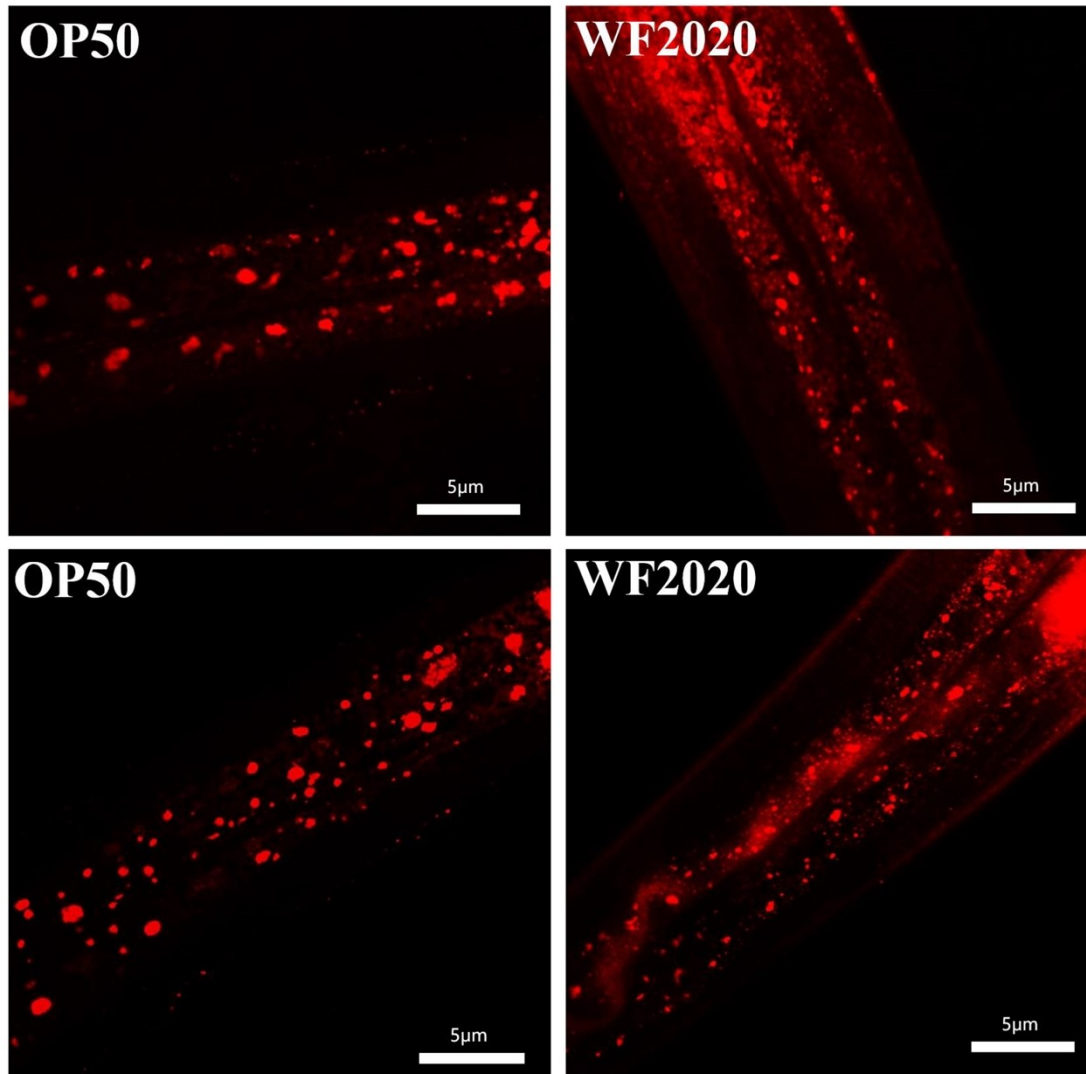


Fig S2. Effect of WF2020 on mitochondria using Mito-Tracker Red CMXRos staining. L1-stage worms incubated on the NGM plates supplemented with a suspension of 100 µL of 4.4×10^{10} CFU/mL WF2020 or OP50 cells. After a 3-day incubation, worms were selected randomly, washed, resuspended in PBS buffer (pH 7.2) supplemented with 4% paraformaldehyde, stained with Mito-Tracker Red CMXRos staining dye at 25 °C in the dark for 6 h, and observed under a confocal laser scanning microscopy (LSM700, Carl Zeiss).

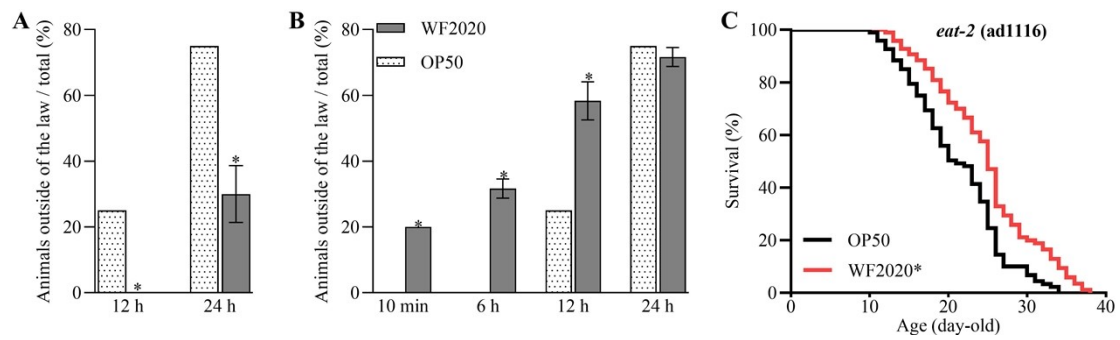


Fig S3. The preference of *C. elegans* N2 for OP50 and WF2020 (A-B) and effect of WF2020 on worm lifespan in *eat-2* mutants (C). Briefly, 30 μ L of bacterial suspension of OP50 or WF2020 (4.4×10^{10} CFU/mL) was dripped onto the center of NGM plates and incubated at 20 $^{\circ}$ C for 24 h. L4-stage worms fed with OP50 were placed on the NGM plates supplemented with OP50 or WF2020 plaques (A) and L4-stage worms fed with OP50 or WF2020 from L1 stage were placed on the NGM plates supplemented with OP50 plaques. During the incubation, the number (N) of nematodes leaving the plaques at 10 min, 6 h, 12 h, and 24 h were recorded to calculate the avoidance index according to the formula: Avoidance index = $N_{\text{outside}} / (N_{\text{outside}} + N_{\text{inside}})$.

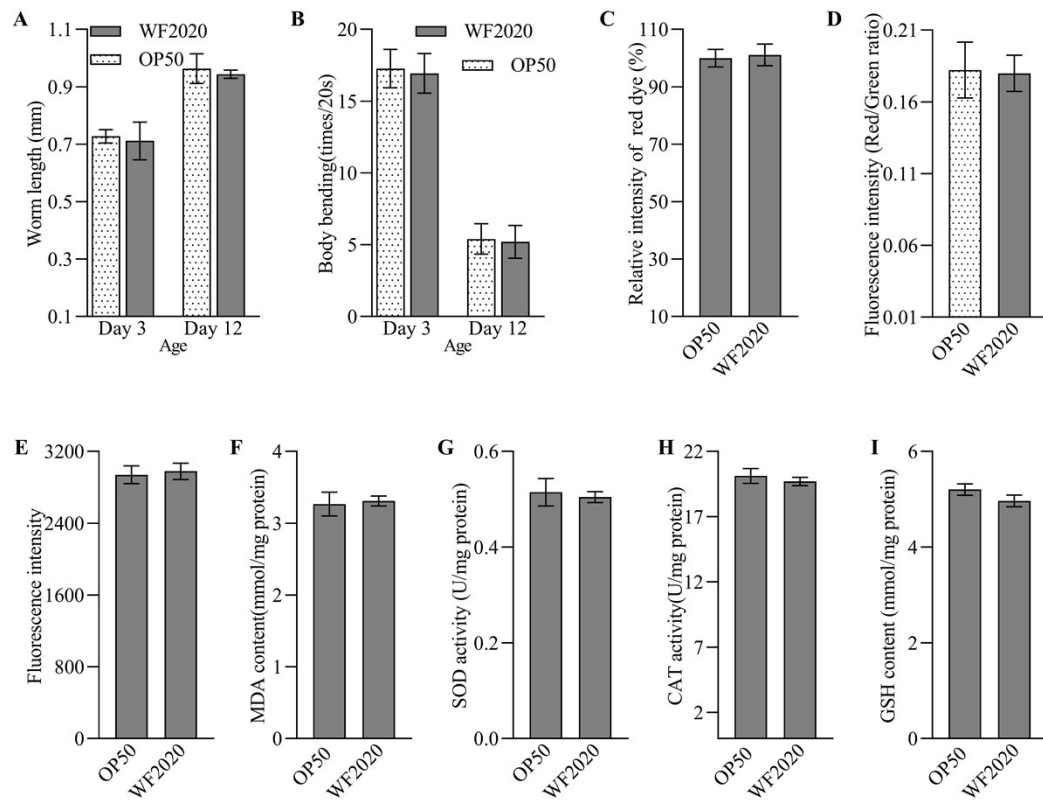


Fig. S4 Changes in body length (A), body bending (B), lipid accumulation (C), mitochondrial membrane potential (D), ROS production (E), MDA content (F), SOD activity (G), CAT activity (H), and GSH content (I) of *pmk-1* mutants caused by WF2020. L1-stage worms incubated on the NGM plates supplemented with a suspension of 100 μ L of 4.4×10^{10} CFU/mL WF2020 or OP50 cells.

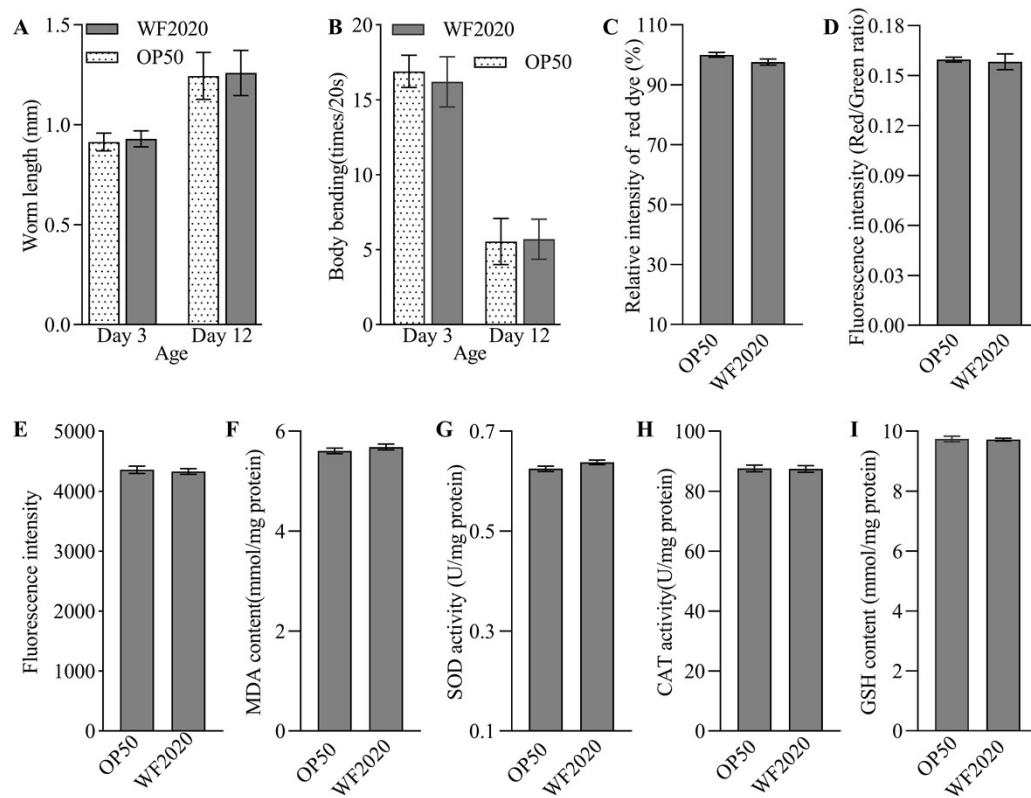


Fig. S5 Changes in body length (A), body bending (B), lipid accumulation (C), mitochondrial membrane potential (D), ROS production (E), MDA content (F), SOD activity (G), CAT activity (H), and GSH content (I) of *jnk-1* mutants caused by WF2020. L1-stage worms incubated on the NGM plates supplemented with a suspension of 100 μ L of 4.4×10^{10} CFU/mL WF2020 or OP50 cells.

Table S1 The primers used for qRT-PCR

Gene	Sequences (5'-3') of paired primers
<i>sek-1</i>	GCCGATGGAAAGTGGTTTTA / TAAACGGCATCGCCAATAAT
<i>pmk-1</i>	CCGACTCCACGAGAAGGATA / AGCGAGTACATTTCAGCAGCA
<i>nsy-1</i>	AGTTCCAATAGTTCAAGT / CTCTCATTCCATCAAGTA
<i>jnk-1</i>	AGAAGCGTGGAAGAGGATCA / ATCCAAAGAGACAGCGTCGT
<i>jkk-1</i>	ATTGATGGCTGTGAAGACTAT / AATATCCGTAACCTGGTGACAA
<i>ced-1</i>	CCTATACTCTCGTCAATCAG / TTATTCTCCAATTCTTCTTCTTC
<i>ced-2</i>	CATTATTTGATTGAAAGAGGAG / CATTATTTGATTGAAAGAGGAG
<i>ced-3</i>	ACGGGAGATCGTGAAAGC / AGAGTTGGCGGATGAAGG
<i>ced-4</i>	AGTCACTCGCAATGGCTCT / GCTGATGAACGACGGAAT
<i>ced-5</i>	GATACCAGAAGAACAACACTCAA / CATCTCAACACATTCAACAA
<i>ced-6</i>	GATTGTTGGATGGATTGG / ACCGAATGGATTATACTCA
<i>sod-1</i>	CGTAGGCGATCTAGGAAATGTG / AACCAACCATAGATCGGCCAACG
<i>sod-2</i>	CTTCAAAACACCGTTCGCTG / CAGTGGAAACAAGTCCAGTT
<i>sod-3</i>	TTCAAAGGAGCTGATGGACACT / AAGTGGGACCATTTCCTTCCAA
<i>sod-4</i>	TTGGGACGCGGTACTTCAG / GCAAGTCGGCTTCCAGCAT
<i>sod-5</i>	GCCTCTTCGGAGCGAACA / TCTCGATCGACGTGGACAAC
<i>ctl-1</i>	CGGATACCGTACTCGTGATGA / CCAAACAGCCACCCAAATCA
<i>ctl-2</i>	TCCGTGACCCTATCCACTTC / TGGGATCCGTATCCATTCAT
<i>ctl-3</i>	GCGGATACCGTACTCGTGAT / GTGGCTGCTCGTAGTTGTGA
<i>gst-5</i>	TTCTTACAAGTTGACCTAC / AGTGA CTCTATTATCCTCGTA
<i>gst-6</i>	AACATTCTGGCATCTAATC / AACATTCTGGCATCTAATC
<i>gst-1</i>	TACCTACAAGAACGGAGA / AACACCATCAAGAGCATT
<i>gst-13</i>	ATGAACAGAAGGATAAAG / CAAGAATCACTGGATAATC
<i>gst-15</i>	AAGAGCAAGTCAGGTTAT / TGTCTTCATCATCATCA
<i>gst-20</i>	ATCAAGCCATACTATTAC / GGTCTCATATTATCACA
<i>gst-27</i>	GATATTGTGATTGTGGAATG / ATCGCATATACCTTCTCT
<i>gst-36</i>	GATATGATTGCTGAAGTTAT / TCTCCTTGAAGAATTGTT
<i>gst-23</i>	TACTACGAATTGAGAAGTTGTT / GTGAGGATGAGGTGAATATC
<i>gst-44</i>	GTTTGAAGAAGCAGAAAAGTC / AAGAAGGAATGTTAGCCAATA
<i>gst-9</i>	CTAGAAGTTGATGGAAGACA / AACACGATAATCCTTGAATTG
<i>gst-22</i>	AAACAATGGAAGAAGTGAAG / CTTGAGATTTCGTCAGAGTAT
<i>gst-34</i>	TCATTCCGTCCATTGCTTTA / TCCGACAAGATACTCCGATT
<i>gst-41</i>	TACATCCGTCTTCTACTAATTG / TTTCTTGCCCATCAACTTTA
<i>gst-35</i>	AAGATAACAAGTCAGGATTC / CTGGTCTGTTCTTAATGTAA
<i>gst-11</i>	TTGTGGCATGTGTTGAGGAT / GCTGATGAACGGAATGATTGTG
<i>gst-26</i>	CGATATTGTAGTTGTGGAGT / ACTTCTTGATAGCTGGAATC
<i>gst-12</i>	TCACCAAGGAAGTTATCAAT / CATTTCAGGAATCCAAAGTTT
<i>gst-14</i>	AAACCTGCTATGGATGTGTA / TTGAATGTTGTCAGCGATAT
<i>gst-3</i>	AACATCCGAGAAGAAGTTAT / GTCAAATTATCAGCAATCACTA
<i>gst-2</i>	GAGGATTAGGTGAAGTCATC / ACTCCATCAATCTCCAATAC
<i>gst-31</i>	TTGAAGACATCCGAATCATC / GATTGCAGTAGATTGAGGAA
<i>fat-1</i>	GTTCAATCCAATTCCATTCT / AGATTCCAGAGATTACACAT
<i>fat-2</i>	CATCACTTCTTCTTACCACAA / AACCAGTTATAGTTGACTTCTC
<i>fat-3</i>	TCTTGGATGGTATATACTTCTG / TGATGAGTGTTATGCTTGTG
<i>fat-4</i>	TGGTGAAGAAGCACGATCAG / AAGCAGAAGATCCGACCAA
<i>fat-5</i>	TGCGATTATATCTAAACAGTTC / CACAAATAAAGCGACATTCT
<i>fat-6</i>	CTTGTGCTGCTTCACTTCTCC / GAAGTTGTGACCTCCCTCTCC
<i>fat-7</i>	ACCCGTGGATTCTTCTTCACT / TAACGGAATGTTCCAGCTACG
<i>lipl-1</i>	CATCTCACTCGGCATTCT / ACTATCTTGTCCAGTAACTTCTAA
<i>lipl-2</i>	GCAGGAACCTTCTACTCAA / AATCATATTCTGGTGGTGA
<i>lipl-3</i>	AGTAAGGAGAATAAGAAGA / ATTGTAGTCAGTGAAGTA
<i>lipl-4</i>	TTGGAGTGATGATTG / AATGTAAGTGGTTGTAGTT
<i>lipl-5</i>	TGAGATGGCTACCTATGA / TCGTTAGAGTTCCTTGTG
<i>lipl-6</i>	AGTTCCTGACCAACAATA / GTGTGCTAAGTATATTCCAA
<i>lipl-7</i>	TGTTCAATCAATCAAGAGTT / TCCATCAGTTCCATAATCA
<i>pept-1</i>	TCCATCATTGTAGTCATCAC / CGTCCTCTTCATCAGTTG

nlp-15 CTCATTGTTCTTCCTGAATC / TTTATTGAATGCTCCGAAAC
nlp-30 AACTTCTTCAATTCTGATTCTC / GTATCCACCATATCCTCTTC
nlp-26 AACTTCTTCGTGATCTTCTC / TTCCATTGAATCCTCCTCTT
nlp-34 TTCTTCATTGTCCTTCTCCTC / ATCCTCCGTATCCTCCGTAT
nlp-35 TGTCAGTTCATTATTGTGTT / ATGTTGGAGGTATTGTGGAG
nlp-43 AATCTACCTTCTACCTTCTA / TTATTCCCTCATTCTTCTTC
nlp-36 CATCTTCTTCGGAGTCCCTCT / AGCTTCATGTCGATGTTCTT
nlp-79 ATGGCTCTCGTTTTACTTAG / GATTTCGTGACATCTTTGGTA
nlp-82 TAATCATCATCCTGCTCATCT / GAATTGTGAACCTCCTCCTA
nlp-76 TCATACACCGTCAAAGTCAA / GTTCTACATTGCCAGAGTCA
nlp-9 TAATCACAACGCCAATTTATTC / AAGTAAACTGGGTACGATGA
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nlp-14 CTTTGGATGGTCTTGATGGA / GAGTTGAGGGCTTTTTATC
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nlp-42 GCAAGTAGTCACTTCTCG / TCCCAACTGATTCCATTCTG
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nlp-60 TTCTTCATCTTCCACATTCAT / AGGTAATCTTCCAACAACAG
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