Supporting Information

Sequential Glaser reaction - diastereoselective cyclocarboxylation of propargylamines with CO₂: A green catalytic access to bis-oxazolidinonedienes and their topochemical polymerization

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1. General Information

All reagents were used as received from commercial sources without further purification. All solvents were dried over activated 4 Å molecular sieves for 24 h. All reactions were analysed by TLC and by GC using a 30 m SE-30 capillary column. Flash column chromatography was performed on silica gel 60 (70–230 mesh). Melting points were measured with an Electrothermal apparatus and are uncorrected. Electron impact mass spectra [m/z, relative intensity (%)] were determined with a GC-MS apparatus at 70 eV ionization energy.

HRMS spectra were obtained with LTQ Orbitrap XL Thermo. Unless otherwise indicated NMR spectra were recorded on Bruker AVANCE 400 and JEOL 600MHz ECZ600R spectrometers in deuterated chloroform, using the solvent residual signals as internal reference (7.26 and 77.00 ppm, respectively for ¹H and ¹³C), or deuterated dimethyl sulfoxide (2.50 and 39.52 ppm, respectively for ¹H and ¹³C). Chemical shifts (δ) and coupling constants (J) are given in ppm and in Hz, respectively.

Single Crystal X-ray Diffraction. SCXRD analyses were performed on single crystal samples of **3b** and **3i**, on a Bruker D8 Venture diffractometer equipped with a kappa goniometer. Data collection was performed using microfocused MoK α radiation ($\lambda = 0.71073$ Å) and CuK α radiation ($\lambda = 1.54178$ Å) under nitrogen flux with Oxford Cryosteam. Lorentz polarization and absorption correction were applied. Data were reprocessed using APEX v4 software. Structures were solved by direct methods using SHELXT¹ and refined by full-matrix least-squares on all F2 using SHELXL² implemented in Olex2.21³. For all samples anisotropic displacement parameters were refined except for hydrogen atoms.

Powder X-ray diffraction. Monomer-to-polymer conversion in batch was checked by PXRD. The data were collected on a Rigaku Smartlab XE diffractometer in θ – θ Bragg–Brentano geometry with Cu K α radiation. The samples were placed on glass supports and exposed to radiation (5° ≤ 2 θ ≤ 50°) at a scan rate of 5 °/min. The diffracted beam was collected on a two-dimensional (2D) Hypix 3000 solid-state detector; 5° radiant Soller slits were used as a compromise for high flux and moderate peak asymmetry at low angles. A beam stopper and antiscatterer air component were used to mitigate the scattering at a low angle. All of the experimental data were refined with a Pawley fit performed with TOPAS v6⁴ against the cell parameters extracted from SCXRD analyses opportunely refined to compensate for the difference in the data collection temperature.

2. Experimental Procedures

Propargylic amines tested in this study.





1q



1r



1s



1t

Figure S1. Propargylic amines used in this work.

General procedures for the synthesis of propargyl amines 1

The propargyl amines were synthesized using two different methods.

Procedure A: synthesis from propargyl bromide (1a, 1j, 1m, 1n, 1s)



Following the literature procedure⁵, the propargyl bromide (1 equiv.) was added dropwise to the corresponding neat amine (6 equiv.) at 0 °C, under nitrogen atmosphere. The mixture was allowed to warm to room temperature and stirred overnight. Then, a mixture of a solution 1 M of NaOH (4 mL/mmol) and Et₂O (4 mL/mmol) was added and stirred for 15 min. The mixture was transferred to a separating funnel, and it was extracted with Et₂O (3x). The combined organic phases were dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude was then purified by flash column chromatography (eluent Hexane:EtOAc).

For amines **1m** and **1n** some modifications were adopted. In a Schlenk tube under nitrogen atmosphere, K₂CO₃ (10 mmol) was added to a stirred suspension of the corresponding amino acid methyl ester hydrochloride (16 mmol) in ACN dry. Then, the mixture was cooled in an ice bath and after 10 min propargyl bromide (4 mmol) was added dropwise. The mixture was allowed to warm to room temperature and stirred overnight. After 16 h, the mixture was diluted with diethyl ether and concentrated under vacuum. Then, the mixture was transferred to a separating funnel, and it was extracted with DCM (3x). The combined organic phases were dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude was then purified by flash column chromatography (eluent Hexane/EtOAc).

Procedure B: reductive amination (1b-1h, 1k, 1l, 1o, 1p and 1r)



According to the literature procedure⁶, one drop of AcOH was added to a solution of propargyl amine (1 equiv.) and aryl aldehyde (1.1 eq.) in MeOH (0.6 M). The resulting mixture was then stirred at room temperature and was monitored with TLC until propargyl imine was formed. NaBH₄ (1.5 equiv.) was added in three different portions at 0 °C and the solution was then stirred for 1 h prior to the evaporation of the solvent. The mixture was diluted with water, extracted with DCM (2 times), and the combined organic layers were then washed with a 1 M HCl solution. Aqueous layers were neutralized, extracted with DCM (2 times), and the resulting organic phase was washed with brine, dried over Na₂SO₄, concentrated under reduced pressure, and finally purified by flash column chromatography (eluent Hexane/EtOAc gradient).

Synthesis of Glaser product 2a



In a 10 mL Schlenk tube open to air, propargyl amine (1 eq.), pyridine (1.1 eq.) and CuCl (0.05 mmol) were dissolved in MeCN (0.5 M) and kept under stirring. The reaction proceeded at room temperature for 4h. The crude was then recovered with DCM and directly purified by flash silica gel column chromatography. Product **2a** was isolated in 67% yield.

General procedure for the synthesis of 3



In a 10 mL Schlenk tube open to air, propargyl amine (1.0 mmol), pyridine (1.1 mmol) and CuCl (0.05 mmol) were dissolved in MeCN dry (2 mL) under stirring. The Glaser reaction (*step 1*) was monitored by TLC until complete conversion of **1**. The solvent was degassed through a freeze-pump-thaw procedure and, then, the tube was filled with CO₂ (balloon) and DBU (2.5 mol%) was added to the mixture (*step 2*). The reaction was kept at 50 °C under stirring for 16 h. The suspension was filtered on a Büchner funnel and washed with a 1:1 ethyl acetate/hexane mixture (2-3 mL) and a saturated NH₄OH solution (2 mL). Finally, the solid product was dried up under reduced pressure, quantified and directly characterized.



Figure S2. Synthesis of compounds 3.

Procedure for Gram Scale synthesis of 3b



In a 25 mL Schlenk tube open to air, propargyl amine (4.3 mmol), pyridine (4.8 mmol) and CuCl (0.215 mmol) were dissolved in MeCN dry (4 mL) under stirring. The Glaser reaction (*step 1*) was monitored by TLC until complete conversion of **1b**. The solvent was degassed through a freeze-pump-thaw procedure and, then, the tube was filled with CO₂ (balloon) and DBU (2.5 mol%) was added to the mixture (*step 2*). The reaction was kept at 50 °C under stirring for 16 h. The suspension was filtered on a Büchner funnel and washed with a 1:1 ethyl acetate/hexane mixture (5 mL) and a saturated NH₄OH solution (5 mL). Finally, the solid was dried up under reduced pressure to give product **3b** (0.82 g, 89% yield).

Procedure for the synthesis of 3q



In a 10 mL Schlenk tube open to air, propargyl amine **1a** (0.5 mmol), propargyl amine **1o** (0.5 mmol), pyridine (1.1 mmol) and CuCl (0.05 mmol) were dissolved in MeCN dry (2 mL) under stirring. The reaction (step 1) was monitored by TLC until complete conversion of the starting materials. The solvent was degassed through a freeze-pump-thaw procedure and, then, the tube was filled with CO₂ (balloon) and DBU (2.5 mol%) was added to the mixture (step 2). The reaction was kept at 50 °C under stirring for 16 h. After that, the crude mixture was recovered with DCM and directly purified by flash column chromatography (Eluents DCM:Hexane:EtOAc 10:8:1) to yield product **3q** (49% yield).

3. Green chemistry metrics calculations

Green chemistry parameters were evaluated as described in literature. ^{7,8}



= 32.0

PMI was calculated for a typical synthesis of 2-oxazolidinones described in literature⁹. In this work, a column chromatography (EtOAc: Petroleum ether 1:20) was necessary to obtain the product with an acceptable purity. We estimated a minimal consumption of eluent of 300 mL (Petroleum ether 285.7 mL, 183 g; EtOAc, 14.3 mL, 12.9 g) and of silica gel (30 g).



4. Characterization Data

Characterization of Substrates 1



N-benzylprop-2-yn-1-amine (1a). Yellow oil (522 mg, 90% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.34 (m, 4H), 7.29 (ddd, J = 6.1, 4.7, 2.4 Hz, 1H), 3.91 (s, 2H), 3.46 (d, J = 2.4 Hz, 2H), 2.29 (t, J = 2.4 Hz, 1H), 1.62 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 139.4, 128.5, 128.4, 127.2, 82.1, 71.6, 52.3, 37.3. The NMR data closely match the ones previously reported in the literature.¹⁰



N-(4-methoxybenzyl)prop-2-yn-1-amine (1b). Yellow oil (399 mg, 57% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.24 (m, 2H), 6.93 – 6.84 (m, 2H), 3.83 (s, 2H), 3.81 (s, 3H), 3.42 (d, J = 2.4 Hz, 2H), 2.28 (t, J = 2.4 Hz, 1H), 1.59 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 158.8, 131.5, 129.6, 113.8, 82.2, 71.5, 55.3, 51.6, 37.2. The NMR data closely match the ones previously reported in the literature.¹⁰



N,N-dimethyl-4-((prop-2-yn-1-ylamino)methyl)aniline (1c). Red oil (376 mg, 50% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.28 – 7.20 (m, 2H), 6.79 – 6.70 (m, 2H), 3.81 (s, 2H), 3.44 (d, J = 2.4 Hz, 2H), 2.96 (s, 6H), 2.27 (t, J = 2.4 Hz, 1H), 1.54 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 150.0, 129.4, 127.3, 112.7, 82.3, 71.3, 51.8, 40.8, 37.2. The NMR data closely match the ones previously reported in the literature.⁶



N-(4-bromobenzyl)prop-2-yn-1-amine (1d). Yellow oil (624 mg, 70% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.52 – 7.42 (m, 2H), 7.29 – 7.20 (m, 2H), 3.42 (dt, J = 2.2, 0.9 Hz, 2H), 2.28 (t, J = 2.4 Hz, 1H), 1.61 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 138.4, 131.5, 130.1, 121.0., 81.8, 71.8, 51.5, 37.2. The NMR data closely match the ones previously reported in the literature.¹⁰



N-(4-chlorobenzyl)prop-2-yn-1-amine (1e). Yellow Oil (644 mg, 90% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.28 (s, 4H), 3.83 (s, 2H), 3.39 (d, J = 2.4 Hz, 2H), 2.28 (t, J = 2.4 Hz, 1H), 1.55 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 137.9, 132.8, 129.8, 128.5, 81.9, 71.8, 51.4, 37.2. The NMR data closely match the ones previously reported in the literature.¹⁰



N-(3-methylbenzyl)prop-2-yn-1-amine (1f). Yellow Oil (369 mg, 58% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.25 (t, J = 7.5 Hz, 1H), 7.20 (s, 1H), 7.17 (d, J = 7.6 Hz, 1H), 7.11 (d, J = 7.4 Hz, 1H), 3.88 (s, 2H), 3.46 (d, J = 2.4 Hz, 2H), 2.38 (s, 3H), 2.29 (t, J = 2.4 Hz, 1H), 1.80 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 139.2, 138.1, 129.2, 128.4, 128.0, 125.5, 82.0, 71.7, 52.3, 37.4, 21.4.



N-(3-nitrobenzyl)prop-2-yn-1-amine (1g). Yellow oil (418 mg, 55% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.24 (s, 1H), 8.12 (d, J = 8.2 Hz, 1H), 7.71 (d, J = 7.6 Hz, 1H), 7.50 (t, J = 7.9 Hz, 1H), 4.00 (s, 2H), 3.45 (d, J = 2.4 Hz, 2H), 2.30 (t, J = 2.4 Hz, 1H), 2.17 (s, 1H).¹³C NMR (101 MHz, CDCl₃) δ 148.4, 141.6, 134.5, 129.3, 123.2, 122.3, 81.4, 72.2, 51.2, 37.3.



N-(2-(benzyloxy)benzyl)prop-2-yn-1-amine (1h). Yellow oil (773 mg, 77% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.44 (m, 2H), 7.45 – 7.38 (m, 2H), 7.37 – 7.31 (m, 2H), 7.31 – 7.21 (m, 2H), 7.03 – 6.92 (m, 2H), 5.14 (s, 2H), 3.97 (s, 2H), 3.44 (d, J = 2.5 Hz, 2H), 2.22 (t, J = 2.4 Hz, 1H), 1.84 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 156.8, 137.1, 130.2, 128.6, 128.5, 127.9, 127.2 (2C), 120.8, 111.8, 82.1, 71.4, 69.9, 48.1, 37.5.

N-(prop-2-yn-1-yl)octan-1-amine (1j). Yellow oil (548 mg, 82% yield). ¹H NMR (400 MHz, CDCl₃) δ 3.44 (s, 2H), 2.69 (t, J = 7.2 Hz, 2H), 2.22 – 2.21 (t, 1H), 1.60 – 1.39 (m, 4H), 1.30 (d, J = 11.2 Hz, 10H), 0.89 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 82.3, 71.2, 48.7, 38.2, 31.8, 29.8, 29.5, 29.3, 27.3, 22.7, 14.1.



N-(2-phenylpropyl)prop-2-yn-1-amine (1k). Yellow Oil (187 mg, 27% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.32 (m, 2H), 7.26 (d, J = 7.5 Hz, 3H), 3.41 (t, J = 2.3 Hz, 2H), 3.00 – 2.92 (m, 2H), 2.89 – 2.81 (m, 1H), 2.22 (t, J = 2.4 Hz, 1H), 1.38 (s, 1H), 1.31 (d, J = 6.6 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 145.0, 128.6, 127.2, 126.5, 82.1, 71.3, 55.7, 40.1, 38.3, 20.2.



N-(thiophen-2-ylmethyl)prop-2-yn-1-amine (11). Yellow oil (574 mg, 95% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.24 (dd, J = 4.8, 1.5 Hz, 1H), 7.03 – 6.94 (m, 2H), 4.11 (s, 2H), 3.47 (d, J = 2.4 Hz, 2H), 2.29 (t, J = 2.4 Hz, 1H), 1.75 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 142.9, 126.7, 125.5, 124.8, 81.7, 71.9, 46.7, 37.0. The NMR data closely match the ones previously reported in the literature.¹⁰



Methyl prop-2-yn-1-ylglycinate (1m). Colourless oil (290 mg, 57% yield). ¹H NMR (400 MHz, CDCl₃) δ 3.74 (s, 3H), 3.53 (s, 2H), 3.49 (d, *J* = 2.1 Hz, 2H), 2.24 (t, *J* = 2.4 Hz, 1H), 1.86 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 172.3, 81.1, 72.1, 51.9, 49.0, 37.6.



Methyl prop-2-yn-1-yl-D-phenylalaninate (1n). Yellow oil (617 mg, 71% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.29 (m, 2H), 7.28 – 7.19 (m, 3H), 3.78 (dd, *J* = 7.2, 6.3 Hz, 1H), 3.70 (s, 3H), 3.46 (dd, *J* = 17.0, 2.5 Hz, 1H), 3.39 (dd, *J* = 17.0, 2.5 Hz, 1H), 3.05 (dd, *J* = 13.6, 6.2 Hz, 1H), 2.98 (dd, *J* = 13.7, 7.3 Hz, 1H), 2.21 (t, *J* = 2.5 Hz, 1H), 1.80 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 174.3, 136.8, 129.2, 128.5, 126.9, 81.1, 71.8, 61.1, 51.8, 39.4, 36.8.



N-benzyl-2-methylbut-3-yn-2-amine (10). Yellow oil (672 mg, 97% yield). ¹**H NMR** (400 MHz, CDCl₃) δ 7.45 – 7.25 (m, 5H), 3.92 (s, 2H), 2.41 (d, *J* = 1.0 Hz, 1H), 1.47 (s, 6H), 1.39 (bs, 1H).¹³**C NMR** (101 MHz, CDCl₃) δ 140.6, 128.5, 127.0, 89.1, 69.9, 50.1, 49.0, 29.6.



N-benzyl-3-ethylpent-1-yn-3-amine (1p). Yellow Oil (716 mg, 89% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.45 – 7.39 (m, 2H), 7.39 – 7.32 (m, 2H), 7.32 – 7.25 (m, 1H), 3.85 (s, 2H), 2.40 (s, 1H), 1.81 – 1.62 (m, 4H), 1.23 (s, 1H), 1.03 (t, *J* = 7.4 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 140.9, 128.5, 128.4, 126.9, 87.8, 71.5, 57.4, 48.1, 30.5, 8.1.



N-(prop-2-yn-1-yl)adamantan-2-amine (1r). White solid (605 mg, 80% yield). ¹H NMR (400 MHz, CDCl₃) δ 3.45 (d, J = 2.4 Hz, 2H), 2.93 (d, J = 2.6 Hz, 1H), 2.20 (t, J = 2.4 Hz, 1H), 2.02 (dd, J = 12.6, 3.1 Hz, 2H), 1.91 – 1.70 (m, 10H), 1.54 – 1.46 (m, 2H), 1.29 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 77.4, 77.0, 76.7, 70.7, 60.5, 37.9, 37.4, 35.5, 31.9, 31.3, 27.8, 27.7.



N-(prop-2-yn-1-yl)aniline (1s). Red oil (472 mg, 90% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.30 (td, J = 7.3, 2.1 Hz, 2H), 6.87 (t, J = 7.4 Hz, 1H), 6.76 (dd, J = 8.7, 1.1 Hz, 2H), 3.98 (d, J = 2.5 Hz, 2H), 2.29 (t, J = 2.4 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 146.9, 129.3, 118.7, 113.6, 81.2, 71.4, 33.7. The NMR data closely match the ones previously reported in the literature.¹⁰

Characterization of 2a



*N*¹,*N*⁶-*dibenzylhexa-2,4-diyne-1,6-diamine (2a).* Red Brown oil (289 mg, 67% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.35 (m, 8H), 7.32 – 7.29 (m, 2H), 3.92 (s, 4H), 3.54 (s, 4H), 1.66 (bs, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 139.2, 128.5, 128.4 (2C), 127.3, 68.2, 52.3, 38.0.

Characterization of Products 3



(*5Z*,*5'Z*)-*5*,*5'*-(*ethane*-*1*,*2*-*diylidene*)*bis*(*3*-*benzyloxazolidin*-*2*-*one*) (*3a*). White solid (175.1 mg, 96% yield); dec. 276 °C. ¹H NMR (400 MHz, DMSO) δ 7.43 – 7.25 (m, 10H), 5.53 (s, 2H), 4.44 (s, 4H), 4.20 (s, 4H). ¹³C NMR (101 MHz, DMSO) δ 155.2, 142.3, 136.2, 129.2, 128.24, 128.22, 95.5, 47.9, 47.6. HRMS (ESI) *m/z* calculated for C₂₂H₂₀N₂NaO₄ [M+Na]⁺: 399.1315, found: 399.1312.



(52,5'2)-5,5'-(ethane-1,2-diylidene)bis(3-(4-methoxybenzyl)oxazolidin-2-one) (3b). White solid (216.5 mg, 90% yield); dec. 230 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.21 (d, J = 8.4 Hz, 4H), 6.91 (d, J = 8.4 Hz, 4H), 5.52 (s, 2H), 4.42 (s, 4H), 4.06 (s, 4H), 3.83 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 159.6, 155.2, 141.2, 129.6, 126.8, 114.4, 95.9, 55.3, 47.4, 47.3. HRMS (ESI) m/z calculated for C₂₄H₂₄N₂NaO₆ [M+Na]⁺: 459.1526, found: 459.1525.



(52,5'2)-5,5'-(ethane-1,2-diylidene)bis(3-(4-(dimethylamino)benzyl)oxazolidin-2-one) (3c). White Solid (212.4 mg, 91% yield); dec. 241 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.12 (m, 4H), 6.75 (d, J = 8.0 Hz, 4H), 5.51 (t, J = 0.8 Hz, 2H), 4.39 (s, 4H), 4.07 – 4.02 (m, 4H), 2.98 (s, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 155.2, 150.4, 141.3, 129.4, 122.5, 112.8, 95.8, 47.5, 47.2, 40.5. HRMS (ESI) m/z calculated for C₂₆H₃₀N₄NaO₄ [M+Na]⁺: 485.2159, found: 485.2156.



(*5Z*,*5'Z*)-*5*,*5'*-(*ethane-1*,*2-diylidene*)*bis*(*3*-(*4-bromobenzyl*)*oxazolidin-2-one*) (*3d*). White Solid (256 mg, 93% yield); dec. 320 °C. ¹H NMR (400 MHz, DMSO) δ 7.61 – 7.53 (m, 4H), 7.31 – 7.23 (m, 4H), 5.52 (s, 2H), 4.41 (s, 4H), 4.20 (s, 4H). ¹³C NMR (101 MHz, DMSO) δ 155.2, 142.3, 135.7, 132.1, 130.5, 121.4, 95.5, 48.0, 47.0. HRMS (ESI) *m*/*z* calculated for C₂₂H₁₈Br₂N₂NaO₄ [M+Na]⁺: 554.9525, found: 554.9523.



(*5Z*,*5'Z*)-*5*,*5'*-(*ethane*-*1*,*2*-*diylidene*)*bis*(*3*-(*4*-*chlorobenzyl*)*oxazolidin*-*2*-*one*) (*3e*). White Solid (216.7 mg, 97% yield); dec. 310 °C. ¹H NMR (400 MHz, DMSO) δ 7.45 – 7.40 (m, 4H), 7.35 – 7.30 (m, 4H), 5.52 (s, 2H), 4.43 (s, 4H), 4.21 (s, 4H). ¹³C NMR (101 MHz, DMSO) δ 155.2, 142.3, 135.3, 133.0, 130.2, 129.2, 95.5, 48.0, 46.9. HRMS (ESI) *m*/*z* calculated for C₂₂H₁₈Cl₂N₂NaO₄ [M+Na]⁺: 467.0536, found: 467.0532.



(52,5'2)-5,5'-(ethane-1,2-diylidene)bis(3-(3-methylbenzyl)oxazolidin-2-one) (3f). White solid (158 mg, 91% yield); dec. 260 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.27 (t, J = 7.5 Hz, 2H), 7.16 (d, J = 7.6 Hz, 2H), 7.10 (s, 2H), 7.08 (d, J = 8.5 Hz, 2H), 5.54 (s, 2H), 4.45 (s, 4H), 4.08 (s, 4H), 2.37 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 155.3, 141.2, 138.9, 134.7, 129.1, 128.9 (2C), 125.2, 96.0, 47.9, 47.4, 21.4. HRMS (ESI) m/z calculated for C₂₄H₂₄N₂NaO₄ [M+Na]⁺: 427.1628, found: 427.1626.



(*5Z*,*5'Z*)-*5*,*5'*-(*ethane*-1,*2*-*diylidene*)*bis*(*3*-(*3*-*nitrobenzyl*)*oxazolidin*-*2*-*one*) (*3g*). Pale yellow solid (206.8 mg, 89% yield); dec. 225 °C. ¹H NMR (400 MHz, DMSO) δ 8.18 (d, J = 8.1 Hz, 2H), 8.15 (s, 2H), 7.78 (d, J = 7.7 Hz, 2H), 7.69 (t, J = 7.8 Hz, 2H), 5.55 (s, 2H), 4.59 (s, 4H), 4.27 (s, 4H). ¹³C NMR (101 MHz, DMSO) δ 160.1, 153.3, 147.1, 143.4, 139.7, 135.5, 127.9, 127.8, 100.3, 52.9, 51.5. HRMS (ESI) *m/z* calculated for C₂₂H₁₈N₄NaO₈ [M+Na]⁺: 489.1017, found: 489.1016.



(*52,5'2*)-*5,5'-(ethane-1,2-diylidene)bis(3-(2-(benzyloxy)benzyl)oxazolidin-2-one) (3h).* Orange solid (279.25 mg, 94% yield); dec. 185 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.36 (m, 10H), 7.32 (t, *J* = 7.5 Hz, 4H), 6.99 (t, *J* = 7.8 Hz, 4H), 5.46 (s, 2H), 5.12 (s, 4H), 4.56 (s, 4H), 4.05 (s, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 156.7, 155.3, 141.4, 136.5, 130.5, 129.7, 128.7, 128.3, 127.5, 123.5, 121.2, 111.9, 95.7, 70.3, 48.1, 42.8. HRMS (ESI) *m/z* calculated for C₃₆H₃₂N₂NaO₆ [M+Na]⁺: 611.2152, found: 611.2151.



(*5Z*,*5'Z*)-*5*,*5'*-(*ethane-1*,*2*-*diylidene*)*bis*(*3*-*methyloxazolidin-2*-*one*) (*3i*). White solid (122 mg, 89%); dec. 264 °C. ¹H NMR (400 MHz, CDCl₃) δ 5.60 (s, 2H), 4.24 (s, 4H), 2.97 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 155.4, 141.0, 95.8, 50.1, 30.6. HRMS (ESI) *m*/*z* calculated for C₁₀H₁₂N₂NaO₄ [M+Na]⁺: 247.0689, found: 247.0685.



(5Z,5'Z)-5,5'-(ethane-1,2-diylidene)bis(3-octyloxazolidin-2-one) (3j). White Solid (192.15 mg, 91% yield); m.p. 186-188 °C. ¹H NMR (400 MHz, CDCl₃) δ 5.59 (s, 2H), 4.22 (s, 4H), 3.32 (t, *J* = 7.3 Hz, 4H), 1.62 – 1.52 (m, 4H), 1.37 – 1.25 (m, 20H), 0.89 (t, *J* = 6.7 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 155.2, 141.2, 95.8, 48.0, 43.9, 31.7, 29.1 (2C), 27.2, 26.6, 22.6, 14.1. HRMS (ESI) *m/z* calculated for C₂₄H₄₀N₂NaO₄ [M+Na]⁺: 443.2880, found: 443.2878.



(52,5'2)-5,5'-(ethane-1,2-diylidene)bis(3-(2-phenylpropyl)oxazolidin-2-one) (3k). White solid (198.6 mg, 96% yield); m.p. 215-217 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.34 (t, J = 7.3 Hz, 4H), 7.28 – 7.18 (m, 6H), 5.40 (s, 2H), 3.95 (d, J = 13.9 Hz, 2H), 3.76 (d, J = 13.8 Hz, 2H), 3.60 (dd, J = 14.0, 6.8 Hz, 2H), 3.34 (dd, J = 14.0, 8.7 Hz, 2H), 3.09 (h, J = 7.0 Hz, 2H), 1.31 (d, J = 7.0 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 155.4, 143.3, 141.1, 128.9, 127.1, 127.0, 95.6, 51.0, 48.7, 38.6, 19.2. HRMS (ESI) *m/z* calculated for C₂₆H₂₈N₂NaO₄ [M+Na]⁺: 455.1941, found: 455.1940.



(52,5'2)-5,5'-(ethane-1,2-diylidene)bis(3-(thiophen-2-ylmethyl)oxazolidin-2-one) (3l). White solid (178 mg, 92% yield); dec. 283°C. ¹H NMR (400 MHz, CDCl₃) δ 7.32 (dd, J = 5.0, 1.3 Hz, 2H), 7.06 – 7.03 (m, 2H), 7.01 (dd, J = 5.0, 3.5 Hz, 2H), 5.56 (t, J = 0.8 Hz, 2H), 4.68 (d, J = 0.7 Hz, 4H), 4.19 – 4.15 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 154.7, 141.1, 136.9, 127.5, 127.2, 126.3, 96.1, 47.3, 42.4. HRMS (ESI) m/z calculated for C₁₈H₁₆N₂NaO₄S₂ [M+Na]⁺: 411.0443, found: 411.0441.



Dimethyl 2,2'-((5Z,5'Z)-ethane-1,2-diylidenebis(2-oxooxazolidin-3-yl-5-ylidene))diacetate (3m). White Solid (112.3 mg, 65% yield); dec. 200 °C. ¹H NMR (400 MHz, DMSO) δ 5.62 (s, 2H), 4.35 (s, 4H), 4.14 (s, 4H), 3.70 (s, 6H). ¹³C NMR (101 MHz, DMSO) δ 169.3, 155.3, 142.3, 95.6, 52.7, 48.8, 45.0. HRMS (ESI) *m/z* calculated for C₁₄H₁₆N₂NaO₈ [M+Na]⁺: 363.0799, found: 363.0798.



Dimethyl 2,2'-((5Z,5'Z)-ethane-1,2-diylidenebis(2-oxooxazolidin-3-yl-5-ylidene))(2R,2'R)-bis(3-phenylpropanoate) (3n). White Solid (232.4 mg, 74% yield); m.p. 175-178 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.28 (m, 6H), 7.21 (dt, J = 7.9, 1.5 Hz, 4H), 5.49 (s, 2H), 4.88 (dd, J = 10.9, 5.3 Hz, 2H), 4.36 (dd, J = 13.3, 3.5 Hz, 2H), 4.13 (dd, J = 13.2, 4.3 Hz, 2H), 3.78 (d, J = 1.5 Hz, 6H), 3.40 (dd, J = 14.8, 5.3 Hz, 2H), 3.01 (ddd, J = 14.8, 10.9, 1.1 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 170.2, 155.2, 141.2, 135.5, 129.0, 128.4, 127.4, 96.0, 56.4, 52.7, 45.5, 35.1. HRMS (ESI) m/z calculated for C₂₈H₂₈N₂NaO₈ [M+Na]⁺: 543.1738, found: 543.1735.



(52,5'2)-5,5'-(ethane-1,2-diylidene)bis(3-benzyl-4,4-dimethyloxazolidin-2-one) (3o). White solid (201.1 mg, 92% yield); m.p. 225-228 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.25 (m, 10H), 5.58 (s, 2H), 4.48 (s, 4H), 1.33 (s, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 154.6, 152.7, 137.4, 128.7, 127.8, 127.7, 93.8, 61.9, 44.1, 27.5. HRMS (ESI) m/z calculated for C₂₆H₂₈N₂NaO₄ [M+Na]⁺: 455.1941, found: 455.1938.



(5Z,5'Z)-5,5'-(ethane-1,2-diylidene)bis(3-benzyl-4,4-diethyloxazolidin-2-one) (3p). White Solid (129.4 mg, 53%) dec. 238 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.41 (m, 4H), 7.40 – 7.26 (m, 6H), 5.49 (s, 2H), 4.35 (s, 4H), 1.75 (dq, J = 14.5, 7.2 Hz, 4H), 1.51 (dq, J = 14.6, 7.3 Hz, 4H), 0.61 (t, J = 7.2 Hz, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 155.9, 148.8, 137.1, 128.9, 128.6, 128.0, 94.3, 70.3, 44.3, 31.6, 7.5. HRMS (ESI) *m/z* calculated for $C_{30}H_{36}N_2NaO_4$ [M+Na]⁺: 511.2567, found: 511.2565.



(Z)-3-benzyl-5-((Z)-2-(3-benzyl-2-oxooxazolidin-5-ylidene)ethylidene)-4,4-dimethyloxazolidin-2-one (3q). Pale yellow oil (99.0 mg, 49% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.24 (m, 10H), 5.57 (s, 2H), 4.50 (s, 2H), 4.47 (s, 2H), 4.10 (s, 2H), 1.32 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 155.3, 154.5, 152.7, 141.1, 137.4, 134.8, 129.0, 128.7, 128.4, 128.2, 127.8, 127.8, 96.3, 93.5, 61.8, 48.0, 47.4, 44.1, 27.5. HRMS (ESI) *m/z* calculated for C₂₄H₂₄N₂NaO₄ [M+Na]⁺: 427.1628, found: 427.1622.

Characterization of 4p



(Z)-3-benzyl-5-(4-(benzylamino)-4-ethylhex-2-yn-1-ylidene)-4,4-diethyloxazolidin-2-one (4p). Pale yellow oil (81.6 mg, 40% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.42 (m, 4H), 7.37 – 7.32 (m, 4H), 7.26 (t, J = 7.3 Hz, 2H), 4.71 (s, 1H), 4.37 (s, 2H), 3.86 (s, 2H), 1.74 (dtq, J = 28.5, 14.3, 7.3 Hz, 6H), 1.48 (dd, J = 14.7, 7.4 Hz, 2H), 1.02 (t, J = 7.4 Hz, 6H), 0.67 (t, J = 7.3 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 159.1, 155.6, 141.1, 137.0, 128.9, 128.6, 128.4, 128.1, 126.8, 99.2, 81.9, 76.1, 70.5, 58.4, 48.3, 44.3, 31.9, 30.6, 8.3, 7.5. HRMS (ESI) *m/z* calculated for C₂₉H₃₆N₂NaO₂ [M+Na]⁺: 467.2669, found: 467.2666.

5. Crystallographic Data

Powder X-Ray Diffraction Data

Reaction conditions tested to promote the polymerization of 3b to 5b. The ball mill grinding experiments were performed by means of a Fritsch Pulverisette 23 shaker mill with a vertical movement. This mill has a fixed amplitude of 9 mm and adjustable frequency from 15 to 50 Hz with an adjustable timer.



Entry	Metodology	Solvent	Oxidant	Milling Frequency (Hz)	Temperature (°C)	Time (h)
1 ^a	Batch	MeCN	O ₂ (1 atm)	-	80	72
2 ^a	Batch	MeCN	O ₂ (1 atm)	-	100	72
3 ^b	Ball Milling	neat	Air (1 atm)	50 Hz	r.t.	1.5

Table S1. Reaction conditions: ^a **3b** (0.1 mmol) was dissolved in MeCN (1.5 mL) and was kept at 80-100 °C under O_2 atmosphere for 72h. ^b **3b** (0.1 mmol) was milled at the frequency of 50 Hz in a 15 mL stainless-steel jar under atmosphere conditions for 1.5h.



Figure S3. Cascade visualization of PXRD pattern of polymer formation from **3b**. Experimental conditions refer to those in TableS1. From lower to upper powder pattern: calculated powder pattern of **3b**; calculated powder pattern of **5b**; experimental data after treating **3b** under conditions of Entry 3; experimental data after treating **3b** under conditions of Entry 1.



Figure S4. Pawley refinement of **3b** after the mechanochemical treatment as reported in the experimental section. Multi-phase model (red line) against experimental data (black line) shows the co-presence of both **3b** and **5b**. Green and Blue ticks reported at the below the powder pattern refers to the modelled peaks positions of **5b** and **3b** respectively.

Identification code	3b	3i	5b
Empirical formula	C24H24N2O6	$C_{10}H_{12}N_2O_4$	C36H36N3O9
Formula weight	436.45	224.22	654.68
Temperature/K	200.00	200.00	293.00(10)
Crystal system	monoclinic	monoclinic	triclinic
Space group	C2/c	$P2_1/n$	P-1
a/Å	40.9802(7)	6.6549(3)	6.0202(3)
b/Å	5.98560(10)	12.8056(5)	13.4077(7)
c/Å	8.7293(2)	6.6575(3)	19.4725(12)
α/°	90	90	97.433(5)
β/°	97.2560(10)	116.973(2)	91.503(4)
γ/°	90	90	92.772(4)
Volume/Å ³	2124.07(7)	505.64(4)	1555.89(15)
Z	4	2	2
$\rho_{calc}g/cm^3$	1.365	1.473	1.397
μ/mm^{-1}	0.818	0.115	0.837
F(000)	920.0	236.0	690.0
Crystal size/mm ³	0.3 imes 0.05 imes 0.05	0.8 imes 0.7 imes 0.7	0.2 imes 0.03 imes 0.03
D = di = ti = ti	CuKa	ΜοΚα	Cu Ka
Kaalallon	$(\lambda = 1.54178)$	$(\lambda = 0.71073)$	$(\lambda = 1.54184)$
2 Θ range for data collection/°	4.348 to 140.084	6.364 to 52.804	6.658 to 133.2
	$-49 \le h \le 49$	$-8 \le h \le 8,$	$-7 \le h \le 7$,
Index ranges	$-7 \le k \le 7$	$-15 \le k \le 13$	$-15 \le k \le 15$,
	$-9 \le 1 \le 10$	$-8 \le 1 \le 8$	$-23 \le 1 \le 23$
Reflections collected	13280	4910	27348
	2013	1022	5448
Independent reflections	$R_{int} = 0.0367$	$R_{int} = 0.0480$	$R_{int} = 0.1104,$
	$R_{sigma} = 0.0205$	$R_{sigma} = 0.0353$	$R_{sigma} = 0.0604$
Data/restraints/parameters	2013/0/193	1022/0/75	5448/2/455
Goodness-of-fit on F^2	1.046	1.103	1.088
Final R indexes $[I > = 2\sigma (I)]$	$R_1 = 0.0343$	$R_1 = 0.0334$	$R_1 = 0.0952,$
1 mull K mulles [1 > -20 (1)]	$wR_2 = 0.0886$	$wR_2 = 0.0860$	$wR_2 = 0.2692$
Final R indexes [all data]	$R_1 = 0.0366$	$R_1 = 0.0347$	$R_1 = 0.1274,$
	$wR_2 = 0.0910$	$wR_2 = 0.0868$	$wR_2 = 0.3203$
Largest diff. peak/hole / e Å ⁻³	0.17/-0.19	0.19/-0.15	0.63/-0.41

Single Crystal X-Ray Diffraction Data

Table S2. Crystal data and structure refinement for 3b, 3i and 5b.



Figure S5 – ORTEP drawing of **3b**. All non-hydrogen atoms shown as ellipsoids at the 50% probability level. H atoms (isotropically refined) are reported in ball-and-stick style for the sake of clarity. Colour code: O=red, N=blue, C=white. Symmetry related atoms are marked with the same label.



Figure S6 – ORTEP drawing of **3i**. All non-hydrogen atoms shown as ellipsoids at the 50% probability level. H atoms (isotropically refined) are reported in ball-and-stick style for the sake of clarity. Colour code: O=red, N=blue, C=white. Symmetry related atoms are marked with the same label.



Figure S7 – ORTEP drawing of asymmetric unit of **5b**. All non-hydrogen atoms shown as ellipsoids at the 50% probability level. H atoms (isotropically refined) are reported in ball-and-stick style for the sake of clarity. Colour code: O=red, N=blue, C=white. Disordered atoms are split in two parts labelled with *a* and *b* suffixes respectively.

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7. Copy of NMR Spectra







S22

























f1 (ppm)



























S46



200 190 f1 (ppm)











f1 (ppm)

