

Supporting information

**Ethanol-assisted mechanochemical asymmetric
cross-dehydrogenative coupling reaction with recoverable
chiral amine/NaCl for accessing chiral α -alkyl α -glycine
derivatives**

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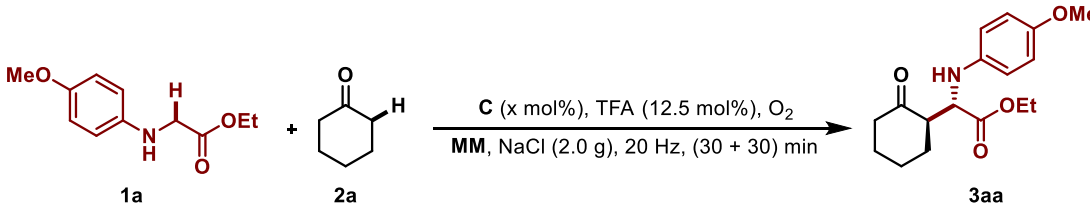
1. General information

All of the ball milling reactions were conducted in a Mixer mill (MM 400 RetschGmbH, Hann, Germany) with 25 mL stainless-steel vessels (equipped with gas inlet and outlet valve) with stainless-steel balls, if not mentioned otherwise. Reactions were monitored by Thin Layer Chromatography (TLC) using UV light (254 nm) for detection. ^1H , ^{13}C and ^{19}F NMR spectra were recorded on Bruker 400, 500 or 600 MHz spectrometer in CDCl_3 with tetramethylsilane (TMS) as internal standard. Chemical shifts are reported in parts per million (ppm). The following abbreviations were used to explain multiplicities: s = singlet, brs = broad singlet, d = doublet, t = triplet, q = quadruplet, m = multiplet and the coupling constants (J) were reported in Hertz unit (Hz). Melting points were measured using an SRS OptiMelt MPA100 apparatus and were uncorrected. High Resolution Mass Spectrometry (HRMS) and Electrospray Ionization-Mass Spectrometry (ESI-MS) were recorded on an Agilent 6210 LC/TOFMS or Agilent 6550 QTOFMS. High Performance Liquid Chromatography (HPLC) were performed on SHIMADZU LC-20AT apparatus, using Daicel Chiralpak AD-H chiral column, eluted with a mixture of hexane and isopropyl alcohol. Optical Rotations were measured with Rudolph Autopol V polarimeter. X-ray crystallographic experiments were performed by the Crystallography Service of the Department of Chemistry, Zhejiang University. The synthesis of *N*-arylglycine esters/amide substrates was meticulously carried out under solvent-free ball-milling conditions, in accordance with our recent publication¹.

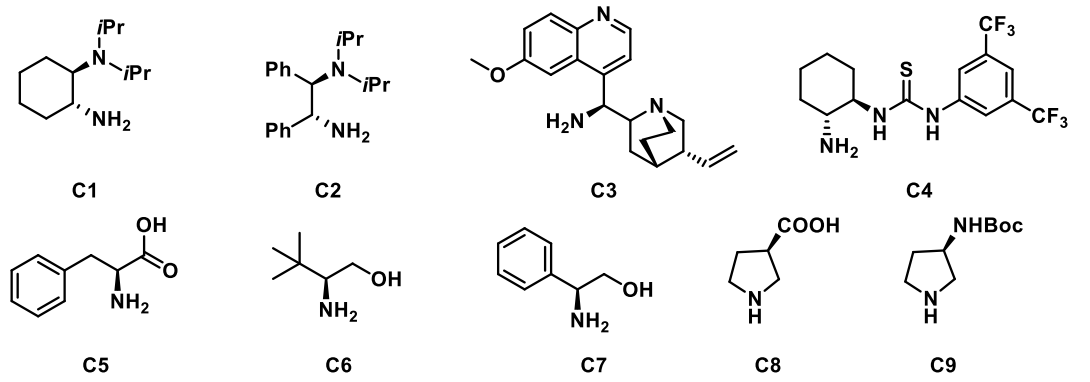
2. Reaction optimization & typical procedures

2.1 Optimization of the reaction conditions

Table S1. Screening of ligands^a



Reaction scheme showing the synthesis of 3aa from 1a and 2a. 1a is ethyl 2-(4-methoxyphenyl)glycidate, and 2a is cyclohexanone. The reaction uses catalyst C (x mol%), TFA (12.5 mol%), O₂, MM, NaCl (2.0 g), 20 Hz, (30 + 30) min to produce 3aa, ethyl 2-(4-methoxyphenyl)cyclohexanecarboxylate.



Chemical structures of ligands C1 through C9:

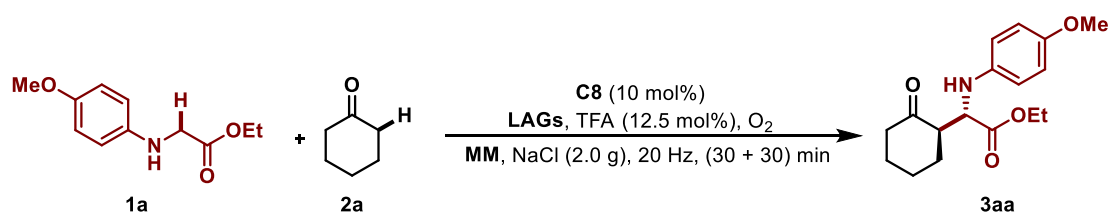
- C1: N,N-diisopropylcyclohexylamine
- C2: N,N-diisopropyl-1,2-diphenylethane-1,2-diamine
- C3: A complex ligand with a quinoline ring, a methoxy group, and a bicyclic amine moiety
- C4: N-(2,4,6-trifluorophenyl)ethane-1,2-diamine
- C5: 1-phenylethylamine
- C6: 1-tert-butylethylamine
- C7: 1-phenylethylamine
- C8: 1-aminopropanoic acid
- C9: N-Boc-1-aminopropanoic acid

entry	catalyst (mol%)	yield (%) ^b	ee (%) ^b	dr ^b
1	C1 (20)	39	<5	96: 4
2	C2 (20)	35	<5	84: 16
3	C3 (20)	31	<5	50: 50

4	C4 (20)	39	43	82: 18
5	C5 (20)	35	16	60: 40
6	C6 (20)	30	13	50: 50
7	C7 (20)	33	10	55: 45
8	C8 (20)	37	58	88: 12
9	C9 (20)	42	<5	50: 50
10	C8 (15)	47	55	85: 15
11	C8 (10)	46	58	87: 13
12	C8 (5)	43	52	75: 25
13 ^c	C8 (10)	51	59	88: 12
14 ^d	C8 (10)	42	55	85: 15
15 ^e	C8 (10)	35	52	81: 19

^a Reaction conditions: **1a** (0.2 mmol), TFA (12.5 mol%), NaCl (2.0 g) were pre-milled at 20 Hz for 30 min under oxygen conditions, using two stainless-steel balls ($d_{MB} = 1.2$ cm) in a 25 mL stainless vial, then **2a** (5 equiv.), **C** (x mol%) were added and milled for another 30 min. ^b Yields were those of the isolated products, ee values were determined by HPLC, *dr* values were determined by ¹H NMR. ^c **2a** (4 equiv.) was used. ^d **2a** (3.5 equiv.) was used. ^e **2a** (3 equiv.) was used.

Table S2. Screening of LAGs^a

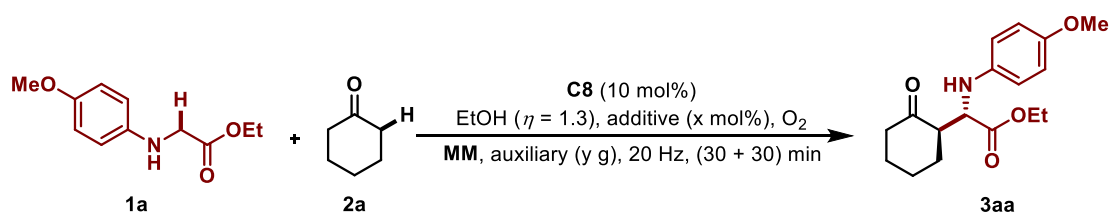


entry	LAGs (η)	yield (%) ^b	ee (%) ^b	<i>dr</i> ^b
1	TFA (1.5)	21	<5	20: 80
2	DMAE (1.5)	56	79	77: 23
3	MeOH (1.5)	53	73	80: 20
4	EtOH (1.5)	62	85	98: 2
5	<i>i</i> -PrOH (1.5)	58	83	98: 2
6	<i>t</i> -BuOH (1.5)	57	82	95: 5
7	BnOH (1.5)	45	74	68: 32
8	HFIP (1.5)	39	75	78: 22
9	H ₂ O (1.5)	42	9	56: 14
10	MeCN (1.5)	52	57	83: 17
11	EtOAc (1.5)	40	<5	50: 50
12	DMF (1.5)	25	54	50: 50

13	DMSO (1.5)	29	53	85: 15
14	NMP (1.5)	20	41	70: 30
15	Acetone (1.5)	49	37	60: 40
16	Et ₂ O (1.5)	32	5	55: 45
17	DBE (1.5)	22	14	50: 50
18	THF (1.5)	27	18	64: 36
19	1,4-Dioxane (1.5)	23	13	50: 50
20	DCM (1.5)	38	<5	55: 45
21	Cyclohexane (1.5)	35	5	50: 50
22	<i>n</i> -Hexane (1.5)	37	<5	43: 57
23	Toluene (1.5)	24	5	50: 50
24	EtOH (1.3)	71	89	98: 2
25	EtOH (1.1)	63	80	95: 5
26	EtOH (1.7)	61	87	96: 4

^a Reaction conditions: **1a** (0.2 mmol), TFA (12.5 mol%), NaCl (2.0 g) were pre-milled at 20 Hz for 30 min under oxygen conditions, using two stainless-steel balls ($d_{MB} = 1.2$ cm) in a 25 mL stainless vial, then **2a** (4 equiv.), **C8** (10 mol%) and LAGs (η) [$\eta = V$ (liquid; μL)/ m (reagents; mg)] were added and milled for another 30 min. ^b Yields were those of the isolated products, *ee* values were determined by HPLC, *dr* values were determined by ¹H NMR. TFA = Trifluoroacetic acid; DMAE = Dimethylaminoethanol; HFIP = Hexafluoroisopropanol; DMF = *N,N*-Dimethylformamide; DMSO = Dimethyl sulfoxide; NMP = *N*-Methylpyrrolidone; DBE=Dibutyl ether; THF = Tetrahydrofuran; DCM = Dichloromethane.

Table S3. Screening of the additives and auxiliaries^a

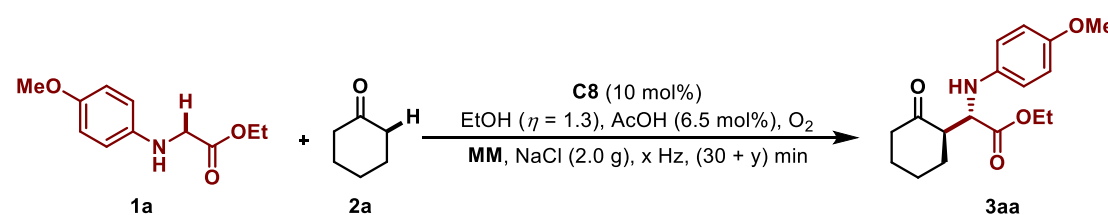


entry	additive (mol%)	auxiliary (g)	yield (%) ^b	<i>ee</i> (%) ^b	<i>dr</i> ^b
1	TFA (12.5)	NaCl (2.0)	71	89	98: 2
2	MeSO ₃ H (12.5)	NaCl (2.0)	n.r.	–	–
3	TfOH (12.5)	NaCl (2.0)	48	80	94: 6
4	AcOH (12.5)	NaCl (2.0)	66	90	97: 3
5	AcOH (9.5)	NaCl (2.0)	69	89	97: 3
6	AcOH (6.5)	NaCl (2.0)	75	91	98: 2
7	AcOH (6.5)	Silica gel (2.0)	53	51	65: 35
8	AcOH (6.5)	Neutral Al ₂ O ₃ (2.0)	n.r.	–	–

9	AcOH (6.5)	Na ₂ SO ₄ (2.0)	72	85	90: 10
10	AcOH (6.5)	NaCl (1.0)	65	90	97: 3
11	AcOH (6.5)	NaCl (3.0)	66	89	97: 3

^a Reaction conditions: **1a** (0.2 mmol), additive, and grinding auxiliary were pre-milled at 20 Hz for 30 min under oxygen conditions, using two stainless-steel balls ($d_{MB} = 1.2$ cm) in a 25 mL stainless vial, then **2a** (4 equiv.), **C8** (10 mol%) and EtOH ($\eta = 1.3$) were added and milled for another 30 min. ^b Yields were those of the isolated products, ee values were determined by HPLC, dr values were determined by ¹H NMR. n.r. = no reaction.

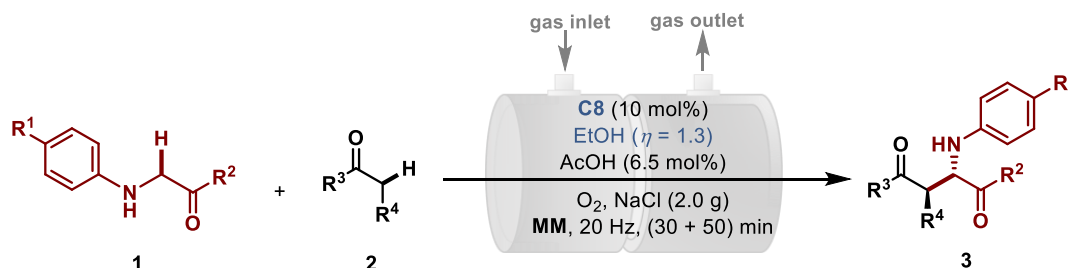
Table S4. Screening of the mechanical parameters^a



entry	Frequency (Hz)	balls (n×mm)	time (min+min)	yield (%) ^b	ee (%) ^b	dr ^b
1	25	2×12	30+30	82	82	88: 12
2	15	2×12	30+30	54	78	76: 24
3	20	2×12	30+30	75	91	98: 2
4	20	2×12	30+40	78	93	99: 1
5	20	2×12	30+50	80	95	>99: 1
6	20	2×12	30+60	83	89	97: 3
7	20	2×14	30+50	78	85	97: 3
8	20	2×10	30+50	76	90	99: 1

^a Reaction conditions: **1a** (0.2 mmol), AcOH (6.5 mol%), NaCl (2.0 g) were pre-milled at x Hz for 30 min under oxygen conditions, using two stainless-steel balls in a 25 mL stainless vial, then **2a** (4 equiv.), **C8** (10 mol%) and EtOH ($\eta = 1.3$) were added and milled for another y min. ^b Yields were those of the isolated products, ee values were determined by HPLC, dr values were determined by ¹H NMR.

2.2 Typical procedures for LAG induced asymmetric CDC reaction



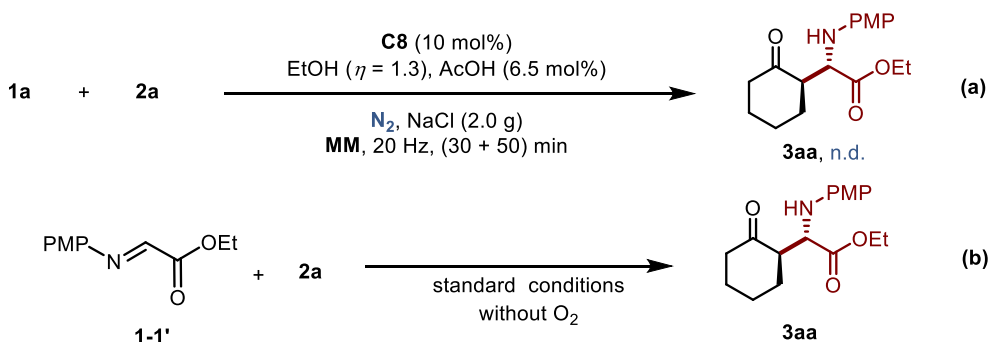
Typical procedure: a mixture of **1** (0.2 mmol), AcOH (6.5 mol%) and NaCl (2.0 g) were placed in a stainless-steel vessel (25 mL, equipped with gas inlet and outlet valve) with two stainless-steel balls ($d_{MB} = 1.2$ cm), and oxygen was filled in through the gas inlet valve. Then, the ball milling vessel was placed in

the mixer mill and pre-milled at 20 Hz for 30 min. After that, **2** (0.8 mmol, 4.0 equiv.), **C8** (0.02 mmol, 0.1 equiv.) and EtOH ($\eta = 1.3$) were added and oxygen was also filled in through the gas inlet valve. The mixtures were milled at 20 Hz for another 2×25 min, then the contents were scratched off the vessel and purified directly by column chromatography on silica gel using petroleum ether/EtOAc as eluent to give the desired products **3**.

The preparation procedure for **3ac**, **3ah**, **3aj**, **3ak** and **3am**, using the LAG method follows the typical procedure. After the ball-milling, the contents were rinsed with petroleum ether/EtOAc (10:1), the filtrate was concentrated and dried under vacuum to give the pure products.

3. Mechanism study

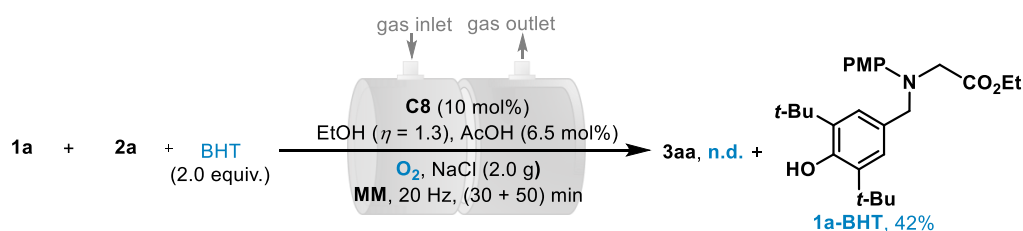
3.1 Control experiments



entry	variation from "standard conditions ^a (without O ₂)"	yield (%) ^b	ee (%) ^b
1	--	85	82
2	without EtOH	78	70
3	without AcOH	86	90
4	without AcOH and EtOH	82	91

Reaction conditions: (a) **1a** (0.2 mmol), AcOH (6.5 mol%) and NaCl (2.0 g) were placed in a stainless-steel vessel (25 mL, equipped with gas inlet and outlet valve) with two stainless-steel balls ($d_{MB} = 1.2$ cm) under nitrogen atmospheres in a mixer mill and pre-milled at 20 Hz for 30 min. Then, **2a** (4.0 equiv.), **C8** (10 mol%) and EtOH ($\eta = 1.3$) were added under nitrogen atmospheres and the mixtures were milled for another 2×25 min under nitrogen atmospheres. (b) **1-1'** (0.2 mmol), AcOH (6.5 mol%) and NaCl (2.0 g) was placed in a stainless-steel vessel (25 mL) with two stainless-steel balls ($d_{MB} = 1.2$ cm). Then, the ball milling vessel was placed in the mixer mill and pre-milled at 20 Hz for 30 min. After that, **2a** (4.0 equiv.), **C8** (10 mol%) and EtOH ($\eta = 1.3$) were added. The mixtures were milled at 20 Hz for another 2×25 min.

3.2 Radical trapping experiments



Reaction conditions: **1a** (0.2 mmol), AcOH (6.5 mol%), BHT (2.0 equiv.) and NaCl (2.0 g) were placed in a stainless-steel vessel (25 mL, equipped with gas inlet and outlet valve) with two stainless-steel balls ($d_{MB} = 1.2$ cm), and oxygen was filled in through the gas inlet valve. Then, the ball milling vessel was placed in the mixer mill and pre-milled at 20 Hz for 30 min. After that, **2a** (4.0 equiv.), **C8** (10 mol%) and EtOH ($\eta = 1.3$) were added and oxygen was also filled in through the gas inlet valve. The mixtures were milled at 20 Hz for another 2×25 min.

3.3 Linear effect experiments

The reactions were conducted following the typical procedure (see section 2.2).

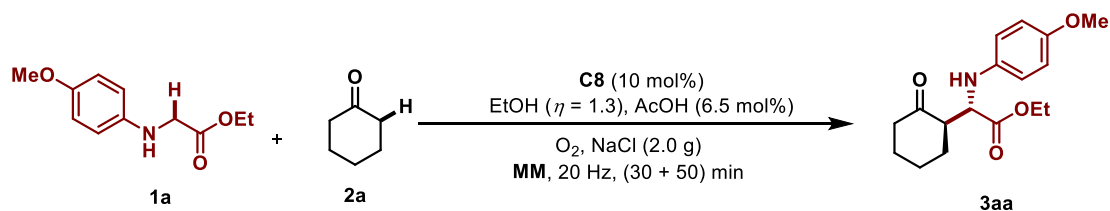


Table S5. Relationship between the ee values of **C8** and **3aa**.

ee % (C8)	0	5	15	25	30	40	50	60	75	85	99
ee % (3aa)	0	3	15	27	33	46	57	65	77	82	95

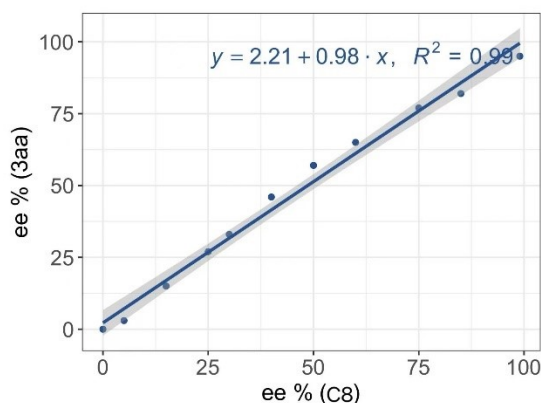
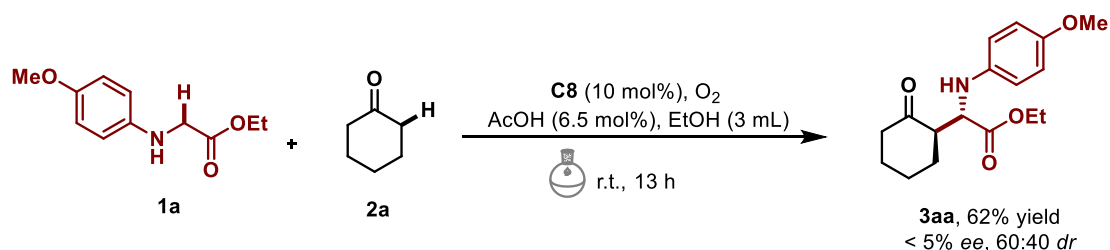


Figure S1. Relationship between the ee values of **C8** and **3aa**.

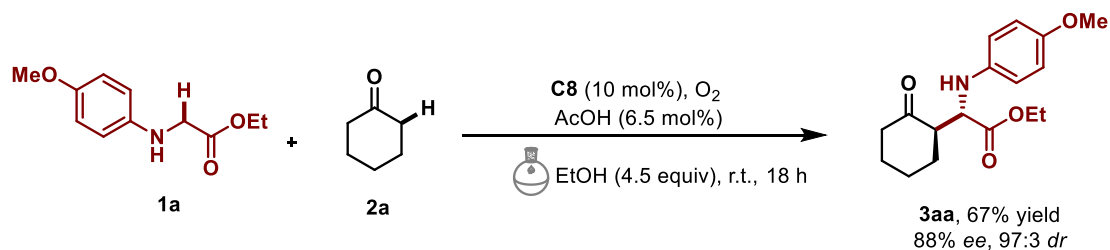
3.4 Comparative experiments

Comparative experiments under solution-based conditions.

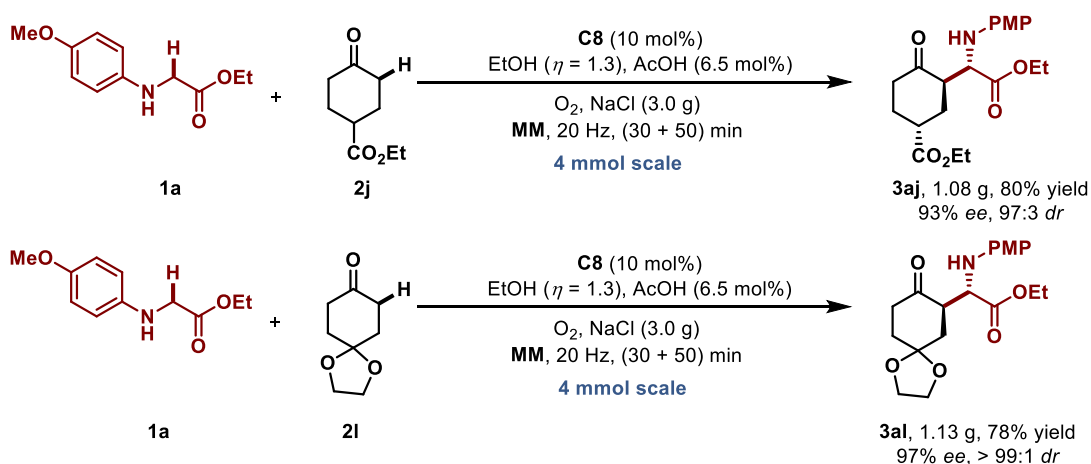


Reaction of **1a** and **2a** under solution-based conditions. Reaction conditions: **1a** (0.2 mmol) and AcOH (6.5 mol%) were placed in a flask (10 mL) with EtOH (3 mL) under oxygen atmosphere pre-stirred at rt for 6 h, then **2a** (0.8 mmol, 4.0 equiv.) and **C8** (10 mol%) were added, and the mixtures were stirring for 7 h.

Comparative experiments under neat-stirring conditions.

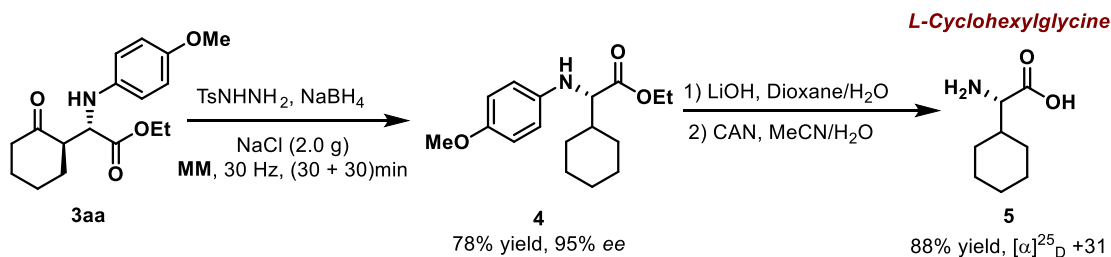


3.5 Gram-scale reactions



A mixture of **1a** (4 mmol), AcOH (6.5 mol%) and NaCl (3.0 g) were placed in a stainless-steel vessel (50 mL, equipped with gas inlet and outlet valve) with two stainless-steel balls ($d_{MB} = 1.2$ cm), and oxygen was filled in through the gas inlet valve. Then, the ball milling vessel was placed in the mixer mill and pre-milled at 20 Hz for 30 min. After that, **2j** (4.0 equiv.) or **2k** (4.0 equiv.), **C8** (10 mmol%) and EtOH ($\eta = 1.3$) were added and oxygen was also filled in through the gas inlet valve. The mixtures were milled at 20 Hz for another 2×25 min. After the ball-milling, the contents were rinsed with petroleum ether/EtOAc (10:1), the filtrate was concentrated and dried under vacuum to give **3aj** (1.08 g) or **3al** (1.13 g).

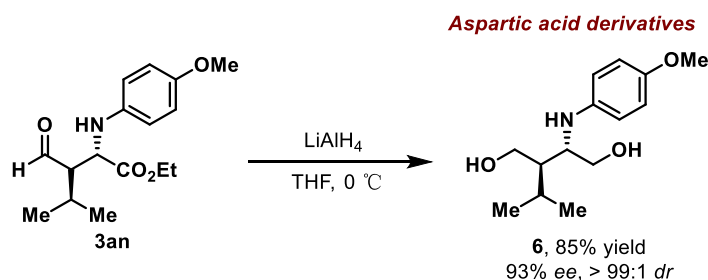
3.6 Synthetic applications



Step1: A mixture of **3aa** (500 mg, 1.637 mmol), tosylhydrazine (427 mg, 1.4 equiv.) and NaCl (2.0 g) were placed in a stainless-steel vessel (25 mL) with two stainless-steel balls ($d_{MB} = 1.2$ cm). Then, the ball milling vessel was placed in the mixer mill and milled at 30 Hz for 30 min. After that, NaBH₄ (620 mg)

were added and the mixtures were milled at 30 Hz for 30 min, then the contents were scratched off the vessel, then brine was added and the mixture was extracted with EtOAc. The combined organic layers were dried and concentrated, and the residue was purified by flash column chromatography to afford ethyl (*R*)-*N*-(*p*-methoxyphenyl)-cyclohexylglycinate **4** in 78% yield with 95% ee.

Step2: To a solution of **4** (291 mg, 1.0 mmol) in dioxane/water (14 mL, 1:1) was added LiOH (280 mg) and the mixture was stirred at room temperature for 14 h. Then, the mixture was acidified with 1 N HCl and extracted with EtOAc. The combined organic layers were dried, filtered, and concentrated. Purification of the residue by flash column chromatography afforded known *N*-PMP-protected cyclohexylglycine. The solution of ceric ammonium nitrate (CAN, 1.1 g, 2 mmol, 2.5 equiv.) distilled water (8.0 mL) was added slowly to the stirred solution of *N*-PMP-protected cyclohexylglycine (200 mg, 0.81 mmol) in MeCN/H₂O (3/1, 16.0 mL) at 0 °C. The reaction mixture was further stirred at 0 °C for about 1 h, till the reaction completed as monitored by TLC. The reaction was then quenched by adding the saturated Na₂SO₄ solution and extracted with EtOAc. The combined organic layer was extracted with 0.1 M HCl. The combine aqueous layer was neutralized by NaHCO₃ (pH = 7) and extracted with EtOAc. The combined organic layer was washed with brine solution, dried over anhydrous Na₂SO₄ and evaporated under reduced pressure, product **5** (138.2 mg, 88% yield) was afforded without further purification.



A solution of **3an** (118 mg, 0.4 mmol) in THF (20 mL) was cooled to 0 °C and LiAlH₄ (10 mL, 1 M solution in THF) was added. After stirring for 30 min, the ice-bath was removed and the mixture was stirred for 1.5 h at room temperature. The mixture was quenched by careful addition of aqueous NH₄Cl solution, followed by 3 M HCl, and then extracted with EtOAc. The combined organic layers were dried, filtered, and concentrated. Purification of the residue by flash column chromatography afforded (2*R*,3*S*)-2-isopropyl-3-((4-methoxyphenyl)amino)butane-1,4-diol **6** (86 mg, 85% yield, 93% ee, >99:1 dr) as pale yellow oil.

3.7 Recycling of grinding auxiliary (NaCl) and C8

After the reaction was completed (see section 2.2), the mixtures were dissolved in EtOAc, then filtered to give the recovered NaCl as offwhite solid, which can be directly used for the next reaction after drying under reduced pressure. *No additional acetic acid or C8 was required when using the recovered NaCl for a fresh reaction, which implies the chiral catalyst can be simultaneously recovered with NaCl, and its inherent acidity was enough to promote the aerobic oxidation of glycinate.*

For the synthesis of **3aa**, NaCl along with **C8** can be recycled and reused for at least 5 times and its recovery yields (mass of recovered solid g/2.0 g) were ranging from 96% to 99%. The average consumption of NaCl for single reaction is 16 mg. (*Fresh NaCl was added to keep its mass at 2.0 g for each reaction.*)

Table S6. Recycling and recovery of NaCl and **C8**.

Recovery of NaCl and C8 (g)	ee (%)	Yield (%)	dr (n:1)
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Run 1	1.98	93	78	99
Run 2	1.97	92	77	99
Run 3	1.95	92	79	99
Run 4	1.94	91	77	99
Run 5	1.92	91	76	98

3.8. Green chemistry metrics evaluation

The green chemistry metrics including Effective Mass Yield (EMY), Atom Economy (AE), Atom Efficiency (AEF), Reaction Mass Efficiency (RME), Optimum Efficiency (OE), Process Mass Intensity (PMI), Mass Intensity (MI), Mass Productivity (MP), *E*-factor, and Solvent Intensity (SI), were calculated according to the reported method.¹ EcoScale was also calculated according to the reported method.²

Based on the mechanochemical LAG method, the green chemistry metrics were calculated as follows (all the calculations are made on the basis that NaCl and **C8** can be recovered for five times) :

$$\begin{aligned} E \text{ factor} &= \frac{\sum m (\text{reactants}) + \sum m (\text{reagents}) + \sum m (\text{solvent}) + \sum m (\text{additives}) - \sum m (\text{products})}{\sum m (\text{products})} \\ &= \frac{39.04 + 78.52 + 0.23 + 44.90 + 0.525 + 16 - 46.03}{46.03} = 2.89 \end{aligned}$$

$$RME = \frac{\sum m (\text{products})}{\sum m (\text{reactants})} \times 100\% = \frac{46.03}{39.04 + 78.52} \times 100\% = 39.15\%$$

$$EMY = \frac{\sum m (\text{products})}{\sum m (\text{toxic reagents})} \times 100\% = \frac{46.03}{39.04 + 78.52 + 0.23 + 44.90 + 0.525 + 16} \times 100\% = 25.68\%$$

$$AE = \frac{\sum MW (\text{products})}{\sum MW (\text{reactants})} \times 100\% = \frac{291.35}{195.22 + 98.15 \times 4} = 49.56\%$$

$$AEF = AE \times Yield\% = 49.56\% \times 79\% = 39.15\%$$

$$OE = \frac{RME}{AE} \times 100\% = \frac{39.15}{49.56} = 80.00\%$$

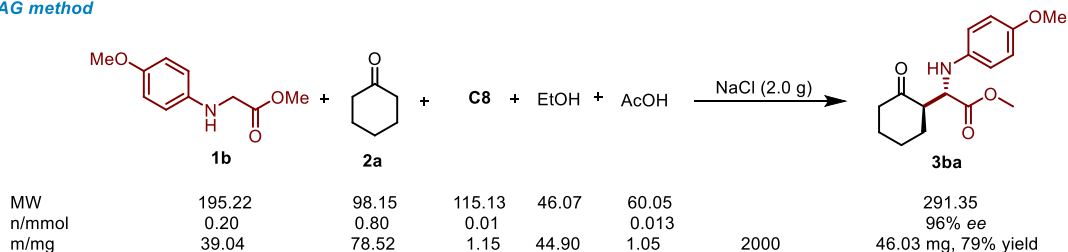
$$\begin{aligned} PMI &= \frac{\sum m (\text{reactants}) + \sum m (\text{reagents}) + \sum m (\text{solvents}) + \sum m (\text{additives})}{\sum m (\text{products})} \\ &= \frac{39.04 + 78.52 + 0.23 + 44.90 + 0.525 + 16}{46.03} = 3.89 \end{aligned}$$

$$\begin{aligned} MI &= \frac{\sum m (\text{reactants}) + \sum m (\text{reagents}) + \sum m (\text{solvents (except water)}) + \sum m (\text{additives})}{\sum m (\text{products})} \\ &= \frac{39.04 + 78.52 + 0.23 + 44.90 + 0.525 + 16}{46.03} = 3.89 \end{aligned}$$

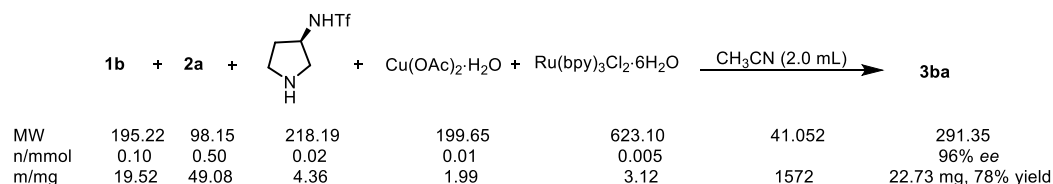
$$MP = \frac{1}{MI} \times 100\% = \frac{1}{3.89} = 25.69\%$$

$$SI = \frac{\sum m(\text{solvents (except water)})}{\sum m(\text{products})} = \frac{44.90}{46.03} = 0.98$$

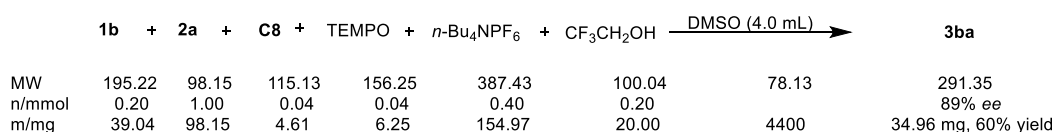
LAG method



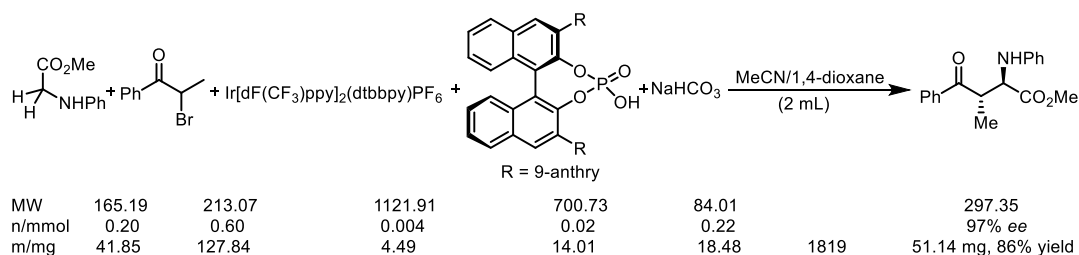
Zhang's method



Mei's method



Wang's method



Tanaka's method

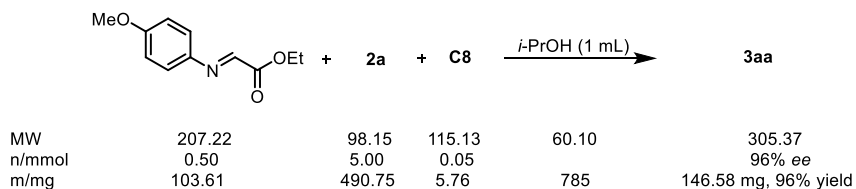


Table S7. Comparison of green chemistry metrics.

Entry		LAG method	Zhang's work	Mei's work	Wang's work	Tanaka's work
Product	Yield (%)	79	78	60	86	96

	ee (%)	96	96	89	97	96
	dr	>99:1	97:3	>99:1	>20:1	>99:1
Effective Mass Yield	EMY (%)	25.68	1.39	0.74	2.52	10.58
Atom Economy	AE (%)	49.56	42.47	42.47	36.97	25.69
Atom Efficiency	AEF (%)	39.15	33.13	25.48	31.79	24.66
Reaction Mass Efficiency	RME (%)	39.15	33.13	25.48	30.14	24.66
Optimum Efficiency	OE (%)	79.00	78.01	60.00	81.53	96.00
Process Mass Intensity	PMI	3.89	71.74	135.10	39.61	9.45
Mass Intensity	MI	3.55	69.16	135.10	39.61	9.45
Mass Productivity	MP (%)	25.69	1.45	0.74	2.52	10.58
E-Factor	E-Factor	2.89	70.74	134.10	38.61	8.45
Solvent Intensity	SI	0.98	69.16	125.86	35.57	5.36
Reaction Time	h	1.33	>3	8	24	12

Table S8. Calculation of eco-scale score of LAG method for the preparation of **3ba**.

Parameters	Penalty points (LAG method)																					
1. Yield (100-x)/2 Mechanochemical synthesis: 79%	10.5																					
2. Price of reaction components (to obtain 10 mmol of end product)																						
<table border="1"> <thead> <tr> <th>Reaction components to get 10 mmol of product</th> <th>Price/g or mL</th> <th>Price to get 10 mmol of product</th> </tr> </thead> <tbody> <tr> <td>a. 1c (2.471 g)</td> <td></td> <td></td> </tr> <tr> <td>b. 2a (5.215 mL)</td> <td>0.08</td> <td>0.42</td> </tr> <tr> <td>c. C8 (0.0146 g)</td> <td>112.70</td> <td>1.646</td> </tr> <tr> <td>d. EtOH (3.6 mL)</td> <td>0.063</td> <td>0.23</td> </tr> <tr> <td>e. AcOH (0.0126 mL)</td> <td>0.020</td> <td>0.00026</td> </tr> <tr> <td>f. NaCl (25.32 g)</td> <td>0.012</td> <td>0.304</td> </tr> </tbody> </table>	Reaction components to get 10 mmol of product	Price/g or mL	Price to get 10 mmol of product	a. 1c (2.471 g)			b. 2a (5.215 mL)	0.08	0.42	c. C8 (0.0146 g)	112.70	1.646	d. EtOH (3.6 mL)	0.063	0.23	e. AcOH (0.0126 mL)	0.020	0.00026	f. NaCl (25.32 g)	0.012	0.304	
Reaction components to get 10 mmol of product	Price/g or mL	Price to get 10 mmol of product																				
a. 1c (2.471 g)																						
b. 2a (5.215 mL)	0.08	0.42																				
c. C8 (0.0146 g)	112.70	1.646																				
d. EtOH (3.6 mL)	0.063	0.23																				
e. AcOH (0.0126 mL)	0.020	0.00026																				
f. NaCl (25.32 g)	0.012	0.304																				
total price (Mechanochemical synthesis) = \$2.596 Expensive (> \$10 and <\$50)	0																					
3. Safety EtOH (F, highly flammable)	5																					
4. Technical setup Mechanochemical synthesis: Unconventional activation technique (mechanical activation)	2																					
5. Temperature/Time Mechanochemical synthesis: Room temperature, <24 h	1																					
6. Workup and purification Mechanochemical synthesis: Classical chromatography	0																					

Total	19.5
Eco-scale score	81.5

Table S9. Calculation of eco-scale score of Zhang's method for the preparation of **3ba**.

Parameters			Penalty points (Zhang's method)
1. Yield (100-x)/2 Synthesis: 78%			11
2. Price of reaction components (to obtain 10 mmol of end product)			
Reaction components to get 10 mmol of product	Price/g or mL	Price to get 10 mmol of product	
a. 1c (2.502 g)			
b. 2a (6.601 mL)	101.44	253.80	
c. C ₅ H ₉ F ₃ N ₂ O ₂ S (0.559 g)	0.08	0.53	
d. Cu(OAc) ₂ ·H ₂ O (0.255 g)	0.039	0.01	
e. Ru(bpy) ₃ Cl ₂ ·6H ₂ O (0.400 g)	18.80	7.52	
f. MeCN (256.6 mL)	0.0083	2.14	
total price (Mechanochemical synthesis) = \$264 Very expensive (> \$50)			5
3. Safety			
a. Cu(OAc) ₂ ·H ₂ O (N, dangerous for environment)			5
b. MeCN (T, toxic)			5
c. MeCN (F+, extremely flammable)			10
4. Technical setup			
Photocatalytic synthesis: Unconventional activation technique (photochemical activation)			2
5. Temperature/Time			
Photocatalytic synthesis: Room temperature, <24 h			1
6. Workup and purification			
Photocatalytic synthesis: Classical chromatography			0
Total			39
Eco-scale score			61

Table S10. Calculation of eco-scale score of Mei's method for the preparation of **3ba**.

Parameters			Penalty points (Mei's method)
1. Yield (100-x)/2 Synthesis: 60%			20
2. Price of reaction components (to obtain 10 mmol of end product)			
Reaction components	Price/g or mL	Price to get 10	

to get 10 mmol of product		mmol of product	
a. 1c (3.254 g)			
b. 2a (8.583 mL)	0.08	0.68	
c. L8 (0.384 g)	112.70	43.27	
d. TEMPO (0.521 g)	3.0596	1.59	
e. <i>n</i> -Bu ₄ NPF ₆ (12.915 g)	0.667	8.62	
f. CF ₃ CH ₂ OH (1.667 g)	4.1722	6.95	
g. DMSO (333.4 mL)	0.0092	3.06	
total price (Mechanochemical synthesis) = \$64.17 Very expensive (> \$50)			5
3. Safety			
a. CF ₃ CH ₂ OH (T, toxic)			5
b. CF ₃ CH ₂ OH (F, highly flammable)			5
c. DMSO (T, toxic)			5
d. DMSO (F+, extremely flammable)			10
4. Technical setup			
Electrochemical synthesis: Unconventional activation technique (electrical activation)			2
5. Temperature/Time			
Electrochemical synthesis: Room temperature, <24 h			1
6. Workup and purification			
Electrochemical synthesis: Classical chromatography			0
Total			53
Eco-scale score			47

Table S11. Calculation of eco-scale score of Wang's method.

Parameters			Penalty points (Wang's method)
1. Yield (100-x)/2 Synthesis: 86%			7
2. Price of reaction components (to obtain 10 mmol of end product)			
Reaction components to get 10 mmol of product	Price/g or mL	Price to get 10 mmol of product	
a. C ₉ H ₁₁ NO ₂ (2.433 g)	0.4033	0.98	
b. C ₉ H ₉ Br (7.4333 g)			
c. Ir[dF(CF ₃)ppy] ₂ (dtb bpy)PF ₆ (0.261 g)	237.65	62.03	
d. Ligand (0.815 g)	624.30	508.80	
e. NaHCO ₃ (1.074 g)	0.0042	0.0045	
f. MeCN (58.14 mL)	0.0083	0.48	
g. 1,4-dioxane (58.14 mL)	0.0078	0.45	

total price (Mechanochemical synthesis) = \$686.47 Very expensive (> \$50)	5
3. Safety a. MeCN (T, toxic) b. MeCN (F+, extremely flammable) c. 1,4-dioxane (F, highly flammable)	5 10 5
4. Technical setup Photocatalytic synthesis: Unconventional activation technique (photochemical activation)	2
5. Temperature/Time Photocatalytic synthesis: Room temperature, <24 h	1
6. Workup and purification Photocatalytic synthesis: Classical chromatography	0
Total	35
Eco-scale score	65

Table S12. Calculation of eco-scale score of Tanaka's method.

Parameters	Penalty points (Tanaka's method)															
1. Yield (100-x)/2 Synthesis: 96%	2															
2. Price of reaction components (to obtain 10 mmol of end product)																
<table border="1"> <thead> <tr> <th>Reaction components to get 10 mmol of product</th> <th>Price/g or mL</th> <th>Price to get 10 mmol of product</th> </tr> </thead> <tbody> <tr> <td>a. C₁₁H₁₃NO₃ (2.158 g)</td> <td></td> <td></td> </tr> <tr> <td>b. 2a (10.73 mL)</td> <td>0.08</td> <td>0.86</td> </tr> <tr> <td>c. C8 (0.112 g)</td> <td>112.70</td> <td>12.62</td> </tr> <tr> <td>d. <i>i</i>-PrOH (3 mL)</td> <td>0.005</td> <td>0.10</td> </tr> </tbody> </table>	Reaction components to get 10 mmol of product	Price/g or mL	Price to get 10 mmol of product	a. C ₁₁ H ₁₃ NO ₃ (2.158 g)			b. 2a (10.73 mL)	0.08	0.86	c. C8 (0.112 g)	112.70	12.62	d. <i>i</i> -PrOH (3 mL)	0.005	0.10	
Reaction components to get 10 mmol of product	Price/g or mL	Price to get 10 mmol of product														
a. C ₁₁ H ₁₃ NO ₃ (2.158 g)																
b. 2a (10.73 mL)	0.08	0.86														
c. C8 (0.112 g)	112.70	12.62														
d. <i>i</i> -PrOH (3 mL)	0.005	0.10														
total price (Mechanochemical synthesis) = \$13.58 Expensive (> \$10 and <\$50)	3															
3.Safety <i>i</i> -PrOH (F, highly flammable)	5															
4. Technical setup: Solution synthesis: Common setup	0															
5. Temperature/Time Solution synthesis: Room temperature, <24 h	1															
6. Workup and purification Solution synthesis: Classical chromatography	0															
Total	11															
Eco-scale score	89															

3.9 Crystal data for 3ha.

Single crystal of minor diastereomers of **3ha** suitable for X-ray analysis was obtained by slow evaporation of 0.01 M solution in 7:3 mixture of petroleum ether/ ethyl acetate at room temperature. A suitable crystal was selected on a Bruker APEX-II CCD diffractometer. The crystal was kept at 296.15 K during data collection. Using Olex2,³ the structure was solved with the SHELXT⁴ structure solution program using Intrinsic Phasing and refined with the SHELXT refinement package using Least Squares minimisation.

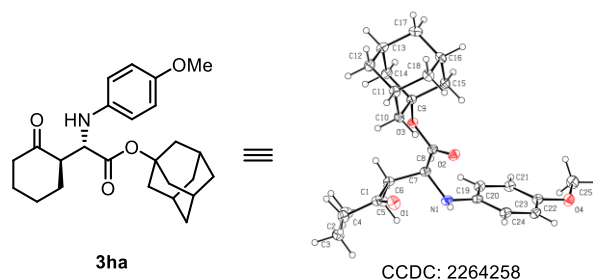


Table S13. Crystal data and structure refinement for compound **3ha**.

Identification code	cu_230519_CH_3ka_0m
Empirical formula	C ₂₅ H ₃₃ NO ₄
Formula weight	411.52
Temperature/K	170.00
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	6.82470(10)
b/Å	15.1975(3)
c/Å	20.6745(4)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	2144.33(7)
Z	4
ρ _{calc} /cm ³	1.275
μ/mm ⁻¹	0.682
F(000)	888.0
Crystal size/mm ³	0.48 × 0.26 × 0.2
Radiation	CuKα (λ = 1.54178)
2θ range for data collection/°	7.218 to 149.136
Index ranges	-8 ≤ h ≤ 7, -19 ≤ k ≤ 19, -25 ≤ l ≤ 25
Reflections collected	22064
Independent reflections	4363 [R _{int} = 0.0621, R _{sigma} = 0.0384]
Data/restraints/parameters	4363/1/275
Goodness-of-fit on F ²	1.032
Final R indexes [I >= 2σ (I)]	R ₁ = 0.0407, wR ₂ = 0.1041

Final R indexes [all data]	R ₁ = 0.0464, wR ₂ = 0.1077
Largest diff. peak/hole / e Å ⁻³	0.25/-0.21
Flack parameter	-0.12(12)

Table S14. Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for cu_230519_CH_3ka_0m. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
O1	6185(3)	3537.6(14)	5434.7(11)	41.4(5)
O2	6052(3)	5381.0(13)	4782.1(8)	34.3(4)
O3	4854(2)	5660.5(11)	5785.7(8)	27.8(3)
O4	-764(3)	5649.6(14)	2380.5(9)	38.0(4)
N1	3115(3)	4213.0(14)	4516.1(10)	30.2(4)
C1	4722(3)	3389.9(16)	5744.8(12)	27.6(5)
C2	4586(4)	2621.7(18)	6204.1(13)	35.7(6)
C3	2669(4)	2112.7(17)	6138.7(14)	35.8(5)
C4	922(4)	2738.8(18)	6187.8(14)	37.3(6)
C5	1036(4)	3439.4(17)	5660.5(14)	35.1(5)
C6	2933(3)	3984.6(14)	5701.9(11)	25.7(4)
C7	3101(3)	4649.9(15)	5144.8(11)	26.4(4)
C8	4873(3)	5262.9(15)	5203.8(11)	26.6(4)
C9	6514(3)	6206.5(15)	6002.7(11)	24.2(4)
C10	8341(3)	5636.7(15)	6070.8(11)	27.5(5)
C11	9989(4)	6202.0(17)	6366.5(12)	30.4(5)
C12	9347(4)	6533.4(19)	7034.4(12)	35.1(5)
C13	7510(4)	7102.6(18)	6959.8(12)	34.1(5)
C14	5858(4)	6540.2(17)	6664.5(11)	29.4(5)
C15	6903(4)	6987.6(15)	5552.6(12)	27.7(5)
C16	8560(4)	7546.2(16)	5848.3(13)	32.0(5)
C17	7949(4)	7882.2(17)	6513.9(14)	36.9(6)
C18	10407(4)	6979.1(17)	5915.5(13)	33.2(5)
C19	2094(3)	4598.6(15)	3997.2(12)	27.5(4)
C20	205(4)	4932.4(16)	4061.6(12)	30.0(5)
C21	-804(3)	5283.5(17)	3536.0(12)	29.8(5)
C22	76(4)	5298.3(16)	2931.0(12)	29.5(5)
C23	1936(4)	4940.2(17)	2852.0(12)	32.3(5)
C24	2929(4)	4603.7(16)	3380.0(12)	31.0(5)
C25	-2535(4)	6115(2)	2461.8(15)	42.3(6)

Table S15. Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for cu_230519_CH_3ka_0m. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^2U_{11}+2hka*b*U_{12}+\dots]$.

Atom	U ₁₁	U ₂₂	U ₃₃	U ₂₃	U ₁₃	U ₁₂
O1	23.6(9)	47.8(11)	52.7(11)	6.9(9)	5.8(8)	5.3(8)
O2	32.7(9)	39.4(9)	30.8(8)	-4.9(7)	7.4(7)	-3.2(8)
O3	24.8(8)	31.8(8)	26.8(7)	-4.8(6)	3.4(6)	-3.6(7)
O4	36.8(10)	44.4(10)	32.9(8)	0.7(8)	-3.0(8)	11.3(8)
N1	28.0(10)	31.6(9)	31.0(10)	-7.7(8)	-2.7(8)	6.3(8)
C1	20.8(11)	31.1(11)	30.8(10)	-4.3(9)	-2.5(9)	0.5(9)
C2	31.1(13)	36.9(13)	38.9(13)	3.4(10)	-1.2(10)	4.9(10)
C3	37.0(14)	29.6(11)	40.8(13)	0.3(10)	2.8(11)	0.7(10)
C4	27.3(12)	34.4(13)	50.1(15)	-3.5(11)	7.0(11)	-3.6(10)
C5	20.6(11)	30.4(12)	54.4(15)	-1.7(11)	0.0(10)	-0.1(9)
C6	19.7(10)	26.4(10)	31.1(10)	-5.1(8)	3.2(8)	-1.0(8)
C7	21.5(10)	27.9(10)	29.7(11)	-4.2(9)	0.1(8)	4.0(9)
C8	26.0(11)	26.2(10)	27.5(10)	-3.3(8)	0.0(9)	2.9(9)
C9	21.6(10)	25.9(10)	25.2(10)	-3.0(8)	1.3(8)	-0.1(8)
C10	27.9(11)	23.9(10)	30.8(10)	1.0(8)	1.9(9)	2.5(9)
C11	22.9(11)	33.4(12)	34.8(12)	2.2(9)	-1.7(10)	5.0(10)
C12	32.8(13)	42.0(13)	30.5(12)	-0.1(10)	-7.6(10)	0.9(11)
C13	30.9(13)	39.0(13)	32.3(12)	-9.2(10)	-1.2(10)	2.4(10)
C14	24.3(10)	35.9(12)	28.1(10)	-4.5(9)	1.8(9)	1.1(9)
C15	25.2(11)	26.6(10)	31.1(11)	2.6(8)	-2.2(9)	0.6(9)
C16	30.6(12)	25.2(10)	40.3(13)	4.0(9)	-3.0(10)	-2.5(9)
C17	32.3(13)	28.4(12)	50.1(15)	-9.1(10)	-7.4(12)	2.8(10)
C18	22.7(12)	35.6(12)	41.4(13)	2.6(10)	1.2(10)	-2.5(9)
C19	24.8(10)	25.2(10)	32.5(11)	-6.8(8)	-2.6(9)	-3.6(9)
C20	24.6(11)	34.8(11)	30.7(11)	-2.6(9)	1.2(9)	-1.9(9)
C21	20.0(10)	33.8(12)	35.6(12)	-5.3(9)	-0.1(9)	1.2(9)
C22	25.8(11)	30.7(11)	31.9(11)	-3.5(9)	-3.9(9)	0.0(9)
C23	31.0(13)	36.7(12)	29.2(11)	-6.1(9)	2.8(10)	3.5(10)
C24	24.3(10)	33.7(11)	35.1(12)	-7.5(9)	-0.2(9)	3.0(10)
C25	36.9(14)	46.3(15)	43.7(14)	1.8(12)	-4.0(11)	11.1(11)

Table S16. Bond Lengths for cu_230519_CH_3ka_0m.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
O1	C1	1.207(3)	C9	C14	1.526(3)
O2	C8	1.200(3)	C9	C15	1.531(3)
O3	C8	1.346(3)	C10	C11	1.541(3)

Table S16 continued. Bond Lengths for cu_230519_CH_3ka_0m.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
O3	C9	1.474(3)	C9	C14	1.526(3)
O4	C22	1.382(3)	C9	C15	1.531(3)
O4	C25	1.411(3)	C10	C11	1.541(3)
N1	C7	1.460(3)	C11	C12	1.534(3)
N1	C19	1.407(3)	C11	C18	1.531(3)
C1	C2	1.508(3)	C12	C13	1.531(4)
C1	C6	1.521(3)	C13	C14	1.541(3)
C2	C3	1.526(4)	C13	C17	1.531(4)
C3	C4	1.529(4)	C15	C16	1.541(3)
C4	C5	1.526(4)	C16	C17	1.526(4)
C5	C6	1.539(3)	C16	C18	1.533(3)
C6	C7	1.537(3)	C19	C20	1.392(3)
C6	C7	1.537(3)	C19	C24	1.398(3)

Table S17. Bond Angles for cu_230519_CH_3ka_0m.

Atom	Atom	Atom	Length/Å	Atom	Atom	Atom	Length/Å
C8	O3	C9	121.16(18)	C9	C10	C11	108.48(18)
C22	O4	C25	116.8(2)	C12	C11	C10	109.4(2)
C19	N1	C7	119.10(19)	C18	C11	C10	108.92(19)
O1	C1	C2	122.0(2)	C18	C11	C12	110.4(2)
O1	C1	C6	121.5(2)	C13	C12	C11	109.2(2)
C2	C1	C6	116.6(2)	C12	C13	C14	109.0(2)
C1	C2	C3	112.9(2)	C17	C13	C12	109.7(2)
C2	C3	C4	110.3(2)	C17	C13	C14	109.5(2)
C5	C4	C3	110.3(2)	C9	C14	C13	108.97(19)
C4	C5	C6	112.3(2)	C9	C15	C16	108.27(19)
C1	C6	C5	111.00(18)	C17	C16	C15	110.0(2)
C1	C6	C7	112.01(19)	C17	C16	C18	109.3(2)
C7	C6	C5	112.0(2)	C18	C16	C15	109.3(2)
N1	C7	C6	111.62(19)	C16	C17	C13	109.7(2)
N1	C7	C8	110.04(19)	C11	C18	C16	109.6(2)
C8	C7	C6	113.55(18)	C20	C19	N1	122.5(2)
O2	C8	O3	126.0(2)	C20	C19	C24	117.6(2)
O2	C8	C7	124.2(2)	C24	C19	N1	119.8(2)
O3	C8	C7	109.68(19)	C19	C20	C21	121.5(2)
O3	C9	C10	109.69(17)	C22	C21	C20	119.7(2)
O3	C9	C14	103.57(17)	O4	C22	C21	124.7(2)
O3	C9	C15	112.63(18)	O4	C22	C23	115.7(2)
C10	C9	C14	110.22(19)	C21	C22	C23	119.6(2)
C10	C9	C15	110.75(18)	C24	C23	C22	119.9(2)
C14	C9	C15	109.75(19)	C23	C24	C19	121.5(2)

Table S18. Torsion Angles for cu_230519_CH_3ka_0m.

A	B	C	D	Angle/°	A	B	C	D	Angle/°
O1	C1	C2	C3	-134.7(3)	C9	C15	C16	C17	-60.2(2)
O1	C1	C6	C5	136.6(3)	C9	C15	C16	C18	59.8(3)
O1	C1	C6	C7	10.6(3)	C10	C9	C14	C13	60.8(3)
O3	C9	C10	C11	-174.07(17)	C10	C9	C15	C16	-60.6(2)
O3	C9	C14	C13	178.12(19)	C10	C11	C12	C13	-61.0(3)
O3	C9	C15	C16	176.10(18)	C10	C11	C18	C16	61.1(3)
O4	C22	C23	C24	177.5(2)	C11	C12	C13	C14	60.7(3)
N1	C7	C8	O2	2.2(3)	C11	C12	C13	C17	-59.2(3)
N1	C7	C8	O3	-179.75(19)	C12	C11	C18	C16	-59.0(3)
N1	C19	C20	C21	-177.7(2)	C12	C13	C14	C9	-60.3(3)
N1	C19	C24	C23	177.0(2)	C12	C13	C17	C16	60.5(3)
C1	C2	C3	C4	-52.4(3)	C14	C9	C10	C11	-60.6(2)
C1	C6	C7	N1	65.2(2)	C14	C9	C15	C16	61.3(2)
C1	C6	C7	C8	-59.9(2)	C14	C13	C17	C16	-59.1(3)
C2	C1	C6	C5	-44.7(3)	C15	C9	C10	C11	61.0(2)
C2	C1	C6	C7	-170.7(2)	C15	C9	C14	C13	-61.4(2)
C2	C3	C4	C5	58.9(3)	C15	C16	C17	C13	59.7(3)
C3	C4	C5	C6	-58.9(3)	C15	C16	C18	C11	-61.0(3)
C4	C5	C6	C1	50.4(3)	C17	C13	C14	C9	59.8(3)
C4	C5	C6	C7	176.4(2)	C17	C16	C18	C11	59.4(3)
C5	C6	C7	N1	-60.3(2)	C18	C11	C12	C13	58.8(3)
C5	C6	C7	C8	174.62(19)	C18	C16	C17	C13	-60.3(3)
C6	C1	C2	C3	46.6(3)	C19	N1	C7	C6	140.8(2)
C6	C7	C8	O2	128.1(2)	C19	N1	C7	C8	-92.2(2)
C6	C7	C8	O3	-53.8(2)	C19	C20	C21	C22	0.5(4)
C7	N1	C19	C20	-45.3(3)	C20	C19	C24	C23	1.0(4)
C7	N1	C19	C24	138.9(2)	C20	C21	C22	O4	-178.3(2)
C8	O3	C9	C10	-63.9(3)	C20	C21	C22	C23	1.8(4)
C8	O3	C9	C14	178.4(2)	C21	C22	C23	C24	-2.5(4)
C8	O3	C9	C15	59.9(3)	C22	C23	C24	C19	1.1(4)
C9	O3	C8	O2	-9.3(4)	C24	C19	C20	C21	-1.8(3)
C9	O3	C8	C7	172.71(18)	C25	O4	C22	C21	7.8(4)
C9	C10	C11	C12	60.4(2)	C25	O4	C22	C23	-172.2(2)
C9	C10	C11	C18	-60.3(2)					

Table S19. Hydrogen Atom Coordinates ($\text{\AA} \times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for cu_230519_CH_3ka_0m.

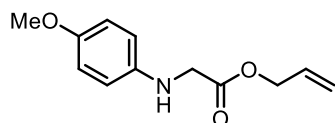
Atom	x	y	z	U(eq)
H2A	4708	2840.48	6653.34	43
H2B	5695.23	2217.11	6121.83	43

Table S19 continued. Hydrogen Atom Coordinates ($\text{\AA} \times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for cu_230519_CH_3ka_0m.

Atom	x	y	z	U(eq)
H3A	2584.32	1663.94	6484.7	43
H3B	2638.27	1805.63	5716.52	43
H4A	-311.54	2402.14	6141.7	45
H4B	918.4	3025.03	6617.9	45
H5A	-105.11	3837.82	5699.76	42
H5B	967.41	3150.87	5231.78	42
H6	2873.24	4329.79	6113.14	31
H7	1902.77	5027.83	5159.69	32
H10A	8748.83	5412.86	5641.73	33
H10B	8062.66	5126.7	6354.16	33
H11	11200.43	5837.14	6412.86	36
H12A	9062.99	6027.18	7320.93	42
H12B	10411.96	6883.59	7232.78	42
H13	7091.7	7325.15	7393.57	41
H14A	5563.09	6036.11	6951.71	35
H14B	4653.96	6898.23	6618.65	35
H15A	5701.06	7346.87	5506.15	33
H15B	7292.67	6774.01	5118.93	33
H16	8842.81	8057.72	5558.66	38
H17A	6769.35	8257	6472.85	44
H17B	9015.22	8243.24	6701.12	44
H18A	10811.41	6758.5	5485.19	40
H18B	11490.61	7338.19	6093.57	40
H20	-410.3	4920.56	4473.72	36
H21	-2089.05	5511.87	3591.44	36
H23	2524.33	4926.79	2435.66	39
H24	4208.01	4370.63	3321.8	37
H25A	-2917.09	6383.82	2049.66	63
H25B	-2354.72	6576.61	2787.65	63
H25C	-3563.17	5709.27	2604.47	63
H1	4320(40)	4090(30)	4400(18)	51

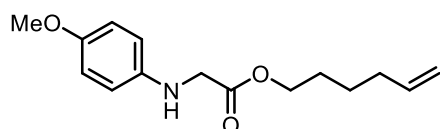
4. Characterization data

For substrates 1:



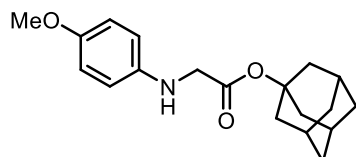
Allyl (4-methoxyphenyl)glycinate (**1e**)⁵

Yellow oil (375.7 mg, 85% yield). **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.79 (d, *J* = 8.9 Hz, 2H), 6.59 (d, *J* = 8.9 Hz, 2H), 6.01–5.83 (m, 1H), 5.35–5.24 (m, 2H), 4.67–4.66 (m, 2H), 3.90 (s, 2H), 3.74 (s, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 171.06, 152.61, 141.10, 131.58, 118.74, 114.83, 114.35, 65.68, 55.63, 46.69.



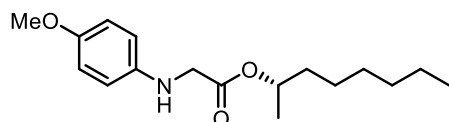
Hex-5-en-1-yl (4-methoxyphenyl)glycinate (**1f**)

Colorless oil (432.3 mg, 82% yield). **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.79 (d, *J* = 9.0 Hz, 2H), 6.58 (d, *J* = 8.9 Hz, 2H), 5.83–5.72 (m, 1H), 5.04–4.95 (m, 2H), 4.17 (t, *J* = 6.6 Hz, 2H), 3.86 (s, 2H), 3.74 (s, 3H), 2.10–2.04 (m, 2H), 1.70–1.62 (m, 2H), 1.44 (m, 2H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 171.44, 152.59, 141.23, 138.15, 114.86, 114.32, 65.06, 55.67, 46.75, 33.15, 27.94, 25.01. **HRMS (ESI)** *m/z*: calcd for C₁₅H₂₂NO₃ [M+H]⁺ 264.1594, found 264.1593.



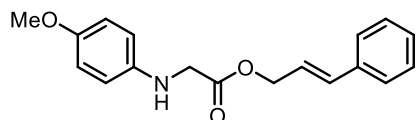
(1S,3S)-Adamantan-1-yl (4-methoxyphenyl)glycinate (**1h**)

White solid (460 mg, 73% yield). m.p. = 86–87 °C. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.78 (d, *J* = 8.9 Hz, 2H), 6.58 (d, *J* = 8.9 Hz, 2H), 3.75 (d, *J* = 5.5 Hz, 5H), 2.14 (m, 9H), 1.66 (d, *J* = 3.1 Hz, 6H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 170.15, 152.50, 141.38, 114.85, 114.39, 81.89, 55.72, 47.56, 41.30, 36.05, 30.81. **HRMS (ESI)** *m/z*: calcd for C₁₉H₂₆NO₃ [M+H]⁺ 316.1907, found 316.1906.



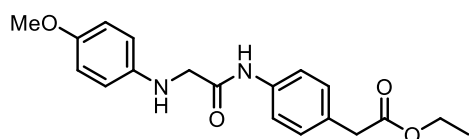
(S)-octan-2-yl (4-methoxyphenyl)glycinate (**1i**)

Colorless oil (439.5 mg, 75% yield). **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.78 (d, *J* = 8.9 Hz, 2H), 6.58 (d, *J* = 8.9 Hz, 2H), 4.99 (m, 1H), 3.83 (s, 2H), 3.73 (s, 3H), 1.64–1.44 (m, 2H), 1.31–1.21 (m, 11H), 0.88 (t, *J* = 6.8 Hz, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 171.02, 152.55, 141.32, 114.83, 114.31, 72.22, 55.66, 46.99, 35.83, 31.65, 29.02, 25.23, 22.51, 19.91, 14.00. **HRMS (ESI)** *m/z*: calcd for C₁₇H₂₈NO₃ [M+H]⁺ 294.2063, found 294.2051.



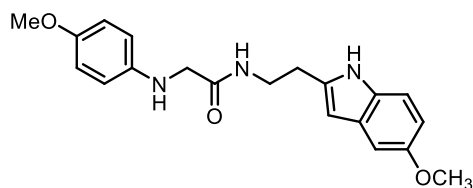
Cinnamyl (4-methoxyphenyl)glycinate (**1n**)⁶

Colorless oil (469.3 mg, 79% yield). **¹H NMR** (600 MHz, Chloroform-*d*) δ 7.39–7.35 (m, 2H), 7.34–7.30 (m, 2H), 7.28–7.24 (m, 1H), 6.81–6.73 (m, 2H), 6.69–6.56 (m, 3H), 6.26 (m, 1H), 4.82 (m, 2H), 3.92 (s, 2H), 3.73 (s, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 171.21, 152.72, 141.06, 135.96, 134.76, 128.59, 128.18, 126.61, 122.45, 114.89, 114.50, 65.69, 55.67, 46.88.



Ethyl 2-(4-(2-((4-methoxyphenyl)amino)acetamido)phenyl)acetate (**1r**)

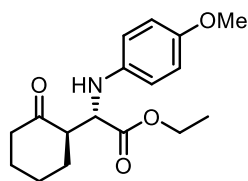
White Solid (266.7 mg, 78% yield). m.p. = 80-81 °C. **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.72 (s, 1H), 7.49 (d, *J* = 8.5 Hz, 2H), 7.22 (d, *J* = 8.5 Hz, 2H), 6.80 (d, *J* = 8.9 Hz, 2H), 6.63 (d, *J* = 8.9 Hz, 2H), 4.14 (t, *J* = 7.1 Hz, 2H), 3.82 (s, 2H), 3.74 (s, 3H), 3.56 (s, 2H), 1.23 (t, *J* = 7.2 Hz, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 171.56, 169.13, 153.48, 140.88, 136.28, 130.16, 129.78, 119.87, 114.98, 114.74, 60.84, 55.65, 50.59, 40.77, 14.11. **HRMS (ESI)** *m/z*: calcd for C₁₉H₂₃N₂O₄ [M+H]⁺ 343.1652, found 343.1645.



N-(2-(5-methoxy-1H-indol-2-yl)ethyl)-2-((4-methoxyphenyl)amino)acetamide (**1t**)

White Solid (254.2 mg, 72% yield). m.p. = 116-117 °C. **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.09 (s, 1H), 7.21 (d, *J* = 8.8 Hz, 1H), 7.00 (d, *J* = 2.5 Hz, 1H), 6.93 (t, *J* = 6.0 Hz, 1H), 6.84 (m, 1H), 6.75 (d, *J* = 8.9 Hz, 2H), 6.71 (d, *J* = 2.4 Hz, 1H), 6.47 (d, *J* = 8.9 Hz, 2H), 3.84 (s, 3H), 3.74 (s, 3H), 3.66 (s, 2H), 3.59 (m, 2H), 2.93–2.84 (m, 2H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 170.81, 153.80, 152.85, 141.13, 131.46, 127.50, 122.98, 114.79, 114.17, 112.05, 111.93, 100.35, 55.85, 55.66, 49.40, 39.00, 25.21. **HRMS (ESI)** *m/z*: calcd for C₂₀H₂₄N₃O₃ [M+H]⁺ 354.1812, found 354.1804.

For products:

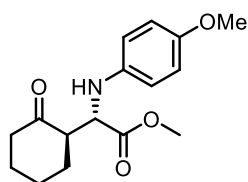
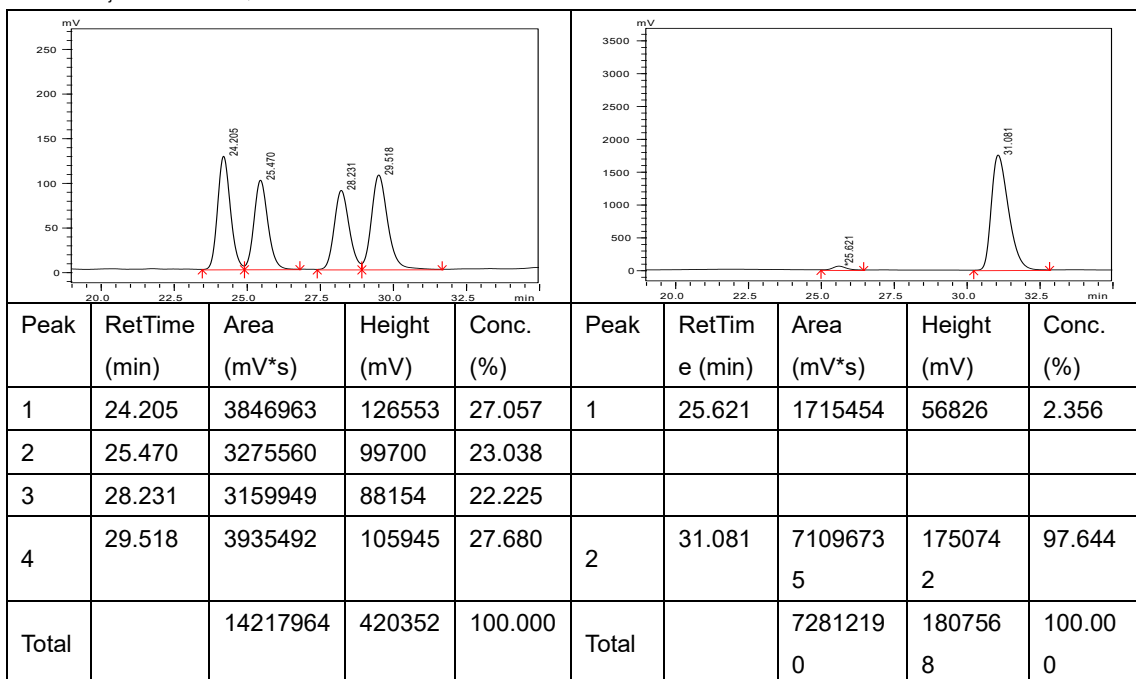


Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3aa**)⁷

Yellow oil (49 mg, 80% yield); > 99:1 *dr*. **¹H NMR** (500 MHz, Chloroform-*d*) δ 6.76 (d, *J* = 8.9 Hz, 2H), 6.63 (d, *J* = 8.9 Hz, 2H), 4.18–4.10 (m, 2H), 3.98 (d, *J* = 4.1 Hz, 1H), 3.73 (s, 3H), 3.14–3.05 (m, 1H), 2.50–2.37 (m, 1H), 2.36–2.27 (m, 1H), 2.17–2.08 (m, 1H), 2.06–2.01 (m, 1H), 1.97–1.87 (m, 2H), 1.77–1.62 (m, 2H), 1.21 (t, *J* = 7.1 Hz, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 211.03, 173.08, 152.74,

142.13, 115.60, 114.74, 61.19, 59.07, 55.68, 53.57, 41.80, 30.52, 26.83, 24.53, 14.09.

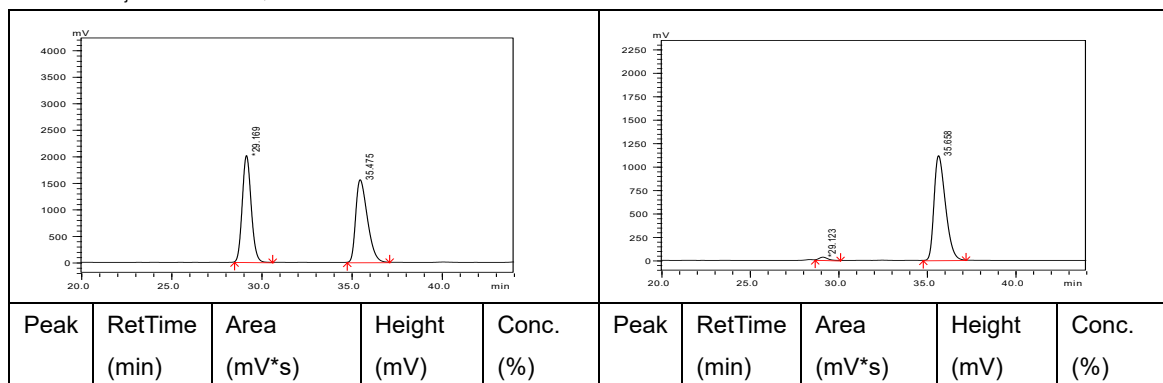
Optical Rotation: $[\alpha]^{25}_D = 29$ ($c = 1.0$, CH_2Cl_2). The ee value was determined by **HPLC analysis:** 97.6:2.4 er, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, $\lambda = 214$ nm, retention time: $t_{\text{major}} = 31.1$ min, $t_{\text{minor}} = 25.6$ min.



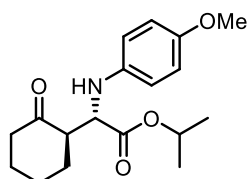
Methyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ba**)⁷

Pale yellow oil (46 mg, 79% yield); > 99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.76 (d, $J = 8.8$ Hz, 2H), 6.62 (d, $J = 8.8$ Hz, 2H), 3.99 (d, $J = 3.9$ Hz, 1H), 3.73 (s, 3H), 3.68 (s, 3H), 3.16–3.08 (m, 1H), 2.45–2.39 (m, 1H), 2.37–2.28 (m, 1H), 2.11–2.02 (m, 2H), 1.97–1.83 (m, 2H), 1.78–1.61 (m, 2H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 211.10, 173.64, 152.74, 142.02, 115.44, 114.77, 58.87, 55.67, 53.65, 52.27, 41.83, 30.56, 26.84, 24.57.

Optical Rotation: $[\alpha]^{25}_D = 24.3$ ($c = 1.0$, CH_2Cl_2). The ee value was determined by **HPLC analysis:** 98.2:1.8 er, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, $\lambda = 214$ nm, retention time: $t_{\text{major}} = 35.7$ min, $t_{\text{minor}} = 29.1$ min.



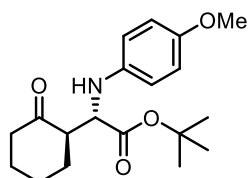
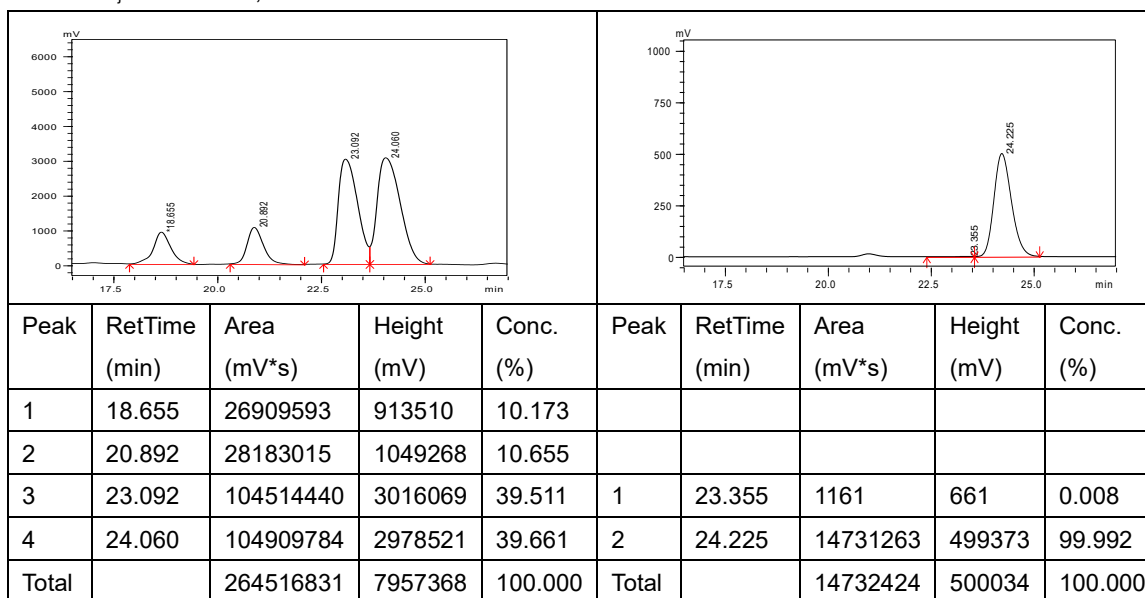
1	29.169	68858297	2006741	49.731	1	29.123	907934	30034	1.795
2	35.475	69603611	1554593	50.269	2	35.658	49673100	1114778	98.205
Total		138461909	3561334	100.000	Total		50581034	1144812	100.000



Isopropyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ca**)⁷

Yellow oil (48.5 mg, 76% yield); > 99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.75 (d, *J* = 8.9 Hz, 2H), 6.63 (d, *J* = 8.9 Hz, 2H), 5.06–4.92 (m, 1H), 3.96 (d, *J* = 4.2 Hz, 1H), 3.73 (s, 3H), 3.10–3.04 (m, 1H), 2.46–2.26 (m, 2H), 2.14–2.00 (m, 2H), 1.98–1.84 (m, 2H), 1.78–1.62 (m, 2H), 1.22 (d, *J* = 6.2 Hz, 3H), 1.13 (d, *J* = 6.2 Hz, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 210.86, 172.49, 152.72, 142.20, 115.68, 114.68, 68.74, 59.18, 55.67, 53.52, 41.73, 30.45, 26.83, 24.47, 21.62.

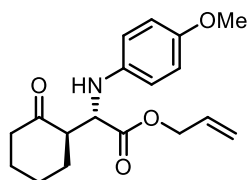
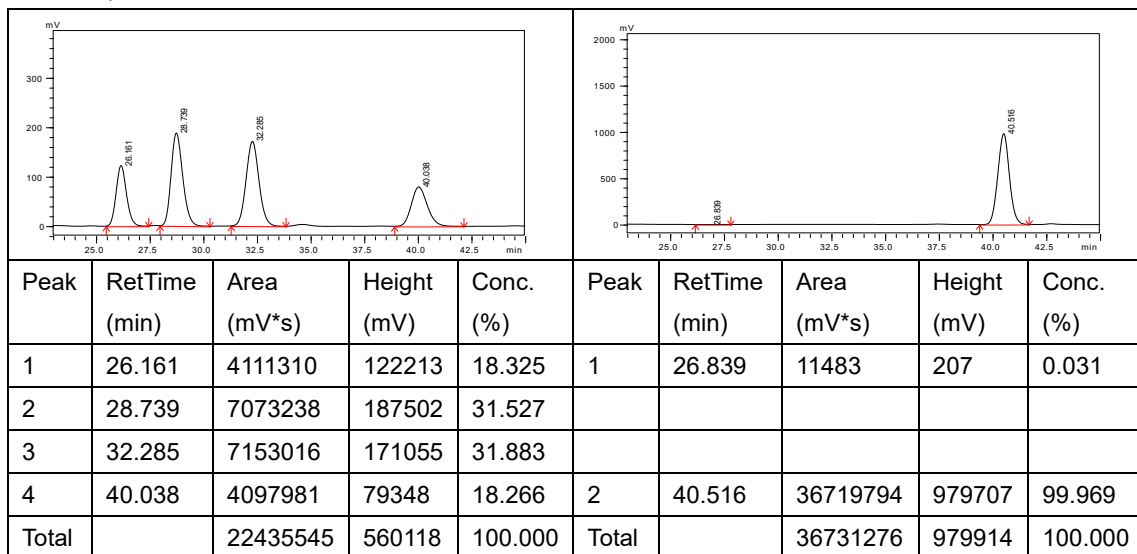
Optical Rotation: $[\alpha]_D^{25} = 18.9$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 99.9:0.1 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, λ = 254 nm, retention time: *t*_{major} = 24.2 min, *t*_{minor} = 21.0 min.



Tert-butyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3da**)⁷

White solid (52 mg, 79% yield); > 99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.75 (d, *J* = 8.8 Hz, 2H), 6.61 (d, *J* = 8.8 Hz, 2H), 3.92 (d, *J* = 4.3 Hz, 1H), 3.73 (s, 3H), 3.07–2.97 (m, 1H), 2.51–2.38 (m, 1H), 2.36–2.24 (m, 1H), 2.15–1.97 (m, 2H), 1.96–1.83 (m, 2H), 1.78–1.63 (m, 2H), 1.39 (s, 9H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 210.84, 172.00, 152.59, 142.32, 115.46, 114.67, 81.54, 59.47, 55.67, 53.45, 41.68, 30.32, 27.88, 26.78, 24.41.

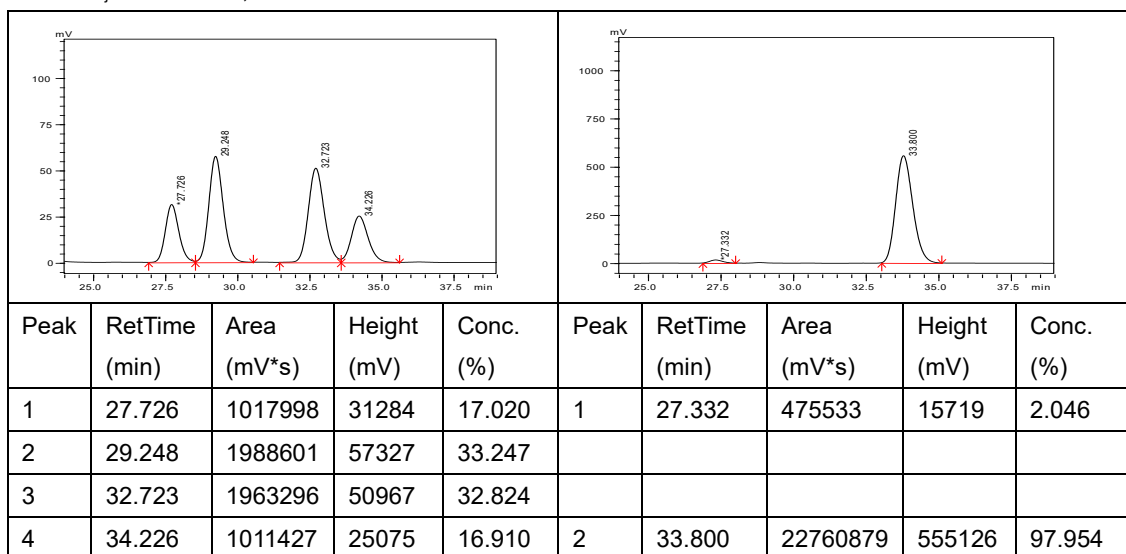
Optical Rotation: $[\alpha]_{25}^D = 26$ ($c = 1.0$, CH_2Cl_2). The ee value was determined by **HPLC analysis:** 99.9:0.1 er , Chiralcel AD-H column, hexane/ i -PrOH = 90/10, flow rate = 0.5 mL/min, $\lambda = 214$ nm, retention time: $t_{\text{major}} = 26.8$ min, $t_{\text{minor}} = 40.5$ min.



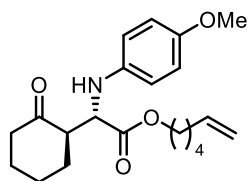
Allyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ea**)⁷

Yellow oil (49.5 mg, 78% yield); > 99:1 dr . **¹H NMR** (400 MHz, Chloroform- d) δ 6.76 (d, $J = 8.4$ Hz, 2H), 6.63 (d, $J = 8.4$ Hz, 2H), 5.95–5.75 (m, 1H), 5.31–5.14 (m, 2H), 4.58 (d, $J = 5.5$ Hz, 2H), 4.01 (d, $J = 3.7$ Hz, 1H), 3.73 (s, 3H), 3.17–3.10 (m, 1H), 2.47–2.40 (m, 1H), 2.37–2.27 (m, 1H), 2.15–2.02 (m, 2H), 1.98–1.87 (m, 2H), 1.80–1.59 (m, 2H). **¹³C NMR** (100 MHz, Chloroform- d) δ 211.02, 172.79, 152.78, 142.06, 131.77, 118.27, 115.55, 114.77, 65.75, 59.04, 55.69, 53.59, 41.81, 30.55, 26.83, 24.56.

Optical Rotation: $[\alpha]_{25}^D = 22.4$ ($c = 1.0$, CH_2Cl_2). The ee value was determined by **HPLC analysis:** 98.0:2.0 er , Chiralcel AD-H column, hexane/ i -PrOH = 90/10, flow rate = 0.7 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 33.8$ min, $t_{\text{minor}} = 27.3$ min.



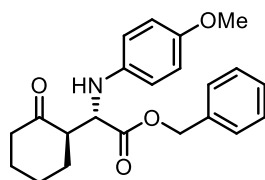
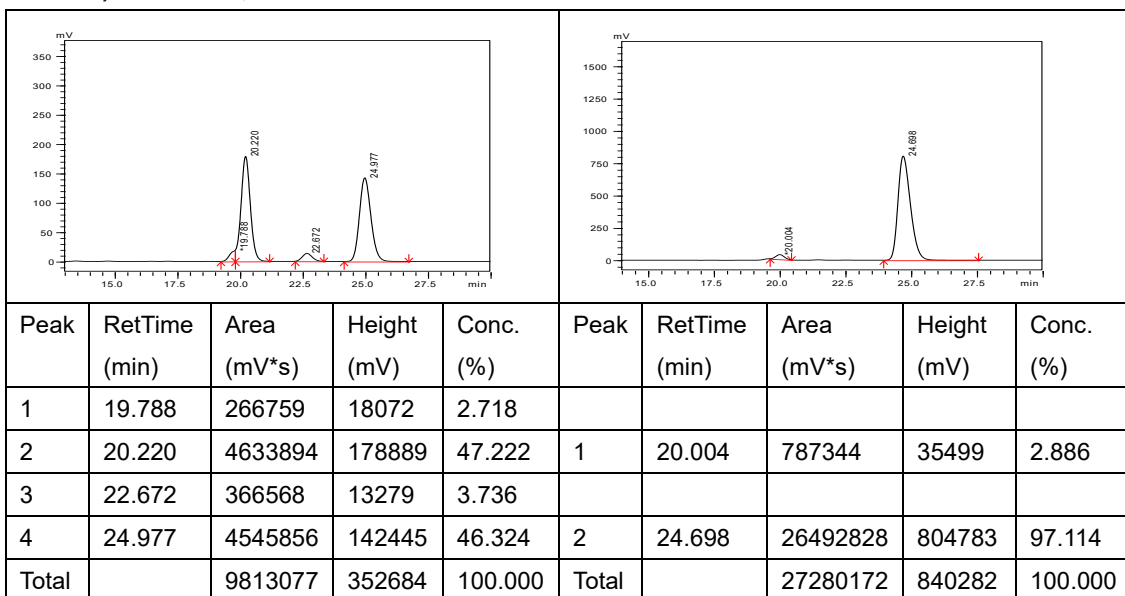
Total		5981322	164653	100.000	Total		23236412	570846	100.000
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Allyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3fa**)⁵

Yellow oil (53.2 mg, 74% yield); > 99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.77 (d, *J* = 9.1 Hz, 2H), 6.73 (d, *J* = 9.1 Hz, 2H), 5.81–5.70 (m, 1H), 5.03–4.92 (m, 2H), 4.22 (d, *J* = 5.2 Hz, 1H), 4.15–4.04 (m, 2H), 3.74 (s, 3H), 2.90–2.76 (m, 1H), 2.51–2.41 (m, 1H), 2.37–2.27 (m, 1H), 2.26–2.16 (m, 1H), 2.12–1.93 (m, 4H), 1.86–1.76 (m, 1H), 1.74–1.64 (m, 2H), 1.64–1.53 (m, 2H), 1.42–1.32 (m, 2H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 210.05, 173.48, 153.11, 140.93, 138.28, 116.12, 114.79, 114.74, 65.04, 58.18, 55.66, 53.54, 41.83, 33.16, 29.66, 27.91, 26.85, 25.07, 24.78.

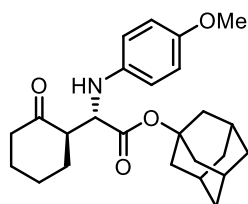
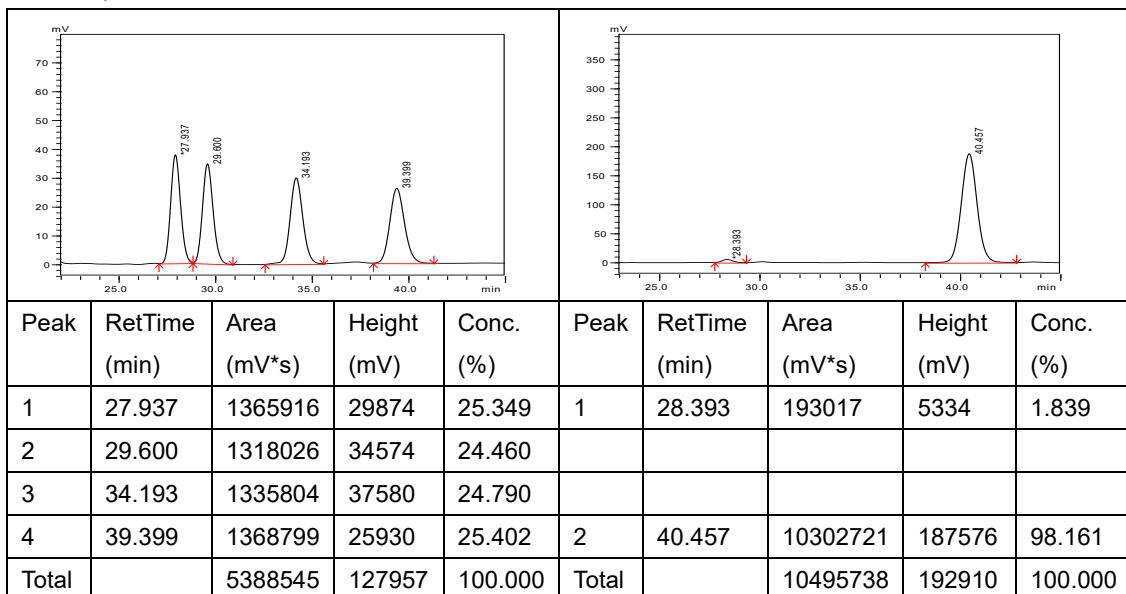
Optical Rotation: $[\alpha]_D^{25} = 16.4$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 97.1:2.9 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, λ = 254 nm, retention time: *t*_{major} = 24.7 min, *t*_{minor} = 20.0 min.



Benzyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ga**)⁷

Yellow oil (58 mg, 79% yield); > 99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.40–7.29 (m, 3H), 7.25–7.20 (m, 2H), 6.75 (d, *J* = 8.9 Hz, 2H), 6.62 (d, *J* = 8.9 Hz, 2H), 5.20–5.08 (m, 2H), 4.06 (d, *J* = 4.0 Hz, 1H), 3.74 (s, 3H), 3.16–3.07 (m, 1H), 2.46–2.37 (m, 1H), 2.35–2.25 (m, 1H), 2.12–1.99 (m, 2H), 1.95–1.85 (m, 2H), 1.74–1.60 (m, 2H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 211.02, 172.96, 152.83, 142.03, 135.58, 128.44, 128.14, 128.01, 115.67, 114.78, 66.88, 59.05, 55.71, 53.52, 41.78, 30.51, 26.82, 24.51.

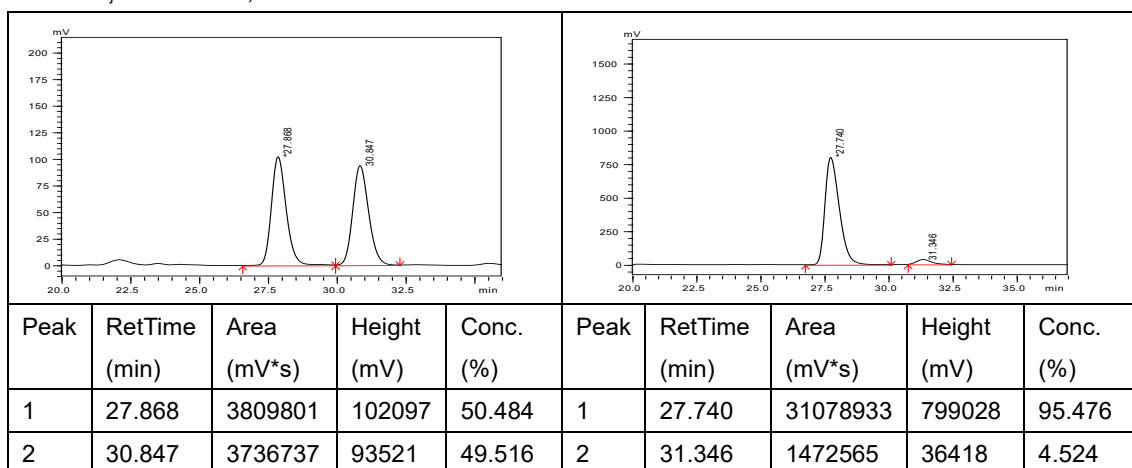
Optical Rotation: $[\alpha]_{25}^D = 25.6$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 98.2:1.8 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 214$ nm, retention time: $t_{\text{major}} = 40.5$ min, $t_{\text{minor}} = 28.3$ min.



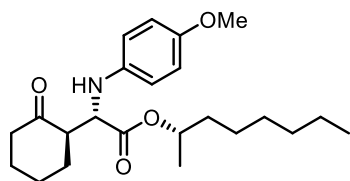
(1S,3R)-Adamantan-1-yl (2S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ha**)⁸

White solid (60.9 mg, 74% yield); > 99:1 *dr*. m.p. = 122–123 °C. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.75 (d, $J = 8.8$ Hz, 2H), 6.62 (d, $J = 8.8$ Hz, 2H), 3.93 (d, $J = 4.4$ Hz, 1H), 3.73 (s, 3H), 3.07–2.96 (m, 1H), 2.47–2.39 (m, 1H), 2.35–2.24 (m, 1H), 2.14–1.99 (m, 11H), 1.96–1.85 (m, 2H), 1.74–1.59 (m, 8H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 210.83, 171.65, 152.63, 142.24, 115.57, 114.67, 81.61, 59.52, 55.67, 53.43, 41.69, 41.15, 36.04, 30.73, 30.26, 26.76, 24.40.

Optical Rotation: $[\alpha]_{25}^D = 8.6$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 95.5:4.5 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 27.7$ min, $t_{\text{minor}} = 31.3$ min.



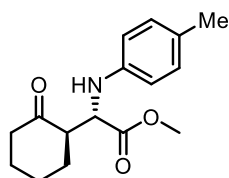
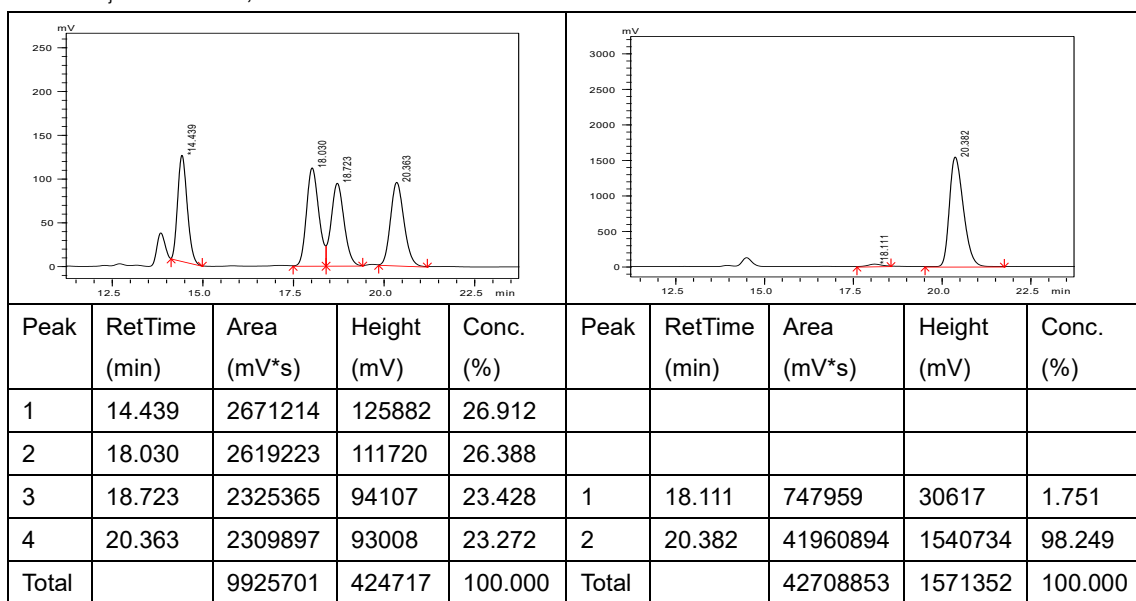
Total		7546538	195617	100.000	Total		32551498	835446	100.000
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(S)-Octan-2-yl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ia**)

Yellow oil (60 mg, 77% yield); 98:2 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.74 (d, *J* = 8.8 Hz, 2H), 6.62 (d, *J* = 8.8 Hz, 2H), 4.95–4.78 (m, 1H), 3.94 (d, *J* = 4.3 Hz, 1H), 3.72 (s, 3H), 3.18–3.00 (m, 1H), 2.49–2.37 (m, 1H), 2.35–2.26 (m, 1H), 2.14–2.00 (m, 2H), 1.99–1.88 (m, 2H), 1.79–1.65 (m, 2H), 1.51–1.35 (m, 2H), 1.27–1.06(m, 11H), 0.85 (t, *J* = 7.1 Hz, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 210.80, 172.72, 152.79, 142.32, 115.63, 114.71, 72.19, 59.32, 55.65, 53.57, 41.75, 35.76, 31.66, 30.61, 29.00, 26.91, 25.05, 24.51, 22.49, 19.80, 13.98. **HRMS (ESI)** *m/z*: calcd for C₂₃H₃₆NO₄ [M+H]⁺ 390.2644, found 390.2645.

Optical Rotation: $[\alpha]^{25}_D = 17.5$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 98.2:1.8 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, λ = 254 nm, retention time: *t*_{major} = 20.4 min, *t*_{minor} = 18.1 min.

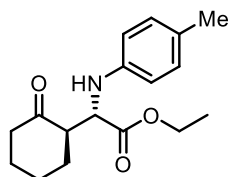
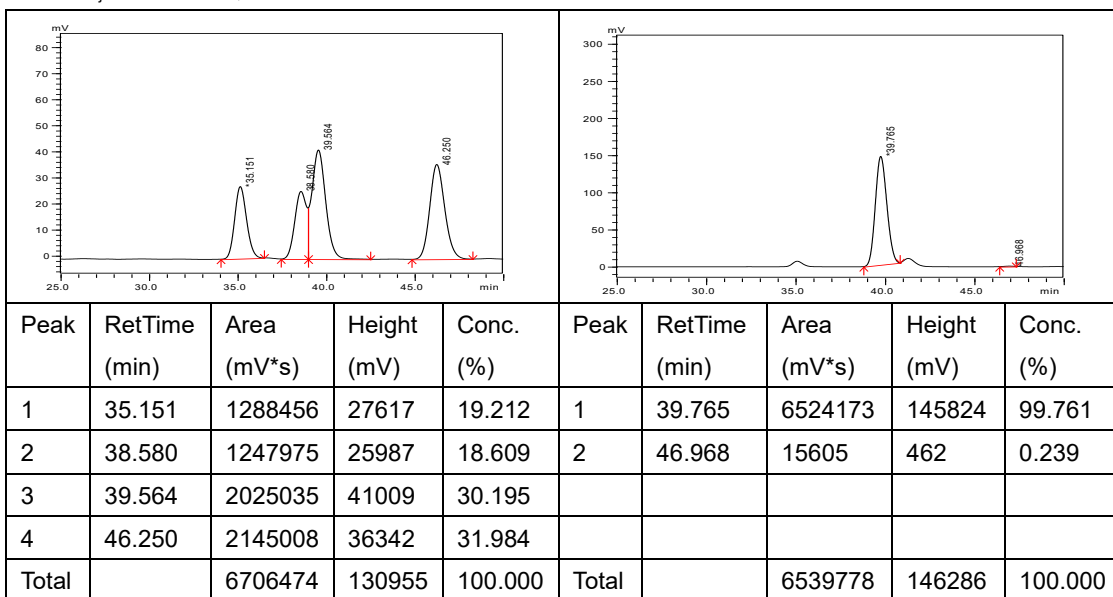


Methyl (S)-2-((R)-2-oxocyclohexyl)-2-(p-tolylamino)acetate (**3ja**)

Yellow oil (38.2 mg, 71% yield); > 99:1 *dr*. **¹H NMR** (600 MHz, Chloroform-*d*) δ 6.98 (d, *J* = 8.4 Hz, 2H), 6.57 (d, *J* = 8.4 Hz, 2H), 4.09 (d, *J* = 3.8 Hz, 1H), 3.69 (s, 3H), 3.19–3.12 (m, 1H), 2.47–2.38 (m, 1H), 2.36–2.31 (m, 1H), 2.23 (s, 3H), 2.14–2.08 (m, 1H), 2.06–2.03 (m, 1H), 1.95–1.85 (m, 2H), 1.76–1.63 (m, 2H). **¹³C NMR** (150 MHz, Chloroform-*d*) δ 211.12, 173.48, 145.50, 129.79, 127.73, 113.95, 57.89, 53.67, 52.32, 41.84, 30.50, 26.84, 24.59, 20.36. **HRMS (ESI)** *m/z*: calcd for C₁₆H₂₂NO₃ [M+H]⁺ 276.1594, found

276.1585.

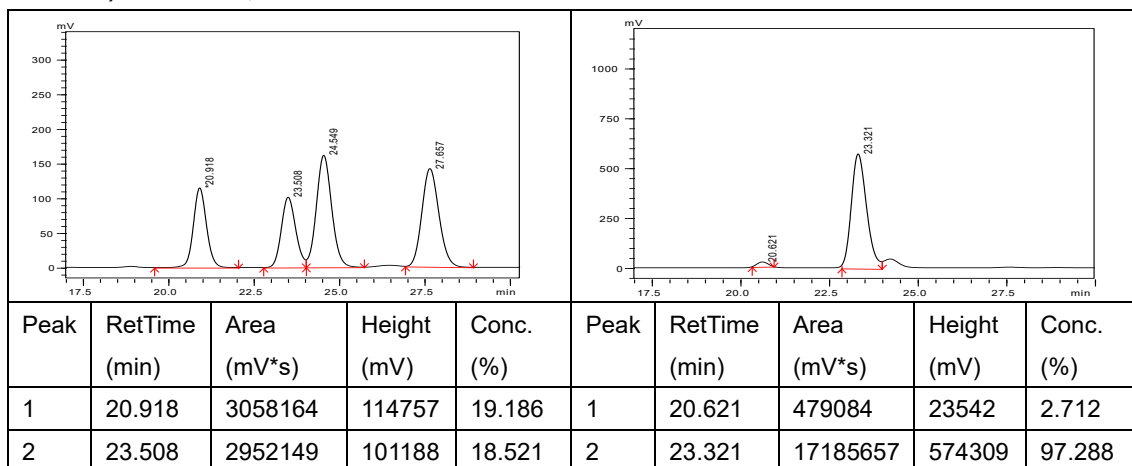
Optical Rotation: $[\alpha]_D^{25} = 14.7$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 99.7:0.3 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 98/2, flow rate = 1.0 mL/min, $\lambda = 240$ nm, retention time: $t_{\text{major}} = 39.8$ min, $t_{\text{minor}} = 47.0$ min.



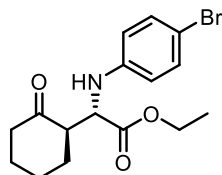
Ethyl (S)-2-((R)-2-oxocyclohexyl)-2-(p-tolylamino)acetate (**3ka**)⁷

Yellow oil (39 mg, 66% yield); 98:2 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.97 (d, $J = 7.8$ Hz, 2H), 6.57 (d, $J = 7.8$ Hz, 2H), 4.23–4.05 (m, 3H), 3.21–3.05 (m, 1H), 2.48–2.37 (m, 1H), 2.32 (m, 1H), 2.22 (s, 3H), 2.16–2.00 (m, 2H), 1.97–1.81 (m, 2H), 1.80–1.61 (m, 2H), 1.25–1.17 (m, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 211.03, 172.90, 145.66, 129.71, 127.56, 113.97, 61.21, 57.95, 53.58, 41.78, 30.41, 26.79, 24.51, 20.34, 14.09.

Optical Rotation: $[\alpha]_D^{25} = 18.0$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 97.3:2.7 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 96/4, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 20.6$ min, $t_{\text{minor}} = 23.3$ min.



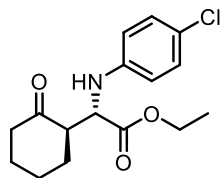
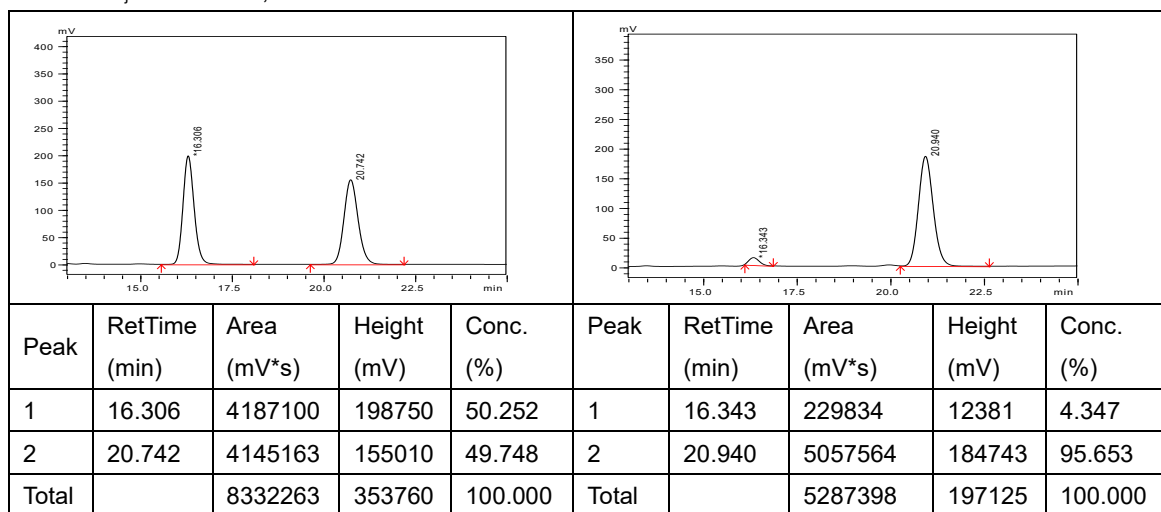
3	24.549	5036013	161456	31.594					
4	27.657	4893371	141462	30.699					
Total		15939697	518863	100.000	Total		17664742	597851	100.000



Ethyl (S)-2-((4-bromophenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**31a**)⁷

Yellow oil (41.1 mg, 58% yield); 80:20 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.24 (d, *J* = 8.8 Hz, 2H), 6.60 (d, *J* = 8.8 Hz, 2H), 4.23 (d, *J* = 5.0 Hz, 1H), 4.13 (m, 2H), 3.19–2.77 (m, 1H), 2.52–2.36 (m, 1H), 2.36–2.04 (m, 3H), 1.95 (m, 1H), 1.89–1.54 (m, 3H), 1.26–1.22 (m, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 209.90, 172.72, 145.97, 131.92, 115.63, 110.22, 61.33, 56.56, 53.35, 41.77, 29.77, 26.80, 24.77, 14.08.

Optical Rotation: $[\alpha]_{25}^D = 30.0$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 95.6:4.4 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 20.9 min, *t*_{minor} = 16.3 min.

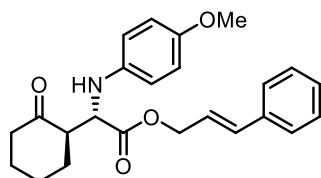
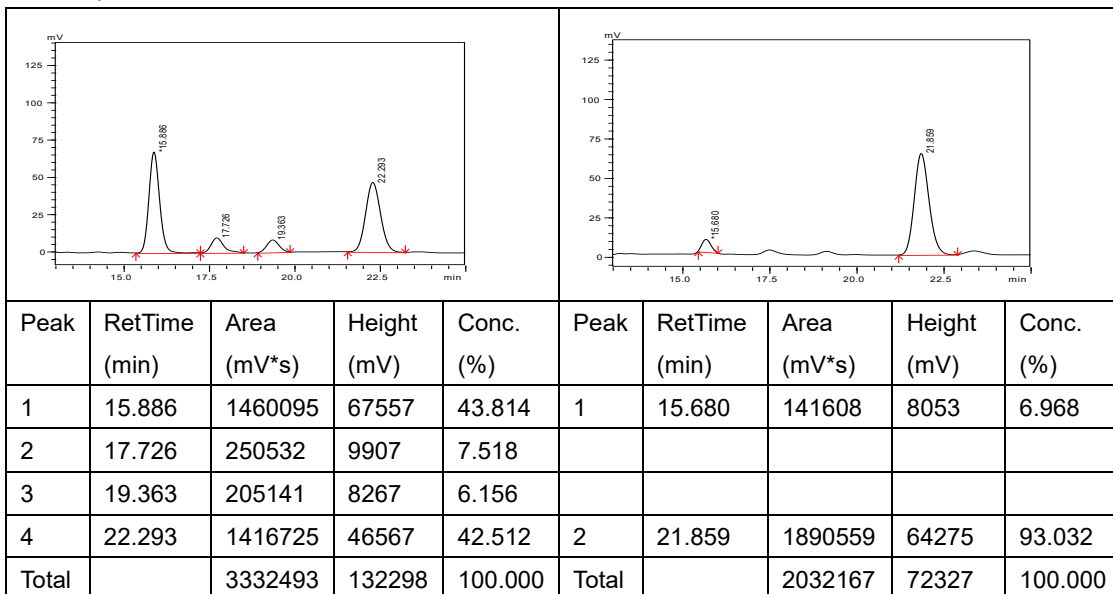


Ethyl (S)-2-((4-chlorophenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ma**)⁷

Yellow oil (37.2 mg, 60% yield); 99:1 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.12 (d, *J* = 8.8 Hz, 2H), 6.66 (d, *J* = 8.8 Hz, 2H), 4.24 (d, *J* = 4.9 Hz, 1H), 4.19–4.11 (m, 2H), 2.91–2.74 (m, 1H), 2.53–2.41 (m, 1H), 2.36–2.26 (m, 1H), 2.23–2.15 (m, 1H), 2.12–2.04 (m, 1H), 2.00–1.91 (m, 1H), 1.86–1.76 (m, 1H), 1.73–1.61 (m, 2H), 1.23 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 209.92, 172.85, 145.60, 129.12, 123.27, 115.30, 61.39, 56.80, 53.45, 41.85, 29.80, 26.85, 24.84, 14.15.

Optical Rotation: $[\alpha]_{25}^D = 42.5$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 93.0:7.0 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention

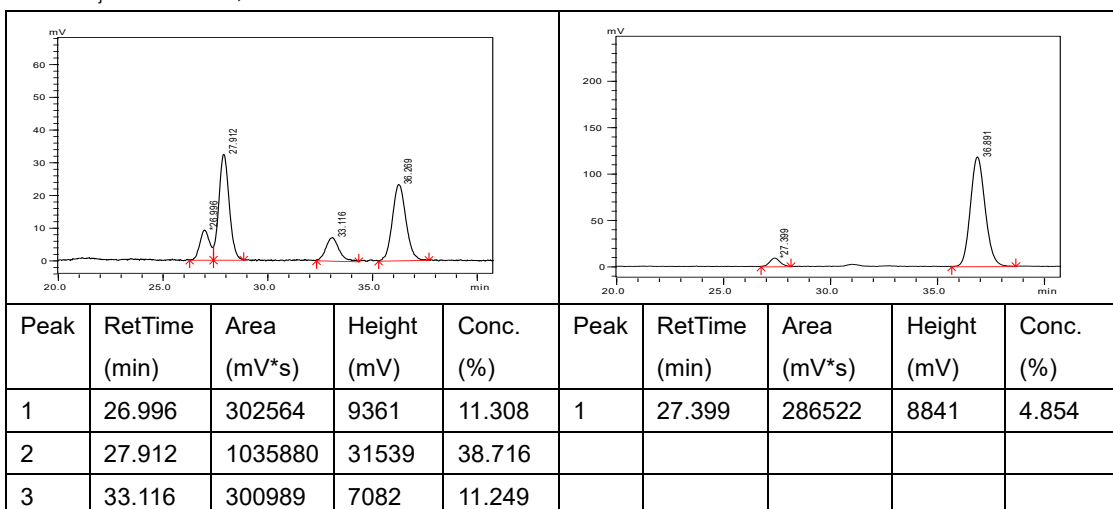
time: $t_{\text{major}} = 21.9 \text{ min}$, $t_{\text{minor}} = 15.7 \text{ min}$.



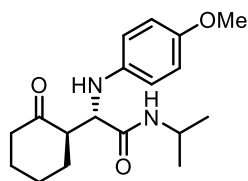
Cinnamyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3na**)

Brown oil (52 mg, 69% yield); 99:1 *dr*. $^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.38–7.31 (m, 4H), 7.30–7.26 (m, 1H), 6.79 (d, $J = 8.9 \text{ Hz}$, 2H), 6.68 (d, $J = 8.9 \text{ Hz}$, 2H), 6.56 (d, $J = 15.9 \text{ Hz}$, 1H), 6.28–6.16 (m, 1H), 4.77 (m, 2H), 4.06 (d, $J = 4.1 \text{ Hz}$, 1H), 3.75 (s, 3H), 3.23–3.13 (m, 1H), 2.51–2.42 (m, 1H), 2.40–2.32 (m, 1H), 2.20–2.04 (m, 2H), 2.03–1.91 (m, 2H), 1.82–1.65 (m, 2H). $^{13}\text{C NMR}$ (100 MHz, Chloroform-*d*) δ 211.07, 172.91, 152.78, 142.06, 133.88, 128.53, 127.98, 126.56, 122.75, 115.59, 114.78, 65.58, 59.07, 55.65, 53.58, 41.83, 30.59, 26.85, 24.54. **HRMS (ESI)** m/z : calcd for $\text{C}_{24}\text{H}_{28}\text{NO}_4$ $[\text{M}+\text{H}]^+$ 394.2013, found 394.2005.

Optical Rotation: $[\alpha]_{\text{D}}^{25} = 22.3$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis**: 95.1:4.9 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 254 \text{ nm}$, retention time: $t_{\text{major}} = 36.9 \text{ min}$, $t_{\text{minor}} = 27.4 \text{ min}$.



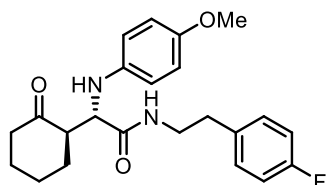
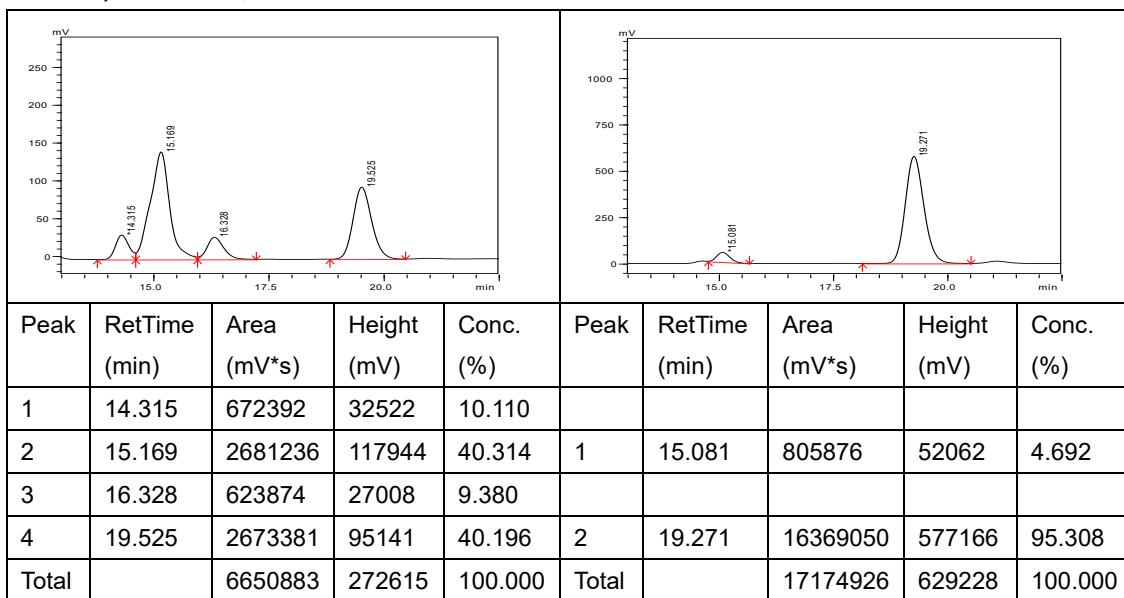
4	36.269	1036173	23235	38.727	2	36.891	5616542	117881	95.146
Total		2675606	71218	100.000	Total		5903064	126722	100.000



(S)-N-isopropyl-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamide (**30a**)

White solid (44.9 mg, 70% yield); 95:5 *dr*. m.p. = 118–120 °C. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.84 (dd, *J* = 12.7, 8.5 Hz, 1H), 6.76 (d, *J* = 8.9 Hz, 2H), 6.55 (d, *J* = 8.9 Hz, 2H), 4.09–3.97 (m, 1H), 3.92 (d, *J* = 3.0 Hz, 1H), 3.73 (s, 3H), 3.36–3.20 (m, 1H), 2.38 (m, 2H), 2.10–2.01 (m, 2H), 1.90–1.82 (m, 1H), 1.71–1.50 (m, 3H), 1.13 (d, *J* = 6.6 Hz, 3H), 1.08 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 213.61, 170.97, 152.53, 141.14, 114.98, 114.46, 58.92, 55.68, 53.22, 42.34, 41.22, 31.08, 27.62, 24.91, 22.57, 22.55. HRMS (ESI) *m/z*: calcd for C₁₈H₂₆N₂NaO₃ [M+Na]⁺ 341.1836, found 314.1835.

Optical Rotation: [α]_D²⁵ = 36.5 (c = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 95.3:4.7 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 19.3 min, *t*_{minor} = 15.1 min.

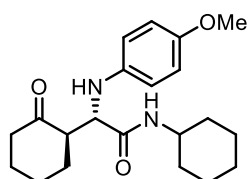
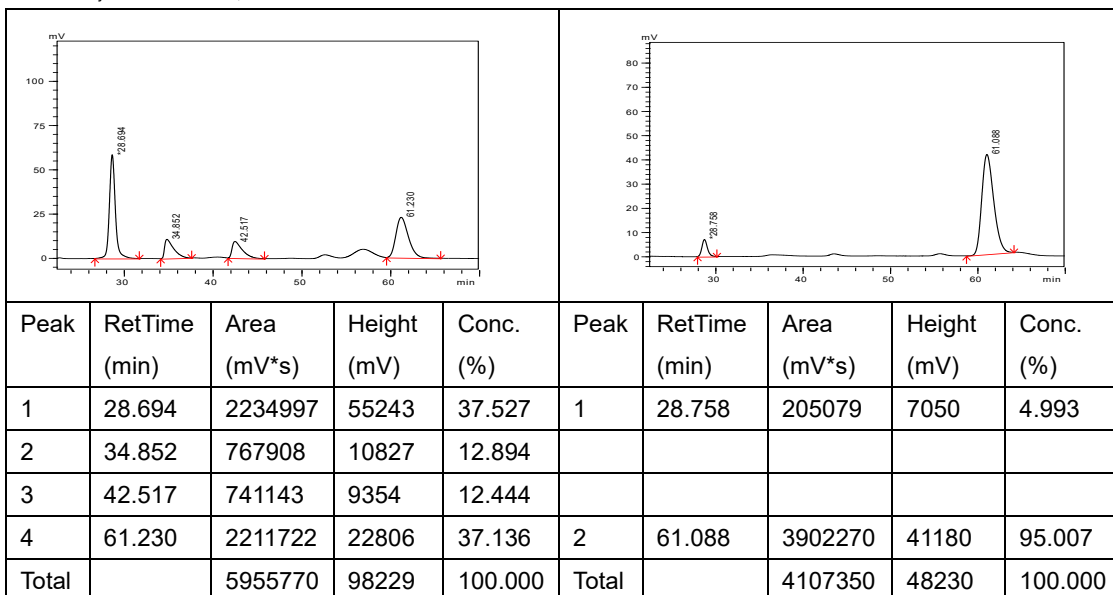


(S)-N-(4-fluorophenethyl)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamide (**3pa**)

White solid (55 mg, 69% yield); >99:1 *dr*. m.p. = 146–147 °C. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.09 (dd, *J* = 8.5, 5.5 Hz, 2H), 7.07–6.97 (m, 1H), 6.93 (t, *J* = 8.7 Hz, 2H), 6.75 (d, *J* = 8.9 Hz, 2H), 6.50 (d, *J* = 8.9 Hz, 2H), 3.87 (d, *J* = 2.9 Hz, 1H), 3.74 (s, 3H), 3.47 (q, *J* = 6.7 Hz, 2H), 3.38–3.26 (m, 1H), 2.74 (t, *J* = 6.9 Hz, 2H), 2.45–2.31 (m, 2H), 2.11–1.97 (m, 2H), 1.90–1.81 (m, 1H), 1.72–1.52 (m, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 213.42, 172.10, 161.52 (d, *J*₁ = 243 Hz, C_q), 152.70, 140.92, 134.47 (d, *J*₄ = 3 Hz,

Cq), 134.44, 130.21 (d, $J_3 = 7$ Hz, CH), 130.13, 115.22 (d, $J_2 = 21$ Hz, CH), 115.03, 114.40, 59.28, 55.69, 53.13, 42.34, 40.51, 34.87, 31.28, 27.59, 24.90. ^{19}F NMR (376 MHz, Chloroform-*d*) δ -116.85. HRMS (ESI) m/z : calcd for $\text{C}_{23}\text{H}_{27}\text{FN}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 421.1903, found 421.1912.

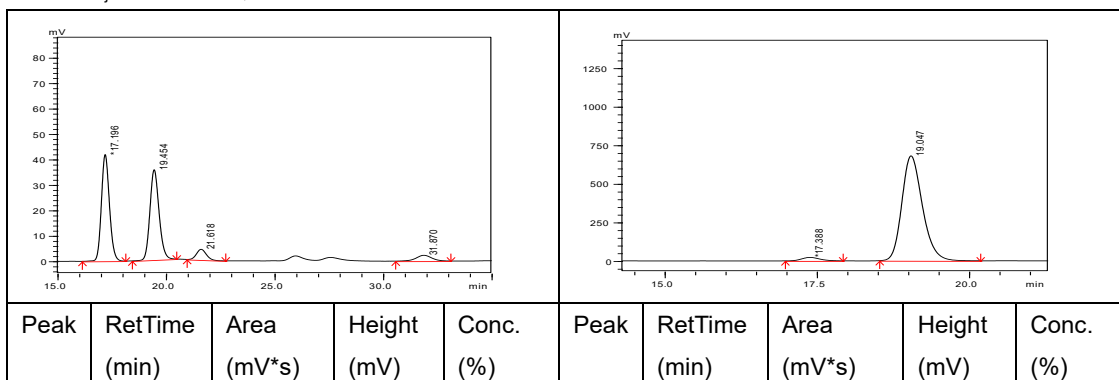
Optical Rotation: $[\alpha]^{25}_{\text{D}} = 42$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by HPLC analysis: 95.0:5.0 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 61.1$ min, $t_{\text{minor}} = 28.8$ min.



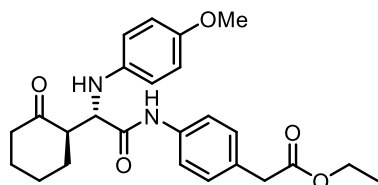
(*S*)-*N*-cyclohexyl-2-((4-methoxyphenyl)amino)-2-((*R*)-2-oxocyclohexyl)acetamide (**3qa**)

Yellow oil (53.8 mg, 75% yield); 99:1 *dr*. ^1H NMR (400 MHz, Chloroform-*d*) δ 6.89 (s, 1H), 6.77 (d, $J = 8.3$ Hz, 2H), 6.58 (d, $J = 8.3$ Hz, 2H), 3.97 (s, 1H), 3.74 (s, 4H), 3.30 (m, 1H), 2.39 (m, 2H), 2.13–2.04 (m, 2H), 1.89–1.55 (m, 9H), 1.39–1.27 (m, 2H), 1.24–1.07 (m, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 213.68, 170.71, 152.67, 141.01, 115.04, 114.62, 58.92, 55.74, 53.25, 47.98, 42.37, 32.83, 30.98, 27.66, 26.51, 25.49, 24.94, 24.65. HRMS (ESI) m/z : calcd for $\text{C}_{21}\text{H}_{30}\text{N}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$ 381.2149, found 381.2142.

Optical Rotation: $[\alpha]^{25}_{\text{D}} = 22.9$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by HPLC analysis: 97.2:2.8 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 19.0$ min, $t_{\text{minor}} = 17.4$ min.



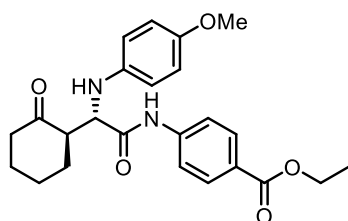
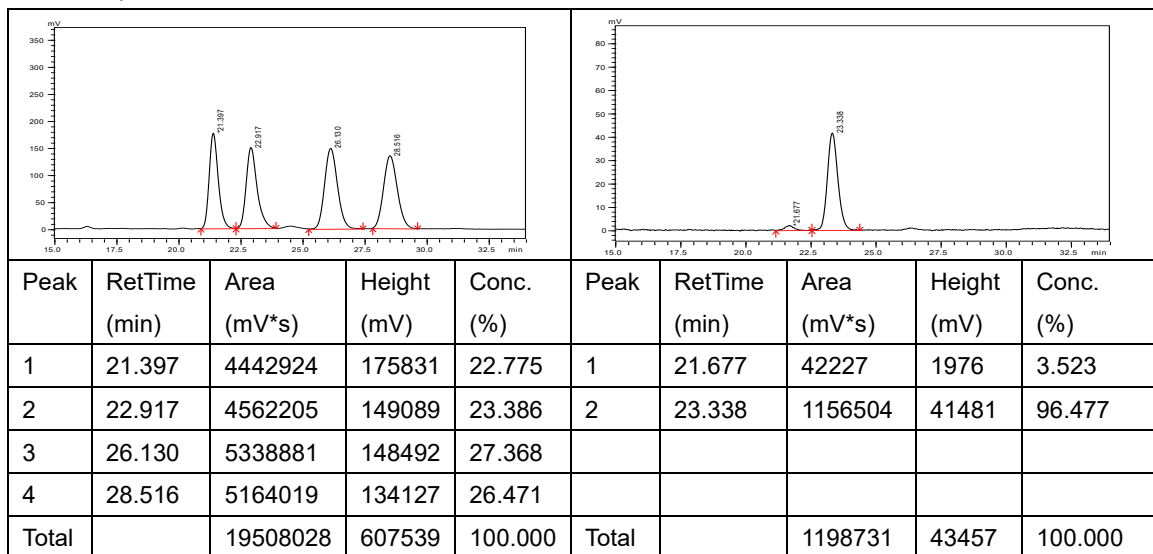
1	17.196	1078515	41896	45.667	1	17.388	460063	22238	2.802
2	19.454	1039284	35459	44.006	2	19.047	15956562	679586	97.198
3	21.618	130158	4175	5.511					
4	31.870	113733	2270	4.816					
Total		2361690	83800	100.000	Total		16416625	701824	100.000



Ethyl 2-(4-((S)-2-((R)-2-oxocyclohexyl)acetamido)phenyl)-acetate (**3ra**)

Brown oil (61.3 mg, 70% yield); 96:4 dr. **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.36–7.27 (m, 4H), 6.77 (d, *J* = 8.9 Hz, 2H), 6.64 (d, *J* = 8.9 Hz, 2H), 4.20–4.09 (m, 3H), 3.74 (s, 3H), 3.63–3.59 (m, 2H), 2.83–2.58 (m, 1H), 2.42–2.17 (m, 1H), 2.16–1.91 (m, 3H), 1.75–1.46 (m, 4H), 1.28–1.25 (m, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 213.48, 175.06, 171.40, 152.31, 141.67, 133.91, 133.60, 130.01, 127.51, 114.88, 114.04, 90.08, 61.01, 57.91, 55.74, 46.97, 40.89, 33.51, 24.70, 23.24, 22.17, 14.12. **HRMS (ESI)** *m/z*: calcd for C₂₅H₃₁N₂O₅ [M+H]⁺ 439.2233, found 439.2231.

Optical Rotation: $[\alpha]_D^{25} = 15.4$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis**: 96.5:3.5 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 95/5, flow rate = 0.7 mL/min, λ = 240 nm, retention time: *t*_{major} = 21.7 min, *t*_{minor} = 23.4 min.

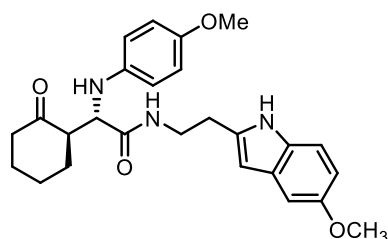
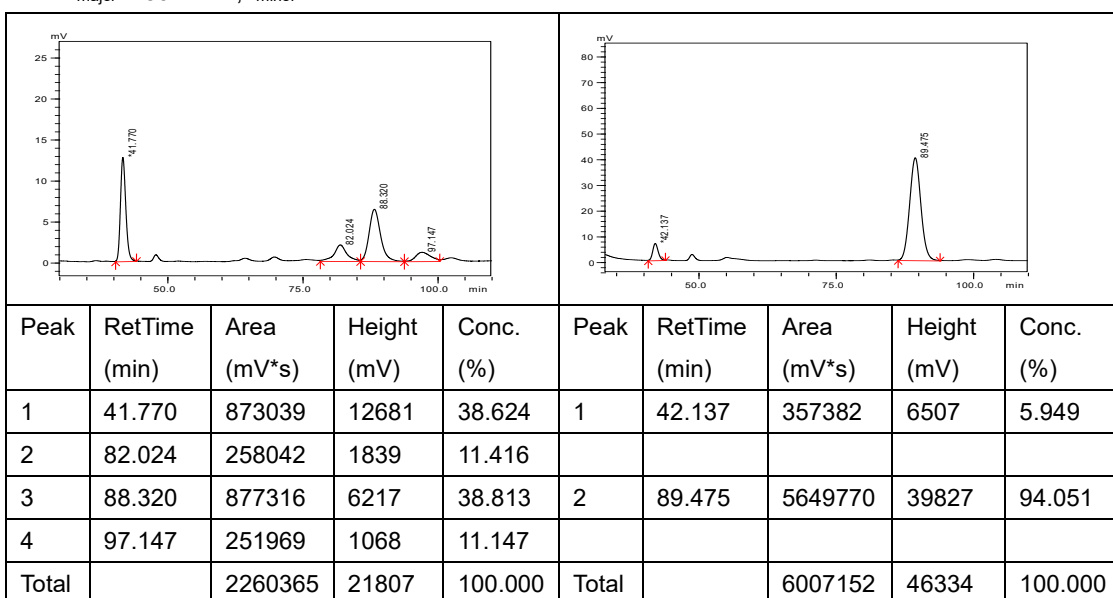


Ethyl 4-((S)-2-((R)-2-oxocyclohexyl)acetamido)benzoate (**3sa**)

Brown oil (57.7 mg, 68% yield); 99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.03 (d, *J* = 8.5 Hz, 2H), 7.43 (d, *J* = 8.6 Hz, 2H), 6.76 (d, *J* = 9.1 Hz, 2H), 6.62 (d, *J* = 8.9 Hz, 2H), 4.64 (d, *J* = 5.9 Hz, 1H),

4.40–4.33 (m, 2H), 3.73 (s, 3H), 2.83–2.62 (m, 1H), 2.17–2.08 (m, 1H), 1.78–1.60 (m, 2H), 1.59–1.41 (m, 2H), 1.39 (t, $J = 7.1$ Hz, 3H), 1.28–1.12 (m, 1H), 1.12–0.96 (m, 1H), 0.96–0.73 (m, 1H). ^{13}C NMR (100 MHz, Chloroform- d) δ 213.67, 174.86, 166.03, 152.41, 141.38, 139.32, 130.68, 130.28, 126.62, 118.85, 114.90, 114.09, 90.26, 61.16, 57.92, 55.72, 47.15, 42.26, 33.32, 24.76, 23.19, 21.96, 14.26. **HRMS (ESI)** m/z : calcd for $\text{C}_{24}\text{H}_{29}\text{N}_2\text{O}_5$ $[\text{M}+\text{H}]^+$ 425.2076, found 425.2056.

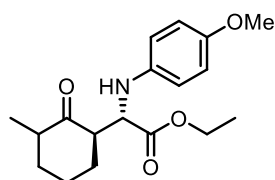
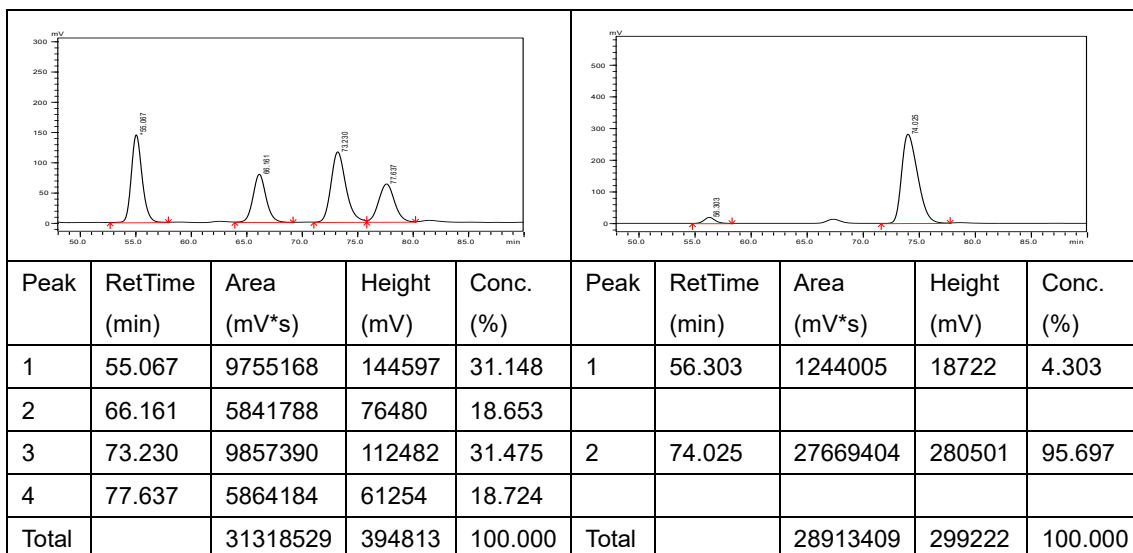
Optical Rotation: $[\alpha]^{25}_{\text{D}} = 16.3$ ($c = 1.0$, CH_2Cl_2). The ee value was determined by **HPLC analysis:** 94.0:6.0 er , Chiralcel AD-H column, hexane/ i -PrOH = 90/10, flow rate = 0.7 mL/min, $\lambda = 240$ nm, retention time: $t_{\text{major}} = 89.4$ min, $t_{\text{minor}} = 42.1$ min.



(*S*)-*N*-(2-(5-methoxy-1H-indol-2-yl)ethyl)-2-((4-methoxyphenyl)amino)-2-((*R*)-2-oxocyclohexyl)acetamide (**3ta**)

Brown oil (67.4mg, 75% yield); 85:15 dr . ^1H NMR (400 MHz, Chloroform- d) δ 8.05 (s, 1H), 7.22 (d, $J = 8.8$ Hz, 1H), 7.09 (t, $J = 5.8$ Hz, 1H), 7.03 (d, $J = 2.5$ Hz, 1H), 6.89 (d, $J = 2.4$ Hz, 1H), 6.84 (m, 1H), 6.72 (d, $J = 8.9$ Hz, 2H), 6.47 (d, $J = 8.9$ Hz, 2H), 3.89 (d, $J = 3.2$ Hz, 1H), 3.85 (s, 3H), 3.73 (s, 3H), 3.56 (m, 2H), 3.36–3.06 (m, 1H), 2.89 (t, $J = 6.8$ Hz, 2H), 2.51–2.24 (m, 2H), 2.09–1.96 (m, 2H), 1.88–1.78 (m, 1H), 1.73–1.48 (m, 3H). ^{13}C NMR (100 MHz, Chloroform- d) δ 213.46, 172.15, 153.89, 152.47, 141.10, 131.40, 127.61, 122.97, 114.96, 114.34, 112.39, 112.20, 111.85, 100.35, 59.16, 55.87, 55.69, 53.08, 42.29, 39.40, 31.21, 27.50, 25.18, 24.86. **HRMS (ESI)** m/z : calcd for $\text{C}_{26}\text{H}_{32}\text{N}_3\text{O}_4$ $[\text{M}+\text{H}]^+$ 450.2393, found 450.2380.

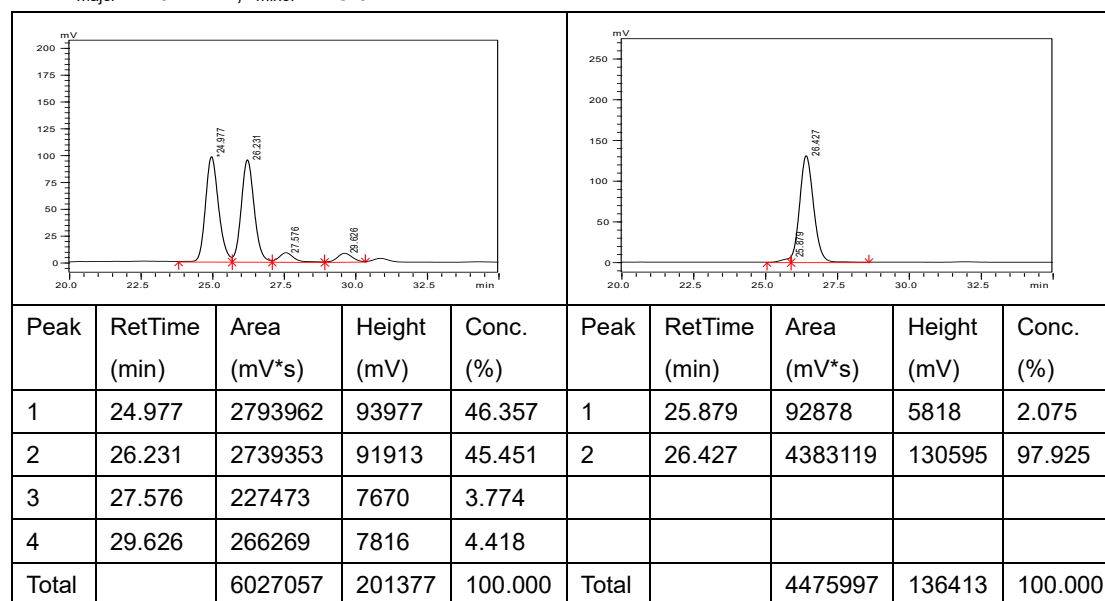
Optical Rotation: $[\alpha]^{25}_{\text{D}} = 63.5$ ($c = 1.0$, CH_2Cl_2). The ee value was determined by **HPLC analysis:** 95.7:4.3 er , Chiralcel AD-H column, hexane/ i -PrOH = 98/2, flow rate = 1.0 mL/min, $\lambda = 240$ nm, retention time: $t_{\text{major}} = 74$ min, $t_{\text{minor}} = 56.3$ min.

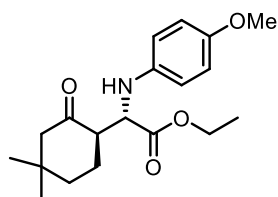


Ethyl (2S)-2-((4-methoxyphenyl)amino)-2-((1R)-3-methyl-2-oxocyclohexyl)acetate (**3ab**)

Yellow oil (42.2 mg, 66% yield); 97:3 *dr*. **¹H NMR** (400 MHz, DMSO-*d*₆) δ 6.71 (d, *J* = 9.0 Hz, 2H), 6.61 (d, *J* = 9.0 Hz, 2H), 5.48 (d, *J* = 10.1 Hz, 1H), 4.34–4.24 (m, 1H), 3.96 (q, *J* = 7.1 Hz, 2H), 3.63 (s, 3H), 2.81–2.66 (m, 2H), 2.07–1.92 (m, 2H), 1.82–1.73 (m, 2H), 1.67–1.58 (m, 1H), 1.49–1.40 (m, 1H), 1.07 (t, *J* = 7.1 Hz, 3H), 0.99 (d, *J* = 6.7 Hz, 3H). **¹³C NMR** (100 MHz, DMSO-*d*₆) δ 212.85, 172.89, 151.38, 141.86, 114.52, 114.06, 60.13, 56.07, 55.27, 51.91, 42.62, 34.55, 28.59, 19.70, 15.24, 14.00. **HRMS (ESI)** *m/z*: calcd for C₁₈H₂₅NNaO₄ [M+Na]⁺ 342.1681, found 342.1670.

Optical Rotation: [α]_D²⁵ = 23.9 (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis**: 97.9:2.1 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, λ = 240 nm, retention time: *t*_{major} = 26.4 min, *t*_{minor} = 25.9 min.

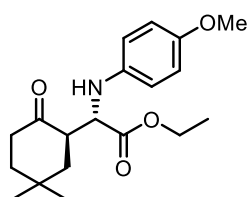
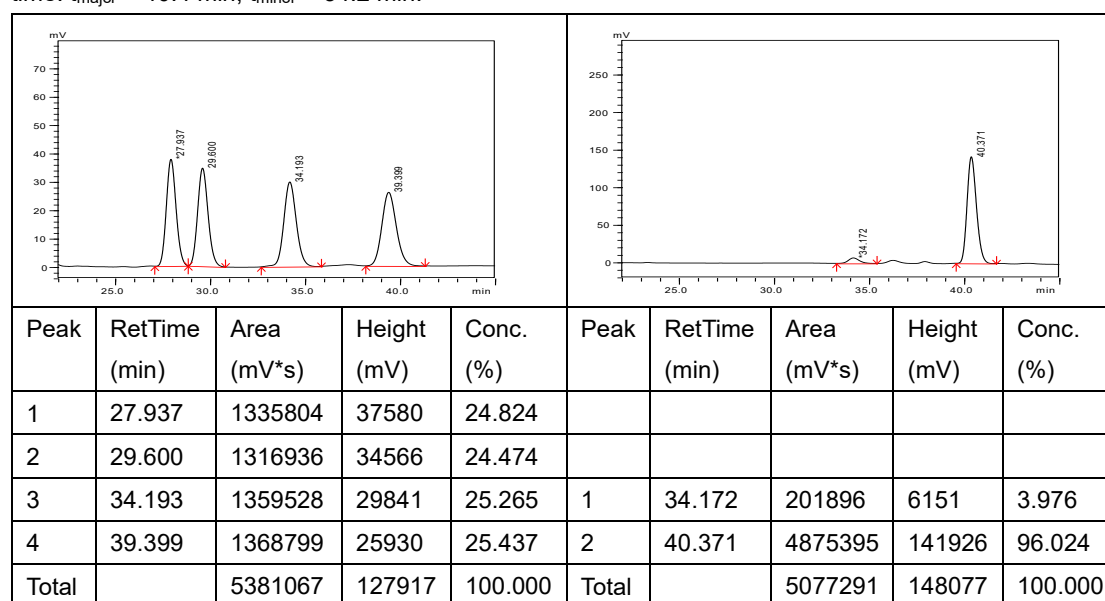




Ethyl (S)-2-((R)-4,4-dimethyl-2-oxocyclohexyl)-2-((4-methoxyphenyl)amino)acetate (**3ac**)

Yellow oil (51.3 mg, 77% yield); >99:1 *dr*. **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.76 (d, *J* = 8.9 Hz, 2H), 6.65 (d, *J* = 8.9 Hz, 2H), 4.18–4.10 (m, 2H), 3.97 (d, *J* = 4.0 Hz, 1H), 3.73 (s, 3H), 3.09–2.95 (m, 1H), 2.26–2.19 (m, 1H), 2.16–2.00 (m, 3H), 1.76–1.62 (m, 2H), 1.21 (t, *J* = 7.1 Hz, 3H), 1.05 (s, 3H), 0.87 (s, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 211.02, 173.11, 152.84, 142.18, 115.83, 114.75, 61.20, 58.97, 55.69, 54.66, 52.44, 37.47, 36.25, 31.41, 26.16, 25.35, 14.09. **HRMS (ESI)** *m/z*: calcd for C₁₉H₂₇NNaO₄ [M+Na]⁺ 356.1838, found 356.1827.

Optical Rotation: $[\alpha]^{25}_D = 16.8$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 96.0:4.0 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, λ = 240 nm, retention time: *t*_{major} = 40.4 min, *t*_{minor} = 34.2 min.

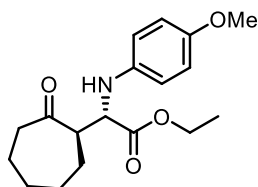
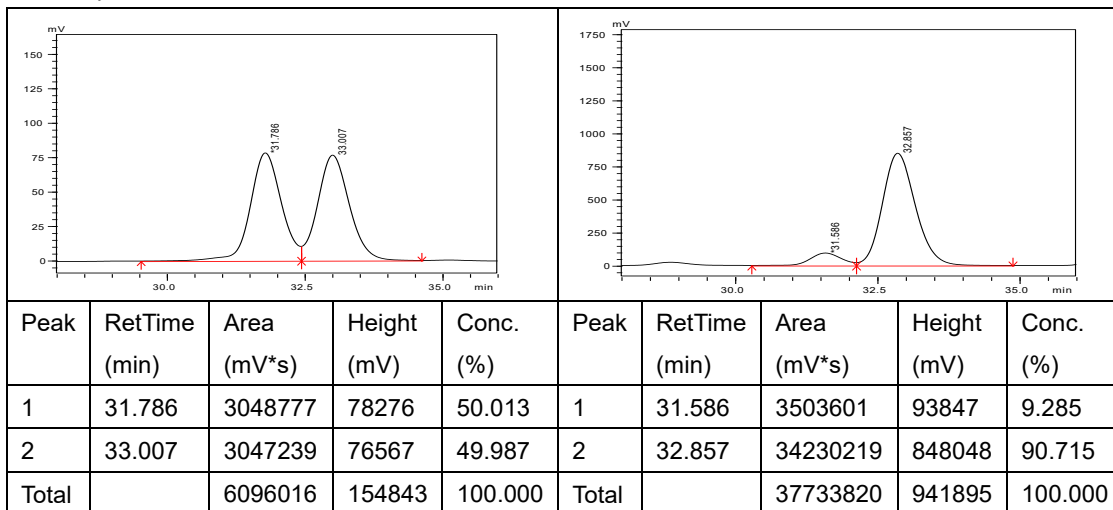


Ethyl (S)-2-((R)-5,5-dimethyl-2-oxocyclohexyl)-2-((4-methoxyphenyl)amino)acetate (**3ad**)⁷

Yellow oil (50.7 mg, 76% yield); 95:5 *dr*. **¹H NMR** (600 MHz, Chloroform-*d*) δ 6.76 (d, *J* = 9.0 Hz, 2H), 6.63 (d, *J* = 9.0 Hz, 2H), 4.17–4.10 (m, 2H), 3.86 (d, *J* = 3.4 Hz, 1H), 3.73 (s, 3H), 3.33–3.21 (m, 1H), 2.51–2.44 (m, 1H), 2.31–2.25 (m, 1H), 1.93 (t, *J* = 13.2 Hz, 1H), 1.78–1.73 (m, 1H), 1.71–1.62 (m, 2H), 1.24 (s, 3H), 1.21 (t, *J* = 7.1 Hz, 4H), 1.04 (s, 3H). **¹³C NMR** (150 MHz, Chloroform-*d*) δ 211.46, 173.12, 152.79, 142.36, 115.72, 114.74, 61.16, 59.42, 55.66, 49.55, 42.88, 38.80, 37.99, 31.48, 30.63, 24.55, 14.06.

Optical Rotation: $[\alpha]^{25}_D = 14.5$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 94.8:5.2 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.5 mL/min, λ = 254 nm, retention

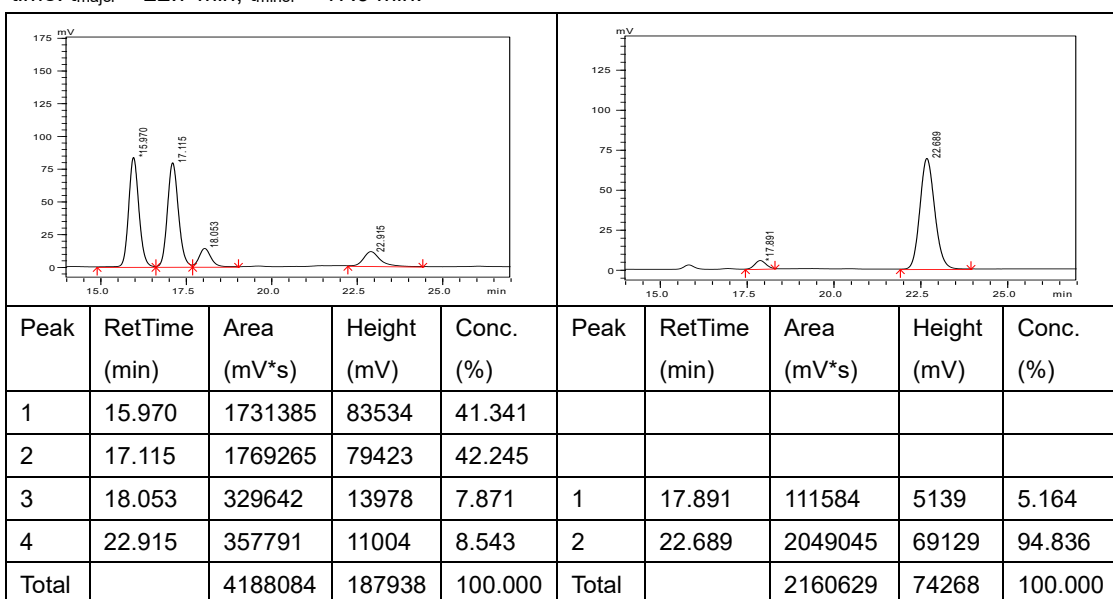
time: $t_{\text{major}} = 22.7 \text{ min}$, $t_{\text{minor}} = 17.9 \text{ min}$.

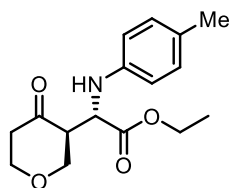


Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocycloheptyl)acetate (**3ae**)⁷

Yellow oil (48 mg, 75% yield); 99:1 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.77 (d, $J = 8.9 \text{ Hz}$, 2H), 6.67 (d, $J = 8.9 \text{ Hz}$, 2H), 4.31 (d, $J = 4.9 \text{ Hz}$, 1H), 4.16 (q, $J = 7.1 \text{ Hz}$, 2H), 3.74 (s, 3H), 3.07–2.98 (m, 1H), 2.61–2.47 (m, 2H), 2.03–1.84 (m, 4H), 1.60–1.49 (m, 2H), 1.43–1.32 (m, 2H), 1.23 (t, $J = 7.1 \text{ Hz}$, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 214.19, 172.55, 152.73, 140.91, 115.23, 114.85, 61.29, 60.69, 55.70, 54.34, 43.88, 29.90, 29.01, 27.16, 24.22, 14.15.

Optical Rotation: $[\alpha]_{\text{D}}^{25} = 15.6$ ($c = 1.0$, CH₂Cl₂). The *ee* value was determined by **HPLC analysis**: 94.8:5.2 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 254 \text{ nm}$, retention time: $t_{\text{major}} = 22.7 \text{ min}$, $t_{\text{minor}} = 17.9 \text{ min}$.

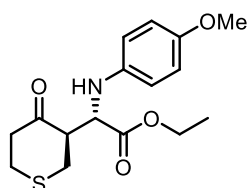
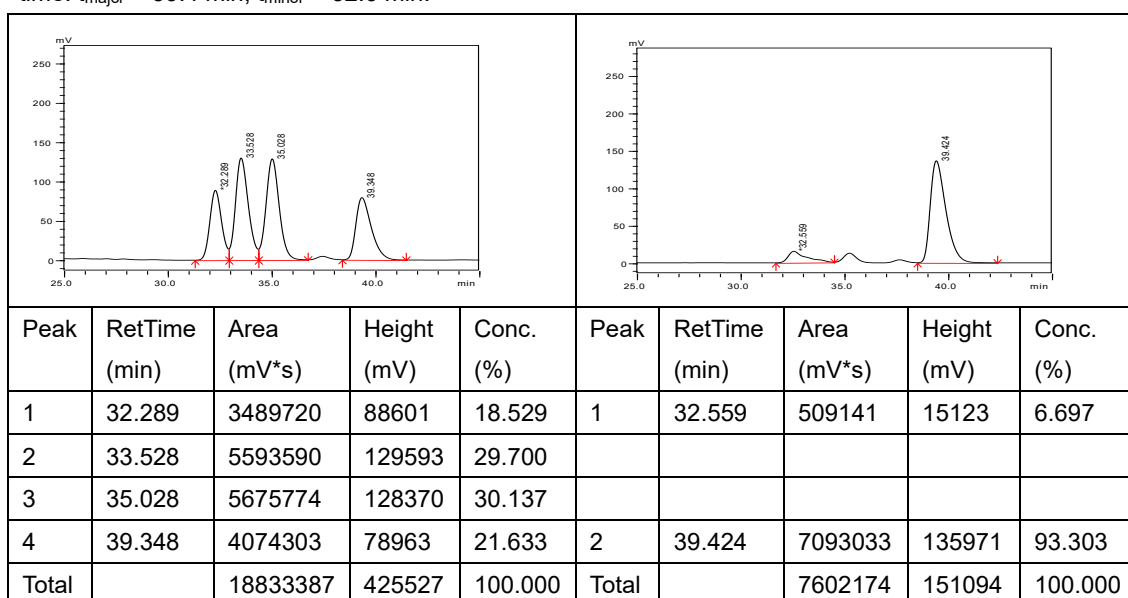




Ethyl (S)-2-((S)-4-oxotetrahydro-2H-pyran-3-yl)-2-(p-tolylamino)acetate (**3kf**)

Yellow oil (42 mg, 72% yield); 85:15 *dr*. $^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 6.98 (d, $J = 8.4$ Hz, 2H), 6.56 (d, $J = 8.4$ Hz, 2H), 4.26–4.08 (m, 5H), 3.97 (t, $J = 5.9$ Hz, 1H), 3.93–3.82 (m, 1H), 3.84–3.72 (m, 1H), 3.32–2.88 (m, 1H), 2.65–2.54 (m, 1H), 2.53–2.43 (m, 2H), [2.23, 2.04] (s, 3H), 1.22 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (100 MHz, Chloroform-*d*) δ 206.21, 171.97, 144.86, 129.77, 128.21, 114.19, 69.94, 67.75, 61.52, 55.32, 53.77, 42.81, 42.02, 20.33, 14.05. **HRMS (ESI)** m/z : calcd for $\text{C}_{16}\text{H}_{21}\text{NNaO}_4$ $[\text{M}+\text{Na}]^+$ 314.1368, found 314.1353.

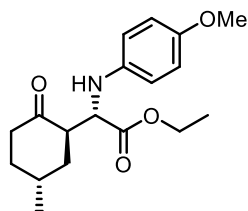
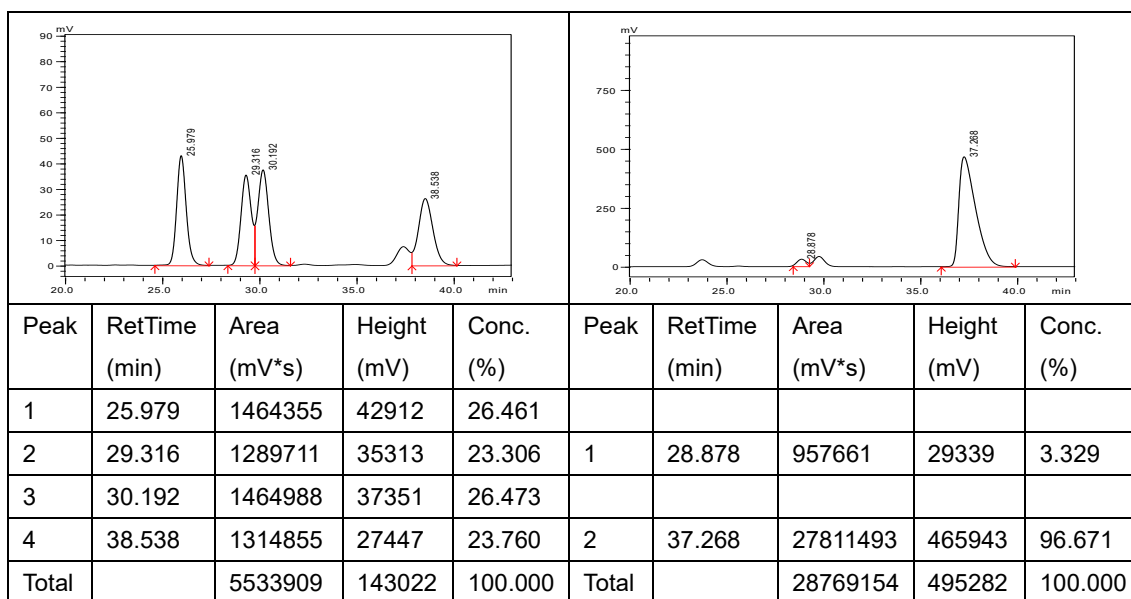
Optical Rotation: $[\alpha]_{25}^{\text{D}} = 8.6$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 93.3:7.7 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 96/4, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 39.4$ min, $t_{\text{minor}} = 32.6$ min.



Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-4-oxotetrahydro-2H-thiopyran-3-yl)acetate (**3ag**)⁷

Yellow oil (49.2mg, 76% yield); 95:5 *dr*. $^1\text{H NMR}$ (400 MHz, DMSO-*d*₆) δ 6.71 (d, $J = 8.9$ Hz, 2H), 6.63 (d, $J = 9.0$ Hz, 2H), 5.17 (d, $J = 10.0$ Hz, 1H), 4.52–4.35 (m, 1H), 4.09–4.03 (m, 2H), 3.64 (s, 3H), 3.26–3.16 (m, 1H), 3.09–3.01 (m, 1H), 2.99–2.88 (m, 3H), 2.71–2.58 (m, 2H), 1.13 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (100 MHz, DMSO-*d*₆) δ 207.42, 171.93, 151.59, 141.83, 114.49, 114.34, 60.50, 56.64, 55.26, 54.50, 43.25, 31.32, 29.18, 14.03.

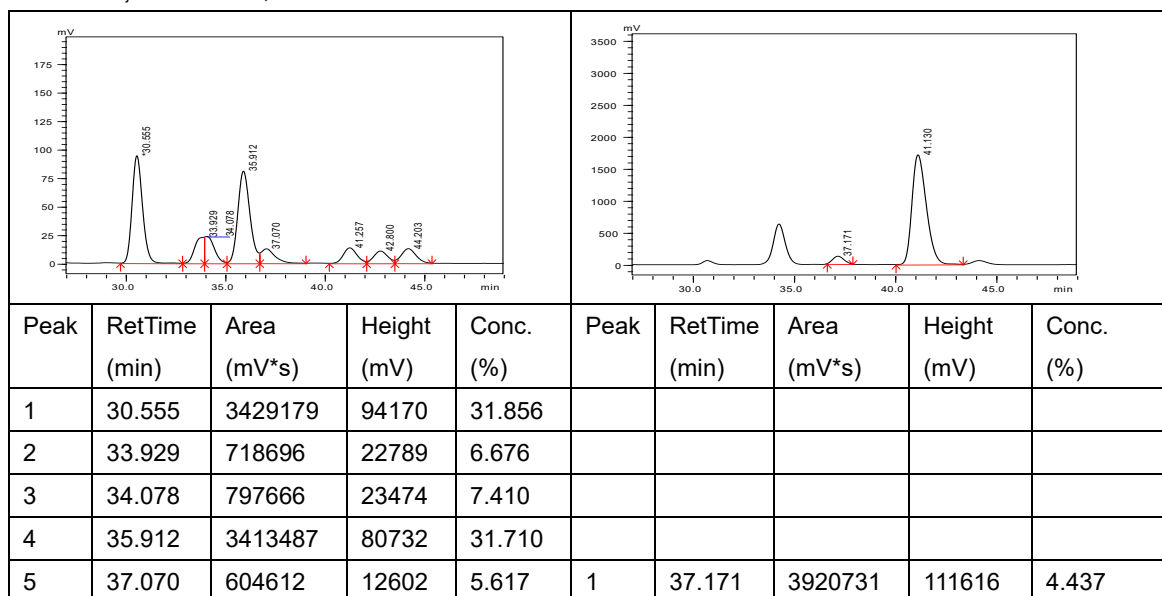
Optical Rotation: $[\alpha]_{25}^{\text{D}} = 32.5$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 96.7:3.3 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 37.3$ min, $t_{\text{minor}} = 28.9$ min.



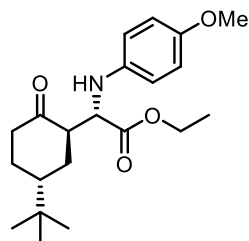
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((1R,5R)-5-methyl-2-oxocyclohexyl)acetate (**3ah**)⁷

Yellow oil (50.5 mg, 79% yield); 80:20 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.73 (d, *J* = 8.9 Hz, 2H), 6.60 (d, *J* = 8.9 Hz, 2H), 4.16–4.07 (m, 3H), 3.69 (s, 3H), 3.20–2.94 (m, 1H), 2.42–2.31 (m, 2H), 2.26–2.09 (m, 1H), 2.06–1.90 (m, 2H), 1.79–1.48 (m, 2H), 1.18 (t, *J* = 7.1 Hz, 3H), 1.11 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 211.17, 172.65, 152.74, 141.23, 115.35, 114.57, 61.00, 58.80, 55.43, 50.23, 37.58, 36.13, 33.16, 26.66, 19.27, 13.99.

Optical Rotation: $[\alpha]_D^{25} = 23.6$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 95.6:4.4 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.5 mL/min, λ = 254 nm, retention time: *t*_{major} = 41.1 min, *t*_{minor} = 37.2 min.



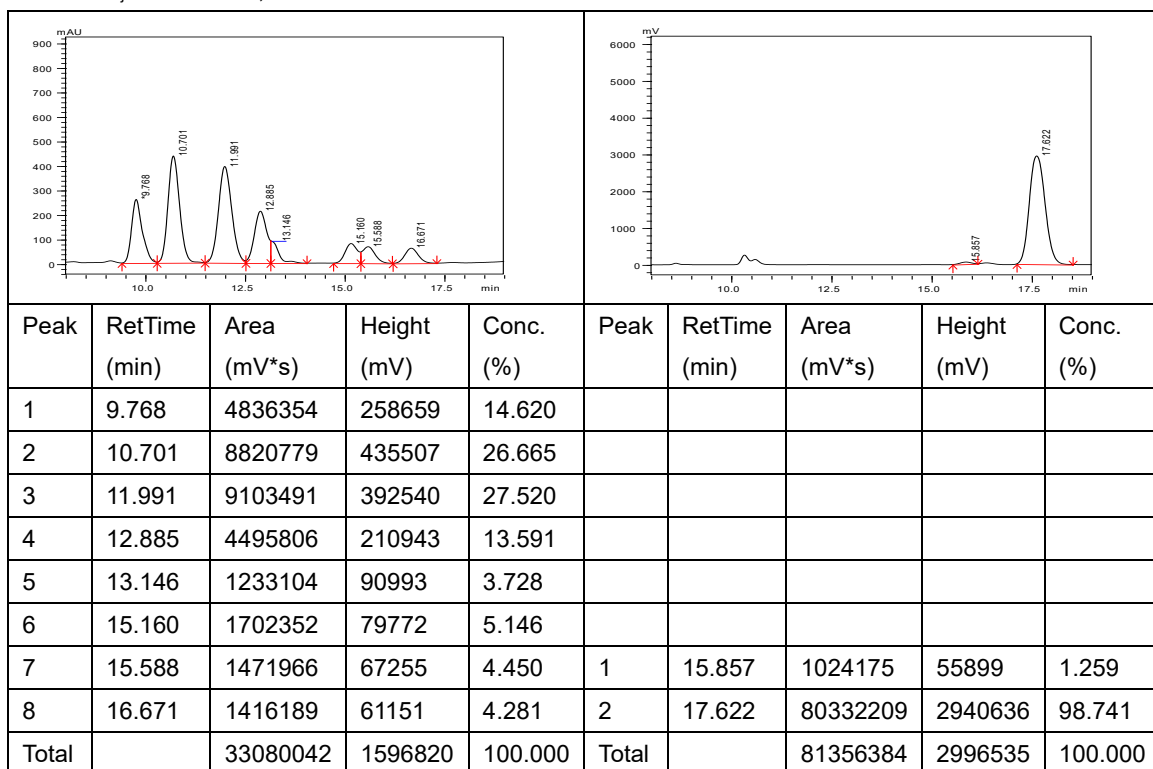
6	41.257	634184	13496	5.891	2	41.130	84444729	1713470	95.563
7	42.800	518317	10633	4.815					
8	44.203	648479	12665	6.024					
Total		10764620	270560	100.000	Total		88365460	1825086	100.000

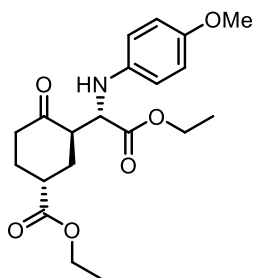


Ethyl (S)-2-((1R,5R)-5-(tert-butyl)-2-oxocyclohexyl)-2-((4-methoxyphenyl)amino)acetate (**3ai**)⁷

Yellow oil (52 mg, 72% yield); 98:2 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.74 (d, *J* = 8.9 Hz, 2H), 6.62 (d, *J* = 8.9 Hz, 2H), 4.32 (d, *J* = 9.2 Hz, 1H), 4.19–4.10 (m, 2H), 3.72 (s, 3H), 2.85–2.66 (m, 1H), 2.46–2.32 (m, 2H), 2.08–1.97 (m, 2H), 1.78–1.62 (m, 2H), 1.55–1.44 (m, 1H), 1.21 (t, *J* = 7.1 Hz, 4H), 0.92 (s, 9H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 210.35, 173.03, 151.72, 142.78, 114.90, 114.43, 60.56, 57.09, 55.69, 52.11, 46.01, 41.08, 32.63, 30.98, 27.85, 27.59, 14.55.

Optical Rotation: [α]_D²⁵ = 26.5 (c = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 98.7:1.3 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 17.6 min, *t*_{minor} = 15.9 min.

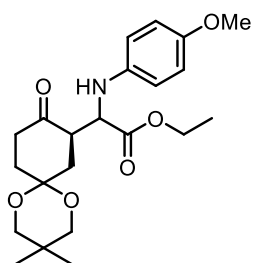
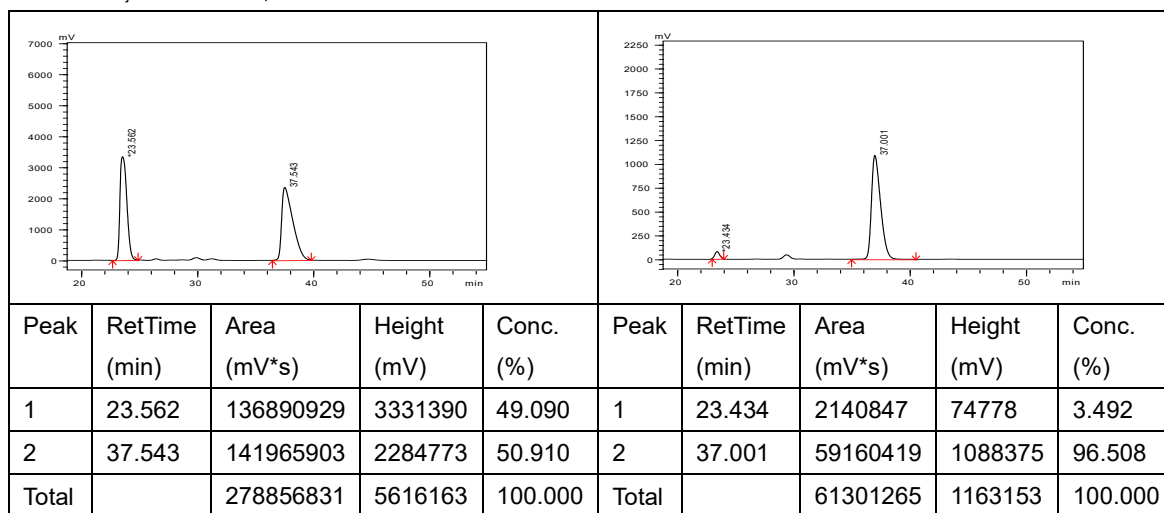




Ethyl (1R,3R)-3-((S)-2-ethoxy-1-((4-methoxyphenyl)amino)-2-oxoethyl)-4-oxocyclohexan-e-1-carboxylate (**3aj**)⁷

Yellow oil (62.6 mg, 83% yield); 97:3 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.78 (d, *J* = 8.9 Hz, 2H), 6.66 (d, *J* = 8.9 Hz, 2H), 4.26 (q, *J* = 7.1 Hz, 2H), 4.21–4.14 (m, 2H), 3.96 (d, *J* = 4.3 Hz, 1H), 3.76 (s, 3H), 3.45–3.30 (m, 1H), 3.02–2.90 (m, 1H), 2.60–2.32 (m, 4H), 2.28–2.19 (m, 1H), 2.01–1.95 (m, 1H), 1.33 (t, *J* = 7.1 Hz, 4H), 1.23 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 210.12, 173.99, 172.67, 152.94, 142.05, 115.75, 114.79, 61.30, 60.95, 59.13, 55.70, 50.20, 38.48, 38.20, 31.05, 27.61, 14.25, 14.10.

Optical Rotation: $[\alpha]^{25}_D = 29.2$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 96.5:3.5 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, λ = 240 nm, retention time: *t*_{major} = 37.0 min, *t*_{minor} = 23.4 min.

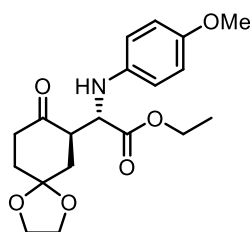
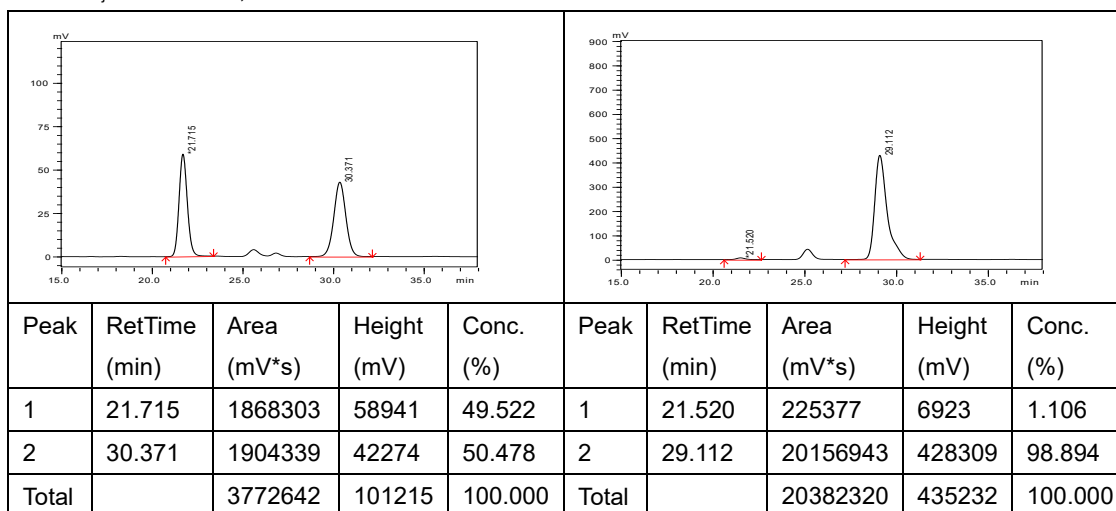


Ethyl 2-((R)-3,3-dimethyl-9-oxo-1,5-dioxaspiro[5.5]undecan-8-yl)-2-((4-methoxyphenyl)amino)acetate (**3ak**)⁷

Yellow oil (63 mg, 78% yield); 97:3 *dr*. ¹H NMR (600 MHz, Chloroform-*d*) δ 6.76 (d, *J* = 8.8 Hz, 2H), 6.63 (d, *J* = 8.9 Hz, 2H), 4.25–4.07 (m, 3H), 3.94 (d, *J* = 3.4 Hz, 1H), 3.73 (s, 3H), 3.57 (s, 2H), 3.55 (d, *J* = 5.1 Hz, 2H), 3.40–3.32 (m, 1H), 2.60–2.50 (m, 3H), 2.36–2.29 (m, 1H), 2.07 (t, *J* = 13.3 Hz, 1H), 1.80–1.72 (m, 1H), 1.21 (t, *J* = 7.1 Hz, 3H), 1.04 (s, 3H), 0.98 (s, 3H). ¹³C NMR (150 MHz, Chloroform-*d*) δ 209.91,

172.69, 152.97, 142.13, 115.89, 114.81, 96.60, 70.74, 70.47, 61.30, 59.02, 55.71, 48.82, 36.85, 35.42, 30.51, 30.24, 22.64, 22.52, 14.10.

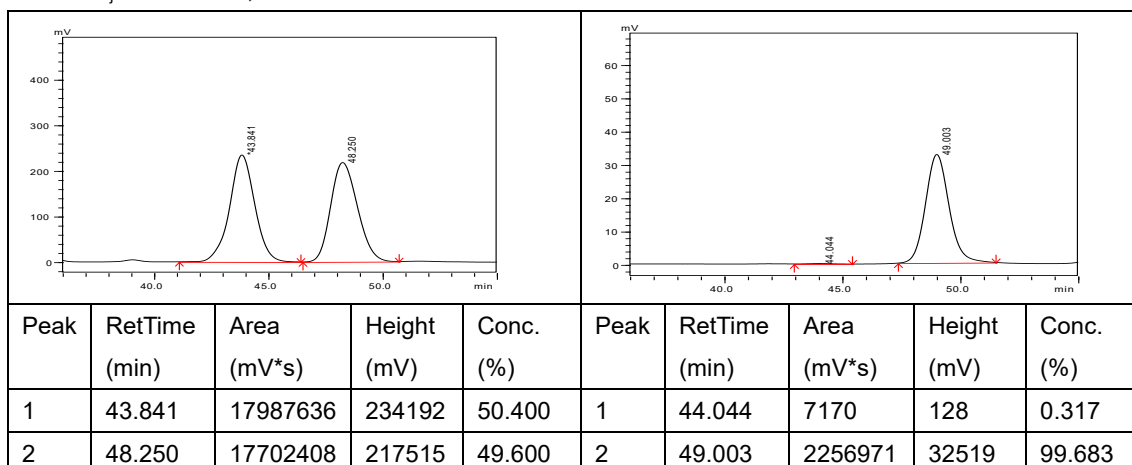
Optical Rotation: $[\alpha]_D^{25} = 20.8$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 98.9:1.1 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 240$ nm, retention time: $t_{\text{major}} = 29.1$ min, $t_{\text{minor}} = 21.5$ min.



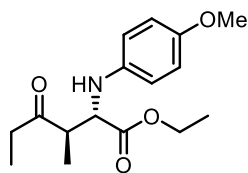
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-8-oxo-1,4-dioxaspiro[4.5]decan-7-yl)acetate (**3a**)⁷

Yellow oil (59.5mg, 82% yield); >99:1 dr. ¹H NMR (400 MHz, Chloroform-*d*) δ 6.75 (d, $J = 8.9$ Hz, 2H), 6.62 (d, $J = 8.9$ Hz, 2H), 4.37–4.09 (m, 3H), 4.09–4.00 (m, 4H), 3.91 (d, $J = 3.6$ Hz, 1H), 3.73 (s, 3H), 3.53–3.42 (m, 1H), 2.72–2.62 (m, 1H), 2.45–2.36 (m, 1H), 2.29 (t, $J = 13.1$ Hz, 1H), 2.14–2.07 (m, 1H), 2.05–1.93 (m, 2H), 1.21 (t, $J = 7.1$ Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 209.58, 172.65, 152.89, 142.10, 115.83, 114.72, 107.47, 64.81, 64.65, 61.29, 58.98, 55.65, 50.00, 37.96, 37.59, 33.81, 14.08.

Optical Rotation: $[\alpha]_D^{25} = 22.0$ ($c = 1.0$, CH_2Cl_2). The *ee* value was determined by **HPLC analysis:** 99.7:0.3 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 49.0$ min, $t_{\text{minor}} = 44.0$ min.



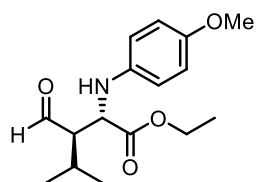
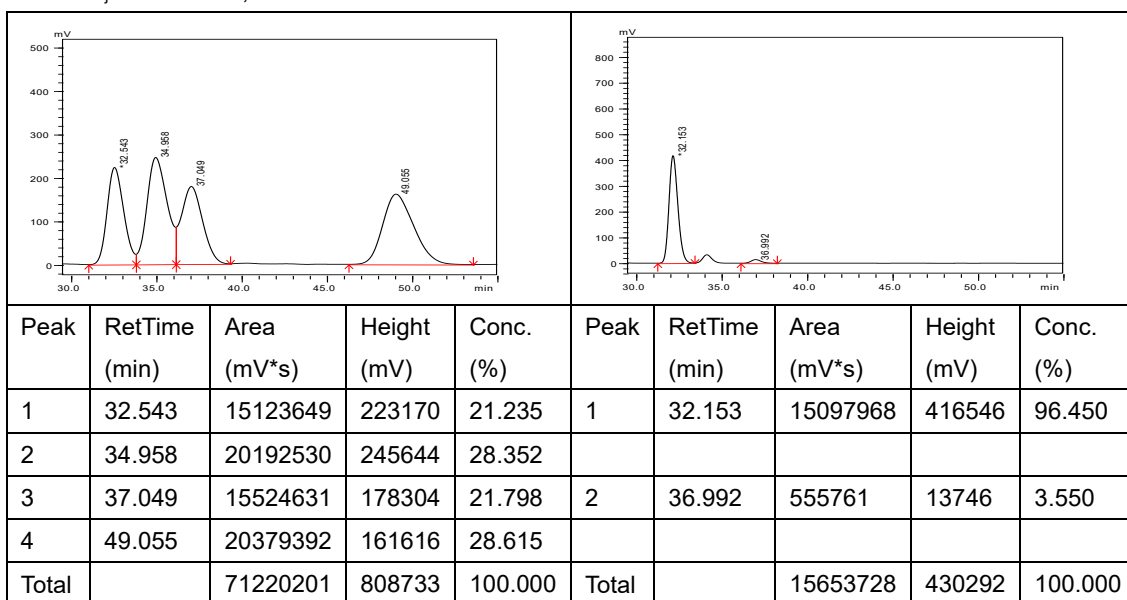
Total		35690044	451707	100.000	Total		2264141	32646	100.000
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Ethyl (2S,3R)-2-((4-methoxyphenyl)amino)-3-methyl-4-oxohexanoate (**3am**)⁷

Yellow oil (37 mg, 63% yield); 99:1 *dr*. ¹H NMR (600 MHz, Chloroform-*d*) δ 6.75 (d, *J* = 9.0 Hz, 2H), 6.64 (d, *J* = 9.0 Hz, 2H), 4.19 (d, *J* = 7.0 Hz, 1H), 4.16–4.10 (m, 2H), 3.73 (s, 3H), 3.08–2.98 (m, 1H), 2.58–2.49 (m, 2H), 1.20 (t, *J* = 7.2 Hz, 3H), 1.17 (d, *J* = 7.1 Hz, 3H), 1.06 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (150 MHz, Chloroform-*d*) δ 212.20, 172.76, 153.05, 140.80, 115.81, 114.79, 61.16, 60.78, 55.64, 48.36, 34.94, 14.12, 13.36, 7.50.

Optical Rotation: $[\alpha]_D^{25} = -19.6$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 96.5:3.5 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 96/4, flow rate = 1.0 mL/min, λ = 240 nm, retention time: *t*_{major} = 32.2 min, *t*_{minor} = 37.0 min.

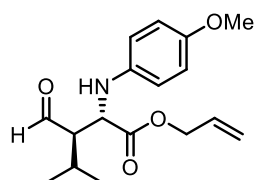
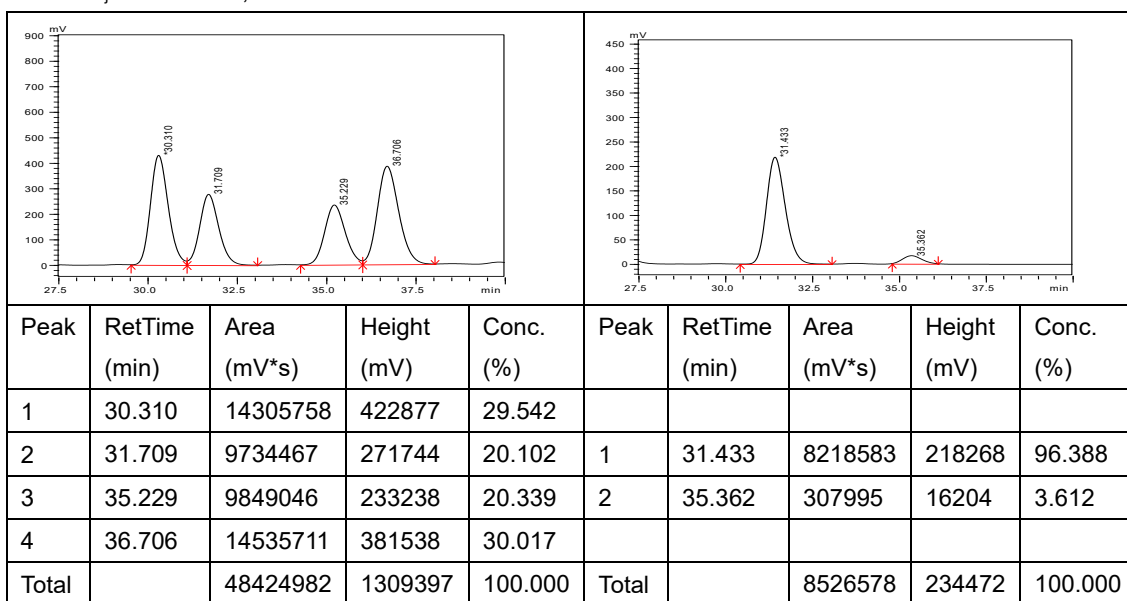


Ethyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)-4-methylpentanoate (**3an**)⁷

Yellow oil (38.7 mg, 66% yield); 90:10 *dr*. ¹H NMR (600 MHz, Chloroform-*d*) δ 9.74 (d, *J* = 3.5 Hz, 1H), 6.76 (d, *J* = 8.9 Hz, 2H), 6.66 (d, *J* = 8.9 Hz, 2H), 4.35 (d, *J* = 7.7 Hz, 1H), 4.14 (q, *J* = 7.2 Hz, 2H), 3.73 (s, 3H), 2.61–2.53 (m, 1H), 2.16–2.05 (m, 1H), 1.20 (t, *J* = 7.1 Hz, 3H), 1.11 (d, *J* = 6.9 Hz, 3H), 1.07 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (150 MHz, Chloroform-*d*) δ 203.20, 172.81, 153.26, 140.46, 115.86, 114.77, 61.30, 59.58, 57.24, 55.59, 27.51, 21.20, 19.12, 14.08.

Optical Rotation: $[\alpha]_D^{25} = -17.8$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:**

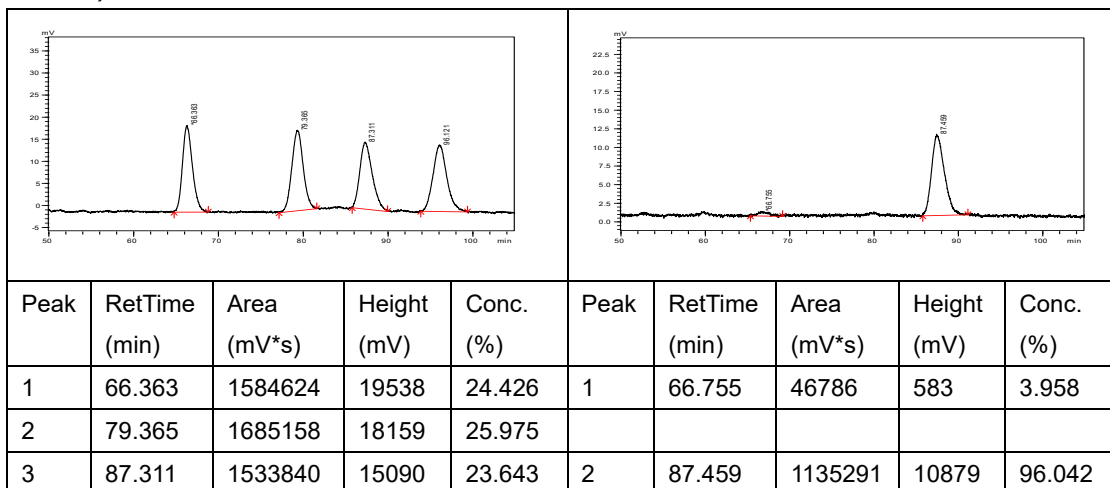
96.4:3.6 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 88/12, flow rate = 0.5 mL/min, λ = 240 nm, retention time: t_{major} = 31.4 min, t_{minor} = 35.4 min.



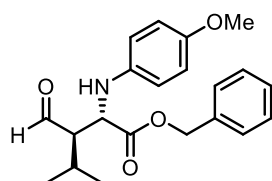
Ethyl (2*S*,3*R*)-3-formyl-2-((4-methoxyphenyl)amino)octanoate (**3en**)⁷

Yellow oil (43.3 mg, 71% yield); 99:1 *dr*. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.66 (d, J = 3.9 Hz, 1H), 6.73 (d, J = 9.0 Hz, 2H), 6.68 (d, J = 9.0 Hz, 2H), 5.83 (m, 1H), 5.66 (d, J = 10.7 Hz, 1H), 5.24 (dd, J = 17.3, 1.7 Hz, 1H), 5.17 (dd, J = 10.5, 1.6 Hz, 1H), 4.56 (d, J = 5.3 Hz, 2H), 4.53–4.47 (m, 1H), 3.64 (s, 3H), 2.50–2.44 (m, 1H), 2.02–1.88 (m, 1H), 1.01 (d, J = 6.9 Hz, 3H), 0.97 (d, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 203.74, 172.27, 151.87, 141.20, 132.19, 117.85, 114.62, 114.48, 64.83, 58.60, 56.02, 55.23, 26.87, 20.97, 18.42.

Optical Rotation: $[\alpha]_{\text{D}}^{25} = -26.5$ (c = 1.0, CH₂Cl₂). The *ee* value was determined by HPLC analysis: 96.1:3.9 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 98/2, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 87.5 min, t_{minor} = 66.8 min.



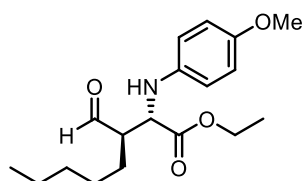
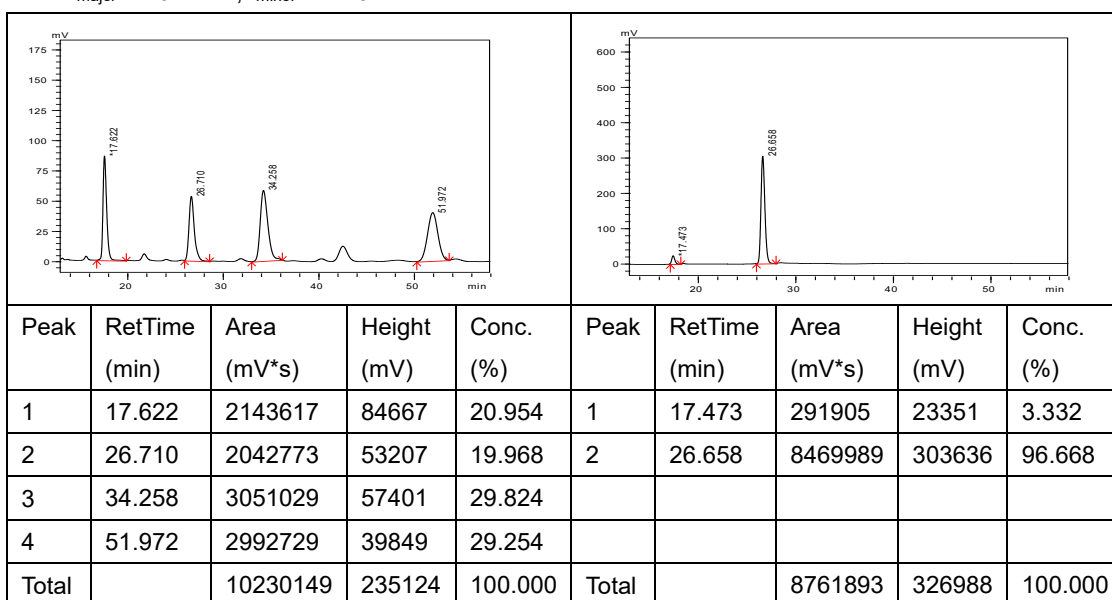
4	96.121	1683905	15029	25.956					
Total		6487528	67816	100.000	Total		1182078	11463	100.000



Benzyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)-4-methylpentanoate (**3gn**)⁷

Yellow oil (48 mg, 68% yield); 92:8 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.72 (d, *J* = 3.4 Hz, 1H), 7.37–7.28 (m, 3H), 7.26–7.14 (m, 2H), 6.76 (d, *J* = 8.9 Hz, 2H), 6.66 (d, *J* = 8.9 Hz, 2H), 5.13 (d, *J* = 1.1 Hz, 2H), 4.43 (d, *J* = 7.9 Hz, 1H), 3.74 (s, 3H), 2.68–2.57 (m, 1H), 2.09–1.98 (m, 1H), 1.10 (d, *J* = 7.0 Hz, 3H), 1.04 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 203.09, 172.66, 153.22, 140.30, 135.11, 128.42, 128.27, 128.13, 115.85, 114.72, 66.95, 59.41, 57.16, 55.52, 27.46, 21.12, 18.98.

Optical Rotation: [α]_D²⁵ = -11.3 (c = 1.0, CH₂Cl₂). The ee value was determined by **HPLC analysis:** 96.7:3.3 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, λ = 254 nm, retention time: *t*_{major} = 26.7 min, *t*_{minor} = 17.5 min.

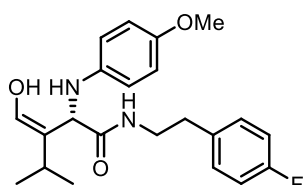
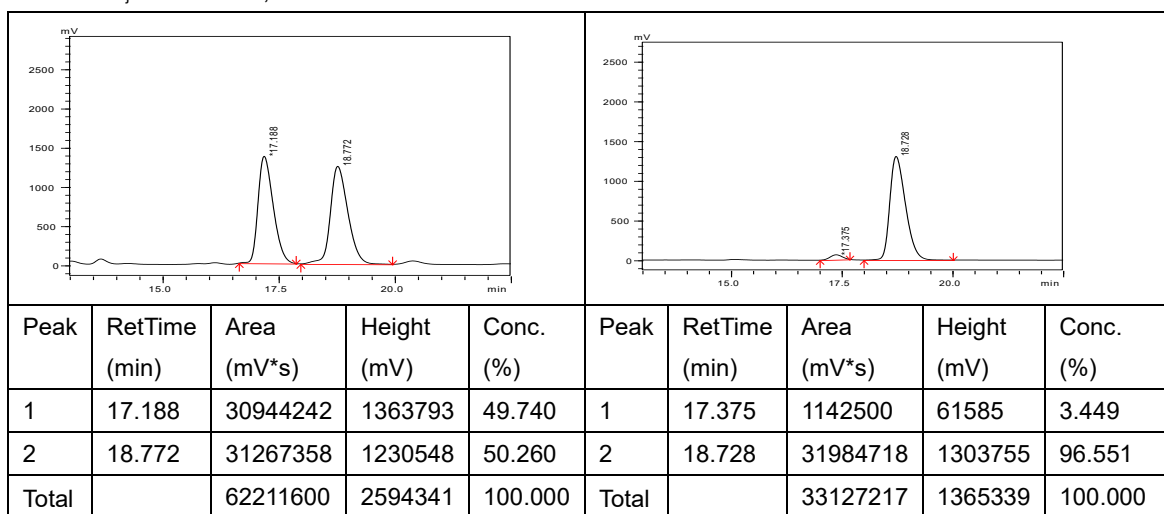


Ethyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)octanoate (**3ao**)⁷

Yellow oil (43.7 mg, 68% yield); 92:8 *dr*. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.65 (d, *J* = 2.5 Hz, 1H), 6.77 (d, *J* = 8.9 Hz, 2H), 6.64 (d, *J* = 8.9 Hz, 2H), 4.26 (d, *J* = 6.5 Hz, 1H), 4.21–4.12 (m, 2H), 3.73 (s, 3H), 2.87–2.64 (m, 1H), 1.78–1.67 (m, 1H), 1.60–1.51 (m, 1H), 1.37–1.26 (m, 6H), 1.22 (t, *J* = 7.1 Hz, 3H), 0.89–0.84 (m, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 202.34, 172.27, 153.20, 140.39, 115.72, 114.88, 61.53, 58.14, 55.68, 53.97, 31.68, 26.99, 25.69, 22.36, 14.18, 13.96.

Optical Rotation: [α]_D²⁵ = -22.6 (c = 1.0, CH₂Cl₂). The ee value was determined by **HPLC analysis:**

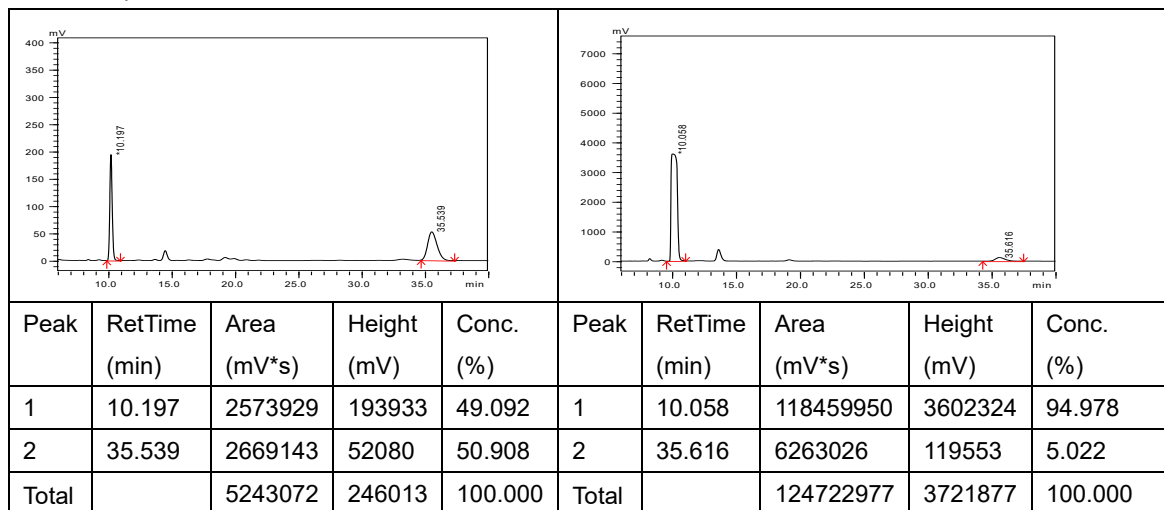
96.6:3.4 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 0.7 mL/min, λ = 254 nm, retention time: t_{major} = 17.4 min, t_{minor} = 18.3 min.

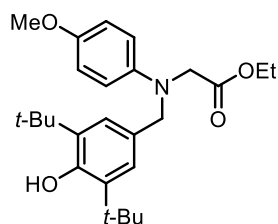


(*S,Z*)-*N*-(4-fluorophenethyl)-3-(hydroxymethylene)-2-((4-methoxyphenyl)amino)-4-methyl-pentanamide (**3pn**)

Yellow oil (47.9 mg, 62% yield); **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.23–7.03 (m, 1H), 7.03–6.86 (m, 5H), 6.75 (d, J = 8.9 Hz, 2H), 6.51 (d, J = 8.9 Hz, 2H), 5.84 (s, 1H), 5.07 (s, 1H), 3.75 (s, 3H), 3.72 (s, 1H), 3.31–3.19 (m, 1H), 3.01–2.77 (m, 2H), 2.75–2.67 (m, 1H), 2.60–2.48 (m, 1H), 1.13 (d, J = 6.7 Hz, 3H), 1.05 (d, J = 7.0 Hz, 3H). **¹³C NMR** (100 MHz, Chloroform-*d*) δ 169.99, 167.85, 161.52 (d, J_1 = 243 Hz, Cq), 153.30, 139.14, 134.74, 134.71 (d, J_4 = 3 Hz, Cq), 130.27 (d, J_3 = 7 Hz, CH), 130.19, 120.58, 115.52, 115.20 (d, J_2 = 21 Hz, CH), 115.00, 72.93, 55.64, 41.40, 34.08, 27.03, 22.27, 20.08. **¹⁹F NMR** (376 MHz, Chloroform-*d*) δ -116.58. **HRMS (ESI) m/z** : calcd for C₂₂H₂₆¹⁹FN₂O₄ [M+H]⁺ 369.1973, found 369.1981.

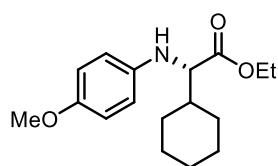
Optical Rotation: $[\alpha]_{\text{D}}^{25} = -19.4$ (c = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis**: 95.0:5.0 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 90/10, flow rate = 1.5 mL/min, λ = 254 nm, retention time: t_{major} = 10.1 min, t_{minor} = 35.5 min.





Ethyl *N*-(3,5-di-*tert*-butyl-4-hydroxybenzyl)-*N*-(4-methoxyphenyl)glycinate (**1a-BHT**)⁹

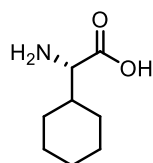
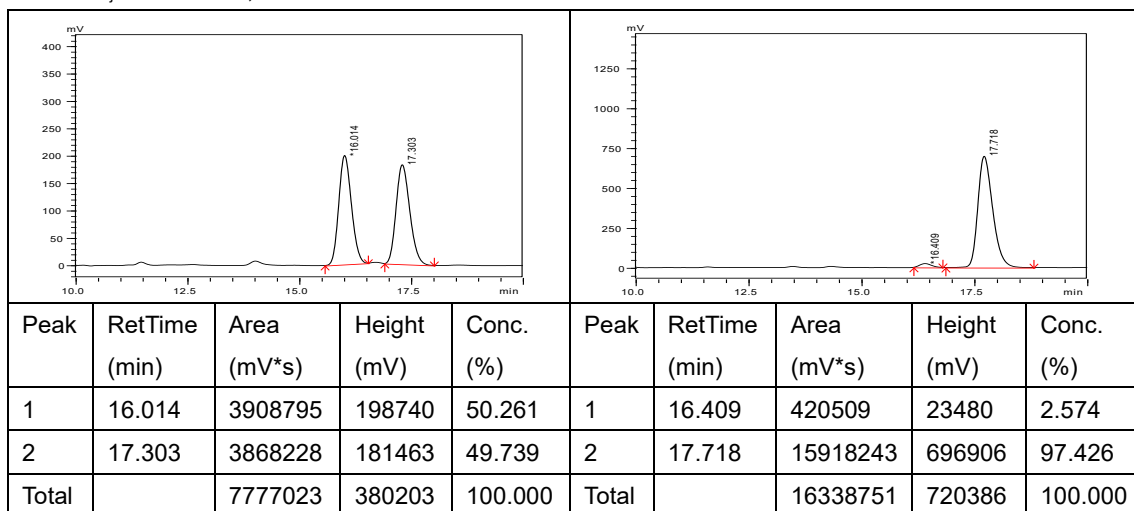
Colorless oil; **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.07 (s, 2H), 6.81 (d, *J* = 9.1 Hz, 2H), 6.73 (d, *J* = 9.2 Hz, 2H), 5.14 (s, 1H), 4.47 (s, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 3.94 (s, 2H), 3.75 (s, 3H), 1.41 (s, 18H), 1.23 (s, 3H).



Ethyl (*S*)-2-cyclohexyl-2-((4-methoxyphenyl)amino)acetate (**4**)¹⁰

Colorless oil (371 mg, 78% yield). **¹H NMR** (400 MHz, DMSO-*d*₆) δ 6.66 (d, *J* = 9.0 Hz, 2H), 6.52 (d, *J* = 8.9 Hz, 2H), 4.04 (m, 2H), 3.63 (m, 1H), 3.60 (s, 3H), 1.83 (m, 1H), 1.73–1.51 (m, 5H), 1.12–1.11 (m, 8H). **¹³C NMR** (100 MHz, DMSO-*d*₆) δ 173.68, 151.07, 142.25, 114.49, 113.62, 62.16, 59.93, 55.26, 40.22, 29.15, 29.13, 25.86, 25.64, 14.24.

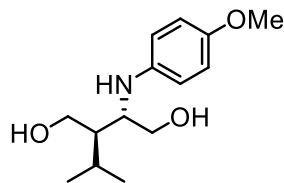
Optical Rotation: $[\alpha]^{25}_{\text{D}} = 26.8$ (*c* = 1.0, CH₂Cl₂). The *ee* value was determined by **HPLC analysis:** 97.4:2.6 *er*, Chiralcel AD-H column, hexane/*i*-PrOH = 88/12, flow rate = 1.0 mL/min, λ = 214 nm, retention time: *t*_{major} = 17.7 min, *t*_{minor} = 16.4 min.



(*S*)-2-amino-2-cyclohexylacetic acid (**5**)¹¹

White powder (138 mg, 88% yield). **¹H NMR** (600 MHz, Methanol-*d*₄) δ 3.75 (m, 1H), 1.93–1.86 (m, 1H), 1.81–1.75 (m, 3H), 1.65 (m, 2H), 1.27 (m, 3H), 1.18–1.08 (m, 2H). **¹³C NMR** (150 MHz, Methanol-*d*₄) δ 171.18, 58.88, 40.24, 29.54, 29.28, 27.05, 26.97, 26.77.

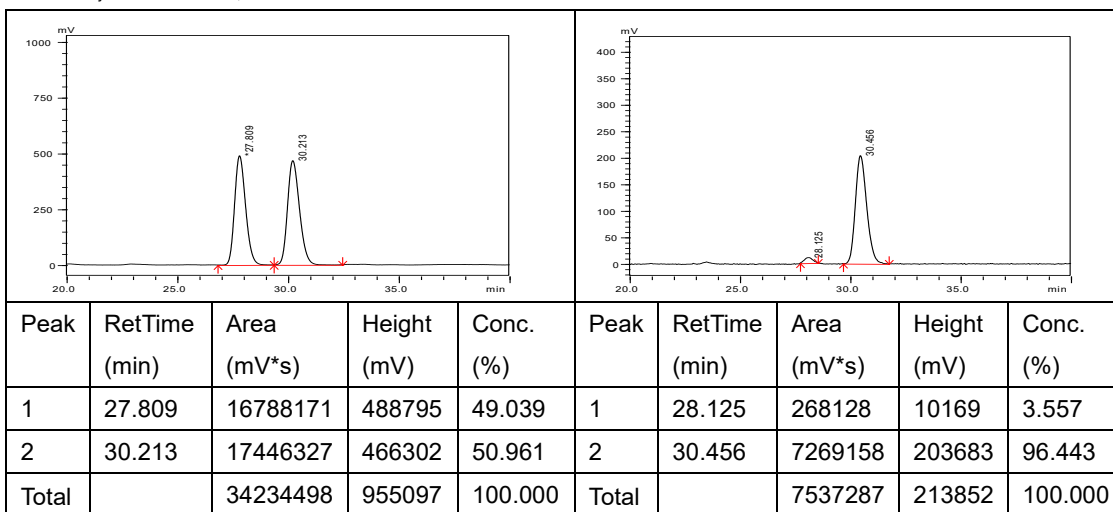
Optical Rotation: $[\alpha]^{25}_D = 31$ (*c* = 1.0, MeOH).



(2R,3S)-2-Isopropyl-3-((4-methoxyphenyl)amino)butane-1,4-diol (**6**)¹²

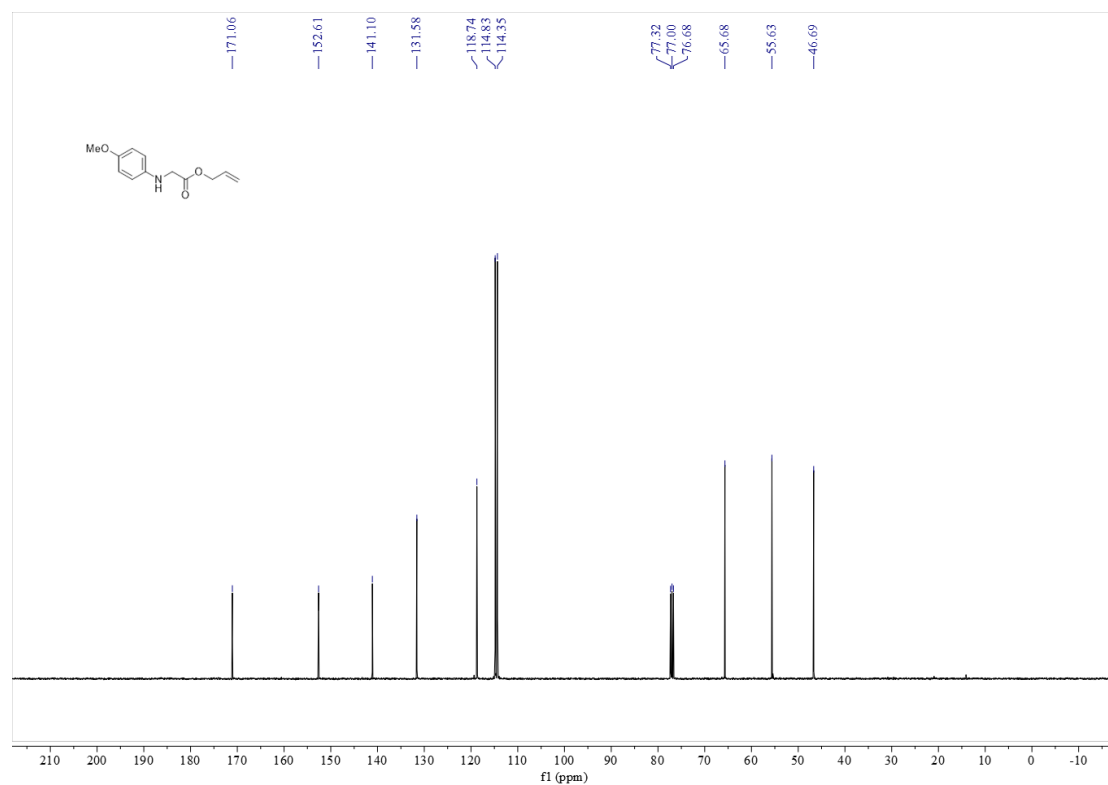
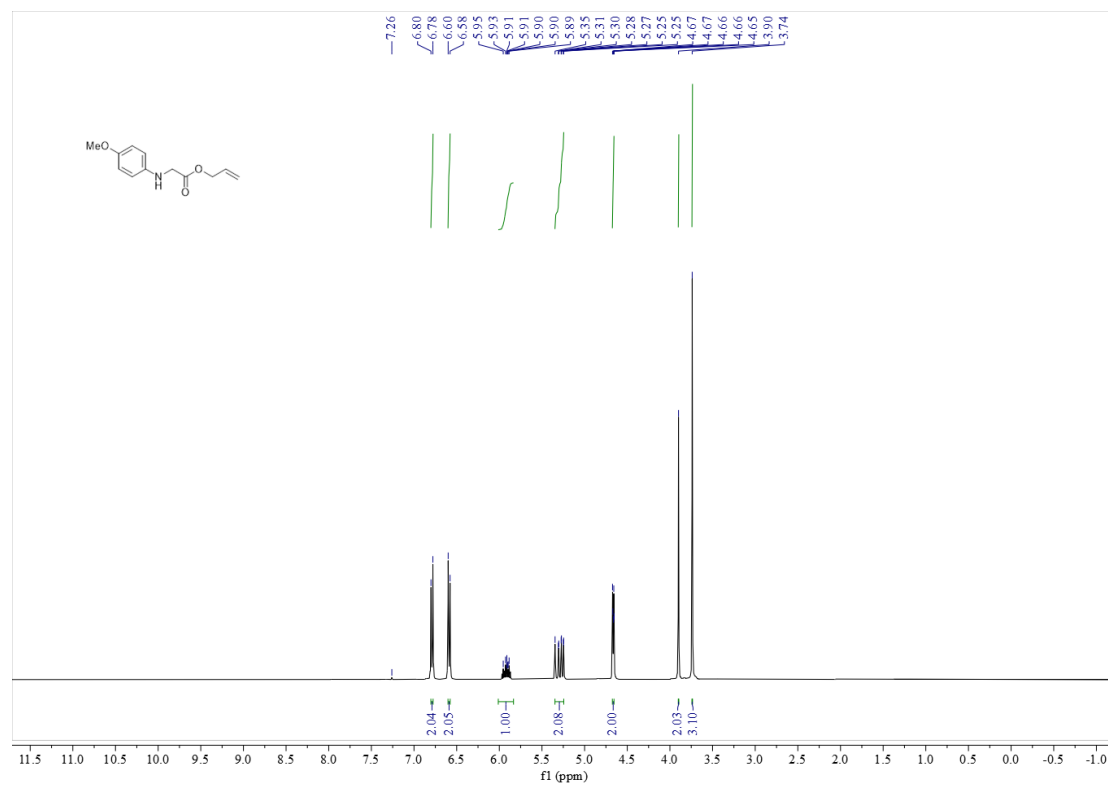
Yellow oil (86 mg, 85% yield); > 99:1 dr. **¹H NMR** (600 MHz, Chloroform-*d*) δ 6.75 (d, *J* = 8.9 Hz, 2H), 6.68 (d, *J* = 8.9 Hz, 2H), 3.83–3.80 (m, 1H), 3.78–3.75 (m, 1H), 3.73 (s, 3H), 3.69 (m, 1H), 3.59 (m, 1H), 3.52 (m, 1H), 1.86 (m, 1H), 1.58 (m, 1H), 0.96 (d, *J* = 6.8 Hz, 3H), 0.89 (d, *J* = 6.9 Hz, 3H). **¹³C NMR** (150 MHz, Chloroform-*d*) δ 152.66, 141.69, 141.67, 116.21, 114.87, 62.47, 61.47, 58.60, 55.66, 47.55, 27.38, 21.20, 18.925.

Optical Rotation: $[\alpha]^{25}_D = 26.3$ (*c* = 1.0, MeOH). The ee value was determined by **HPLC analysis:** 96.4:3.6 er, Chiralcel AD-H column, hexane/*i*-PrOH = 93/7, flow rate = 0.5 mL/min, λ = 254 nm, retention time: *t*_{major} = 30.5 min, *t*_{minor} = 28.1 min.

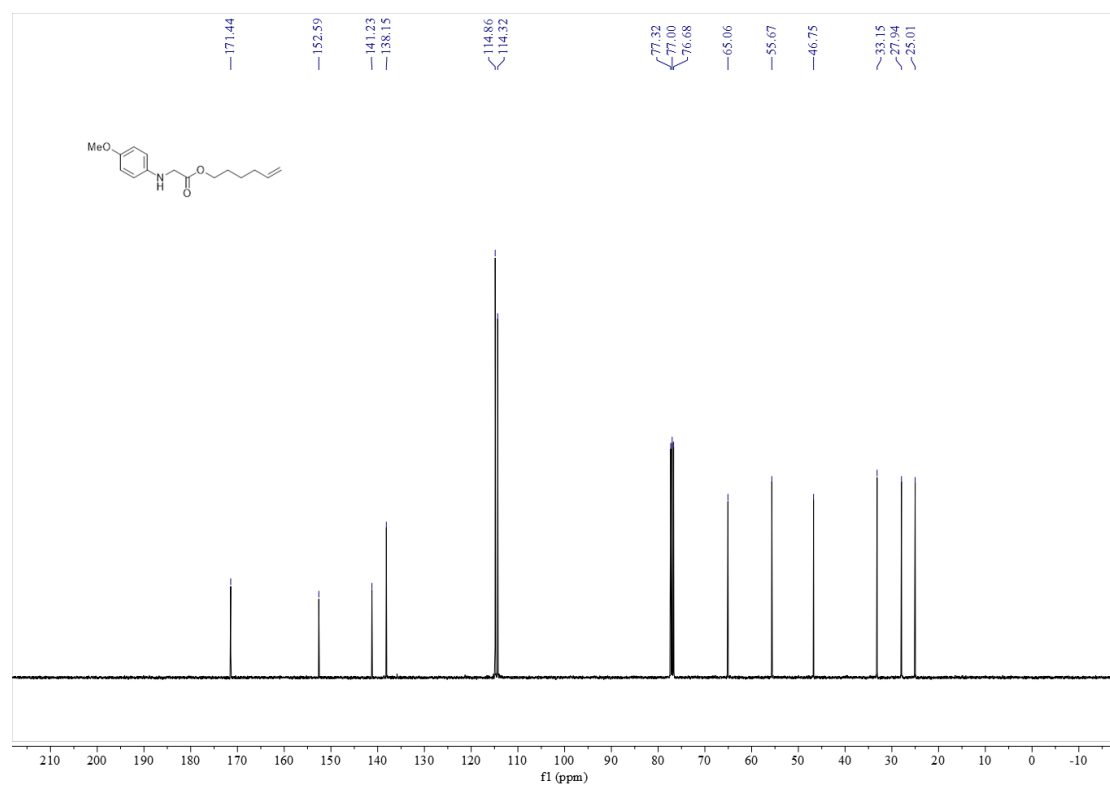
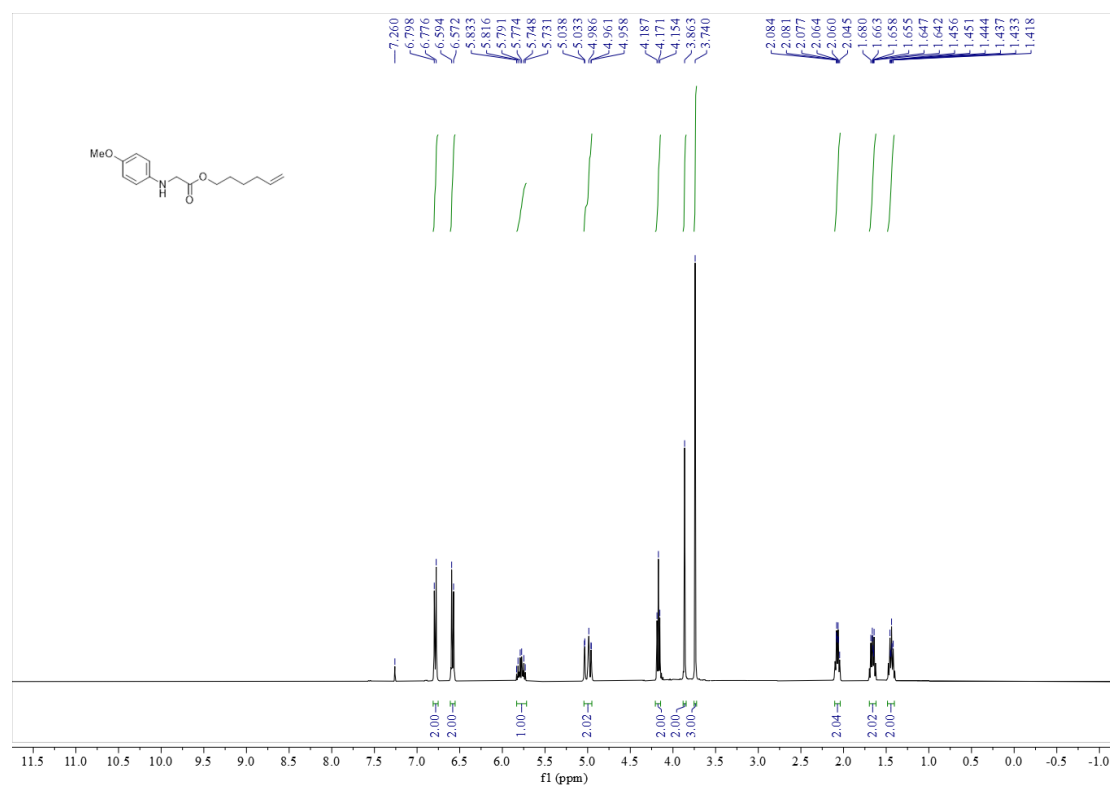


5. NMR spectra

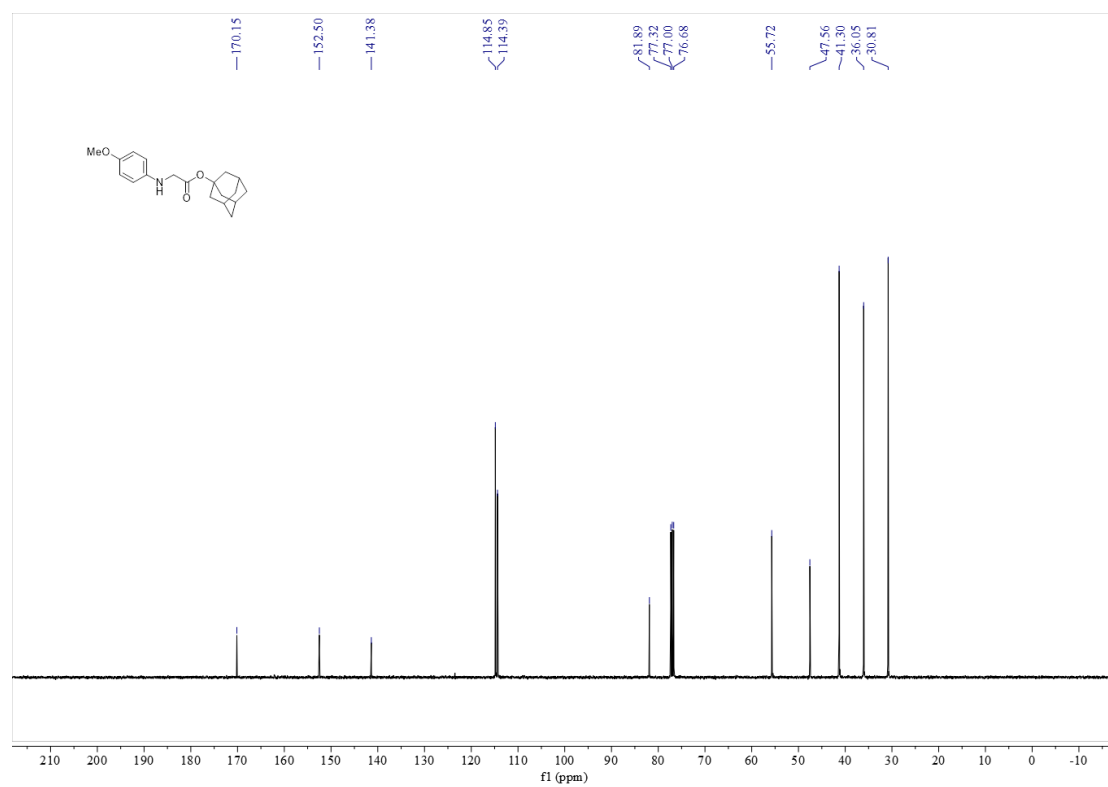
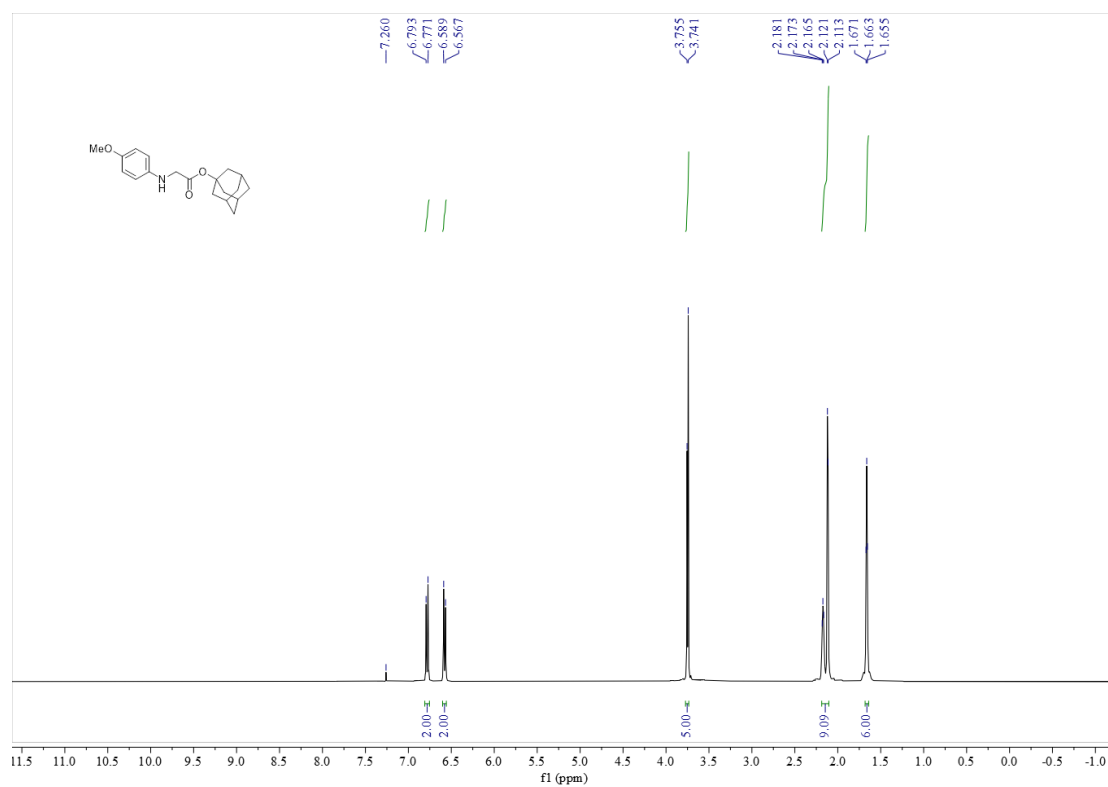
Allyl (4-methoxyphenyl)glycinate (**1e**)



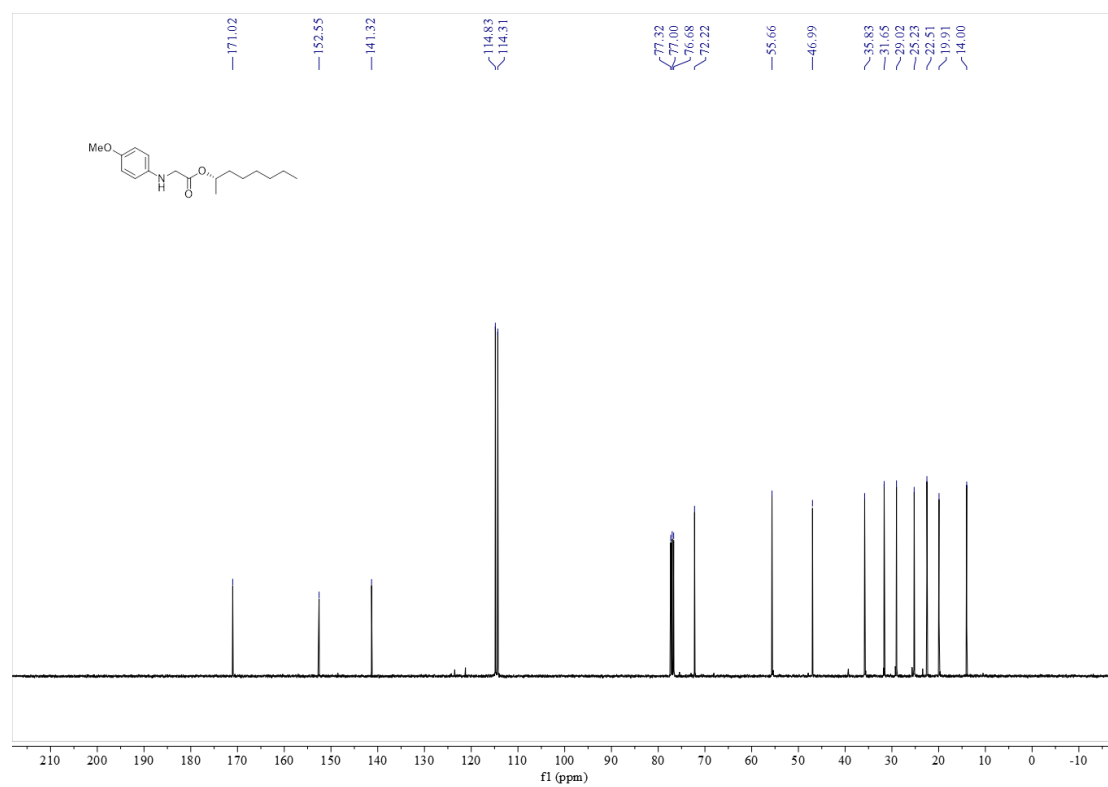
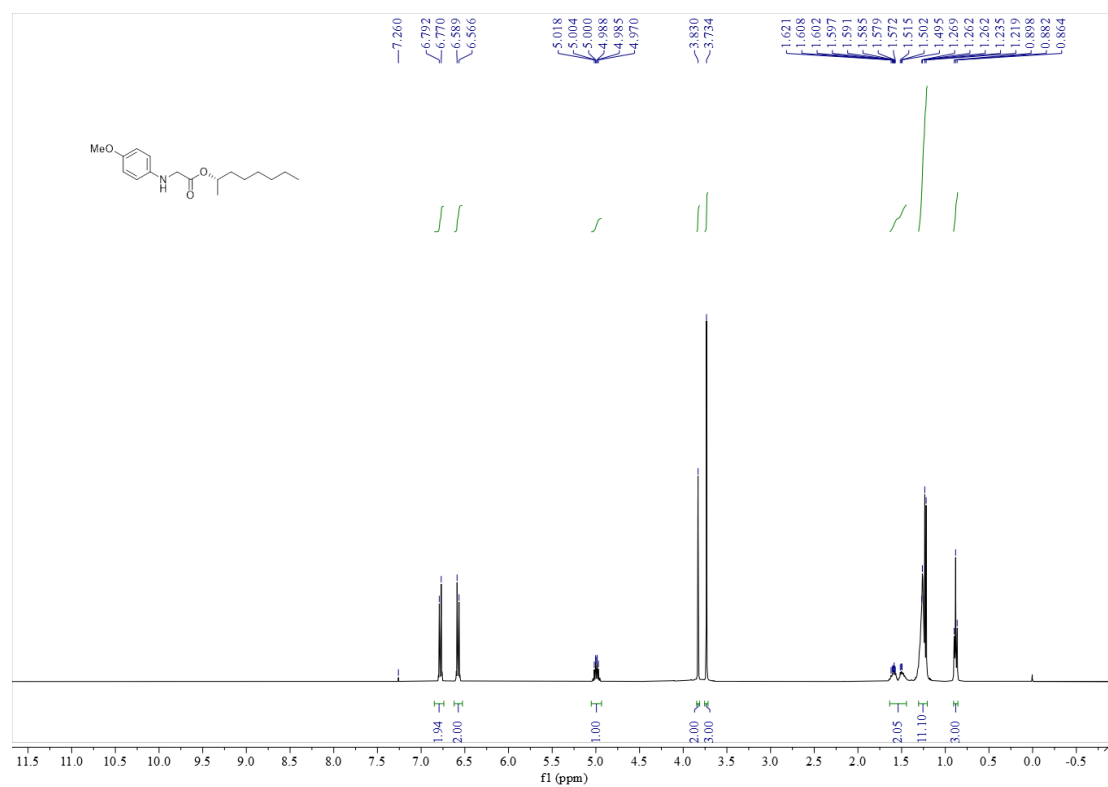
Hex-5-en-1-yl (4-methoxyphenyl)glycinate (**1f**)



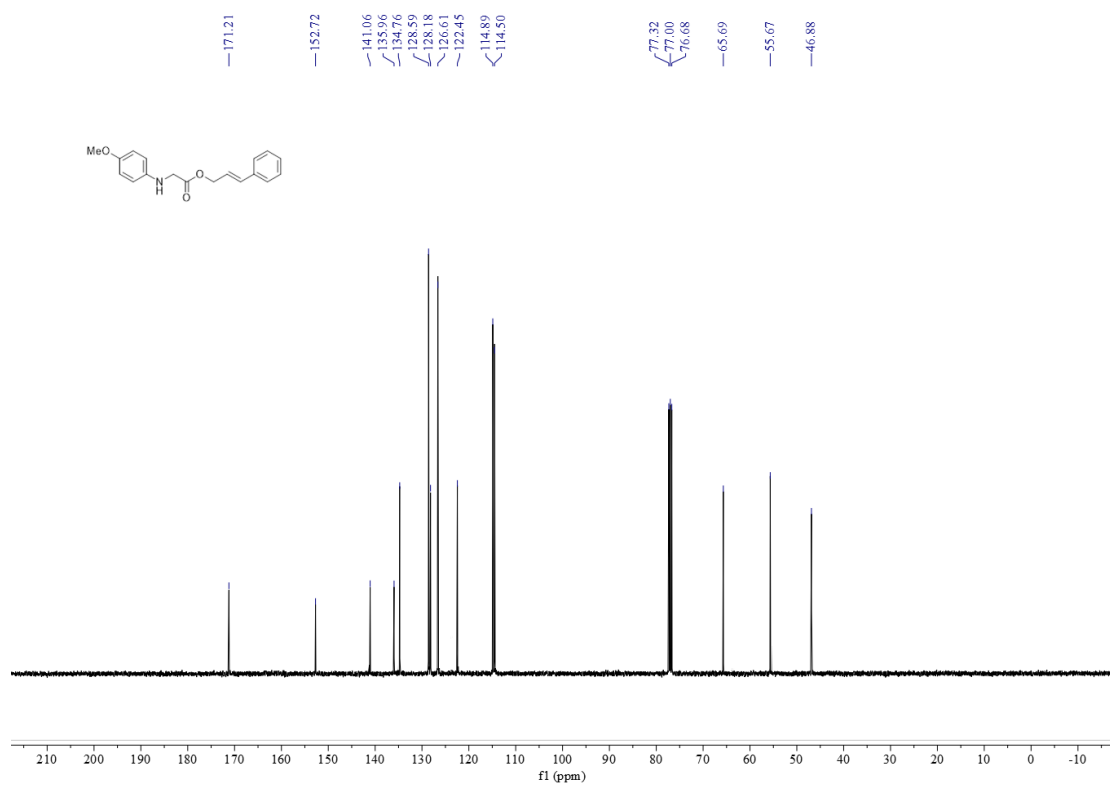
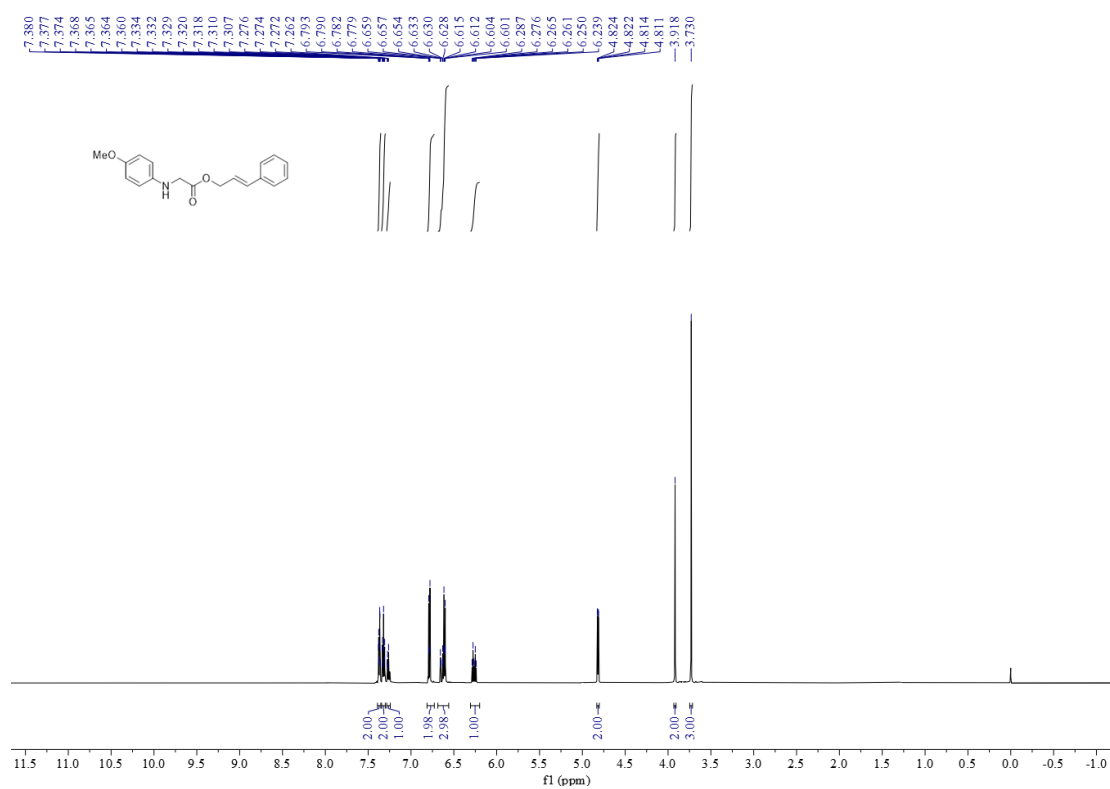
(1S,3S)-Adamantan-1-yl (4-methoxyphenyl)glycinate (**1h**)



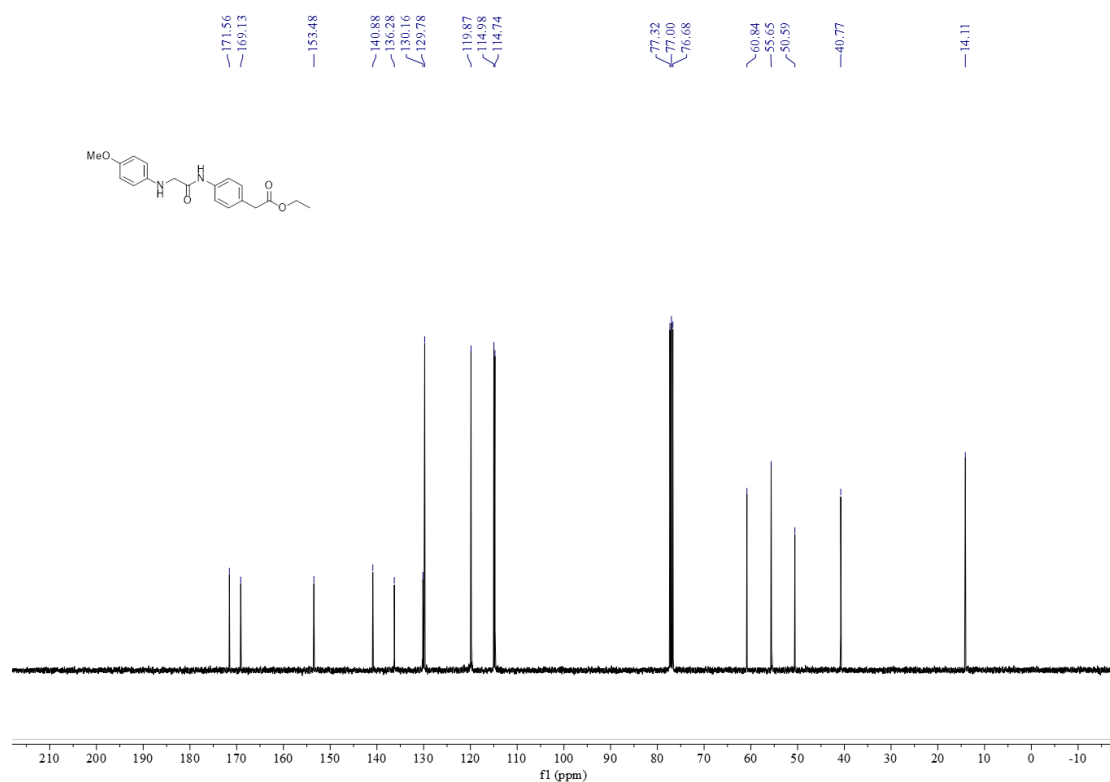
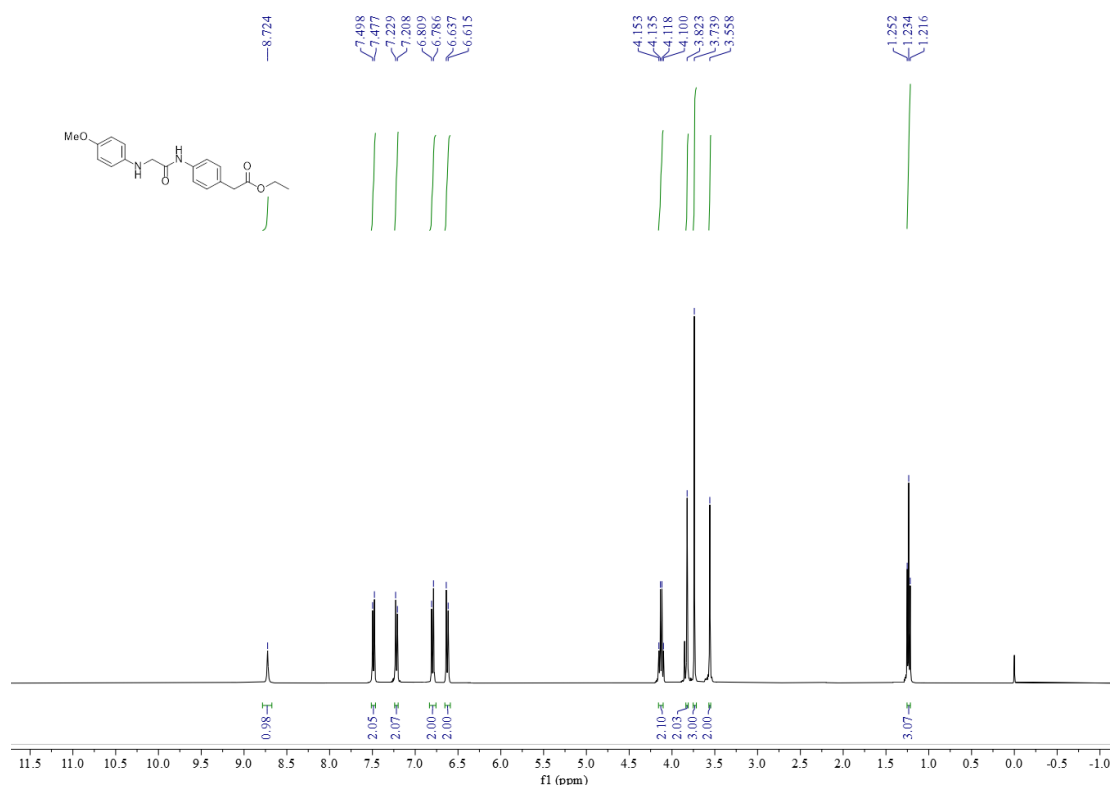
(S)-octan-2-yl (4-methoxyphenyl)glycinate (**1i**)



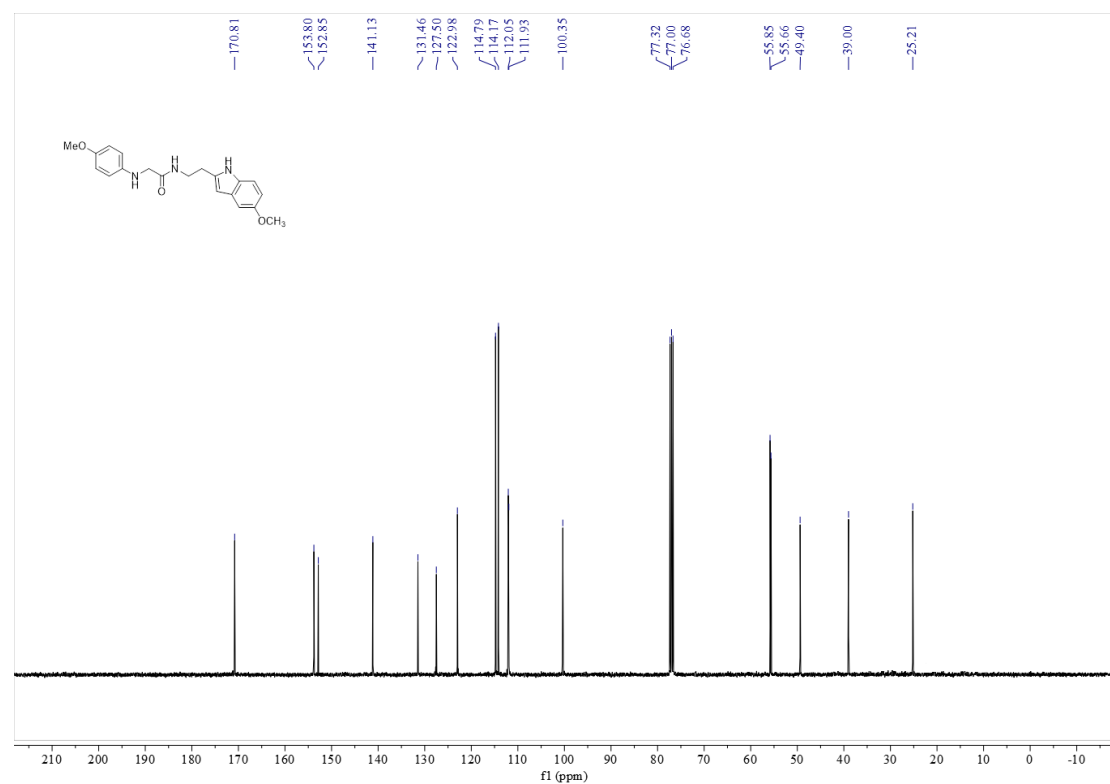
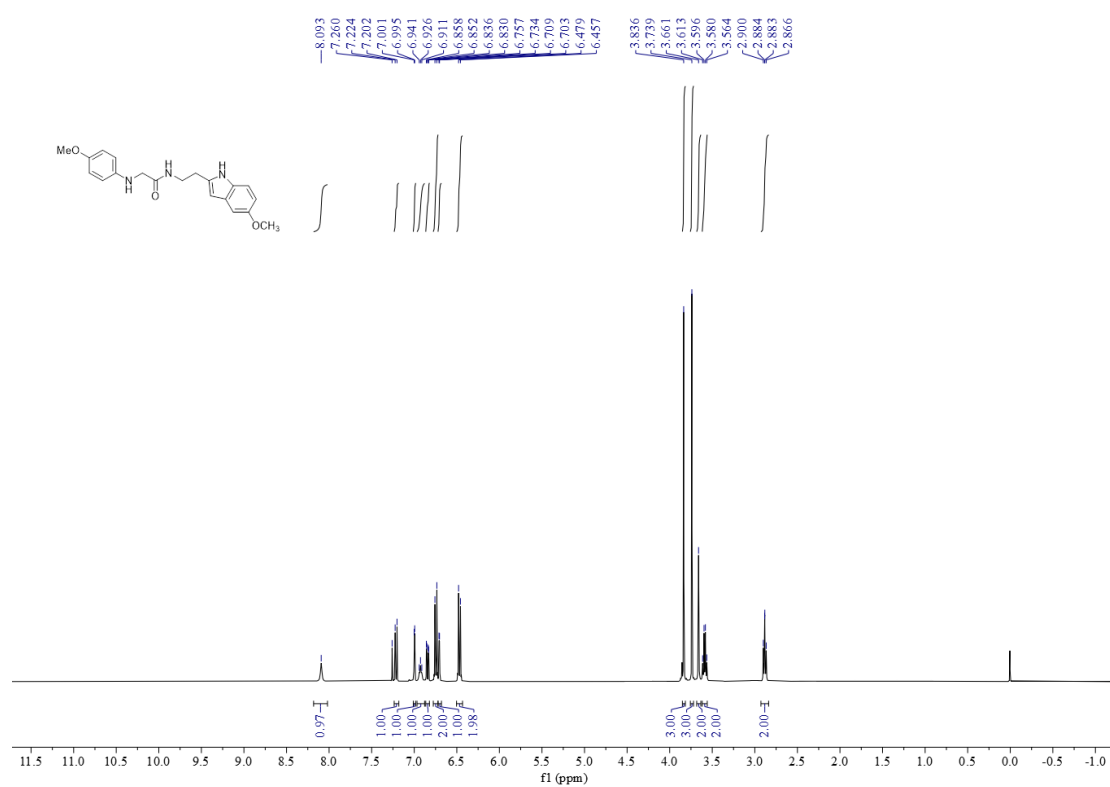
Cinnamyl (4-methoxyphenyl)glycinate (**1n**)



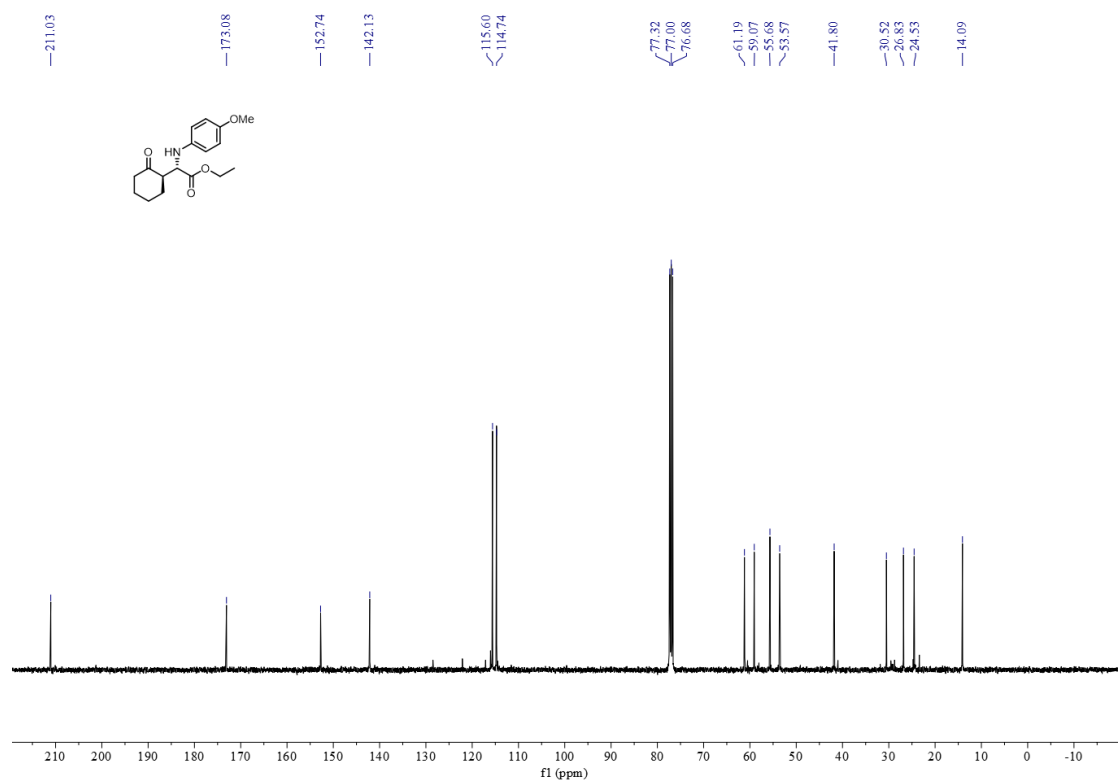
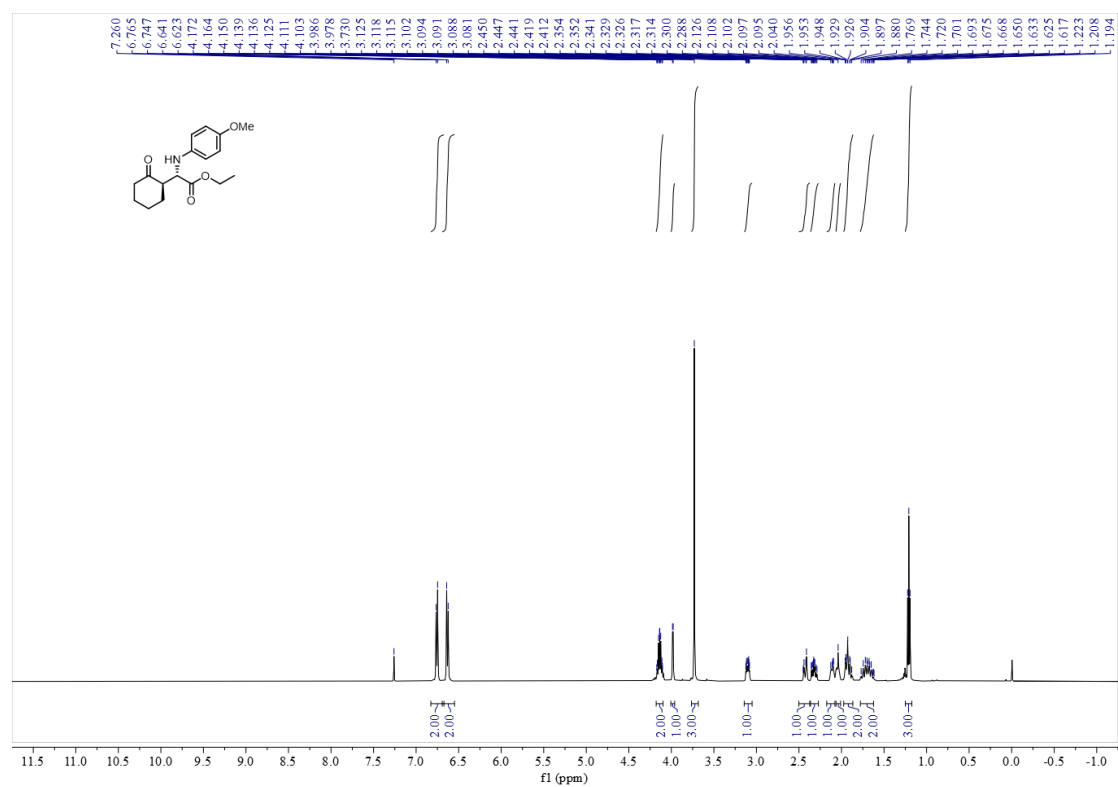
Ethyl 2-(4-(2-((4-methoxyphenyl)amino)acetamido)phenyl)acetate (**1r**)



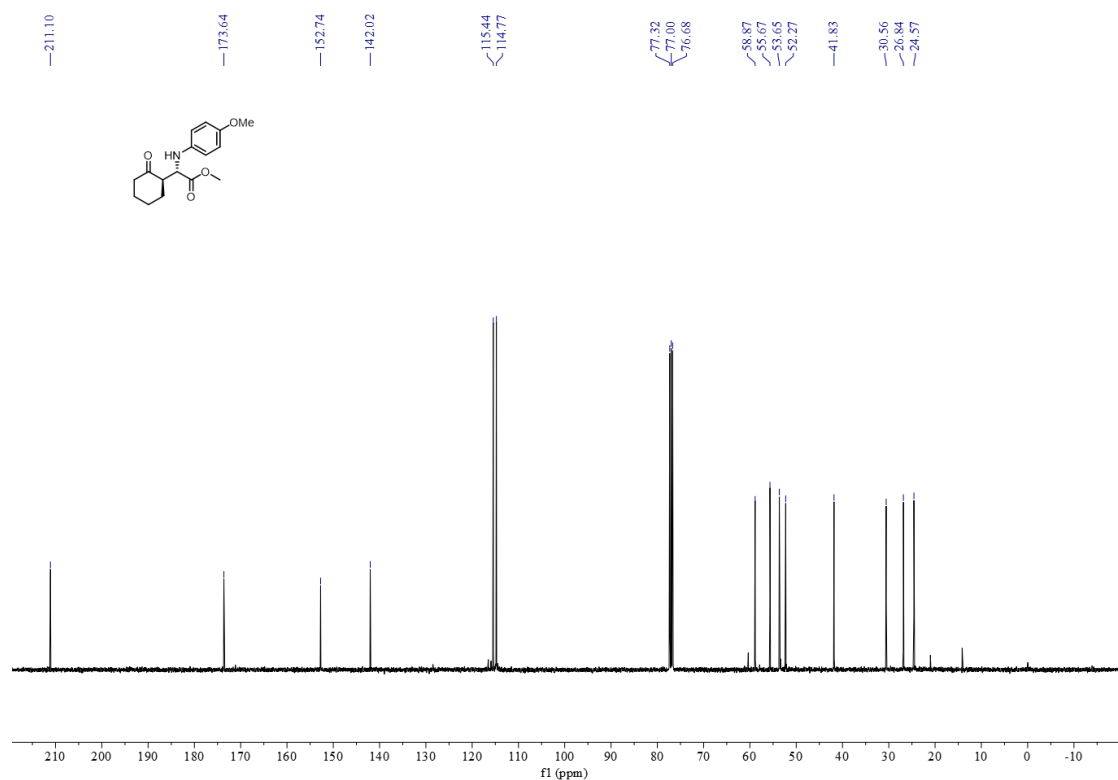
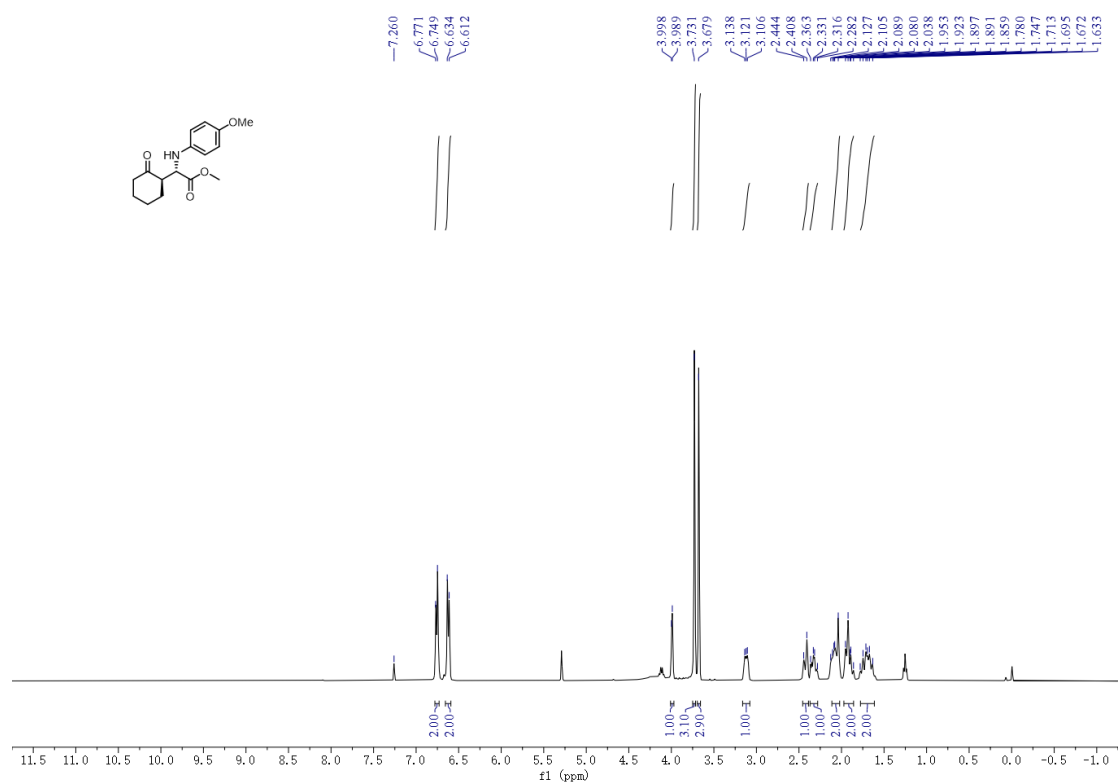
N-(2-(5-methoxy-1H-indol-2-yl)ethyl)ethanamide (1t)



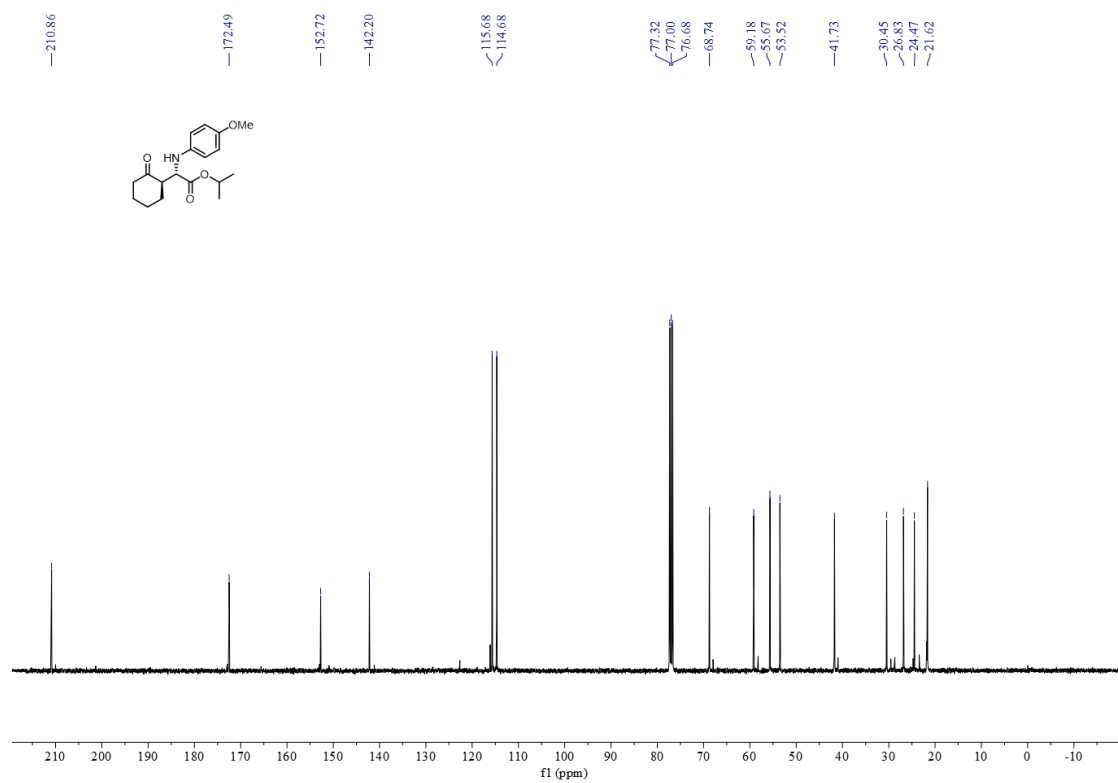
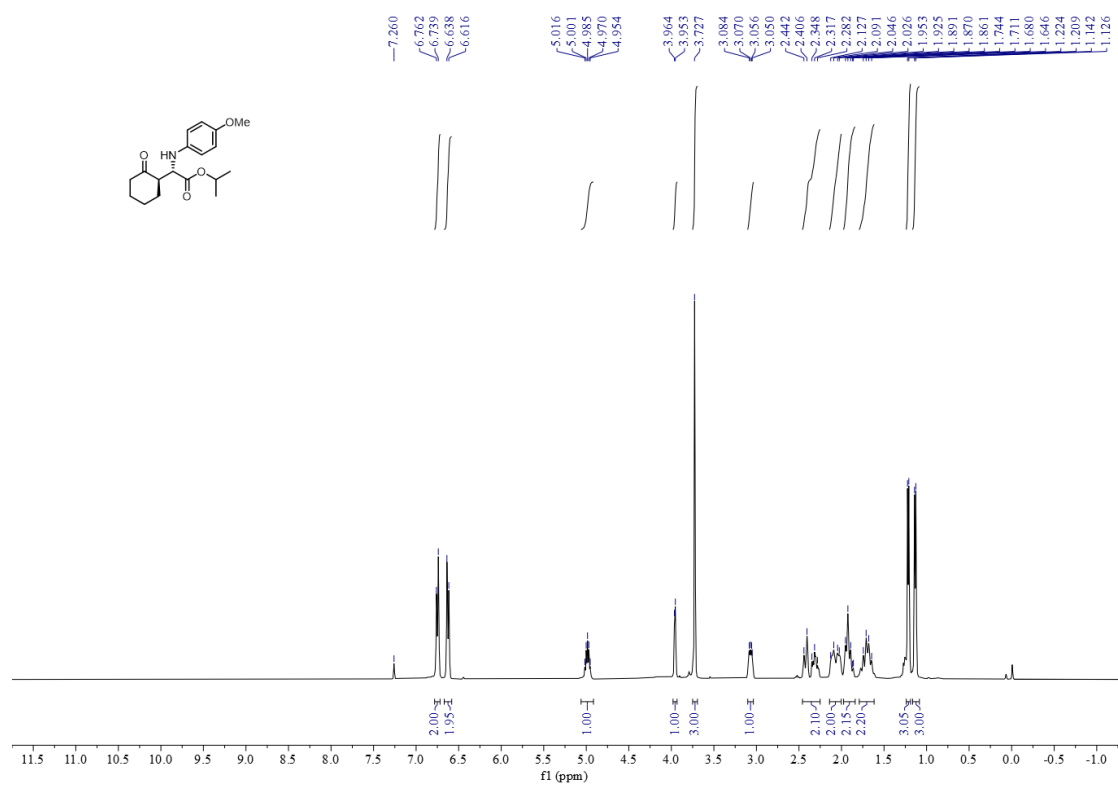
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3aa**)



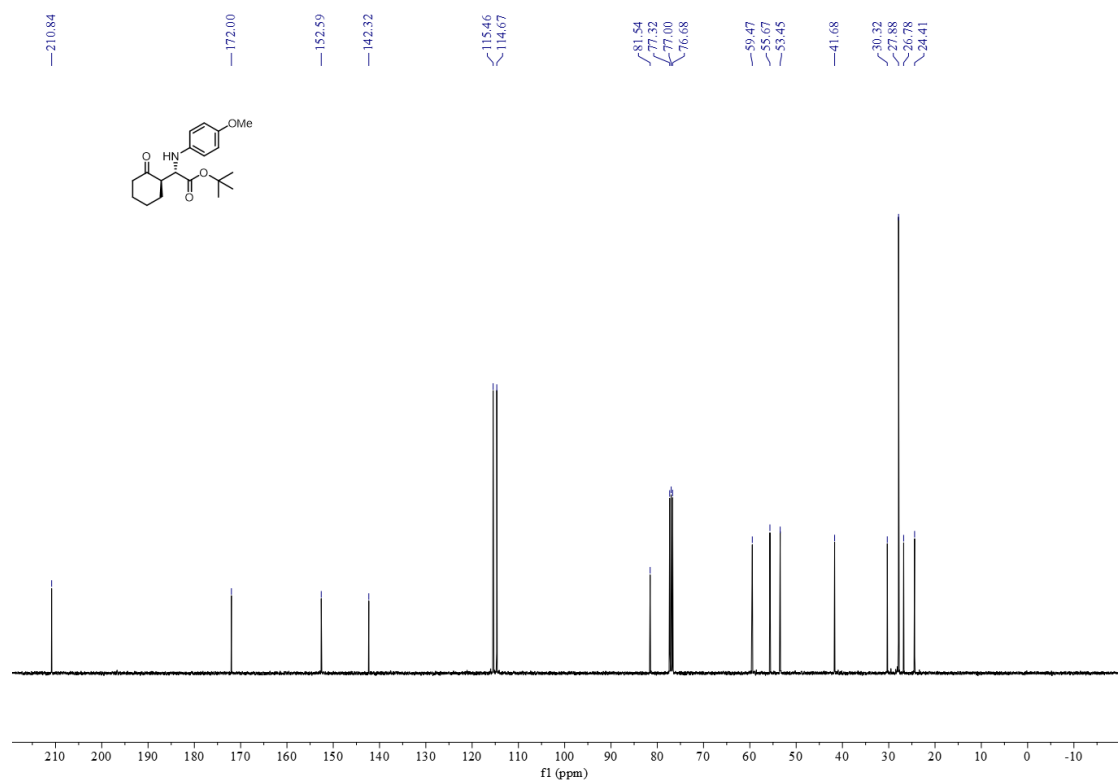
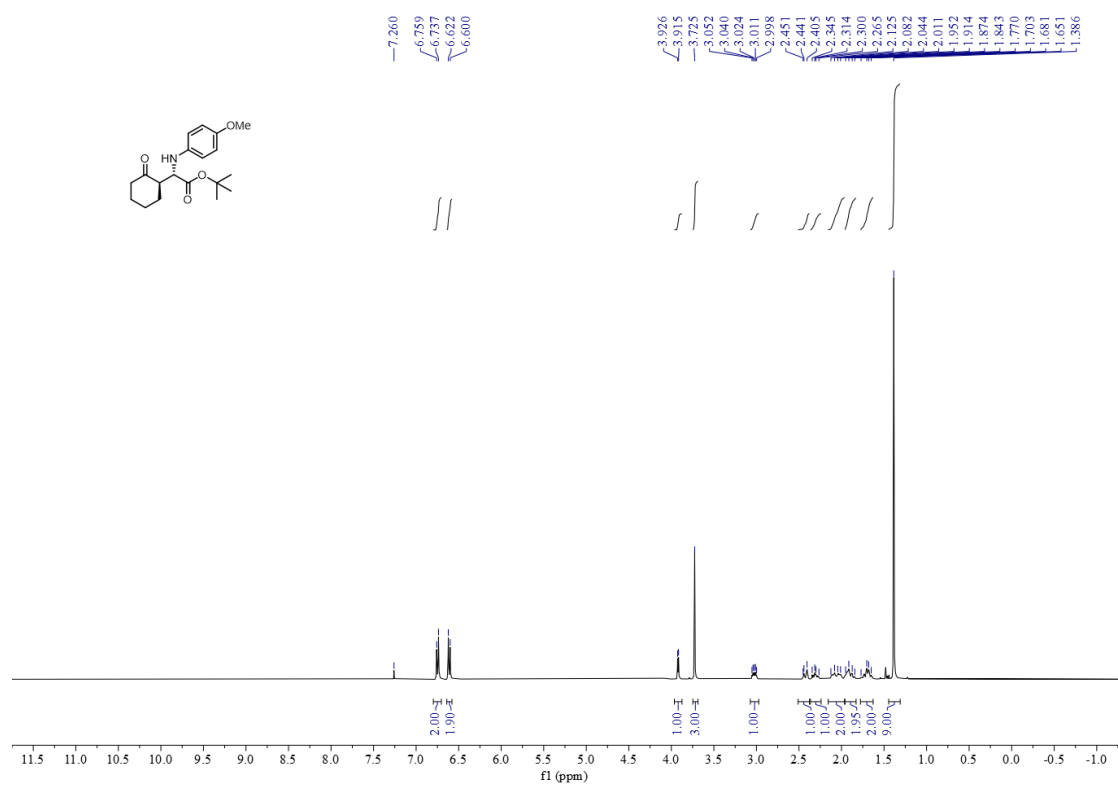
Methyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ba**)



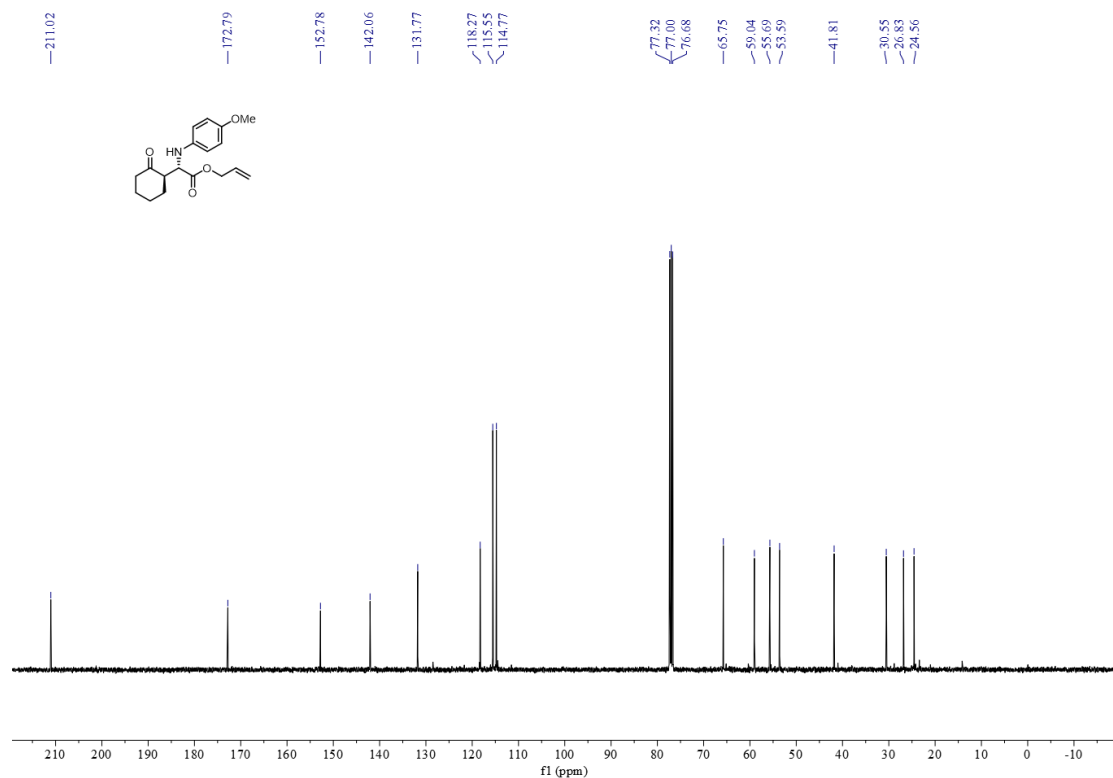
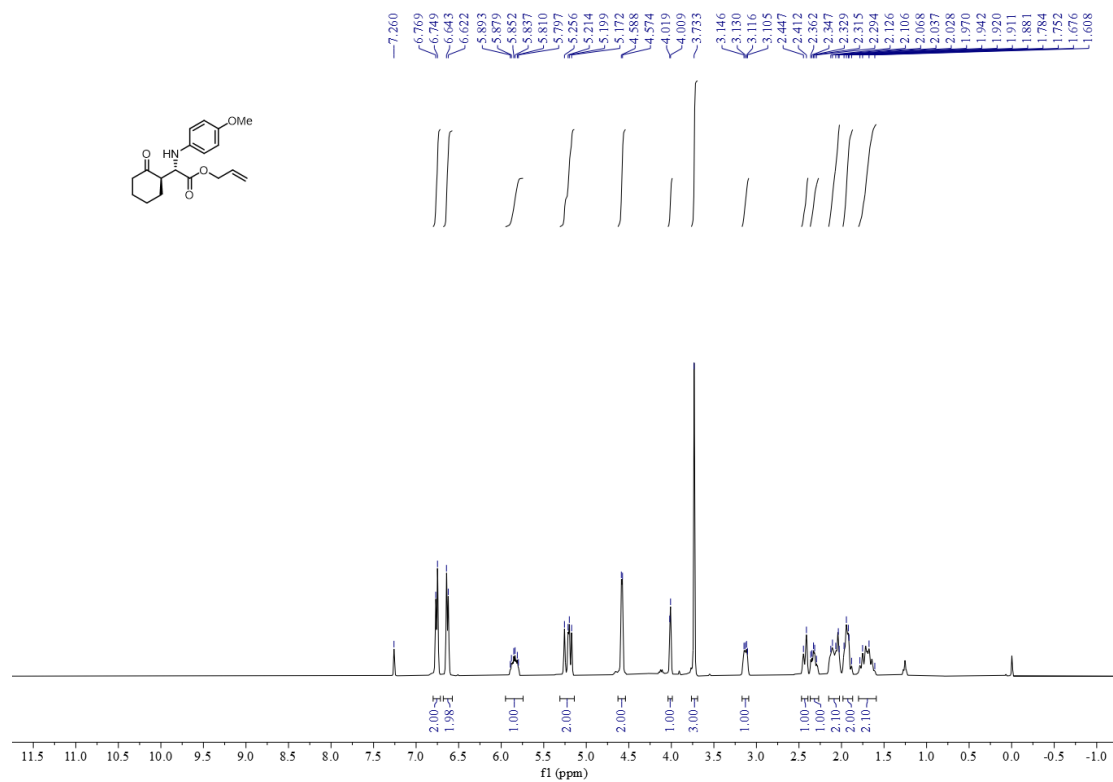
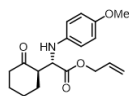
Isopropyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ca**)



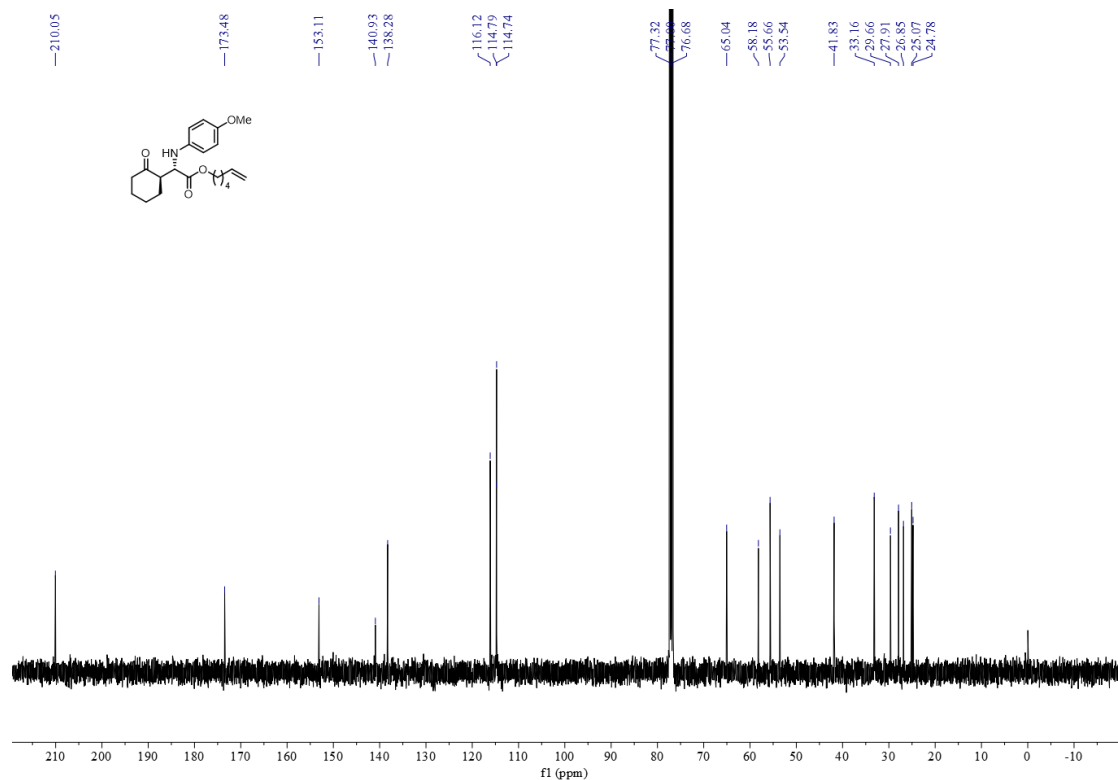
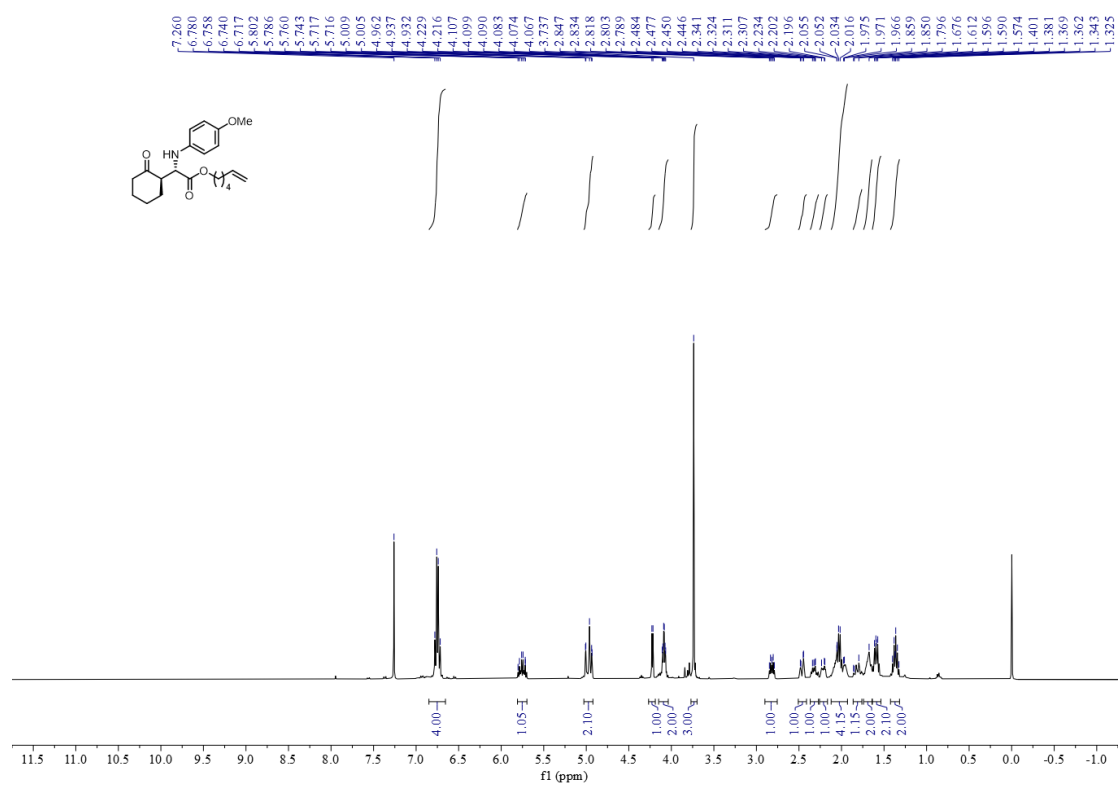
Tert-butyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3da**)



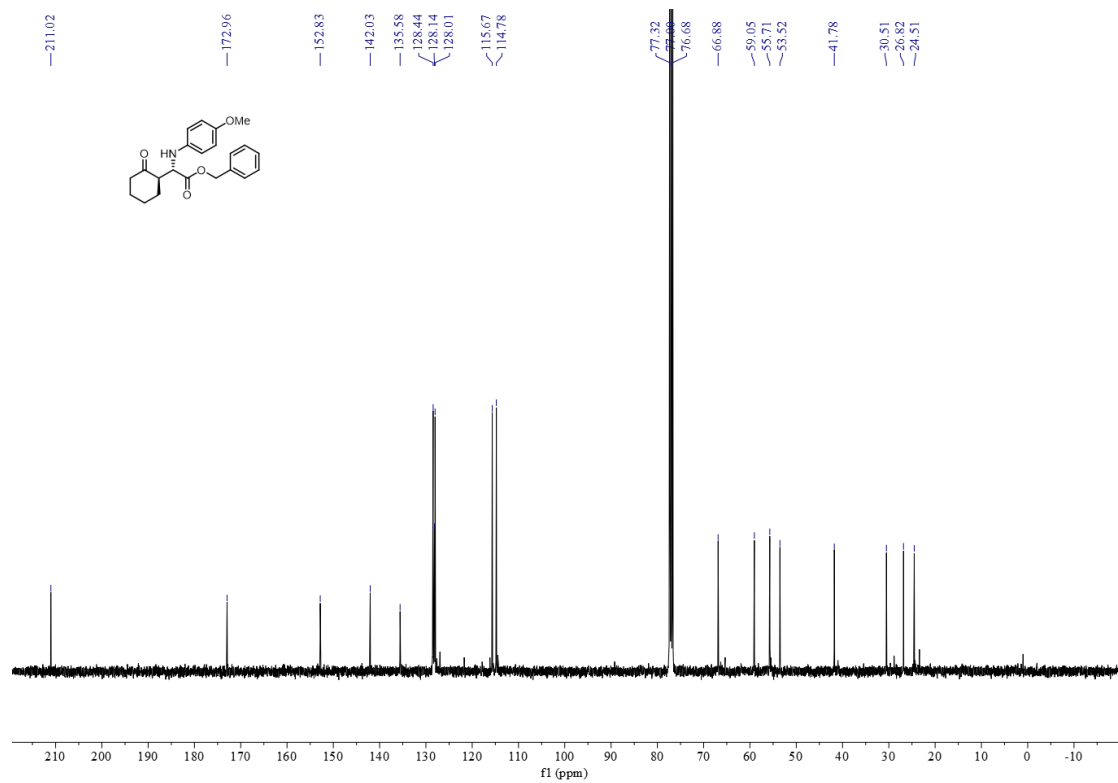
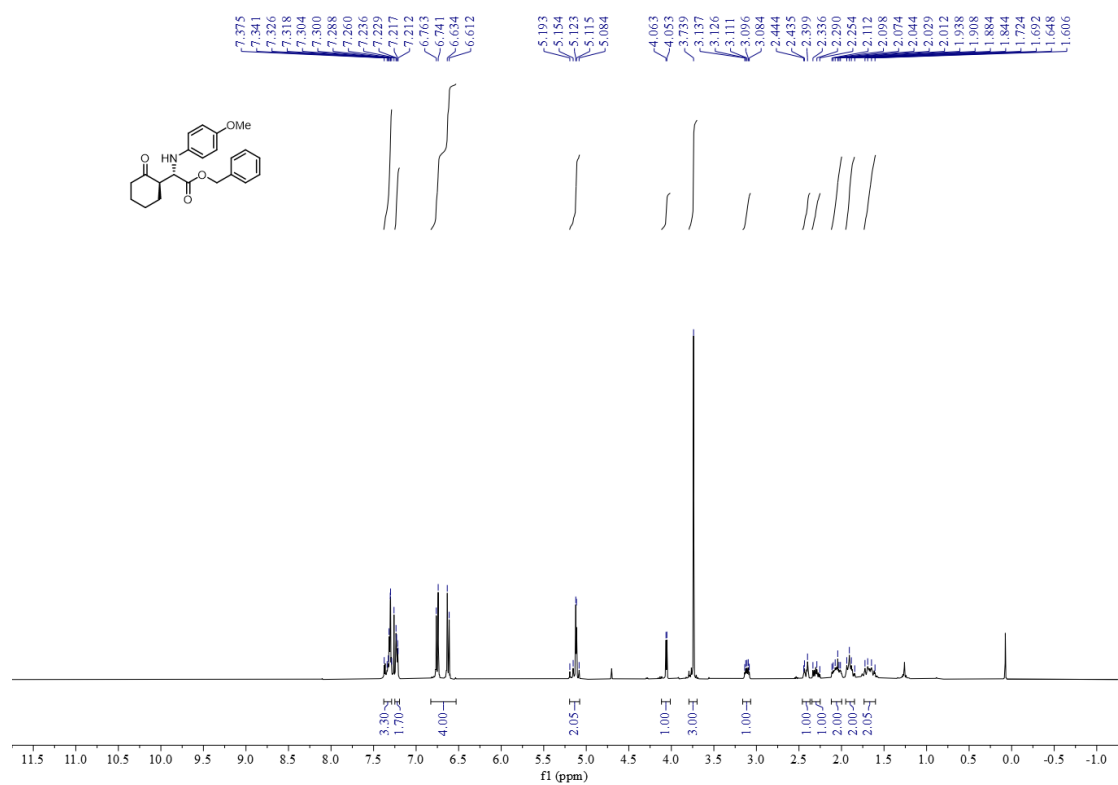
Allyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ea**)



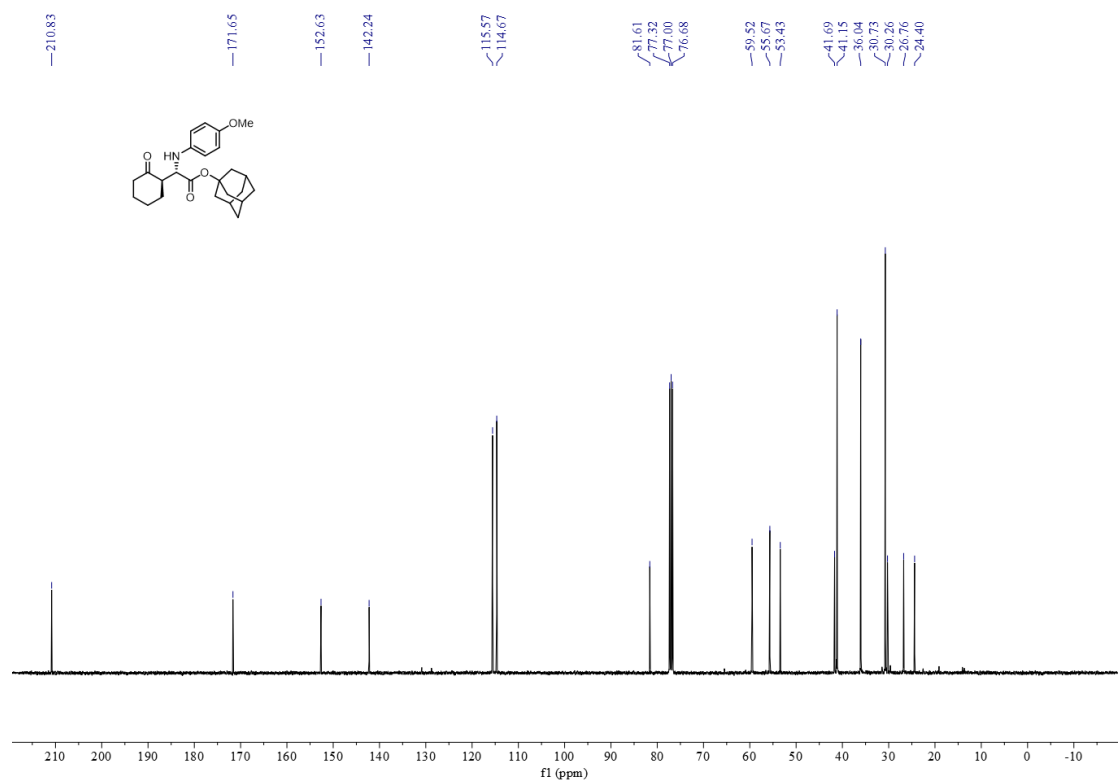
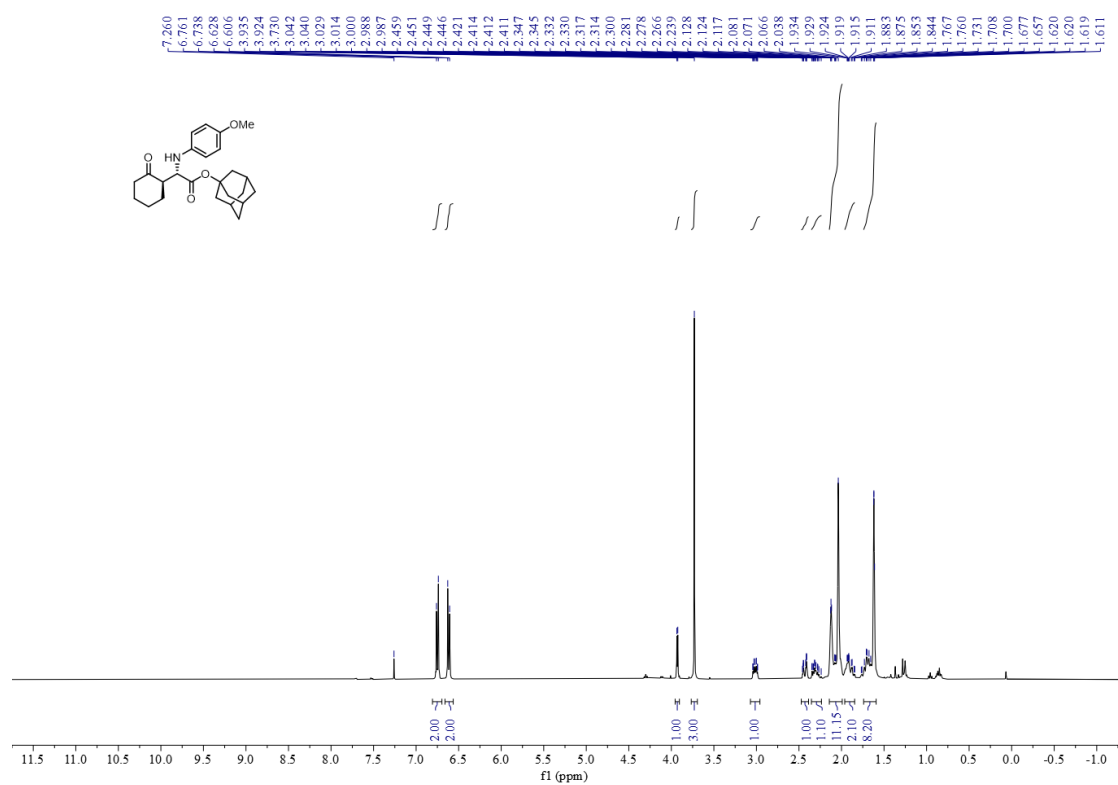
Allyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3fa**)



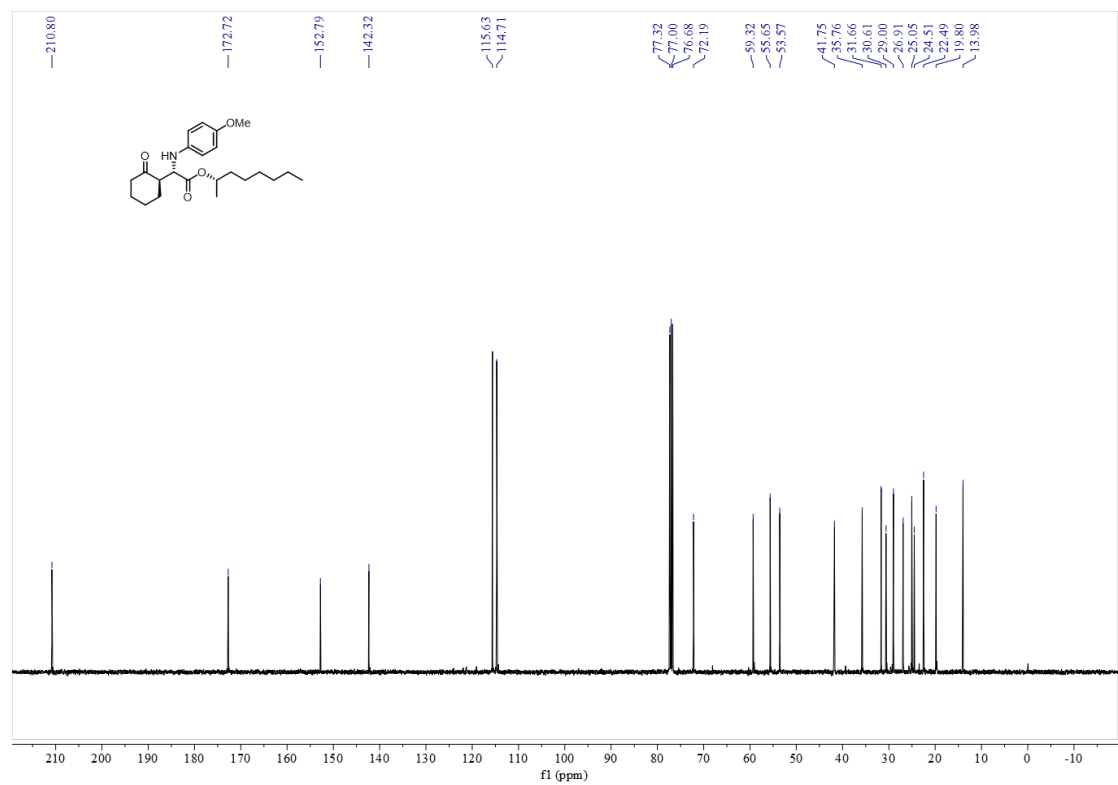
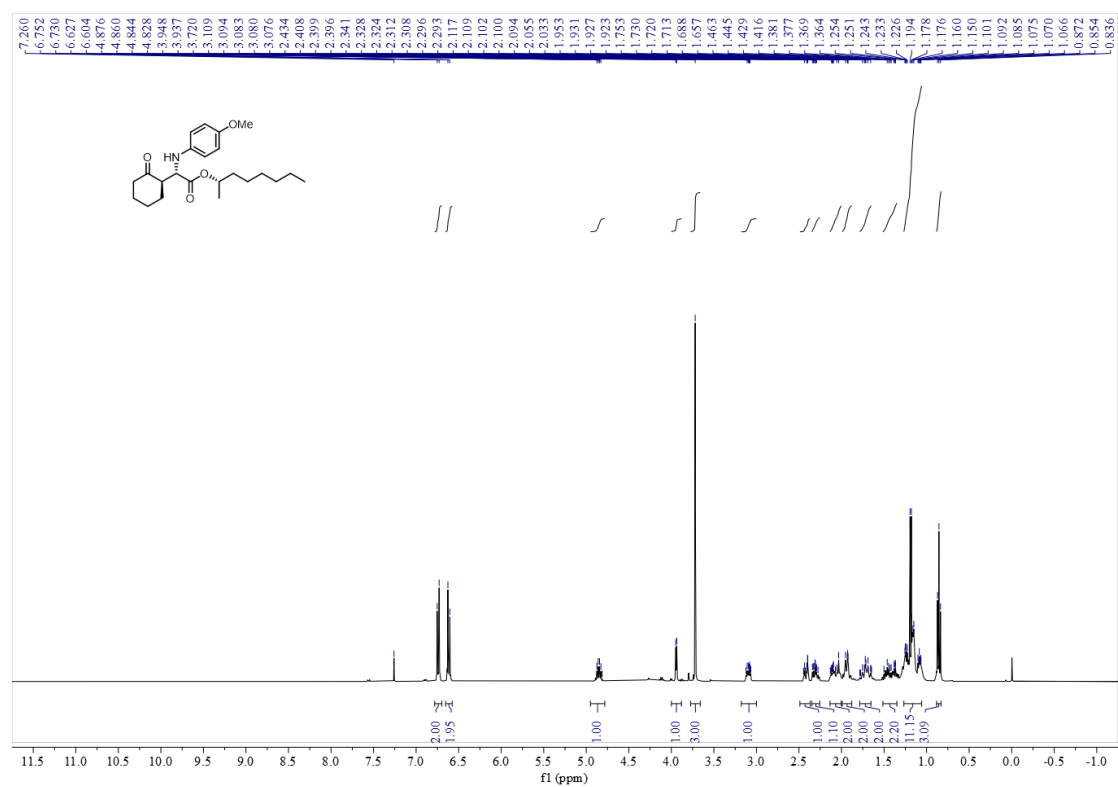
Benzyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ga**)



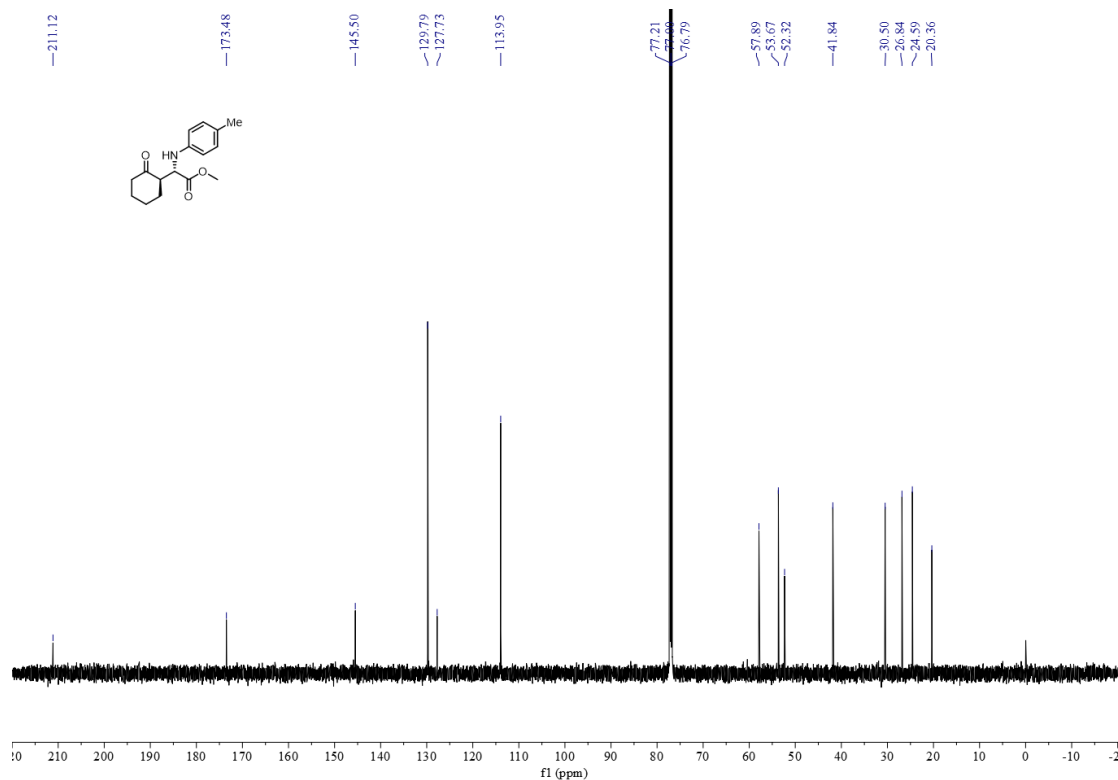
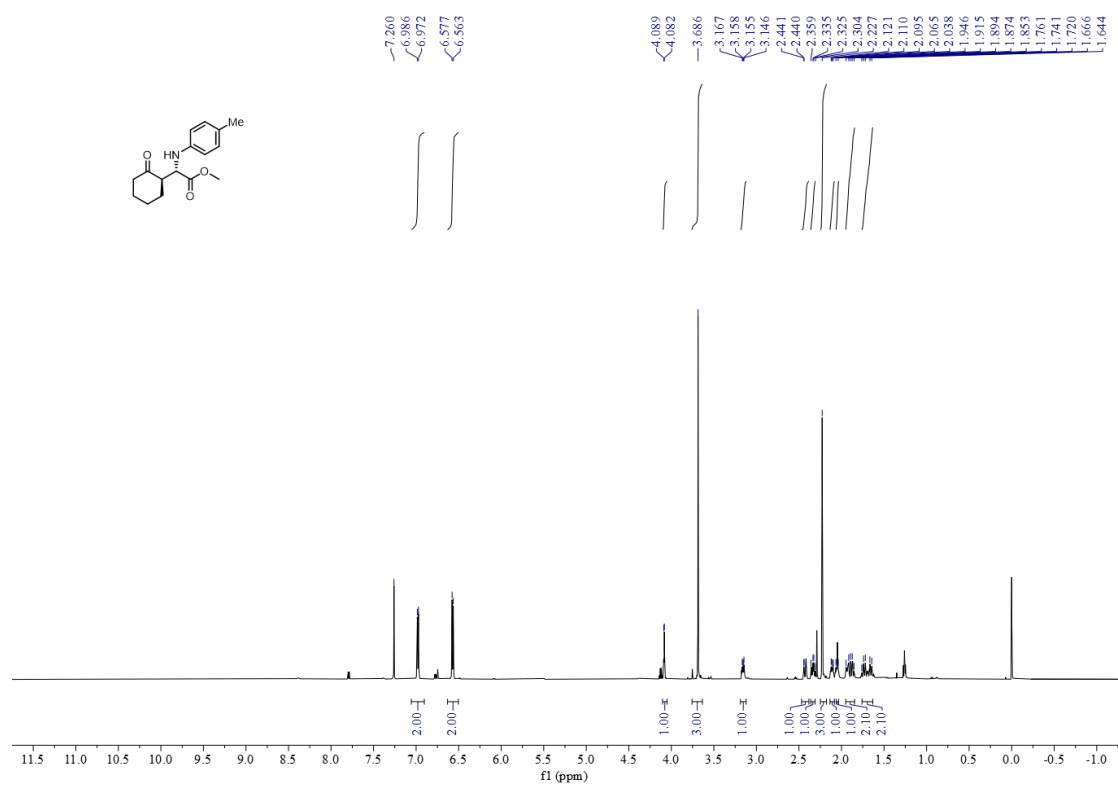
(1S,3R)-Adamantan-1-yl (2S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)aceta-te (**3ha**)



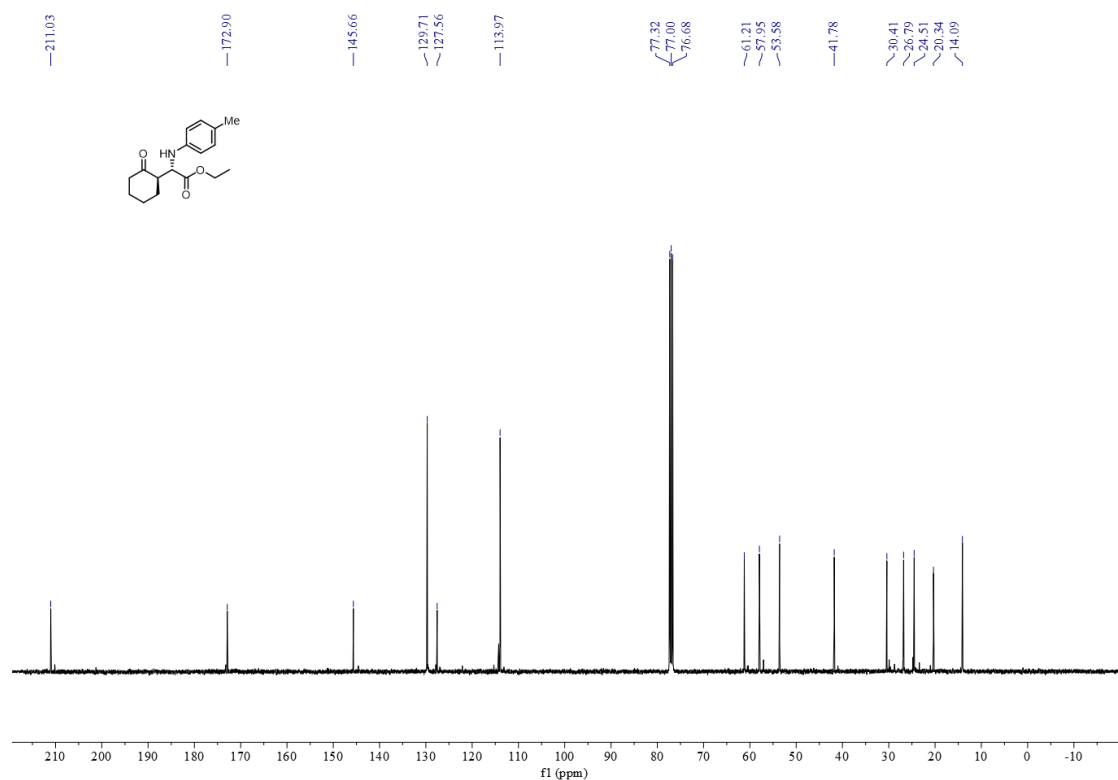
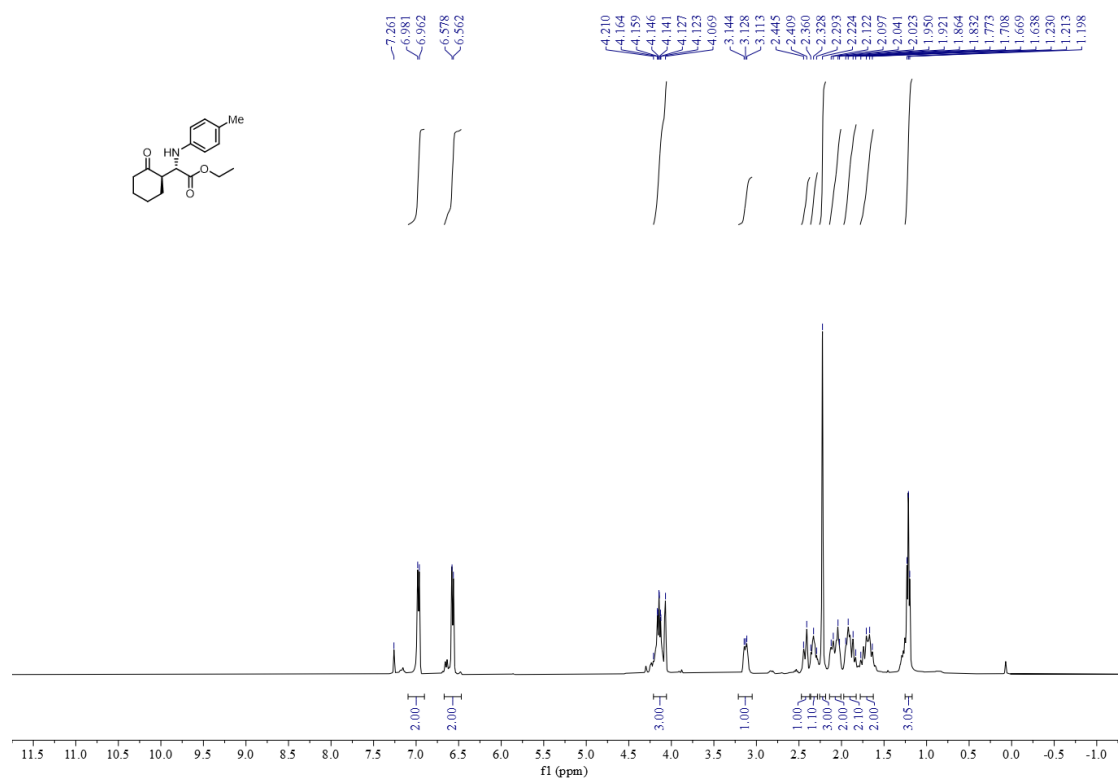
(S)-Octan-2-yl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ia**)



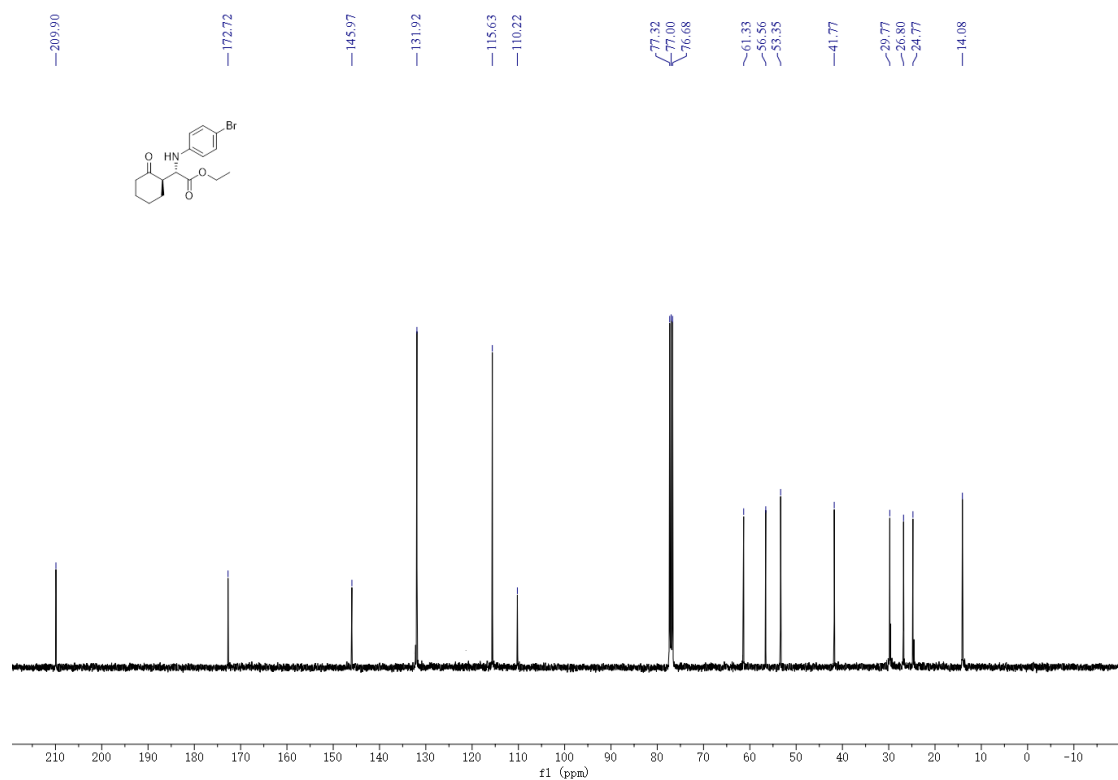
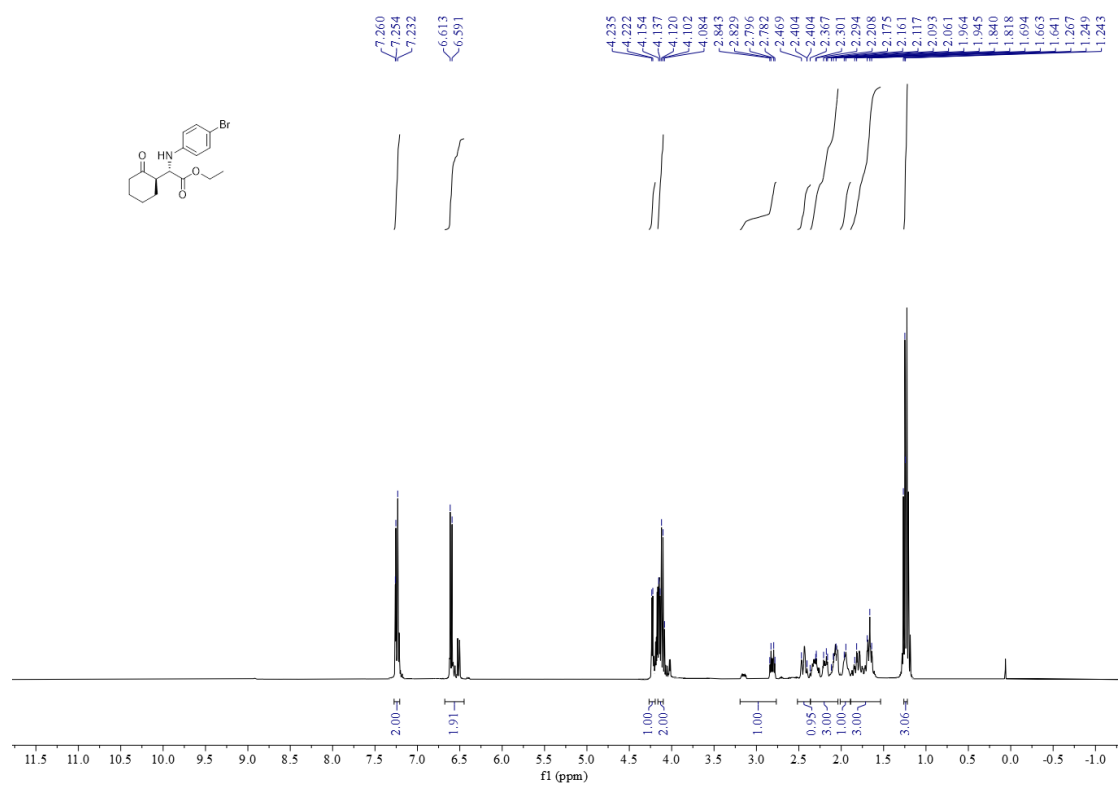
Methyl (S)-2-((R)-2-oxocyclohexyl)-2-(p-tolylamino)acetate (**3ja**)



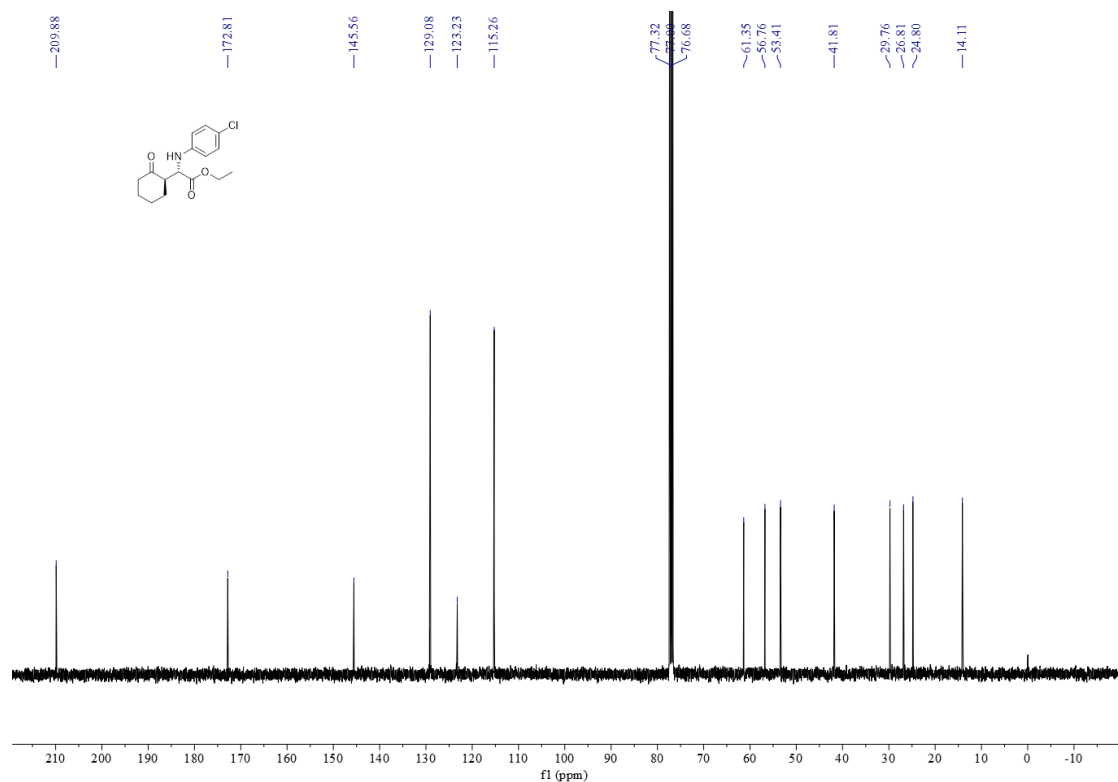
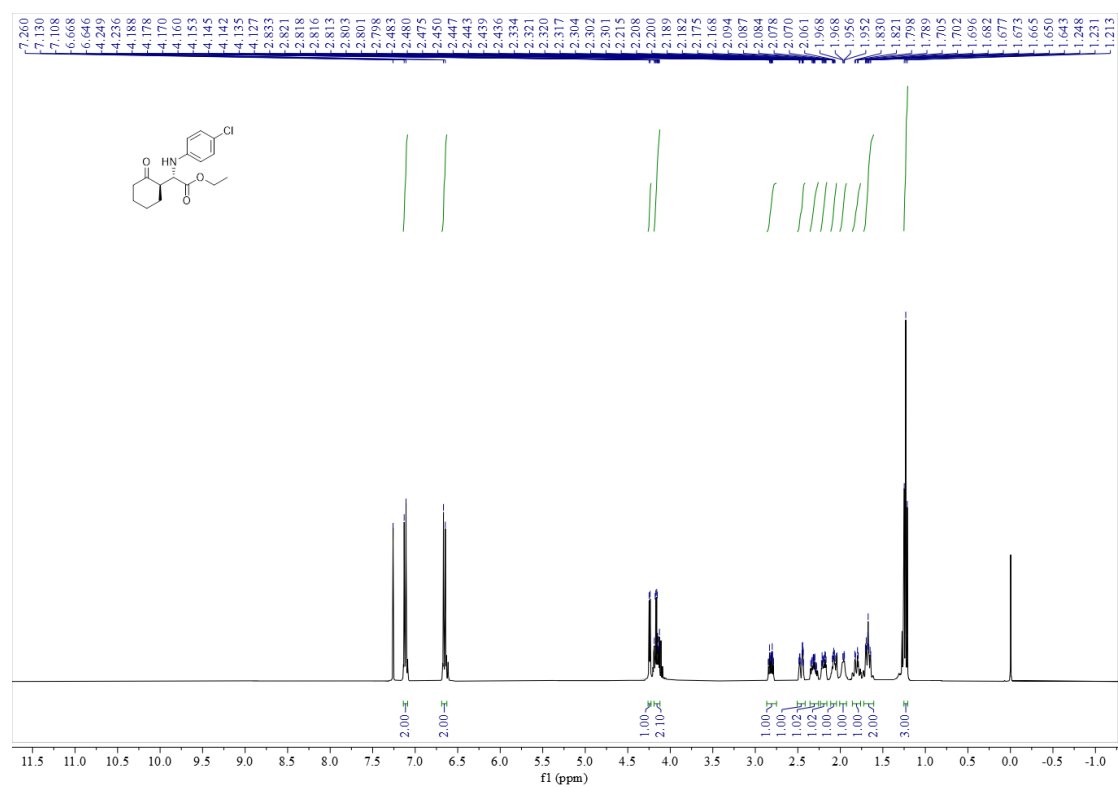
Ethyl (S)-2-((R)-2-oxocyclohexyl)-2-(p-tolylamino)acetate (**3ka**)



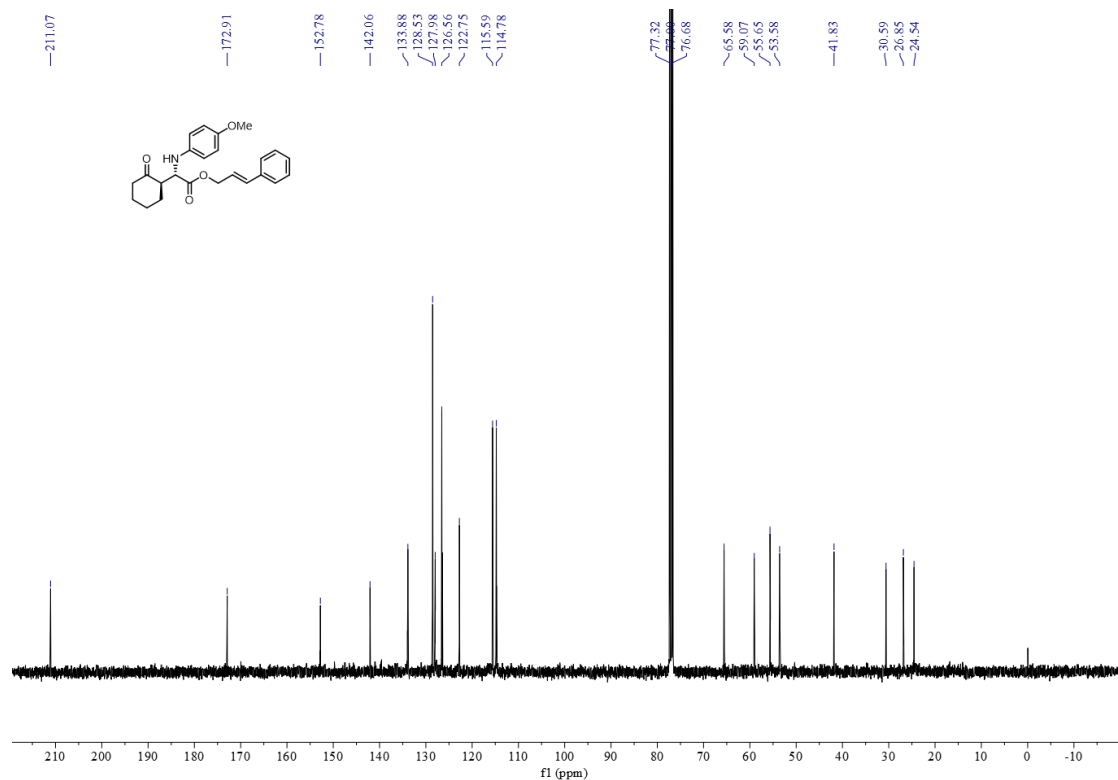
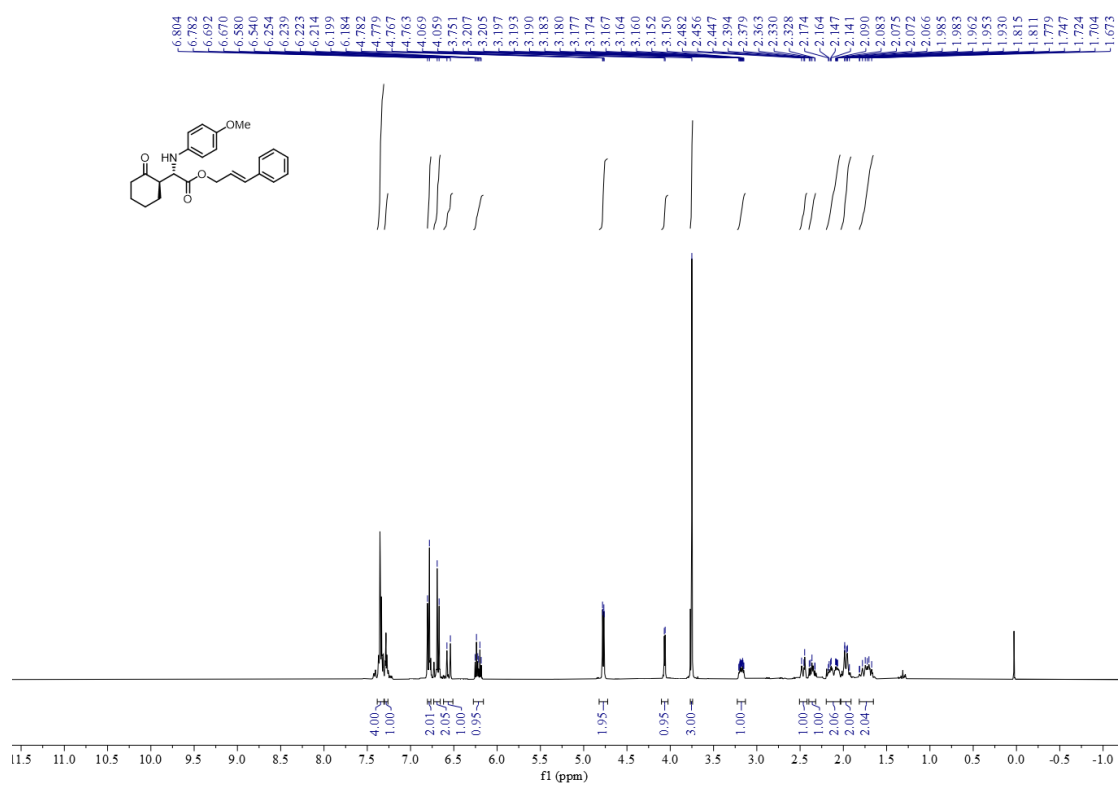
Ethyl (S)-2-((4-bromophenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**31a**)



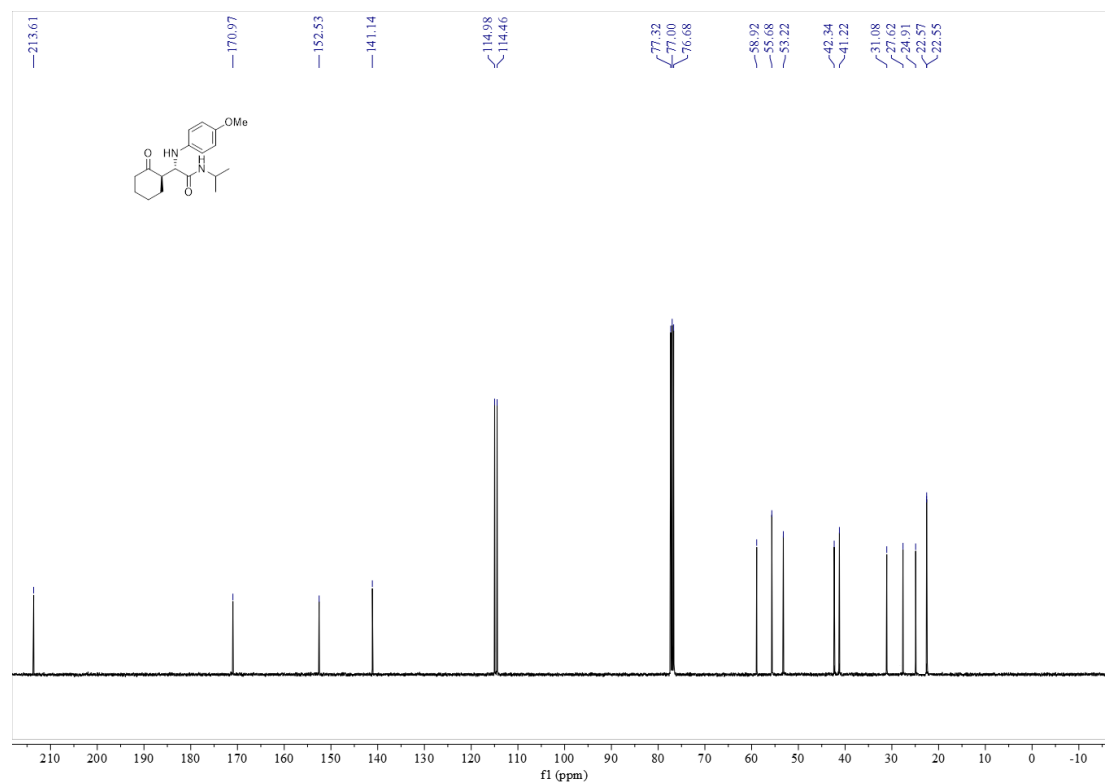
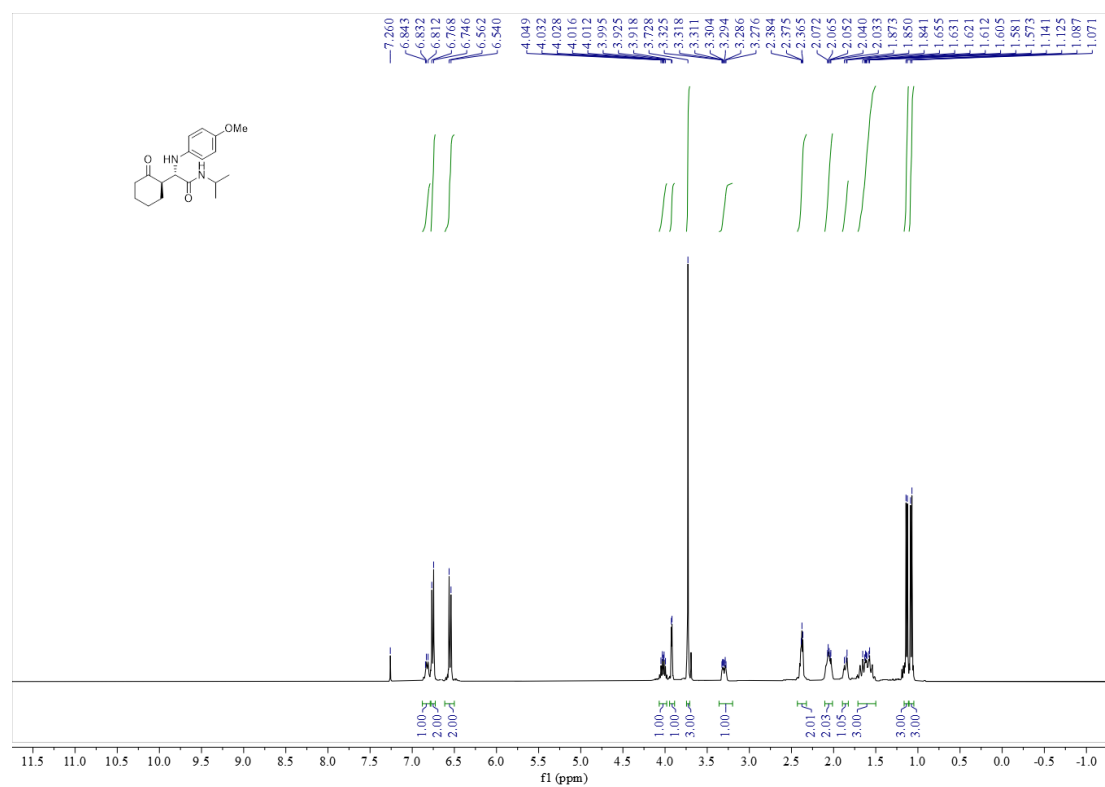
Ethyl (S)-2-((4-chlorophenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3ma**)



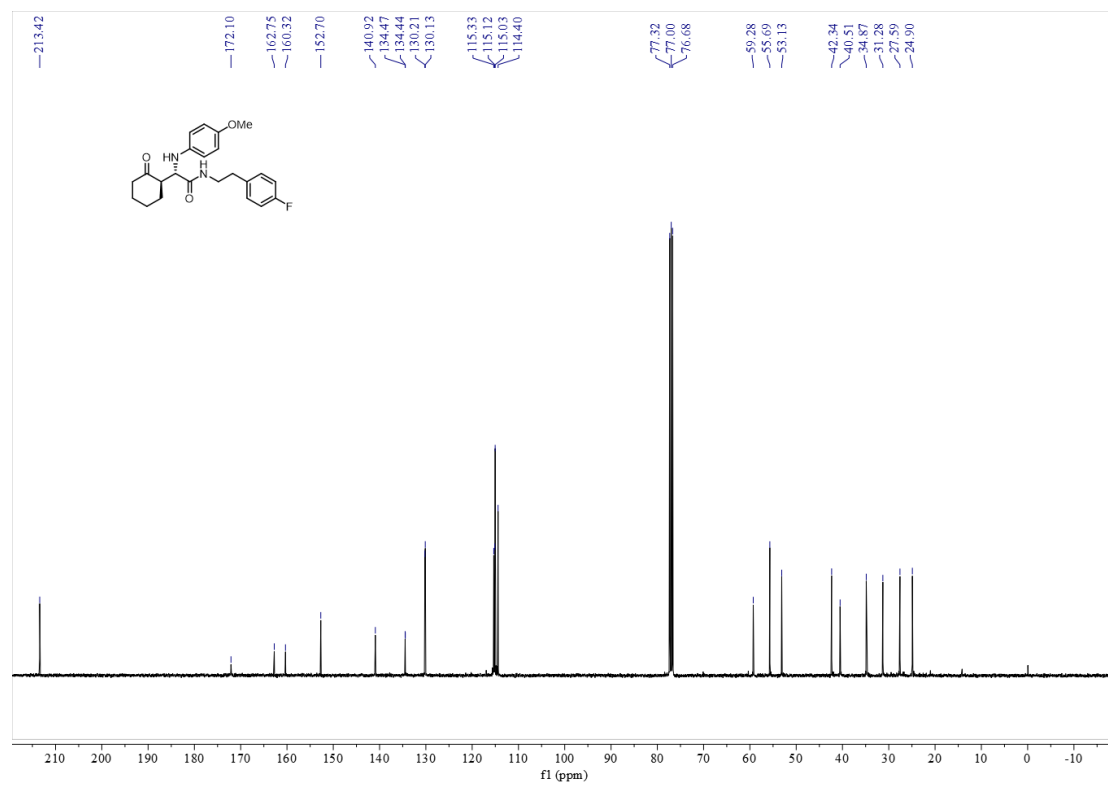
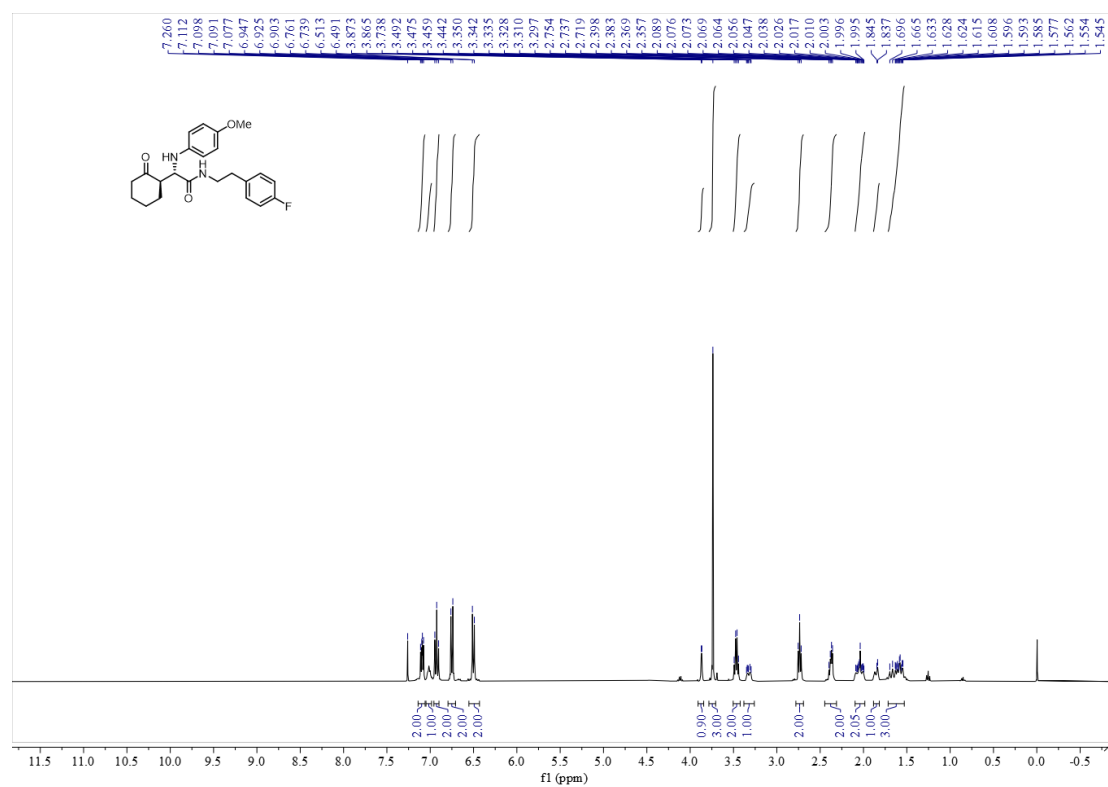
Cinnamyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetate (**3na**)

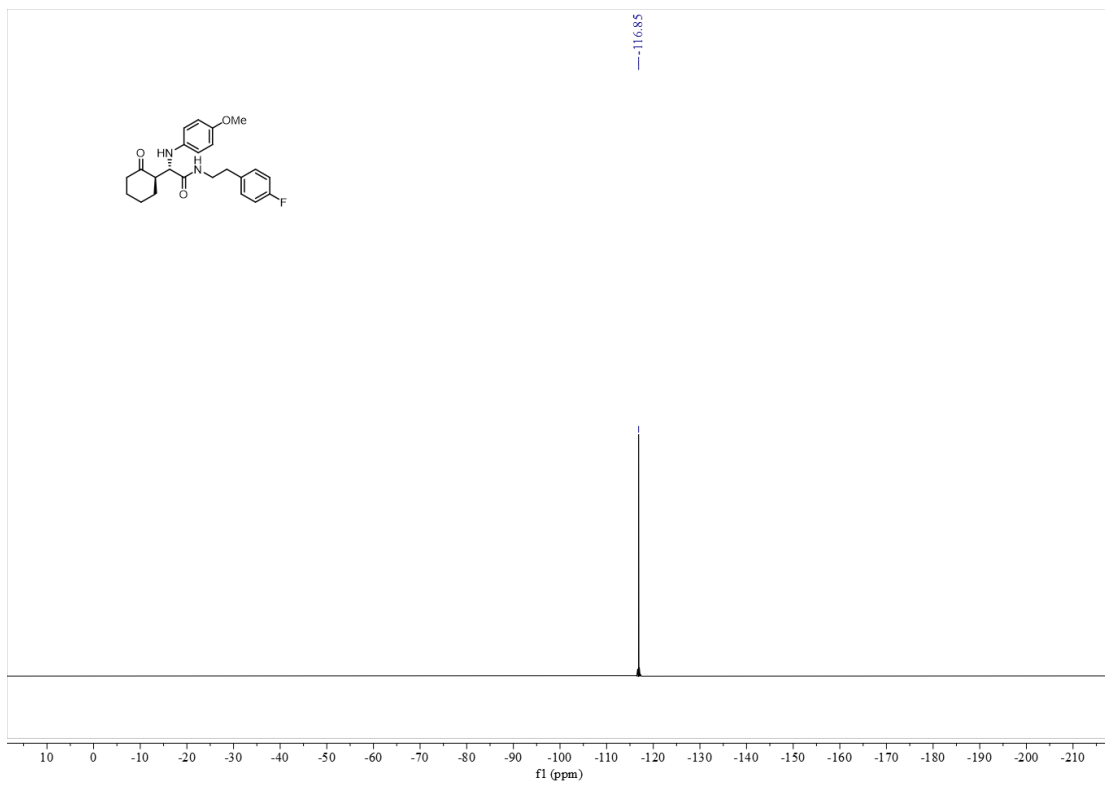


(S)-N-isopropyl-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamide (**30a**)

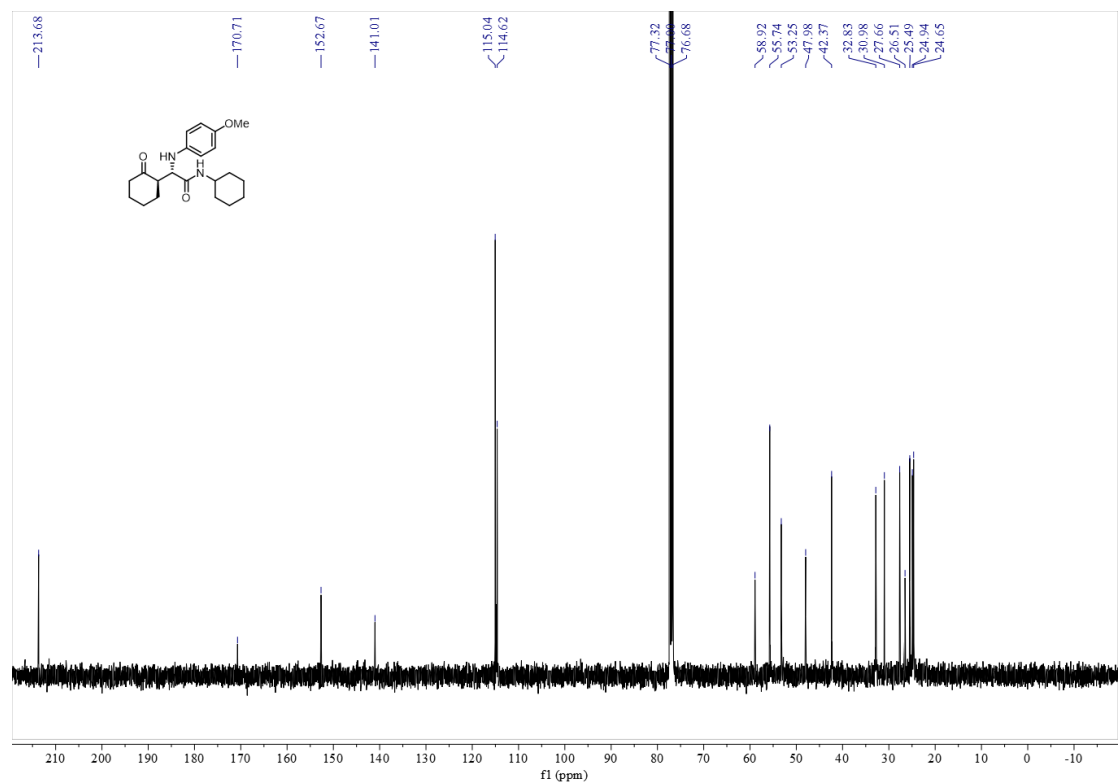
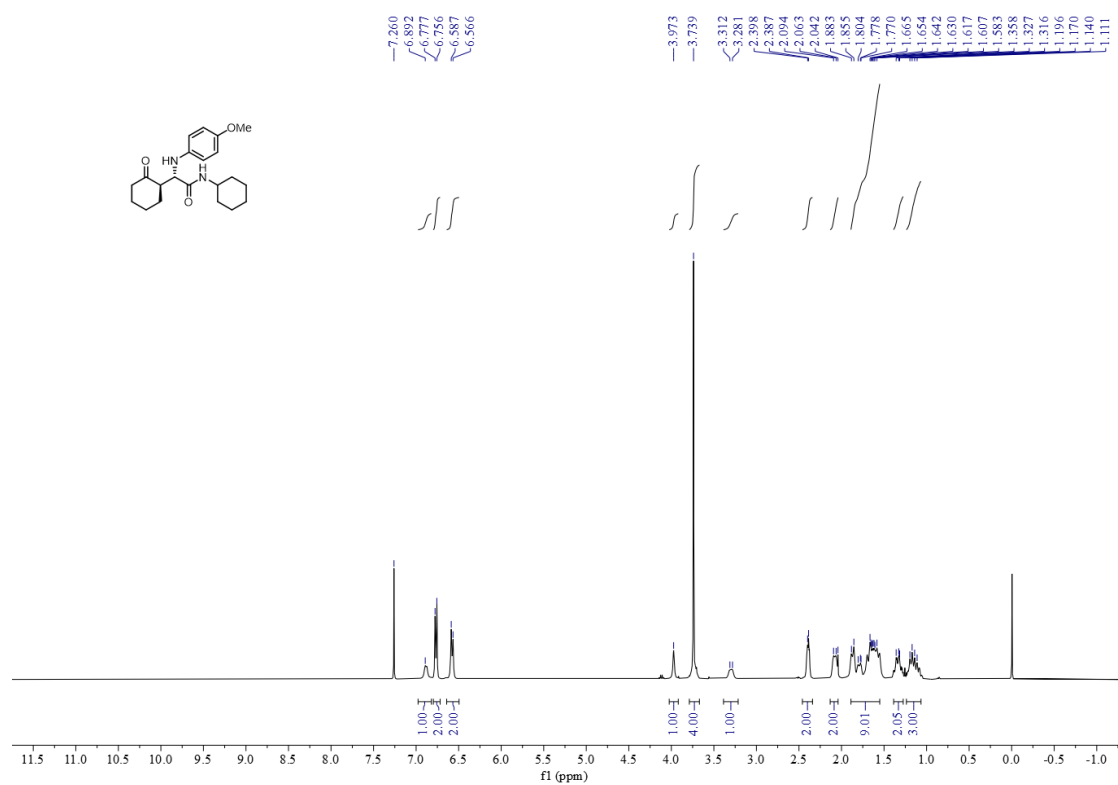


(S)-N-(4-fluorophenethyl)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamide (3pa)

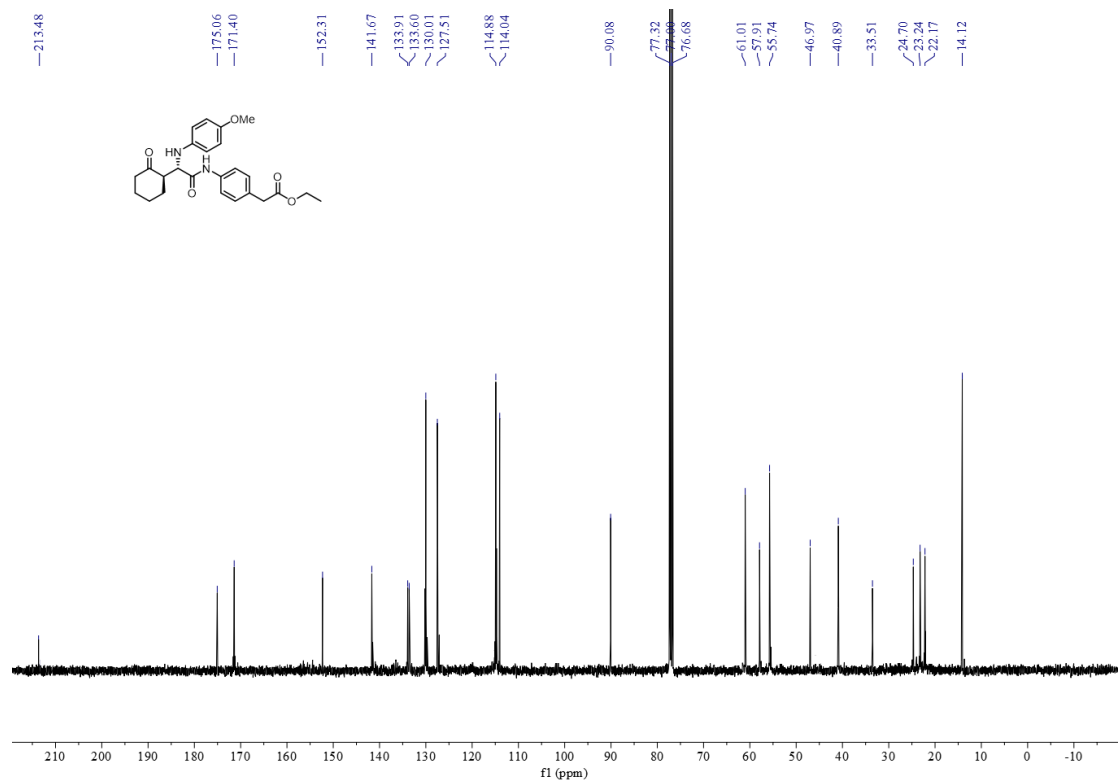
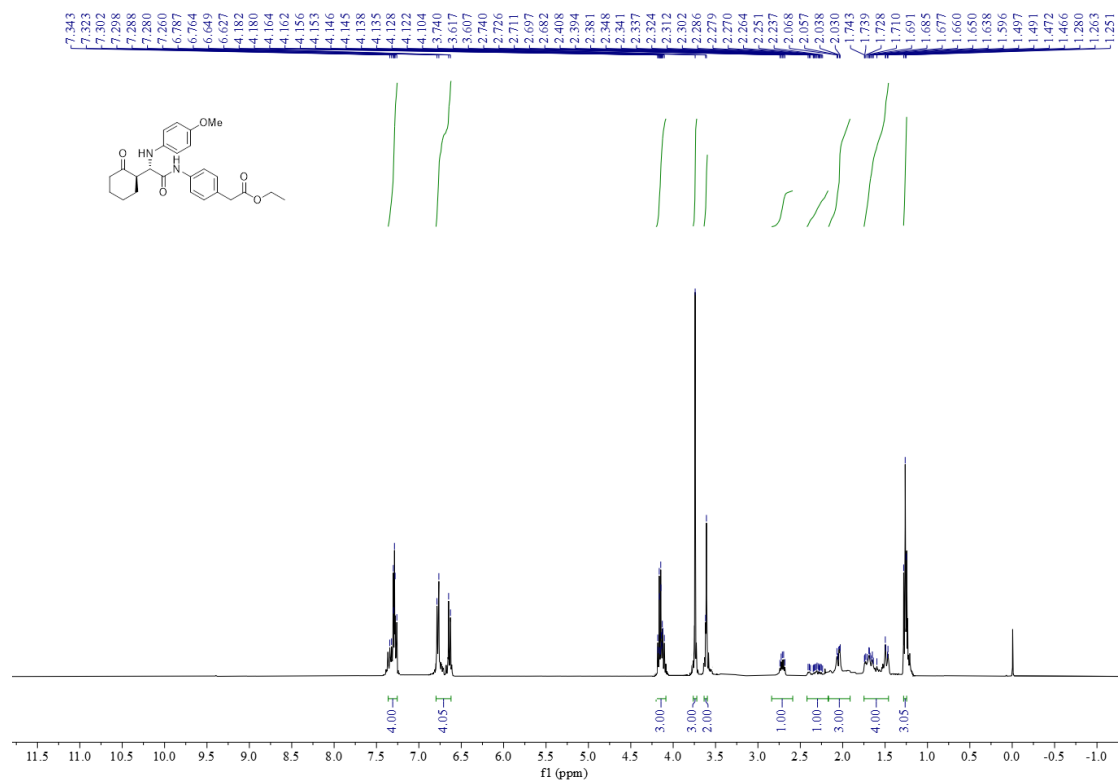




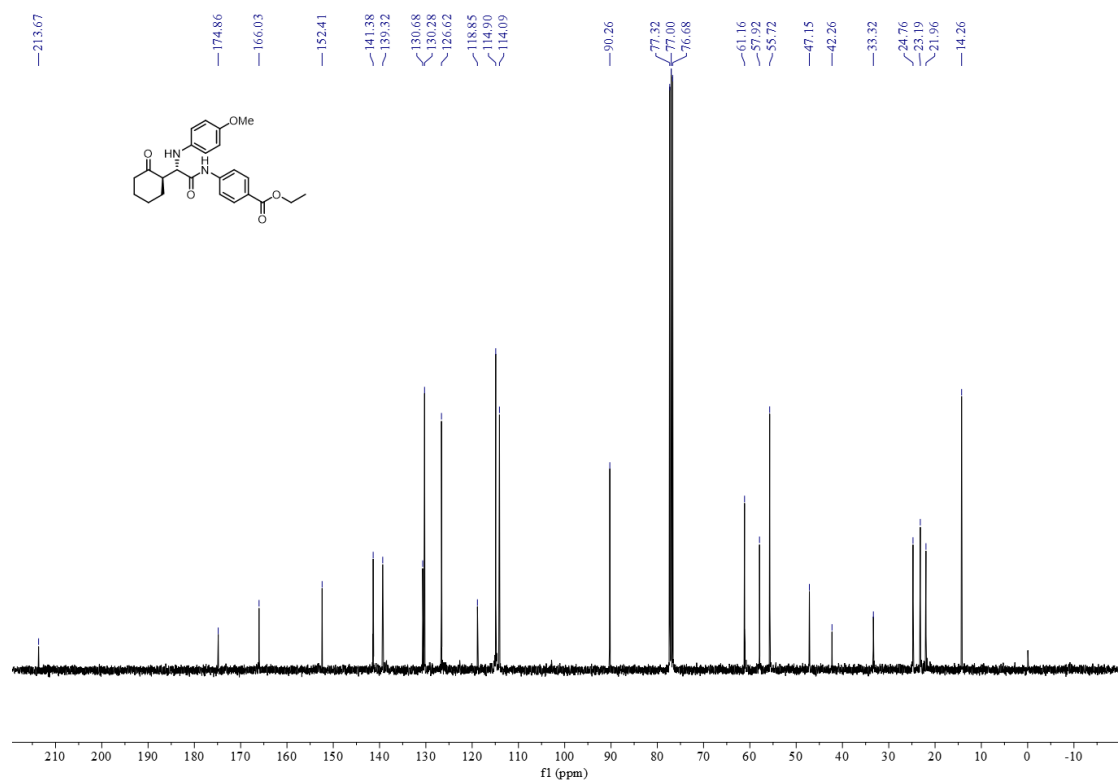
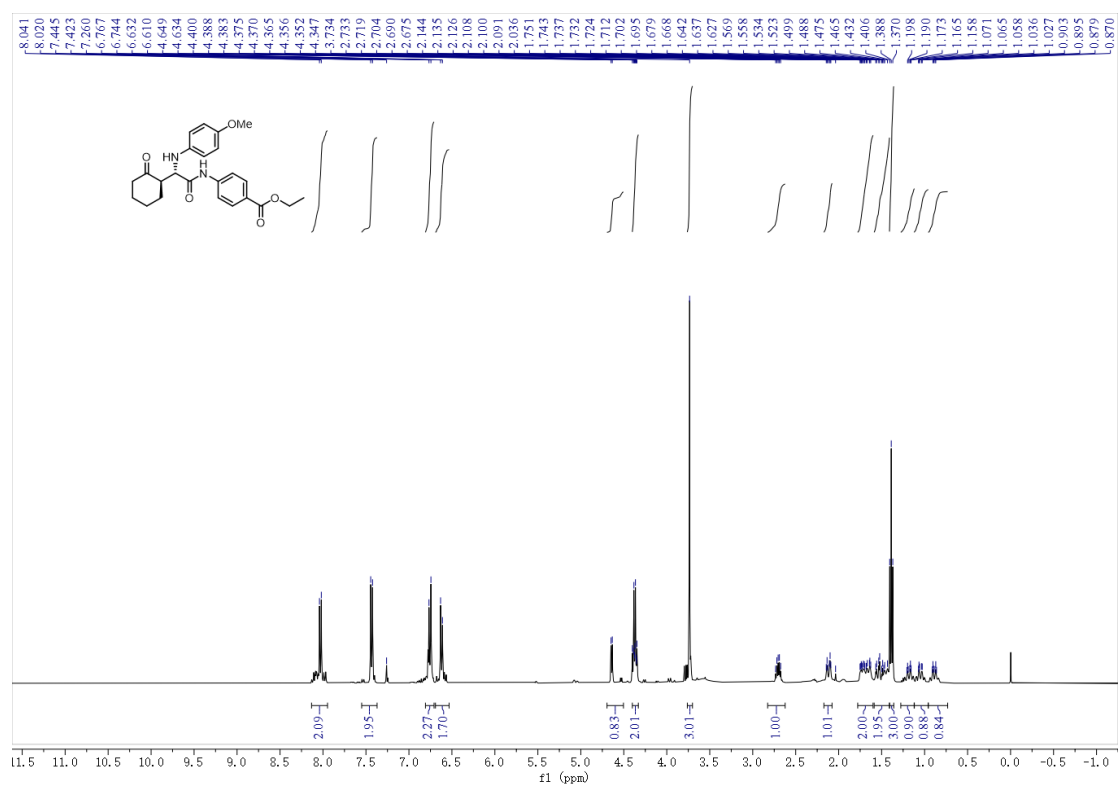
(S)-N-cyclohexyl-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamide (**3qa**)



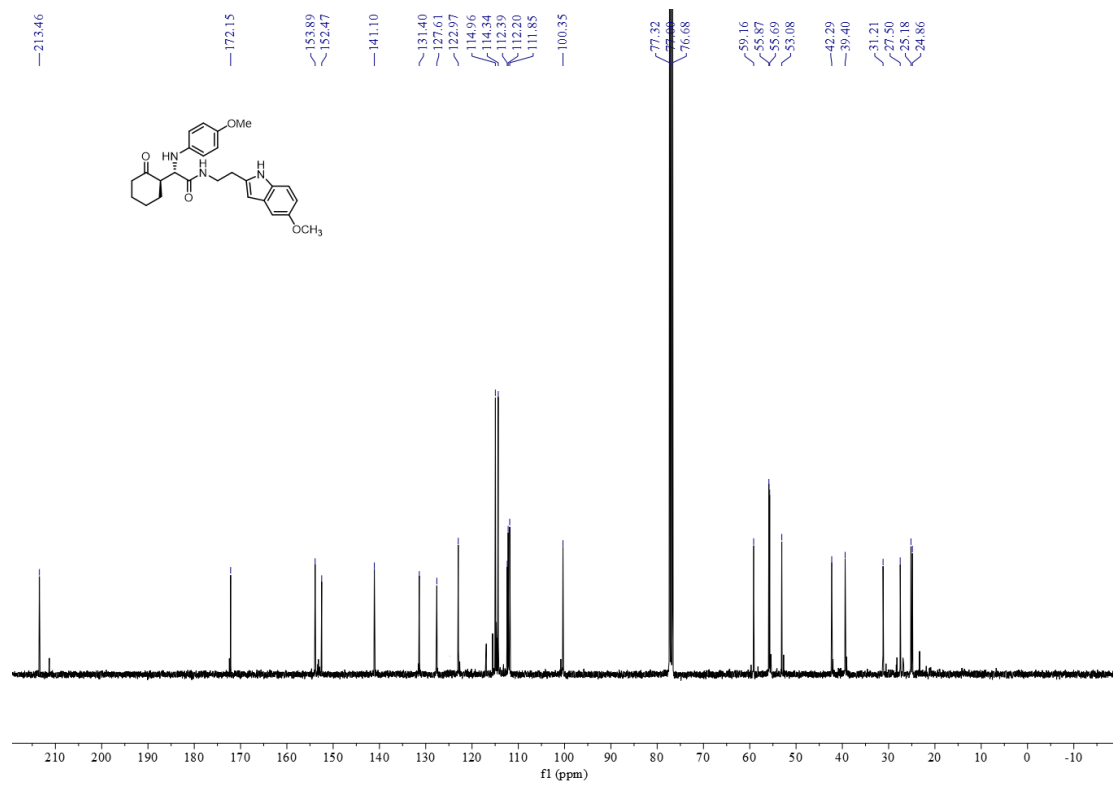
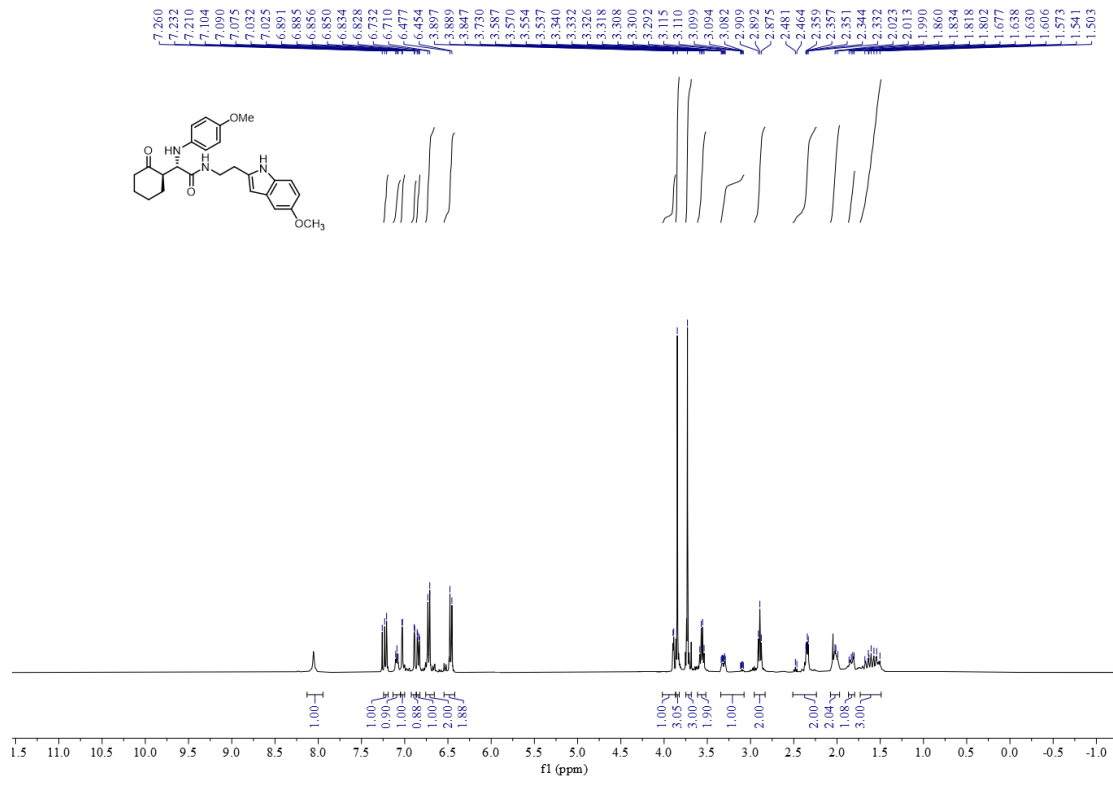
Ethyl 2-(4-((S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamido)phenyl)ac-etate (**3ra**)



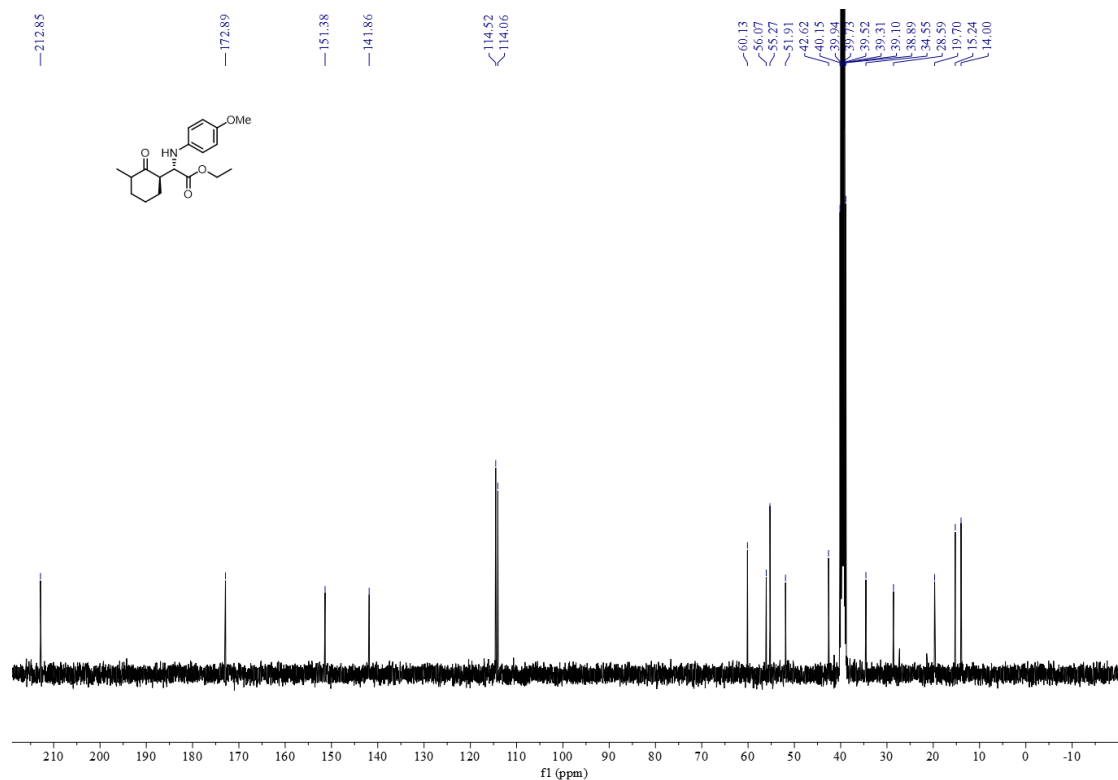
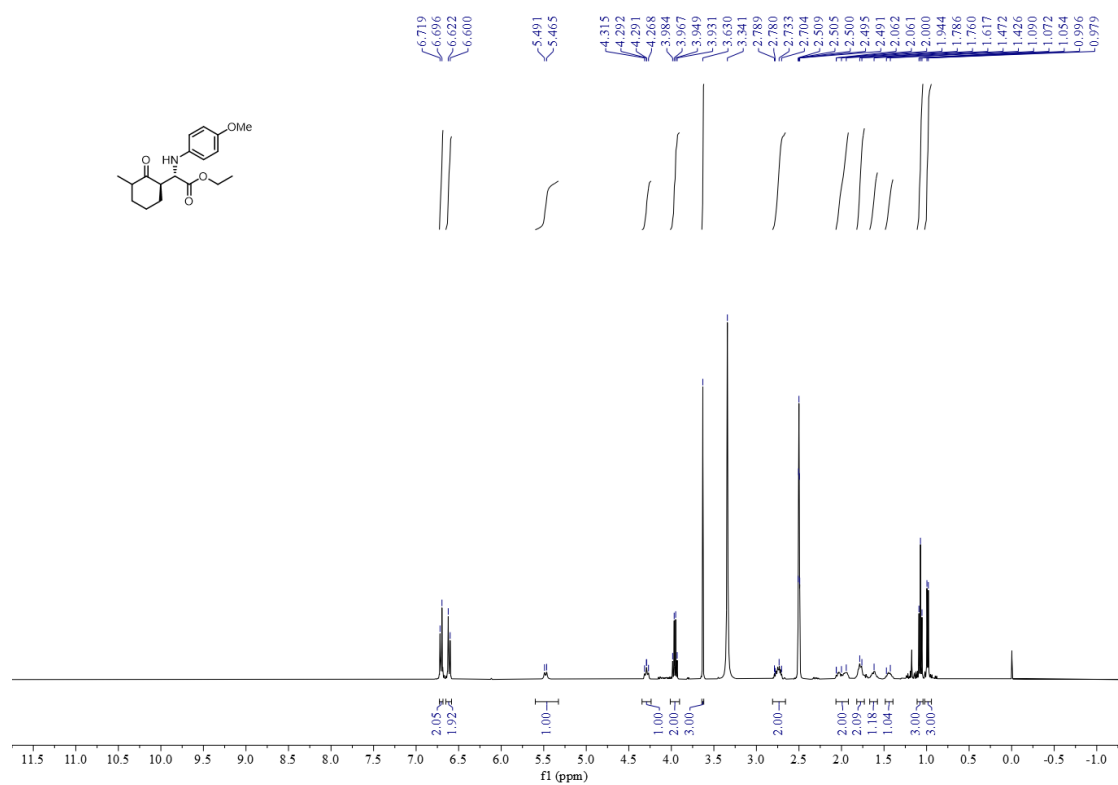
Ethyl 4-((S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamido)benzoate (**3sa**)



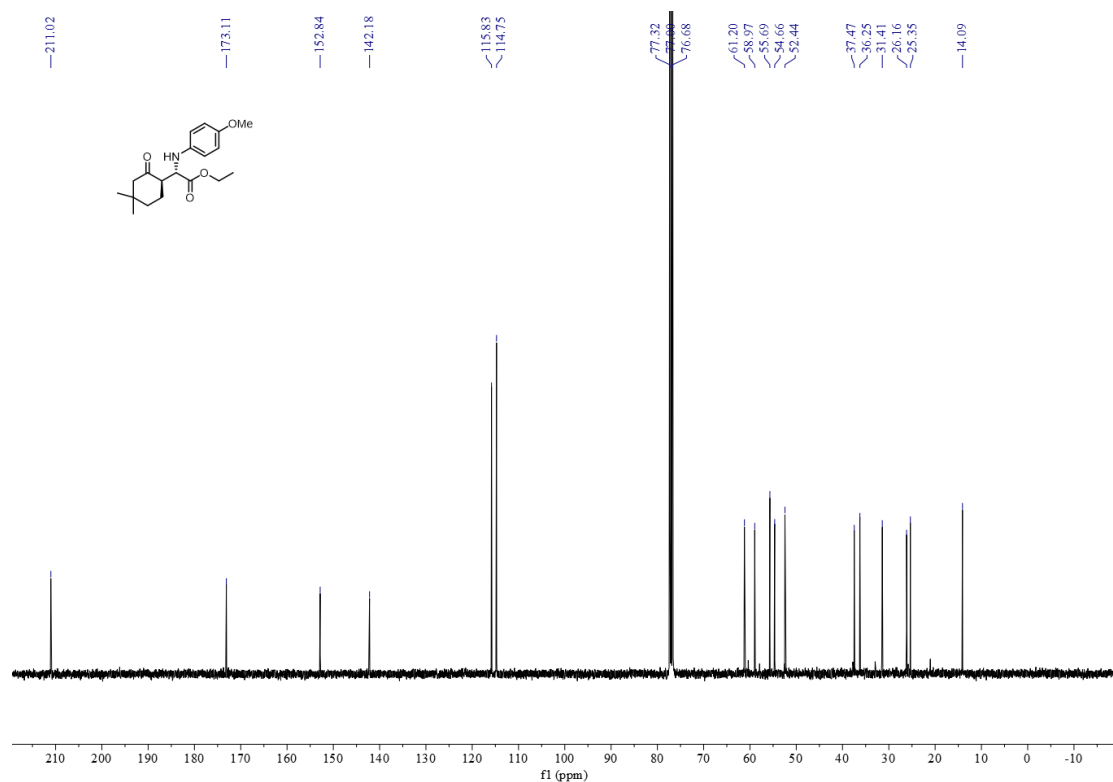
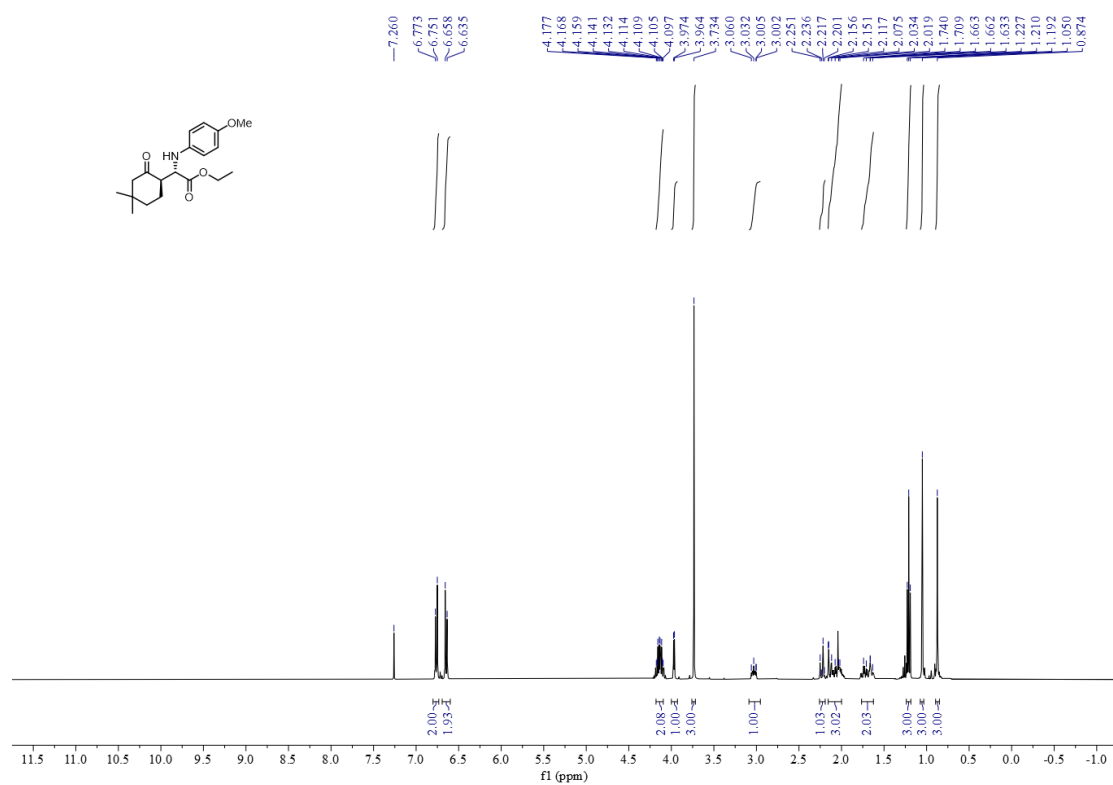
(S)-N-(2-(5-methoxy-1H-indol-2-yl)ethyl)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocyclohexyl)acetamide (3ta)



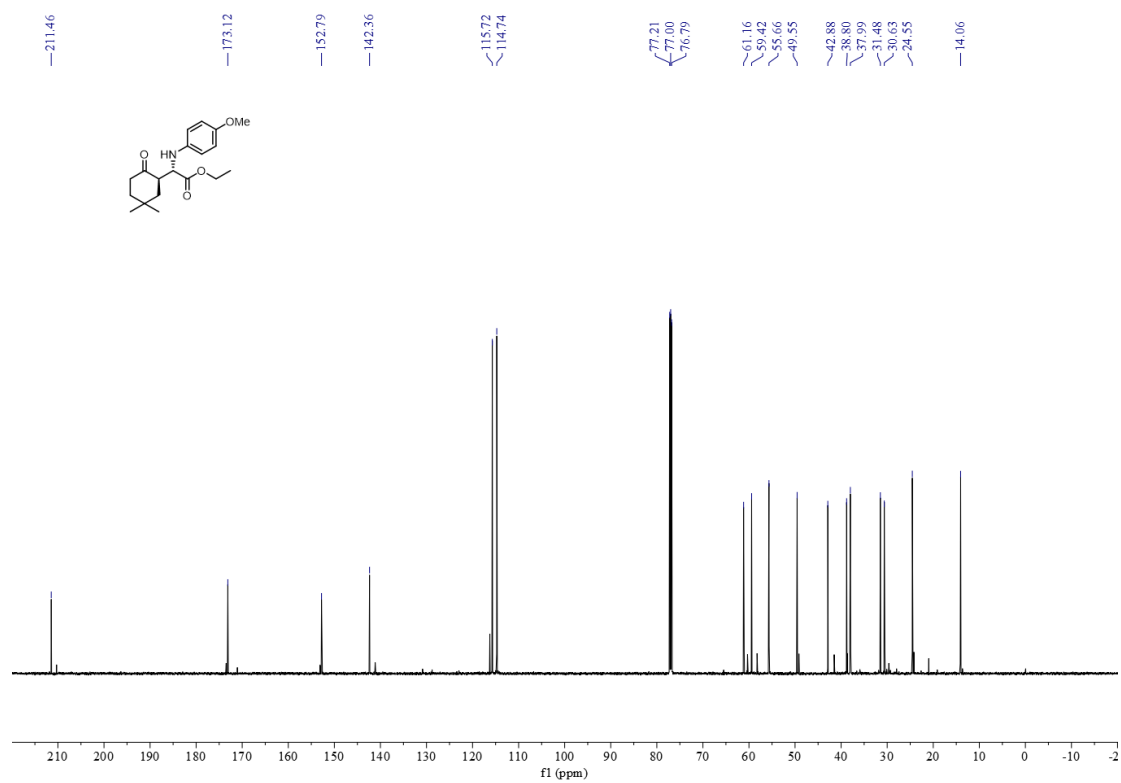
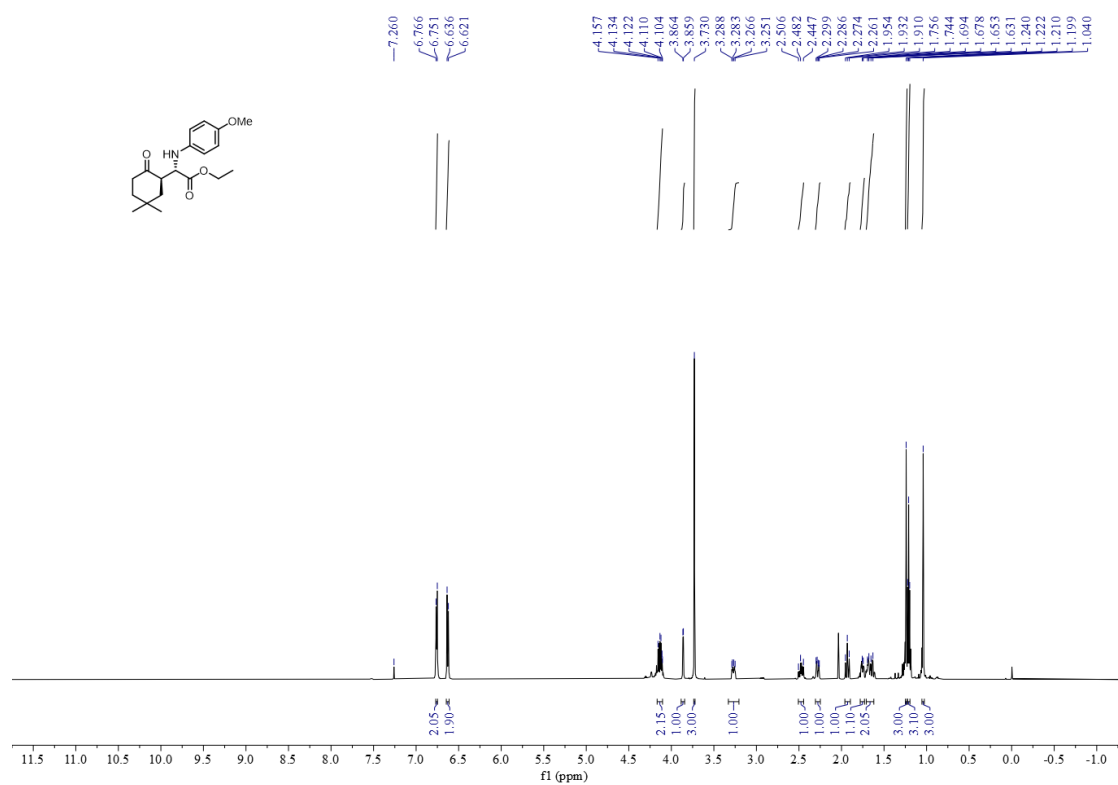
Ethyl (2S)-2-((4-methoxyphenyl)amino)-2-((1R)-3-methyl-2-oxocyclohexyl)acetate (**3ab**)



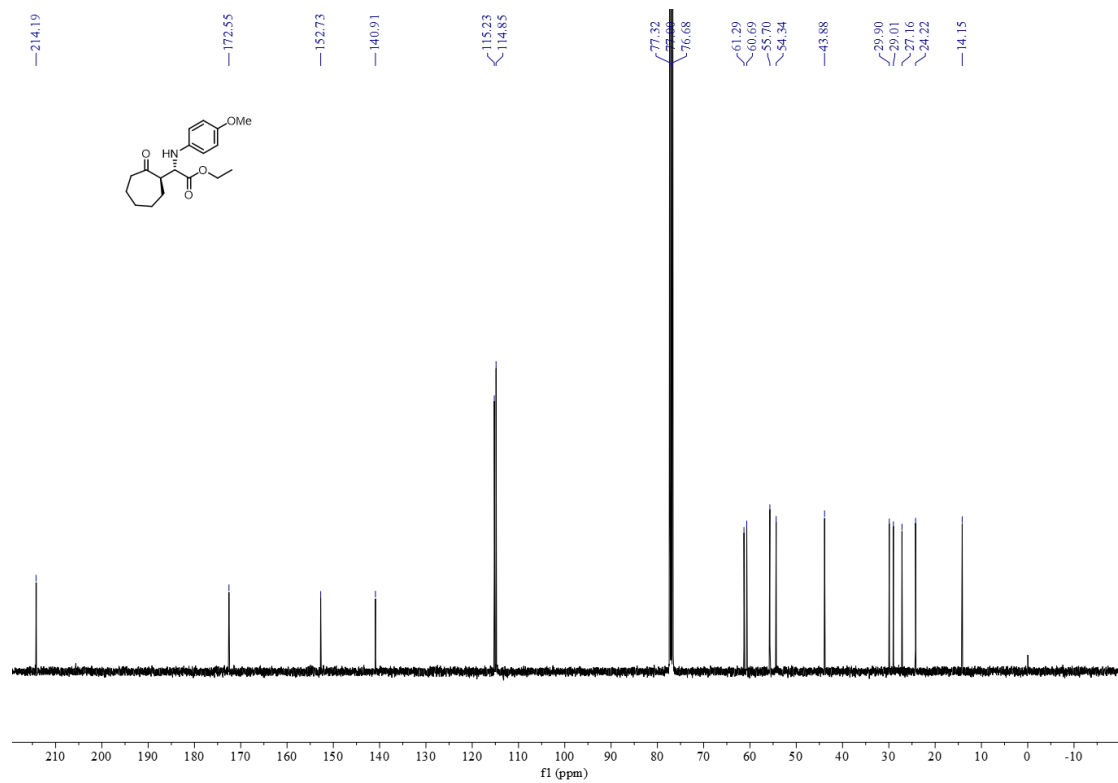
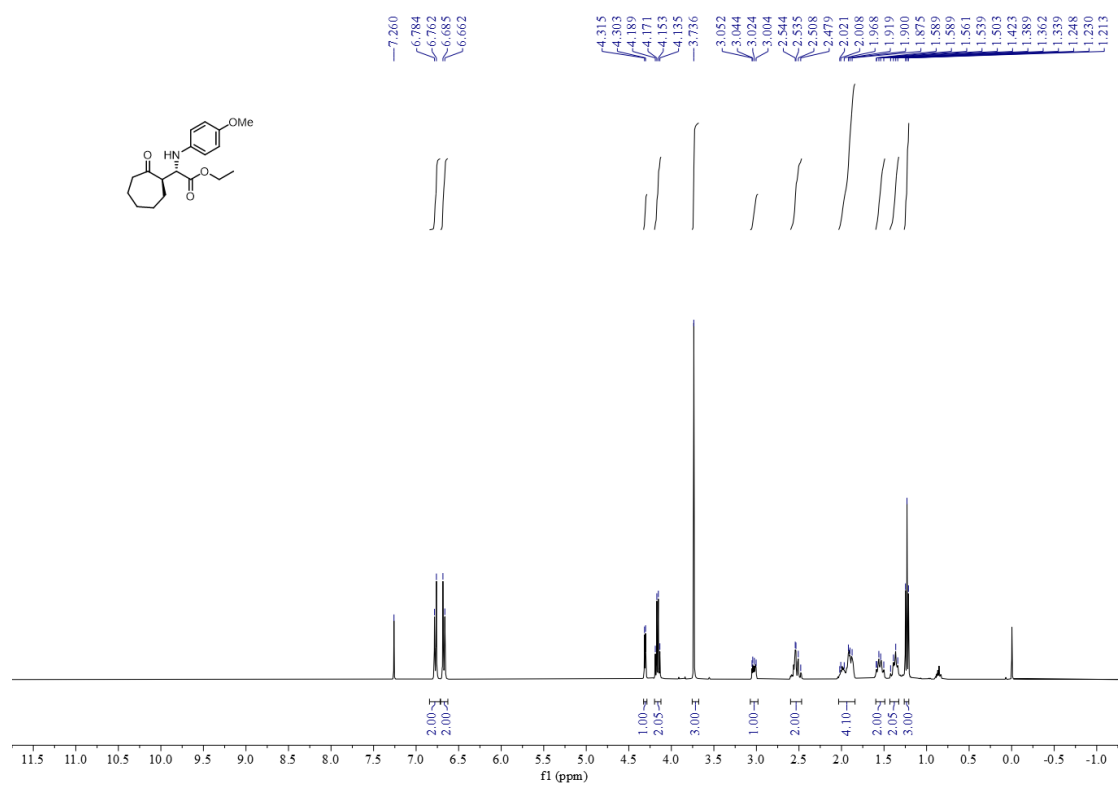
Ethyl (S)-2-((R)-4,4-dimethyl-2-oxocyclohexyl)-2-((4-methoxyphenyl)amino)acetate (**3ac**)



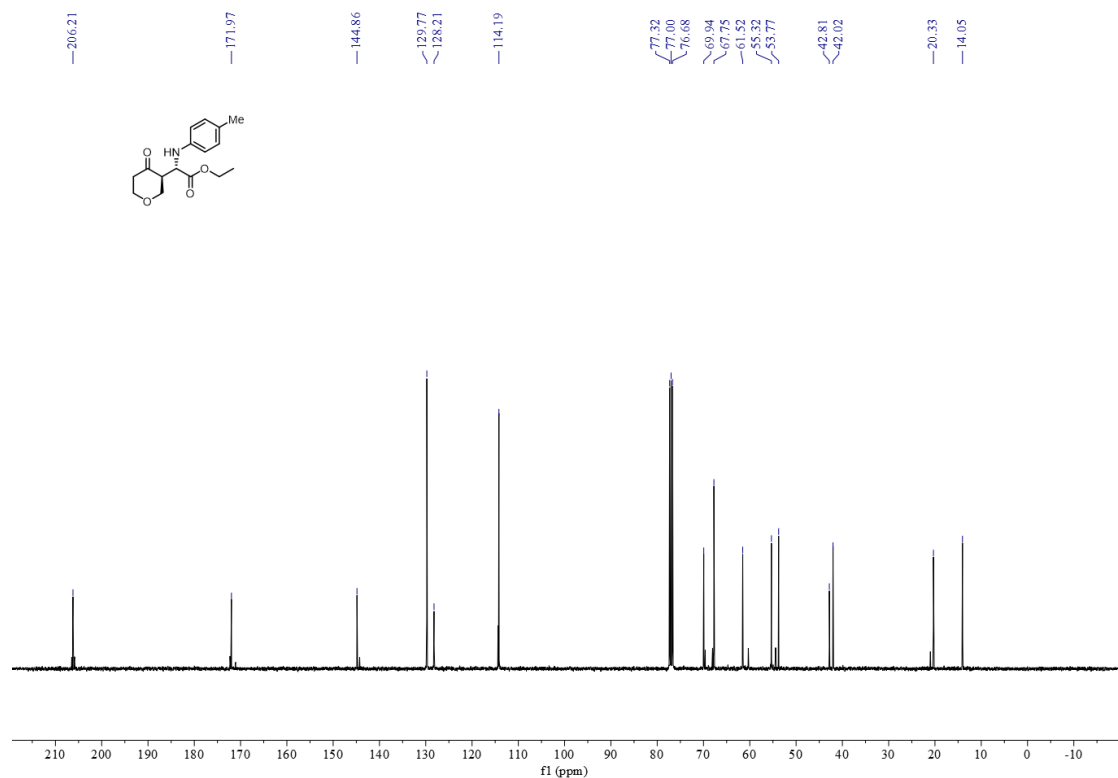
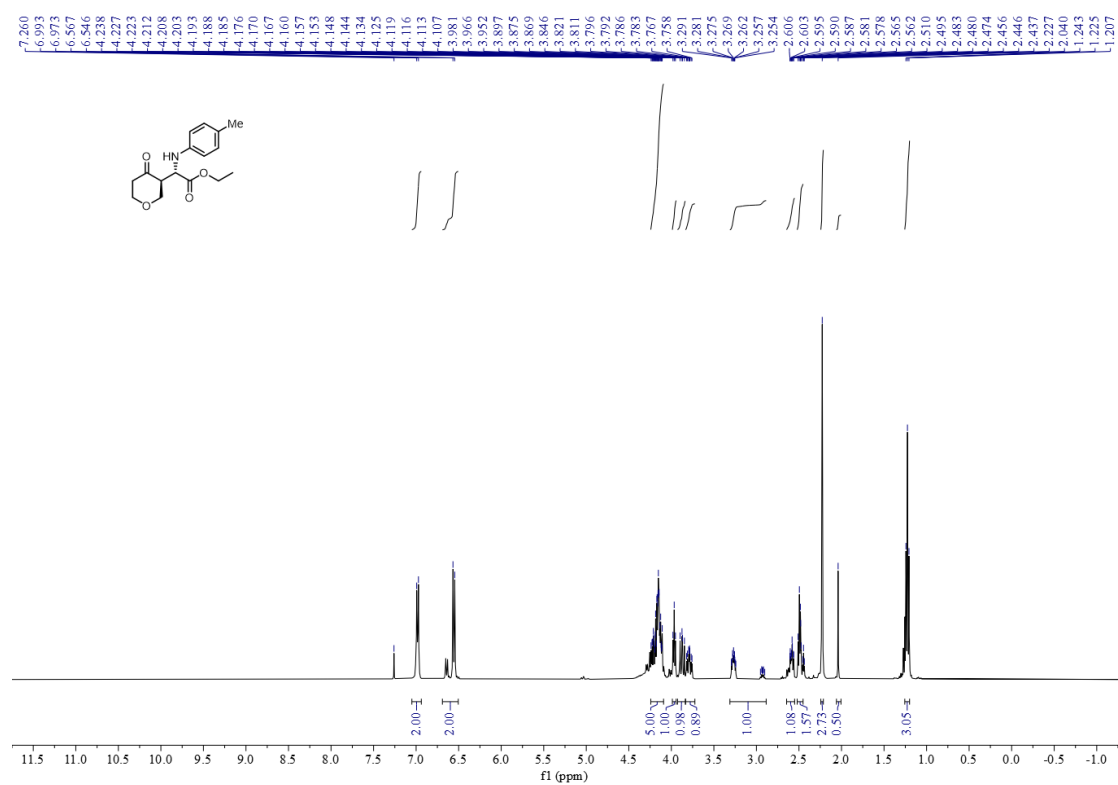
Ethyl (S)-2-((R)-5,5-dimethyl-2-oxocyclohexyl)-2-((4-methoxyphenyl)amino)acetate (**3ad**)



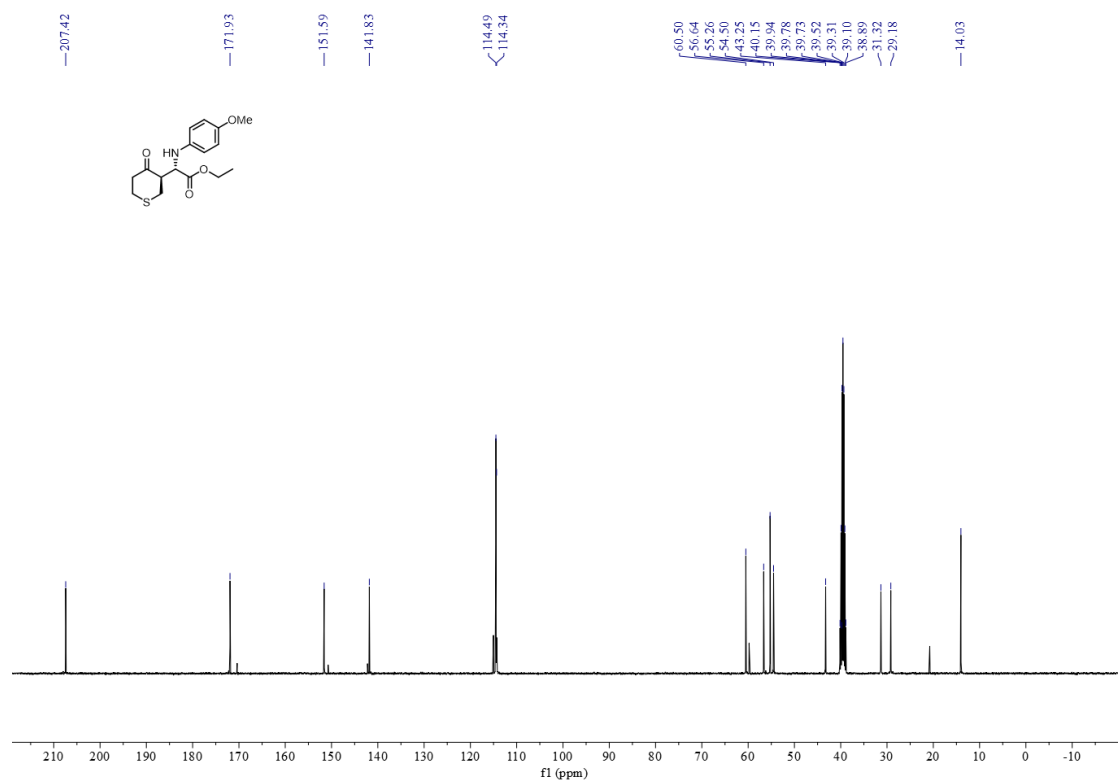
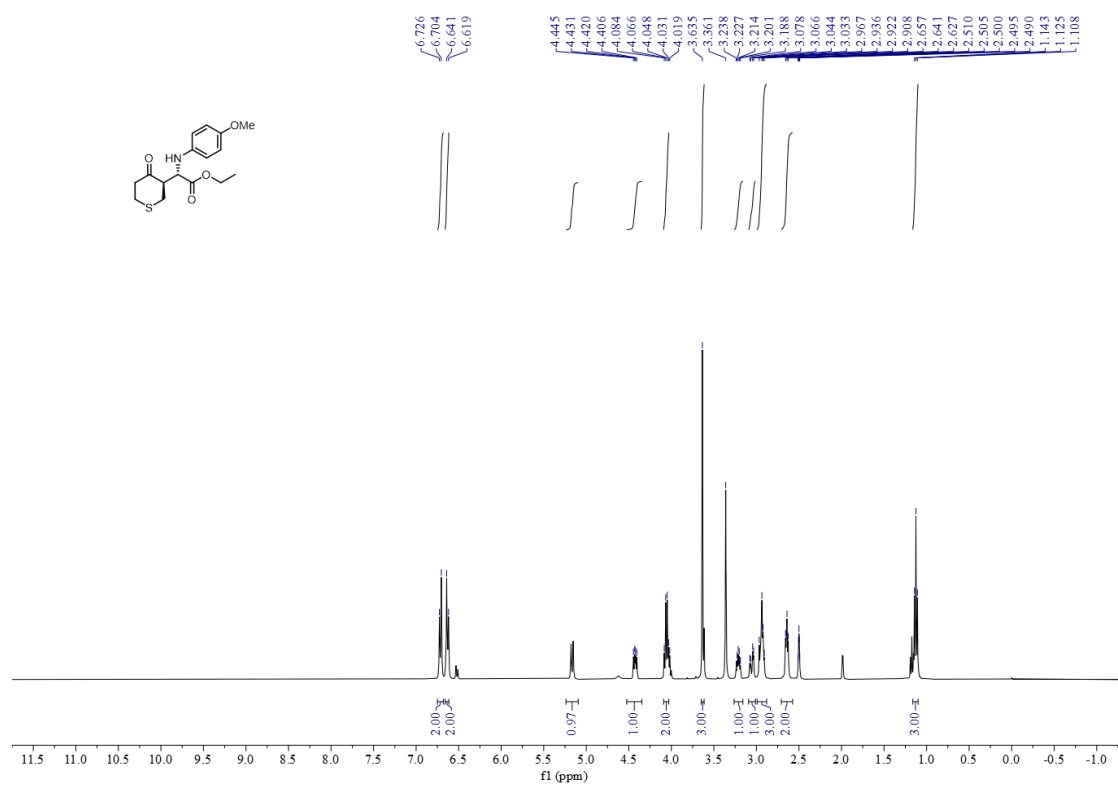
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-2-oxocycloheptyl)acetate (**3ae**)



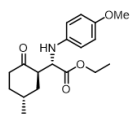
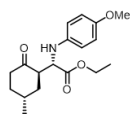
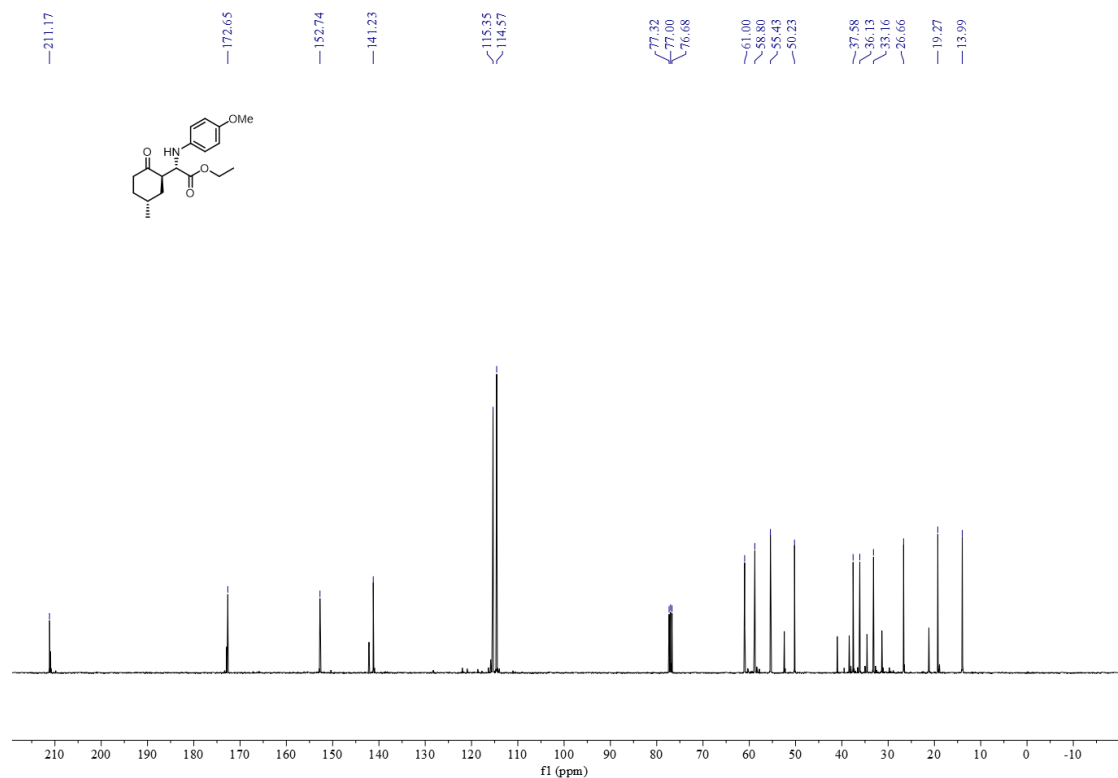
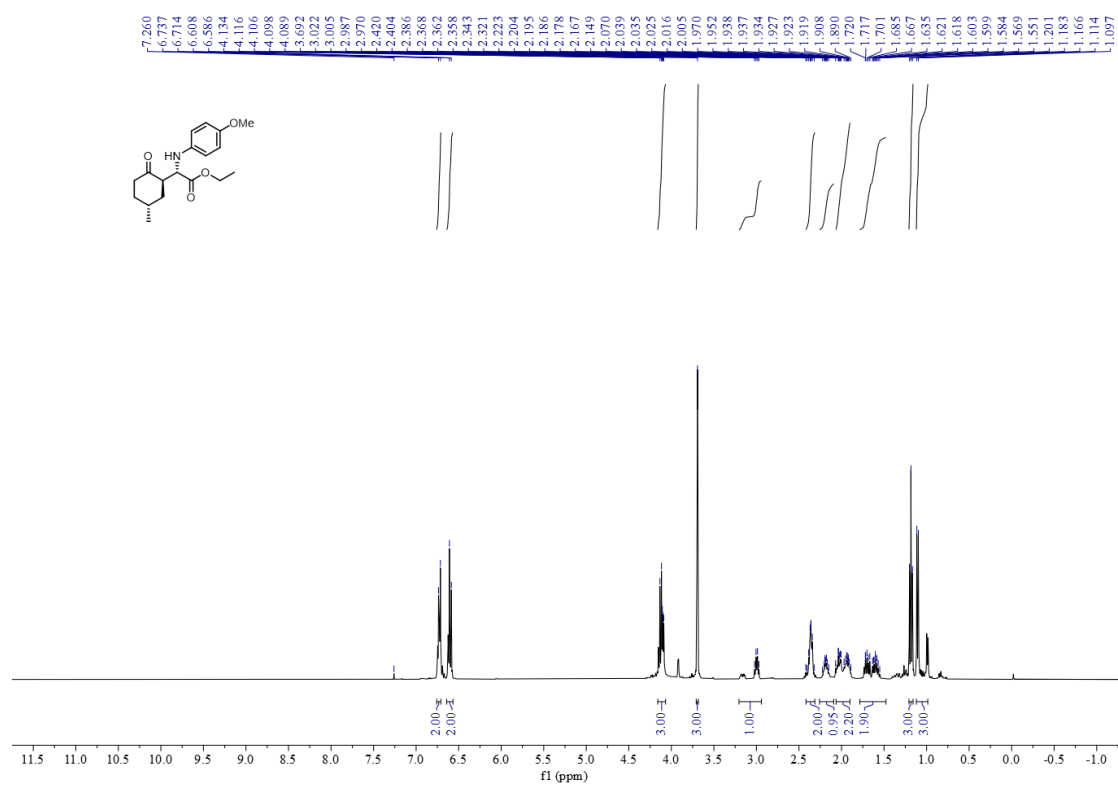
Ethyl (S)-2-((S)-4-oxotetrahydro-2H-pyran-3-yl)-2-(p-tolylamino)acetate (**3kf**)



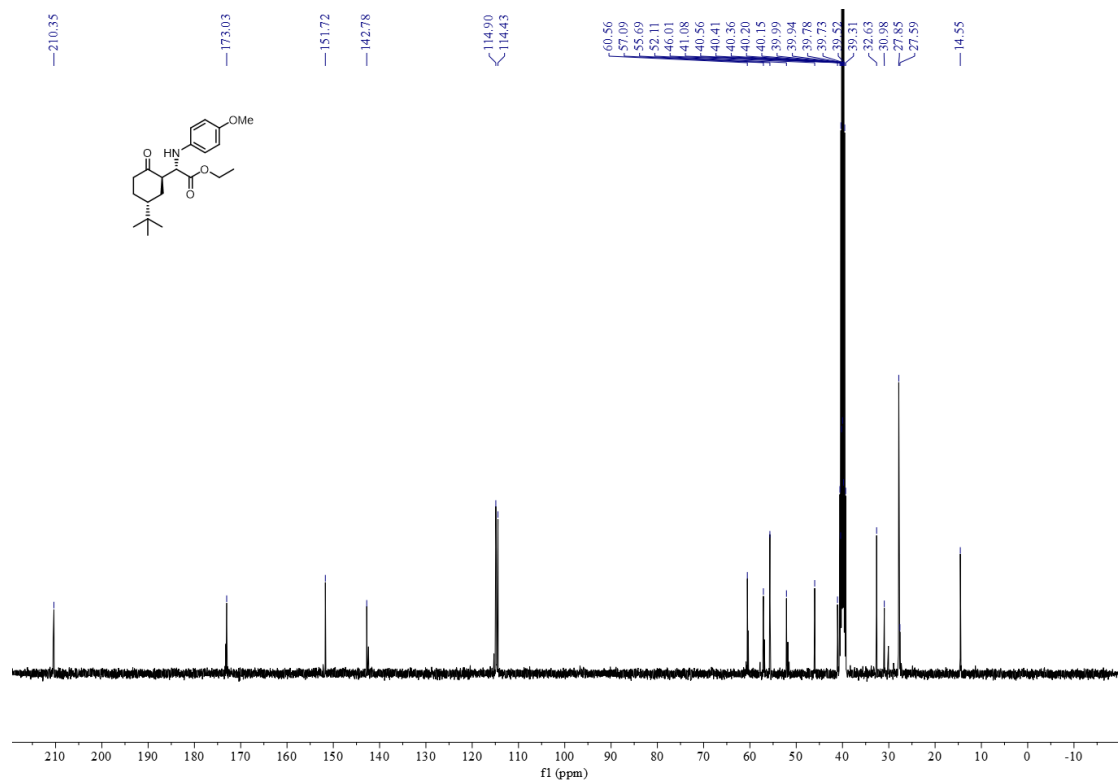
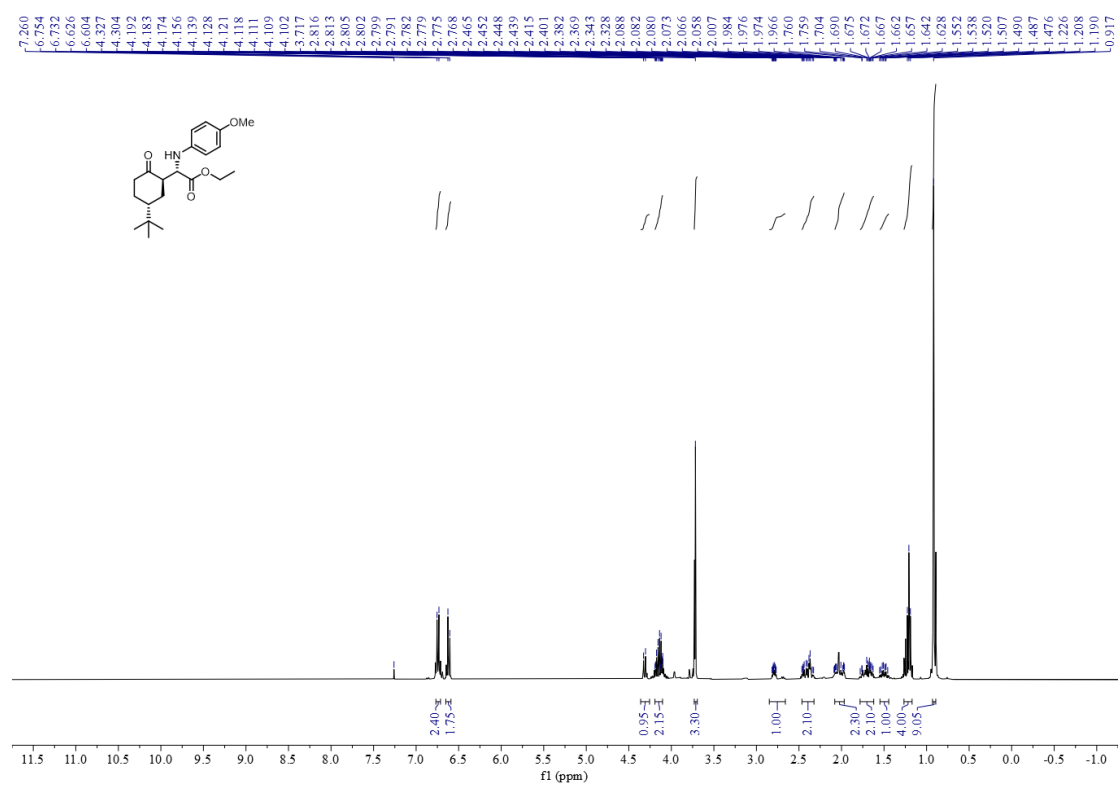
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-4-oxotetrahydro-2H-thiopyran-3-yl)acetate (**3ag**)



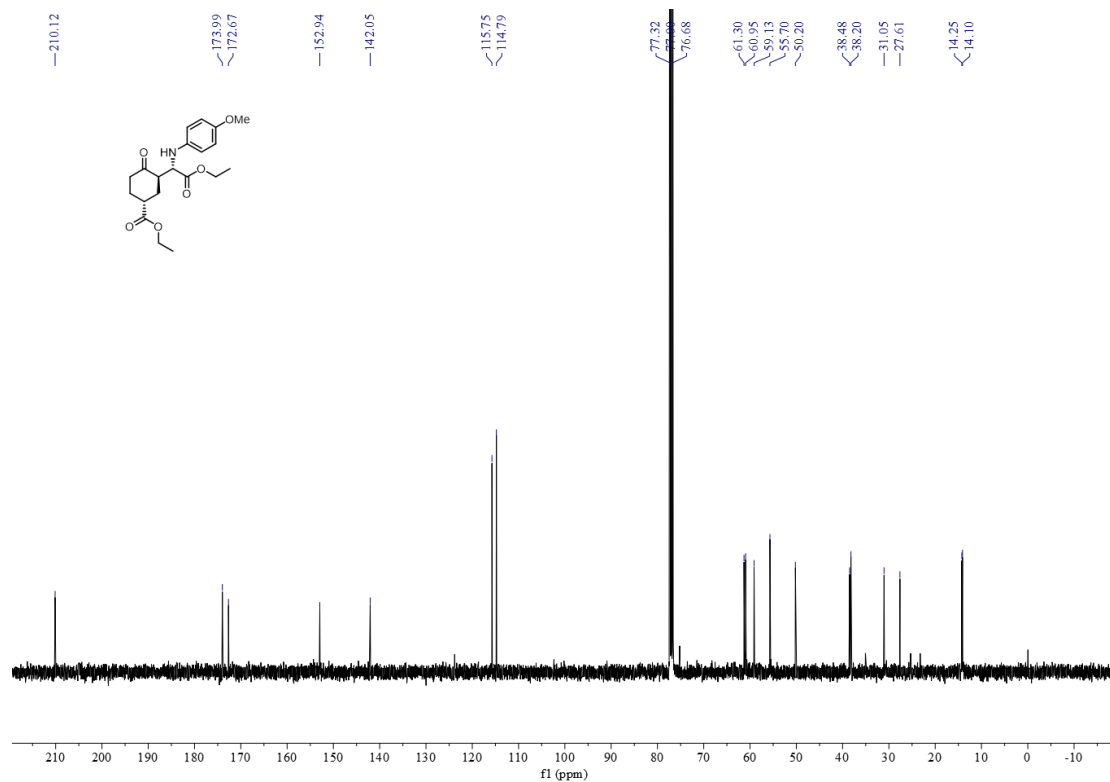
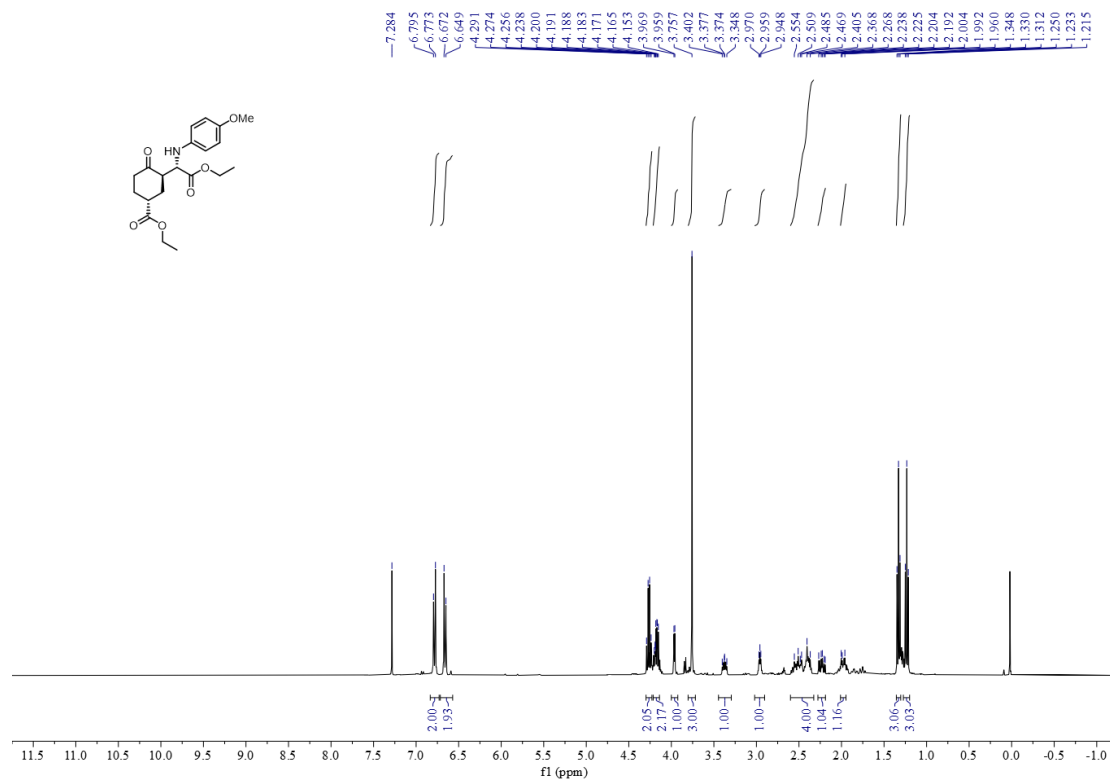
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((1R,5R)-5-methyl-2-oxocyclohexyl)acetate (**3ah**)



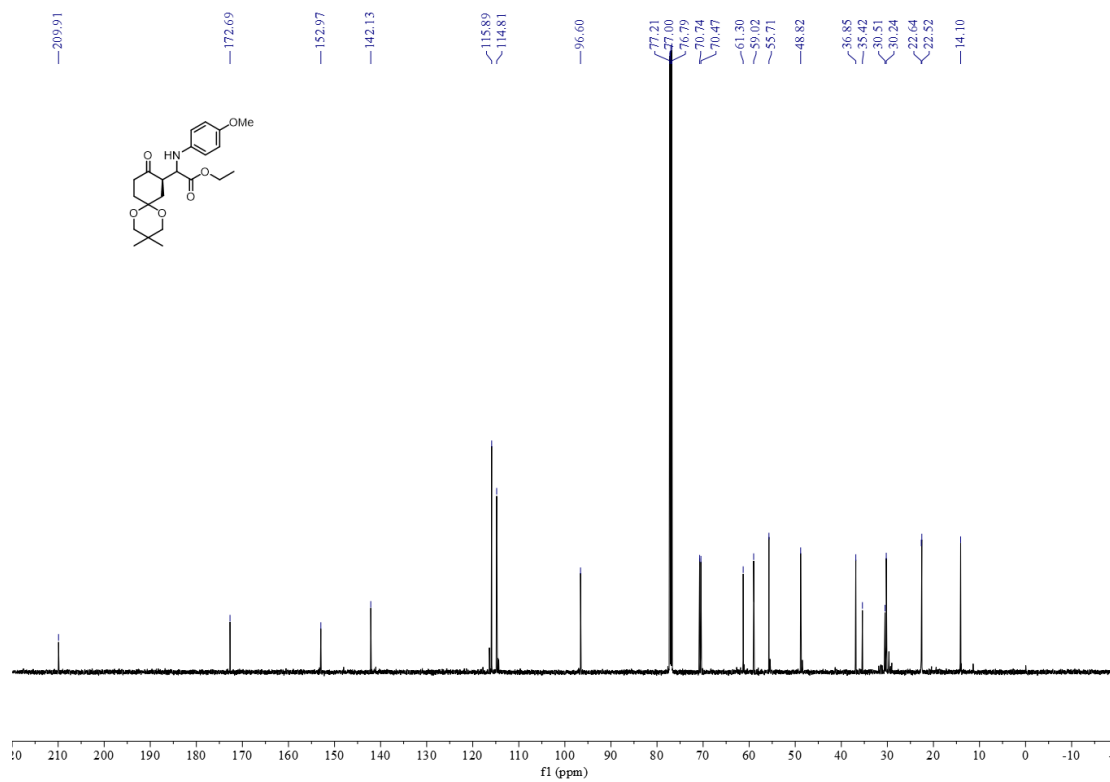
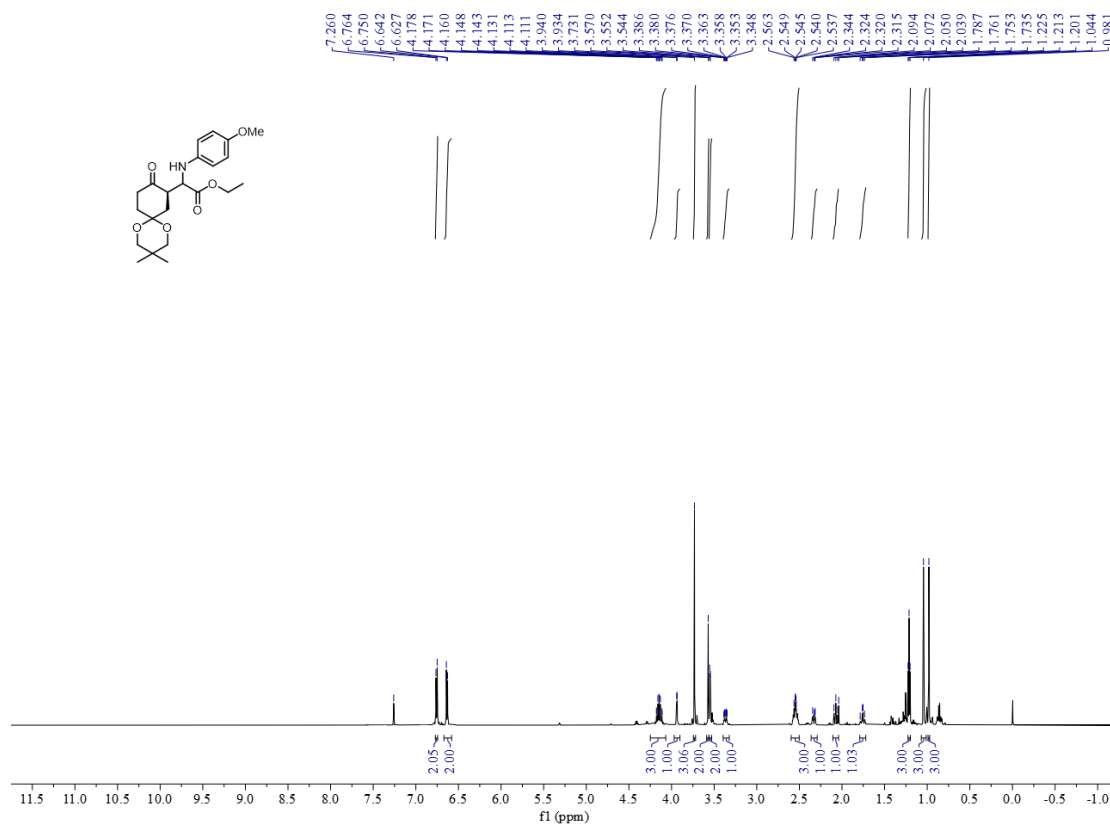
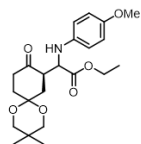
Ethyl (S)-2-((1R,5R)-5-(tert-butyl)-2-oxocyclohexyl)-2-((4-methoxyphenyl)amino)acetate (**3ai**)



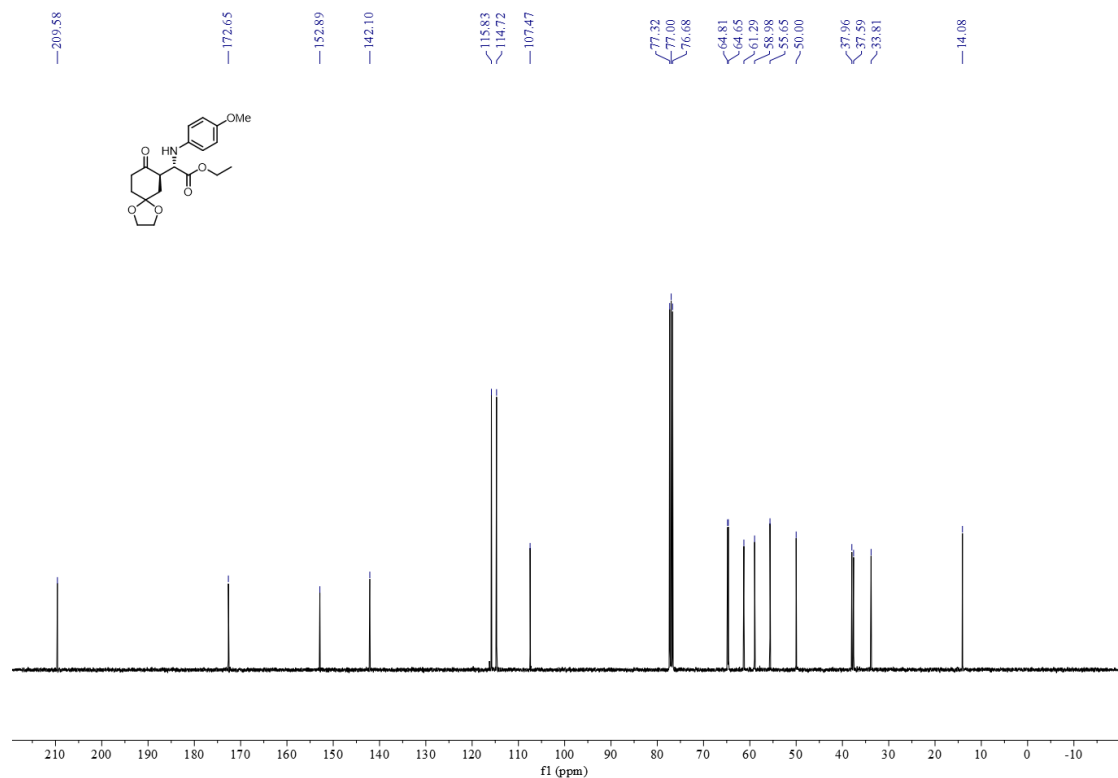
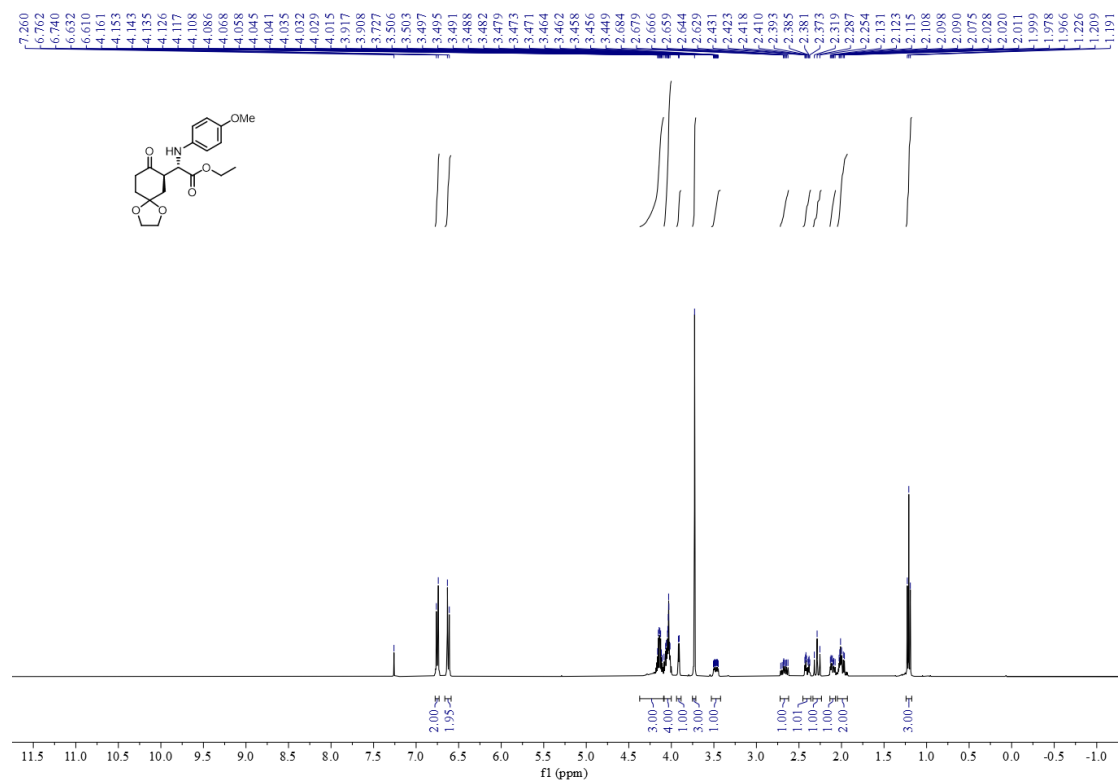
Ethyl (1R,3R)-3-((S)-2-ethoxy-1-((4-methoxyphenyl)amino)-2-oxoethyl)-4-oxocyclohexan-e-1-carboxylate (**3aj**)



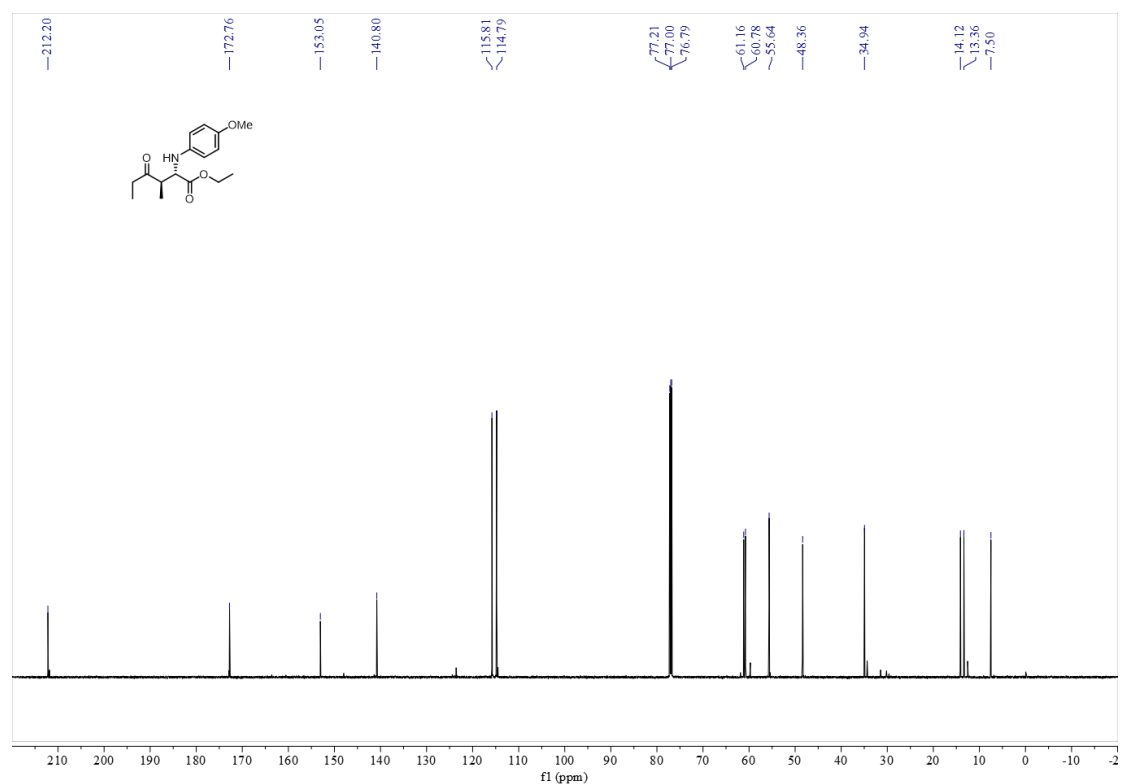
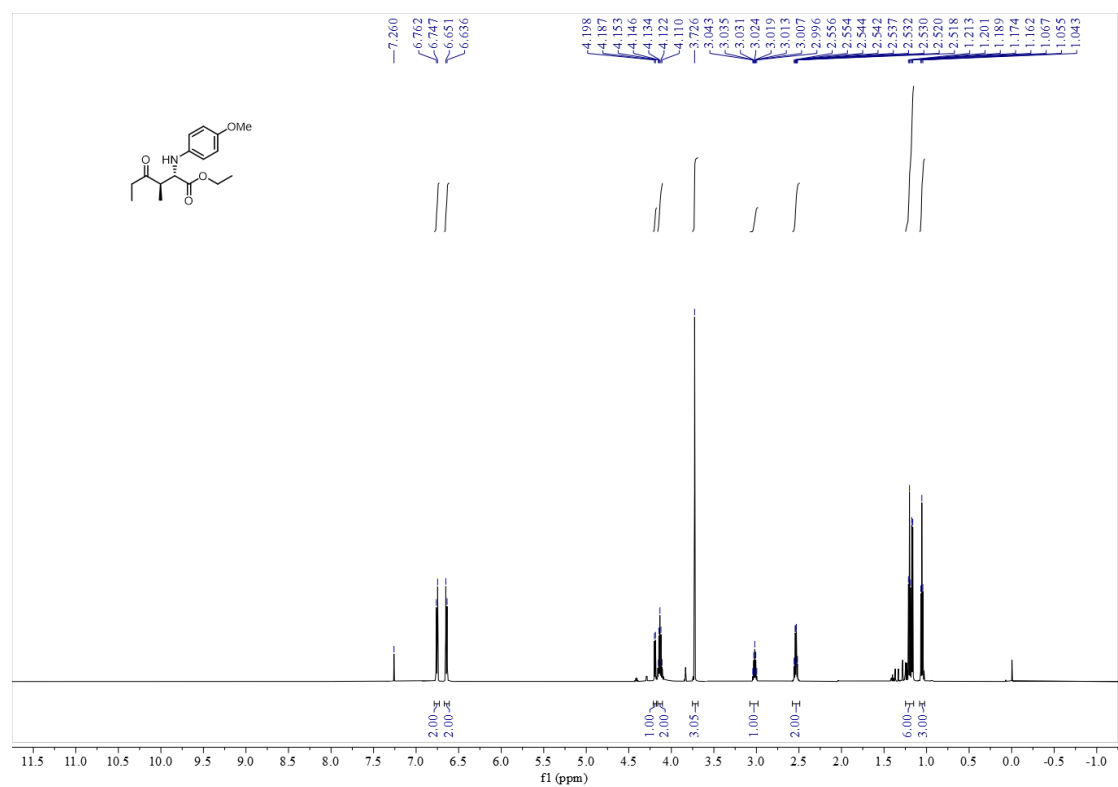
Ethyl 2-((R)-3,3-dimethyl-9-oxo-1,5-dioxaspiro[5.5]undecan-8-yl)-2-((4-methoxyphenyl)amino)acetate
(3ak)



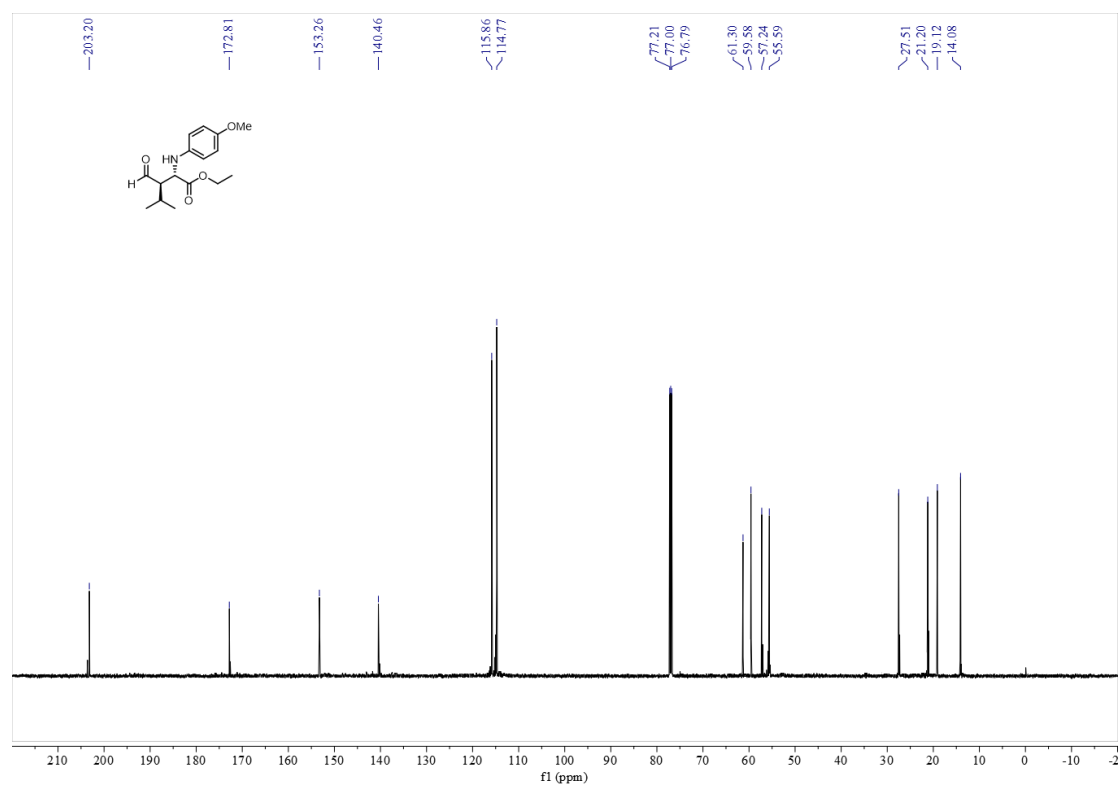
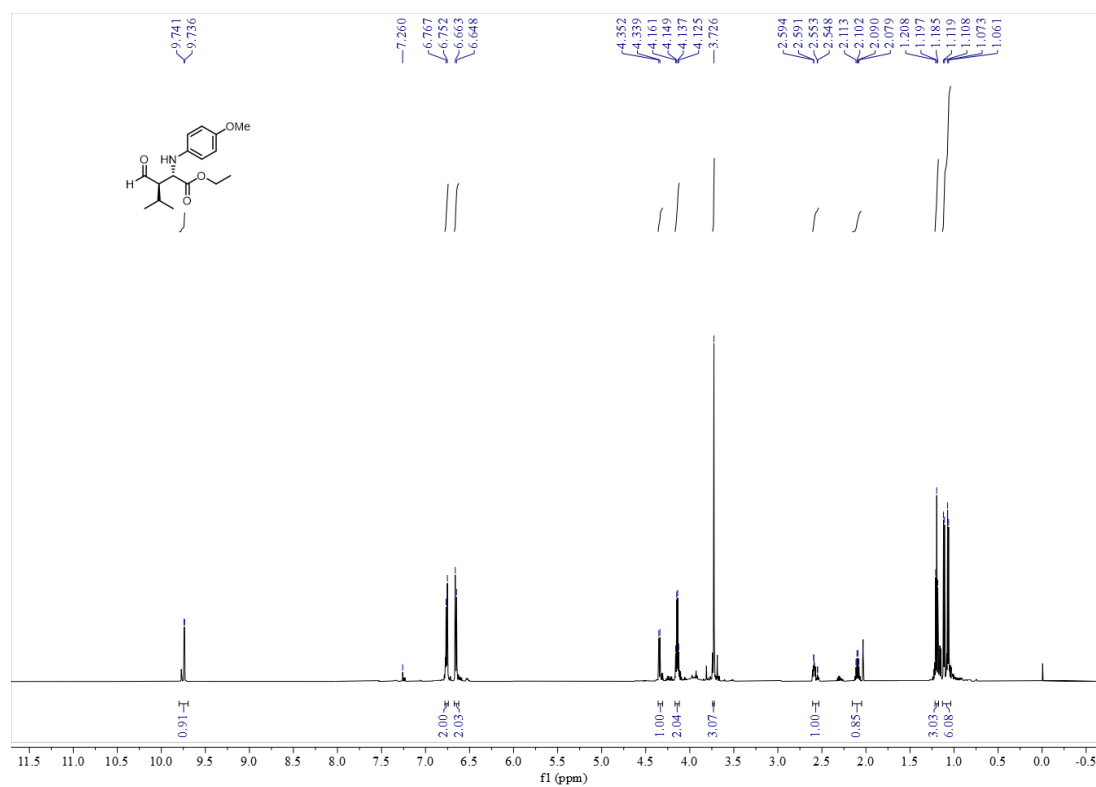
Ethyl (S)-2-((4-methoxyphenyl)amino)-2-((R)-8-oxo-1,4-dioxaspiro[4.5]decan-7-yl)acetate (**3a1**)



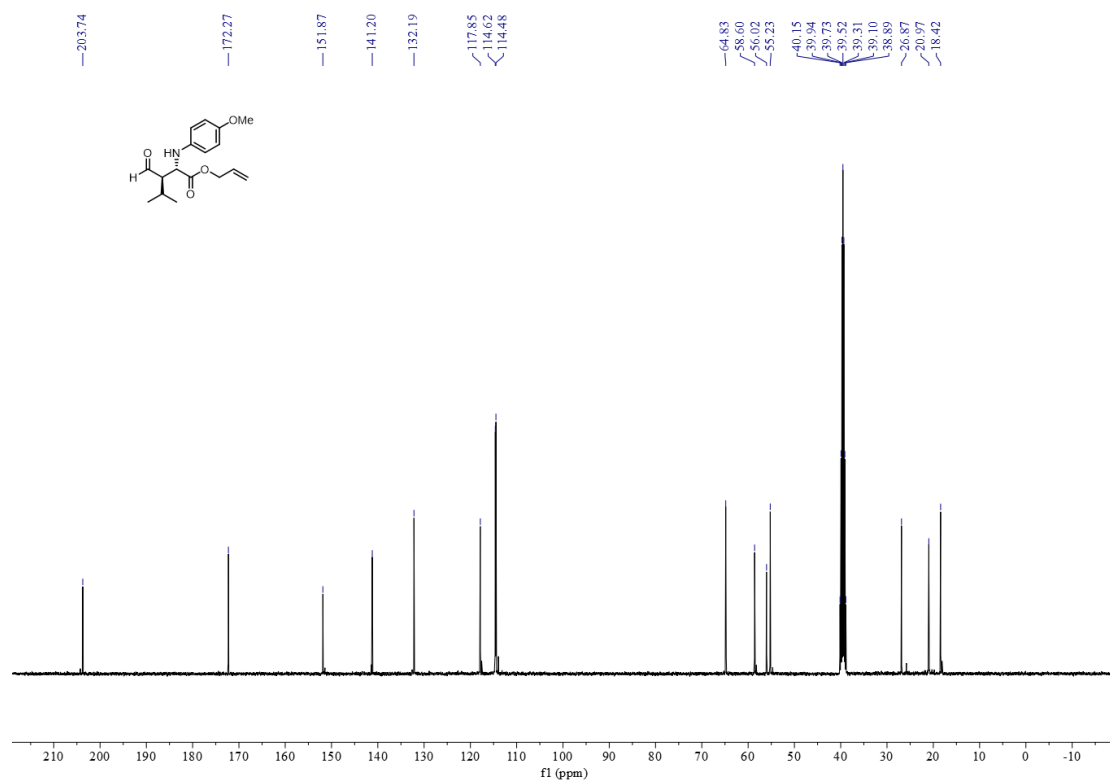
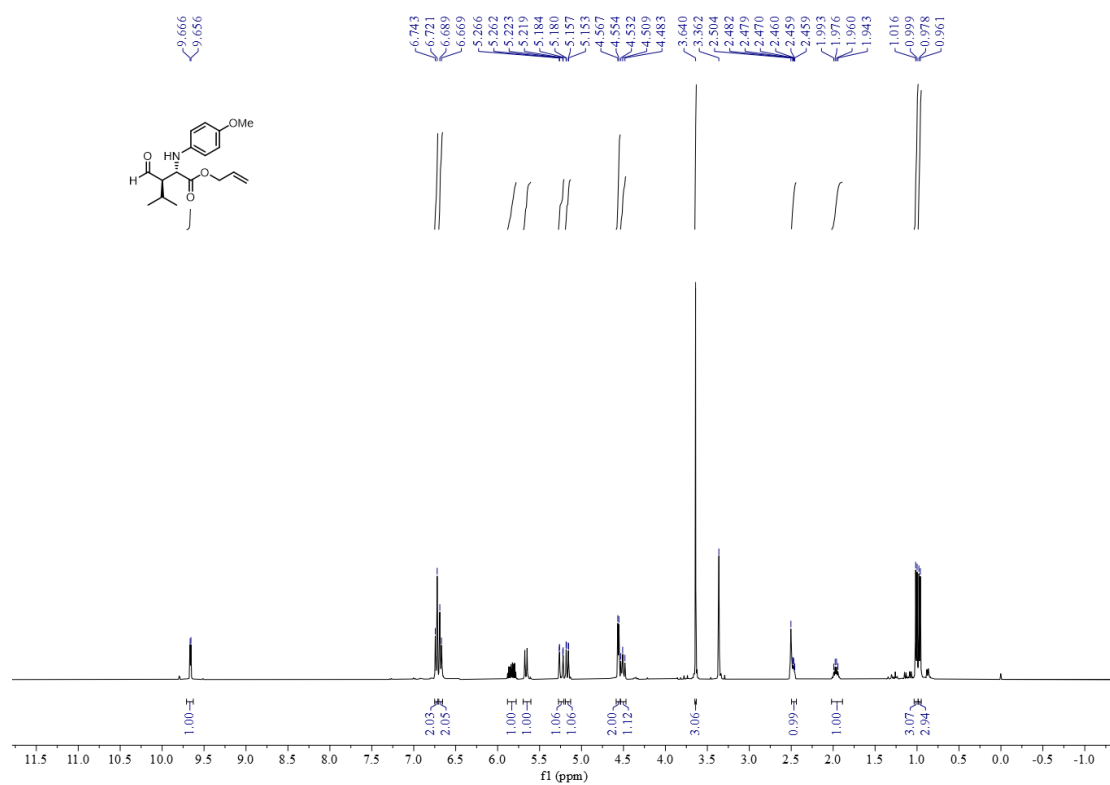
Ethyl (2S,3R)-2-((4-methoxyphenyl)amino)-3-methyl-4-oxohexanoate (**3am**)



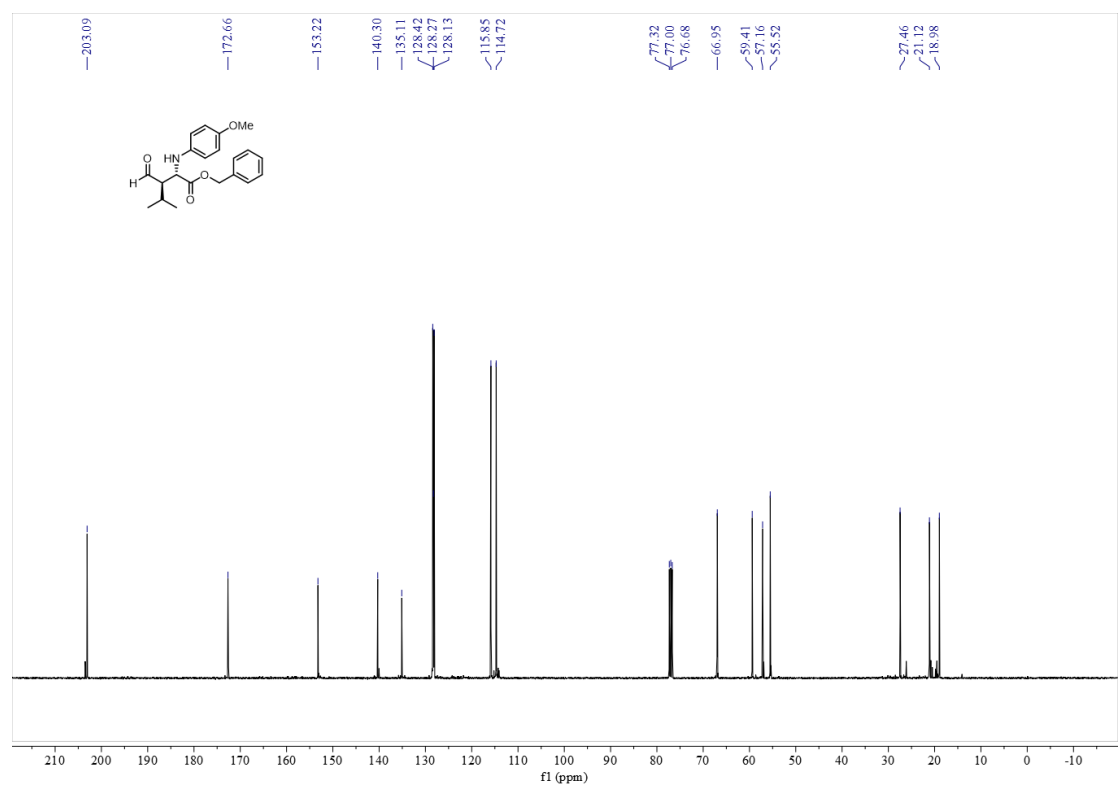
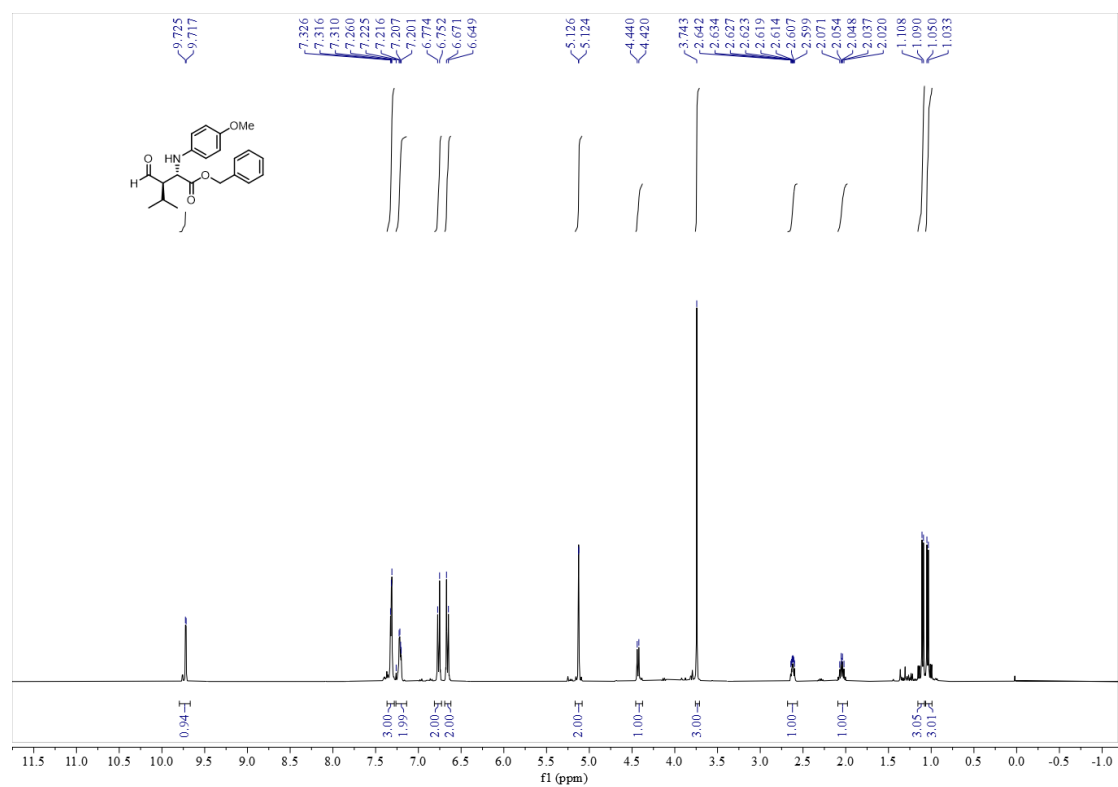
Ethyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)-4-methylpentanoate (**3an**)



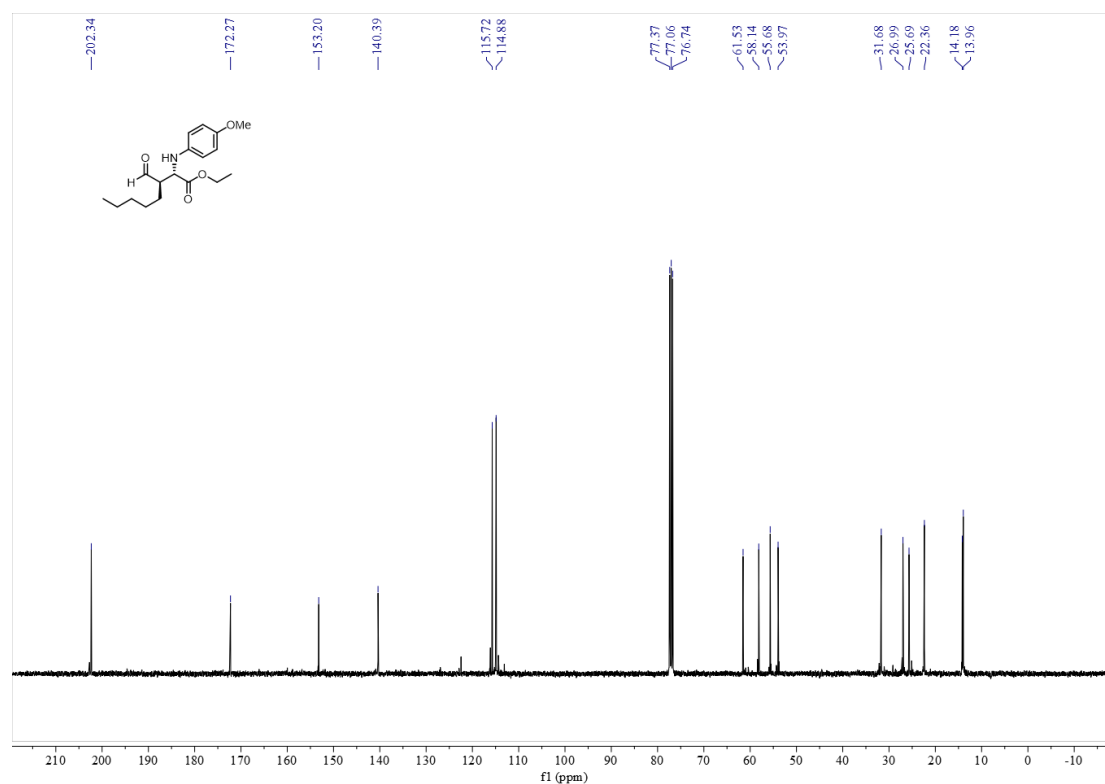
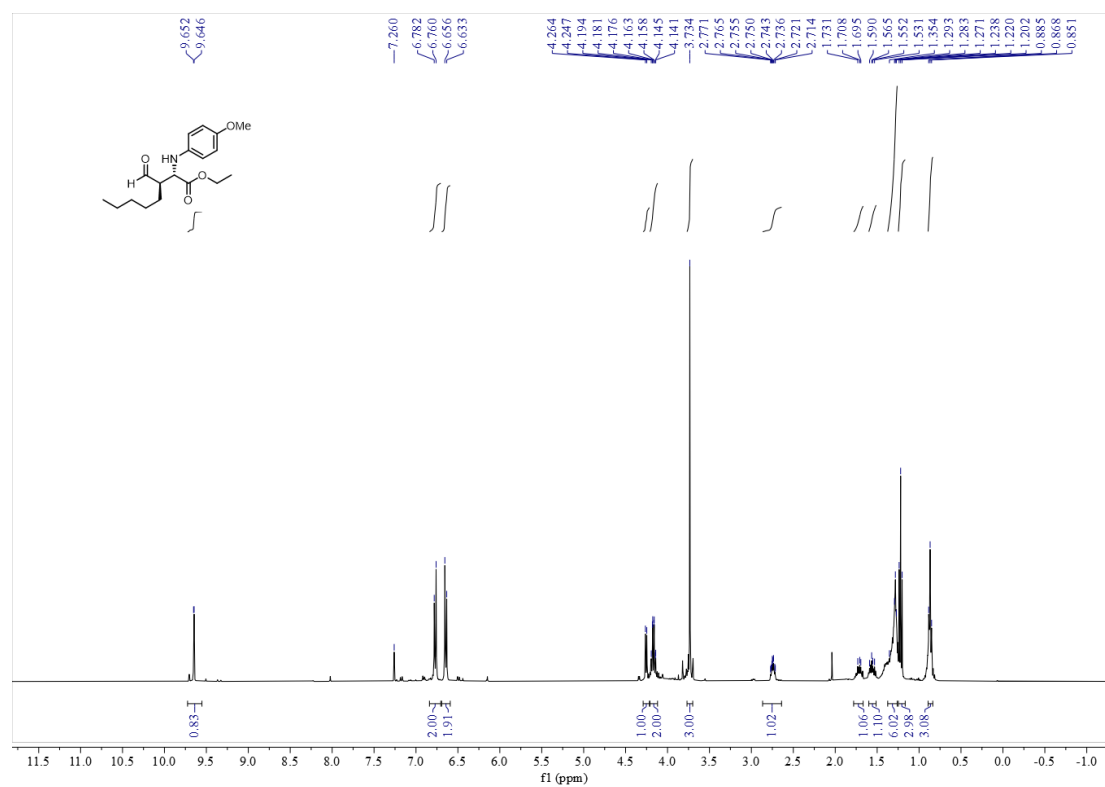
Ethyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)octanoate (**3en**)



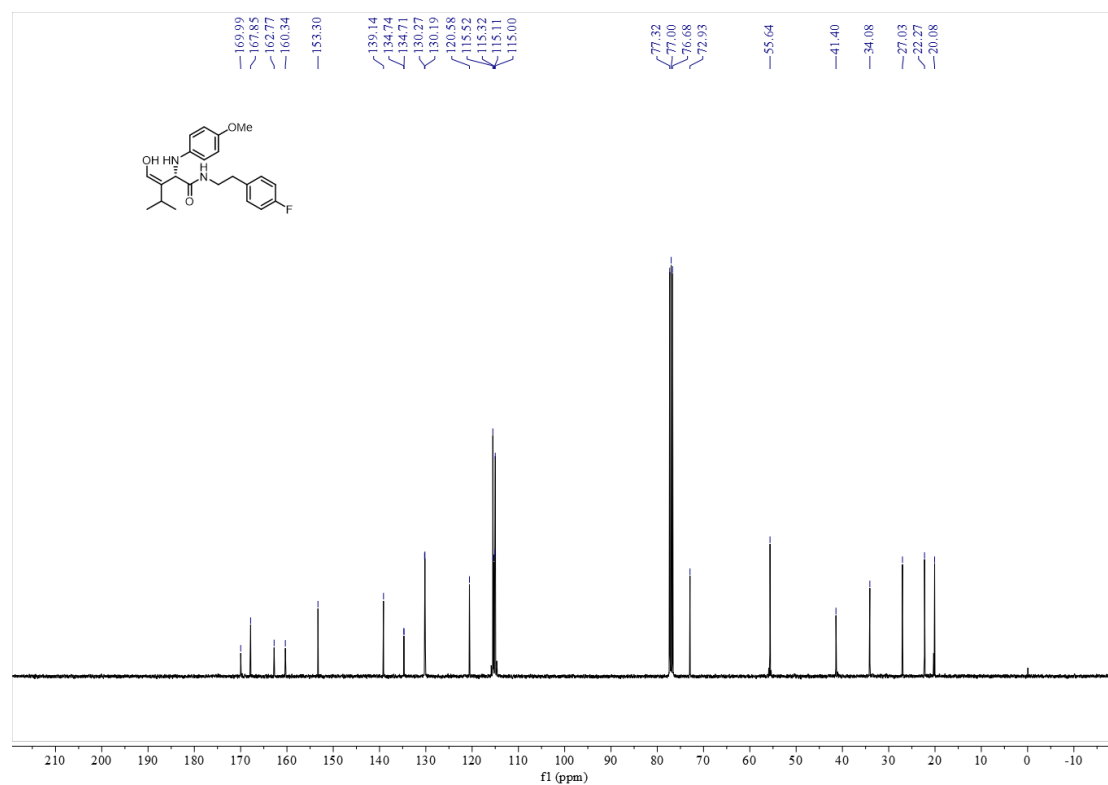
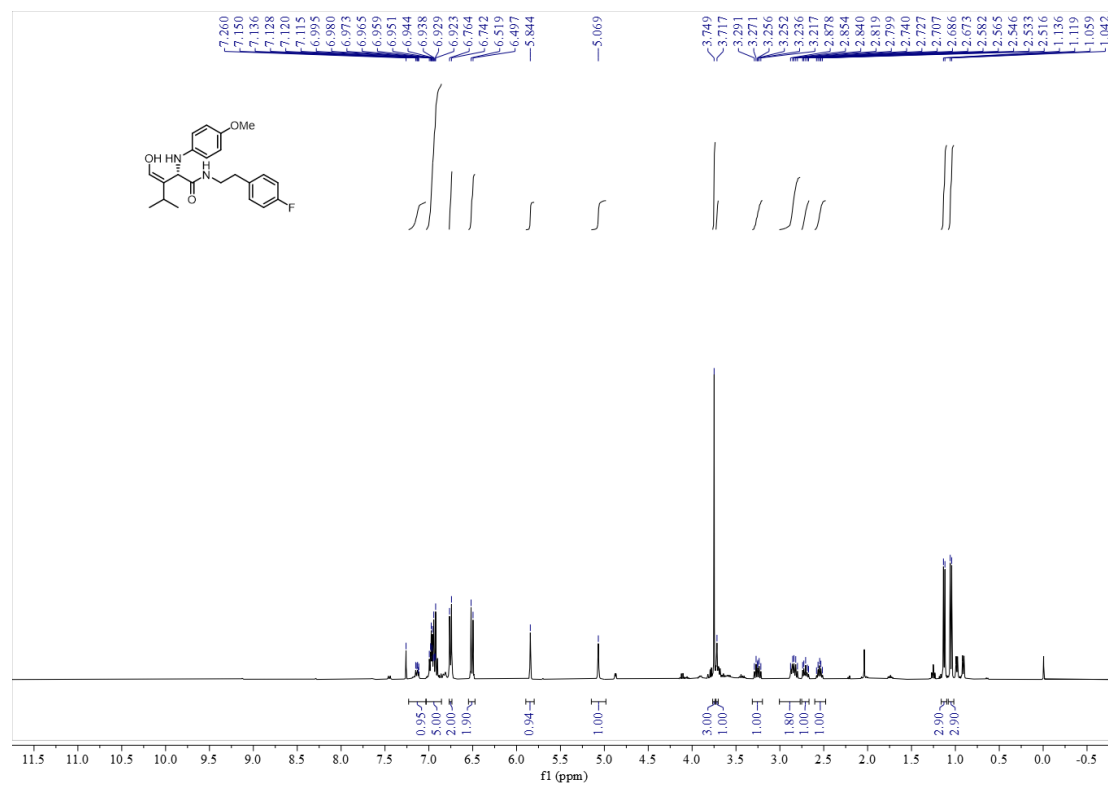
Benzyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)-4-methylpentanoate (**3gn**)

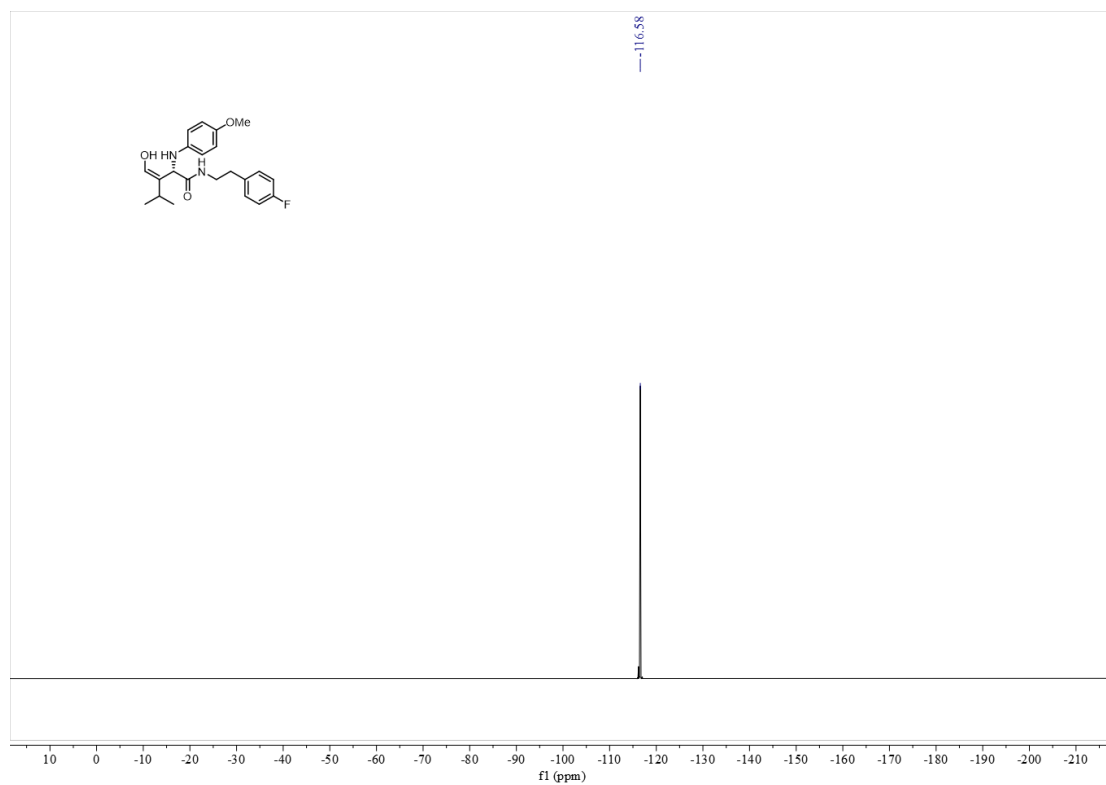


Ethyl (2S,3R)-3-formyl-2-((4-methoxyphenyl)amino)octanoate (**3ao**)

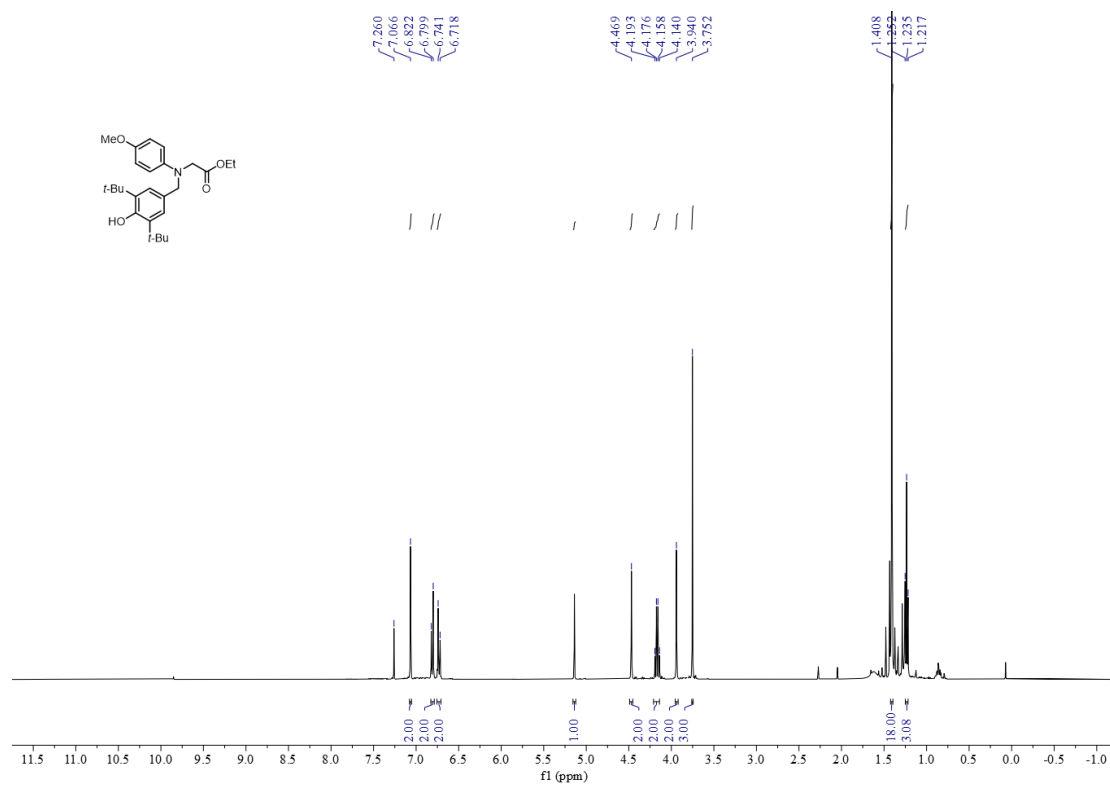


(S,Z)-N-(4-fluorophenethyl)-3-(hydroxymethylene)-2-((4-methoxyphenyl)amino)-4-methylpentanamide
(3pn)

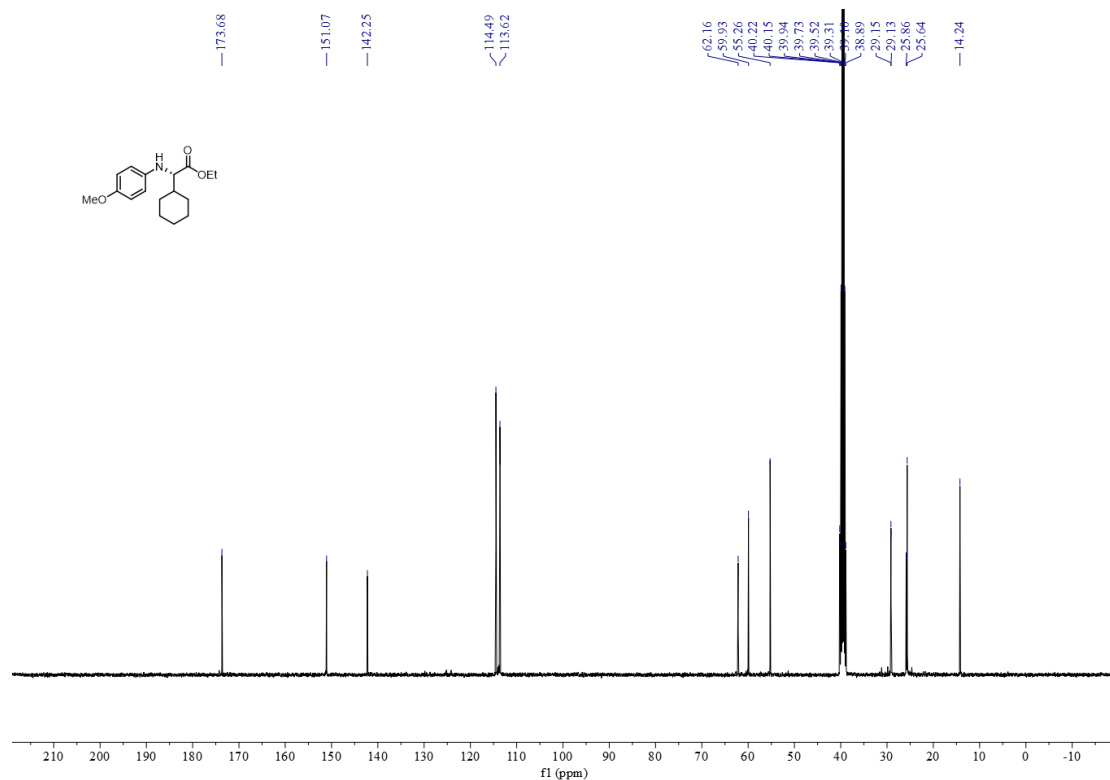
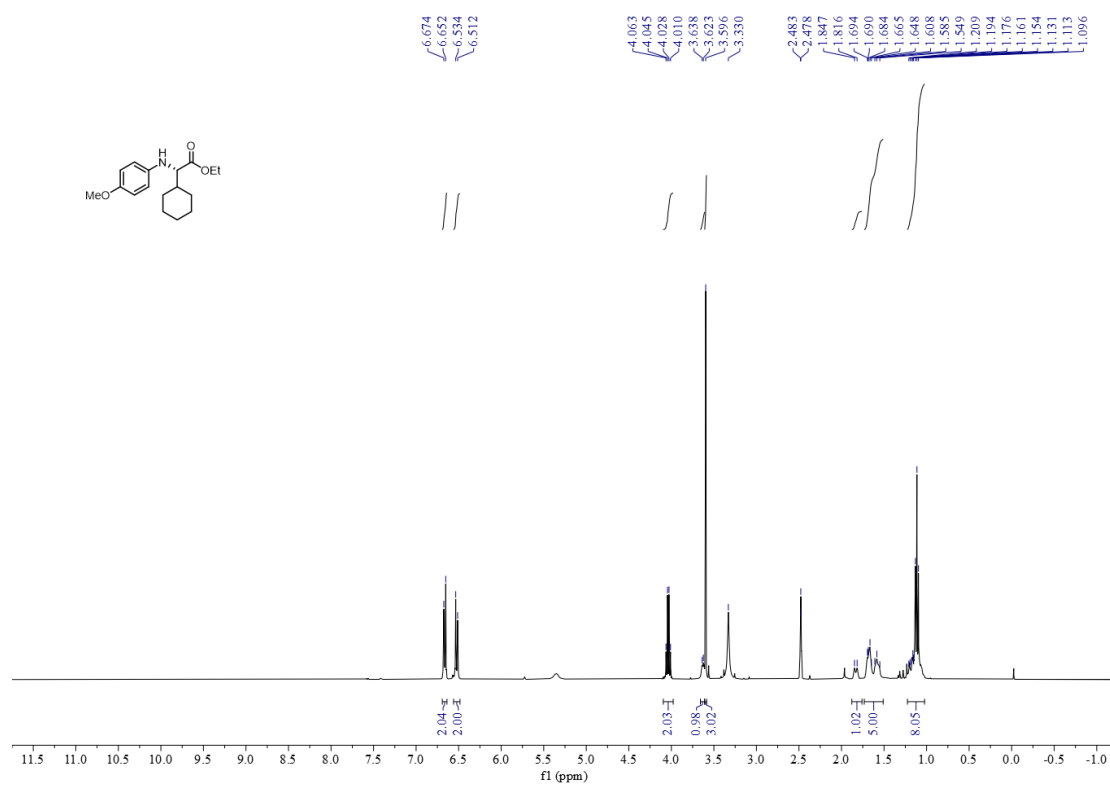




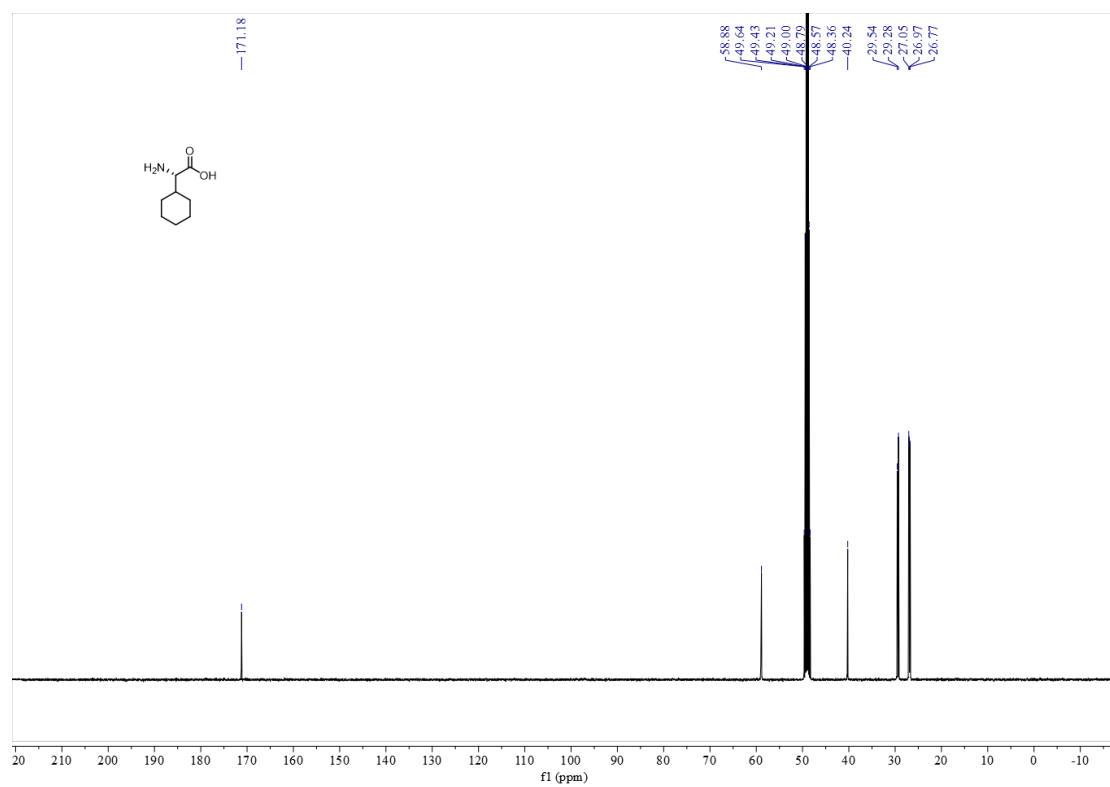
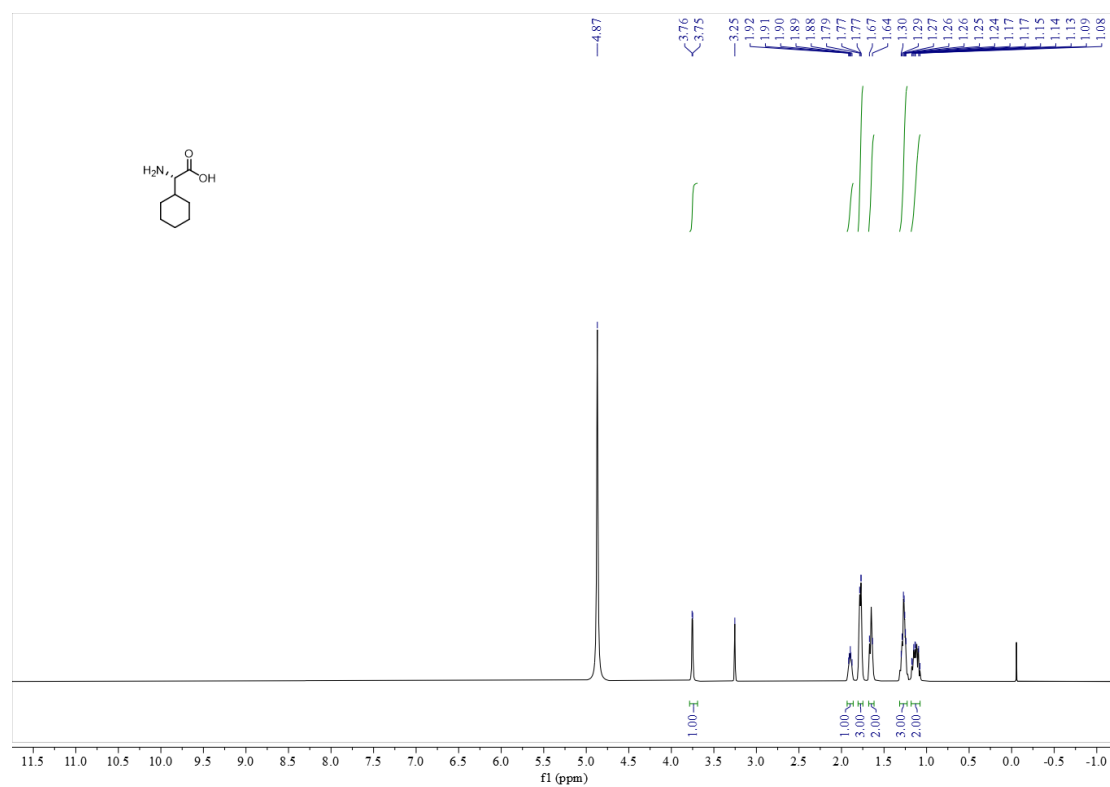
Ethyl N-(3,5-di-tert-butyl-4-hydroxybenzyl)-N-(4-methoxyphenyl)glycinate (**1a-BHT**)



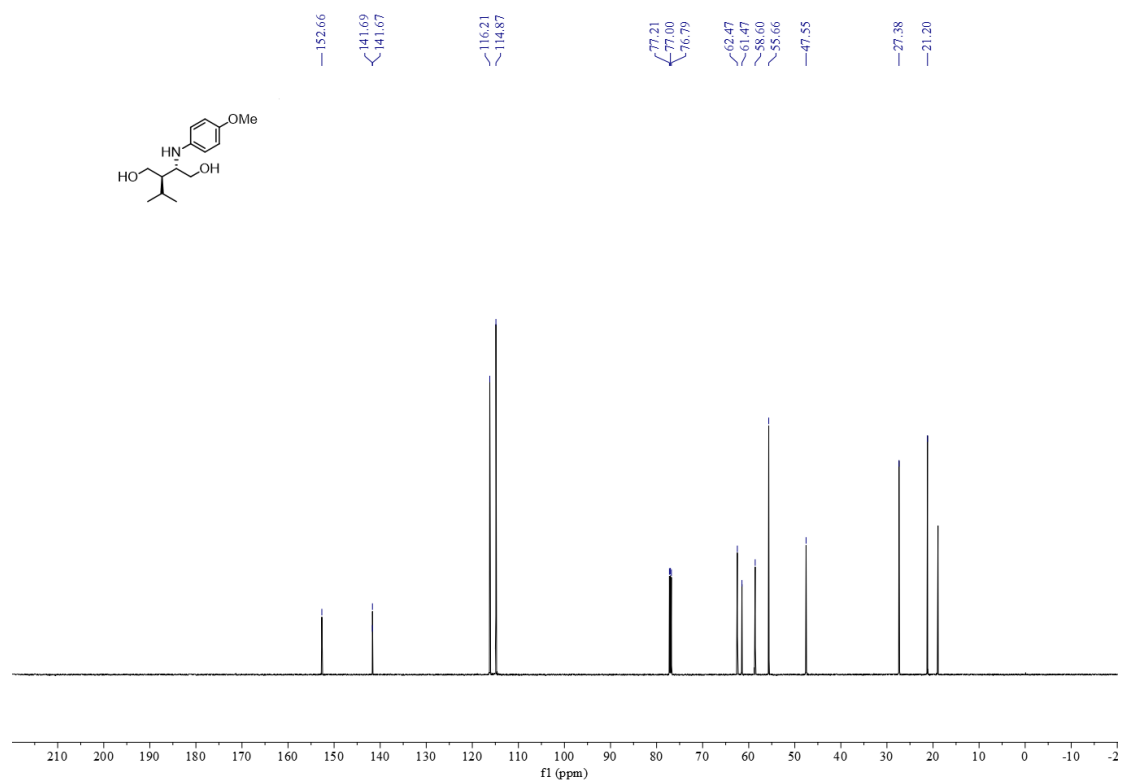
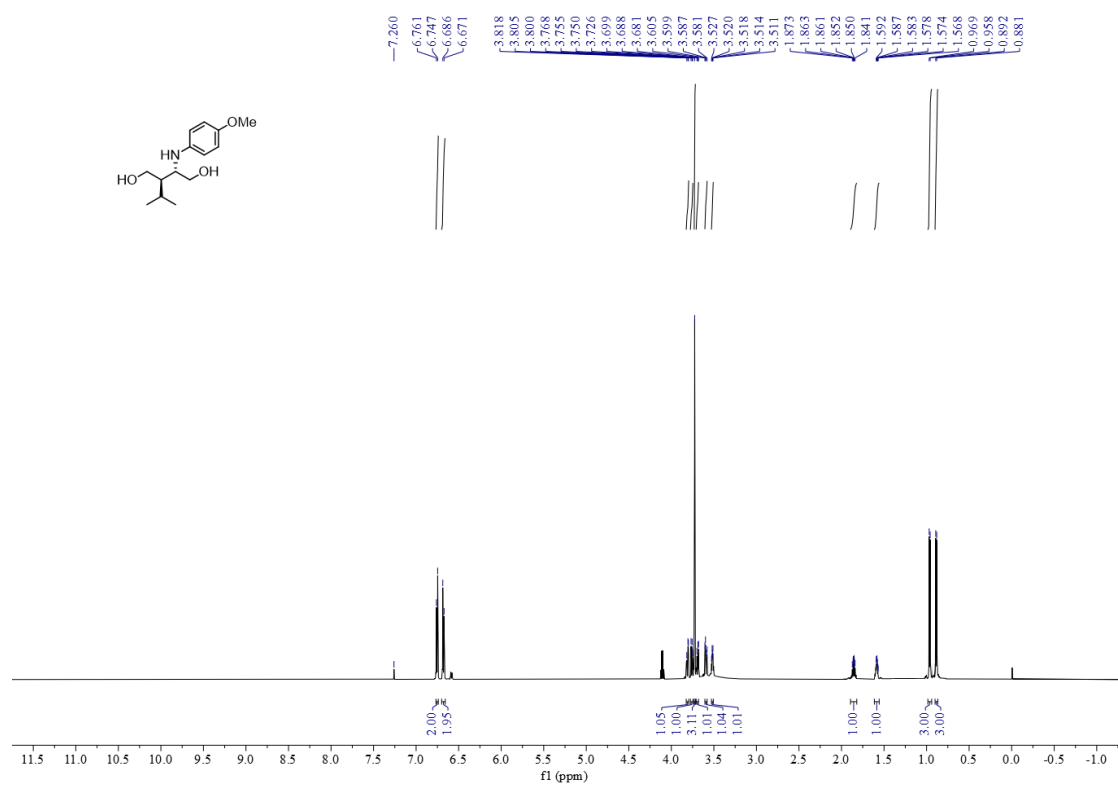
Ethyl (S)-2-cyclohexyl-2-((4-methoxyphenyl)amino)acetate (**4**)



(S)-2-amino-2-cyclohexylacetic acid (**5**)



(2R,3S)-2-Isopropyl-3-((4-methoxyphenyl)amino)butane-1,4-diol (**6**)



6. References

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