

C-H Fluorination Promoted by Pyridine *N*-Oxyl Radical

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1. General Information

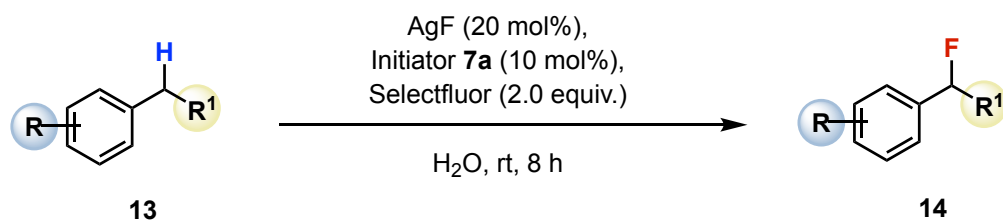
Flash chromatography was performed using silica gel purchased from Qingdao Haiyang. Mixtures of petroleum ether/ethyl acetate or dichloromethane/methanol were used as eluting solvents. AgF and Selectfluor were purchased from Energy Chemical and used as received. H₂O and *tert*-butanol served as reaction solvents, purchased from Adamas, and used as received.

All new compounds were characterized by NMR spectroscopy and high-resolution mass spectroscopy (HRMS). NMR spectra were recorded on a Bruker AMX 400 spectrometer and calibrated using TMS or residual deuterated solvent as an internal reference [CDCl₃: 7.26 ppm or 0.00 ppm (TMS) for ¹H NMR and 77.16 ppm for ¹³C NMR]. The tabulated data were reported in ppm. HRMS spectra were recorded on a Waters Q-TOF Premier.



Figure S1. A Picture of Reaction Vials Used in this Study.

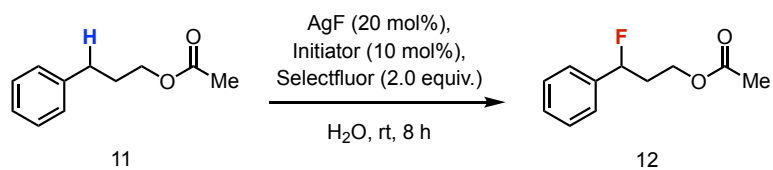
2. General Procedure for C-H Fluorination



General Procedure:

13 (1.0 equiv.), pyridine *N*-oxides **7** as initiator (10 mol%), Selectfluor (2.0 equiv.) and AgF (20 mol %) were weighed into an 8 mL screw-capped vial equipped with a magnetic stir bar. The vial was loosely capped and transferred into a nitrogen-filled glovebox. The solvent was added to the vial. The vial was tightly sealed with a Teflon-lined cap and wrapped with black tape to stir for 8 hours. The reaction was extracted with ethyl acetate (2 mL x 3). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was subsequently subjected to silica gel chromatograph to give desired product **14**.

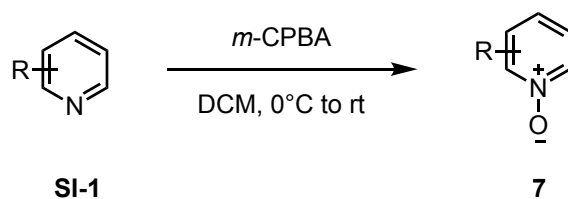
3. Condition Optimization



Entry	Deviation from standard situation	NMR Yield
1	none	85%
2	under air	n. d
3	with natural light	75%
4	2 h	34%
5	KF instead of Selectfluor	n. d
6	KF instead of AgF	n. d
7	add 2 equiv. KF	77%
8	<i>t</i> BuOH/H ₂ O = 1:4	84%
9	<i>t</i> BuOH/H ₂ O = 1:1	75%
10	MeCN	n. d
11	MeCN/H ₂ O = 1:1	22%
12	DCM	n. d

[a] Reactions were performed under N₂ atmosphere with 11 (0.1 mmol), Selectfluor (2.0 equiv.), AgF (20 mol%) and Initiator (10 mol%) in H₂O (1.0 mL) for 8 h. The reaction vial was encased with black tape to establish light-shielding conditions. NMR yields were determined by ¹H NMR spectroscopy with trimethyl benzene-1,3,5-tricarboxylate as an internal standard.

4. Procedure for the Preparation of Pyridine *N*-oxides

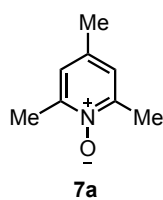


The *N*-oxides used in this study were prepared following the reported procedures as shown below. All the *N*-oxides are known compounds, and the characterization data for these compounds were presented below. Data align consistently with previous reports. ^[1]

Procedure for the Preparation of Pyridine *N*-oxides:

m-CPBA (1.1 equiv.) was added slowly to a solution of pyridine (1.0 equiv.) in DCM at 0 °C, and the reaction mixture was stirred for 12 h at room temperature. The reaction mixture was concentrated, and the residue was subjected to flash-column chromatography (SiO₂, eluted with petroleum ether/ethyl acetate = 1:1 initially, then 5% MeOH in DCM, then 10% MeOH in DCM) to afford pyridine *N*-oxides.

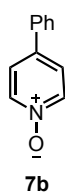
5. Characterization Data for Pyridine *N*-oxides



2,4,6-trimethylpyridine 1-oxide (7a)

Following the **Procedure for the Preparation of Pyridine *N*-oxides**, **SI-1a** (1.2 g, 10 mmol), *m*-CPBA (2.4 g, 12 mmol), and DCM (30 mL) were used. The residue was purified by silica gel column chromatography (hexane: ethyl acetate = 1:1 initially, then 5% MeOH in DCM, then 10% MeOH in DCM) to afford the product as yellow oil.

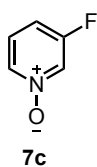
$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 6.96 (s, 2H), 2.51 (s, 6H), 2.28 (s, 3H). $^{13}\text{C NMR}$ (101 MHz, Chloroform-*d*) δ 148.32, 136.39, 132.15, 129.43, 127.90, 124.86, 20.28, 18.27.



4-phenylpyridine 1-oxide (7b)

Following the **Procedure for the Preparation of Pyridine *N*-oxides**, **SI-1b** (1.6 g, 10 mmol), *m*-CPBA (2.4 g, 12 mmol), and DCM (30 mL) were used. The residue was purified by silica gel column chromatography to afford the product as brown solid.

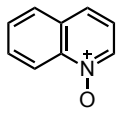
$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 8.26 (d, $J = 7.1$ Hz, 2H), 7.79–7.37 (m, 7H).



3-fluoropyridine 1-oxide (7c)

Following the **Procedure for the Preparation of Pyridine *N*-oxides**, **SI-1c** (970 mg, 10 mmol), *m*-CPBA (2.4 g, 12 mmol), and DCM (30 mL) were used. The residue was purified by silica gel column chromatography to afford the product as white solid.

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 8.19 (dt, $J = 4.3, 2.0$ Hz, 1H), 8.10 (d, $J = 8.7$ Hz, 1H), 7.33–7.25 (m, 1H), 7.15–7.07 (m, 1H).



7d

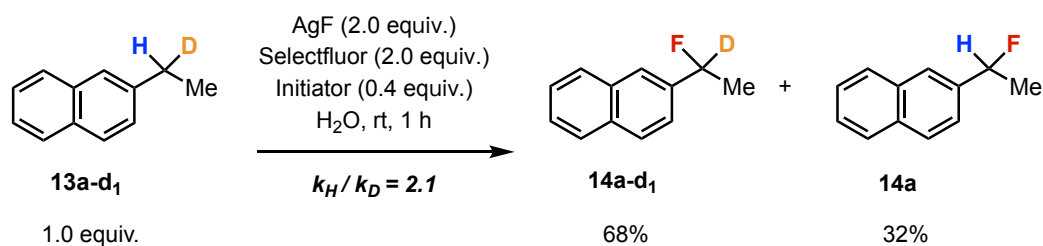
3-(trifluoromethyl)pyridine 1-oxide (7d)

Following the **Procedure for the Preparation of Pyridine *N*-oxides**, **SI-1d** (1.3 g, 10 mmol), *m*-CPBA (2.4 g, 12 mmol), and DCM (30 mL) were used. The residue was purified by silica gel column chromatography (hexane: ethyl acetate =1:1) to afford the product as brown solid.

¹H NMR (400 MHz, Chloroform-*d*) δ 8.75 (d, $J = 8.8$ Hz, 1H), 8.54 (d, $J = 6.0$ Hz, 1H), 7.87 (d, $J = 8.2$ Hz, 1H), 7.82–7.70 (m, 2H), 7.64 (t, $J = 7.6$ Hz, 1H), 7.33–7.25 (m, 1H).

6. Mechanistic Studies

The Intramolecular Kinetic Isotope Effect Study



13a-d₁ (1.0 equiv.), **7a** (0.4 equiv.), Selectfluor (2.0 equiv.) and AgF (2.0 equiv.) were weighed into an 8 mL screw-capped vial containing a magnetic stir bar. The vial was loosely capped and transferred into a nitrogen-filled glovebox. H₂O was added to the vial, and it was tightly sealed with a Teflon-lined cap. The entire setup was then wrapped with black tape and stirred for 1 hour. The ratio of **14a-d₁** : **14a** was determined by ¹⁹F NMR on a Bruker AMX 400 spectrometer instrument to be 68:32.

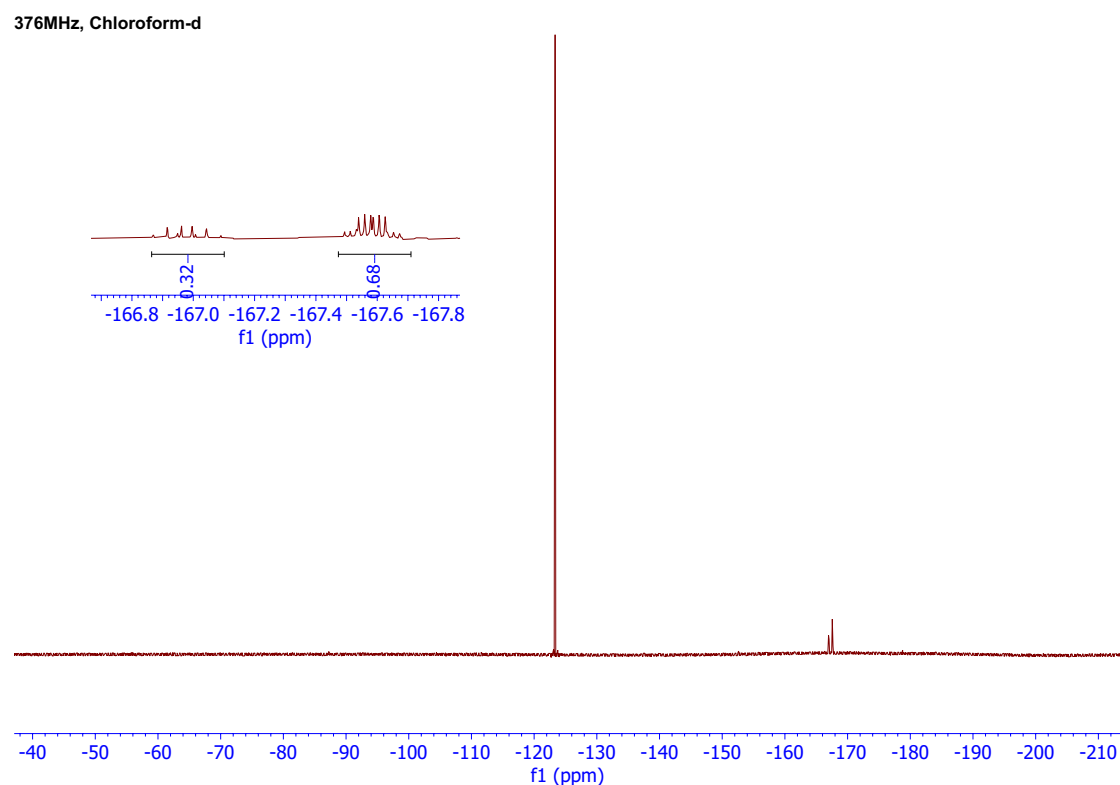
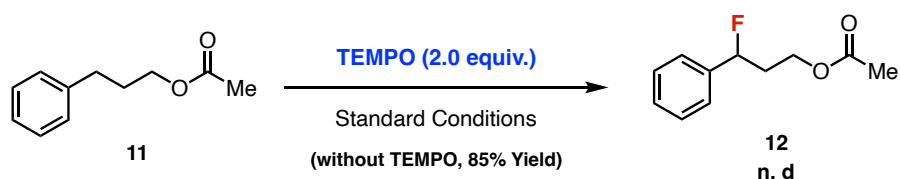


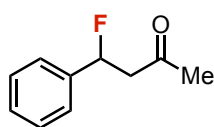
Figure S2. ¹⁹F NMR of the Intramolecular Kinetic Isotope Effect Study.

Radical Trap Experiments



According to the **general procedure**, **11** (1.0 equiv.), pyridine *N*-oxides **7a** as initiator (10 mol%), Selectfluor (2.0 equiv.), AgF (20 mol %) and TEMPO(2.0 equiv.) were weighed into an 8 mL screw-capped vial equipped with a magnetic stir bar. The vial was loosely capped and transferred into a nitrogen-filled glovebox. The solvent was added to the vial. The vial was tightly sealed with a Teflon-lined cap and wrapped with black tape to stir for 8 hours. The reaction was extracted with ethyl acetate (2 mL x 3). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. Fluorination product **12** was not detected.

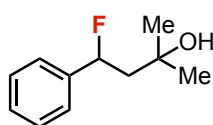
7. Characterization Data for C-H Fluorination Products



4-fluoro-4-phenylbutan-2-one (14a)

Following the **General Procedure**, **13a** (14.8 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. **14a** was obtained as yellow solid. (94% NMR yield). Once the residue was purified by silica gel column chromatography (hexane: EtOAc = 10:1), a 1:1 mixture of **14a** and trans-4-phenyl-3-buten-2-one (elimination product) was obtained when purified by silica gel chromatograph.^[2]

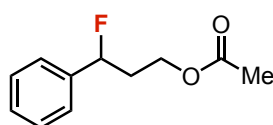
¹H NMR (400 MHz, Chloroform-*d*) δ 7.41–7.33 (m, 5H), 5.95 (ddd, $J = 46.9, 8.8, 4.0$ Hz, 1H), 3.20 (ddd, $J = 16.7, 14.7, 8.8$ Hz, 1H), 2.82 (ddd, $J = 32.1, 16.7, 4.0$ Hz, 1H), 2.22 (s, 3H). **¹⁹F NMR (376MHz, Chloroform-*d*)** δ -173.87 (d, $J = 32.5$ Hz), -173.96 – -174.46 (m). Characterization data are consistent with previous reports.^[3]



4-fluoro-2-methyl-4-phenylbutan-2-ol (14b)

Following the **General Procedure**, **13b** (16.4 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 10:1) to afford the product as colorless oil (12.7 mg, 70% yield).

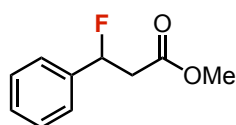
¹H NMR (400 MHz, Chloroform-*d*) δ 7.45–7.24 (m, 5H), 5.79 (ddd, $J = 49.1, 10.2, 2.4$ Hz, 1H), 2.24 (td, $J = 15.7, 10.2$ Hz, 1H), 2.01–1.67 (m, 2H), 1.36 (d, $J = 6.2$ Hz, 6H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 139.67, 139.47, 127.53, 127.49, 127.38, 127.34, 124.45, 124.39, 92.50, 92.48, 90.83, 90.81, 69.14, 49.30, 49.09, 29.73, 29.23. **¹⁹F NMR (376MHz, Chloroform-*d*)** δ -173.31. Characterization data are consistent with previous reports.^[4]



3-fluoro-3-phenylpropyl acetate (14c)

Following the **General Procedure**, **13c** (17.8 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 10:1) to afford the product as colorless oil (16.4mg, 84% yield).

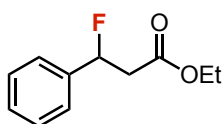
¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 (m, 5H), 5.57 (ddd, $J = 47.8, 8.8, 4.3$ Hz, 1H), 4.32–4.15 (m, 2H), 2.37–2.09 (m, 2H), 2.05 (s, 3H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 170.95, 139.58, 139.39, 128.61, 125.52, 125.45, 92.22, 90.59, 60.50, 60.45, 36.36, 36.12, 20.92. **¹⁹F NMR (376MHz, Chloroform-*d*)** δ -177.39. Characterization data are consistent with previous reports. ^[3]



Methyl-3-fluoro-3-phenylpropanoate (14d)

Following the **General Procedure**, **13d** (16.4 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 10:1) to afford the product as colorless oil. (14.6mg, 80% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.47–7.31 (m, 5H), 5.93 (ddd, $J = 46.9, 9.2, 4.1$ Hz, 1H), 3.74 (s, 3H), 3.04 (ddd, $J = 16.0, 13.5, 9.1$ Hz, 1H), 2.80 (ddd, $J = 32.6, 16.0, 4.1$ Hz, 1H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 170.26, 170.22, 138.80, 138.61, 128.98, 128.96, 128.79, 125.71, 125.65, 91.55, 89.84, 52.15, 42.50, 42.23. **¹⁹F NMR (376MHz, Chloroform-*d*)** δ -173.18. Characterization data are consistent with previous reports. ^[3]

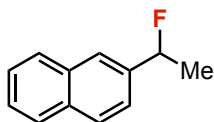


Ethyl-3-fluoro-3-phenylpropanoate (14e)

Following the **General Procedure**, **13e** (17.8 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 10:1) to afford the product as colorless oil (10.4mg, 53% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.38 (d, $J = 6.0$ Hz, 5H), 5.92 (ddd, $J = 47.0, 9.1, 4.3$ Hz, 1H), 4.19 (qd, $J = 7.2, 1.8$ Hz, 2H), 3.03 (ddd, $J = 16.0, 13.6, 9.2$ Hz, 1H), 2.79 (ddd, $J = 32.3, 16.0, 4.2$ Hz, 1H), 1.26 (t, $J = 7.2$ Hz, 3H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 169.74, 138.88, 138.69, 128.93, 128.75, 125.75, 125.68, 91.62, 89.90, 61.08, 42.74, 42.47, 14.22. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -173.02 (ddd, $J = 46.4, 32.4, 13.2$ Hz). Characterization data are consistent with previous reports. ^[5]

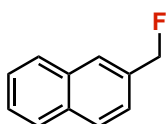
2-(1-fluoroethyl)naphthalene (14f)



Following the **General Procedure, 13f** (15.6 mg, 0.1 mmol), AgF (6.3 mg, 50 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane) to afford the product as yellow oil (7.1mg, 41% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.95–7.75 (m, 4H), 7.48 (ddd, *J* = 9.2, 5.8, 2.0 Hz, 3H), 5.79 (dq, *J* = 47.6, 6.4 Hz, 1H), 1.73 (dd, *J* = 23.8, 6.4 Hz, 3H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 137.89, 137.70, 132.14, 132.06, 127.35, 127.06, 126.69, 125.29, 125.17, 123.19, 123.11, 122.12, 122.06, 90.95, 89.28, 22.07, 21.81. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -166.99 (d, *J* = 2.2 Hz). Characterization data are consistent with previous reports.^[6]

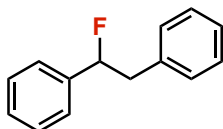
2-(fluoromethyl)naphthalene (14g)



Following the **General Procedures, 13g** (14.2 mg, 0.1 mmol), AgF (6.3 mg, 50 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane) to afford the product as yellow oil (27% NMR yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.92–7.81 (m, 4H), 7.56–7.44 (m, 3H), 5.54 (d, *J* = 47.8 Hz, 2H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 133.80, 133.63, 133.47, 133.21, 128.60, 128.20, 127.87, 126.86, 126.79, 126.58, 126.50, 125.10, 85.71, 84.06. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -206.55, -206.62, -206.76, -206.82, -206.89, -207.03, -207.09. Characterization data are consistent with previous reports.^[7]

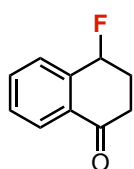
(1-fluoroethane-1,2-diyl)dibenzene (14h)



Following the **General Procedure, 13h** (18.2 mg, 0.1 mmol), AgF (12.1 mg, 1.0 equiv.), **7a** (1.4 mg, 10 mol%), and *t*BuOH/H₂O = 1:1 (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 49:1) to afford the product as colorless oil. (7.8mg, 39% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.38–7.20 (m, 8H), 7.22–7.15 (m, 2H), 5.61 (ddd, *J* = 47.3, 8.1, 4.8 Hz, 1H), 3.27 (ddd, *J* = 17.5, 14.2, 8.1 Hz, 1H), 3.10 (ddd, *J* =

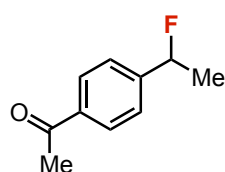
28.5, 14.2, 4.8 Hz, 1H); ^{13}C NMR (101 MHz, Chloroform-*d*) δ 138.86, 138.66, 135.68, 135.64, 128.49, 127.36, 127.35, 127.33, 125.67, 124.67, 124.60, 94.74, 94.71, 93.01, 92.98, 43.04, 42.80; ^{19}F NMR (376 MHz, Chloroform-*d*) δ -173.17. Characterization data are consistent with previous reports^[8]



4-fluoro-3,4-dihydronaphthalen-1(2H)-one (14i)

Following the **General Procedure**, **13i** (14.6 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 8:1) to afford the product as yellow oil (5.9 mg, 36% yield).

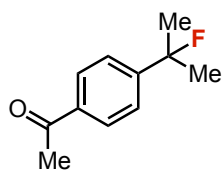
^1H NMR (400 MHz, Chloroform-*d*) δ 8.07 (d, J = 7.8 Hz, 1H), 7.65 (t, J = 7.5 Hz, 1H), 7.59–7.47 (m, 2H), 5.74 (dt, J = 50.4, 5.1 Hz, 1H), 3.03–2.92 (m, 1H), 2.65 (dt, J = 17.3, 6.4 Hz, 1H), 2.55–2.41 (m, 2H). ^{13}C NMR (101 MHz, Chloroform-*d*) 196.68, 140.33, 140.15, 134.24, 131.48, 129.80, 129.77, 128.16, 128.10, 127.22, 88.65, 86.94, 34.03, 33.96, 29.80, 29.45, 29.23,. ^{19}F NMR (376 MHz, Chloroform-*d*) δ -170.27 – -170.58 (m). Characterization data are consistent with previous reports.^[9]



1-(4-(1-fluoroethyl)phenyl)ethan-1-one (14j)

Following the **General Procedure**, **13j** (14.8 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 97:3) to afford the product as yellow oil. (13.4 mg, 81% yield).

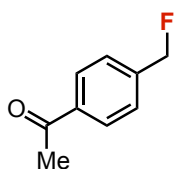
^1H NMR (400 MHz, Chloroform-*d*) δ 7.97 (d, J = 8.1 Hz, 2H), 7.44 (d, J = 8.1 Hz, 2H), 5.68 (dq, J = 47.6, 6.5 Hz, 1H), 2.61 (s, 3H), 1.65 (dd, J = 24.0, 6.5 Hz, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 197.84, 146.88, 146.68, 136.86, 128.71, 125.23, 125.15, 91.25, 89.56, 26.79, 23.23, 22.99. ^{19}F NMR (376 MHz, Chloroform-*d*) -171.22, -170.88– -171.83 (m). Characterization data are consistent with previous reports. ^[2]



1-(4-(2-fluoropropan-2-yl)phenyl)ethan-1-one (14k)

Following the **General Procedure**, **13k** (16.2mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 97:3) to afford the product as yellow oil. (7.4 mg, 41% yield).

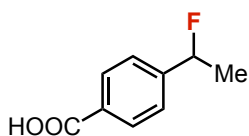
¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, J = 8.2 Hz, 2H), 7.57–7.44 (m, 2H), 2.61 (s, 3H), 1.70 (d, J = 21.9 Hz, 6H). **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -138.79, -138.84, -138.89, -138.94, -138.99, -139.04, -139.09. Characterization data are consistent with previous reports. ^[2]



1-(4-(fluoromethyl)phenyl)ethan-1-one (14l)

Following the **General Procedure**, **13l** (13.4 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 97:3) to afford the product as yellow oil. (9.3mg, 61% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.04–7.87 (m, 2H), 7.45 (dd, J = 8.3, 1.6 Hz, 2H), 5.44 (d, J = 47.2 Hz, 2H), 2.61 (s, 3H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 197.76, 141.53, 141.36, 137.21, 137.18, 128.70, 126.90, 126.84, 84.56, 82.88, 26.76. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -212.86 (d, J = 3.4 Hz), -213.02, -213.06 (t, J = 3.4 Hz), -213.25 (d, J = 3.4 Hz). Characterization data are consistent with previous reports. ^[10]

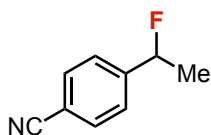


4-(1-fluoroethyl)benzoic acid (14m)

Following the **General Procedure**, **13m** (15.0 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by preparative TLC (hexane: EtOAc = 7:3) to afford the product as white solid. (41% NMR yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.13 (d, J = 7.9 Hz, 2H), 7.44 (s, 2H), 5.70 (dq, J = 47.4, 6.3 Hz, 1H), 1.66 (dd, J = 24.1, 6.5 Hz, 3H). **δ ¹⁹F NMR (376 MHz,**

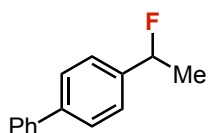
Chloroform-*d*) δ -171.66 (d, J = 4.2 Hz). Characterization data are consistent with previous reports.^[4]



4-(1-fluoroethyl)benzonitrile (14n)

Following the **General Procedure**, **13n** (13.1 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7b** (1.7 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 20:1) to afford the product as yellow oil. (9.4mg, 63% yield).

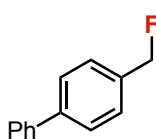
¹H NMR (400 MHz, Chloroform-*d*) δ 7.68 (d, J = 8.1 Hz, 2H), 7.45 (d, J = 8.1 Hz, 2H), 5.67 (dq, J = 47.4, 6.5 Hz, 1H), 1.64 (dd, J = 24.0, 6.4 Hz, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 146.91, 146.71, 132.50, 125.68 (d, J = 7.8 Hz), 118.69, 112.06, 90.82, 89.12, 23.18, 22.94. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -172.67 (d, J = 2.8 Hz). Characterization data are consistent with previous reports. ^[2]



4-(1-fluoroethyl)-1,1'-biphenyl (14o)

Following the **General Procedures**, **13o** (18.2 mg, 0.1 mmol), AgF (2.5 mg, 20 mol%), **7a** (1.4 mg, 10 mol%), and *t*BuOH/H₂O = 1:1 (1 mL) were used. The residue was purified by silica gel column chromatography (hexane) to afford the product as white solid. (69% NMR yield).

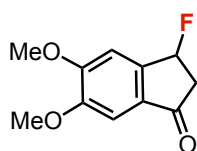
¹H NMR (400 MHz, Chloroform-*d*) δ 7.60 (dd, J = 8.0, 5.8 Hz, 4H), 7.44 (t, J = 7.9 Hz, 4H), 7.41–7.30 (m, 1H), 5.68 (dq, J = 47.7, 6.4 Hz, 1H), 1.69 (dd, J = 23.8, 6.4 Hz, 3H). ¹⁹F NMR (376 MHz, Chloroform-*d*) -166.61 (dq, J = 15.2, 10.1 Hz). Characterization data are consistent with previous reports.^[6]



4-(1-fluoroethyl)-1,1'-biphenyl (14p)

Following the **General Procedure**, **13p** (16.8 mg, 0.1 mmol), AgF (6.3 mg, 50 mol%), **7a** (1.4 mg, 10 mol%), and *t*BuOH/H₂O=1:1 (1mL) were used. The residue was purified by silica gel column chromatography (hexane) to afford the product as white solid. (66% NMR yield).

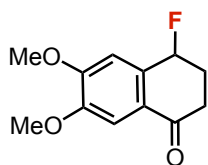
¹H NMR (400 MHz, Chloroform-*d*) δ 7.66–7.55 (m, 4H), 7.50–7.42 (m, 4H), 7.40–7.34 (m, 1H), 5.42 (d, J = 47.9 Hz, 2H). **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -206.03 (d, J = 4.0 Hz), -206.23 (t, J = 3.8 Hz), -206.42 (d, J = 4.1 Hz). Characterization data are consistent with previous reports. ^[7]



3-fluoro-5,6-dimethoxy-2,3-dihydro-1*H*-inden-1-one (14q)

Following the **General Procedure**, **13q** (19.2 mg, 0.1 mmol), AgF (6.3 mg, 50 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 8:1) to afford the product as white solid. (9.2mg, 44% yield).

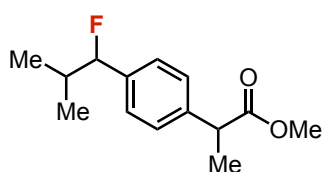
¹H NMR (400 MHz, Chloroform-*d*) δ 7.24–7.10 (m, 2H), 6.10 (ddd, J = 56.4, 6.4, 1.9 Hz, 1H), 4.01 (s, 3H), 3.95 (s, 3H), 3.10 (ddd, J = 18.3, 11.5, 6.4 Hz, 1H), 2.85 (ddd, J = 23.2, 18.9, 2.0 Hz, 1H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 198.78, 154.80, 154.77, 150.90, 144.32, 144.14, 129.39, 129.37, 106.12, 102.63, 87.90, 86.14, 55.44, 55.40, 55.27, 55.23, 43.23, 43.02. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -167.69. **HRMS (ESI)** calculated for C₁₁H₁₂FO₃⁺ [M+H]⁺ m/z 211.0765, found 211.0766.



4-fluoro-6,7-dimethoxy-3,4-dihydronaphthalen-1(2*H*)-one (14r)

Following the **General Procedure**, **13r** (20.6 mg, 0.1 mmol), AgF (6.3 mg, 50 mol%), **7a** (1.4 mg, 10 mol%), and H₂O (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 8:1) to afford the product as yellow solid (8.9mg, 40% yield).

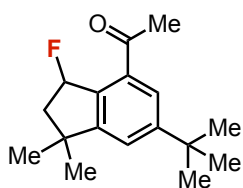
¹H NMR (400 MHz, Chloroform-*d*) δ 7.52 (s, 1H), 6.98 (s, 1H), 5.84–5.57 (m, 1H), 3.97 (d, J = 16.6 Hz, 6H), 3.01–2.80 (m, 1H), 2.61 (dt, J = 10.3, 5.7 Hz, 1H), 2.52–2.33 (m, 2H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 195.59, 154.01, 135.09, 134.92, 125.19, 109.53, 109.47, 108.44, 88.75, 87.04, 56.36, 56.24, 33.72, 33.65, 29.84, 29.62. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -169.30– -169.55 (m). Characterization data are consistent with previous reports. ^[11]



Methyl-2-(4-(1-fluoro-2-methylpropyl)-phenyl)propanoate (14s)

Following the **General Procedure**, **13s** (22.0 mg, 0.1 mmol), AgF (6.3 mg, 50 mol %), **7a** (1.4 mg, 10 mol %), and *t*BuOH/H₂O = 1:1 (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 10:1) to afford the product as colorless oil (16.7mg, 70% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.37–7.13 (m, 4H), 5.08 (dd, *J* = 47.0, 6.8 Hz, 1H), 3.73 (q, *J* = 7.2 Hz, 1H), 3.66 (s, 3H), 2.09 (ddt, *J* = 23.7, 13.6, 6.8 Hz, 1H), 1.50 (d, *J* = 7.2 Hz, 2H), 1.02 (d, *J* = 7.7 Hz, 2H), 0.85 (d, *J* = 6.8 Hz, 3H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 173.88, 139.33, 139.31, 137.37, 137.16, 129.61, 126.30, 125.48, 125.41, 98.93, 98.91, 97.21, 97.18, 51.07, 51.04, 51.01, 50.98, 44.14, 44.11, 33.34, 33.12, 17.55, 17.35, 17.29, 16.54, 16.49. **¹⁹F NMR (376 MHz, Chloroform-*d*)** -179.64 (d, *J* = 8.4 Hz). Characterization data are consistent with previous reports. ^[11]



1-(6-(tert-butyl)-3-fluoro-1,1-dimethyl-2,3-dihydro-1H-inden-4-yl)ethan-1-one (14t)

Following the **General Procedure**, **13t** (24.4 mg, 0.1 mmol), AgF (6.3 mg, 50 mol%), **7a** (1.4 mg, 10 mol%), and *t*BuOH/H₂O = 1:1 (1 mL) were used. The residue was purified by silica gel column chromatography (hexane: EtOAc = 9:1) to afford the product as white solid. (11.0mg, 42% yield)^[11]

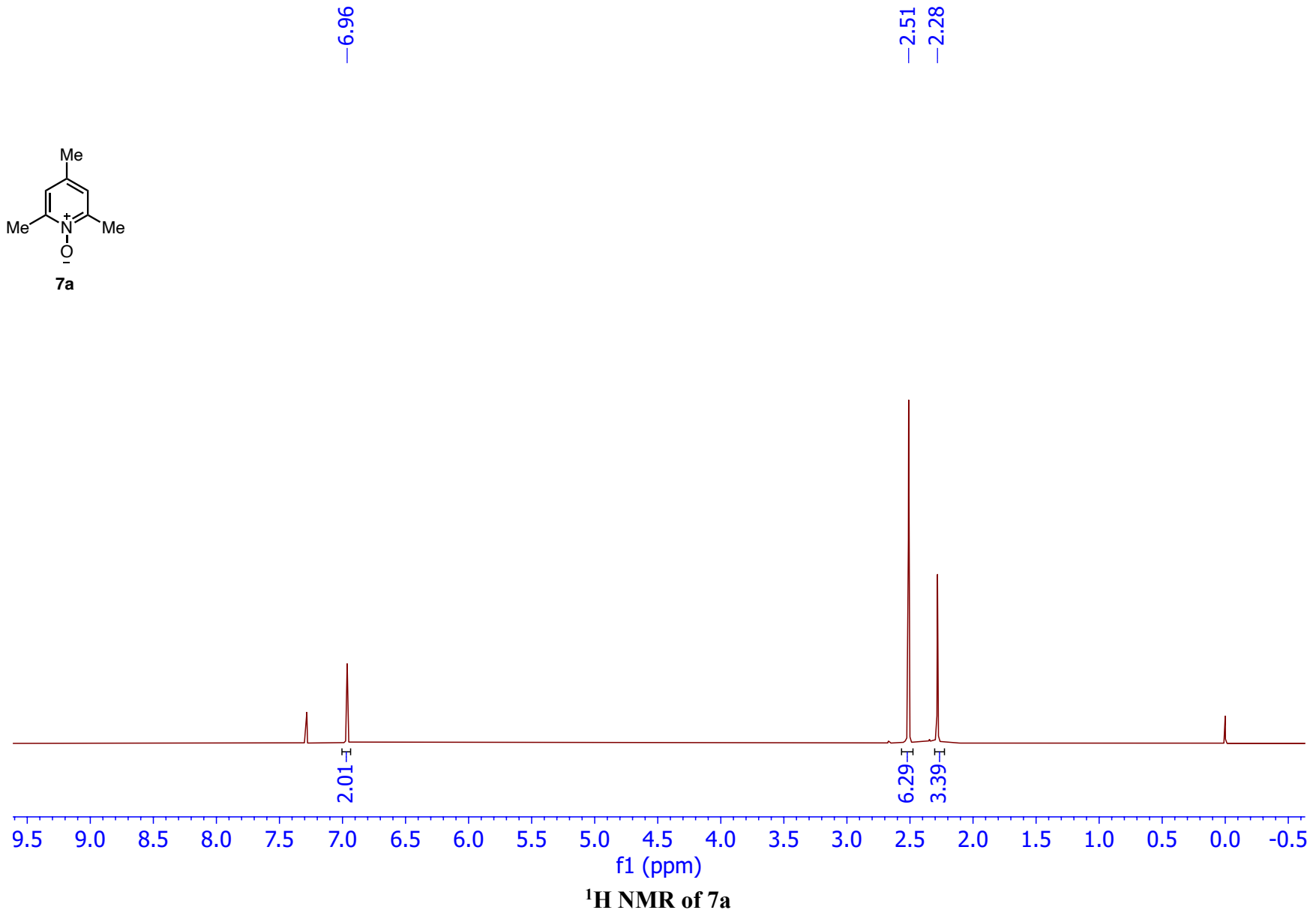
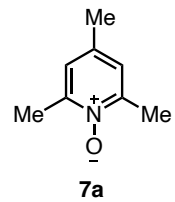
¹H NMR (400 MHz, Chloroform-*d*) δ 7.76 (d, *J* = 1.8 Hz, 1H), 7.43 (t, *J* = 1.6 Hz, 1H), 6.44 (ddd, *J* = 54.0, 6.1, 1.5 Hz, 1H), 2.66 (s, 3H), 2.38–2.08 (m, 2H), 1.36 (d, *J* = 13.6 Hz, 15H). **¹³C NMR (101 MHz, Chloroform-*d*)** δ 198.64, 154.82, 154.77, 153.15, 153.11, 134.12, 133.96, 133.87, 124.56, 124.53, 122.55, 122.52, 93.56, 91.85, 47.46, 47.24, 41.70, 41.68, 34.07, 30.48, 30.34, 28.00, 27.95, 27.61, 27.57. **¹⁹F NMR (376 MHz, Chloroform-*d*)** δ -158.61. Characterization data are consistent with previous reports. ^[11]

8. Reference

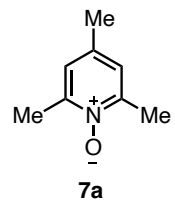
- [1] (a) F. Yang, X. Zhang, F. Li, Z. Wang, L. Wang, *Green Chem.*, **2016**, *18*, 3518-3521; (b) K. D. Kim, J. H. Lee, *Org. Lett.*, **2018**, *20*, 7712-7716; (c) J. Schießl, P. M. Stein, J. Stirn, K. Emler, M. Rudolph, F. Rominger, A. S. K. Hashmi, *Adv. Synth. Catal.*, **2019**, *361*, 725-738.
- [2] J.-B. Xia, C. Zhu, C. Chen, *J. Am. Chem. Soc.*, **2013**, *135*, 17494-17500.
- [3] S. Bloom, C. R. Pitts, R. Woltornist, A. Griswold, M. G. Holl, T. Lectka, *Org. Lett.*, **2013**, *15*, 1722-1724.
- [4] D. Cantillo, O. de Frutos, J. A. Rincón, C. Mateos, C. O. Kappe, *J. Org. Chem.*, **2014**, *79*, 8486-8490.
- [5] A. Mandal, J. Jang, B. Yang, H. Kim, K. Shin, *Org. Lett.*, **2023**, *25*, 195-199.
- [6] E. Emer, L. Pfeifer, J. M. Brown, V. Gouverneur, *Angew. Chem. Int. Ed.*, **2014**, *53*, 4181-4185.
- [7] Y.-M. Su, G.-S. Feng, Z.-Y. Wang, Q. Lan, X.-S. Wang, *Angew. Chem. Int. Ed.*, **2015**, *54*, 6003-6007.
- [8] J. Sheng, H.-Q. Ni, H.-R. Zhang, K.-F. Zhang, Y.-N. Wang, X.-S. Wang, *Angew. Chem. Int. Ed.*, **2018**, *57*, 7634-7639.
- [9] V. Dinoiu, T. Fukuhara, K. Miura, N. Yoneda, *J. Fluorine Chem.*, **2003**, *121*, 227-231.
- [10] L. An, Y.-L. Xiao, Q.-Q. Min, X. Zhang, *Angew. Chem. Int. Ed.*, **2015**, *54*, 9079-9083.
- [11] W. Liu, J. T. Groves, *Angew. Chem. Int. Ed.*, **2013**, *52*, 6024-6027.

9. NMR Spectra

400 HMz, Chloroform-d



101 MHz, Chloroform-d

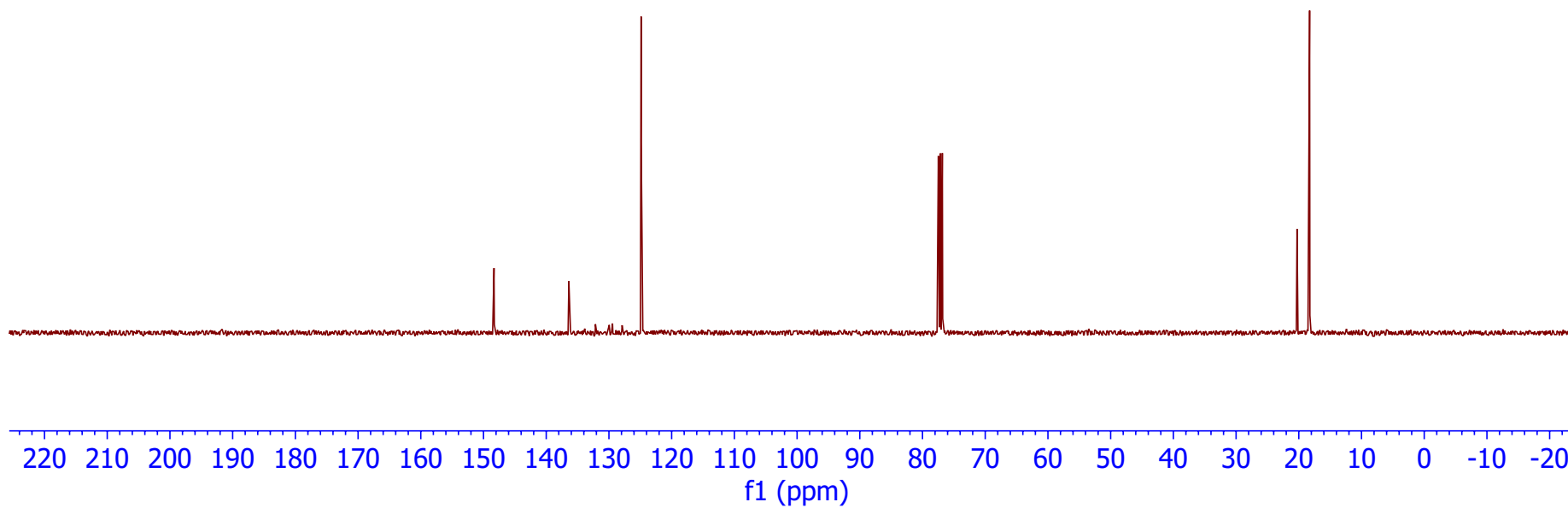


-148.32

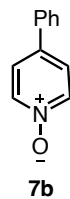
-136.39

-124.86

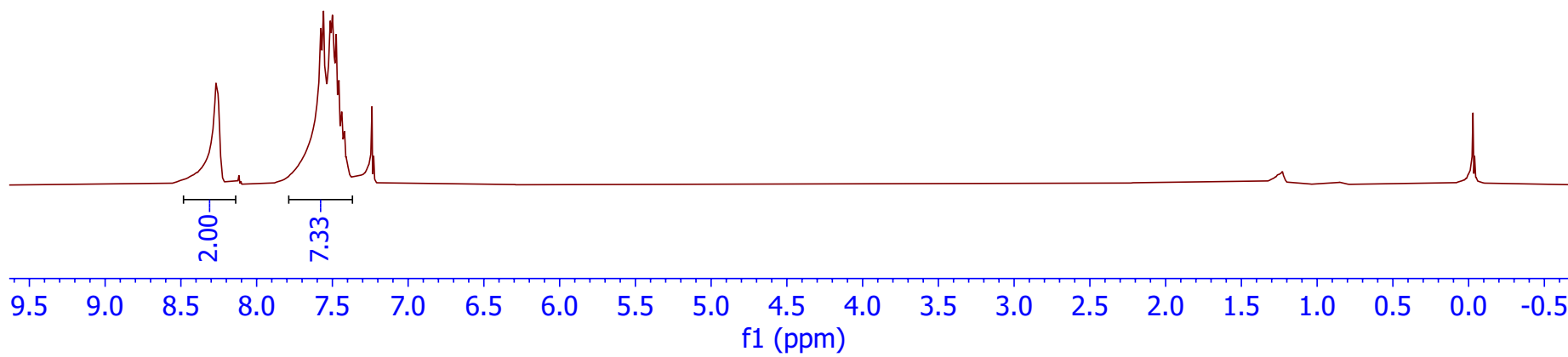
20.28
18.27



400 HMz, Chloroform-d



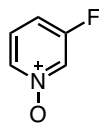
8.27
8.25
7.57
7.56
7.52
7.50
7.47
7.45
7.44
7.42



¹H NMR of 7b

400 HMz, Chloroform-d

8.20
8.19
8.19
8.18
8.18
8.11
8.09
7.31
7.29
7.27
7.25
7.13
7.12
7.11
7.10
7.09
7.08



7c

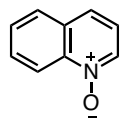


9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5

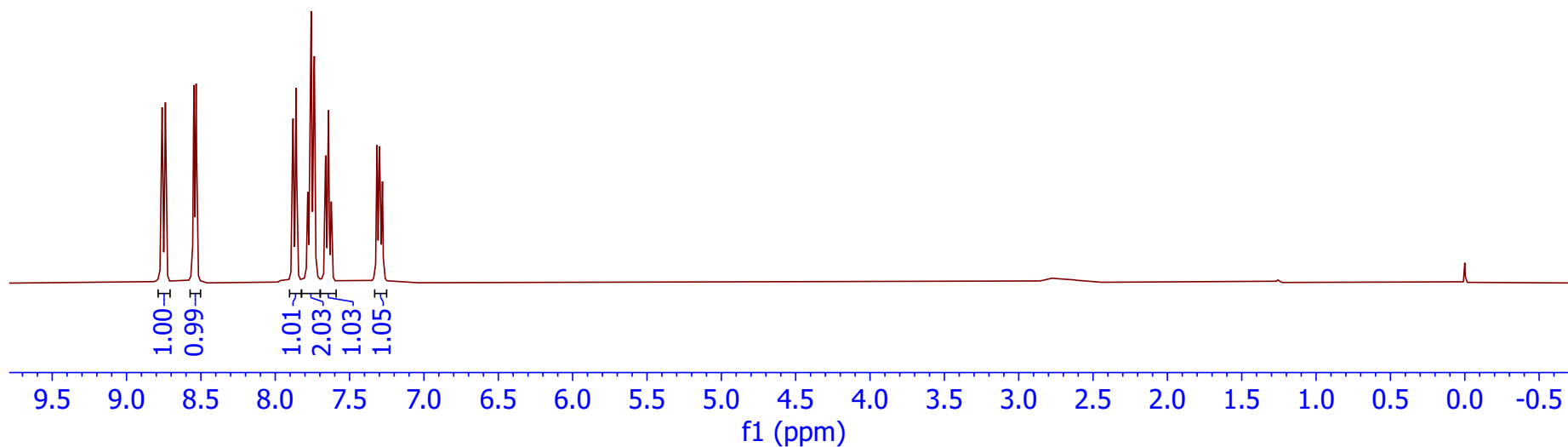
f1 (ppm)
¹H NMR of 7c

400 HMz, Chloroform-d

8.76
8.74
8.55
8.53
7.88
7.86
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7.76
7.76
7.74
7.74
7.66
7.64
7.62
7.32
7.30
7.28

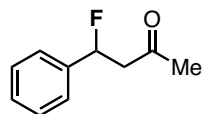


7d

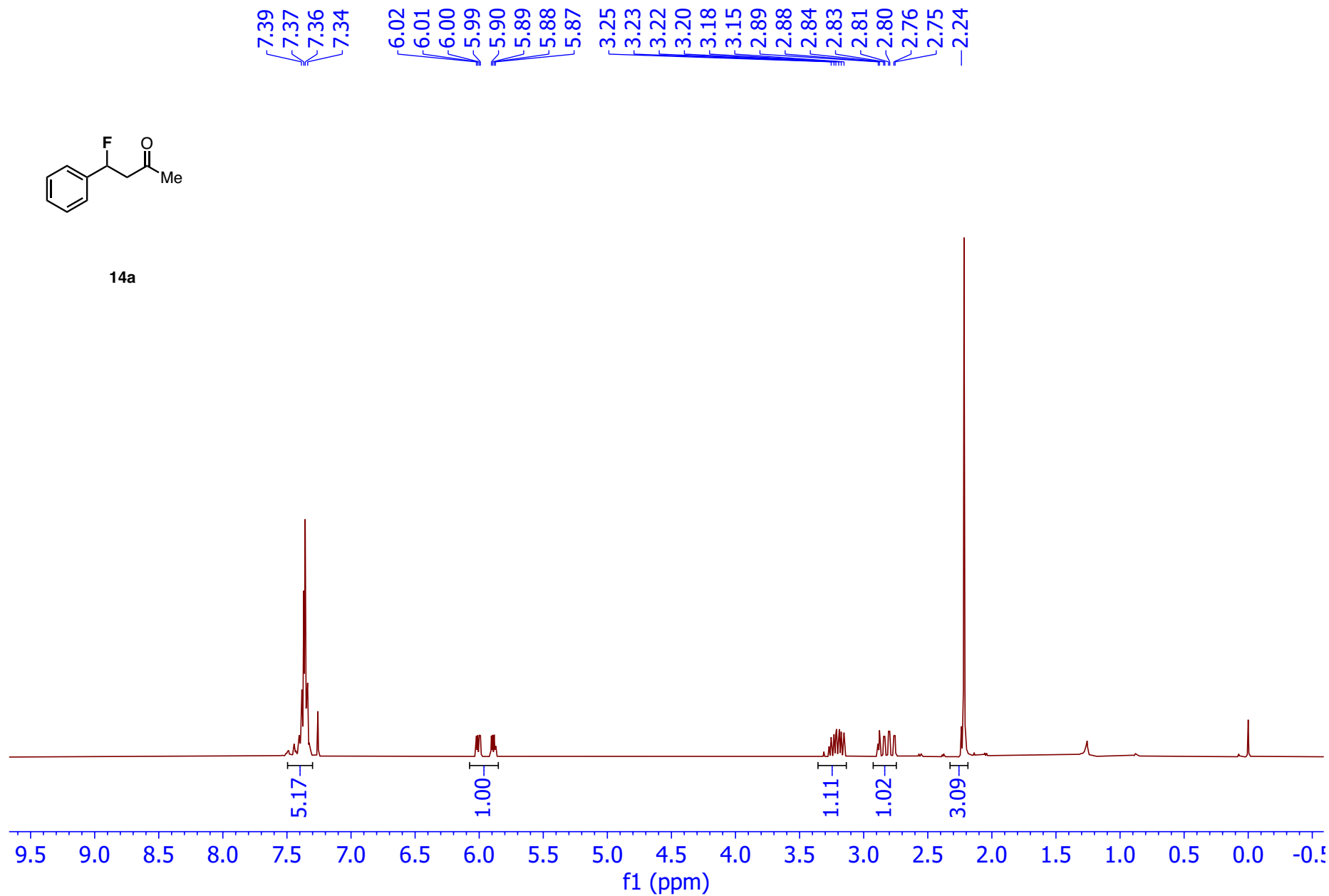


¹H NMR of 7d

400 HMz, Chloroform-d

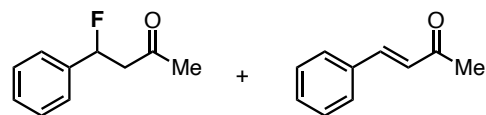


14a



¹H NMR of 14a

400 HMz, Chloroform-d

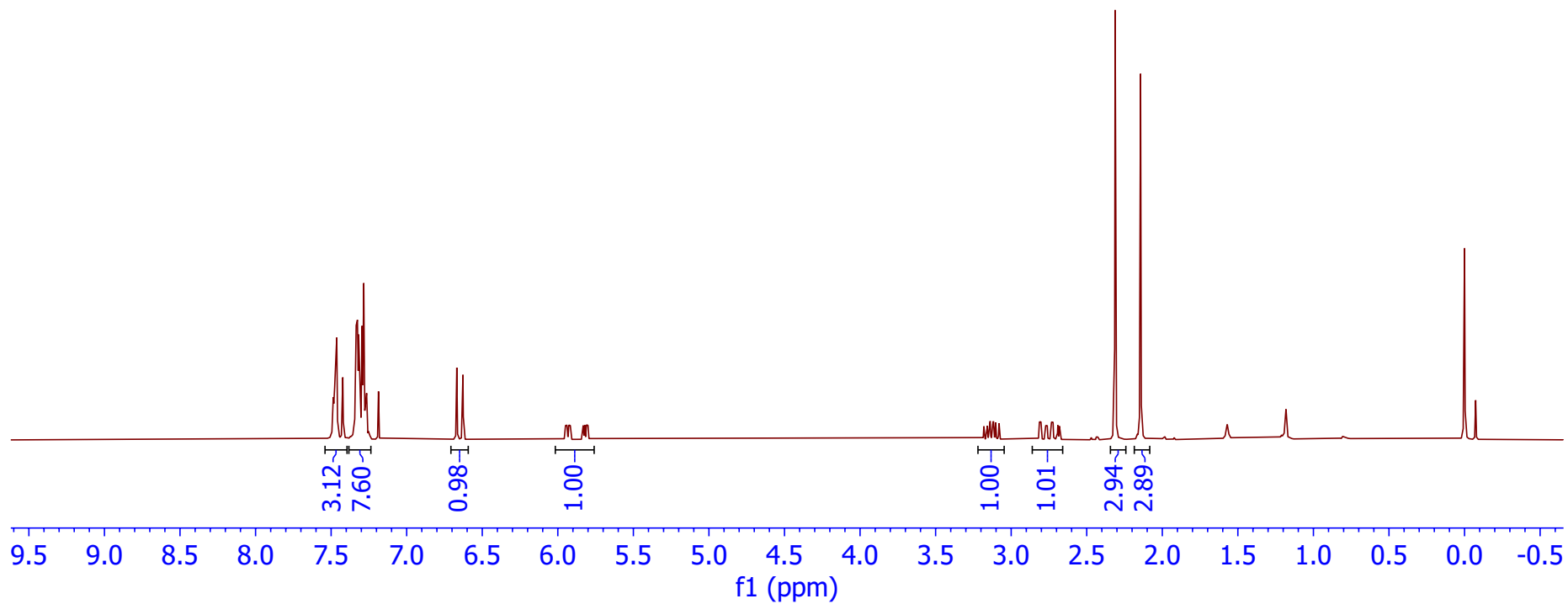


14a

14a-byproduct

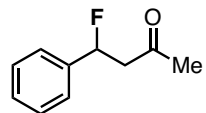
7.46
7.42
7.33
7.33
7.30
7.28
7.26
6.67
6.63
5.95
5.94
5.93
5.92
5.83
5.82
5.81
5.80

3.18
3.16
3.14
3.14
3.12
3.12
3.10
3.08
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2.80
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2.76
2.73
2.72
2.69
2.68
2.31
2.14



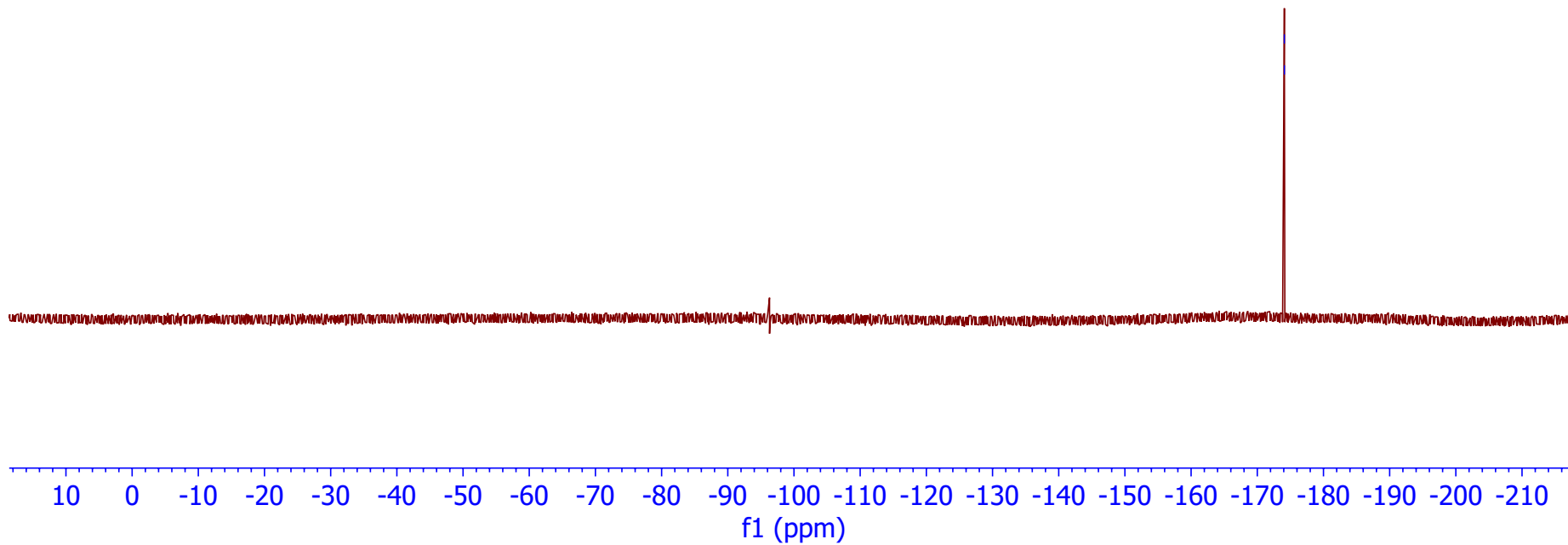
¹H NMR of 14a

376MHz, Chloroform-d



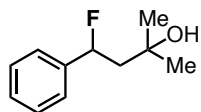
14a

-174.09
-174.10



^{19}F NMR of 14a

400 HMz, Chloroform-d

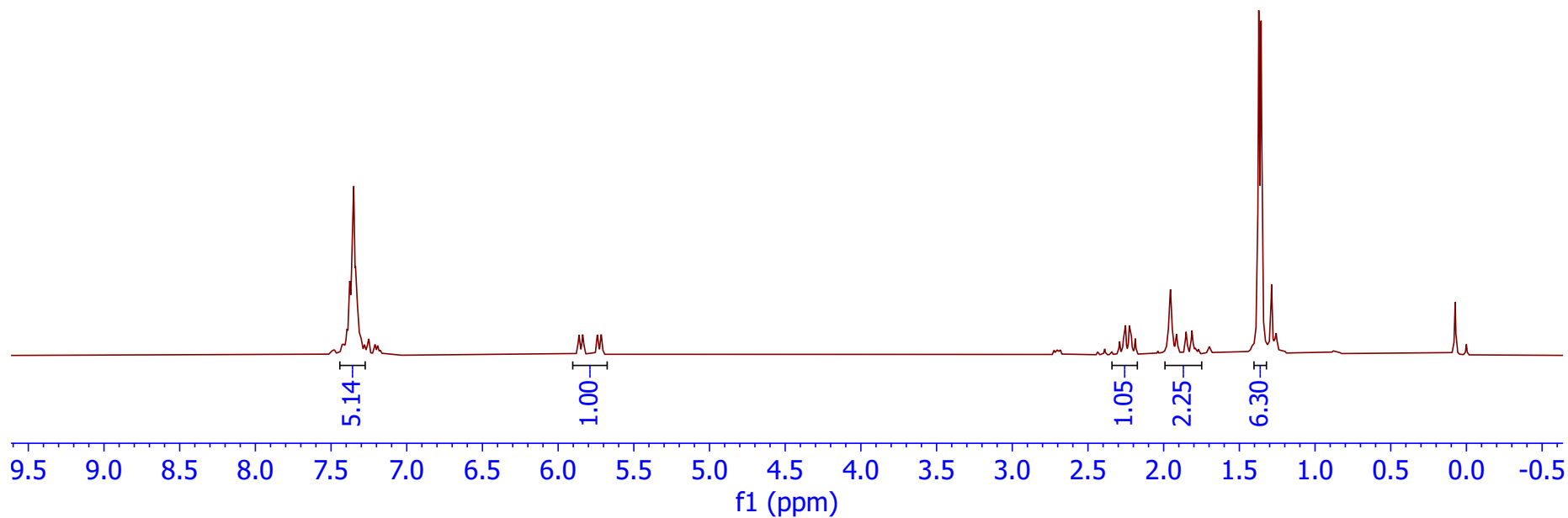


14b

7.38
7.36
7.35
7.34
7.33
7.32

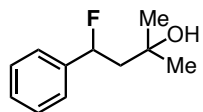
5.86
5.86
5.84
5.83
5.74
5.74
5.72
5.71

2.29
2.27
2.25
2.23
2.21
2.19
2.00
1.96
1.95
1.92
1.91
1.89
1.42
1.41
1.37
1.36



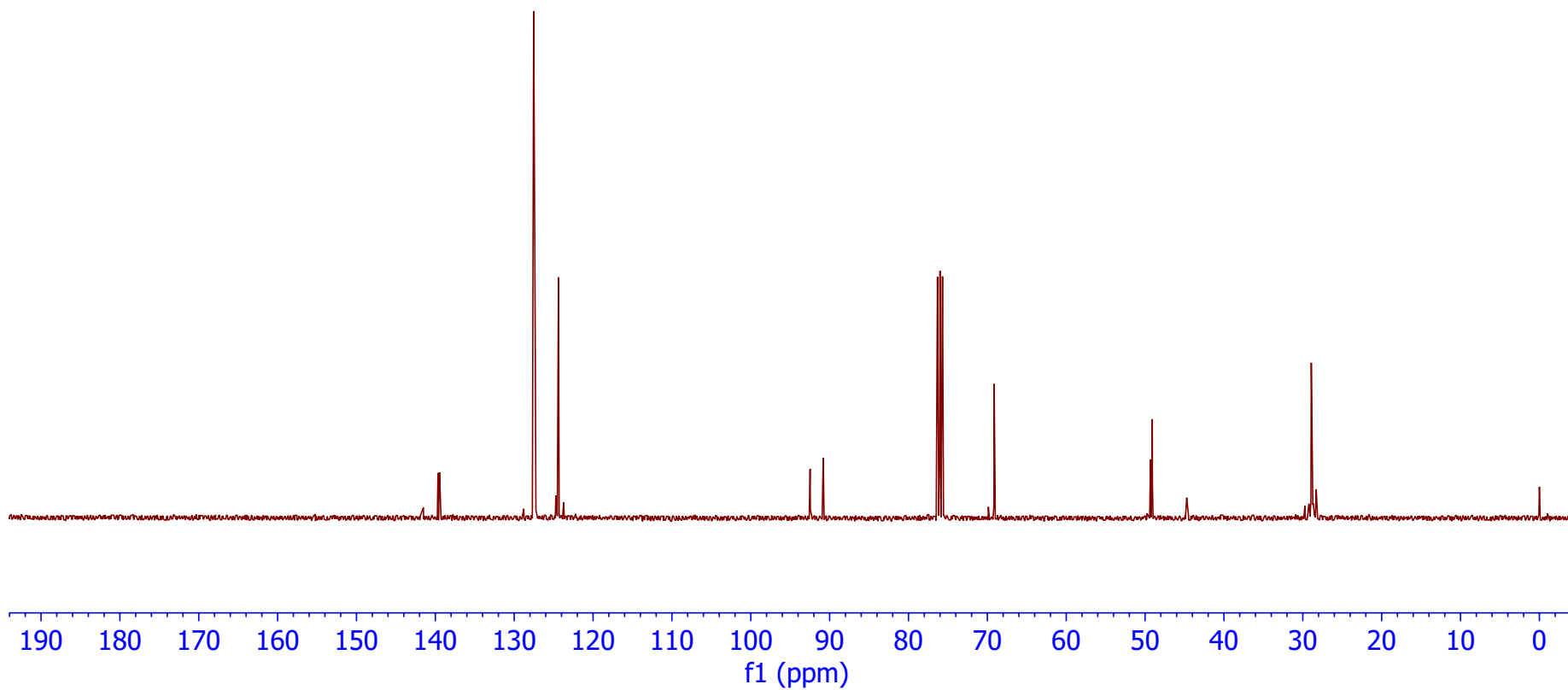
¹H NMR of 14b

101 MHz, Chloroform-d



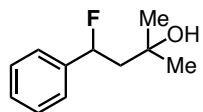
14b

139.67
139.47
128.79
127.53
127.49
127.38
127.34
127.32
127.29
124.45
124.39
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92.48
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69.14
68.93
49.32
49.30
49.09
28.92
28.80



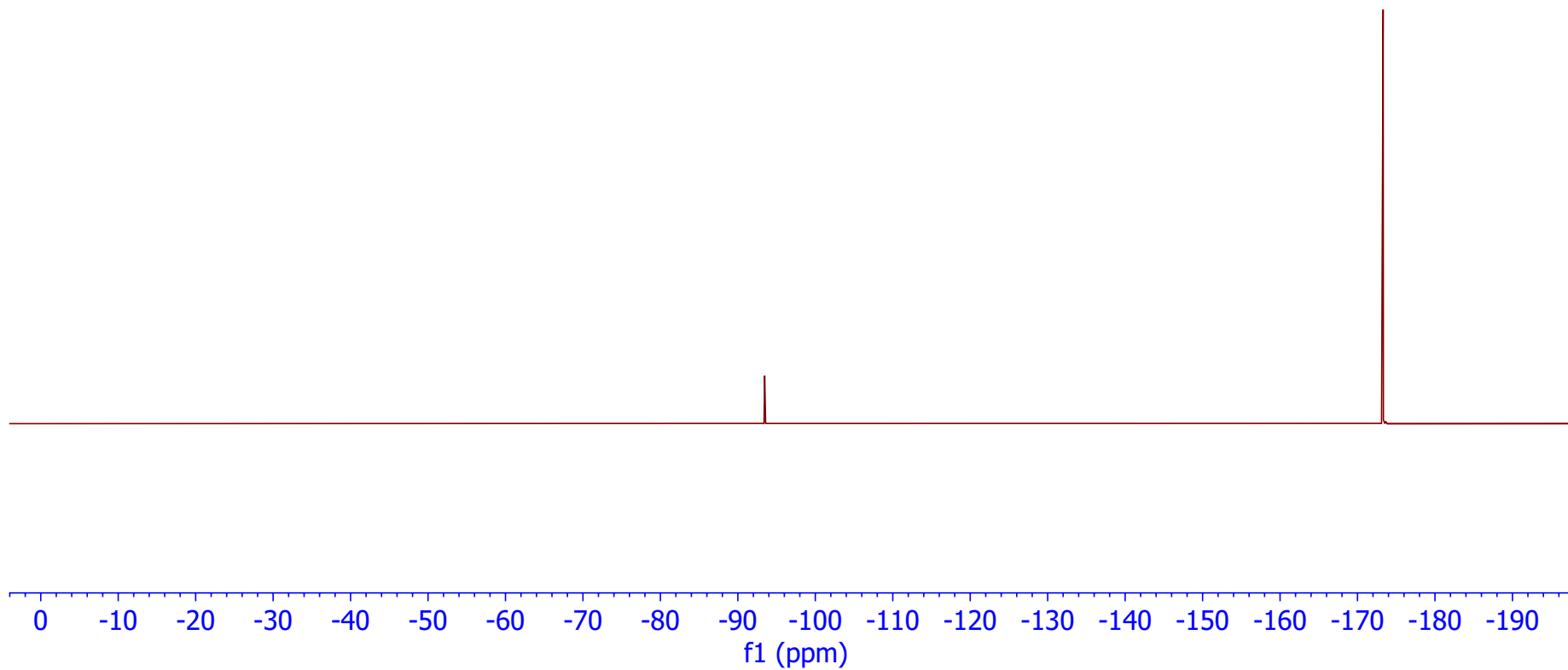
¹³C NMR of 14b

376MHz, Chloroform-d



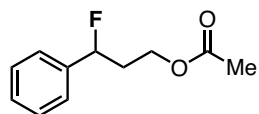
14b

--173.19
--173.30
--173.31
--173.34
--173.35

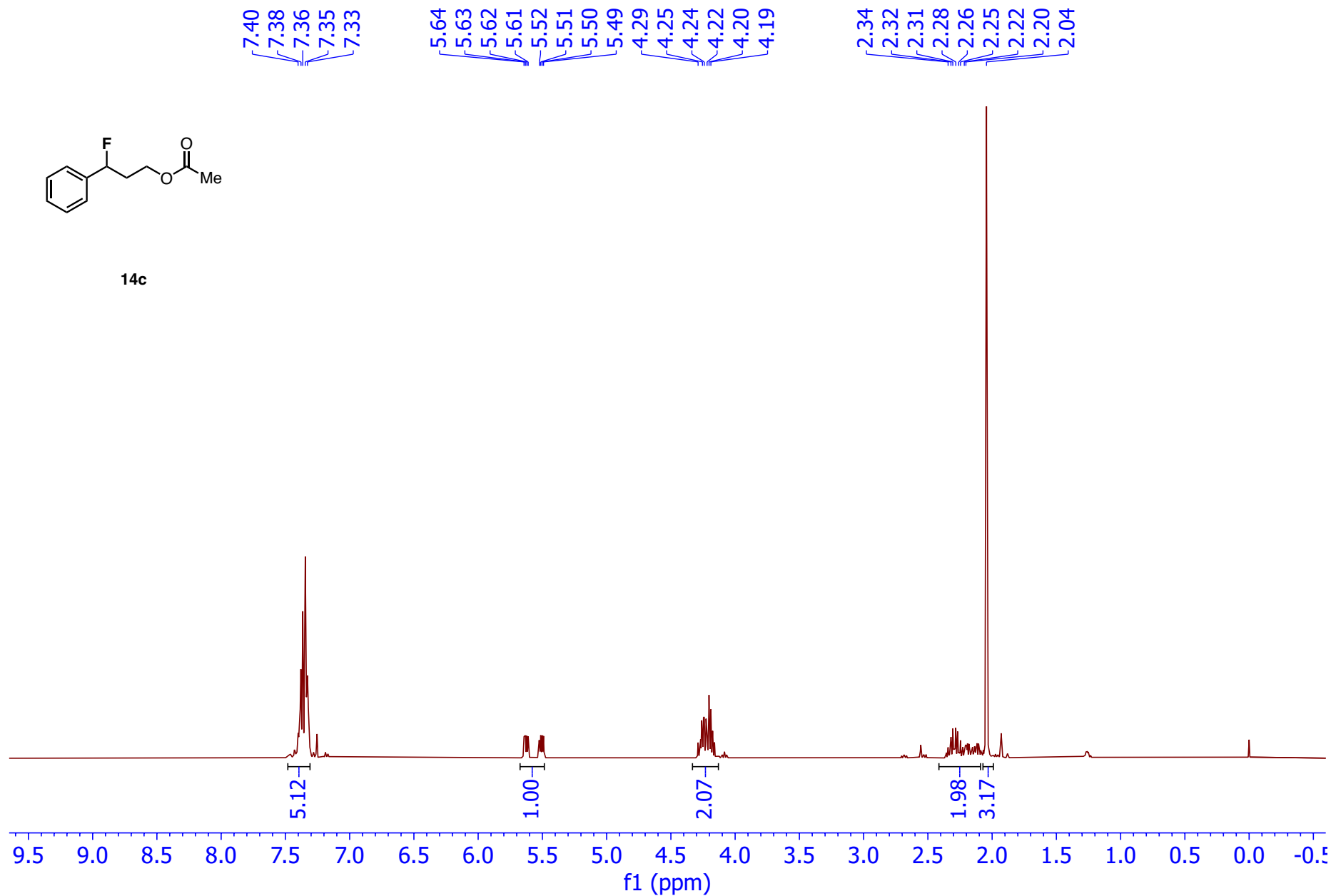


^{19}F NMR of 14b

400 HMz, Chloroform-d

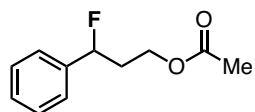


14c

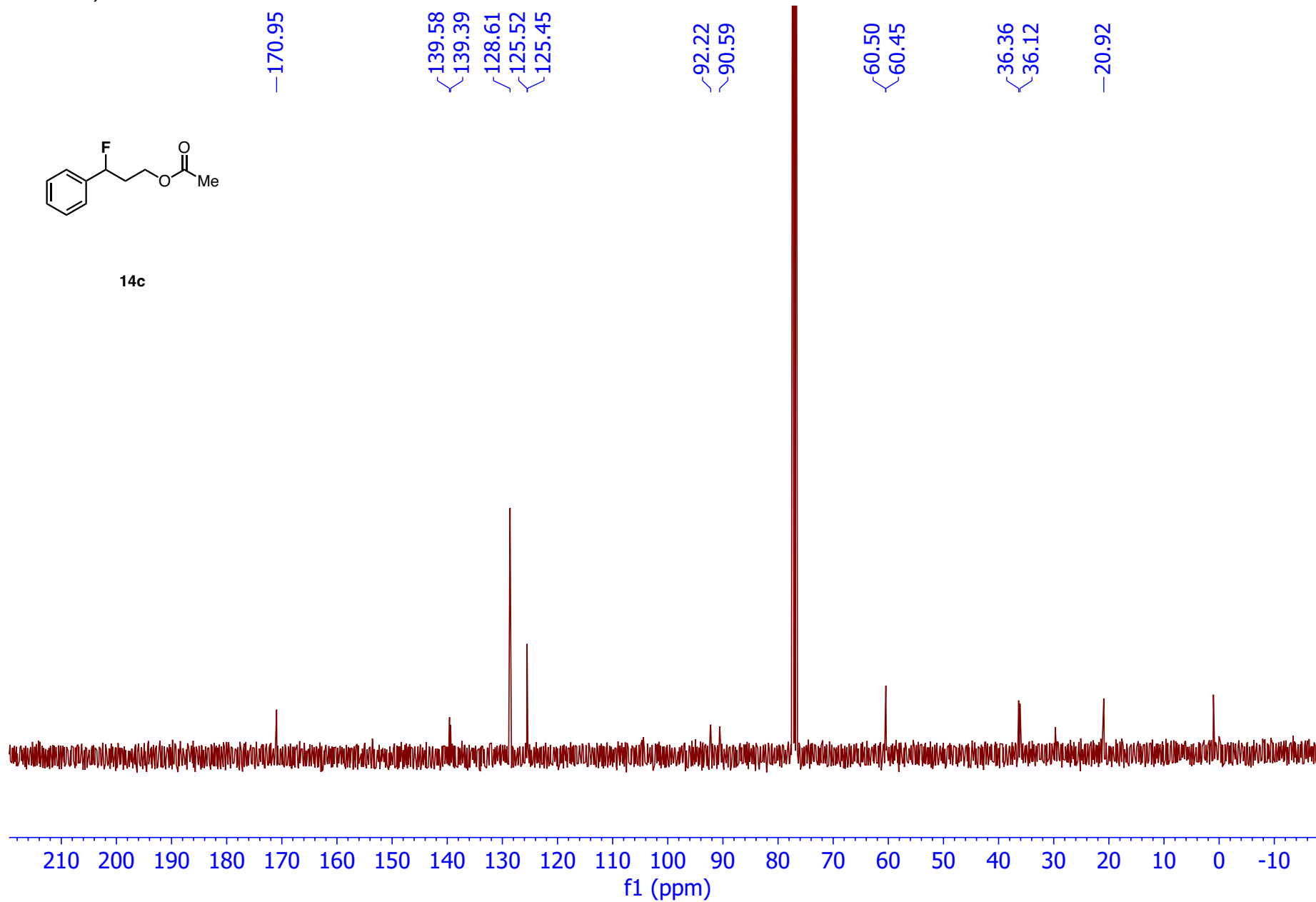


¹H NMR of 14c

101 MHz, Chloroform-d

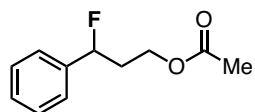


14c



¹³C NMR of 14c

376MHz, Chloroform-d



14c

--177.39

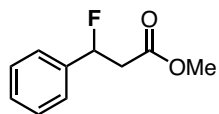


10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210

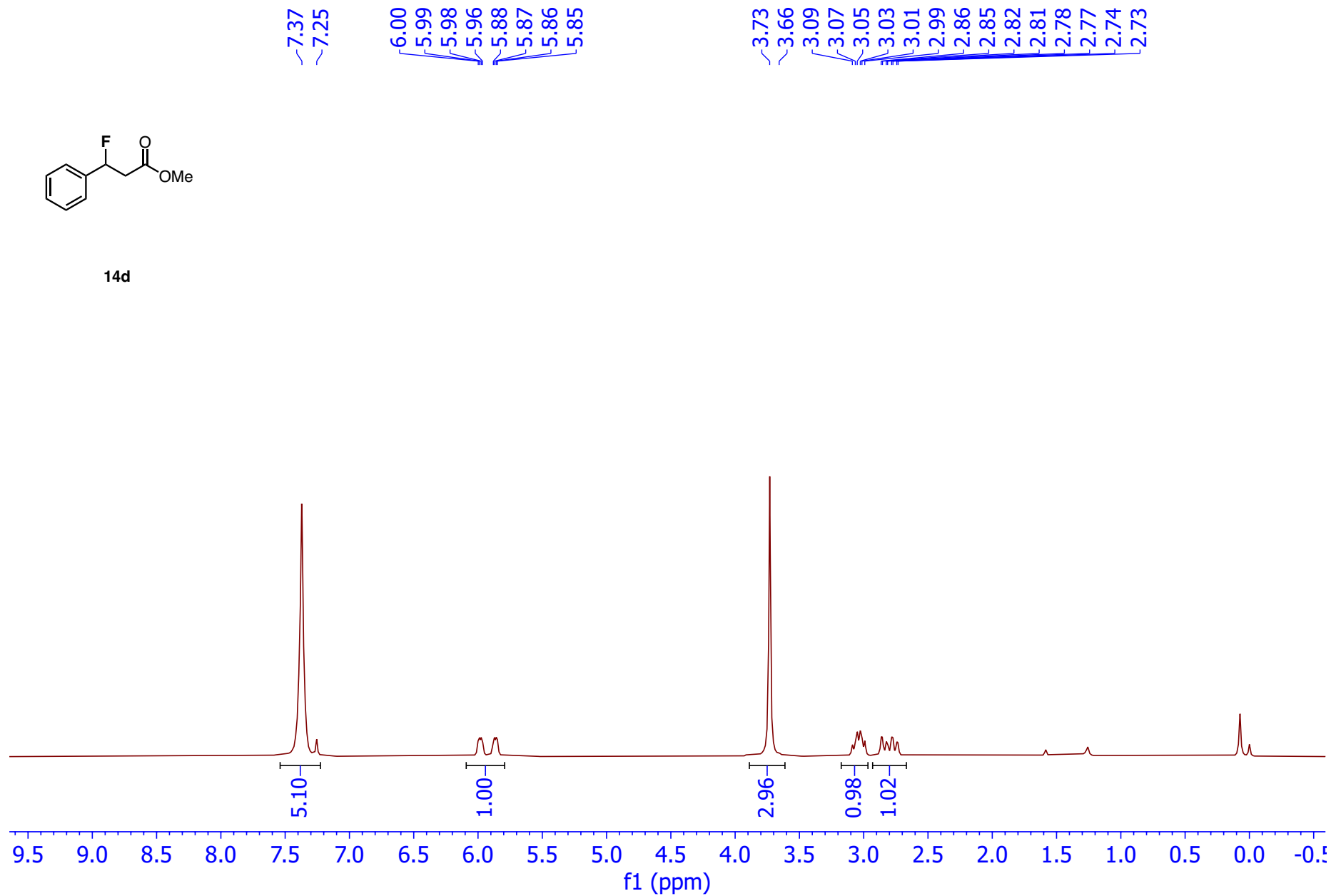
f1 (ppm)

⁹F NMR of 14c

400 HMz, Chloroform-d

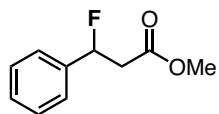


14d

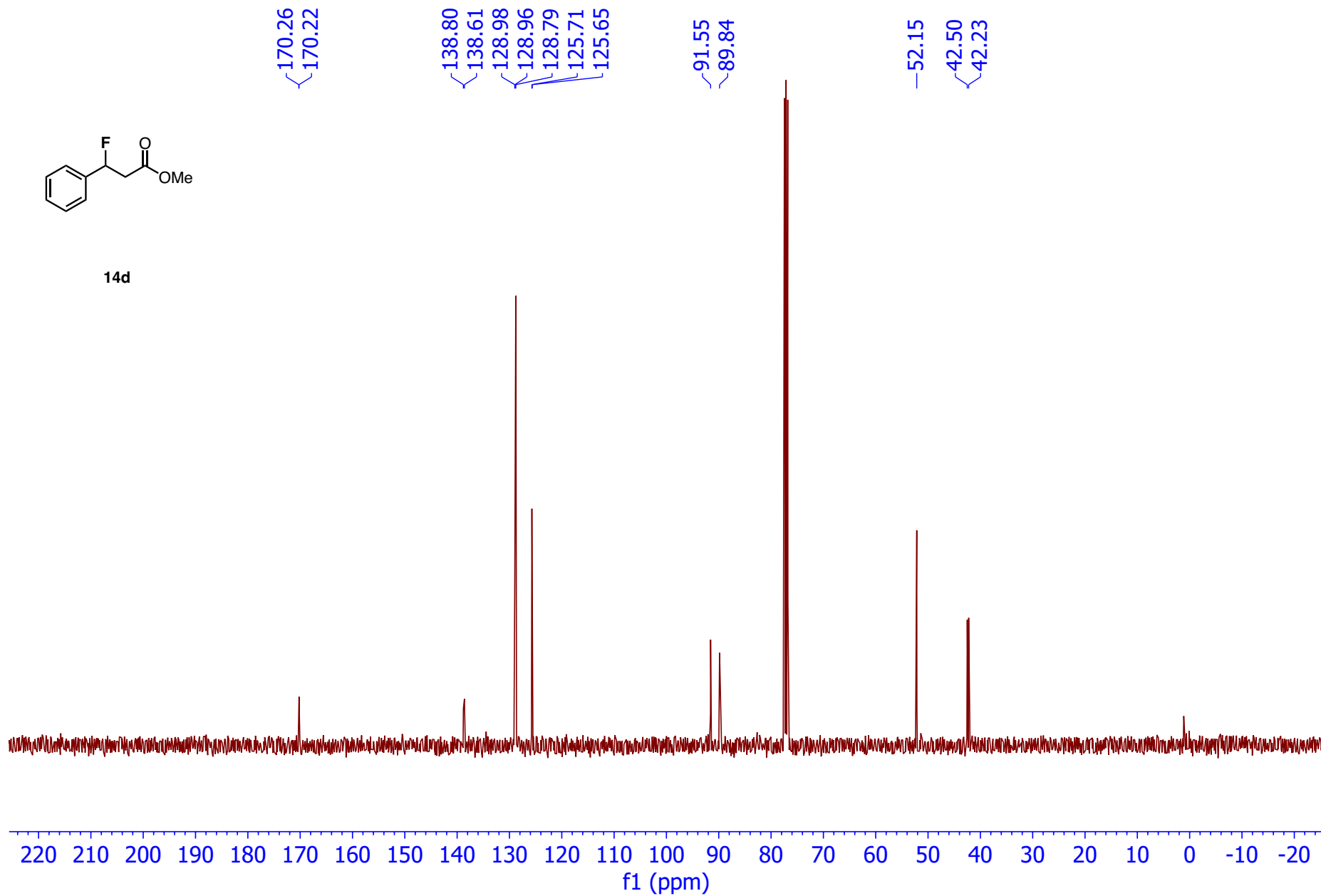


¹H NMR of 14d

101 MHz, Chloroform-d

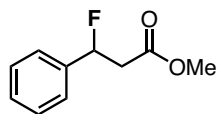


14d



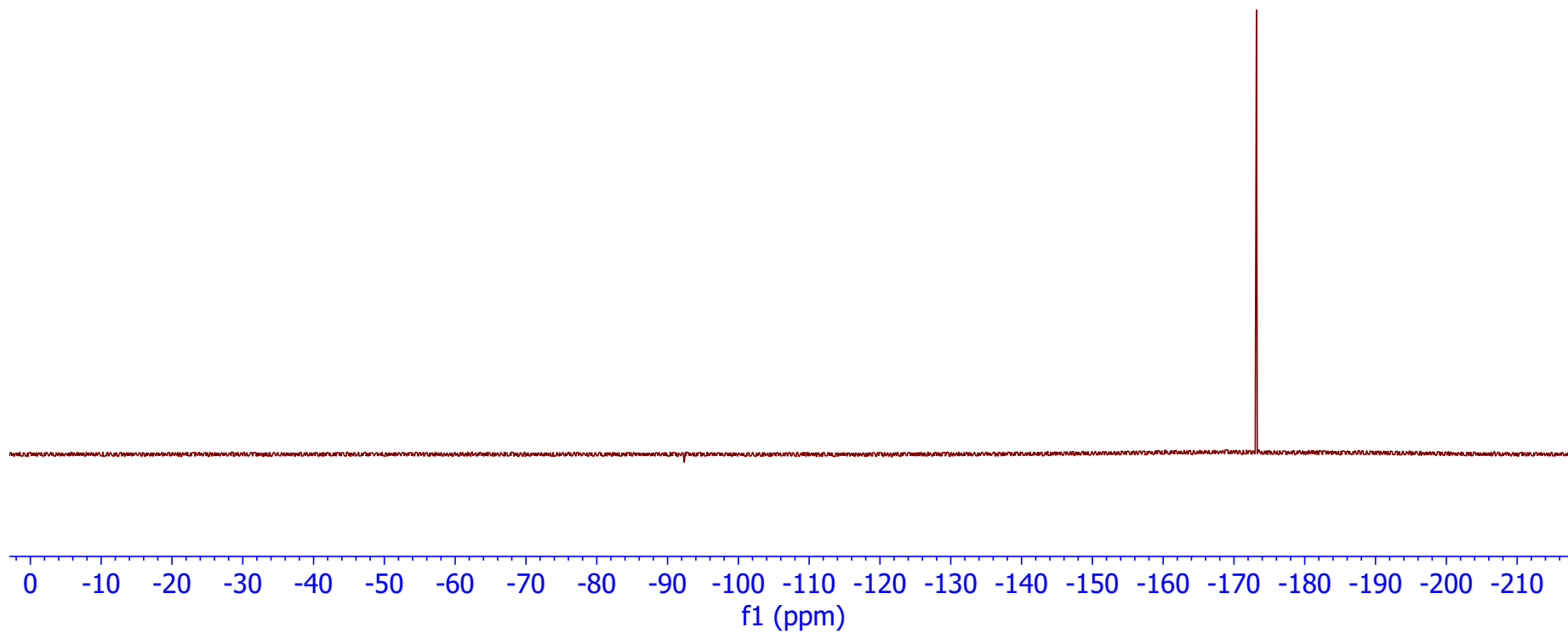
¹³C NMR of 14d

376MHz, Chloroform-d



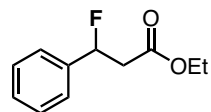
14d

--173.18

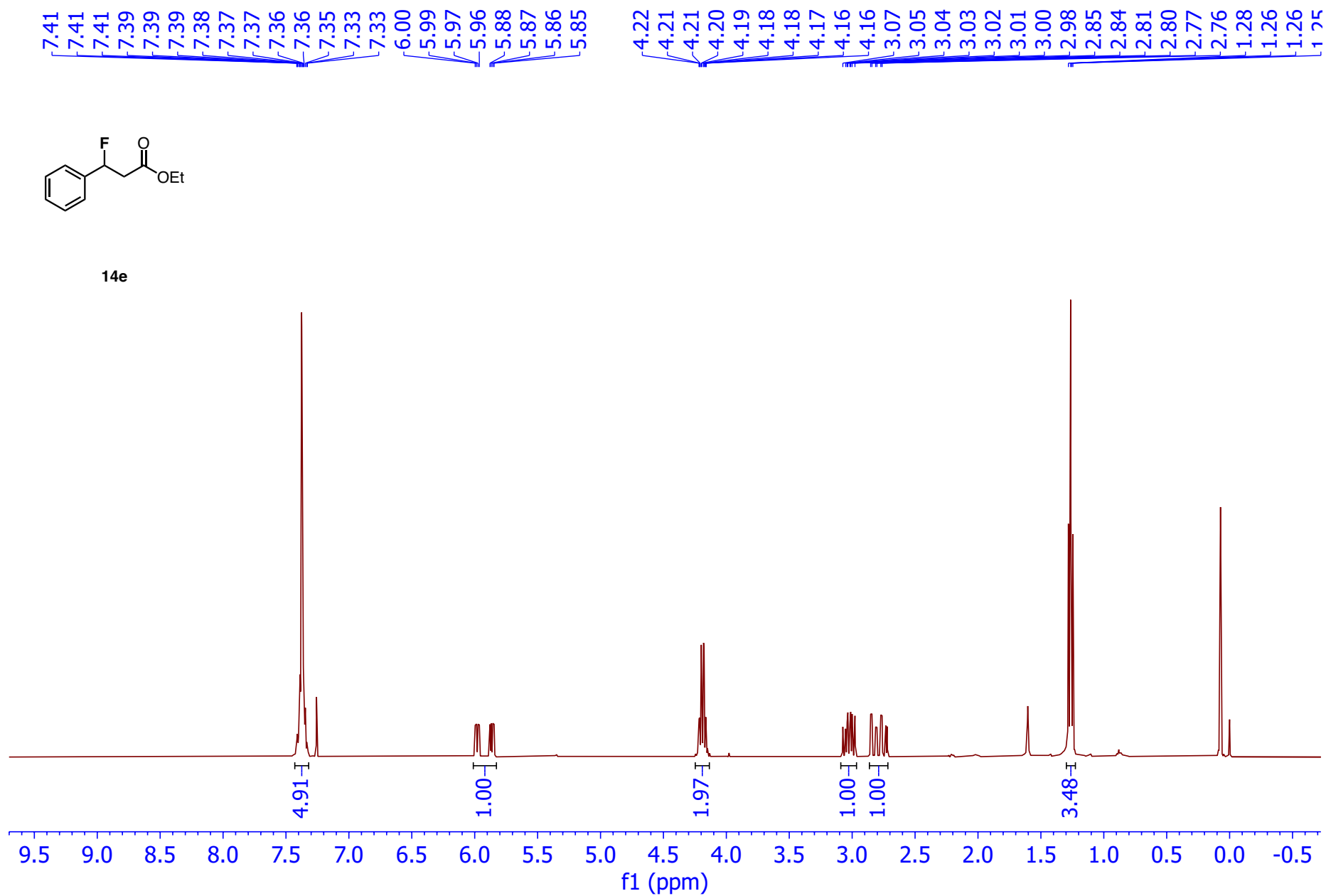


¹⁹F NMR of 14d

400 HMz, Chloroform-d

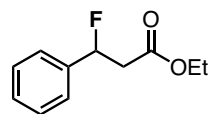


14e

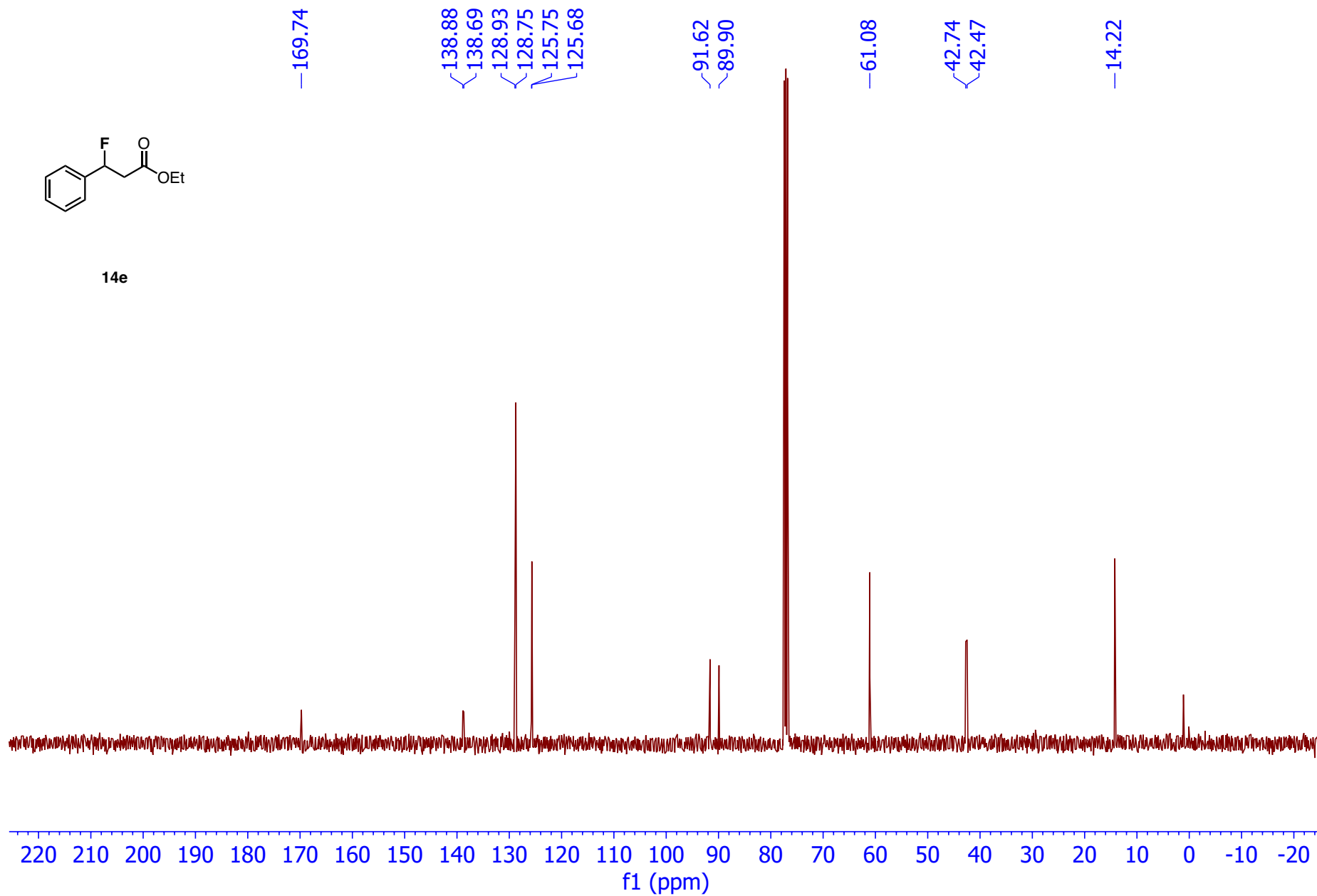


¹H NMR of 14e

101 MHz, Chloroform-d

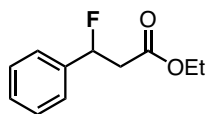


14e



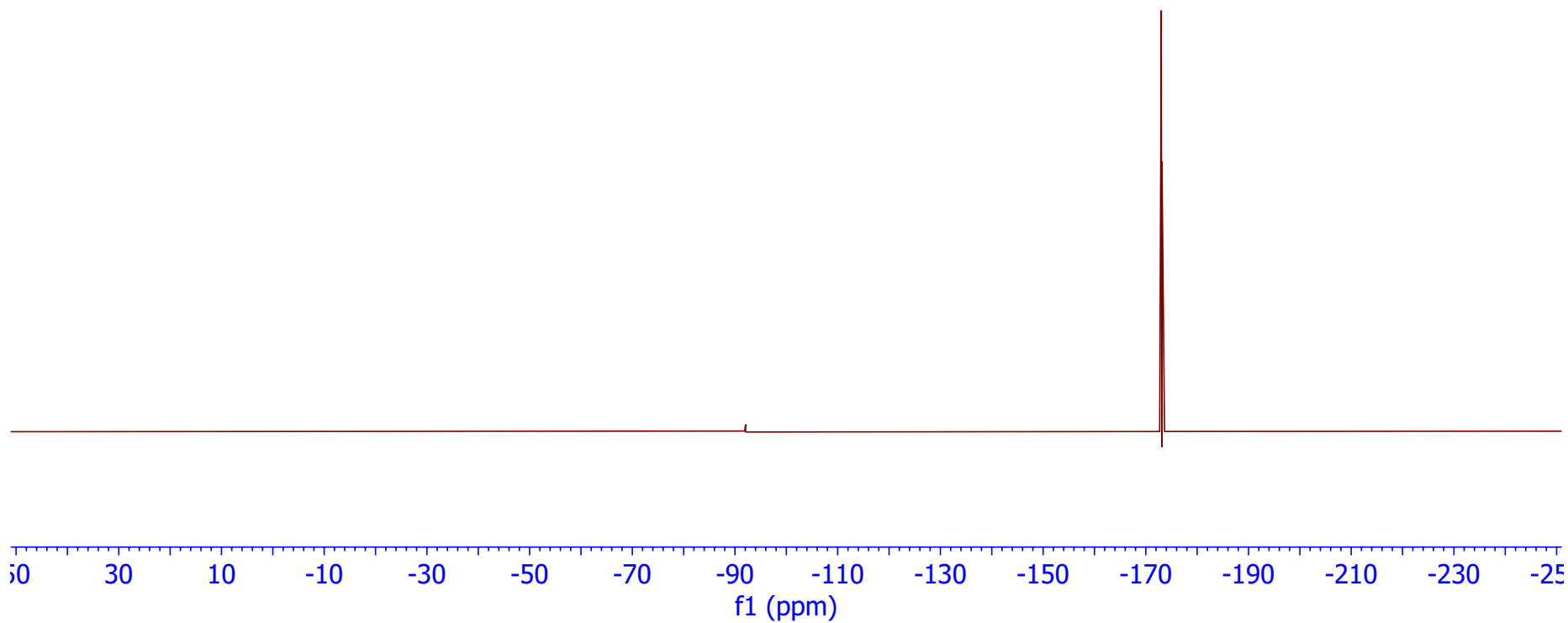
¹³C NMR of 14e

376MHz, Chloroform-d



14e

-172.90
-172.93
-172.96
-172.98
-173.02
-173.06
-173.09
-173.11
-173.14
-173.17
-173.24
-173.36
-173.40

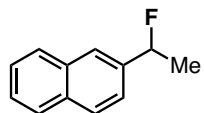


¹⁹F NMR of 14e

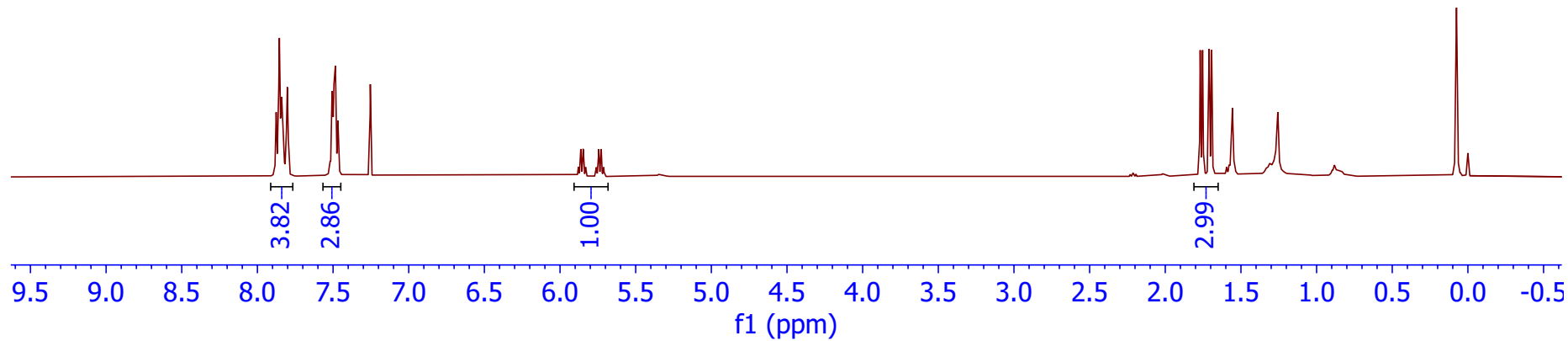
400 HMz, Chloroform-d

7.88
7.86
7.86
7.84
7.84
7.83
7.80
7.52
7.51
7.50
7.49
7.49
7.48
7.47
7.46
5.88
5.86
5.85
5.83
5.76
5.74
5.73
5.71
5.35

1.77
1.75
1.71
1.69

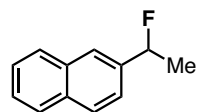


14f

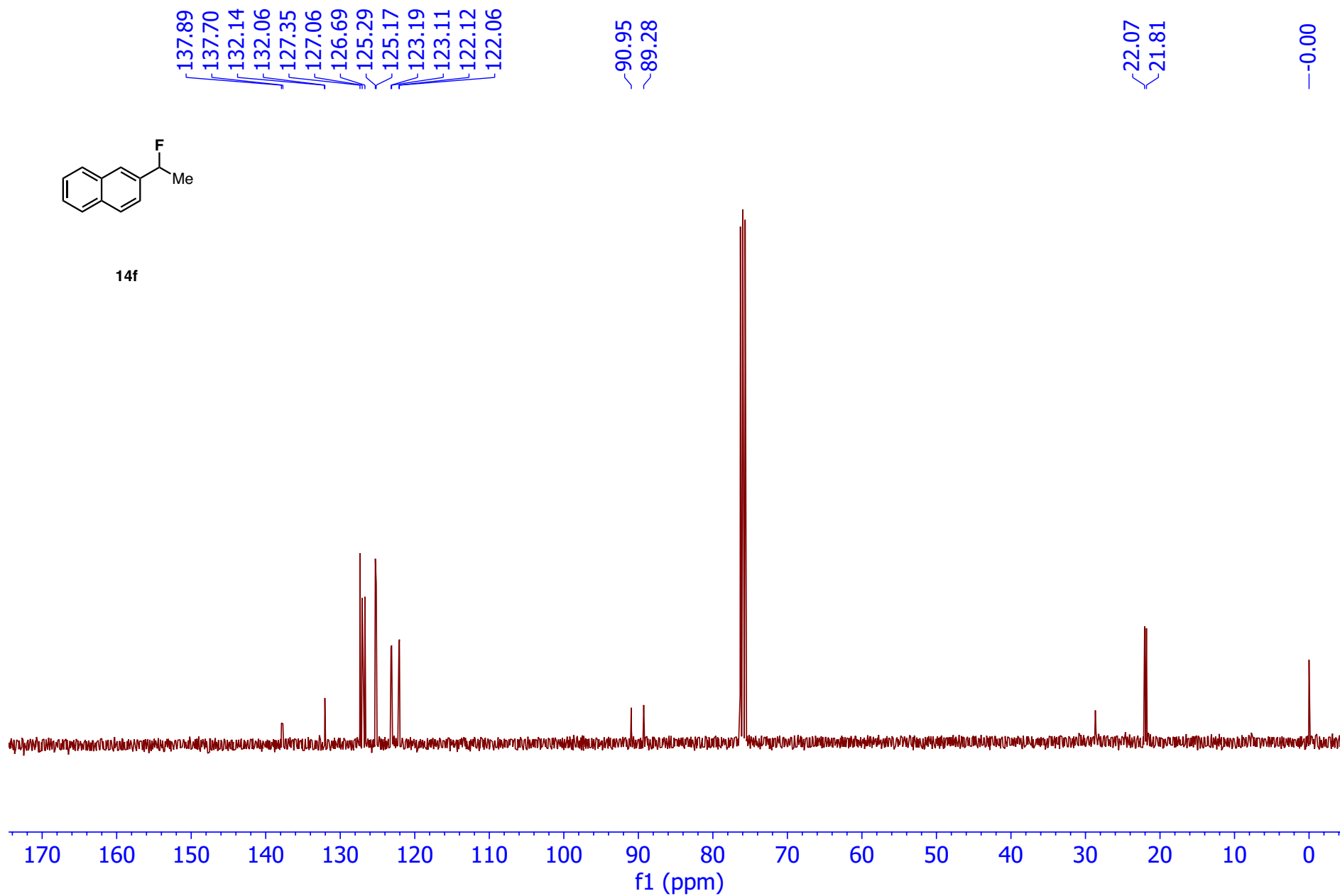


¹H NMR of 14f

101 MHz, Chloroform-d

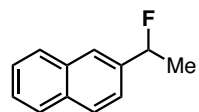


14f



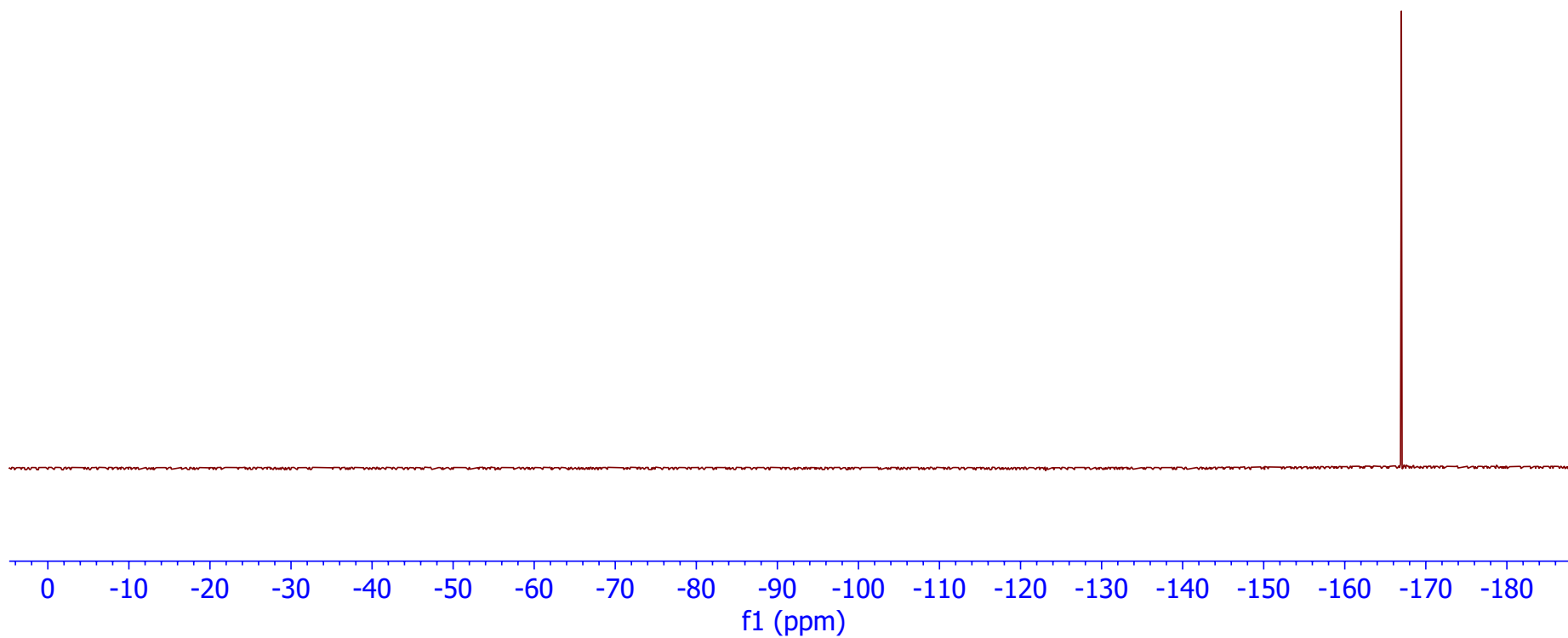
¹³C NMR of 14f

376MHz, Chloroform-d



14f

-166.97
-166.98
-166.99
-167.03
-168.49

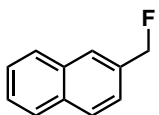


^{19}F NMR of 14f

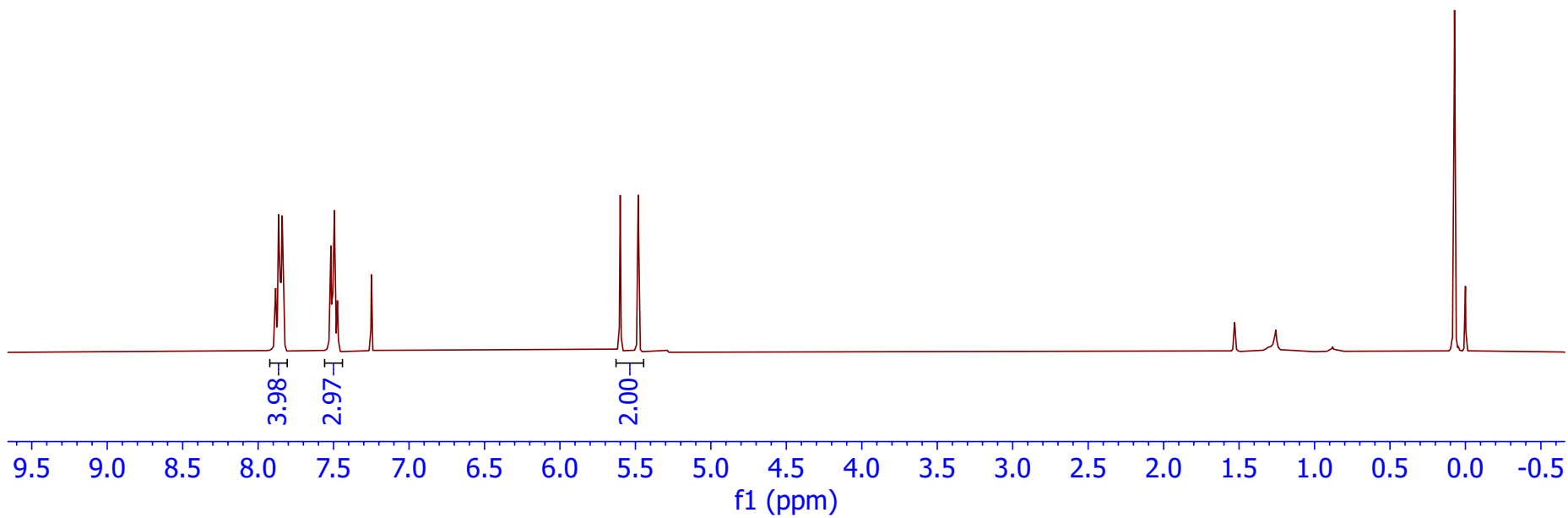
400 HMz, Chloroform-d

7.88
7.86
7.86
7.85
7.84
7.83
7.83
7.53
7.52
7.51
7.50
7.49
7.49
7.47

5.60
5.48

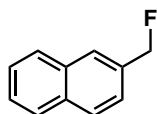


14g

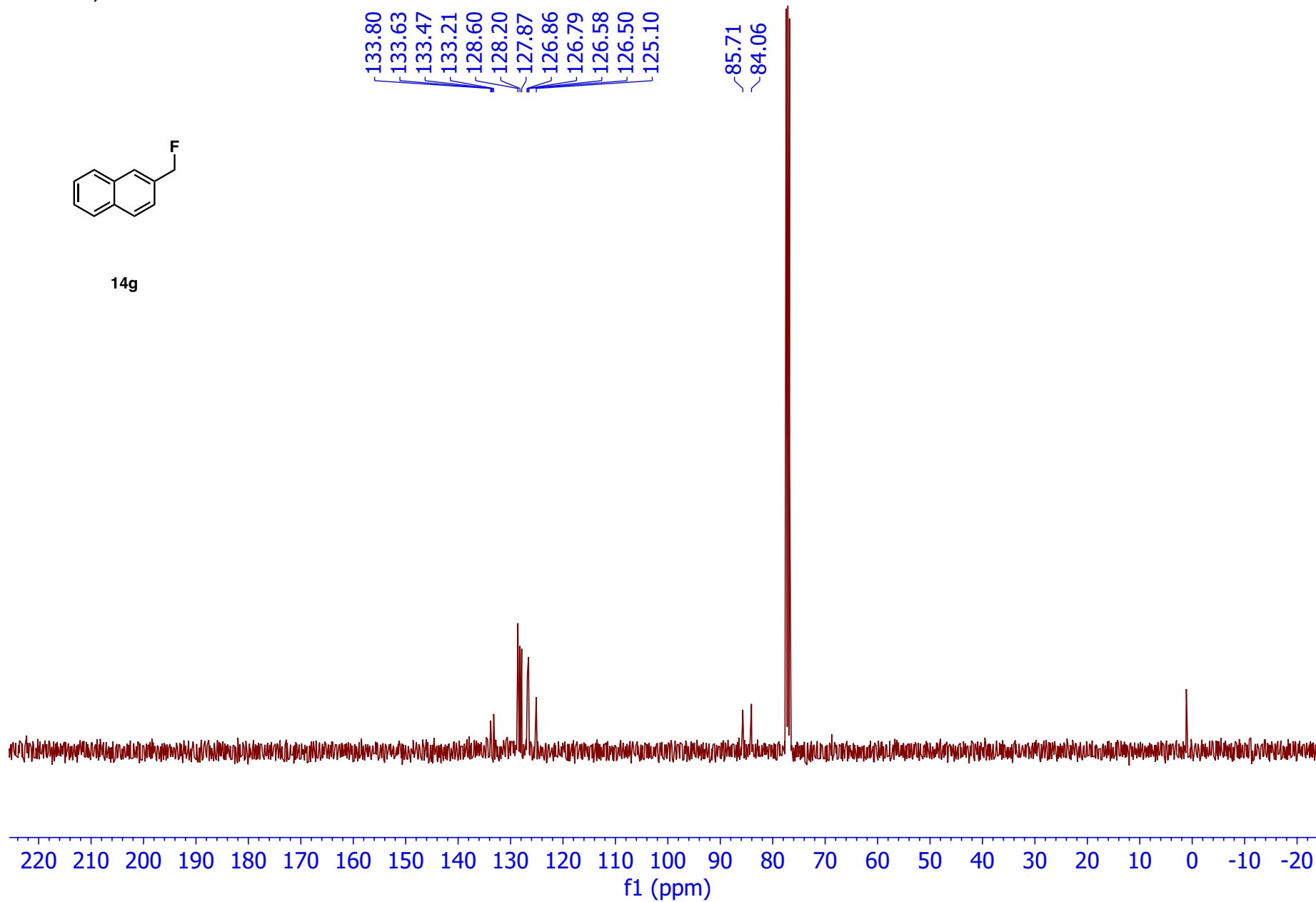


¹H NMR of 14g

101 MHz, Chloroform-d

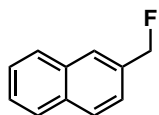


14g



¹³C NMR of 14g

376MHz, Chloroform-d



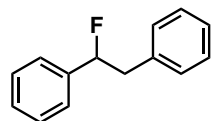
14g

-206.55
-206.62
-206.76
-206.82
-206.89
-207.03
-207.09

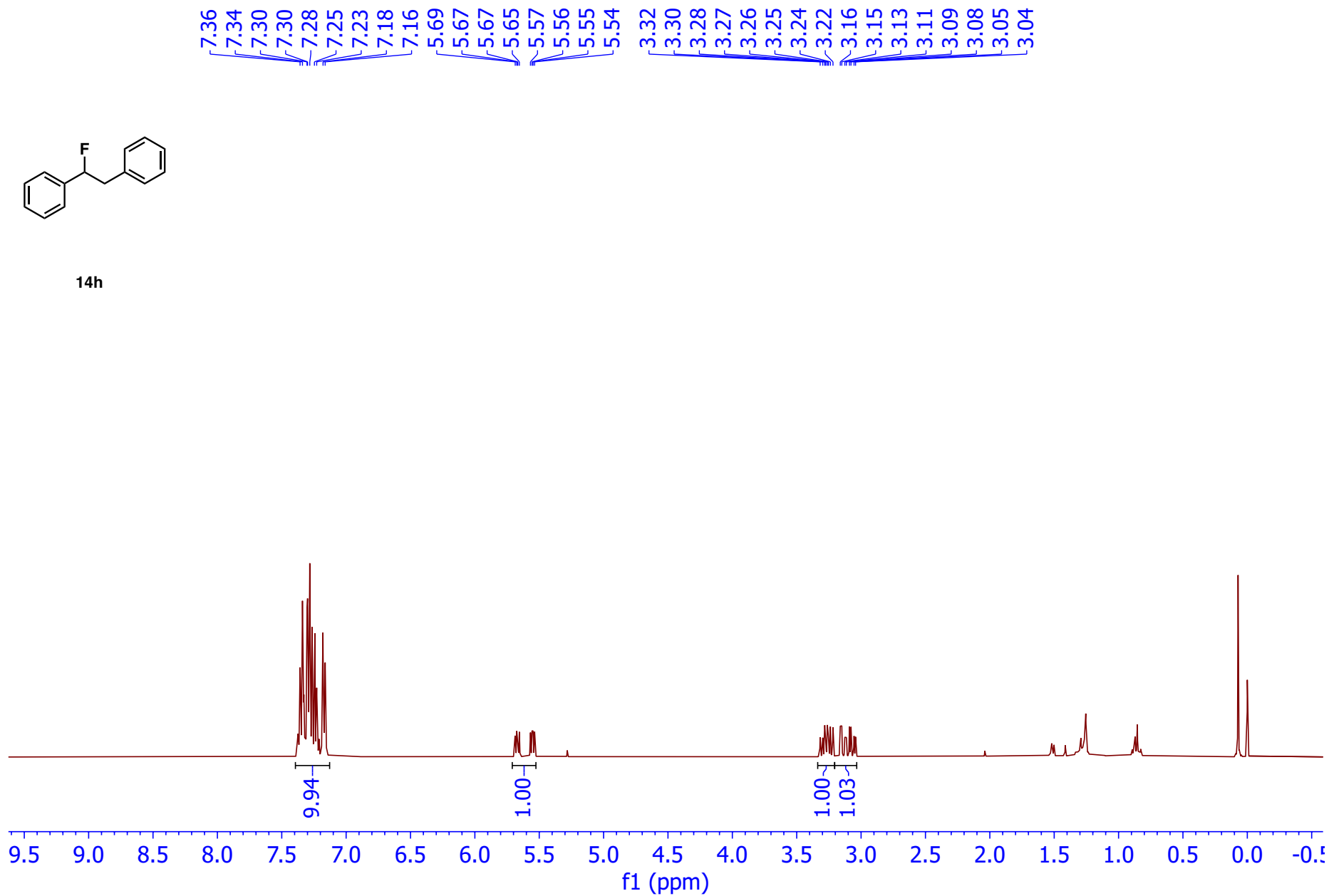


^{19}F NMR of 14g

400 HMz, Chloroform-d

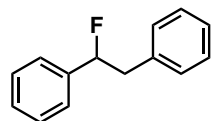


14h



¹H NMR of 14h

101 MHz, Chloroform-d

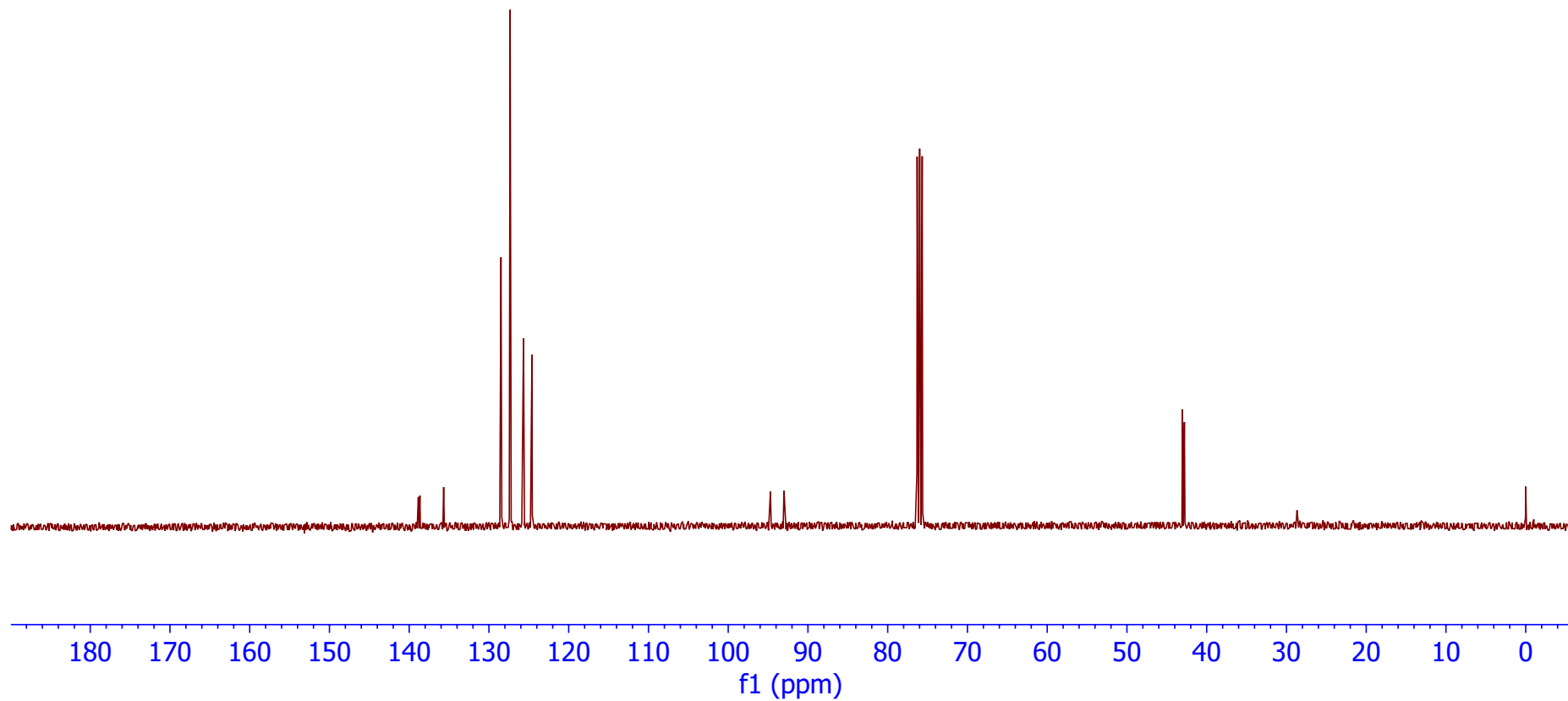


14h

138.86
138.66
135.68
135.64
128.49
127.36
127.35
127.33
125.67
124.67
124.60

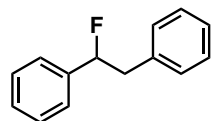
94.74
94.71
93.01
92.98

43.04
42.80



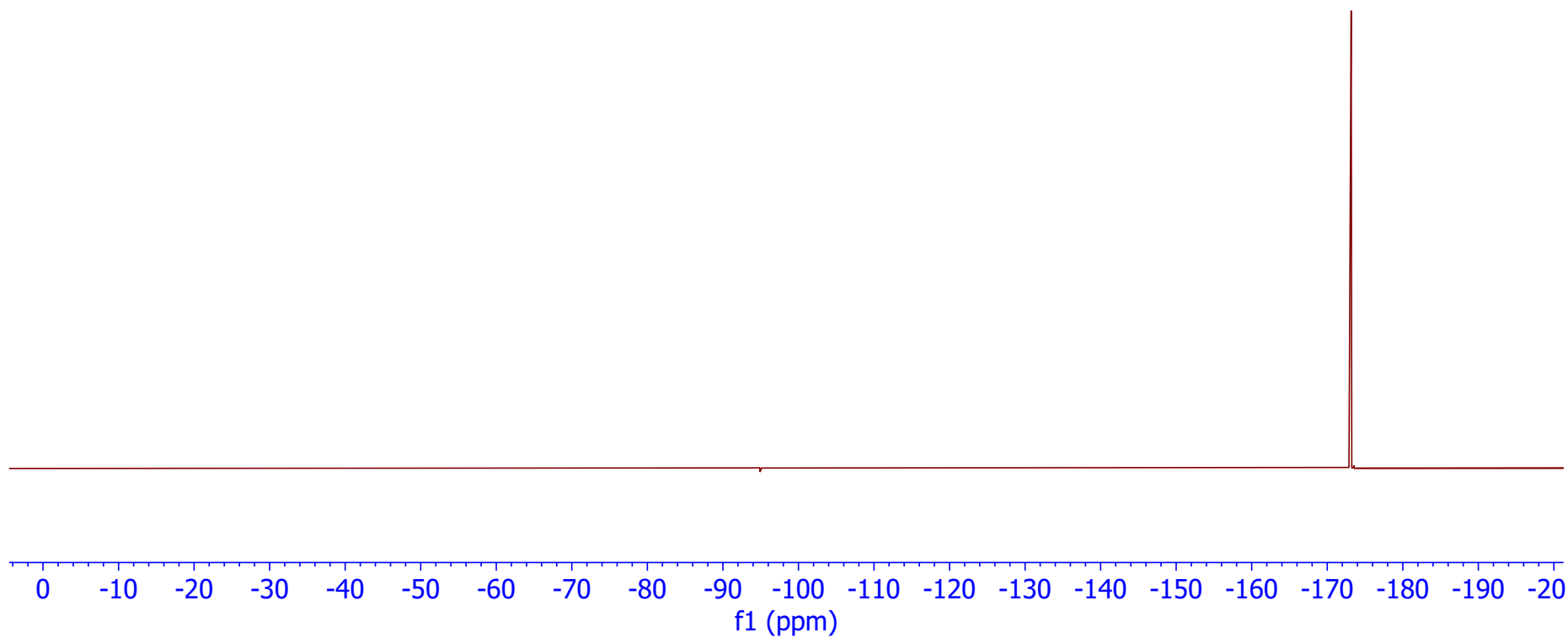
¹³C NMR of 14h

376MHz, Chloroform-d



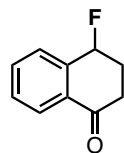
14h

173.04
173.15
173.17
173.20
173.21
186.66



¹⁹F NMR of 14h

400 HMz, Chloroform-d

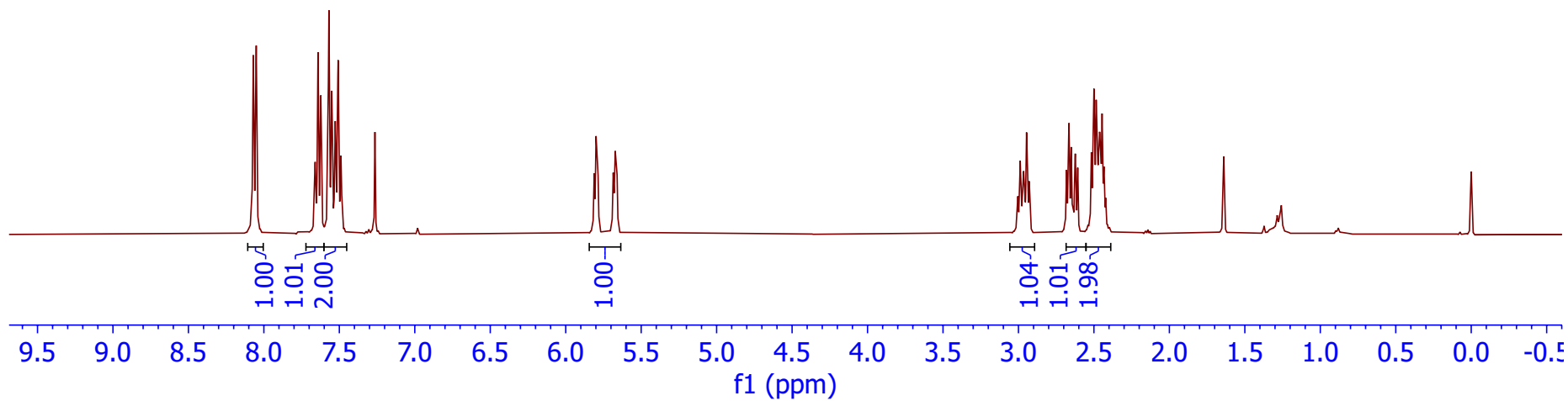


14i

8.27
8.07
8.05
7.66
7.64
7.62
7.57
7.55
7.53
7.51
7.49

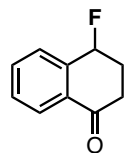
5.81
5.80
5.79
5.66

3.65
3.01
2.99
2.96
2.95
2.93
2.68
2.66
2.65
2.64
2.62
2.61
2.52
2.50
2.49
2.46
2.45
2.43



¹H NMR of 14i

101 MHz, Chloroform-d



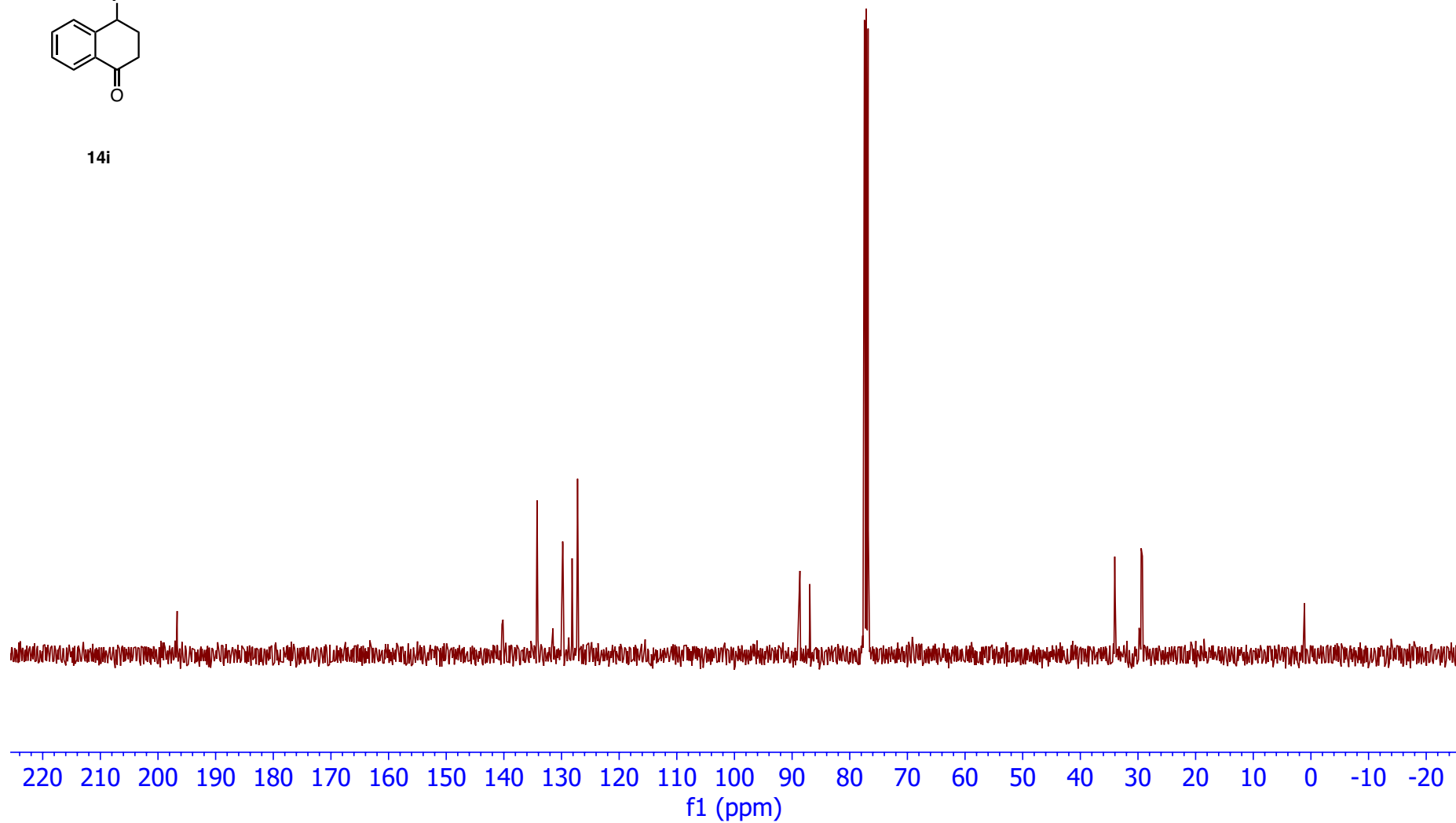
14i

196.68

140.33
140.15
134.24
131.48
129.80
129.77
128.16
128.10
127.22

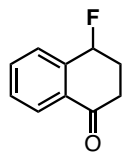
88.65
86.94

34.03
33.96
29.80
29.45
29.23



¹³C NMR of 14i

376MHz, Chloroform-d



14i

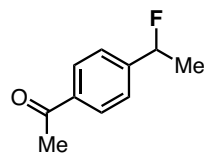
170.31
170.35
170.36
170.40
170.44
170.49
170.53



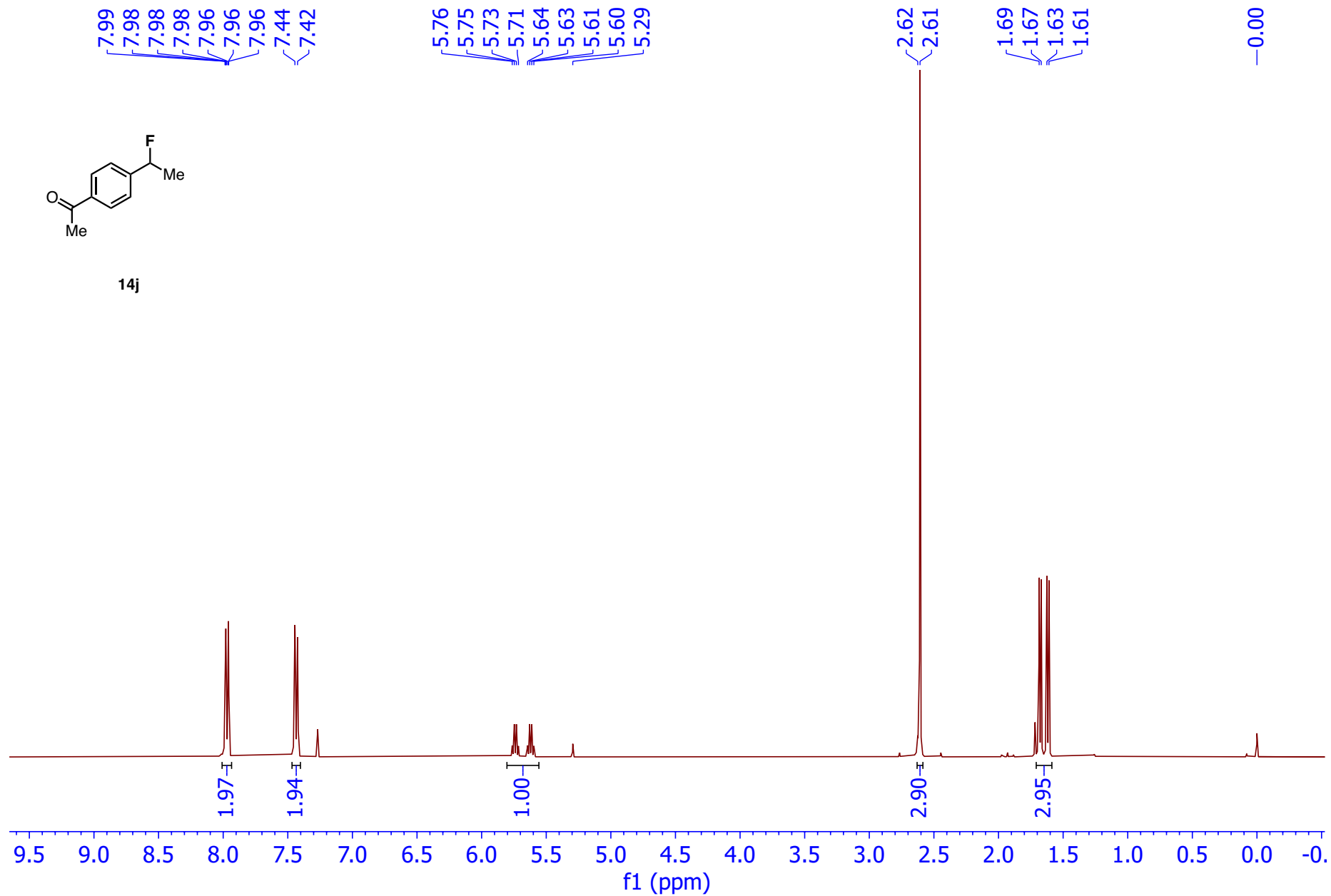
50 30 10 -10 -30 -50 -70 -90 -110 -130 -150 -170 -190 -210 -230 -250
f1 (ppm)

^{19}F NMR of 14i

400 HMz, Chloroform-d

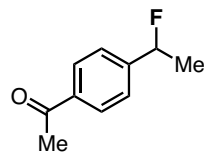


14j

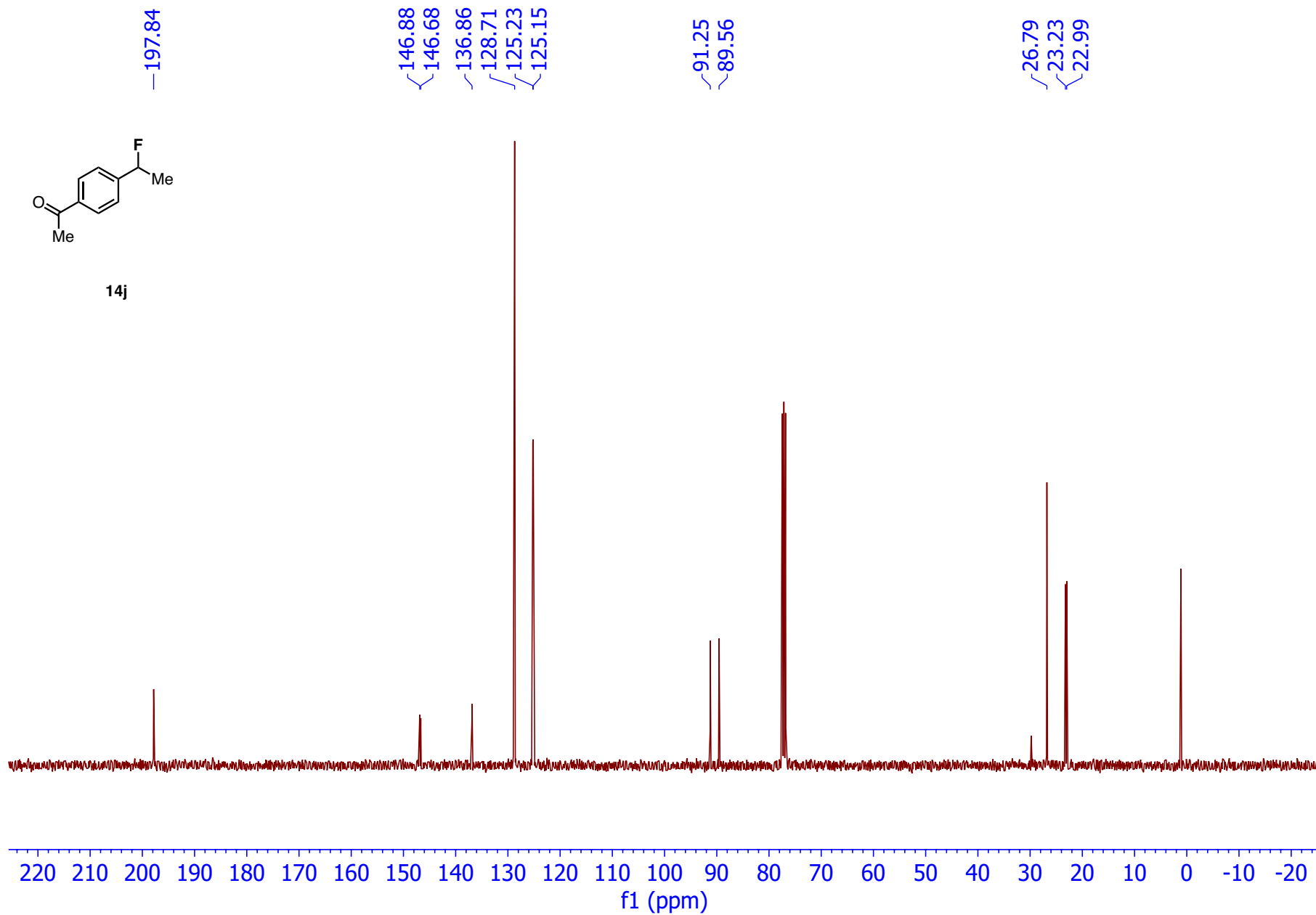


¹H NMR of 14j

101 MHz, Chloroform-d

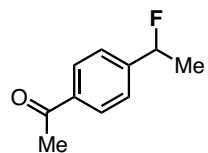


14j



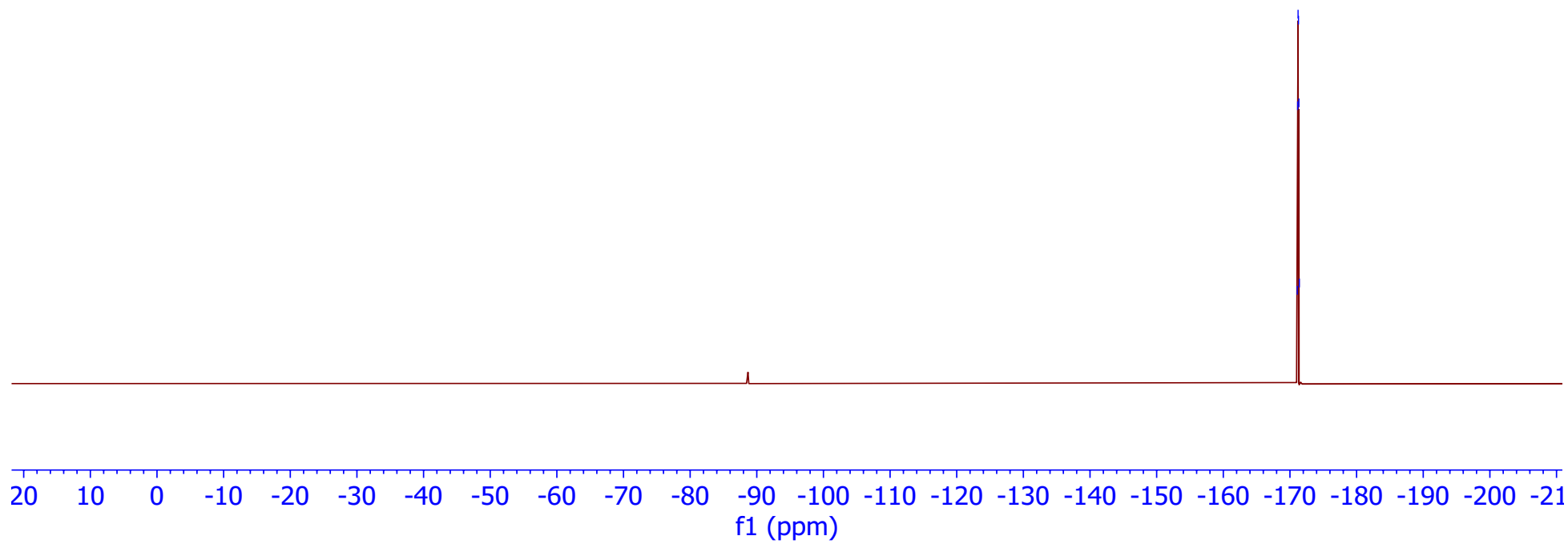
¹³C NMR of 14j

376MHz, Chloroform-d



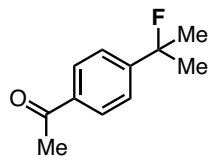
14j

-171.09
-171.16
-171.22
-171.28
-171.35
-171.41



¹⁹F NMR of 14j

400 HMz, Chloroform-d

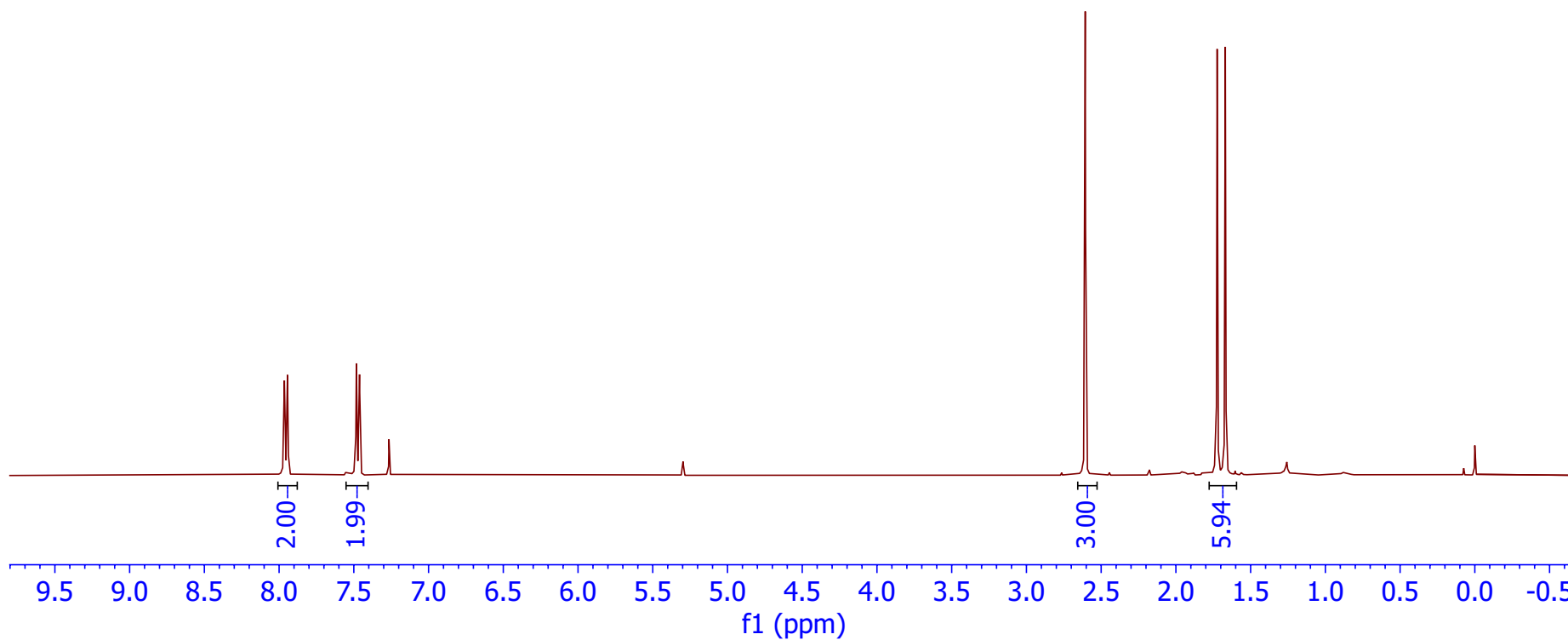


14k

7.96
7.95
7.94
7.48
7.48
7.47
7.46

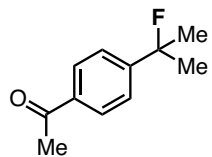
2.63
2.61

1.74
1.72
1.69
1.67



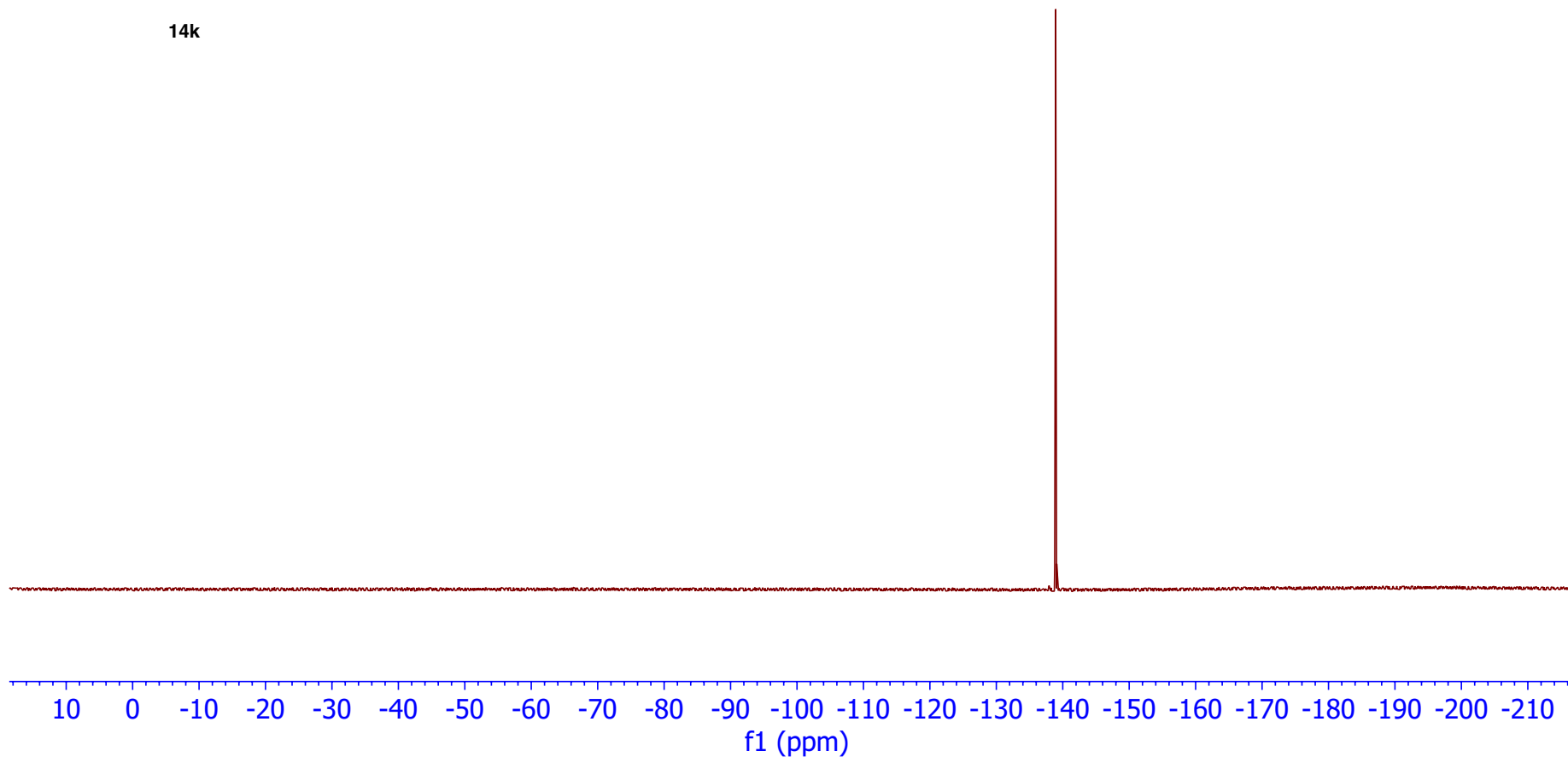
¹H NMR of 14k

376MHz, Chloroform-d



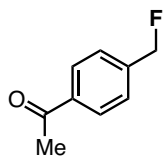
14k

-138.79
-138.84
-138.89
-138.92
-138.94
-138.96
-138.99
-139.01
-139.04
-139.09



¹⁹F NMR of 14k

400 HMz, Chloroform-d

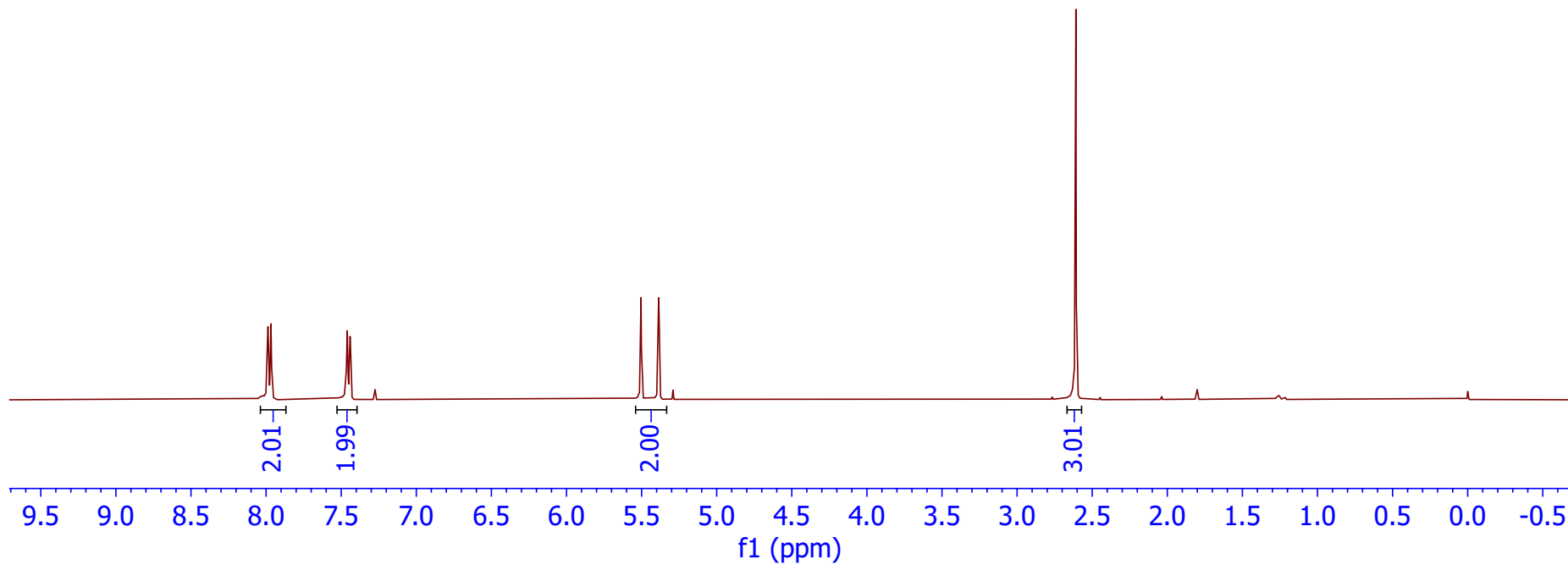


14l

8.04
8.02
7.99
7.99
7.98
7.97
7.97
7.97
7.46
7.46
7.44
7.44

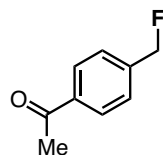
5.50
5.39

2.63
2.62
2.61



¹H NMR of 14l

101 MHz, Chloroform-d

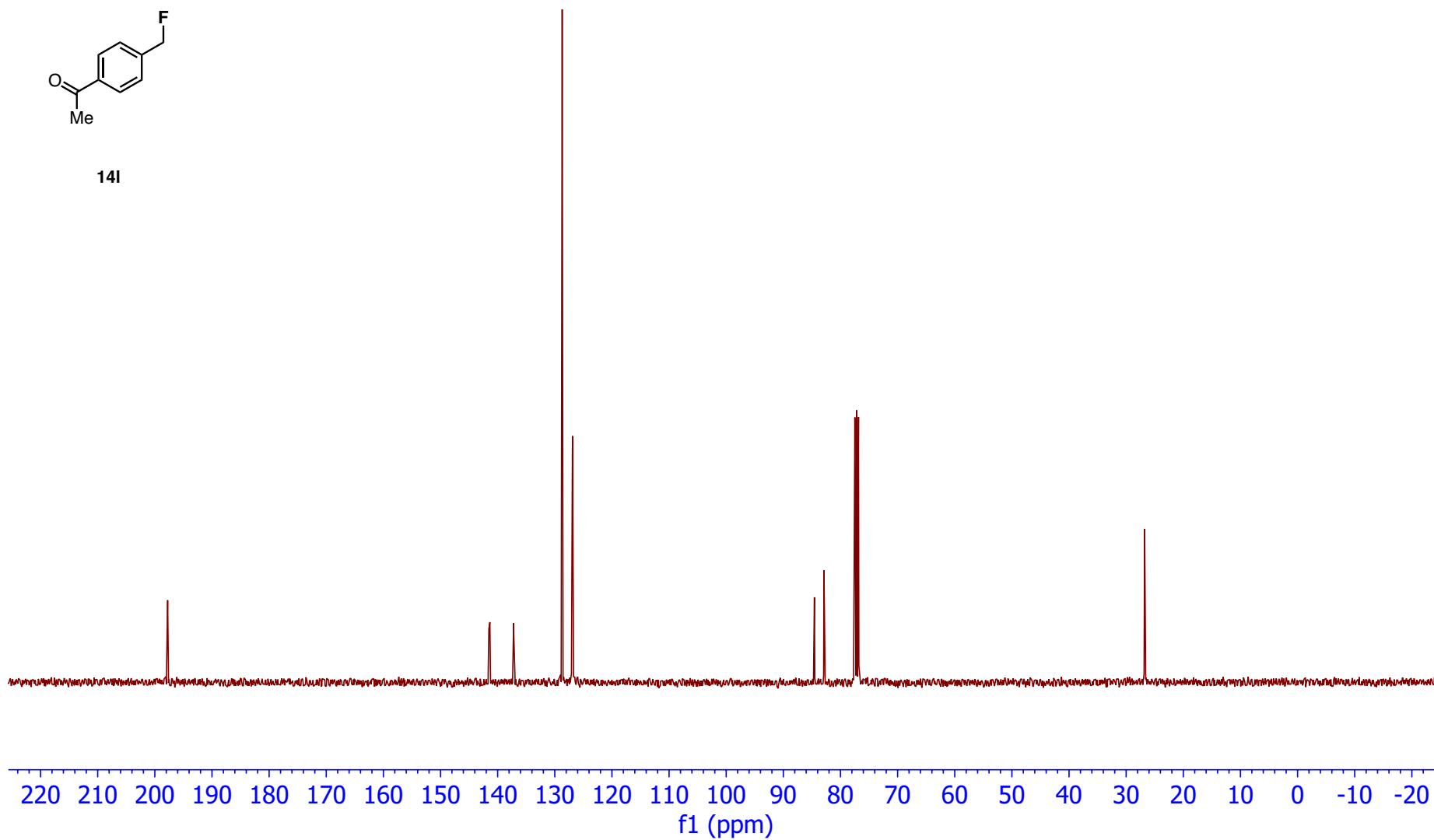


14I

197.76

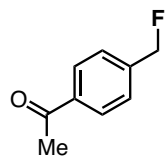
141.53
141.36
137.21
137.18
128.70
126.90
126.84

84.56
82.88



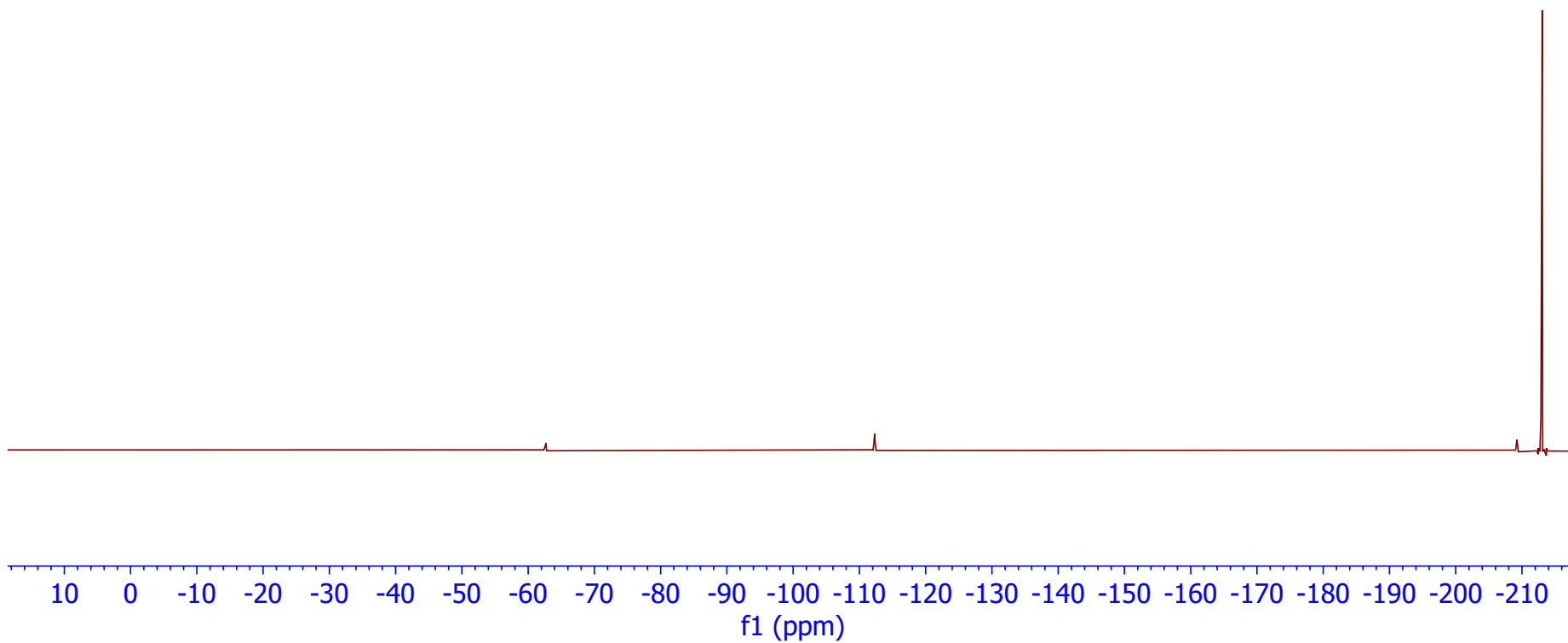
¹³C NMR of 14I

376MHz, Chloroform-d



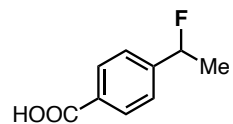
14I

-212.43
-212.45
-212.66
-212.85
-212.86
-212.93
-213.02
-213.05
-213.06
-213.06
-213.25
-213.26
-213.38



¹⁹F NMR of 14I

400 HMz, Chloroform-d

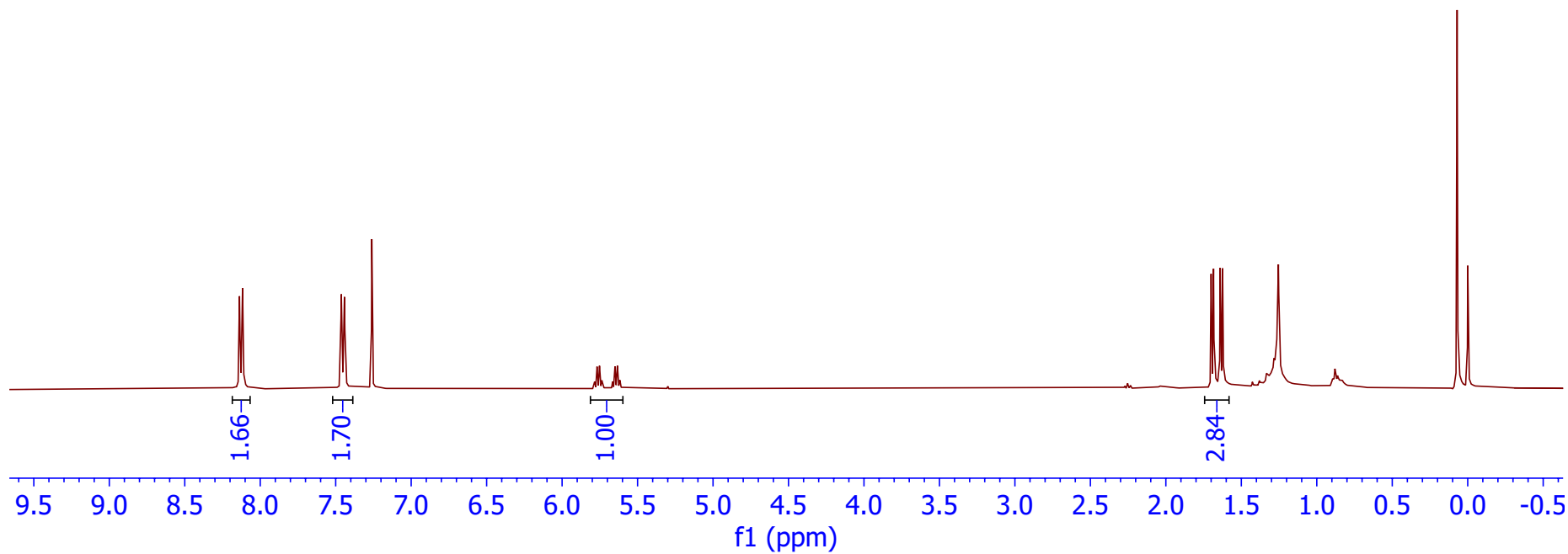


14m

8.14
8.13
8.12
8.04
7.46
7.44

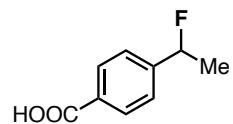
5.78
5.77
5.75
5.74
5.67
5.65
5.63
5.62

1.70
1.68
1.64
1.62



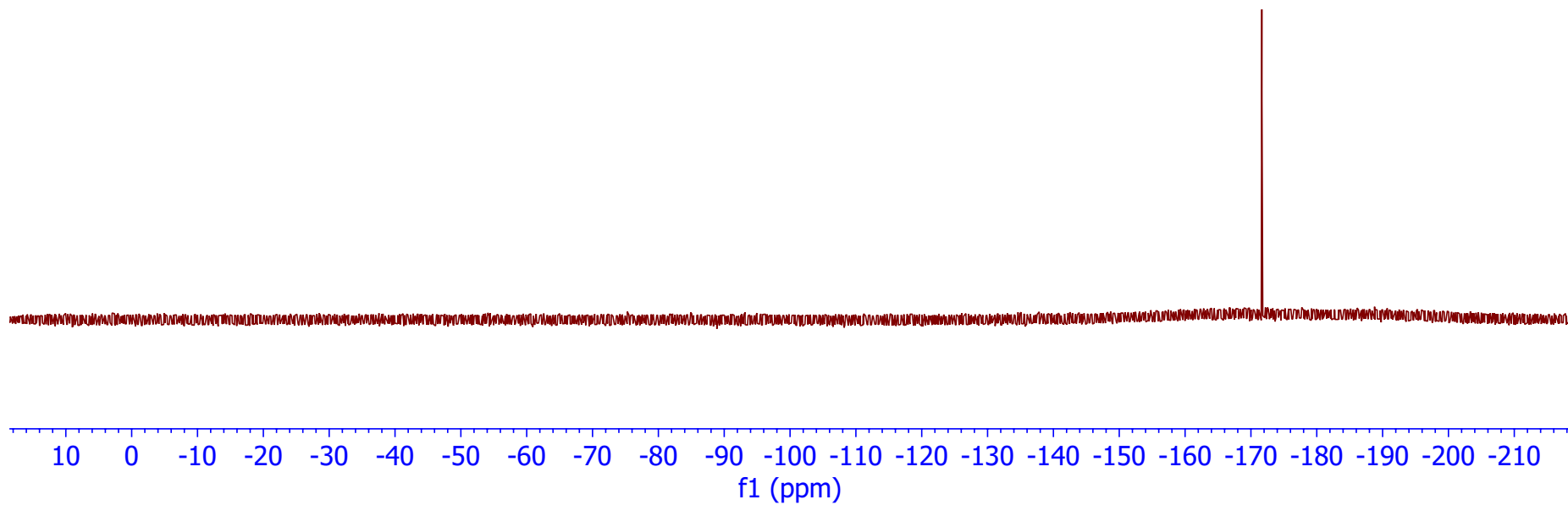
¹H NMR of 14m

376MHz, Chloroform-d



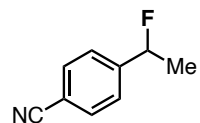
14m

-171.65
-171.66



^{19}F NMR of 14m

400 HMz, Chloroform-d

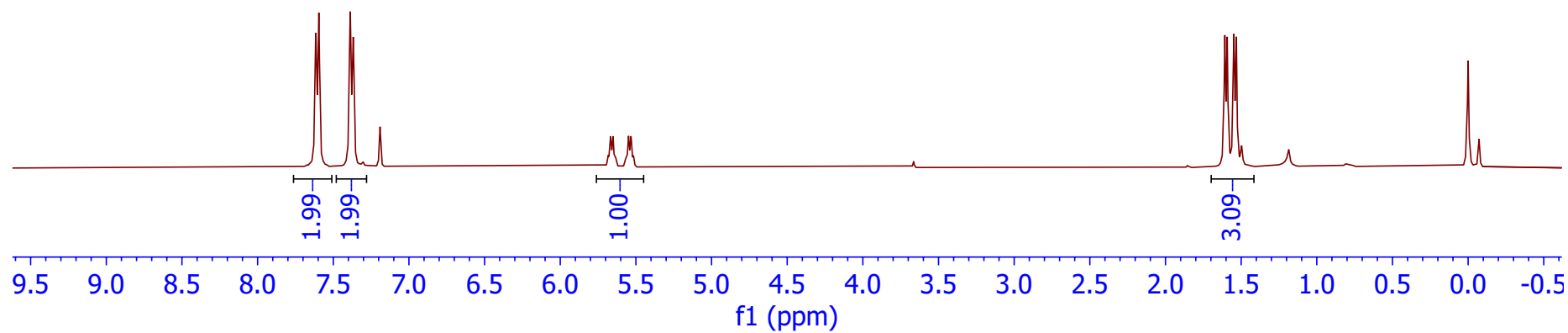


14n

7.62
7.59
7.39
7.37

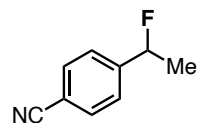
5.68
5.67
5.65
5.63
5.56
5.55
5.53
5.52

1.61
1.59
1.55
1.53
1.18

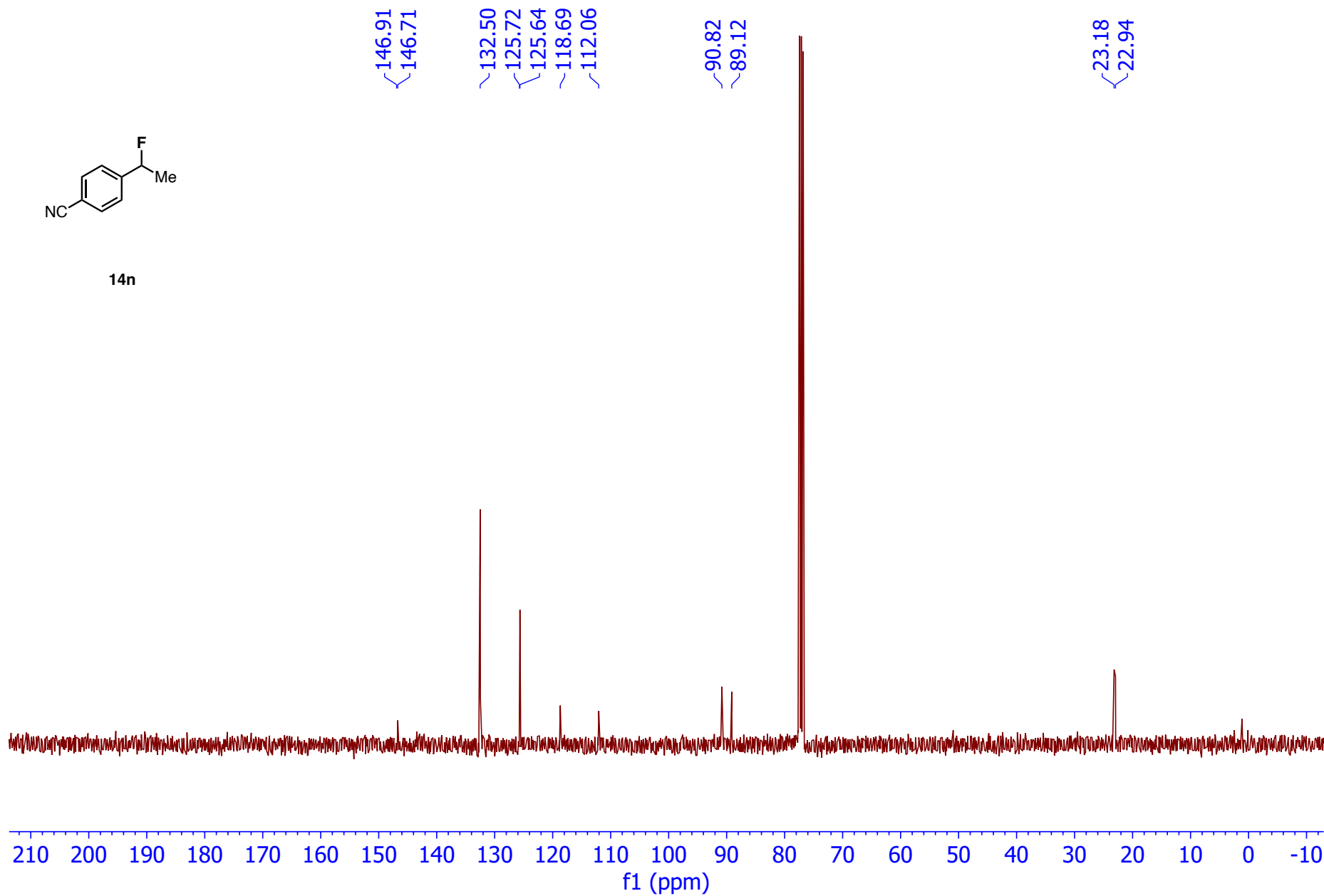


¹H NMR of 14n

101 MHz, Chloroform-d

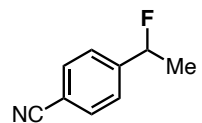


14n



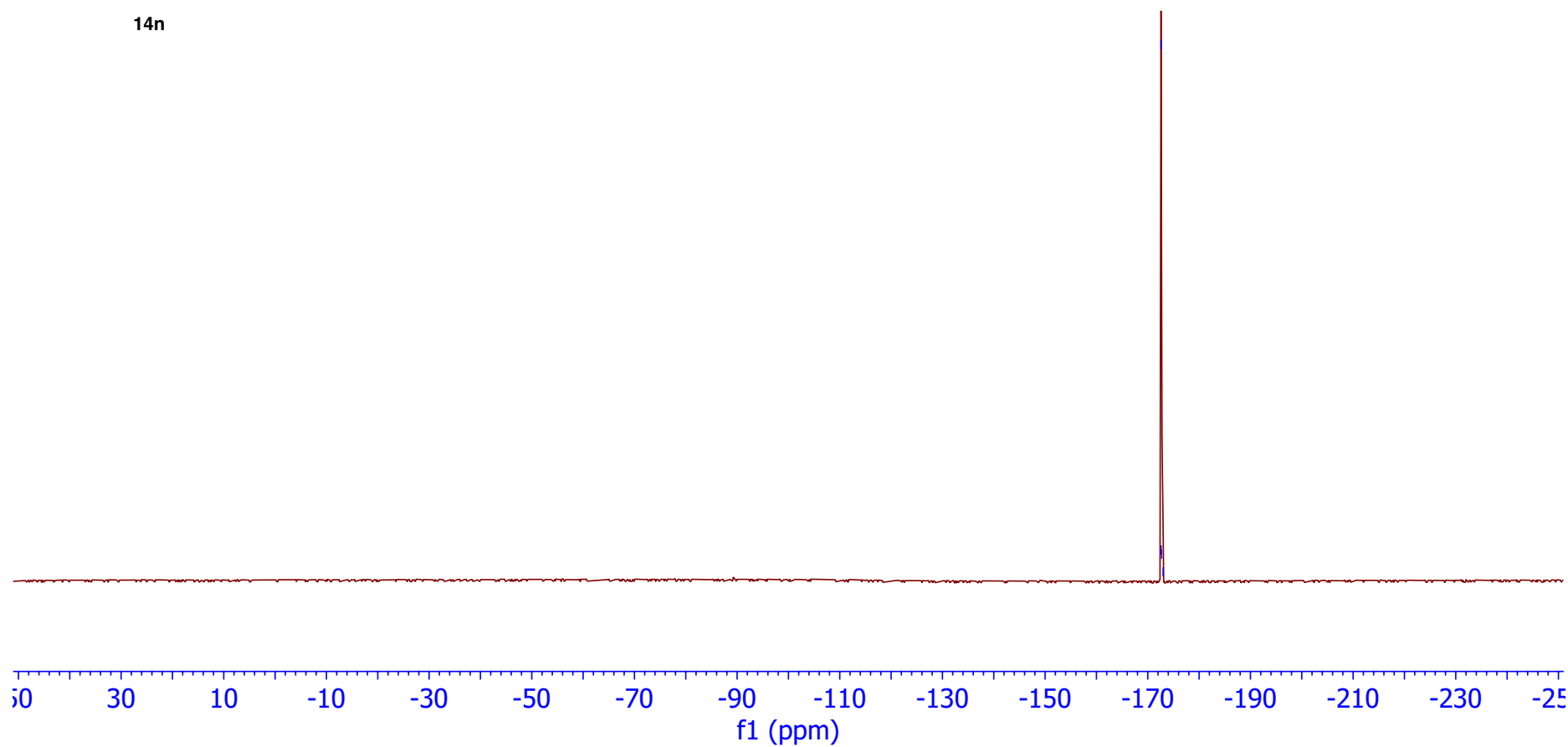
¹³C NMR of 14n

376MHz, Chloroform-d



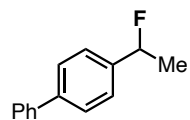
14n

-172.57
-172.62
-172.69
-172.97



^{19}F NMR of 14n

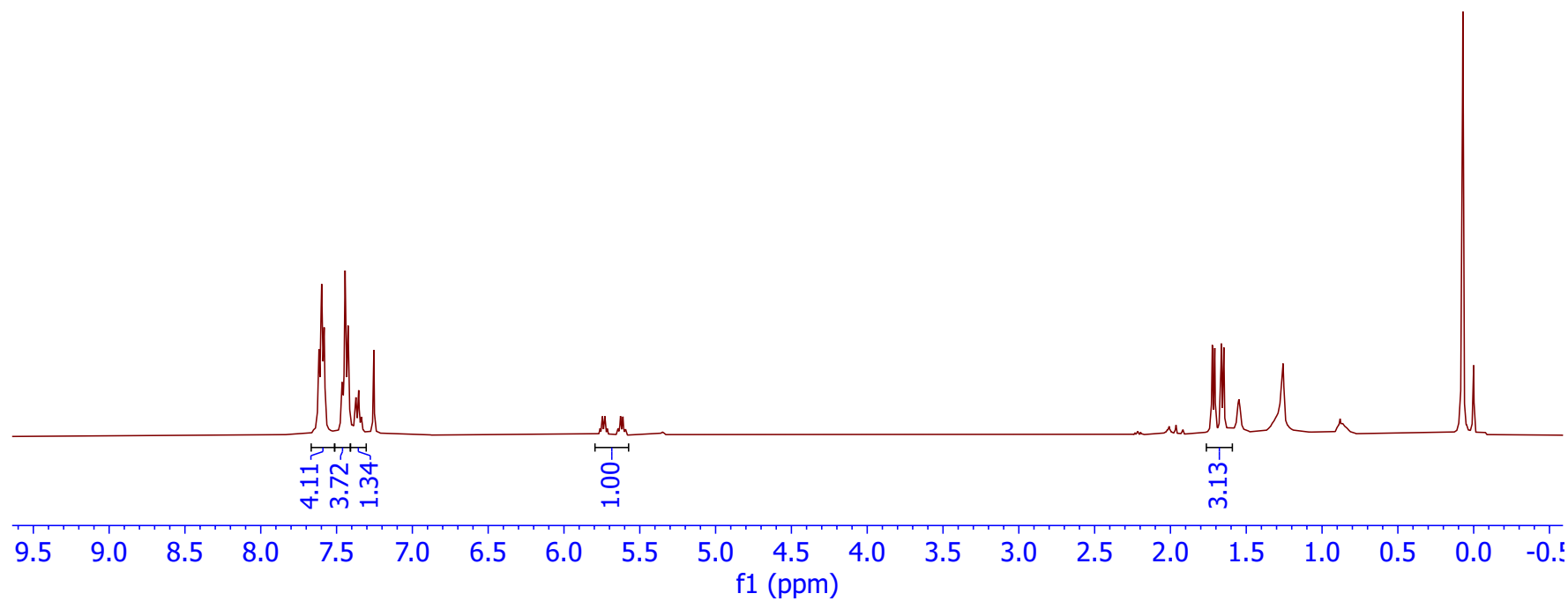
400 HMz, Chloroform-d



14o

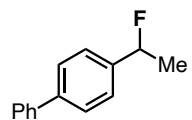
7.62
7.60
7.60
7.58
7.46
7.44
7.42
7.39
7.38
7.37
7.35
5.76
5.75
5.73
5.71
5.64
5.63
5.61
5.60

1.72
1.71
1.66
1.65



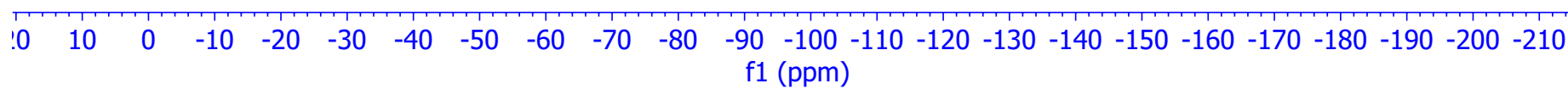
¹H NMR of 14o

376MHz, Chloroform-d



14o

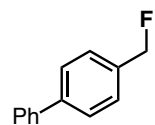
---166.61



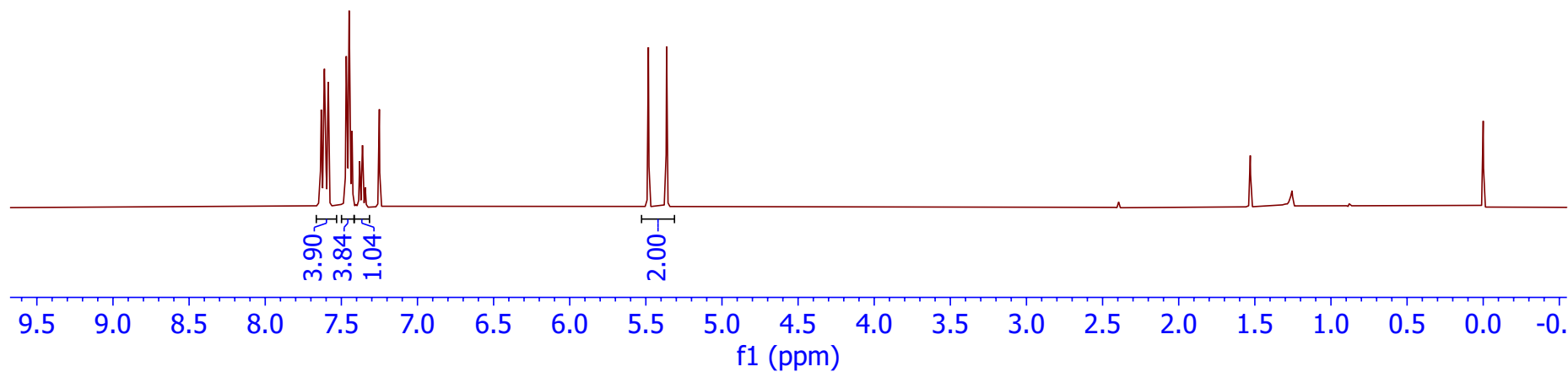
^{19}F NMR of 14o

400 HMz, Chloroform-d

7.63
7.61
7.61
7.60
7.59
7.59
7.47
7.46
7.45
7.44
7.43
7.43
7.38
7.38
7.38
7.37
7.36
7.36
7.35
7.34
5.48
5.36

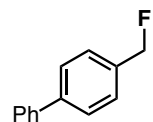


14p

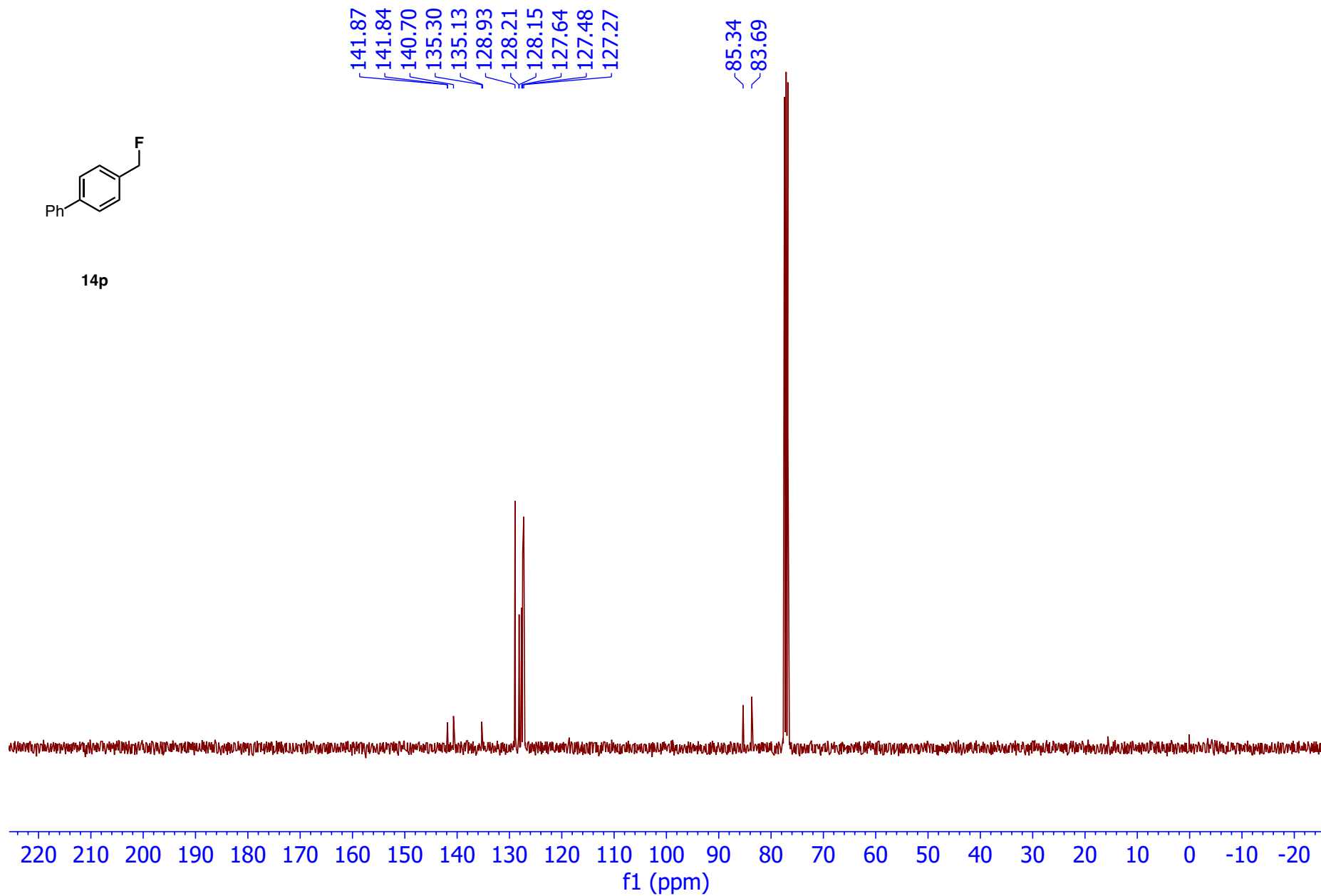


¹H NMR of 14p

101 MHz, Chloroform-d

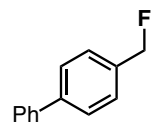


14p



^{13}C NMR of 14p

376MHz, Chloroform-d



14p

-206.02
-206.04
-206.22
-206.23
-206.24
-206.42
-206.43

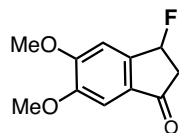


10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210

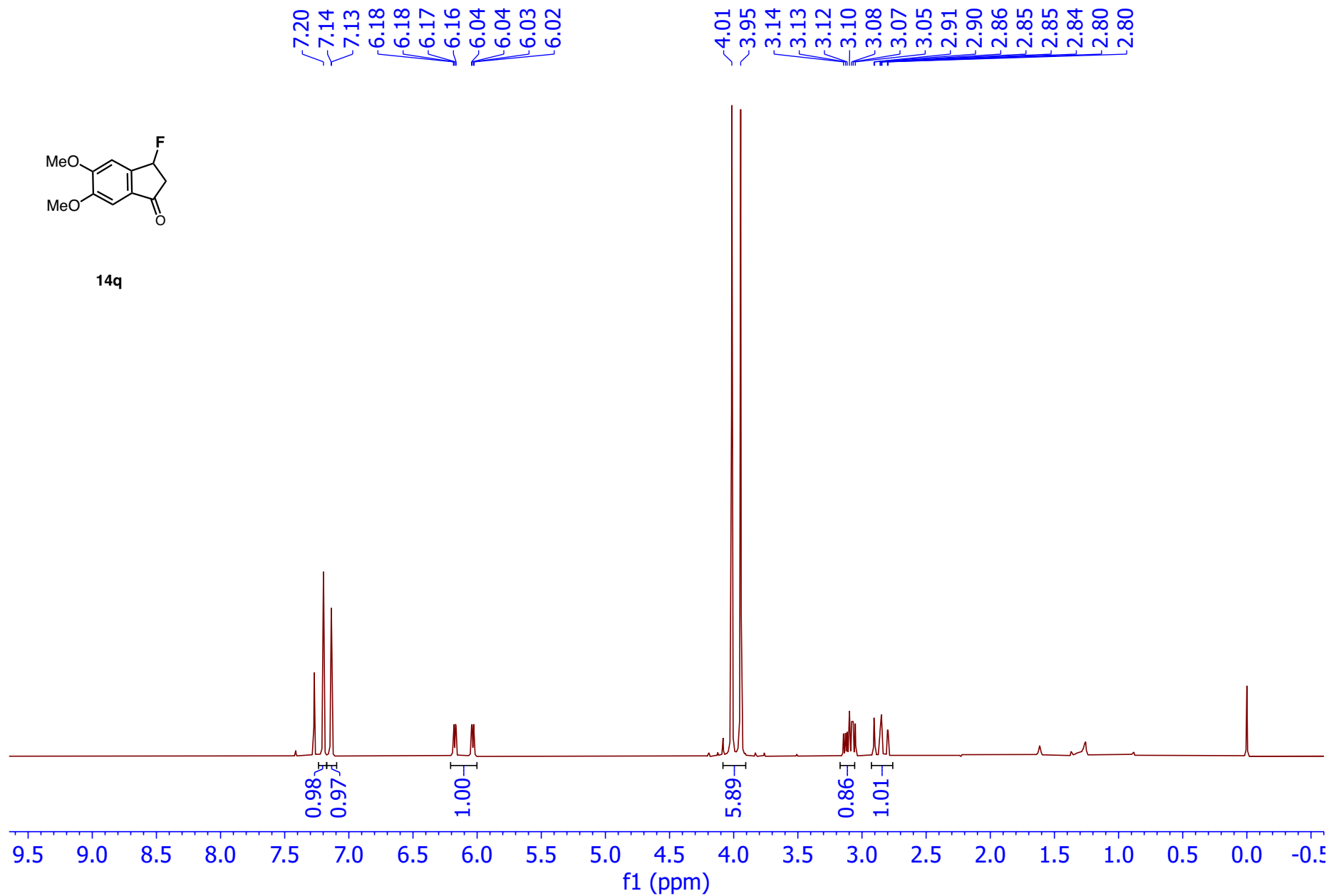
f1 (ppm)

¹⁹F NMR of 14p

400 HMz, Chloroform-d



14q



¹H NMR of 14q

101 MHz, Chloroform-d

198.78

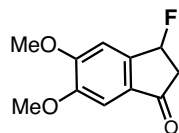
154.80
154.77
150.90
144.32
144.14

129.39
129.37

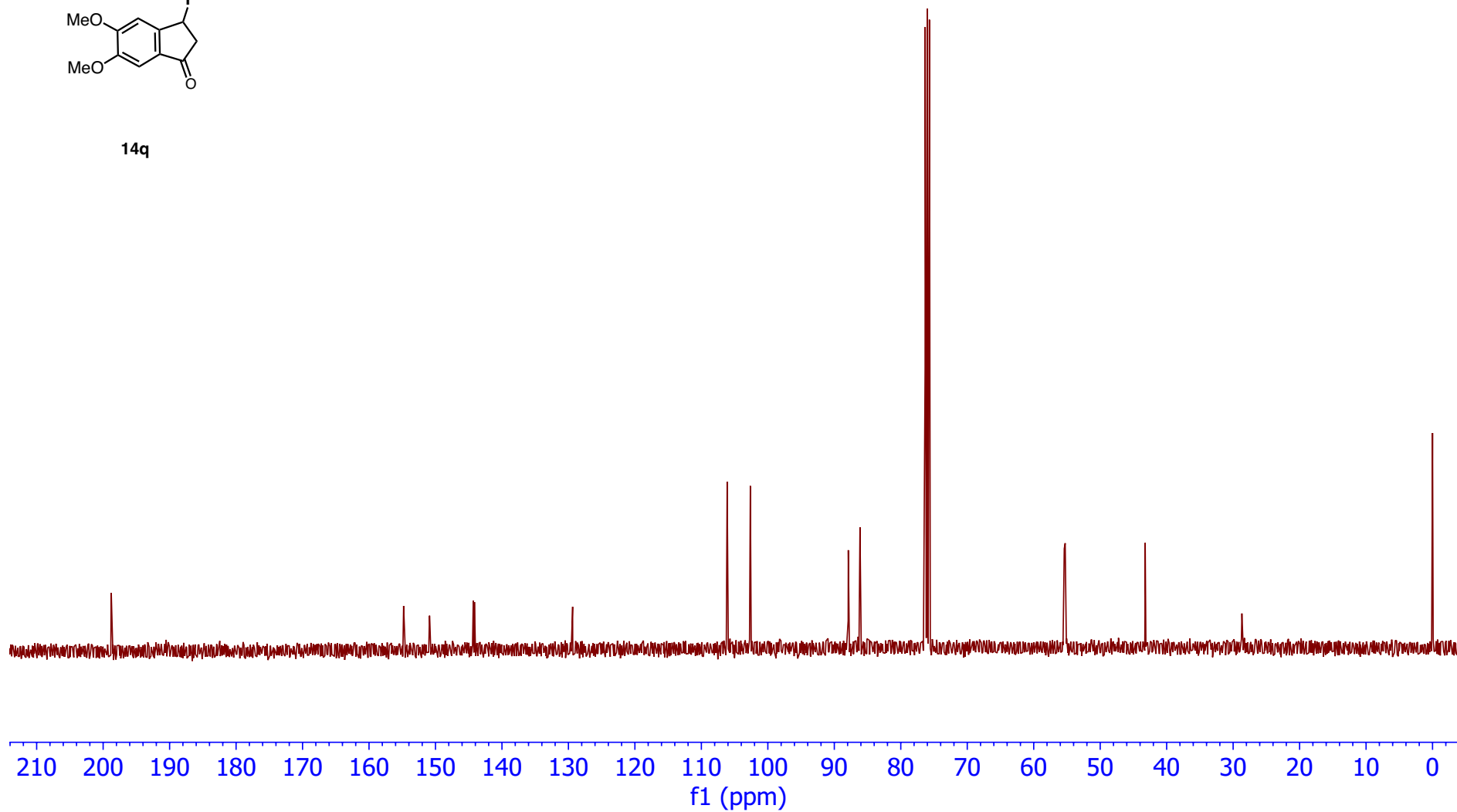
106.12
102.63

87.90
86.14

55.44
55.40
55.27
55.23
43.23
43.02

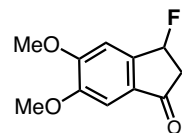


14q

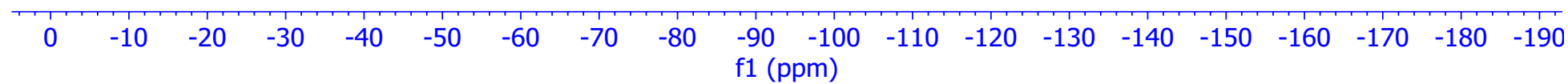


¹³C NMR of 14q

376MHz, Chloroform-d

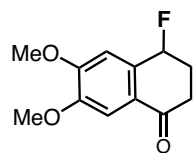


14q

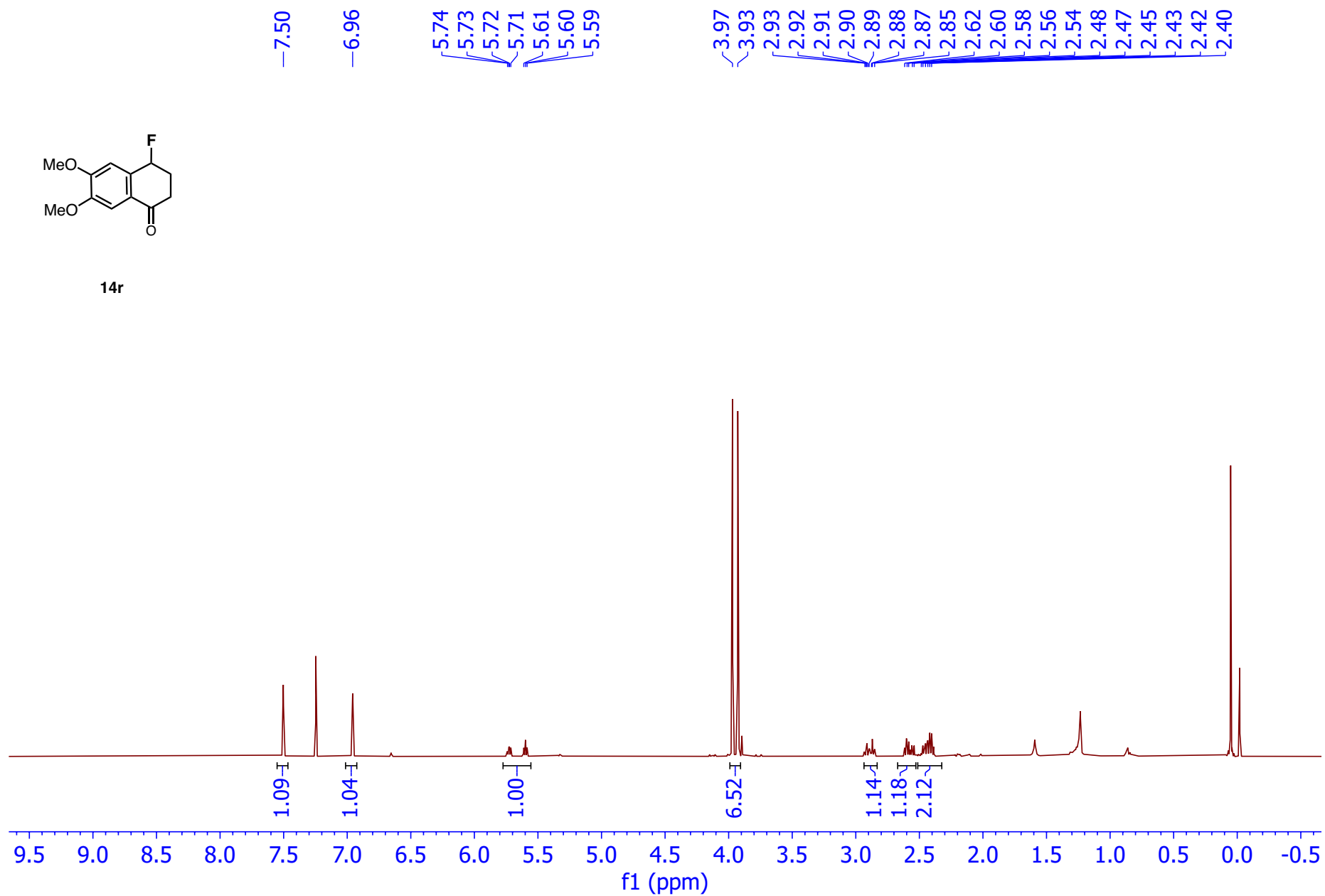


¹⁹F NMR of 14q

400 HMz, Chloroform-d

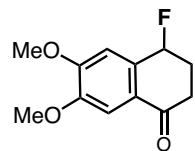


14r

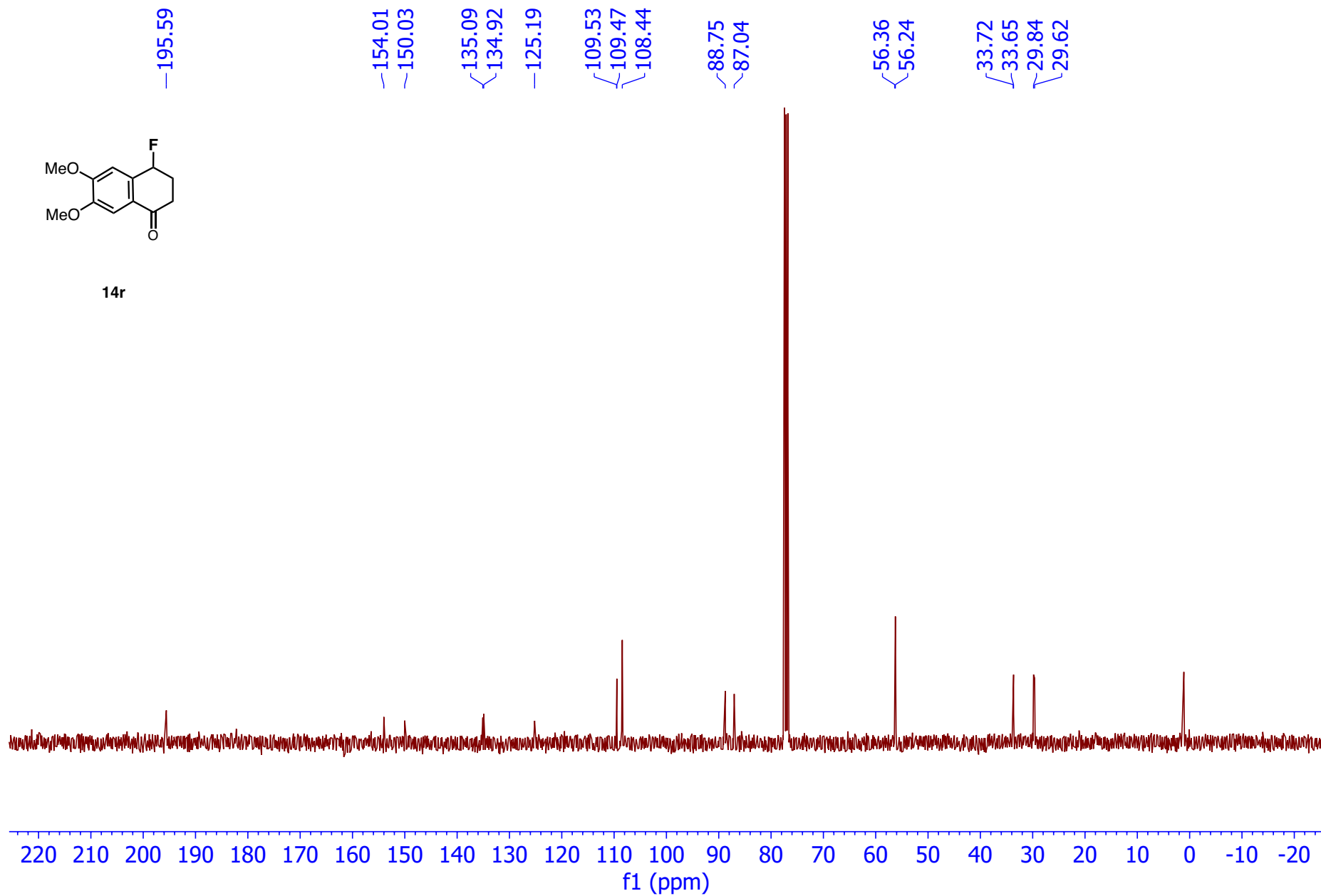


¹H NMR of 14r

101 MHz, Chloroform-d

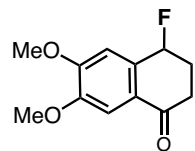


14r



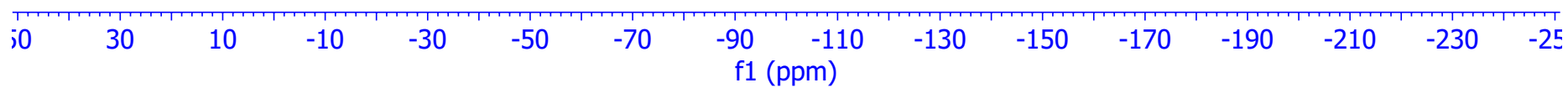
¹³C NMR of 14r

376MHz, Chloroform-d



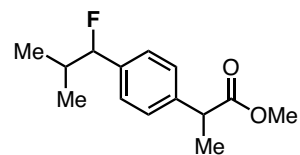
14r

-169.33
-169.34
-169.38
-169.42
-169.46
-169.52

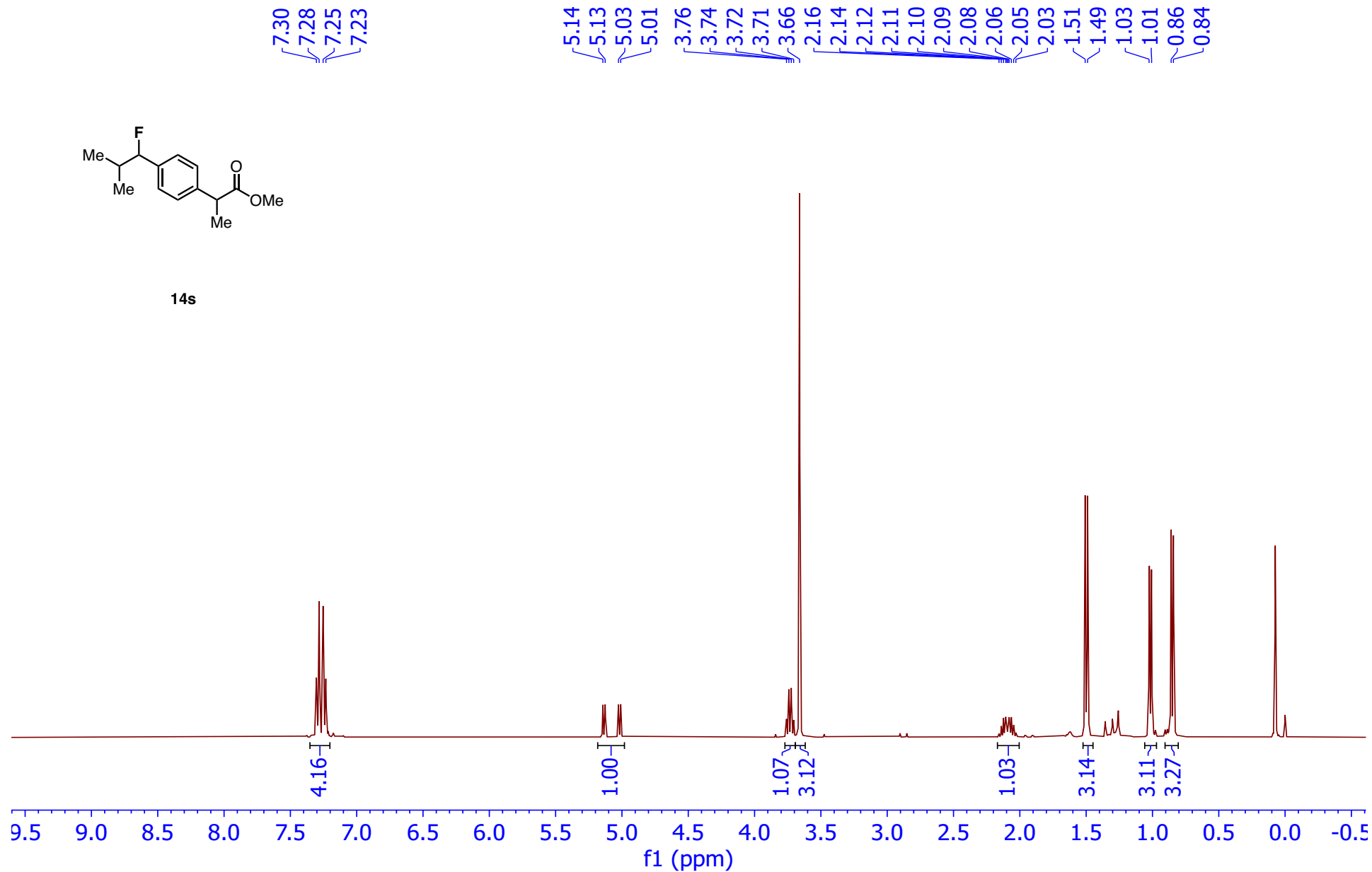


¹⁹F NMR of 14r

400 HMz, Chloroform-d

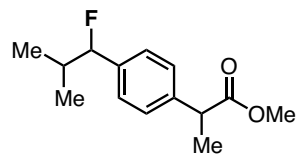


14s

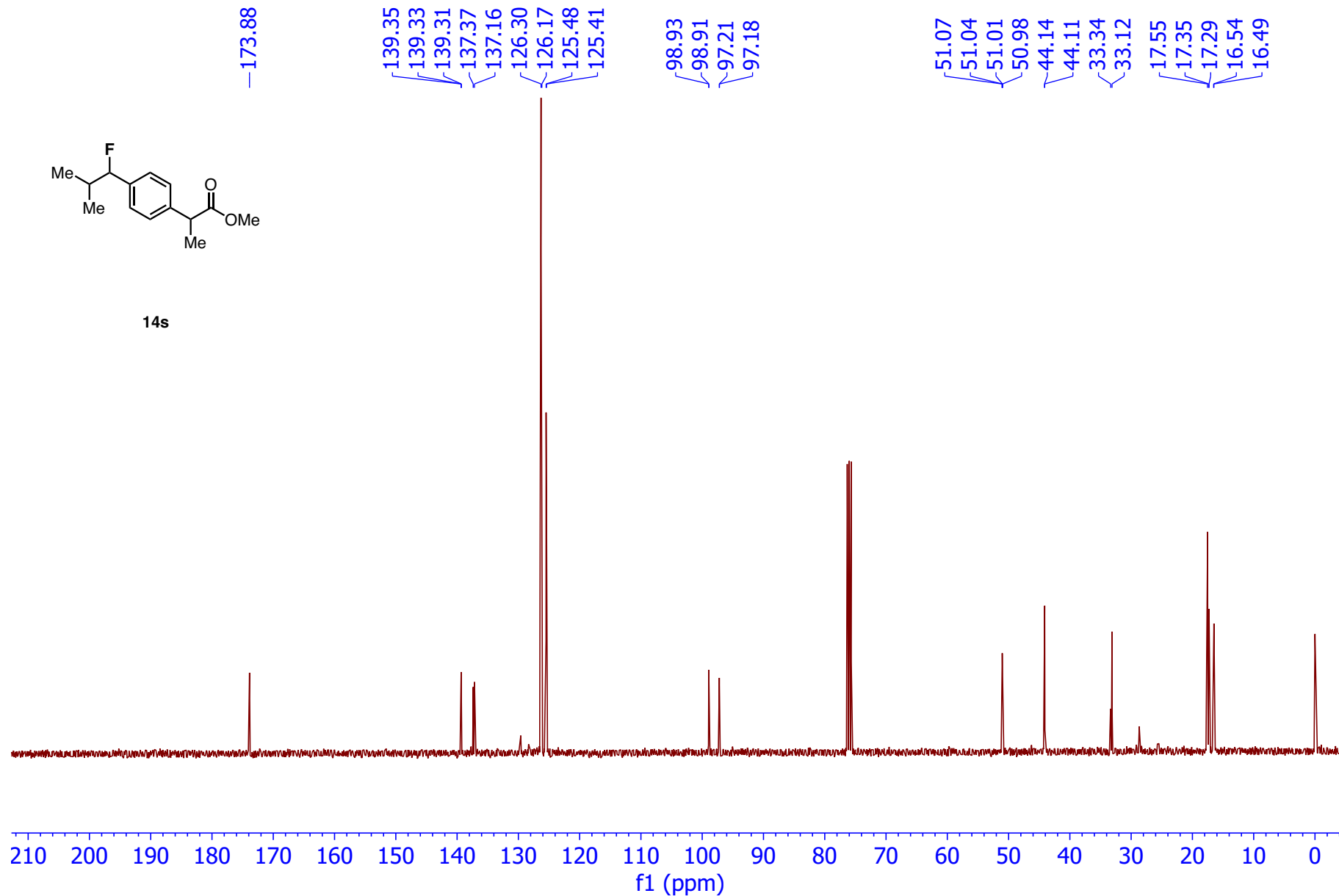


¹H NMR of 14s

101 MHz, Chloroform-d

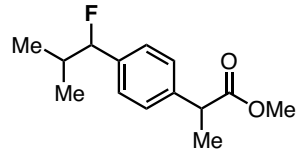


14s



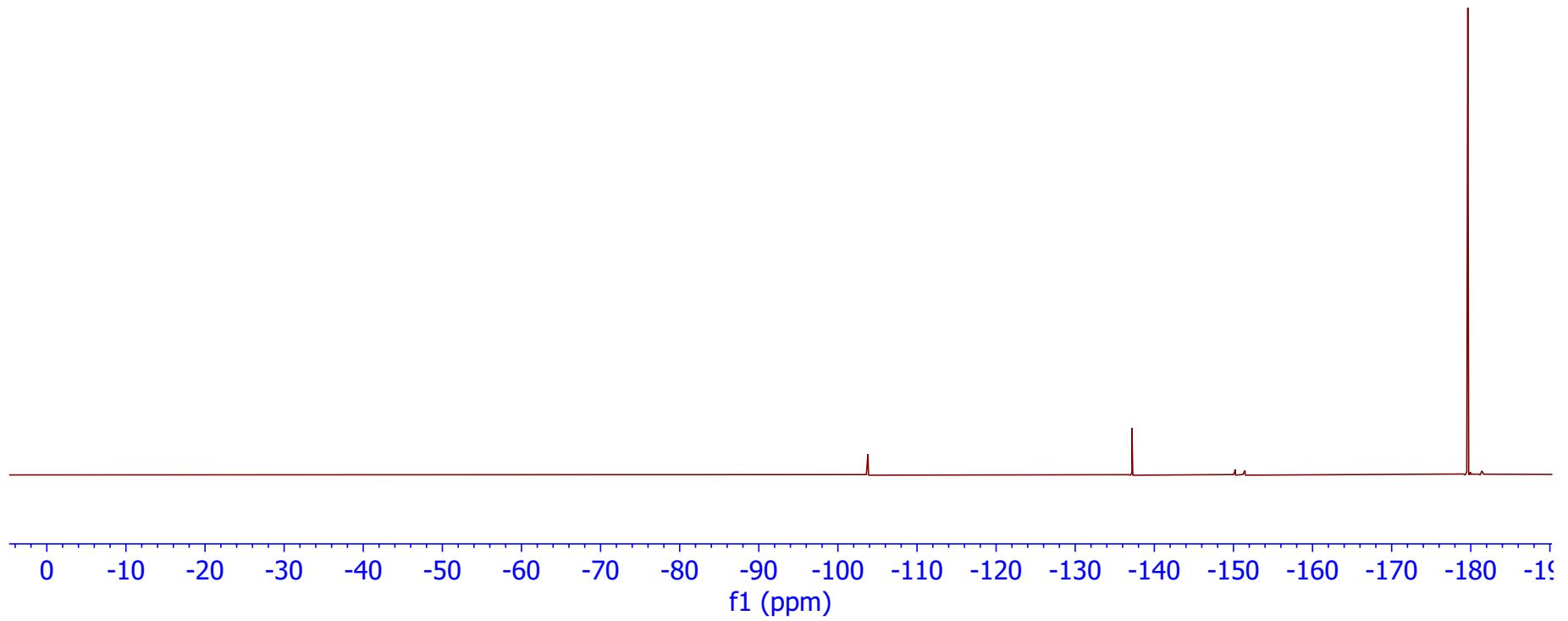
13C NMR of 14s

376MHz, Chloroform-d



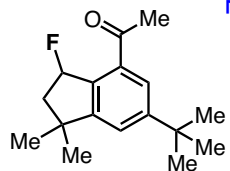
14s

-179.63
-179.65

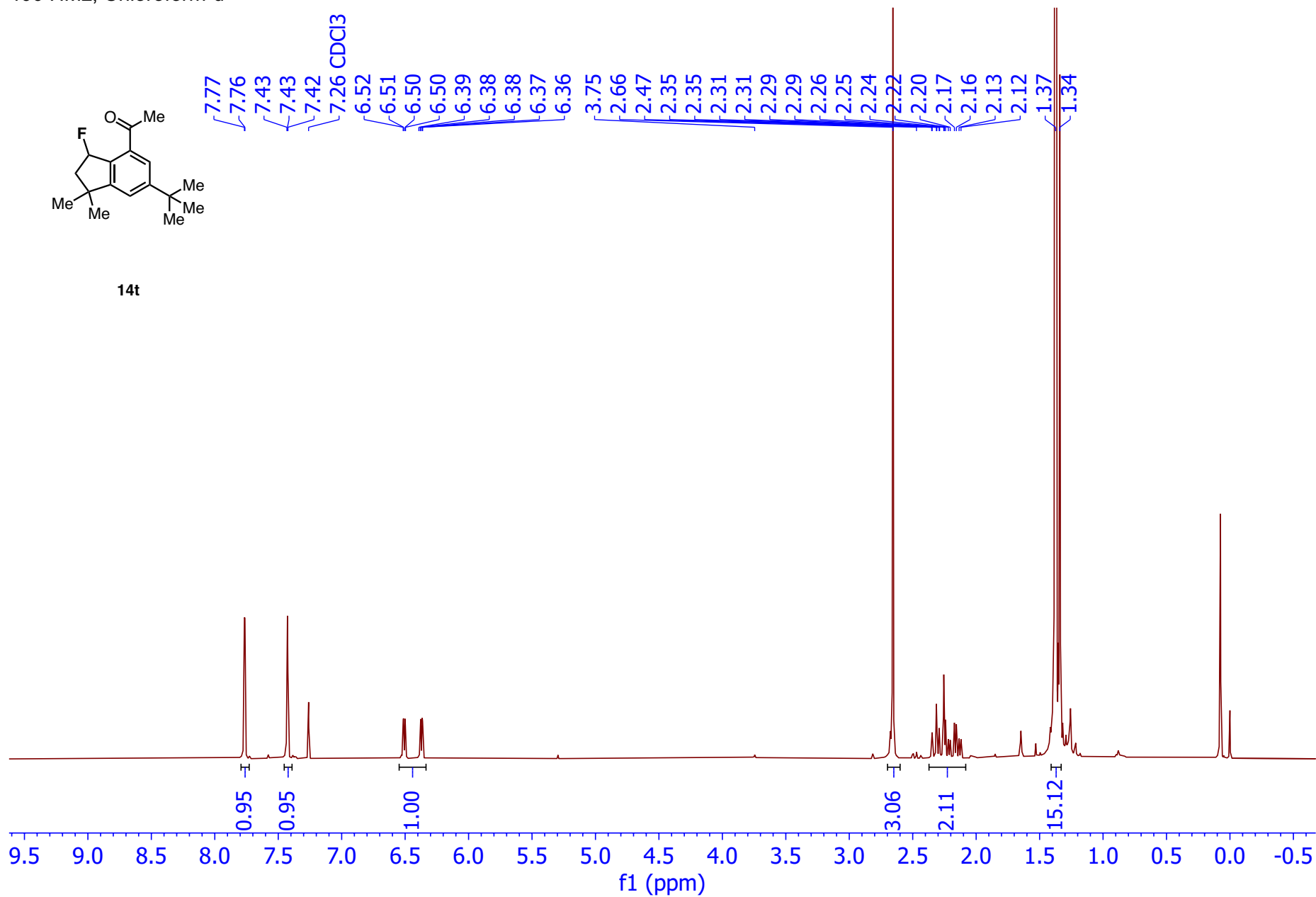


^{19}F NMR of 14s

400 HMz, Chloroform-d

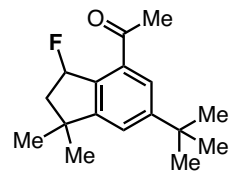


14t

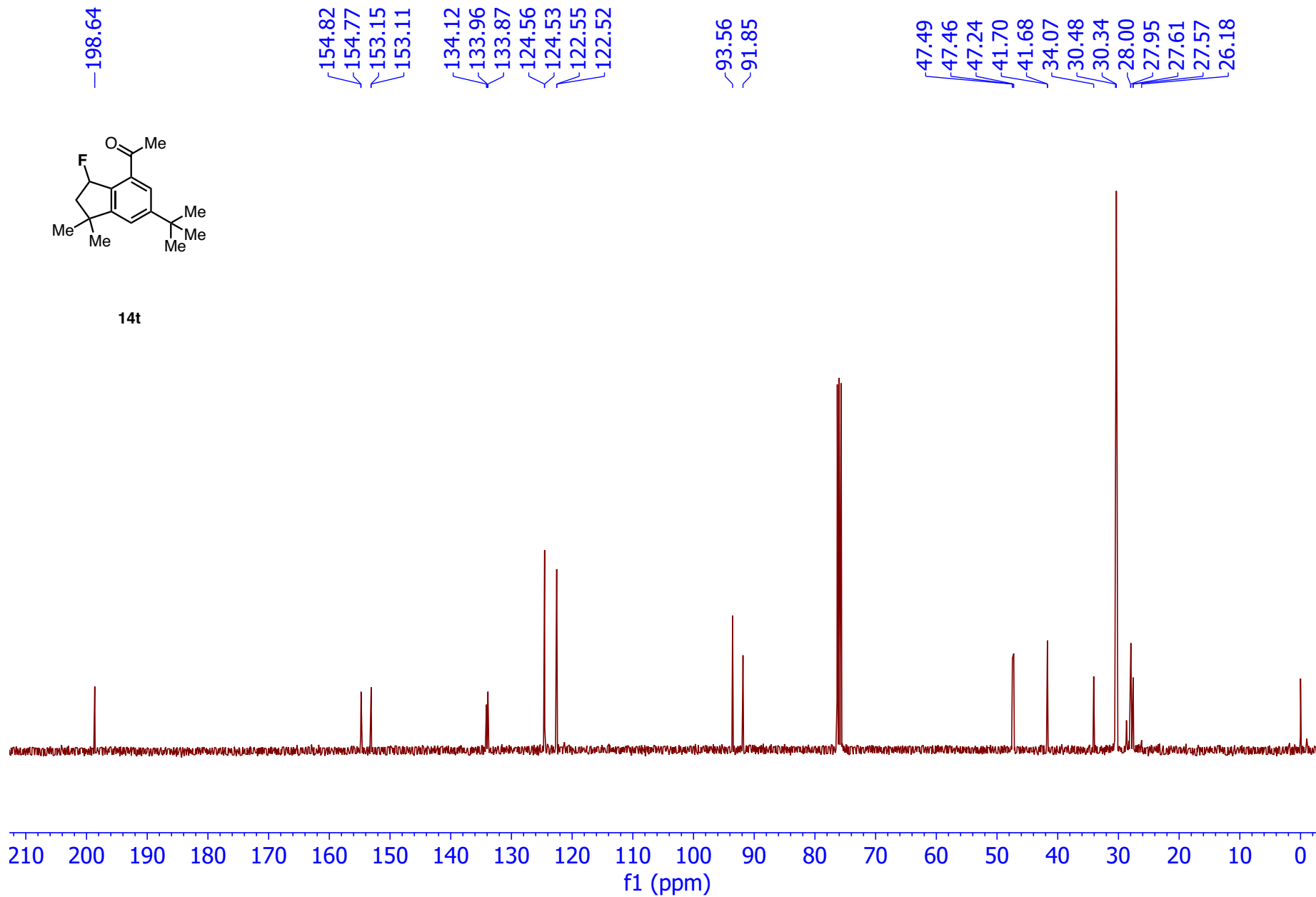


¹H NMR of 14t

101 MHz, Chloroform-d

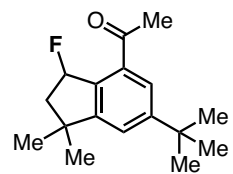


14t



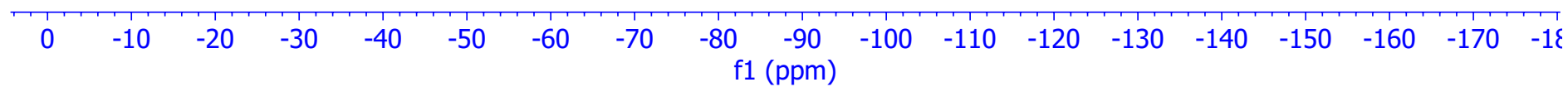
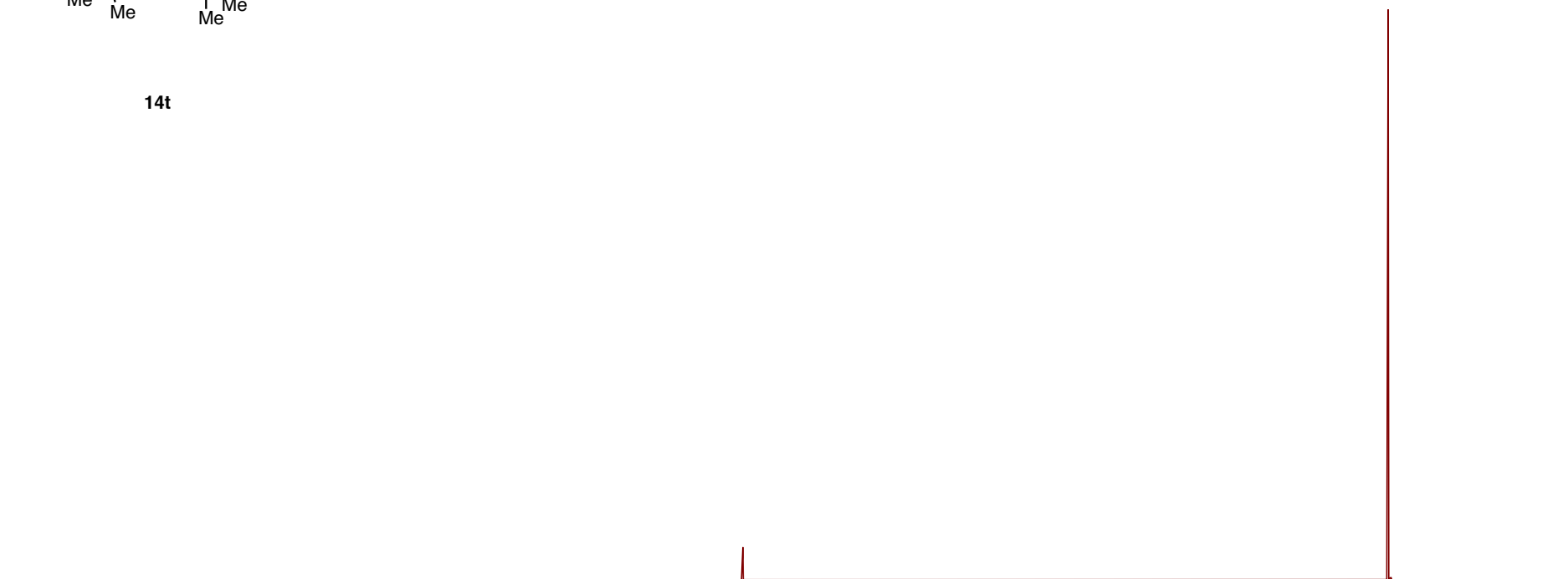
¹³C NMR of 14t

376MHz, Chloroform-d



14t

--158.61



¹⁹F NMR of 14t