

## *Supporting Information*

### **Metal-free stereoselective intramolecular oxyamination of alkynes using a robotic and rapid photocatalytic synthesis and screening system**

Jia-Min Lu<sup>1,2†</sup>, Yihui Mao<sup>1†</sup>, Qihang Guo<sup>1,2</sup>, Chengcheng Zhong<sup>1</sup>, Xian-Ge Gao<sup>2</sup>, Qun Fang<sup>1,2,\*</sup>,  
Zhan Lu<sup>1,\*</sup>

<sup>1</sup> *Department of Chemistry, Zhejiang University, Hangzhou 310058, China.*

<sup>2</sup> *Institute of Intelligent Chemical Manufacturing and iChemFoundry Platform, ZJU-Hangzhou  
Global Scientific and Technological Innovation Center, Hangzhou 311200, China*

*† These authors contributed equally*

Corresponding author: [luzhan@zju.edu.cn](mailto:luzhan@zju.edu.cn); [fangqun@zju.edu.cn](mailto:fangqun@zju.edu.cn)

<b>I. General Information and method of the Automated System.....</b>	<b>S2</b>
<b>II. Optimizations of Reaction Conditions.....</b>	<b>S4</b>
<b>III. Procedures for the Synthesis of Starting Materials.....</b>	<b>S5</b>
<b>IV. Visible-Light-Promoted Intramolecular Oxyamidation of Alkynes.....</b>	<b>S12</b>
<b>V. Synthetic Applications and Mechanistic Studies.....</b>	<b>S24</b>
<b>VII. References.....</b>	<b>S33</b>
<b>IX. NMR Spectra.....</b>	<b>S34</b>

## **I. General Information and Method of the Robotic System.**

THF and toluene were distilled from sodium benzophenoneketyl prior to use. MeCN [Extra Dry, with molecular sieves, Water  $\leq$  50 ppm (by K.F.)], MeCN, EA, HCl, NH<sub>2</sub>OH·HCl, iPr<sub>2</sub>NEt, Neutral Red (NR), Sodium sulfate (Na<sub>2</sub>SO<sub>4</sub>), Triethylamine (Et<sub>3</sub>N), Imidazole and *N,N*-Carbonyldiimidazole (CDI) were purchased from Energy and used as received. The other commercially available chemicals were used as received without mentioned. 5 W blue LEDs were used as the light source for the scale-up reaction. NMR spectra were recorded on a Bruker-400 instrument or Oxford instrument. <sup>1</sup>H NMR chemical shifts were referenced to tetramethylsilane signal (0 ppm), <sup>13</sup>C NMR chemical shifts were referenced to the solvent resonance (77.00 ppm, CDCl<sub>3</sub>), <sup>19</sup>F NMR chemical shifts were referenced to the solvent resonance. The following abbreviations (or combinations thereof) were used to explain multiplicities: s = singlet, d = doublet, t = triplet, m = multiplet, br = broad, q = quadruplet, PE = petroleum ether, EA = ethyl acetate, THF = tetrahydrofuran, DCM = dichloromethane. IR spectra were recorded on a Perkin-Elmer Spectrum One FTIR spectrometer with diamond ATR accessory. High-resolution mass spectra (HRMS) were recorded on LCMS-IT-TOF (ESI-TOF) and EI-TOF (electro-spray ionization-time of flight). Melting points were obtained using an X-4 melting point apparatus (Laboratory Devices, Beijing Taike Co.).

### **General Information of the Robotic System**

The light source module consisted of 4 lasers (5.5 W,  $\lambda_{\max}$  = 450 nm, Oxlasers, Shanghai, China) and 4 optical fibers (400- $\mu$ m diameter, Lanwin Technology, Zhongshan, China) for conducting the light into the LCW photocatalytic microreactor. The reaction mixture was prepared using the liquid handling module with a syringe pump (20738325 Cavro XCalibur Pump, Tecan, Männedorf, Switzerland) and a 10-port selective valve (C5-3000EUTF, VICI, Schenkkan, Switzerland), connected with a PFA capillary (1512L, IDEX, Northbrook, USA). The LCW photocatalytic microreactor was designed as a cannula configuration with an inner fused-silica capillary (530  $\mu$ m i.d. and 690  $\mu$ m o.d., Refined Chromatography Co., Yongnian, China) and an outer glass capillary (1.5 mm i.d. and 2.5 mm o.d., Jingke, Guangzhou, China), installed with two polypropylene tees (3.2 mm, 1/8, Jieliantech Technology Co., Beijing, China). The reaction channel and the PFA capillary were connected using unions (P-702, IDEX, Northbrook, USA) or tees (P-712, IDEX, Northbrook,

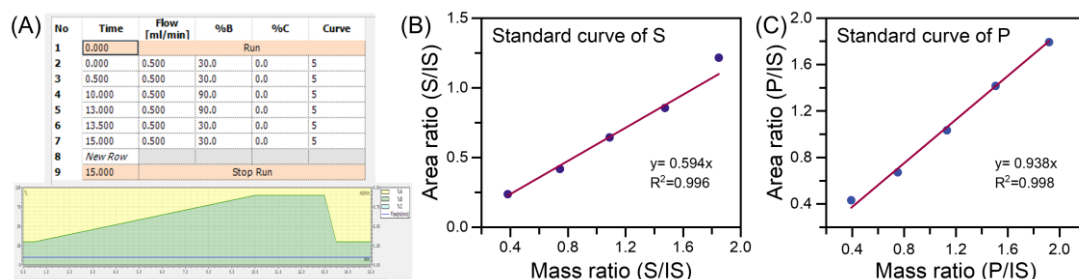
USA). In order to perform the synthesis at the set temperatures, the cooling solution was circulated in the outer capillary using a diaphragm pump (KVP004, Kamoer Fluid Technology Co., Shanghai, China), and the temperature of the circulating cooling solution was monitored in real time by a digital temperature controller (XH-W3002, Xinghe Electronic Technology Co., Suqian, China), which could control the on and off of the semiconductor chilling plate (TEC1-12706, Xinghe Electronic Technology Co., Suqian, China). The self-developed control program was written with LabView (8.0, National Instruments, Austin, USA).

### **Basic Operation of the Robotic System**

Utilizing the self-developed control program, the operation of the 10-port selective valve, the syringe pump, the LCW photocatalytic microreactor, and the lasers was automatically controlled, and different photocatalyst species and solvents were selected. For the optimization of photocatalysts, the desired volumes of the stock solutions (0.2 M substrate, 0.2 M internal standard, 0.02 M photocatalyst and 0.004 M DIPEA in MeCN) stored in argon were aspirated using the 10-port selective valve and the syringe pump to prepare the reaction mixtures before synthesis. After fast and homogeneous mixing, the reaction mixture was delivered into the reaction channel of the LCW photocatalytic microreactor for rapid synthesis. For the optimization of solvents, the reaction mixtures were prepared and stored in argon, and then delivered to the LCW photocatalytic microreactor for synthesis as well. Unless otherwise mentioned, the difunctionalization of alkynes were performed in the LCW photocatalytic microreactor with 2.5 min residence time under automated temperature control (e.g.,  $40 \pm 2$  °C) and uniform light irradiation from the 3 W 450 nm lasers. A parallel photoreactor (PL-SX100A, Beijing Pulinsaisi Technology Co., Beijing, China) was used for the synthesis of different substrates. After synthesis, the reaction mixture was collected, diluted and filtered. Then, the product yields of the reaction mixture were obtained using the HPLC (Vanquish Core, C<sub>18</sub> column, Thermo Fisher Scientific, Waltham, USA) analysis method with a UV-Vis detector at the detection wavelength of 220 nm. During the HPLC analysis, the mobile phase A was the ultrapure water obtained from a water purification system (Barnstead MicroPure, Thermo Fisher Scientific, Waltham, USA), and the mobile phase B was MeCN.

The quantitative results of the condition screening were obtained by HPLC analysis method. For the difunctionalization of alkynes, the HPLC gradient elution method was optimized (from 30%B to 90%B) and the detection time for each sample was 15 min, which could match the time

consumed for condition screening of 14 min (**Fig. S1A**). The standard curves of the model substrate and its corresponding product were made, respectively, using 1,3,5-trimethylbenzene as the internal standard substance (**Fig. S1B, S1C**). When screening the conditions of the model substrate, only a small amount of reaction mixture (c.a. 5  $\mu\text{L}$ ) was collected, diluted and then analyzed by the HPLC system to obtain information such as product yields and substrate recovery.



**Fig. S1.** Quantitative analysis using HPLC method. (A) Optimized HPLC gradient elution method. (B) Standard curve of the model substrate. (C) Standard curves of the model product.

## II. Optimizations of Reaction Conditions.

It took only 14 min to complete each condition screening rapidly, among which only 2.5 min residence time was needed for synthesis, 2 min was for sample collection after synthesis, 5.5 min was for rinsing and replacing sample of the reaction channel, and 4 min was for sample preparation, including reactants introducing, diluting, mixing, and so on.

Utilizing the self-developed system for automated photocatalytic synthesis and screening, the optimized conditions and results on different photocatalysts are shown in **Figure 2**. The optimized conditions and results on different solvents are shown in **Figure S2**. In most solvents, such as MeOH, THF, DCM, DMF, acetone, dioxane, and so on, the reaction gave **2a** in 8-73% yield. The reaction using MeCN as a solvent could afford **2a** in 81% yield, so MeCN was chosen as the optimal solvent. The optimization results of the amount of DIPEA and reaction temperature are also shown in **Figure S2**. The reaction could give **2a** in 76% yield with 2 mol% DIPEA, in 45% with 1 e.q. DIPEA, and in 60% without DIPEA. What's more, the reaction could deliver **2a** in 59% yield at 20 °C. As a result, 10 mol% was chosen as the appropriate DIPEA amount, and 40 °C was chosen as the optimal reaction temperature. The reaction was also carried out using MeCN (SCR) and the yield of **2a** was

74%. Addition of 10 e.q. of water to the reaction mixture slightly decreased the yield of **2a** to 59%, suggesting that the synthesis strategy can tolerate the presence of water to some extent.

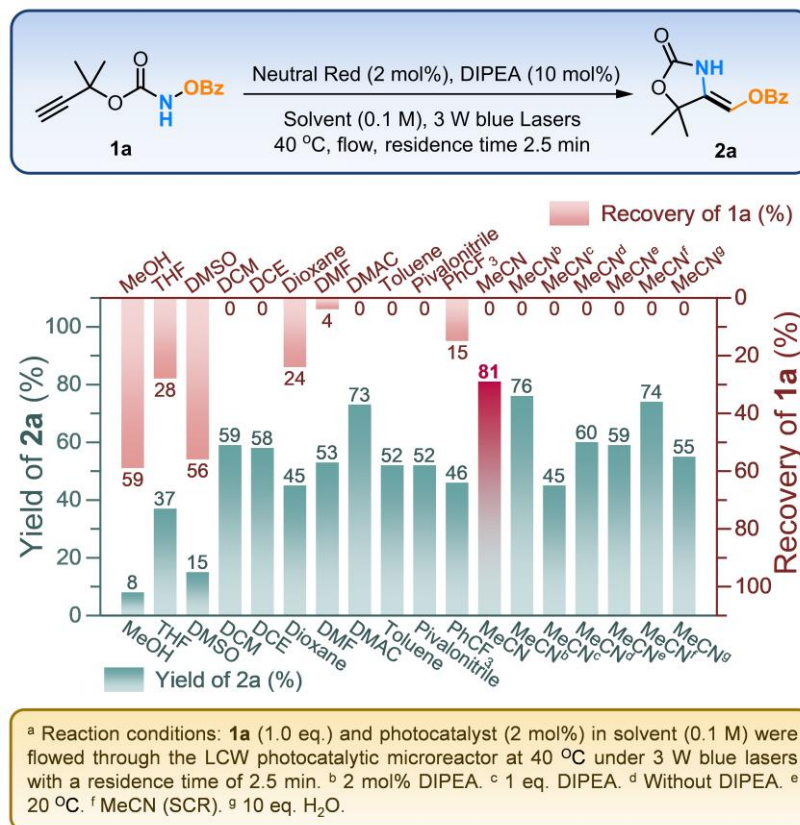
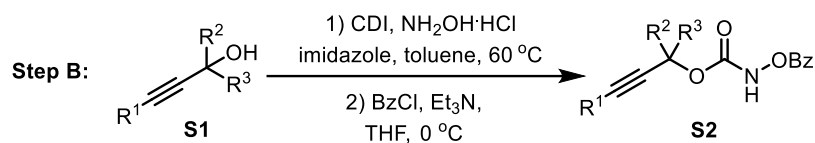


Fig. S2. Optimizations of solvents, DIPEA amount and reaction temperature. <sup>a</sup>

### III. Procedures for the Synthesis of Starting Materials

Starting materials were prepared by the following steps:

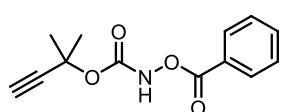
**Step A.** Substituted propargyl alcohol **S1** were prepared according to the previously reported procedures.<sup>1</sup>



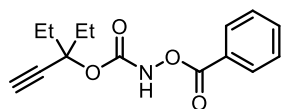
**Step B.** Alkyl benzoyloxycarbamate **S2** were prepared according to a reported procedure.<sup>1</sup> A 250

mL round-bottom flask was charged with **S1** (60 mmol), toluene (100 mL), *N,N*-Carbonyldiimidazole (78 mmol, 1.3 equiv). The mixture was stirred at 60 °C, after **S2** was fully converted to corresponding intermediate (monitored by TLC), the reaction mixture was concentrated by rotary evaporation. Then the resulting suspension was dissolved in DCM, and washed with water, brine, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated by rotary evaporation. Then the resulting suspension was dissolved in MeCN, added with imidazole (120 mmol, 2 equiv) and NH<sub>2</sub>OH·HCl (180 mmol, 3 equiv). After the intermediate was fully consumed (monitored by TLC), the reaction mixture was concentrated by rotary evaporation. Then the resulting suspension was dissolved in HCl (60 mL, 1 M), the aqueous phase was extracted with EA (50 mL x 3). The organic layers were combined, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated by rotary evaporation, and further purified by flash chromatography on silica gel (PE/EA = 10/1~1/1) to afford the corresponding hydroxylamine.

Hydroxylamine (1 equiv) was dissolved in THF (0.2 M) in a 250 mL round-bottom flask, added with Et<sub>3</sub>N (1.0 equiv), then added benzoyl chloride (1.0 equiv, 1 M in THF) slowly at 0 °C. The mixture was stirred at room temperature for 1~2 hours (monitored by TLC), quenched with H<sub>2</sub>O, and separated. The aqueous phase was extracted with DCM (40 mL x 3). The organic layers were combined, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated by rotary evaporation, and further purified by flash chromatography on silica gel (PE/EA = 10/1~5/1) to afford the corresponding product **S2**.

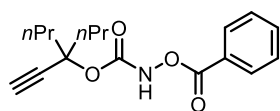


**2-Methylbut-3-yn-2-yl (benzoyloxy)carbamate (1a):** 2.85 g, 77% yield, white solid; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.43 (brs, 1H), 8.14-8.04 (m, 2H), 7.63 (t, *J* = 7.6 Hz, 1H), 7.48 (t, *J* = 7.6 Hz, 2H), 2.60 (s, 1H), 1.74 (s, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 165.8, 154.5, 134.2, 129.9, 128.6, 126.7, 83.7, 74.4, 73.2, 28.9; The characterization data of the product is consistent with the reported literature.<sup>1</sup>

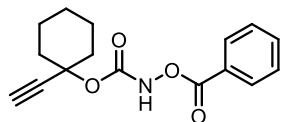


**3-Ethylpent-1-yn-3-yl (benzoyloxy)carbamate (1b):** 4.46 g, 85% yield, white solid, m.p. = 95-96 °C, IR (neat): 3300, 2980, 1746, 1456, 1231 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.43 (brs, 1H), 8.15-8.02 (m, 2H), 7.62 (t, *J* = 7.6 Hz, 1H), 7.47 (dd, *J* = 7.6 Hz, 2H), 2.62 (s, 1H), 2.17-2.04 (m, 2H), 2.04-1.91

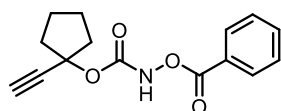
(m, 2H), 1.01 (t,  $J = 7.4$  Hz, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 154.5, 134.1, 129.8, 128.6, 126.7, 82.2, 81.9, 74.9, 30.9, 8.1; HRMS (ESI) calculated for  $[\text{C}_{15}\text{H}_{17}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  298.1050, found:  $m/z$  298.1049.



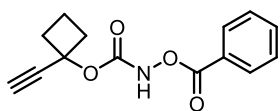
**4-Ethynylheptan-4-yl (benzoyloxy)carbamate (1c):** 4.22 g, 70% yield, white solid, m.p. = 63-66 °C, IR (neat): 3300, 2923, 1747, 1453, 1233  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.31 (brs, 1H), 8.10 (d,  $J = 7.4$  Hz, 2H), 7.63 (t,  $J = 7.6$  Hz, 1H), 7.48 (t,  $J = 7.6$  Hz, 2H), 2.62 (s, 1H), 2.12-1.98 (m, 2H), 1.96-1.84 (m, 2H), 1.56-1.42 (m, 4H), 0.94 (t,  $J = 7.4$  Hz, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 154.5, 134.1, 129.9, 128.6, 126.7, 82.4, 81.4, 74.8, 40.5, 17.2, 13.9; HRMS (ESI) calculated for  $[\text{C}_{17}\text{H}_{21}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  326.1363, found:  $m/z$  326.1364.



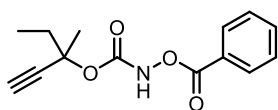
**1-Ethynylcyclohexyl (benzoyloxy)carbamate (1d):** 4.05 g, 64% yield, white solid, m.p. = 135-137 °C, IR (neat): 3294, 2936, 2860, 1743, 1451, 1227  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.38 (brs, 1H), 8.13-8.27 (m, 2H), 7.67-6.60 (m, 1H), 7.48 (t,  $J = 7.6$  Hz, 2H), 2.67 (s, 1H), 2.21-2.09 (m, 2H), 2.22-1.89 (m, 2H), 1.72-1.57 (m, 5H), 1.55-1.46 (m, 1H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 154.4, 134.2, 129.9, 128.6, 126.7, 82.6, 77.9, 75.0, 36.9, 24.8, 22.3; HRMS (ESI) calculated for  $[\text{C}_{16}\text{H}_{17}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  310.1050, found:  $m/z$  310.1051.



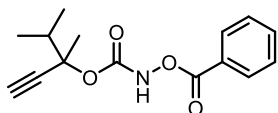
**1-Ethynylcyclopentyl (benzoyloxy)carbamate (1e):** 3.17 g, 77% yield, white solid, m.p. = 110-112 °C, IR (neat): 3291, 2957, 1744, 1452, 1234  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.31 (brs, 1H), 8.10 (d,  $J = 7.6$  Hz, 2H), 7.64 (t,  $J = 7.6$  Hz, 1H), 7.48 (t,  $J = 7.6$  Hz, 2H), 2.64 (s, 1H), 2.46-2.26 (m, 2H), 2.23-2.13 (m, 2H), 1.90-1.62 (m, 4H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 154.8, 134.2, 129.9, 128.7, 126.6, 83.0, 82.9, 73.9, 40.3, 23.1; HRMS (ESI) calculated for  $[\text{C}_{15}\text{H}_{15}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  296.0893, found:  $m/z$  296.0894.



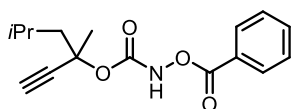
**1-Ethynylcyclobutyl (benzoyloxy)carbamate (1f):** 3.74 g, 41% yield, white solid, m.p. = 100-102 °C, IR (neat): 3271, 1740, 1453, 1229, 1103  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.39 (brs, 1H), 8.13-8.07 (m, 2H), 7.67-7.60 (m, 1H), 7.53-7.45 (m, 2H), 2.66 (s, 1H), 2.65-2.57 (m, 2H), 2.56-2.45 (m, 2H), 2.04-1.89 (m, 2H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.7, 154.4, 134.3, 129.9, 128.7, 126.5, 82.9, 73.6, 73.5, 36.4, 14.1; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{13}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  282.0737, found:  $m/z$  282.0738.



**3-Methylpent-1-yn-3-yl (benzoyloxy)carbamate (1g):** 0.88 g, 90% yield, white solid, m.p. = 90-91 °C, IR (neat): 3289, 2976, 1744, 1455, 1235  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.31 (brs, 1H), 8.10 (d,  $J$  = 7.2 Hz, 2H), 7.64 (t,  $J$  = 7.2 Hz, 1H), 7.49 (t,  $J$  = 7.6 Hz, 2H), 2.62 (s, 1H), 2.10-1.97 (m, 1H), 1.97-1.84 (m, 1H), 1.75 (s, 3H), 1.05 (t,  $J$  = 7.2 Hz, 3H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 154.5, 134.2, 129.9, 128.7, 126.7, 82.7, 78.3, 74.1, 34.4, 26.0, 8.4; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{15}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  284.2893, found:  $m/z$  284.0895.



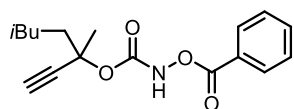
**3,4-Dimethylpent-1-yn-3-yl (benzoyloxy)carbamate (1h):** 0.44 g, 77% yield, white solid, m.p. = 88-90 °C, IR (neat): 3274, 2973, 1746, 1455, 1236  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.30 (brs, 1H), 8.13-8.06 (m, 1H), 7.64 (t,  $J$  = 7.6 Hz, 1H), 7.49 (t,  $J$  = 7.6 Hz, 2H), 2.61 (s, 1H), 2.30-2.18 (m, 1H), 1.74 (s, 3H), 1.05 (d,  $J$  = 6.8 Hz, 3H), 1.02 (d,  $J$  = 6.8 Hz, 3H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.9, 154.6, 134.2, 129.9, 128.7, 126.7, 81.8, 81.5, 74.8, 37.4, 23.3, 17.3, 17.0; HRMS (ESI) calculated for  $[\text{C}_{15}\text{H}_{17}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  298.1050, found:  $m/z$  298.1050.



**3,5-Dimethylhex-1-yn-3-yl (benzoyloxy)carbamate (1i):** 1.33 g, 74% yield, white solid, m.p. = 71-73 °C, IR (neat): 3302, 2957, 1747, 1457, 1234  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.28 (brs, 1H), 8.10 (d,  $J$  = 7.6 Hz, 2H), 7.64 (t,  $J$  = 7.6 Hz, 1H), 7.48 (t,  $J$  = 7.6 Hz, 2H), 2.63 (s, 1H), 2.06-1.87 (m, 2H), 1.83-1.74 (m, 4H), 0.98 (d,  $J$  = 6.8 Hz, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 154.5, 134.2, 129.9, 128.6, 126.7, 83.1, 77.9, 74.4, 49.4, 27.3, 24.8, 24.0, 23.7; HRMS (ESI) calculated for

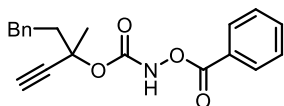


$[C_{16}H_{19}NNaO_4]^+$  ( $M+Na^+$ ) requires  $m/z$  312.1206, found:  $m/z$  312.1208.



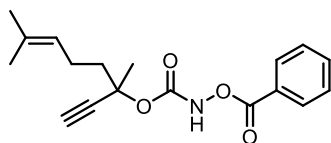
**3,6-Dimethylhept-1-yn-3-yl (benzyloxy)carbamate (1j):** 0.23 g, 59%

yield, white solid, m.p. = 49-52 °C, IR (neat): 3290, 2956, 1748, 1459, 1234  $cm^{-1}$ ;  $^1H$  NMR: (400 MHz,  $CDCl_3$ )  $\delta$  8.29 (brs, 1H), 8.10 (d,  $J$  = 7.2 Hz, 2H), 7.64 (t,  $J$  = 7.6 Hz, 1H), 7.49 (t,  $J$  = 7.6 Hz, 2H), 2.61 (s, 1H), 2.05-1.93 (m, 1H), 1.90-1.80 (m, 1H), 1.76 (s, 3H), 1.60-1.47 (m, 1H), 1.44-1.31 (m, 2H), 0.92-0.85 (m, 6H);  $^{13}C$  NMR: (100 MHz,  $CDCl_3$ )  $\delta$  165.8, 154.5, 134.2, 129.9, 128.7, 126.7, 83.0, 77.9, 74.1, 39.4, 32.8, 27.9, 26.5, 22.5, 22.4; HRMS (ESI) calculated for  $[C_{17}H_{21}NNaO_4]^+$  ( $M+Na^+$ ) requires  $m/z$  326.1363, found:  $m/z$  326.1361.



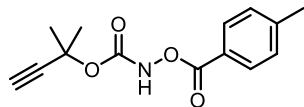
**3-Methyl-5-phenylpent-1-yn-3-yl (benzyloxy)carbamate (1k):**

7.16 g, 85% yield, white solid, m.p. = 102-103 °C, IR (neat): 3300, 2922, 1747, 1453, 1233  $cm^{-1}$ ;  $^1H$  NMR: (400 MHz,  $CDCl_3$ )  $\delta$  8.36 (brs, 1H), 8.15-8.04 (m, 2H), 7.62 (t,  $J$  = 7.6 Hz, 1H), 7.47 (t,  $J$  = 7.6 Hz, 2H), 7.26 (t,  $J$  = 7.6 Hz, 2H), 7.21-7.10 (m, 3H), 2.83 (t,  $J$  = 8.4 Hz, 2H), 2.68 (s, 1H), 2.35-2.22 (m, 1H), 2.20-2.07 (m, 1H), 1.82 (s, 3H);  $^{13}C$  NMR: (100 MHz,  $CDCl_3$ )  $\delta$  165.8, 154.5, 141.0, 134.2, 129.9, 128.7, 128.39, 128.37, 126.6, 126.0, 82.5, 77.3, 74.6, 43.4, 30.5, 26.6; HRMS (ESI) calculated for  $[C_{20}H_{19}NNaO_4]^+$  ( $M+Na^+$ ) requires  $m/z$  360.1206, found:  $m/z$  360.1204.



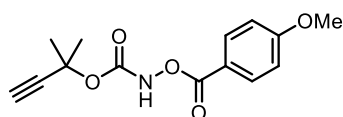
**3,7-Dimethyloct-6-en-1-yn-3-yl (benzyloxy)carbamate (1l):**

5.09 g, 78% yield, white solid, m.p. = 55-57 °C, IR (neat): 3295, 2924, 1747, 1452, 1234  $cm^{-1}$ ;  $^1H$  NMR: (400 MHz,  $CDCl_3$ )  $\delta$  8.31 (brs, 1H), 8.10 (d,  $J$  = 8.0 Hz, 2H), 7.64 (t,  $J$  = 7.6 Hz, 1H), 7.48 (t,  $J$  = 7.6 Hz, 2H), 5.09 (t,  $J$  = 6.8 Hz, 1H), 2.63 (s, 1H), 2.25-2.13 (m, 2H), 2.07-1.96 (m, 1H), 1.92-1.81 (m, 1H), 1.77 (s, 3H), 1.68 (s, 3H), 1.59 (s, 3H);  $^{13}C$  NMR: (100 MHz,  $CDCl_3$ )  $\delta$  165.8, 154.5, 134.2, 132.5, 129.9, 128.7, 126.7, 122.8, 82.7, 77.6, 74.3, 41.4, 26.5, 25.6, 22.8, 17.6; HRMS (ESI) calculated for  $[C_{18}H_{21}NNaO_4]^+$  ( $M+Na^+$ ) requires  $m/z$  338.1363, found:  $m/z$  338.1366.



**2-Methylbut-3-yn-2-yl ((4-methylbenzoyl)oxy)carbamate (1m):**

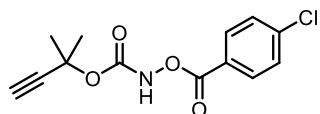
0.47 g, 60% yield, white solid, m.p. = 74-76 °C, IR (neat): 3287, 1741, 1611, 1465, 1237  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.31 (brs, 1H), 7.99 (d,  $J = 7.6$  Hz, 2H), 7.27 (d,  $J = 7.6$  Hz, 2H), 2.60 (s, 1H), 2.43 (s, 3H), 1.75 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.9, 154.5, 145.2, 130.0, 129.4, 123.8, 83.7, 74.4, 73.2, 28.9, 21.8; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{15}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  284.0893, found:  $m/z$  284.0895.



**2-Methylbut-3-yn-2-yl ((4-methoxybenzoyl)oxy)carbamate**

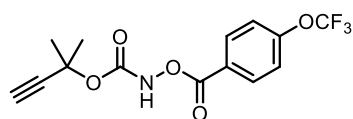
**(1n):** 0.48 g, 58% yield, colorless oil, IR (neat): 3275, 1739, 1606,

1511, 1463, 1242  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.32 (brs, 1H), 8.05 (d,  $J = 8.8$  Hz, 2H), 6.95 (d,  $J = 8.8$  Hz, 2H), 3.88 (s, 3H), 2.60 (s, 1H), 1.75 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.5, 164.3, 154.6, 132.1, 118.8, 114.0, 83.8, 74.4, 73.1, 55.5, 28.9; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{15}\text{NNaO}_5]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  300.0842, found:  $m/z$  300.0843.



**2-Methylbut-3-yn-2-yl ((4-chlorobenzoyl)oxy)carbamate (1o):**

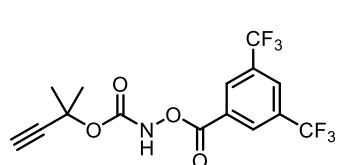
0.76 g, 83% yield, white solid, m.p. = 76-77 °C, IR (neat): 3300, 1745, 1596, 1483, 1239  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.30 (brs, 1H), 8.04 (d,  $J = 7.2$  Hz, 2H), 7.47 (d,  $J = 7.2$  Hz, 2H), 2.61 (s, 1H), 1.75 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.0, 154.4, 140.9, 131.3, 129.1, 125.1, 83.6, 74.7, 73.3, 28.9; HRMS (ESI) calculated for  $[\text{C}_{13}\text{H}_{12}\text{ClNNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  304.0347, found:  $m/z$  304.0348.



**2-Methylbut-3-yn-2-yl ((4-**

**(trifluoromethoxy)benzoyl)oxy)carbamate (1p):** 0.86 g, 89%

yield, white solid, m.p. = 76-78 °C, IR (neat): 3299, 1749, 1607, 1471, 1241  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.30 (brs, 1H), 8.16 (d,  $J = 8.0$  Hz, 2H), 7.31 (d,  $J = 8.0$  Hz, 2H), 2.61 (s, 1H), 1.75 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  164.7, 154.4, 153.5, 132.1, 124.9, 120.4, 120.2 (q,  $J = 259.5$  Hz), 83.6, 74.7, 73.3, 28.9;  $^{19}\text{F}$  NMR: (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -57.6; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{12}\text{F}_3\text{NNaO}_5]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  356.0560, found:  $m/z$  356.0561.

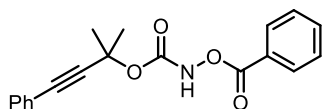


**2-Methylbut-3-yn-2-yl**

**((3,5-**

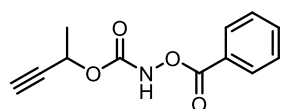
**bis(trifluoromethyl)benzoyloxy)carbamate (1q):** 0.63 g, 82%

yield, white solid, m.p. = 75-77 °C, IR (neat): 3274, 1752, 1382, 1282, 1222  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.55 (s, 2H), 8.40 (brs, 1H), 8.15 (s, 1H), 2.63 (s, 1H), 1.77 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  163.4, 154.1, 132.3 (q,  $J = 34.2$  Hz), 130.1 (q,  $J = 3.0$  Hz), 129.0, 127.5 (q,  $J = 3.7$  Hz), 122.6 (q,  $J = 271.2$  Hz), 83.4, 75.1, 73.5, 28.8;  $^{19}\text{F}$  NMR: (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -63.0; HRMS (ESI) calculated for  $[\text{C}_{15}\text{H}_{11}\text{F}_6\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  406.0484, found:  $m/z$  406.0482.



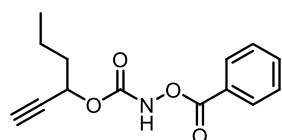
**2-Methyl-4-phenylbut-3-yn-2-yl (benzoyloxy)carbamate (1r):**

1.55 g, 48% yield, white solid, m.p. = 111-114 °C, IR (neat): 2921, 2855, 1748, 1456, 1235  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.36 (brs, 1H), 8.14-8.08 (m, 2H), 7.64 (t,  $J = 7.2$  Hz, 1H), 7.52-7.42 (m, 4H), 7.33-7.25 (m, 3H), 1.84 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.9, 154.5, 134.2, 131.9, 129.9, 128.7, 128.5, 128.1, 126.7, 122.3, 89.1, 84.8, 75.5, 29.1; HRMS (ESI) calculated for  $[\text{C}_{19}\text{H}_{17}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  346.1050, found:  $m/z$  346.1051.



**But-3-yn-2-yl (benzoyloxy)carbamate (1s):** 6.00 g, 74% yield, white

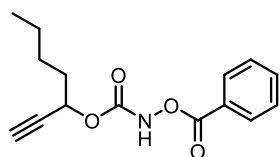
solid, m.p. = 82-84 °C, IR (neat): 3275, 2925, 1743, 1454, 1232  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.50 (brs, 1H), 8.13-8.06 (m, 2H), 7.68-7.60 (m, 1H), 7.52-7.44 (m, 2H), 5.53-5.45 (m, 1H), 2.54 (d,  $J = 2.0$  Hz, 1H), 1.57 (d,  $J = 6.8$  Hz, 3H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.6, 155.3, 134.3, 129.9, 128.7, 126.4, 81.0, 74.0, 62.9, 21.2; HRMS (ESI) calculated for  $[\text{C}_{12}\text{H}_{11}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  256.0580, found:  $m/z$  256.0581.



**Hex-1-yn-3-yl (benzoyloxy)carbamate (1t):** 1.62 g, 69% yield, white

solid, m.p. = 63-66 °C, IR (neat): 3292, 2963, 1743, 1456, 1232  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.41 (brs, 1H), 8.10 (d,  $J = 7.6$  Hz, 2H), 7.64 (t,  $J = 7.2$  Hz, 1H), 7.49 (t,  $J = 7.2$  Hz, 2H), 5.41 (t,  $J = 6.4$  Hz, 1H), 2.52 (s, 1H), 1.90-1.75 (m, 2H), 1.66-1.42 (m, 2H), 0.95 (t,  $J = 7.2$  Hz, 3H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.7, 155.5, 134.3, 130.0, 128.7, 126.5, 80.2, 74.6, 66.5, 36.6, 18.0, 13.5; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{15}\text{NNaO}_4]^+$

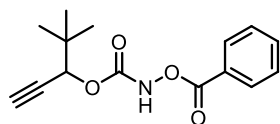
(M+Na<sup>+</sup>) requires m/z 284.0893, found: m/z 284.0892.



**Hept-1-yn-3-yl (benzyloxy)carbamate (1u):** 18.29 g, 84% yield,

white soild, m.p. = 49-52 °C, IR (neat): 3291, 2926, 1741, 1456, 1230 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.52 (brs, 1H), 8.13-8.06 (m, 2H),

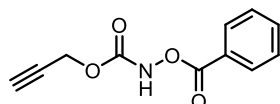
7.52-7.44 (m, 1H), 7.52-7.45 (m, 2H), 5.40 (td, *J* = 6.8, 2.0 Hz, 1H), 2.54 (d, *J* = 2.0 Hz, 1H), 1.91-1.76 (m, 2H), 1.50-1.28 (m, 4H), 0.90 (t, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 165.7, 155.5, 134.3, 129.9, 128.7, 126.5, 80.2, 74.6, 66.6, 34.3, 26.7, 22.1, 13.8; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 298.1050, found: m/z 298.1052.



**4,4-Dimethylpent-1-yn-3-yl (benzyloxy)carbamate (1v):** 4.23 g, 77%

yield, white soild, m.p. = 50-53 °C, IR (neat): 3296, 2965, 1741, 1456, 1370, 1231 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.59 (brs, 1H), 8.12-

8.07 (m, 2H), 7.64 (t, *J* = 7.6 Hz, 1H), 7.48 (t, *J* = 7.6 Hz, 2H), 5.14 (d, *J* = 2.4 Hz, 1H), 2.51 (d, *J* = 2.0 Hz, 1H), 1.03 (s, 9H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 165.7, 155.8, 134.2, 129.9, 128.7, 126.5, 78.9, 75.2, 74.3, 35.2, 25.3; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 298.1050, found: m/z 298.1052.



**Prop-2-yn-1-yl (benzyloxy)carbamate (1w):** 3.59 g, 82% yield, white

soild, m.p. = 63-65 °C, IR (neat): 3294, 2920, 1744, 1453, 1233 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.55 (br, 1H), 8.12-8.05 (m, 2H), 7.68-7.61

(m, 1H), 7.53-7.44 (m, 2H), 4.82 (d, *J* = 2.4 Hz, 2H), 2.55 (t, *J* = 2.4 Hz, 1H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 165.5, 155.5, 134.3, 129.9, 128.7, 126.3, 75.9, 54.0; HRMS (ESI) calculated for [C<sub>11</sub>H<sub>9</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 242.0424, found: m/z 242.0423.

## IV. Visible-Light-Promoted Intramolecular Oxyamidation of Alkynes

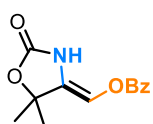
Materials used for set-up: 10 mL Schlenk flask. IKA RCT digital. Blue LED lamp (450 nm, 5

W/m, 1 m). Water bath. The LED lamp was set aside the Schlenk flask.

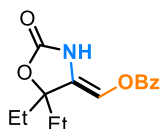
General procedure for visible-light-promoted alkyne carboamination: a 10 mL flame-dried Schlenk flask was cooled at room temperature under argon, charged with **1** (0.2 mmol), NR (0.004 mmol), DIPEA (0.02 mmol) and MeCN (2 mL). The mixture was degassed for 3 times, and then warmed to room temperature under argon, stirred at 40 °C under 5 W blue LEDs in the parallel batch photoreactor for 12 hours (**Fig. S3**). The reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel to afford the corresponding product **2**.



**Fig. S3.** Photograph of the parallel batch photoreactor.

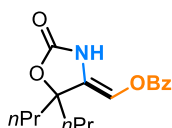


**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)methyl benzoate (2a):** prepared according to the general procedure, using 0.0496 g of benzoyloxycarbamate **1a** (0.20 mmol), 0.0014g of NR (0.002 mmol), 0.0026 g of DIPEA (0.02 mmol, 3.4 uL) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0366 g, 74% yield, white solid; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.99 (brs, 1H), 8.21 (d, *J* = 8.0 Hz, 2H), 7.62 (t, *J* = 7.6 Hz, 1H), 7.50 (t, *J* = 7.6 Hz, 2H), 6.95 (s, 1H), 1.64 (s, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.8, 156.3, 133.7, 130.6, 130.2, 128.7, 128.4, 111.2, 82.9, 28.3; The characterization data of the product is consistent with the reported literature.<sup>1</sup>



**(Z)-(5,5-diethyl-2-oxooxazolidin-4-ylidene)methyl benzoate (2b):** prepared according to the general procedure, using 0.0554 g of benzoyloxycarbamate **1b** (0.20 mmol), 0.0012 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and

2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0428 g, 78% yield, colorless oil; IR (neat): 2977, 1763, 1717, 1373, 1251 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 9.27 (brs, 1H), 8.24 (d, *J* = 7.6 Hz, 2H), 7.61 (t, *J* = 7.6 Hz, 1H), 7.51 (t, *J* = 7.6 Hz, 2H), 6.90 (s, 1H), 2.01-1.89 (m, 2H), 1.78-1.67 (m, 2H), 1.01 (t, *J* = 7.2 Hz, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.9, 157.1, 133.6, 130.2, 128.6, 128.5, 127.3, 111.5, 88.7, 32.6, 7.2; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 298.1050, found: *m/z* 298.1051.



**(Z)-(2-oxo-5,5-dipropoxyloxazolidin-4-ylidene)methyl benzoate (2c):** prepared according to the general procedure, using 0.0611 g of benzoyloxycarbamate **1c** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and

2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0468 g, 77% yield, white solid, m.p. = 174-177 °C; IR (neat): 2961, 1762, 1718, 1454, 1251 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.70 (brs, 1H), 8.20 (d, *J* = 8.0, 2H), 7.63 (t, *J* = 7.6 Hz, 1H), 7.51 (t, *J* = 7.6 Hz, 2H), 6.89 (s, 1H), 1.94-1.82 (m, 2H), 1.71-1.59 (m, 2H), 1.56-1.38 (m, 4H), 0.95 (t, *J* = 7.2 Hz, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.9, 156.7, 133.7, 130.1, 128.6, 128.5, 128.0, 111.5, 88.1, 42.2, 16.1, 13.9; HRMS (ESI) calculated for [C<sub>17</sub>H<sub>21</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 326.1363, found: *m/z* 326.1363.

**(CCDC: 2329040)**

Bond precision:	C-C = 0.0019 Å	Wavelength=0.71073	
Cell:	<i>a</i> / Å =12.2627(5)	<i>b</i> / Å =8.4254(4)	<i>c</i> / Å =16.0986(6)
	alpha/°=90	beta/°=101.590(1)	gamma/°=90

Temperature: 170 K

	Calculated	Reported
Volume	1629.36(12)	1629.36(12)
Space group	P 21/c	P 1 21/c 1
Hall group	-P 2ybc	-P 2ybc
Moiety formula	C <sub>17</sub> H <sub>21</sub> NO <sub>4</sub>	C <sub>17</sub> H <sub>21</sub> NO <sub>4</sub>
Sum formula	C <sub>17</sub> H <sub>21</sub> NO <sub>4</sub>	C <sub>17</sub> H <sub>21</sub> NO <sub>4</sub>
Mr	303.35	303.35
Dx,g cm <sup>-3</sup>	1.237	1.237
Z	4	4
Mu (mm <sup>-1</sup> )	0.088	0.088
F000	648.0	648.0
F000'	648.33	
h,k,lmax	16,11,21	16,11,21
Nref	4073	4056
Tmin,Tmax	0.967,0.983	0.680,0.746
Tmin'	0.958	

Correction method= # Reported T Limits: Tmin=0.680 Tmax=0.746

AbsCorr = MULTI-SCAN

Data completeness= 0.996

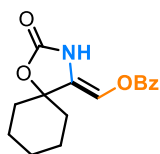
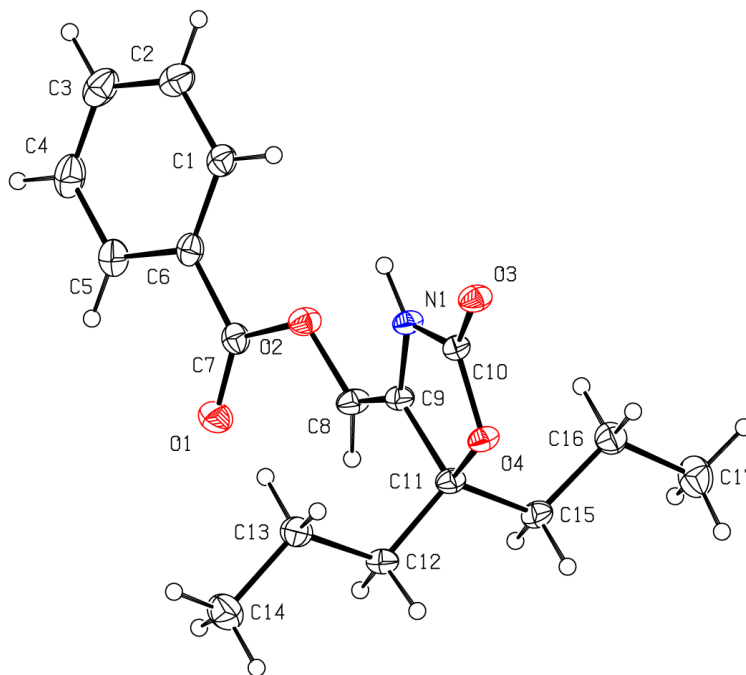
Theta(max)= 28.342

R(reflections)= 0.0456( 3342)

wR2(reflections)= 0.1046( 4056)

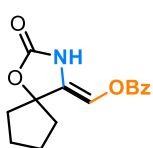
S = 1.092

Npar= 201



**(Z)-(2-oxo-1-oxa-3-azaspiro[4.5]decan-4-ylidene)methyl benzoate (2d):**

prepared according to the general procedure, using 0.0576 g of benzoyloxycarbamate **1d** (0.20 mmol), 0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0389 g, 68% yield, white solid, m.p. = 260-262 °C; IR (neat): 2935, 1754, 1710, 1274, 1142  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.04 (brs, 1H), 8.42 (d,  $J = 8.0$  Hz, 2H), 7.62 (t,  $J = 7.2$  Hz, 1H), 7.51 (t,  $J = 7.2$  Hz, 2H), 6.95 (s, 1H), 2.28 (d,  $J = 13.6$  Hz, 2H), 1.82-1.66 (m, 5H), 1.64-1.54 (m, 2H), 1.40-1.30 (m, 1H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.9, 156.4, 133.7, 130.7, 130.1, 128.6, 128.5, 111.5, 84.6, 37.3, 24.5, 21.6; HRMS (ESI) calculated for  $[\text{C}_{16}\text{H}_{17}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  310.1050, found:  $m/z$  310.1051.

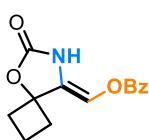


**(Z)-(2-oxo-1-oxa-3-azaspiro[4.4]nonan-4-ylidene)methyl benzoate (2e):**

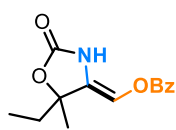
prepared according to the general procedure, using 0.0549 g of benzoyloxycarbamate **1e** (0.20 mmol), 0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary



evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0396 g, 72% yield, light yellow soild, m.p. = 184-186 °C; IR (neat): 3271, 2952, 1719, 1367, 1259, 1126  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.92 (brs, 1H), 8.20 (d,  $J = 7.2$  Hz, 2H), 7.62 (t,  $J = 7.2$  Hz, 1H), 7.50 (t,  $J = 7.2$  Hz, 2H), 6.97 (s, 1H), 2.38-2.24 (m, 2H), 2.00-1.78 (m, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.8, 156.6, 133.7, 130.2, 129.5, 128.6, 128.5, 111.2, 92.6, 41.3, 24.1; HRMS (ESI) calculated for  $[\text{C}_{15}\text{H}_{15}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  296.0893, found:  $m/z$  296.0895.

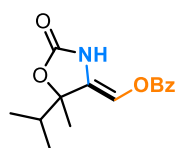


**(Z)-(6-oxo-5-oxa-7-azaspiro[3.4]octan-8-ylidene)methyl benzoate (2f):** repared according to the general procedure, using 0.0518 g of benzoyloxycarbamate **1f** (0.20 mmol), 0.0015 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0243 g, 47% yield, white soild, m.p. = 182-184 °C; IR (neat): 3271, 1763, 1720, 1327, 1256, 1133  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.53 (brs, 1H), 8.19 (d,  $J = 7.6$  Hz, 2H), 7.63 (t,  $J = 7.6$  Hz, 1H), 7.52 (t,  $J = 7.6$  Hz, 2H), 7.26 (s, 1H), 2.82-2.70 (m, 2H), 2.52-2.42 (m, 2H), 2.14-2.00 (m, 1H), 1.94-1.78 (m, 1H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.8, 156.3, 133.8, 130.4, 130.2, 128.7, 128.4, 111.6, 84.1, 36.8, 12.8; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{13}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  282.0737, found:  $m/z$  282.0735.



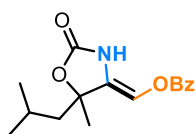
**(Z)-(5-ethyl-5-methyl-2-oxooxazolidin-4-ylidene)methyl benzoate (2g):** repared according to the general procedure, using 0.0525 g of benzoyloxycarbamate **1g** (0.20 mmol), 0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0395 g, 76% yield, white soild, m.p. = 176-178 °C; IR (neat): 3219, 2983, 1764, 1719, 1256, 1131  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.27 (brs, 1H), 8.23 (d,  $J = 7.6$  Hz, 2H), 7.61 (t,  $J = 7.6$  Hz, 1H), 7.50 (t,  $J = 7.6$  Hz, 2H), 6.92 (s, 1H), 1.98-1.87 (m, 1H), 1.86-1.73 (m, 1H), 1.60 (s, 3H), 1.02 (t,  $J = 7.2$  Hz, 3H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.9, 156.7, 133.7, 130.2, 129.2, 128.6, 128.5, 111.4, 85.7, 34.0, 26.8, 7.5; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{15}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  284.0893, found:

m/z 284.0892.



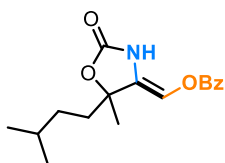
**(Z)-(5-isopropyl-5-methyl-2-oxooxazolidin-4-ylidene)methyl benzoate (2h):**

repared according to the general procedure, using 0.0555 g of benzoyloxycarbamate **1h** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0394 g, 72% yield, white soild, m.p. = 209-212 °C; IR (neat): 2924, 1761, 1717, 1375, 1258, 1128 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 9.15 (brs, 1H), 8.23 (d, *J* = 7.6 Hz, 2H), 7.62 (t, *J* = 7.2 Hz, 1H), 7.51 (t, *J* = 7.6 Hz, 2H), 6.92 (s, 1H), 2.01-1.88 (m, 1H), 1.59 (s, 3H), 1.09-1.02 (m, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.9, 156.7, 133.7, 130.2, 129.0, 128.6, 128.4, 111.6, 88.0, 37.5, 25.0, 16.4, 16.2; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 298.1050, found: m/z 298.1048.



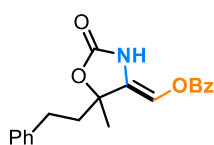
**(Z)-(5-isobutyl-5-methyl-2-oxooxazolidin-4-ylidene)methyl benzoate (2i):**

repared according to the general procedure, using 0.0580 g of benzoyloxycarbamate **1i** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0415 g, 72% yield, light yellow soild, m.p. = 159-162 °C; IR (neat): 2958, 1762, 1718, 1256, 1128 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 9.33 (brs, 1H), 8.24 (d, *J* = 8.0 Hz, 2H), 7.62 (t, *J* = 7.6 Hz, 1H), 7.50 (t, *J* = 7.6 Hz, 2H), 6.93 (s, 1H), 1.97-1.78 (m, 2H), 1.74-1.65 (m, 1H), 1.60 (s, 3H), 0.99 (d, *J* = 5.2 Hz, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.9, 156.7, 133.7, 130.2, 130.1, 128.6, 128.5, 111.5, 85.6, 49.3, 27.8, 24.3, 24.1, 24.0; HRMS (ESI) calculated for [C<sub>16</sub>H<sub>19</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 312.1206, found: m/z 312.1206.



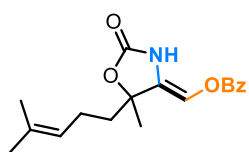
**(Z)-(5-isopentyl-5-methyl-2-oxooxazolidin-4-ylidene)methyl benzoate**

**(2j):** prepared according to the general procedure, using 0.0620 g of benzoyloxycarbamate **1j** (0.20 mmol), 0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0463 g, 76% yield, colorless oil; IR (neat): 3214, 2955, 1759, 1718, 1257, 1127 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 9.28 (brs, 1H), 8.24 (d, *J* = 7.2 Hz, 2H), 7.62 (t, *J* = 7.6 Hz, 1H), 7.51 (t, *J* = 7.6 Hz, 2H), 6.94 (s, 1H), 1.94-1.84 (m, 1H), 1.77-1.67 (m, 1H), 1.61 (s, 3H), 1.59-1.49 (m, 1H), 1.40-1.24 (m, 2H), 0.90 (d, *J* = 6.8 Hz, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.8, 156.8, 133.7, 130.2, 129.4, 128.6, 128.4, 111.4, 85.5, 39.0, 31.7, 27.9, 27.2, 22.5, 22.4; HRMS (ESI) calculated for [C<sub>17</sub>H<sub>21</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 326.1363, found: *m/z* 326.1363.



**(Z)-(5-methyl-2-oxo-5-phenethyloxazolidin-4-ylidene)methyl benzoate**

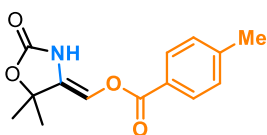
**(2k):** prepared according to the general procedure, using 0.0681 g of benzoyloxycarbamate **1k** (0.20 mmol), 0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0472 g, 70% yield, colorless oil; IR (neat): 2923, 1761, 1719, 1454, 1256 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.42 (brs, 1H), 8.18 (d, *J* = 7.6 Hz, 2H), 7.64 (t, *J* = 7.6 Hz, 1H), 7.52 (t, *J* = 7.6 Hz, 2H), 7.33-7.26 (m, 2H), 7.23-7.14 (m, 3H), 6.97 (s, 1H), 2.86-2.68 (m, 2H), 2.28-2.16 (m, 1H), 2.10-1.97 (m, 1H), 1.66 (s, 3H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 162.8, 156.5, 140.5, 133.8, 130.2, 129.1, 128.7, 128.5, 128.4, 128.3, 126.2, 111.6, 85.0, 43.0, 29.5, 27.3; HRMS (ESI) calculated for [C<sub>20</sub>H<sub>19</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 360.1206, found: *m/z* 360.1206.



**(Z)-(5-methyl-5-(4-methylpent-3-en-1-yl)-2-oxooxazolidin-4-**

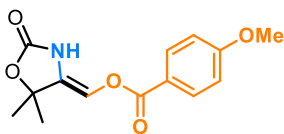
**ylidene)methyl benzoate (2l):** prepared according to the general procedure, using 0.0662 g of benzoyloxycarbamate **1l** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12

hours, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0249 g, 38% yield, colorless oil; IR (neat): 2980, 1725, 1454, 1378, 1269  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.99 (brs, 1H), 8.21 (d,  $J = 7.6$  Hz, 2H), 7.62 (t,  $J = 7.2$  Hz, 1H), 7.51 (t,  $J = 7.6$  Hz, 2H), 6.93 (s, 1H), 5.09 (t,  $J = 6.8$  Hz, 1H), 2.22-2.02 (m, 2H), 1.98-1.86 (m, 1H), 1.80-1.68 (m, 1H), 1.67 (s, 3H), 1.64-1.57 (m, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.8, 156.4, 133.7, 132.8, 130.1, 129.3, 128.6, 128.4, 122.4, 111.4, 85.2, 40.9, 27.3, 25.6, 21.9, 17.7; HRMS (ESI) calculated for  $[\text{C}_{18}\text{H}_{21}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  338.1363, found:  $m/z$  338.1364.



**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)methyl 4-methylbenzoate (2m):** repared according to the general procedure, using 0.0522 g of benzoyloxycarbamate **1m** (0.20 mmol), 0.0013 g of NR

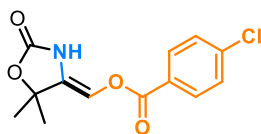
(0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40  $^\circ\text{C}$  for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0320 g, 61% yield, light yellow soild, m.p. = 277-280  $^\circ\text{C}$ ; IR (neat): 2991, 1760, 1714, 1417, 1267  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.99 (brs, 1H), 8.09 (d,  $J = 8.0$  Hz, 2H), 7.29 (d,  $J = 8.0$  Hz, 2H), 6.94 (s, 1H), 2.43 (s, 3H), 1.63 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.9, 156.4, 144.6, 130.3, 130.2, 129.4, 125.6, 111.2, 82.9, 28.3, 21.7; HRMS (ESI) calculated for  $[\text{C}_{14}\text{H}_{15}\text{NNaO}_4]^+$  ( $\text{M}+\text{Na}^+$ ) requires  $m/z$  284.0893, found:  $m/z$  284.0894.



**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)methyl 4-methoxybenzoate (2n):** repared according to the general procedure, using 0.0536 g of benzoyloxycarbamate **1n** (0.20 mmol), 0.0013 g of NR

(0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40  $^\circ\text{C}$  for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $\text{Et}_3\text{N}$ ), the reaction afforded 0.0277 g, 50% yield, light yellow soild, m.p. = 174-177  $^\circ\text{C}$ ; IR (neat): 2987, 1755, 1606, 1513, 1257  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR: (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.97 (brs, 1H), 8.16 (d,  $J = 8.8$  Hz, 2H), 6.97 (d,  $J = 8.8$  Hz, 2H), 6.93 (s, 1H), 3.89 (s, 3H), 1.63 (s, 6H);  $^{13}\text{C}$  NMR: (100 MHz,  $\text{CDCl}_3$ )  $\delta$  164.0, 162.6,

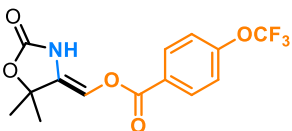
156.5, 132.3, 130.1, 120.7, 113.9, 111.3, 83.0, 55.5, 28.3; HRMS (ESI) calculated for  $[C_{14}H_{15}NNaO_4]^+$  ( $M+Na^+$ ) requires  $m/z$  300.0842, found:  $m/z$  300.0842.



**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)methyl 4-**

**chlorobenzoate (2o):** repared according to the general procedure, using 0.0564 g of benzoyloxycarbamate **1o** (0.20 mmol), 0.0013 g of NR (0.004

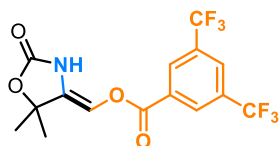
mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1H$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $Et_3N$ ), the reaction afforded 0.0184 g, 33% yield, white soild, m.p. = 225-228 °C; IR (neat): 3529, 2921, 1716, 1263, 1178, 1124  $cm^{-1}$ ;  $^1H$  NMR: (400 MHz,  $CDCl_3$ )  $\delta$  9.33 (brs, 1H), 8.18 (d,  $J$  = 8.0 Hz, 2H), 7.49 (d,  $J$  = 8.0 Hz, 2H), 6.95 (s, 1H), 1.64 (s, 6H);  $^{13}C$  NMR: (100 MHz,  $CDCl_3$ )  $\delta$  162.0, 156.5, 140.3, 131.6, 130.8, 129.0, 126.8, 111.1, 83.1, 28.3; HRMS (ESI) calculated for  $[C_{13}H_{12}ClNNaO_4]^+$  ( $M+Na^+$ ) requires  $m/z$  304.0347, found:  $m/z$  304.0345.



**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)methyl 4-**

**(trifluoromethoxy)benzoate (2p):** repared according to the general procedure, using 0.0665 g of benzoyloxycarbamate **1p** (0.20 mmol),

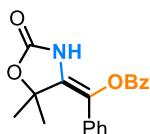
0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by  $^1H$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1%  $Et_3N$ ), the reaction afforded 0.0322 g, 49% yield, white soild, m.p. = 218-220 °C; IR (neat): 2926, 1755, 1712, 1271, 1174  $cm^{-1}$ ;  $^1H$  NMR: (400 MHz,  $CDCl_3$ )  $\delta$  9.46 (brs, 1H), 8.31 (d,  $J$  = 7.6 Hz, 2H), 7.34 (d,  $J$  = 7.6 Hz, 2H), 6.96 (s, 1H), 1.65 (s, 6H);  $^{13}C$  NMR: (100 MHz,  $CDCl_3$ )  $\delta$  161.7, 156.6, 153.2, 132.3, 130.9, 126.8, 120.4, 120.3, (q,  $J$  = 258.1 Hz), 111.2, 83.1, 28.3;  $^{19}F$  NMR: (376 MHz,  $CDCl_3$ )  $\delta$  -57.5; HRMS (ESI) calculated for  $[C_{14}H_{12}F_3NNaO_5]^+$  ( $M+Na^+$ ) requires  $m/z$  354.0560, found:  $m/z$  354.0561.



**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)methyl 3,5-**

**bis(trifluoromethyl)benzoate (2q):** prepared according to the general procedure, using 0.0769 g of benzoyloxycarbamate **1q** (0.20 mmol),

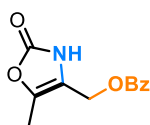
0.0012 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0465 g, 61% yield, white solid, m.p. = 256-258 °C; IR (neat): 2957, 1723, 1282, 1244, 1139 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 10.30 (brs, 1H), 8.75 (s, 2H), 8.12 (s, 1H), 6.99 (s, 1H), 1.66 (s, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 160.5, 157.2, 132.4 (q, *J* = 33.4 Hz), 132.3, 130.8, 130.5, 127.1-126.9 (m), 122.8 (q, *J* = 271.6 Hz), 111.0, 83.1, 28.1; <sup>19</sup>F NMR: (376 MHz, CDCl<sub>3</sub>) δ -62.8; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>11</sub>F<sub>6</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 406.0484, found: m/z 406.0483.



**(Z)-(5,5-dimethyl-2-oxooxazolidin-4-ylidene)(phenyl)methyl benzoate (2r):**

prepared according to the general procedure, using 0.0649 g of benzoyloxycarbamate **1r** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and

2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0431 g, 67% yield, white solid, m.p. = 210-213 °C; IR (neat): 3352, 2981, 1747, 1390, 1277 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.11 (d, *J* = 8.0 Hz, 2H), 7.65 (t, *J* = 6.8 Hz, 2H), 7.51 (t, *J* = 7.6 Hz, 2H), 7.47-7.35 (m, 4H), 7.32-7.25 (m, 1H), 1.69 (s, 6H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 165.2, 155.5, 134.0, 133.6, 132.6, 130.0, 129.1, 128.8, 128.7, 128.5, 126.1, 124.0, 84.9, 25.7; HRMS (ESI) calculated for [C<sub>19</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires m/z 346.1050, found: m/z 346.1051.

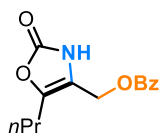


**(5-Methyl-2-oxo-2,3-dihydrooxazol-4-yl)methyl benzoate (2s):** prepared

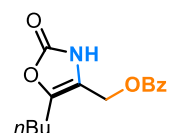
according to the general procedure, using 0.0466 g of benzoyloxycarbamate **1s** (0.20 mmol), 0.0012 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL

of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA

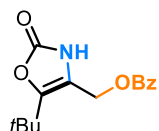
= 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0159 g, 34% yield, yellow solid, m.p. = 96-99 °C; IR (neat): 3266, 2960, 1723, 1452, 1271 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.47 (br, 1H), 8.03 (d, *J* = 7.6 Hz, 2H), 7.59 (t, *J* = 7.6 Hz, 1H), 7.45 (t, *J* = 7.6 Hz, 2H), 5.03 (s, 2H), 2.18 (s, 3H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 167.0, 155.7, 137.4, 133.5, 129.8, 129.1, 128.5, 116.2, 55.3, 10.1; HRMS (ESI) calculated for [C<sub>12</sub>H<sub>11</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 256.0580, found: *m/z* 256.0578.



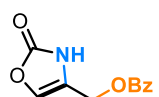
**(2-Oxo-5-propyl-2,3-dihydrooxazol-4-yl)methyl benzoate (2t):** prepared according to the general procedure, using 0.0526 g of benzoyloxycarbamate **1t** (0.20 mmol), 0.0012 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0192 g, 37% yield, colorless oil; IR (neat): 3352, 2972, 1722, 1452, 1382, 1270 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.55 (brs, 1H), 8.03 (d, *J* = 7.2 Hz, 2H), 7.59 (t, *J* = 7.2 Hz, 1H), 7.45 (t, *J* = 7.6 Hz, 2H), 5.04 (t, 2H), 2.48 (t, *J* = 7.2 Hz, 2H), 1.70-1.58 (m, 2H), 0.96 (t, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 167.1, 155.6, 141.1, 133.6, 129.8, 129.0, 128.5, 116.3, 55.2, 26.3, 20.7, 13.4; HRMS (ESI) calculated for [C<sub>14</sub>H<sub>15</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 284.0893, found: *m/z* 284.0894.



**(5-Butyl-2-oxo-2,3-dihydrooxazol-4-yl)methyl benzoate (2u):** prepared according to the general procedure, using 0.0549 g of benzoyloxycarbamate **1u** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0170 g, 31% yield, colorless oil; IR (neat): 3355, 2974, 1722, 1382, 1272 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.73 (brs, 1H), 8.04 (d, *J* = 8.0 Hz, 2H), 7.59 (t, *J* = 7.6 Hz, 1H), 7.45 (t, *J* = 7.6 Hz, 2H), 5.04 (s, 2H), 2.50 (t, *J* = 7.2 Hz, 2H), 1.68-1.54 (m, 2H), 1.44-1.22 (m, 2H), 0.92 (t, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 167.0, 155.8, 141.2, 133.6, 129.8, 129.1, 128.5, 116.0, 55.2, 29.4, 24.1, 21.9, 13.6; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M+Na<sup>+</sup>) requires *m/z* 298.1050, found: *m/z* 298.1050.



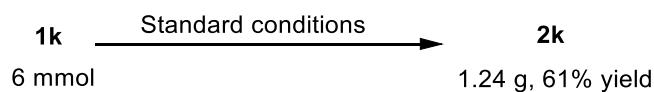
**(5-(*Tert*-butyl)-2-oxo-2,3-dihydrooxazol-4-yl)methyl benzoate (2v):** prepared according to the general procedure, using 0.0551 g of benzoyloxycarbamate **1v** (0.20 mmol), 0.0013 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0276 g, 50% yield, colorless oil; IR (neat): 3283, 2977, 1722, 1453, 1270, 1113 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.68 (brs, 1H), 8.05 (d, *J* = 7.6 Hz, 2H), 7.59 (t, *J* = 7.2 Hz, 1H), 7.45 (t, *J* = 7.2 Hz, 2H), 5.18 (s, 2H), 1.34 (s, 9H); <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>) δ 166.7, 155.5, 147.3, 133.5, 129.8, 129.2, 128.5, 114.3, 56.1, 32.9, 28.8; HRMS (ESI) calculated for [C<sub>15</sub>H<sub>17</sub>NNaO<sub>4</sub>]<sup>+</sup> (M<sup>+</sup>Na<sup>+</sup>) requires *m/z* 298.1050, found: *m/z* 298.1051.



**(2-Oxo-2,3-dihydrooxazol-4-yl)methyl benzoate (2w):** prepared according to the general procedure, using 0.0443 g of benzoyloxycarbamate **1w** (0.20 mmol), 0.0014 g of NR (0.004 mmol), 0.0026 g of DIPEA (0.02 mmol) and 2 mL of MeCN. After reaction at 40 °C for 12 hours, the reaction was concentrated by rotary evaporation and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1-2/1, 1% Et<sub>3</sub>N), the reaction afforded 0.0101 g, 23% yield, IR (neat): 2923, 2853, 1727, 1457, 1375, 1273 cm<sup>-1</sup>; <sup>1</sup>H NMR: (400 MHz, CDCl<sub>3</sub>) δ 8.37 (brs, 1H), 8.04 (d, *J* = 7.2 Hz, 2H), 7.60 (t, *J* = 6.8 Hz, 1H), 7.46 (t, *J* = 6.8 Hz, 2H), 6.89 (s, 1H), 5.05 (s, 2H). HRMS (ESI) calculated for [C<sub>11</sub>H<sub>10</sub>NO<sub>4</sub>]<sup>+</sup> (M+H<sup>+</sup>) requires *m/z* 220.0604, found: *m/z* 220.0605. The characterization data of the product is consistent with the reported literature.<sup>2</sup>

## V. Synthetic Applications and Mechanistic Studies

### a) Gram-scale reaction and synthetic applications:

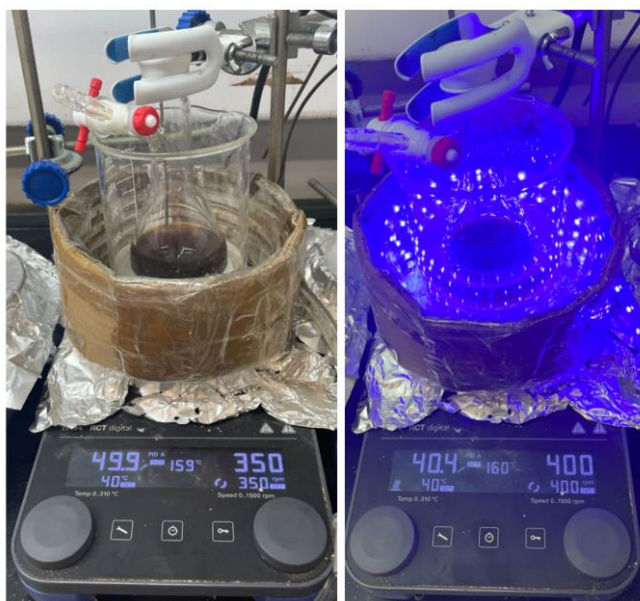


Materials used for set-up: 100 mL Schlenk flask. IKA RCT digital. Blue LED lamp strip (454 nm, 5 W/m, 1 m). Water bath (95 mm). The LED lamp strap was wrapped around the outside of the



oil bath.

A 100 mL flame-dried Schlenk flask was cooled at room temperature under argon, then charged with **1k** (2.0244 g, 6.0 mmol), NR (0.0346 g, 0.12 mmol), DIPEA (0.0776 g, 0.6 mmol) and MeCN (60 mL). The mixture was degassed for three times, warmed to room temperature under argon, and stirred at 40 °C while being irradiated with 5 W blue LEDs for 12 hours. The reaction mixture was concentrated by rotary evaporation, and monitored by <sup>1</sup>H NMR, and then further purified by flash chromatography on silica gel (PE/EA = 10/1-5/1, 1% Et<sub>3</sub>N) to afford 1.24 g **2k** in 61% yield.



**Fig. S4.** Photograph of the scale-up reaction.

The E-factor takes into account waste byproducts, leftover reactants, solvent losses, spent catalysts and catalyst supports, and anything else that can be regarded as a waste.<sup>3</sup> Since the solvent MeCN can be recovered, it is not regarded as the waste. The  $m_{product}$  is the mass of the product purified by flash chromatography. The  $m_{byproduct}$  is calculated as the difference between the mass of the product in 100% theoretical yield and the mass of the purified product. The  $m_{NR}$  and  $m_{DIPEA}$  are the mass of the NR and DIPEA used in the reaction, respectively. Herein, the E-factor of the **Gram-scale reaction** is calculated by:<sup>3</sup>

$$\begin{aligned} E &= \frac{\text{Total mass of waste from process}}{\text{Total mass of product}} = \frac{m_{byproduct} + m_{NR} + m_{DIPEA}}{m_{product}} \\ &= \frac{0.7884 + 0.0346 + 0.0776 \text{ g}}{1.2368 \text{ g}} = 0.728 \end{aligned}$$

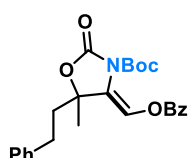
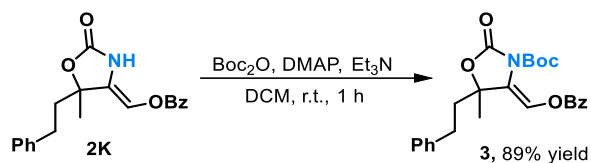
The atom utilization (AU) measures the conversion of atoms in the feedstock to the target product, regardless of factors such as yield (all assumed to be 100%) and selectivity, as well as the various types of catalysts and additives used in the synthesis process. Herein, the  $A_{\text{substrate}}$  and the  $A_{\text{product}}$  are the relative molecular masses of the substrate **1k** and product **2k**, respectively. The AU of the **Gram-scale reaction** is calculated by:<sup>3</sup>

$$\text{AU (\%)} = \frac{A_{\text{product}}}{A_{\text{substrate}}} \times 100 \% = \frac{337.4}{337.4} \times 100 \% = 100 \%$$

The reaction mass efficiency (RME) is also a measure of feedstock utilization, and refers to how much of the feedstock mass is converted into the product. Herein, the mass of the product is the mass of product **2k** purified by flash chromatography, and the total mass of reactants is the mass of substrate **1k**. The RME of the **Gram-scale reaction** is calculated by:<sup>3</sup>

$$\text{RME (\%)} = \frac{\text{Mass of product}}{\text{Total mass of reactants}} \times 100 \% = \frac{1.2368 \text{ g}}{2.0252 \text{ g}} \times 100 \% = 61 \%$$

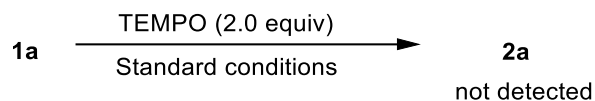
### Synthetic applications



**Tert-butyl (Z)-4-((benzoyloxy)methylene)-5-methyl-2-oxo-5-phenethyloxazolidine-3-carboxylate (3):** repared according to the general procedure,<sup>4</sup> a mixture of **2k** (0.169 g, 0.5 mmol),  $\text{Boc}_2\text{O}$  (0.160 g, 0.7 mmol),

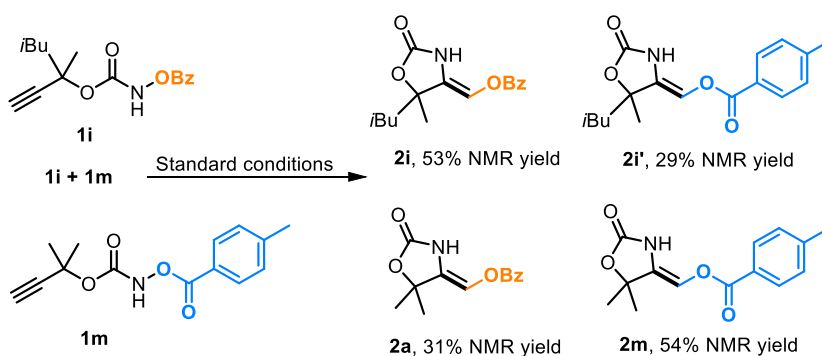
$\text{Et}_3\text{N}$  (0.076 g, 105  $\mu\text{L}$ , 0.75 mmol) and a catalytic amount of DMAP (0.0072 g, 0.05 mmol) in  $\text{CH}_2\text{Cl}_2$  (10 mL) was stirred at room temperature for 1 hour, the reaction was concentrated by rotary evaporation and monitored by  $^1\text{H}$  NMR, and then further purified by flash chromatography on silica gel (PE/EA = 5/1), the reaction afforded 0.196 g, 89% yield, colorless oil;  $^1\text{H}$  NMR: (600 MHz,  $\text{CDCl}_3$ )  $\delta$  8.12 (d,  $J = 7.8$  Hz, 2H), 7.61 (t,  $J = 7.2$  Hz, 1H), 7.45 (t,  $J = 7.8$  Hz, 2H), 7.32-7.26 (m, 2H), 7.21-7.16 (m, 3H), 7.08 (s, 1H), 2.80-2.70 (m, 2H), 2.25-2.15 (m, 1H), 2.11-2.02 (m, 1H), 1.66 (s, 3H), 1.57 (s, 9H);  $^{13}\text{C}$  NMR: (150 MHz,  $\text{CDCl}_3$ )  $\delta$  162.7, 150.7, 147.1, 140.4, 133.9, 130.2, 128.6, 128.5, 128.3, 128.3, 126.2, 124.4, 117.2, 85.2, 83.2, 42.9, 29.5, 27.8, 26.5; HRMS (ESI) calculated for  $[\text{C}_{25}\text{H}_{27}\text{NNaO}_6]^+$  ( $\text{M} + \text{Na}^+$ ) requires  $m/z$  460.1731, found:  $m/z$  460.1732.

### b) Radical-inhibition experiments:



According to the general procedure, a 10 mL flame-dried Schlenk flask was cooled at room temperature under argon, then charged with 0.0742 g (0.3 mmol) of **1a**, 0.0017 g (0.006 mmol) of NR, 0.0039 g (0.03 mmol) of DIPEA and 3 mL of MeCN, and 0.0937 g (0.6 mmol) of TEMPO. The mixture was degassed for 3 times, then warmed to room temperature under argon, stirred at 40 °C under blue LED for 12 hours. The mixture was concentrated by rotary evaporation, and no product **2a** was monitored by <sup>1</sup>H NMR.

### c) Cross-over reaction:



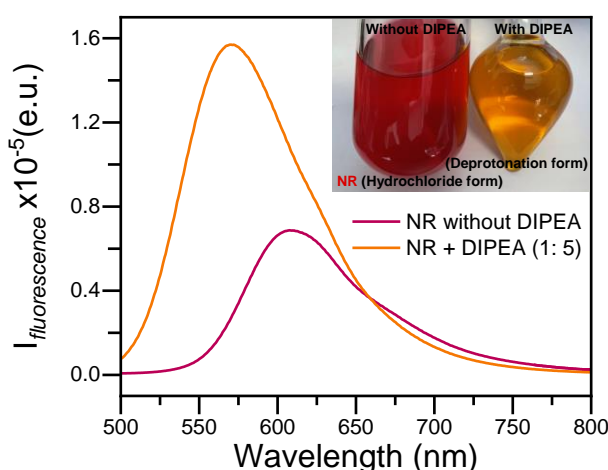
According to the general procedure, a 10 mL flame-dried Schlenk flask was cooled at room temperature under argon, then charged with a 1:1 N-acyloxycarbamates mixture of 0.0586 g **1i** (0.2 mmol) and 0.0526 g **1m** (0.2 mmol), 0.0024 g (0.008 mmol) of NR, 0.0052 g (0.04 mmol) of DIPEA and 4 mL of MeCN. The mixture was degassed for 3 times, then warmed to room temperature under argon, stirred at 40 °C under blue LED for 12 hours. The mixture was concentrated by rotary evaporation, and 4 cross-over products were observed, including **2i** (53%), **2i'** (29%), **2a** (31%), **2m** (54%) (NMR yield, using Trimethylphenylsilane as an internal standard).

### d) Stern-Volmer quenching experiments

#### Spectrum Data of NR:

**Experimental Details:** All fluorescence data was acquired on a Shimadzu RF6000. Stock solutions of  $5 \times 10^{-5}$  M NR, and  $5 \times 10^{-5}$  M 1:5 mixture of NR and DIPEA were prepared in HPLC-grade MeCN. After preparing these solutions in separate vials, the volume was transferred to a 1 cm cuvette, which was fitted with an appropriate septum, and sparged for 10 minutes prior to data acquisition. Absorption experiment samples were excited in the 300-700 nm wavelength range, then collected at 590 nm. Emission experiment samples were excited at 450 nm, then collected in the 460-700 nm wavelength range.

The commercially available hydrochloride form of NR is a good acid-base indicator, which appears red in acetonitrile solution. When DIPEA is added to the solution, the solution becomes alkaline and the color changes to yellow. The data unveiled that the pinnacle of absorption wavelength for NR stood at 478.4 and 559.9 nm, while the maximum emission wavelength reached 614 nm. In the case of NR+DIPEA (1:5), the maximum absorption wavelength settled at 444.2 nm, with the maximum emission wavelength at 570 nm. The results of fluorescence spectroscopy imply that the maximum absorption and emission wavelength of NR exhibit fluctuations contingent upon the acidity and alkalinity of the solution. Considering the alkaline solution of our reaction, it follows that the maximum absorption wavelength of NR should approximately hover around 444.2 nm, while the maximum emission wavelength should manifest at approximately 570 nm. The fluorescence intensity of NR is significantly enhanced with the addition of DIPEA (**Fig. S5**).



**Fig. S5.** Emission data of NR, NR+DIPEA (1:5). The color change of NR in acetonitrile at different pH values.

## Fluorescence Quenching Data

**Table S1.** Sample, contents:  $5 \times 10^{-5}$  M NR+DIPEA (1:5), indicated concentration **1b**

Entry	Concentration, mM	Intensity	Average	I <sub>0</sub> /I
blank1	0	164769		
blank2	0	156982	159579	1
blank3	0	156987		
S1-1	5	153194		
S1-2	5	149432	151851	1.05089
S1-3	5	152928		
S2-1	10	148452		
S2-2	10	150042	146994	1.08562
S2-3	10	142488		
S3-1	15	141843		
S3-2	15	142063	142307	1.12137
S3-3	15	143015		
S4-1	20	139484		
S4-2	20	140941	139229	1.14616
S4-3	20	137263		
S5-1	25	134419		
S5-2	25	136015	134272	1.18848
S5-3	25	132383		

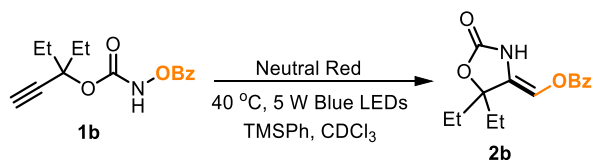
**Experimental Details:** All fluorescence data was acquired on a Shimadzu RF6000. Stock solutions of a 1:5 mixture of NR+DIPEA, and **1b** were prepared in HPLC-grade MeCN. Stock solutions were combined by volume to give 4.0 mL samples which each contained  $5 \times 10^{-5}$  M NR+DIPEA (1:5), as well as variable concentrations (5, 10, 15, 20, 25 mM) of **1b** indicated in the “concentration” column of the table. After preparing these solutions in separate vials, the volume was transferred to a 1 cm cuvette, which was fitted with an appropriate septum, and sparged for 10

minutes prior to data acquisition. Samples were excited at 450 nm, then collected in the 460-700 nm wavelength range. Intensities were recorded at 570 nm ( $\lambda_{\text{max}}$  of NR in alkaline solution), repeated three times, and then averaged. The intensity of the emission peak at 570 nm ( $\lambda_{\text{ex}} = 444$  nm) expressed as the ratio  $I_0/I$ , where  $I_0$  is the emission intensity of NR at 570 nm in the absence of a quencher and  $I$  is the observed intensity, as a function of the quencher concentration was measured. The Stern-Volmer emission spectra and plot of NR (0.05 mM) at different concentrations of **1b** are shown in **Scheme 3d**, the data are shown in **Table S1**.

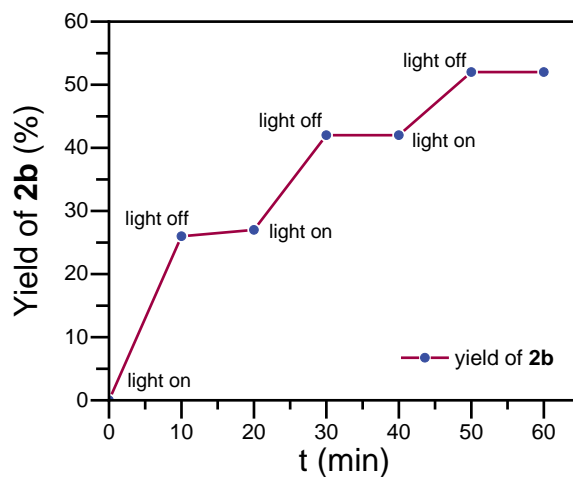
#### e) Light on and off experiments

A 25 mL flame-dried Schlenk flask was cooled at room temperature under nitrogen, charged with **1b** (0.2 mmol), NR (0.002 mmol), DIPEA (0.02 mmol) and  $\text{CDCl}_3$  (4 mL). The mixture was degassed for 3 times, and then warmed to room temperature under nitrogen, stirred at 40 °C under 5 W blue LEDs. The reaction yields were detected by NMR (**Table S2**, **Fig. S6**).

**Table S2.** Light on and off experiments.



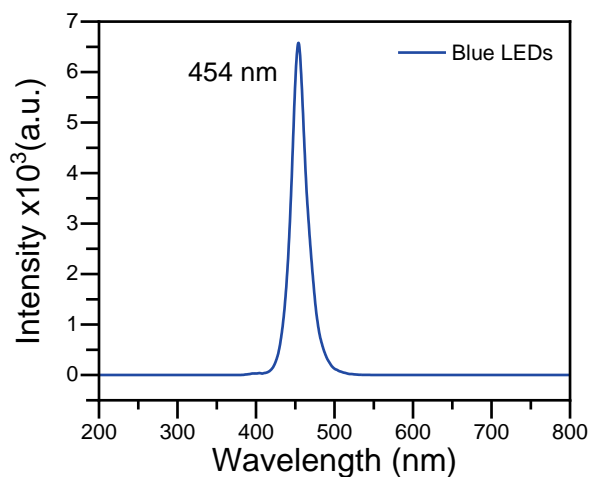
Entry	Time (min)	Light	<b>2b</b> (%)	Recovery of <b>1b</b> (%)
<b>1</b>	0	On	0	100
<b>2</b>	10	off	26	61
<b>3</b>	20	On	27	61
<b>4</b>	30	off	42	42
<b>5</b>	40	On	42	42
<b>6</b>	50	off	52	36
<b>7</b>	60	/	52	36



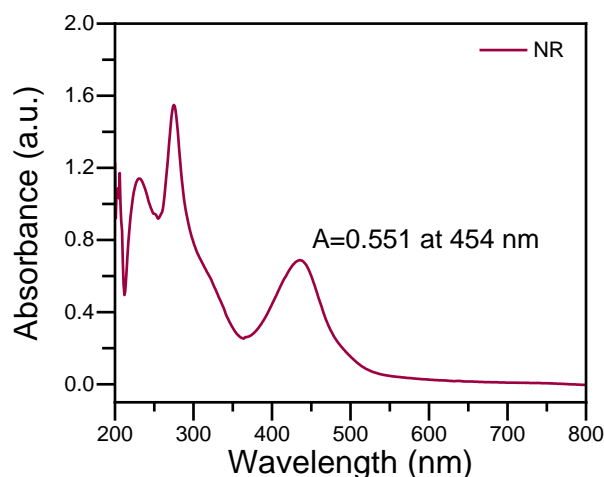
**Fig. S6.** Light on and off experiments.

#### f) Quantum yield experiments

A 25 mL flame-dried Schlenk flask was cooled at room temperature under nitrogen, charged with **1b** (0.2 mmol), NR (0.002 mmol), DIPEA (0.02 mmol) and  $\text{CDCl}_3$  (4 mL). The mixture was degassed for 3 times, and then warmed to room temperature under nitrogen, stirred at 40 °C under 5 W blue LEDs for 1200 s. The reaction yields were detected by NMR.



**Fig. S7.** The emission spectra of the light source (5 W blue LEDs,  $\lambda_{\text{max}} = 454$  nm).



**Fig. S8.** The absorbance spectra of NR ( $5 \times 10^{-5}$  mol/L) in the presence of DIPEA ( $2.5 \times 10^{-4}$  mol/L).

The quantum yield is calculated by:<sup>5</sup>

$$\Phi = \frac{\text{moles of product formed}}{\text{moles of light absorbed}} = \frac{\text{moles of product formed}}{\text{moles of photon} \cdot f}$$

The absorbance (A) of NR ( $c=5 \times 10^{-5}$  mol/L) at 454 nm is 0.551 (**Fig. S7**, **Fig. S8**), and the optical length of the cuvette (l) is 1 cm, so the molar absorption coefficient  $\epsilon$  of NR in 454 nm is calculated by:

$$\epsilon = \frac{A}{c \cdot l} = \frac{0.551}{5 \times 10^{-5} \times 1} = 11020 \text{ L} \cdot \text{mol}^{-1} \cdot \text{cm}^{-1}$$

The fraction of light absorbed (f) is calculated by:

The molar absorption coefficient  $\epsilon$  of NR in 454 nm is  $11020 \text{ L} \cdot \text{mol}^{-1} \cdot \text{cm}^{-1}$ . The diameter of the 25 mL Schlenk flask is 2 cm. The NR concentration is 0.0005 mol/L. So the absorbance  $A = \epsilon \cdot c \cdot l = 11020 \times 2 \times 0.0005 = 11.02$ , and the fraction of light absorbed  $f = 1 - 10^{-A} > 0.9999$ . The moles of photon is calculated by:

$$\text{moles of photon} = \frac{\text{total number of photons}}{NA} = \frac{\frac{E_{\text{total}}}{h\nu}}{NA} = \frac{\frac{P \cdot t}{h\nu}}{NA} = \frac{\frac{p \cdot S \cdot t}{\frac{hc}{\lambda}}}{NA}$$

The NA is Avogadro constant. The h is Planck constant. The c is lightspeed. The  $\lambda$  is light wavelength. The t is reaction time. The p is light intensity. The S is the illumination area. The diameter of the 25 mL Schlenk flask is 2 cm. So, when the reaction solution is 2 mL, the solution height is about  $2/\pi$  cm. And the illumination area S is about  $4 \text{ cm}^2$ .

Determination of the light intensity at the 454 nm LED systems. The light intensity was



detected by Sanwa laser power meter LP1. The instrument probe was placed in the center of the oil bath, and the results were as follows (**Table S3**).

**Table S3.** The light intensity at the 454 nm LED systems

Entry	Light intensity (mW/cm <sup>2</sup> )
1	2.412
2	2.342
3	2.425
4	2.372
5	2.217
6	2.435
<b>Average</b>	<b>2.367</b>

Experiment: 55.4 mg (0.2 mmol) **1b**, 1.4 mg (0.002 mmol) NR, 2.6 mg of DIPEA (0.02 mmol, 3.4  $\mu$ L), 4.0 mL (0.05 M) CDCl<sub>3</sub> after 1200 s yielded 42% of **2b**.  
 $\Phi(42\%) = 0.9743$ .

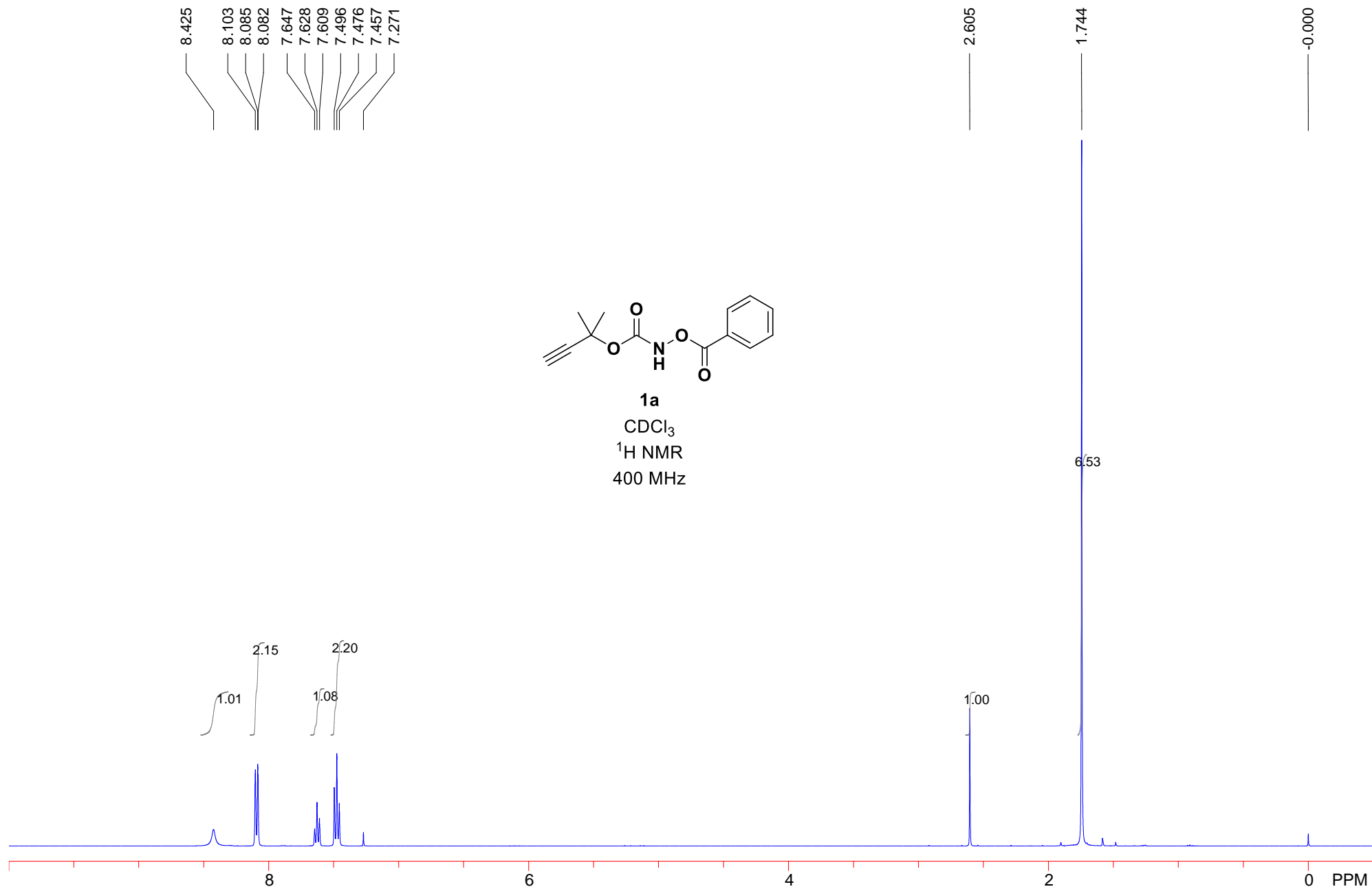
Sample quantum yield calculation:

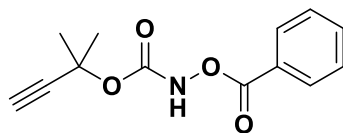
$$\Phi = \frac{8.4 \times 10^{-5} \text{ mol}}{\frac{2.367 \times 8 \times 10^{-3} \times 1200}{\frac{6.626 \times 10^{-34} \times 3 \times 10^8}{\frac{454 \times 10^{-9}}{6.02 \times 10^{23}}}} \times 1.00} = 0.9743$$

## **VII. References**

1. Q. Guo, D. Lu, Y. Mao and Z. Lu, *Chem. Commun.*, 2023, **59**, 1979-1982.
2. S. Su, T. Wu, Y. Xia, D. J. Wink and D. Lee, *Chem. - Eur. J.*, 2023, **29**, e202203371.
3. R. A. Sheldon, *ACS Sustainable Chem. Eng.*, 2017, **6**, 32-48.
4. Y. Tan, F. Han, M. Hemming, J. Wang, K. Harms, X. Xie, and E. Meggers, *Org. Lett.*, 2020, **22**, 16, 6653-6656.
5. M. A. Cismesia, T. P. Yoon, *Chem. Sci.*, 2015, **6**, 5426-5434.

## IX. NMR Spectra





**1a**

CDCl<sub>3</sub>  
13C NMR  
100 MHz

165.774

154.494

134.151

129.893

128.624

126.655

83.686

77.314

77.000

76.679

74.433

73.194

28.854

150

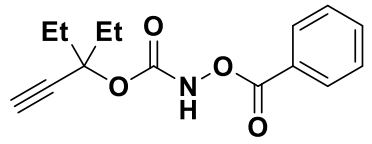
100

50

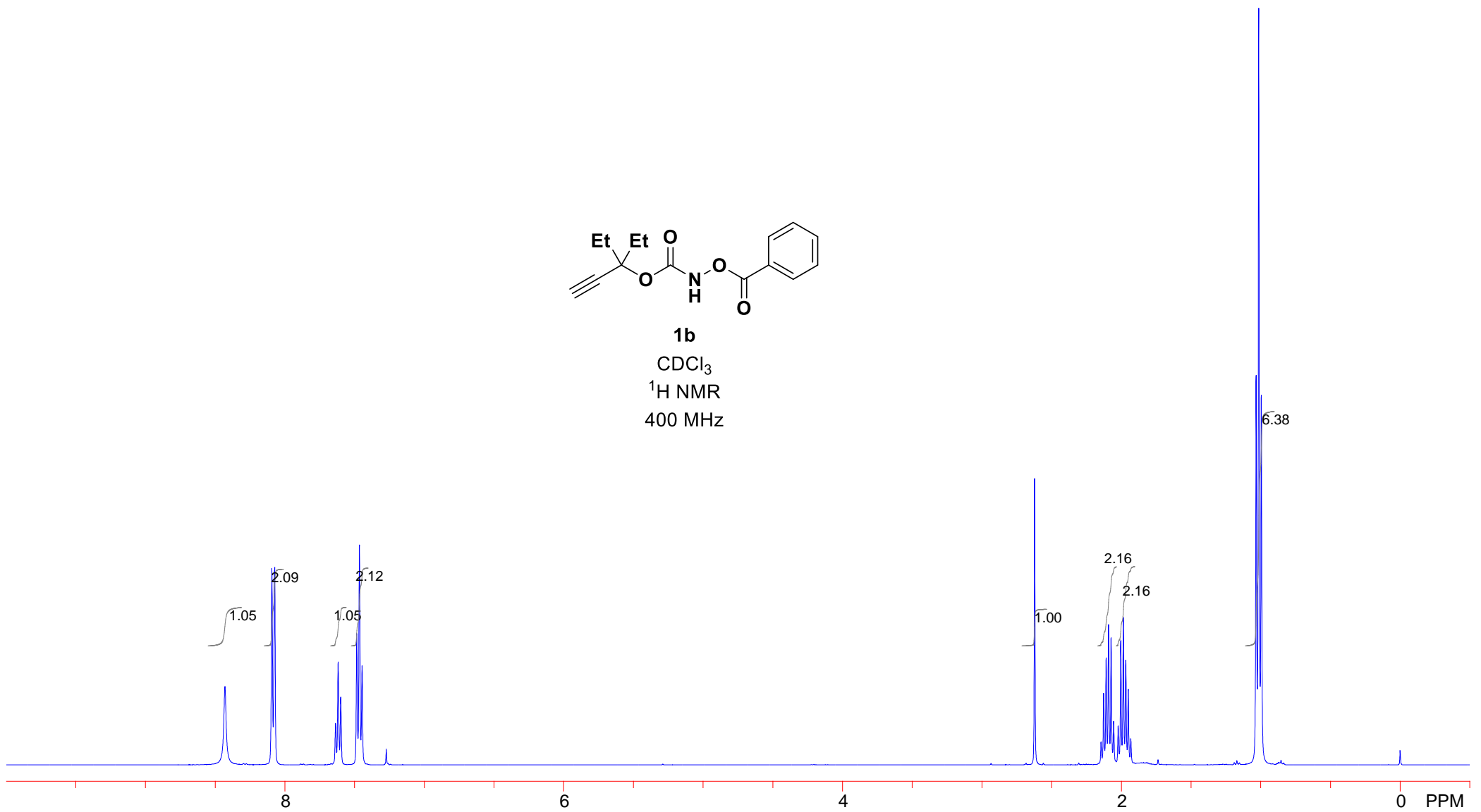
0 PPM

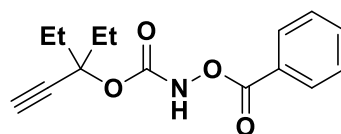
8.430  
8.093  
8.074  
8.073  
7.637  
7.618  
7.600  
7.485  
7.465  
7.447  
7.274

2.145  
2.622  
2.127  
2.109  
2.092  
2.073  
2.055  
2.023  
2.004  
1.986  
1.968  
1.950  
1.932  
1.033  
1.014  
0.995  
0.000



**1b**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





**1b**

CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

165.825

154.468

134.085

129.849

128.585

126.672

82.248

81.940

77.316

77.000

76.684

74.874

30.869

8.075

150

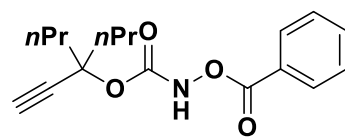
100

50

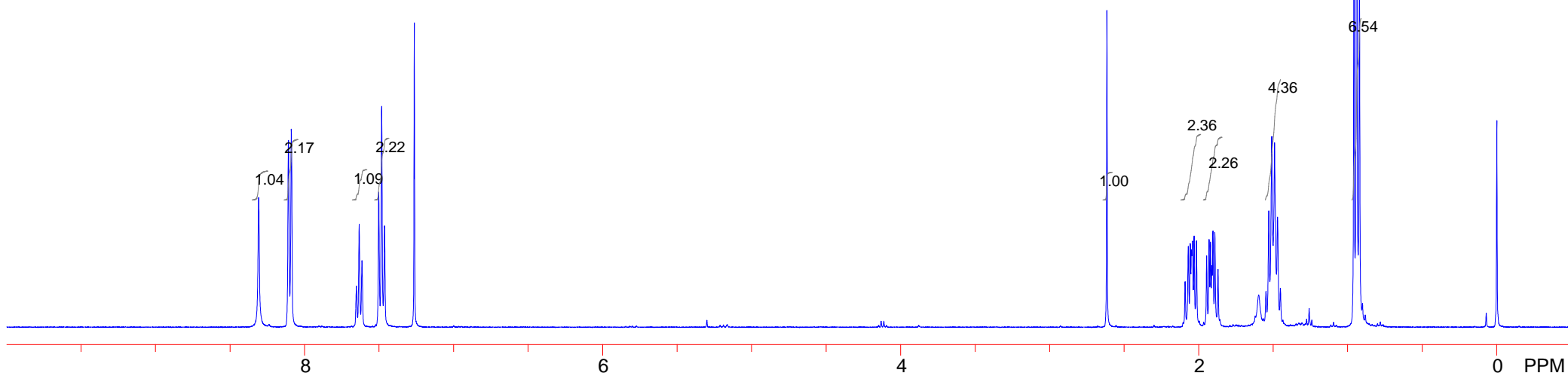
0 PPM

8.308  
8.108  
8.089  
7.652  
7.634  
7.615  
7.502  
7.483  
7.464  
7.264

2.092  
2.070  
2.057  
2.050  
2.617  
2.042  
2.031  
2.015  
1.947  
1.931  
1.921  
1.912  
1.905  
1.892  
1.871  
1.549  
1.530  
1.511  
1.491  
1.471  
1.453  
0.959  
0.941  
0.922  
0.000



**1C**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



165.833

154.452

134.109

129.873

128.601

126.704

82.351

81.386

77.316

77.000

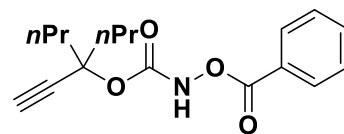
76.676

74.779

40.511

17.172

13.932



**1c**

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

150

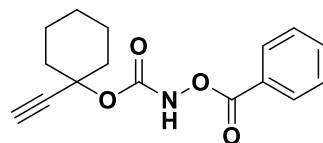
100

50

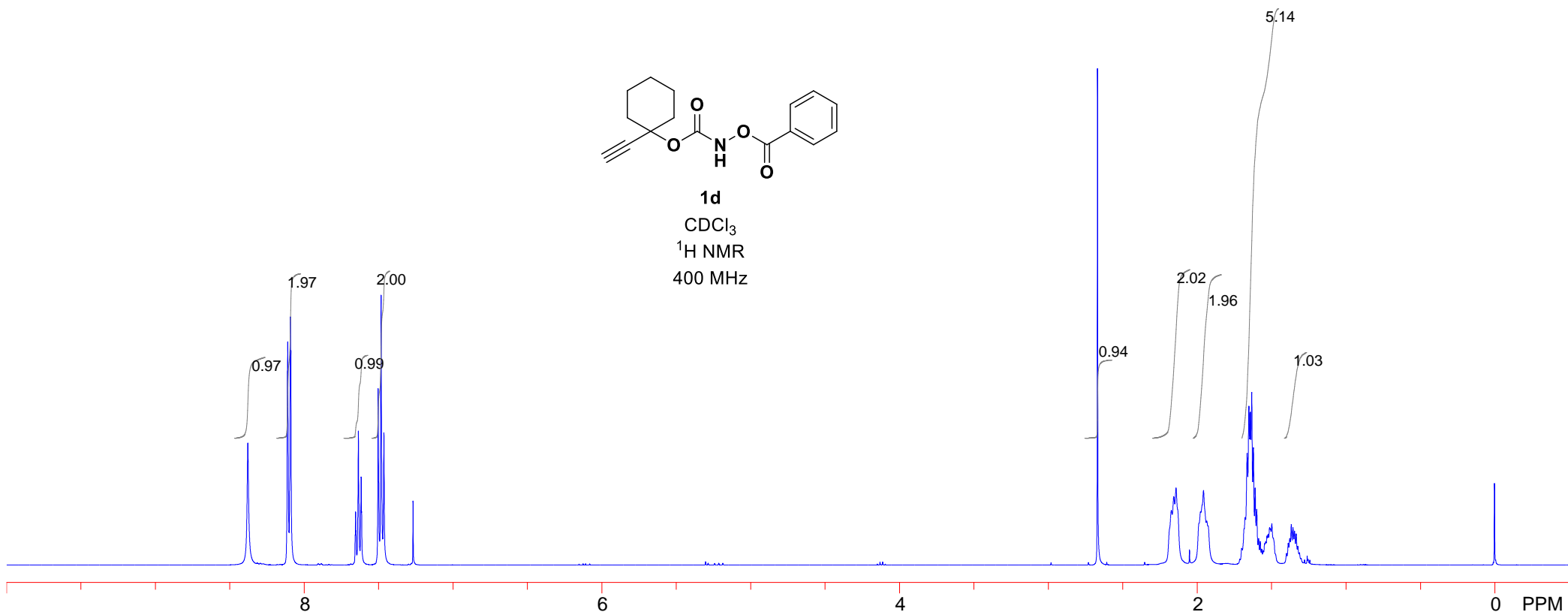
0 PPM

8.378  
8.110  
8.092  
8.089  
7.657  
7.654  
7.651  
7.636  
7.620  
7.617  
7.614  
7.503  
7.483  
7.464  
7.269

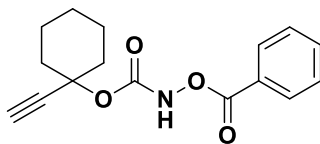
2.172  
2.156  
2.141  
1.976  
1.956  
1.934  
2.668  
1.925  
1.683  
1.676  
1.663  
1.650  
1.641  
1.632  
1.621  
1.609  
1.598  
1.585  
1.529  
1.522  
1.512  
1.506  
1.496  
1.487  
1.376  
1.366  
1.354  
1.344  
1.333  
1.322



**1d**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz







**1d**

CDCl<sub>3</sub>  
13C NMR  
100 MHz

165.847

154.377

134.173

129.907

128.646

126.655

82.644

77.868

77.321

77.000

76.679

75.046

36.919

24.829

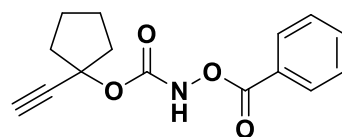
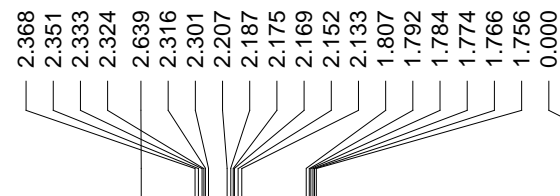
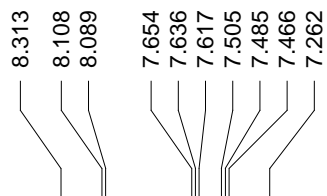
22.336

150

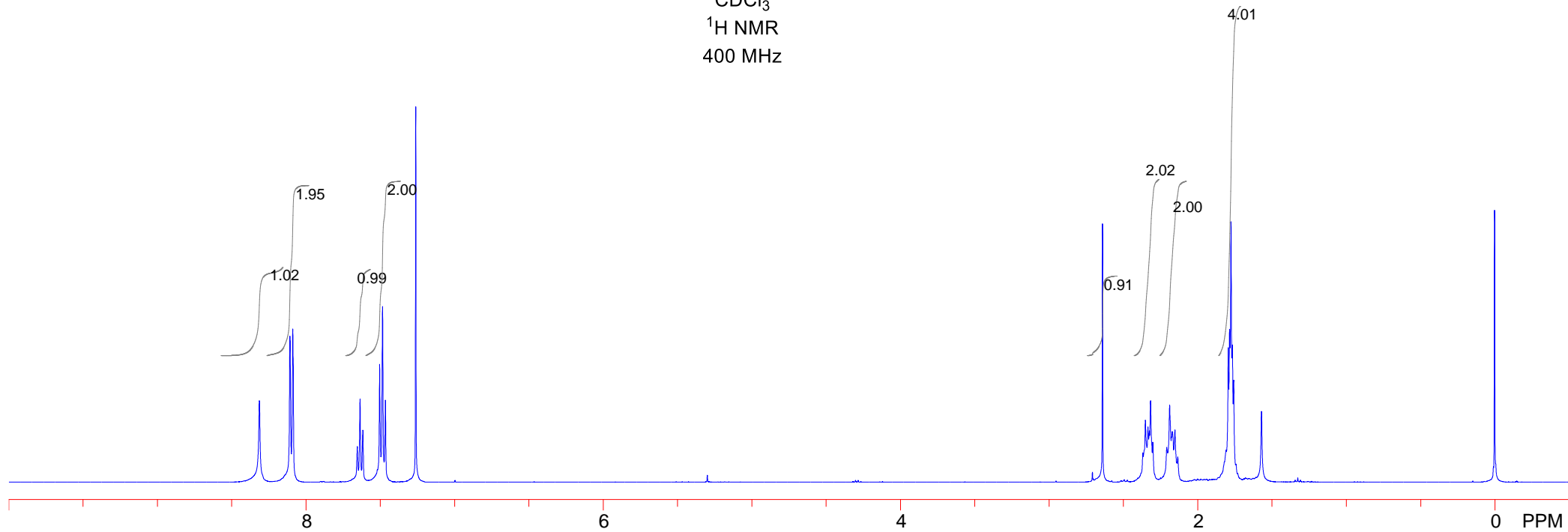
100

50

0 PPM



**1e**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



165.818

154.822

134.202

129.900

128.660

126.597

82.964

82.855

77.314

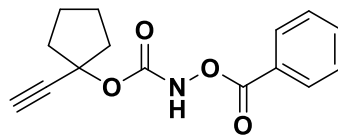
77.000

76.679

73.879

40.324

23.145

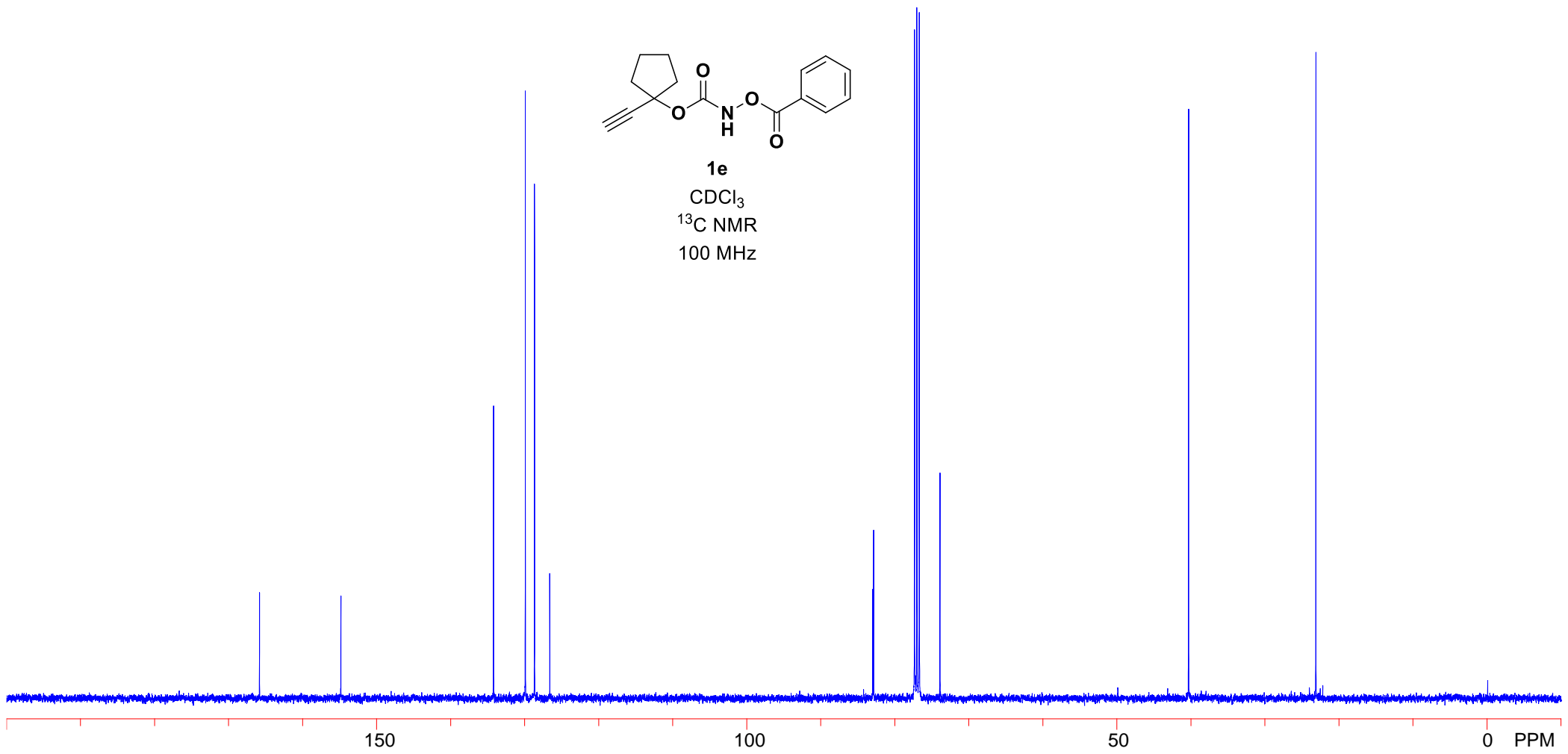


**1e**

CDCl<sub>3</sub>

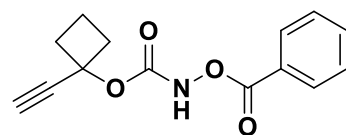
<sup>13</sup>C NMR

100 MHz

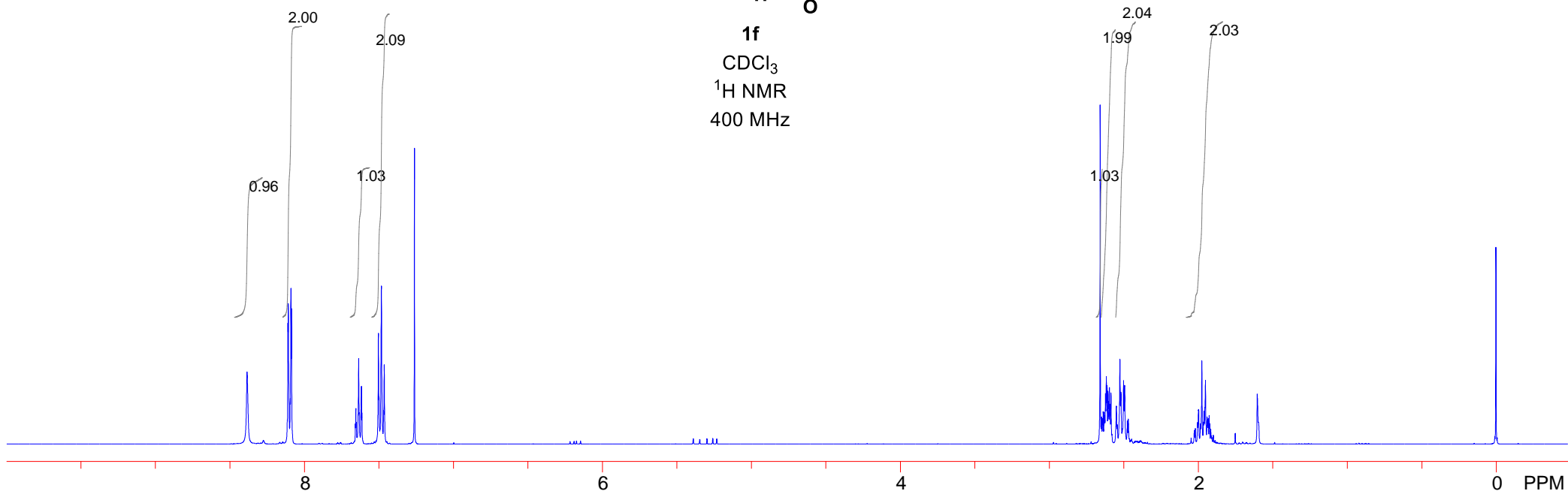


8.385  
8.111  
8.108  
8.095  
8.090  
8.087  
7.655  
7.640  
7.636  
7.632  
7.621  
7.617  
7.614  
7.504  
7.500  
7.483  
7.469  
7.465  
7.262

2.657  
2.637  
2.630  
2.628  
2.622  
2.616  
2.611  
2.605  
2.602  
2.595  
2.592  
2.590  
2.585  
2.548  
2.524  
2.518  
2.517  
2.501  
2.493  
1.998  
1.996  
1.975  
1.960  
1.954  
1.950  
1.925  
-0.000



**1f**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



165.723

154.363

134.253

129.944

128.668

126.524

82.913

77.321

77.000

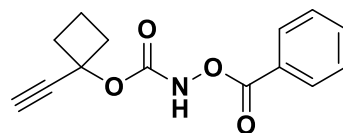
76.686

73.631

73.515

36.357

14.140

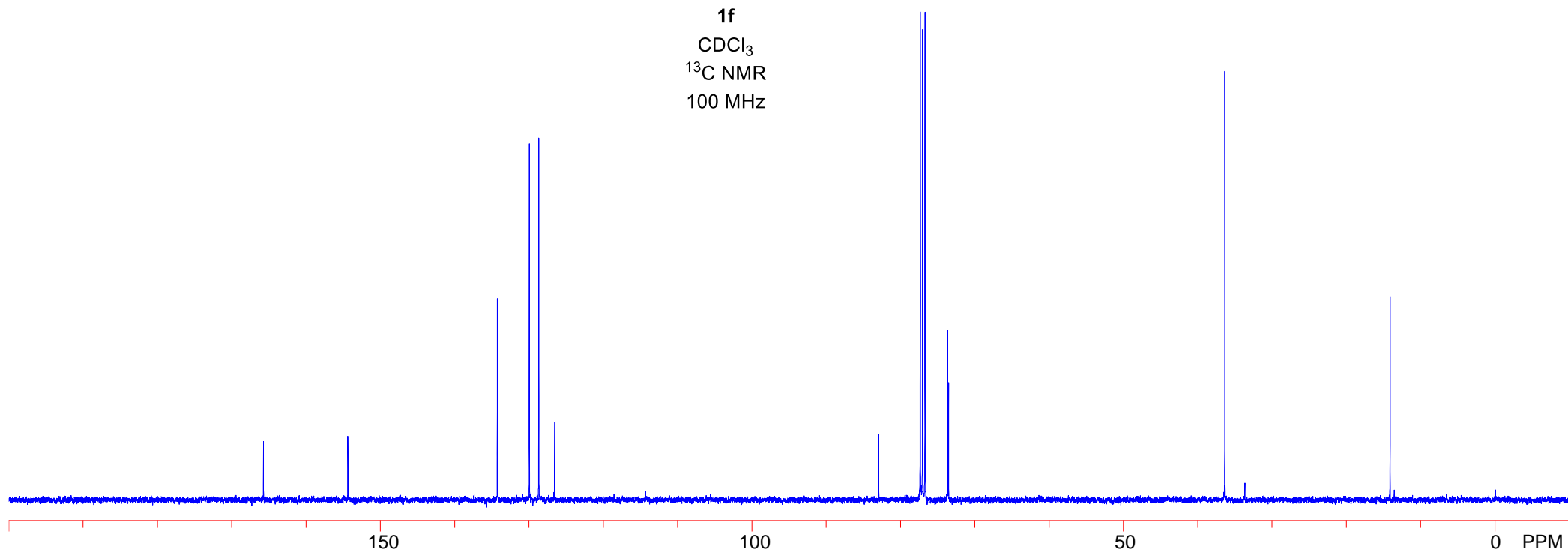


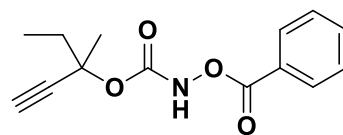
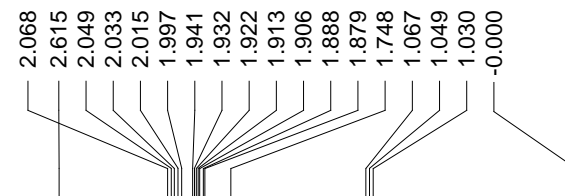
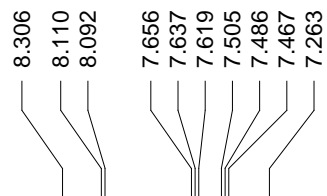
**1f**

CDCl<sub>3</sub>

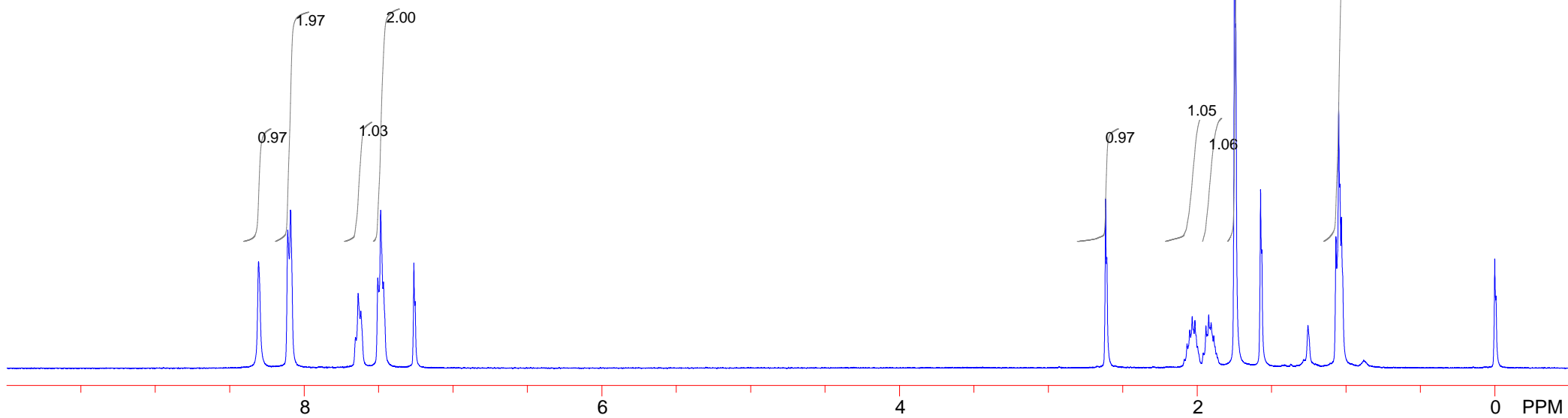
<sup>13</sup>C NMR

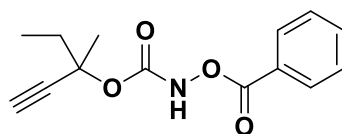
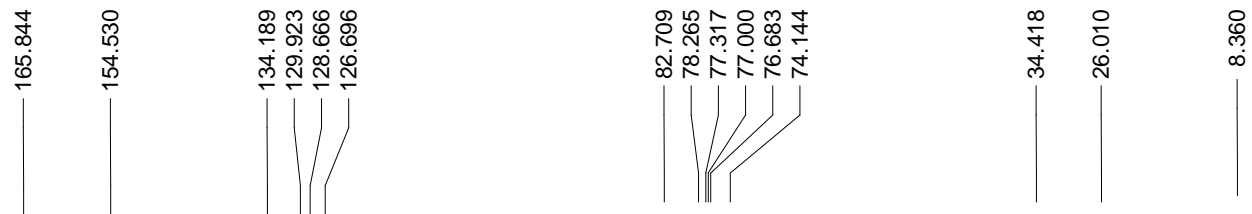
100 MHz



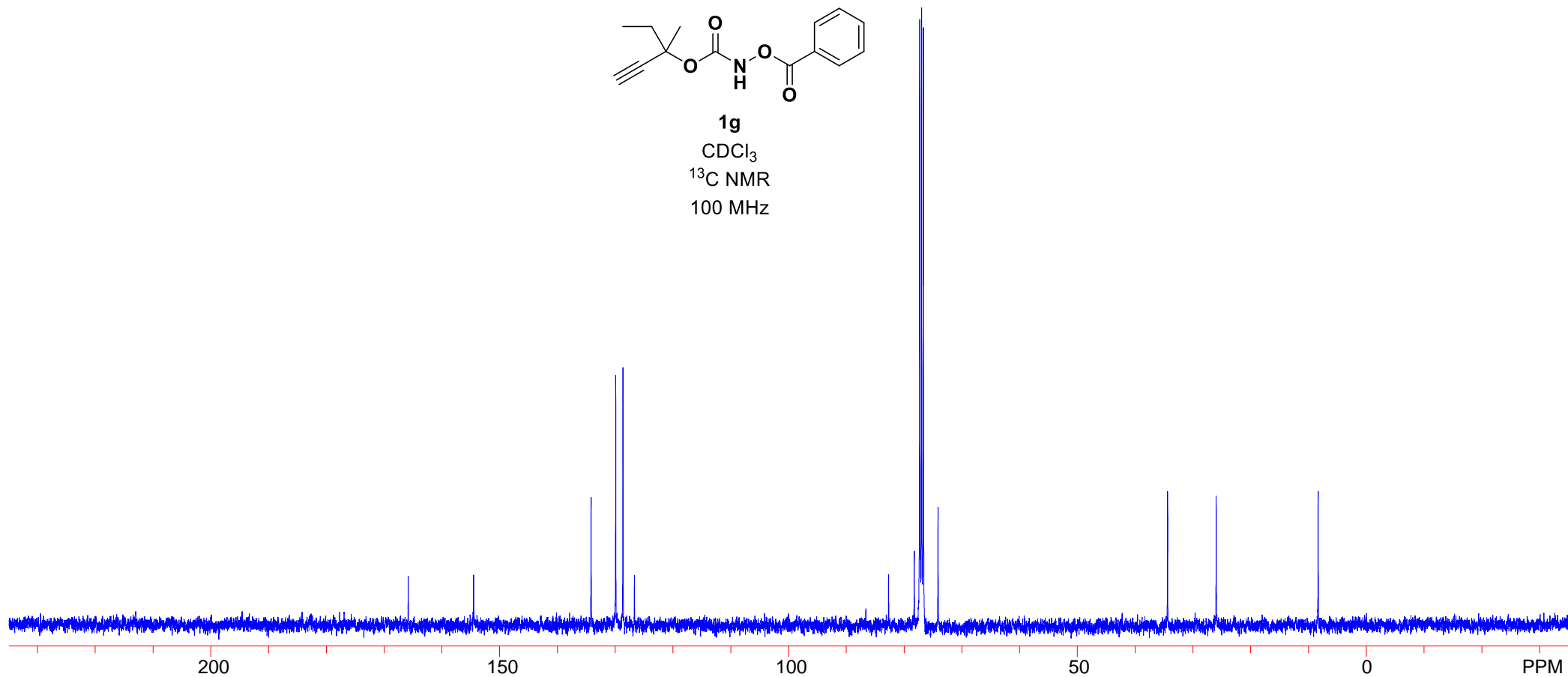


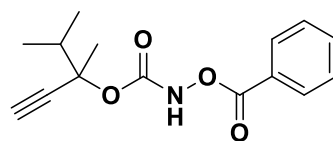
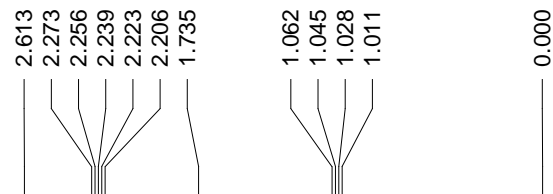
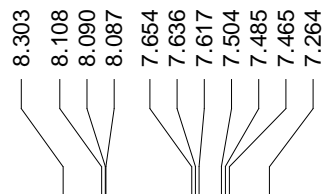
**1g**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



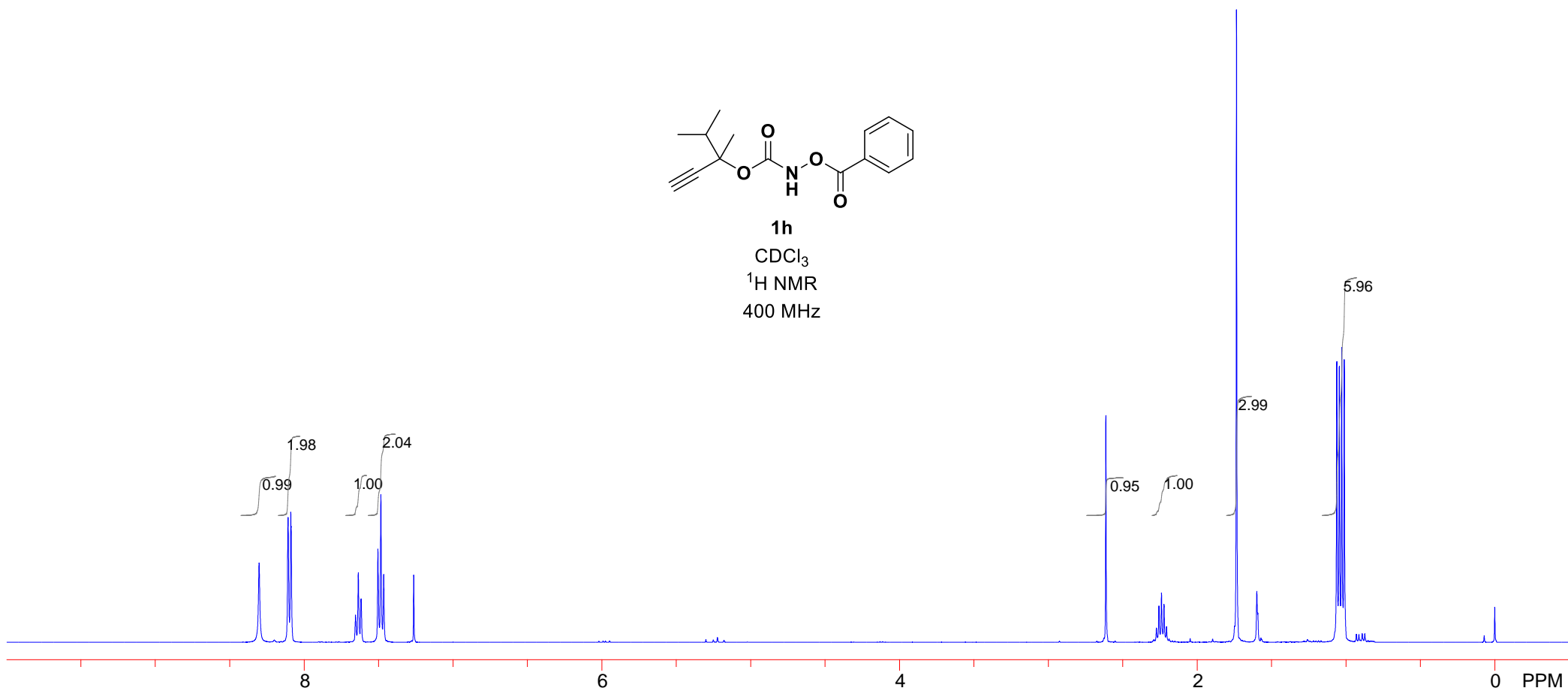


**1g**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

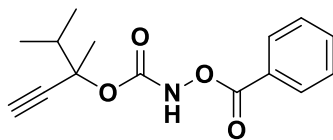




**1h**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz







**1h**

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

165.853

154.563

134.189

129.915

128.679

126.717

81.818

81.534

77.317

77.000

76.683

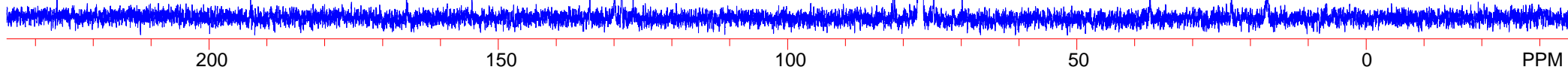
74.782

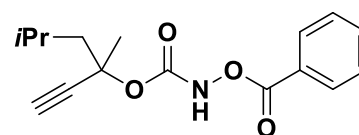
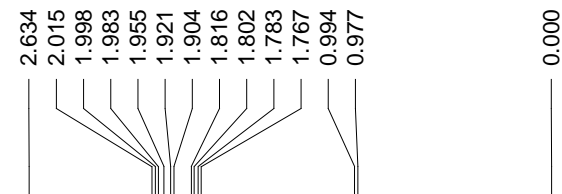
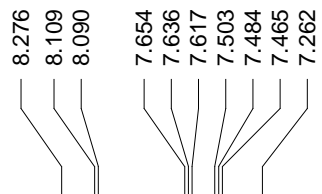
37.353

23.302

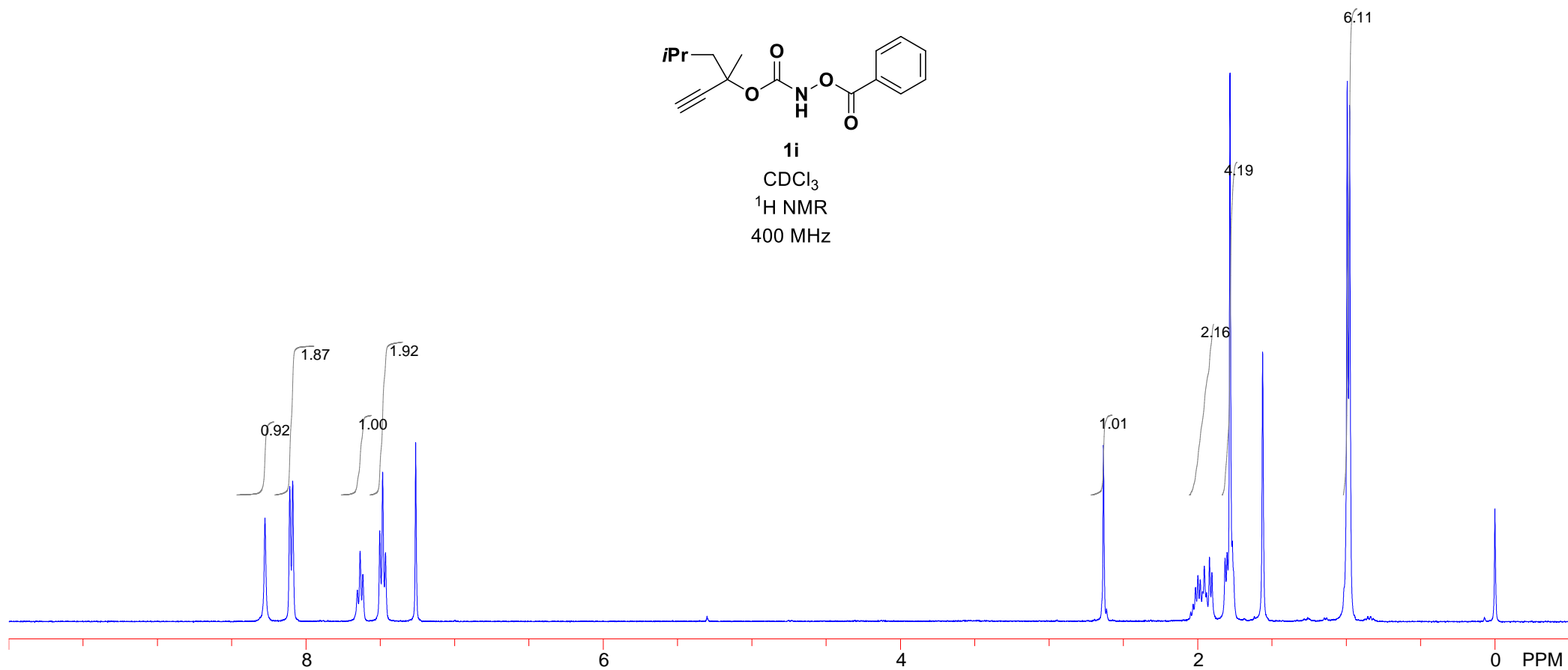
17.317

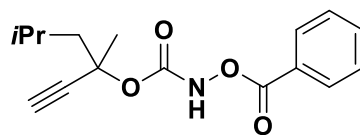
17.045





**1i**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





**1i**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

165.810

154.475

134.176

129.922

128.645

126.683

83.100

77.948

77.317

77.000

76.683

74.407

49.397

27.288

24.774

23.974

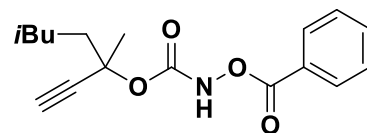
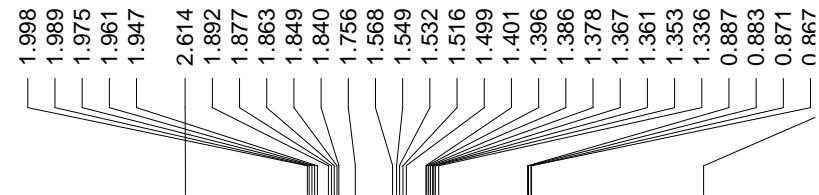
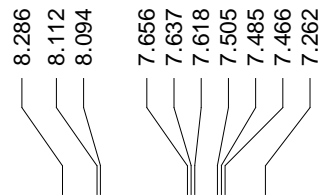
23.711

150

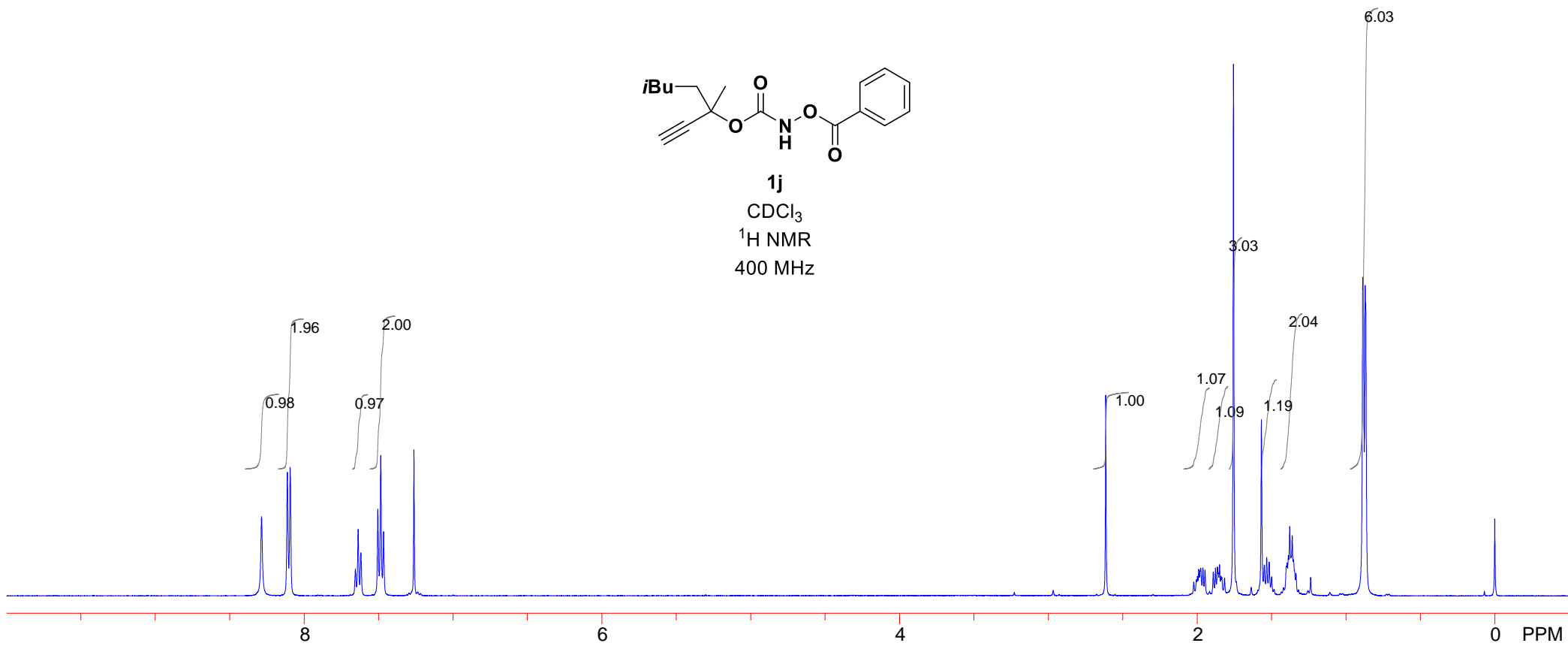
100

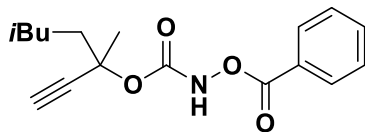
50

0 PPM



1j  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





1j

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

165.778

154.530

134.202

129.944

128.674

126.708

82.960

77.911

77.317

77.000

76.683

74.111

39.443

32.790

27.931

26.504

22.477

22.436

150

100

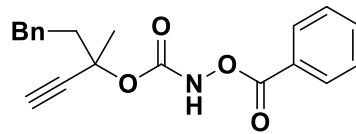
50

0 PPM

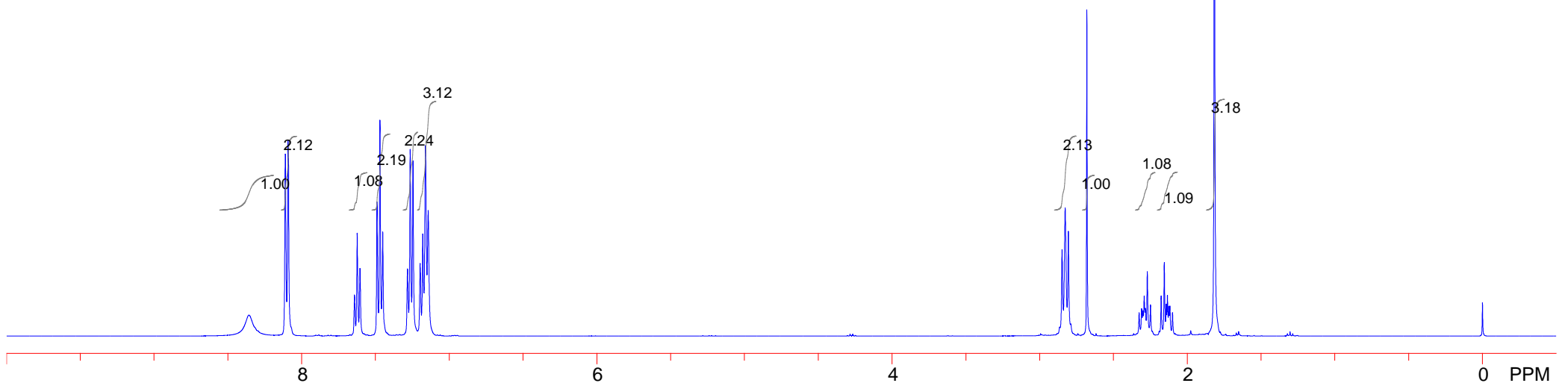
8.357  
8.110  
8.092  
8.089  
7.642  
7.624  
7.605  
7.490  
7.470  
7.451  
7.283  
7.265  
7.245  
7.197  
7.179  
7.161  
7.143

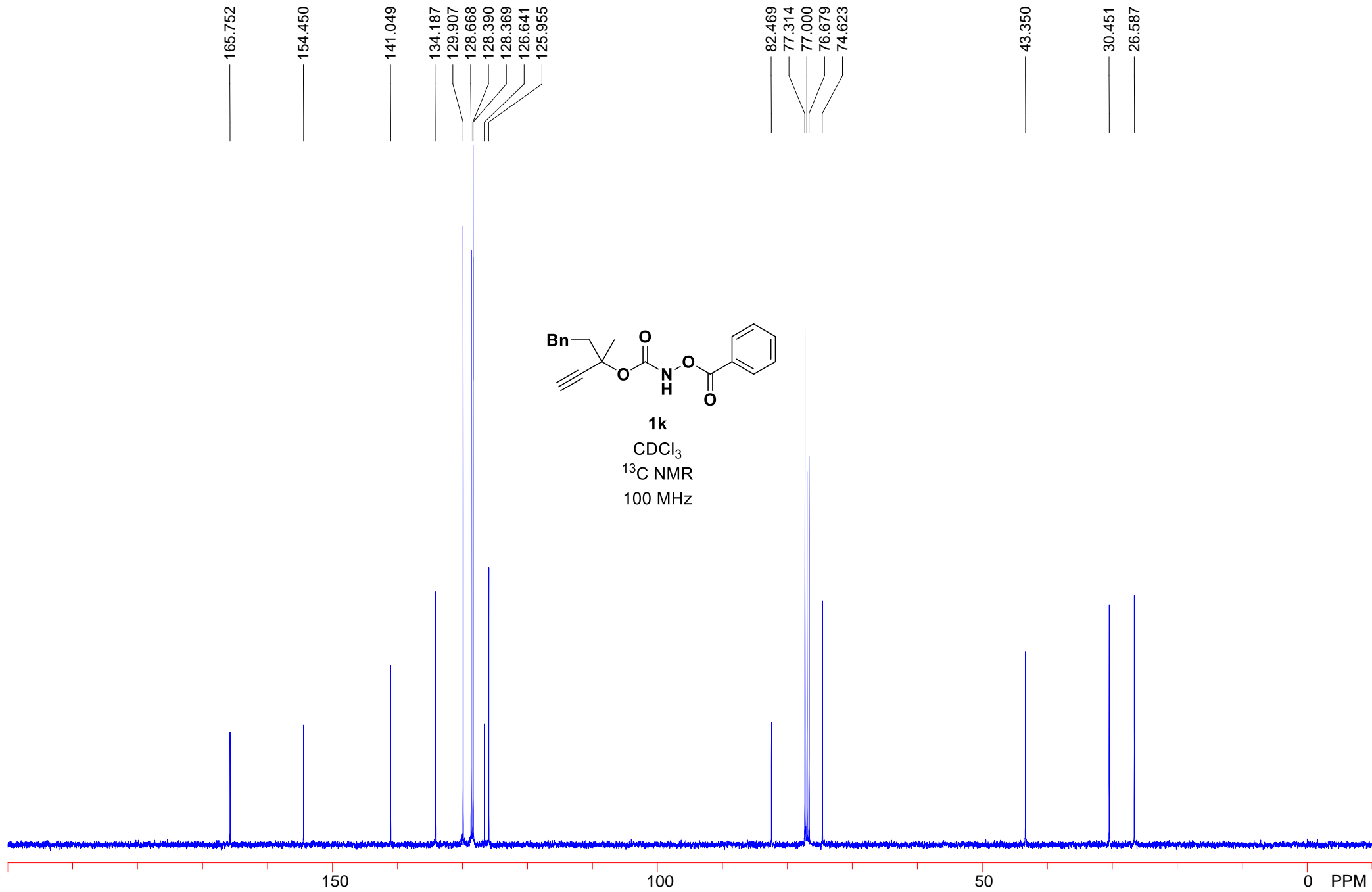
2.848  
2.827  
2.806  
2.680  
2.326  
2.309  
2.301  
2.292  
2.284  
2.271  
2.249  
2.177  
2.155  
2.142  
2.134  
2.125  
2.118  
2.100  
1.816

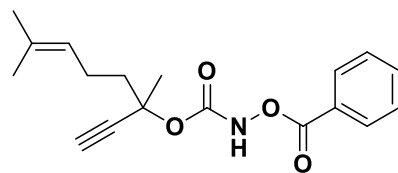
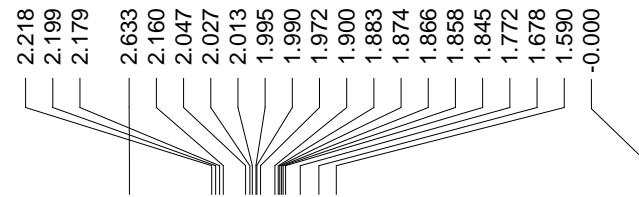
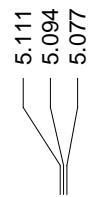
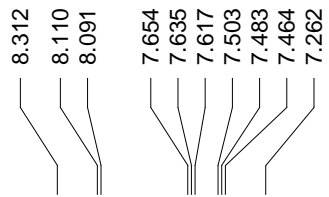
0.000



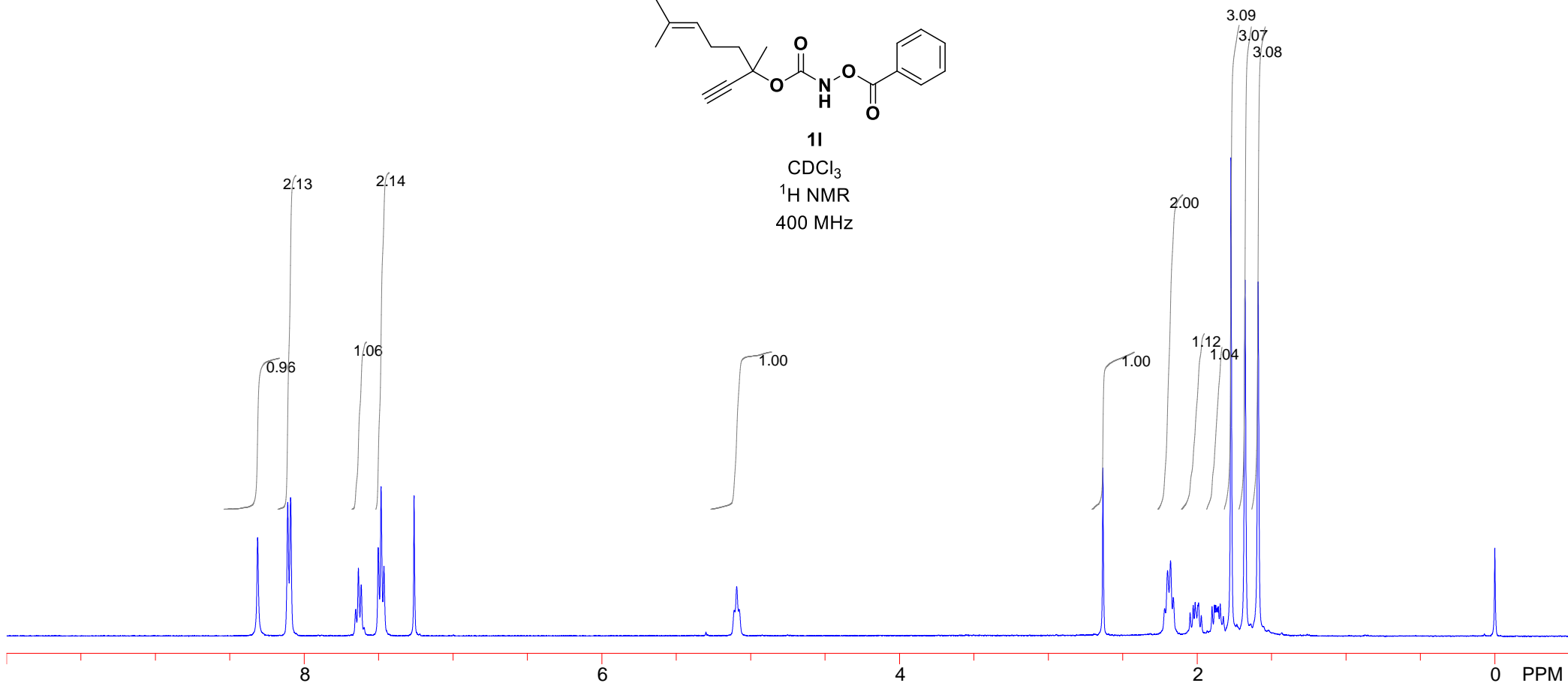
**1k**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz







**11**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





165.814

154.541

134.193

132.523

129.935

128.674

126.683

122.817

82.729

77.631

77.321

77.000

76.683

74.288

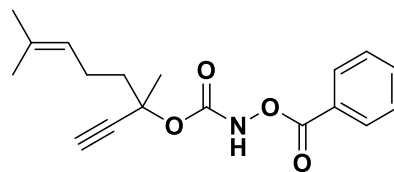
41.380

26.489

25.615

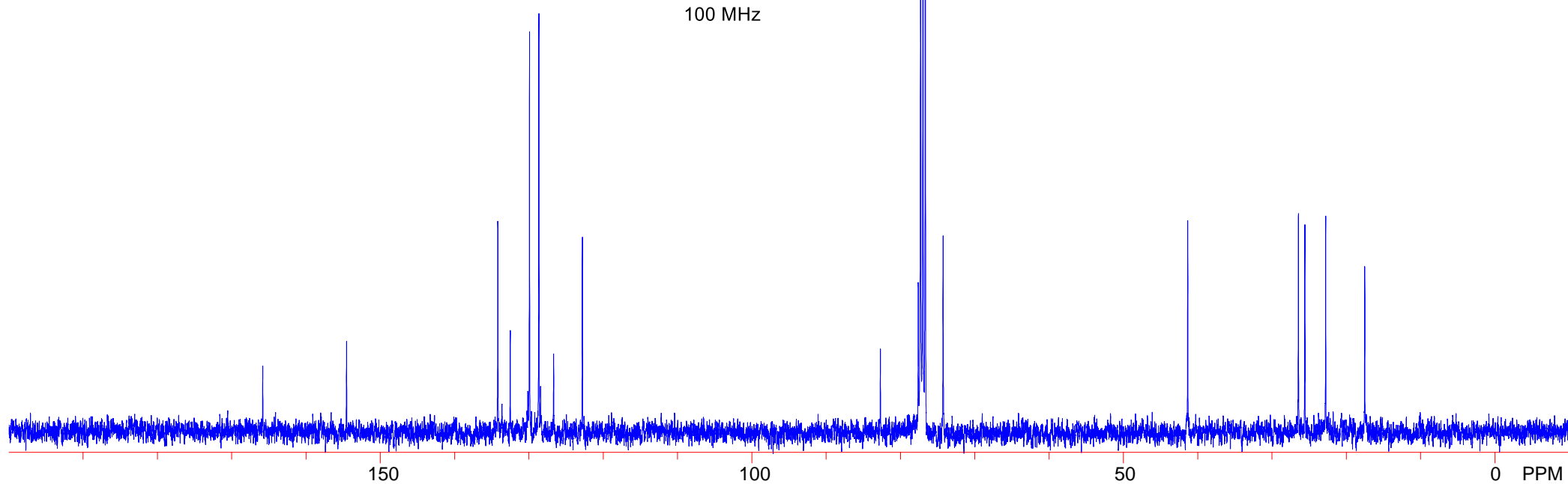
22.816

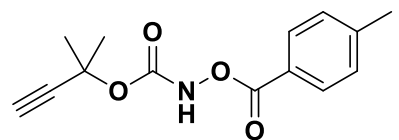
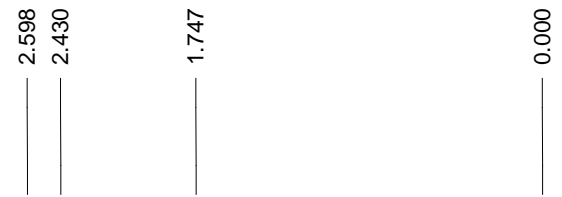
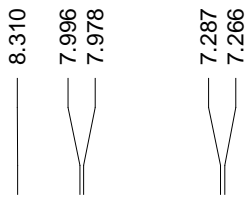
17.561



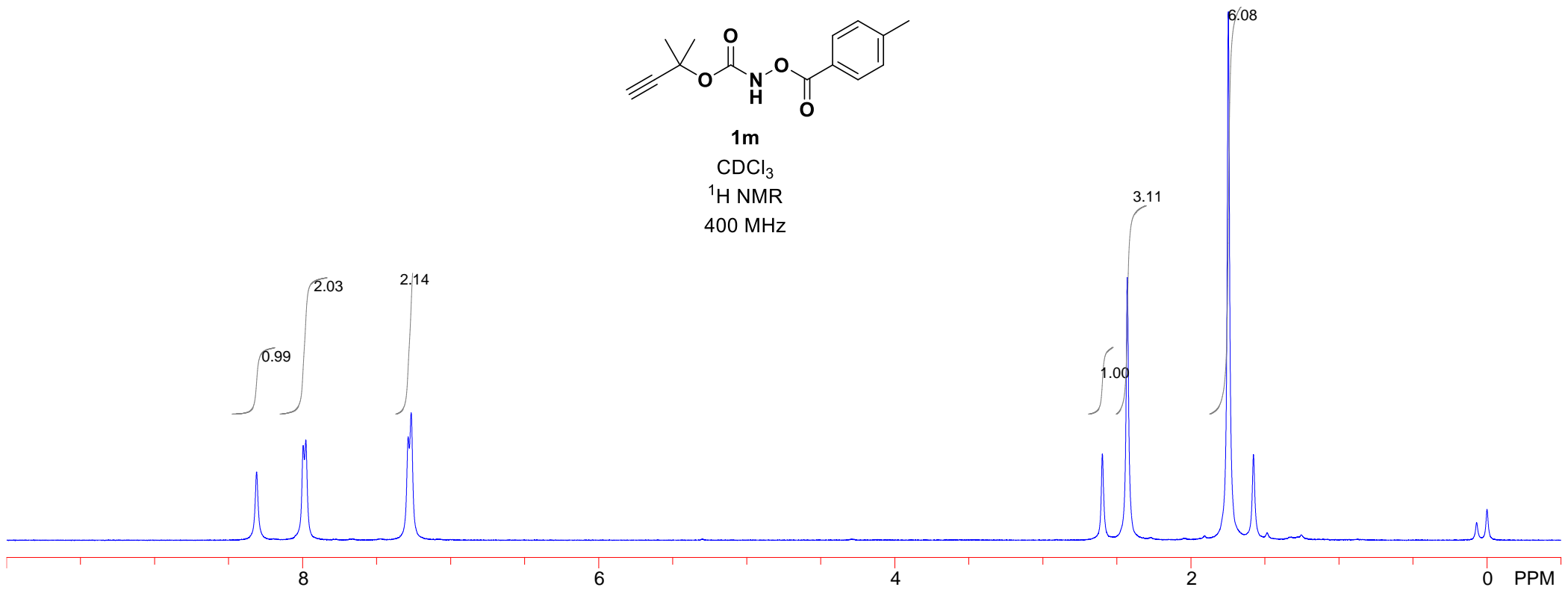
**11**

CDCl<sub>3</sub>  
13C NMR  
100 MHz

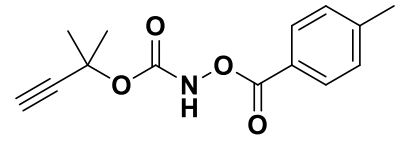




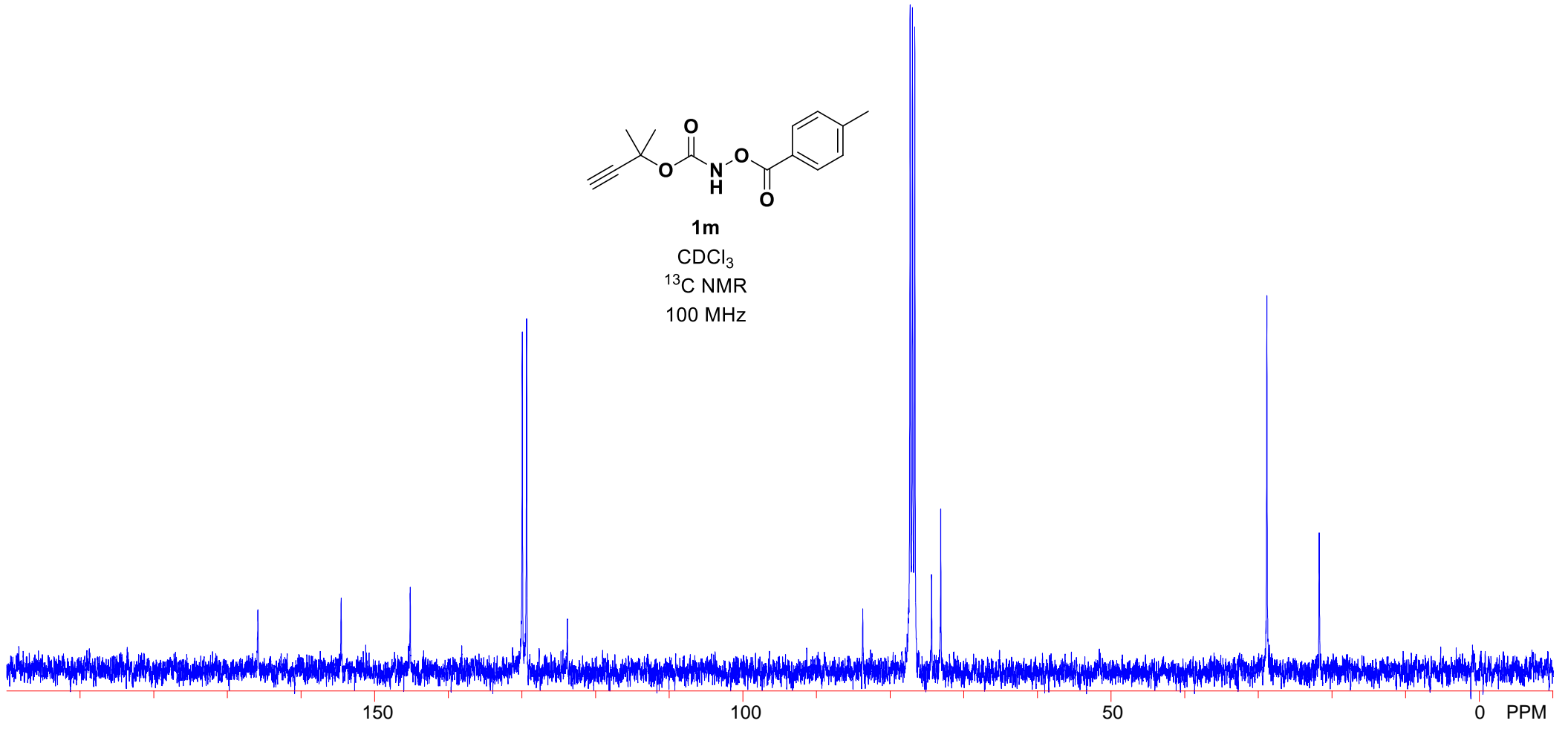
**1m**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

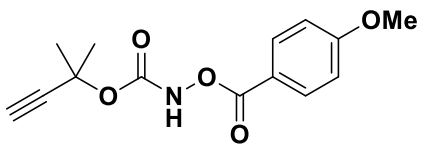
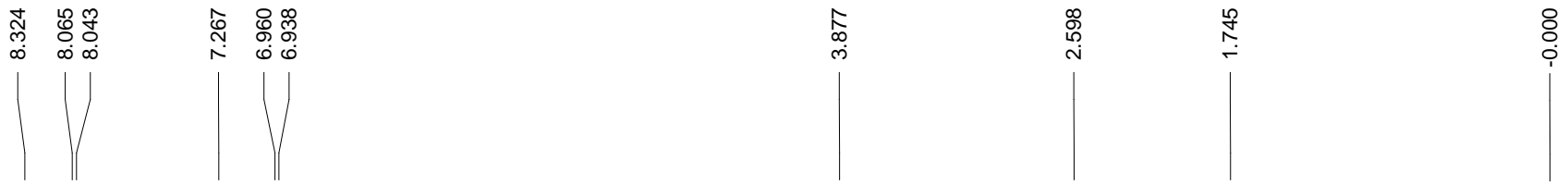


165.859  
154.550  
145.169  
129.960  
129.374  
123.831  
83.747  
77.313  
77.000  
76.679  
74.399  
73.159  
28.879  
21.778

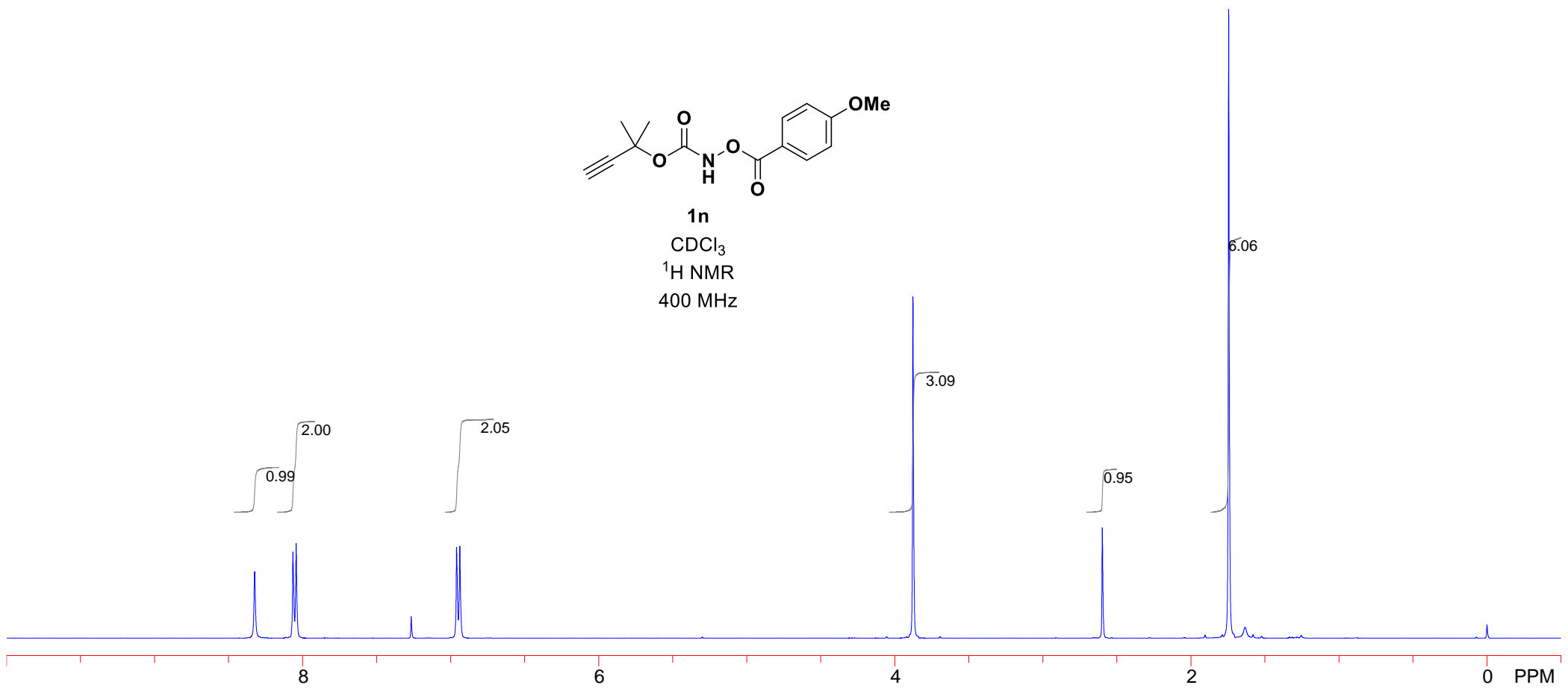


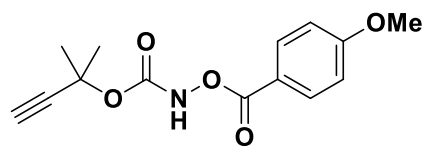
**1m**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz





**1n**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





**1n**

CDCl<sub>3</sub>  
13C NMR  
100 MHz

165.517  
164.322

154.640

132.132

118.786

113.980

83.768

77.317

77.000

76.683

74.362

73.142

55.505

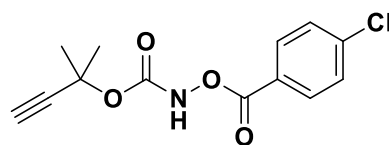
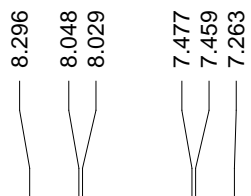
28.896

150

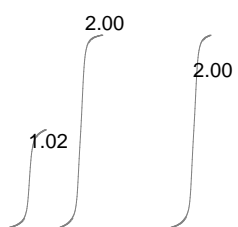
100

50

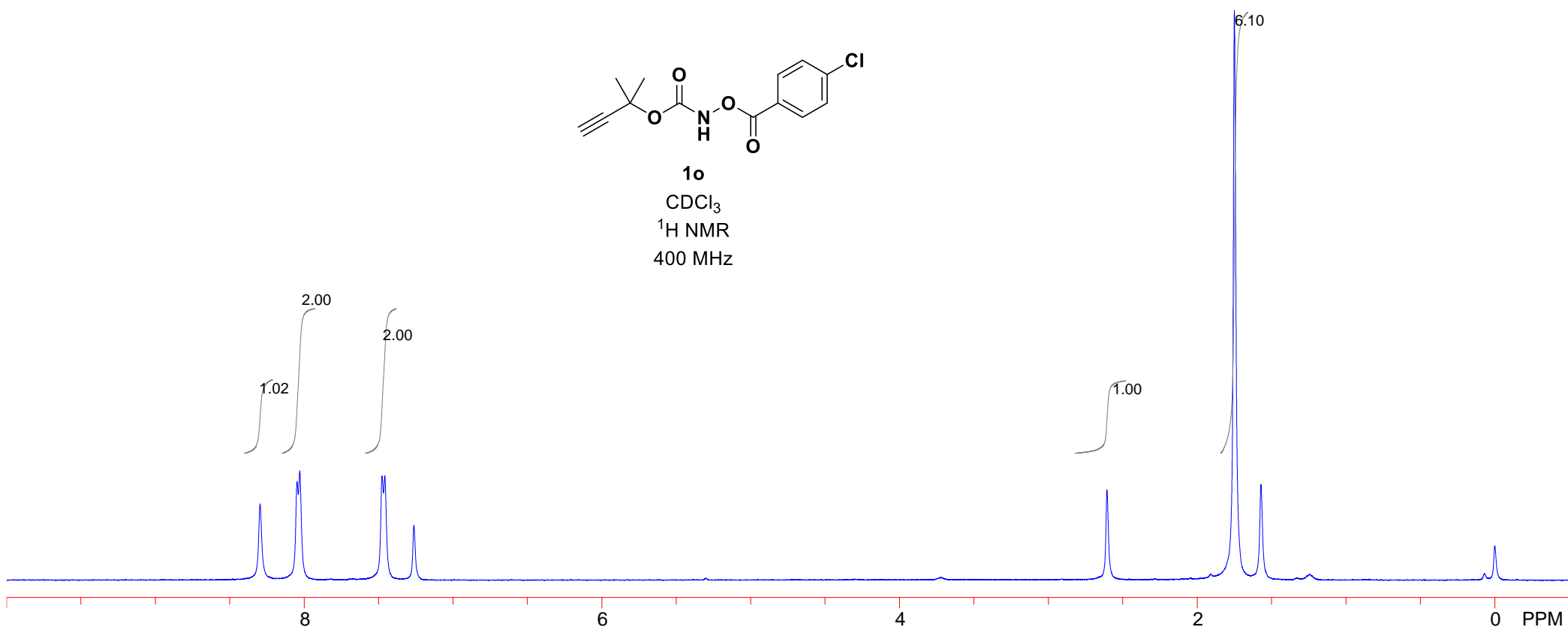
0 PPM

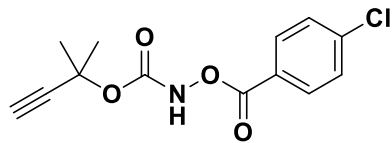


**1o**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



6.10





**1o**

CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

165.014

154.381

140.874

131.299

129.127

125.117

83.624

77.317

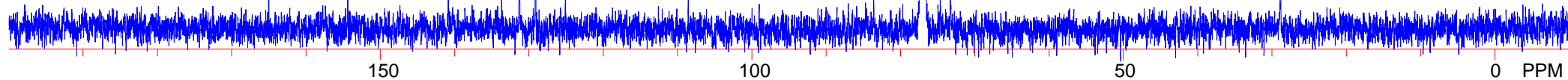
77.000

76.683

74.671

73.295

28.896



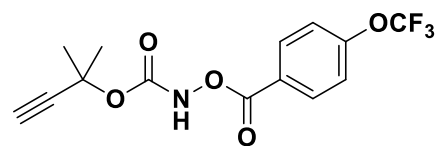
8.301  
8.169  
8.149

7.324  
7.305  
7.263

2.607

1.752

0.000



**1p**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

1.02  
2.00

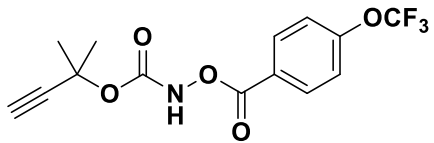
2.01

1.00

6.00

8 6 4 2 0 PPM





**1p**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

164.673

154.392

153.473

132.058

124.934

124.052

121.485

120.414

118.890

116.075

83.570

77.321

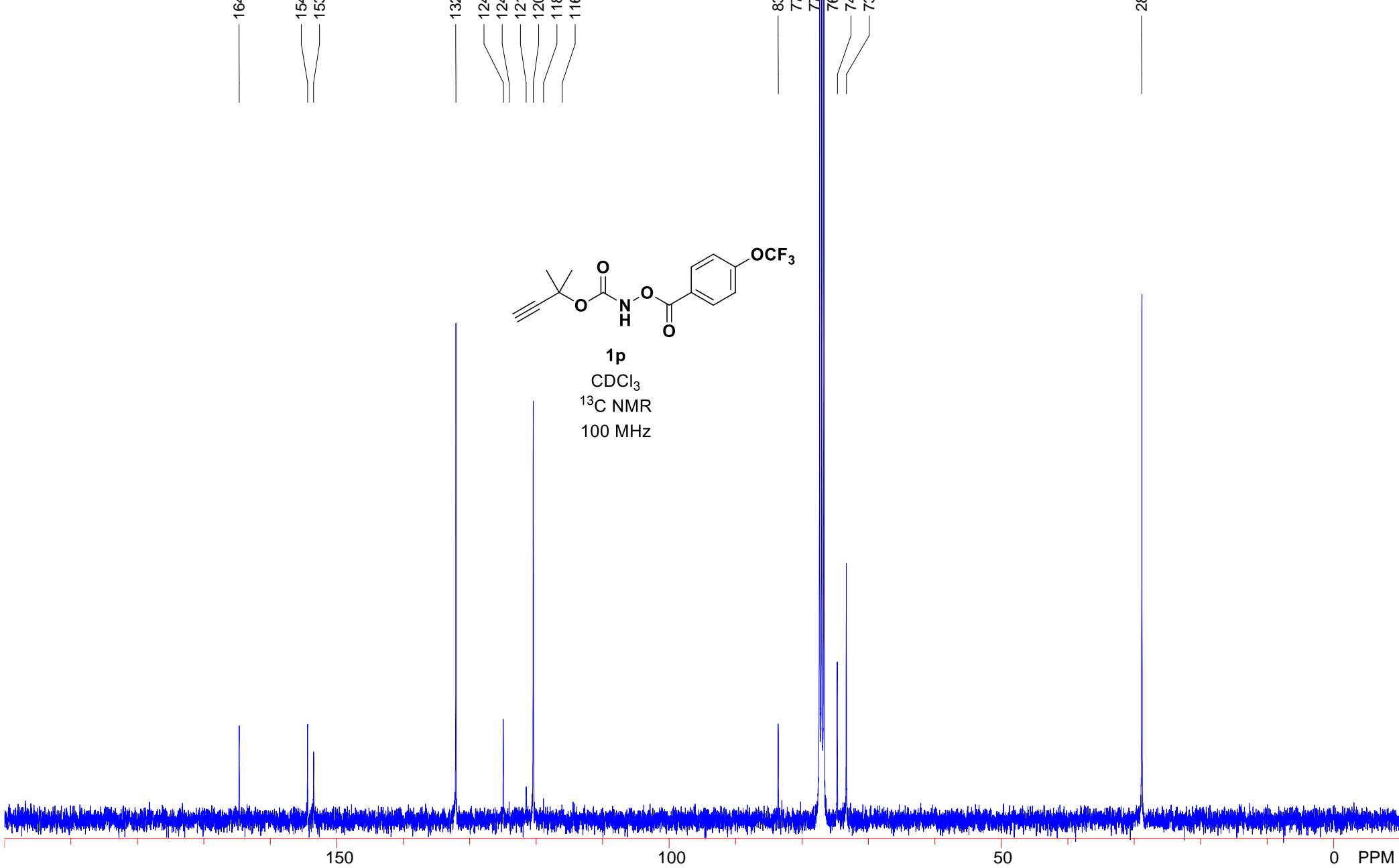
77.000

76.686

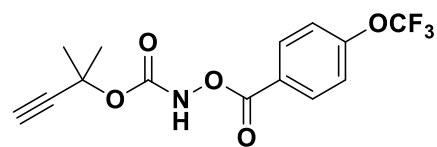
74.674

73.318

28.869



-57.597

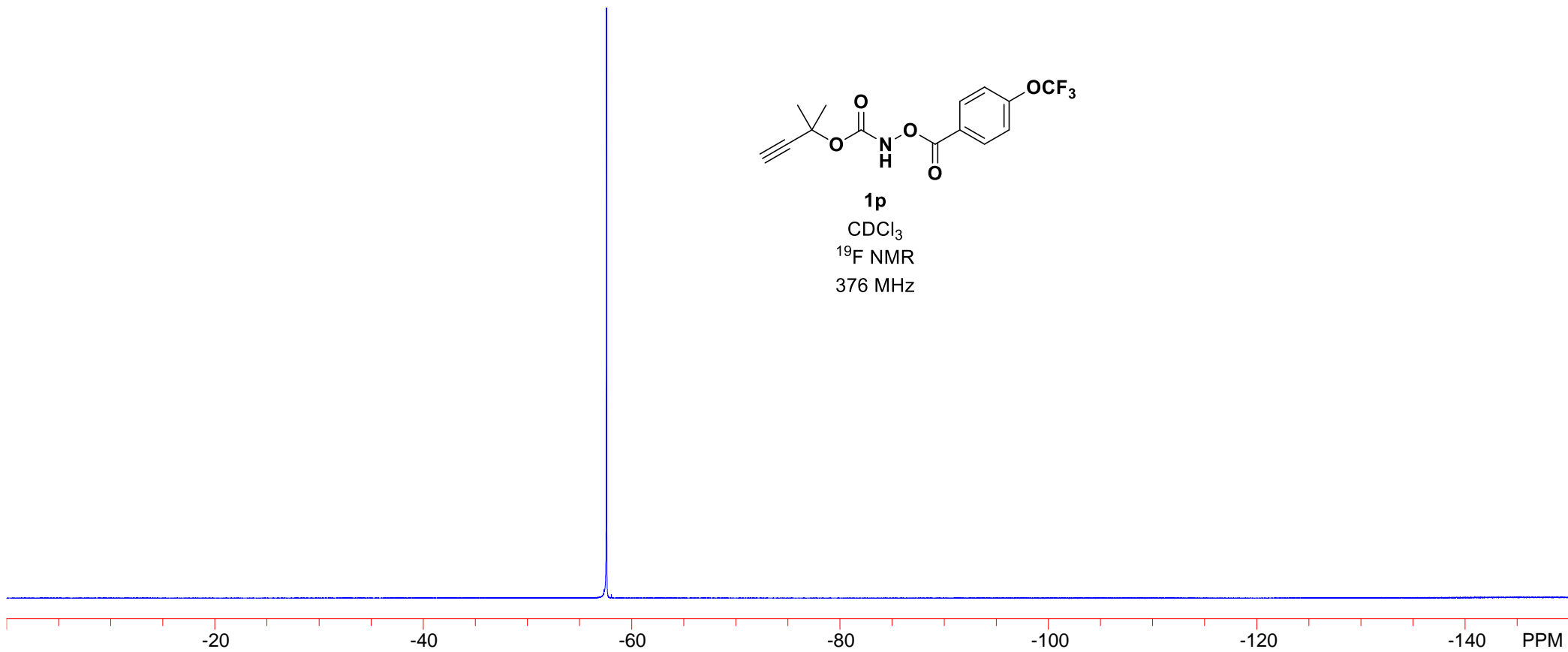


**1p**

CDCl<sub>3</sub>

<sup>19</sup>F NMR

376 MHz



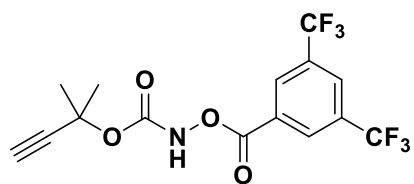
8.554  
8.401  
8.148

7.271

2.632

1.766

-0.000

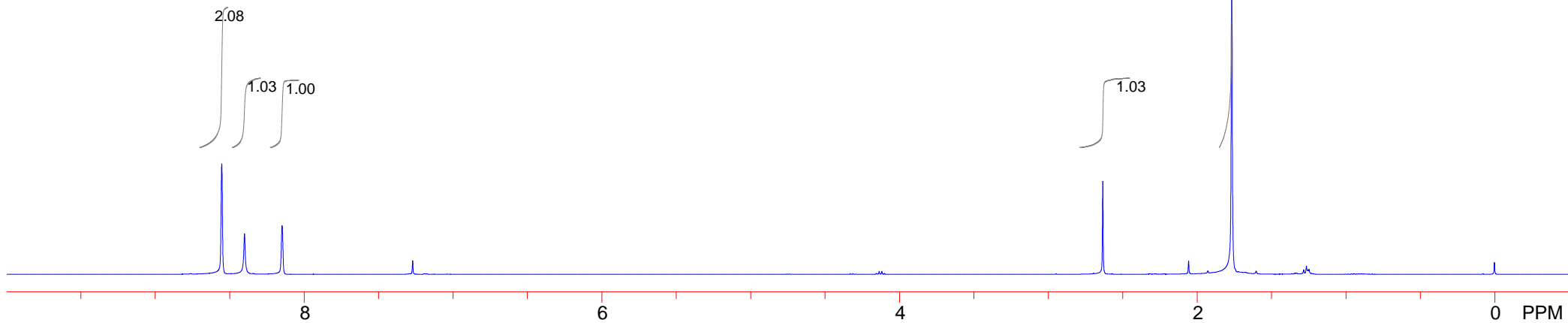


**1q**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

2.08  
1.03  
1.00

1.03

6.12



163.375

154.144

133.108

132.765

132.423

132.080

130.068

130.038

128.974

127.588

127.552

127.515

127.479

126.626

123.906

121.194

118.481

83.365

77.314

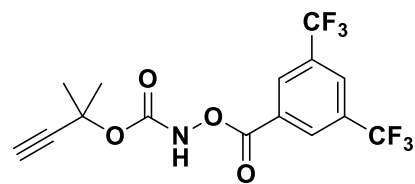
77.000

76.679

75.126

73.515

28.840



**1q**

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz



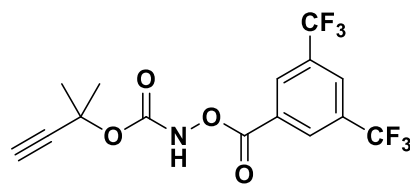
150

100

50

0 PPM

-63.046



**1q**

CDCl<sub>3</sub>

<sup>19</sup>F NMR

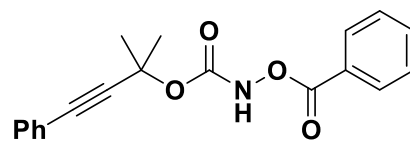
376 MHz



8.359  
8.121  
8.103  
8.100  
7.654  
7.635  
7.617  
7.505  
7.485  
7.466  
7.458  
7.453  
7.445  
7.441  
7.434  
7.314  
7.310  
7.303  
7.299  
7.291  
7.286  
7.277  
7.274  
7.262

1.835

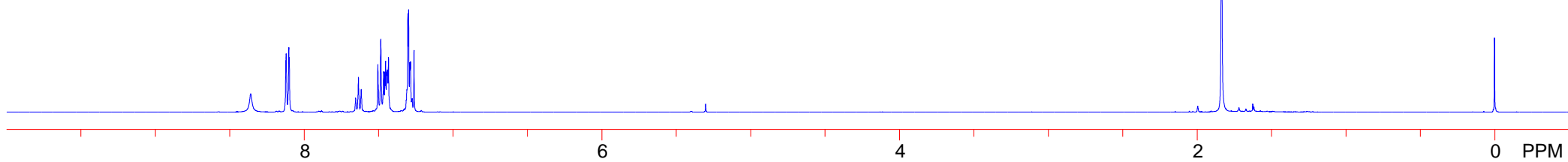
-0.000

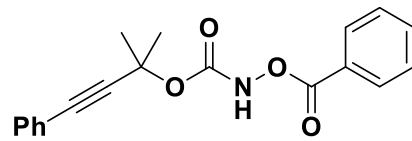


**1r**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

0.94  
2.00  
1.07  
4.24  
3.01

6.13



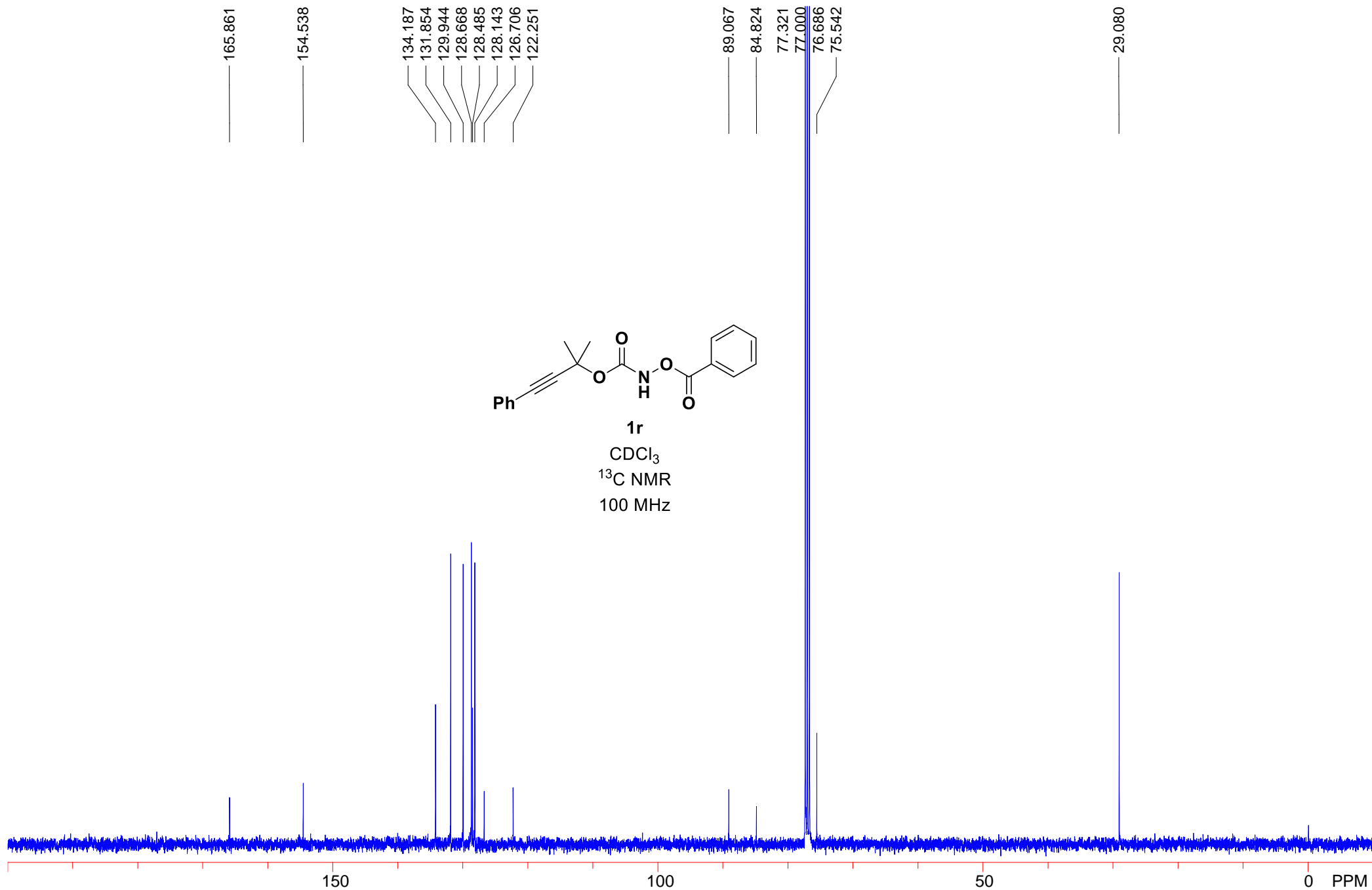


**1r**

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz



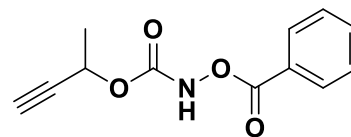
8.498  
8.109  
8.106  
8.093  
8.088  
8.085  
7.665  
7.662  
7.659  
7.643  
7.627  
7.624  
7.621  
7.508  
7.488  
7.473  
7.469  
7.268

5.518  
5.513  
5.501  
5.496  
5.485  
5.479  
5.468  
5.463

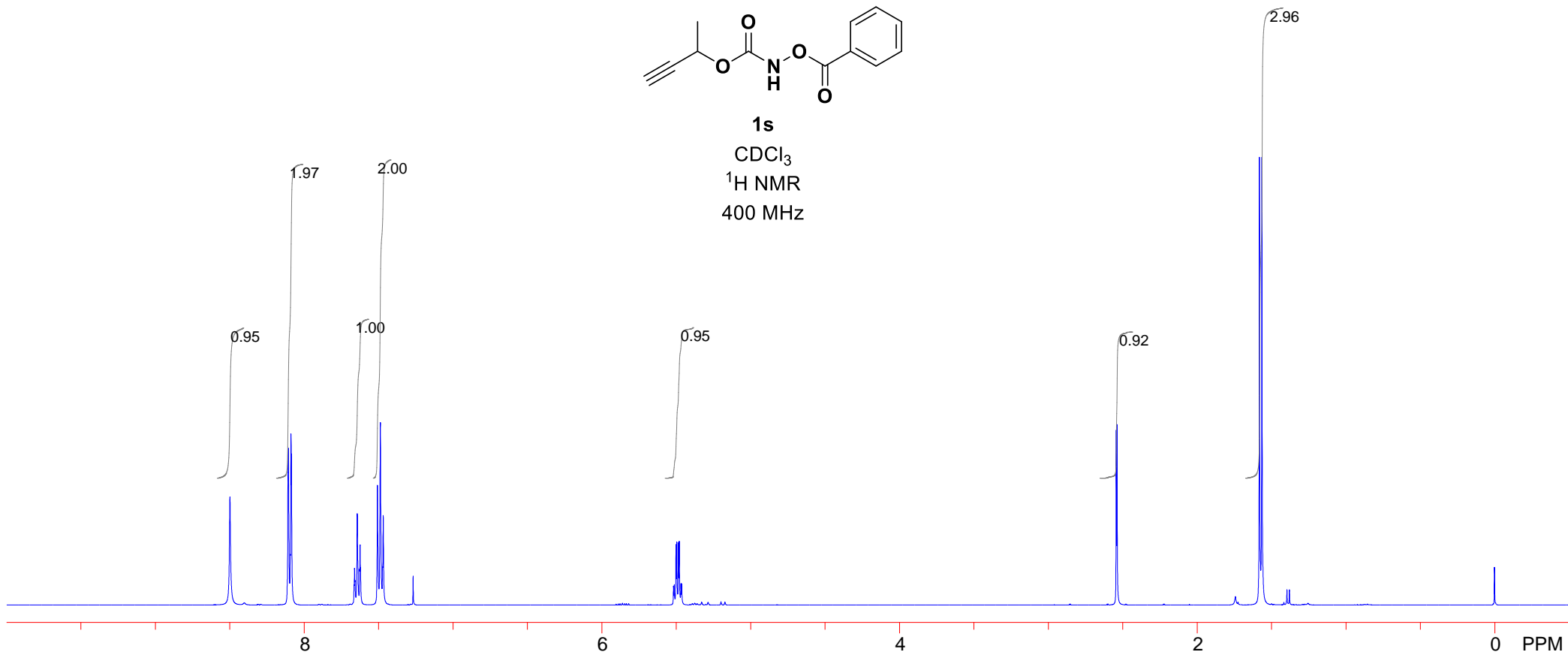
2.542  
2.537

1.580  
1.563

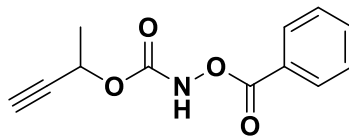
-0.000



**1s**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz







**1s**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

165.628

155.289

134.275

129.944

128.682

126.436

81.047

77.321

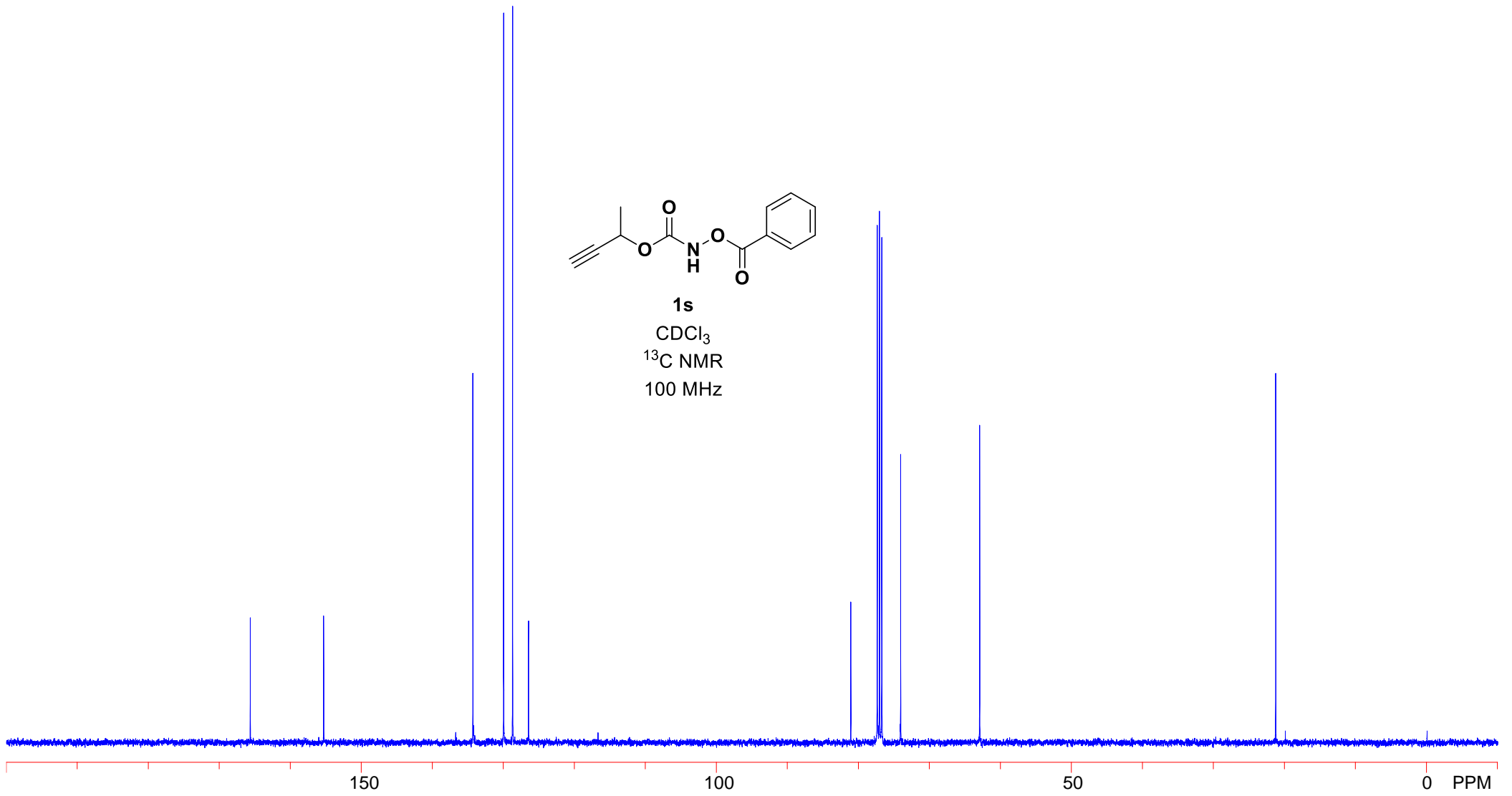
77.000

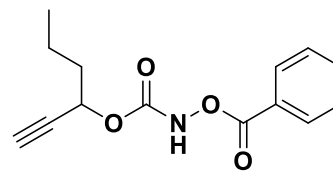
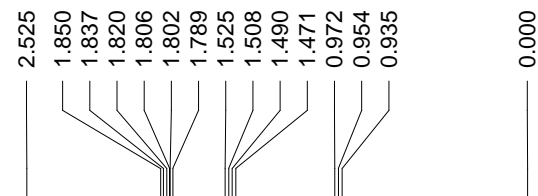
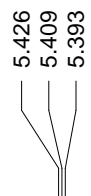
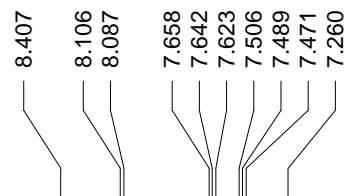
76.686

74.018

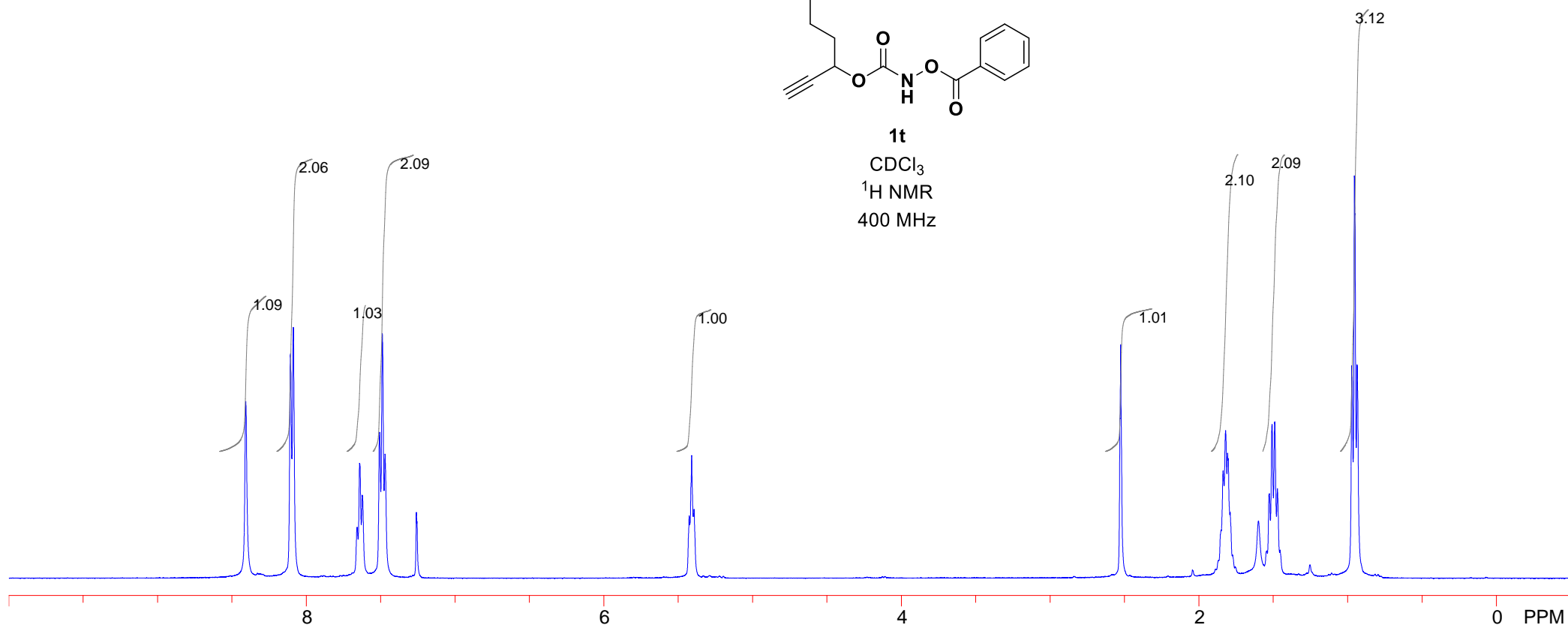
62.898

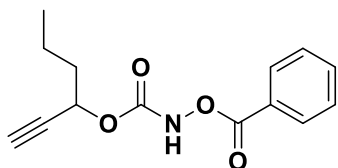
21.242





**1t**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





1t

CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

165.692

155.470

134.284

129.969

128.712

126.539

80.256

77.317

77.000

76.683

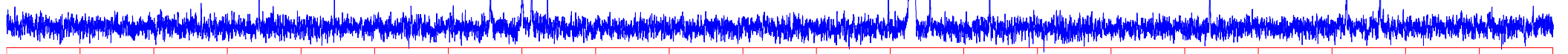
74.613

66.494

36.615

18.047

13.492



150

100

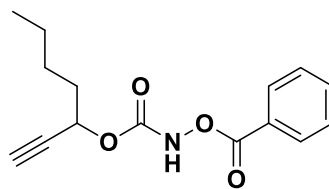
50

0 PPM

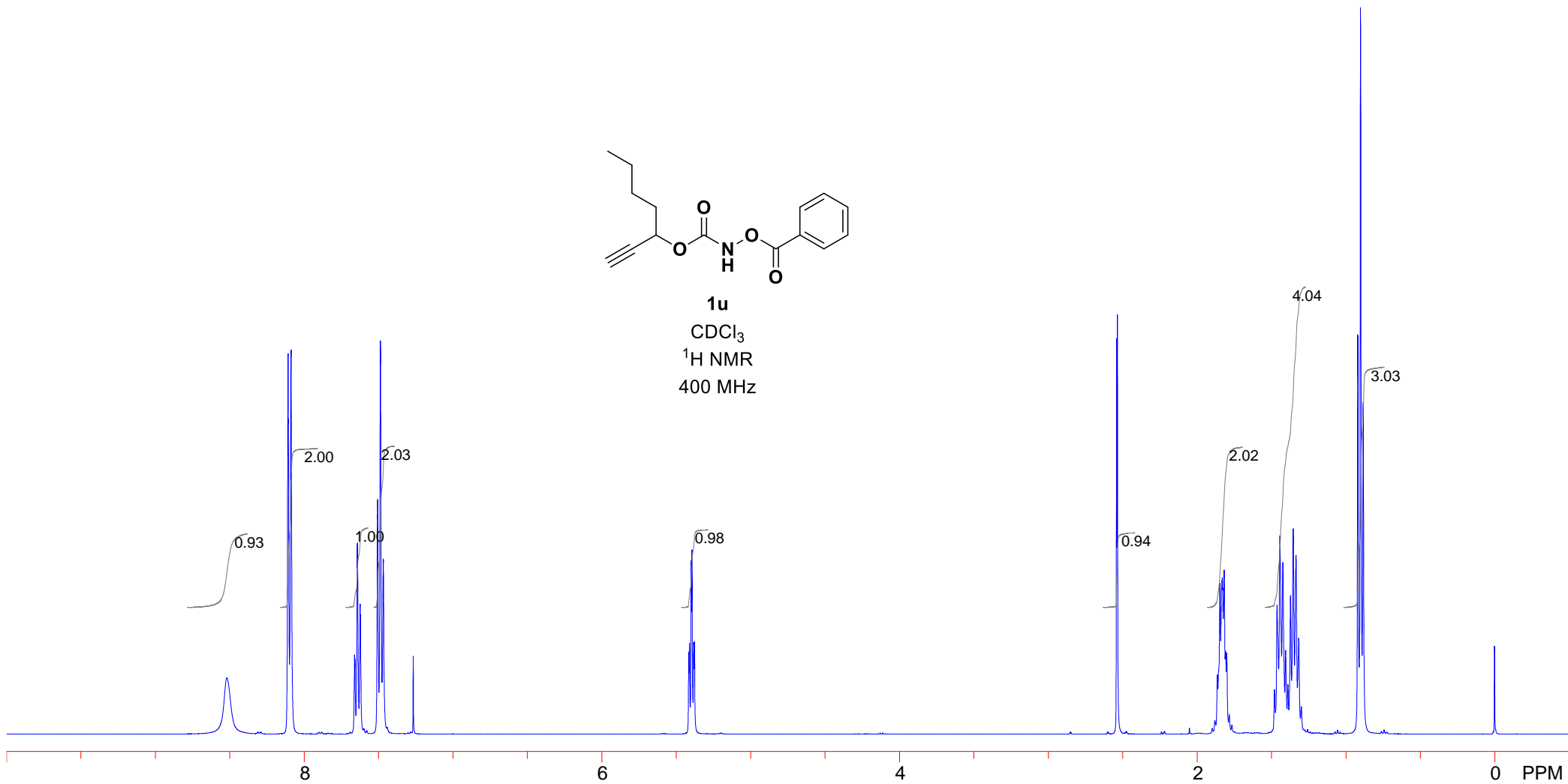
8.519  
8.108  
8.106  
8.090  
8.087  
8.085  
7.662  
7.659  
7.656  
7.643  
7.641  
7.624  
7.622  
7.508  
7.506  
7.488  
7.469  
7.468  
7.268

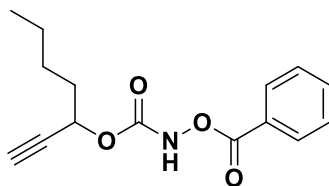
5.416  
5.411  
5.399  
5.394  
5.383  
5.377

2.540  
2.534  
1.864  
1.862  
1.855  
1.846  
1.835  
1.832  
1.830  
1.827  
1.817  
1.808  
1.800  
1.479  
1.461  
1.442  
1.423  
1.404  
1.389  
1.371  
1.353  
1.334  
1.316  
0.918  
0.900  
0.882  
-0.000



**1u**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





**1u**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

165.657

155.508

134.260

129.936

128.675

126.466

80.237

77.321

77.000

76.686

74.616

66.631

34.257

26.740

22.088

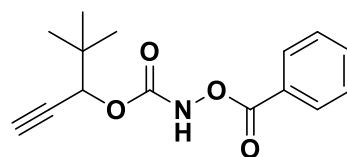
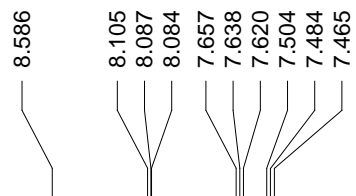
13.834

150

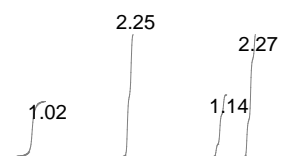
100

50

0 PPM



**1v**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



165.657

155.777

134.231

129.893

128.668

126.480

78.888

77.321

77.000

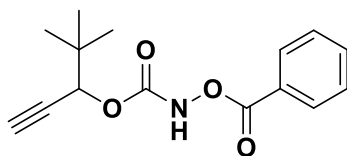
76.686

75.163

74.324

35.220

25.252

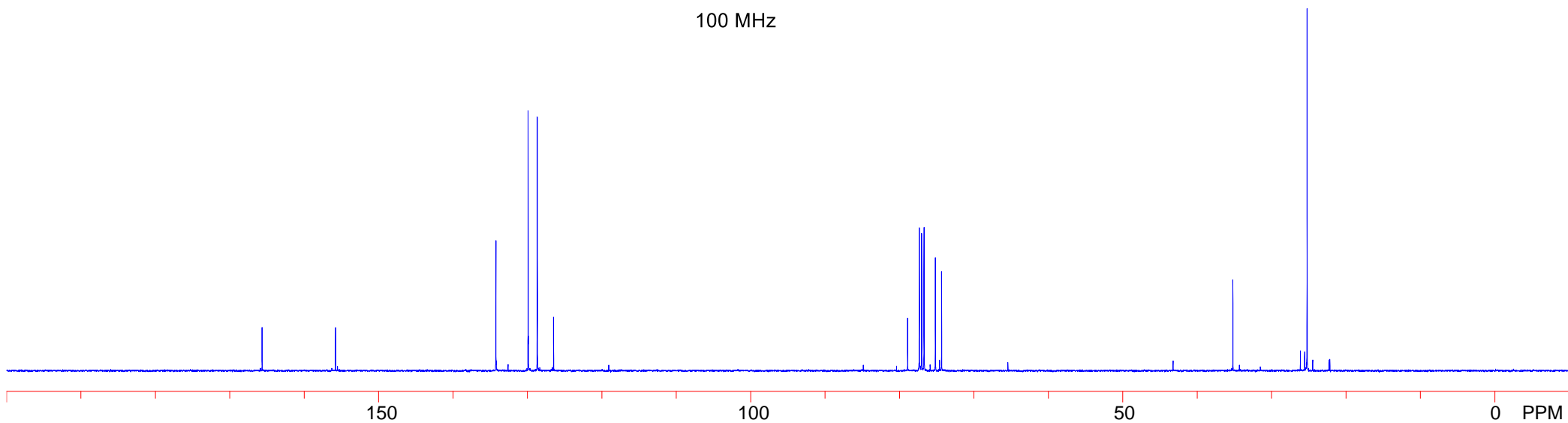


1v

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

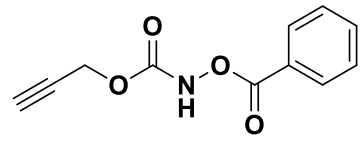


8.552  
8.108  
8.106  
8.092  
8.088  
8.085  
7.669  
7.666  
7.663  
7.658  
7.647  
7.632  
7.628  
7.625  
7.511  
7.491  
7.476  
7.472  
7.267

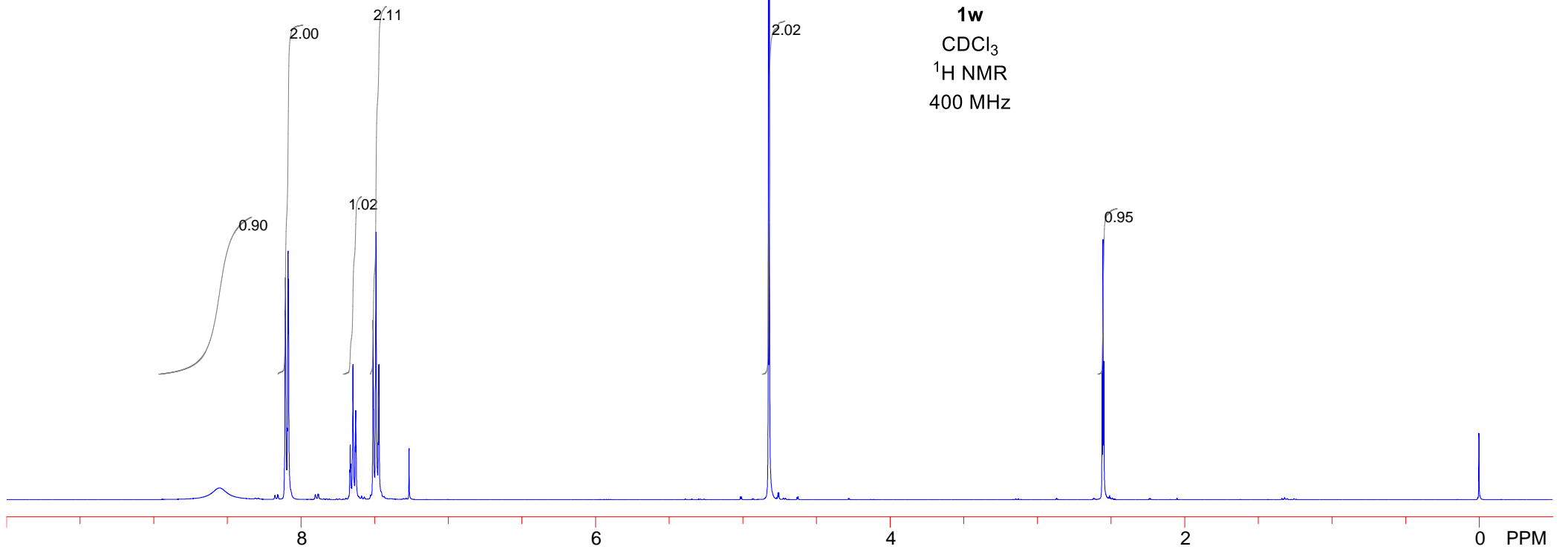
4.826  
4.820

2.559  
2.553  
2.547

-0.000



**1w**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz





165.516

155.483

134.279

129.912

128.650

126.288

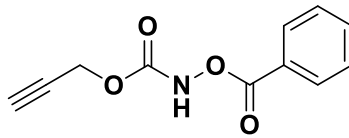
77.260

76.946

76.625

75.860

54.000

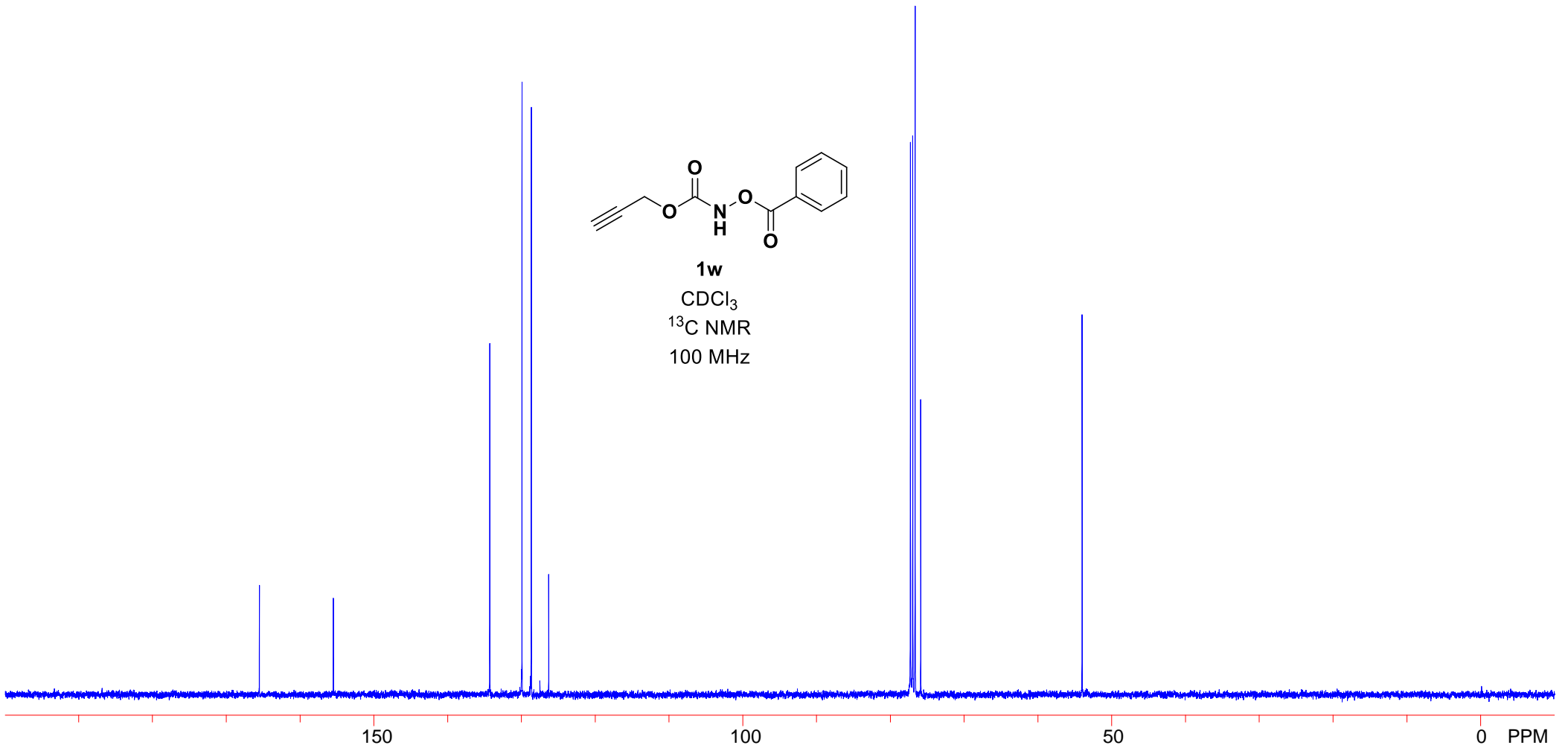


**1w**

CDCl<sub>3</sub>

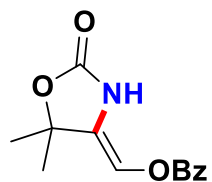
<sup>13</sup>C NMR

100 MHz



8.986  
8.218  
8.198  
7.635  
7.616  
7.598  
7.524  
7.504  
7.485  
7.262  
6.953

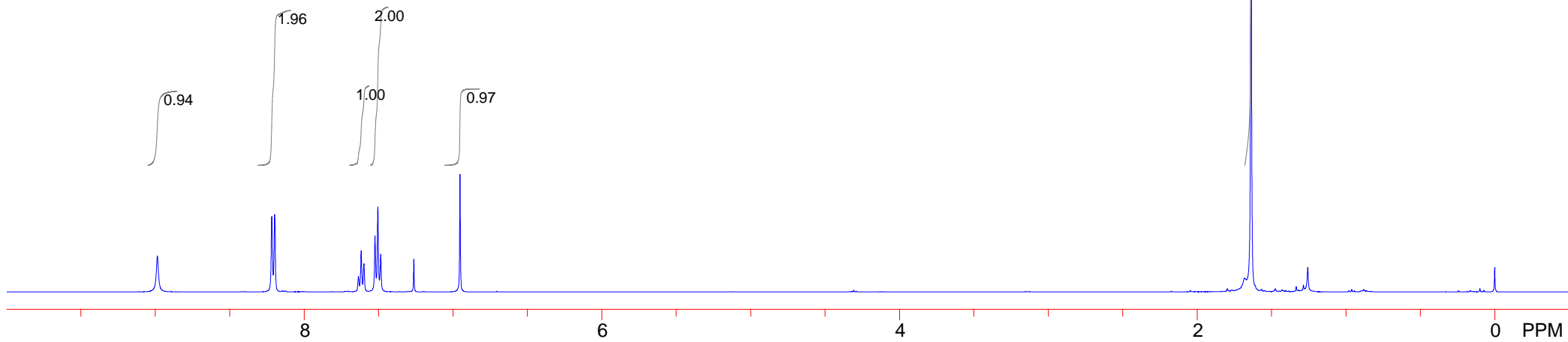
1.638  
0.000



**2a**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

0.94  
1.96  
2.00  
1.00  
0.97

6.05



162.877

156.348

133.716

130.576

130.172

128.655

128.412

111.204

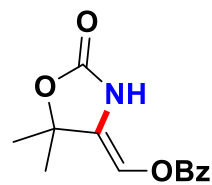
82.912

77.323

77.006

76.689

28.275

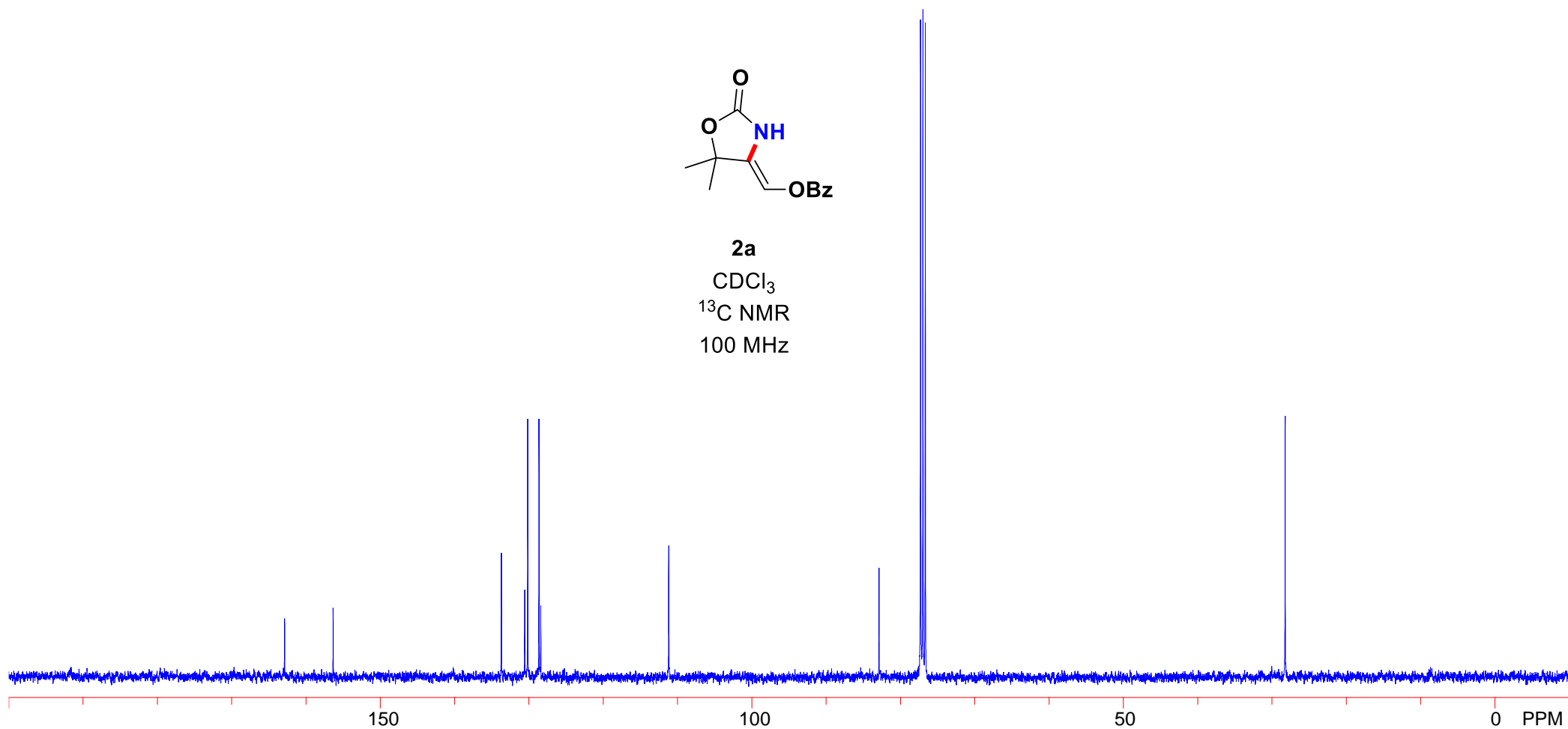


**2a**

CDCl<sub>3</sub>

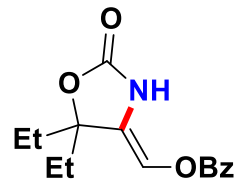
<sup>13</sup>C NMR

100 MHz

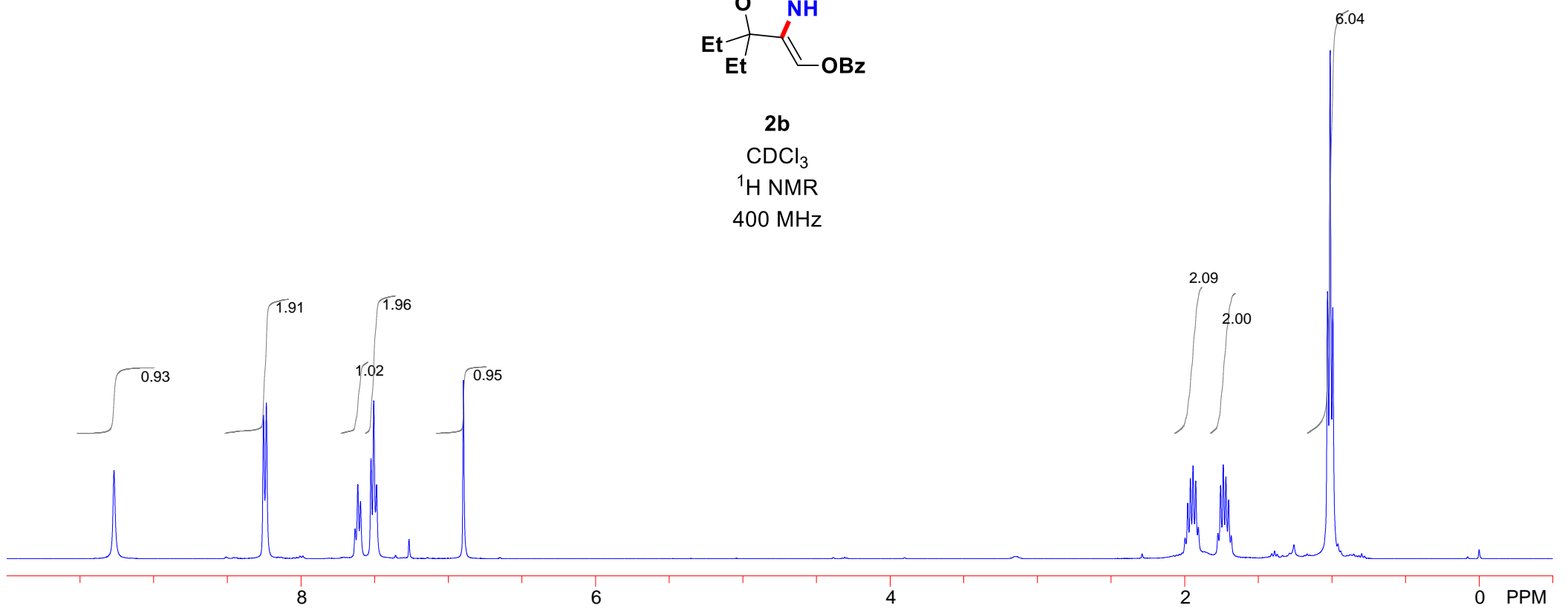


9.269  
8.254  
8.235  
7.632  
7.614  
7.595  
7.525  
7.506  
7.487  
7.266  
6.897

1.997  
1.979  
1.961  
1.943  
1.925  
1.907  
1.774  
1.756  
1.738  
1.720  
1.702  
1.683  
1.030  
1.012  
0.994  
-0.000



**2b**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



162.893

157.061

133.645

130.195

128.617

128.485

127.343

111.524

88.747

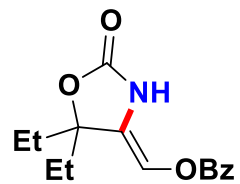
77.317

77.000

76.683

32.642

7.177

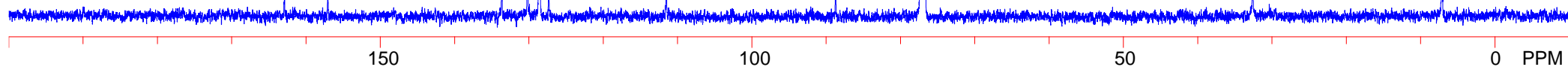


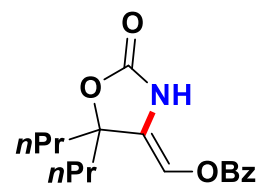
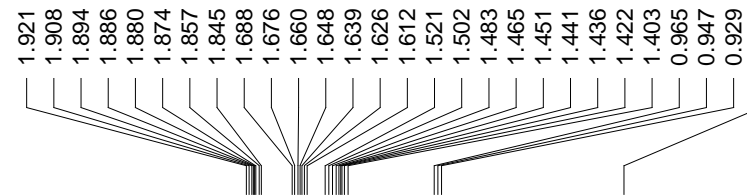
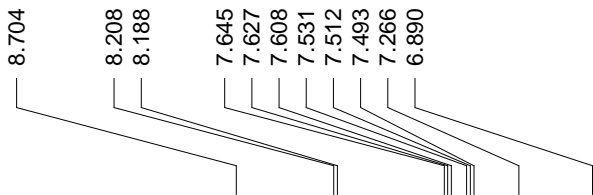
**2b**

CDCl<sub>3</sub>

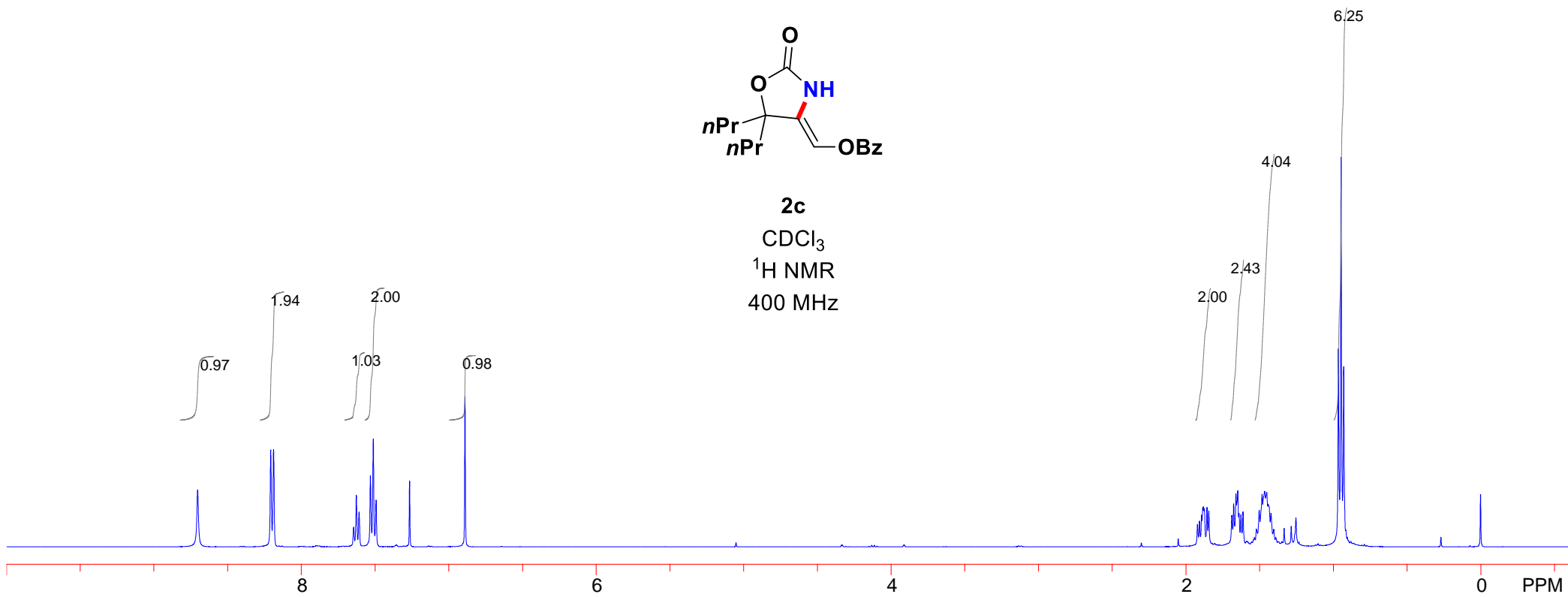
<sup>13</sup>C NMR

100 MHz





**2c**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



162.872

156.718

133.684

130.133

128.631

128.493

128.026

111.460

88.068

77.314

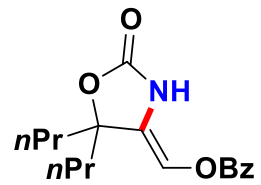
77.000

76.679

42.198

16.094

13.936



2c

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

150

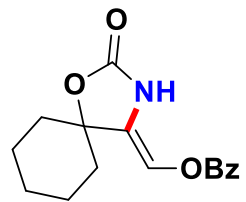
100

50

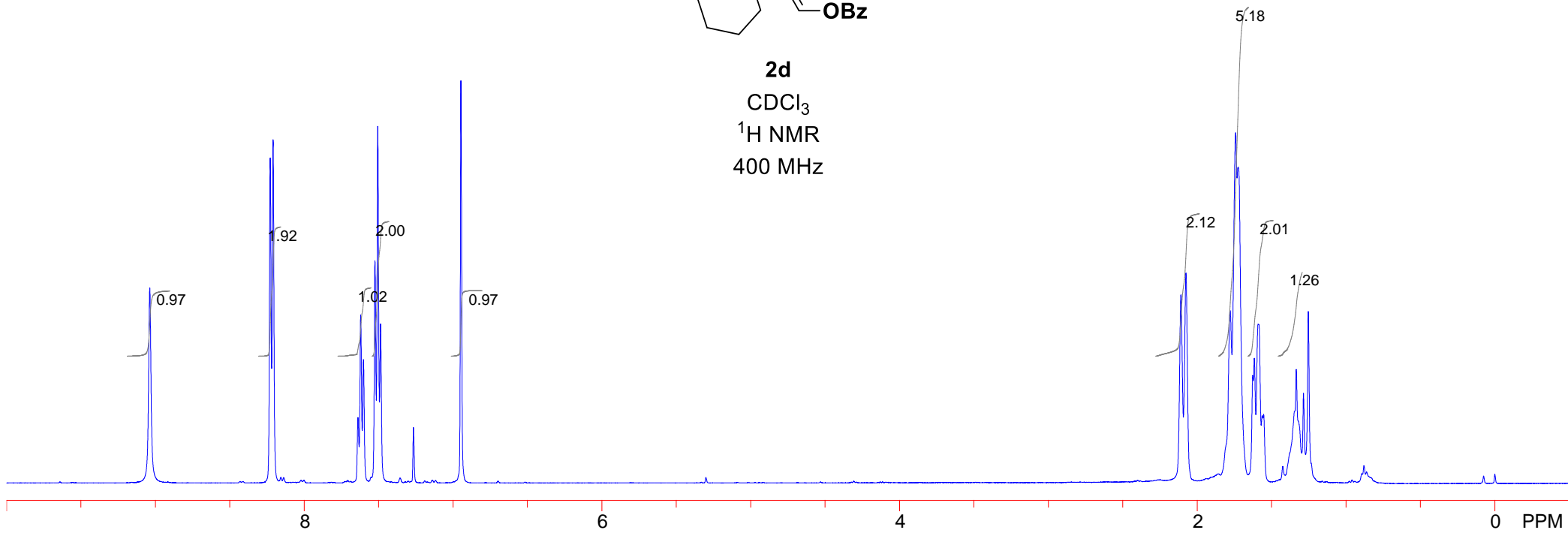
0 PPM

9.037  
8.228  
8.208  
7.638  
7.620  
7.601  
7.524  
7.506  
7.487  
7.266  
6.947

2.109  
2.075  
1.778  
1.741  
1.724  
1.626  
1.616  
1.590  
1.562  
1.553  
1.334  
1.285  
1.253  
-0.000

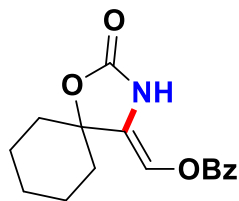


**2d**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

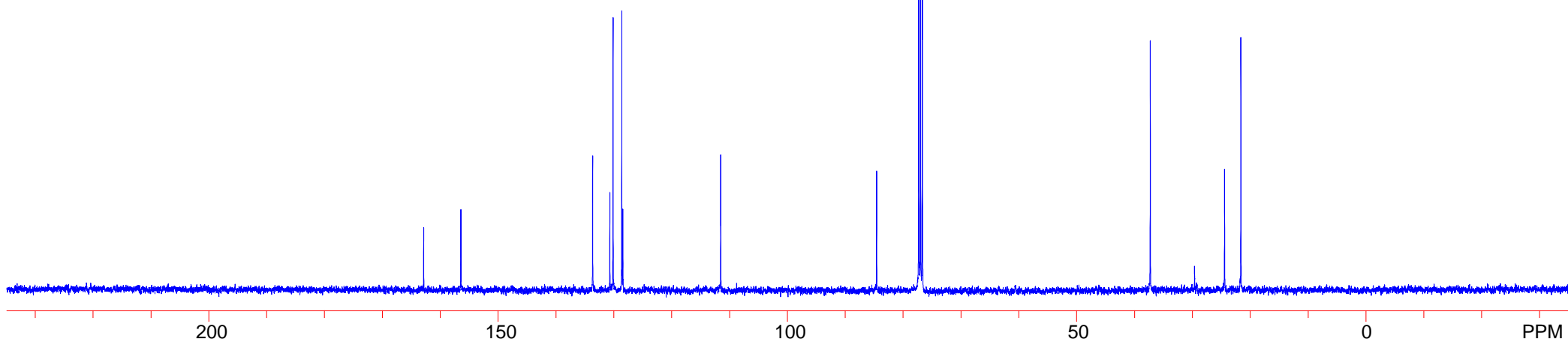




162.871  
156.441  
133.669  
130.693  
130.141  
128.624  
128.463  
111.540  
84.600  
77.317  
77.000  
76.683  
37.304  
24.469  
21.646



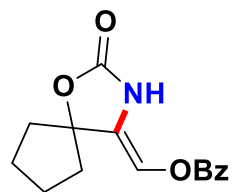
**2d**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz



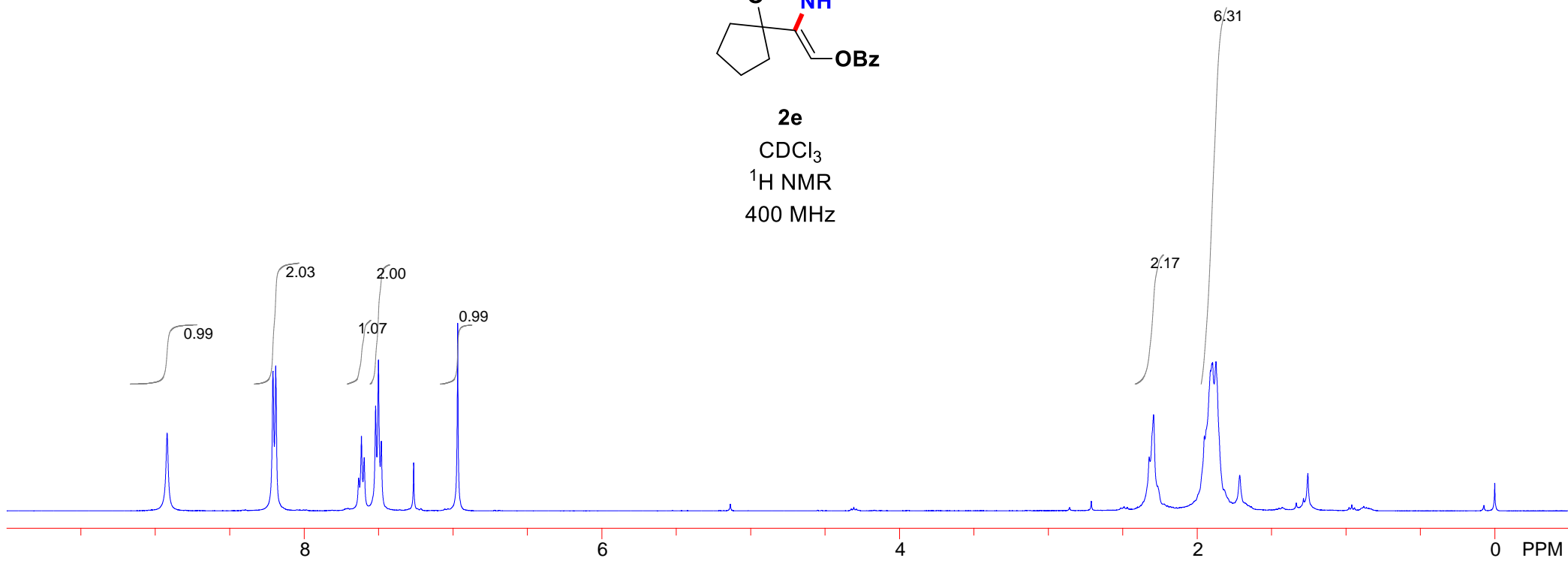
8.921  
8.209  
8.191  
7.633  
7.615  
7.596  
7.520  
7.500  
7.482  
7.264  
6.968

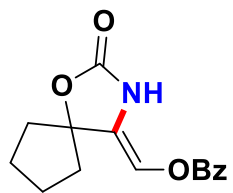
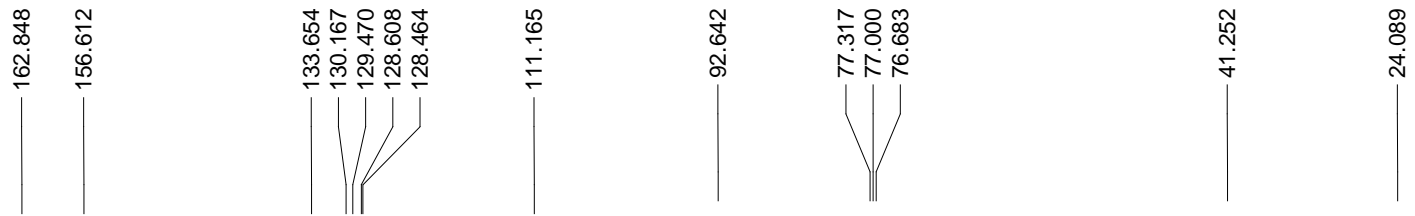
2.322  
2.293  
2.264  
1.951  
1.911  
1.901  
1.896  
1.873

-0.000

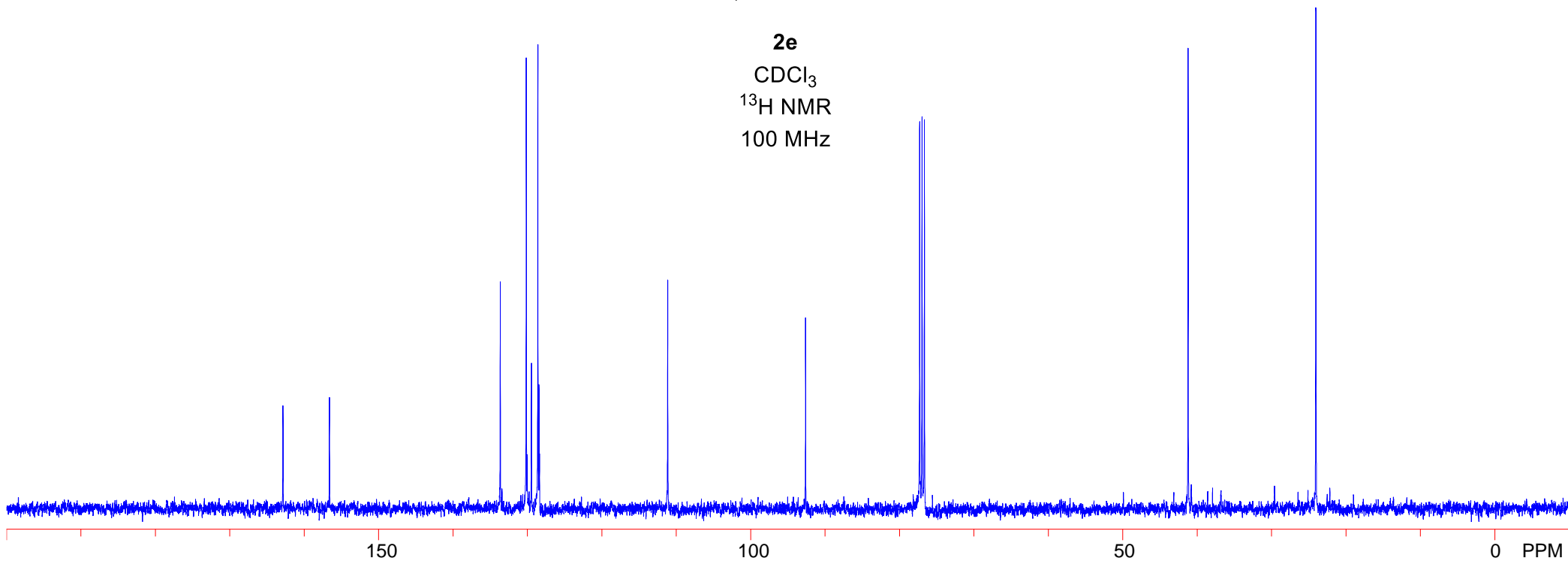


**2e**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



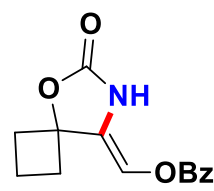


**2e**  
CDCl<sub>3</sub>  
 $^{13}\text{H}$  NMR  
100 MHz

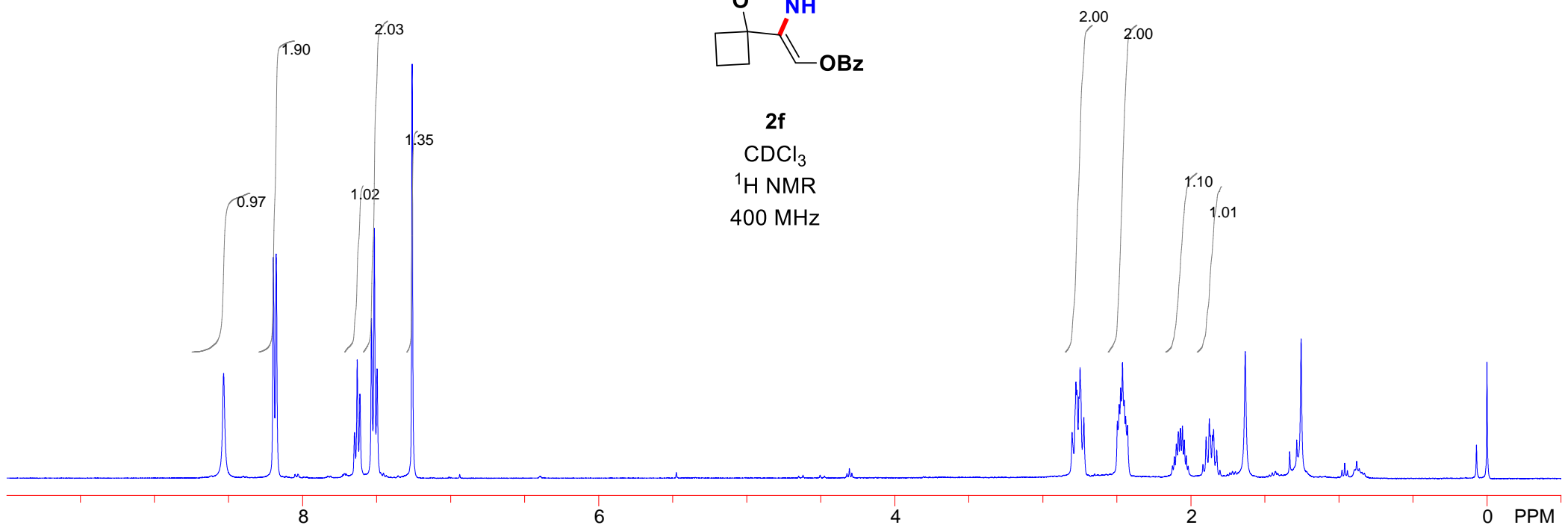


8.533  
8.197  
8.178  
7.649  
7.631  
7.612  
7.534  
7.515  
7.496  
7.260

2.802  
2.776  
2.769  
2.756  
2.748  
2.722  
2.497  
2.484  
2.475  
2.462  
2.450  
2.440  
2.428  
2.124  
2.111  
2.098  
2.085  
2.070  
2.057  
2.044  
2.031  
2.018  
1.918  
1.897  
1.875  
1.869  
1.854  
1.846  
1.832  
1.825  
-0.000



**2f**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



162.827

156.298

133.769

130.418

130.175

128.662

128.439

111.598

84.094

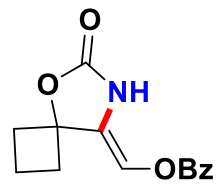
77.317

77.000

76.683

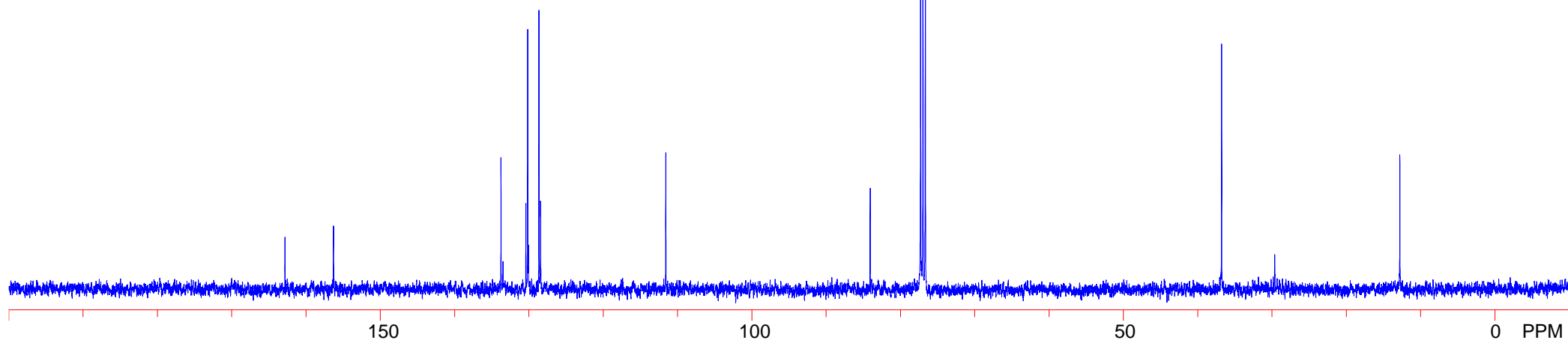
36.817

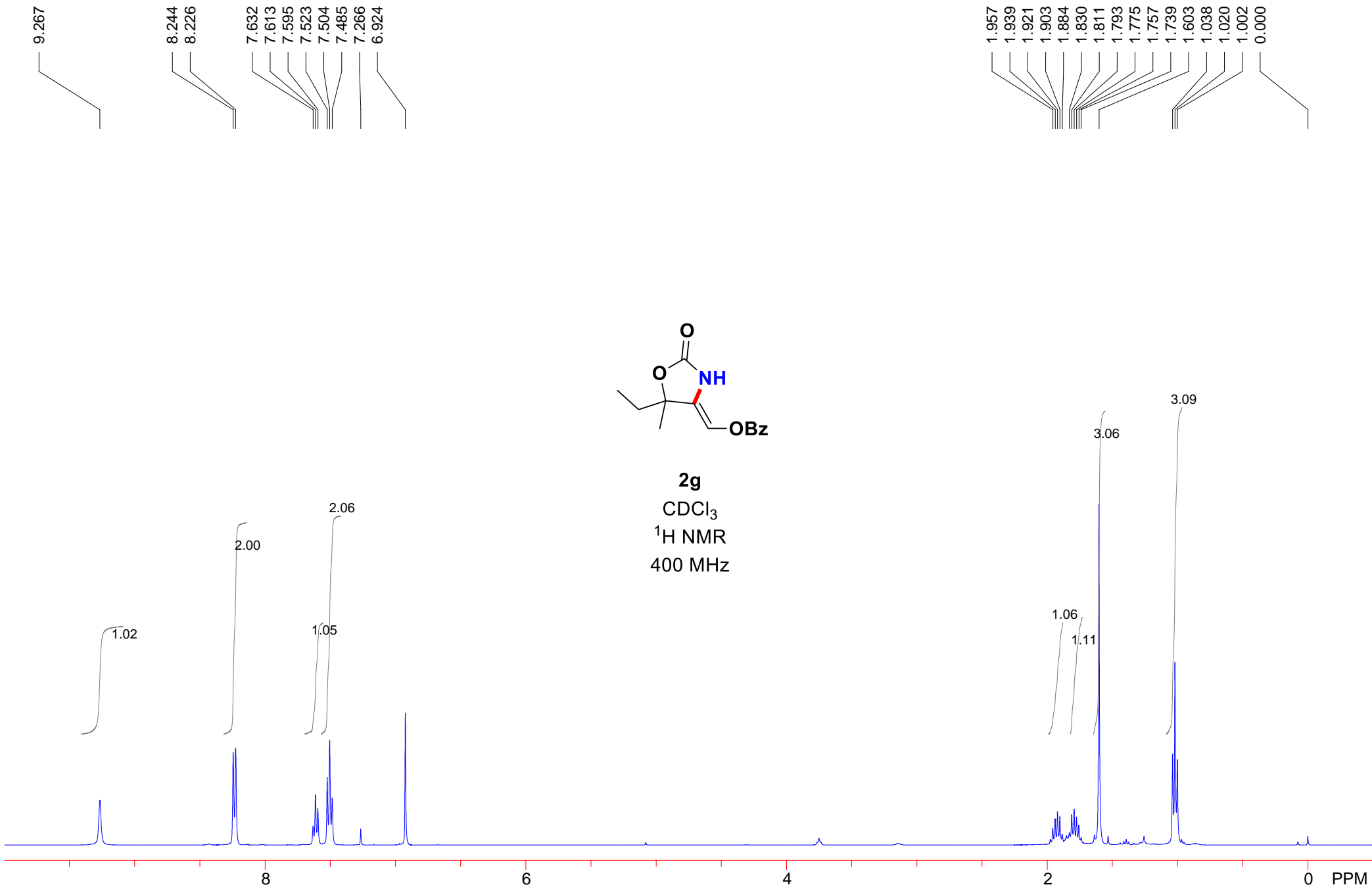
12.841



**2f**

CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz





162.889  
156.694

133.662  
130.183  
129.153  
128.625  
128.460

111.375

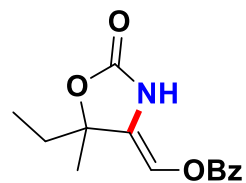
85.689

77.317  
77.000  
76.683

33.989

26.772

7.462

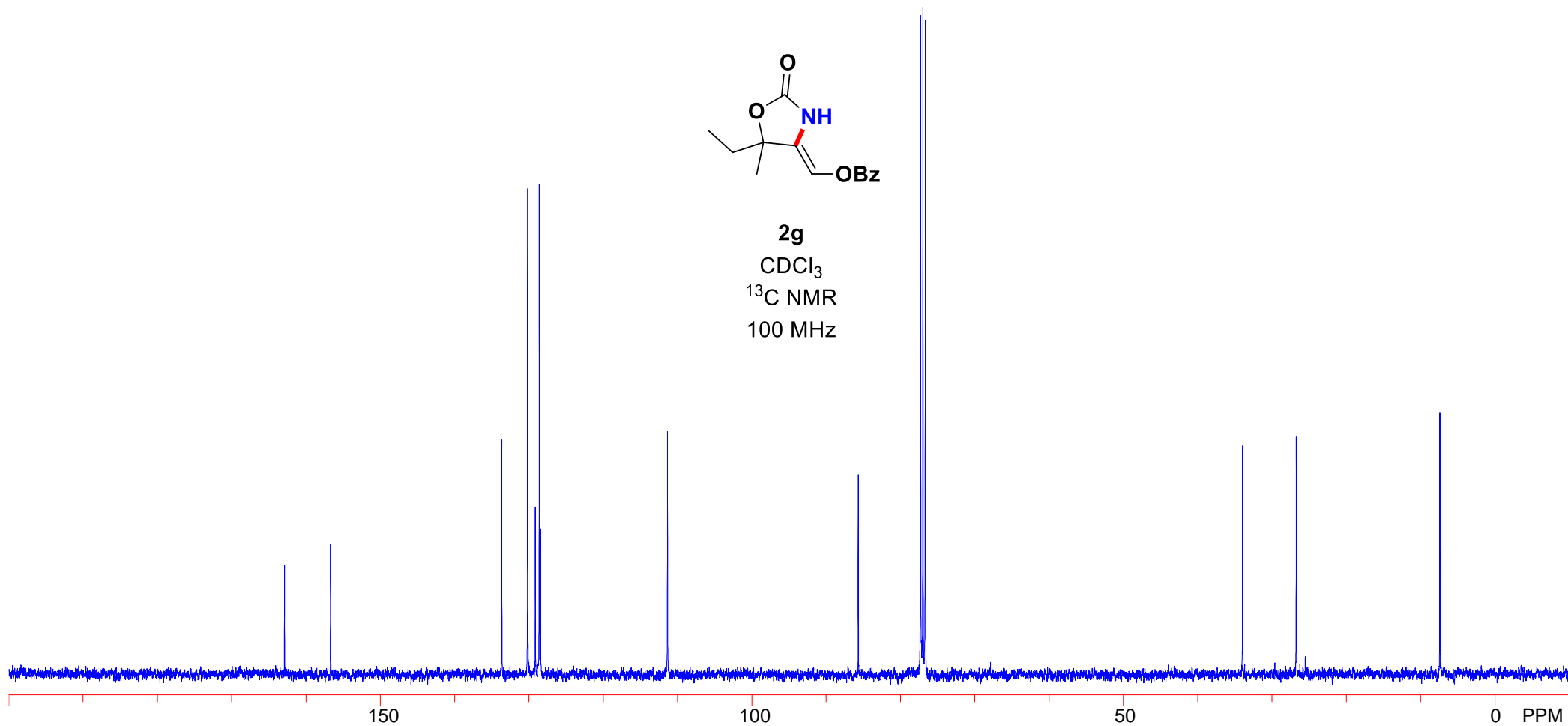


**2g**

CDCl<sub>3</sub>

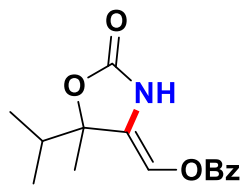
<sup>13</sup>C NMR

100 MHz

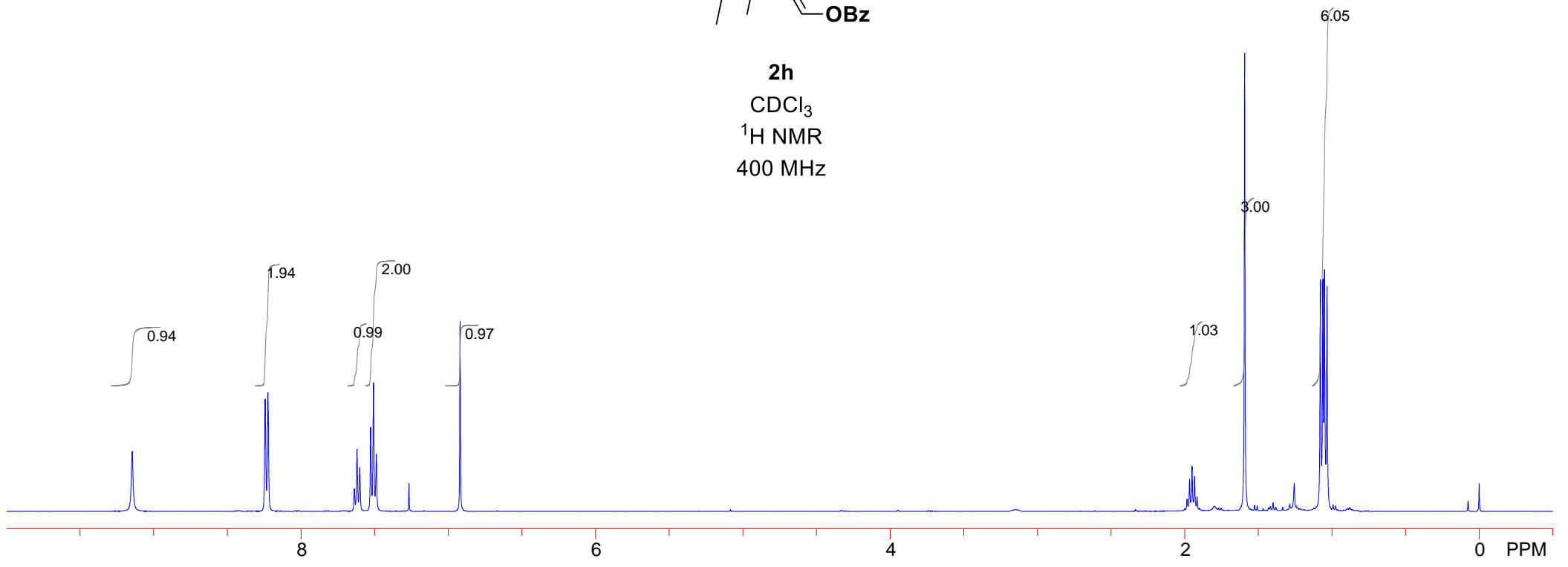


9.146  
 8.243  
 8.224  
 7.637  
 7.619  
 7.601  
 7.527  
 7.507  
 7.488  
 7.267  
 6.920

1.984  
 1.967  
 1.950  
 1.933  
 1.916  
 1.592  
 1.078  
 1.061  
 1.050  
 1.033  
 -0.000



**2h**  
 CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
 400 MHz





162.889

156.744

133.674

130.171

129.004

128.625

128.444

111.590

87.951

77.317

77.000

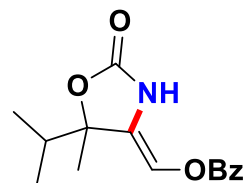
76.683

37.464

24.975

16.398

16.229



**2h**

CDCl<sub>3</sub>

<sup>13</sup>C NMR

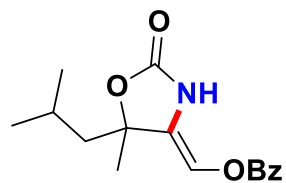
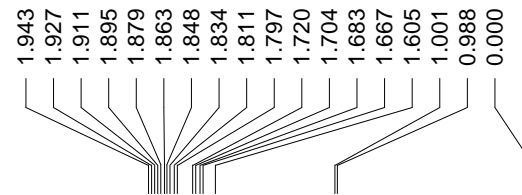
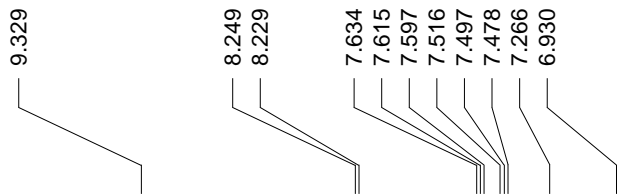
100 MHz

150

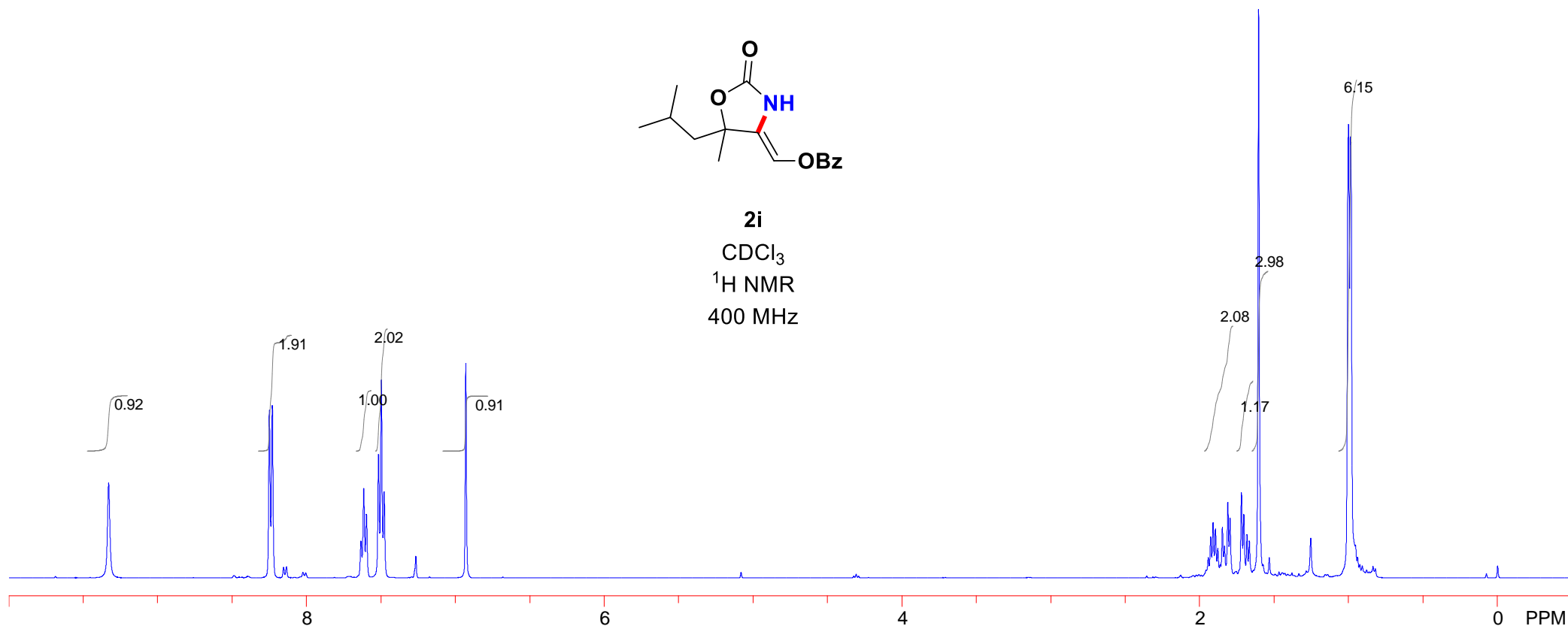
100

50

0 PPM



**2i**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



162.863  
156.722

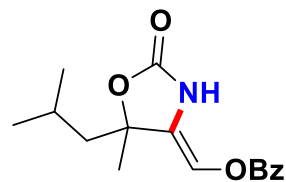
133.690  
130.207  
128.608

111.503

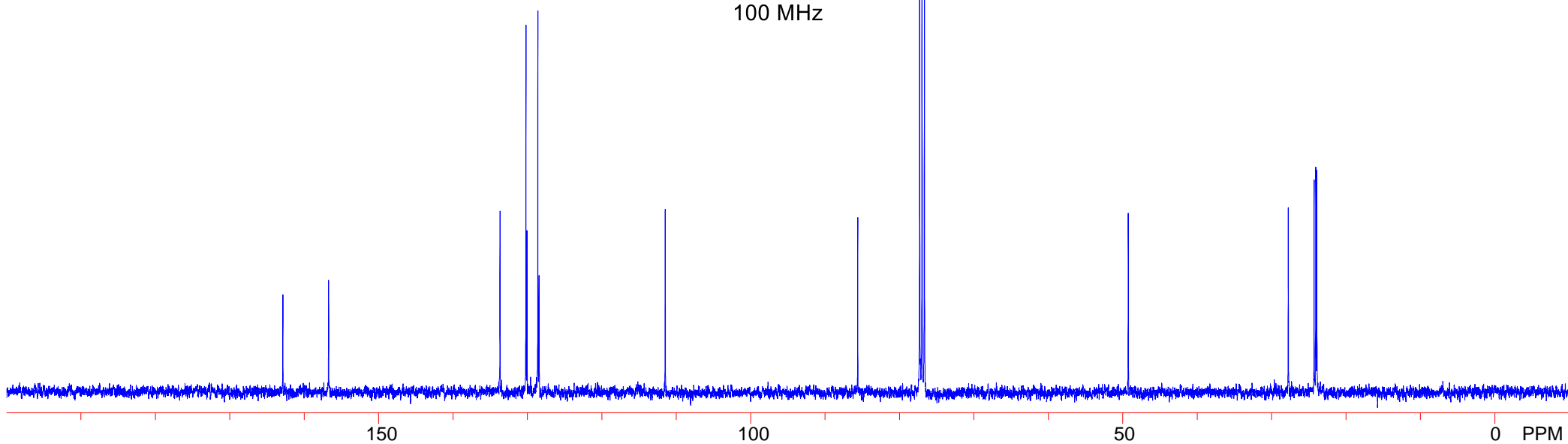
85.631  
77.317  
77.000  
76.683

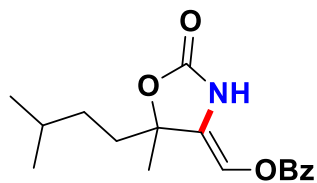
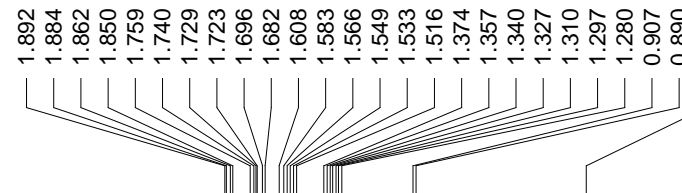
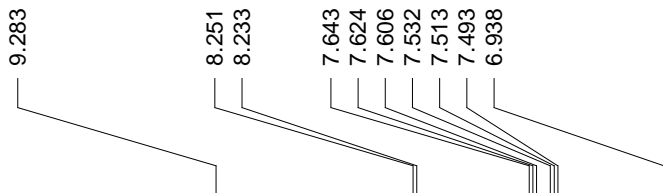
49.294

27.795  
24.329  
24.094  
23.958

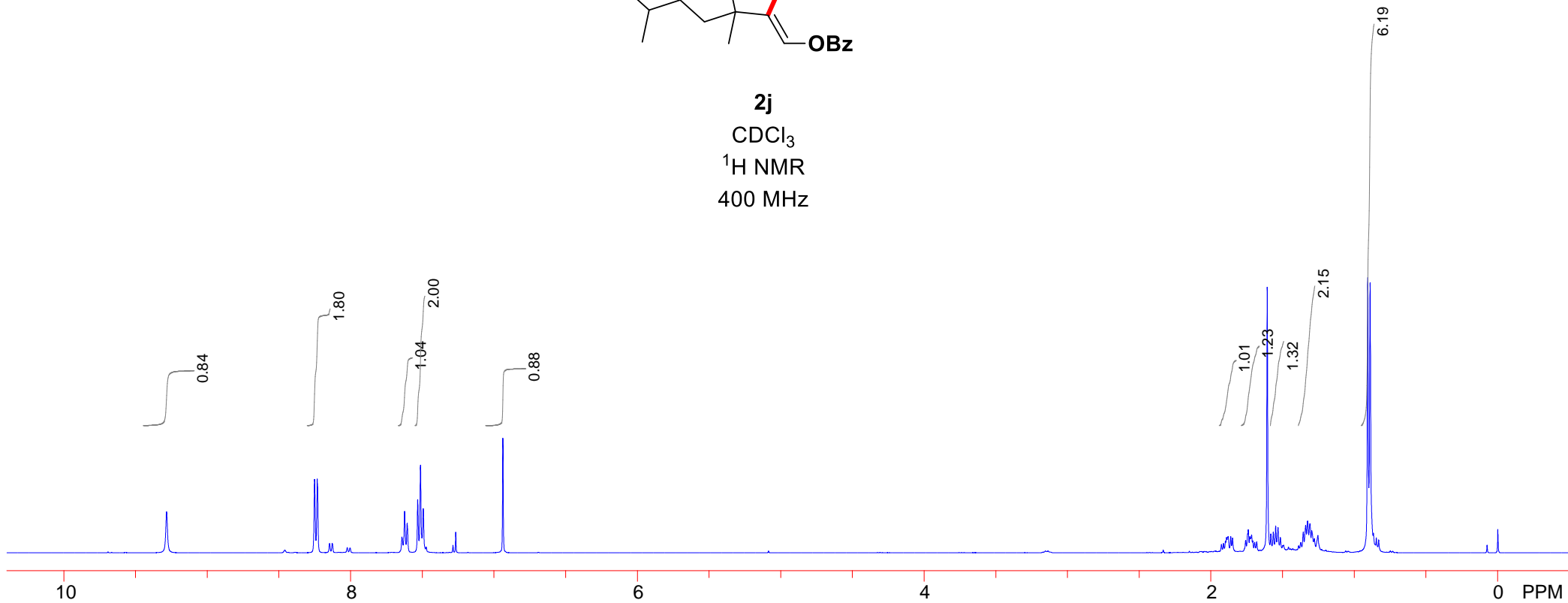


**2i**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz





**2j**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



162.840  
156.772

133.703  
130.175  
129.416  
128.637  
128.444

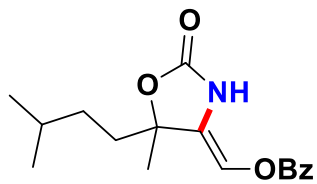
111.380

85.549

77.317  
77.000  
76.683

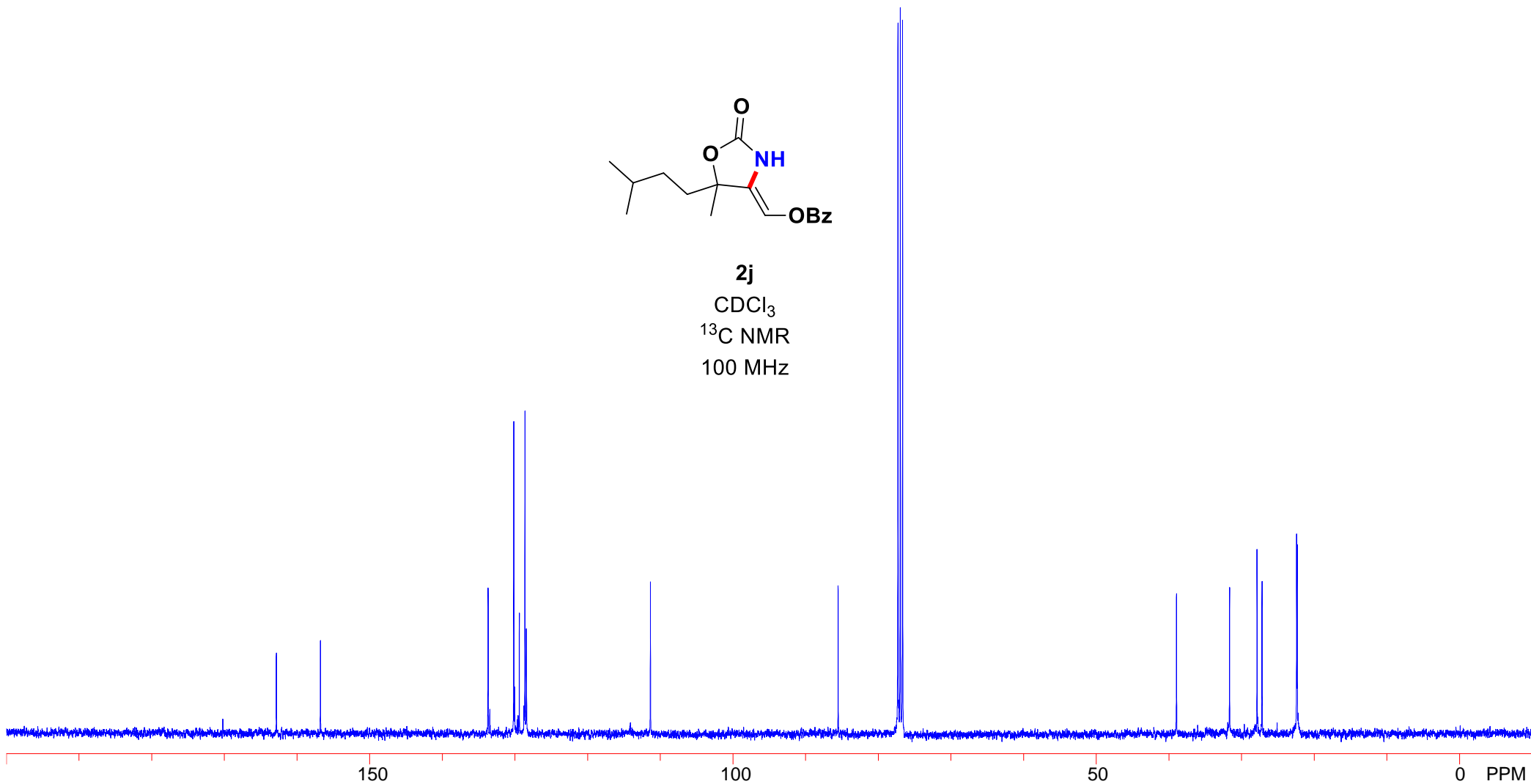
38.993

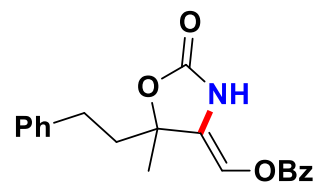
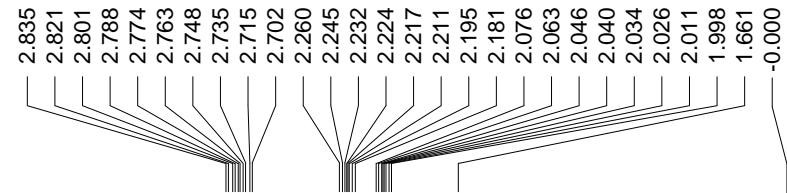
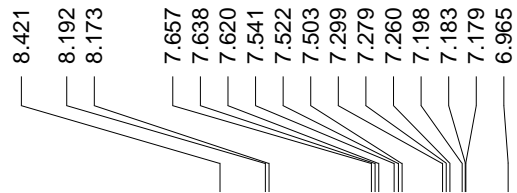
31.690  
27.910  
27.222  
22.461  
22.383



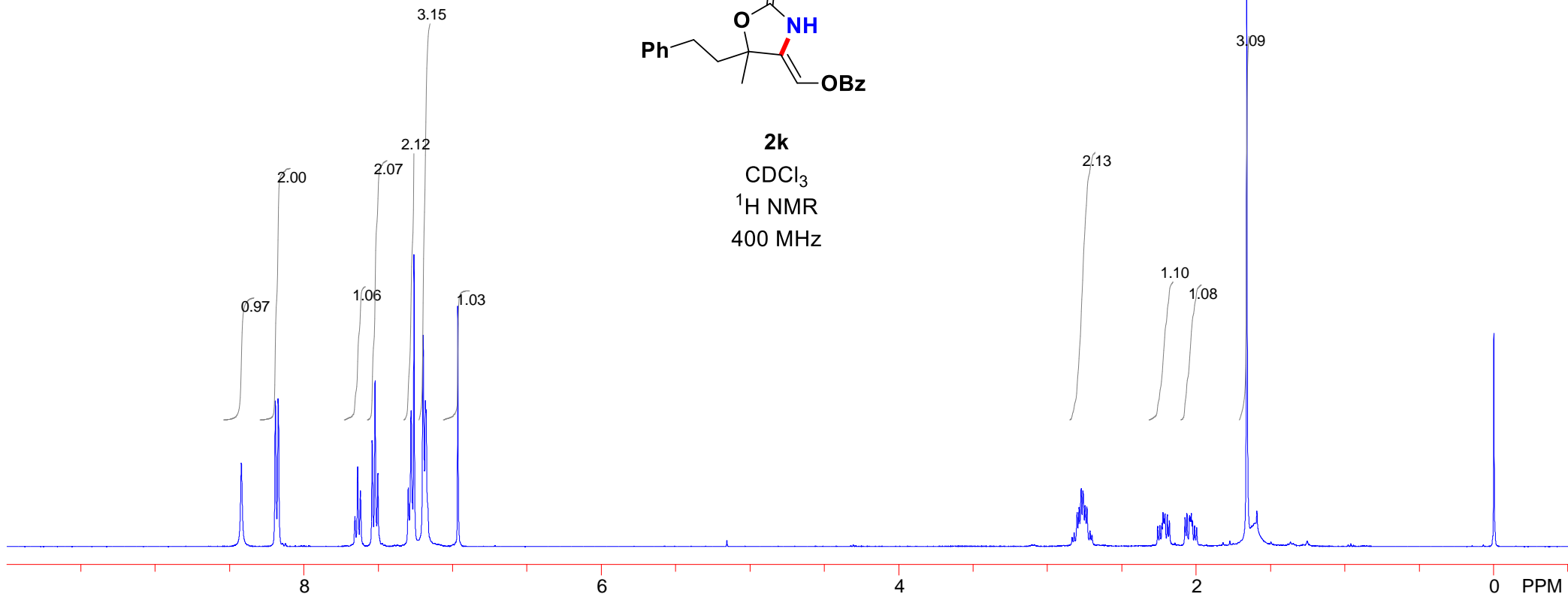
2j

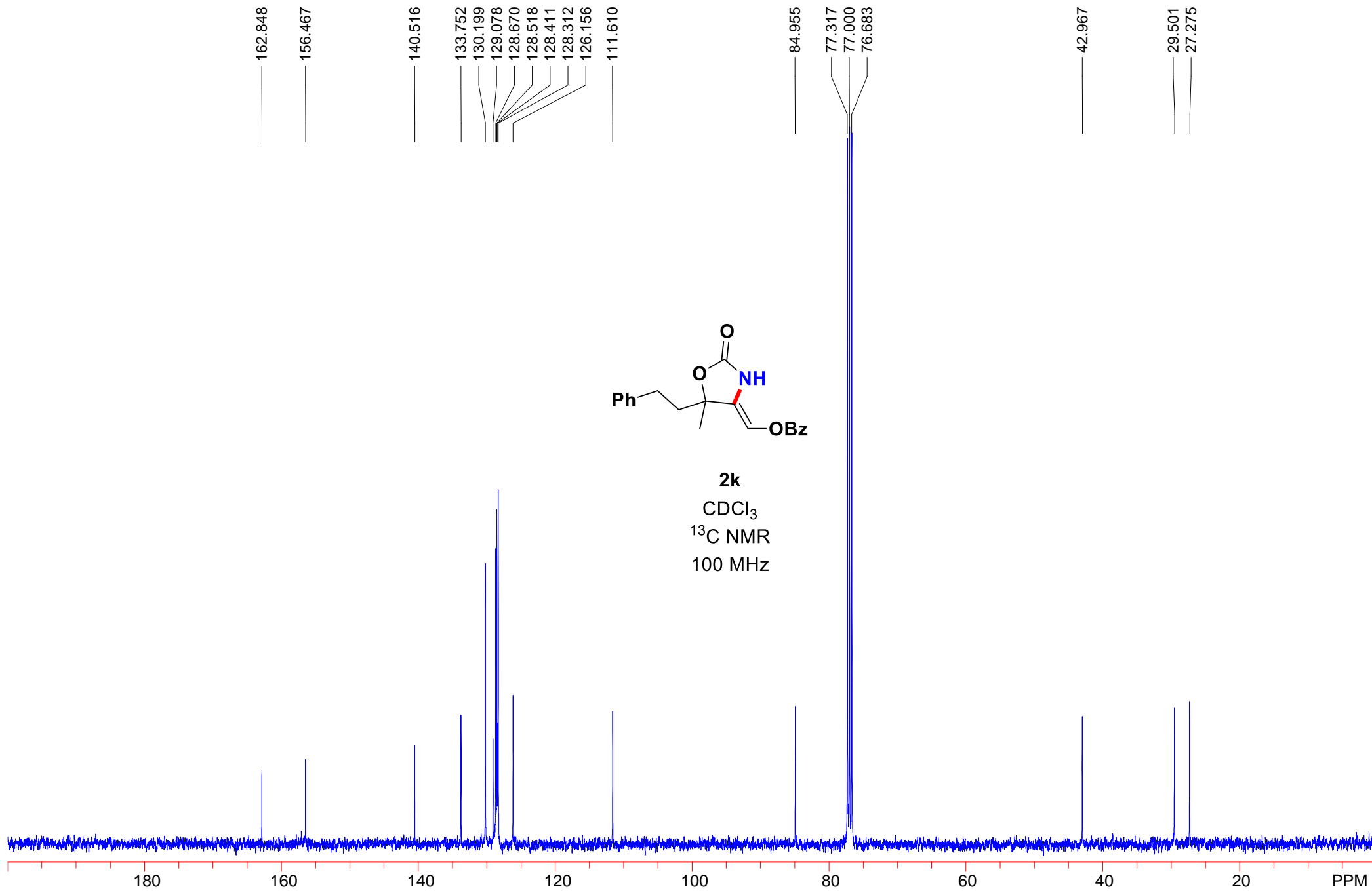
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz





**2k**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

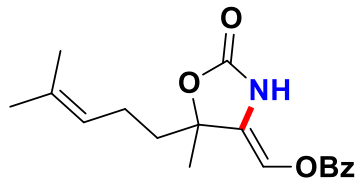




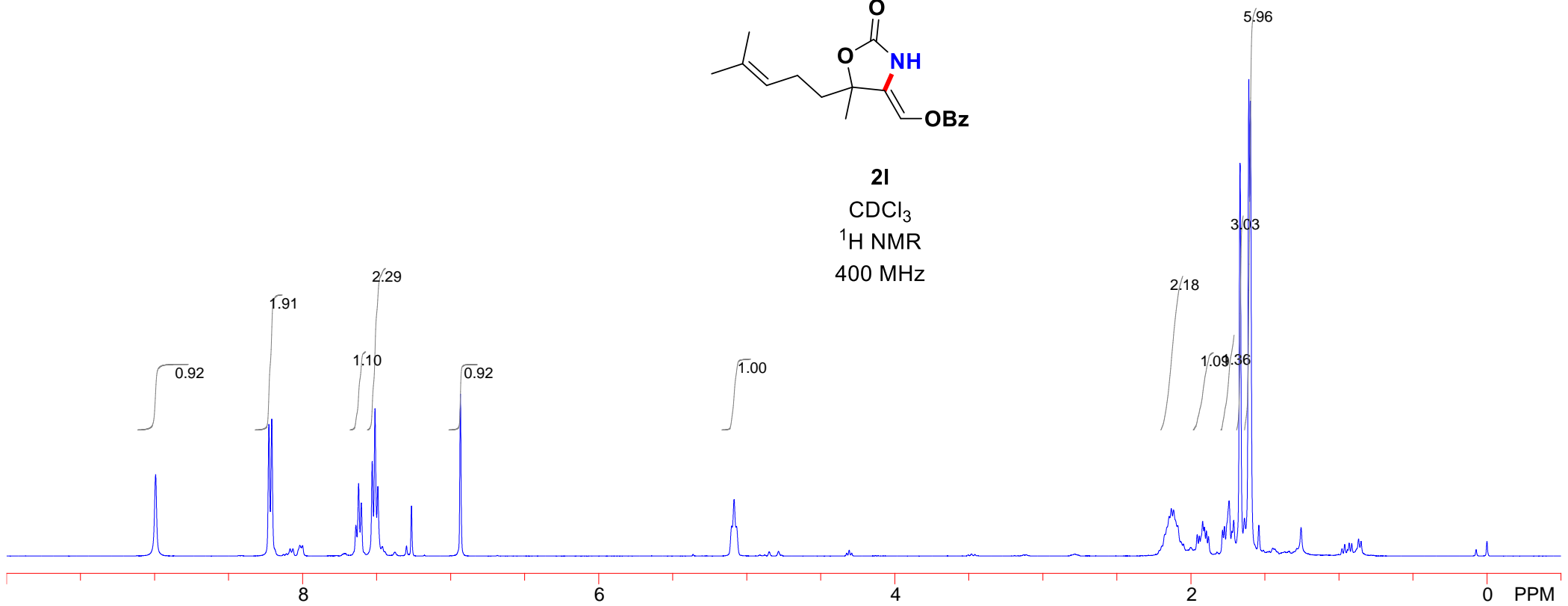
8.993  
8.227  
8.208  
7.639  
7.621  
7.603  
7.530  
7.511  
7.491  
7.264  
6.934

5.102  
5.085  
5.069

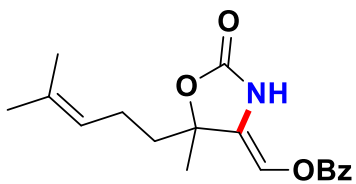
2.176  
2.147  
2.132  
2.117  
2.088  
2.068  
2.050  
1.957  
1.943  
1.921  
1.908  
1.895  
1.881  
1.786  
1.773  
1.742  
1.723  
1.711  
1.668  
1.638  
1.609  
1.598  
-0.000



**2I**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz







**2I**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

162.822

156.381

133.706

132.820

130.150

129.268

128.646

128.427

122.427

111.407

85.201

77.317

77.000

76.683

40.924

27.267

25.639

21.946

17.673

150

100

50

0 PPM

8.985

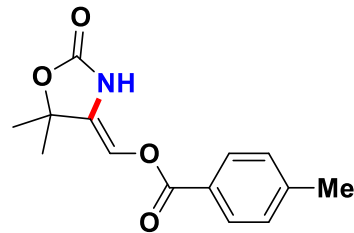
8.101  
8.081

7.306  
7.286  
7.265  
6.943

2.431

1.633

0.000



2m

CDCl<sub>3</sub>

<sup>1</sup>H NMR

400 MHz

0.99

2.05

2.07

1.00

3.16

6.04

8

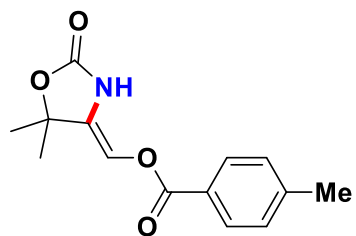
6

4

2

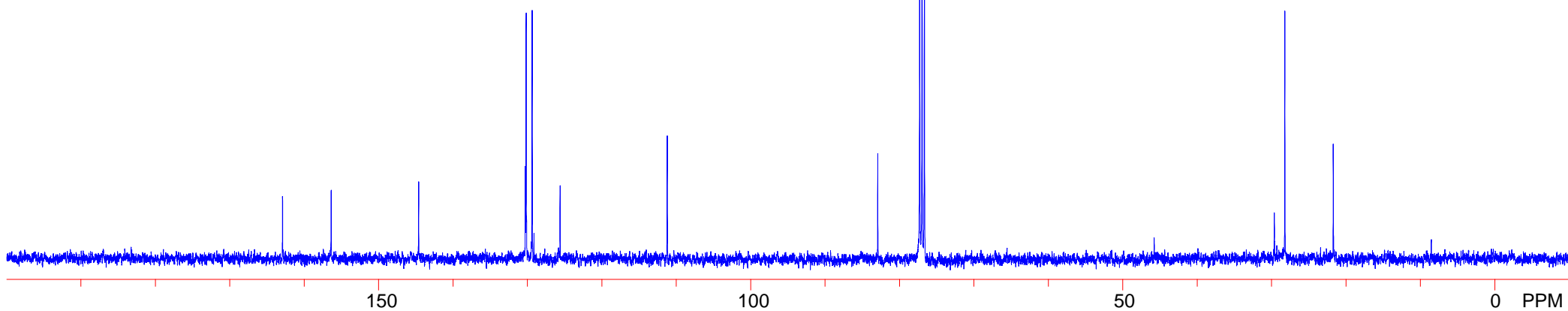
0 PPM

162.904  
156.371  
144.625  
130.289  
130.186  
129.374  
125.619  
111.231  
82.948  
77.317  
77.000  
76.683  
28.261  
21.749



2m

CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz



8.972

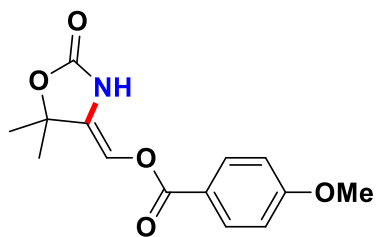
8.170  
8.148

7.266  
6.982  
6.961  
6.934

3.889

1.631

-0.000



2n

CDCl<sub>3</sub>

<sup>1</sup>H NMR

400 MHz

0.94

2.00

2.10

1.08

3.12

6.01

8

6

4

2

0 PPM

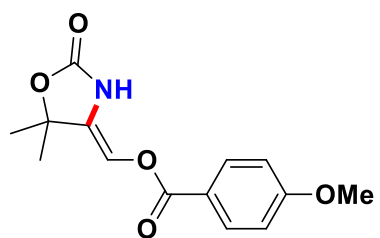
164.013  
162.583  
156.491

132.334  
130.116  
120.731  
113.930  
111.338

83.001  
77.317  
77.000  
76.683

55.489

28.257

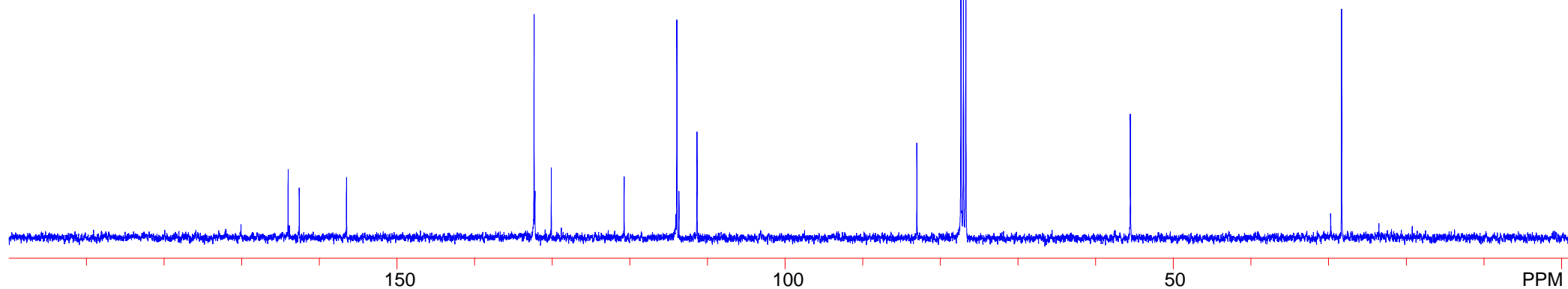


2n

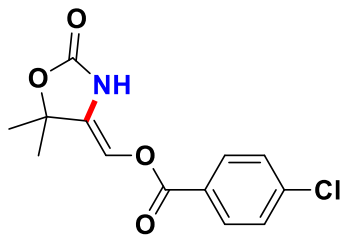
CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

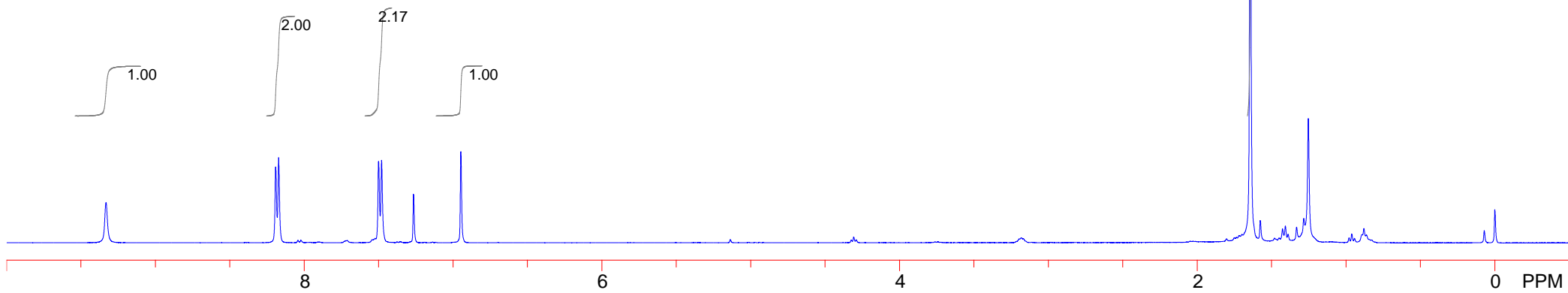


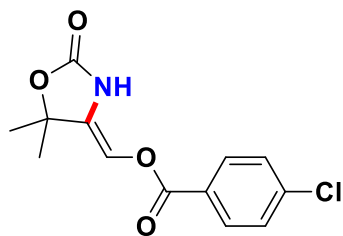
9.331  
8.192  
8.171  
7.500  
7.480  
7.266  
6.947  
1.644  
-0.000



**2o**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

6.11





2o

CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

162.010

156.524

140.284

131.596

130.788

129.036

126.823

111.123

83.055

77.317

77.000

76.683

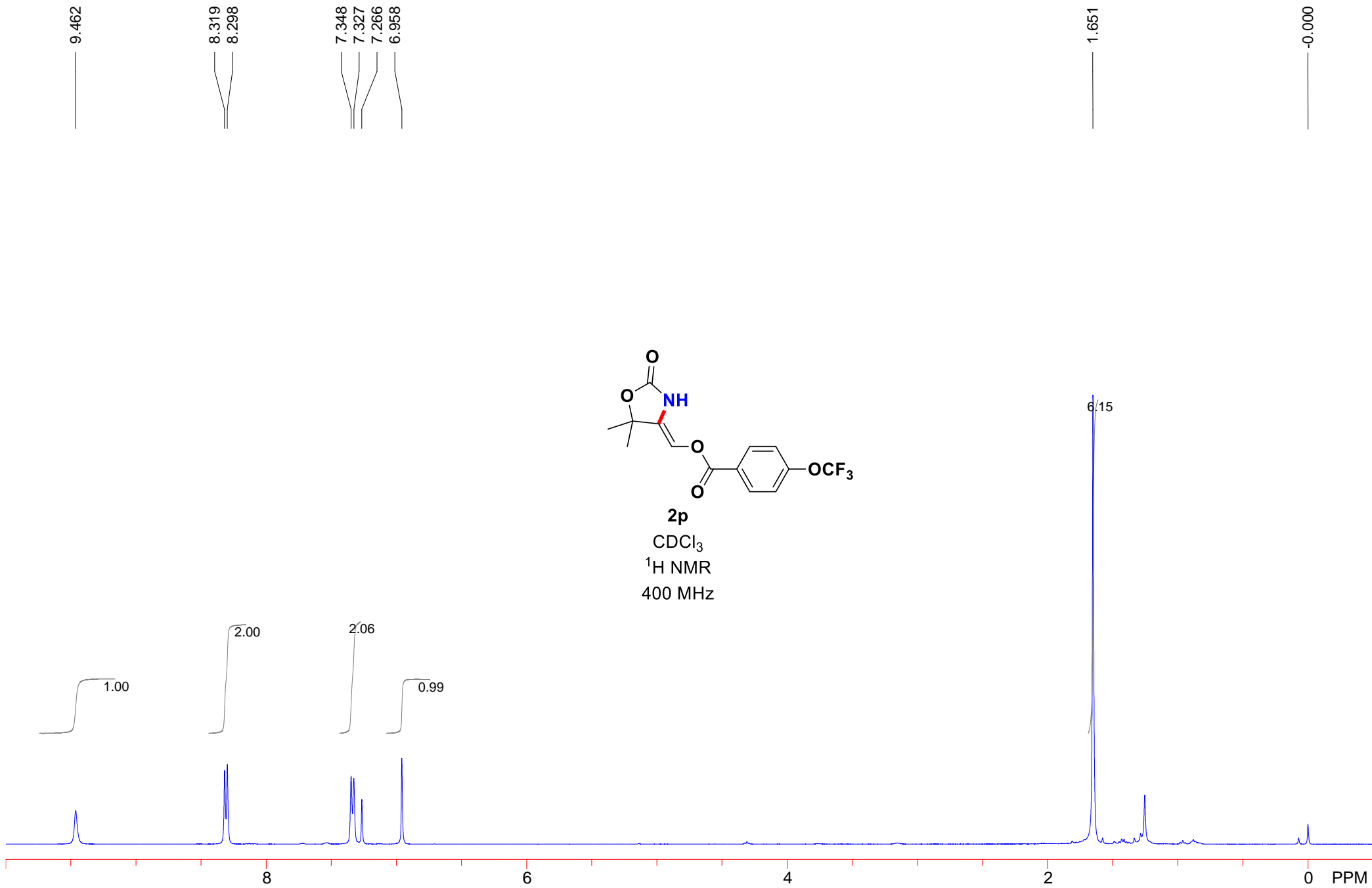
28.261

150

100

50

0 PPM



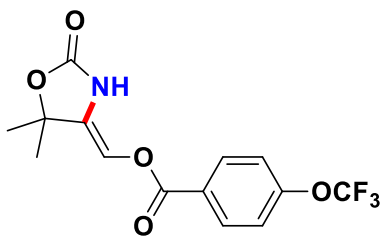


161.698  
156.601  
153.174

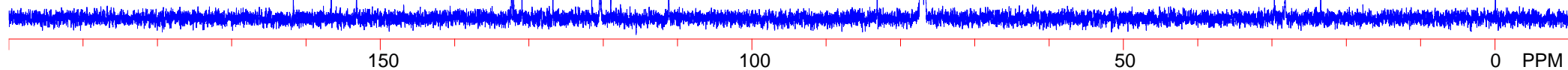
132.313  
130.921  
126.757  
124.110  
121.544  
120.435  
118.963  
116.389  
111.190

83.125  
77.321  
77.000  
76.686

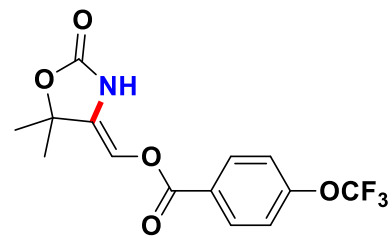
28.256



**2p**  
CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz

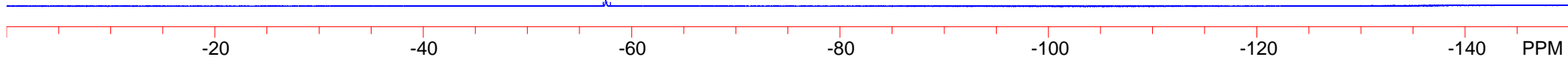


-57.514



**2p**

CDCl<sub>3</sub>  
<sup>19</sup>F NMR  
376 MHz



10.298

8.746

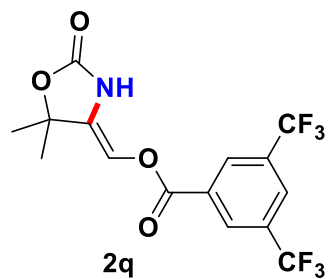
8.122

7.266

6.986

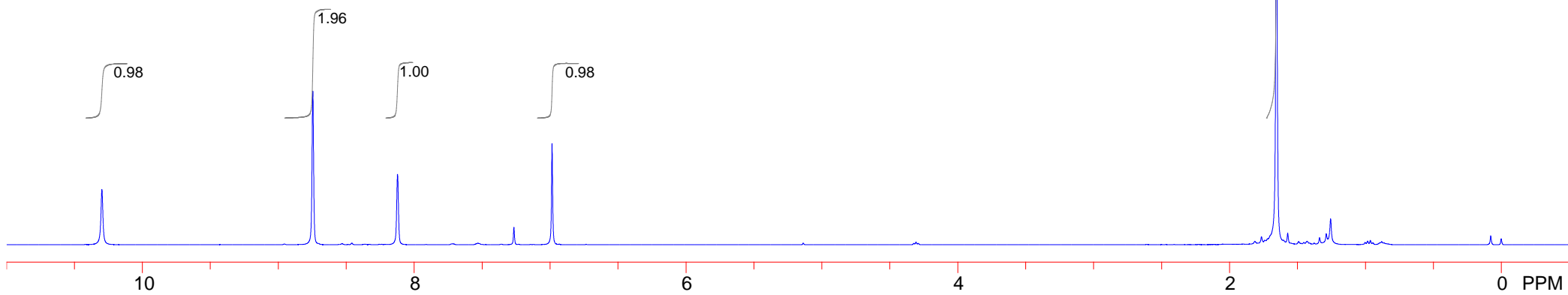
1.655

-0.000



CDCl<sub>3</sub>  
1H NMR  
400 MHz

6.07

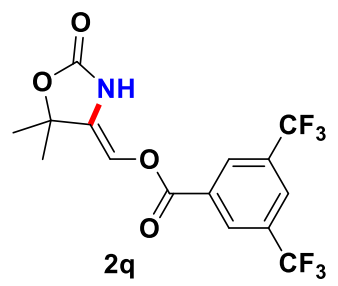


160.534  
157.225

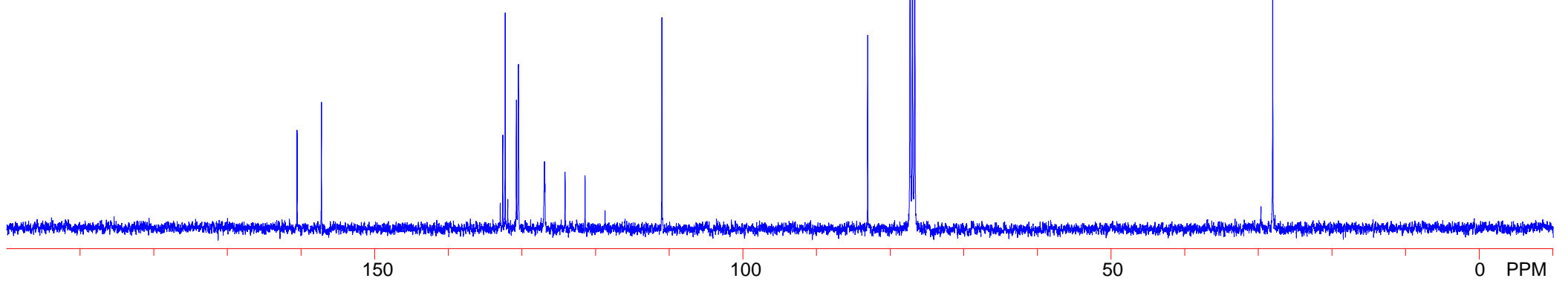
132.952  
132.614  
132.280  
131.934  
130.767  
130.483  
126.988  
126.951  
126.918  
126.872  
124.156  
121.440  
118.728  
111.008

83.071  
77.317  
77.000  
76.683

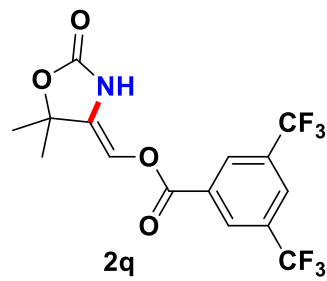
28.088



CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
100 MHz



-62.786



2q

CDCl<sub>3</sub>

<sup>19</sup>F NMR

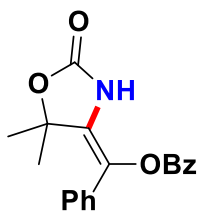
376 MHz



8.120  
8.100  
7.668  
7.651  
7.634  
7.532  
7.513  
7.494  
7.445  
7.425  
7.408  
7.389  
7.370  
7.309  
7.291  
7.274  
7.258

1.687

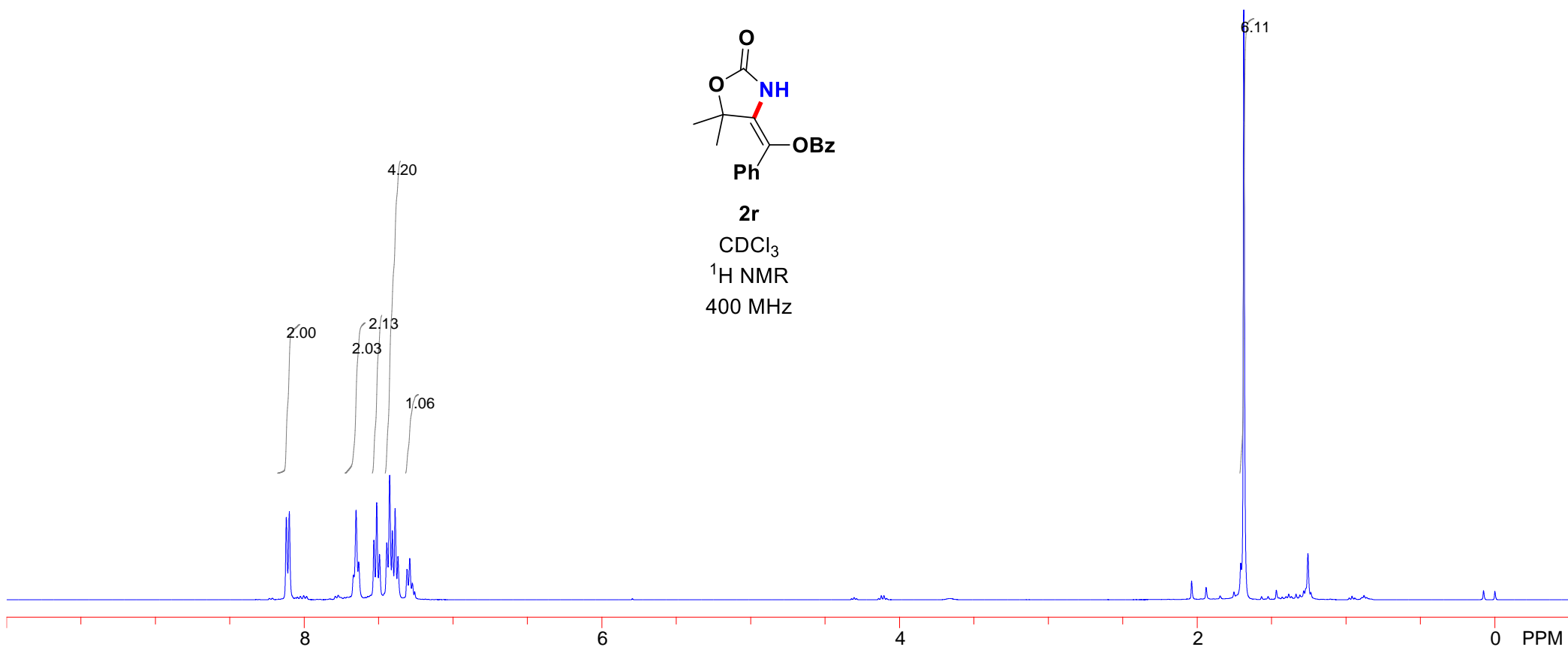
-0.000



**2r**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

2.00  
2.03  
2.13  
4.20  
1.06

6.11



165.155

155.469

134.003

133.615

132.630

129.980

129.135

128.838

128.707

128.476

126.098

123.963

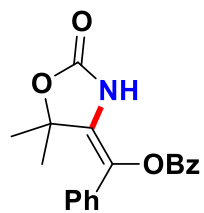
84.856

77.317

77.000

76.683

25.747

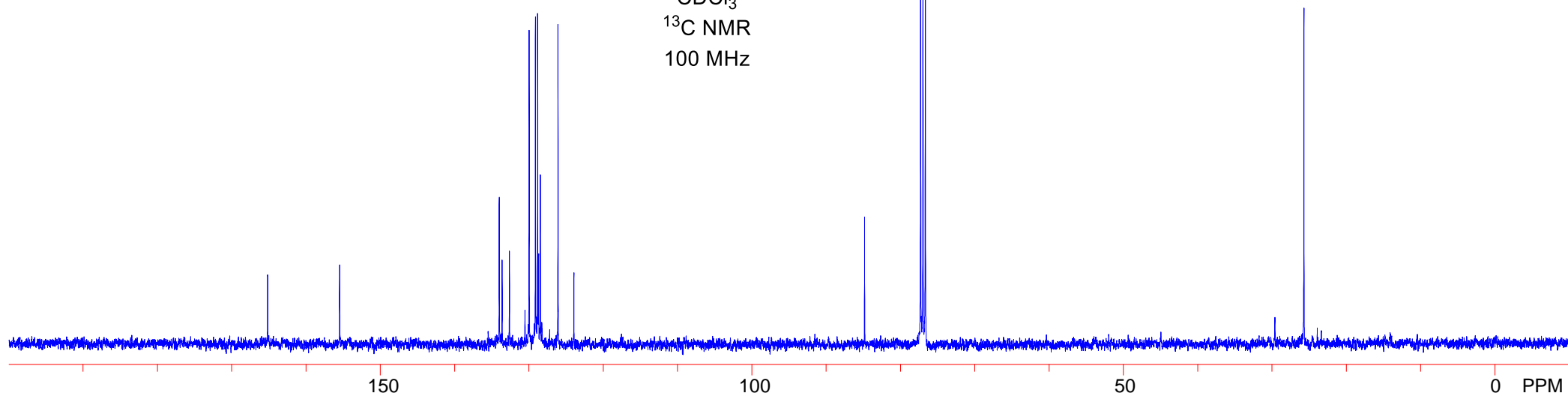


2r

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

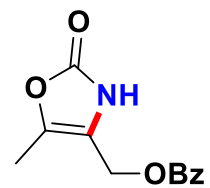


8.471  
8.043  
8.024  
7.605  
7.587  
7.568  
7.468  
7.449  
7.430  
7.264

5.029

2.177

0.000



**2s**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

2.04  
0.90  
2.08  
1.00

1.98

3.07

8 6 4 2 0 PPM

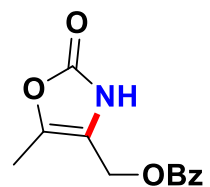


8.043  
8.024  
7.591  
7.472  
7.453  
7.434  
7.264

5.029

2.179

0.000



**2s**

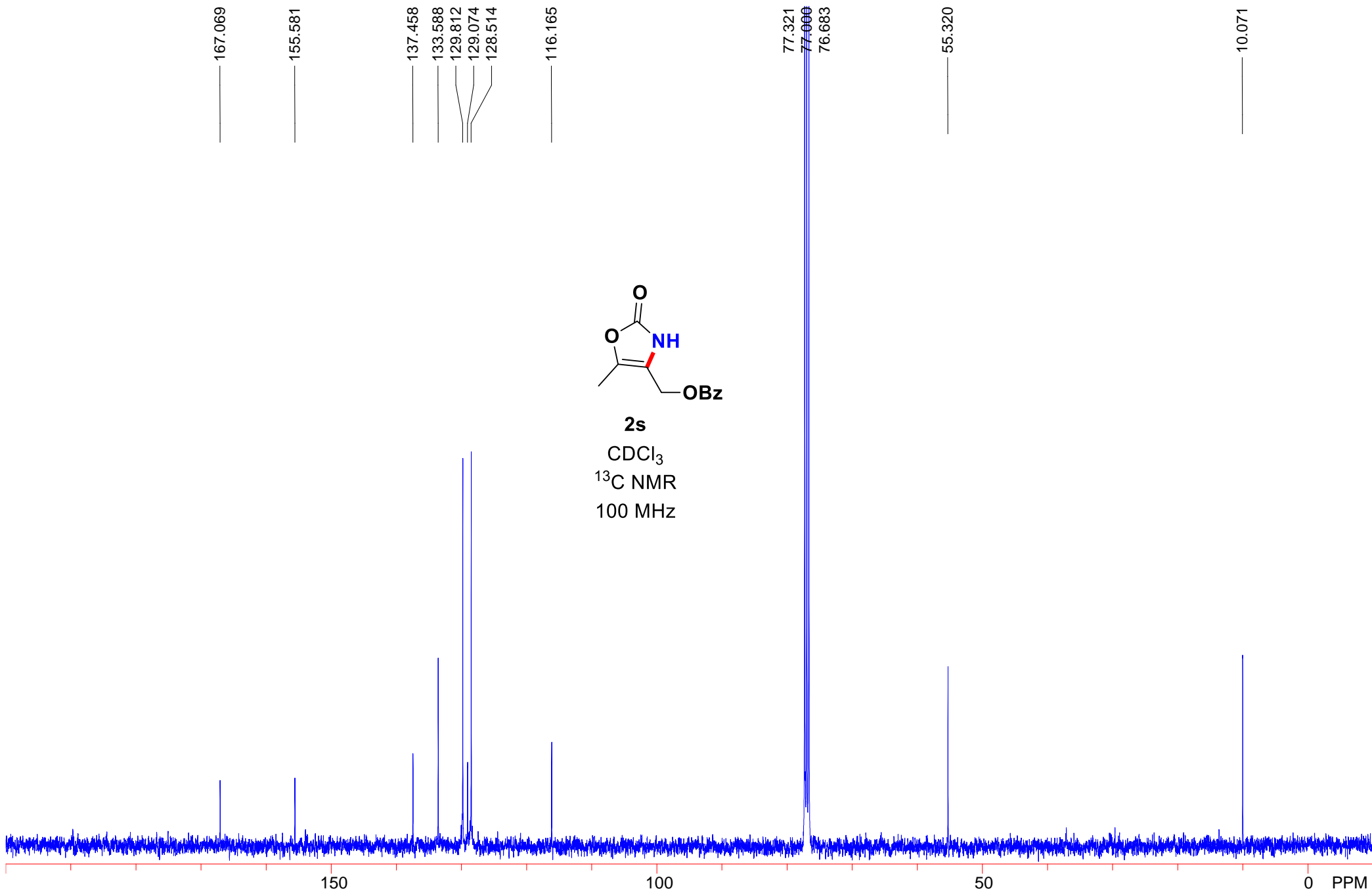
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

0.95  
2.00  
1.08  
2.19

2.00

3.01

8 6 4 2 0 PPM



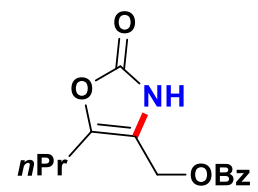
8.552  
8.042  
8.024  
7.604  
7.586  
7.567  
7.467  
7.448  
7.429  
7.266

5.039

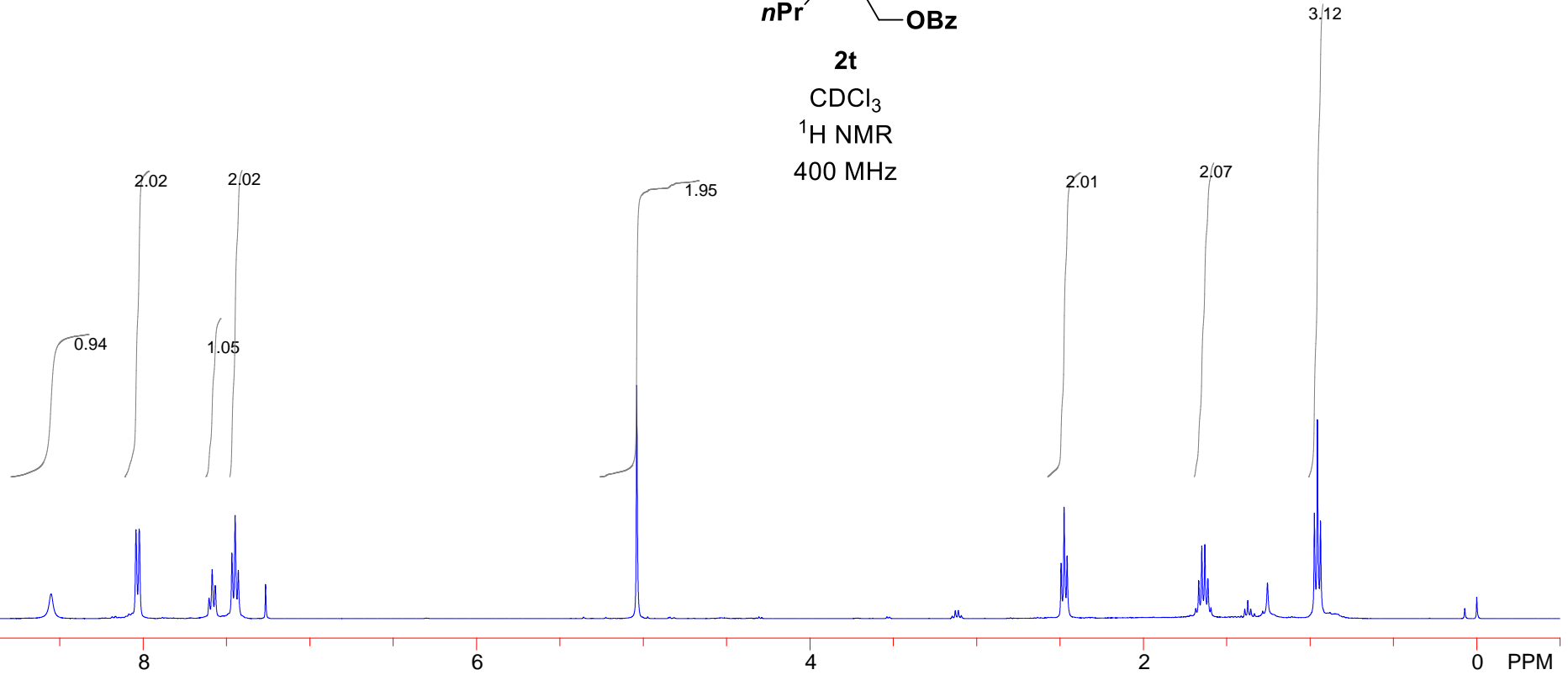
2.494  
2.475  
2.457

1.668  
1.649  
1.631  
1.613  
1.594  
1.374  
1.255  
0.974  
0.955  
0.937

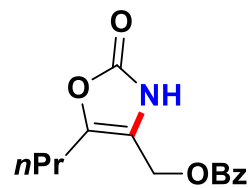
-0.000



**2t**  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



167.093  
155.627  
141.097  
133.592  
129.796  
129.045  
128.514  
116.268  
77.317  
77.000  
76.683  
55.249  
26.311  
20.709  
13.356

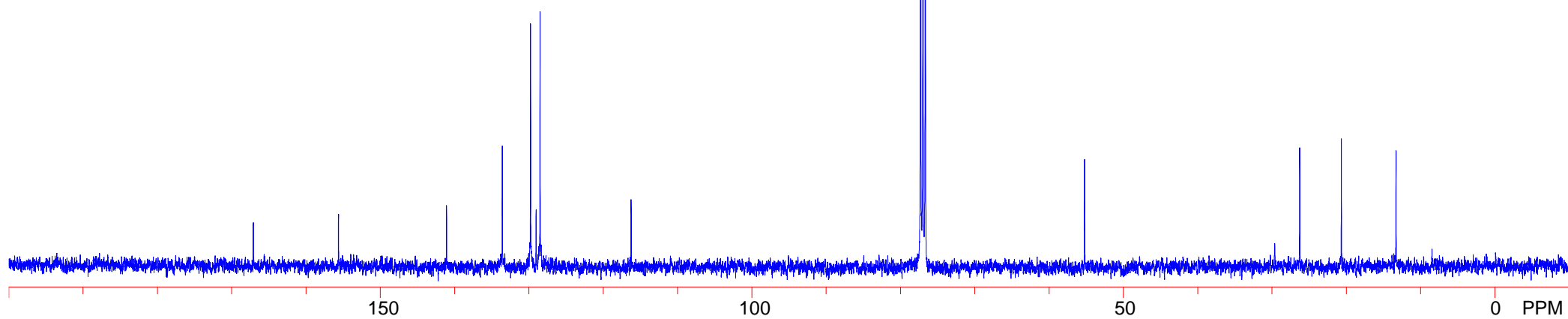


**2t**

CDCl<sub>3</sub>

<sup>13</sup>C NMR

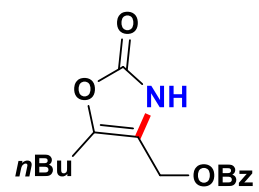
100 MHz



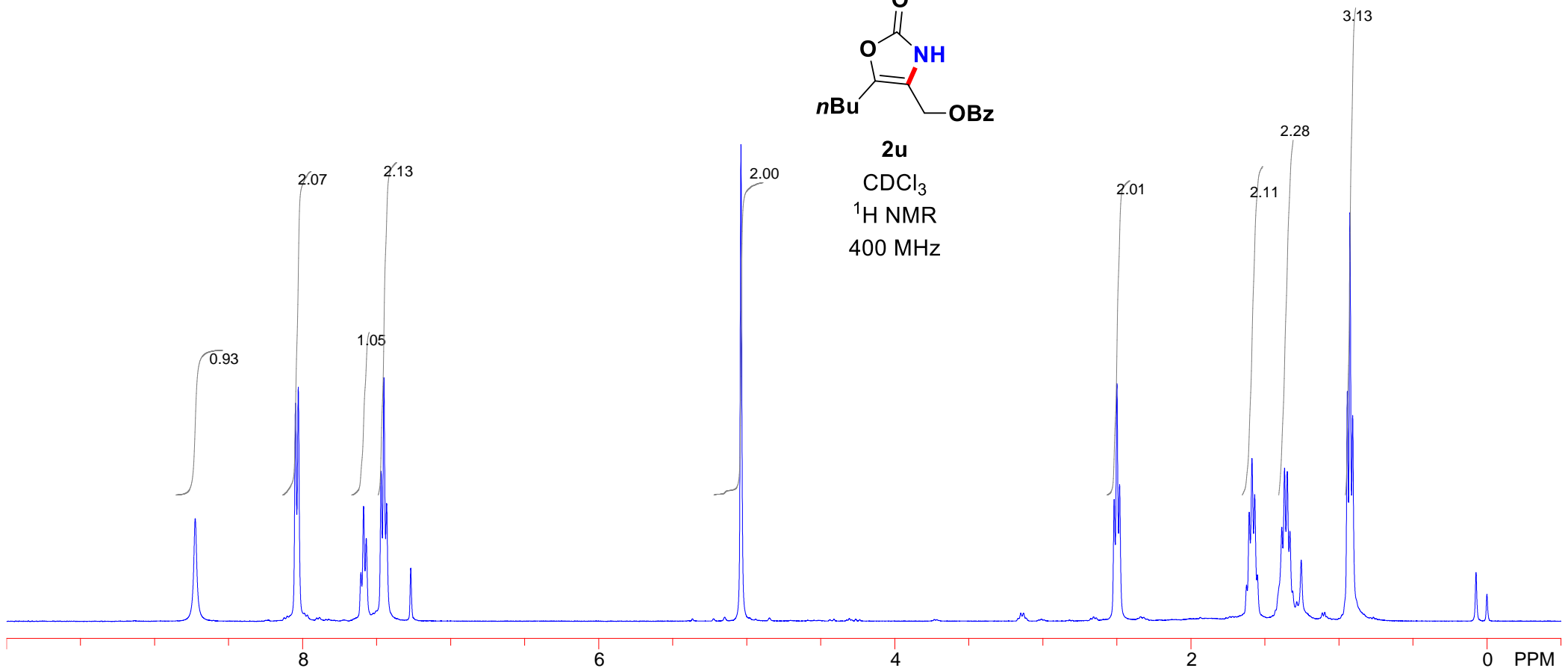
8.725  
8.048  
8.028  
7.606  
7.588  
7.569  
7.469  
7.451  
7.432  
7.269

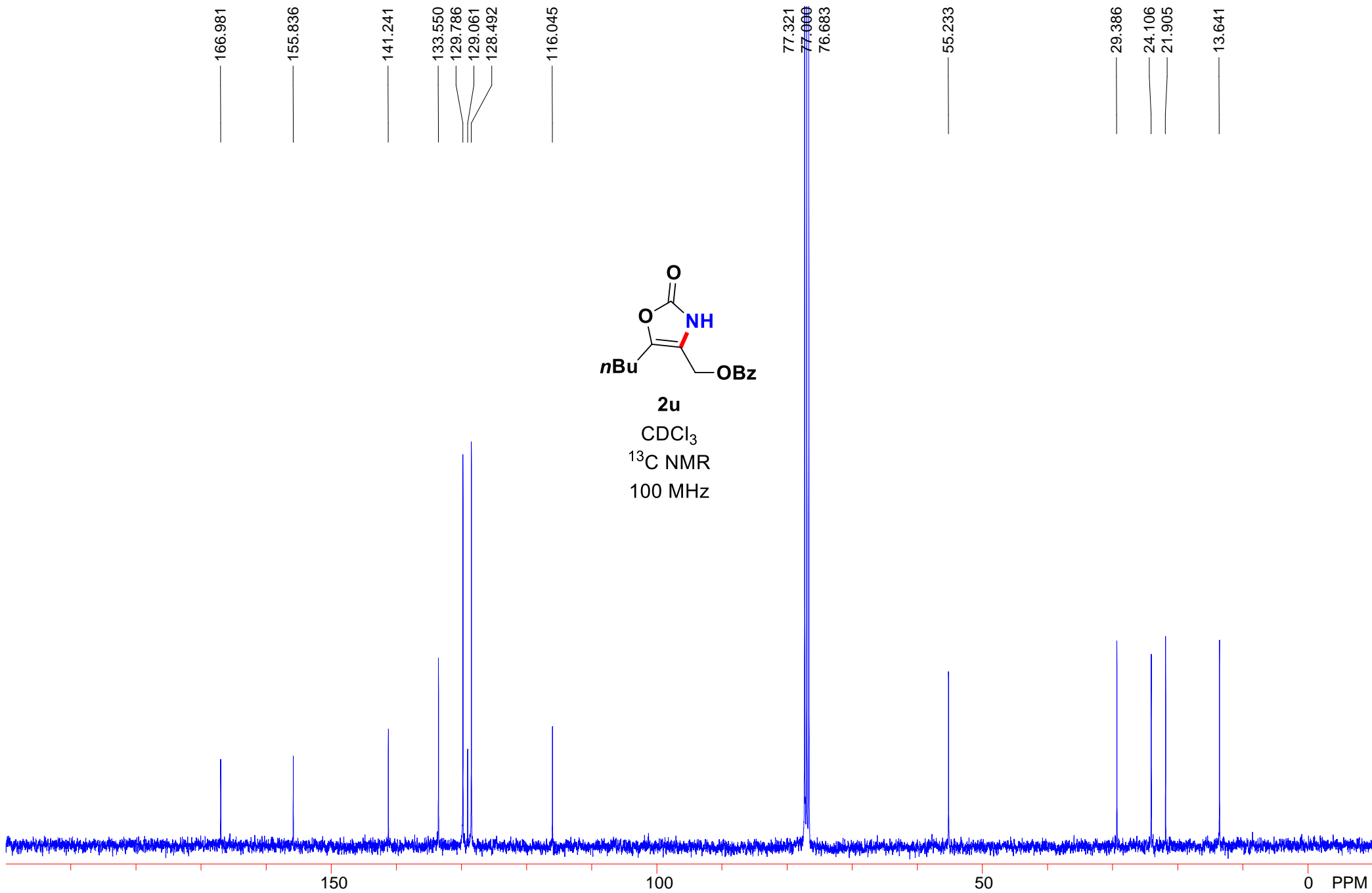
5.039

2.518  
2.500  
2.481  
1.624  
1.606  
1.587  
1.569  
1.551  
1.386  
1.367  
1.348  
1.331  
1.312  
1.255  
0.943  
0.925  
0.907  
-0.000



CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz



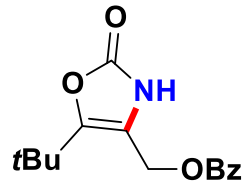


8.675  
8.064  
8.043  
7.604  
7.587  
7.569  
7.471  
7.453  
7.433  
7.267

5.184

1.336

0.000

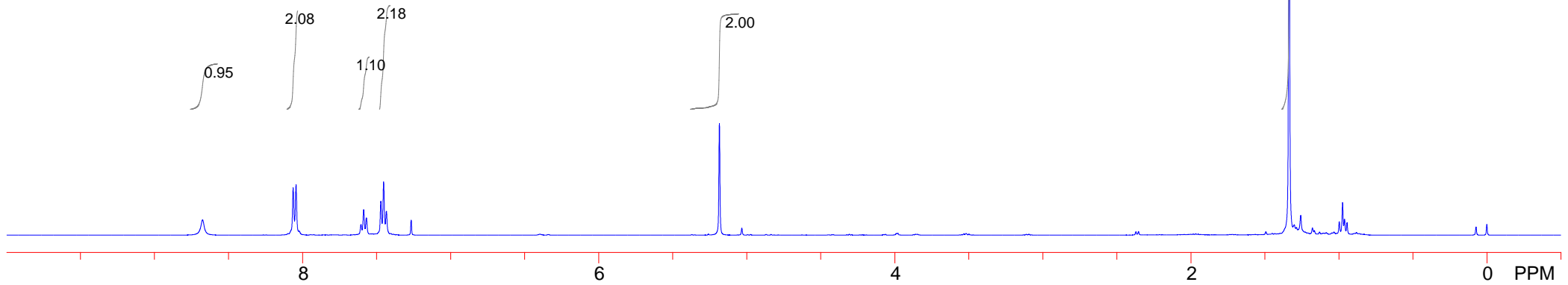


2v  
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
400 MHz

9.06

0.95  
2.08  
1.10  
2.18

2.00



166.702

155.482

147.268

133.476

129.816

129.173

128.477

114.285

77.317

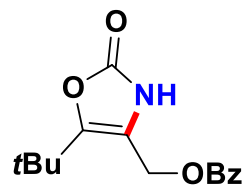
77.000

76.683

56.099

32.947

28.775



2v

CDCl<sub>3</sub>

<sup>13</sup>C NMR

100 MHz

150

100

50

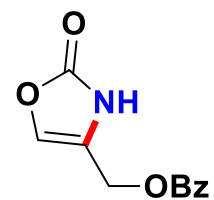
0 PPM



8.369  
8.046  
8.028  
7.622  
7.605  
7.588  
7.480  
7.463  
7.445  
7.263  
6.893

5.054

-0.000

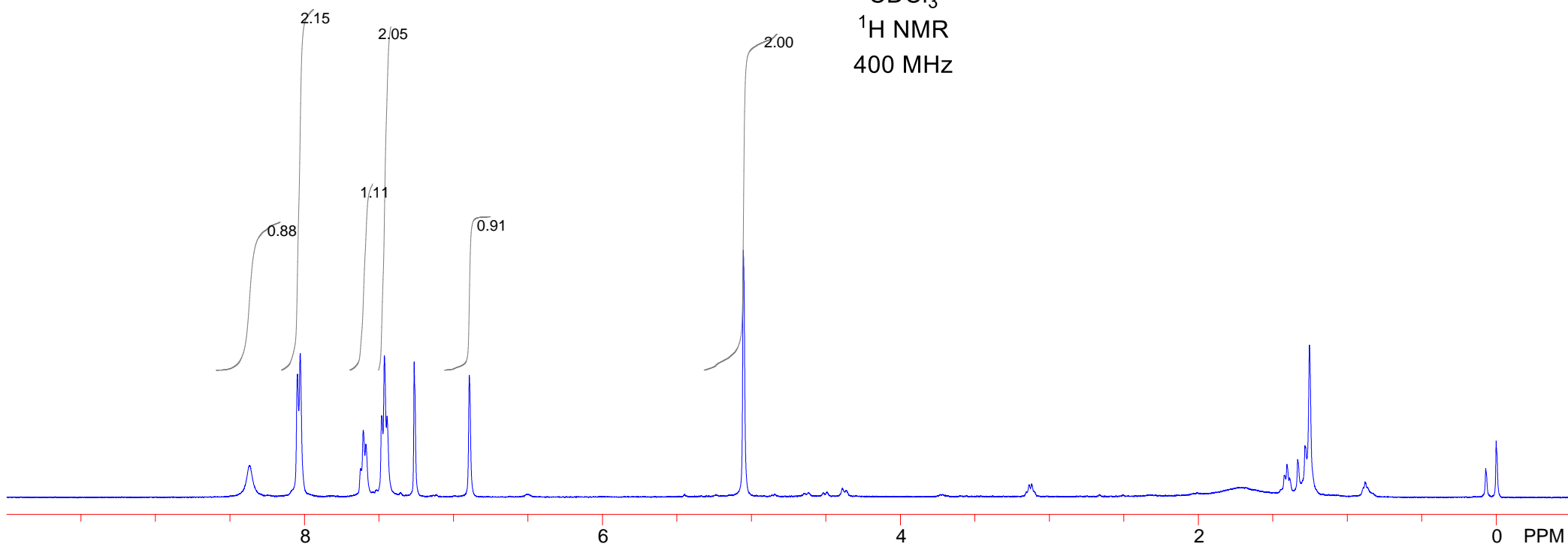


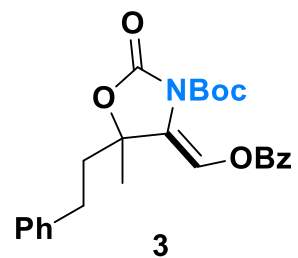
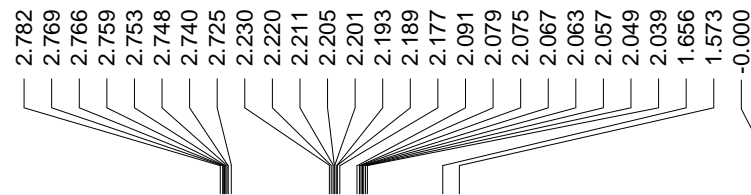
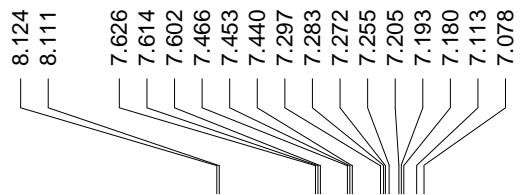
**2w**

CDCl<sub>3</sub>

<sup>1</sup>H NMR

400 MHz

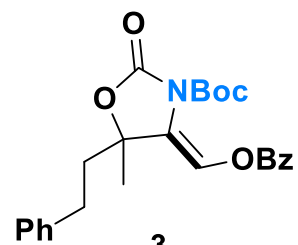




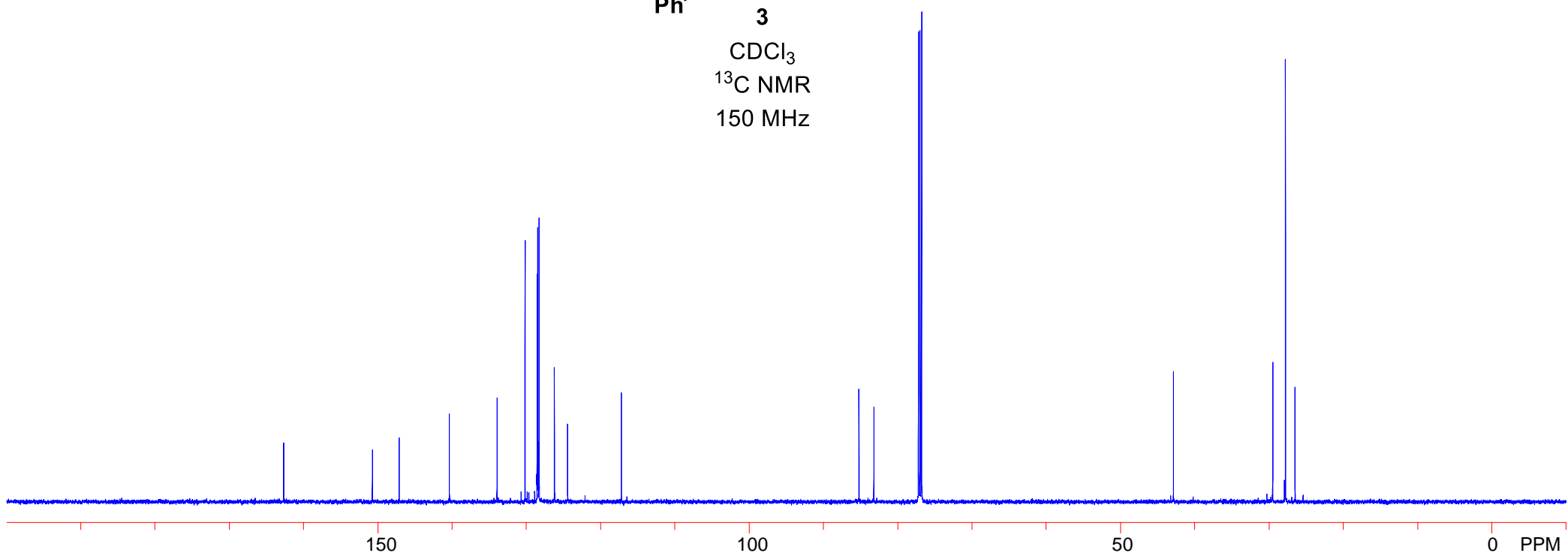
CDCl<sub>3</sub>  
<sup>1</sup>H NMR  
 600 MHz



162.668  
150.735  
147.132  
140.354  
133.943  
130.175  
128.551  
128.479  
128.341  
128.305  
126.219  
124.443  
117.210  
85.230  
83.230  
77.217  
77.000  
76.791  
42.869  
29.492  
27.774  
26.489



CDCl<sub>3</sub>  
<sup>13</sup>C NMR  
150 MHz

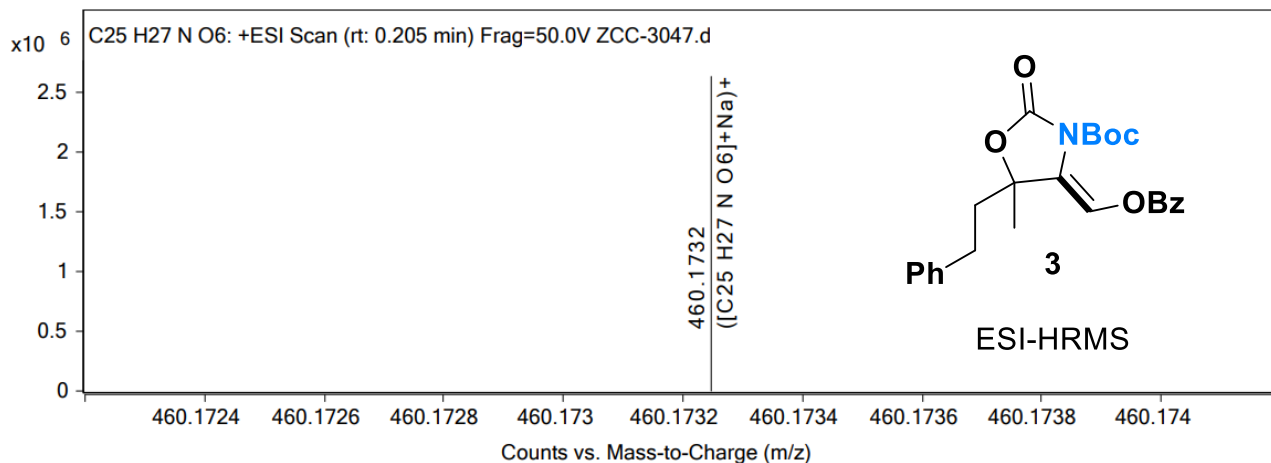


## Qualitative Analysis Report

<b>Data File</b>	ZCC-3047.d	<b>Sample Name</b>	Sample48
<b>Sample Type</b>	Sample	<b>Position</b>	P2-E4
<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Acq Method</b>	a3-50V-100%CH3OH.m	<b>Acquired Time</b>	11/22/2024 12:02:31 PM (UTC+08:00)
<b>DA Method</b>	ZJU.m	<b>Comment</b>	
<b>Sample Group</b>		<b>Info.</b>	
<b>Stream Name</b>	LC 1	<b>Acquisition Time (Local)</b>	11/22/2024 12:02:31 PM (UTC+08:00)
<b>Acquisition SW Version</b>	6200 series TOF/6500 series Q-TOF B.09.00 (B9044.0)	<b>QTOF Driver Version</b>	8.00.00
<b>QTOF Firmware Version</b>	25.723	<b>Tune Mass Range Max.</b>	3200

### Spectra

**Fragmentor Voltage**      **Collision Energy**      **Ionization Mode**  
 50                                  0                                  ESI



#### Formula Calculator Element Limits

Element	Min	Max
C	0	100
H	0	100
O	0	6
N	0	5

#### Formula Calculator Results

Formula	Best	Measured Mass	Tgt Mass	Diff (ppm)	Score
C25 H27 N Na O6	True	460.1732	460.1731	-0.48	99.79

--- End Of Report ---