

Supplementary information

Rewiring methanol assimilation and reductive glycine pathways in *Saccharomyces cerevisiae* to increase one-carbon recovery

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Supplementary Methods:

Whole-genome resequencing and data analysis

The control strain M01 and three evolved strains of M29 were chosen for genome sequencing. The whole genome was sequenced using Illumina HiSeq/Novaseq/MGI2000 at Genewiz Biotech (Suzhou, China). Data analysis was used by fastp (V0.23.0) and annotation for SNV/InDel was performed by Annovar (V21 Apr 2018).

¹³C Metabolite tracer analysis

For ¹³C metabolite tracer analysis, strain M45 was cultivated in YPD overnight and washed twice with PBS (pH=7.2) before being transferred to YNB medium with 0.1% yeast extract and 0.4% ¹³C-methanol (Sigma-Aldrich), initial OD₆₀₀ was around 0.3, then cultured for 144 h. The yeast (about 10⁷ cells) was taken, mixed with 1000 μL of extraction solution consisting of methanol, acetonitrile, and water in a ratio of 2:2:1 (v/v). The mixture was vortexed for 30 s and then incubated in liquid nitrogen for 1 min. Subsequently, the samples were allowed to thaw at room temperature and vortexed for 30 s. This freeze-thaw cycle was repeated three times. Following that, the samples were sonicated for 10 min in a 4°C water bath and incubated for 1 h at -40°C to precipitate proteins. The samples were centrifuged at 12000 rpm for 15 min at 4°C. The supernatant was collected and evaporated to dryness by a vacuum concentrator. The dried extracts were then reconstituted in 100 μL of a mixture of acetonitrile and water in a 1:1 ratio (v/v), followed by sonication for 10 min at 50 Hz and 4°C, and centrifugation for 15 min at 16,200 g and 4°C to eliminate insoluble material. The supernatants were transferred to glass vials for subsequent analysis.

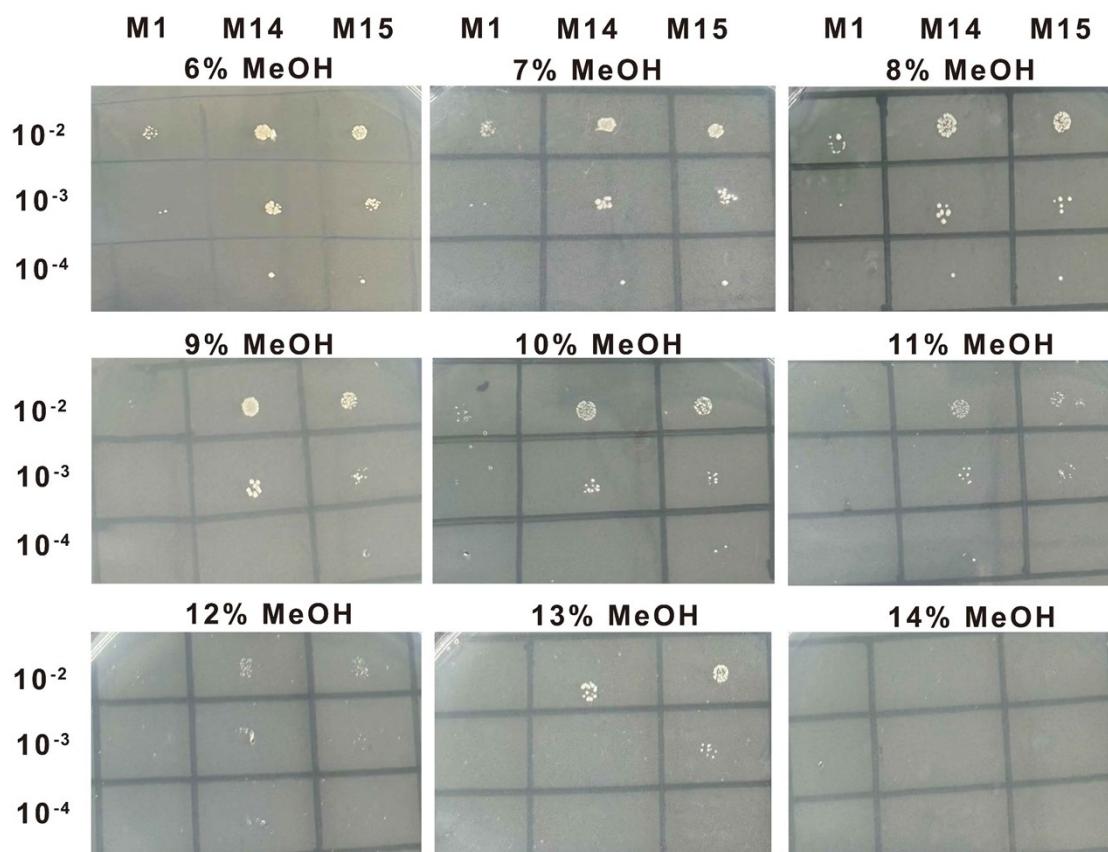
For the analysis of polar metabolites, LC-MS/MS was conducted using an UHPLC system (Vanquish, Thermo Fisher Scientific) coupled with a Waters ACQUITY UPLC BEH Amide column (2.1 mm×100 mm, 1.7 μm) and an Orbitrap Exploris 120 mass

spectrometer (Orbitrap MS, Thermo Fisher Scientific). The mobile phase comprised 25 mmol/L of ammonium acetate and 25 mmol/L of ammonia hydroxide in water (pH=9.75) (A) and acetonitrile (B). The auto-sampler temperature was 4°C, and the injection volume was 2 µL. For the analysis of non-polar metabolites, LC-MS/MS was conducted using an UHPLC system (Vanquish, Thermo Fisher Scientific) coupled with a Phenomenex Kinetex C18 column (2.1 mm×100 mm, 2.6 µm) and an Orbitrap Exploris 120 mass spectrometer (Orbitrap MS, Thermo Fisher Scientific). The mobile phase consisted of 0.01% acetic acid in water (A) and a mixture of isopropanol and acetonitrile (1:1, v/v) (B). The auto-sampler temperature was 4°C, and the injection volume was 2 µL. The ESI source conditions were set as follows: sheath gas flow rate at 50 Arb, auxiliary gas flow rate at 15 Arb, capillary temperature at 320°C, full MS resolution at 60,000, MS/MS resolution at 15,000, collision energy set to 20/30/40 eV in NCE mode, and spray voltage at 3.8 kV (positive) or -3.4 kV (negative), respectively.

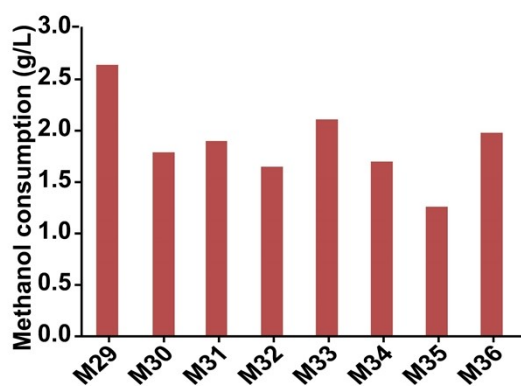
GC-MS was used for the detection of metabolites in the reductive glycine pathway. Firstly, strain M47 was cultivated in YPD overnight and washed twice with PBS (pH = 7.2) before being transferred to YNB medium with 0.1% yeast extract, 2% methanol, 250 mM sodium formate, 100 mM (NH₄)₂SO₄, and 35 mM NaH¹³CO₃ (Sigma-Aldrich), initial OD₆₀₀ was around 0.2, then cultured for 144 h. The metabolites were subsequently derivatized for GC-MS analysis following a previously published protocol ¹. The cells were ground in liquid nitrogen, re-suspended in 1 mL of cold (-40°C) 50% aqueous methanol, and placed in dry ice for 30 min before thawing the samples on ice. Then, 0.4 mL of chloroform was added, and the mixture was vortexed for 30 s, and centrifuged for 15 min at 14,000 rpm (4°C). The supernatant was then transferred to new 1.5 mL tubes for evaporation and storage at -80°C prior to analysis.

Metabolites were derivatized for GC-MS analysis as follows: First, 70 μ L of O-Isobutylhydroxylamine hydrochloride was added to the dried pellet and incubated for 20 min at 85°C. After cooling, 30 μ L of *N-tert*-butyldimethylsilyl-*N*-methyltrifluoroacetamide (MTBSTFA) was added and samples were re-incubated for 60 min at 85°C before centrifugation for 15 min at 12,000 rpm (4°C). The supernatant was transferred to an autosampler vial for GC-MS analysis.

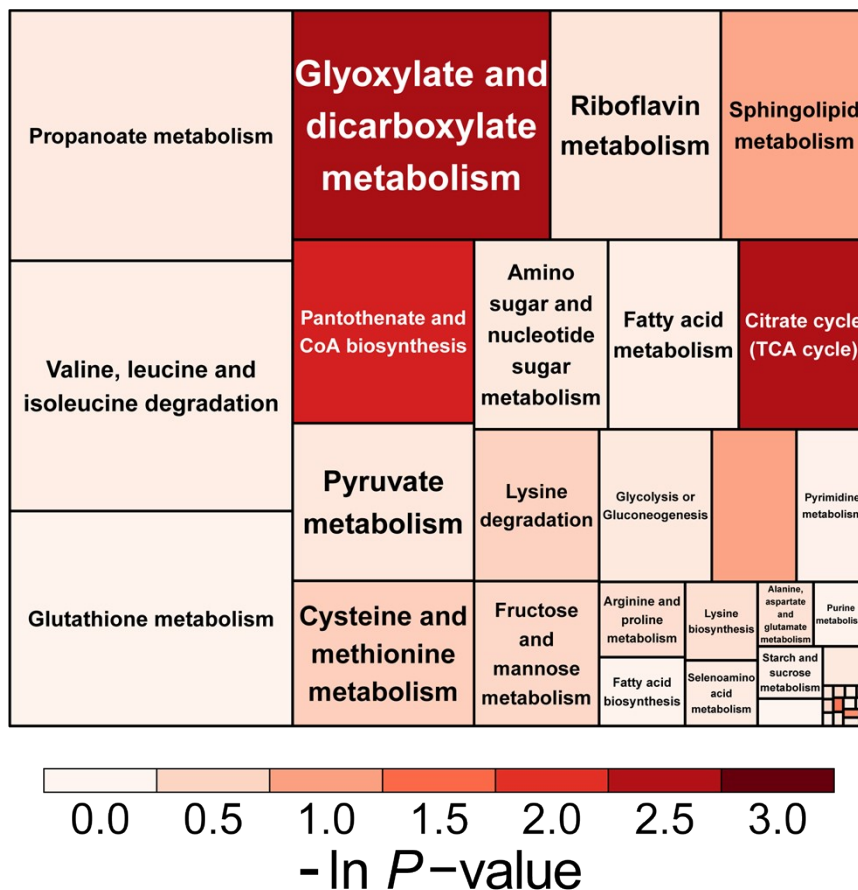
A Shimadzu QP-2020 GC-MS was programmed with an injection temperature of 250°C. The GC column used was a 30 m \times 0.25 mm \times 0.25 mm DB-5ms. GC-MS interface temperature was 300°C and ion source temperature was set at 200°C, with 70 V ionization voltage. The mass spectrometer was set to scan *m/z* range 50-700, with 1 kV detector. GC flow rate with helium carrier gas was 0.92 ml/min. The gradient elution of GC oven temperature was list as follows: 120°C for 3 min, followed by a ramp of 4 °C min⁻¹ to 170°C, 0 min hold; 2°C min⁻¹ to 182°C, 0 min hold; 20°C min⁻¹ to 195°C, 0 min hold; 3°C min⁻¹ to 230°C, 0 min hold; 10°C min⁻¹ to 265°C, 0 min hold; 3°C min⁻¹ to 280°C, 5 min hold, and the injection volume was 5 μ L.



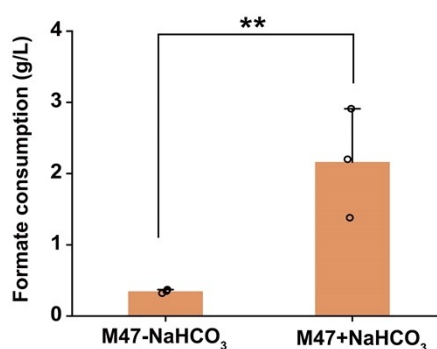
Supplementary Fig. S1 Growth in different concentrations of methanol to evaluate their tolerance to methanol. Growth on solid YNB medium with 0.1% yeast extract, and different concentrations of methanol, methanol (MeOH). Images were taken after incubating at 30°C for 6 days.



Supplementary Fig. S2 Fermentation validation of 8 single clones from the evolved strains G2-80 plate. Strains were cultured in YNB medium with 2% methanol, 0.1% yeast extract at 30°C, 220 rpm, and the initial OD₆₀₀ was around 0.15.

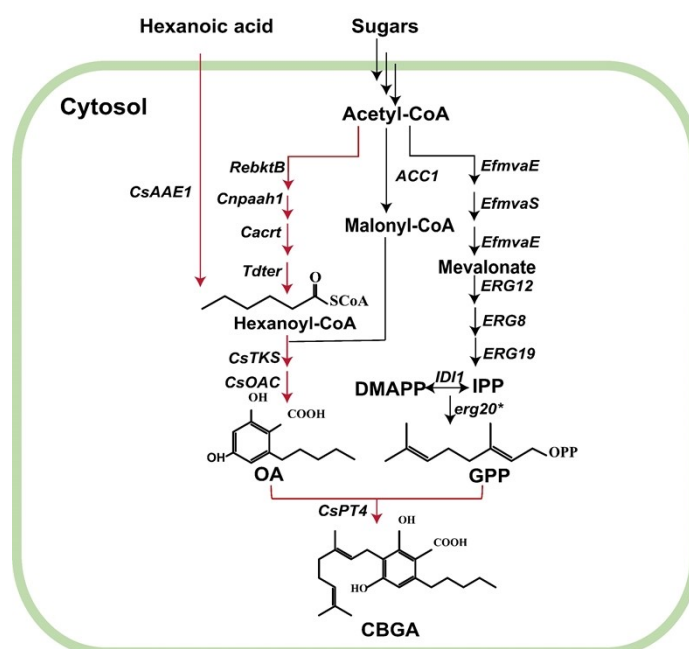


Supplementary Fig. S3 Pathway analysis of ^{13}C -labelled metabolites in strain M045. The labelled metabolites are mainly involved in glyoxylate metabolism and TCA cycle, suggesting that we have successfully rewired the central carbon metabolism of *S. cerevisiae* through modularization and ALE strategies, engineering it into a methylotrophic yeast.



Supplementary Fig. S4 Comparison of formate consumption in strain M47 in media with and without NaHCO₃. The consumption of formate in the medium with NaHCO₃

was significantly higher than that without NaHCO₃, proving the activity of RGP. Strains were cultured in YNB medium with 0.1% yeast extract, 2% methanol, 250 mM sodium formate, 100 mM (NH₄)₂SO₄, and 100 mM NaHCO₃ at 30°C, 220 rpm.



Supplementary Fig. S5 Biosynthetic pathway of cannabigerolic acid (CBGA) in *S. cerevisiae*. Biosynthetic pathway of CBGA contains heterologous pathway of CBGA (red arrows) and the native pathway of *S. cerevisiae* (black arrows). CsAAE1, acyl activating enzyme from *Cannabis sativa*; ACC1, acetyl-CoA carboxylase; RebktB, β -ketothiolase from *Ralstonia eutropha*; RepaaH1, 3-hydroxybutyryl coenzyme A dehydrogenase from *Ralstonia eutropha*; Cacrt, crotonase from *Clostridium acetobutylicum*; Tdter, trans-2-enoyl-CoA reductase from *Treponema denticola*; CsTKS, polyketide synthase from *C. sativa*; CsOAC, olivetolic acid cyclase from *C. sativa*; CsPT4, geranyl transferase from *C. sativa*; EfmvaE, acetoacetyl-CoA thiolase from *Enterococcus faecalis*; EfmvaS, HMG-CoA synthase from *E. faecalis*; ERG12, mevalonate kinase; ERG8, phosphomevalonate kinase; ERG19, mevalonate pyrophosphate decarboxylase; IDI1, isopentenyl-pyrophosphate delta isomerase; erg20*, farnesyl pyrophosphate synthetase with F96W/N127W mutation. IPP, isopentenyl diphosphate; DMAPP, dimethylallyl diphosphate; GPP, geranylgeranyl pyrophosphate, OA, olivetolic acid, CBGA, cannabigerolic acid.

Supplementary Table 1 Thermodynamic analysis of different glycine biosynthetic pathways

Pathway	Reactions	$\Delta_r G'^m$ (kJ/mol)
RGP	Formate+THF+ATP \rightleftharpoons 10-formyl-THF+ADP+Orthophosphate	-2.1
	10-formyl-THF \rightleftharpoons 5,10-methenyl-THF+H ₂ O	5.4
	5,10-methenyl-THF+NADPH \rightleftharpoons 5,10-methylene-THF+NADP ⁺	-6.5
	5,10-methylene-THF+NH ₄ +CO ₂ +NADH \rightleftharpoons Glycine+THF+NAD ⁺	-4.9
Methanol-RGP	Methanol+O ₂ \rightleftharpoons Formaldehyde+H ₂ O ₂	-98.9
	Formaldehyde+THF \rightleftharpoons 5,10-methylene-THF+H ₂ O	-5.2
	5,10-methylene-THF+NH ₄ +CO ₂ +NADH \rightleftharpoons Glycine+THF+NAD ⁺	-4.9

Supplementary Table 2 Plasmids used in this study

Plasmids	Description	Source
p423-SpSgH	2 μ ; <i>HIS3</i> ; <i>AmpR</i> ; <i>pSNR52-BsaI-BsaI-SpSgRNA-tSUP4</i>	2
p426-SpSgH	2 μ ; <i>URA3</i> ; <i>AmpR</i> ; <i>pSNR52-BsaI-BsaI-SpSgRNA-tSUP4</i>	3
p423*-ccdB	2 μ ; <i>HIS3</i> ; <i>AmpR</i> ; <i>pSNR52-BsaI-ccdB-BsaI-tSUP4</i>	2
p426*-ccdB	2 μ ; <i>URA3</i> ; <i>AmpR</i> ; <i>pSNR52-BsaI-ccdB-BsaI-tSUP4</i>	3
p41K-iCas9	2 μ ; <i>KanMX</i> ; <i>AmpR</i> ; <i>pTEF1-iCas9-tADH1</i>	4
pESC-URA	2 μ ; <i>URA3</i> ; <i>AmpR</i> ; <i>pGAL1-MCS1-tCYC1</i> ; <i>pGAL10-MCS2-tADH1</i>	2
pH23	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>tADH1-pGPD1-eGFP-tCYC1-pENO2</i>	2
pH5	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF1-eGFP-TEF1</i>	2
pH66	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>tTEF1-pPGK1-eGFP-tHXT7</i>	2
pH5-PFK1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF1-PFK1-tTEF1</i>	this study
pH5-PFK2	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF1-PFK2-tTEF1</i>	this study
pH5-FBA1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF1-FBA1-tTEF1</i>	this study
pH7-RPE1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTP11-RPE1-tTEF1</i>	this study
pH7-Bmplgx	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTP11-Bmplgx-tTEF1</i>	this study
pH10-BsMDH	2 μ ; <i>URA3</i> ; <i>AmpR</i> ; <i>pGPM1-BsMDH-tTEF1</i>	this study
pH9-ADK1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF2-ADK1-tHXT7</i>	this study
pH9-FLS	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF2-FLS-tTEF1</i>	this study
pH5-HPS	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF1-HPS-tTEF1</i>	this study
pH5-PHI	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTEF1-PHI-tTEF1</i>	this study
pH10-AOX1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pGPM1-AOX1-tTEF1</i>	this study
pH10-DAK1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pGPM1-DAK1-tTEF1</i>	this study
pH11-DAS2	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pENO2-DAS2-tTEF1</i>	this study
pH11-CAT1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pENO2-CAT1-tTEF1</i>	this study
pH7-TKL1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pTP11-TKL1-tTEF1</i>	this study
pH23-FBP1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pGPD1-FBP1-tCYC1</i>	this study
pH23-ZWF1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pGPD1-ZWF1-tCYC1</i>	this study
pH12-GCV1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pFBA1-GCV1-tTEF1</i>	this study
pH12-GCV2	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pFBA1-GCV2-tTEF1</i>	this study
pH13-GCV3	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pPDC1-GCV3-tCYC1</i>	this study
pH13-MIS1	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pPDC1-MIS1-tCYC1</i>	this study
pH23-SHM2 ^{A194C/S248T}	2 μ ; <i>LEU2</i> ; <i>AmpR</i> ; <i>pGPD1-SHM2^{A194C/S248T}-tCYC1</i>	this study

Supplementary Table 3 Strains used in this study

Strains	Description	Source
INVSc1	<i>MATa/MATa his 3Δ1 leu2 trp 1-289 ura3-52</i>	Lian's lab
M01	INVSc1[<i>YPRCδ15c::pPGK1-Tdter-tHXT7; SAP155b::pTEF1-RebktB-tTEF1;1414a::pGAL1-EfmvaS-tCYC1/pGAL10-EfmvaE-tADH1; 1114a::pGAL1-ERG12-tCYC1/pGAL10-IDI-tADH1;SAP155c::pGAL1-RePaaH1-tCYC1/pGAL10-CaCrt-tADH1;1014a::pGAL1-ERG19-tCYC1/pGAL10-ERG8-tADH1; 1622b:: pGAL1-CsOAC-tCYC1/pGAL10-CsTKS-tADH1;911b:: pGPD1-CsAAE1-tCYC1; 1021b::pGAL1-tHMG1-tCYC1/pGAL10-erg20*-tADH1; 416d::pGAL1-CsPT4-tCYC1; 308a::pGAL1-CBCAS-tCYC1; X4:: pGAL1-CsOAC-tCYC1/pGAL10-CsTKS-tADH1;XI3:: pGAL1-CsOAC-tCYC1/pGAL10-CsTKS-tADH1;XII5:: pGAL1-CsOAC-tCYC1/pGAL10-CsTKS-tADH1;208a:: pGPD1-CsAAE1-tCYC1;106a:: pGPD1-CsAAE1-tCYC1; CAN1y::pGAL1-CsPT4-tCYC1;YOLCd1b::pGAL1-CsPT4-tCYC1; 1021b:: pGAL10-erg20*-tADH1; XII::pTEF1-CHK-tTEF1;X12::pTEF1-AuPK-tTEF1; XII4::pPGK1-FAA2-tHXT7; X2::pGAL1-IDI1-tCYC1; X3:: pTEF1-ERO1-tTEF1; NS7:: pPGK1-IRE1*-tHXT7; NS8:: pPGK1-PDI1-tHXT7; NS9:: pPGK1-CNE1-tHXT7; NS2:: pTEF1-INO2-tTEF1; NS3::pPGK1-KAR2-tHXT7; NS14:: pPGK1-HAC1s*-tHXT7; YGLCτ3:: pTEF1-ERO1-tTEF1; Gal80Δ]</i>	This study
M02	M01[<i>NS18:: pTEF1-HPS-tTEF1; NS19:: pTEF1-PHI-tTEF1</i>]	This study
M06	M02[<i>NS22:: pTEF1-PFK2-tTEF1; NS23:: pTEF1-FBA1-tTEF1;YORWΔ22:: pTEF1-PFK1-tTEF1;NS24:: pTPI1-RPE1-tTEF1; NS25:: pTPI1-Bmg1px-tTEF1; NS10:: pGPM1-BsMDH-tTEF1; NS11:: pGPM1-BsMDH-tTEF1</i>]	This study
M07	M06[<i>NS5:: pTEF2-ADK1-tTEF1; NS6:: pTEF2-FLS-tTEF1</i>]	This study
M09	M06[<i>XI7:: pGPM1-AOX1-tTEF1;XI8:: pGPM1-DAK1-tTEF1;YCRWδ12:: pENO2-DAS2-tTEF1;YNRCΔ9::pENO2-CAT1-tTEF1</i>]	This study
M14	M09[<i>V1::pGPD1-FBP1-tCYC1;V3::pGPD1-ZWF1-tCYC1; NS17:: pGPD1-TKL1-tCYC1; YGR067C:: tYGR067C</i>]	This study
M15	M14[<i>LPL1Δ;IZH3Δ</i>]	This study
M27	M15-G80-2	This study
M29	M15-G80-2-1	This study
M45	M29[<i>4-OH:: pGPD1-SHM2^{A194CS248T}-tCYC1</i>]	This study
M47	M45[<i>NS20::pFBA1-GCV1-tTEF1;NS21::pFBA1-GCV2-tTEF1; XII3:: pPDC1-GCV3-tCYC1; XI5::pPDC1-MIS1-tCYC1</i>]	This study

Supplementary Table 4 List of primers used in this study

primers	sequences (5'-3')
for plasmids construction	
IDP2-F	ACACCAGAACTTAGTTTCGACGGATGGATCCATGACAAAGATTAAGGTAGCTAAC CC
IDP2-R	GCGTGACATAACTAATTACATGATCTCGAGTTACAATGCAGCTGCCTCGAA
PFK1-F	CTAATCTAAGTTTTAATTACAAAGGATCCATGCAATCTCAAGATTCATGCT
PFK1-R	TATGCAACTAGAAAAGTCTTATCAATCTCCCTCGAGTCATTTGTTTCAGCGGCTA AAG
PFK2-F	TCTAATCTAAGTTTTAATTACAAAGGATCCATGACTGTTACTACTCCTTTGTGAA TG
PFK2-R	ACTAGAAAAGTCTTATCAATCTCCCTCGAGTTAATCAACTCTCTTCTTCCAACCA A
FBA1-F	CTAATCTAAGTTTTAATTACAAAGGATCCATGGGTGTTGAACAAATCTTAA
FBA1-R	AGAAAACGTCTTATCAATCTCCCTCGAGTTATAAAGTGTTAGTGGTACGGAAAG
GCV1-F	AACCAAGTAATACATATTCAAAAGGATCCATGTCTATAATCAAAAAAATTGTGTT TAAG
GCV1-R	AGAAAAGTCTTATCAATCTCCCTCGAGTTACTGCTTGTAGTAATGTGTGG
GCV2-F	AACCAAGTAATACATATTCAAAAGGATCCATGCTTAGGACAAGAGTGACT
GCV2-R	AAAGTCTTATCAATCTCCCTCGAGTCATTCAGTTTCGTTTCGCAATTC
GCV3-F	AATCTAAGTTTTAATTACAAAGGATCCATGTTACGCACTACTAGACTATG
GCV3-R	AAAAGTCTTATCAATCTCCCTCGAGTCAGTCATCATGAACCAGTGTCT
MIS1-F	ACACAGTCAAATCAATCAAAGGATCCATGTTGTCGAGACTATCTTTATTGAGT
MIS1-R	TGACATAACTAATTACATGATCTCGAGTTAAAATAGACCTTCAATTCACC
for genome integration	
HPS-HOMO-F	GCAAAATAGTATACCATCTTTGGTTCCTGGACCACATTTTCGCGCGTAATACGAC TCAC
HPS-HOMO-R	AAAGGGTTCGCTCAATGGCAGATGCTGCCATACGAGGACAATAGCGCCGATCAA AGTAT
PHI-HOMO-F	AACTCCGTCCACAGTACCCGATCAGCTGGCTCATCGTTATGCGCGCGTAATACGA CTCA
PHI-HOMO-R	CTTGCAACGCTAGTAACGCCGATCCACAGAGAAACCGGGATAGCGCCGATCAA AGTAT
PFK2-HOMO-F	AAGGTTGGCCCGCCAAAATCGTAAGTCTCGGTGCTCTGTTGCGCGCGTAATACGA CTCA
PFK2-HOMO-R	TTACTGGAGTAATGCTTATATTTCGCGTCCCCCACGCATAGCGCCGATCAAA GTAT
FBA1-HOMO-F	TGTGGTGAATGCACCTGGTGCACGCTGCGAAGGTGACGTAGAGCGCGCGTAATA CGACT
FBA1-HOMO-R	TGGCTTTTAACTTATTACCCCAAGAGAGTGTCTACAGAAATAGCGCCGATCAAA GTAT
PFK1-HOMO-F	TCTTTTAAATACTTATTAACGTACTCAAACAACACTACTTTGAGCGCGCGTAATA CGAC
PFK1-HOMO-R	GGTCCCTATTCCGATAATCTTAGCAGAGTGAATAGTAATAATAGCGCCGATCAAA GTAT
RPE1-HOMO-F	CATTAGATAGAGAGGGGCGAGATGTTCAAGCTATACCCATTCTACTTATTCCCTTC GAGA
RPE1-HOMO-R	ACTGAAATGAAAATTCATATTTACTTTTTATTGTTACTGACCATGATTACGCCA AGC
Bmg1px-HOMO-F	CCTAATTAGTAGGAAGCGGAAAATAATAATATAAGAAAGTCTACTTATTCCCTTC GAGA
Bmg1px-HOMO-R	TTGCGCGCTCAAGCGGCCCTATACTGCACACCTATTACTTGACCATGATTACGCC AAGC
BsMDH-HOMO-F	ACTTTTTACCATCCTTTAGCTTTACCTAATATAATGAAATTAGTCGTGCAATGTAT GAC
BsMDH-HOMO-R	TTCGAGAAATAGTTGGTATAAATAACTATAAATAACGTTTATAGCGCCGATCAAA GTAT
BsMDH-HOMO-	AACTAACCTAAGAATTTTGAAATCACAACAAAAATAATTAGTCGTGCAATGTATG

F	AC
BsMDH-HOMO-R	ACAAAAAAGTAGTAATAAATAGGTCCAAATCTTCTTTATTATAGCGCCGATCAAA GTAT
ADK1-HOMO-F	CATTTGAATTTTAGTAGTAATAATAAGATCCCATCCGGGCGCCATAACCAA GGTA
ADK1-HOMO-R	TCATCTATAAAGACTGTACGCATATTTGGAACAACTGCTATAGCGCCGATCAAA GTAT
FLS-HOMO-F	ACATTTCAAGTGGTTTCTCAAGGGAGAATCATAGTTTAGCGGGCGCCATAACCAA GGTA
FLS-HOMO-R	CTTCGTATACTTATTACGCAGGTAGGAGTGCAATAGTTGAATAGCGCCGATCAAA GTAT
AOX1-HOMO-F	CTTGACATGTAACGTAAGAAAAGAAAAAGAGATGGCAGATAGTCGTGCAATGT ATGAC
AOX1-HOMO-R	AATCATTGCTATCCCCACAAAAACAGTGCATGTACTTTGATAGCGCCGATCAAA GTAT
DAK1-HOMO-R	TTTTTGCCATACATTTAGCGCTCCACACTCTAAATAAAGTGTGTCGACGCTGCGG GTAT
DAK1-HOMO-F	TTTATAATATGCAAAAGGCGGTAACGATATGCCGCGCAAAATAGCGCCGATCAA AGTAT
DAS2-HOMO-F	GCCGATGAAATAAAATCCTGATATCATCTATATAGTAGTGTGTCGACGCTGCGGG TAT
DAS2-HOMO-R	TGAAGATATATGAATCTACAAGAGAGAGTCAATATTTCTAATAGCGCCGATCAA AGTAT
CAT1-HOMO-F	AGACATTTTTTGGGATTAATTGTTTATAAAAAGCTATGAACGTGTCGACGCTGCGG GTAT
CAT1-HOMO-R	GACATAATTGATGGGAAACAGTTATCAAAGTTATTGGATGATAGCGCCGATCAA AGTAT
TKL1-HOMO-R	GATCTGTGAAGGTTTTGAGAGAATGAGGCGAAAACACAGGCTACTTATTCCCTTC GAGA
TKL1-HOMO-F	AAGCGAGTGATTCATCATGAAAAGAATTTATCACTTCGAAATAGCGCCGATCAA AGTAT
YGR067C-HOMO-F	TTTTATTTAAACCTTCTCTCGGGAGAAGACAATTGTTGAATGGCTGCGGGTCAA AAAA
YGR067C-HOMO-R	GCTCTTGCGGATAAGTCTTCTTTATTATAAAAATACACTTACGCGGAATCTGCCAA AATA
FBP1-HOMO-F	GTGGAAAATGACCATAATGATAATTATCAATAGATAAAATCCACACCTCTACCG GCATG
FBP1-HOMO-R	CTAAAATGGTAGGCATGAGTGTCTCTCTTGTGTGCTGCAAATTAAGCCTTC GAG
ZWF1-HOMO-F	TGAAGACTACTGAACCATGGCAGCTAATACAATCACGCCCCACACCTCTACCGG CATG
ZWF1-HOMO-R	AAAAAGGCTAGGTACTTTAAAAATTTCTACAAATATAGAGCAAATTAAGCCTT CGAG
GCV1-HOMO-F	ATAAGCCCTAGAAACCTTACACCCTAATTTGCACAAGAAATAACAATACTGACA GTACTAA
GCV1-HOMO-R	CATCTGGAATATAATTCCCCCTCTGAAGCAAATTTTTCATAGCGCCGATCAAA GTAT
GCV2-HOMO-F	CATCATATAGGGACATACCTCTCAAGTTATTGTCTTGATAACAATACTGACAGT ACTAAAT
GCV2-HOMO-R	TCCAGGCAGGACGACACGAGAATAGACGGGCTGATCCCGTATAGCGCCGATCAA AGTAT
GCV3-HOMO-F	GACACATCTCTAAGCTGAAACTGAGAATACTGTTGTA AAAACATGCGACTGGGTG AGCAT
GCV3-HOMO-R	AAGTCCATTACCCTTAAGGTTGTTGTCACAACCCACGGAGGCAAATTAAGCCTT CGAG
MIS1-HOMO-F	AAGCTTCGAAGAATATGTAAATATAGTAGTATGAATCTAACATGCGACTGGGTG AGCAT
MIS1-HOMO-R	GAGTGT CATATATCCCTCCTTTAAATTTTTTACACTTACGCAAATTAAGCCTTC GAG
<i>SHM2^{A194C/S248T}</i> -HOMO-F	TACATACAACCTTTTTAAACTAATATACACATTTTAGCAGACCACACCTCTACCGG CATG
<i>SHM2^{A194C/S248T}</i> -HOMO-R	ATCCTCATAAGCAGCAATCAATTCTATCTATACTTTAAAAGCAAATTAAGCCTT CGAG
LPL1-UP-F	CCTCAACATTTATCAAATAGTTTAGCAATG
LPL1-UP-R	TCGAATTTACGGCCAATTAAGTAGC

LPL1-DOWN-F	CTAGTAATTGGGCCGTAAATTCGATGAAGCGCTGTATAATATATATATGGTTG
LPL1-DOWN-R	TTGAAGCAAGATATTGGAAAGACAAGGG
IZH3-UP-F	ACTACTGCATGAAATCACTTAAGTTCG
IZH3-UP-R	GGTATGACTGACTACCTTTACTTACAA
IZH3-DOWN-F	TAAGTAAAGGTAGTCAGTCATACCTACTTAATTTGATTGTCTGTTGGGA
IZH3-DOWN-R	CTACGAAATTAGCGTATCCGAAAA

Supplementary Table 5 Mutations in different evolved strains

Protein	M29-1	M29-2	M29-3	Annotation
AAD4	C23F		C23F	Enables aryl-alcohol dehydrogenase (NADP+) activity. Predicted to be involved in cellular aldehyde metabolic process.
AAD10	F33Y		F33Y	Enables aryl-alcohol dehydrogenase (NADP+) activity. Predicted to be involved in cellular aldehyde metabolic process.
DAN4	P216S		P216S	Predicted to be involved in fungal-type cell wall organization.
GDA1	V208I		V208I	Enables GDP phosphatase activity and UDP phosphatase activity. Involved in protein glycosylation.
ADP1	K26I		K26I	Predicted to enable ATPase-coupled transmembrane transporter activity.
RPL30	I92T		I92T	Involved in negative regulation of mRNA splicing, via spliceosome and rRNA processing.
TGL3	A12D		A12D	Involved in cell budding and triglyceride catabolic process.
COS2		R311K	R311K	Predicted to be involved in protein transport to vacuole involved in ubiquitin-dependent protein catabolic process via the multivesicular body sorting pathway.
YHR219W	S251P	S251P		Predicted to be active in cytoplasm.
YMR317W	E323A	E323A		No found.
COS6	V89F			Predicted to be involved in protein transport to vacuole involved in ubiquitin-dependent protein catabolic process via the multivesicular body sorting pathway.
PAU4	I24L			Predicted to be involved in fungal-type cell wall organization.
PFF1		V372F		Predicted to enable metalloexopeptidase activity and zinc ion binding activity. Predicted to be involved in proteolysis.
YPS5		I39T		Predicted to enable aspartic-type endopeptidase activity.
KEX1		A200V		Involved in apoptotic process.
LSG1		G494W		Involved in ribosomal large subunit assembly and ribosomal subunit export from nucleus.

NAM8	A363G	Enables mRNA binding activity. Involved in mRNA splice site recognition and positive regulation of mRNA splicing, via spliceosome. Located in cytoplasm and nucleus.
INA1	D42N	Predicted to be involved in fungal-type cell wall organization.
RAS2	K124T	Involved in several processes, including cytoplasm to vacuole targeting by the Cvt pathway; positive regulation of transcription by galactose; and protein localization to bud neck.

Supplementary Table 6 Integration site used in this study

site	20 bp spacer (5'-3')
NS5	GAGAGAATGACATTGCTCAG
NS6	TAGTTAATATCATCTAGATG
NS10	ATATAAAAAAATTCTTACTA
NS11	TATTCTTAGGAAAATCAACA
NS17	CTTGCGAAAAGAAAAGGGCA
NS18	AACTGCTCAGGGCGGATAAC
NS22	ATCTTAAATGAAAAGACAGAG
NS24	ATTATGAAAGTTTCAACTA
NS25	CACCATTCAAGTTACCGAGA
YORWΔ22	CGTTTGGGTAGTTCAGCTAA
YCRWδ12	TTTAATGGAAACGAAATGCA
YNRCΔ9	GCACGGAACCTCGGACCTAG
XI7	GTCAGTAACAGTGATTGCTG
XI8	GATGAAATAGCCTCAGTTAC
V1	CGTTTATAGACGGCACTGTC
V3	GTGCTCTTAAGTCGTAATGA
X1	GTGCCTGCTGCTATGCTCAA
III3	AGACATATGATAATCGAGCA
4-OH	GTTTCGTGAAGCATTCTTAGC

Supplementary Table 7 Heterologous genes used in this study

genes	gene sequence (5'-3')
<i>BsMDH</i>	ATGAAAGCTGCTGTTGTTAATGAATTTAAGAAAGCTTTGGAAATTAAGAAGTTGAAA GACCAAAATTGGAAGAAGGTGAAGTTTTGGTTAAAATTGAAGCTTGTGGTGTGTTGTC TACTGATTTGCATGCTGCTCATGGTGATTGGCCAATTAACCAAAAATTGCCATTGATTC CTGGTCATGAAGGTGTTGGTATTGTTGTTGAAGTTGCTAAAGGTGTTAAATCTATTA GTTGGTGATAGAGTTGGTATTCCATGGTTGATTCTGCTTGTGGTGAATGTGAATATTG TTGACTGGTCAAGAACTTTGTGTCCACATCAATTGAATGGTGGTTATTCTGTTGATG GTGGTTATGCTGAATATTGTAAGCTCCTGCTGATTATGTTGCTAAAATTCCTGATAAT TTAGATCCTGTTGAGGTCGCTCCTATTTTGTGTGCTGGTGTACTACTATAAAGCTTTG AAAGTTTCTGGTGTAGACCTGGTGAATGGGTTGCTATTTATGGTATTGGTGGTTGGG TCATATTGCTTTGCAATATGCTAAAGCTATGGGTTGAATGTTGTTGCTGTTGATATTT TGATGAAAAATCTAAATTGGCTAAAGATTTGGGTGCTGATATTGCTATTAATGGTTGA AAGAAGATCCTGTTAAAGCTATTCATGATCAAGTTGGTGGTGTTCATGCTGCTATTTCT GTTGCTGTTAATAAAAAGGCTTTTGAACAAGCTTATCAATCTGTTAAAAGAGGTGGTA

	<p>CTTTGGTTGTCGTTGGTTTGCCAAATGCTGATTTGCCAATTCCAATTTTTGATACTGTTT TGAATGGTGTCTCTGTTAAAGGTTCTATTGTTGGTACTAGAAAAGATATGCAAGAAGCT TTGGATTTTGCTGCTAGAGGTAAGTTAGACCAATTGTTGAAACTGCTGAGTTGGAAG AAATTAATGAAGTTTTTGAGAGAATGGAGAAAAGGTAATAATGGTAGAATTGTTTT GAAATTGAAAGAAGACTAA</p>
<i>BmHPS</i>	<p>ATGGAATTGCAATTGGCTTTGGATTTGGTTAATATTGAAGAAGCAAAACAAGTTGTTG CTGAAGTTCAAGAATATGTTGATATTGTTGAAATTGGTACTCCTGTTATTAATAATTTGG GGTTTGCAAGCTGTTAAGGCTGTTAAAGATGCTTTTCCACATTTGCAAGTTTTGGCTGA TATGAAAACATATGGATGCTGCAGCTTATGAAGTTGCTAAAGCTGCTGAACATGGTGT GATATTGTTACTATTTTGGCTGCAGCTGAGGACGTTTCTATTAAGGTTGCTGTTGAGGA AGCTAAAAAATTGGGTAAAAAAATTTTGGTTGATATGATTGCTGTTAAAAATTTGGAA GAAAGAGCAAAGCAAGTTGACGAAATGGGTGTTGATTATATTTGTGTTTCATGCTGGTT TGATTTACAAGCTGTTGGTAAGAATCCATTGGATGATTTGAAAAGAATTAAGCTGT TTGTTAAAAATGCTAAAACCTGCTATTGCTGGTGGTATTAAATTGGAAACTTTGCTGAA TTATTAAGCTGAACCTGATTTGGTTATTGTTGGTGGAGGTATTGCTAATCAAACCTGAT AAAAAGCTGCAGCTGAAAAAATTAATAAATTGGTTAAACAAGGTTTGTAAACTTTAC GCCGAGAGGCTTTATAGGAAGGACGGTACCATCCCAGTCGACGAGGAAAATAGAATT AGAATCGATGACTGGGAACCTTGAGGAGGACGTCCAGAAAAGCCGTTTCTGCCTTGATGG AGAAGGTCACCGGTGAAAATGCCGAGAGTTTGACCGACTTGCCGGTTACAGACACG ACTTCTTGCCAGTAACGGTTTTGACGTTGAGGGAATCAACTACGAGGCCGAAGTTGA GAGGTTCCACAGAATCTAA</p>
<i>BmPHI</i>	<p>ATGTTGACTACTGAATTTTTGGCTGAAATTGTTAAAGAATTGAATTCTTCTGTTAATCA AATTGCTGATGAAGAAGCTGAAGCTTTGGTTAATGGTATTTTGCAATCTAAAAAAGTTT TTGTTGCTGGTGTCTGGTAGATCTGGTTTTATGGCTAAATCTTTGCTATGAGAATGATG CATATGGGTATTGATGCTTATGTTGTTGGTGAACCTGTTACTCCAAATTATGAAAAAGA AGATATTTTGATTATTGGTTCTGGTTCTGGTGAACCTAAATCTTTGGTTTCTATGGCTCA AAAAGCTAAATCTATTGGTGGTACTATTGCTGCTGTTACTATTAATCCTGAATCTACTA TTGGTCAATTGGCTGATATTGTTATTAATAATGCCTGGTTCTCCAAAAGATAAATCTGAA GCTAGAGAAACTATTCAACCAATGGGATCTTTGTTTGAACAGACATTGTTATTGTTCTA TGATGCTGTTATCTTGAGATTTATGAAAAAAAAGGTTTGGATACTAAAACCTATGTAT GGTAGACATGCTAATTTGGAATAA</p>
<i>Bmgpx</i>	<p>ATGAGAGAATTGAAGTCAGAAAAAAGAGTTCAATCTTTGGCTATGGAATTTTTGTCTG TTGCTCAACAAGCTGCTTTGGCTCTTATCCATGGATTGGTAAAGGTAATAAAAAATGAA GTTGATAGAGCTGGTACTGAAGCTATGAGAAAATAGATTGAATTTGATTGATATGTCTG GTTTGATTGTTATTGGTGAAGGTGAAATGGATGAAGCTCCAATGTTGATATTTGGTGAA GAATTGGTACTGGTAAAGGTCCACAATTGGATATTGCTGTTGATCCTTGTTGATGGTGA TGGTTTGATGGCTAAAGGTATGGATAATTCTATTGCTGTTATTGCTGCTTCTACTAGAG GTTCTTTGTTGCATGCTCCTGATATGTATATGGAAAAAATTGCTGTTGGTCCAAAAGCT AAAGGTTGTGTTAATTTGGATGCTTCTTTGACTGAAAATATGAAATCTGTTGCTAAAGC TTTGGGTAAAGATTTGAGAGAATTAACCTGTTATGATTCAAGATAGACCAAGACATGAT CATTTGATTCAACAAGTTAGAGATGTTGGTGTAGATTGAAAATGTTTTCTGATGGTGA TGTTACTAGAGCTATTGGTACTGCTTTGGAAGAAGTTGATGTTGATATTTGGTTGGTA CTGGTGGTCTCCTGAAGGTGTTATCGCTGCTACTGCTTTGAAATGTTTGGTGGTGAT TTTCAAGGTAGATTGGCTCCTCAAAATGAAGAGGAATTTGACAGATGTATCACTATGG GTATTACTGACCCTAGAAAAATCTTACTATTGATGAAATTTGTTAAATCAGATGATTGT TTTTTTGTTGCTACTGGTATTACAGATGGTTTGTGATTAATGGTATTAGAAAAAAGA AGACGGTTTGATGCAAACCTCATT</p>
<i>PpAOX</i>	<p>ATGGCTATCCCCGAAGAGTTTGATATCCTAGTTCTAGGTGGTGGATCCAGTGGATCCTG TATTGCCGGAAGATTGGCAAACCTTGACCCTCTTGAAGTTGGTCTTATCGAAGCA GGTGAGAACAACCTCAACAACCCATGGTCTACCTCCAGGTATTTACCAAGAAACA TGAAGTTGGACTCCAAGACTGCTTCTTCTACACTTCAACCCATCCTCACTTGAAT GGTAGAAGAGCCATTGTTCCATGTGCTAACGTCTTGGGTGGTGGTTCTTCTATCAACTT CATGATGTACACCAGAGGTTCTGCTTCTGATTACGATGACTTCCAAGCCGAGGGCTGG AAAACCAAGGACTTGCTTCCATTGATGAAAAAGACTGAGACCTACCAAGAGCTTGCA ACAACCTGACATTCACGGTTTTCGAAGGTCCAATCAAGGTTTCTTTCCGGTAACTACACC TACCCAGTTTGCCAGGACTTCTTGAGGGCTTCTGAGTCCCAAGGTATTCCATACGTTGA CGACTTGAAGACTTGGTTACTGCTCAGGTGCTGAACACTGGTTGAAGTGGATCAAC AGAGACTGGTCTGCTTCCGACTCTGCTCATGCTCATTTGTCATTTGTCCTACTATGAGAAA CCACGACAACCTTGTACTTGTACTGTAACACGAAGGTGACAAAATTTGTCGAAGAC GGAAGAGCTGCTGCTGTTAGAACCGTTCCAAGCAAGCCTTTGAACCCAAAGAAGCCAA GTCACAAGATCTACCGTGCTAGAAAGCAAATCGTTTGTCTTGTGGTACCATCTCCTCT CCATTGGTTTTGCAAAGATCCGGTTTTGGTGACCCAATCAAGTTGAGAGCCGCTGGTGT TAAGCCTTTGGTCAACTTGCCAGGTGTCGGAAGAAAATTTCAAGACCACTACTGTTTCT TCAGTCTTACAGAATCAAGCCTCAGTACGAGTCTTTTCGATGACTTCGTCGGTGGTGT GCTGAGATTCAAAAGAGAGTCTTTGACCAATGGTACGCCAATGGTACTGCTCCTCTTG CCACTAACGGTATCGAAGCTGGTGTCAAGATCAGACCAACACCAGAAGAACTCTCTCA AATGGACGAATCCTTCCAGGAGGTTACAGAGAATACTTCCAAGACAAGCCAGACAA</p>

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FLS

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