

## Solid-State Aromatic Nucleophilic Fluorination: A Rapid, Practical, and Environmentally Friendly Route to *N*-Heteroaryl Fluorides

Koji Kubota,<sup>\*a,b</sup> Tetsu Makino,<sup>a</sup> Keisuke Kondo,<sup>a</sup> Tamae Seo,<sup>a</sup> Mingoo Jin<sup>b</sup> and Hajime Ito<sup>\*a,b</sup>

<sup>a</sup>Division of Applied Chemistry, Graduate School of Engineering, Hokkaido University, Sapporo, Hokkaido, 060-8628, Japan.

<sup>b</sup>Institute for Chemical Reaction Design and Discovery (WPI-ICReDD), Hokkaido University, Sapporo, Hokkaido, 001-0021, Japan.

e-mail: hajito@eng.hokudai.ac.jp, kbt@eng.hokudai.ac.jp

### Table of Contents

- 1. Chemicals and Instrumentation**
- 2. Substrate Preparation**
- 3. General Experimental Procedure**
- 4. Scaled-up Synthesis of 2r**
- 5. Thermography Observation for Reaction Temperature**
- 6. Calculation of E-factors**
- 7. Calculation of Reagents Cost**
- 8. Additional Optimization Study**
- 9. Comparison with Solution-based Protocols**
- 10. Purification Without Using Dichloromethane**
- 11. Characterization of Obtained Fluorinated Products**
- 12. References**
- 13. NMR spectra**

## 1. Chemicals and Instrumentation

Solvents and liquid additives were purchased from commercial suppliers and further dried over molecular sieve (MS 4Å). KF (spray-dried) was purchased from Wako Chemicals (166-13241). All mechanochemical reactions were carried out using grinding vessels in a Retsch MM400 mill. Both jars (1.5 mL, 5.0 mL, and 10 mL) and balls (7 mm, 10 mm, and 15 mm, diameter) are made of stainless (SUS400B and SUS420J2, respectively) (Figure S1 and S2). The heat gun Takagi, HG-1450B, with temperature control function was used (Figure S3). NMR spectra were recorded on JEOL JNM-ECZ400S and JNM-ECS400 spectrometers ( $^1\text{H}$ : 396 or 399 or 401 MHz,  $^{13}\text{C}$ : 99 or 100 or 101 MHz,  $^{19}\text{F}$ : 369 or 373 or 375 or 377 MHz). Tetramethylsilane ( $^1\text{H}$ ,  $\delta$ : 0.00),  $\text{CDCl}_3$  ( $^{13}\text{C}$ ,  $\delta$ : 77.0), and fluorobenzene ( $^{19}\text{F}$ ,  $\delta$ : -113.6) were employed as external standards, respectively. Multiplicity was recorded as follows: s = singlet, brs = broad singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sept = septet, o = octet, m = multiplet. Recycle preparative gel permeation chromatography (GPC) was conducted with a JAI LC-9101 using  $\text{CHCl}_3$  as an eluent with JAIGEL-1H. Thermography was recorded with an NEC Avio Thermo GEAR G120. High-resolution mass spectra were recorded at the Global Facility Center, Hokkaido University.



**Figure S1.** Retsch MM 400 used in this study.

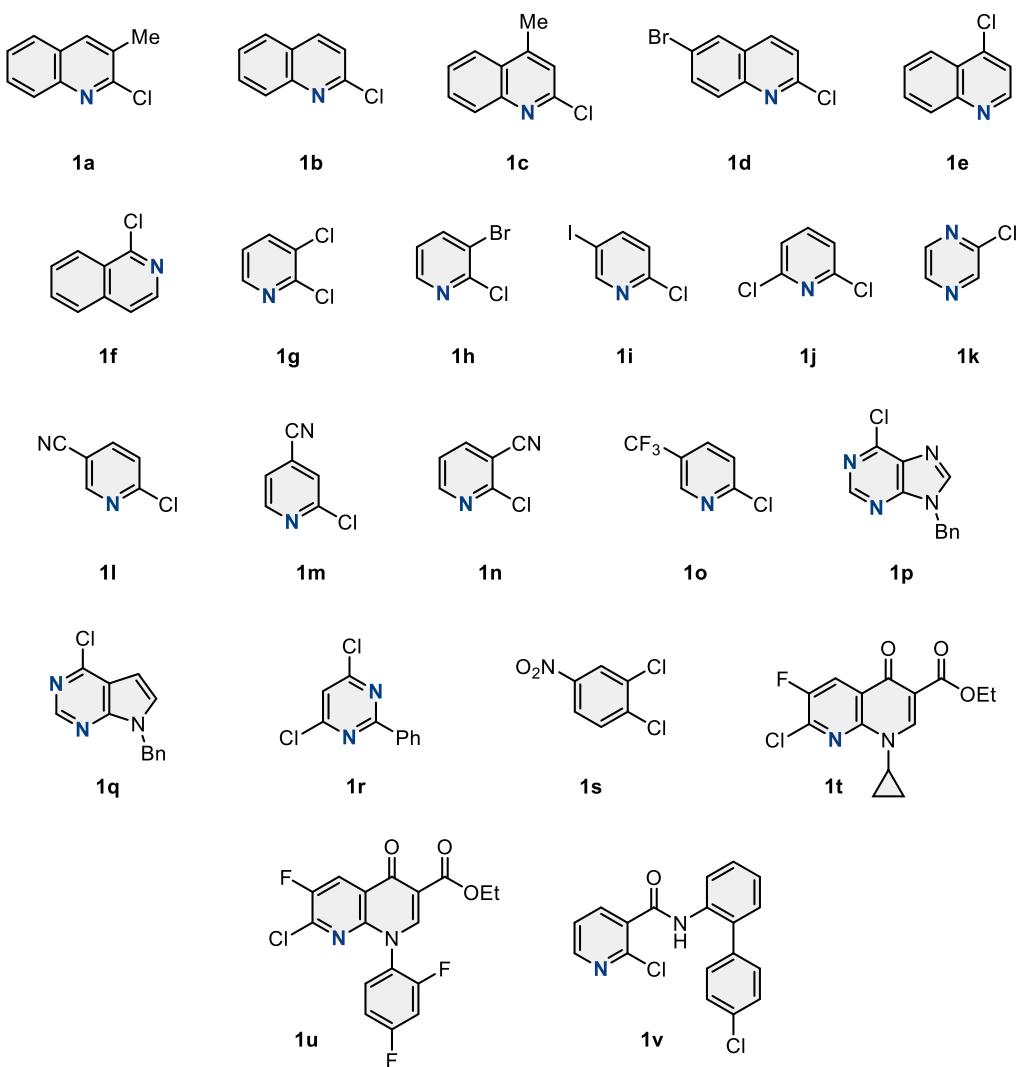


**Figure S2.** Stainless jars and balls used in this study.



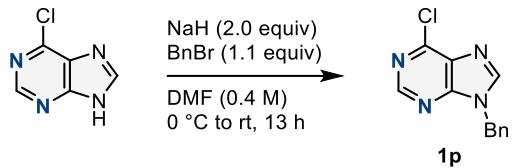
**Figure S3.** The temperature-controllable heat gun used in this study.

## 2. Substrate Preparation



**1a–1o, 1r, 1s, 1u and 1v** were purchased from commercial suppliers and used as received. **1p, 1q, and 1t** were prepared through the following procedures.

### Preparation of 9-benzyl-6-chloro-9H-purine (1p).

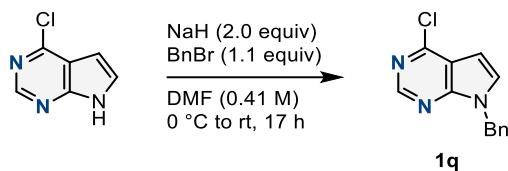


In a vacuum-dried reaction vial equipped with a magnetic stirring bar, 6-chloro-9H-purine (463.8 mg, 3.0 mmol) was dissolved in DMF (7.5 mL). NaH (60% dispersal in mineral oil, 358.5 mg, 6.5 mmol)

was added to the solution at 0 °C, and then stirred for 30 min. After 30 min, benzyl bromide (0.39 mL, 3.3 mmol) was added to the mixture. The mixture was allowed to warm to room temperature and stirred for 13 h. After the reaction, the mixture was quenched with water and extracted with 20% EtOAc/hexane three times. The organic layer was washed with brine and dried over anhydrous MgSO<sub>4</sub>. After filtration, the solvent was removed under vacuum, and the residue was purified by silica gel column chromatography (EtOAc/hexane, 30:70–40:60) to afford compound **1p** (250.9 mg, 1.0 mmol, 34%) as a white solid. <sup>1</sup>H and <sup>13</sup>C NMR were in agreement with the literature.<sup>1</sup>

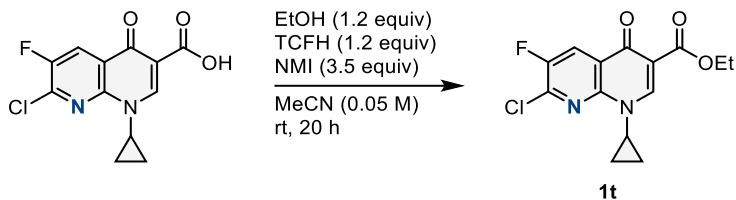
<sup>1</sup>H NMR (396 MHz, CDCl<sub>3</sub>, δ): 5.47 (s, 2H), 7.30–7.41 (m, 5H), 8.11 (s, 1H), 8.80 (s, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 47.8 (CH<sub>2</sub>), 127.9 (CH), 128.8 (CH), 129.2 (CH), 131.5 (C), 134.5 (C), 144.9 (CH), 151.1 (C), 151.8 (C), 152.1 (CH). HRMS-ESI (m/z): [M+Na]<sup>+</sup> calcd for C<sub>12</sub>H<sub>9</sub>ClN<sub>4</sub>Na, 267.0408; found, 267.0405.

#### Preparation of 7-benzyl-4-chloro-7H-pyrrolo[2,3-d]pyrimidine (**1q**).



In a vacuum-dried reaction vial equipped with a magnetic stirring bar, 4-chloro-1H-pyrrolo[2,3-d]pyrimidine (500.3 mg, 3.3 mmol) was dissolved in DMF (8.0 mL). NaH (60% dispersal in mineral oil, 262.8 mg, 6.6 mmol) was added to the solution at 0 °C, and then stirred for 30 min. After 30 min, benzyl bromide (0.43 mL, 3.6 mmol) was added to the mixture. The mixture was allowed to warm to room temperature and stirred for 17 h. After the reaction, the mixture was quenched with water and extracted with 20% EtOAc/hexane three times. The organic layer was washed with brine and dried over anhydrous MgSO<sub>4</sub>. After filtration, the solvent was removed under vacuum, and the residue was purified by silica gel column chromatography (Et<sub>2</sub>O/hexane, 0:100–15:85) to afford compound **1q** (561.5 mg, 2.3 mmol, 71%) as a white solid. <sup>1</sup>H and <sup>13</sup>C NMR were in agreement with the literature.<sup>2</sup> <sup>1</sup>H NMR (401 MHz, CDCl<sub>3</sub>, δ): 5.46 (s, 2H), 6.62 (d, *J* = 3.6 Hz, 1H), 7.21–7.23 (m, 3H), 7.30–7.36 (m, 3H), 8.68 (s, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 48.2 (CH<sub>2</sub>), 99.7 (CH), 117.2 (C), 127.4 (CH), 127.9 (CH), 128.7 (CH), 128.9 (CH), 136.0 (C), 150.6 (CH), 150.9 (C), 151.9 (C). HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd for C<sub>13</sub>H<sub>11</sub>ClN<sub>3</sub>, 244.0636; found, 244.0635.

**Preparation of ethyl-7-chloro-6-fluoro-1-cyclopropyl-1,4-dihydro-4-oxo-1,8-naphthyridine-3-carboxylate (**1t**).**

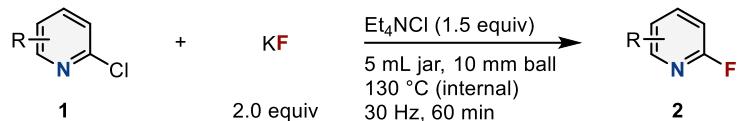


In a 100 mL two-necked round-bottomed flask equipped with a magnetic stirring bar, 7-chloro-1-cyclopropyl-6-fluoro-1,4-dihydro-4-oxo-1,8-naphthyridine-3-carboxylic acid (565.4 mg, 2.0 mmol) was dissolved in MeCN (38 mL). EtOH (0.14 mL, 2.4 mmol), *N*-methyimidazole (NMI) (0.56 mL, 7.0 mmol), and chloro-*N,N,N',N'*-tetramethylformamidinium hexafluorophosphate (TCFH) (673.9 mg, 2.4 mmol) were added to the solution, then stirred at room temperature for 20 h. After the reaction, the solution was diluted with water and extracted with CH<sub>2</sub>Cl<sub>2</sub> three times. The organic layer was washed with brine and dried over MgSO<sub>4</sub>. After filtration, the solvent was removed under vacuum. The residue was washed with water to remove tetramethylurea and purified by silica gel column chromatography (EtOAc/hexane, 20:80–100:0) to afford compound **1t** (533.4 mg, 1.7 mmol, 86%) as a white powder. <sup>1</sup>H NMR was in agreement with the literature.<sup>3</sup>

<sup>1</sup>H NMR (401 MHz, CDCl<sub>3</sub>, δ): 1.04–1.09 (m, 2H), 1.29–1.36 (m, 2H), 1.41 (t, *J* = 7.2 Hz, 3H), 3.66 (tt, *J* = 3.2, 4.0 Hz, 1H), 4.41 (q, *J* = 7.2 Hz, 2H), 8.44 (d, *J* = 7.2 Hz, 1H), 8.66 (s, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 7.5 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>), 34.3 (CH), 61.0 (CH<sub>2</sub>), 111.8 (C), 122.8 (d, *J* = 21.0 Hz, CH), 123.6 (d, *J* = 2.8 Hz, C), 142.2 (d, *J* = 21.9 Hz, C), 145.7 (d, *J* = 1.9 Hz, C), 148.8 (CH), 152.4 (d, *J* = 261.3 Hz, C), 164.5 (C), 173.0 (C). HRMS-EI (m/z): [M]<sup>+</sup> calcd for C<sub>14</sub>H<sub>12</sub>ClFN<sub>2</sub>O<sub>3</sub>, 310.0515; found, 310.0516.

### 3. General Experimental Procedure

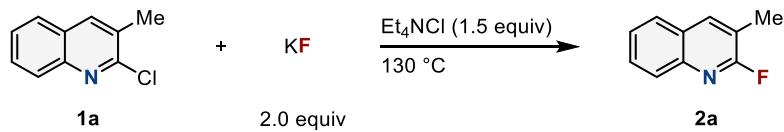
#### 1) Mechanochemical S<sub>N</sub>Ar fluorination



**Procedure A:** Heteroaryl chloride **1** (0.50 mmol), KF (1.0 mmol, 2.0 equiv), and Et<sub>4</sub>NCl (0.75 mmol, 1.5 equiv) were placed in a ball milling vessel (stainless, 5 mL) loaded with one grinding ball (stainless, diameter: 10 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 60 min at 30 Hz) and a heat gun (preset temperature at 250 °C). After 60 min, the jar was cooled rapidly with cold water and opened. The mixture was passed through a short silica gel column eluting with 50% EtOAc/CH<sub>2</sub>Cl<sub>2</sub>. The crude mixture was purified by flash column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane, typically 0:100–5:95) to give the corresponding fluorinated product **2**.

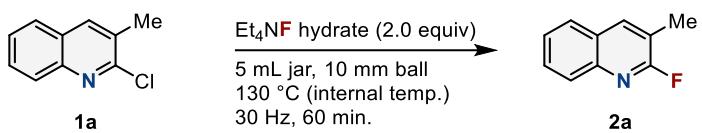
**Procedure B:** Heteroaryl chloride **1** (0.50 mmol), KF (1.0 mmol, 2.0 equiv), and Et<sub>4</sub>NCl (0.75 mmol, 1.5 equiv) were placed in a ball milling vessel (stainless, 1.5 mL) loaded with one grinding ball (stainless, diameter: 7 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 30–60 min at 30 Hz) and a heat gun (preset temperature at 250 °C). After 30–60 min, the jar was then cooled and opened. The mixture was passed through a short silica gel column eluting with 50% EtOAc/CH<sub>2</sub>Cl<sub>2</sub>. The crude mixture was purified by flash column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane, typically 0:100–20:80) to give the corresponding fluorinated product **2**.

#### 2) S<sub>N</sub>Ar fluorination reaction under neat conditions



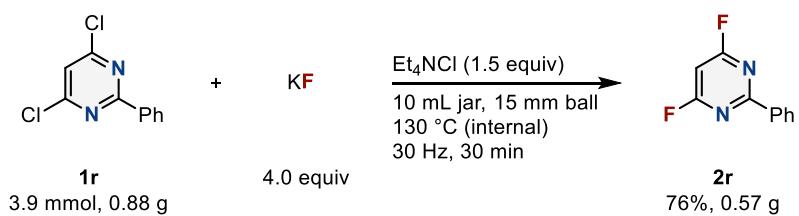
In a vacuum-dried test tube equipped with a magnetic stirring bar, 2-chloro-3-methylquinoline **1a** (0.50 mmol), KF (1.0 mmol, 2.0 equiv) and Et<sub>4</sub>NCl (0.75 mmol, 1.5 equiv) were added. The mixture was warmed to 130 °C and stirred for 60 min or 24 h. After the reaction, the mixture was cooled to room temperature and diluted with water, then extracted with Et<sub>2</sub>O three times. The organic layer was washed with brine and dried over anhydrous MgSO<sub>4</sub>. The yield was determined by <sup>19</sup>F NMR analysis with fluorobenzene as an internal standard.

### 3) Mechanochemical S<sub>N</sub>Ar fluorination reaction using ammonium fluoride



2-Chloro-3-methylquinoline **1a** (88.6 mg, 0.50 mmol) and Et<sub>4</sub>NF hydrate (150 mg, 1.0 mmol, 2.0 equiv based on anhydrous) were placed in a ball milling vessel (stainless, 5 mL) loaded with one grinding ball (stainless, diameter: 10 mm) in a glovebox. After the vessel was closed purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 60 min at 30 Hz) and heat gun (preset temperature at 250 °C). After 60 min, the jar was then cooled rapidly with cold water and opened. The mixture was passed through a short silica gel column eluting with 50% EtOAc/CH<sub>2</sub>Cl<sub>2</sub>. The crude mixture was analyzed using <sup>19</sup>F NMR with fluorobenzene (15.5 mg) as an internal standard to obtain a NMR yield of **2a** in 61% yield.

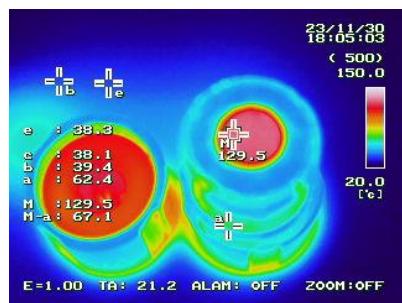
#### 4. Scaled-up Synthesis of **2r**



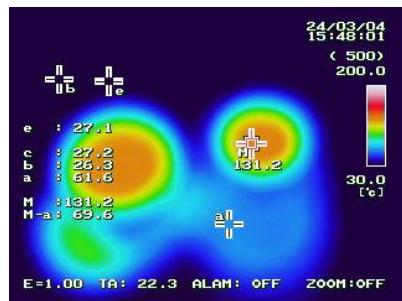
Fenclorim **1r** (0.877 g, 3.9 mmol), KF (0.907 g, 15.6 mmol, 4.0 equiv) and Et<sub>4</sub>NCl (0.970 g, 5.85 mmol, 1.5 equiv) were placed in a ball milling vessel (stainless, 10 mL) loaded with grinding ball (stainless, diameter: 15 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 30 min at 30 Hz) and a heat gun (preset temperature at 250 °C). After 30 min, the jar was then cooled rapidly with cold water and opened. The mixture was diluted with water and then extracted with EtOAc three times. The organic layer was combined and dried over anhydrous MgSO<sub>4</sub>. After filtration, the solvent was removed under vacuum, and the residue was purified by silica gel column chromatography (Et<sub>2</sub>O/hexane, 0:100–2:98) to afford compound **2r** (569.6 mg, 2.96 mmol, 76%) as a white solid.

## 5. Thermography Observation for Reaction Temperature

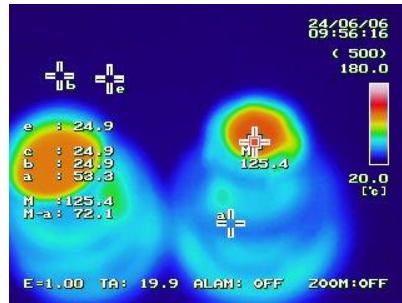
The temperature inside the milling jar after the mechanochemical S<sub>N</sub>Ar fluorination reactions were confirmed by observation with a thermography camera immediately after opening the milling jar. When the pre-set temperature of the heat gun was 250 °C for a 5 mL stainless jar with a 10 mm ball (30 Hz, 60 min), the internal temperature was determined to be 129.5 °C (Figure S4). Similarly, for a 10 mL stainless jar with a 15 mm ball (30 Hz, 30 min), the internal temperature was 131.2 °C (Figure S5). For a 1.5 mL stainless jar with a 7 mm ball (30 Hz, 30 min), the internal temperature was 125.4 °C (Figure S6).



**Figure S4.** Thermography image inside the milling jar (5 mL) after grinding for 60 min at 30 Hz.



**Figure S5.** Thermography image inside the milling jar (10 mL) after grinding for 30 min at 30 Hz.

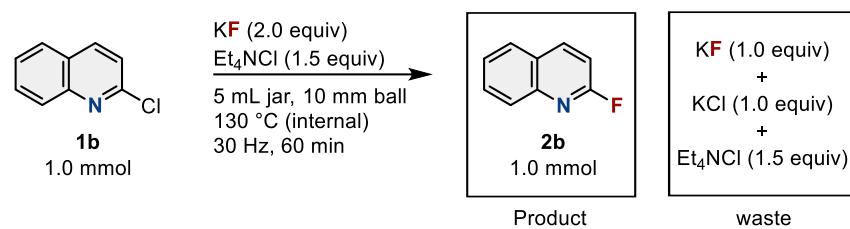


**Figure S6.** Thermography image inside the milling jar (1.5 mL) after grinding for 30 min at 30 Hz.

## 6. Calculation of E-factors

The E-factors were calculated for the present solid-state conditions and compared to those of Bland's and Sanford's conditions following literature procedures.<sup>4,5</sup>

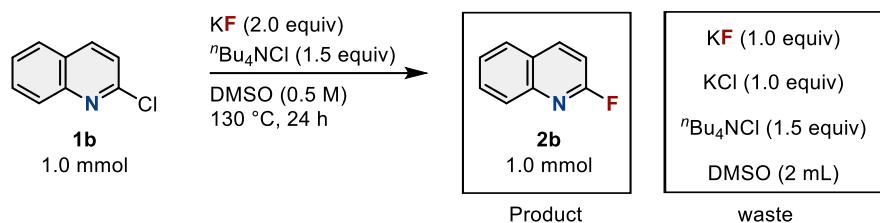
### < Our solid-state fluorination conditions >



**Table S1.** E-factor of our solid-state fluorination conditions.

this work	Mw	mmol	mg
<b>product</b>			
<b>2b</b>	147.15	1.0	147.15
<b>waste</b>			
KF	58.1	1.0	58.1
KCl	74.55	1.0	74.55
Et <sub>4</sub> NCl	165.71	1.5	248.565
total			381.215
E-factor			2.6

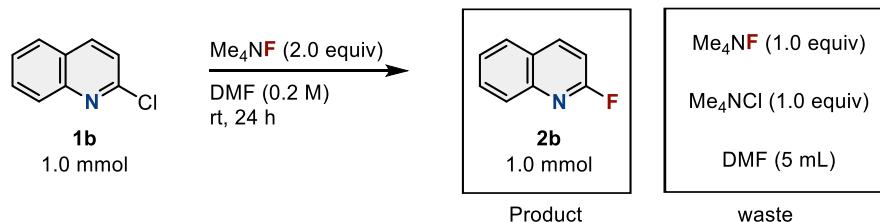
< Bland's fluorination conditions<sup>4</sup> >



**Table S2.** E-factor of Bland's fluorination conditions.

Bland, 2014	Mw	mmol	mg
<b>product</b>			
<b>2b</b>	147.15	1.0	147.15
<b>waste</b>			
KF	58.1	1.0	58.1
KCl	74.55	1.0	74.55
$n\text{Bu}_4\text{NCl}$	277.92	1.5	416.88
DMSO			2200
total			2749.52
E-factor			18.7

< Sanford's fluorination conditions<sup>5</sup> >

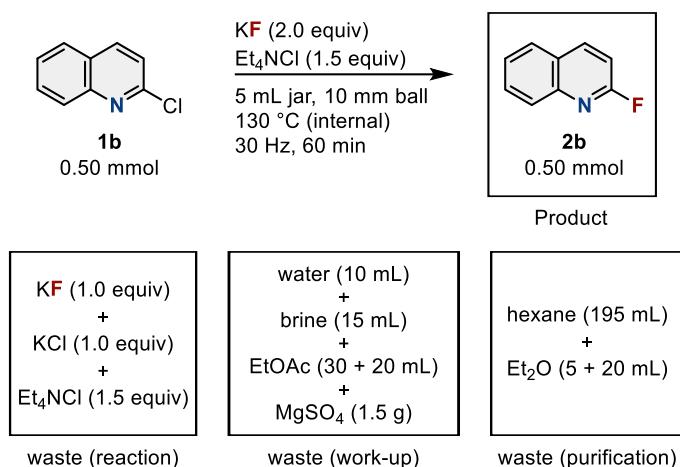


**Table S3.** E-factor of Sanford's fluorination conditions.

Sanford, 2015	Mw	mmol	mg
<b>product</b>			
<b>2b</b>	147.15	1.0	147.15
<b>waste</b>			
Me <sub>4</sub> NF	93.14	1.0	93.14
Me <sub>4</sub> NCl	109.6	1.0	109.6
DMF			4720
total			4922.74
E-factor			33.5

We recalculated the E-factor for the reaction of **1b**, taking into account the purification step. As shown below, the E-factor is 3019.92. We also calculated the E-factor for the large-scale mechanochemical synthesis of **2r** and its purification, which is 1252.73. Importantly, it should be noted here that our research aims to reduce the quantity of reaction solvents, and the current workup/purification procedure is not optimal from a sustainability perspective. Although this was not the focus of this study, it must be considered when developing industrial mechanochemical protocols.

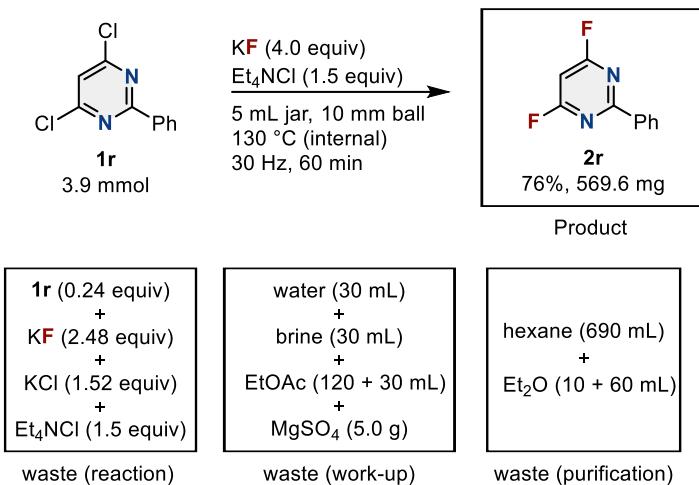
< E-factor for the reaction of **1b** considering the purification process >



**Table S4.** E-factor of our solid-state fluorination conditions considering the purification step.

	Mw	mmol	g
<b>product</b>			
<b>2b</b>	147.15	0.50	0.074
<b>waste (Reaction)</b>			
KF	58.10	0.50	0.029
KCl	74.55	0.50	0.037
Et <sub>4</sub> NCl	165.71	0.75	0.124
<b>waste (work-up)</b>			
water		10	
brine		18	
EtOAc		45	
MgSO <sub>4</sub>		1.5	
<b>waste (purification)</b>			
hexane		130	
Et <sub>2</sub> O		17.5	
<b>total waste</b>		222.191	
E-factor (total)		3019.92	

< E-factor for the reaction of **1r** considering the purification process >



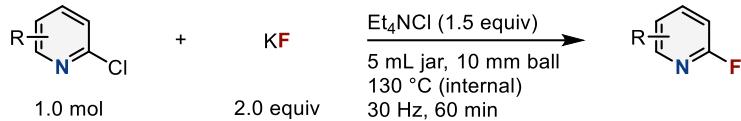
**Table S5.** E-factor of our solid-state fluorination conditions on a gram scale considering the purification step.

this work	Mw	mmol	g
<b>product</b>			
<b>2r</b>	192.17	2.96	0.569
<b>waste (Reaction)</b>			
<b>1r</b>	225.07	0.94	0.211
KF	58.10	9.67	0.562
KCl	74.55	5.93	0.442
Et <sub>4</sub> NCl	165.71	5.85	0.969
<b>waste (work-up)</b>			
water		30	
brine		36	
EtOAc		135	
MgSO <sub>4</sub>		5	
<b>waste (purification)</b>			
hexane		455.4	
Et <sub>2</sub> O		49	
<b>total waste</b>		712.584	
E-factor (reaction)		3.839	
E-factor (total)		1252.73	

## 7. Calculation of Reagents Cost

The reagent costs were calculated on a 1.0 mol scale reaction under the present solid-state conditions and compared to those of Bland's and Sanford's conditions.<sup>4,5</sup> Retail prices are for July 2024. The results showed that our method is much less inexpensive S<sub>N</sub>Ar fluorination system.

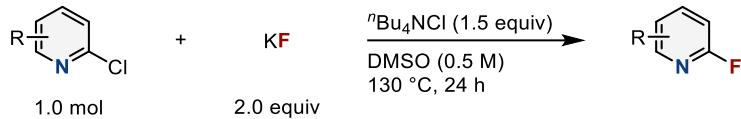
### < Our solid-state fluorination conditions >



**Table S6. Total reagents cost of our solid-state fluorination conditions.**

This work	amount (mol or L)	cost (JPY)
KF (Wako, 166-13241)	2	2092
Et <sub>4</sub> NCl (TCI, T0095)	1.5	12279
Total cost		14371 (ca. US \$89)

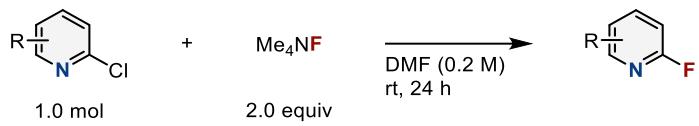
### < Bland's fluorination conditions<sup>4</sup> >



**Table S7. Total reagents cost of Bland's conditions.**

Bland, 2014	amount (mol or L)	cost (JPY)
KF (Wako, 166-13241)	2	2092
<sup>n</sup> Bu <sub>4</sub> NCl (TCI, T0055)	1.5	121729
DMSO (KANTO, 10380-05)	2	34000
Total cost		157821 (ca. US \$977)

< Sanford's fluorination conditions<sup>5</sup> >



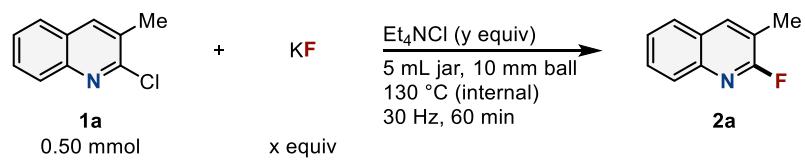
**Table S8. Total reagents cost of Sanford's conditions.**

Sanford, 2015	amount (mol or L)	cost (JPY)
Me <sub>4</sub> NF (Sigma-Aldrich, 459135)	2	1818093
DMF (KANTO, 11339-05)	5	51000
Total cost	1869093 (ca. US \$11571)	

## 8. Additional Optimization Study

2-Chloro-3-methylquinoline (**1a**, 0.50 mmol), KF (1.0–2.0 equiv), and Et<sub>4</sub>NCl (0–1.5 equiv) were placed in a ball-milling vessel (stainless steel, 5 mL) loaded with one grinding ball (stainless steel, diameter: 10 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 60 min, 30 Hz) and a heat gun (preset temperature: 250 °C). After 60 min, the jar was cooled rapidly with cold water and opened. The mixture was passed through a short column of silica gel using EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (1/1, v/v). The crude mixture was analyzed using <sup>19</sup>F NMR spectroscopy with fluorobenzene as the internal standard. When reducing the amount of KF and Et<sub>4</sub>NCl, the yield of **2a** decreased. The use of KF (2.0 equiv) and Et<sub>4</sub>NCl (1.5 equiv) is required for an efficient fluorination.

**Table S9.** Additional optimization study.



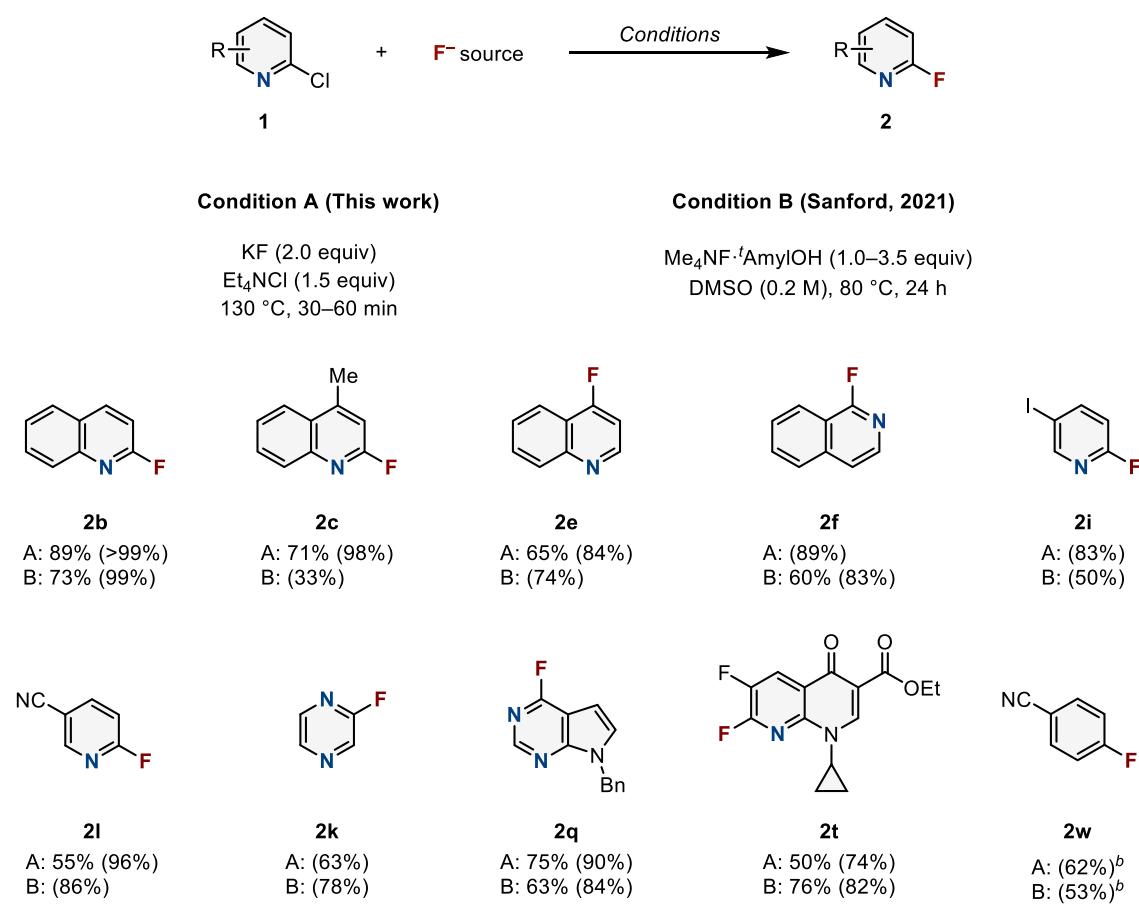
Entry	KF (x equiv)	Et <sub>4</sub> NCl (y equiv)	yield of <b>2a</b> (%) <sup>a</sup>
1	2.0	1.5	>99
2	1.5	1.5	70
3	1.0	1.5	27
4	2.0	0.5	79
5	2.0	0.1	29
6	2.0	0	n.d.

<sup>a</sup><sup>19</sup>F NMR yields determined with fluorobenzene as an internal standard.

## 9. Comparison with Solution-based Protocols

We compared the yields of the present mechanochemical conditions, and the solution-based conditions reported by Sanford, which is one of the most practical S<sub>N</sub>Ar fluorination protocols.<sup>6</sup> As shown below, in these ten examples, there are no significant differences in yield between mechanochemical (condition A) and solution-based (condition B) reactions. For **2c** and **2i**, the mechanochemical protocol is much better than the solution-based method.

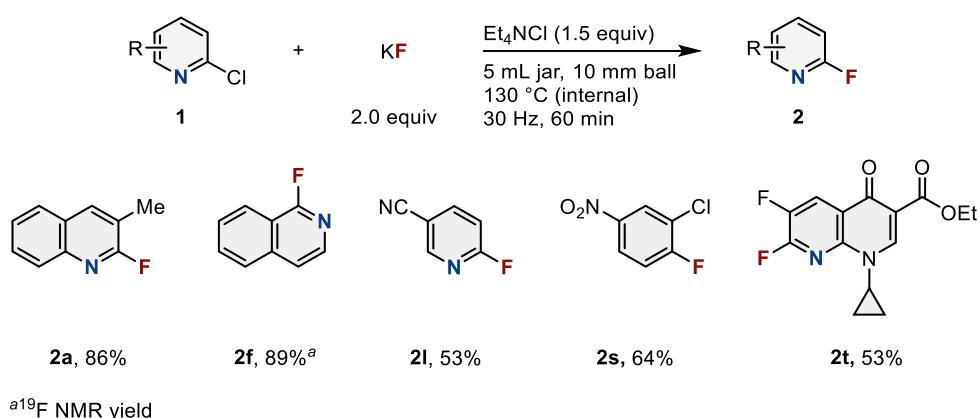
**Table S10.** Comparison with solution-based protocols.



<sup>a</sup>Isolated yields. <sup>19</sup>F NMR yields are given in parentheses. <sup>b</sup>From nitroarene

## 10. Purification Without Using Dichloromethane

To enhance the sustainability of this mechanochemical protocol, we conducted the purification process without using dichloromethane. We used EtOAc for the purification step in reactions involving five different substrates. The results showed that the yields of these products were similar to those of the reactions using dichloromethane ( $\pm 5\%$ ), suggesting that dichloromethane can be replaced by EtOAc in the purification step.



**Procedure C (for 1a and 1t):** Heteroaryl chloride **1** (0.50 mmol), KF (1.0 mmol, 2.0 equiv), and Et<sub>4</sub>NCl (0.75 mmol, 1.5 equiv) were placed in a ball-milling vessel (stainless steel, 5 mL) loaded with one grinding ball (stainless steel, diameter: 10 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 60 min, 30 Hz) and a heat gun (preset temperature: 250 °C). After 60 min, the jar was cooled rapidly with cold water and opened. The mixture was transferred to a separatory funnel with EtOAc (10 mL) and water (10 mL) and then extracted with EtOAc (3 × 10 mL). The combined organic layer was washed with brine (15 mL) and dried over anhydrous MgSO<sub>4</sub>. After filtration, the solvent was removed from the filtrate under reduced pressure. The crude mixture was purified by flash column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane) to give the corresponding fluorinated product (**2**).

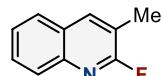
**Procedure D (for 1f):** **1f** (0.50 mmol), KF (1.0 mmol, 2.0 equiv), and Et<sub>4</sub>NCl (0.75 mmol, 1.5 equiv) were placed in a ball milling vessel (stainless steel, 5 mL) loaded with one grinding ball (stainless steel, diameter: 10 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 60 min, 30 Hz) and a heat gun (preset temperature: 250 °C). After 60 min, the jar was cooled rapidly with cold water and opened. The mixture was transferred to a separatory funnel with EtOAc (10 mL) and water (10 mL), then extracted with EtOAc (3 × 10 mL). The combined organic layer was washed with brine (15 mL) and dried over anhydrous MgSO<sub>4</sub>. After

filtration, the solvent was removed from the filtrate under reduced pressure. The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR spectroscopy with fluorobenzene as the internal standard.

**Procedure E (for **1l** and **1s**):** Heteroaryl chloride **1** (0.50 mmol), KF (1.0 mmol, 2.0 equiv), and Et<sub>4</sub>NCl (0.75 mmol, 1.5 equiv) were placed in a ball-milling vessel (stainless steel, 1.5 mL) loaded with one grinding ball (stainless steel, diameter: 7 mm). After the vessel was closed in air without purging with inert gas, the vessel was placed in the ball mill (Retsch MM400, 30 or 45 min, 30 Hz) and a heat gun (preset temperature: 250 °C). After 30 or 45 min, the jar was then cooled and opened. The mixture was transferred to a separatory funnel with EtOAc (10 mL) and water (10 mL), and extracted with EtOAc (3 × 10 mL). The combined organic layers were washed with brine (15 mL) and dried over anhydrous MgSO<sub>4</sub>. After filtration, the solvent was removed from the filtrate under reduced pressure. The crude mixture was purified by flash column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane) to give the corresponding fluorinated product (**2**).

## 11. Characterization of Obtained Fluorinated Products

### 2-Fluoro-3-methylquinoline (2a).

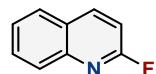


**2a**

The reaction was performed according to the general procedure A. The reaction was carried out with 88.9 mg (0.50 mmol) of **1a**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.3 mg) as an internal standard to obtain the NMR yield of **2a** in >99% yield. Product **2a** was obtained as a white solid (68.8 mg, 0.43 mmol, 85% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–5:95).

$^1\text{H}$  NMR (396 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 2.45 (s, 3H), 7.48–7.54 (m, 1H), 7.63–7.70 (m, 1H), 7.77 (d,  $J$  = 7.9 Hz, 1H), 7.91 (d,  $J$  = 8.7 Hz, 1H), 8.02 (d,  $J$  = 9.9 Hz, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 15.1 ( $\text{CH}_3$ ), 120.0 (d,  $J$  = 36.2 Hz, C), 125.9 (d,  $J$  = 2.0 Hz, CH), 126.6 (CH), 127.3 (d,  $J$  = 2.0 Hz, C), 127.6 (CH), 129.3 (CH), 140.4 (d,  $J$  = 7.6 Hz, CH), 144.2 (d,  $J$  = 17.2 Hz, C), 160.4 (d,  $J$  = 243.2 Hz, C).  $^{19}\text{F}$  NMR (373 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -66.5 (s). HRMS-APCI (m/z):  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{10}\text{H}_9\text{FN}$ , 161.0714; found, 161.0708.

### 2-Fluoroquinoline (2b).



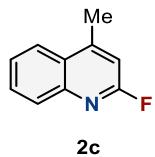
**2b**

The reaction was performed according to the general procedure A. The reaction was carried out with 81.8 mg (0.50 mmol) of **1b**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (14.6 mg) as an internal standard to obtain the NMR yield of **2b** in >99% yield. Product **2b** was obtained as a colorless oil (65.3 mg, 0.44 mmol, 89% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–10:90) and recycling preparative GPC.  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>6</sup>

The reaction was carried out with 104.2 mg (0.50 mmol) of 2-bromoquinoline (**1b'**). The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.2 mg) as an internal standard to obtain the NMR yield of **2b** in >99% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>6</sup>  $^1\text{H}$  NMR (401 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.09 (dd,  $J$  = 2.8, 8.8 Hz, 1H), 7.55 (t,  $J$  = 7.6 Hz, 1H), 7.74 (t,  $J$  = 7.4 Hz, 1H), 7.85 (d,  $J$  = 8.0 Hz, 1H), 7.96 (d,  $J$  = 8.8 Hz, 1H), 8.26 (t,  $J$  = 8.6 Hz, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 110.0 (d,  $J$  = 42.2 Hz, CH), 126.1 (d,  $J$  = 2.9 Hz, CH), 126.8 (d,  $J$  = 1.9 Hz, C), 127.5 (CH), 128.0 (CH), 130.6 (CH), 141.9 (d,  $J$  = 10.5 Hz, CH), 145.7 (d,  $J$  = 16.3 Hz, C), 161.1 (d,  $J$  = 241.6 Hz, C).  $^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -62.2 (s). HRMS-EI (m/z):  $[\text{M}]^+$  calcd for  $\text{C}_9\text{H}_6\text{FN}$ ,

147.0479; found, 147.0479.

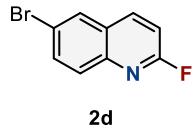
**2-Fluoro-4-methylquinoline (2c).**



The reaction was performed according to the general procedure A. The reaction was carried out with 88.8 mg (0.50 mmol) of **1c**. The resulting crude mixture was analyzed by <sup>19</sup>F NMR with fluorobenzene (15.2 mg) as an internal standard to obtain the NMR yield of **2c** in 98% yield. Product **2c** was obtained as a colorless oil (57.6 mg, 0.36 mmol, 71% yield, isolated along with approximately 6% of starting material) after purification by silica-gel column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane, 0:100–5:95). <sup>1</sup>H and <sup>19</sup>F NMR were in agreement with the literature.<sup>7</sup>

<sup>1</sup>H NMR (396 MHz, CDCl<sub>3</sub>, δ): 2.73 (s, 3H), 6.93 (s, 1H), 7.52–7.57 (m, 1H), 7.72 (t, *J* = 7.5 Hz, 1H), 7.96 (t, *J* = 9.3 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 18.9 (d, *J* = 2.9 Hz, CH<sub>3</sub>), 109.9 (d, *J* = 41.9 Hz, CH), 123.7 (CH), 125.73 (CH), 125.75 (C), 128.5 (CH), 130.2 (CH), 145.5 (d, *J* = 17.2 Hz, C), 151.0 (d, *J* = 10.0 Hz, C), 160.9 (d, *J* = 241.3 Hz, C). <sup>19</sup>F NMR (377 MHz, CDCl<sub>3</sub>, δ): -63.5 (s). HRMS-EI (m/z): [M]<sup>+</sup> calcd for C<sub>10</sub>H<sub>8</sub>FN, 161.0635; found, 161.0636.

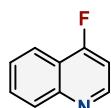
**6-Bromo-2-fluoroquinoline (2d).**



The reaction was performed according to the general procedure A. The reaction was carried out with 121.9 mg (0.50 mmol) of **1d**. The resulting crude mixture was analyzed by <sup>19</sup>F NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2d** in 88% yield. Product **2d** was obtained as a white solid (80.2 mg, 0.35 mmol, 71% yield) after purification by silica-gel column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane, 0:100–5:95). <sup>1</sup>H, <sup>13</sup>C, and <sup>19</sup>F NMR were in agreement with the literature.<sup>6</sup>

<sup>1</sup>H NMR (396 MHz, CDCl<sub>3</sub>, δ): 7.13 (dd, *J* = 2.8, 9.1 Hz, 1H), 7.79–7.85 (m, 2H), 8.02 (d, *J* = 2.0 Hz, 1H), 8.18 (t, *J* = 8.5 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 111.0 (d, *J* = 42.0 Hz, CH), 119.8 (d, *J* = 2.9 Hz, C), 127.8 (d, *J* = 1.9 Hz, C), 129.5 (CH), 129.6 (d, *J* = 16.2 Hz, CH), 133.9 (CH), 140.9 (d, *J* = 9.6 Hz, CH), 144.3 (d, *J* = 17.2 Hz, C), 161.1 (d, *J* = 244.1 Hz, C). <sup>19</sup>F NMR (373 MHz, CDCl<sub>3</sub>, δ): -61.3 (s). HRMS-EI (m/z): [M]<sup>+</sup> calcd for C<sub>9</sub>H<sub>5</sub>BrFN, 224.9584; found, 224.9589.

**4-Fluoroquinoline (2e).**

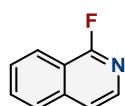


**2e**

The reaction was performed according to the general procedure A. The reaction was carried out with 81.8 mg (0.50 mmol) of **1e**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.3 mg) as an internal standard to obtain the NMR yield of **2e** in 84% yield. Product **2e** was obtained as a yellow oil (48.2 mg, 0.33 mmol, 65% yield, isolated along with approximately 3% of starting material) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–20:80).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>8</sup>

$^1\text{H}$  NMR (396 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.11 (dd,  $J = 5.1, 9.5$  Hz, 1H), 7.59–7.65 (m, 1H), 7.76–7.82 (ddd,  $J = 1.6, 6.9, 8.5$  Hz, 1H), 8.11–8.15 (m, 2H), 8.88 (dd,  $J = 4.9, 8.1$  Hz, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 105.6 (d,  $J = 15.4$  Hz, CH), 119.5 (d,  $J = 12.5$  Hz, C), 120.4 (d,  $J = 4.8$  Hz, CH), 126.8 (CH), 129.1 (d,  $J = 3.8$  Hz, CH), 130.5 (CH), 150.4 (d,  $J = 4.8$  Hz, C), 151.4 (d,  $J = 7.7$  Hz, CH), 165.2 (d,  $J = 268.4$  Hz, C).  $^{19}\text{F}$  NMR (373 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –113.2 (s). HRMS-EI (m/z): [M]<sup>+</sup> calcd for  $\text{C}_9\text{H}_6\text{FN}$ , 147.0479; found, 147.0482.

**1-Fluoroisoquinoline (2f).**



**2f**

The reaction was performed according to the general procedure A. The reaction was carried out with 81.8 mg (0.50 mmol) of **1f**. The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2f** in 89% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>6</sup>

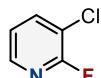
The reaction was carried out with 104.1 mg (0.50 mmol) of 1-bromoisoquinoline (**1f'**). The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.0 mg) as an internal standard to obtain the NMR yield of **2f** in 82% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>6</sup>

The reaction was carried out with 127.6 mg (0.50 mmol) of 1-iodoisoquinoline (**1f''**). The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.4 mg) as an internal standard to obtain the NMR yield of **2f** in >99% yield. Product **2f** was obtained as a yellow oil (62.7 mg, 0.43 mmol, 85% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{pentane}$ , 0:100–10:90).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>6</sup>

$^1\text{H}$  NMR (399 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.53 (dd,  $J = 1.4, 5.8$  Hz, 1H), 7.63–7.69 (m, 1H), 7.74–7.80 (m, 1H),

7.87 (d,  $J$  = 8.0 Hz, 1H), 8.06 (dd,  $J$  = 1.2, 5.6 Hz, 1H), 8.17 (d,  $J$  = 8.4 Hz, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 117.6 (d,  $J$  = 31.7 Hz, C), 119.3 (d,  $J$  = 4.8 Hz, CH), 123.0 (CH), 126.3 (d,  $J$  = 3.8 Hz, CH), 127.9 (CH), 131.4 (CH), 139.1 (d,  $J$  = 16.3 Hz, CH), 139.5 (d,  $J$  = 5.7 Hz, C), 159.9 (d,  $J$  = 246.3 Hz, C).  $^{19}\text{F}$  NMR (375 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -71.7 (s). HRMS-EI (m/z): [M]<sup>+</sup> calcd for  $\text{C}_9\text{H}_6\text{FN}$ , 147.0479; found, 147.0483.

### 3-Chloro-2-fluoropyridine (**2g**).

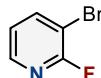


**2g**

The reaction was performed according to the general procedure A. The reaction was carried out with 73.8 mg (0.50 mmol) of **1g**. The resulting crude mixture was diluted with  $\text{CDCl}_3$  and then analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.5 mg) as an internal standard to obtain the NMR yield of **2g** in 66% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>7</sup> Since **2g** is a volatile compound, only the NMR yield was reported.

$^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -71.1 (s).

### 3-Bromo-2-fluoropyridine (**2h**).

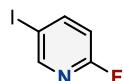


**2h**

The reaction was performed according to the general procedure A. The reaction was carried out with 96.0 mg (0.50 mmol) of **1h**. The resulting crude mixture was diluted with  $\text{CDCl}_3$  and then analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (14.9 mg) as an internal standard to obtain the NMR yield of **2h** in 82% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>9</sup> Since **2h** is a volatile compound, only the NMR yield was reported.

$^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -65.1 (s).

### 2-Fluoro-4-iodopyridine (**2i**).



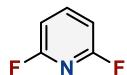
**2i**

The reaction was performed according to the general procedure A. The reaction was carried out with 119.7 mg (0.50 mmol) of **1i**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2i** in 83% yield.  $^{19}\text{F}$  NMR was in agreement with an authentic sample (F0773, Tokyo Chemical Industry Co.). Since **2i** is a

volatile compound, only the NMR yield was reported.

$^{19}\text{F}$  NMR (373 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –70.1 (s).

### 2,6-Difluoropyridine (2j).



2j

The reaction was performed according to the general procedure A. The reaction was carried out with 74.0 mg (0.50 mmol) of **1j**, 116.7 mg (2.0 mmol) of KF, and 248.8 mg (1.5 mmol) of  $\text{Et}_4\text{NCl}$ . The resulting crude mixture was diluted with  $\text{CDCl}_3$  and then analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2j** in 75% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>10</sup> Since **2j** is a volatile compound, only the NMR yield was reported.

$^{19}\text{F}$  NMR (373 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –68.1 (s).

### 2-Fluoropyrazine (2k).

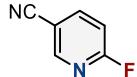


2k

The reaction was performed according to the general procedure A. The reaction was carried out with 57.4 mg (0.50 mmol) of **1k**. The resulting crude mixture was diluted with  $\text{CDCl}_3$  and then analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (14.9 mg) as an internal standard to obtain the NMR yield of **2k** in 63% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>11</sup> Since **2k** is a volatile compound, only the NMR yield was reported.

$^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –79.4 (s).

### 5-Cyano-2-fluoropyridine (2l).



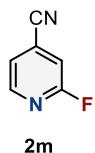
2l

The reaction was performed according to the general procedure B. The reaction was carried out with 69.2 mg (0.50 mmol) of **1l** for 30 min. The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2l** in 96% yield. Product **2l** was obtained as a colorless solid (33.5 mg, 0.27 mmol, 55% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–20:80).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>12</sup>

$^1\text{H}$  NMR (396 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.08–7.13 (m, 1H), 8.05–8.13 (m, 1H), 8.59 (d,  $J$  = 2.4 Hz, 1H).  $^{13}\text{C}$

NMR (100 MHz, CDCl<sub>3</sub>, δ): 108.0 (d, *J* = 4.8 Hz, *C*), 110.8 (d, *J* = 37.3 Hz, CH), 115.5 (*C*), 144.4 (d, *J* = 9.6 Hz, CH), 152.2 (d, *J* = 17.3 Hz, CH), 164.9 (d, *J* = 249.2 Hz, *C*). <sup>19</sup>F NMR (373 MHz, CDCl<sub>3</sub>, δ): -57.8 (s). HRMS-APCI (m/z): [M+H]<sup>+</sup> calcd for C<sub>6</sub>H<sub>4</sub>FN<sub>2</sub>, 123.0353; found, 123.0354.

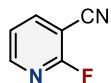
#### 4-Cyano-2-fluoropyridine (2m).



The reaction was performed according to the general procedure B. The reaction was carried out with 69.3 mg (0.50 mmol) of **1m** for 30 min. The resulting crude mixture was analyzed using <sup>19</sup>F NMR with fluorobenzene (15.2 mg) as an internal standard to obtain a NMR yield of **2m** in a 73% yield. <sup>19</sup>F NMR was in agreement with an authentic sample (C3383, Tokyo Chemical Industry Co.). Since **2m** is a volatile compound, only the NMR yield was reported.

<sup>19</sup>F NMR (369 MHz, CDCl<sub>3</sub>, δ): -63.5 (s).

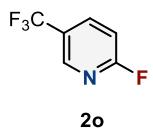
#### 3-Cyano-2-fluoropyridine (2n).



The reaction was performed according to the general procedure B. The reaction was carried out with 69.3 mg (0.50 mmol) of **1n** for 30 min. The resulting crude mixture was analyzed using <sup>19</sup>F NMR with fluorobenzene (15.3 mg) as an internal standard to obtain a NMR yield of **2n** in 92% yield. Product **2n** was obtained as a colorless solid (32.9 mg, 0.27 mmol, 54% yield) after purification by silica-gel column chromatography (SiO<sub>2</sub>, Et<sub>2</sub>O/hexane, 0:100–30:70). <sup>1</sup>H, <sup>13</sup>C, and <sup>19</sup>F NMR were in agreement with the literature.<sup>9</sup>

<sup>1</sup>H NMR (396 MHz, CDCl<sub>3</sub>, δ): 7.38 (ddd, *J* = 1.6, 4.9, 7.3 Hz, 1H), 8.12 (ddd, *J* = 1.6, 6.9, 9.1 Hz, 1H), 8.48–8.50 (m, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 97.3 (d, *J* = 31.6 Hz, *C*), 112.5 (d, *J* = 5.7 Hz, *C*), 121.6 (d, *J* = 3.8 Hz, CH), 144.4 (CH), 152.0 (d, *J* = 14.3 Hz, CH), 162.6 (d, *J* = 247.2 Hz, *C*). <sup>19</sup>F NMR (373 MHz, CDCl<sub>3</sub>, δ): -60.4 (s). HRMS-APCI (m/z): [M+H]<sup>+</sup> calcd for C<sub>6</sub>H<sub>4</sub>FN<sub>2</sub>, 123.0353; found, 123.0354.

**2-Fluoro-5-(trifluoromethyl)pyridine (2o).**



The reaction was performed according to the general procedure B. The reaction was carried out with 90.6 mg (0.50 mmol) of **1o** for 30 min. The resulting crude mixture was analyzed using <sup>1</sup>H NMR with dibromomethane (18.5 mg) as an internal standard to obtain a <sup>1</sup>H NMR yield of **2o** in 76% yield. <sup>1</sup>H NMR was in agreement with an authentic sample (F0995, Tokyo Chemical Industry Co.). Since **2o** is a volatile compound, only the NMR yield was reported.

**9-Benzyl-6-floro-9H-purine (2p).**



The reaction was performed according to the general procedure B. The reaction was carried out with 122.0 mg (0.50 mmol) of **1p** for 60 min. The resulting crude mixture was analyzed using <sup>19</sup>F NMR with fluorobenzene (15.0 mg) as an internal standard to obtain a NMR yield of **2p** in 46% yield. Product **2p** was obtained as a white solid (46.4 mg, 0.20 mmol, 41% yield) after purification by silica-gel column chromatography (SiO<sub>2</sub>, EtOAc/hexane, 25:75–35:65). <sup>1</sup>H, <sup>13</sup>C, and <sup>19</sup>F NMR were in agreement with the literature.<sup>8</sup>

<sup>1</sup>H NMR (396 MHz, CDCl<sub>3</sub>, δ): 5.48 (s, 2H), 7.29–7.42 (m, 5H), 8.08 (s, 1H), 8.68 (s, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, δ): 47.9 (CH<sub>2</sub>), 120.1 (d, *J* = 29.6 Hz, C), 127.9 (CH), 128.8 (CH), 129.2 (CH), 134.5 (C), 144.8 (CH), 152.0 (d, *J* = 14.2 Hz, CH), 155.6 (d, *J* = 11.4 Hz, C), 159.8 (d, *J* = 260.3 Hz, C). <sup>19</sup>F NMR (373 MHz, CDCl<sub>3</sub>, δ): -70.3 (s). HRMS-ESI (m/z): [M+H]<sup>+</sup> calcd for C<sub>12</sub>H<sub>10</sub>FN<sub>4</sub>, 229.0884; found, 229.0882.

**7-Benzyl-4-fluoro-7H-pyrrolo[2,3-d]pyrimidine (2q).**

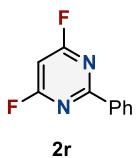


The reaction was performed according to the general procedure B. The reaction was carried out with 121.4 mg (0.50 mmol) of **1q** for 60 min. The resulting crude mixture was analyzed using <sup>19</sup>F NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2q** in 90% yield. Product **2q** was obtained as a white solid (85.1 mg, 0.37 mmol, 75% yield) after purification by silica-

gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–20:80).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>6</sup>

$^1\text{H}$  NMR (401 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 5.48 (s, 2H), 6.61 (d,  $J = 3.2$  Hz, 1H), 7.18–7.24 (m, 3H), 7.29–7.38 (m, 3H), 8.57 (d,  $J = 0.8$  Hz, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 48.6 ( $\text{CH}_2$ ), 98.0 (d,  $J = 4.8$  Hz, CH), 104.2 (d,  $J = 33.3$  Hz, C), 127.5 (CH), 128.1 (CH), 128.4 (d,  $J = 2.0$  Hz, CH), 128.9 (CH), 136.3 (C), 150.6 (d,  $J = 14.3$  Hz, CH), 154.7 (d,  $J = 12.3$  Hz, C), 162.3 (d,  $J = 252.7$  Hz, C).  $^{19}\text{F}$  NMR (373 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –66.5 (s). HRMS-ESI (m/z):  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{13}\text{H}_{11}\text{FN}_3$ , 228.0932; found, 228.0928.

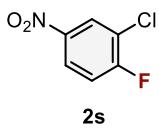
#### 4,6-Difluoro-2-phenylpyrimidine (2r).



The reaction was performed according to the general procedure B. The reaction was carried out with 112.6 mg (0.50 mmol) of **1r**, 116.1 mg (2.0 mmol) of KF, and 123.9 mg (0.75 mmol) of  $\text{Et}_4\text{NCl}$  for 30 min. The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2r** in 65% yield. Product **2r** was obtained as a white solid (53.2 mg, 0.28 mmol, 55% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–2:98).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>6</sup>

$^1\text{H}$  NMR (401 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 6.44 (t,  $J = 2.2$  Hz, 1H), 7.45–7.60 (m, 3H), 8.39–8.47 (m, 2H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 89.9 (t,  $J = 37.1$  Hz, CH), 128.73 (CH), 128.75 (CH), 132.4 (CH), 134.7 (C), 166.0 (t,  $J = 17.3$  Hz, C), 172.2 (dd,  $J = 18.8, 256.7$  Hz, C).  $^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –56.5 (s). HRMS-APCI (m/z):  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{10}\text{H}_7\text{F}_2\text{N}_2$ , 193.0572; found, 193.0566.

#### 3-Chloro-4-fluoronitrobenzene (2s).

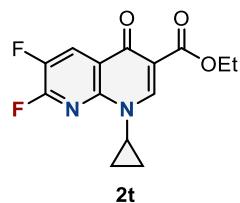


The reaction was performed according to the general procedure B. The reaction was carried out with 95.8 mg (0.50 mmol) of **1s** for 45 min. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.2 mg) as an internal standard to obtain an NMR yield of **2s** in 80% yield. Product **2s** was obtained as a yellow solid (58.0 mg, 0.33 mmol, 66% yield, isolated along with approximately 3% of starting material) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{pentane}$ , 0:100–3:97).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>13</sup>

$^1\text{H}$  NMR (401 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.33 (dd,  $J = 8.2, 9.0$  Hz, 1H), 8.19 (ddd,  $J = 2.7, 4.3, 9.1$  Hz, 1H),

8.37 (dd,  $J = 2.8, 6.0$  Hz, 1H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 117.2 (d,  $J = 24.1$  Hz, CH), 122.5 (d,  $J = 19.2$  Hz, C), 124.0 (d,  $J = 8.7$  Hz, CH), 126.7 (CH), 144.2 (C), 161.9 (d,  $J = 260.0$  Hz, C).  $^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -104.4 (s). HRMS-EI (m/z): [M]<sup>+</sup> calcd for  $\text{C}_6\text{H}_3\text{ClFNO}_2$ , 174.9831; found, 174.9832.

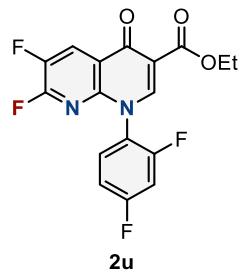
**Ethyl 1-cyclopropyl-6,7-difluoro-4-oxo-1,4-dihydro-1,8-naphthyridine-3-carboxylate (2t).**



The reaction was performed according to the general procedure A. The reaction was carried out with 155.3 mg (0.50 mmol) of **1t**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (14.7 mg) as an internal standard to obtain the NMR yield of **2t** in 74% yield. Product **2t** was obtained as a white solid (73.1 mg, 0.25 mmol, 50% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{EtOAc/hexane}$ , 40:60–50:50).  $^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR were in agreement with the literature.<sup>6</sup>

$^1\text{H}$  NMR (399 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 1.04–1.08 (m, 2H), 1.32 (q,  $J = 6.8$  Hz, 2H), 1.41 (t,  $J = 7.0$  Hz, 3H), 3.59 (tt,  $J = 3.7, 7.4$  Hz, 1H), 4.41 (q,  $J = 7.0$  Hz, 2H), 8.55 (t,  $J = 9.0$  Hz, 1H), 8.65 (s, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.6 (CH<sub>2</sub>), 14.3 (CH<sub>3</sub>), 34.3 (CH), 61.2 (CH<sub>2</sub>), 112.3 (C), 122.8 (d,  $J = 2.8$  Hz, C), 126.0 (dd,  $J = 5.2, 16.8$  Hz, CH), 143.5 (dd,  $J = 27.8, 261.6$  Hz, C), 144.3 (d,  $J = 14.3$  Hz, C), 148.7 (CH), 152.3 (dd,  $J = 16.7, 249.7$  Hz, C), 164.6 (C), 172.8 (C).  $^{19}\text{F}$  NMR (375 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -142.4 (d,  $J = 23.3$  Hz, 1F), -77.3 (d,  $J = 34.5$  Hz, 1F). HRMS-EI (m/z): [M]<sup>+</sup> calcd for  $\text{C}_{14}\text{H}_{12}\text{F}_2\text{N}_2\text{O}_3$ , 294.0811; found, 294.0814.

**Ethyl 1-(2,4-difluorophenyl)-6,7-difluoro-4-oxo-1,4-dihydro-1,8-naphthyridine-3-carboxylate (2u).**

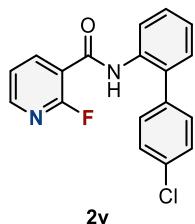


The reaction was performed according to the general procedure A. The reaction was carried out with 191.3 mg (0.50 mmol) of **1u**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.3 mg) as an internal standard to obtain the NMR yield of **2u** in 78% yield. Product

**2u** was obtained as a white solid (116.2 mg, 0.32 mmol, 63% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{EtOAc/hexane}$ , 40:60).

$^1\text{H}$  NMR (399 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 1.41 (t,  $J = 7.0$  Hz, 3H), 4.41 (q,  $J = 7.2$  Hz, 2H), 7.08–7.15 (m, 2H), 7.41–7.46 (m, 1H), 8.53(s, 1H), 8.60 (t,  $J = 8.8$  Hz, 1H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 14.2 ( $\text{CH}_3$ ), 61.4 ( $\text{CH}_2$ ), 105.6 (dd,  $J = 23.0, 26.8$  Hz, CH), 112.6 (dd,  $J = 3.4, 22.5$  Hz, CH), 113.6 (C), 122.3 (d,  $J = 2.9$  Hz, C), 123.3 (dd,  $J = 3.8, 13.4$  Hz, C), 126.2 (dd,  $J = 5.3, 16.8$  Hz, CH), 129.9 (d,  $J = 10.6$  Hz, CH), 143.2 (d,  $J = 13.4$  Hz, C), 143.8 (dd,  $J = 27.3, 262.2$  Hz, C), 149.1 (CH), 152.6 (dd,  $J = 17.3, 252.1$  Hz, C), 157.9 (dd,  $J = 12.4, 255.9$  Hz, C), 163.4 (dd,  $J = 10.5, 254.1$  Hz, C), 164.1 (C), 172.9 (C).  $^{19}\text{F}$  NMR (375 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –141.4 (d,  $J = 34.9$  Hz, 1F), –115.0 (s, 1F), –105.2 (s, 1F), –76.8 (d,  $J = 22.9$  Hz, 1F). HRMS-EI (m/z): [M]<sup>+</sup> calcd for  $\text{C}_{17}\text{H}_{10}\text{F}_4\text{N}_2\text{O}_3$ , 366.0622; found, 366.0629.

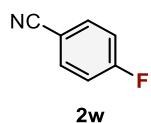
#### N-(4'-Chlorobiphenyl-2-yl)-2-fluoronicotinamide (2v).



The reaction was performed according to the general procedure A. The reaction was carried out with 171.5 mg (0.50 mmol) of **1v**. The resulting crude mixture was analyzed using  $^{19}\text{F}$  NMR with fluorobenzene (15.5 mg) as an internal standard to obtain a NMR yield of **2v** in 55%. Product **2v** was obtained as a white solid (48.2 mg, 0.15 mmol, 30% yield) after purification by silica-gel column chromatography ( $\text{SiO}_2$ ,  $\text{Et}_2\text{O}/\text{hexane}$ , 0:100–25:75) and recycling preparative GPC.

$^1\text{H}$  NMR (396 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 7.21–7.31 (m, 2H), 7.31–7.42 (m, 3H), 7.42–7.52 (m, 3H), 8.31–8.33 (m, 1H), 8.49 (d,  $J = 8.2$  Hz, 1H), 8.58 (brs, 1H), 8.62 (ddd,  $J = 2.2, 7.5, 9.9$  Hz, 1H).  $^{13}\text{C}$  NMR (99 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): 116.2 (d,  $J = 28.2$  Hz, C), 121.9 (CH), 122.6 (d,  $J = 4.7$  Hz, CH), 125.1 (CH), 128.8 (CH), 129.3 (CH), 130.2 (CH), 130.6 (CH), 132.0 (C), 134.4 (C), 134.5 (C), 136.0 (C), 143.7 (d,  $J = 1.9$  Hz, CH), 150.6 (d,  $J = 16.9$  Hz, CH), 159.4 (d,  $J = 235.8$  Hz, C), 159.5 (d,  $J = 8.5$  Hz, C).  $^{19}\text{F}$  NMR (377 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): –65.0 (s). HRMS-EI (m/z): [M]<sup>+</sup> calcd for  $\text{C}_{18}\text{H}_{12}\text{ClFN}_2\text{O}$ , 326.0617; found, 326.0617.

#### 4-Fluorobenzonitrile (2w).



The reaction was performed according to the general procedure B. The reaction was carried out with 68.7 mg (0.50 mmol) of 4-chlorobenzonitrile (**1w**) for 30 min. The resulting crude mixture was

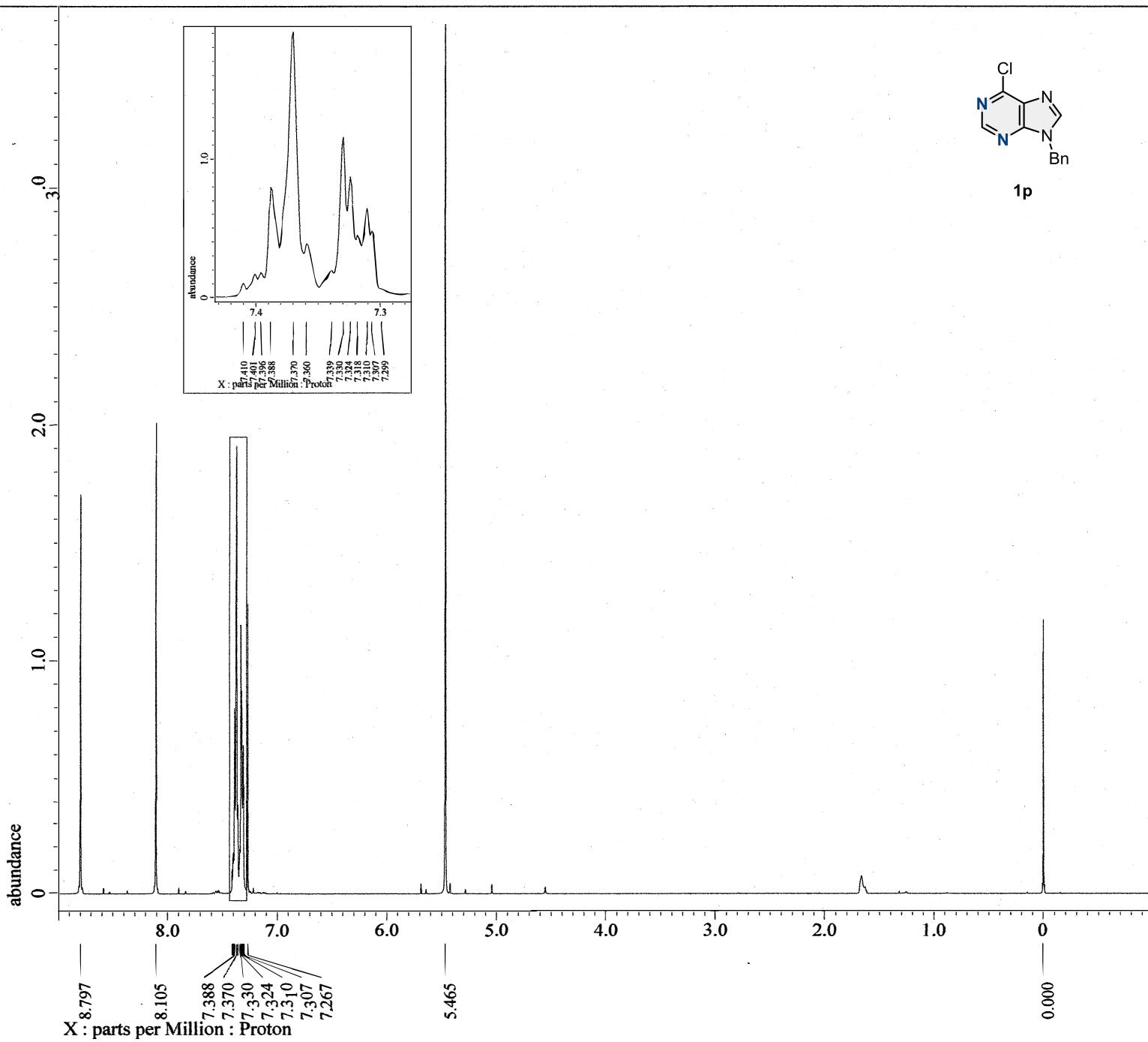
analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.1 mg) as an internal standard to obtain the NMR yield of **2w** in 3% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>14</sup>

The reaction was carried out with 74.0 mg (0.50 mmol) of 4-nitrobenzonitrile (**1w'**) for 30 min. The resulting crude mixture was analyzed by  $^{19}\text{F}$  NMR with fluorobenzene (15.3 mg) as an internal standard to obtain the NMR yield of **2w** in 62% yield.  $^{19}\text{F}$  NMR was in agreement with the literature.<sup>14</sup> Since **2w** is a volatile compound, only the NMR yield was reported.

$^{19}\text{F}$  NMR (373 MHz,  $\text{CDCl}_3$ ,  $\delta$ ): -102.8 (s).

## 12. References

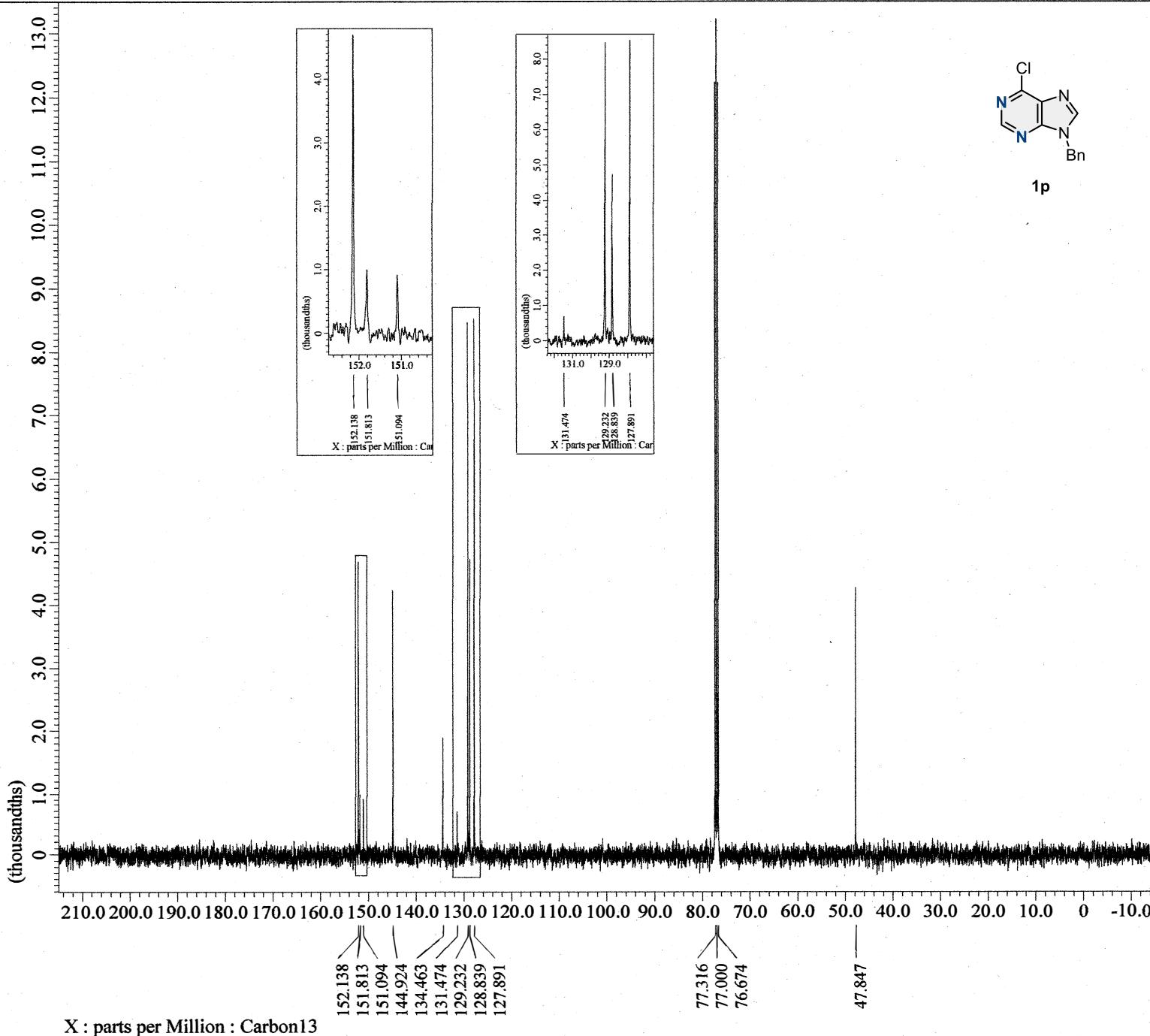
1. Bliman, D.; Pettersson, M.; Bood, M.; Grothi, M. *Tetrahedron Lett.* **2014**, *55*, 2929.
2. Wood, A. B.; Nandiwale, K. Y.; Mo, Y.; Jin, B.; Pomberger, A.; Schultz, V.; Gallou, F.; Jensen, K. F.; Lipshutz, B. H. *Green Chem.* **2020**, *22*, 3441.
3. Karoli, T.; Mamidyal, S. K.; Zuegg, J.; Fry, S. R.; Tee, E. H. L.; Bradford, T. A.; Madala, P. K.; Huang, J. X.; Ramu, S.; Butler, M. S.; Cooper, M. A. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 2428.
4. Allen, L. J.; Lee, S. H.; Cheng, Y.; Hanley, P. S.; Muhuhi, J. M.; Kane, E.; Powers, S. L.; Anderson, J. E.; Bell, B. M.; Roth, G. A.; Sanford, M. S.; Bland, D. C. *Org. Process Res. Dev.* **2014**, *18*, 1045.
5. Schimler, S. D.; Ryan, S. J.; Bland, D. C.; Anderson, J. E.; Sanford, M. S. *J. Org. Chem.* **2015**, *80*, 12137.
6. Morales-Colón, M. T.; See, Y. Y.; Lee, S. J.; Scott, P. J. H.; Bland, D. C.; Sanford, M. S. *Org. Lett.* **2021**, *23*, 4493.
7. Roger, J.; Royer, S.; Cattey, H.; Savateev, A.; Smaliy, R. V.; Kostyuk, A. N.; Hierso, J.-C. *Eur. J. Inorg. Chem.* **2017**, *2017*, 330.
8. Taylor, N. J.; Emer, E.; Preshlock, S.; Schedler, M.; Tredwell, M.; Verhoog, S.; Mercier, J.; Genicot, C.; Gouverneur, V. *J. Am. Chem. Soc.* **2017**, *139*, 8267.
9. Katoh, T.; Tomata, Y.; Tsukamoto, T.; Nakada, Y. *Tetrahedron Lett.* **2015**, *56*, 6043.
10. Ryan, S. J.; Schimler, S. D.; Bland, D. C.; Sanford, M. S. *Org. Lett.* **2015**, *17*, 1866.
11. Plé, N.; Turck, A.; Heynderickx, A.; Quéguiner, G. *Tetrahedron* **1998**, *54*, 4899.
12. Johansen, M. B.; Lindhardt, A. T. *Chem. Commun.* **2018**, *54*, 825.
13. Lacour, M.-A.; Zablocka, M.; Duhayon, C.; Majoral, J.-P.; Taillefer, M. *Adv. Synth. Catal.* **2008**, *350*, 2677.
14. Wood, A. B.; Kincaid, J. R. A.; Lipshutz, B. H. *Green Chem.* **2022**, *24*, 2853.



----- PROCESSING PARAMETERS -----  
 sexp( 0.2[Hz], 0.0[s] )  
 trapezoid( 0[%], 0[%], 80[%], 100[%] )  
 zerofill( 1, TRUE )  
 fft( 1, TRUE, TRUE )  
 ma chiphase  
 ppm

Derived from: MKN278-pure Proton-1-1.jdf

Filename	= MKN278-pure_Proton-1-2
Author	= element
Experiment	= proton_a_utojxp
Sample_Id	= MKN278-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 18-JAN-2024 10:47:11
Revision_Time	= 15-JUN-2024 12:17:08
Comment	= single pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Doma in	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.2982153[T] (400[MHz])
X_Acq_Duration	= 2.20725248[s]
X_Domain	= Proton
X_Freq	= 395.88430144[MHz]
X_Offset	= 5[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45305193[Hz]
X_Sweep	= 7.422802851[kHz]
X_Sweep_Clipped	= 5.93824228[kHz]
Irr_Domain	= Proton
Irr_Freq	= 395.88430144[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= Proton
Tri_Freq	= 395.88430144[MHz]
Tri_Offset	= 5[ppm]
B1_angle	= 2.0[us]
Cli_ppe_d	= FALSE
Scans	= 8
TotalScans	= 8
Relaxation_Delay	= 5[s]
Recv_Gain	= 56
Temp_Get	= 19.1[dC]
X_90_Width	= 6.34[us]
X_Acq_Time	= 2.20725248[s]
X_Angle	= 45[deg]
X_Att	= 5[dB]
X_Pulse	= 3.17[us]
Irr_Mode	= Off
Tri_Mode	= Off
DaRe_Loop	= 500
Dante_Preset	= FALSE
Decimation_Rate	= 0
Initia_lWait	= 1[s]
Phase	= {0, 90, 270, 180, 180,



```
---- PROCESSING PARAMETERS ----
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN278-pure Carbon-1-1.jdf

```

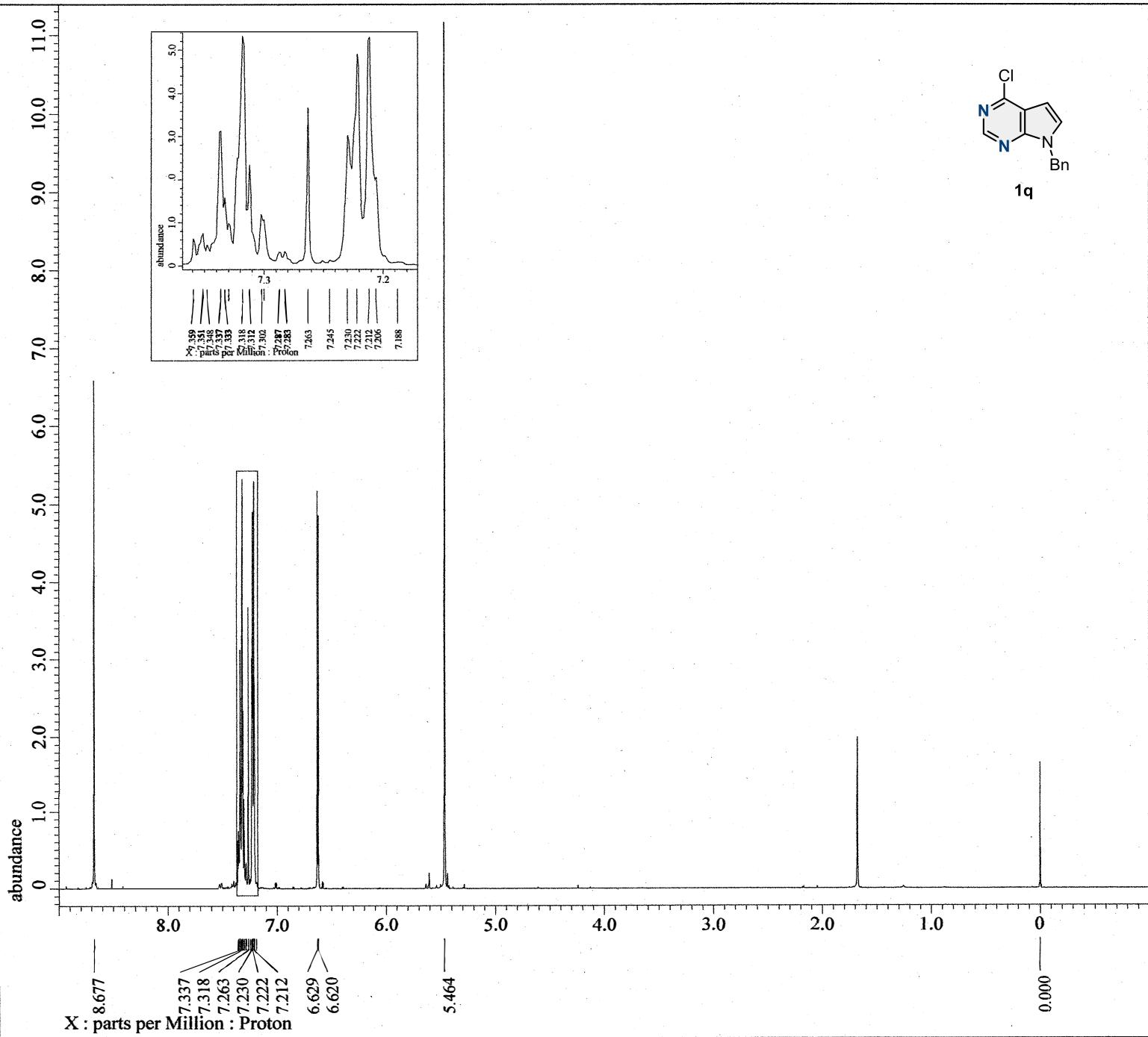
Filename = MKN278-pure_Carbon-
Author = element
Experiment = carbon_auto.jxp
Sample_Id = MKN278-pure
Solvent = CHLOROFORM-D
Actual_Start_Time = 18-JAN-2024 11:01:3
Revision_Time = 15-JUN-2024 14:30:4

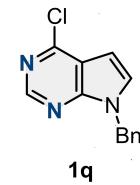
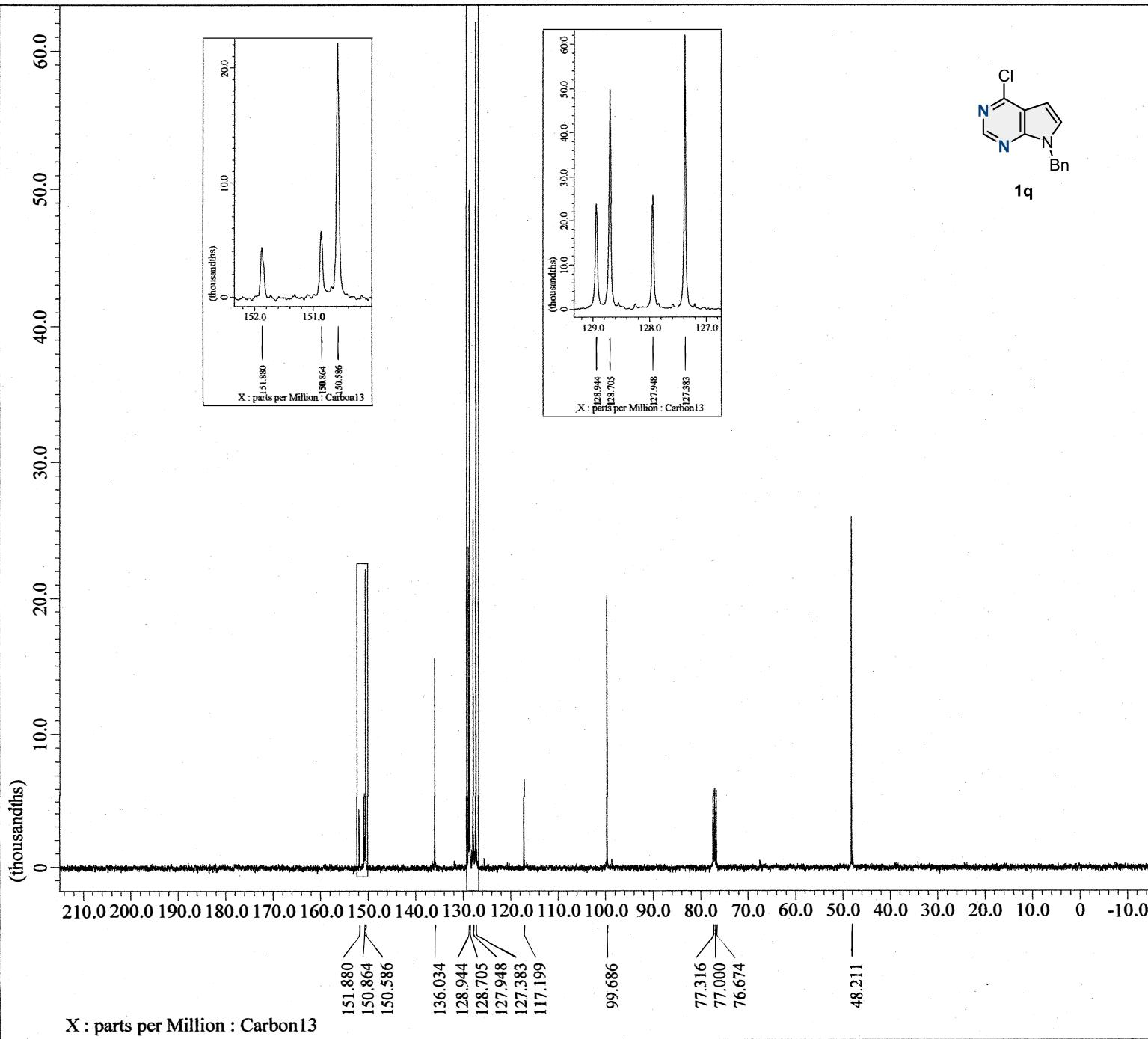
Comment = single pulse decoupl
Data_Format = 1D COMPLEX
Dim_Size = 26214
X_Domain = Carbon13
Dim_Title = Carbon13
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[M
X_Acq_Duration = 1.048576[s]
X_Domain = Carbon13
X_Freq = 99.54517646[MHz]
X_Offset = 100[ppm]
X_Points = 32768
X_Prescans = 4
X_Resolution = 0.95367432[Hz]
X_Sweep = 31.25[kHz]
X_Sweep_Clipped = 25[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Blanking = 5.0[us]
Clipped = FALSE
Scans = 128
Total_Scans = 128

Relaxation_Delay = 2[s]
Recv_Gain = 50
Temp_Get = 18.6[dC]
X_90_Width = 11.5[us]
X_Acq_Time = 1.048576[s]
X_Angle = 30[deg]
X_Atn = 9[dB]
X_Pulse = 3.83333333[us]
Irr_Atn_Dec = 30.172[dB]
Irr_Atn_Dec_Calc = 30.172[dB]
Irr_Atn_Dec_Default_Calc = 30.172[dB]
Irr_Atn_Noe = 30.172[dB]
Irr_Dec_Bandwidth_Hz = 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm = 12.08082432[ppm]
Irr_Dec_Freq = 395.88430144[MHz]
Irr_Dec_Merit_Factor = 2.2
Irr_Decoupling = TRUE
Irr_Noe = TRUE

```





```
---- PROCESSING PARAMETERS ----
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN140-pure Carbon-1-1.jdf

```

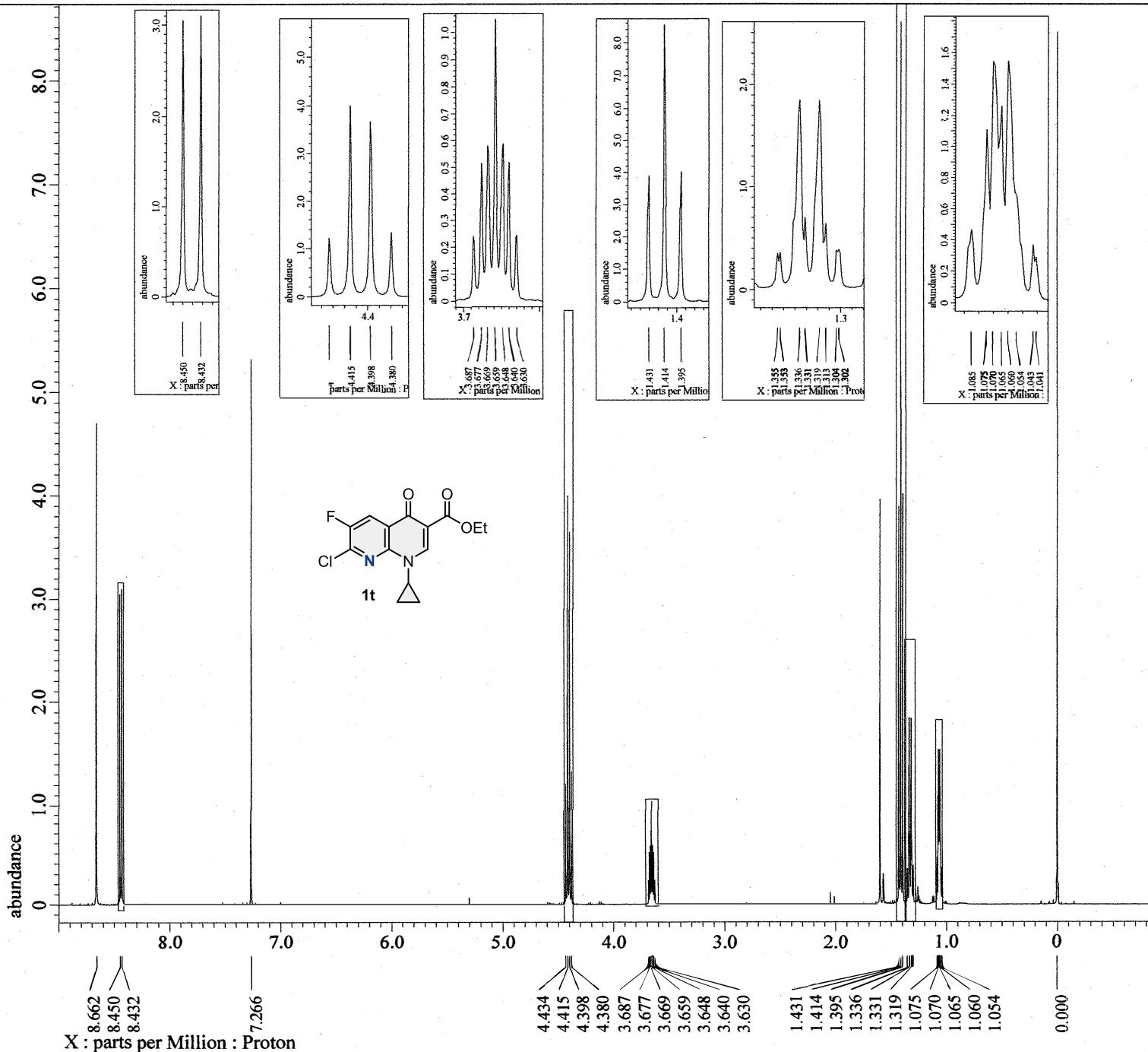
Filename = MKN140-pure_Carbon-
Author = element
Experiment = carbon_auto.jxp
Sample_Id = MKN140-pure
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-JAN-2024 16:56:5
Revision_Time = 15-JUN-2024 15:04:5

Comment = single pulse decoupl
Data_Format = 1D_COMPLEX
Dim_Size = 26214
X_Domain = Carbon13
Dim_Title = Carbon13
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[M
X_Acq_Duration = 1.048576[s]
X_Domain = Carbon13
X_Freq = 99.54517646[MHz]
X_Offset = 100[ppm]
X_Points = 32768
X_Prescans = 4
X_Resolution = 0.95367432[Hz]
X_Sweep = 31.25[kHz]
X_Sweep_Clipped = 25[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Blanking = 5.0[us]
Clipped = FALSE
Scans = 64
Total_Scans = 64

Relaxation_Delay = 2[s]
Recv_Gain = 50
Temp_Get = 18.7[dC]
X_90_Width = 11.5[us]
X_Acq_Time = 1.048576[s]
X_Angle = 30[deg]
X_Atn = 9[dB]
X_Pulse = 3.833333333[us]
Irr_Atn_Dec = 30.172[dB]
Irr_Atn_Dec_Calc = 30.172[dB]
Irr_Atn_Dec_Default_Calc = 30.172[dB]
Irr_Atn_Noe = 30.172[dB]
Irr_Dec_Bandwidth_Hz = 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm = 12.08082432[ppm]
Irr_Dec_Freq = 395.88430144[MHz]
Irr_Dec_Merit_Factor = 2.2
Irr_Decoupling = TRUE
Irr_Noe = TRUE

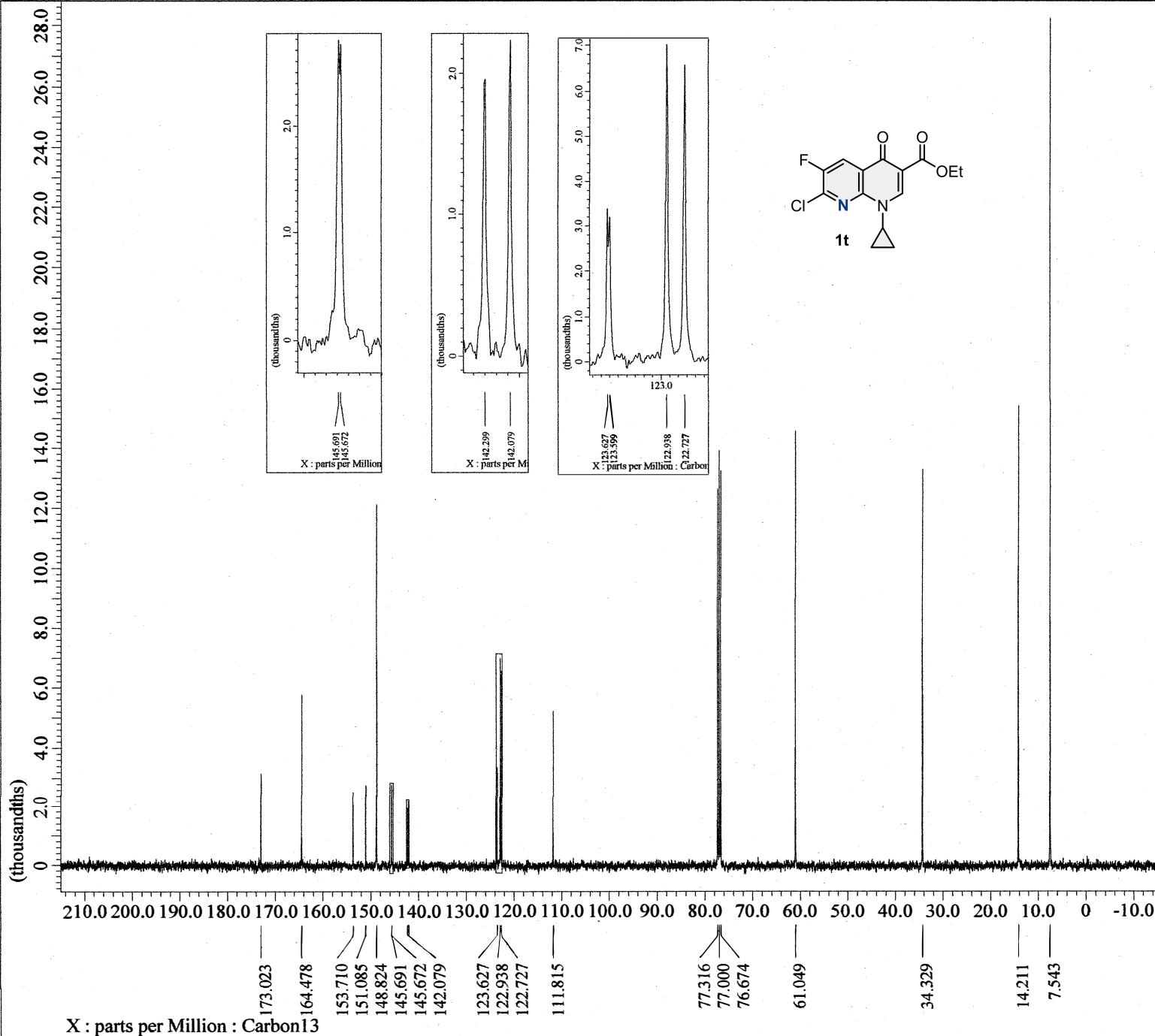
```



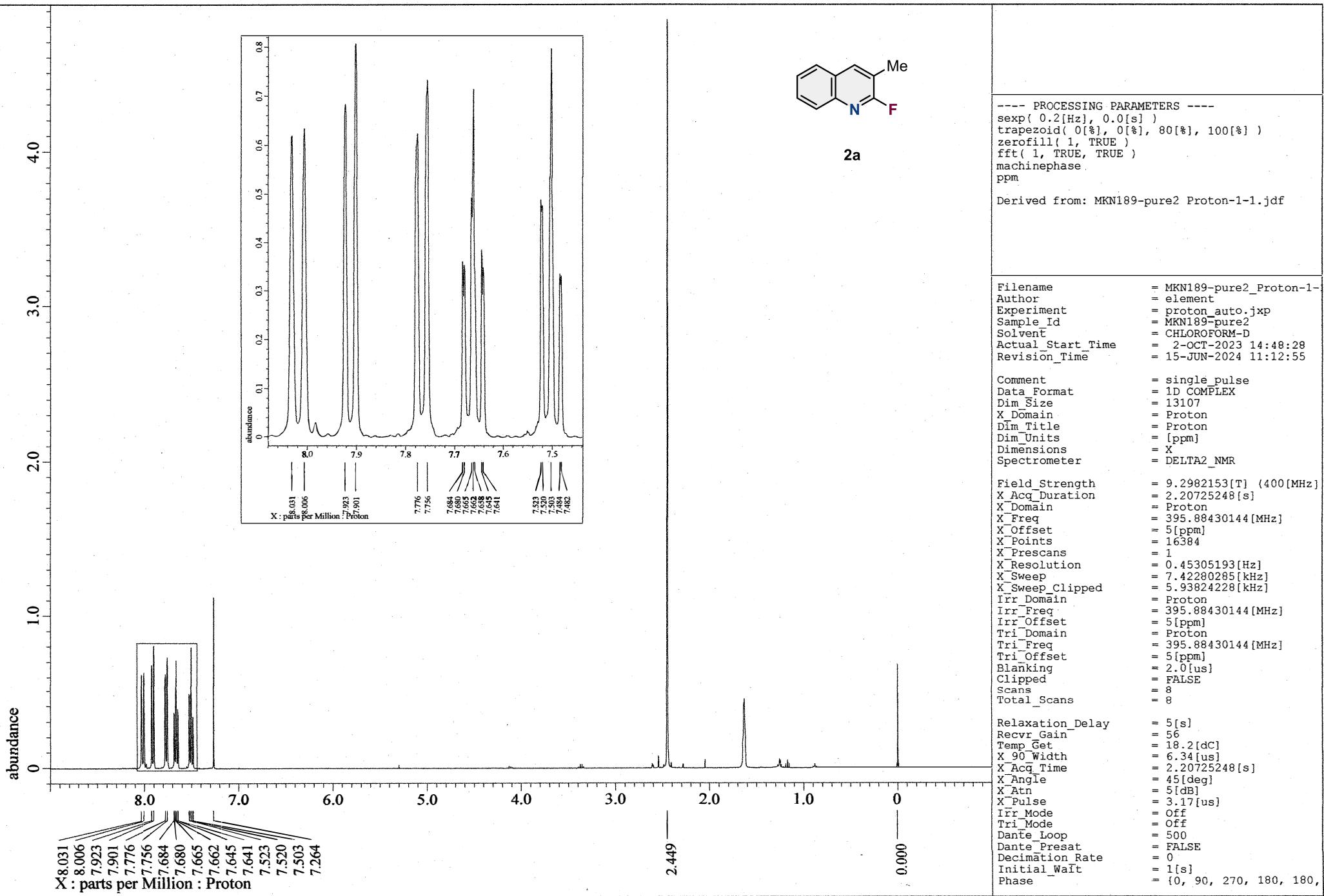
```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

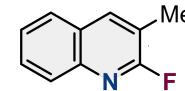
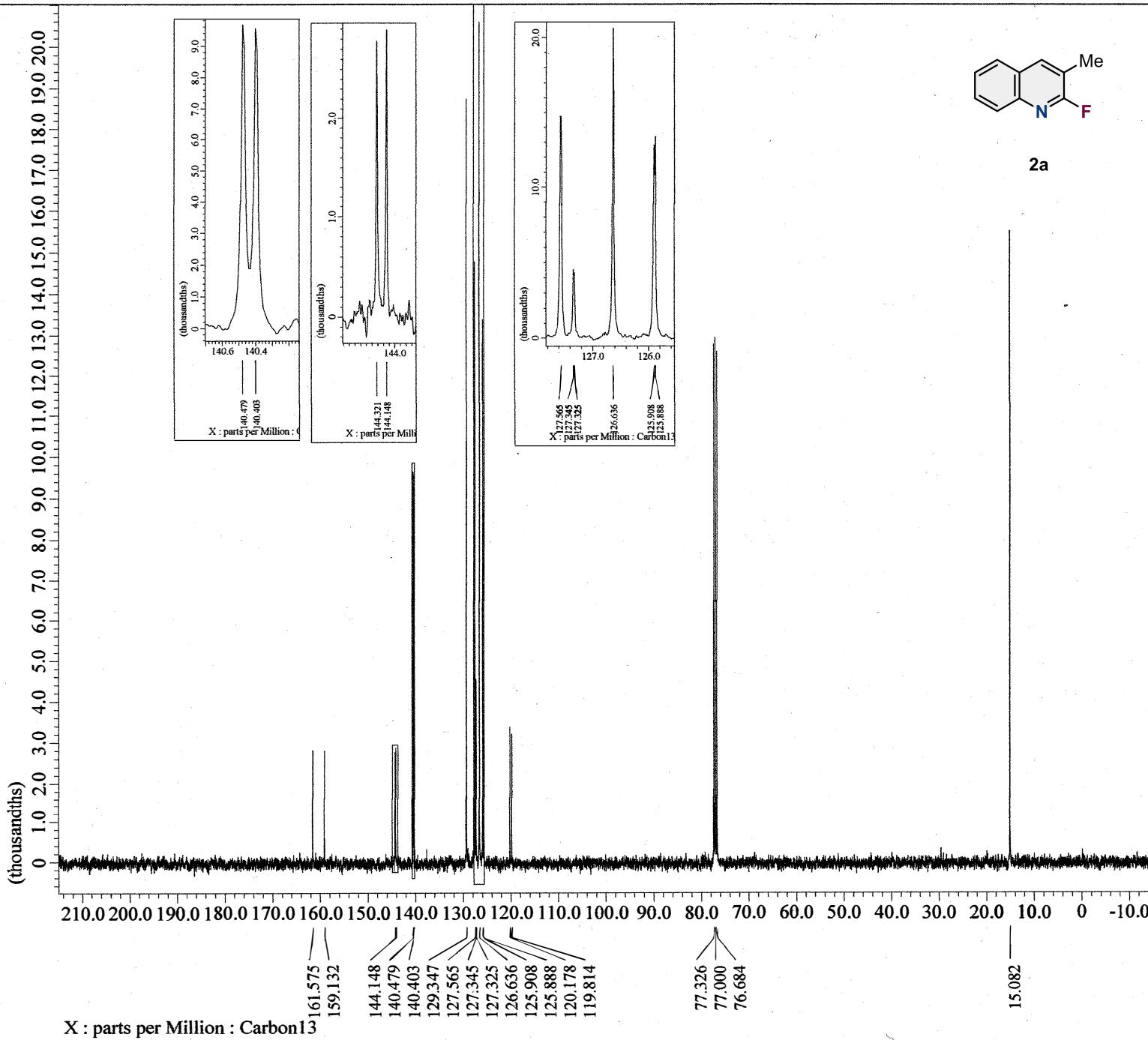
Derived from: MKN218-pure2 Proton-1-1.jdf

Filename	= MKN218-pure2_Proton-1-2.jd
Author	= element
Experiment	= proton.jxp
Sample_Id	= MKN218-pure2
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 16-NOV-2023 12:42:18
Revision_Time	= 15-JUN-2024 15:13:57
Comment	= single_pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Domain	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.4073814[T] (400[MHz])
X_Acc_Duration	= 2.18103808[s]
X_Domain	= 1H
X_Freq	= 400.53219825[MHz]
X_Offset	= 5[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45849727[Hz]
X_Sweep	= 7.51201923[kHz]
X_Sweep_Clipped	= 6.00961538[kHz]
Irr_Domain	= Proton
Irr_Freq	= 400.53219825[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= Proton
Tri_Freq	= 400.53219825[MHz]
Tri_Offset	= 5[ppm]
Clipped	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recvr_Gain	= 46
Temp_Get	= 19.7[dC]
X_90_Width	= 6.7[us]
X_Acc_Time	= 2.18103808[s]
X_Angle	= 45[deg]
X_Atn	= 0.8[dB]
X_Pulse	= 3.35[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Presat	= FALSE
Initial_Wait	= 1[s]
Repetition_Time	= 7.18103808[s]



Filename	= MKN218-pure_Carbon-
Author	= element
Experiment	= carbon_auto.jxp
Sample_Id	= MKN218-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 20-JAN-2024 16:38:4
Revision_Time	= 15-JUN-2024 15:33:2
Comment	
Data_Format	= single pulse decoup
Dim_Size	= 1D COMPLEX
X_Domain	= 26214
Dim_Title	= Carbon13
Dim_Units	= Carbon13
Dimensions	= [ppm]
Spectrometer	= X
Field_Strength	= DELTA2_NMR
X_Acq_Duration	= 9.2982153[T] (400[M])
X_Domain	= 1.048576[s]
X_Freq	= Carbon13
X_Offset	= 99.54517646[MHz]
X_Points	= 100[ppm]
X_Prescans	= 32768
X_Resolution	= 4
X_Sweep	= 0.95367432[Hz]
X_Sweep_Clipped	= 31.25[kHz]
Irr_Domain	= 25[kHz]
Irr_Freq	= Proton
Irr_Offset	= 395.88430144[MHz]
Blanking	= 5[ppm]
Clipped	= 5.0[us]
Scans	= FALSE
Total_Scans	= 128
Relaxation_Delay	= 128
Recvr_Gain	= 2[s]
Temp_Get	= 50
X_90_Width	= 18.7[dC]
X_Acq_Time	= 11.5[us]
X_Angle	= 1.048576[s]
X_Atn	= 30[deg]
X_Pulse	= 9[dB]
Irr_Atn_Dec	= 3.83333333[us]
Irr_Atn_Dec_Calc	= 30.172[dB]
Irr_Atn_Dec_Default_Calc	= 30.172[dB]
Irr_Atn_Noe	= 30.172[dB]
Irr_Dec_Bandwidth_Hz	= 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm	= 12.08082432[ppm]
Irr_Dec_Freq	= 395.88430144[MHz]
Irr_Dec_Merit_Factor	= 2.2
Irr_Decoupling	= TRUE
Irr_Noe	= TRUE





2a

```
---- PROCESSING PARAMETERS ----
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN189-pure2 Carbon-1-1.jdf

```

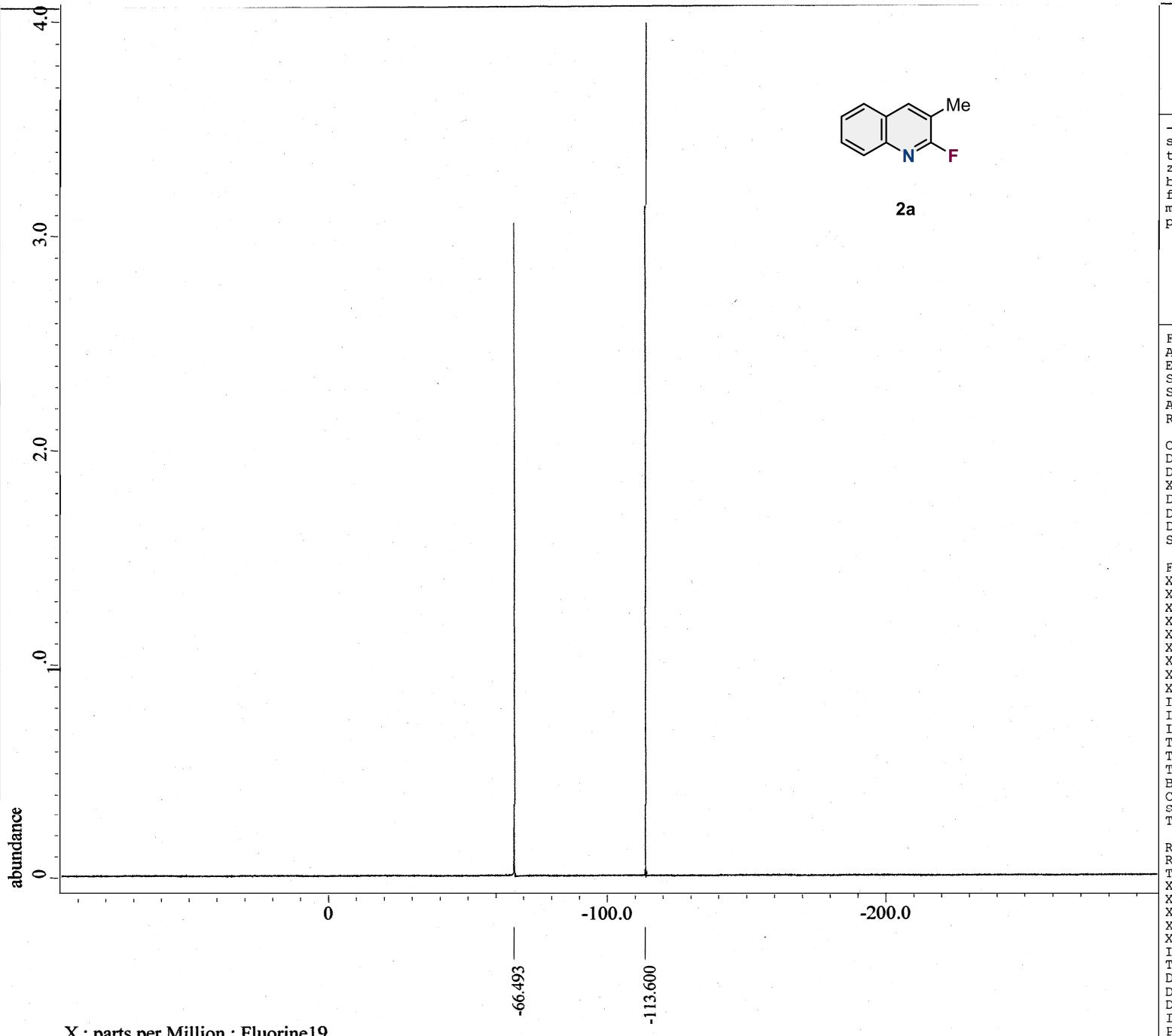
Filename = MKN189-pure2_Carbon
Author = element
Experiment = Carbon_auto.jxp
Sample_Id = MKN189-pure2
Solvent = CHLOROFORM-D
Actual_Start_Time = 13-OCT-2023 17:43:01
Revision_Time = 15-JUN-2024 12:04:22

Comment = single pulse decoupl
Data_Format = 1D COMPLEX
Dim_Size = 26214
X_Domain = Carbon13
Dim_Title = Carbon13
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[M
X_Acq_Duration = 1.048576[s]
X_Domain = Carbon13
X_Freq = 99.54517646[MHz]
X_Offset = 100[ppm]
X_Points = 32768
X_Prescans = 4
X_Resolution = 0.95367432[Hz]
X_Sweep = 31.25[KHz]
X_Sweep_Clipped = 25[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Blanking = 5.0[us]
Clipped = FALSE
Scans = 128
Total_Scans = 128

Relaxation_Delay = 2[e]
Recvr_Gain = 50
Temp_Get = 17.4[dC]
X_90_Width = 11.5[us]
X_Acq_Time = 1.048576[s]
X_Angle = 30[deg]
X_Atn = 9[dB]
X_Pulse = 3.83333333[us]
Irr_Atn_Dec = 30.172[dB]
Irr_Atn_Dec_Calc = 30.172[dB]
Irr_Atn_Dec_Default_Calc = 30.172[dB]
Irr_Atn_Noe = 30.172[dB]
Irr_Dec_Bandwidth_Hz = 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm = 12.08082432[ppm]
Irr_Dec_Freq = 395.88430144[MHz]
Irr_Dec_Merit_Factor = 2.2
Irr_Decoupling = TRUE
Irr_Noe = TRUE

```



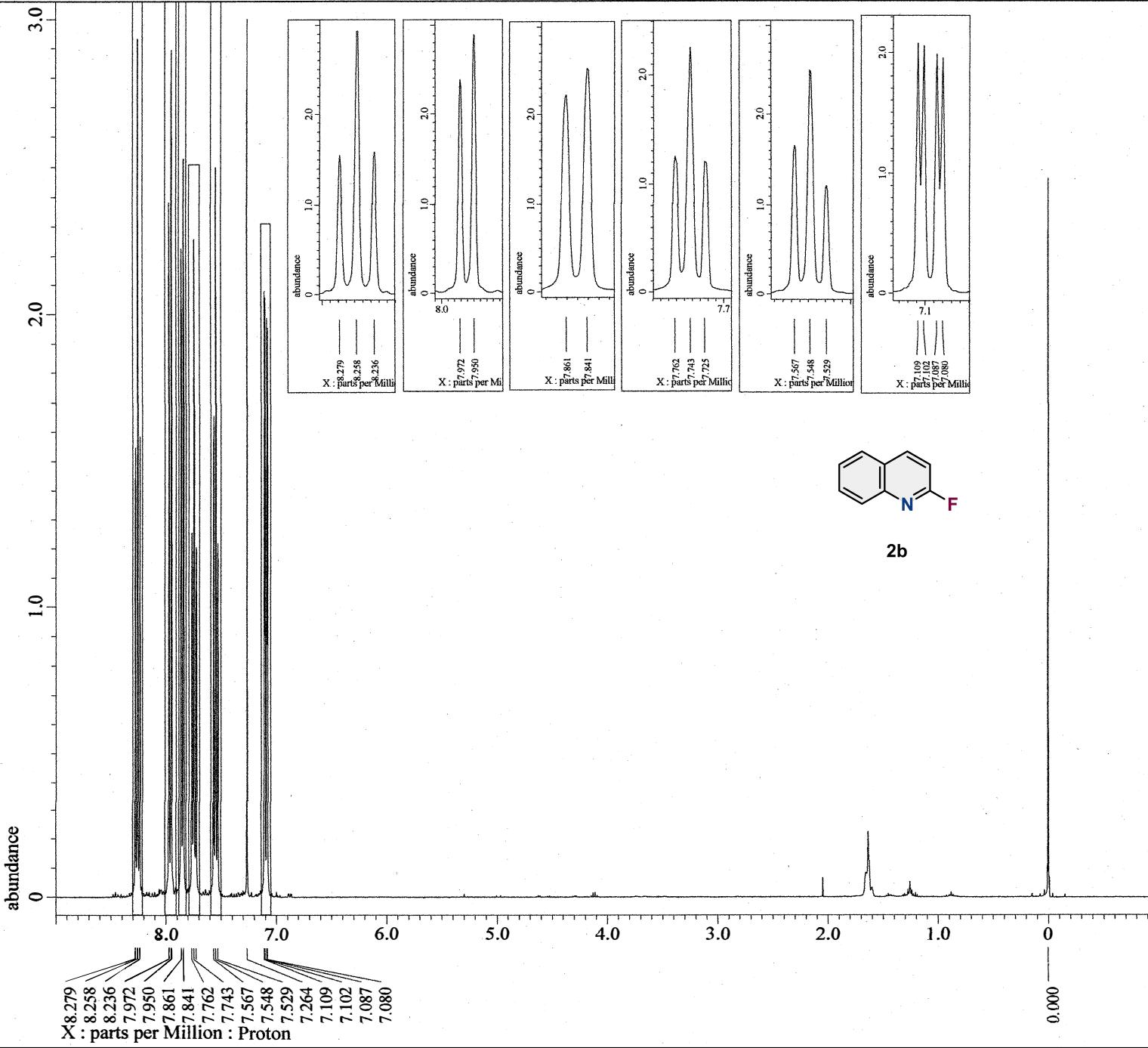
```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

```
Filename = MKN189-pure2_int_singl
Author = element
Experiment = single_pulse.jxp
Sample_Id = MKN189-pure2_int
Solvent = CHLOROFORM-D
Actual_Start_Time = 14-OCT-2023 11:09:07
Revision_Time = 24-JAN-2024 11:39:29

Comment = single pulse
Data_Format = 1D_COMPLEX
Dim_Size = 13107
X_Domain = Fluorine19
Dim_Title = Fluorine19
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 89.12896[ms]
X_Domain = Fluorine19
X_Freq = 372.50336686[MHz]
X_Offset = -100[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 11.21969784[Hz]
X_Sweep = 183.82352941[kHz]
X_Sweep_Clipped = 147.05882353[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 372.50336686[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Fluorine19
Tri_Freq = 372.50336686[MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

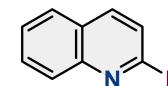
Relaxation_Delay = 5[s]
Recv_Gain = 56
Temp_Get = 18.9[dC]
X_90_Width = 8.03[us]
X_Acq_Time = 89.12896[ms]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 4.015[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180},
```



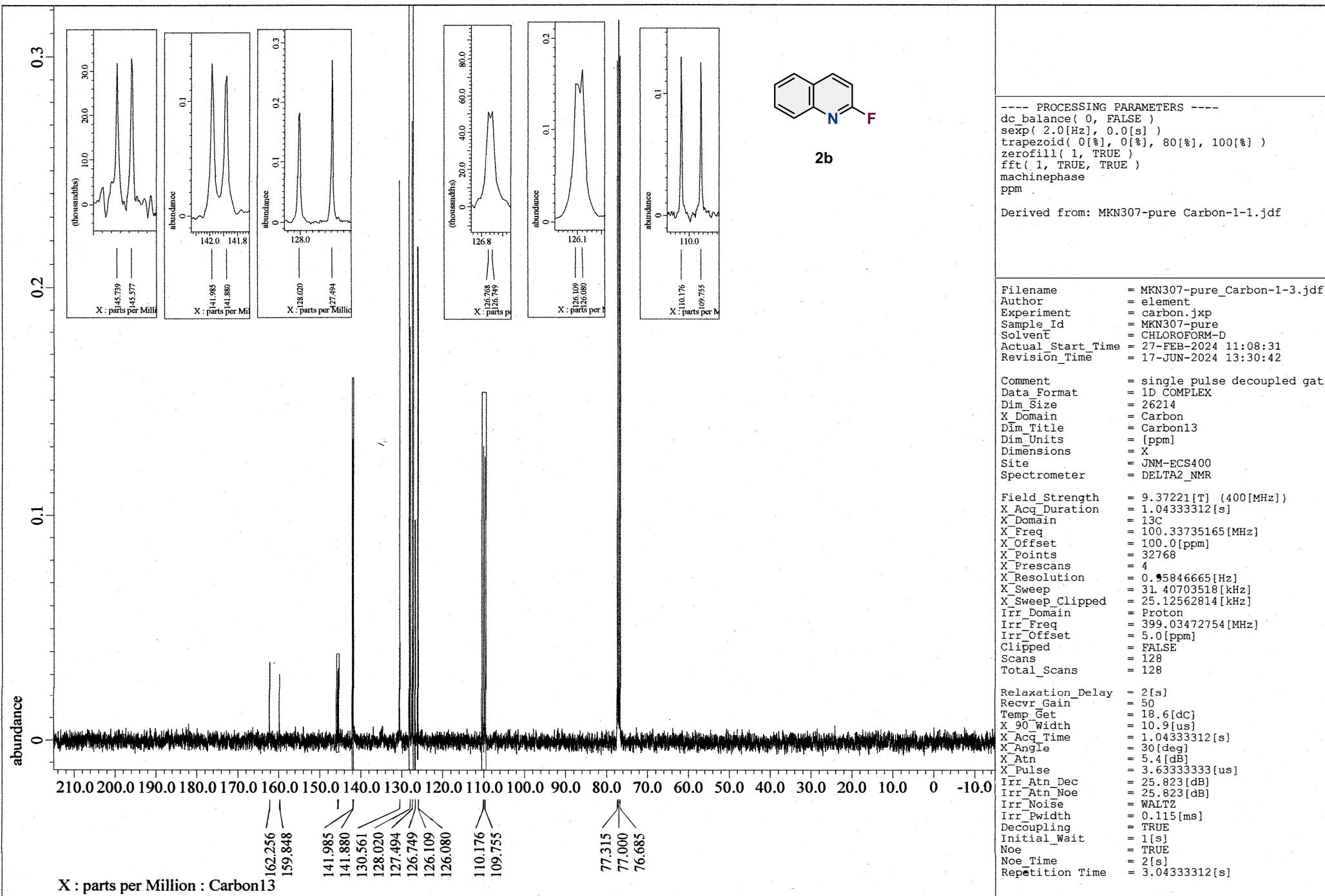
```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

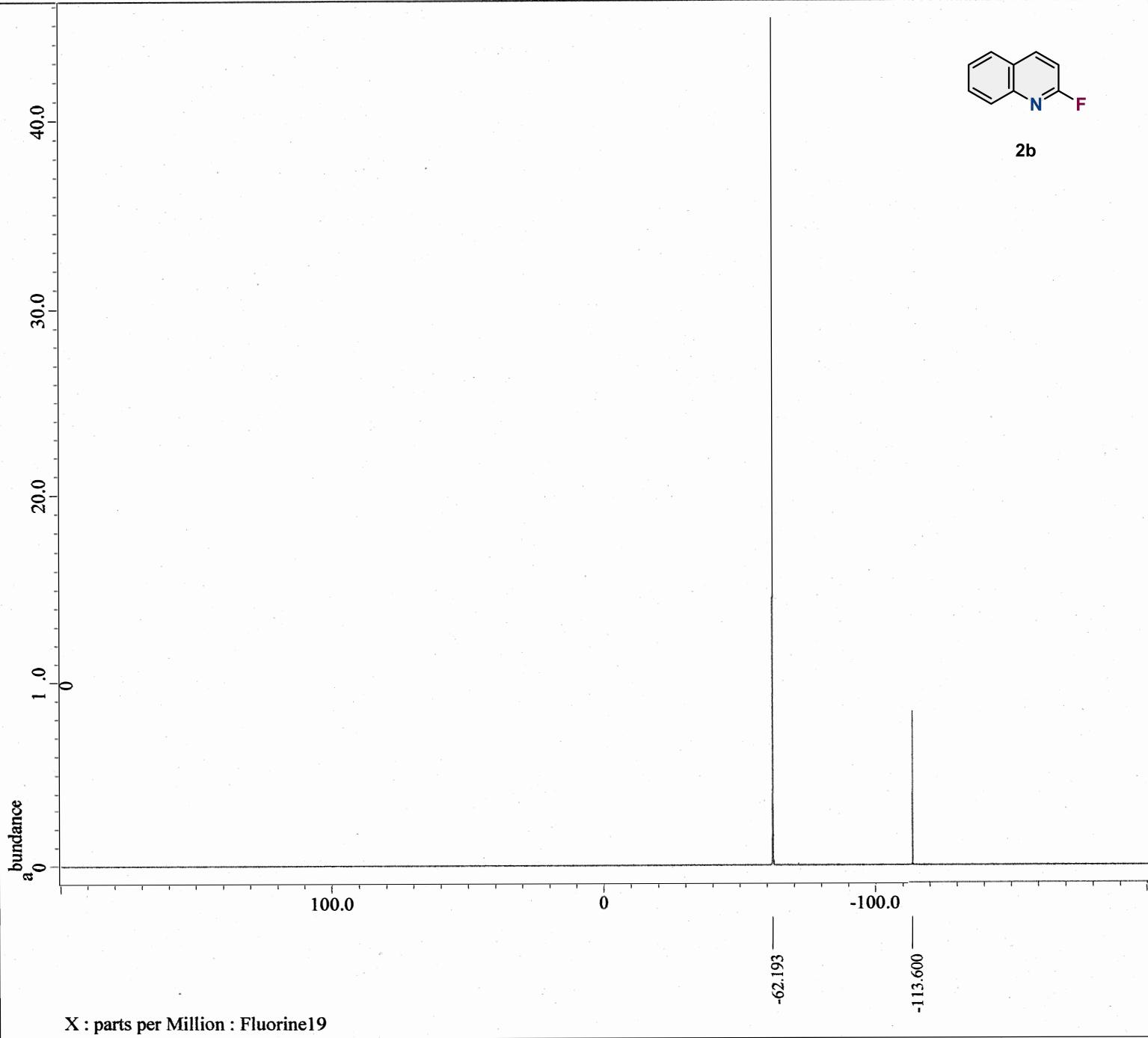
Derived from: MKN307-pure Proton-1-1.jdf

Filename = MKN307-pure\_Proton-1-2.jdf  
Author = element  
Experiment = proton.jxp  
Sample\_Id = MKN307-pure  
Solvent = CHLOROFORM-D  
Actual\_Start\_Time = 27-FEB-2024 10:07:26  
Revision\_Time = 17-JUN-2024 13:15:45



2b



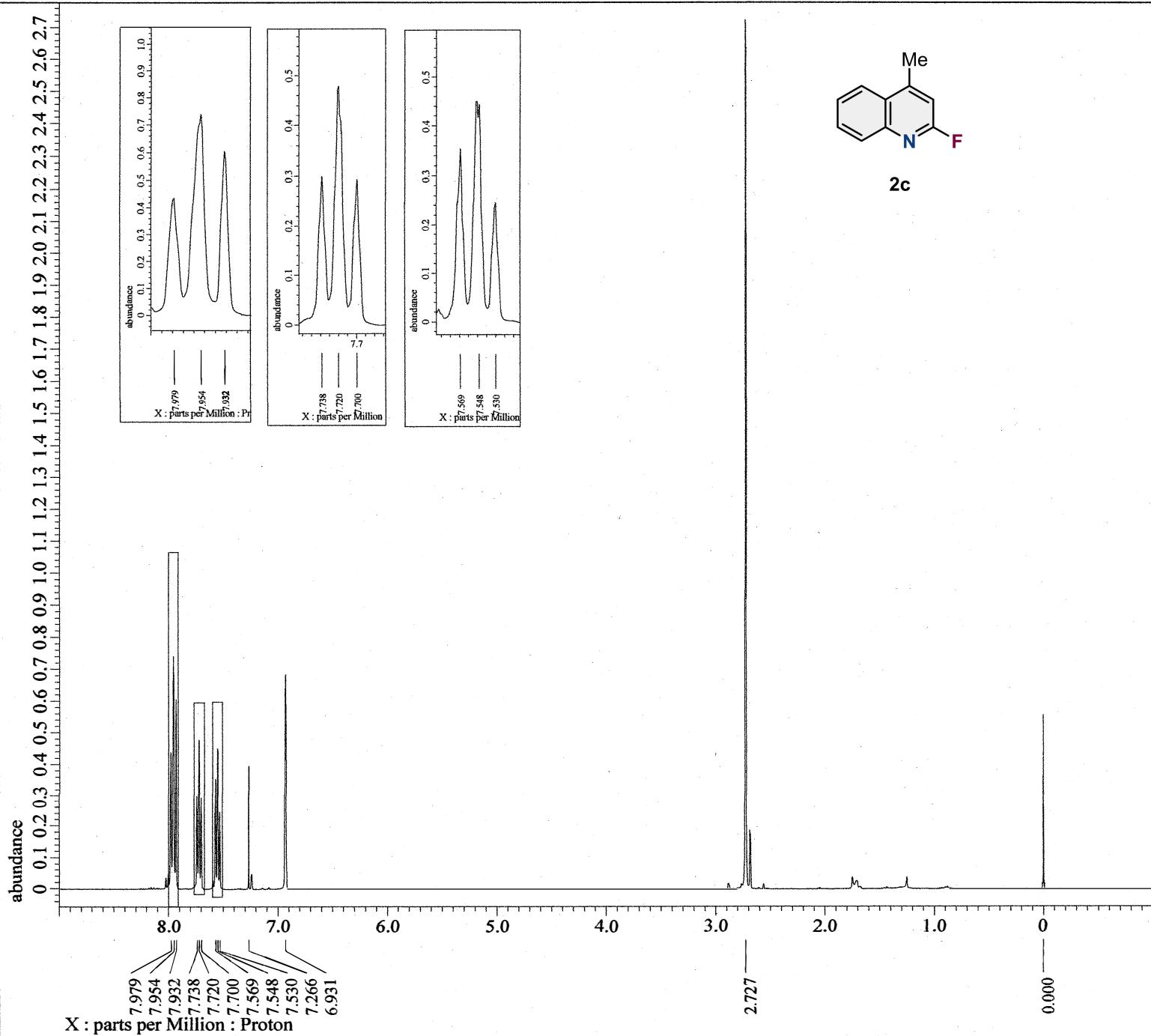


```

---- PROCESSING PARAMETERS ---
dc_balance( 0, FALSE )
sexp{ 0.2[Hz], 0.0[s] )
trap zeid( 0[%], 0[%], 80[%], 100[%] )
zer dfl{ 1, TRUE )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm
phase( 6.31854, -50.40248, 78.10163[%] )

Filename      = MKN307-pure-int_single_pul
Author        = element
Experiment   = single_pulse.jxp
Sample_Id    = MKN307-pure-int
Solvent       = CHLOROFORM-D
Actual_Start_Time = 27-FEB-2024 10:18:18
Revision_Time = 27-FEB-2024 10:58:38
Comment       = single_pulse
Data_Format  = 1D COMPLEX
Dim_Size     = 13107
X_Domain     = Fluori
Dim_Title    = Fluorine19
DimUnits     = [ppm]
Dimensions   = X
Spectrometer = DELTA2_NMR
Field_Strength = 9.4073814[T] (400[MHz])
X_Acq_Duration = 86.50752[ms]
X_Domain     = 19F
X_Freq        = 376.87675879[MHz]
X_Offset      = 0[ppm]
X_Points      = 16384
X_Prescans   = 1
X_Resolution = 11.55968868[Hz]
X_Sweep       = 189.39393939[kHz]
X_Sweep_Clipp_de = 151.51515152[kHz]
Irr_Domain   = Fluorine19
Irr_Freq      = 376.87675879[MHz]
Irr_Offset    = 5[ppm]
Tri_Domain   = Fluorine19
Tri_Freq      = 376.87675879[MHz]
Tri_Offset    = 5[ppm]
Clipped      = F ABE
Scans         = 8
Total_Scans   = 8
Relaxation_Delay = 5[s]
Recvr_Gain    = 46
Temp_Get      = 19.3[dC]
X_90_Width   = 7.59[us]
X_Acc_Time   = 86.50752[ms]
X_Angle       = 45[deg]
X_Atn         = 3[dB]
X_Pulse       = 3.795[us]
Irr_Mode      = Off
Tri_Mode      = Off
Dante_Presat = FALSE
Initial_Wait  = 1[s]
Repetition_Time = 5.08650752[s]

```



```
----- PROCESSING PARAMETERS -----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN196-pure2 Proton-1-1.jdf

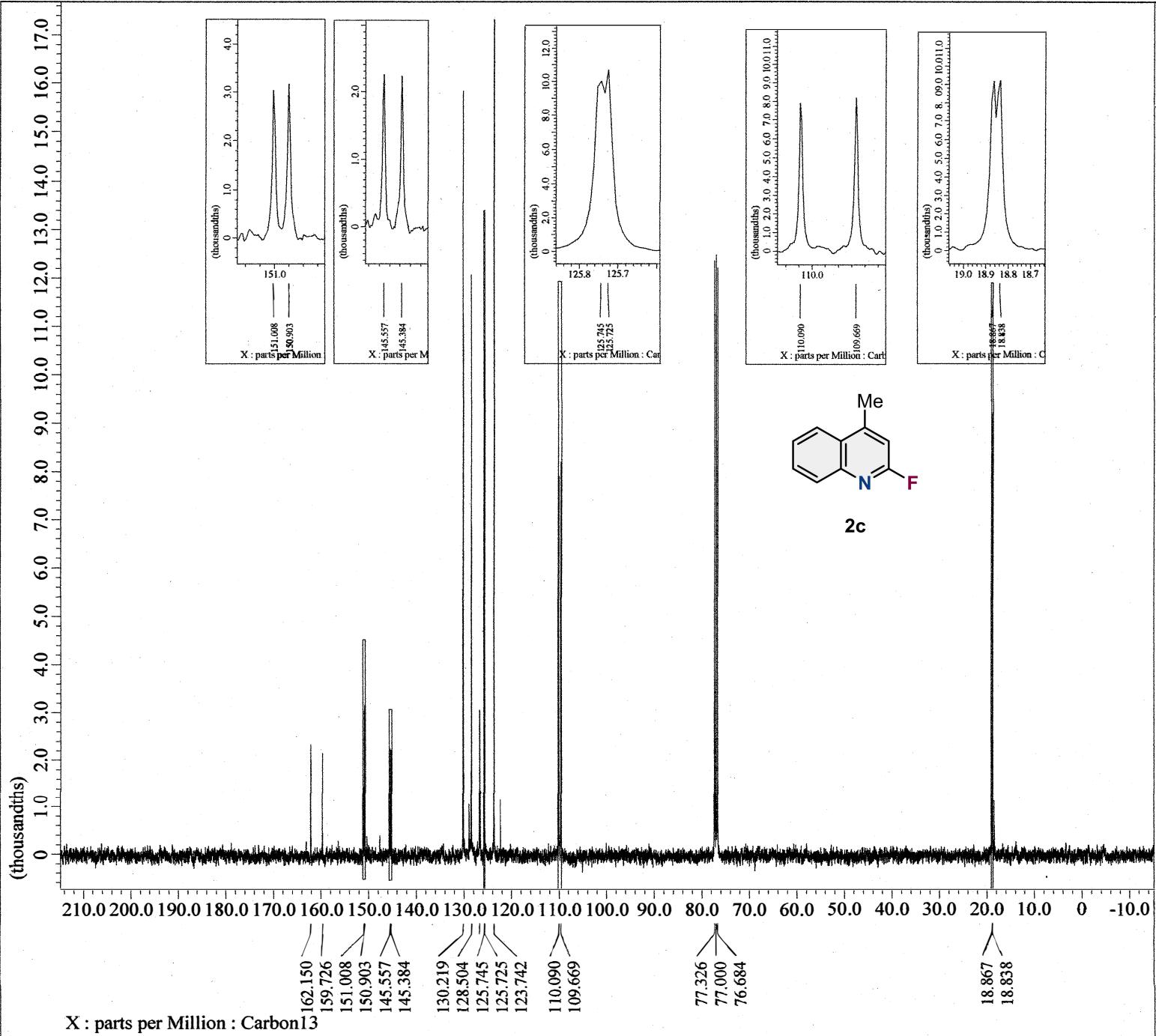
```

Filename = MKN196-pure2_Proton-1-
Author = element
Experiment = proton_auto.jxp
Sample_Id = MKN196-pure2
Solvent = CHLOROFORM-D
Actual_Start_Time = 13-OCT-2023 14:56:10
Revision_Time = 15-JUN-2024 17:34:36

Comment = single pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Proton
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions =
Spectrometer = X
= DELTA2_NMR

Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 2.20725248[s]
X_Domain = Proton
X_Freq = 395.88430144[MHz]
X_Offset =
X_Points = 16384
X_Prescans =
X_Resolution = 0.45305193[Hz]
X_Sweep = 7.42280285[kHz]
X_Sweep_Clipped = 5.93824228[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset =
Tri_Domain = Proton
Tri_Freq = 395.88430144[MHz]
Tri_Offset =
Blanking =
Clipped =
Scans = 8
Total_Scans = 8

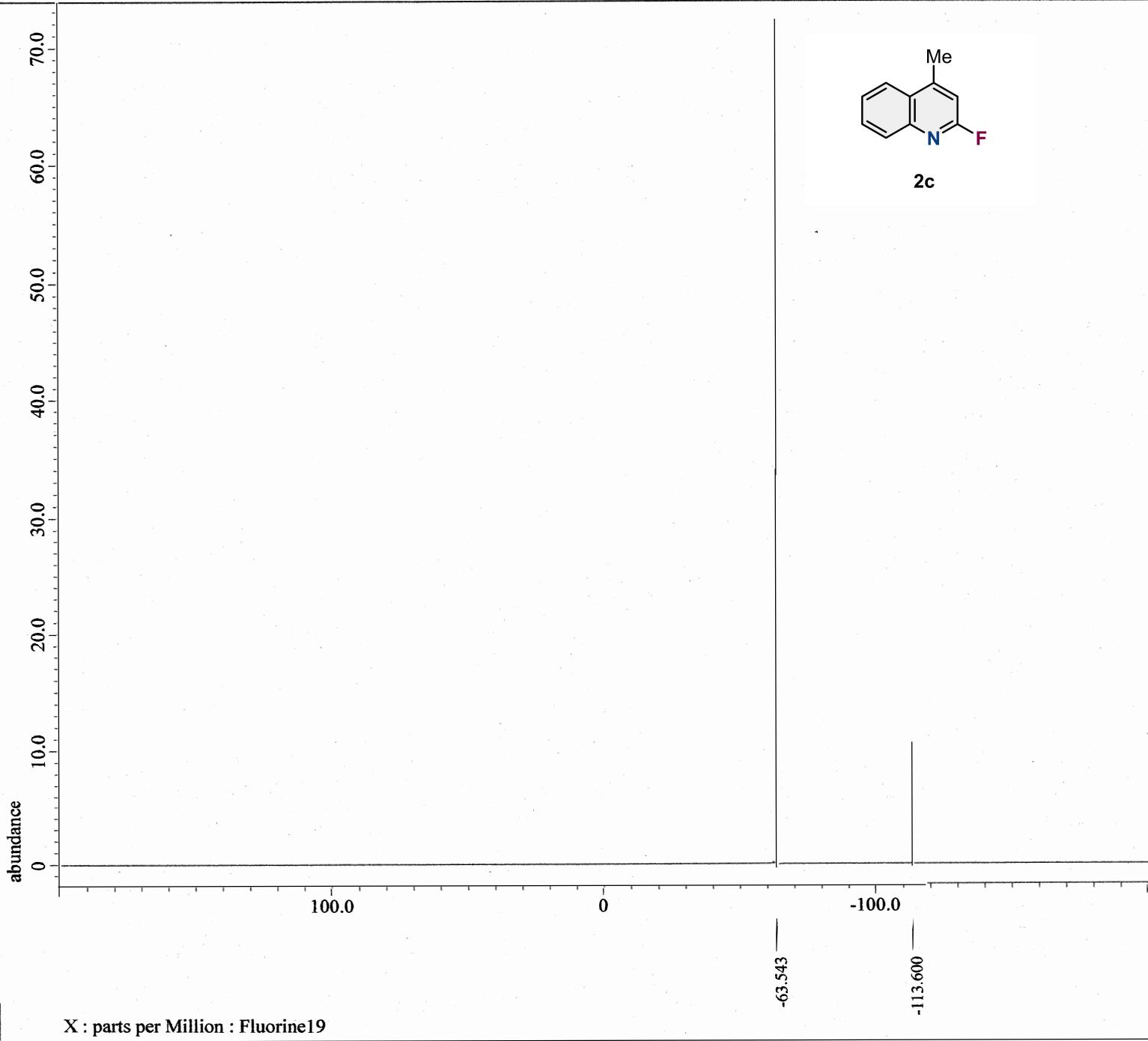
Relaxation_Delay = 5[s]
Recvr_Gain = 46
Temp_Get = 17.5[dC]
X_90_Width = 6.34[us]
X_Acq_Time = 2.20725248[s]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 3.17[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180},
```



```
---- PROCESSING PARAMETERS ----
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN196-pure2 Carbon-1-1.jdf

Filename	= MKN196-pure2_Carbon
Author	= element
Experiment	= carbon auto.jxp
Sample_Id	= MKN196-pure2
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 13-OCT-2023 16:47:1
Revision_Time	= 15-JUN-2024 17:49:4
Comment	= single pulse decoup
Data_Format	= 1D COMPLEX
Dim_Size	= 26214
X_Domain	= Carbon13
Dim_Title	= Carbon13
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.2982153[T] (400[M)
X_Acc_Duration	= 1.048576[s]
X_Domain	= Carbon13
X_Freq	= 99.54517646[MHz]
X_Offset	= 100 [ppm]
X_Points	= 32768
X_Prescans	= 4
X_Resolution	= 0.95367432[Hz]
X_Sweep	= 31.25[kHz]
X_Sweep_Clipped	= 25[kHz]
Irr_Domain	= Proton
Irr_Freq	= 395.88430144[MHz]
Irr_Offset	= 5[ppm]
Blanking	= 5.0[us]
Clipped	= FALSE
Scans	= 128
Total_Scans	= 128
Relaxation_Delay	= 2[s]
Recvr_Gain	= 50
Temp_Get	= 17.5[dC]
X_90_Width	= 11.5[us]
X_Acc_Time	= 1.048576[s]
X_Angle	= 30[deg]
X_Atn	= 9[dB]
X_Pulse	= 3.83333333[us]
Irr_Atn_Dec	= 30.172[dB]
Irr_Atn_Dec_Calc	= 30.172[dB]
Irr_Atn_Dec_Default_Calc	= 30.172[dB]
Irr_Noe	= 30.172[dB]
Irr_Dec_Bandwidth_Hz	= 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm	= 12.08082432[ppm]
Irr_Dec_Freq	= 395.88430144[MHz]
Irr_Dec_Merit_Factor	= 2.2
Irr_Decoupling	= TRUE
Irr_Noe	= TRUE



```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
t_rapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

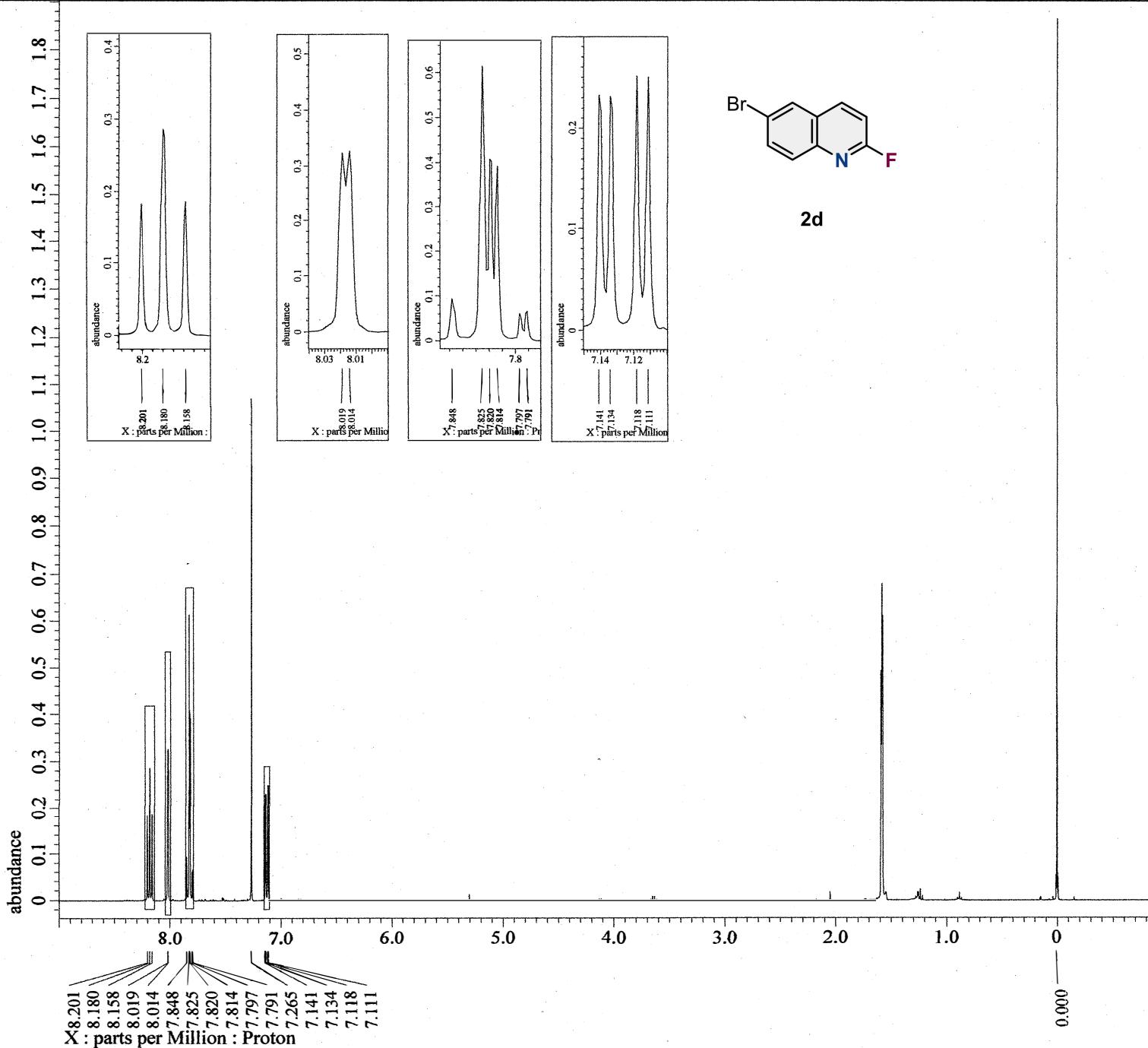
```

Filename      = MKN196-pure_int_singlepulse
Author        = element
Experiment   = single_pure_int.jxp
Sample_Id    = MKN196-pure_int
Solvent       = CHLOROFORM-D
Actual_Start_Time = 13-OCT-2023 15:58:23
Revision_Time = 24-JAN-2024 11:41:04

Comment       = single_pulse
Data_Format  = 1D COMPLEX
Dim_Size     = 13107
X_Domain    = Fluori
Dim_Title   = Fluorine19
Dim_Units   = [ppm]
Dimensions  = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.4073814[T] (400[MHz])
X_Acq_Duration = 86.50752[ms]
X_D_main      = 19F
X_Freq         = 376.87675879[MHz]
X_Offset       = 0[ppm]
X_Points      = 16384
X_Prescans    = 1
X_Resolution  = 11.55968868[Hz]
X_Sweep       = 189.39393939[kHz]
X_Sweep_Clipped = 151.51515152[kHz]
Irr_Domain   = Fluorine19
Irr_Freq      = 376.87675879[MHz]
Irr_Offset    = 5[ppm]
Tri_Domain   = Fluorine19
Tri_Freq      = 376.87675879[MHz]
Tri_Offset    = 5[ppm]
Clipped      = FALSE
Scans         = 8
Total_Scans   = 8

Relaxation_Delay = 5[s]
Recvr_Gain     = 44
Temp_Get        = 19.6[dC]
X_90_Width     = 7.59[us]
X_Acq_Time     = 86.50752[ms]
X_Angle         = 45[deg]
X_Atn          = 3[db]
X_Pulse         = 3.795[us]
Irr_Mode        = Off
Tri_Mode        = Off
Dante_Presat   = FALSE
Initial_Wait   = 1[s]
Repetition_Time = 5.08650752[s]
```



```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN175-pure2 Proton-1-1.jdf

```

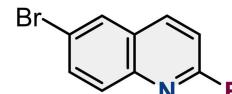
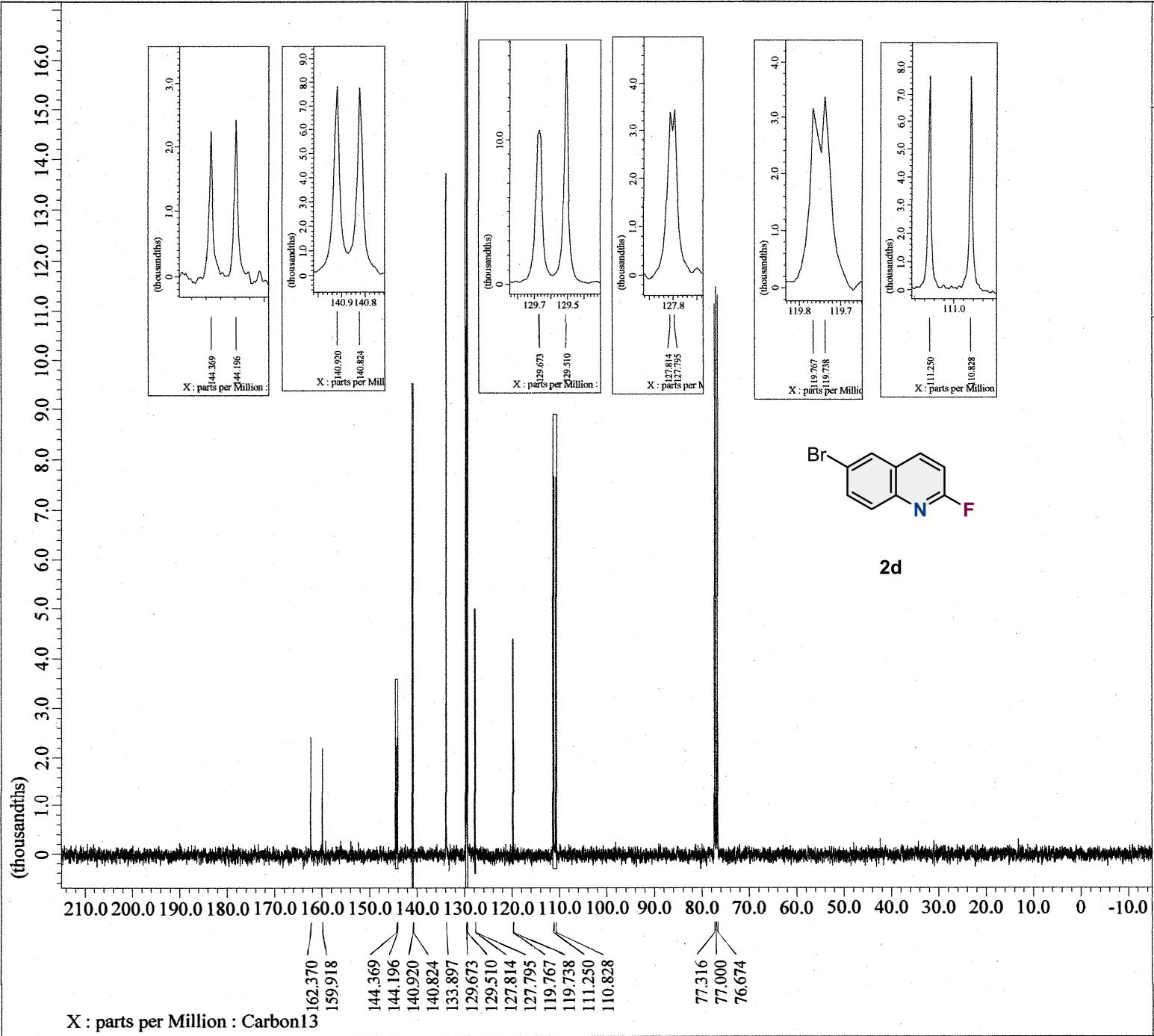
Filename = MKN175-pure2_Proton-1-
Author = element
Experiment = proton_auto.jxp
Sample_Id = MKN175-pure2
Solvent = CHLOROFORM-D
Actual_Start_Time = 29-SEP-2023 13:08:04
Revision_Time = 15-JUN-2024 16:06:04

Comment = single_pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Proton
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions =
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 2.20725248[s]
X_Domain = Proton
X_Freq = 395.88430144[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans =
X_Resolution = 0.45305193[Hz]
X_Sweep = 7.42280285[kHz]
X_Sweep_Clipped = 5.93824228[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 395.88430144[MHz]
Tri_Offset = 5[ppm]
Blanking =
Clipped =
Scans = 8
Total_Scans = 8

Relaxation_Delay = 5[s]
Recv_Gain = 56
Temp_Get = 19[dC]
X_90_Width = 6.34[us]
X_Acq_Time = 2.20725248[s]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 3.17[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180,

```



**2d**

```
---- PROCESSING PARAMETERS ----
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN175-pure2 Carbon-1-1.jdf

```

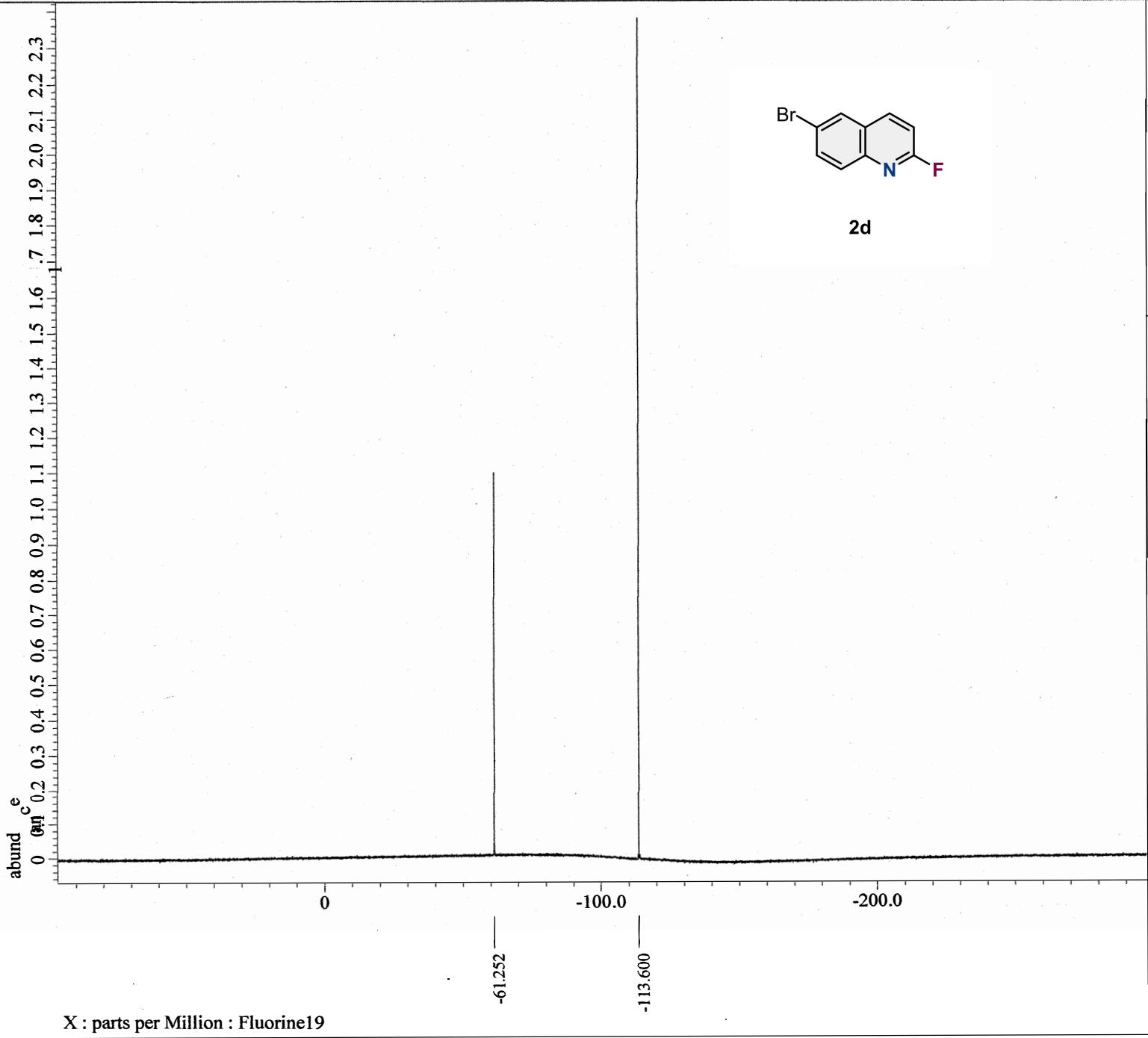
Filename = MKN175-pure2_Carbon
Author = element
Experiment = carbon_auto.jxp
Sample_Id = MKN175-pure2
Solvent = CHLOROFORM-D
Actual_Start_Time = 14-OCT-2023 11:17:5
Revision_Time = 15-JUN-2024 16:19:3

Comment = single pulse decoupl
Data_Format = 1D COMPLEX
Dim_Size = 26214
X_Domain = Carbon13
Dim_Title = Carbon13
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[M
X_Acq_Duration = 1.048576[s]
X_Domain = Carbon13
X_Freq = 99.54517646[MHz]
X_Offset = 100[ppm]
X_Points = 32768
X_Prescans = 4
X_Resolution = 0.95367432[Hz]
X_Sweep = 31.25[kHz]
X_Sweep_Clipped = 25[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Blanking = 5.0[us]
Clipped = FALSE
Scans = 128
Total_Scans = 128

Relaxation_Delay = 2[s]
Recvr_Gain = 50
Temp_Get = 19.5[dC]
X_90_Width = 11.5[us]
X_Acq_Time = 1.048576[s]
X_Angle = 30[deg]
X_Atn = 9[dB]
X_Pulse = 3.83333333[us]
Irr_Atn_Dec = 30.172[dB]
Irr_Atn_Dec_Calc = 30.172[dB]
Irr_Atn_Dec_Default_Calc = 30.172[dB]
Irr_Atn_Noe = 30.172[dB]
Irr_Dec_Bandwidth_Hz = 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm = 12.08082432[ppm]
Irr_Dec_Freq = 395.88430144[MHz]
Irr_Dec_Merit_Factor = 2.2
Irr_Decoupling = TRUE
Irr_Noe = TRUE

```



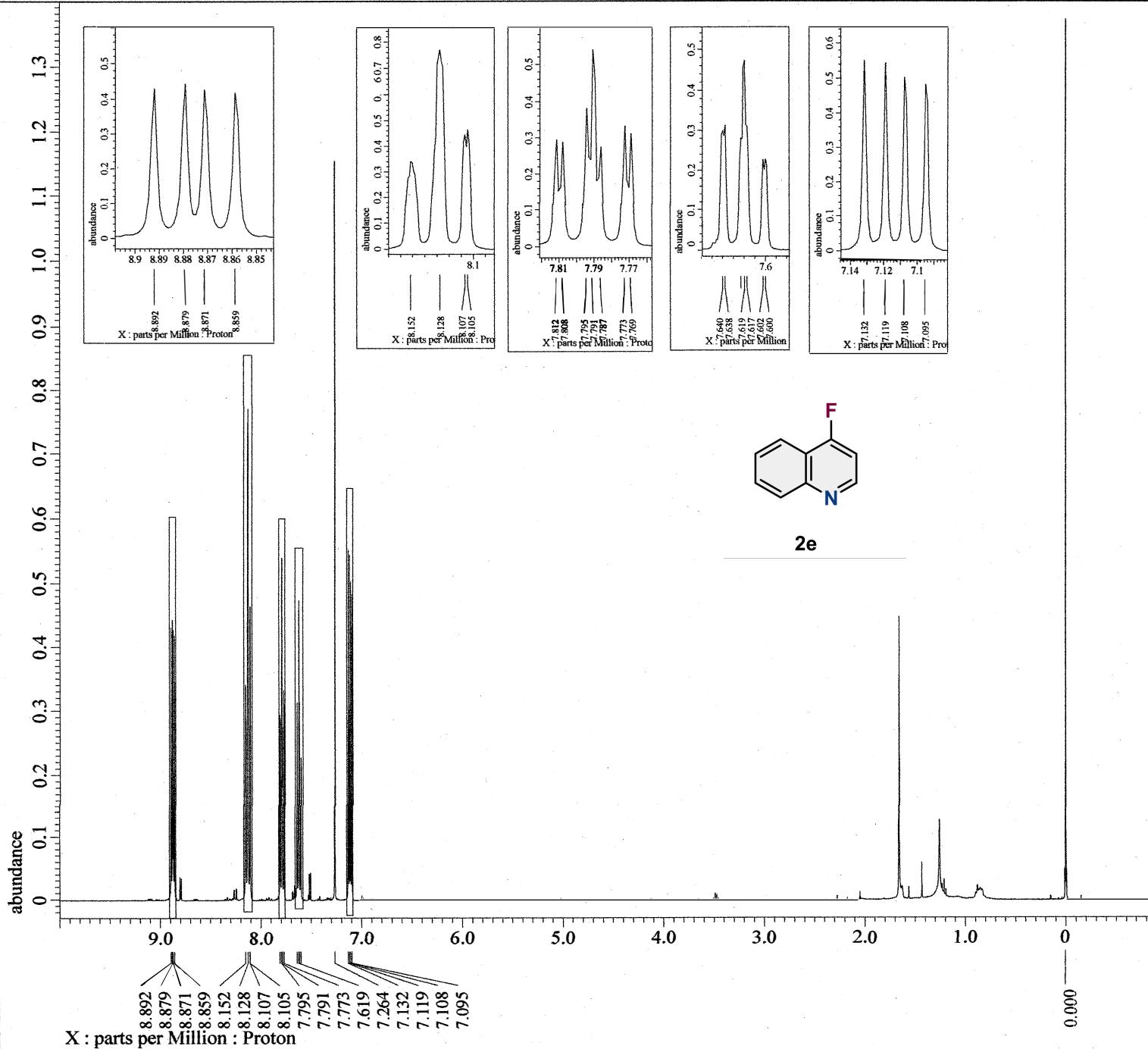
```
----- PROCESSING PARAMETERS -----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
fft( 1, TRUE, TRUE )
machinephase
ppm
phase( 83.95055, 0, 50[%] )

Filename = MKN175-pure2_int_singl
Author = element
Experiment = single_pulse.jxp
Sample_Id = MKN175-pure2_int
Solvent = CHLOROFORM-D
Actual_Start_Time = 14-OCT-2023 11:13:30
Revision_Time = 24-JAN-2024 11:35:11

Comment = single_pulse
Data_Format = 1D COMP EX
Dim_Size = 13107
X_Domain = Fluorine19
Dim_Tittle = Fluorine19
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T ] (400[MHz])
X_Acq_Duration = 89.12896[ms]
X_Domain = Fluorine19
X_Freq = 372.50336686[MHz]
X_Offset = -100[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 11.21969784[Hz]
X_Sweep = 183.82352941[kHz]
X_Sweep_Clipped = 147.05882353[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 372.50336686[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Fluorine19
Tri_Freq = 372.50336686[MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

Relaxation_Delay = 5[s]
Recvr_Gain = 56
Temp_Get = 19.1[dC]
X_90_Width = 8.03[us]
X_Acq_Time = 89.12896[ms]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 4.015[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180,
```



```

----- PROCESSING PARAMETERS -----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm

```

Derived from: MKN281-pure Proton-1-1.jdf

```

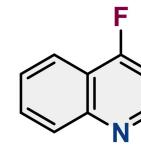
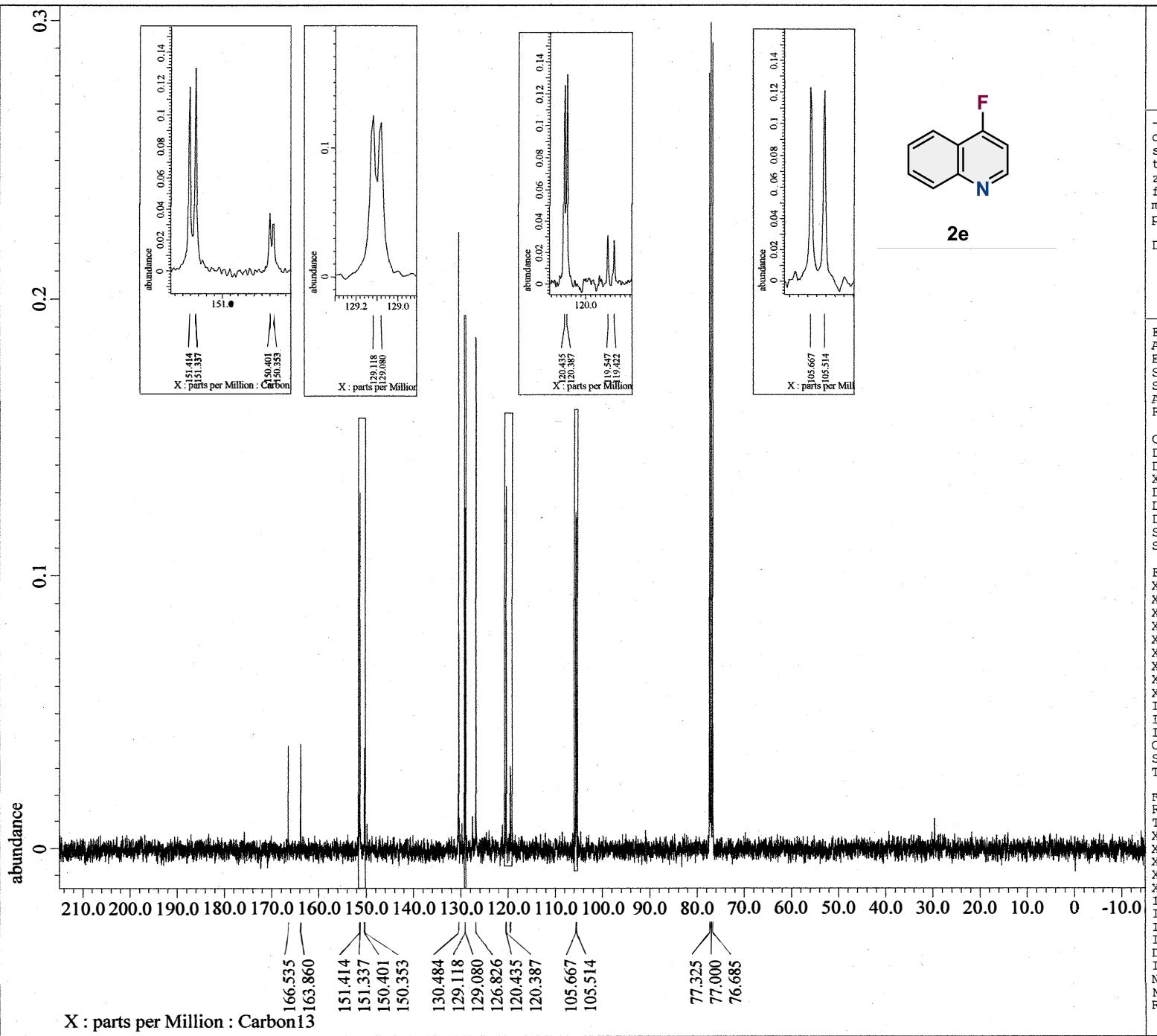
Filename = MKN281-pure_Proton-1-2
Author = element
Experiment = proton_auto.jxp
Sample_Id = MKN281-column
Solvent = CHLOROFORM-D
Actual_Start_Time = 25-JAN-2024 20:22:45
Revision_Time = 15-JUN-2024 16:42:20

Comment = single pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Proton
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 2.20725248[s]
X_Domain = Proton
X_Freq = 395.88430144[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 0.45305193[Hz]
X_Sweep = 7.42280285[kHz]
X_Sweep_Clipped = 5.93824228[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 395.88430144[MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

Relaxation_Delay = 5[s]
Recvr_Gain = 56
Temp_Get = 19.6[dC]
X_90_Width = 6.34[us]
X_Acq_Time = 2.20725248[s]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 3.17[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180,

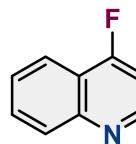
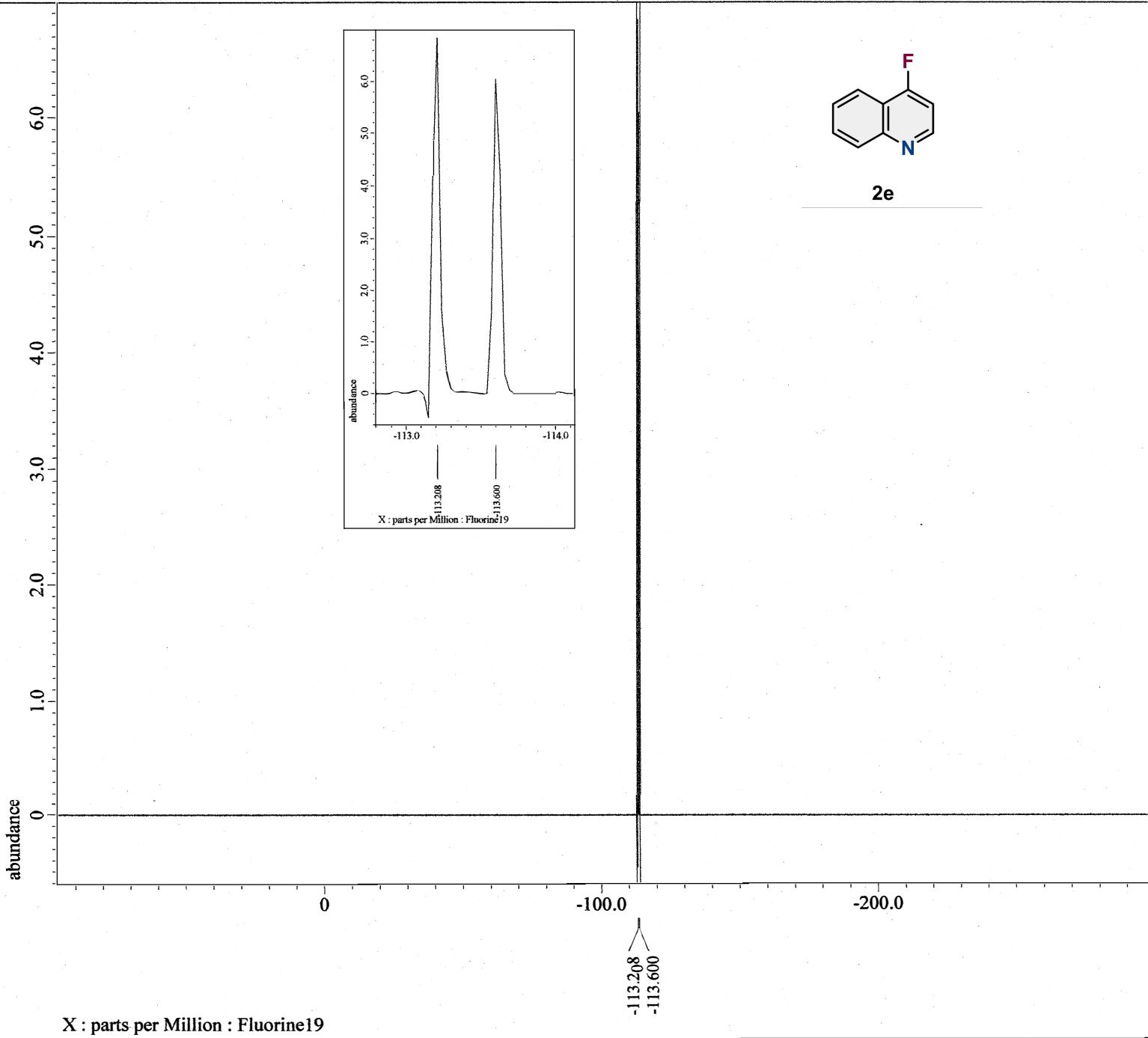
```



```
----- PROCESSING PARAMETERS -----
dc_balance( 0, FALSE )
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN281-pure Carbon-1-1.jdf

```
Filename      = MKN281-pure_Carbon-1-3.jdf
Author        = element
Experiment   = carbon.jxp
Sample_Id    = MKN281-pure
Solvent       = CHLOROFORM-D
Actual_Start_Time = 3-FEB-2024 12:56:29
Revision_Time = 15-JUN-2024 17:12:39
Comment       = single pulse decoupled gat
Data_Format  = 1D COMPLEX
Dim_Size     = 26214
X_Domain    = Carbon
Dim_Title   = Carbon13
Dim_Units   = [ppm]
Dimensions  = X
Site         = JNM-ECS400
Spectrometer = DELTA2_NMR
Field_Strength = 9.37221[T] (400[MHz])
X_Acq_Duration = 1.04333312[s]
X_Domain    = 13C
X_Freq       = 100.33735165[MHz]
X_Offset     = 100.0[ppm]
X_Points    = 32768
X_Prescans  = 4
X_Resolution = 0.95846665[Hz]
X_Sweep      = 31.40703518[kHz]
X_Sweep_Clipped = 25.12562814[kHz]
Irr_Domain  = Proton
Irr_Freq    = 399.03472754[MHz]
Irr_Offset  = 5.0[ppm]
Clipped     = FALSE
Scans        = 128
Total_Scans = 128
Relaxation_Delay = 2[s]
Recvr_Gain   = 50
Temp_Get     = 19.1[dC]
X_90_Width  = 10.9[us]
X_Acq_Time  = 1.04333312[s]
X_Angle      = 30[deg]
X_Atn        = 5.4[dB]
X_Pulse      = 3.633333333[us]
Irr_Atn_Dec = 25.823[dB]
Irr_Atn_Noe = 25.823[dB]
Irr_Noise    = WALTZ
Irr_Pwidth   = 0.115[ms]
Decoupling   = TRUE
Initial_Wait = 1[s]
Noe          = TRUE
Noe_Time    = 2[s]
Repetition_Time = 3.04333312[s]
```

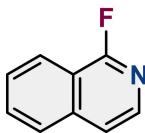
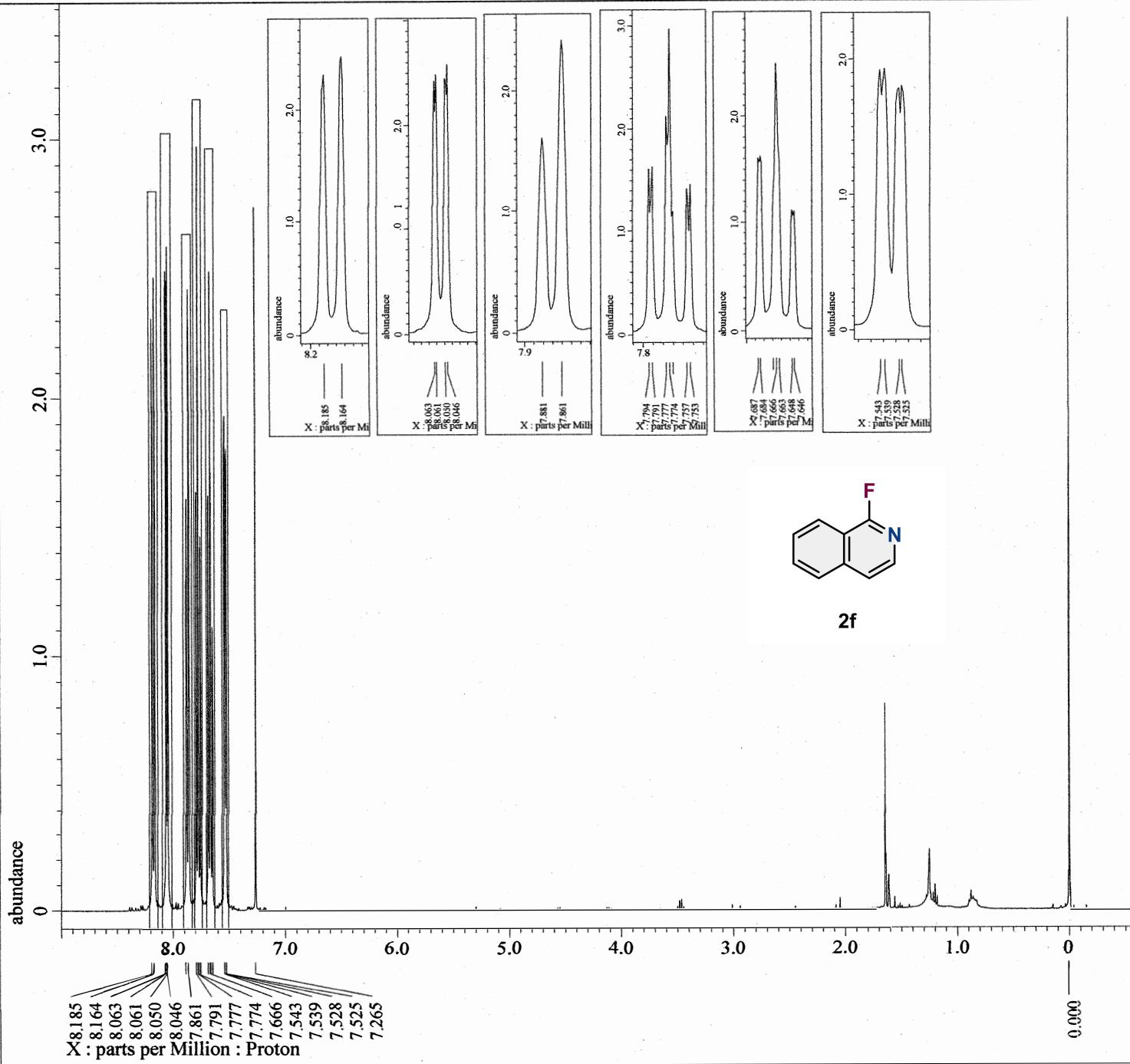


**2e**

```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm
Derived from: MKN281-pure-int single pulse-1-1
```

Filename	= MKN281-pure-int_single
Author	= element
Experiment	= single_pulse.jxp
Sample_Id	= MKN281-pure-int
Solvent	= CHLOROF EM-D
Actual_Start_Time	= 1-FEB-2024 15:22:36
Revision_Time	= 15-JUN-2024 17:19:58
Comment	= single_pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Domain	= F_luorine19
Dim_Title	= Fluorine19
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Feld_Strength	= 9.2982153[T] (400[MHz])
X_AccDuration	= 89.12896[ms]
X_Domain	= F_luorine19
X_Freq	= 372.50336686[MHz]
X_Offset	= -100[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 11.21969784[Hz]
X_Sweep	= 183.82352941[kHz]
X_Sweep_Clipped	= 147.05882353[kHz]
Irr_Domain	= Fl_uorine19
Irr_Freq	= 372.50336686[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= F_luorine19
Tri_Freq	= 372.50336686[MHz]
Tri_Offset	= 5[ppm]
Blaking	= 2.0[us]
ClipEd	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recv_Gain	= 56
Temp_Get	= 18.8[dC]
X_90_Width	= 8.03[us]
X_Acq_Time	= 89.12896[ms]
X_Angle	= 45[deg]
X_Atn	= 5[dB]
X_Pulse	= 4.015[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Loop	= 500
Dante_Presat	= F_ALSE
Decimation_Rate	= 0
Initial_Wait	= 1[s]
Phase	= {0, 90, 270, 180, 180,

X : parts per Million : Fluorine19



**2f**

```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[], 0[], 80[], 100[] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

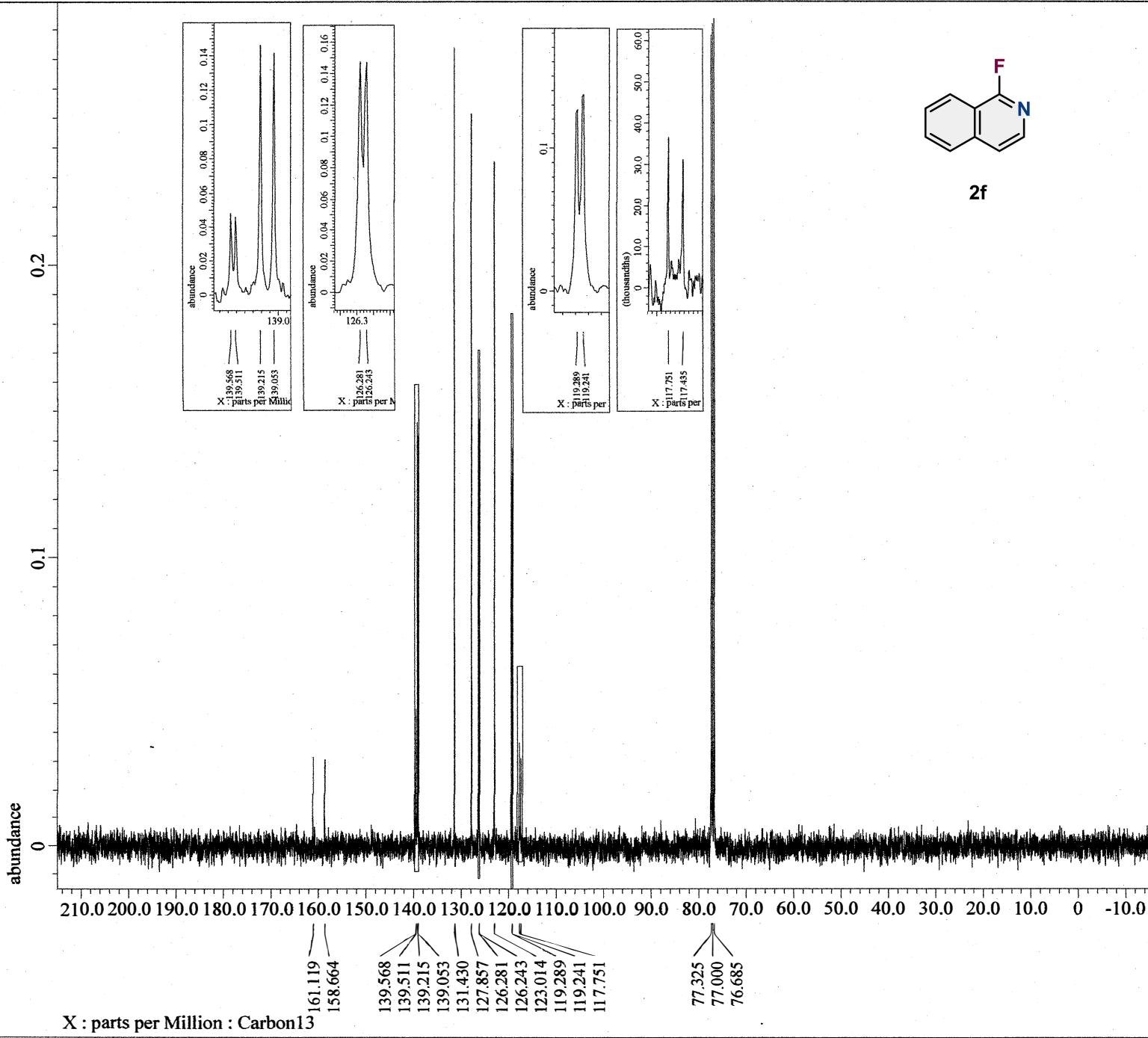
Derived from: MKN306 pure Proton-1-1.jdf

```
Filename      = MKN306_pure_Proton-1-3.jdf
Author        = element
Experiment   = proton.jpx
Sample_Id    = MKN306 column
Solvent       = CHLOROFORM-D
Actual_Start_Time = 22-FEB-2024 19:59:20
Revision_Time = 17-JUN-2024 12:12:47
```

```
Comment       = single pulse
Data_Format  = 1D COMPLEX
Dim_Size     = 13107
X_Domain    = Proton
Dim_Title   = Proton
Dim_Units   = [ppm]
Dimensions  = X
Site         = JNM-ECS400
Spectrometer = DELTA2_NMR

Field_Strength = 9.37221[T] (400[MHz])
X_Acq_Duration = 2.1889024[s]
X_Domain     = 1H
X_Freq        = 399.03472754[MHz]
X_Offset      = 5.0[ppm]
X_Points      = 16384
X_Prescans   = 1
X_Resolution  = 0.45684997[Hz]
X_Sweep       = 7.48502994[kHz]
X_Sweep_Clipped = 5.98802395[kHz]
Irr_Domain   = Proton
Irr_Freq     = 399.03472754[MHz]
Irr_Offset   = 5.0[ppm]
Tri_Domain   = Proton
Tri_Freq     = 399.03472754[MHz]
Tri_Offset   = 5.0[ppm]
Clipped      = FALSE
Scans        = 8
Total_Scans  = 8
```

```
Relaxation_Delay = 5[s]
Recvr_Gain      = 40
Temp_Get         = 18.2[dC]
X_90_Width      = 6.6[us]
X_Acq_Time      = 2.1889024[s]
X_Angle          = 45[deg]
X_Atn            = 1[dB]
X_Pulse          = 3.3[us]
Irr_Mode         = Off
Tri_Mode         = Off
Dante_Presat    = FALSE
Initial_Wait    = 1[s]
Repetition_Time = 7.1889024[s]
```



```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

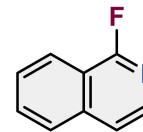
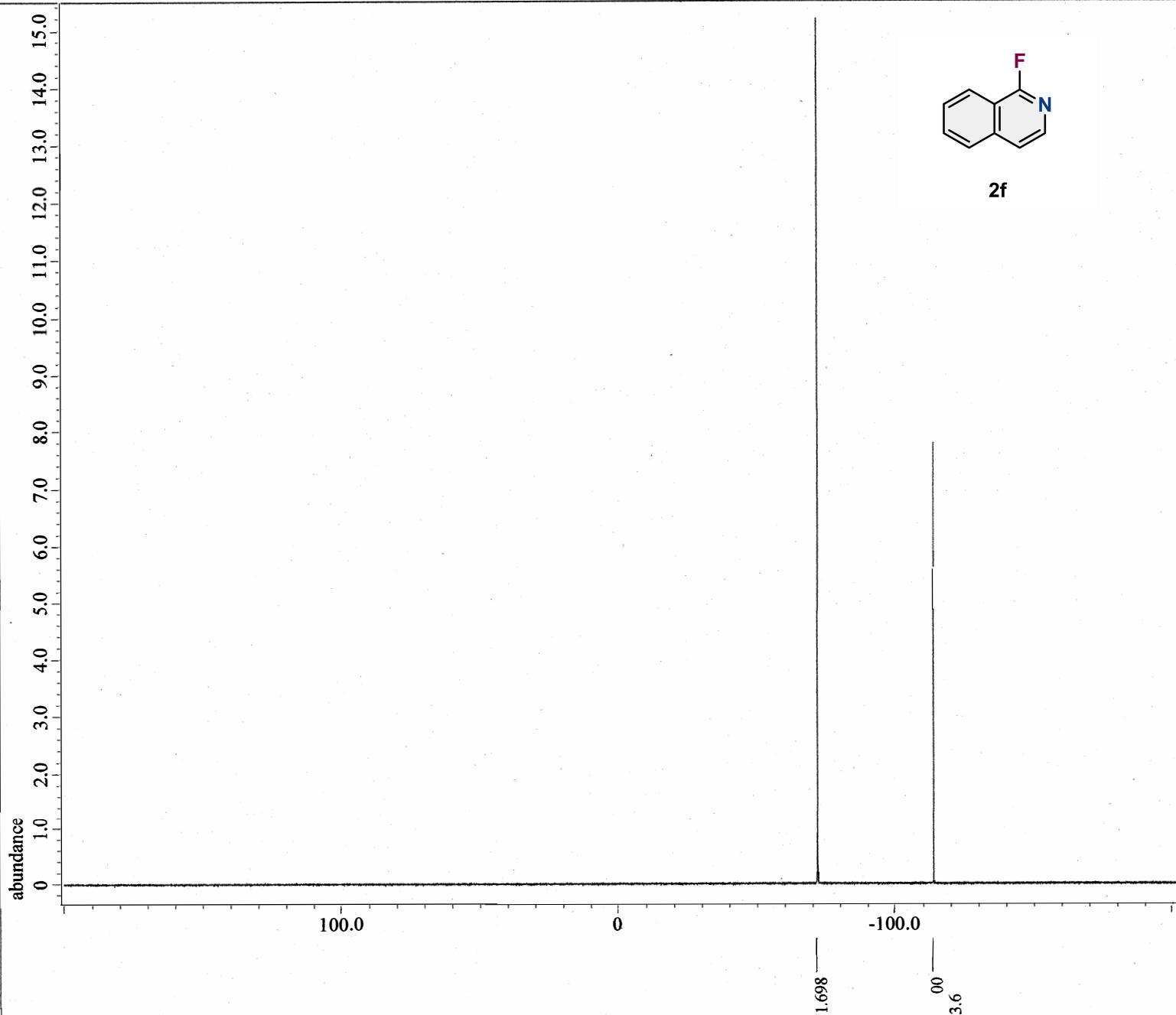
Derived from: MKN306-pure Carbon-3-1.jdf

```
Filename = MKN306-pure_Carbon-3-2.jdf
Author = element
Experiment = carbon.jxp
Sample_Id = MKN306-pure
Solvent = CHLOROFORM-D
Actual_Start_Time = 24-FEB-2024 14:10:38
Revision_Time = 17-JUN-2024 12:25:05

Comment = single pulse decoupled gat
Data_Format = 1D COMPLEX
Dim_Size = 26214
X_Domain = Carbon
Dim_Title = Carbon13
Dim_Units = [ppm]
Dimensions = X
Site = JNM-ECS400
Spectrometer = DELTA2_NMR

Field_Strength = 9.37221[T] (400[MHz])
X_Acq_Duration = 1.04333312[s]
X_Domain = 13C
X_Freq = 100.33735165[MHz]
X_Offset = 100.0[ppm]
X_Points = 32768
X_Prescans = 4
X_Resolution = 0.95846665[Hz]
X_Sweep = 31.40703518[kHz]
X_Sweep_Clipped = 25.12562814[kHz]
Irr_Domain = Proton
Irr_Freq = 399.03472754[MHz]
Irr_Offset = 5.0[ppm]
Clipped = FALSE
Scans = 64
Total_Scans = 64

Relaxation_Delay = 2[s]
Recvr_Gain = 50
Temp_Get = 18.3[dC]
X_90_Width = 10.9[us]
X_Acq_Time = 1.04333312[s]
X_Angle = 30[deg]
X_Atn = 5.4[dB]
X_Pulse = 3.633333333[us]
Irr_Atn_Dec = 25.823[dB]
Irr_Atn_Noe = 25.823[dB]
Irr_Noise = WALTZ
Irr_Pwidth = 0.115[ms]
Decoupling = TRUE
Initial_Wait = 1[s]
Noe = TRUE
Noe_Time = 2[s]
Repetition_Time = 3.04333312[s]
```



**2f**

```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

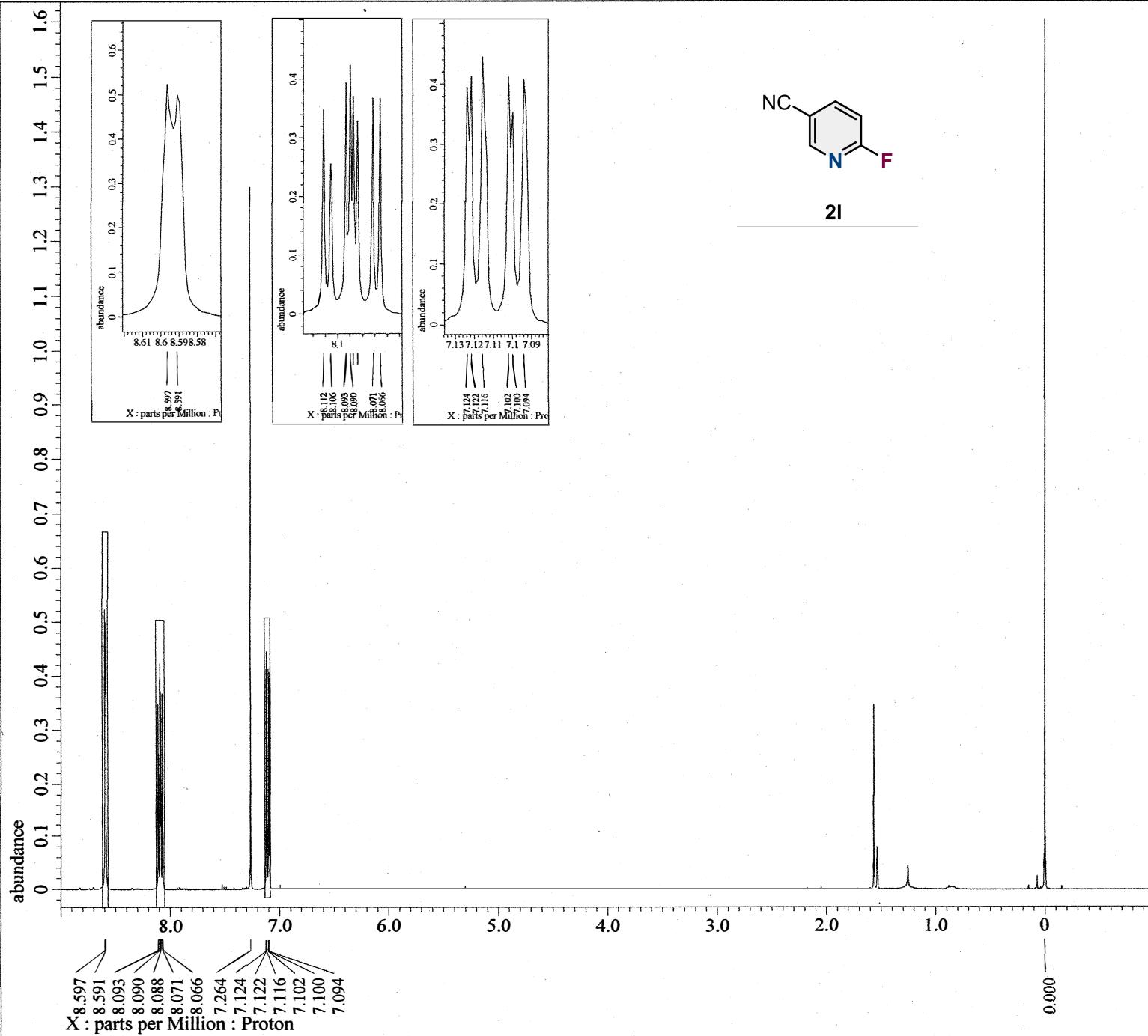
Derived from: MKN 306\_pure-int single pulse-1-1

```
Filename = MKN 306_pure-int_single_pul
Author = element
Experiment = single_pulse.jxp
Sample_Id = MKN 306_pure-int
So went = CHLOROFORM-D
Actual_Start_Time = 24-FEB-2024 13:18:27
Revision_Time = 17-JUN-2024 12:29:14

Comment = single_pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Fluori
Dim_Title = Fluorine19
Dim_Units = [ppm]
Dimensions = X
Site = JNM-ECS400
Spectrometer = DELTA2_NMR

Field_Strength = 9.37221[T] (400[MHz])
X_Acq_Duration = 86.50752[ms]
X_Domain = 19F
X_Freq = 375.46772873[MHz]
X_Offset = 0[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 11.55968868[Hz]
X_Sweep = 189.39393939[kHz]
X_Sweep_Clipped = 151.51515152[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 375.46772873[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Fluorine19
Tri_Freq = 375.46772873[MHz]
Tri_Offset = 5[ppm]
Clipped = FALSE
Scans = 8
Total_Scans = 8

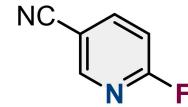
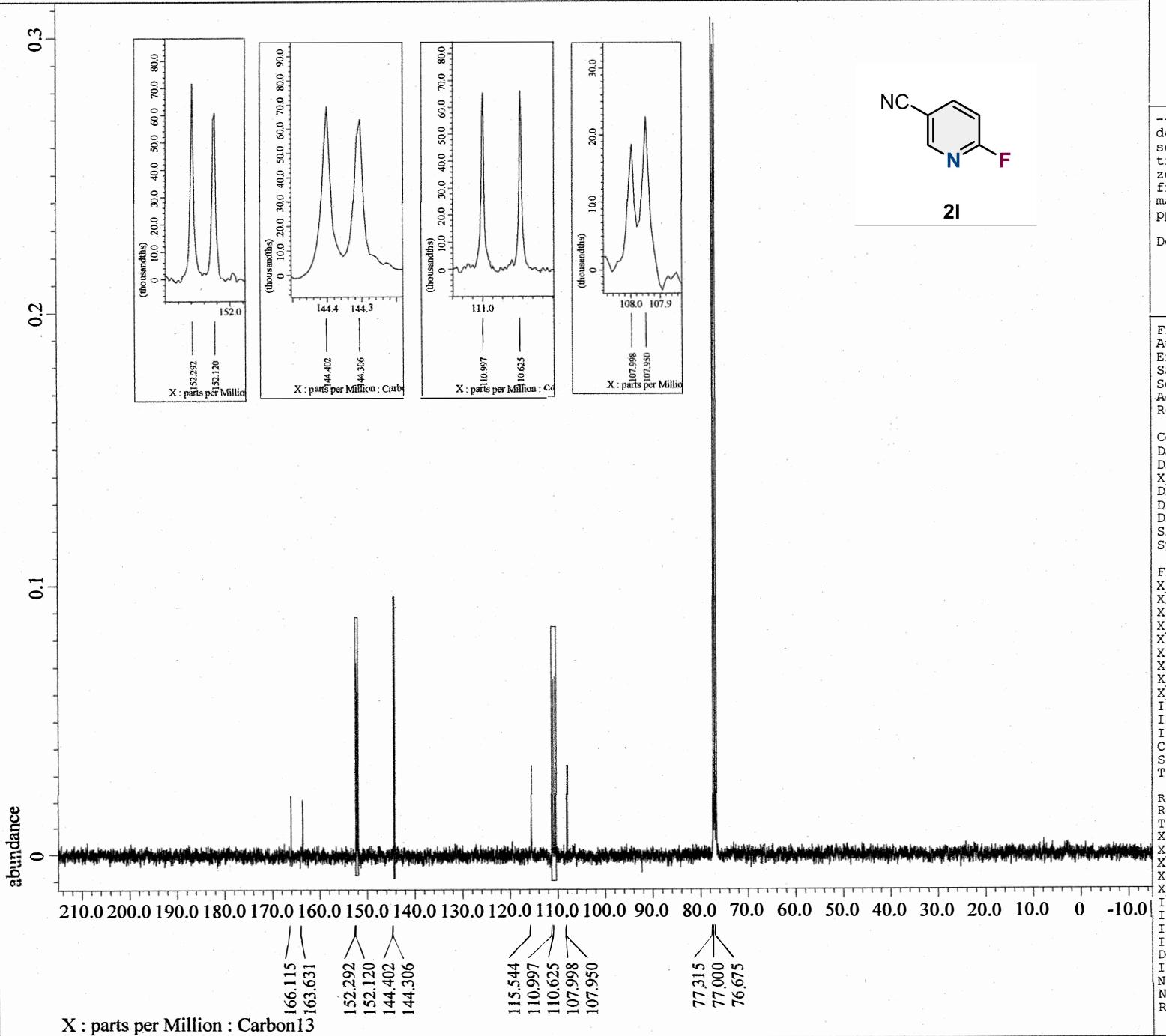
Relaxation_Delay = 5[s]
Recvr_Gain = 44
Temp_Get = 18.4[dC]
X_90_Width = 7.6[us]
X_Acq_Time = 86.50752[ms]
X_Angle = 45[deg]
X_Atn = 2.5[dB]
X_Pulse = 3.8[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Presat = FALSE
Initial_Wait = 1[s]
Repetition_Time = 5.08650752[s]
```



```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN221-pure Proton-1-1.jdf

Filename	= MKN221-pure_Proton-1-2
Author	= element
Experiment	= proton_auto.jxp
Sample_Id	= MKN221-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 23-JAN-2024 17:15:19
Revision_Time	= 17-JUN-2024 11:44:47
Comment	= single pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Domain	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.2982153[T] (400[MHz])
X_Acq_Duration	= 2.20725248[s]
X_Domain	= Proton
X_Freq	= 395.88430144[MHz]
X_Offset	= 5[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45305193[Hz]
X_Sweep	= 7.42280285[kHz]
X_Sweep_Clipped	= 5.93824228[kHz]
Irr_Domain	= Proton
Irr_Freq	= 395.88430144[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= Proton
Tri_Freq	= 395.88430144[MHz]
Tri_Offset	= 5[ppm]
Blanking	= 2.0[us]
Clipped	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recv_Gain	= 56
Temp_Get	= 19.3[dc]
X_90_Width	= 6.34[us]
X_Acq_Time	= 2.20725248[s]
X_Angle	= 45[deg]
X_Atn	= 5[dB]
X_Pulse	= 3.17[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Loop	= 500
Dante_Presat	= FALSE
Decimation_Rate	= 0
Initial_Wait	= 1[s]
Phase	= {0, 90, 270, 180, 180,

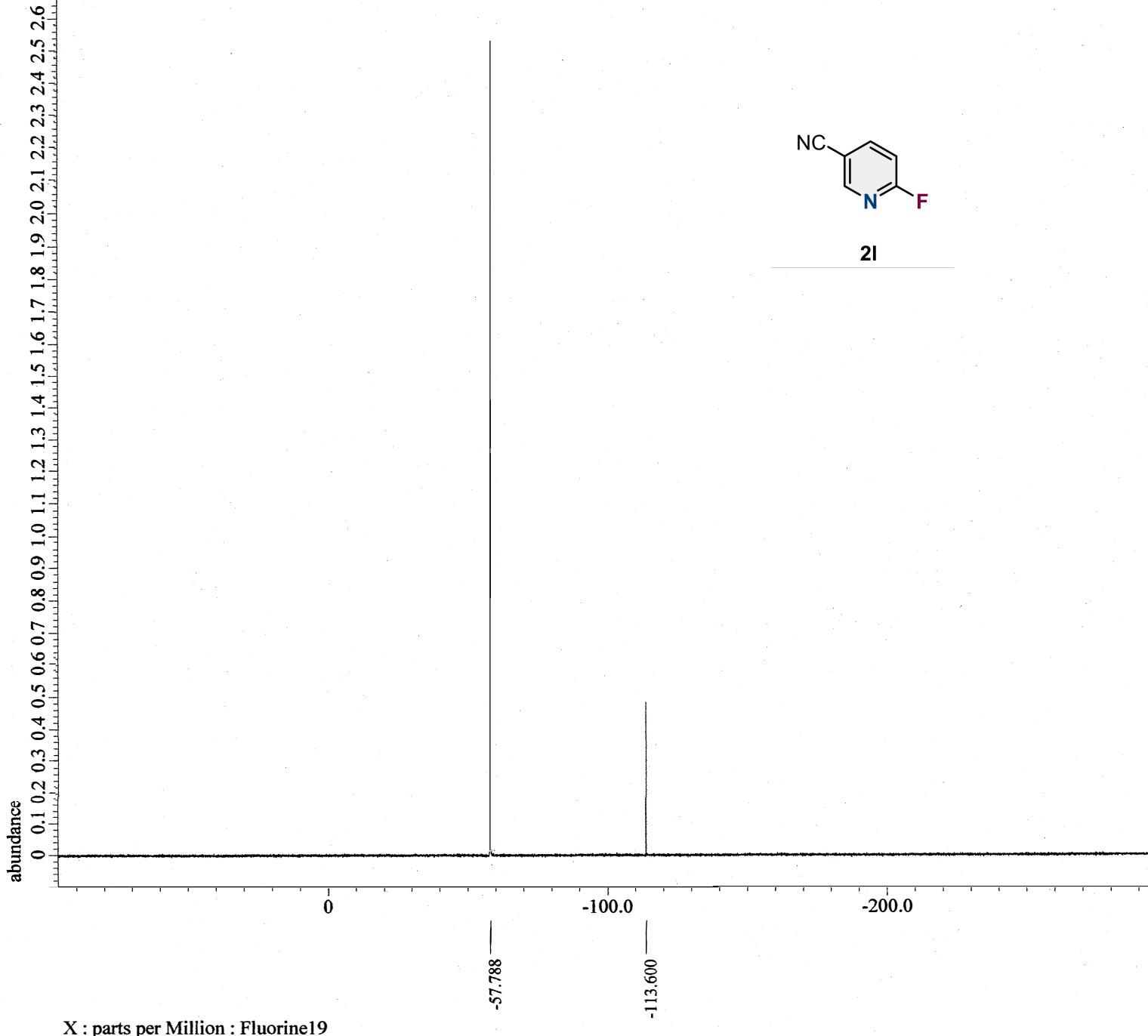


**2I**

```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN221-pure Carbon-1-1.jdf

Filename	= MKN221-pure_Carbon-1-2.jdf
Author	= element
Experiment	= carbon.jxp
Sample_Id	= MKN221-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 23-JAN-2024 18:56:15
Revision_Time	= 17-JUN-2024 11:53:19
Comment	= single pulse decoupled gat
Data_Format	= 1D COMPLEX
Dim_Size	= 26214
X_Domain	= Carbon
Dim_Title	= Carbon13
Dim_Units	= [ppm]
Dimensions	= X
Site	= JNM-ECS400
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.37221[T] (400[MHz])
X_Acq_Duration	= 1.04333312[s]
X_Domain	= 13C
X_Freq	= 100.33735165[MHz]
X_Offset	= 100.0[ppm]
X_Points	= 32768
X_Prescans	= 4
X_Resolution	= 0.95846665[Hz]
X_Sweep	= 31.40703518[kHz]
X_Sweep_Clipped	= 25.12562814[kHz]
Irr_Domain	= Proton
Irr_Freq	= 399.03472754[MHz]
Irr_Offset	= 5.0[ppm]
Clipped	= FALSE
Scans	= 256
Total_Scans	= 256
Relaxation_Delay	= 2[s]
Recvr_Gain	= 50
Temp_Get	= 19.7[dC]
X_90_Width	= 10.9[us]
X_Acq_Time	= 1.04333312[s]
X_Angle	= 30[deg]
X_Atn	= 5.4[dB]
X_Pulse	= 3.63333333[us]
Irr_Atn_Dec	= 25.823[dB]
Irr_Atn_Noe	= 25.823[dB]
Irr_Noise	= WALTZ
Irr_Fwidth	= 0.115[ms]
Decoupling	= TRUE
Initial_Wait	= 1[s]
Noe	= TRUE
Noe_Time	= 2[s]
Repetition_Time	= 3.04333312[s]



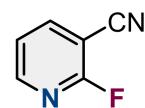
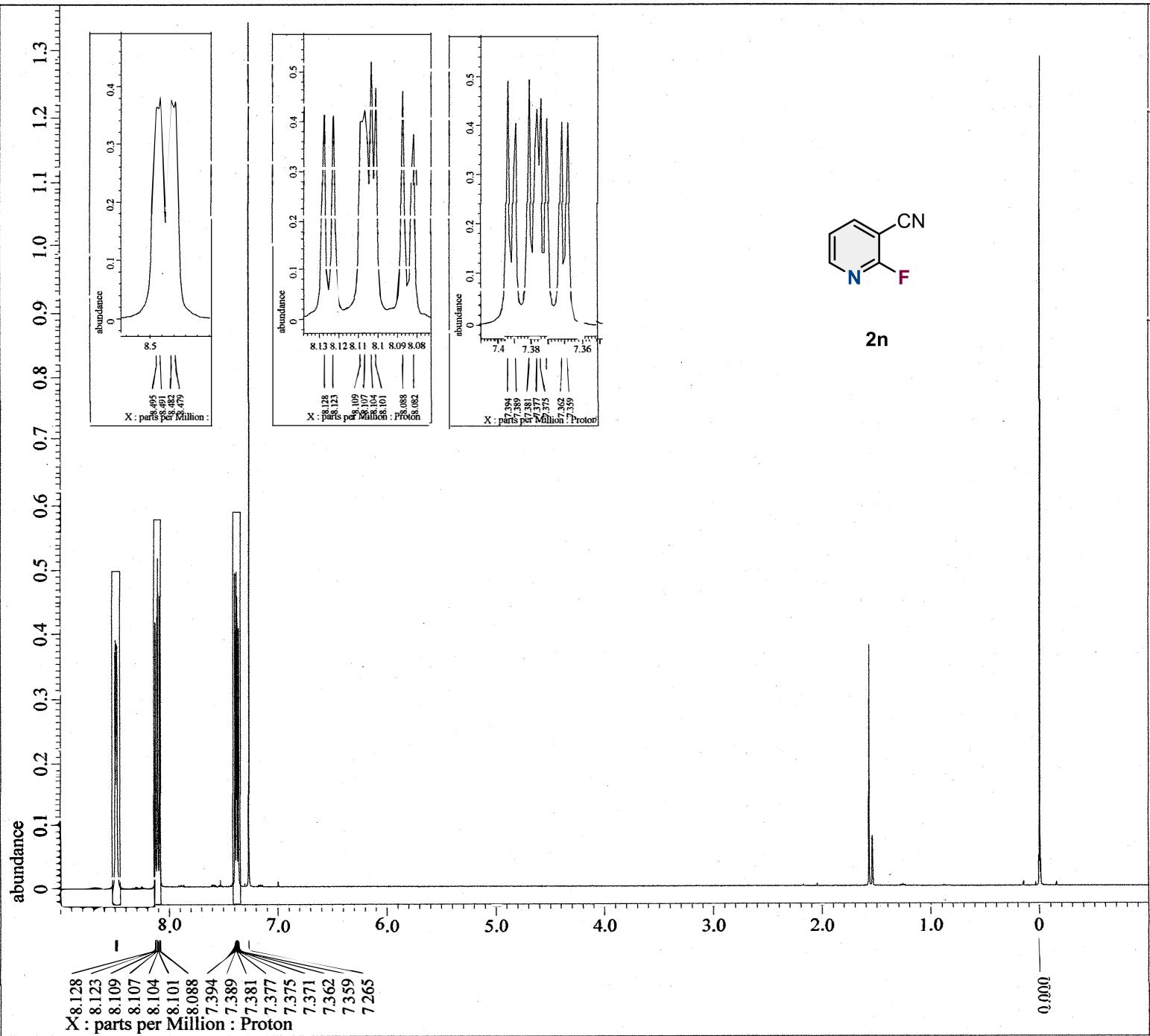
```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm

Filename = MKN221-pure-int_sigle
Autho_r = element
Experiment = single_pulse.jxp
Sample_Id = MKN221-pure-int
Solvent = CHLOROFORM-D
Actual_Start_Time = 23-JAN-2024 17:28:53
Revision_Tme = 17-JUN-2024 11:57:29

Comment = single_pulse
Data_Format = 1D_COMPLEX
Dim_Size = 13107
X_Domain = Fluorine19
Dim_Title = Fluorine19
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.2982153[T] (400[MHz]
X_Acc_Duration = 89.12896[ms]
X_Domain = Fluorine19
X_Freq = 372.50336686[MHz]
X_Offset = -100[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 11.21969784[Hz]
X_Sweep = 183.82352941[kHz]
Xsweep_Clipped = 147.05882353[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 372.50336686[MHz]
Irr_Offset = 5[ppm]
T_rdDomain = Fluorine19
T_rdFreq = 372.50336686[MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

Relaxation_Day = 5[s]
Recvr_Gain = 56
Temp_Get = 18.8[dC]
X_90_Width = 8.03[us]
X_Acq_Time = 89.12896[ms]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 4.015[us]
Irr_Mode = Off
T_rd_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180},
```



2n

```
---- PROCESSING PARAMETERS ----  
sexp( 0.2[Hz], 0.0[s] )  
trapezoid( 0[%], 0[%], 80[%], 100[%] )  
zerofill( 1, TRUE )  
fft( 1, TRUE, TRUE )  
machinephase  
ppm
```

Derived from: MKN240-pure Proton-1-1.jdf

```

File: MKN240-pure_Proton-1.jdx
Filepath: C:\Users\Public\Documents\Bruker\NMR\2024\01\2024-01-23\17:22:40\

[File]
Filepath = C:\Users\Public\Documents\Bruker\NMR\2024\01\2024-01-23\17:22:40\

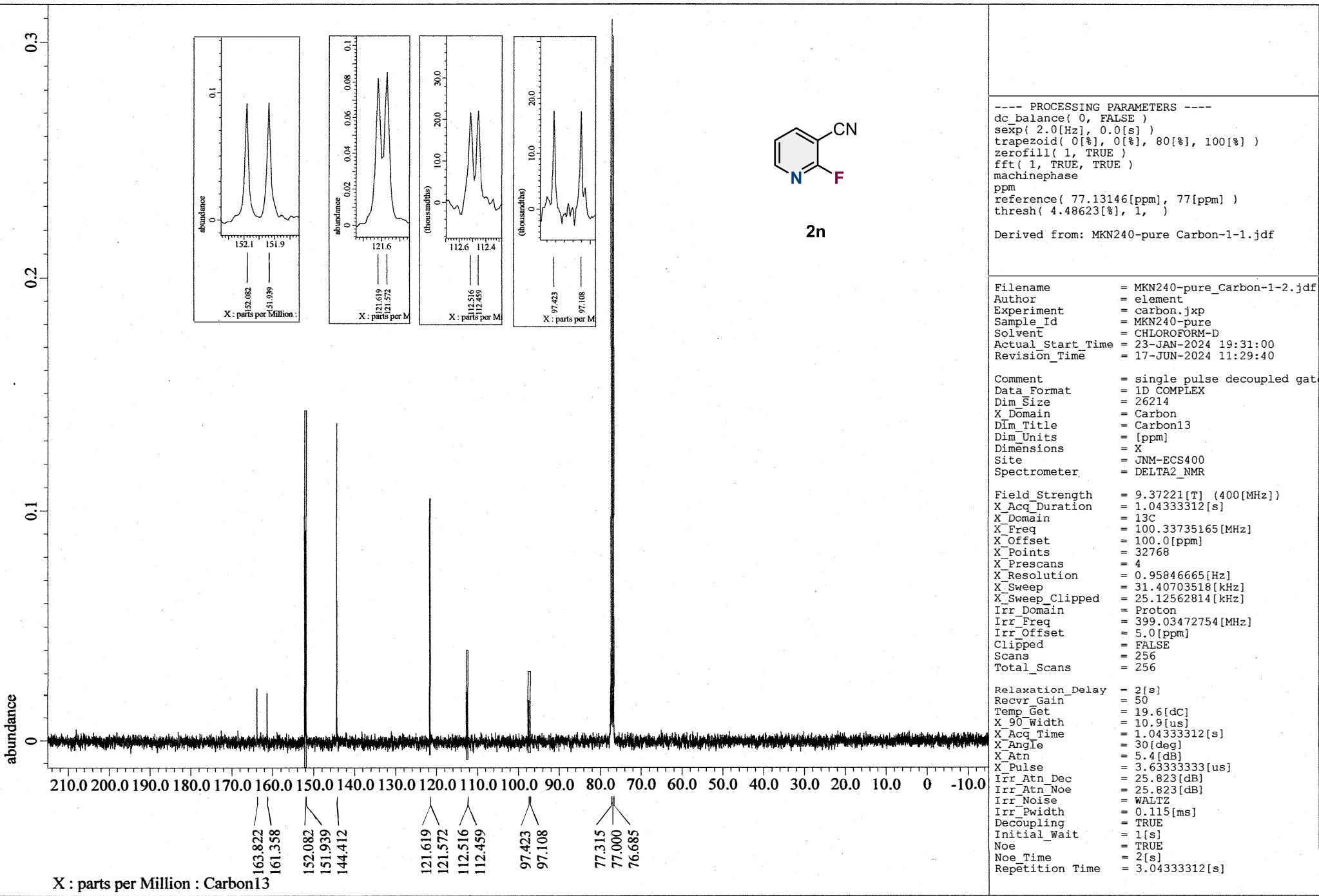
[General]
Experiment = MKN240-pure
Sample_Id = pure
Solvent = CHLOROFORM-D
Actual_Start_Time = 23-JAN-2024 17:22:40
Revision_Time = 17-JUN-2024 11:22:06

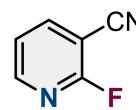
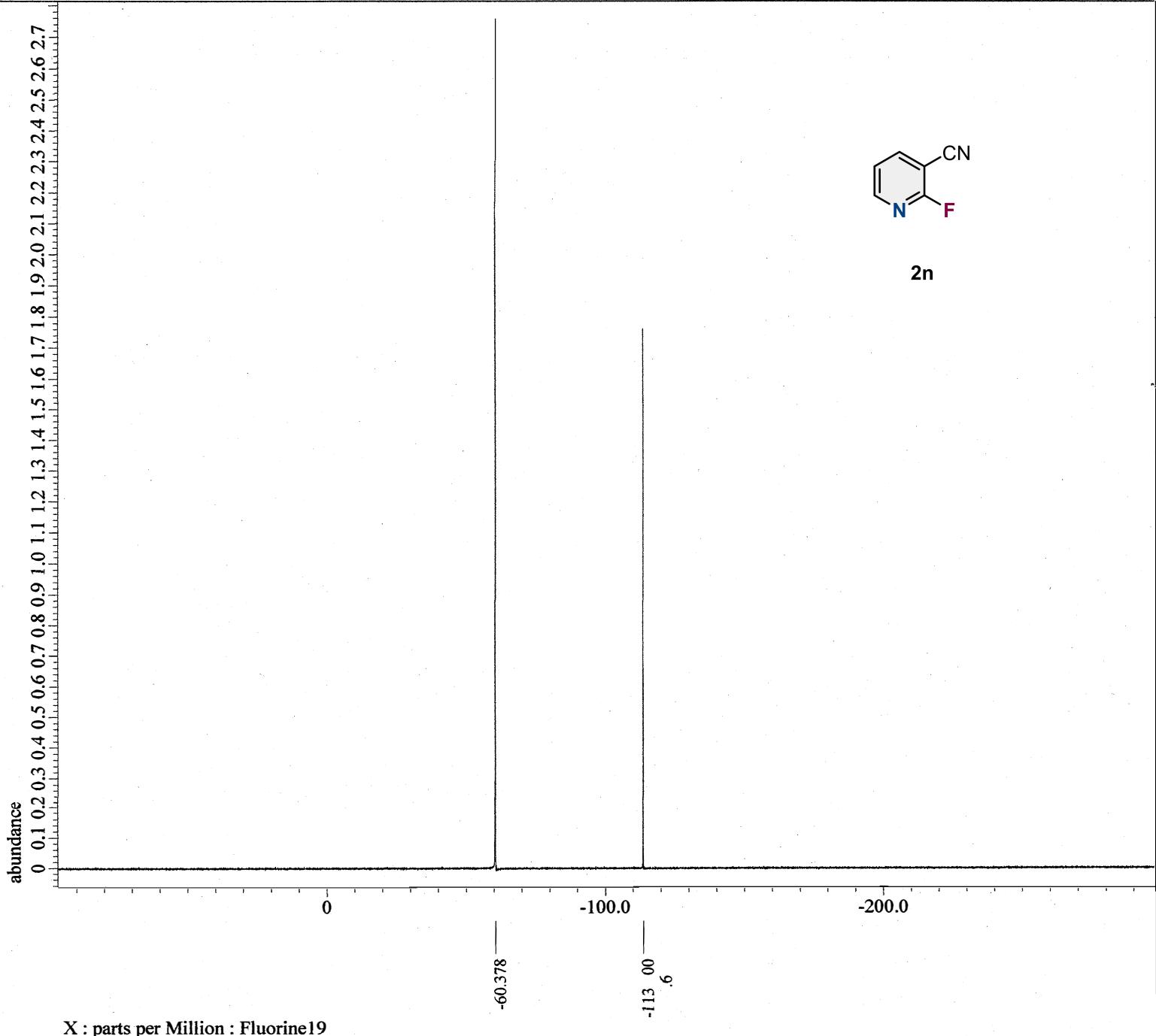
[Comment]
Comment = single_pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Proton
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

[Field_Strength]
Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 2.20725248[s]
X_Domain = Proton
X_Freq = 395.88430144[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 0.45305193[Hz]
X_Sweep = 7.42280285[kHz]
X_Sweep_Clipped = 5.93824228[kHz]
Irr_Domain = Proton
Irr_Freq = 395.88430144[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 395.88430144[MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

[Relaxation_Delay]
Relaxation_Delay = 5[s]
Recvr_Gain = 56
Temp_Get = 19[dC]
X_90_Width = 6.34[us]
X_Acq_Time = 2.20725248[s]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 3.17[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180,

```





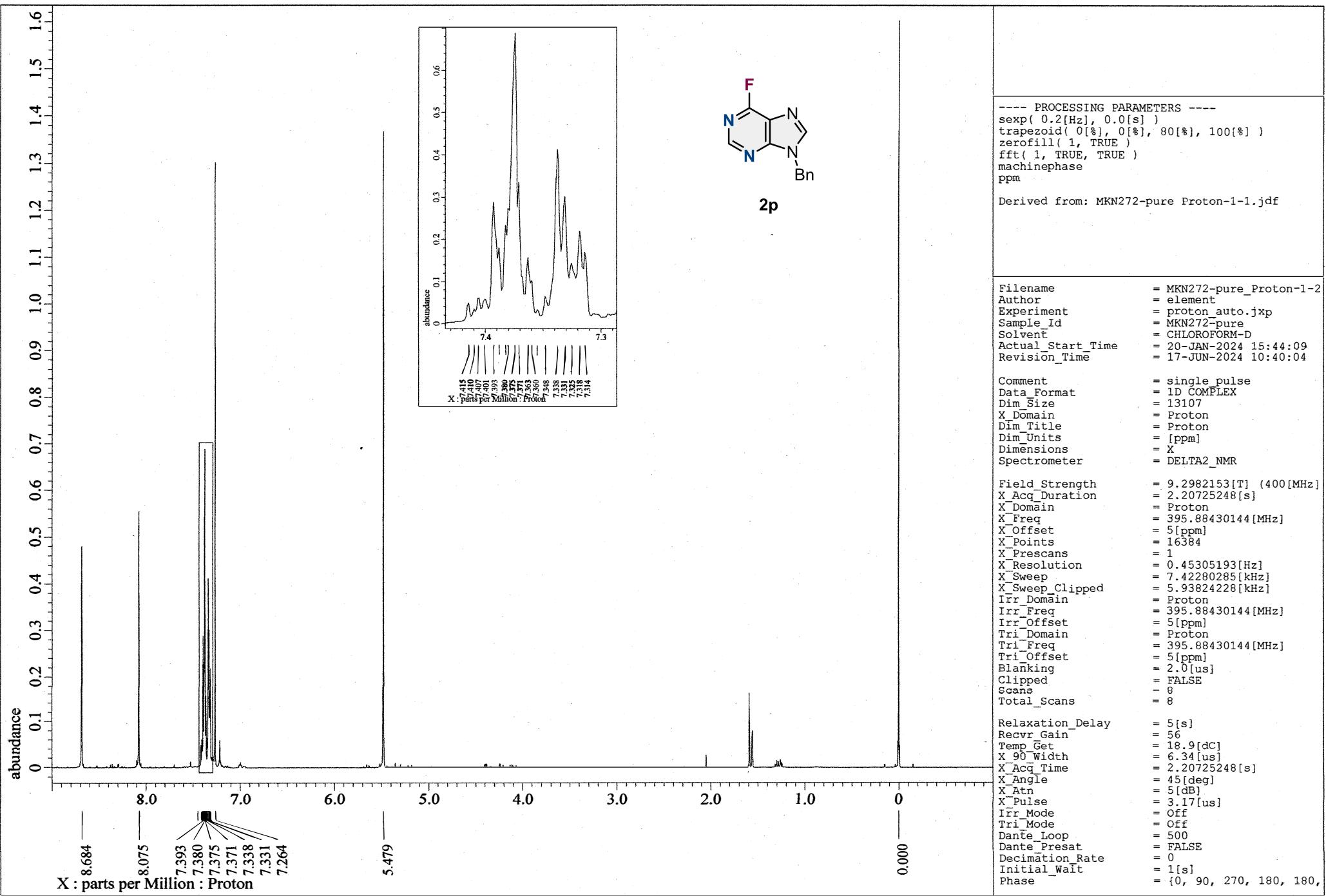
```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[ %],0[ %],80[ %],100[%] )
zerofill( 1 )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm

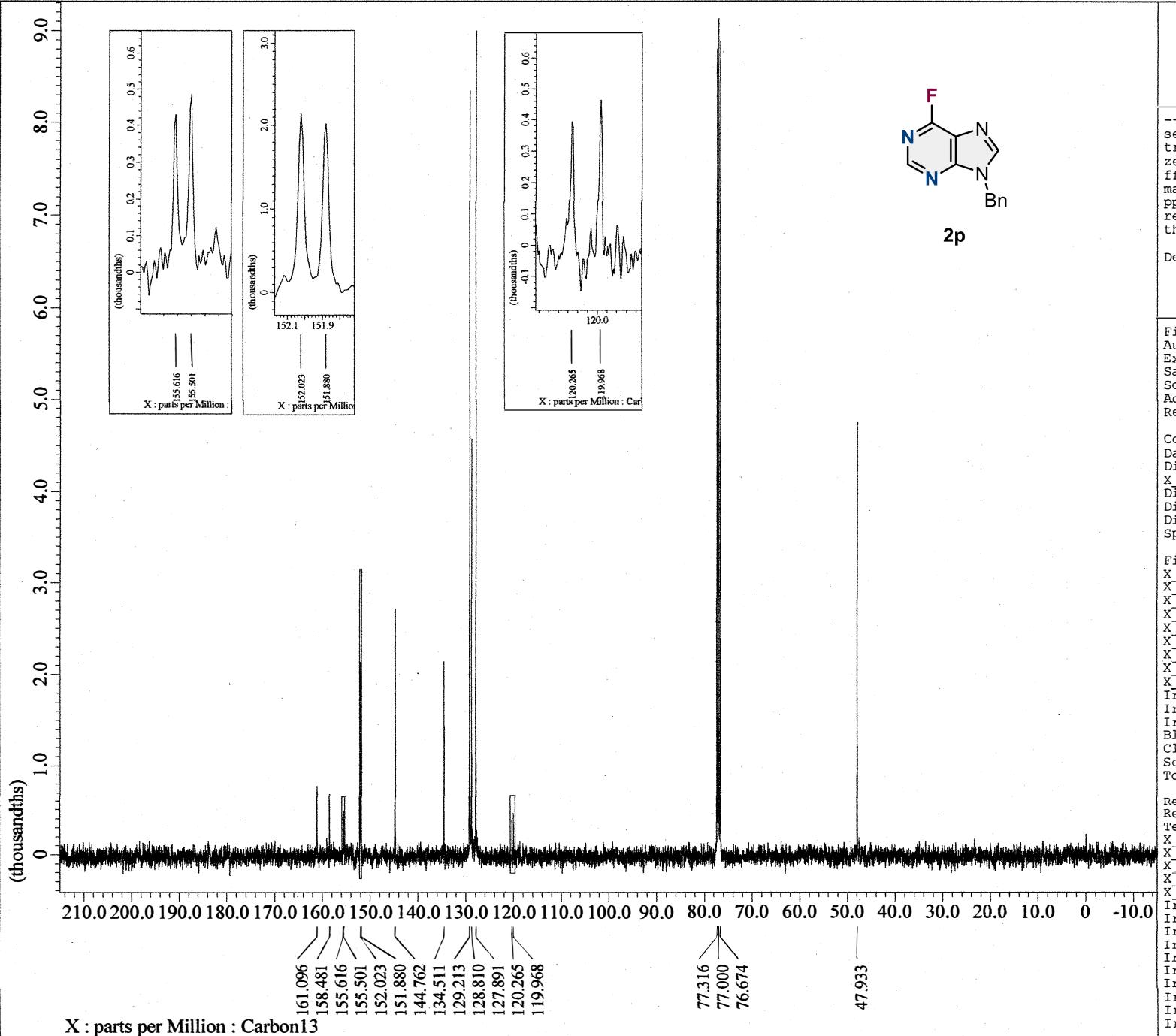
Filename = MKN240-pure-int_single
Author = element
Experiment = single_pulse.jxp
Sample_Id = MKN240-pure-int
Solvent = CHLOROFORM-D
Actual_Start_Time = 23-JAN-2024 17:33:35
Revision_Time = 24-JAN-2024 11:59:08

Comment = single_pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Fluorine19
Dim_Title = Fluorine19
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9. 298215[T] (400[MHz])
X_Acc_Duration = 89. 12896 [ms]
X_Domain = Fluorine19
X_Freq = 372.50336686[MHz]
X_Offset = -100[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 11.21969784[Hz]
X_Sweep = 183.82352941[ kHz]
X_Sweep_Clipped = 147.05 882353[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 372.50336686[ MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Fluorine19
Tri_Freq = 372.50336686[ MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 6
Total_Scans = 8

Relaxation_Delay = 5[ ]
Recvr_Gain = 56
Temp_Get = 18.7[dC]
X_90_Width = 8.03[us]
X_Acq_Time = 89.12896[ ms]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 4.0 15[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Loop = 500
Dante_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[ s]
Phase = {0, 90, 270, 180, 180},
```

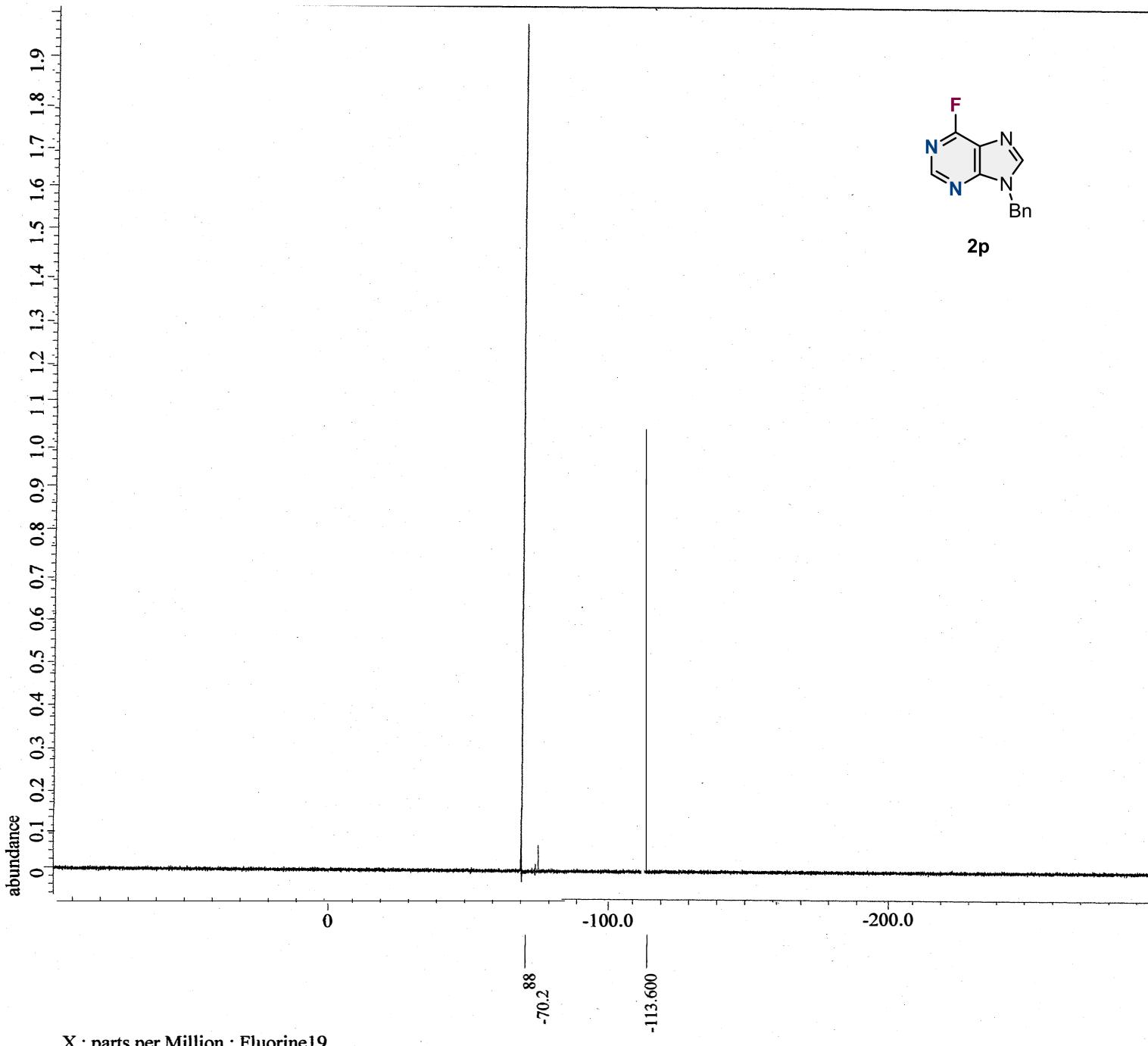




```
---- PROCESSING PARAMETERS ----
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
reference( 77.15094[ppm], 77[ppm] )
thresh( 2.87842[%], 1, )
```

Derived from: MKN272-pure Carbon-1-1.jdf

Filename	= MKN272-pure_Carbon-
Author	= element
Experiment	= carbon_auto.jxp
Sample_Id	= MKN272-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 20-JAN-2024 16:03:2
Revision_Time	= 17-JUN-2024 11:12:5
Comment	= single pulse decoup
Data_Format	= 1D COMPLEX
Dim_Size	= 26214
X_Domain	= Carbon13
Dim_Title	= Carbon13
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.2982153[T] (400[M
X_Acq_Duration	= 1.048576[s]
X_Domain	= Carbon13
X_Freq	= 99.54517646[MHz]
X_Offset	= 100[ppm]
X_Points	= 32768
X_Prescans	= 4
X_Resolution	= 0.95367432[Hz]
X_Sweep	= 31.25[kHz]
X_Sweep_Clipped	= 25[kHz]
Irr_Domain	= Proton
Irr_Freq	= 395.88430144[MHz]
Irr_Offset	= 5[ppm]
Blanking	= 5.0[us]
Clipped	= FALSE
Scans	= 256
Total_Scans	= 256
Relaxation_Delay	= 2[s]
Recv_Gain	= 5.0
Temp_Get	= 18.8[dC]
X_90_Width	= 11.5[us]
X_Acq_Time	= 1.048576[s]
X_Angle	= 30[deg]
X_Atn	= 9[dB]
X_Pulse	= 3.833333333[us]
Irr_Atn_Dec	= 30.172[dB]
Irr_Atn_Dec_Calc	= 30.172[dB]
Irr_Atn_Dec_Default_Calc	= 30.172[dB]
Irr_Atn_Noe	= 30.172[dB]
Irr_Dec_Bandwidth_Hz	= 4.7826087[kHz]
Irr_Dec_Bandwidth_Ppm	= 12.08082432[ppm]
Irr_Dec_Freq	= 395.88430144[MHz]
Irr_Dec_Merit_Factor	= 2.2
Irr_Decoupling	= TRUE
Irr_Noe	= TRUE



```
---- PROCESSING PARAMETERS ----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm

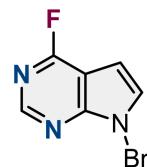
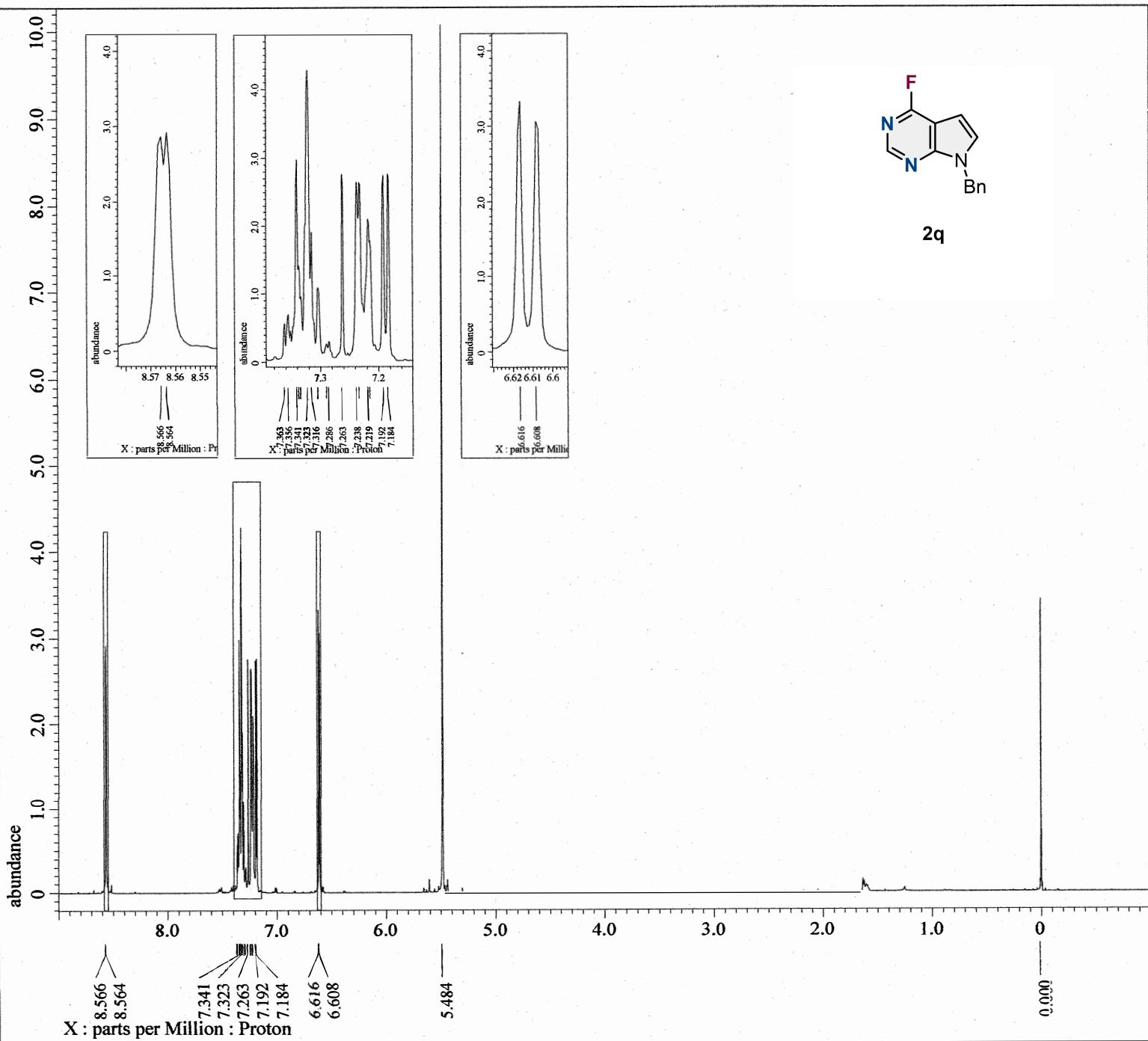
Derived from: MKN272-pure-int single pulse-1-1

Filename = MKN272-pure-int_single
Author = element
Experiment = single_pulse.jxp
Sample_Id = MKN272-pure-int
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-JAN-2024 15:58:51
Revision_Time = 17-JUN-2024 10:45:48

Comment = single pulse
Data_Format = 1D COMF EXL
Dim_Size = 13107
X_Domain = Fluorine19
Dim_Title = Fluorine19
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2 NMR

Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 89.12896[ms]
X_Domain = Fluorine19
X_Freq = 372.50336686[MHz]
X_Offset = -100[ppm]
X_Points = 16384
X_Presca_ns = 1
X_Resolution = 11.21969784[Hz]
X_Sweep = 183.82352941[kHz]
X_Sweep_Clipped = 147.05882353[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 372.50336686[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Fluorine19
Tri_Freq = 372.50336686[MHz]
Tri_Offset = 5[ppm]
Blinking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

RelaxationDelay = 5[s]
Recv_Gain = 56
Temp_Get = 18.3[dC]
X_90_Width = 8.03[us]
X_Acq_Time = 89.12896[ms]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 4.015[us]
Irr_Mode = Off
Tri_Mode = Off
Da_Rate_Presat = 500
Data_Rate_Presat = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180},
```



```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN274-pure Proton-1-1.jdf

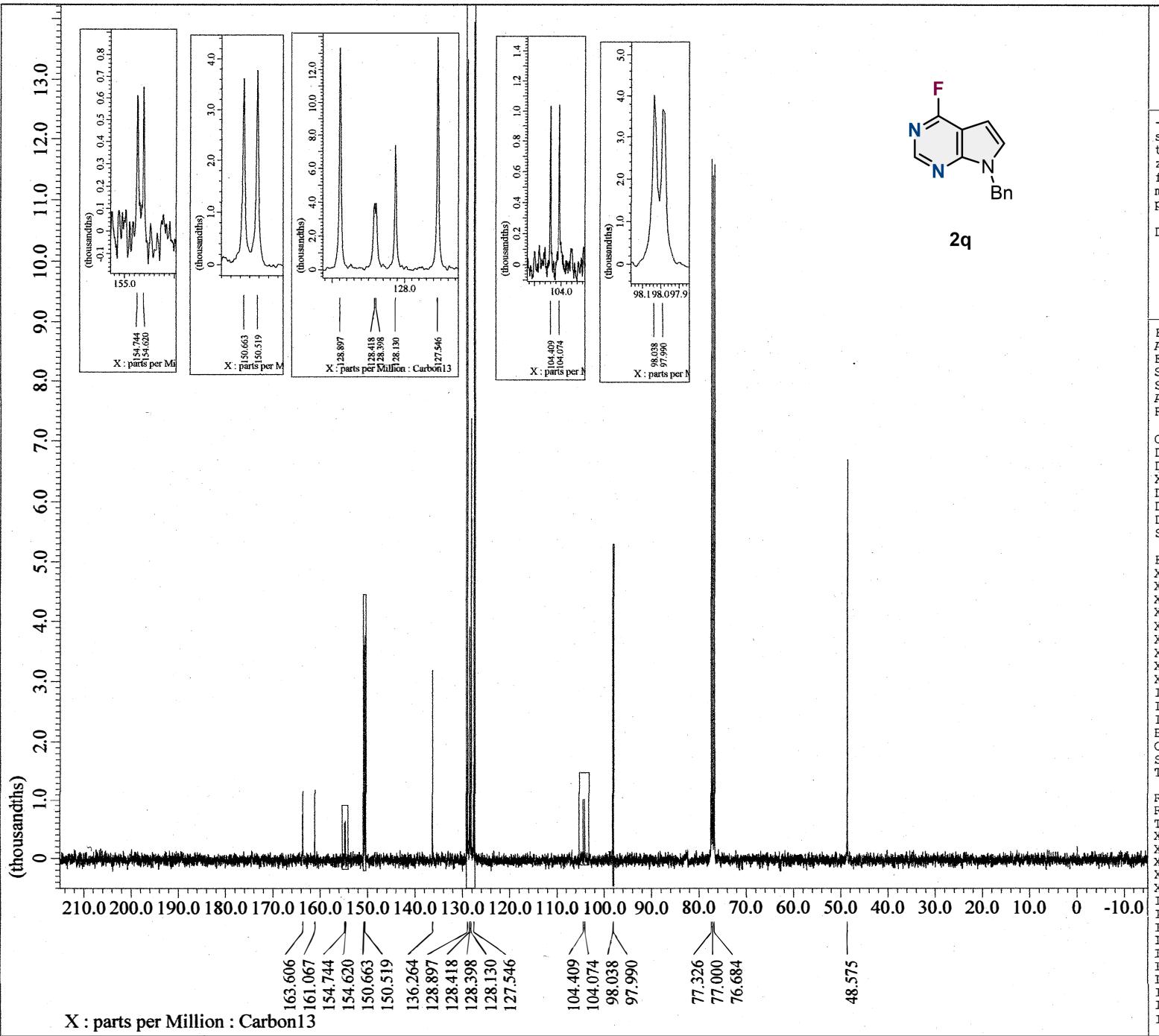
```

Filename = MKN274-pure_Proton-1-2.jdf
Author = element
Experiment = proton.jxp
Sample_Id = MKN274-pure
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-JAN-2024 14:22:21
Revision_Time = 17-JUN-2024 10:17:13

Comment = single_pulse
Data_Format = 1D COMPLEX
Dim_Size = 13107
X_Domain = Proton
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = DELTA2_NMR

Field_Strength = 9.4073814[T] (400[MHz])
X_Acq_Duration = 2.18103808[s]
X_Domain = 1H
X_Freq = 400.53219825[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 1
X_Resolution = 0.45849727[Hz]
X_Sweep = 7.51201923[kHz]
X_Sweep_Clipped = 6.00961538[kHz]
Irr_Domain = Proton
Irr_Freq = 400.53219825[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 400.53219825[MHz]
Tri_Offset = 5[ppm]
Clipped = FALSE
Scans = 8
Total_Scans = 9

Relaxation_Delay = 5[s]
Recvr_Gain = 42
Temp_Get = 19.3[dC]
X_90_Width = 6.7[us]
X_Acq_Time = 2.18103808[s]
X_Angle = 45[deg]
X_Atn = 0.8[dB]
X_Pulse = 3.35[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Presat = FALSE
Initial_Wait = 1[s]
Repetition_Time = 7.18103808[s]
```



----- PROCESSING PARAMETERS -----  
 sexp( 2.0[Hz], 0.0[s] )  
 trapezoid( 0[%], 0[%], 80[%], 100[%] )  
 zerofill( 1, TRUE )  
 fft( 1, TRUE, TRUE )  
 machinephase  
 ppm

Derived from: MKN274-pure Carbon-1-1.jdf

Filename = MKN274-pure\_Carbon-  
 Author = element  
 Experiment = carbon\_auto.jxp  
 Sample\_Id = MKN274-pure  
 Solvent = CHLOROFORM-D  
 Actual\_Start\_Time = 20-JAN-2024 15:16:4  
 Revision\_Time = 17-JUN-2024 10:28:0  
 Comment = single pulse decoupl  
 Data\_Format = 1D COMPLEX  
 Dim\_Size = 26214  
 X\_Domain = Carbon13  
 Dim\_Title = Carbon13  
 Dim\_Units = [ppm]  
 Dimensions = X  
 Spectrometer = DELTA2\_NMR  
 Field\_Strength = 9.2982153[T] (400[M]  
 X\_Acq\_Duration = 1.048576[s]  
 X\_Domain = Carbon13  
 X\_Freq = 99.54517646[MHz]  
 X\_Offset = 100[ppm]  
 X\_Points = 32768  
 X\_Prescans = 4  
 X\_Resolution = 0.95367432[Hz]  
 X\_Sweep = 31.25[kHz]  
 X\_Sweep\_Clipped = 25[kHz]  
 Irr\_Domain = Proton  
 Irr\_Freq = 395.88430144[MHz]  
 Irr\_Offset = 5[ppm]  
 Blanking = 5.0[us]  
 Clipped = FALSE  
 Scans = 256  
 Total\_Scans = 256  
 Relaxation\_Delay = 2[s]  
 Recvr\_Gain = 50  
 Temp\_Get = 18.5[dC]  
 X\_90\_Width = 11.5[us]  
 X\_Acq\_Time = 1.048576[s]  
 X\_Angle = 30[deg]  
 X\_Atn = 9[dB]  
 X\_Pulse = 3.833333333[us]  
 Irr\_Atn\_Dec = 30.172[dB]  
 Irr\_Atn\_Dec\_Calc = 30.172[dB]  
 Irr\_Atn\_Dec\_Default\_Calc = 30.172[dB]  
 Irr\_Atn\_Noe = 30.172[dB]  
 Irr\_Dec\_Bandwidth\_Hz = 4.7826087[kHz]  
 Irr\_Dec\_Bandwidth\_Ppm = 12.08082432[ppm]  
 Irr\_Dec\_Freq = 395.88430144[MHz]  
 Irr\_Dec\_Merit\_Factor = 2.2  
 Irr\_Decoupling = TRUE  
 Irr\_Noee = TRUE



```
----- PROCESSING PARAMETERS -----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
blip( 16, 64 ,30 )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

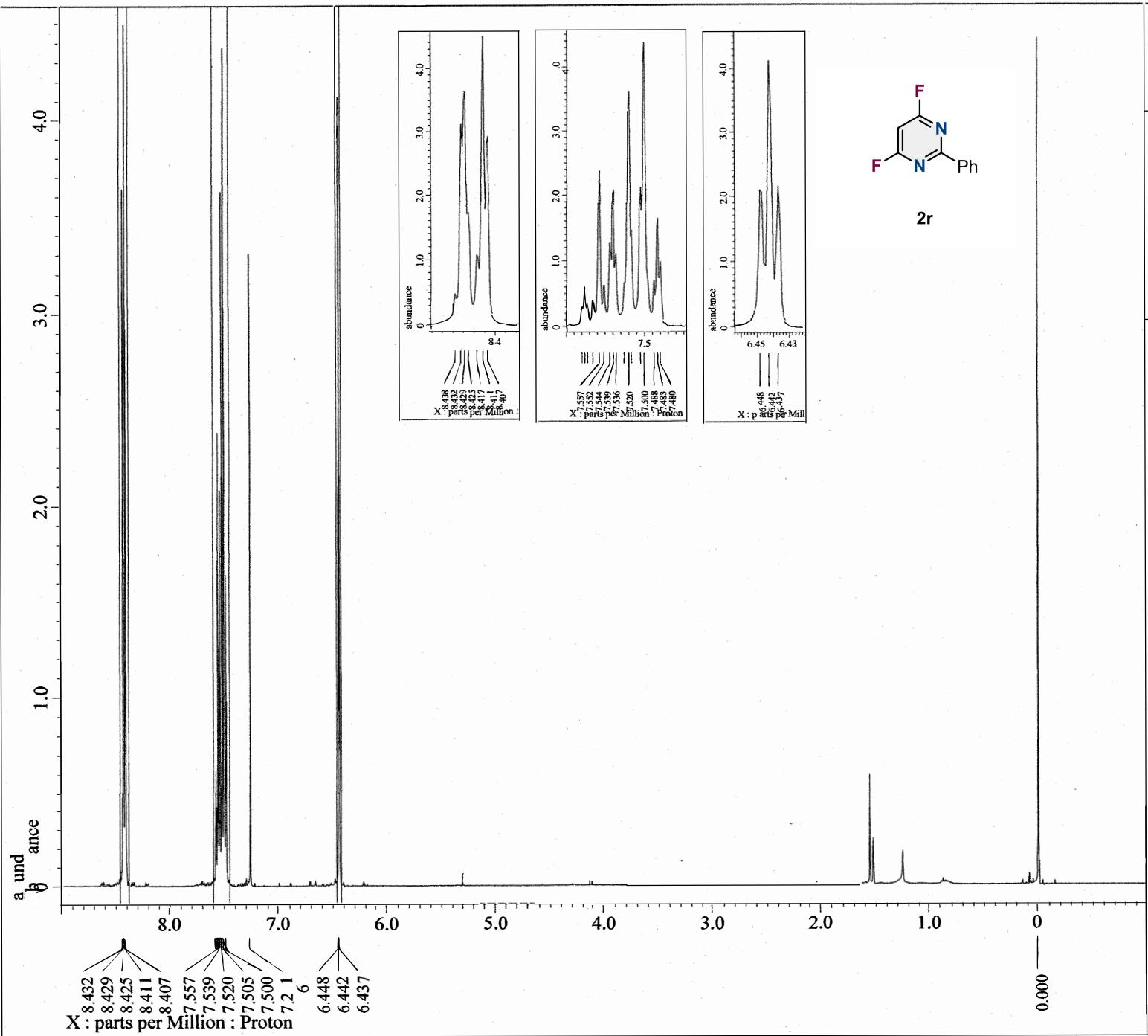
```

Filename = MKN274_pure -mt_single
Author = eleme nt
Experiment = s inge_pulse.jxp
Sample_Id = MKN274-pure-int
Solv_at = CHLOROFORM-D
Actual_Start_Time = 20 -JAN-202415:12:22
Revision_Time = 17 -JUN -202410:23:33

Comment = single_pulse
Data_Format = 1D COMP EX
Dim_Size = 13107
X_Domain = Fluorine19
Dim_Title = Fluorine 19
Dim_Units = [ppm]
Dimensions = X
Specrometer = DELTA2_N_MR

Field_Strength = 9.2982153[T] (400[MHz])
X_Acq_Duration = 89.12896[ms]
X_Domain = Fluorine19
X_Freq = 372.50336686[MHz]
X_Offset = -100[ppm]
X_Points = 16384
X_Pre_scans = 1
X_Resolution = 11.21969784[Hz]
X_Sweep = 183.82352941[k Hz]
X_Sweep_Clipped = 147.05882353[kHz]
Irr_Domain = Fluorine19
Irr_Freq = 372.50336686[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Fluorine19
Tri_Freq = 372.50336686[ MHz]
Tri_Offset = 5[ppm]
Blanking = 2.0[us]
Clipped = FALSE
Scans = 8
Total_Scans = 8

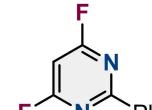
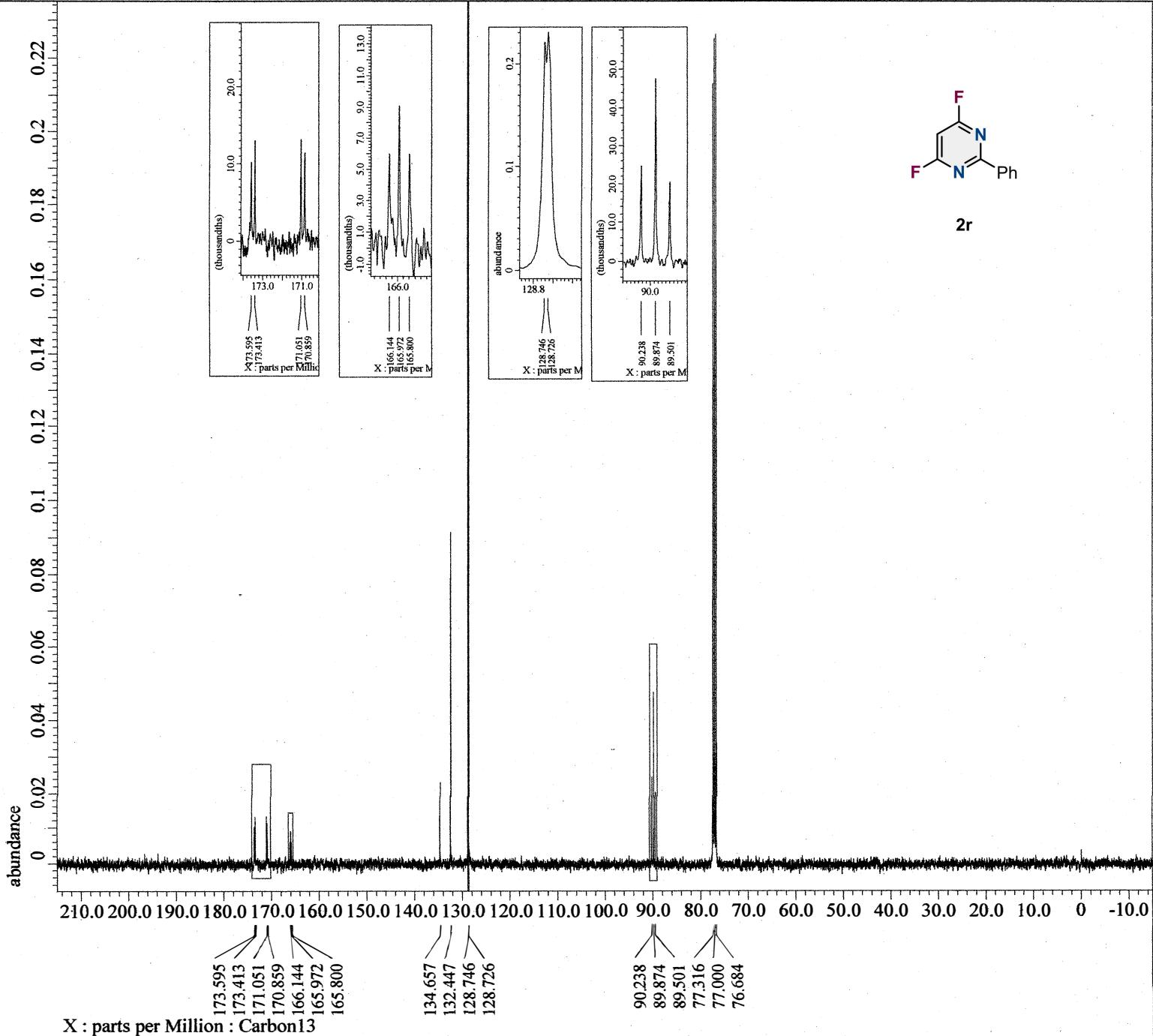
Relaxation_Delay = 5[s]
Recv_Gain = 56
Temp_Get = 18.8[dC]
X_90_Width = 8.03[us]
X_Acq_Time = 89.12896[ms]
X_Angle = 45[deg]
X_Atn = 5[dB]
X_Pulse = 4.015[us]
Irr_Mode = Off
Tri_Mode = Off
Dante_Lo_B = 500
Dante_Preset = FALSE
Decimation_Rate = 0
Initial_Wait = 1[s]
Phase = {0, 90, 270, 180, 180,
```



```
--PROCESSING PARAMETERS--
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN270-pure Proton-1-1.jdf

Filename	= MKN270-pure_Proton-1-2.jdf
Author	= element
Experiment	= proton.jxp
Sample_Id	= MKN270 -pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 22 -JUN-2024 11:13:58
Revision_Time	= 17-JUN-2024 09:47:51
Comment	= single_pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Domain	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_N MR
Field_Strength	= 9.4073814[T] (400[MHz])
X_Acq_Duration	= 2.18103808[s]
X_Domain	= 1H
X_Freq	= 400.53219825[MHz]
X_Offset	= 5[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45849727[Hz]
X_Sweep	= 7.51201923[kHz]
X_Sweep_Clipped	= 6.00961538[kHz]
Irr_Domain	= Proton
Irr_Freq	= 400.53219825[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= Proton
Tri_Freq	= 400.53219825[MHz]
Tri_Offset	= 5[ppm]
Clipped	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recv_Gain	= 44
Temp_Get	= 19.6[dC]
X_90_Width	= 6.7[us]
X_Acq_Time	= 2.18103808[s]
X_Angle	= 45[deg]
X_Atn	= 0.8[db]
X_Pulse	= 3.35[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Presat	= FALSE
Initial_Wait	= 1[s]
Repetition_Time	= 7.18103808[s]

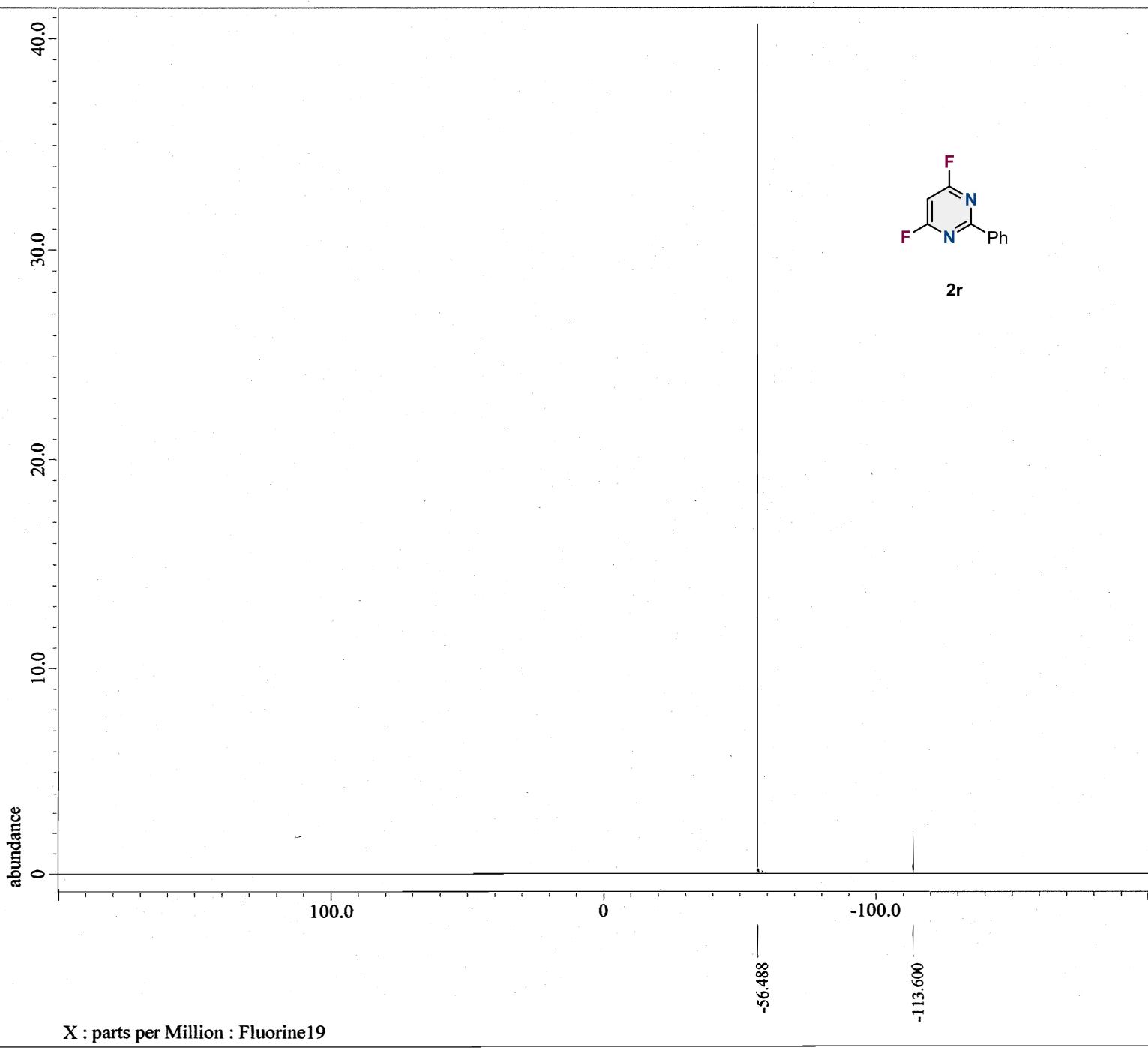


2r

```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN270-pure Carbon-1-1.jdf

Filename	= MKN270-pure_Carbon-1-2.jdf
Author	= element
Experiment	= carbon.jxp
Sample_Id	= MKN270-di-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 22-JAN-2024 11:39:51
Revision_Time	= 17-JUN-2024 09:54:20
Comment	= single pulse decoupled gat
Data_Format	= 1D COMPLEX
Dim_Size	= 26214
X_Domain	= Carbon
Dim_Title	= Carbon13
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.4073814[T] (400[MHz])
X_Acq_Duration	= 1.03809024[s]
X_Domain	= 13C
X_Freq	= 100.71389092[MHz]
X_Offset	= 100[ppm]
X_Points	= 3 2 6
X_Prescans	= 4
X_Resolution	= 0.96330739[Hz]
X_Sweep	= 31.56565657[kHz]
X_Sweep_Clipped	= 25.25252525[kHz]
Irr_Domain	= Proton
Irr_Freq	= 400.53219825[MHz]
Irr_Offset	= 5[ppm]
Clipped	= FALSE
Scans	= 512
Total_Scans	= 512
Relaxation_Delay	= 2[s]
Recvr_Gain	= 50
Temp_Get	= 19.6[dC]
X_90_Width	= 12.68[us]
X_Acq_Time	= 1.03809024[s]
X_Angle	= 30[deg]
X_Atn	= 4[dB]
X_Pulse	= 4.22666667[us]
Irr_Atn_Dec	= 26.45[dB]
Irr_Atn_Noe	= 26.45[dB]
Irr_Noise	= WALTZ
Irr_Pwidth	= 0.115[ms]
Decoupling	= TRUE
Initial_Wait	= 1[s]
Noe	= TRUE
Noe_Time	= 2[s]
Repetition_Time	= 3.03809024[s]

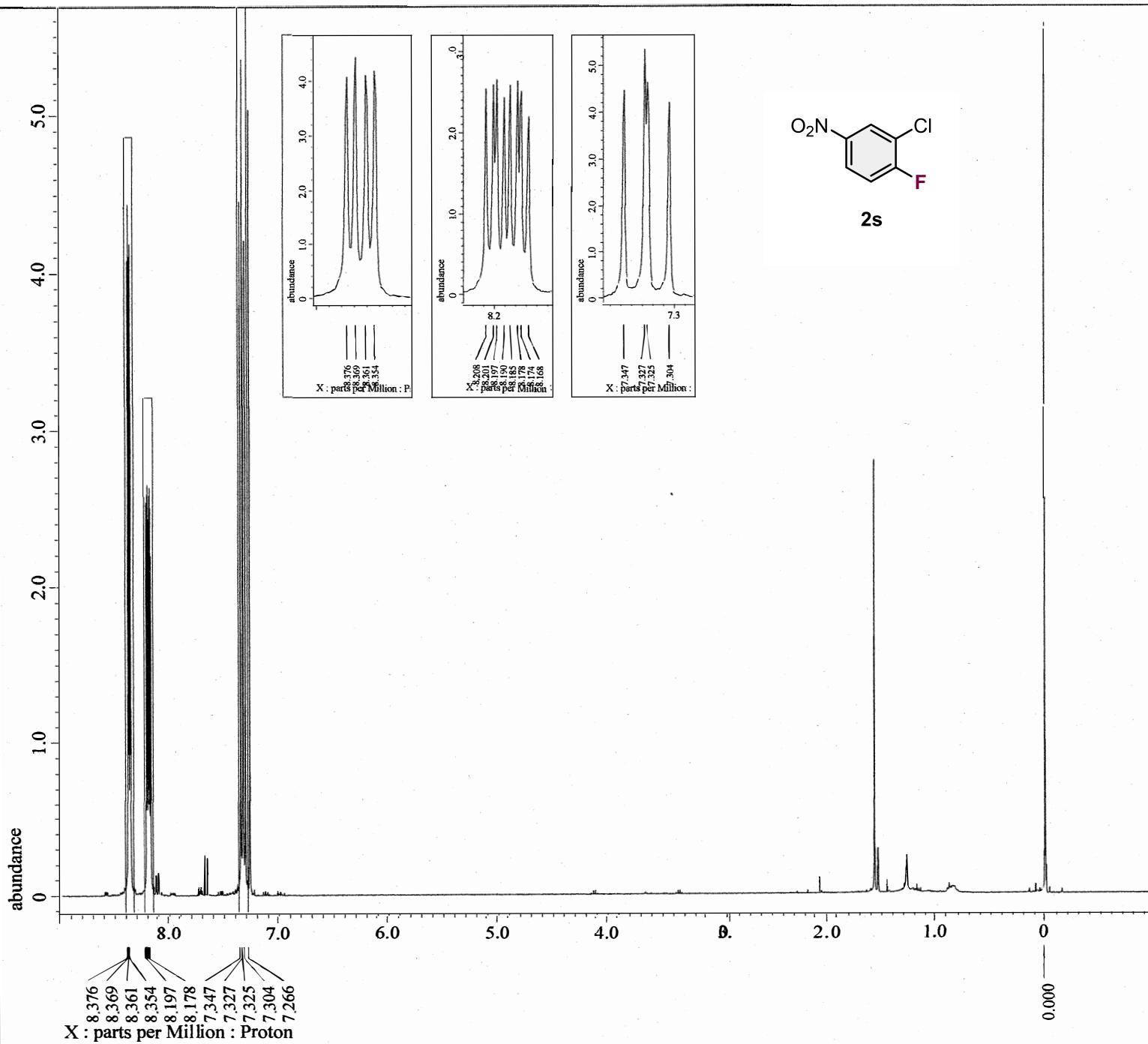


```

---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[ Hz ], 0.0[ s ] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machinephase
ppm

Filename      = MKN270-pur_eint_single_pul
Author        = element
Experiment   = single_pulse.jxp
Sample_Id    = MKN270-di-pure-int
Solvent       = CHLOROFORM-D
Actual_Start_Time = 22-JAN-2024 11:25:33
Revision_Time = 24-JAN-2024 12:15:35
Comment       = single_pulse
Data_Format  = 1D COMP_IEX
Dim_Size     = 13107
X_Domain    = Fluori
Dim_Title   = Fluorine19
Dim_Units   = [ppm]
Dimensions  = X
Sp_ectrmeter = DELTA2_NMR
Field_Strength = 9.4073814[T] (400[MHz])
X_Acq_Duration = 86.50752[ms]
X_Domain     = 19F
X_Freq        = 376.87675879[MHz]
X_Offset      = 0[ppm]
X_Po_ints    = 16384
X_Prescans   = 1
X_Resolution = 11.55968868[Hz]
X_Sweep       = 189.39393939[kHz]
X_Sweep_Clipped = 151.51515152[kHz]
Irr_Domain   = Fluorine19
Irr_Freq     = 376.87675879[MHz]
Irr_Offset   = 5[ppm]
Tri_Domain   = Fluorine19
Tri_F_sq     = 376.87675879[MHz]
Tri_Offset   = 5[ppm]
Clipped      = FALSE
Scans         = 8
Total_Scans  = 8
Relaxation_Delay = 5[s]
Recvr_Gain   = 42
Temp_Get     = 19.2[dC]
X_90_Width  = 7.59[us]
X_Acq_Time  = 86.50752[ms]
X_Angle      = 45[deg]
X_Atn        = 3[dB]
X_Pulse      = 3.795[us]
Irr_Mode     = Off
Tri_Mode     = Off
Dante_Presat = FALSE
Initial_Wait = 1[s]
Repetition_Time = 5.08650752[s]

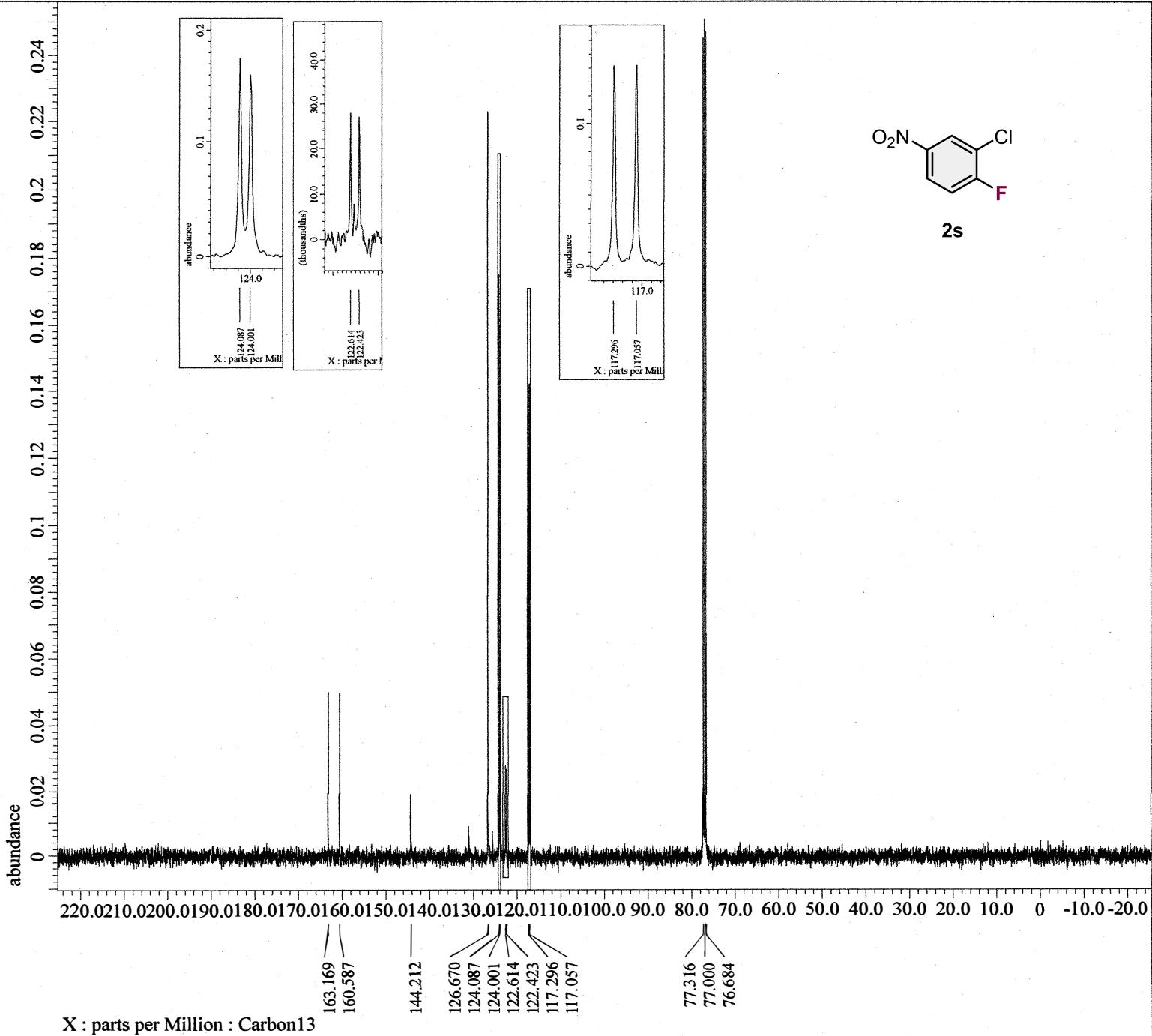
```



```
---- PROCESSING PARAMETERS ----
dc_balance(0, FALSE)
sexp(0.2[Hz], 0.0[s])
trapezoid(0[%], 0[%], 80[%], 100[%])
zerofill(1, TRUE)
fft(1, TRUE, TRUE)
mainlinephase
ppm
```

Derived from: MKN283-pure\_Proton-1-1.jdf

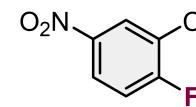
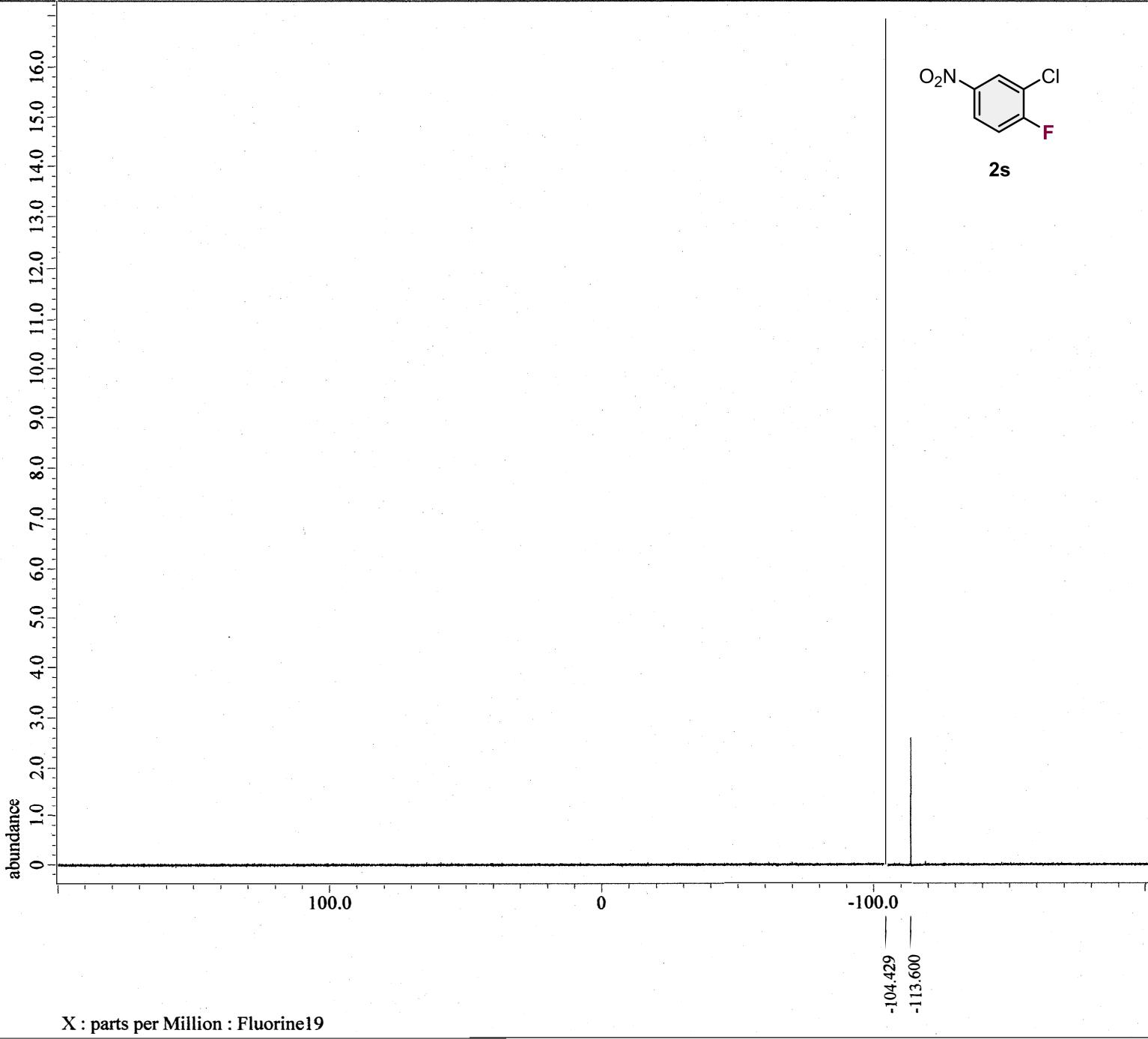
Filename	= MKN283-pure_Proton-1-2.jdf
Author	= element
Experiment	= proton.jxp
Sample_Id	= MKN283-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 25-JAN-2024 13:49:52
Revision_Time	= 17-JUN-2024 09:15:08
Comment	= single_pulse
Data_Format	= 1D COMP_EX
Dim_Size	= 13107
X_Domain	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.4073814[T] (400[MHz])
X_Acq_Duration	= 2.18103808[s]
X_Domain	= 1H
X_Freq	= 400.53219825[MHz]
X_Offset	= 5[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45849727[Hz]
X_Sweep	= 7.51201923[kHz]
X_Sweep_Clipped	= 6.00961538[kHz]
Irr_Domain	= Proton
Irr_Freq	= 400.53219825[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= Proton
Tri_Freq	= 400.53219825[MHz]
Tri_Offset	= 5[ppm]
Clipped	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recvr_Gain	= 48
Temp_Get	= 19[dC]
X_90_Width	= 6.7[us]
X_Acq_Time	= 2.18103808[s]
X_Angle	= 45[deg]
X_Atn	= 0.8[dB]
X_Pulse	= 3.35[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Resat	= FALSE
Initial_Wait	= 1[s]
Report_Echo_Time	= 7.18103808[s]



```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sep( 2.0[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN283-pure Carbon-1-1.jdf

Filename	= MKN283-pure_Carbon-1-2.jdf
Author	= element
Experiment	= carbon.jxp
Sample Id	= MKN283-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 25-JAN-2024 14:19:32
Revision_Time	= 17-JUN-2024 09:34:36
Comment	= single pulse decoupled gat
Data_Format	= 1D COMPLEX
Dim_Size	= 26214
X_Domain	= Carbon
Dim_Title	= Carbon13
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.4073814[T] (400[MHz])
X_Acq_Duration	= 1.03809024[s]
X_Domain	= 13C
X_Freq	= 100.71389092[MHz]
X_Offset	= 100[ppm]
X_Points	= 32768
X_Prescans	= 4
X_Resolution	= 0.96330739[Hz]
X_Sweep	= 31.56565657[kHz]
X_Sweep_Clipped	= 25.25252525[kHz]
Irr_Domain	= Proton
Irr_Freq	= 400.53219825[MHz]
Irr_Offset	= 5[ppm]
Clipped	= FALSE
Scans	= 257
Total_Scans	= 257
Relaxation_Delay	= 2[s]
Recv_Gain	= 50
Temp_Get	= 19.5[dC]
X_90_Width	= 12.68[us]
X_Acq_Time	= 1.03809024[s]
X_Angle	= 30[deg]
X_Atn	= 4[dB]
X_Pulse	= 4.22666667[us]
Irr_Atn_Dec	= 26.45[dB]
Irr_Atn_Noe	= 26.45[dB]
Irr_Noise	= WALTZ
Irr_Pwidth	= 0.115[ms]
Decoupling	= TRUE
Initial_Wait	= 1[s]
Noe	= TRUE
Noe_Time	= 2[s]
Repetition_Time	= 3.03809024[s]



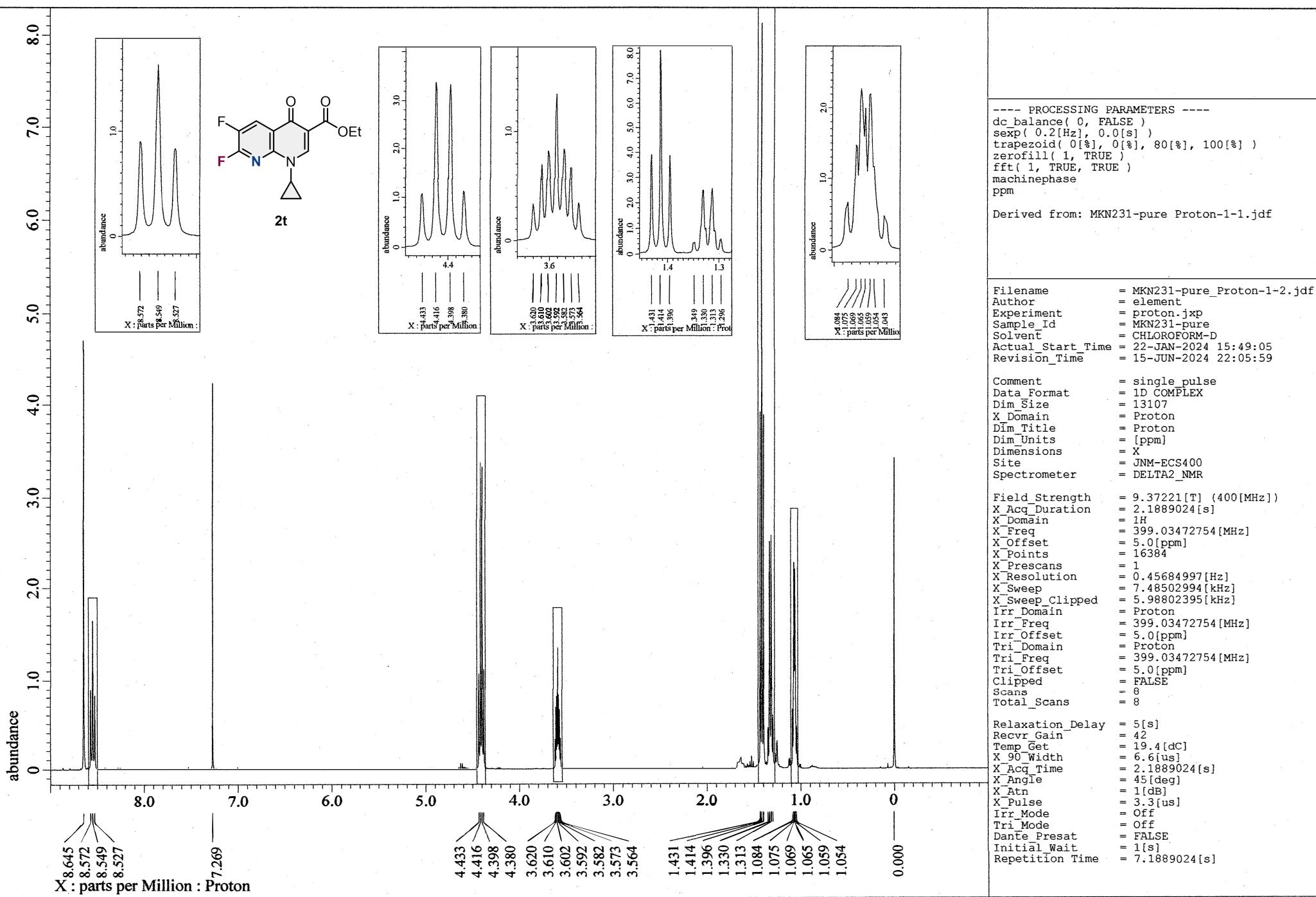
**2s**

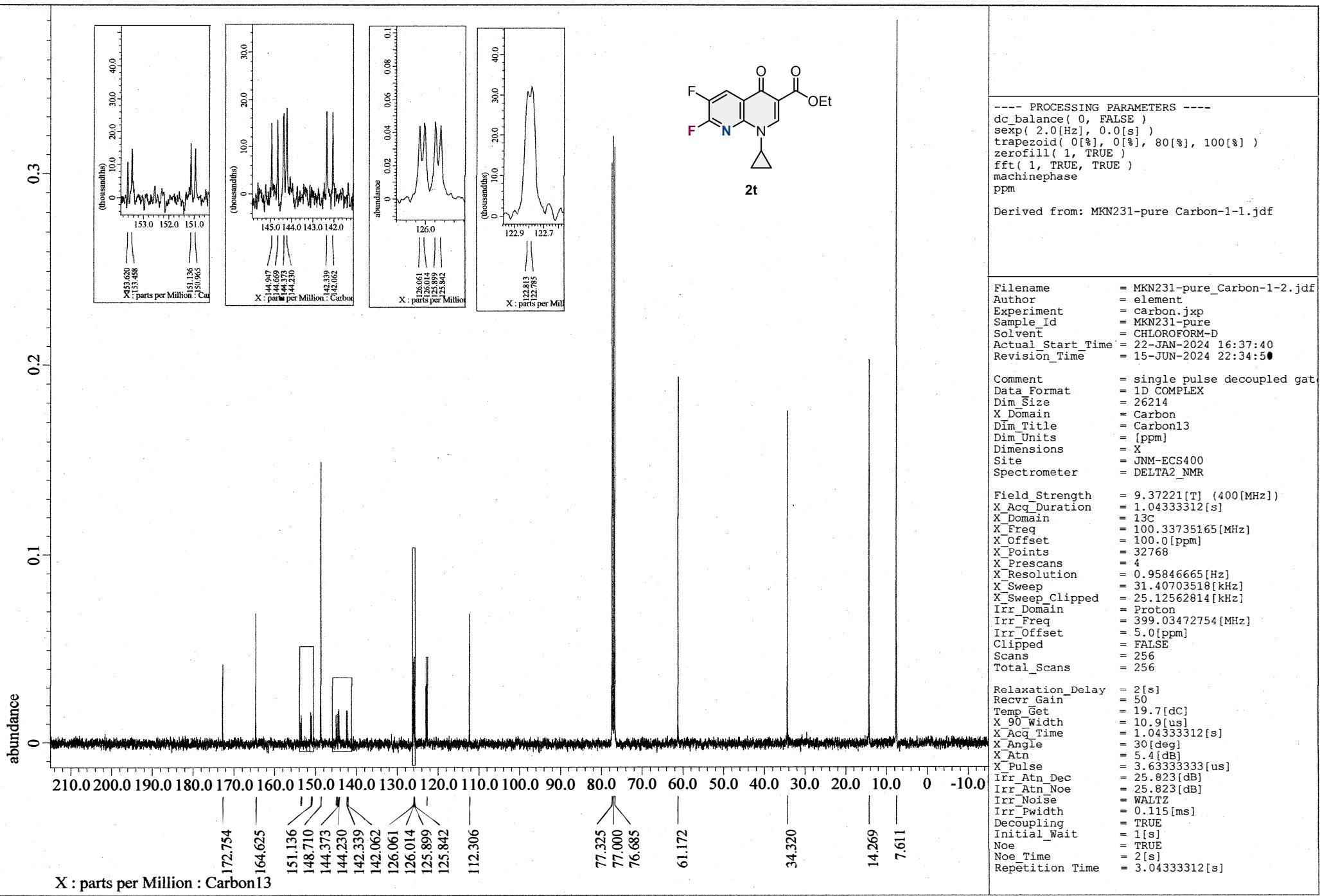
```
---- P ROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
s_ew( 0.2[Hz], 0.0[s] )
t_ragezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1 )
blip( 16, 64, 30 )
fft( 1, TRUE, TRUE )
machin phas e
ppm
```

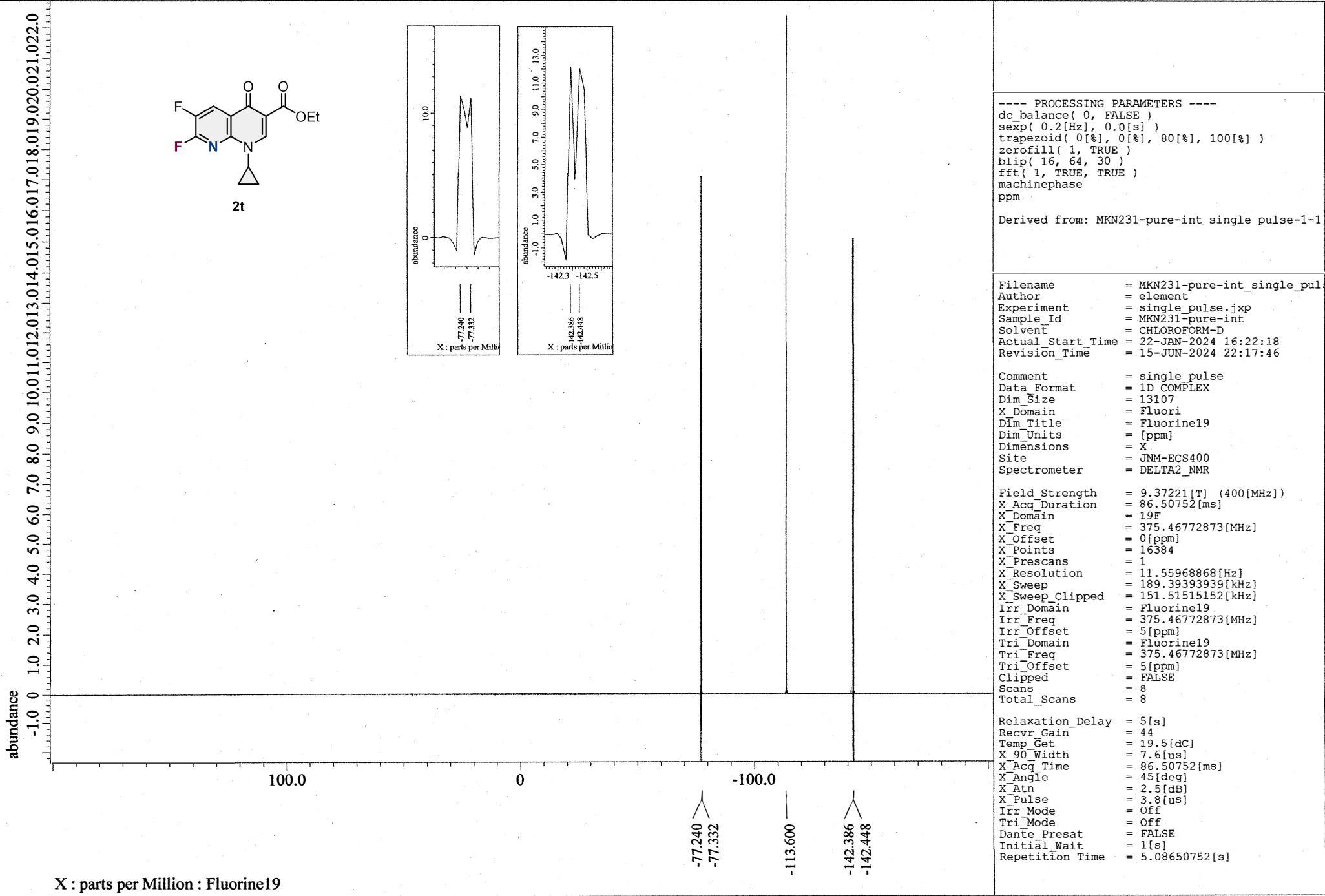
```
Filename      = MKN283-pure-int_single_pul
Aut_har       = element
E_per_ment   = single_pulse.jxp
Sample_Id    = MKN283-pure-int
Solvent       = C HL OROFORM-D
Actual_Start_Tim e= 25-JAN-2024 13:28:28
Revision_Tim e = 17-JUN-2024 09:39:52
```

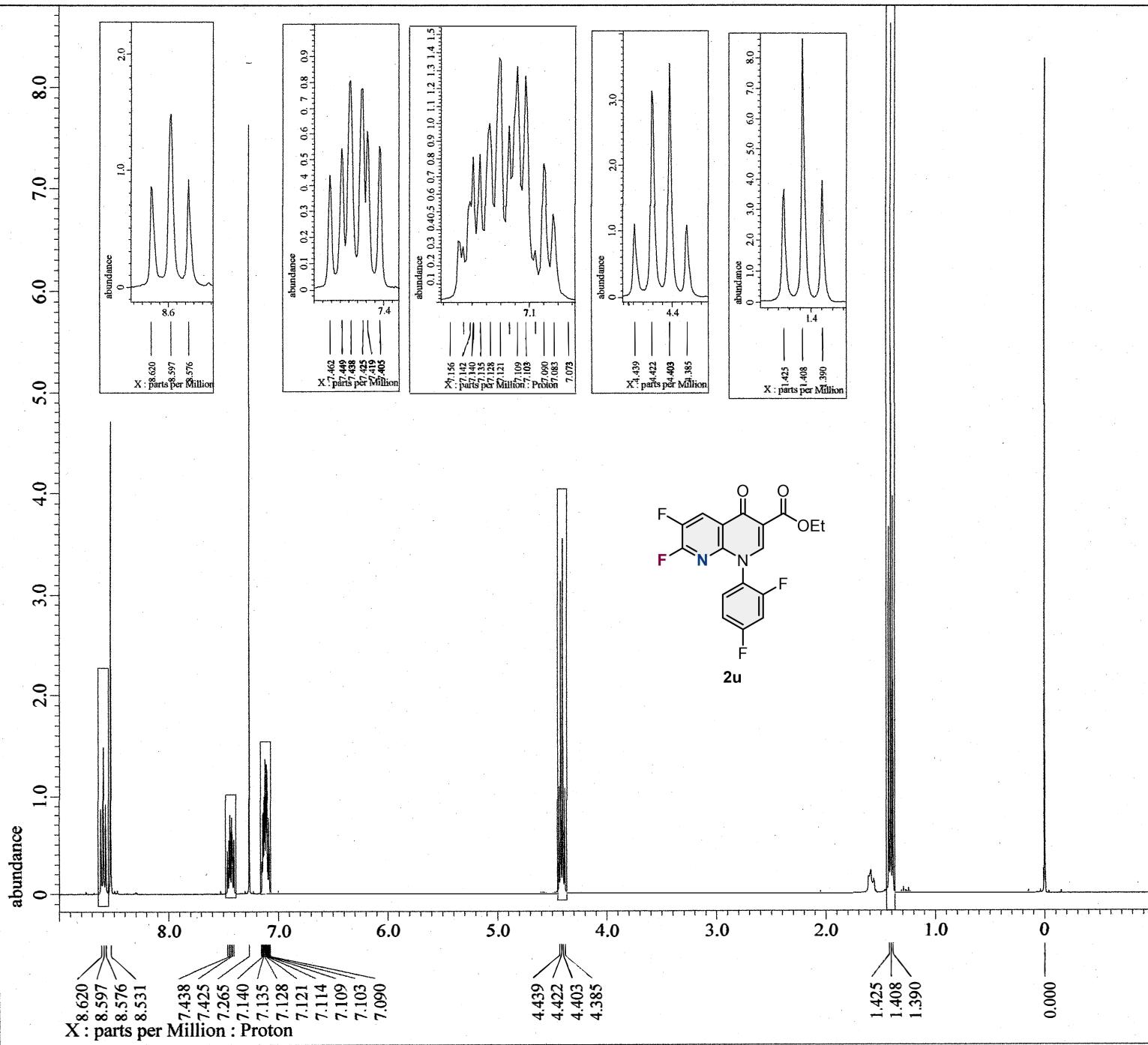
```
Comment       = single_pulse
Data_Format  = 1D COMPLEX
Dim_Size     = 13107
X_Domain    = Fluori
Dim_Title   = Fluorin e19
Dim_Units   = [ppm]
Dimension   = X
Spectrom_e er = DELTA2_NM_R
Field_St rag th = 9.4073814[T] (400[M Hz])
X_Acq_Durati o = 86.50752[ms]
X_Domain    = 19F
X_Freq       = 376.87675879[MHz]
X_Offset    = 0[ppm]
X_Points    = 16384
X_Prescans  = 1
X_Resolution = 11.55968868[ Hz ]
X_Sweep      = 189.39393939[k Hz]
X_Sweep_Clipped = 151.51515152[k Hz]
Irr_Domain  = Fluorine19
Irr_Freq     = 376.87675879[M Hz]
Irr_Offset  = 5[ppm]
Tri_Domain  = Fluorine19
Tri_Freq     = 376.87675879[M Hz]
Tri_Offset  = 5[ppm]
Clipped     = FALSE
Scans        = 8
Total_Scans  = 8
```

```
Relaxation_Delay = 5[s]
Recvr_Gain      = 46
Temp_Get         = 19[dc]
X_90_Width      = 7.59[us]
X_Acq_Time     = 86.50752[ms]
X_Angle         = 45[ deg ]
X_Atn          = 3[dB]
X_Pulse         = 3.795[us]
Irr_Mode        = Off
Tri_Mode        = Off
Dante_Presat   = FALSE
Initial_Wait   = 1[s]
Repetit_IonTim e = 5.08650752[s]
```







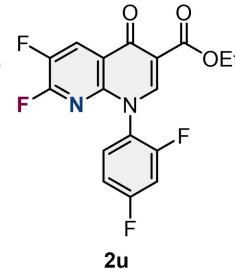
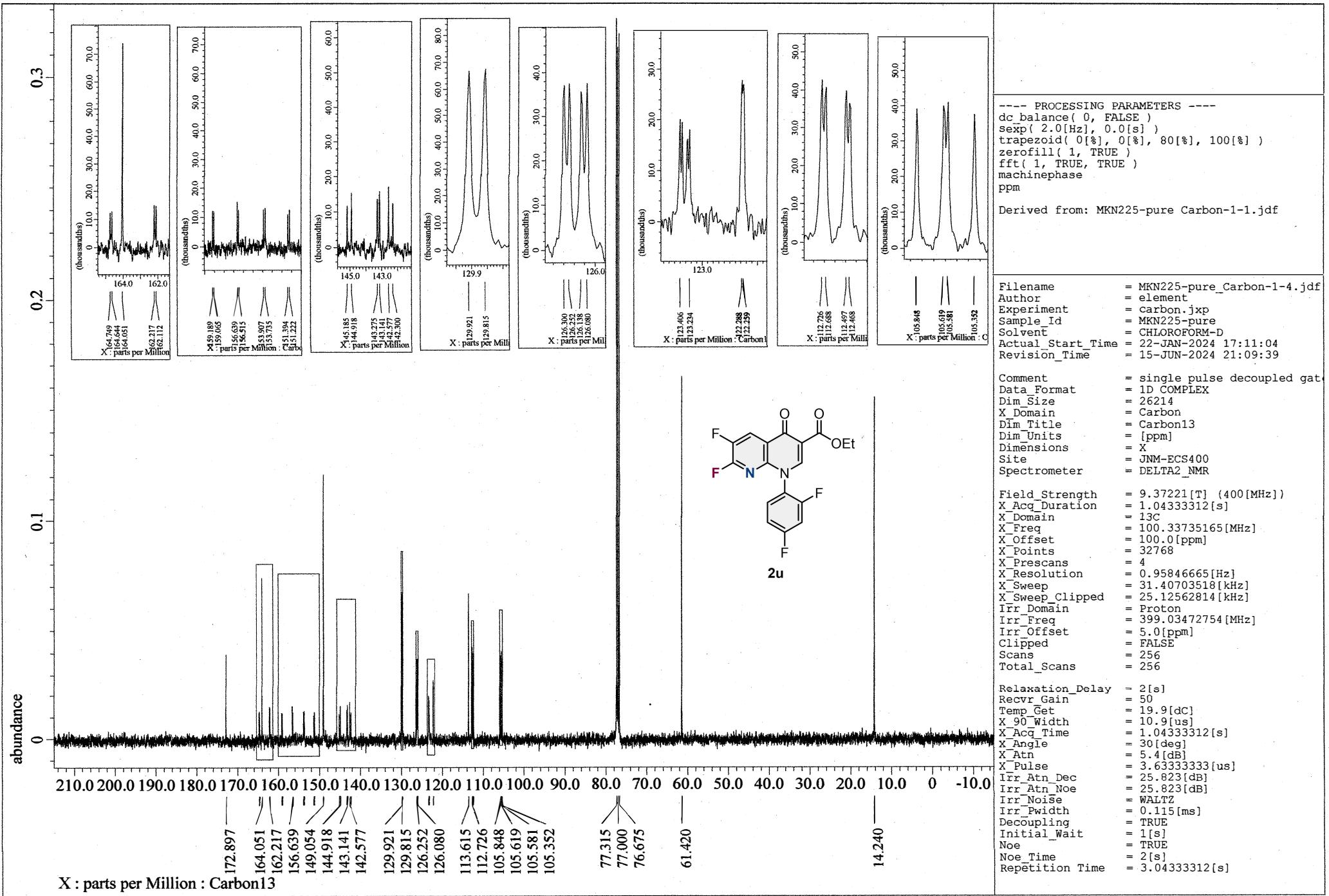


```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

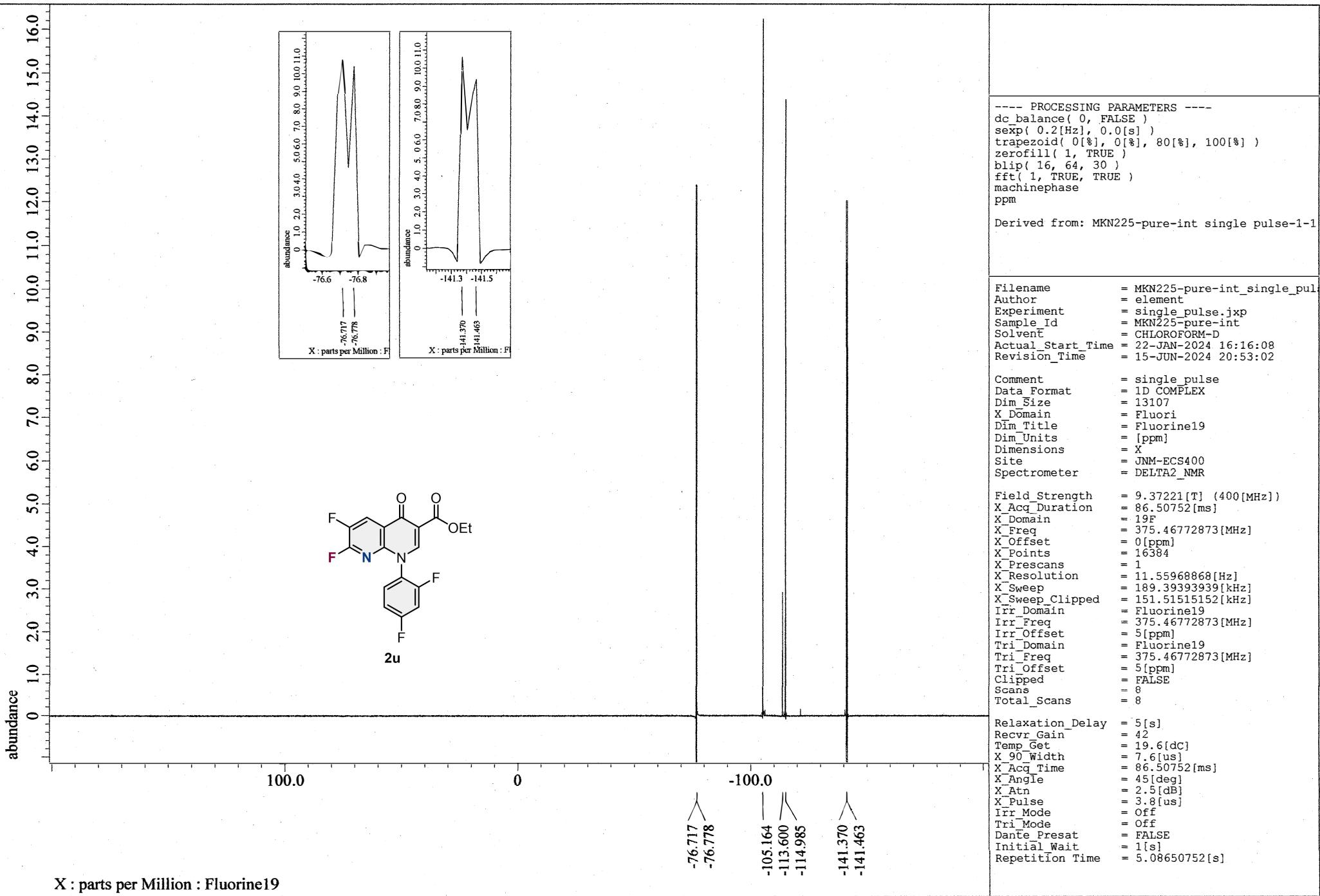
Derived from: MKN225-pure Proton-1-1.jdf

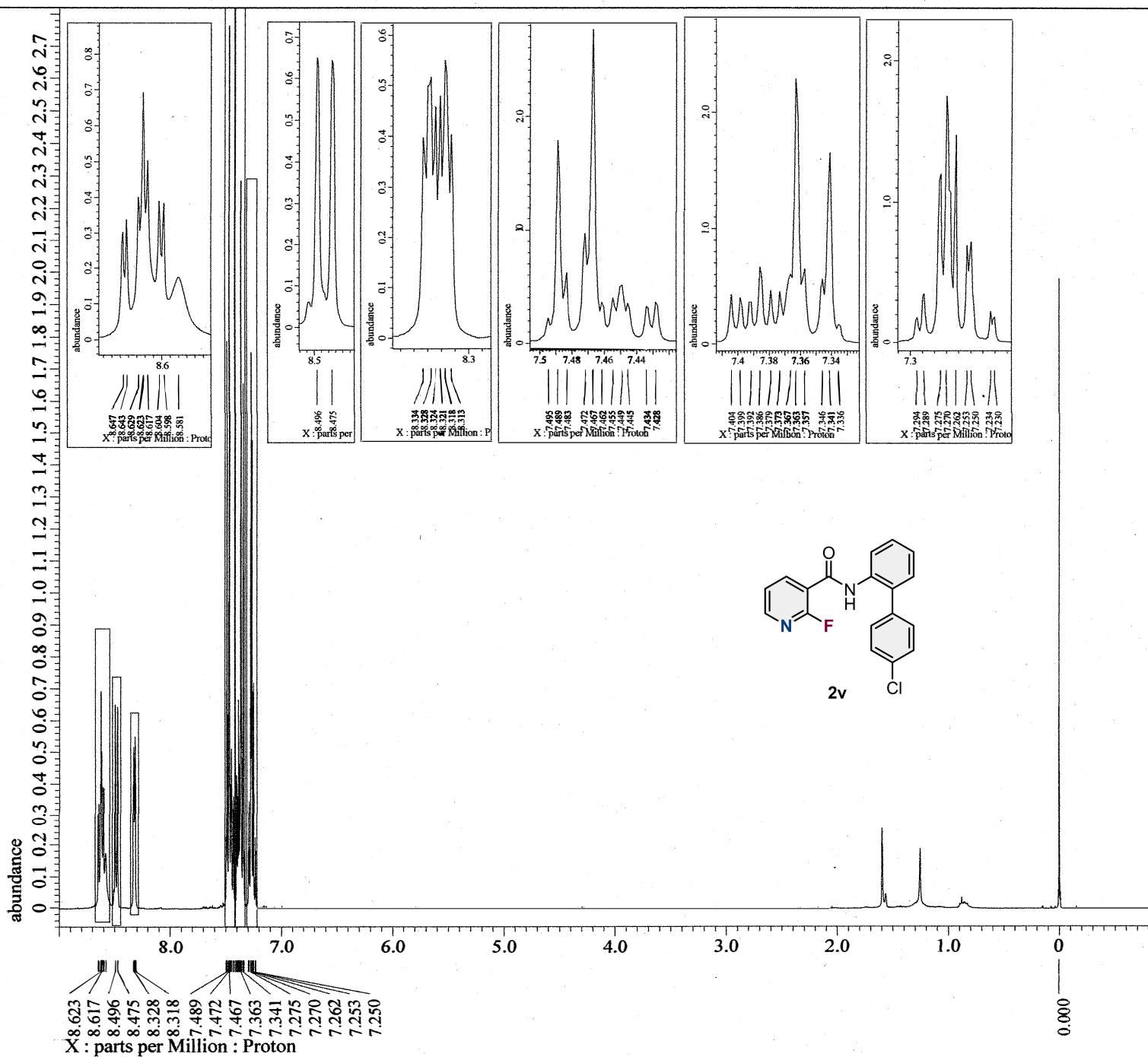
Filename	= MKN225-pure_Proton-1-2.jdf
Author	= element
Experiment	= proton.jxp
Sample_Id	= MKN225-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 22-JAN-2024 16:29:35
Revision_Time	= 15-JUN-2024 19:46:44

Comment	= single_pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Domain	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Site	= JNM-ECS400
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.37221[T] (400[MHz])
X_Acq_Duration	= 2.1889024[s]
X_Domain	= 1H
X_Freq	= 399.03472754[MHz]
X_Offset	= 5.0[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45684997[Hz]
X_Sweep	= 7.48502994[kHz]
X_Sweep_Clipped	= 5.98802395[kHz]
Irr_Domain	= Proton
Irr_Freq	= 399.03472754[MHz]
Irr_Offset	= 5.0[ppm]
Tri_Domain	= Proton
Tri_Freq	= 399.03472754[MHz]
Tri_Offset	= 5.0[ppm]
Clipped	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recv_Gain	= 46
Temp_Get	= 19.3[dC]
X_90_Width	= 6.6[us]
X_Acq_Time	= 2.1889024[s]
X_Angle	= 45[deg]
X_Atn	= 1[dB]
X_Pulse	= 3.3[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Presat	= FALSE
Initial_Wait	= 1[s]
Repetition_Time	= 7.1889024[s]



2u

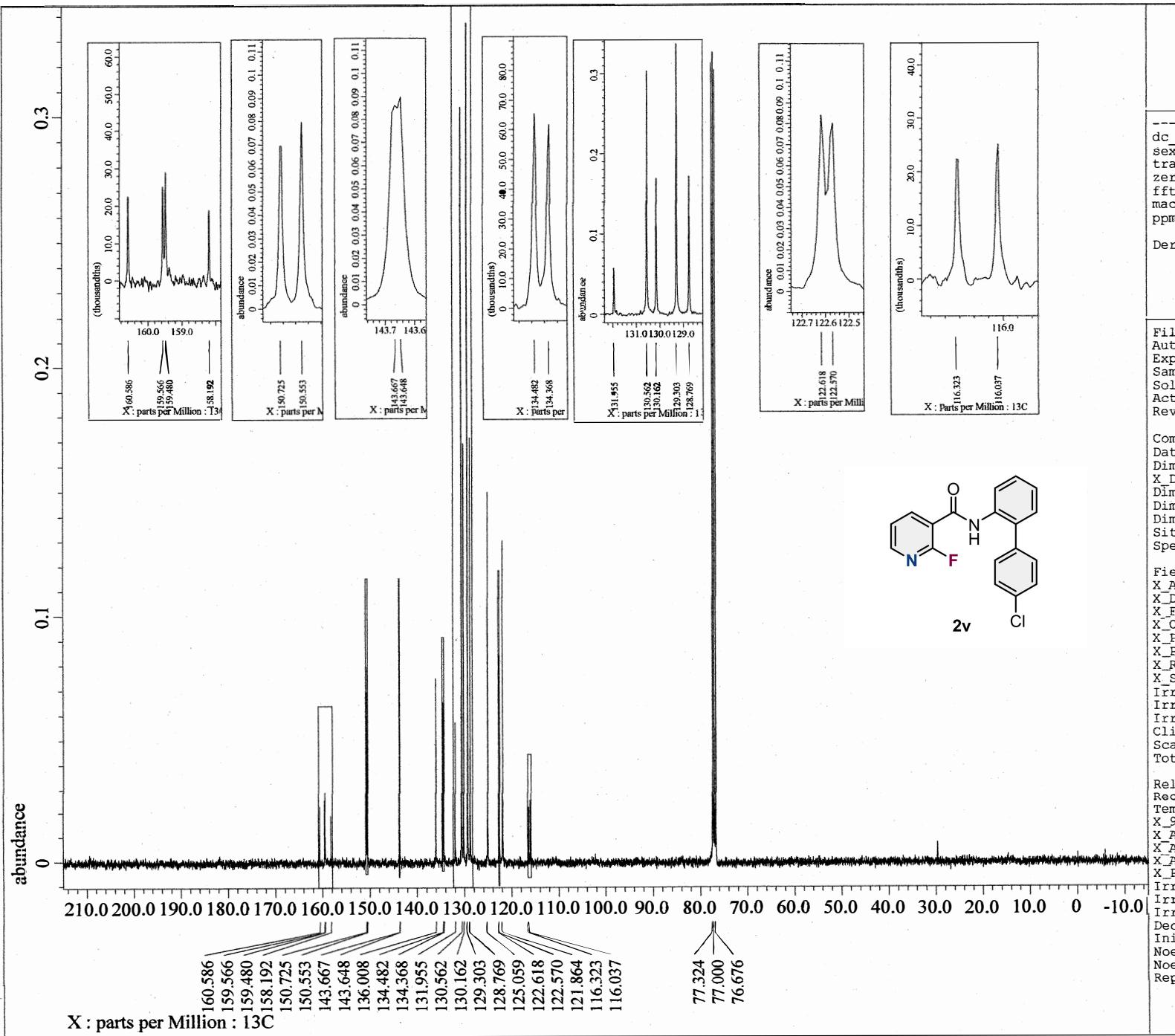




```
----- PROCESSING PARAMETERS -----
sexp( 0.2[Hz], 0.0[s] )
trapezoid( 0[%], 0[%], 80[$], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN219-pure Proton-1-1.jdf

Filename	= MKN219-pure_Proton-1-1.jdf
Author	= element
Experiment	= proton auto.jxp
Sample_Id	= MKN219-pure
Solvent	= CHLOROFORM-D
Actual_Start_Time	= 23-JAN-2024 12:32:19
Revision_Time	= 15-JUN-2024 18:20:55
Comment	= single pulse
Data_Format	= 1D COMPLEX
Dim_Size	= 13107
X_Domain	= Proton
Dim_Title	= Proton
Dim_Units	= [ppm]
Dimensions	= X
Spectrometer	= DELTA2_NMR
Field_Strength	= 9.29821 53[T] (400[MHz])
X_Acq_Duration	= 2.20725248[s]
X_Domain	= Proton
X_Freq	= 395.88430144[MHz]
X_Offset	= 5[ppm]
X_Points	= 16384
X_Prescans	= 1
X_Resolution	= 0.45305193[Hz]
X_Sweep	= 7.42280285[kHz]
X_Sweep_Clipped	= 5.93824228[kHz]
Irr_Domain	= Proton
Irr_Freq	= 395.88430144[MHz]
Irr_Offset	= 5[ppm]
Tri_Domain	= Proton
Tri_Freq	= 395.88430144[MHz]
Tri_Offset	= 5[ppm]
Blanking	= 2.0[us]
Clipped	= FALSE
Scans	= 8
Total_Scans	= 8
Relaxation_Delay	= 5[s]
Recv_Gain	= 56
Temp_Get	= 18.8[dc]
X_90_Width	= 6.34[us]
X_Acq_Time	= 2.20725248[s]
X_Angle	= 45[deg]
X_Atn	= 5[dB]
X_Pulse	= 3.17[us]
Irr_Mode	= Off
Tri_Mode	= Off
Dante_Loop	= 500
Dante_Presat	= FALSE
Decimation_Rate	= 0
Initial_Wait	= 1[s]
Phase	= {0, 90, 270, 180, 180,}



```
---- PROCESSING PARAMETERS ----
dc_balance( 0, FALSE )
sexp( 2.0[Hz], 0.0[s] )
trapezoid3( 0[%], 80[%], 100[%] )
zerofill( 1, TRUE )
fft( 1, TRUE, TRUE )
machinephase
ppm
```

Derived from: MKN219-pure Carbon-1.jdf

```

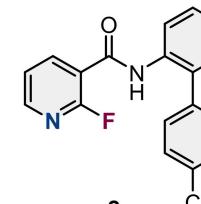
Filepath = MKN219_pure_Carbon-2.jdf
Author = element
Experiment = single_pulse_dec
Sample_Id = 1
Solvent = CHLOROFORM-D
Actual_Start_Time = 23-JAN-2024 19:20:17
Revision_Time = 15-JUN-2024 18:49:15

Comment = single pulse decoupled gat
Data_Format = 1D COMPLEX
Dim_Size = 26214
X_Domain = 13C
Dim_Title = 13C
Dim_Units = [ppm]
Dimensions = X
Site = ECS 400
Spectrometer = JNM-ECS400

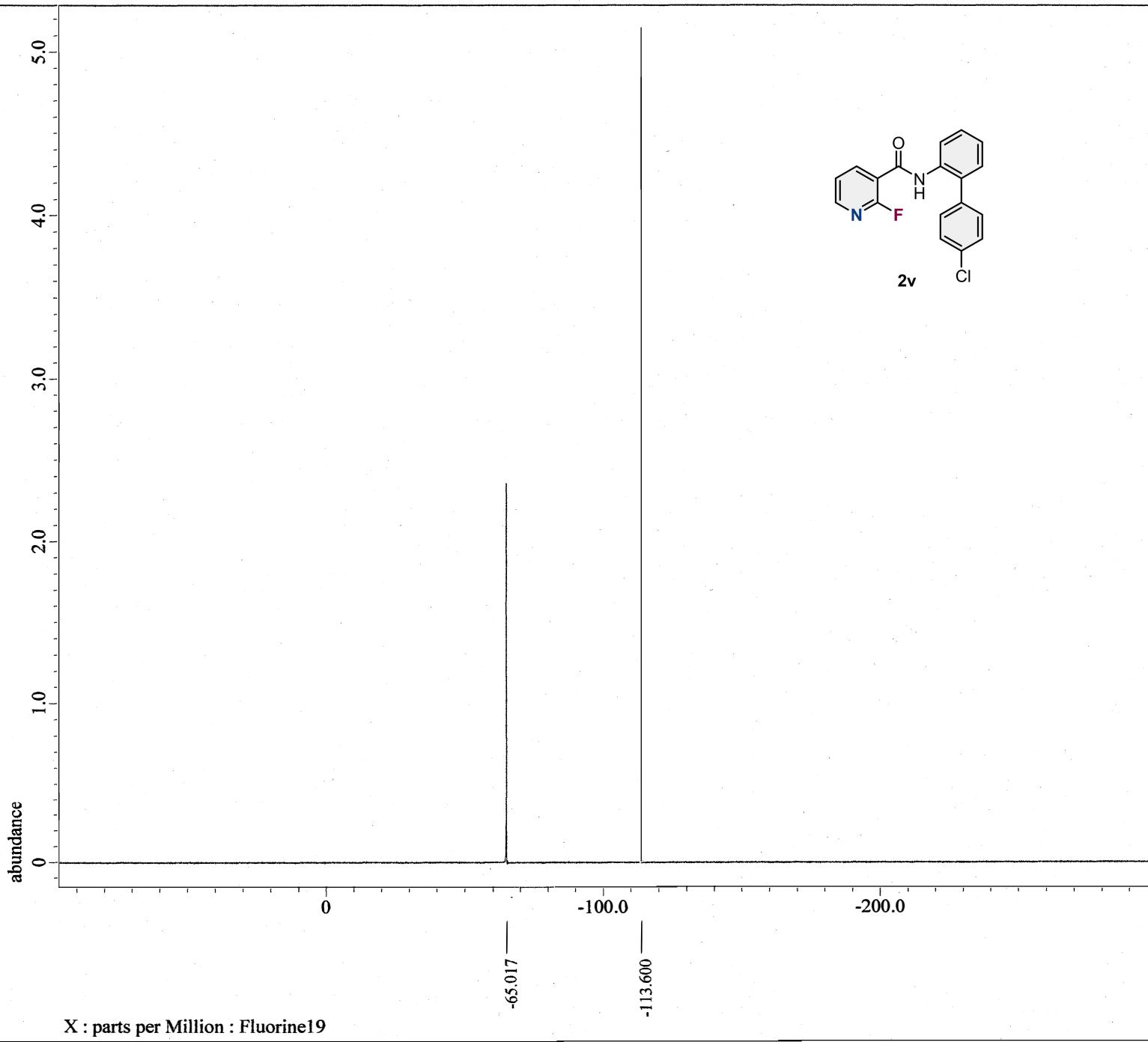
Field_Strength = 9.20197068[T] (390[MHz])
X_Acq_Duration = 1.06430464[s]
X_Domain = 13C
X_Freq = 98.51479726[MHz]
X_Offset = 100[ppm]
X_Points = 32768
X_Prescans = 4
X_Resolution = 0.93958061[Hz]
X_Sweep = 30.78817734[kHz]
Irr_Domain = 1H
Irr_Freq = 391.78655441[MHz]
Irr_Offset = 5[ppm]
Clipped = FALSE
Scans = 509
Total_Scans = 509

Relaxation_Delay = 2[s]
Recvr_Gain = 50
Temp_Get = 19.9[dC]
X_90_Width = 9.46[us]
X_Acq_Time = 1.06430464[s]
X_Angle = 30[deg]
X_Atn = 4.9[dB]
X_Pulse = 3.15333333[us]
Irr_Atn_Dec = 22.45[dB]
Irr_Atn_Noe = 22.45[dB]
Irr_Noise = WALTZ
Decoupling = TRUE
Initial_Wait = 1[s]
Noe = TRUE
Noe_Time = 2[s]
Repetition_Time = 3.06430464[s]

```



2



X : parts per Million : Fluorine19