

Supplementary Information

Bioconversion of a lignin-derived biphenyl dimer into the strategic building block 5-carboxyvanillic acid in *Pseudomonas putida* KT2440

Helena Gómez-Álvarez^{a c 1*}, Carlos del Cerro-Sánchez^{a c 1}, Pablo Iturbe^a, Virginia Rivero-Buceta^{a c}, Juan Nogales^{b c}, Timothy D. H. Bugg^d and Eduardo Díaz^{a c *}

^a Department of Biotechnology, Margarita Salas Center for Biological Research, Spanish National Research Council, Ramiro de Maeztu 9, 28040 Madrid, Spain

^b Department of Systems Biology, National Centre for Biotechnology, Spanish National Research Council, Darwin 3, 28049 Madrid, Spain

^c Interdisciplinary Platform for Sustainable Plastics towards a Circular Economy-Spanish National Research Council (SusPlast-CSIC), Madrid, Spain

^d Department of Chemistry, University of Warwick, Coventry CV4 7AL, United Kingdom

* Corresponding authors at: Department of Biotechnology, Margarita Salas Center for Biological Research, Spanish National Research Council, Ramiro de Maeztu 9, 28040 Madrid, Spain. E-mail addresses: ediaz@cib.csic.es (E. Díaz), hgomalv@cib.csic.es (H. Gómez-Álvarez).

¹ These authors contributed equally to this work.

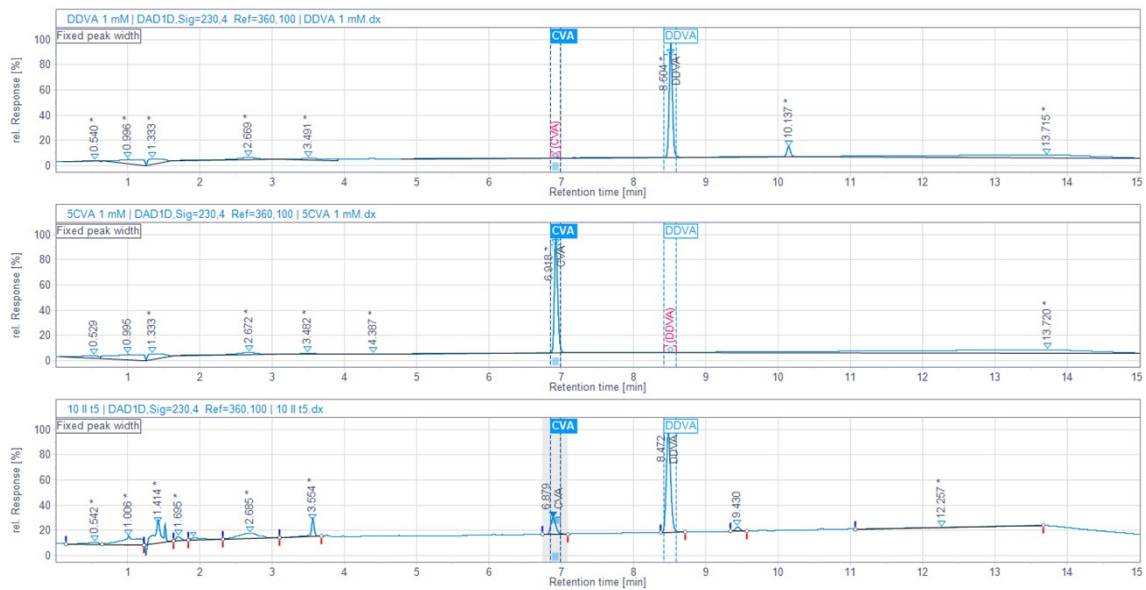
Supplementary Figures

Figure S1. Schematic representation of plasmids pIZDDVA_K, pIZDDVA and p254DDVT. Generated with SnapGene 7.2.

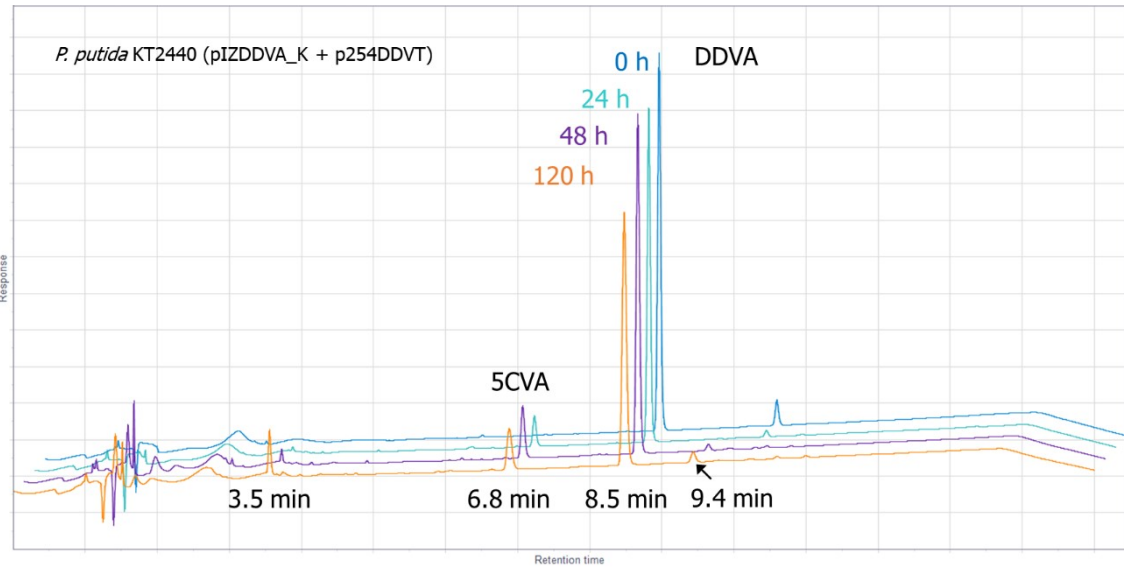
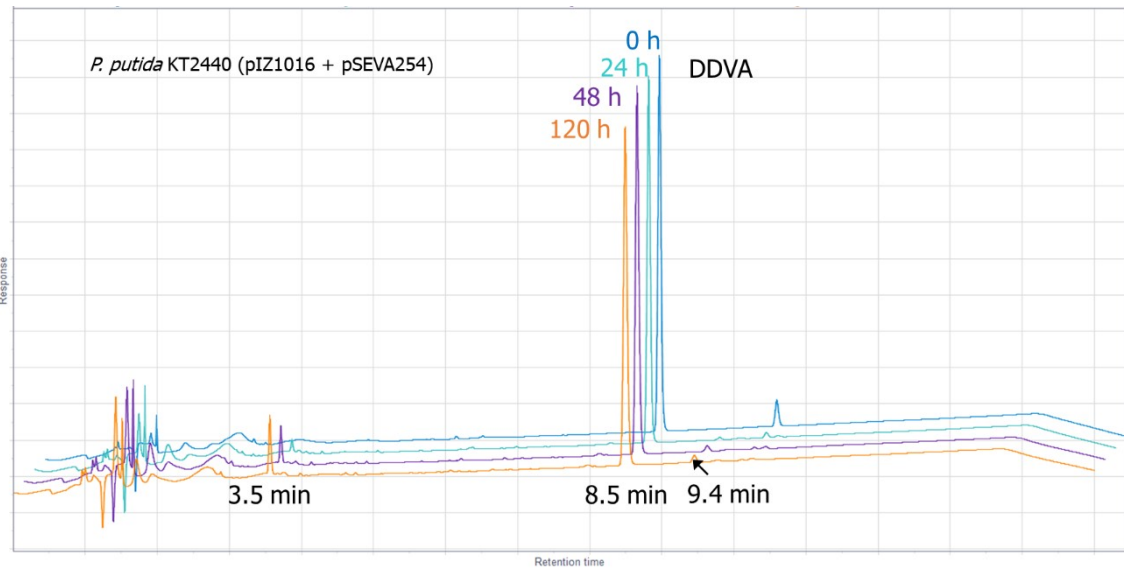


Figure S2. LC/MS representative chromatograms. (A) HPLC Chromatograms showing DAD signals at 230 nm. From top to bottom: signals corresponding to authentic DDVA, authentic 5CVA and sample taken from *P. putida* KT2440 (pIZDDVA_K + p254DDVT) resting cells incubated during 5 days in the presence of 1 mM DDVA at pH 7.5. (B) HPLC chromatograms showing evolution of DAD signals at 230 nm over time of samples taken from resting cells experiments (1 mM DDVA, pH 7.5) with *P. putida* KT2440 bearing either empty plasmids pIZ1016 and pSEVA254 (top) or pIZDDVA_K and p254DDVT (bottom). (C) MS spectrum corresponding to peaks at retention times 3.5 (top), 6.8 (middle) and 9.4 (bottom) minutes from the sample shown in panel B (down). Expected m/z values in this MS method for DDVA pathway intermediaries and final product are: OH-DDVA, 321; DCHM-HOPDA, 350.23; 5CVA, 213.16.

A



B



C

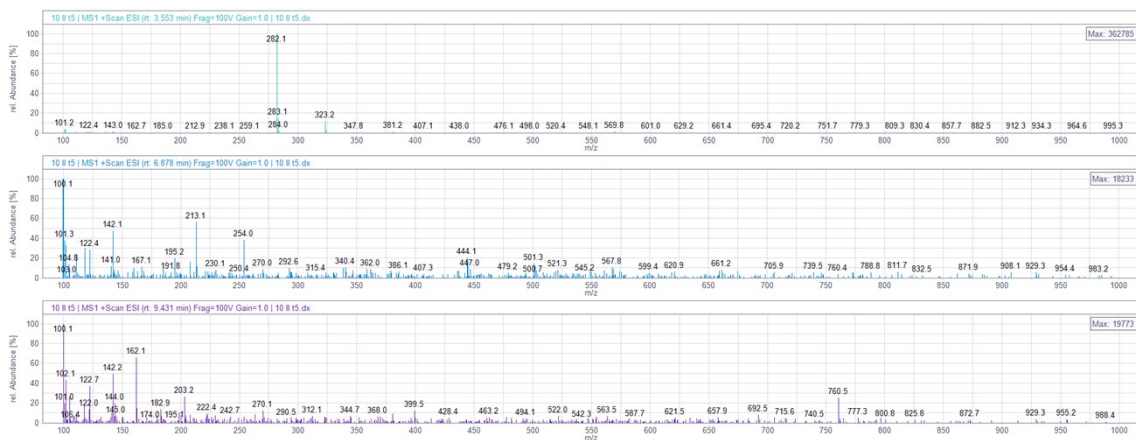


Figure S3 Identification of a consensus TonB box in *P. putida* and functional analysis of a chimeric DdvT* transporter. (A) Amino acid sequence alignment of 11 putative TonB boxes of TonB-dependent transporters from *P. putida* KT2440 and the BtuB receptor from *P. aeruginosa* with the TonB box region of the DdvT transporter from *S. lignivorans* SYK6 (highlighted in purple). Colors indicate similarity (dark pink indicates similar residues in all sequences, light pink indicates similar residues in more than 80% sequences, and white indicates similar residues in less than 80% sequences) considering a Blossum62 score matrix. Logo image was modified from the obtained using WebLogo 3 tool.⁵⁵ (B) 5CVA production profile from 1 mM DDVA using resting cells of *P. putida* KT2440 containing plasmid pIZDDVA_K and plasmid p254DDVTB (encodes the chimeric DdvT* transporter). Samples were taken at 0, 4, 24, 48, 72, and 120 hours. DDVA consumption and 5CVA production are indicated with discontinuous and continuous lines, respectively. All the resting cells experiments were performed in triplicates. Error bars represent standard deviation of the triplicates.

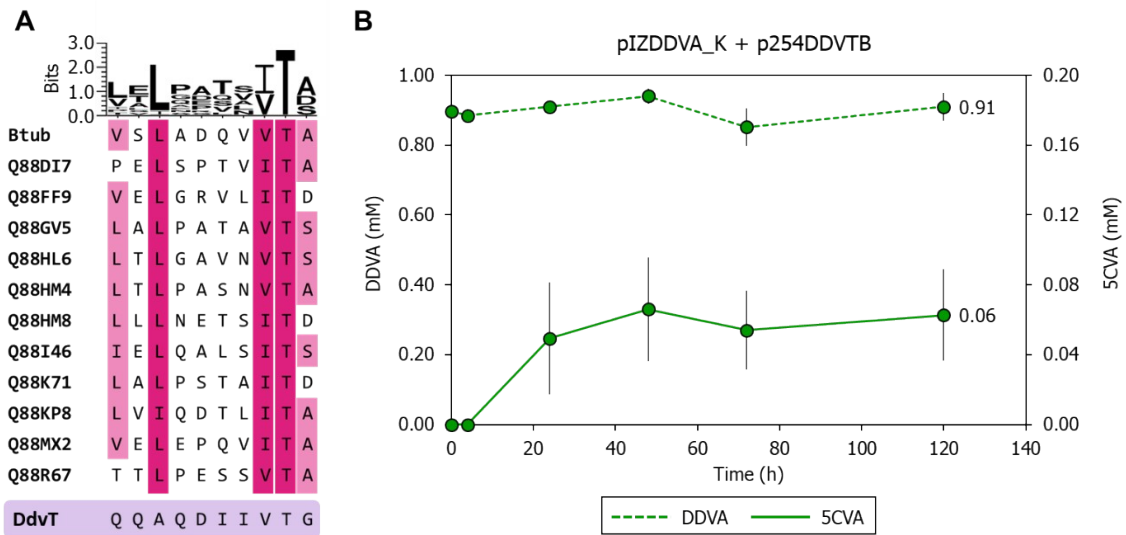
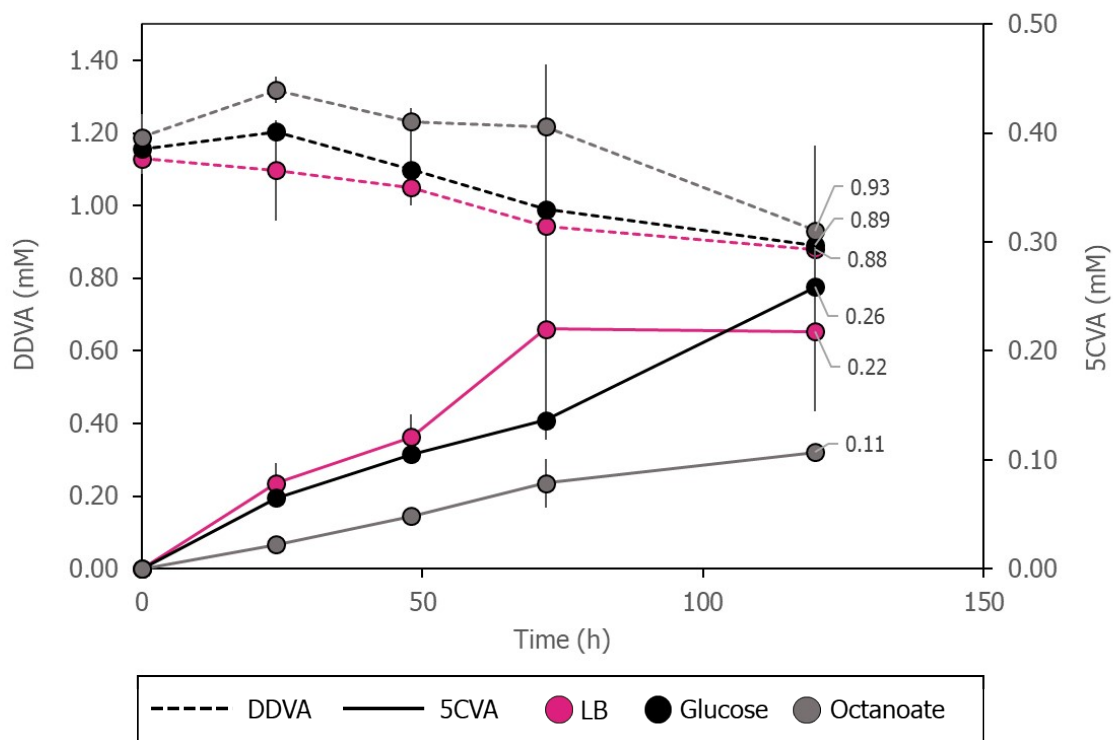


Figure S4. 5CVA production over time with *P. putida* KT2440 (pIZDDVA_K, p254DDVT) cells. Cells were grown in LB medium, M63 medium with 20 mM glucose, or M63 medium with 15 mM octanoate, and then collected (OD_{600} 24) for biotransformation of 1 mM DDVA in resting cells conditions.



Supplementary Tables

Table S1. Oligonucleotides used in this study.

Primer	Sequence (5'-3')
5 ligX	CGGCCGCGCGAATTCGAGCTCGGTACCCTAAGGAGGTGATCTAATGCTGTCCGCAGACCAGAACGACAAGCTG
3 ligX	TATCCTCTCCCAACCTCCTCTTCGGCACCGCATTAGATCACCTCCTTATCACGCGTCGACATAGCCCTTGT TATCCTG
5 ligY	TTCGTGGCCTGGCAATAAGCGAACCGCTTTAAGGAGGTGATCTAATGATCATCGACTGCCATGGTCACGT CAGCGCTCCGGTGGGA
3 ligY	ATGCCTGCAGGTCGACTCTAGAGGATCCCCTCAAACGTCCAAATTAACACCTTCCGCGCATTATC
5 ligZ	CAGGATAACAAGGGCTATGTCGACGCGTGATAAGGAGGTGATCTAATGCGGTGCCGAAGAGGAGTTGGGA GAGGATA
3 ligZ	TCCACCGGAGCGCTGACGTGACCATGGCAGTCGATGATCATTAGATCACCTCCTTAAACGCGTTCGCTTA TTGCCAGGCCACGAA
ligXc_Fw	ATATATCTAGAAAGGAGGTGATCTAATGGCGCAGCTGAAGGTCG
ligXc_Rv	ATATAAAGCTTTCATCCTCCGGGCGATGGT
ligXd_Fw	ATATAAAGCTTAAGGAGGTGATCTAATGCCGCATTTTGATTGCCTTA
ligXd_Rv	ATATAACTAGTTCATCCCTGCGCGCGGTCA
ligXa tune Fw KpnI	CGGGGTACCCTAAGGAGGTGATCTAATGCTGTCCGCAG
ligXa tune Rv NheI NdeI	AAGCAAGCTAGCCATATGATCACCTCCTTATCACGCGTCGACAT
ligZ tune Fw NdeI	ATATATCATATGGCCGAAATCGTGTGGGT
ligZ tune Rv XhoI	TCCGCTCGAGCGTTCGCTTATTGCCAGGCCACGAAGC
ligY tune Fw NheI XhoI	AAGCAAGCTAGCCTCGAGAAGGAGGTGATCTAATGATCATCGACTGCCATGG
ligY tune Rv BamHI	CCGGGATCCCCTCAAACGTCCAAATTAACAC
MFS Fw AvrII	ATATCCTAGGAAGGAGGTGATCTAATGGAGAGGATGACGATGGCA
MFS Rv EcoRI	TATATGAATTCTCAGTCCGGGATTTTGCGCG
galBC Fw AvrII EcoRI	TATCCTAGGAATTC AAGGAGGTGATCTAATGACATCCTGCGCCACCC
galBC Rv SacI	ATAGAGCTCTCAGCCTTCCAGGTCGGTGA
XmaI lig frag Fw	ATATATCCCGGAAGGAGGTGATCTAATGGAGAG
SpeI lig frag Rv	ATATATACTAGTTCATCCCTGCGCGCGGTC
oDCS7	GGGTGGGCGCAGGATGTCATTAGATCACCTCCTTCCCGGGC
oDCS8	ATGACATCCTGCGCCACCCC
ddvT_Fw_SD_XbaI	TATATTCTAGATGACCTAAGGAGGTAATAATGGCTCGCGGAACGTAATTG
ddvT Rv NheI	TATATGCTAGCTCAGAAGCCGAACCGCAC
LELPATVITA_Rv	CGCGGTGATGACGGTGGCCGGCAGCTCCAGCGCCGTCATTTCTGAGC
LELPATVITA_Fw	CTGGAGTGCCTGGCCACCGTCATCACCGCTCGCGCGTGGCCCGAAGC
ddvT_Rv_HindIII	TGACTGAAGCTTTCAGAAGCCGAACCGCAC