## SUPPLEMENTARY INFORMATION

## ElectrochemCap: an integrated detection of loop-mediated isothermal amplification reactions

P. Rioboó-Legaspi, E. Costa-Rama\*, M.T. Fernández-Abedul\*

Departamento de Química Física y Analítica, Facultad de Química, Universidad de Oviedo, Spain.

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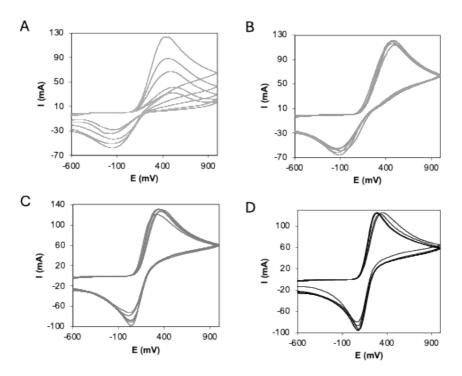
## Procedure 1. Detailed LAMP assay.

The reaction mix used contained: 1x master mix (WarmStart<sup>®</sup> Colorimetric LAMP 2X, including Bst 2.0 DNA Polymerase and phenol red, New England Biolabs), primer mix (1.6  $\mu$ M each FIP and BIP primer, 0.2  $\mu$ M each F3 and B3 primer and 0.4  $\mu$ M each LoopF and LoopB primer and PCR-grade dH<sub>2</sub>O (Invitrogen UltraPureTM Distilled Water (DNase/RNAse free), Life Technologies) up to 48  $\mu$ L of final volume. A volume of 2  $\mu$ L of template SARS-CoV-2 RNA (VR-1986D, ATCC<sup>®</sup>), if required, was added at the desired concentration. All concentrations are indicated as copies· $\mu$ L<sup>-1</sup> of standard or sample solution. In the case of negative (non-template) control reactions, 2  $\mu$ L of PCR-grade dH<sub>2</sub>O were added. All reactions were performed on low-retention, nuclease-free PCR grade tubes (PCR-02-L-C, AxygenTM) and were carried out under sterile conditions in a laminar flow hood. LAMPs were performed in a thermal block (MSC-100, Lan Technics) at 65°C for 30 min.

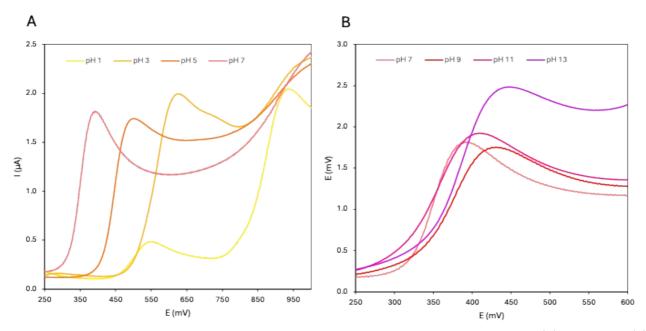
Fragment	Primer	Sequence	Position
N1	F3	TGGACCCCAAAATCAGCG	28278 -28295
	B3	GCCTTGTCCTCGAGGGAAT	28479 - 28461
	FIP	CCACTGCGTTCTCCATTCTGGTAAATGCACCCCGCATTACG	28367 – 28340/ 28296 - 28314
	BIP	CGCGATCAAAACAACGTCGGCCCTTGCCATGTTGAGTGAG	28370 – 28392/ 28450 - 28431
	Loop F	TGAATCTGAGGGTCCACCAAA	28335 - 28315
	Loop B	GGTTTACCCAATAATACTGCGTCTT	28396 – 28420

Table S2. Economic costs of the ElectrochemCap.

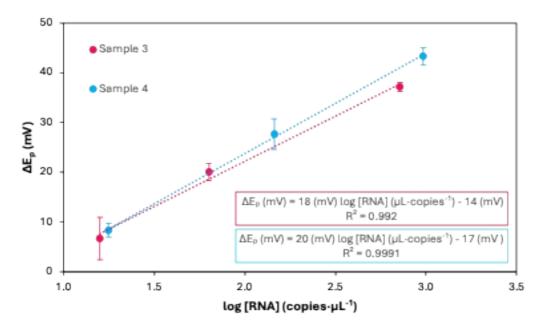
Component	Size per platform	Price of bulk material	Price/platform (EUR)
SPE	1 unit	76 EUR/50 units	1.52
PSA	1.6 cm <sup>2</sup>	0.014 EUR/cm <sup>2</sup>	0.02
TPU adapter	0.54 g	19.33 EUR/kg	0.01
Microcentrifuge tube	1 unit	70.2 EUR/1000 units	0.07
	1.62		
Reaction mix	50 μL	0.081 €/μL	4.05€
TOT	5.67€		



**Figure S1.** Electrochemical characterization of the ElectroCap using different reaction volumes. CVs (n = 5) recorded using A: 25  $\mu$ L; B: 50  $\mu$ L; C: 100  $\mu$ L; D: 200  $\mu$ L of 5 mM K<sub>3</sub>[Fe(CN)<sub>6</sub>]. Scan rate 0.1 V·s<sup>-1</sup>.



**Figure S2.** Phenol red linear sweep voltammograms recorded in 100  $\mu$ M solutions, in more acidic (A) and alkaline (B) conditions. A: in 0.1 M HCl and in 0.1 M BR buffers of pH 3, 5 and 7. B: LSVs recorded in 100  $\mu$ M PR solutions in 0.1 M BR buffers of pH 7, 9 and 11, and in 0.1 M NaOH. Scan rate 0.1 V·s<sup>-1</sup>.

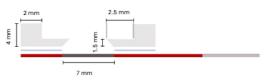


**Figure S3.** Sample dilution results. The shift in peak potential is represented *vs.* logarithm of the expected copy number. Regression equations and correlation coefficient are shown in a red or blue box for samples 3 and 4, respectively. All error bars represent the standard deviation of three replicates.

A 1.5 mL tube

3.25 mm 16 mm 7 mm 3.75 r Ι 7 mm

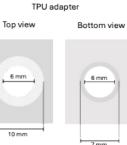
В Flat-bottomed 1.5 mL tube



PSA layer 7 mm 16 mm 14 mm

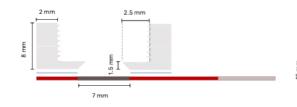
PSA layer

10 mm

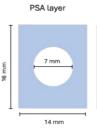


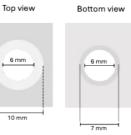
Screw-cap 1.5 mL tube

С

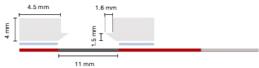








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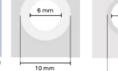


Ε 8- tube 0.5 mm 4.5 4 mm ĒI

7 mm

11 mn 8.7 mm 16 mm

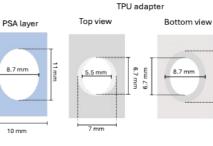
16 mm



20 mm

7 mm

TPU adapter

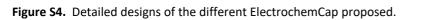


6 mm

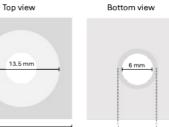
10 mm

PSA layer 112 mm TPU adapter Top view

Bottom view



TPU adapter



7 mm