# **SUPPLEMENTARY INFORMATION**

# **Exploring the influence of mesoporosity in hard carbon-templated hierarchical SAPO-5 for ethanol dehydration**

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### <span id="page-1-0"></span>**Experimental details**

**Powder Diffraction (XRD):** Powder X-ray diffraction was per-formed on a Bruker D2 Phaser diffractometer using a 1D LYNXEYE detector and 0.6 mm slits, with Cu K<sub>a1</sub> radiation. Patterns were run over a 2θ range of 5 to 40 ° with a scan speed of 3 ° min-1 and an increment of 0.01 °. Powdered samples were placed into a plastic holder with roughly 3 mm thickness.

**Inductively Coupled Plasma (ICP):** ICP was performed by MEDAC *via* their commercial mail-in service. For further details see: https://medacltd.com/services-view/metals-icp/.

**Elemental Analysis (CHN):** CHN was performed by London Metropolitan University *via* their commercial mail-in service. For further details see: https://www.londonmet.ac.uk/services-andfacilities/elemental-analysis-service/.

**Nitrogen Physisorption (BET):** BET surface area measurements were performed at 77 K, on a sample dried under 20 mTorr of vacuum at 120 °C overnight. Analysis was per-formed on a Micromeritics Gemini 2375 surface area analyser. Surface area was calculated using the BET model, $1$  while the pore width distribution was calculated with the BJH model, which assumed cylindrical pores.<sup>2</sup> The "total pore volume" was calculated by converting the volume of gaseous nitrogen absorbed to the volume of liquid nitrogen adsorbed, using a conversion factor of 647. The BJH distribution plot<sup>2</sup> showed the variation in pore volume with respect to pore size, the total pore volume calculated for pores above 20 Å was taken as the "mesopore pore volume". The difference between the "total pore volume" and "mesopore pore volume" was calculated as being the "micropore pore volume".

**Solid State Nuclear Magnetic Resonance (ssNMR):** Samples were packed into 4 mm pencil-style zirconium oxide rotors and acquired at a spinning rate of 8 kHz using a 4 mm RevolutionNMR probe.  $^{1}$ H, <sup>27</sup>Al, and  $^{31}$ P spectra were acquired in triple resonance mode, and <sup>29</sup>Si in double resonance on a Bruker Avance Neo widebore Spectrometer, using a 9.4 T field, in air. The sample <sup>1</sup>H, <sup>27</sup>Al, and <sup>31</sup>P, and <sup>29</sup>Si T<sub>1</sub> relaxation time was assessed using a saturation recovery pulse technique, which was used for the respective direct acquisitions. Specifically the following conditions were used:

 $27$ Al NMR: 16 scans, pulse delay of 7 seconds, short pulse excitation

<sup>31</sup>P NMR: 4 scans, pulse delay of between 30 and 50 seconds

 $^{29}$ Si NMR: 4096 scans, pulse delay of 31 seconds, contact time of 6 milliseconds

<sup>1</sup>H NMR: 8 scans, pulse delay of 3 seconds

**Scanning Electron Microscopy (SEM):** SEM images were acquired at the Southampton Biomedical Imaging Unit, using an FEI Quanta 250 FEG scanning electron microscope. Samples were sputter coated with platinum prior to imaging.

**Transmission Electron Microscopy (TEM)**: TEM images were acquired at the Electron Microscopy Centre of the Utrecht University using a Thermo Fisher Scientific TalosF200X electron microscope

**Temperature Programmed Desorption (TPD):** NH<sub>3</sub>-TPD was performed at the Research Complex at Harwell, as part of the UK Catalysis Hub, on a Quantachrome ChemBET Pulsar TPR/TPD Automated Chemisorption Analyser. 0.2 g of pelletised and sieved catalyst (150 – 425 μm) was used for each analysis. The sample was first recalcined at 550 °C for 2 hours, ramping at 10 °C min<sup>-1</sup>, under a 30 mL min<sup>-1</sup> flow of 20 %  $O_2$  in N<sub>2</sub>. The system was then cooled to 150 °C, and held for 2 hours whilst under a flow of 30 mL min<sup>-1</sup> of 5% NH<sub>3</sub> in He. The flow was then changed to 30 mL min<sup>-1</sup> of He, and held at 150 °C for a further 2 hours. After which the system was then heated to 600 °C at a rate of 5 °C min<sup>-1</sup> and held for 1 hour. The data was then normalised and background subtracted to give values as a function of signal (mV  $g^{-1}$ ) versus temperature (°C).

#### <span id="page-2-0"></span>**Structural characterisation data**

#### <span id="page-2-1"></span>**Chemical composition**

**Table S1:** Showing the variation in elemental composition of the three calcined SAPO-5 systems "Calc", contrasting the carbon and nitrogen content with the as-synthesised, uncalcined systems "AS".



#### <span id="page-2-2"></span>**Nitrogen physisorption metrics**

**Table S2:** Nitrogen physisorption values for the three SAPO-5 systems.



<span id="page-2-3"></span>**ssNMR spectra**





Fig. S1: ssNMR of the three SAPO-5 species, focussing on A)<sup>27</sup>Al, B)<sup>31</sup>P, C)<sup>29</sup>Si and D)<sup>1</sup>H nuclei.

<span id="page-4-0"></span>

**Fig. S2:** NH3-TPD profiles of the three SAPO-5 species, with the associated temperature ramp. Data is normalised per gram of sample for ease of comparison.



**Table S3:** Integrals and percentages of the three SAPO-5 systems, showing the influence of carbon templates on acidity.

<span id="page-5-0"></span>**Scanning electron microscopy (SEM) images**





**Fig. S3:** Showing the particle size, shape and uniformity of the three SAPO-5 systems *via* SEM images, at different magnifications, focussing on A & B) MP-SAPO-5 AS, C & D) MP-SAPO-5 Calc, E & F) CNP-SAPO-5 AS, G & H) CNP-SAPO-5 Calc, I & J) CNT-SAPO-5 AS, and K & L) CNT-SAPO-5 Calc.

## <span id="page-7-0"></span>**Transmission electron microscopy (TEM) images**



**Fig. S4:** Electron microscopy images of SAPO-5. A-C) Uncalcined and D-F) calcined. A,D) High resolution TEM images of the crystalline systems. B,E) Low resolution HAADF-STEM images show slight surface roughening after calcination. C,F) High resolution HAADF-STEM images show the presence of 1-2 nm nanoparticles.



**Fig. S5:** Electron microscopy images of uncalcined CNT-SAPO-5 at different magnifications, showing the affinity between the SAPO-5 and CNT.

## <span id="page-8-0"></span>**Comparison of MP-SAPO-5 Morphology**

Whilst the same synthesis procedure was followed in this work as in our previous work<sup>3</sup>, different batches of reagents were used, and this seems to have resulted in subtle morphological differences between the MP-SAPO-5 samples, see Figure S5. The crystals from the earlier work appear to be more discrete and better defined. There are consequently differences in the SANS between the two materials, most notable in the region  $0.01 \le Q \le 0.1$  Å<sup>-1</sup>. Whilst the two samples were collected on different instruments, they have been calibrated in an identical manner, so the difference in intensity between the samples (roughly a factor of 2) is likely due to the difference in packing between the two samples. Thus, the previous (more intense) data is likely packed more densely in the cells, and thus more is illuminated by the neutron beam, than in the current data.



**Fig. S6:** SEM micrographs comparing the crystallinity of the MP-SAPO-5 species synthesized for A) our previous work described in reference 3 and B) this work, along with C) the corresponding SANS data (measured on the LOQ and SANS2D instruments at the ISIS Neutron and Muon Source, respectively).

Q (Å-1)

 $1E-1$ 

-MP SAPO-5 Previous Work

 $1E-2$ 

ॾ  $2E-2$ 

 $2E-3$ 

 $1E+0$ 

 $1F-1$ 

 $1E-2$ 

 $1E-3$ 

# <span id="page-9-0"></span>**Small angle neutron scattering (SANS) data fitting**

#### <span id="page-9-1"></span>**Power law – Fractal – Gaussian Peak model**

**Table S4:** Fitting parameters and values, with associated uncertainties, achieved with a model including a power law, fractal aggregate and Gaussian peak for the three SAPO-5 systems. Grey cells denote parameters that were not optimised.



a) HWHM of Lognormal distribution at the median radius divided by the median radius.

b) One standard deviation = FWHM/2.354

This model is described by a linear combination of the following Q-dependent functions: a power law (to account for interfacial scattering), a fractal aggregate of polydisperse spheres (to account for micropores), and a Gaussian peak (to account for the Bragg peak):

 $Intensity(Q)_{P+F(PD)+G} = Background + A.f(Q)_{PowerLaw} + B.f(Q)_{Fractal(PD)} + D.f(Q)_{Gaussian}$ 

Here, A, B & D are scalar quantities that are proportional to the volume fraction of that component in the model. Links to the individual model descriptions may be found below.





**Fig. S7:** (A-C) Fits of the Power law – Fractal - Gaussian model as defined above to the SANS data from MP-SAPO-5, CNP-SAPO-5 and CNT-SAPO-5, respectively. (D) Residual intensity (observed – calculated) from these fits, serving as a measure of fit quality.

#### <span id="page-11-0"></span>**Power law – Fractal – Monodisperse Sphere – Gaussian Peak Model**

**Table S5:** Fitting parameters and values, with associated uncertainties, achieved with a model including a power law, fractal aggregate, sphere and Gaussian peak for the three SAPO-5 systems. Grey cells denote parameters that were not optimised.





a) HWHM of Lognormal distribution at the median radius divided by the median radius.

b) One standard deviation = FWHM/2.354

This model is described by a linear combination of the following Q-dependent functions: a power law (to account for interfacial scattering), a fractal aggegrate of polydisperse spheres (to account for micropores), monodisperse spheres (to account for mesopores), and a Gaussian peak (to account for the Bragg peak):

 $Intensity(Q)_{P+F(PD)+S+G} = Background + A.f(Q)_{PowerLaw} + B.f(Q)_{Fraction}(PD) + C.f(Q)_{Sphere} + D.f(Q)_{Gaussian}$ 

Here, A, B, C & D are scalar quantities that are proportional to the volume fraction of that component in the model. Links to the individual model descriptions may be found below.





**Fig. S8:** (A-C) Fits of the Power law – Fractal – Sphere - Gaussian model as defined above to the SANS data from MP-SAPO-5, CNP-SAPO-5 and CNT-SAPO-5, respectively. (D) Residual intensity (observed – calculated) from these fits, serving as a measure of fit quality.





**Fig. S9:** Fits of the Power law – Fractal – Sphere - Gaussian model as defined above to the SANS data from MP-SAPO-5, CNP-SAPO-5 and CNT-SAPO-5, respectively. (A-C) Showing the contributions of the individual component functions. (D) Comparing just the Sphere contributions.

## <span id="page-16-0"></span>**Power law – Fractal – Polydisperse Sphere – Gaussian Peak Model**

**Table S6:** Fitting parameters and values, with associated uncertainties, achieved with a model including a power law, fractal aggregate, sphere (with variable polydispersity) and Gaussian peak for the three SAPO-5 systems. Grey cells denote parameters that were not optimised.





a) HWHM of Lognormal distribution at the median radius divided by the median radius. b) One standard deviation = FWHM/2.354

This model is described by a linear combination of the following Q-dependent functions: a power law (to account for interfacial scattering), a fractal aggregate of polydisperse spheres (to account for micropores), polydisperse spheres (to account for mesopores), and a Gaussian peak (to account for the Bragg peak):

 $Intensity(Q)_{P+F(PD)+S+G} = Background + A.f(Q)_{PowerLaw} + B.f(Q)_{Fractal(PD)} + C.f(Q)_{Sphere} + D.f(Q)_{Gaussian}$ 

Here, A, B, C & D are scalar quantities that are proportional to the volume fraction of that component in the model. Links to the individual model descriptions may be found below.

In this last set of model fits the uniformity of the mesopore sizes in the three samples was explored by including polydispersity on the spherical form factor. As can be seen, this had little influence on the parameters for the MP-SAPO-5 system, where  $\chi^2$  improved slightly from 8.4 to 8.0. For CNP-SAPO-5,  $\chi^2$  improved from 48.4 to 39.1. However, for CNT-SAPO-5 the fit improved significantly, with  $\chi^2$  falling from 35.0 to 11.8. This reinforces the theory that the mesopores in the CNT-SAPO-5 species are less uniform than the CNP-SAPO-5 species.

# <span id="page-17-0"></span>**Descriptions of model components**



<span id="page-18-0"></span>

**Fig. S10:** Catalytic data for ethanol dehydration with the three SAPO-5 systems showing A) ethanol conversion and B) diethyl ether yield. Conditions: 230 °C, 25 mL/min N<sub>2</sub> carrier gas, liquid feedstock of 10% heptane (internal standard) in ethanol, liquid flow varied from 3.13 to 12.51 μL min-1 as the WHSV varies from 0.5 to 2.0 hr<sup>-1</sup>. Errors are not shown but are calculated as  $\pm$  3 mol% based on multiple injections.

**Table S7:** Numerical catalytic data for ethanol dehydration with the three SAPO-5 systems showing Conditions: 230 °C, 25 mL/min N<sub>2</sub> carrier gas, liquid feedstock of 10% heptane (internal standard) in ethanol, liquid flow varied from 3.13 to 12.51  $\mu$ L min<sup>-1</sup> as the WHSV varies from 0.5 to 2.0 hr<sup>-1</sup>. Errors are not shown but calculated as  $\pm$  3 mol% based on multiple injections.



Note: Diethyl Ether Yield cannot surpass 50 mol%, as it is calculated based on moles of ethanol, as described in the experimental section of the main part of the paper.

## <span id="page-19-0"></span>**References**

- 1. S. Brunauer, P. H. Emmett and E. Teller, *Journal of the American Chemical Society*, 1938, **60**, 309-319.
- 2. E. P. Barrett, L. G. Joyner and P. P. Halenda, *Journal of the American Chemical Society*, 1951, **73**, 373-380.
- 3. M. E. Potter, A. E. Oakley, J. J. M. Le Brocq, L. N. Riley, M. Carravetta, S. M. King, C. M. Doherty, B. D. Vandegehuchte and R. Raja, *Journal of Materials Chemistry A*, 2023, **11**, 22822-22834.