

Colorless Copper-Containing Coatings With High Antimicrobial Efficacy and Formulation Versatility

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Copper Quantification Method Development

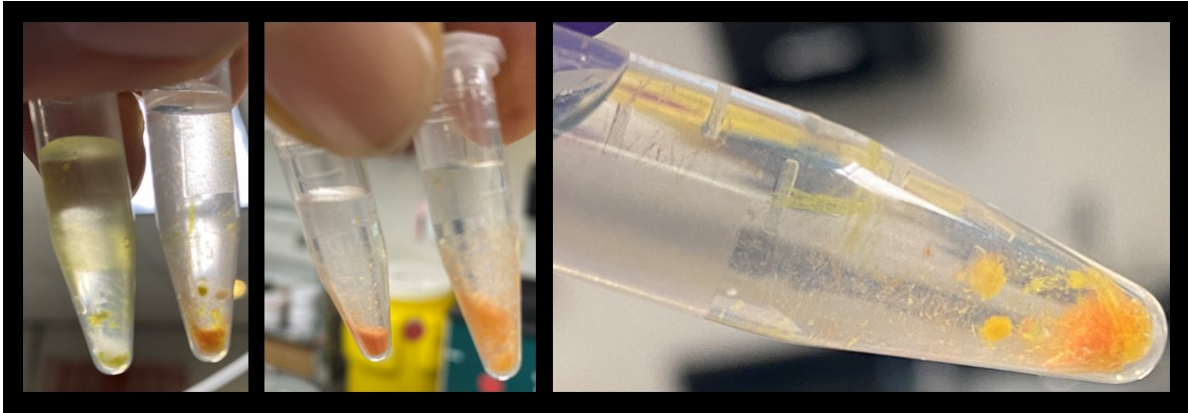


Figure S1: Visual example of CGC + Phosphite extraction samples prepared by the Neocuproine method showing crystallization and precipitation of the copper in microcentrifuge tubes.

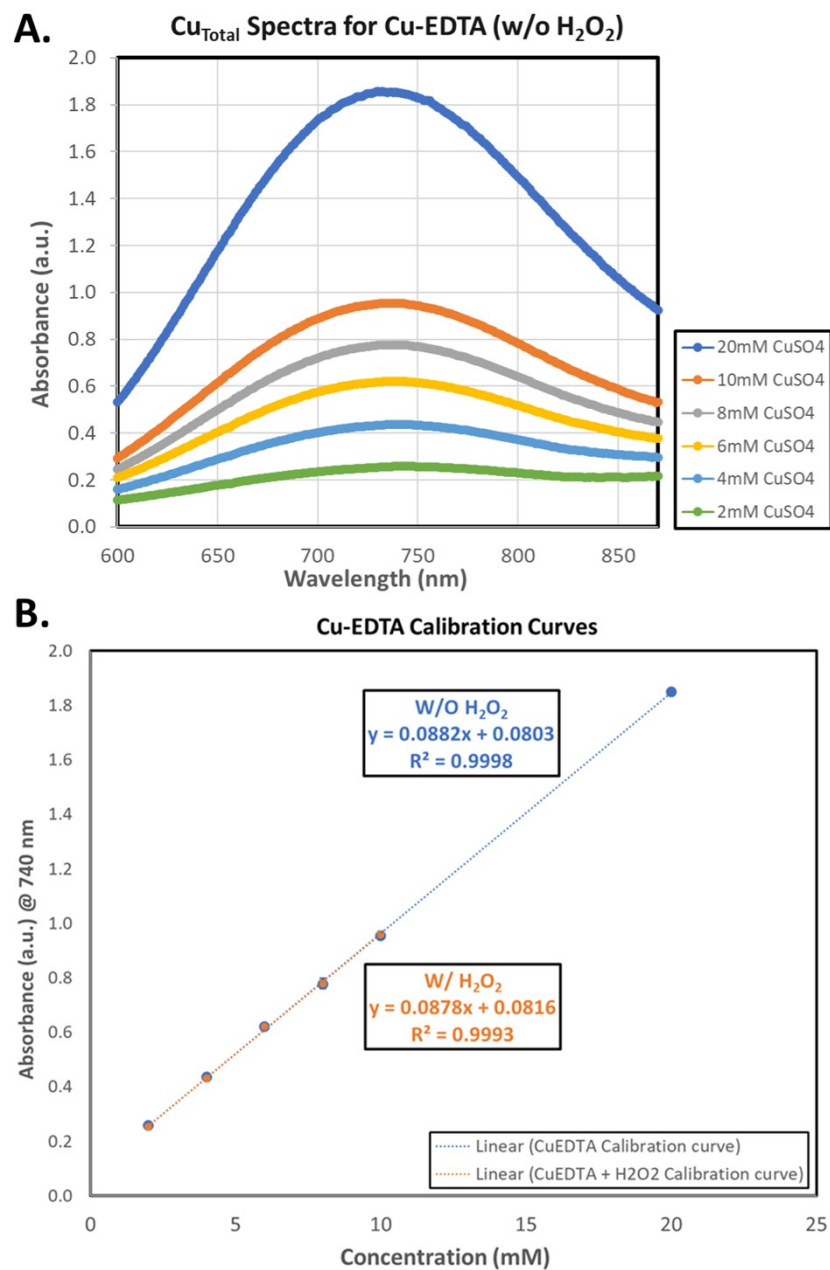


Figure S2: A. UV-vis data for the copper sulfate/EDTA calibration curve. B. Absorbance at 740 nm versus Cu²⁺ concentration calibration curves and line of best fit.

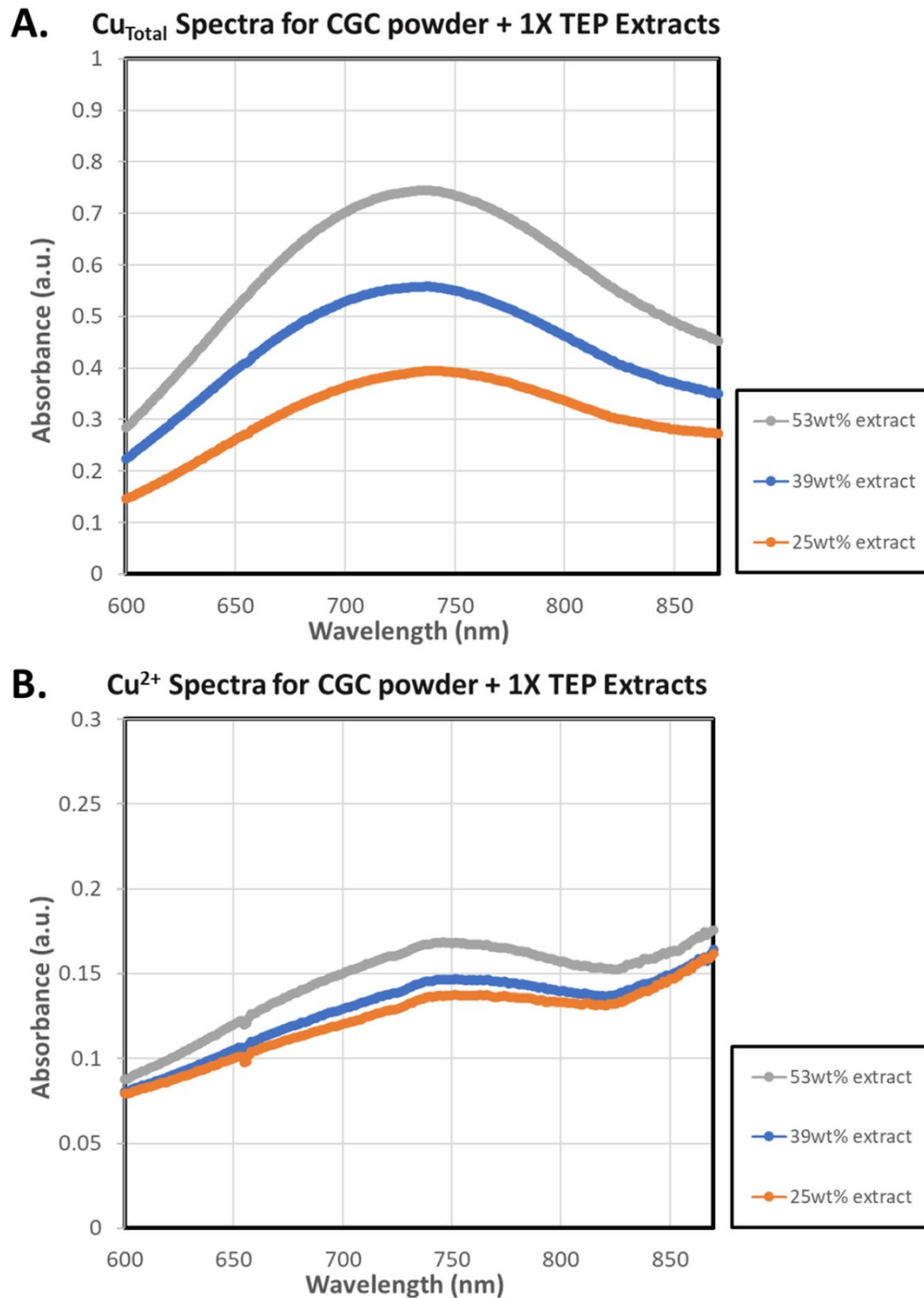


Figure S3: UV-vis spectrum for CGC powder extracts with TEP. **A.** Sample Cu_{Tot} curves. **B.** Curves used to derive $[\text{Cu}^{2+}]$. $[\text{Cu}_{\text{Tot}}]$ determination was made by adding 700 μl of 20 mM EDTA, 388 μl H_2O , 42 μl of copper extract solution, 20 μL of H_2O_2 . $[\text{Cu}^{2+}]$ determination was made by adding 700 μl of 20 mM EDTA, 398 μl H_2O , 42 μl of copper extract solution. $[\text{Cu}^{1+}]$ is calculated by taking the difference between $[\text{Cu}_{\text{Tot}}]$ and $[\text{Cu}^{2+}]$.

$^{31}\text{P}\{^1\text{H}\}$ NMR Parameters for Copper(I) Coordination Environment

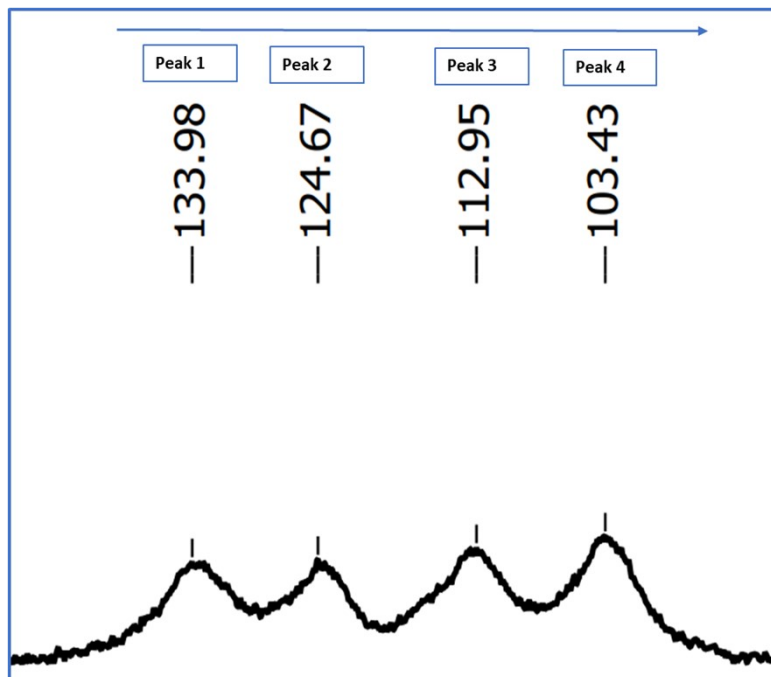


Figure S4: $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum of CGC extract with TEP showing an asymmetric quartet structure due to spin-spin coupling of the phosphorus nuclei to the copper nucleus (^{63}Cu , ^{65}Cu ; $I = 3/2$) in water.

The following expressions were used to investigate the possible relationship between the ^{31}P NMR parameters and the coordination environment about the copper(I) atom [1-5].

$$\Delta V1 \text{ (kHz)} = \frac{[121.41 \times (\text{peak 1} - \text{peak 2})]}{1000}$$

$$\Delta V2 \text{ (kHz)} = \frac{[121.41 \times (\text{peak 2} - \text{peak 3})]}{1000}$$

$$\Delta V3 \text{ (kHz)} = \frac{[121.41 \times (\text{peak 3} - \text{peak 4})]}{1000}$$

$$\Delta Vi \text{ (kHz)} = \frac{[\Delta V1 + \Delta V2 + \Delta V3]}{3}$$

Center δ	Peak 1	Peak 2	Peak 3	Peak 4
ppm	ppm	ppm	ppm	ppm
118.71	133.98	124.67	112.95	103.43

Table 1. Summary calculations of $^{31}\text{P}\{^1\text{H}\}$ NMR parameters used to determine number of phosphorous atoms coordinated to the copper(I) center.

				118.71	Center (ppm)		
9.31	ppm	1130.33	Hz	1.13	ΔV_1	kHz	Low field line spacing
11.72	ppm	1422.93	Hz	1.42	ΔV_2	kHz	
9.52	ppm	1155.82	Hz	1.16	ΔV_3	kHz	High field line spacing
				1.24	ΔV_i (average)	kHz	*Coordination number information
				1.02	$\Delta V_3/V_1$ (ratio)		Asymmetry of the line spacing

*How to interpret the line spacing $\langle \Delta V_i \rangle$ of the quartet?		
~1.6	kHz	One coordinated phosphine
1.2	kHz	Two coordinated phosphine
0.9	kHz	Three coordinated phosphine

Interpretation of results	
1.24	The derived $\Delta V_{i_{Av}}$ value suggests a two coordinate phosphorous-copper(I) complex is present.

Antimicrobial Glass Film Surface Metrology

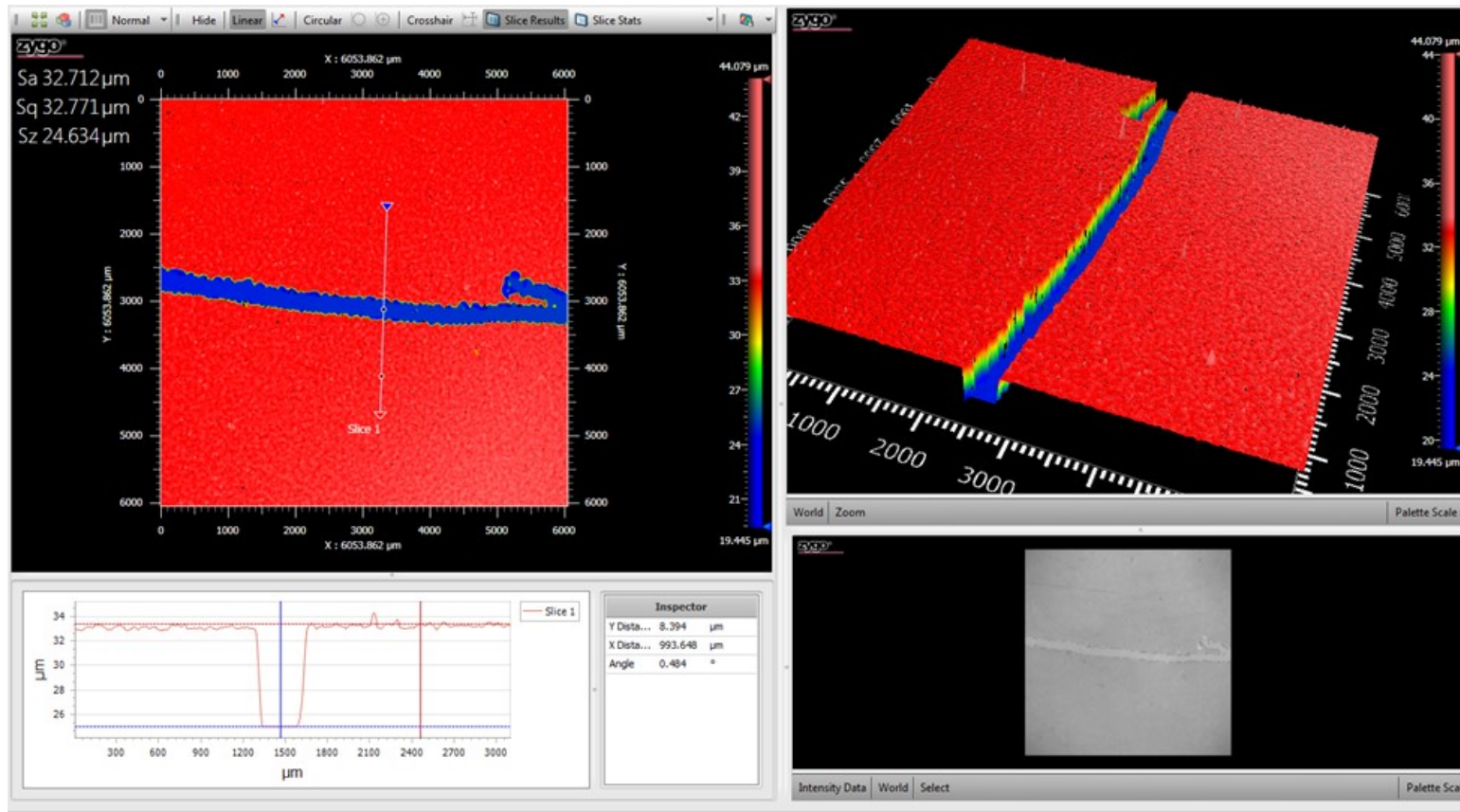


Figure S5: Zygo film thickness measurement for Ferro 221 series coating on Corning® EAGLE XG® Glass with no copper glass-ceramic (CGC) powder or CGC powder extract, flash cured at 150 °C.

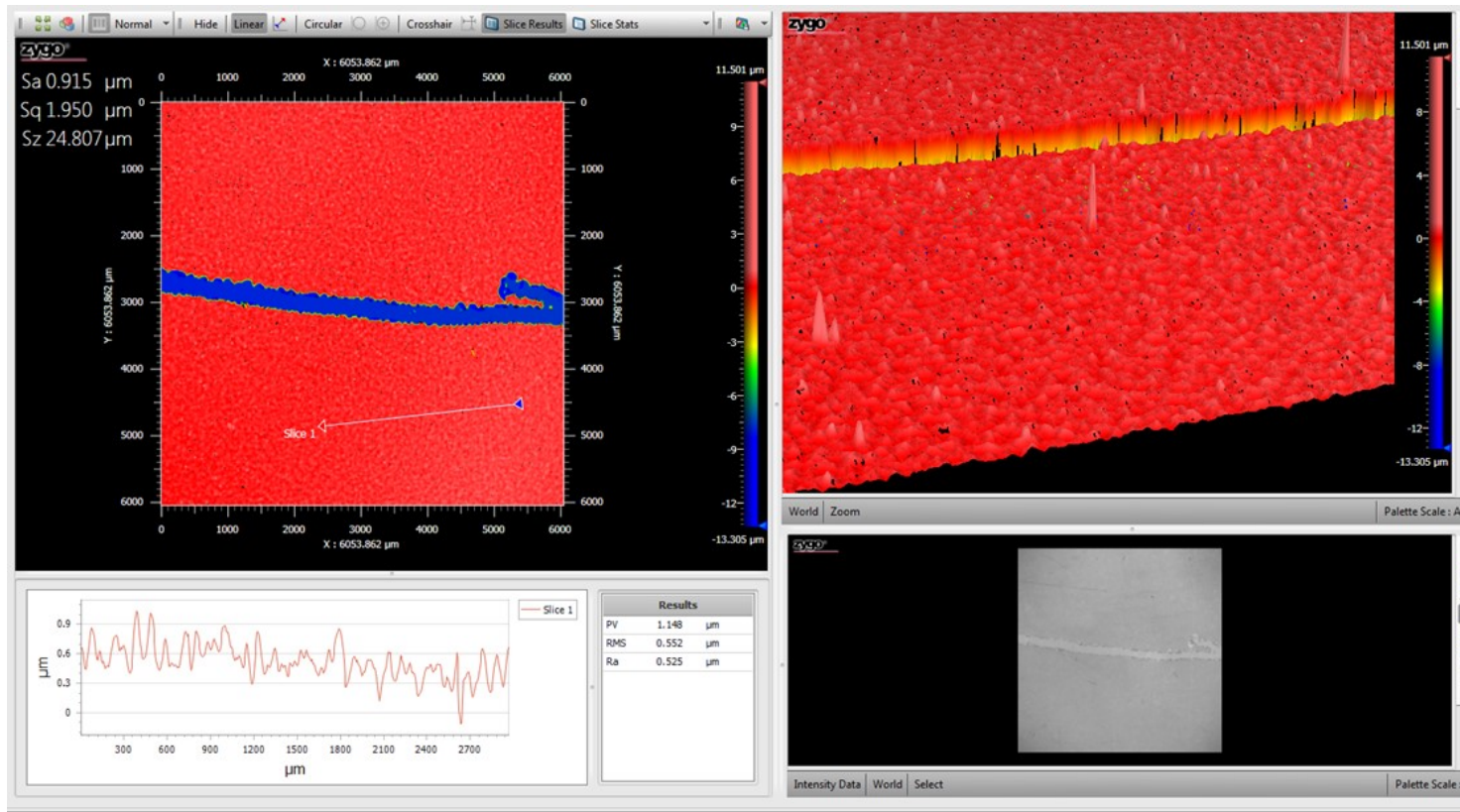


Figure S6: Zygo film surface roughness measurement for Ferro 221 series coating on Corning EAGLE XG Glass with no copper glass-ceramic (CGC) powder or CGC powder extract, flash cured at 150 °C.

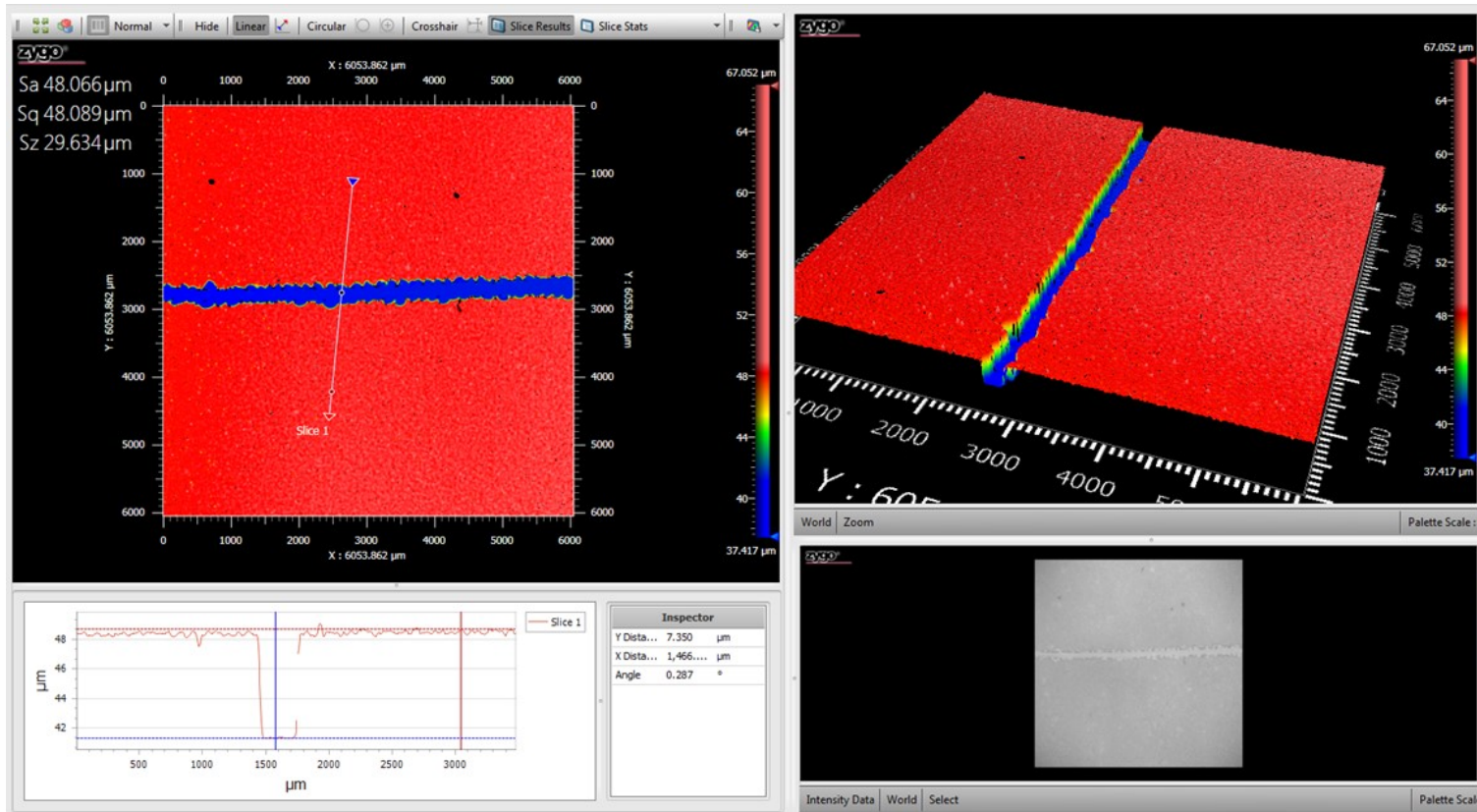


Figure S7: Zygo film thickness measurement for Ferro 221 glass coating on Corning EAGLE XG Glass dosed with copper glass-ceramic (CGC) powder extracts with triethyl phosphite (TEP) ($[\text{Cu}]_{\text{Tot}} = 600\text{ppm}$) and cured at 120 $^{\circ}\text{C}$.

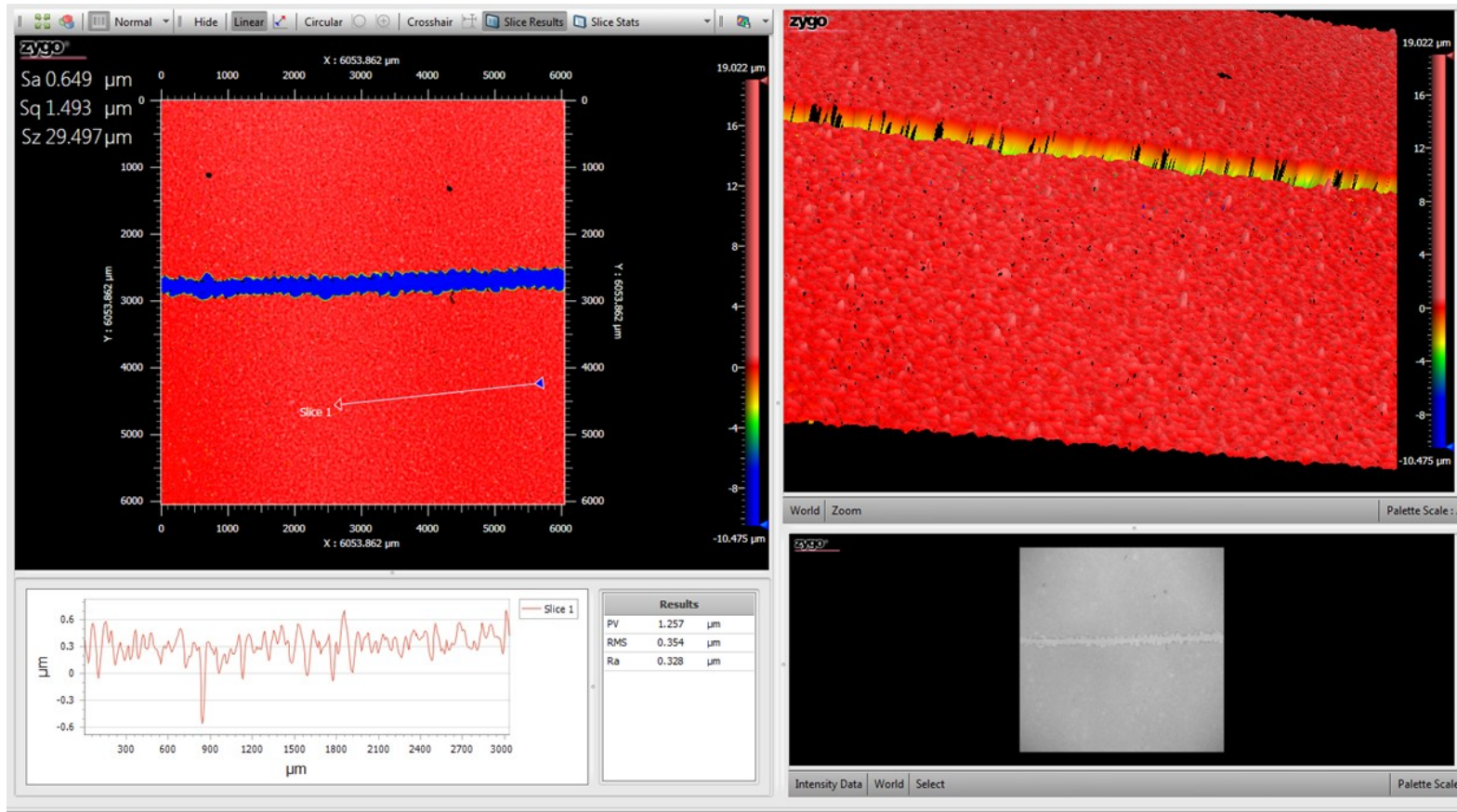


Figure S8: Zygo film surface roughness measurement for Ferro 221 glass coating on Corning EAGLE XG Glass dosed with copper glass-ceramic (CGC) powder extracts with triethyl phosphite (TEP) ($[Cu]_{Tot} = 600\text{ppm}$) and cured at 120 °C.

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