

## Supplementary Information

### ***Functional polysaccharide-coated SPIONs for in vitro mRNA delivery in breast cancer cells***

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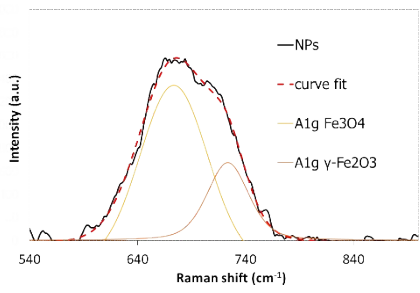
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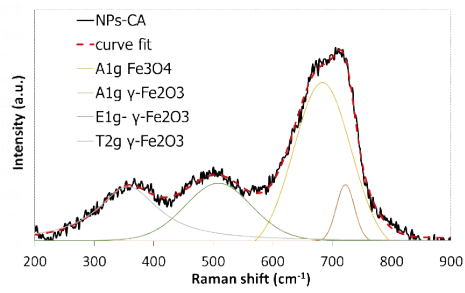
\* *Corresponding author.* (Assist. Professor. C. Chatzidoukas)

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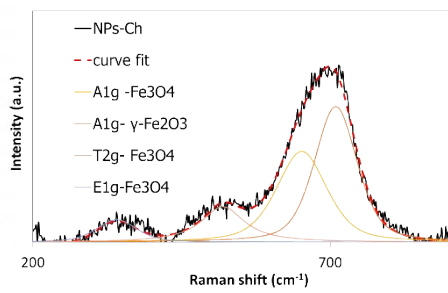
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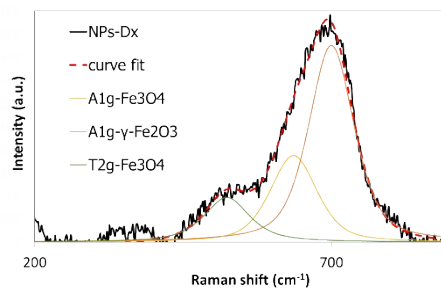
(a)



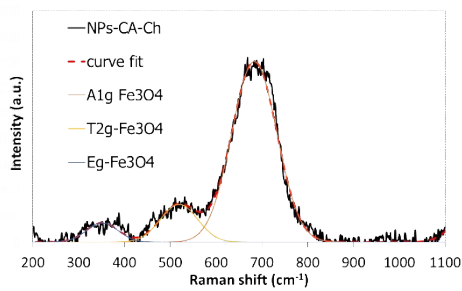
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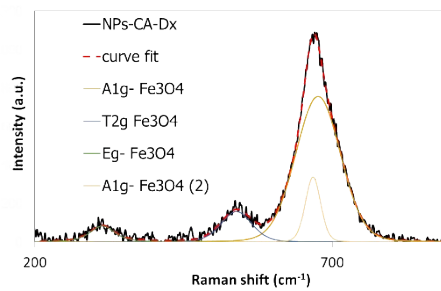
(c)



(d)

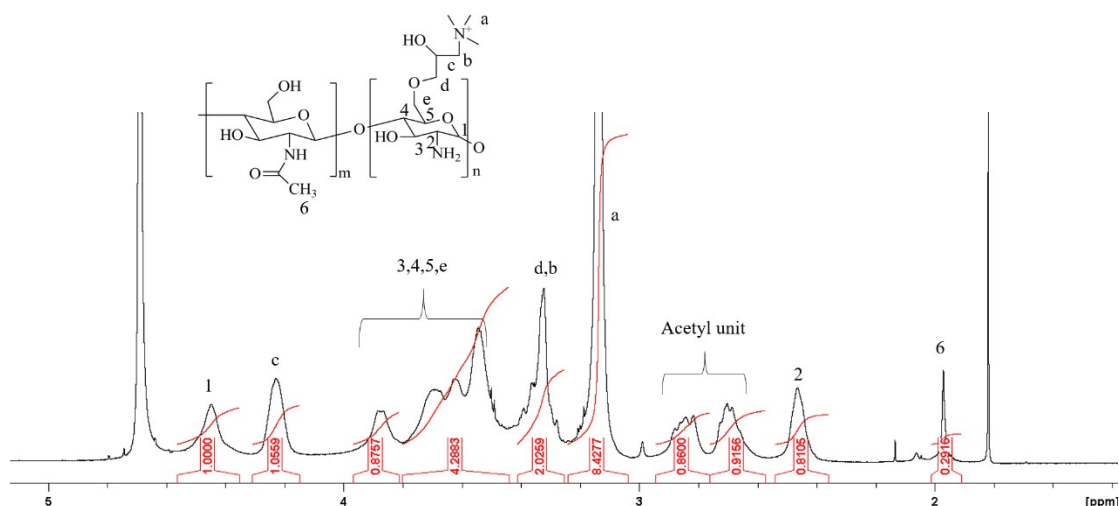


(e)



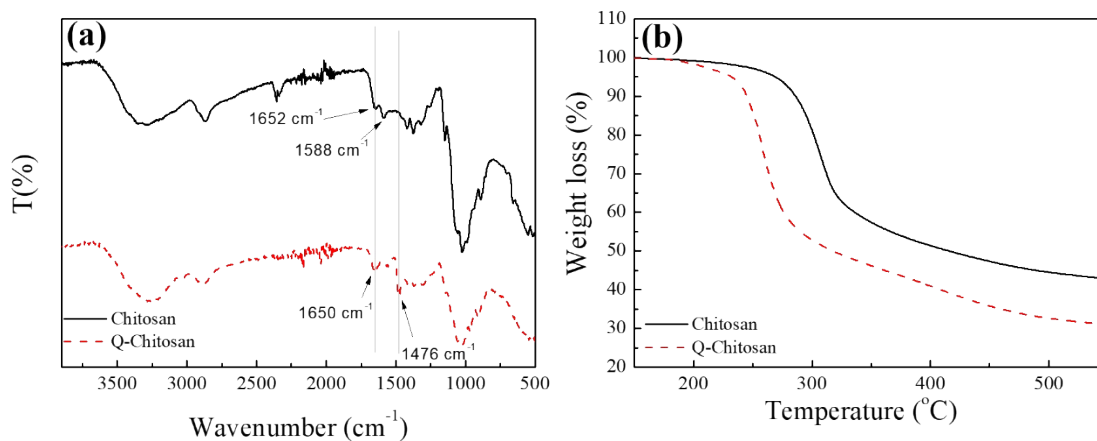
(f)

**Fig. S1:** Raman spectra and the deconvolution curves attributed to magnetite and maghemite for the (a) NPs, (b) NPs-CA, (c) NPs-Ch, (d) NPs-Dx, (e) NPs-CA-Ch and (f) NPs-CA-Dx samples.



**Fig. S2:**  $^1\text{H}$  NMR spectrum of Q-chitosan in  $\text{D}_2\text{O}$ .

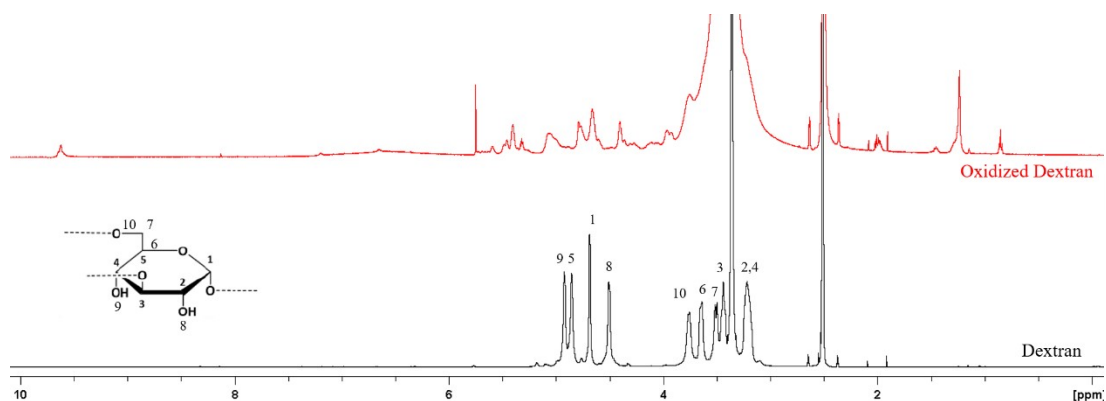
The appearance of the characteristic chemical shift at 3.15 ppm in the  $^1\text{H}$  NMR spectrum of Q-chitosan, assigned to the protons of the newly incorporated trimethyl ammonium groups (Fig. S2), verified the successful quaternization of the polymer.



**Fig. S3:** (a) FTIR spectra and (b) TGA curves of chitosan (black solid line) and Q-chitosan (red dashed line).

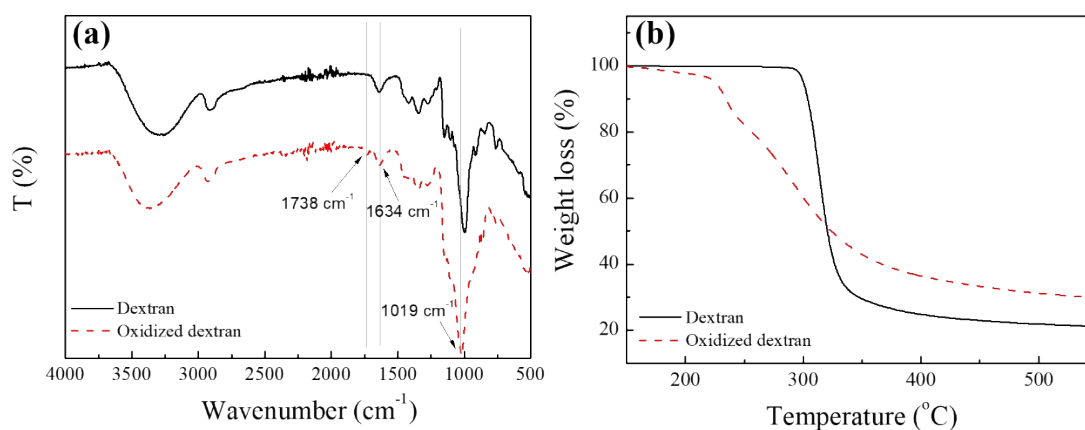
The ATR-FTIR spectra of chitosan (black solid line) before quaternization, showed the vibration band at 1588  $\text{cm}^{-1}$ , attributed to the amino groups and the shoulder at 1652  $\text{cm}^{-1}$ , attributed to the amide bonds of the chitin units of chitosan [i]. After quaternization (Q-chitosan, red dashed line), a strong vibration band at 1476  $\text{cm}^{-1}$ , which corresponds to the C-H bonds of the newly incorporated trimethyl ammonium groups, appeared, confirming the

successful quaternization of the biopolymer. Moreover, a peak at  $1650\text{ cm}^{-1}$ , which corresponds to the primary amine groups of chitosan, which were not modified, was evident.



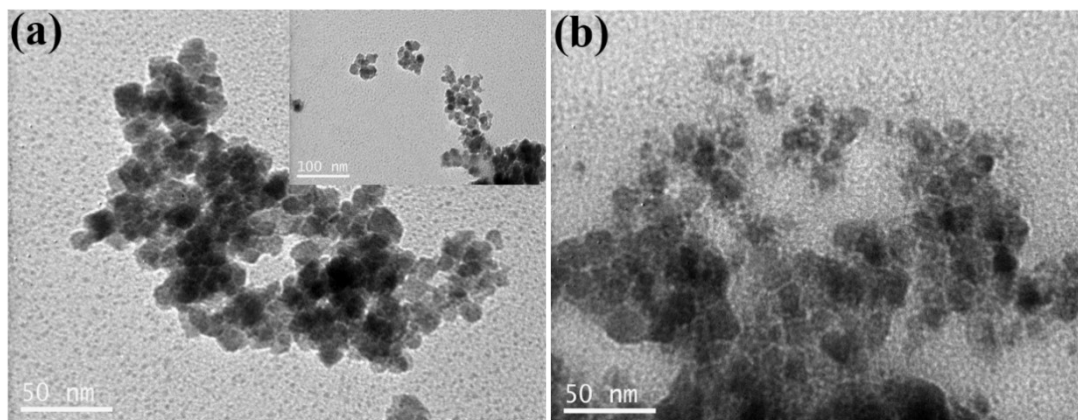
**Fig. S4:**  $^1\text{H}$  NMR spectra of dextran and Ox-dextran in  $\text{d}_6$ -DMSO.

The appearance of a new peak at 9.3 ppm in the  $^1\text{H}$  NMR spectrum of Ox-dextran denoted the successful modification of the polymer.

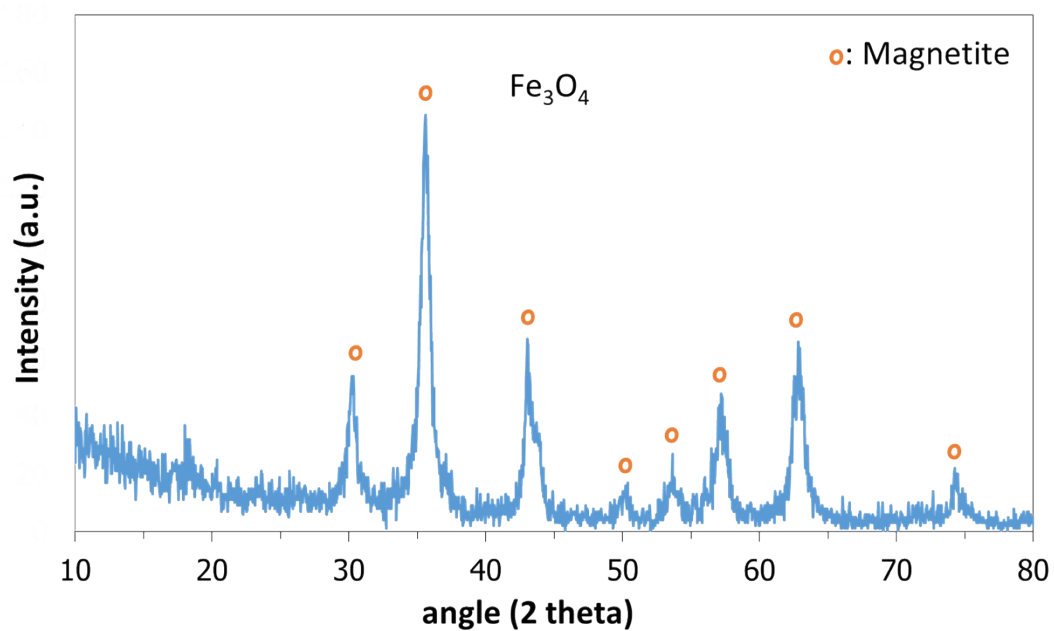


**Fig. S5:** (a) FTIR spectra and (b) TGA curves of dextran (black solid line) and Ox-dextran (red dashed line).

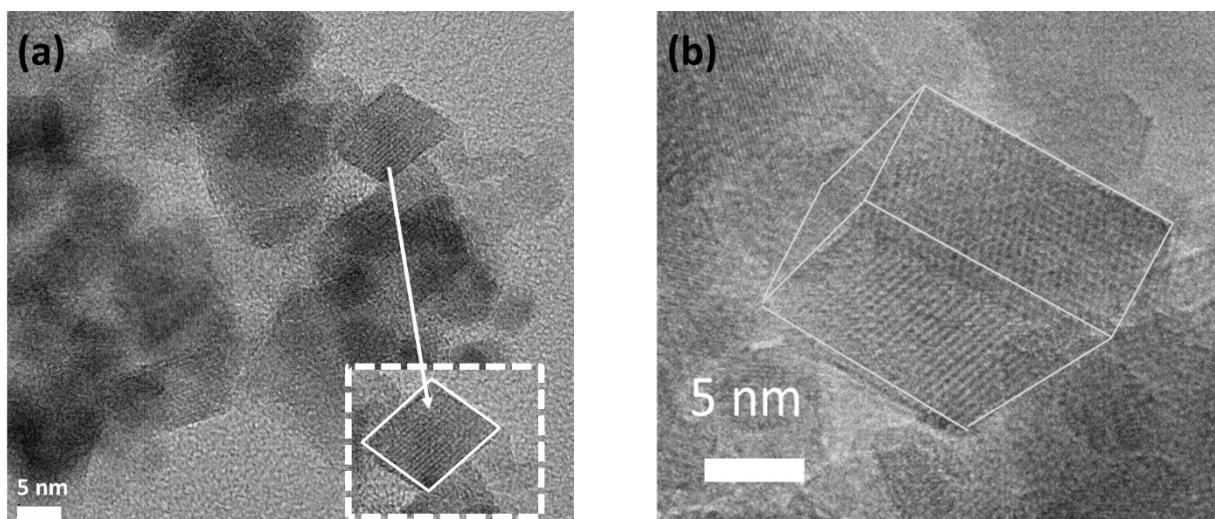
The ATR-FTIR spectrum of Ox-dextran showed the bands at  $1019$  and  $1634\text{ cm}^{-1}$ , which were attributed to the vibrations of the C-O bonds and a characteristic vibration band of the carbonyl bond of the aldehyde group at  $1738\text{ cm}^{-1}$ , verifying the successful oxidation of the polymer.



**Fig. S6:** TEM images of the (a) NPs-CA-Ch and (b) NPs-CA-Dx samples.

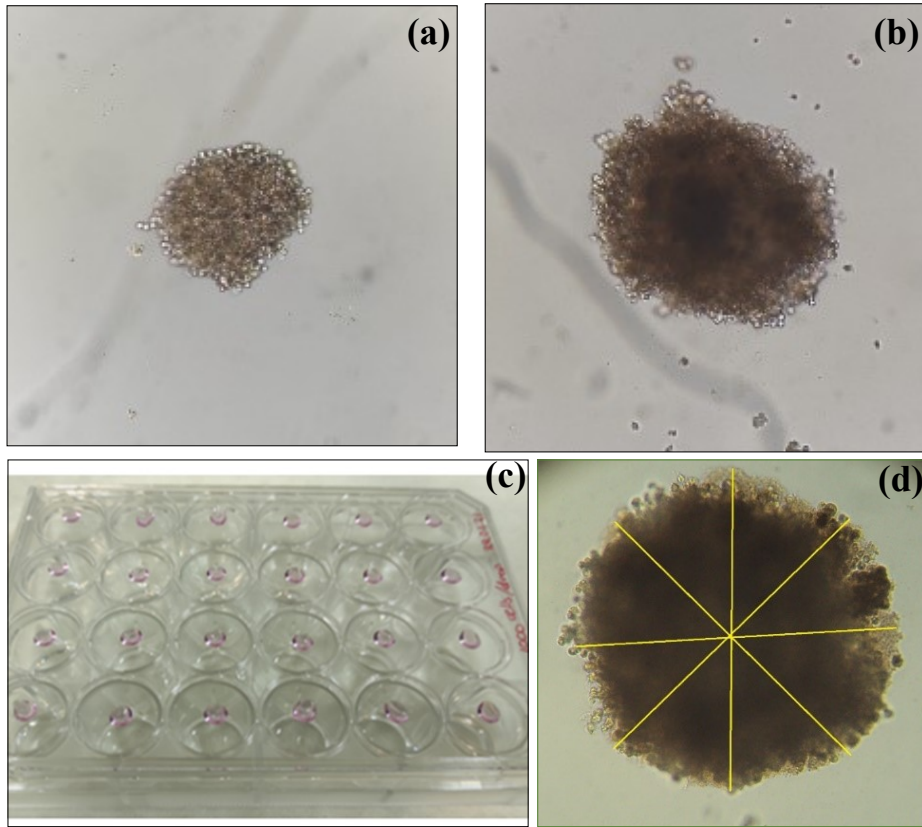


**Fig. S7:** XRD diagram of the bare  $\text{Fe}_3\text{O}_4$  SPIONs.

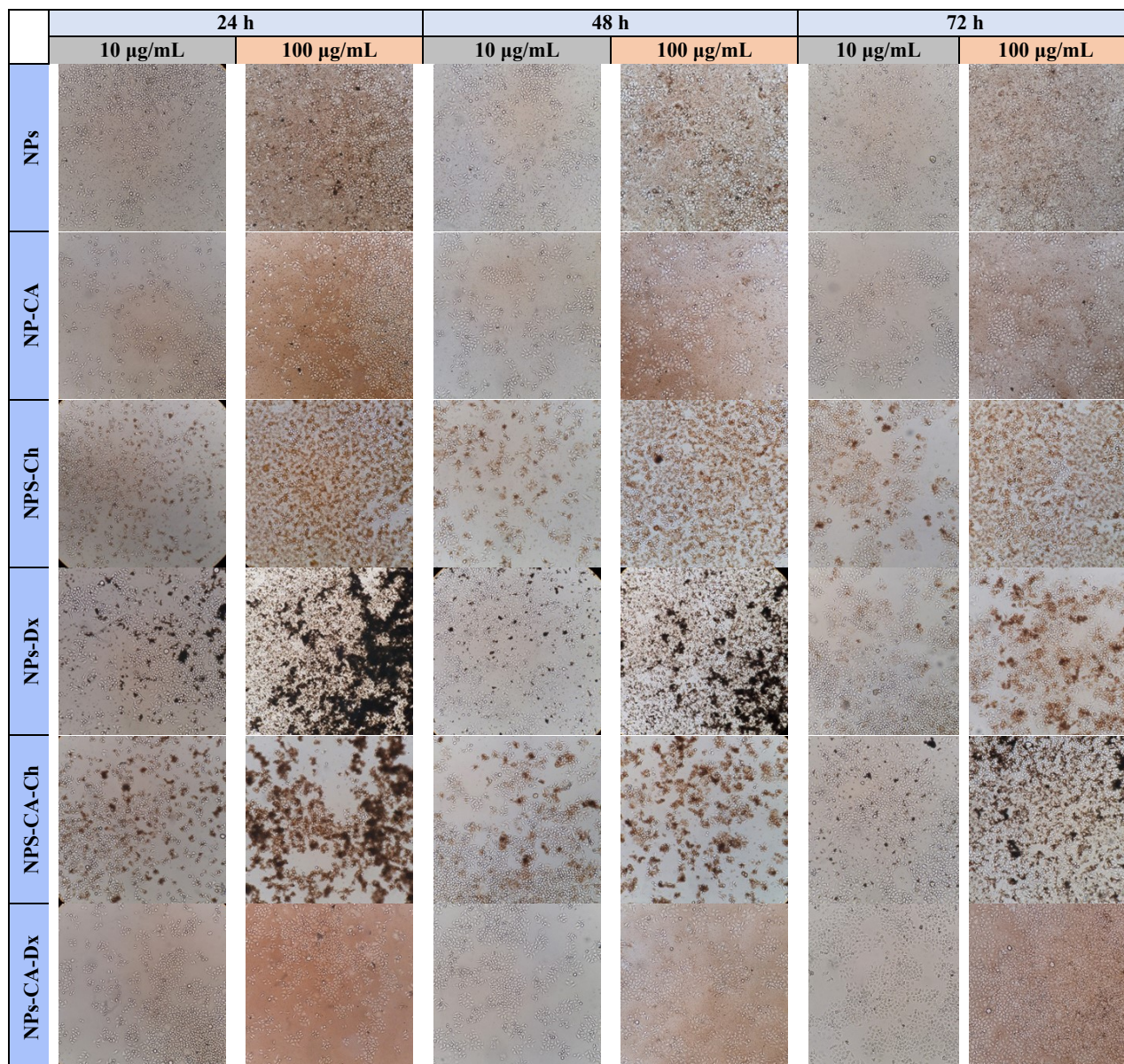


**Fig. S8:** (a) TEM image depicting agglomerates of NPs; the dashed line image shows a primary (single) particle defining with white outline the particle perimeter, (b) a high-resolution TEM image of a primary particle, defining with white outline the crystal planes of the cubic monocrystalline NP; the superparamagnetic NPs are monocrystalline

Fig. S8a illustrates the agglomerates of NPs as observed in TEM. In Fig. S8b, a primary cubic nanoparticle is depicted, revealing its monocrystalline nature through the orientation of planes in the three-dimensional structure. This observation aligns with the anticipated expectation, as SPIONs are known to be monocrystalline.

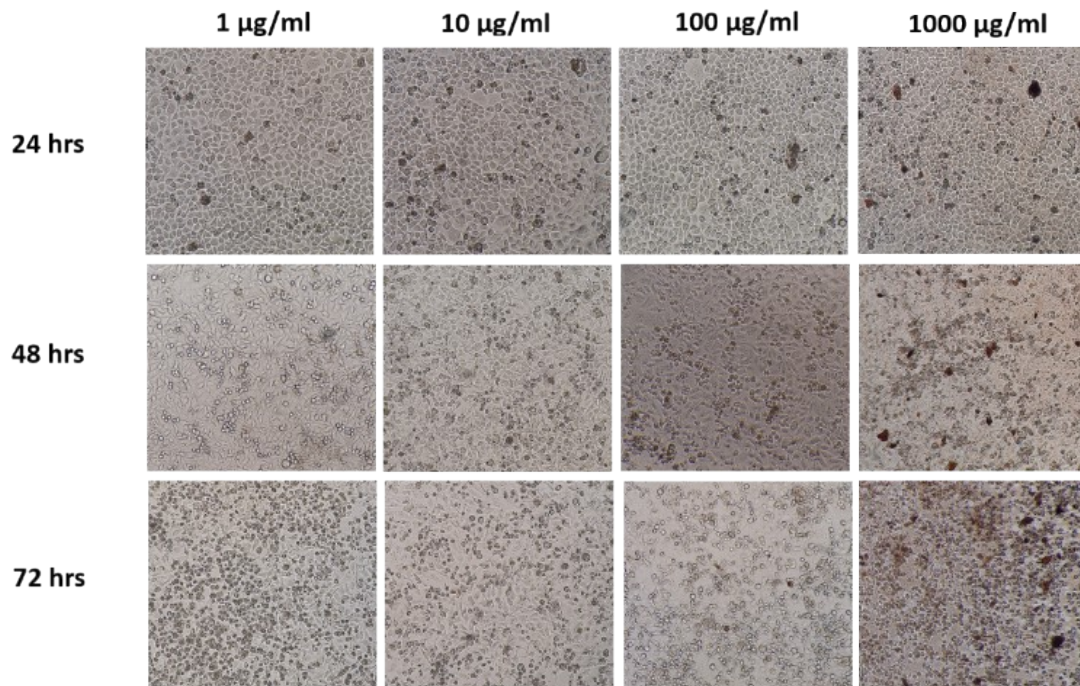


**Fig. S9:** (a), (b) Cell spheroid formation on days 3 and 12. (c) The hanging drop method. (d) Schematic representation of the diameter of each aggregate obtained as an average of 5 measurements.

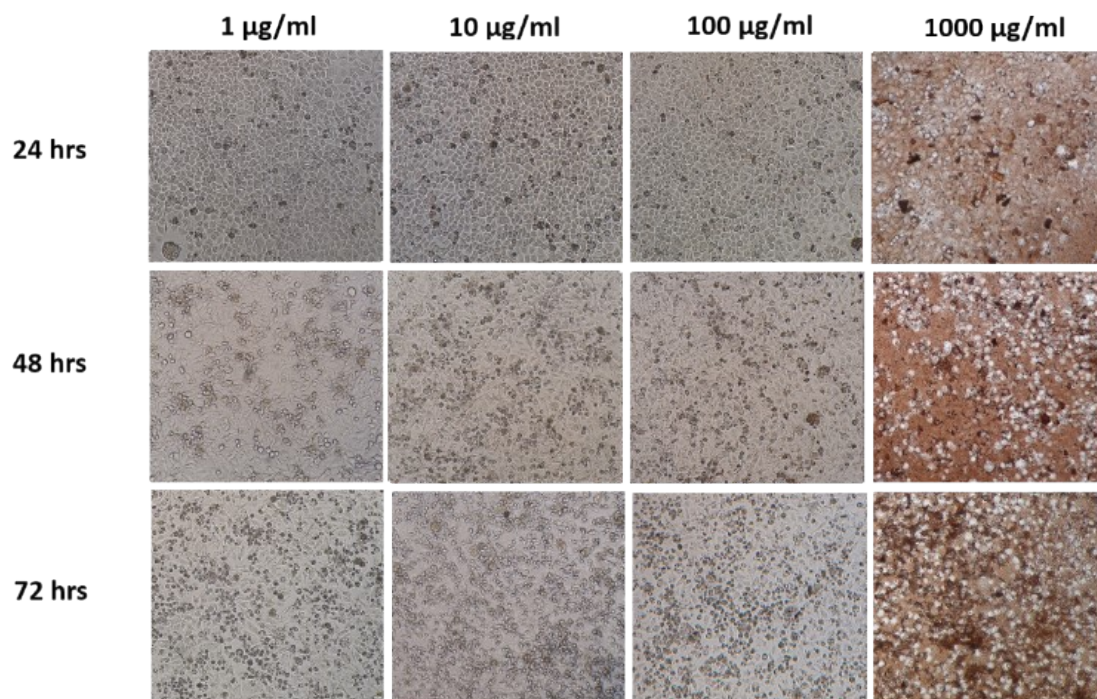


**Fig. S10:** Representative micrographs of cell adhesion of T47D cells following their treatment with SPIONs (photographs obtained using a LEICA inverted microscope, with a 10x phase contrast. Images were captured at several time points).

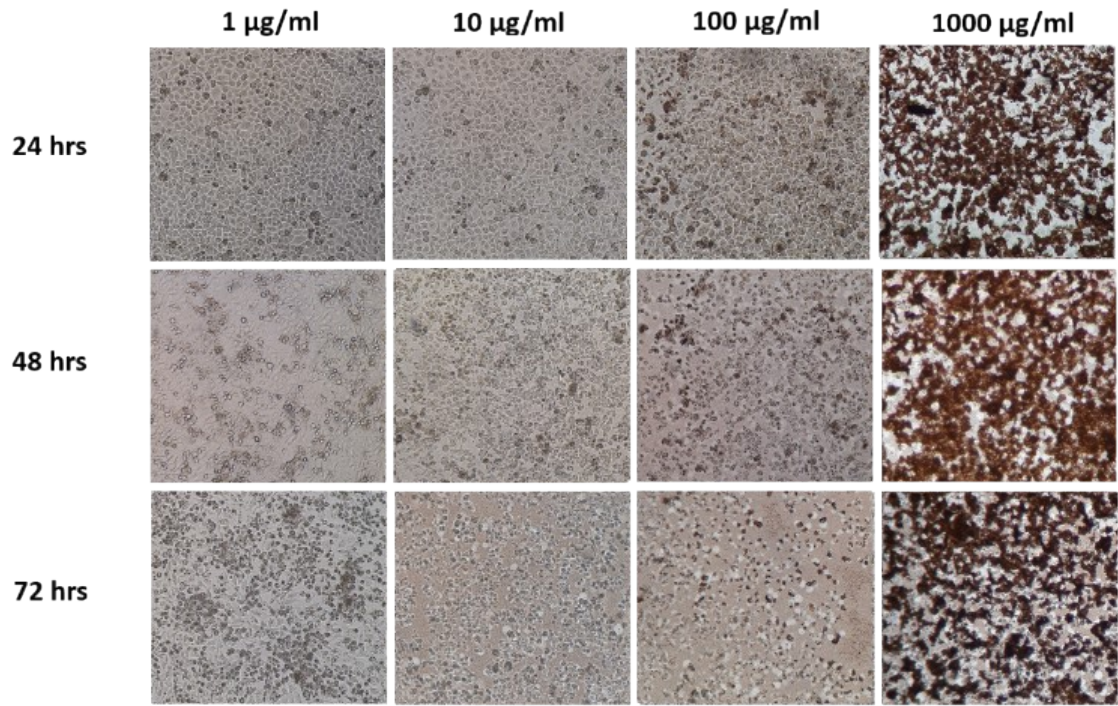




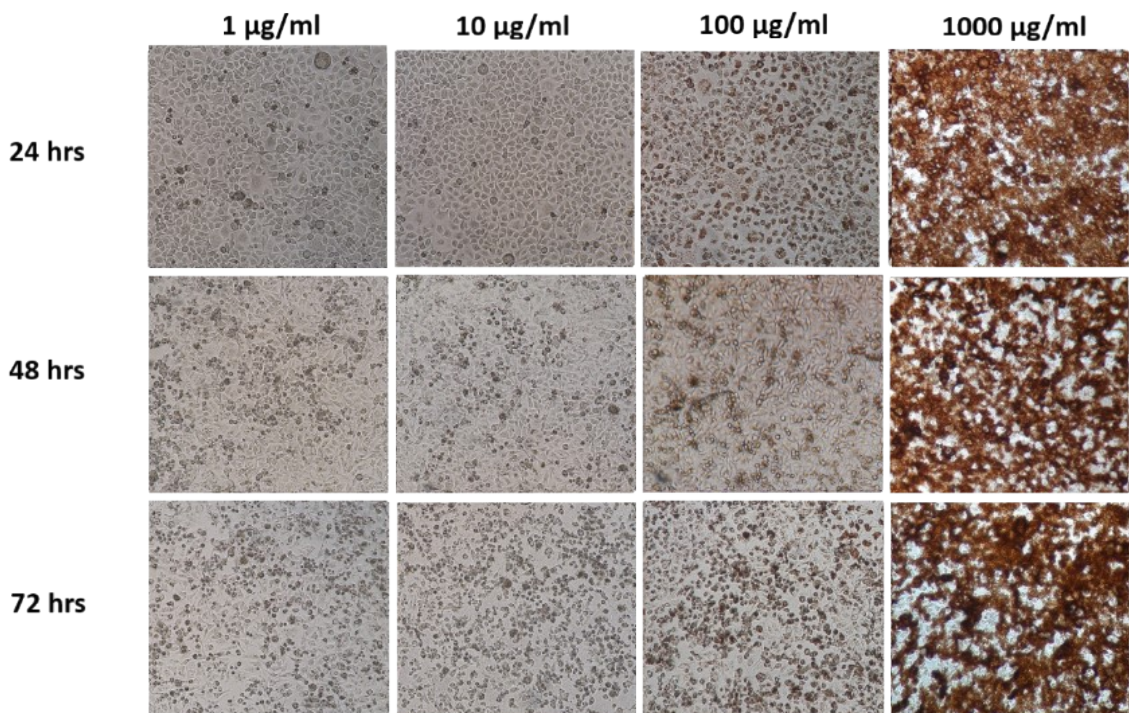
**NPs**



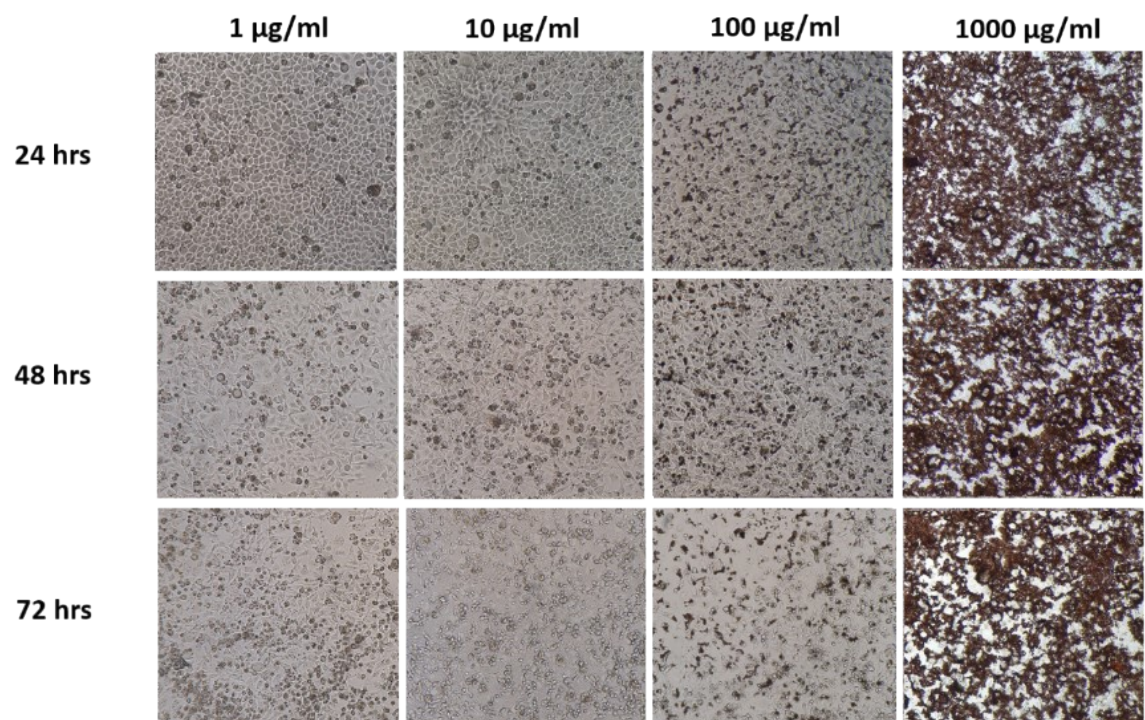
**NPs-CA**



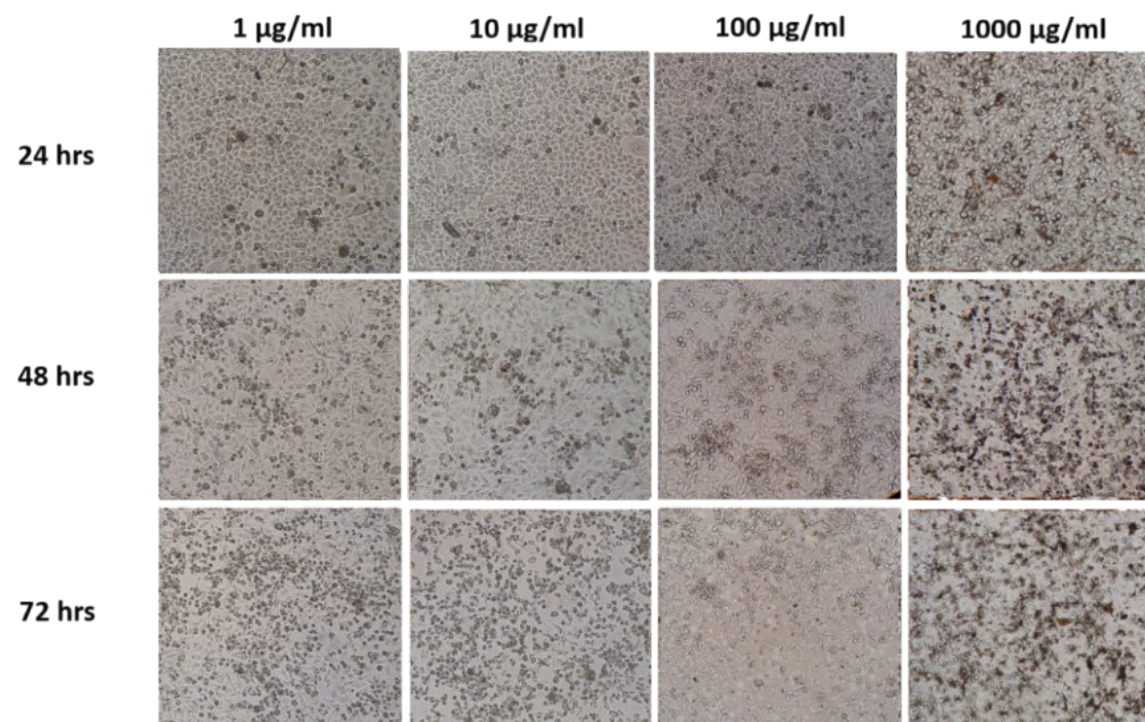
**NPs-Ch**



**NPs-CA-Ch**

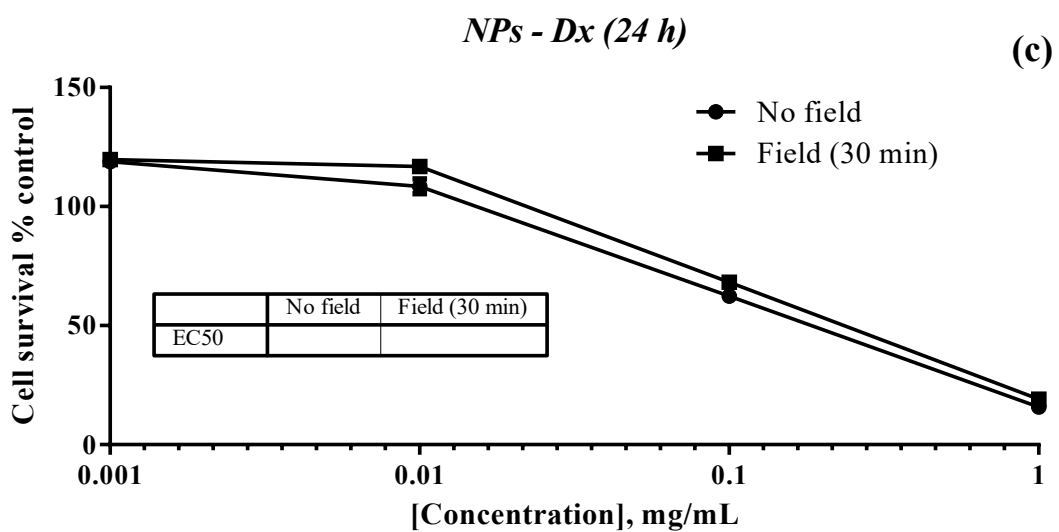
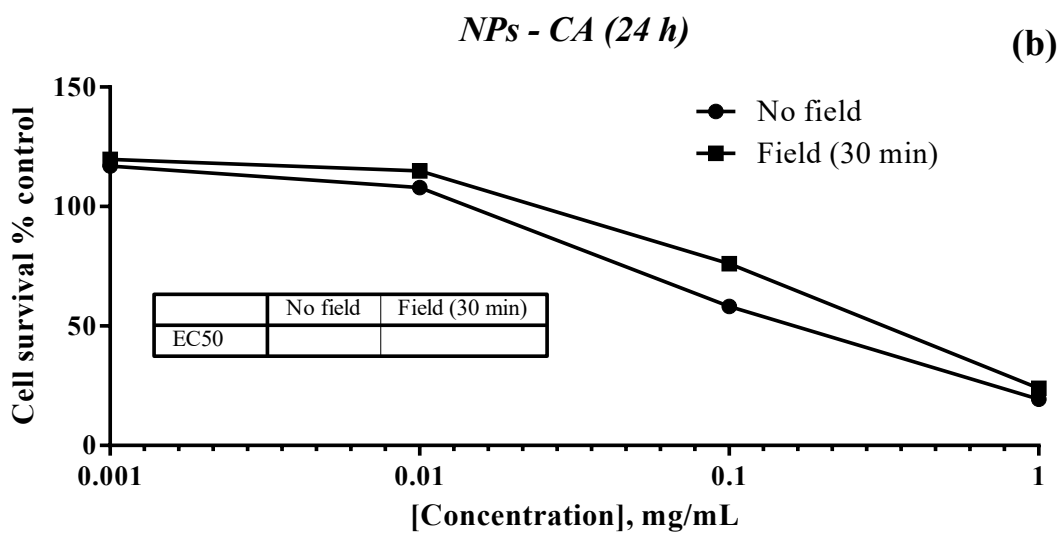
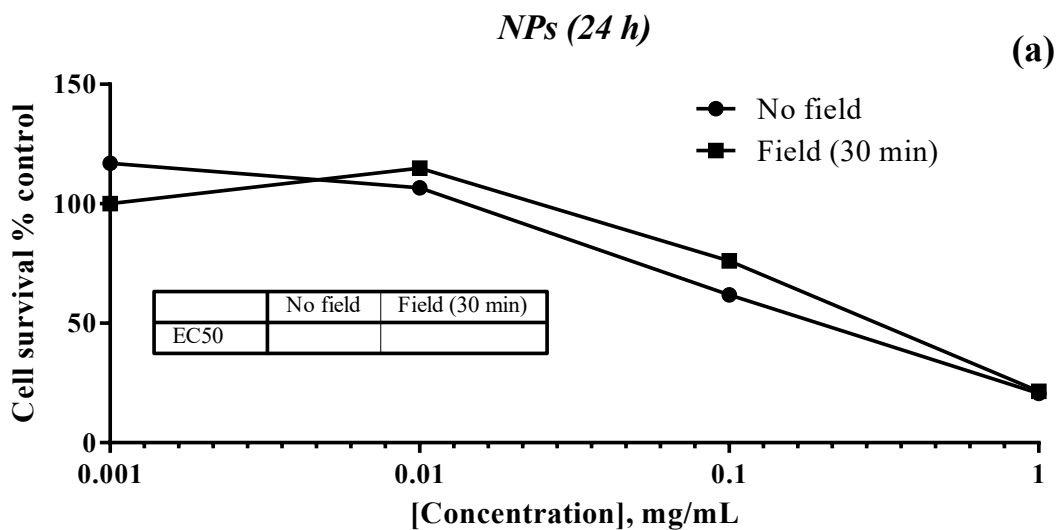


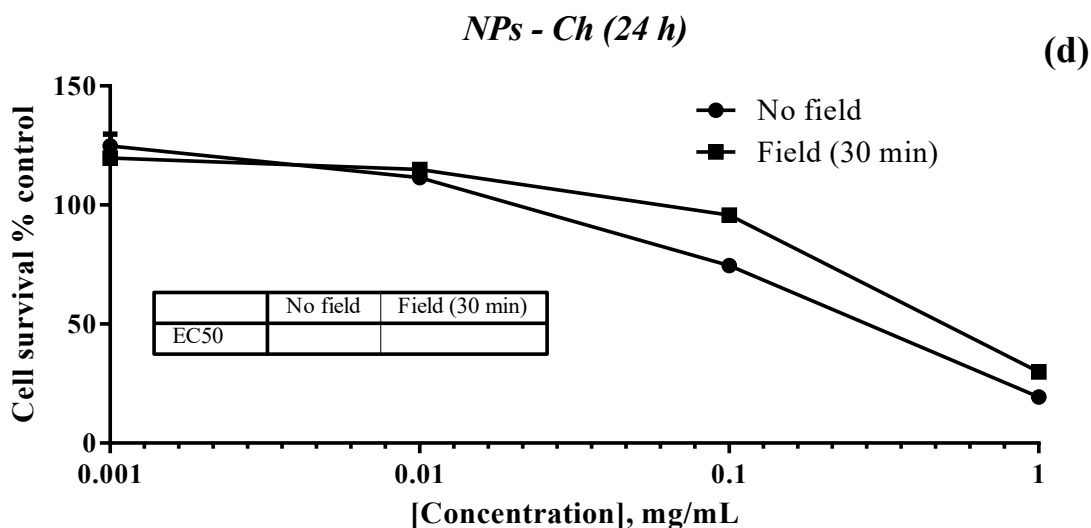
**NPs-Dx**



**NPs-CA-Dx**

**Fig. S11:** Representative micrographs of cell morphology of T47D cells following their treatment with the SPIONs (photographs obtained using a LEICA inverted microscope, with a 10x phase contrast. Images were captured at several time points).

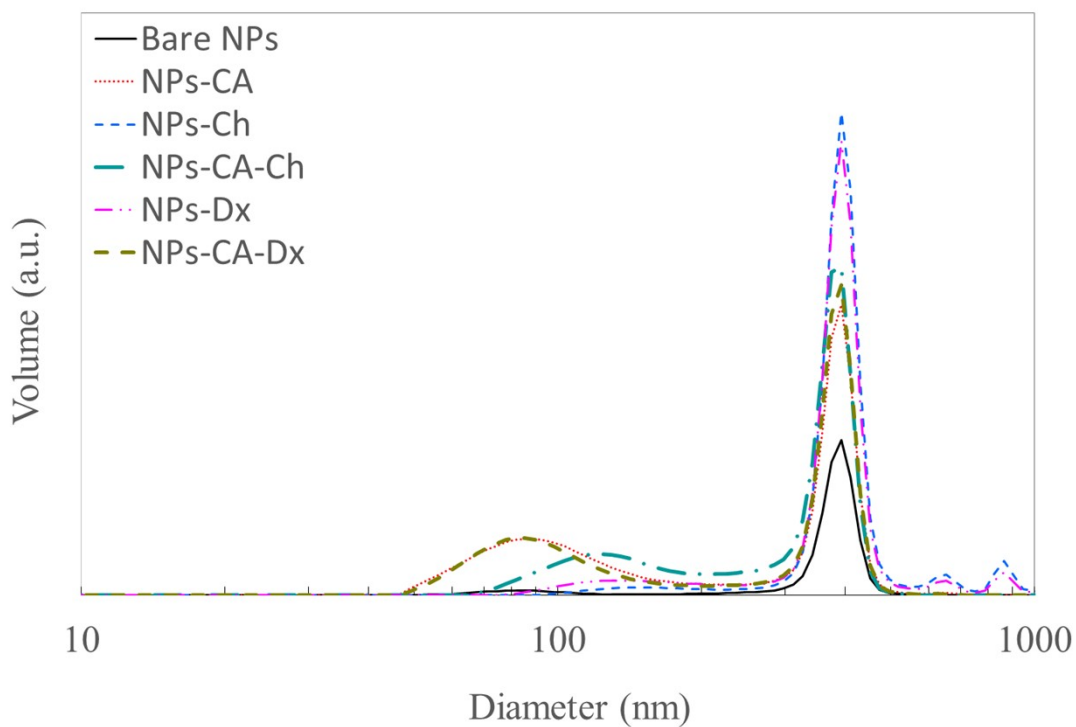




**Fig. S12:** Percentage change of cell survival of the T47D cells (2D monolayers), following treatment with various concentrations (1-1000  $\mu$ M) of a) NPs, b) NPs-CA, c) NPs-Dx, and d) NPs-Ch for 24 h in the presence and absence of a magnetic field.



**Fig. S13.** Gel electrophoresis in 1% w/v agarose gel-to assess the efficient binding of GFP-mRNA onto the magnetic nanoparticles. Lane 1: NPs-Ch<sup>GFPmRNA</sup> (ratio: 1/0.5); Lane 2: NPs-Dx<sup>GFPmRNA</sup> (ratio: 1/0.5); Lane 3: NPs-Dx<sup>GFPmRNA</sup> (ratio: 1/3); Lane 4: NPs-Ch<sup>GFPmRNA</sup> (ratio: 1/3); Lane 5: NPs-CA-Ch<sup>GFPmRNA</sup> (ratio: 1/0.5); Lane 6: NPs-CA-Ch<sup>GFPmRNA</sup> (ratio: 1/1.5); Lane 7: NPs-CA-Ch<sup>GFPmRNA</sup> (ratio: 1/3); Lane 8: NPs-CA-Ch<sup>GFPmRNA</sup> (ratio: 1/5); Lane 9: NPs-CA-Dx<sup>GFPmRNA</sup> (ratio: 1/0.5); Lane 10: NPs-CA-Dx<sup>GFPmRNA</sup> (ratio: 1/1.5); Lane 11: NPs-CA-Dx<sup>GFPmRNA</sup> (ratio: 1/3); Lane 12: NPs-CA-Dx<sup>GFPmRNA</sup> (ratio: 1/5).



**Figure S14:** DLS Volume measurements of the bare NPs (black solid line), NPs-CA (red dotted line), NPs-Ch (blue short-dashed line), NPs-CA-Ch (green dashed-dotted line), NPs-Dx (magenta dashed dotted-dotted line) and NPs-CA-Dx (dark yellow dashed line).

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i Wan, Y.; Creber, K.A.M.; Peppley, B.; Bui, V.T. Ionic conductivity of chitosan membranes. *Polymer (Guildf)*. 2003, 44, 1057–1065, doi:10.1016/S0032-3861(02)00881-9.