

Supplementary materials

New pyrazole-pyridazine hybrids as selective COX-2 inhibitors: Design, synthesis, molecular docking, *in silico* studies and investigation of the anti-inflammatory potential by evaluation of TNF- α , IL-6, PGE-2 and NO in LPS-induced RAW264.7 macrophages

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Supplementary data

(Experimental of pharmacological activity studies and molecular docking , in addition to IR, ^1H NMR and ^{13}C NMR spectra of new compounds)

1. Experimental

1.1 Pharamcological activity studies

1.1.1 Cell viability assay

RAW264.7 cells (6×10^3 cells/per) were placed into a 96-well plate and kept for 24 h. Five different concentrations of the investigated compounds were used (100, 25, 6.3, 1.6, and 0.4 $\mu\text{M/L}$). The cells were pretreated with the different concentrations for 1 h and then incubated with LPS (0.5 $\mu\text{g/ml}$) for 24 h. To each well, MTT solution (5 mg/ml) was added and then incubated for 4 h at 37 °C. The media containing MTT was removed, then treated with 150 μL of DMSO. The absorbance was detected by a microplate reader at 500-600 nm .

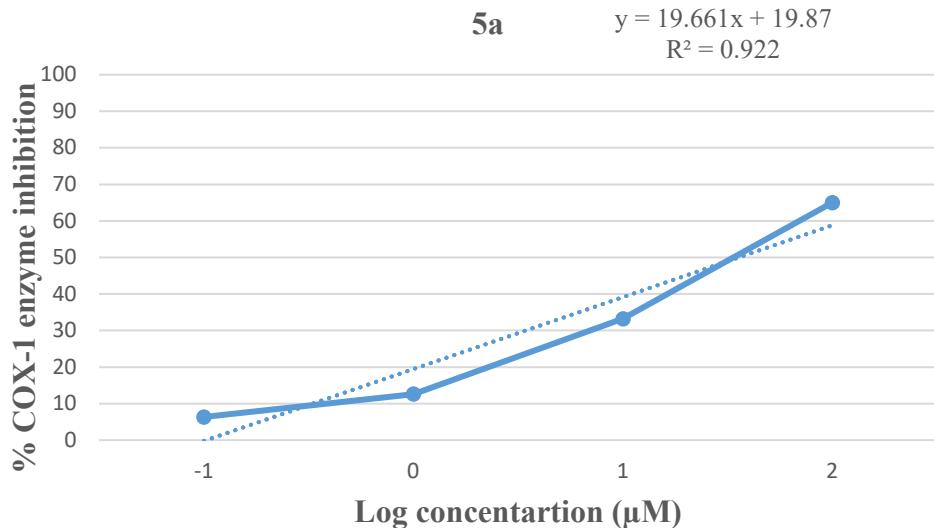
***Cytotoxicity results**

Code	IC₅₀ (μM)	SD (±)
5a	246	13
5b	61.6	3.2
5c	100	5.2
5d	158	8.2
5e	265	14
5f	328	17
6a	311	16
6b	138	7.2
6c	97.5	5.1
6d	212	11
6e	265	14
6f	169	8.8
Celecoxib	201	10

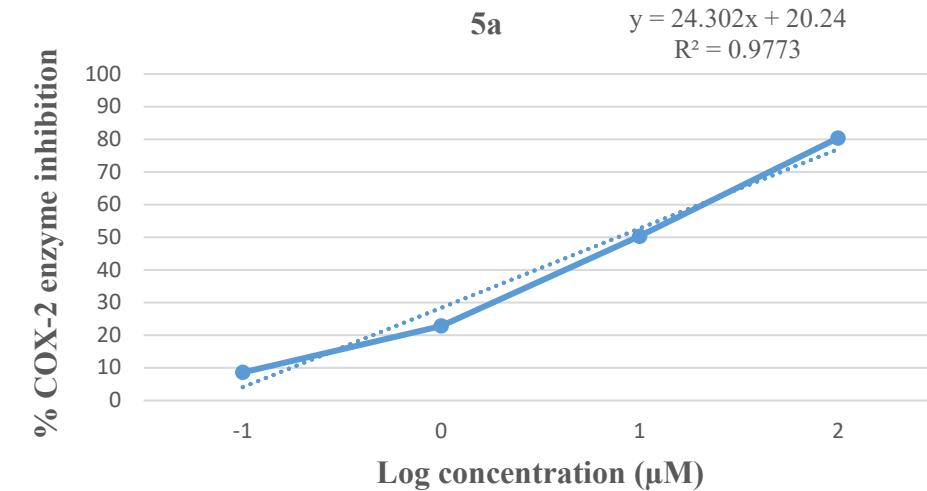
1.1.2 *In vitro* COX-1 and COX-2 inhibitory assay

In this assay, celecoxib and indomethacin were used as reference medications to investigate the ability of the new compounds to suppress human COX-1 and COX-2 using ten folds serial dilutions (1, 0.1, 0.01, 0.001 µg/mL) (**Table 1**). This was achieved using human COX-1 and COX-2 inhibitor screening kit supplied by Cayman chemicals (catalog numbers 701,070 and 701080, respectively, Ann Arbor, MI, USA). Dimethylsulfoxide (DMSO) was used to solubilize the new derivatives. Briefly, a mixture containing COX-1 or COX-2 enzyme (10 µL), heme (10 µL) and DMSO solutions of tested compounds samples (20 µL) was added to the supplied reaction buffer solution [160 µL, 0.1 M Tris–HCl, pH 8 containing 5 mM ethylenediamine tetra acetate (EDTA) and 2 mM phenol] then incubated for 10 min at 37 °C. Meanwhile, arachidonic acid was added (10 µL). To initiate the reaction, the final reaction mixture concentration was 100 µM. Later, stannous chloride (30 µL) was added to stop the COX reactions, then incubated at an ambient temperature for 5 min. Thereafter, quantification of PGF_{2α} produced in the samples by COX reactions was carried out by adopting an enzyme-linked immunosorbent assay (ELISA). The samples were transferred to a 96-well plate and then incubated at room temperature for 18 h. Any unbound reagent was removed by washing. Ellman's reagent (200 µL) was added and incubated for 60–90 min at room temperature. Finally, the plate was read by an ELISA plate reader, and the IC₅₀ values for inhibition of both COX-1 and COX-2 enzymes were determined.

COX-1

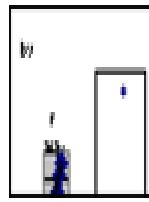
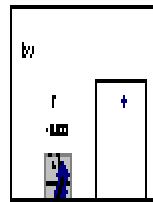


COX-2

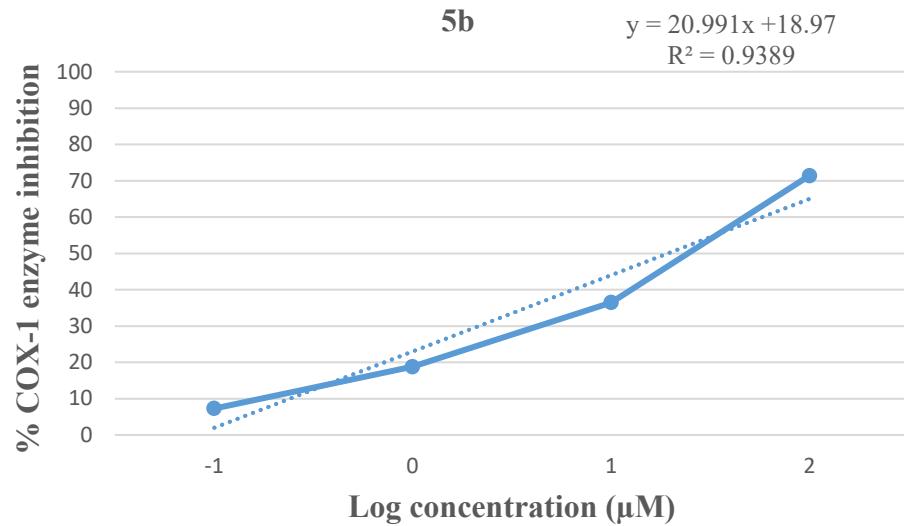


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5a	20.11	100	2	65	42.0402
		10	1	33.2	80.192
		1	0	12.6	10.938
	0.1	-1	6.33	112.343	
EC				0	120

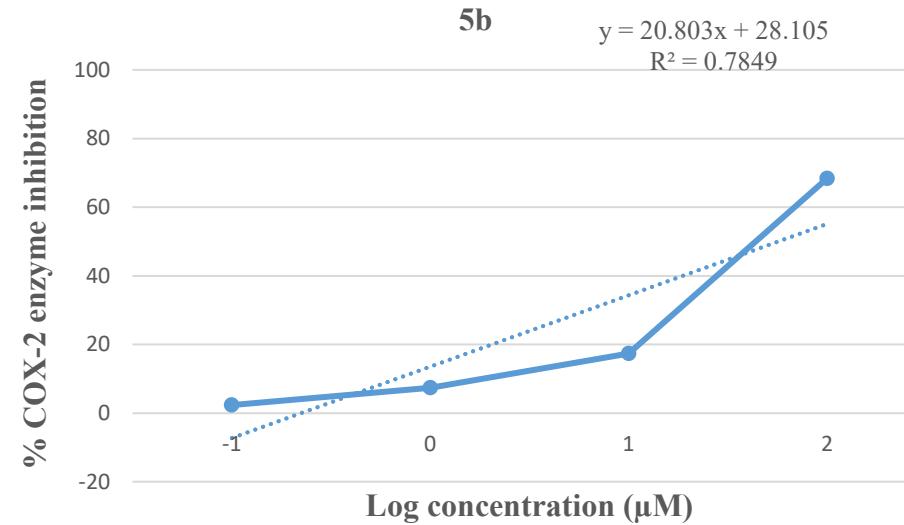
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5a	5.40	100	2	80.4	23.4623
		10	1	50.3	59.682
		1	0	92.6253	97.6658
	0.1	-1	8.56	109.727	
EC				0	120



COX-1

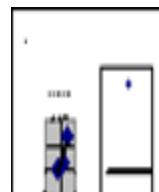
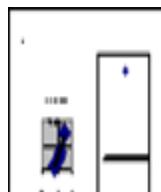


COX-2

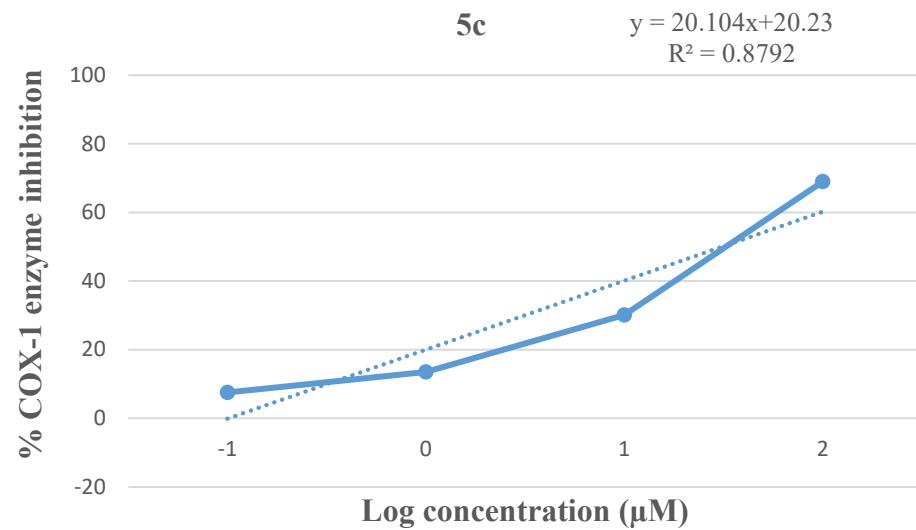


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5b	4.58	100	2	73.9	34.2874
		10	1	40.5	76.2196
		1	0	21.4	97.4377
	0.1	-1	8.96	111.203	
EC				0	120

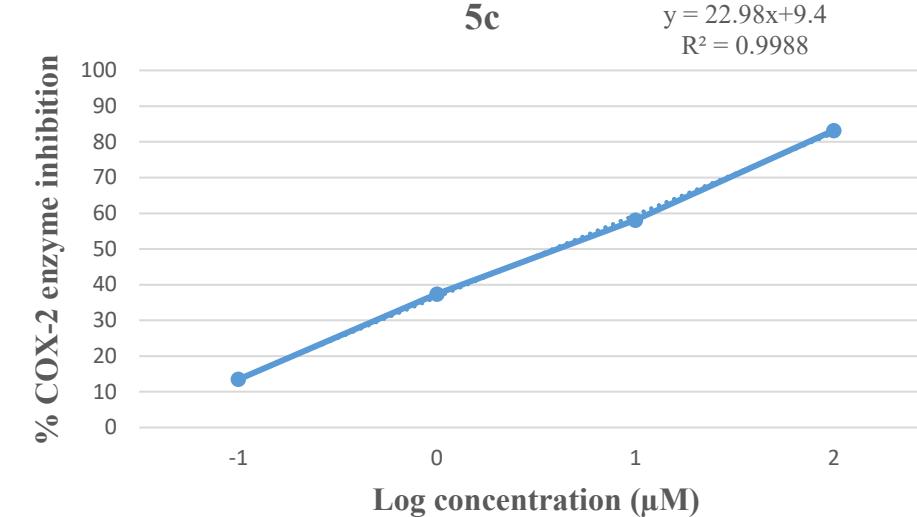
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5b	4.58	100	2	68.4	37.3638
		10	1	17.4	99.1659
		1	0	7.43	111.083
	0.1	-1	2.38	117.144	
EC				0	120



COX-1

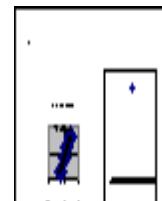
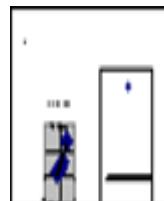


COX-2

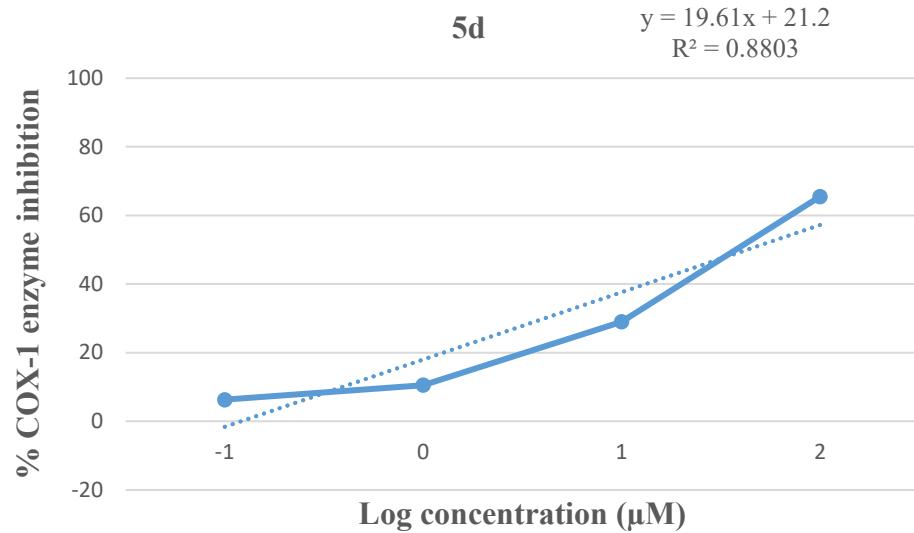


Code	IC_{50}	Conc.	Log	% Inhibition	K activity
5c	18.00	100	2	69	37.2277
		10	1	30.1	83.8644
		1	0	13.5	103.858
	0.1	-1	7.52	110.975	
EC			0	120	

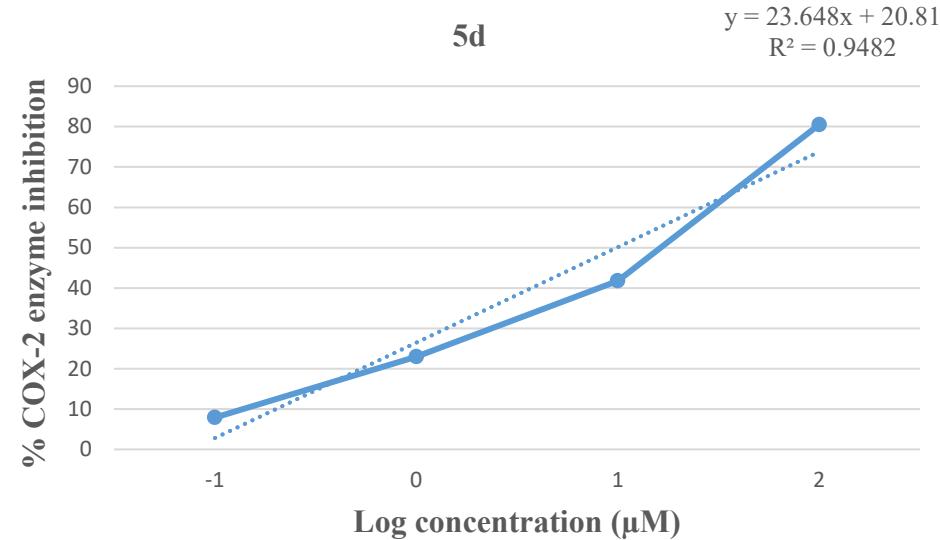
Code	IC_{50}	Conc.	Log	% Inhibition	K activity
5c	9.04	100	2	83.2	20.15
		10	1	58.1	50.309
		1	0	37.4	75.1395
	0.1	-1	13.5	103.846	
EC			0	120	



COX-1

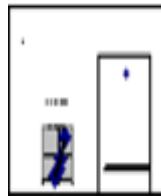
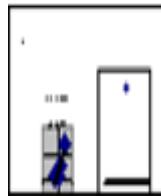


COX-2

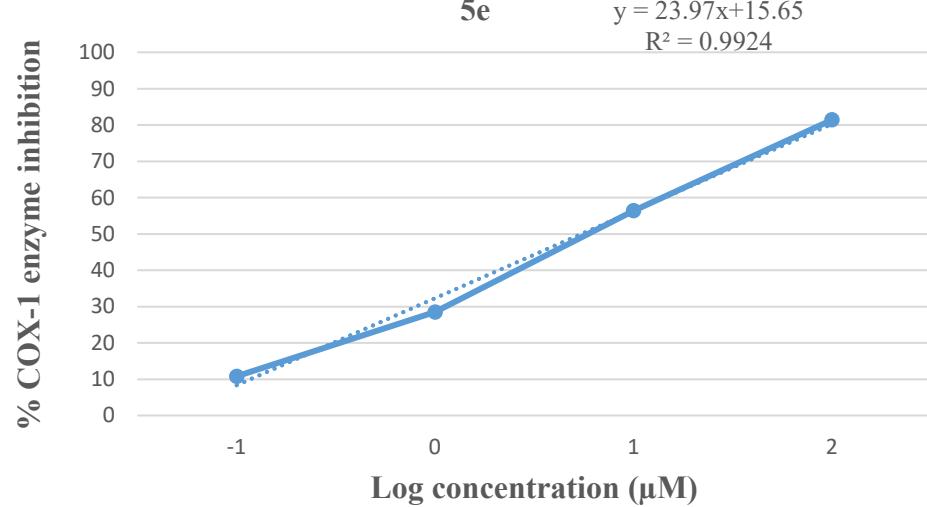


Code	IC_{50}	Conc.	Log	% Inhibition	K activity
5d	4.15	100	2	65.5	41.4281
		10	1	29	85.2445
		1	0	10.5	107.459
	0.1	-1	6.3	112.439	
EC				0	120

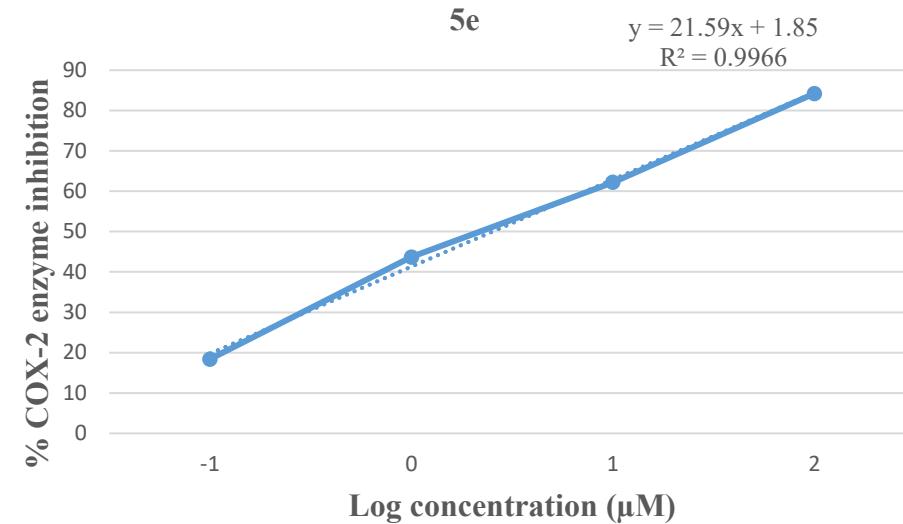
Code	IC_{50}	Conc.	Log	% Inhibition	K activity
5d	20.71	100	2	80.5	23.4143
		10	1	41.8	69.835
		1	0	23	92.3972
	0.1	7.94	35.2	110.471	
EC				0	120



COX-1

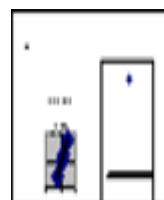
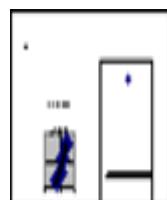


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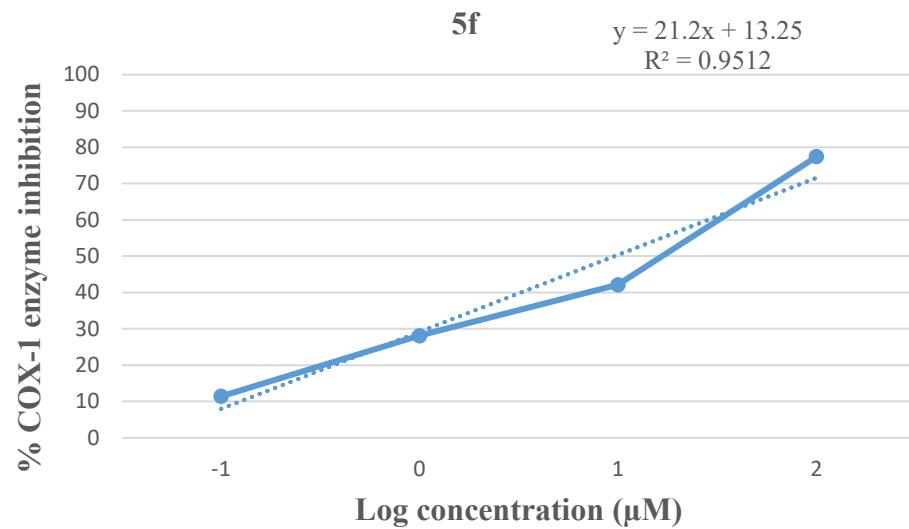


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5e	2.42	100	2	81.4	22.3342
		10	1	56.4	52.3372
		1	0	28.5	85.8326
		0.1	-1	10.8	107.003
EC				0	120

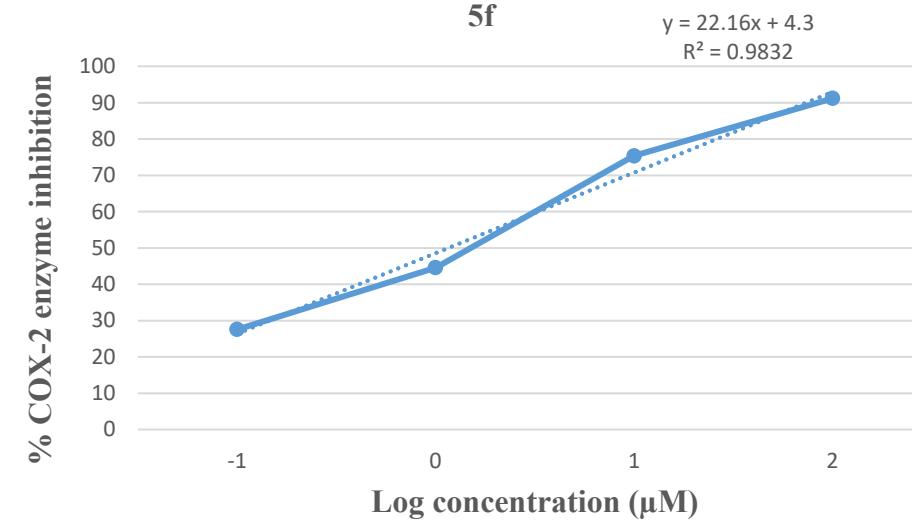
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5e	3.92	100	2	84.2	18.9019
		10	1	62.2	45.3885
		1	0	43.7	67.5428
		0.1	-1	18.4	97.9178
EC				0	120



COX-1

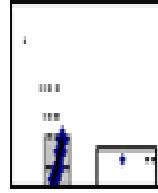
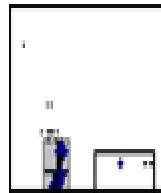


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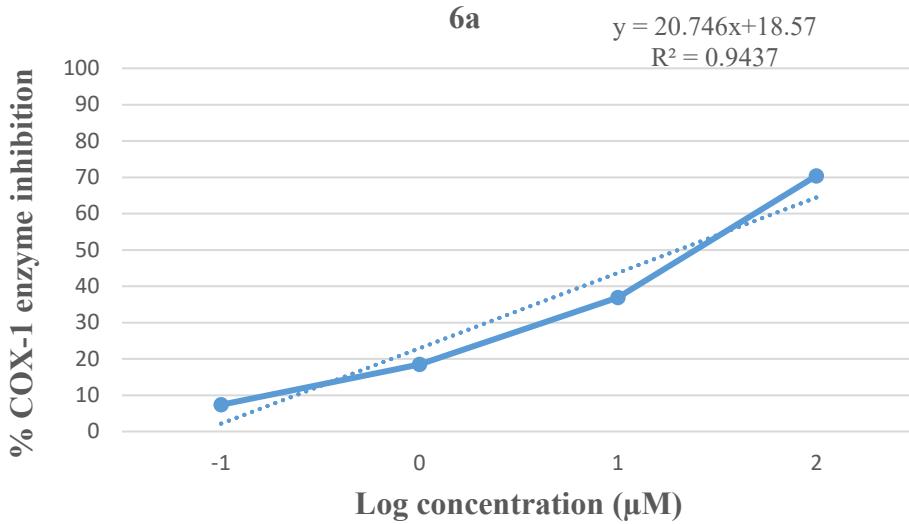


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
5f	14.38	100	2	77.4	27.0987
		10	1	42.1	69.4989
		1	12.6	28.1	86.2286
	0.1	-1	11.4	106.343	
EC			0	120	

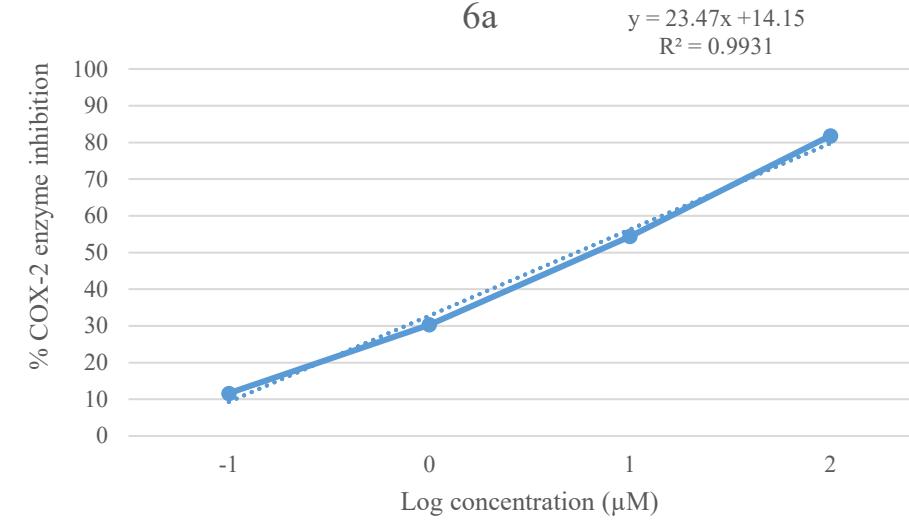
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5f	1.50	100	2	91.2	10.6091
		10	1	75.4	29.535
		1	0	44.6	66.4746
	0.1	-1	27.6	86.9367	
EC			0	120	



COX-1

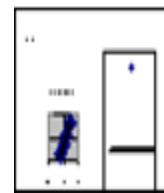
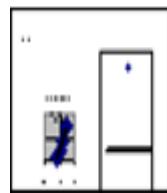


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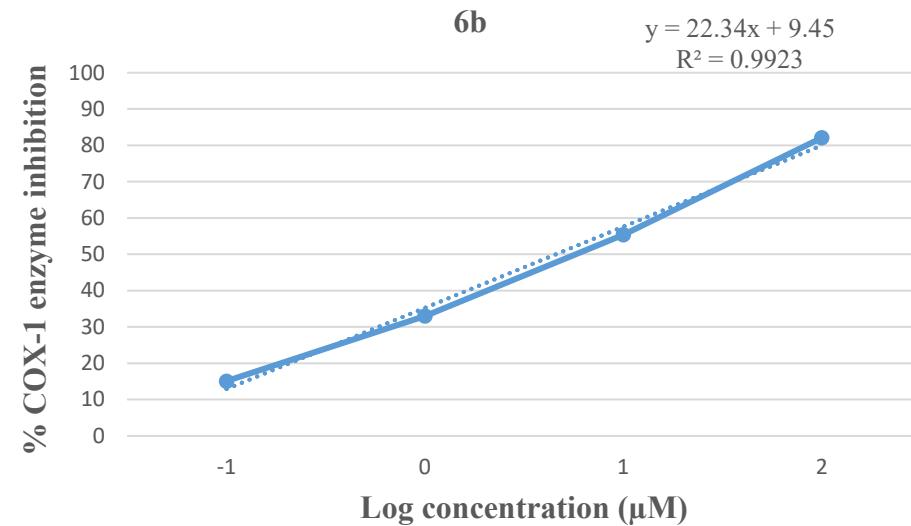


Code	IC_{50}	Conc.	Log	% Inhibition	K activity
6a	35.95	100	2	70.4	35.4635
		10	1	36.9	75.6796
		1	12.6	18.5	97.7618
		0.1	-1	7.38	111.143
EC				0	120

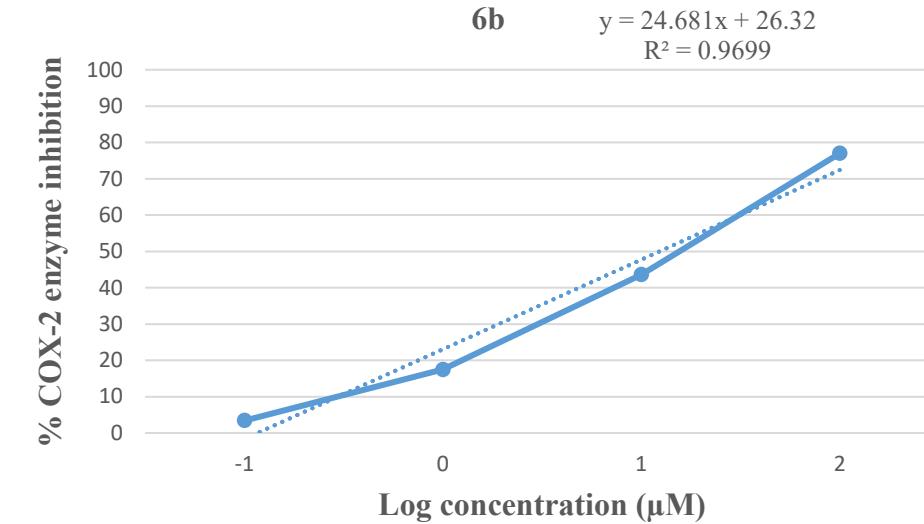
Code	IC_{50}	Conc.	Log	% Inhibition	K activity
6a	7.76	100	2	81.8	21.7942
		10	1	54.4	54.7375
		1	0	30.3	83.6244
		0.1	-1	11.6	106.127
EC				0	120



COX-1

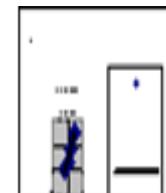
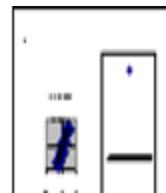


COX-2

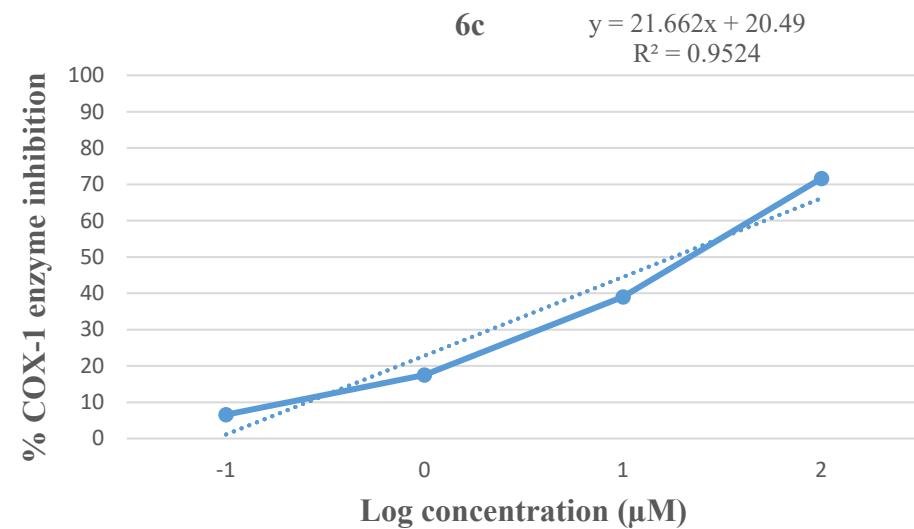


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
6b	19.29	100	2	82.1	21.4341
		10	1	55.4	53.5014
		1	12.6	33	80.444
		0.1	-1	15.1	101.902
EC				0	120

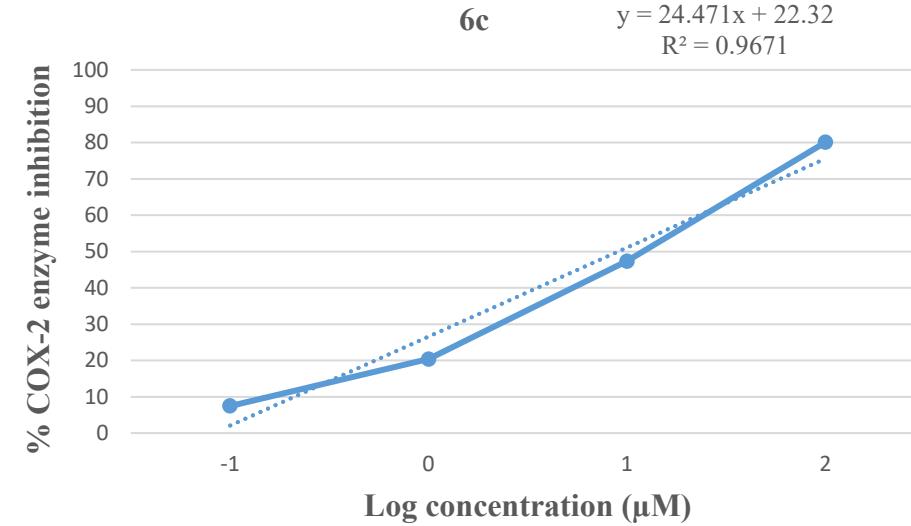
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
6b	56.3	100	2	77	27.6268
		10	1	43.6	67.7228
		1	0	17.5	99.0219
		0.1	-1	3.43	115.884
EC				0	120



COX-1

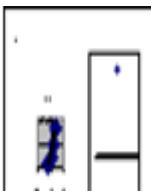
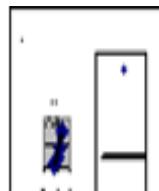


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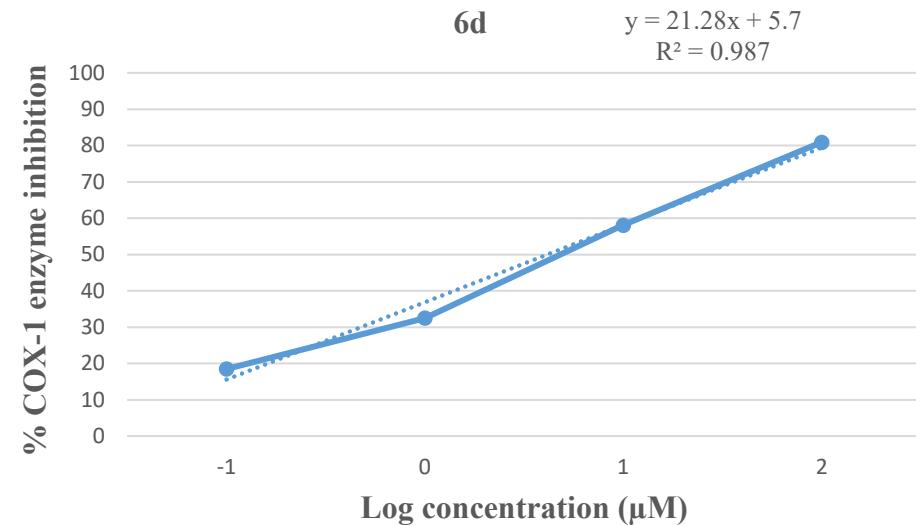


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
6c	31.20	100	2	71.6	34.0954
		10	1	39	73.2553
		1	0	17.5	99.0219
	0.1	-1	6.56	112.127	
EC				0	120

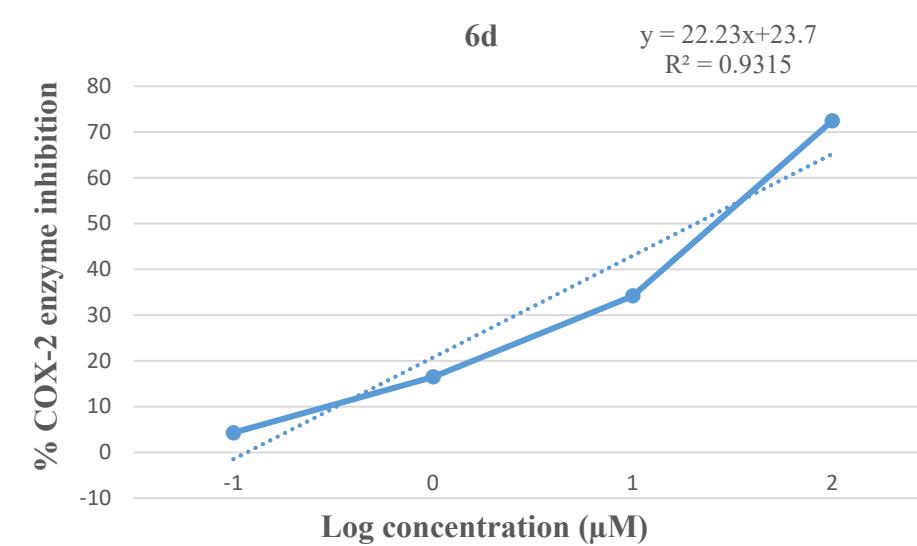
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
6c	3.85	100	2	80.1	23.9064
		10	1	47.4	63.1383
		1	0	20.4	95.5776
	0.1	-1	7.53	110.963	
EC				0	120



COX-1

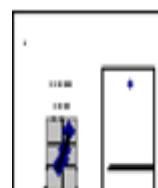
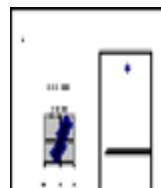


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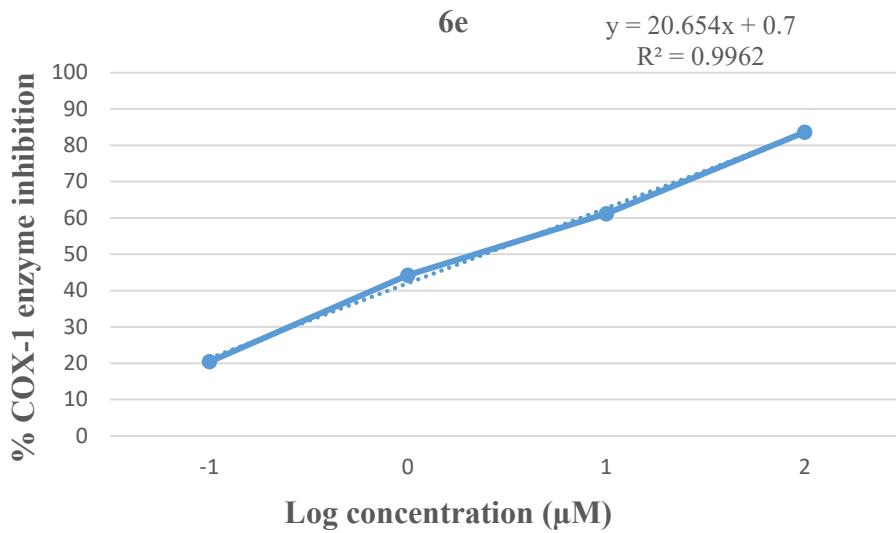


Code	IC_{50}	Conc.	Log	% Inhibition	K activity
6d	42.91	100	2	80.9	22.9703
		10	1	58.1	50.261
		1	0	32.5	81.0321
	0.1	-1	18.5	97.8218	
EC			0	120	

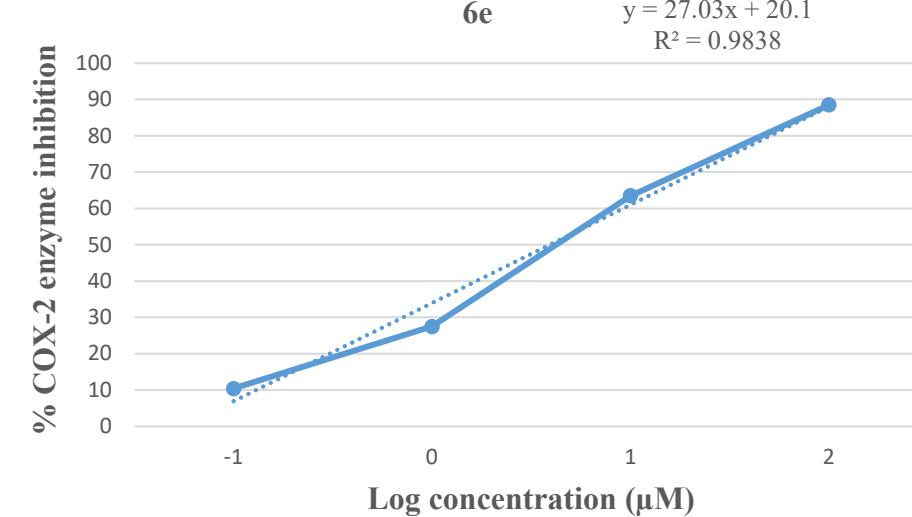
Code	IC_{50}	Conc.	Log	% Inhibition	K activity
6d	9.87	100	2	72.5	33.0153
		10	1	34.2	78.9919
		1	0	16.5	100.222
	0.1	-1	4.3	114.839	
EC			0	120	



COX-1

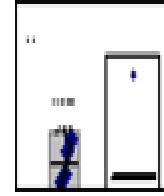
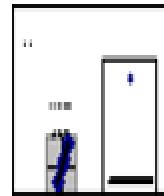


COX-2

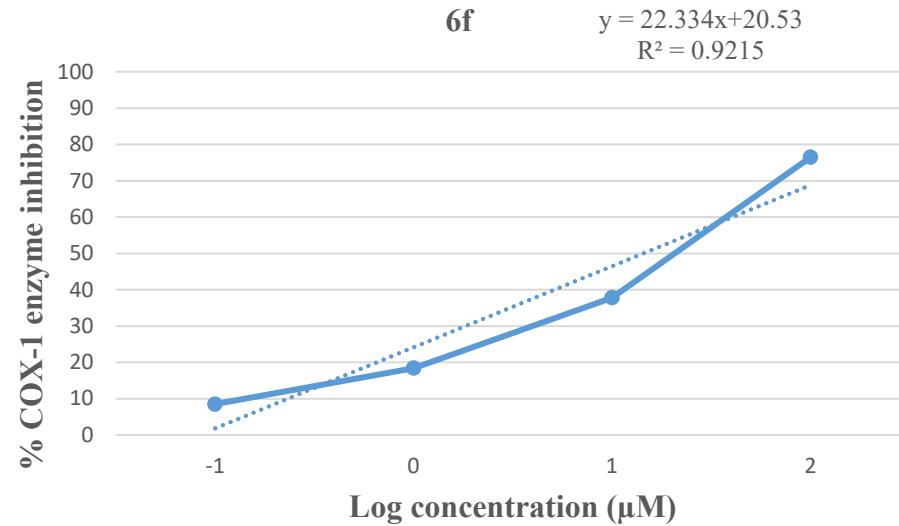


Code	IC_{50}	Conc.	Log	% Inhibition	K activity
6e	5.48	100	2	83.6	19.634
		10	1	61.4	46.3366
		1	12.6	44.2	66.9547
	0.1	-1	20.4	95.5416	
EC			0	120	

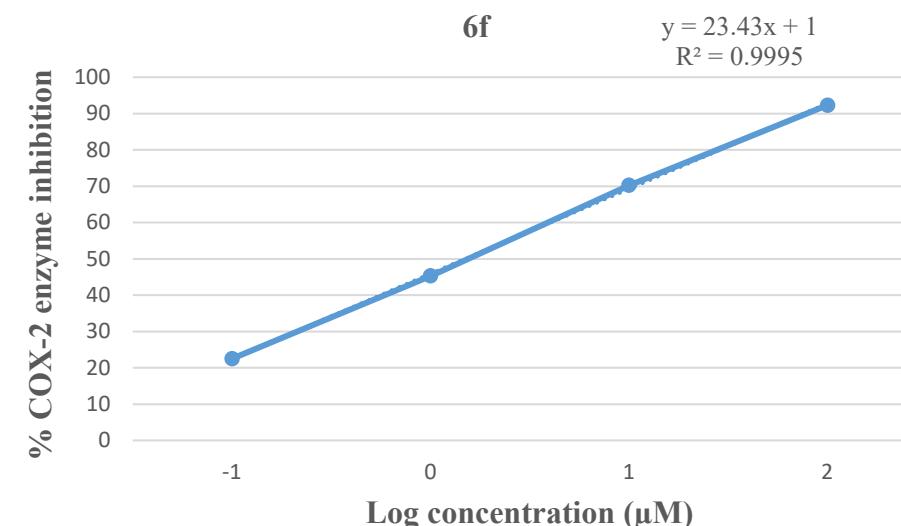
Code	IC_{50}	Conc.	Log	% Inhibition	K activity
6e	2.51	100	2	88.5	13.8254
		10	63.5	83	43.8284
		1	0	27.5	87.0207
	0.1	-1	10.4	107.543	
EC			0	120	



COX-1

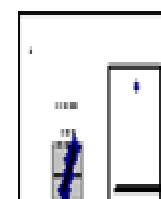
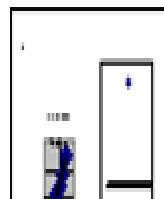


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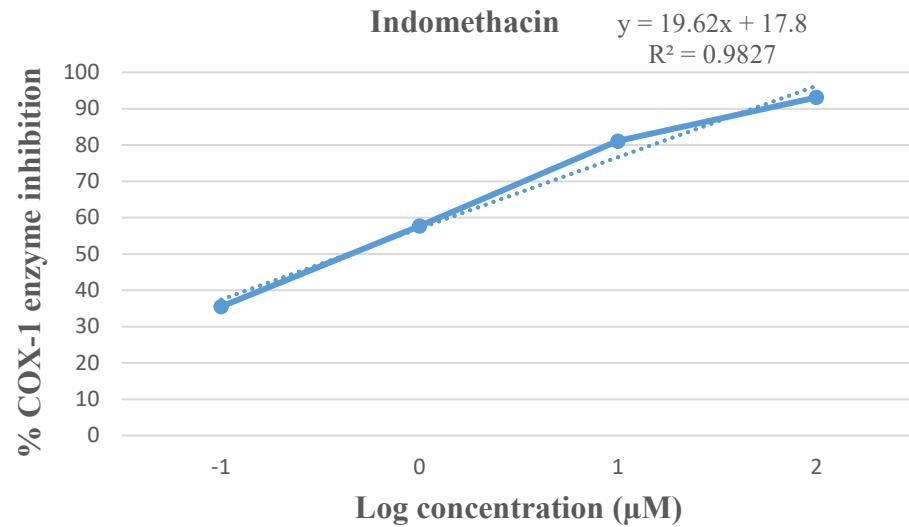


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
6f	9.61	100	2	76.5	28.2148
		10	1	37.8	74.5995
		1	12.6	18.4	97.9178
	0.1	-1	8.52	109.775	
EC			0	120	

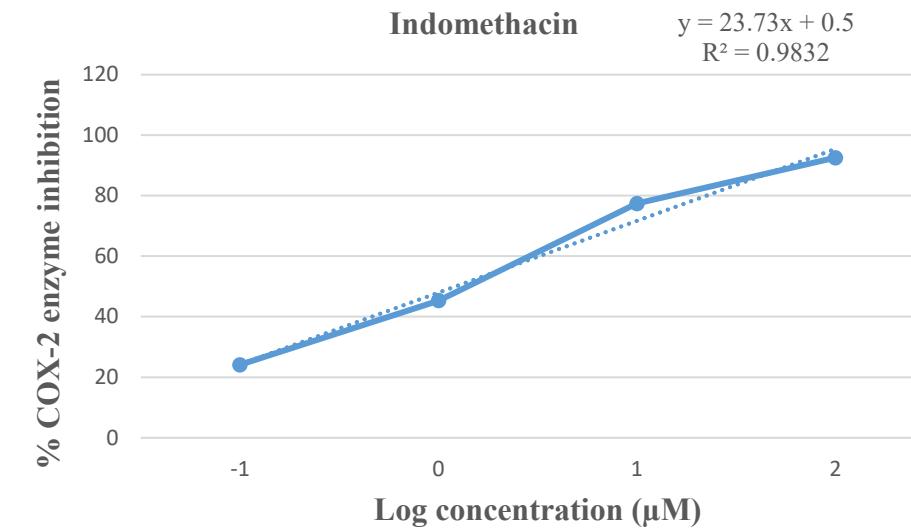
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
6f	1.15	100	2	92.3	9.25293
		10	1	70.2	35..7756
		1	0	45.3	65.6706
	0.1	-1	22.5	92.9973	
EC			0	120	



COX-1

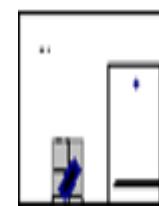
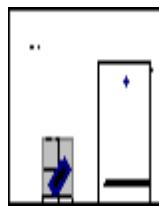


COX-2

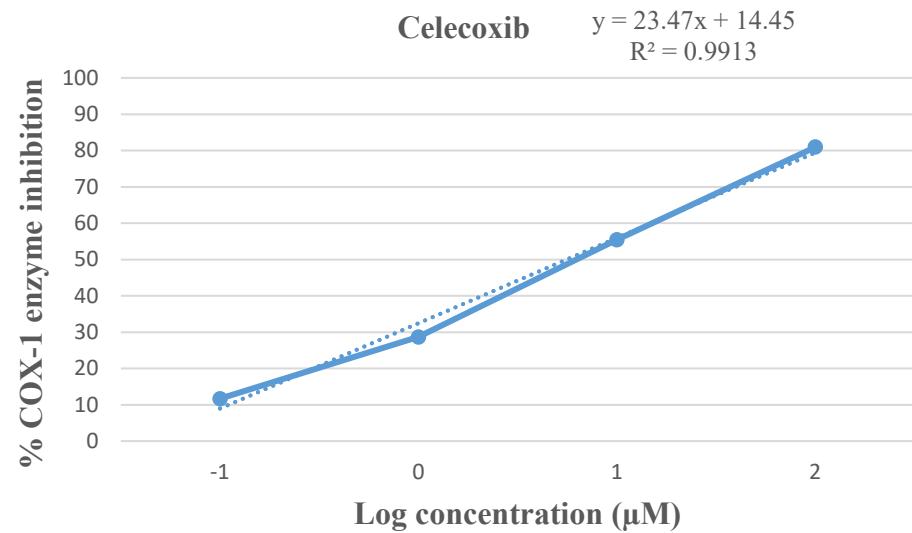


Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
Indomethacin	0.43	100	2	93.2	8.25683
		10	1	81.1	22.7303
		1	12.6	57.7	50.7531
		0.1	-1	35.5	77.4557
EC			0	0	120

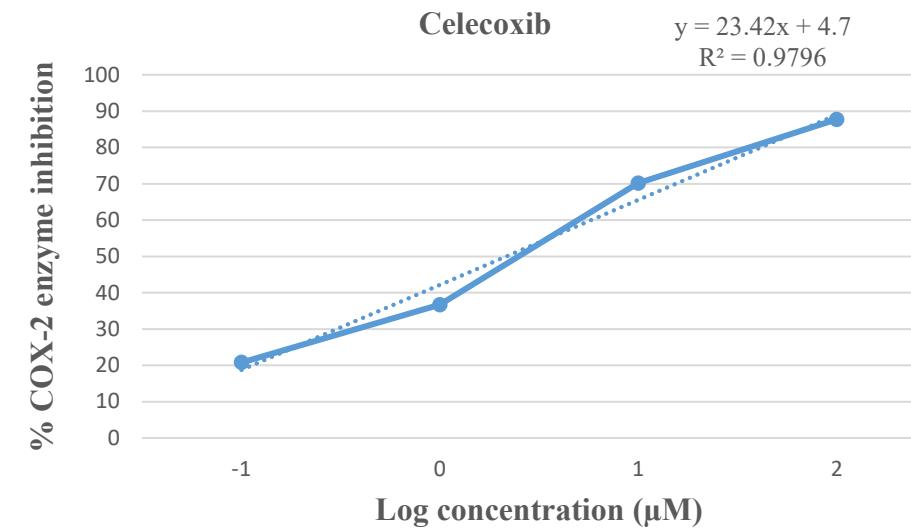
Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
Indomethacin	1.22	100	2	92.5	9.0129
		10	1	77.4	27.1587
		1	0	45.3	65.6346
		0.1	-1	24.1	91.1131
EC			0	0	120



COX-1

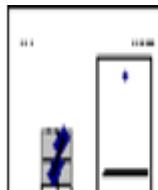
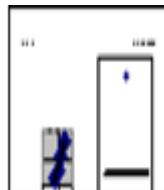


COX-2



Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
Celecoxib	5.43	100	2	81.8	21.8542
		10	1	55.5	53.4413
		1	12.6	28.7	85.5206
		0.1	-1	11.7	105.983
EC			0	0	120

Code	IC ₅₀	Conc.	Log	% Inhibition	K activity
Celecoxib	2.16	100	2	87.7	14.7975
		10	1	70.2	35.7516
		1	0	36.7	75.9196
		0.1	-1	20.8	95.0255
EC			0	0	120



1.1.3 Inhibition of TNF- α , IL-6, and PGE-2 expression in LPS-induced RAW264.7 cell

RAW 264.7 cells were incubated in 96 well-plates (5×10^3 cells/well) in 5% CO₂ at 37 °C for 24 h. Thereafter, the cells were treated with the tested compounds (50 μM) in the presence of LPS (1 μg/mL). The untreated cells, acted as the blank control, whereas the negative control group (LPS group) was represented by the treated cells with LPS + DMSO (final concentration of 0.1% (v/v)). TNF- α , PGE-2, and IL-6 were quantified 24 h after treatment, utilizing specified ELISA kits.

1.1.4 Inhibition of NO expression LPS-induced RAW264.7 cells

Aiming to measure NO expression in the culture medium, RAW264.7 cells pretreated with the test compounds for 2 h were treated with LPS (1 μg/mL) for 24 h. To the culture medium (100 L), Griess reagent (100 L) (Sigma-Aldrich, St. Louis, MO, USA) was added, then incubated at room temperature for 10 min. ELISA reader at 540 nm was used to measure the optical density (OD) as an indicator of NO.

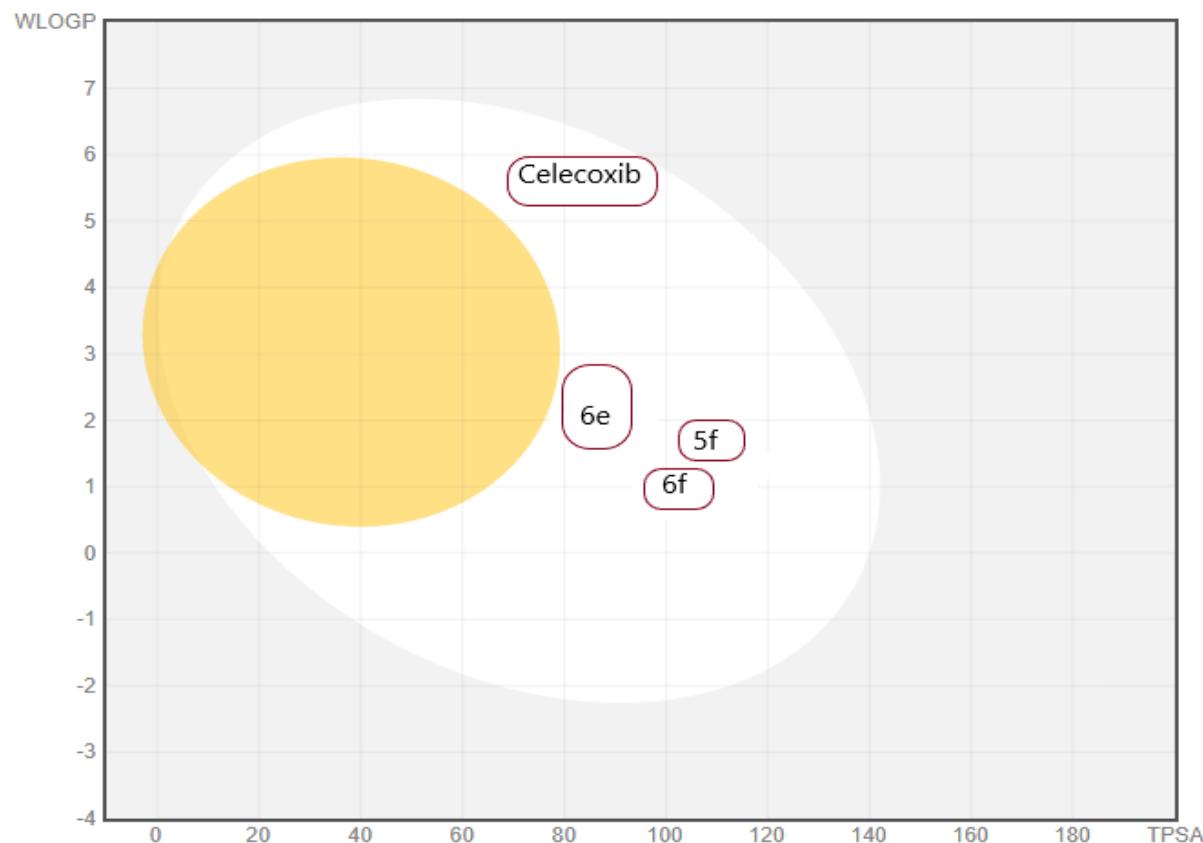
1.2 Molecular docking of compounds **5f, **6e** and **6f** in the active site of COX-2**

Molecular Operating Environment (MOE Version 2015.10) software was used to study the molecular modeling for the new active compounds **5f**, **6e** and **6f**. The COX-2 enzyme X-ray crystal structure (PDB entry 3LN1) was downloaded from the RCSB protein data bank website (<http://www.rcsb.org>). Aiming to prepare protein structure, the repeating chains and water molecules were removed from the enzyme active site. Protonate 3D process was applied to add hydrogen atoms to the atoms of the receptor. The determined pocket was isolated, and the partial charges were calculated. Hiding backbone was performed to give a final prepared protein. Before performing the docking, the tested compounds and celecoxib were prepared automatically by certain steps including 3D protonation, calculating partial charges and minimizing energy with Merck Molecular force field.

Ligands were outlined using ChemDraw office (ChemDraw office version 12.0.2) and saved as mol extension, then prepared by protonation and energy minimization.

2.7. Physicochemical, ADME, and pharmacokinetic properties in *silico* prediction

The BOILED-Egg graphical representation of the WLOGP vs. topological polar surface area for the tested compounds



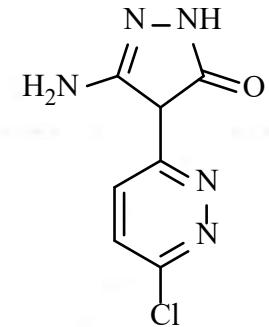
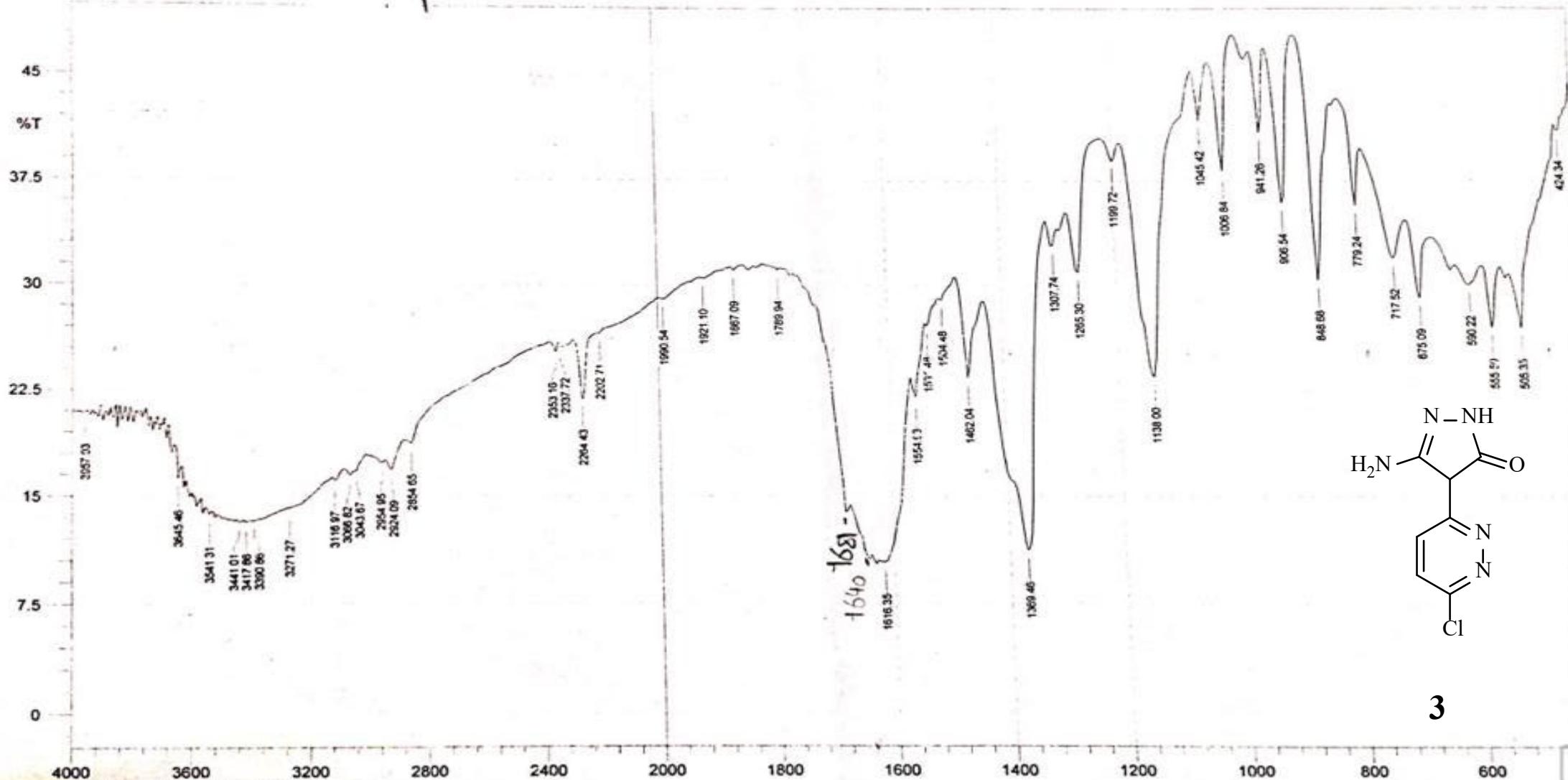
3. IR, ¹H NMR and ¹³C NMR spectra of the new derivatives.



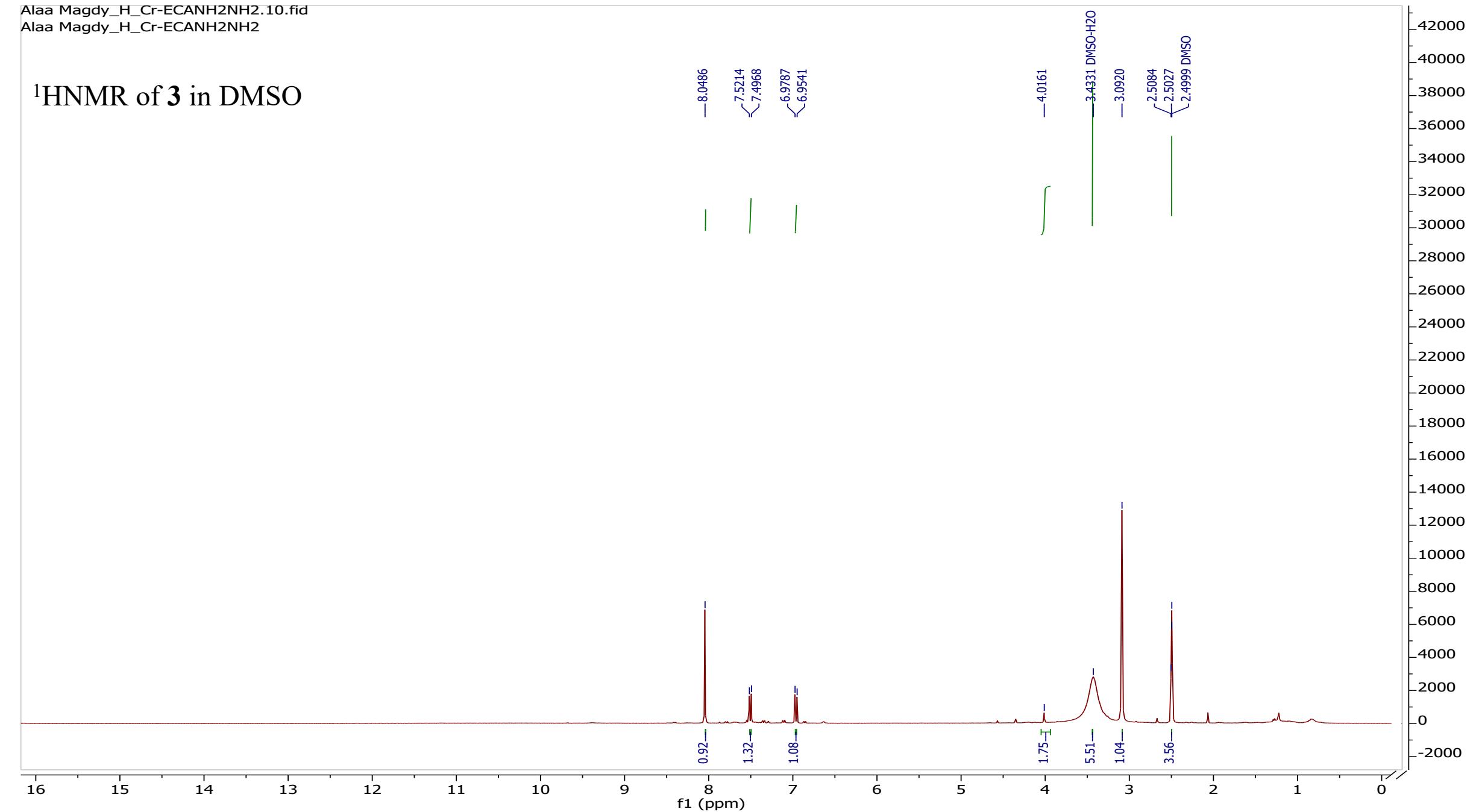
Microanalytical Unit-FOPCU
وحدة التحليل الدقيقة
لأشعة تحت الحمراء



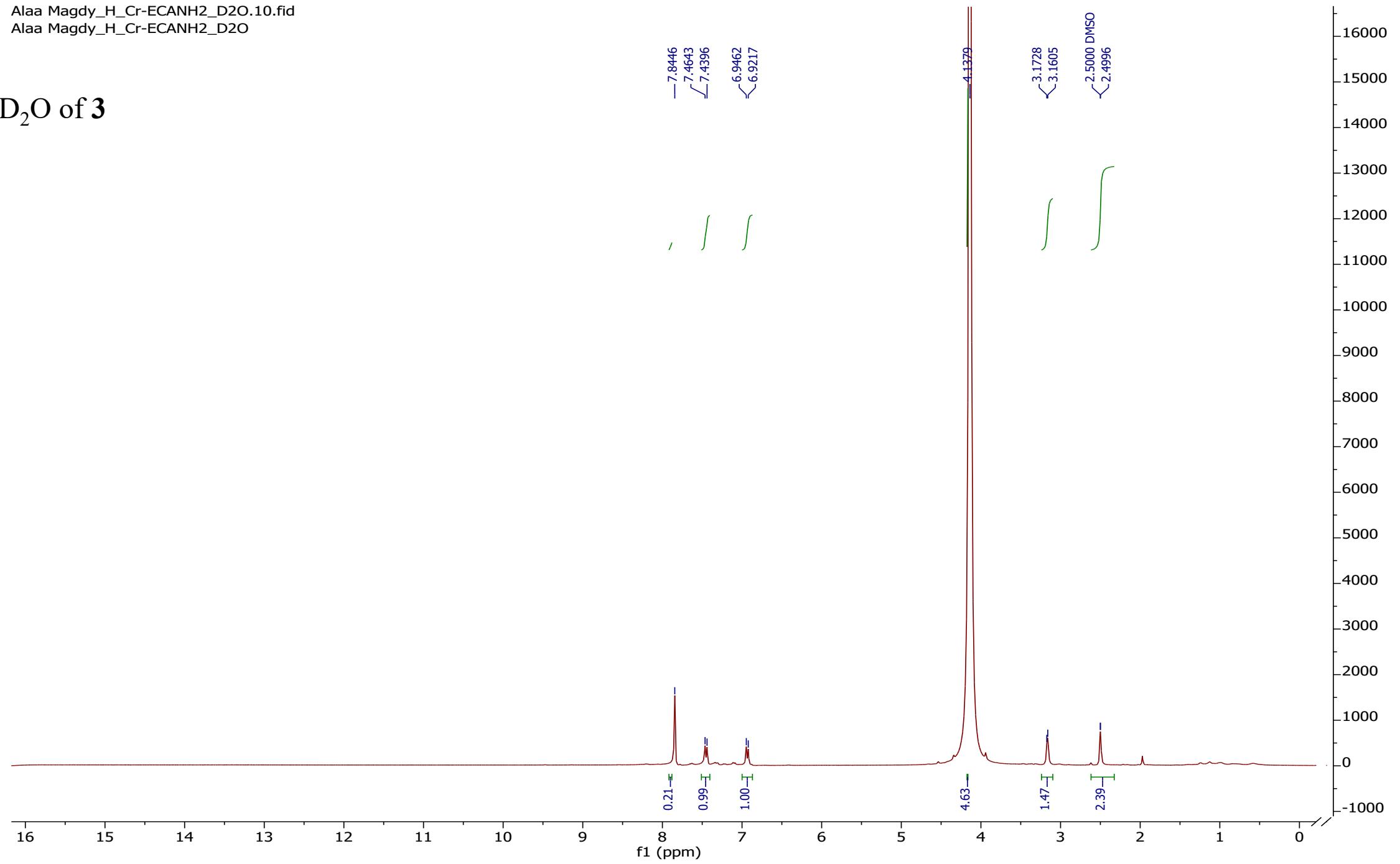
IR of compound 3



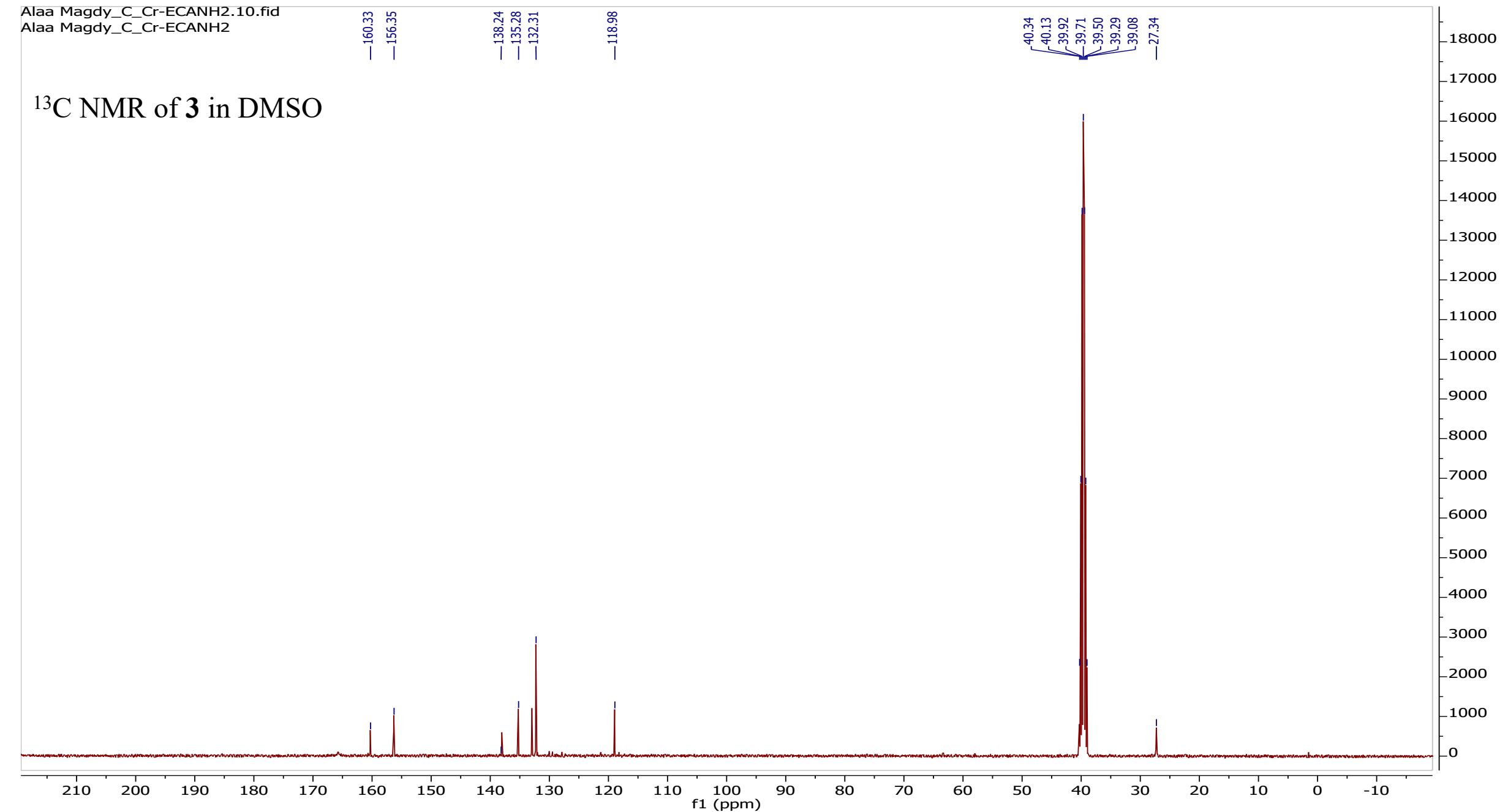
3

¹HNMR of 3 in DMSO

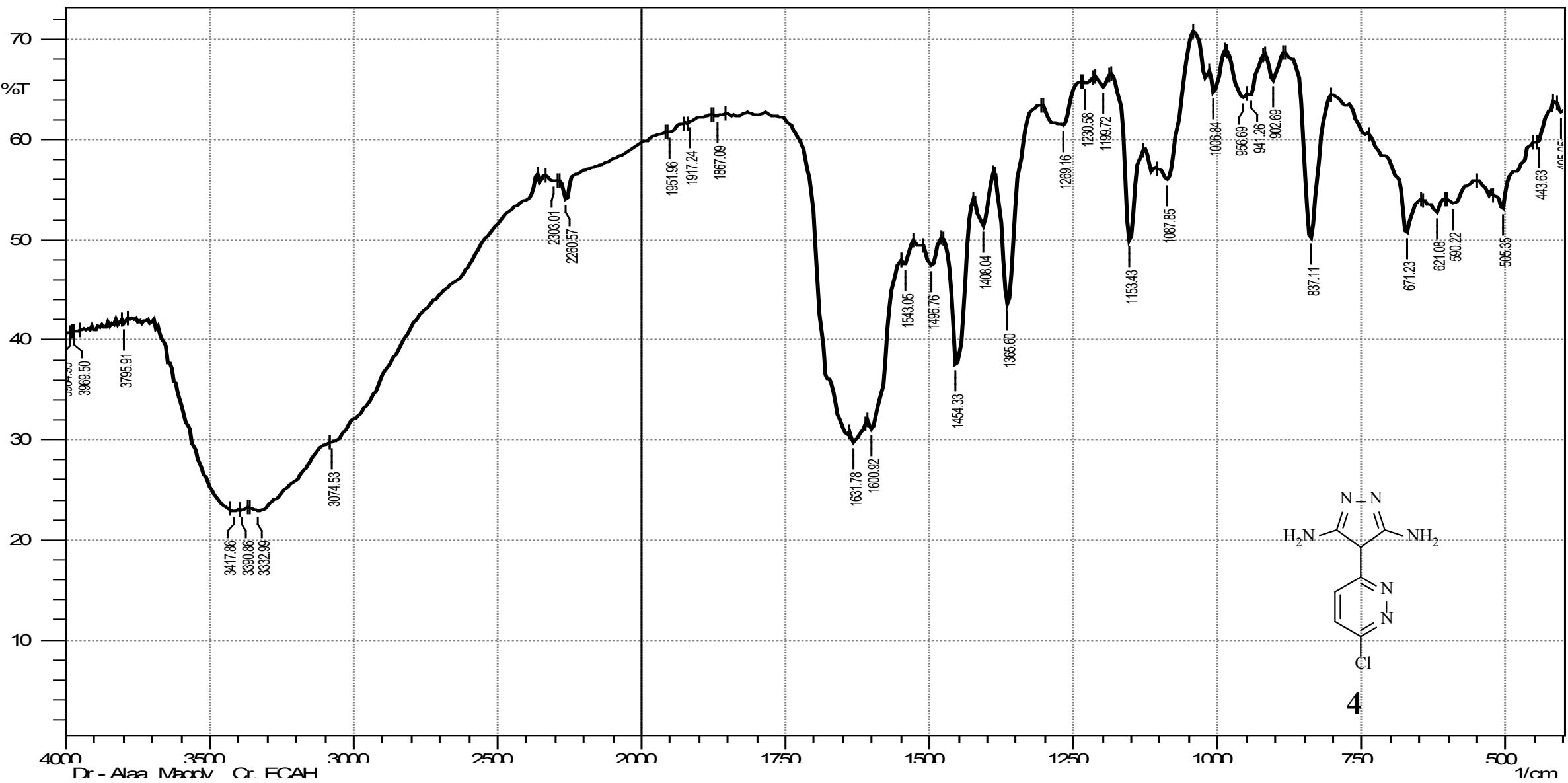
D₂O of 3



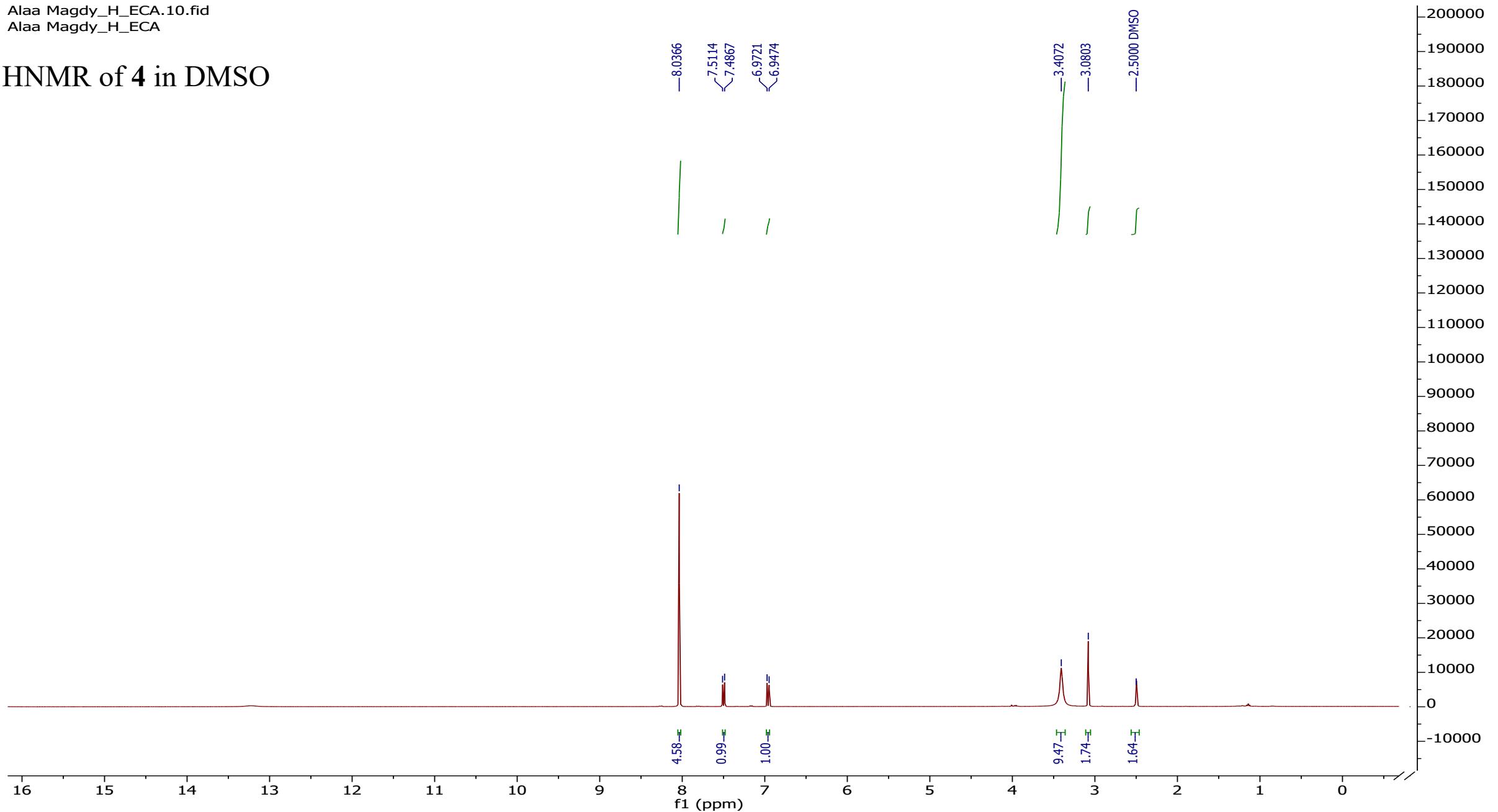
^{13}C NMR of 3 in DMSO



IR of compound 4

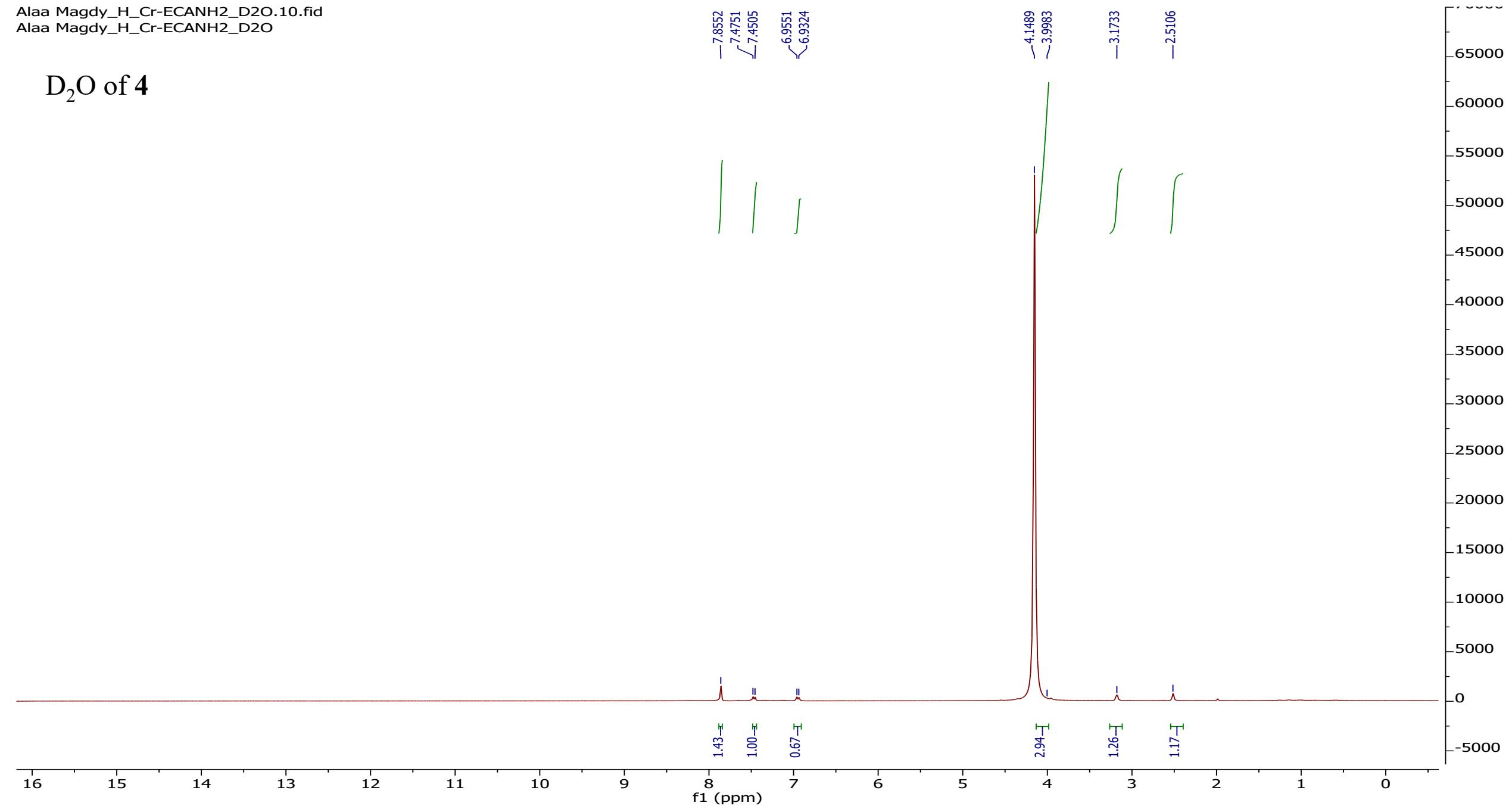


^1H NMR of 4 in DMSO

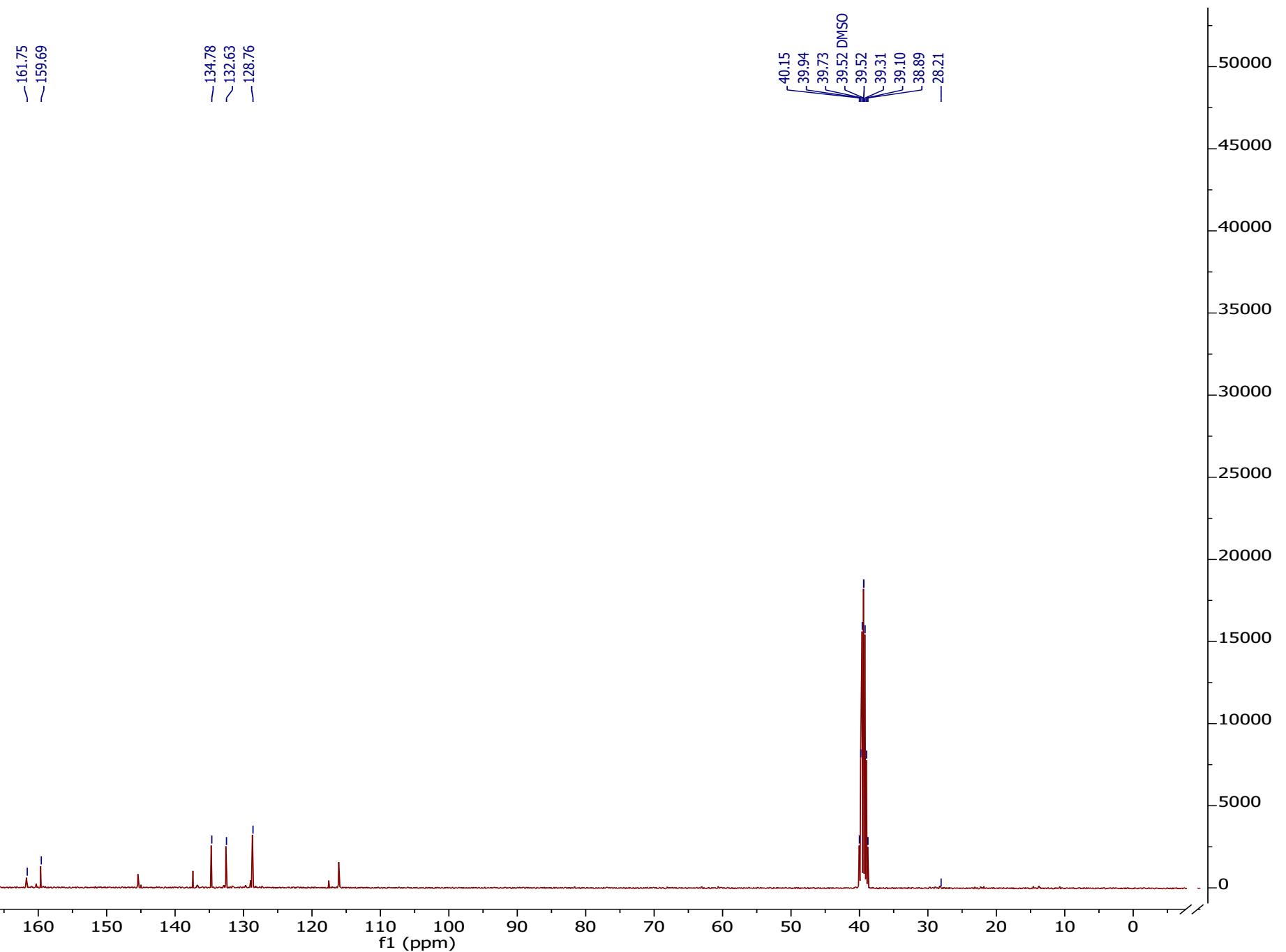


Alaa Magdy_H_Cr-ECANH2_D2O.10.fid
Alaa Magdy_H_Cr-ECANH2_D2O

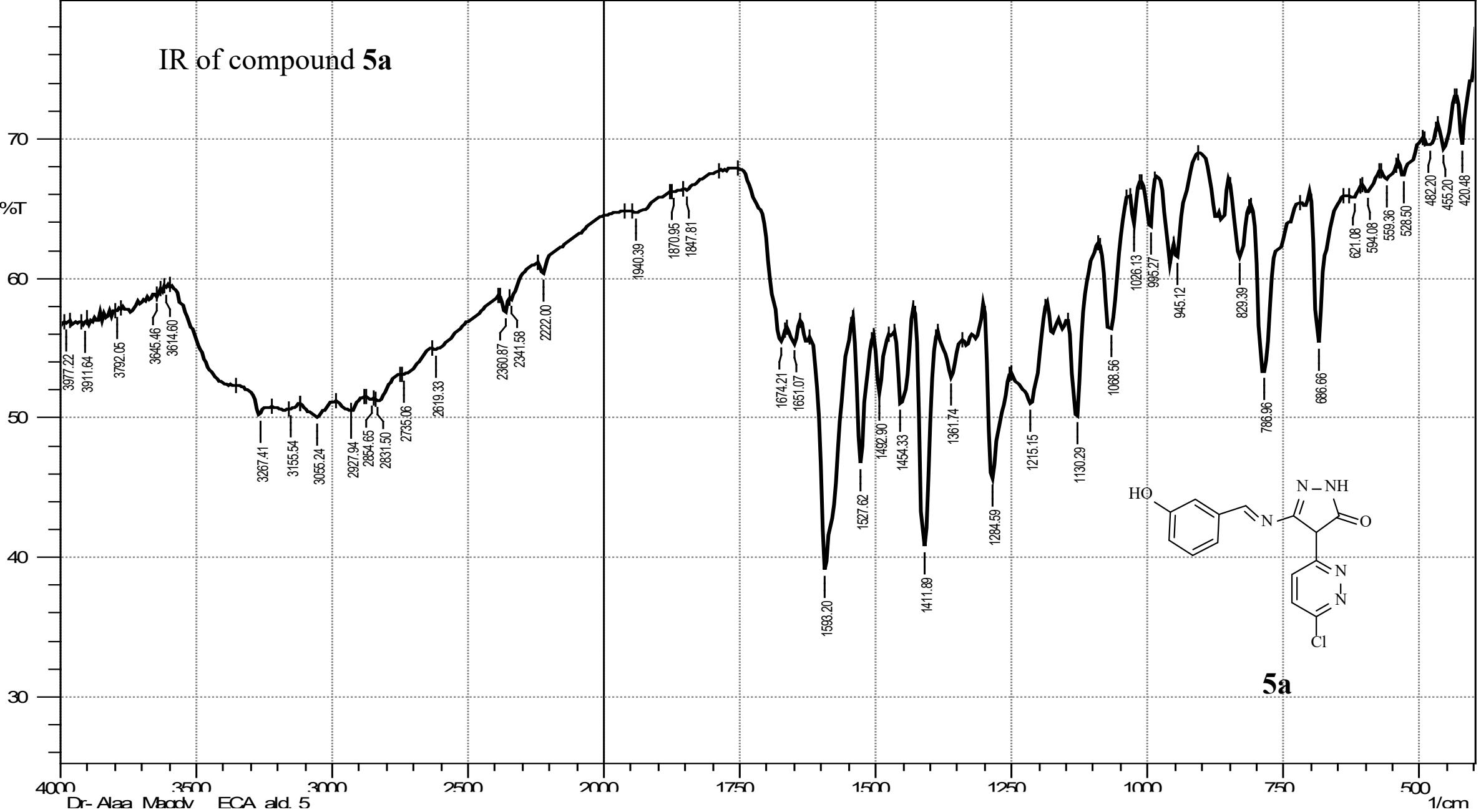
D₂O of 4

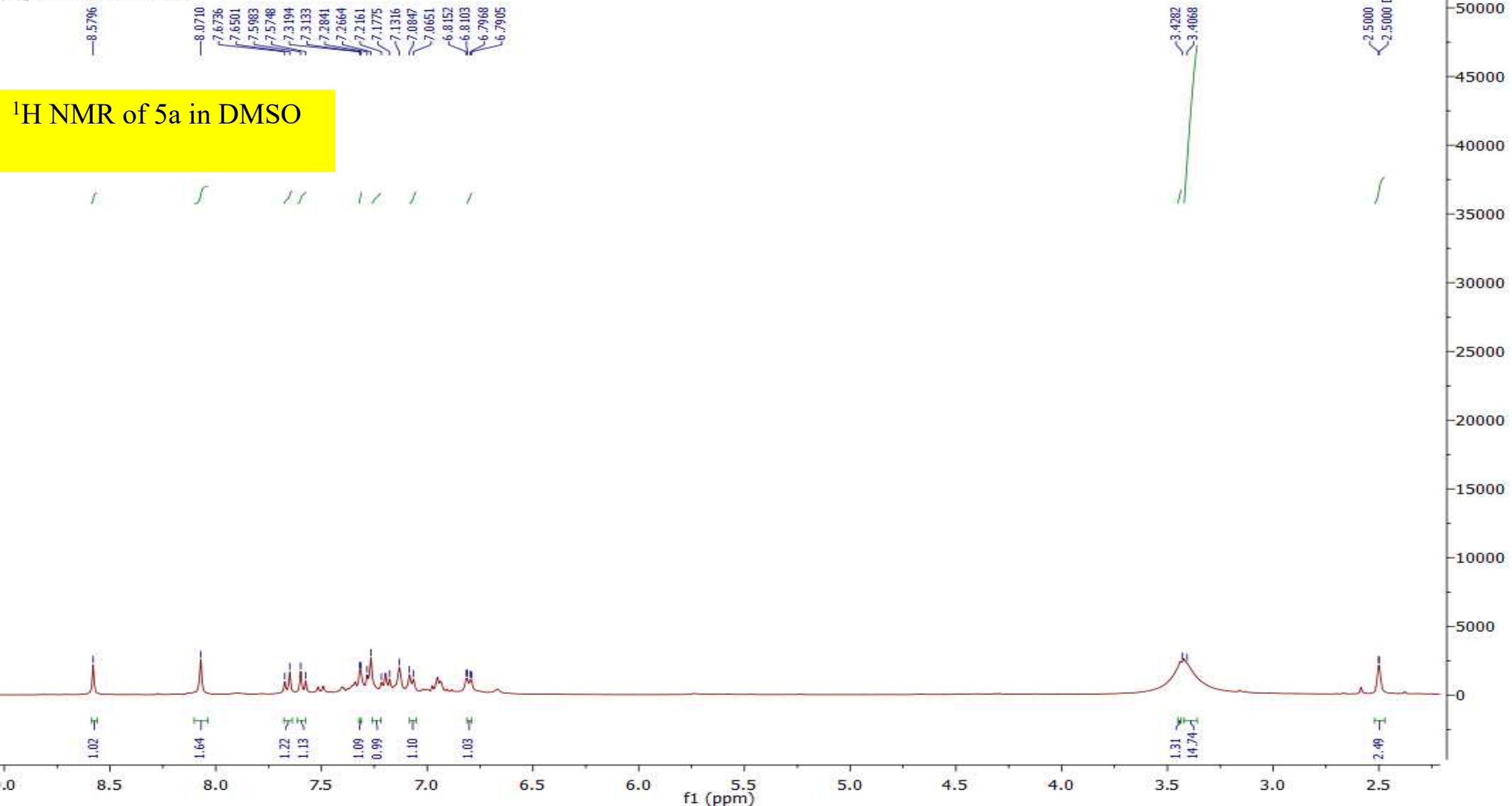


¹³C NMR of 4 in DMSO

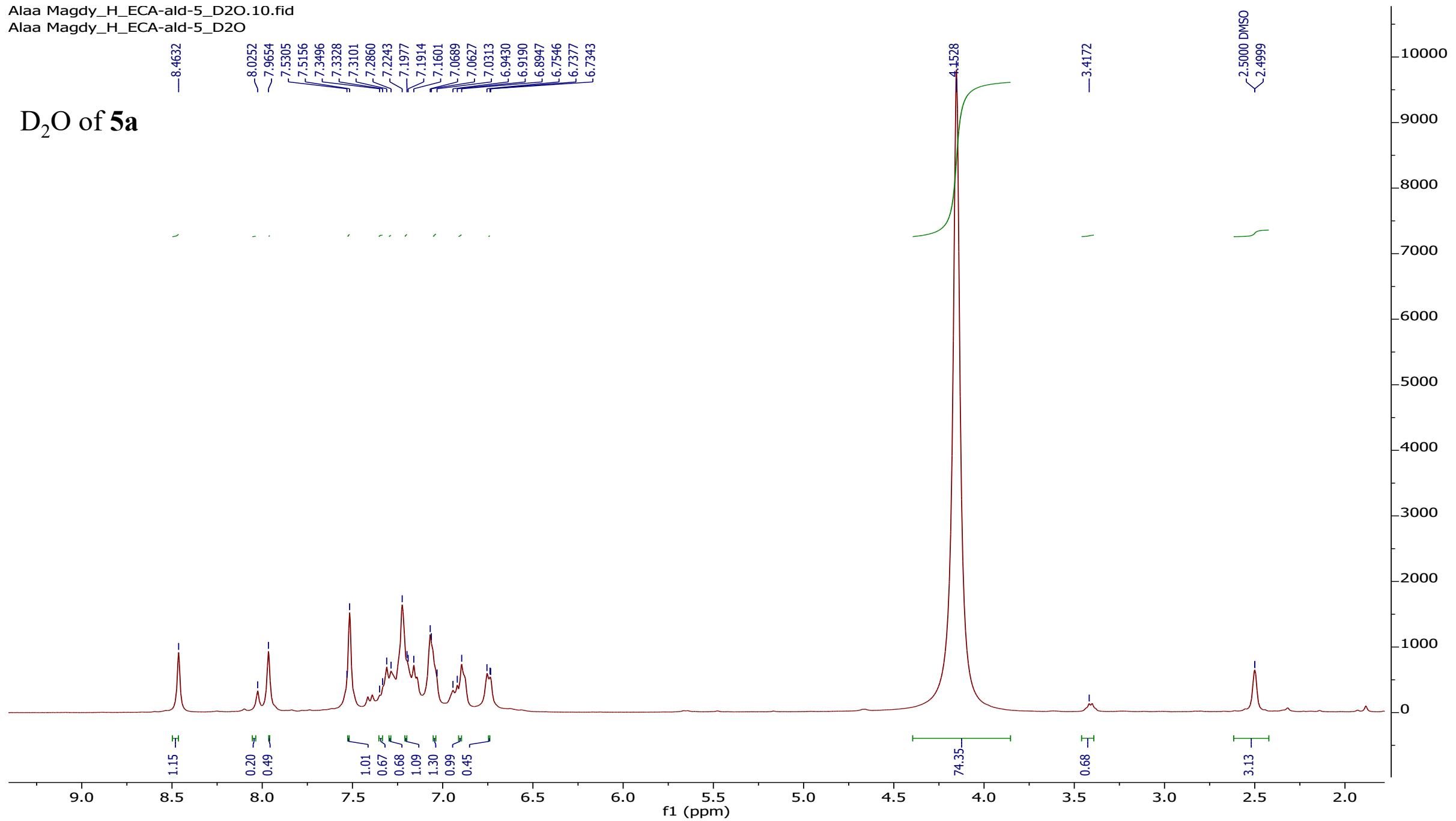


IR of compound 5a



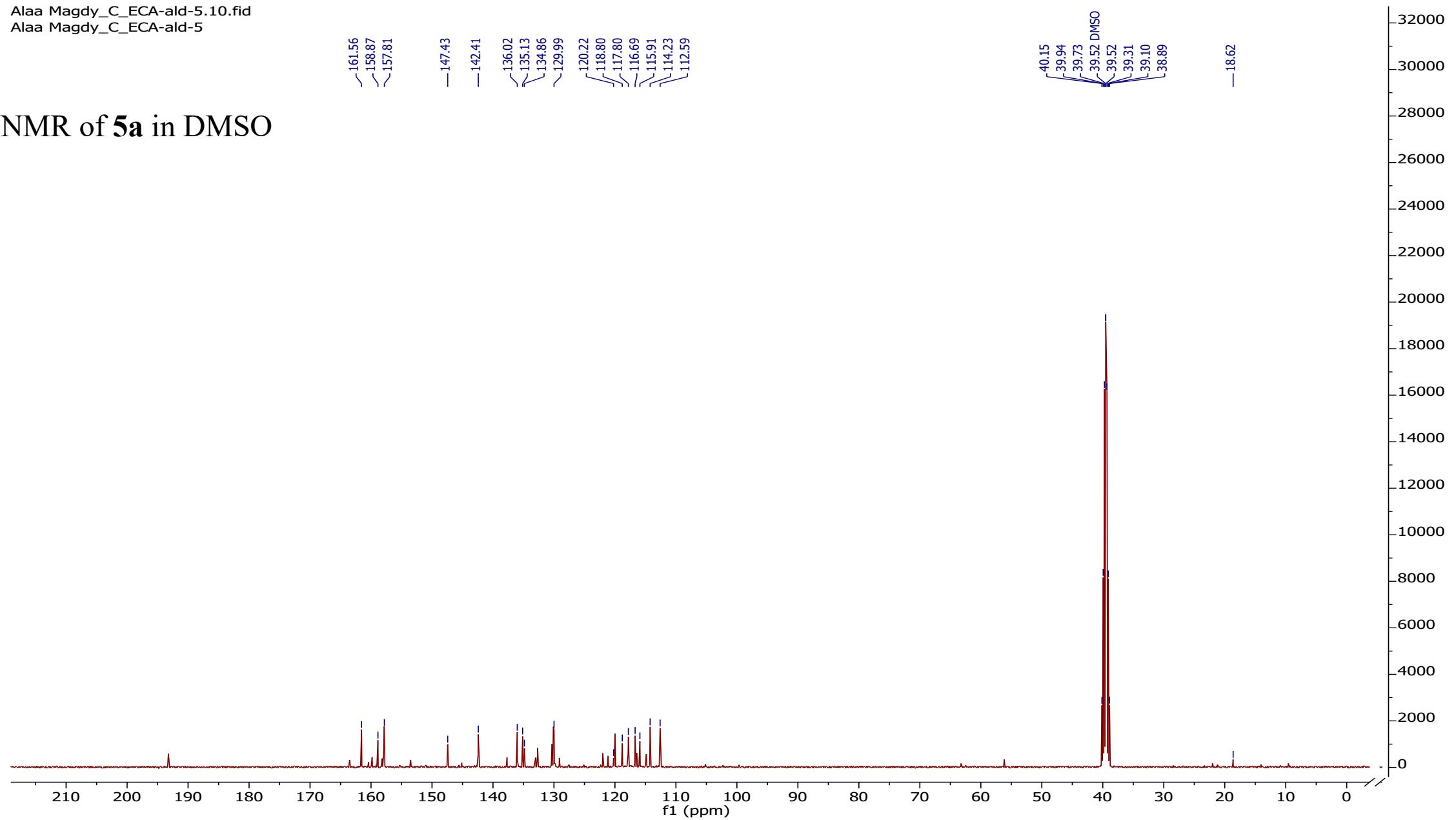


D₂O of 5a



Alaa Magdy_C_ECA-ald-5.10.fid
Alaa Magdy_C_ECA-ald-5

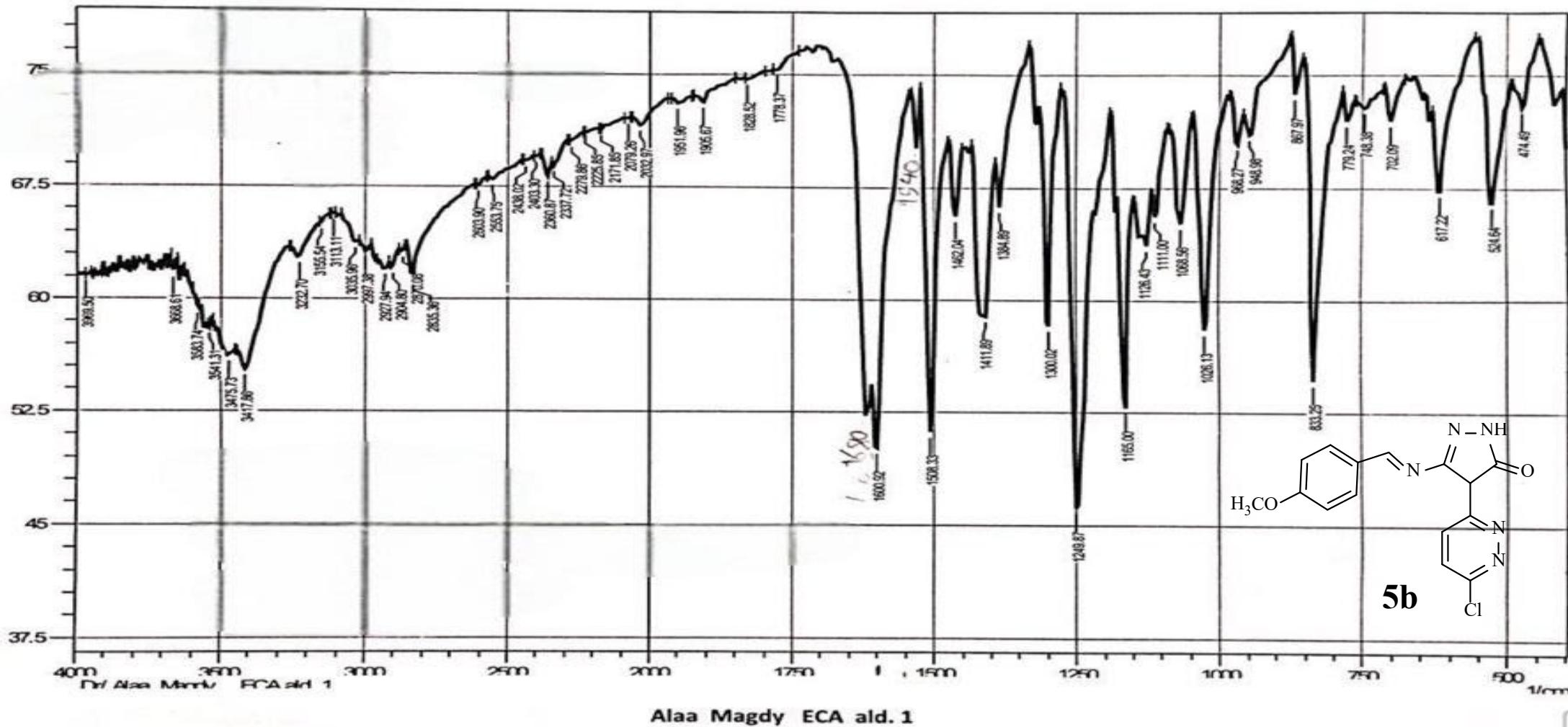
¹³C NMR of **5a** in DMSO

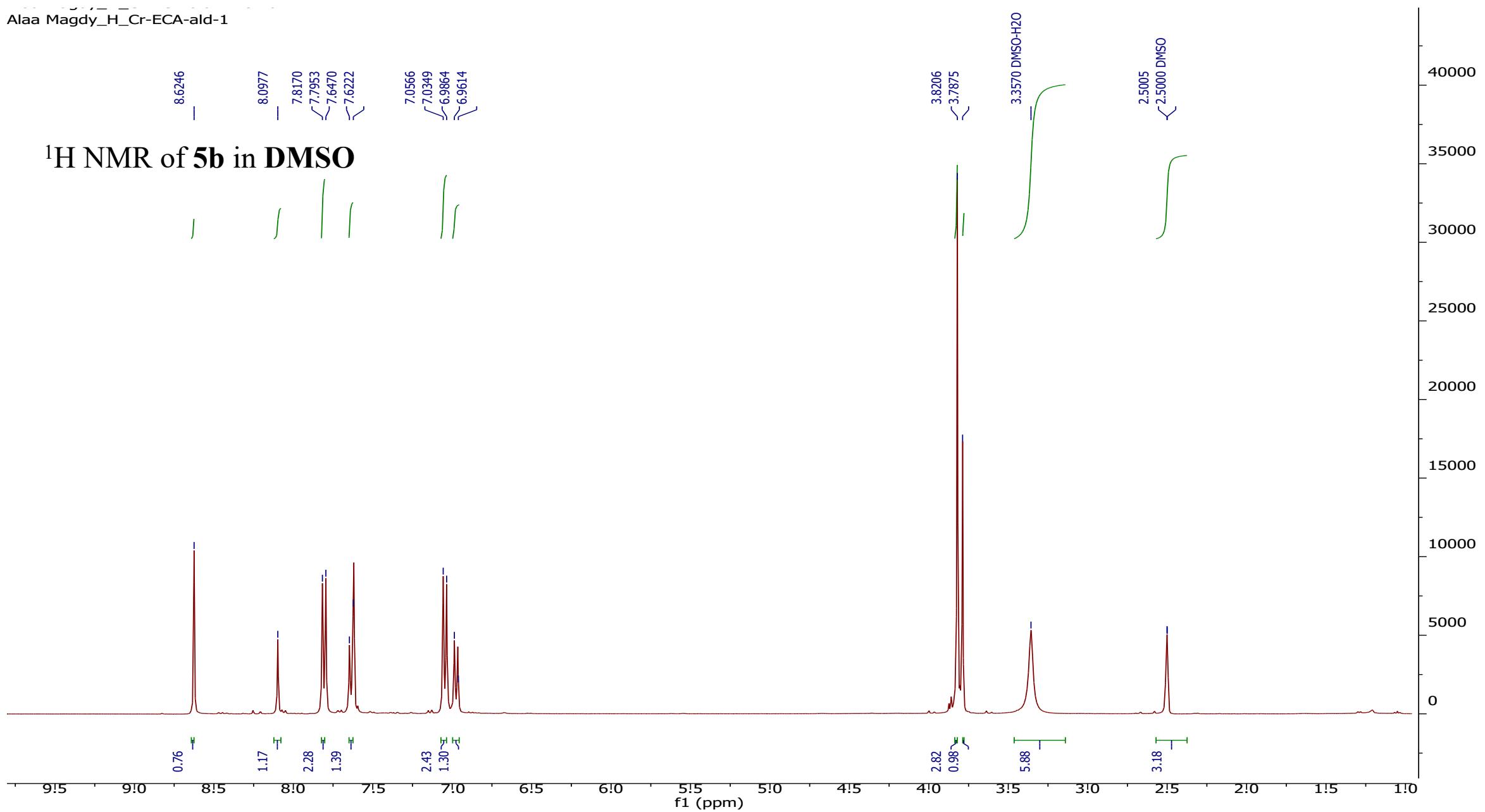


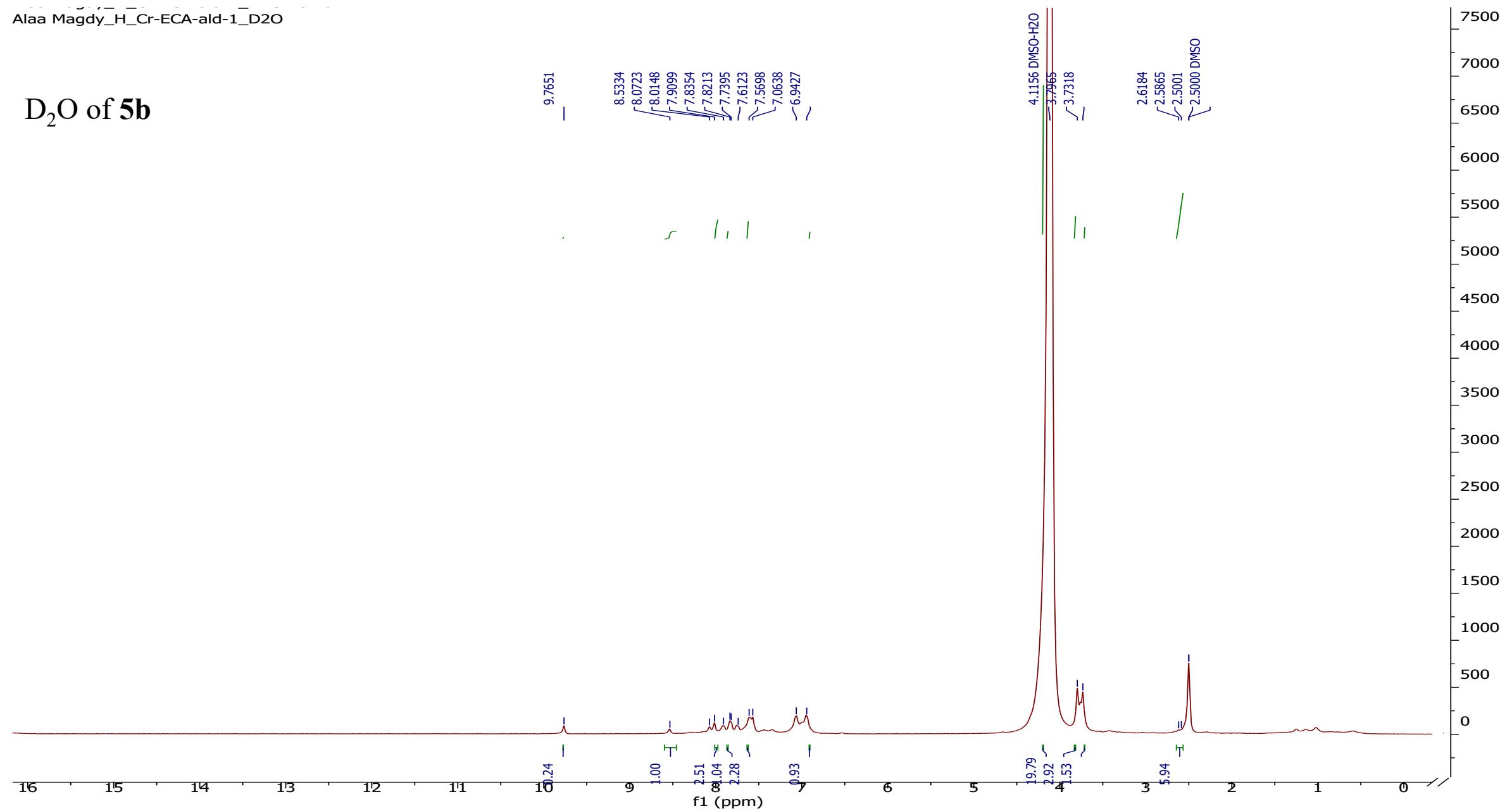
IR of compound 5b

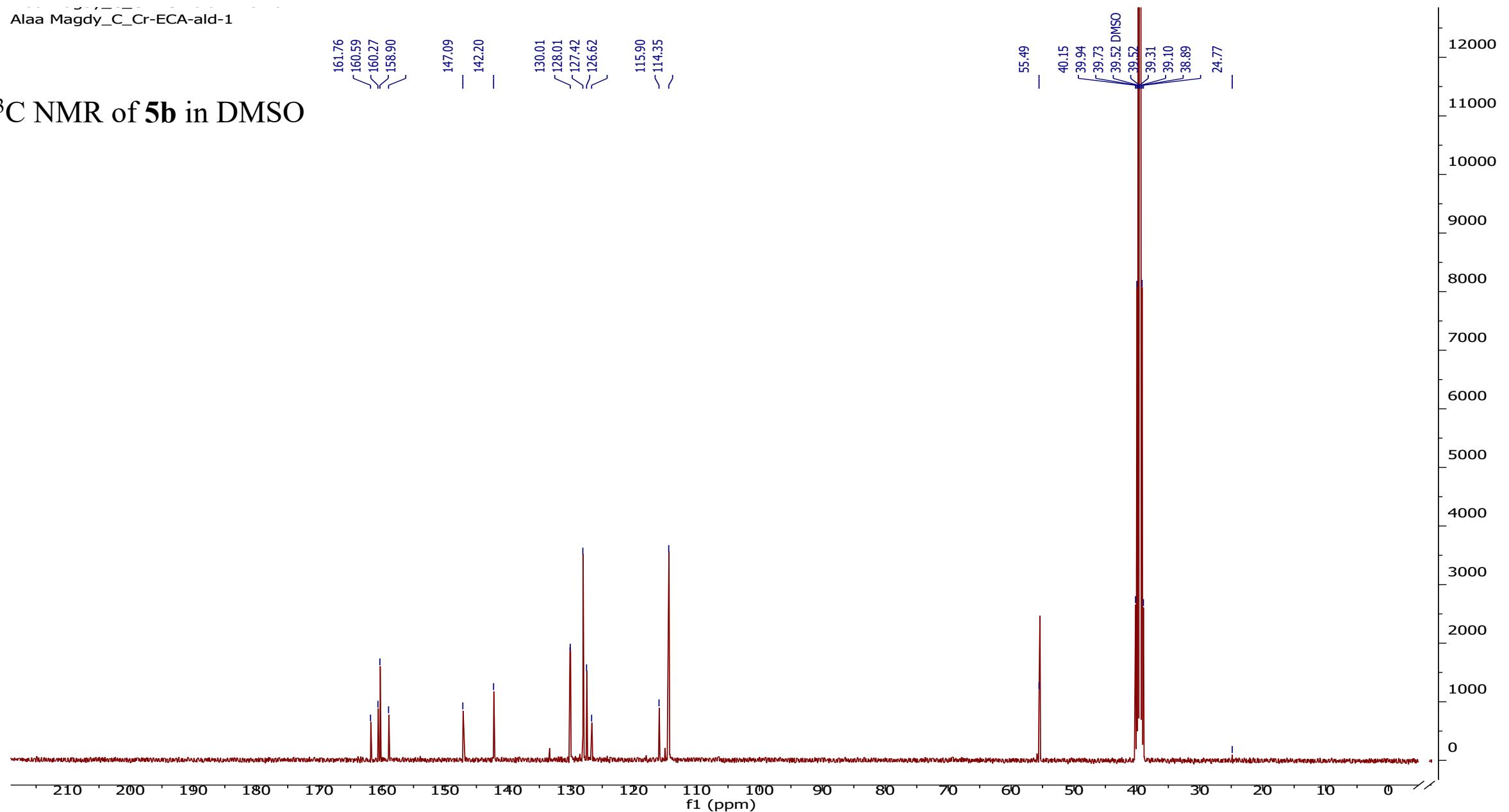


Alaa



¹H NMR of **5b** in DMSO

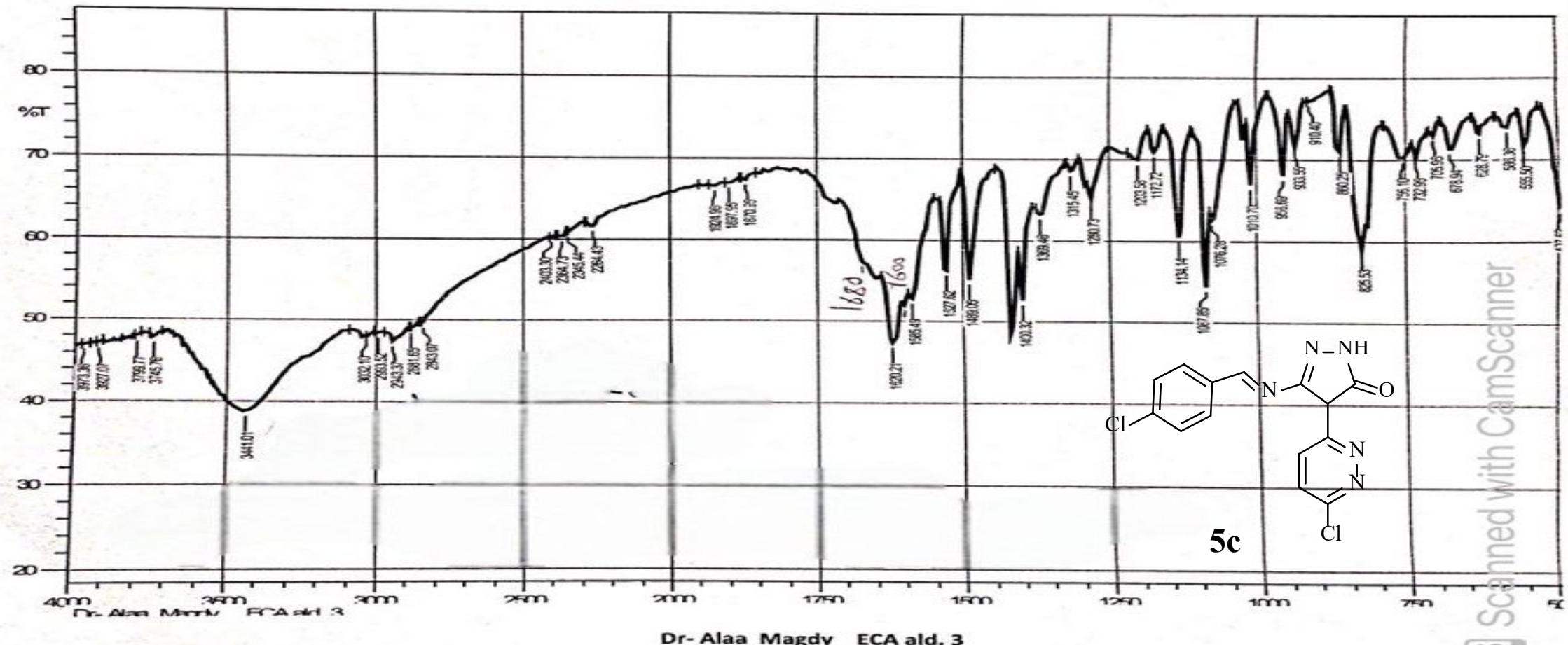
D₂O of 5b

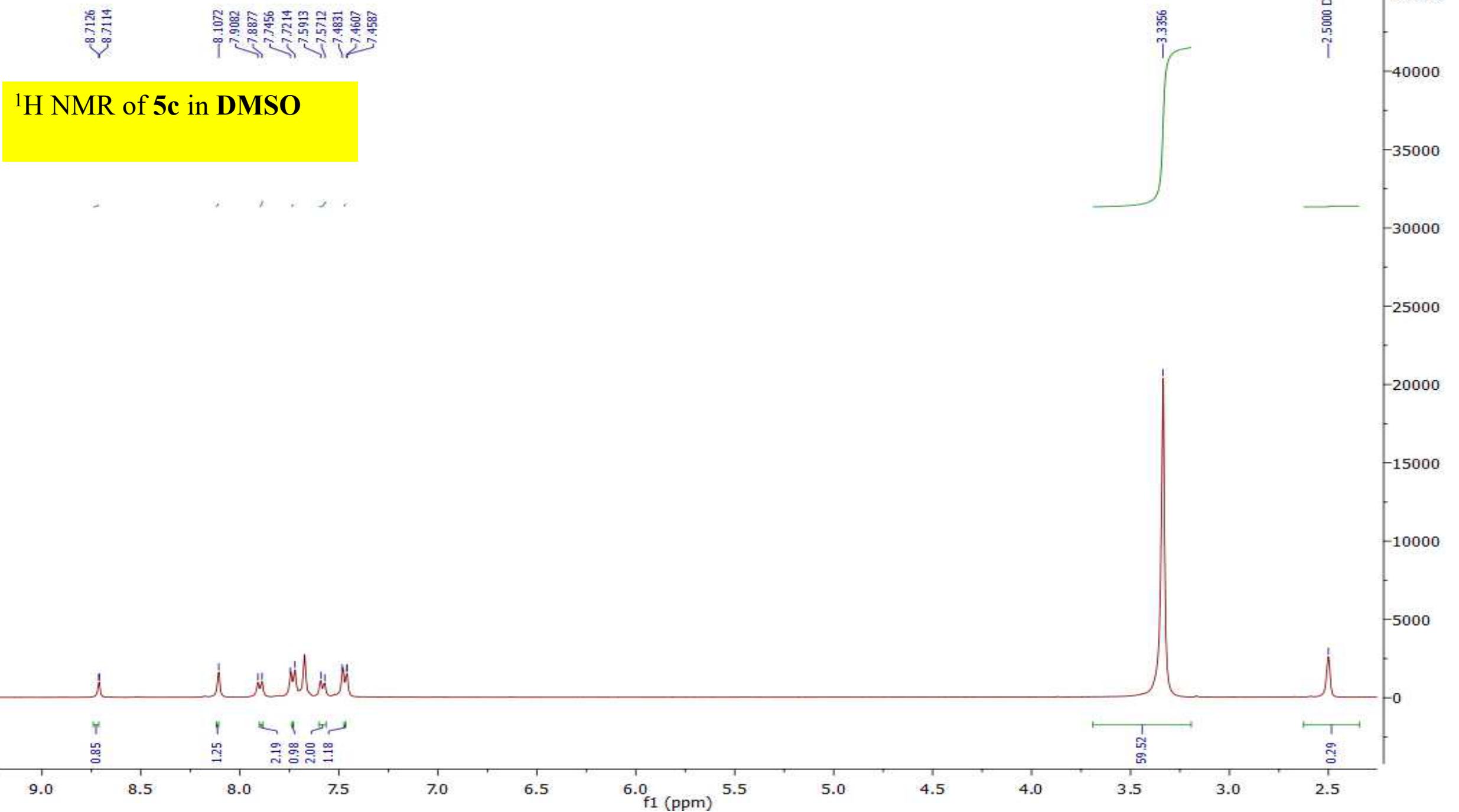
¹³C NMR of **5b** in DMSO

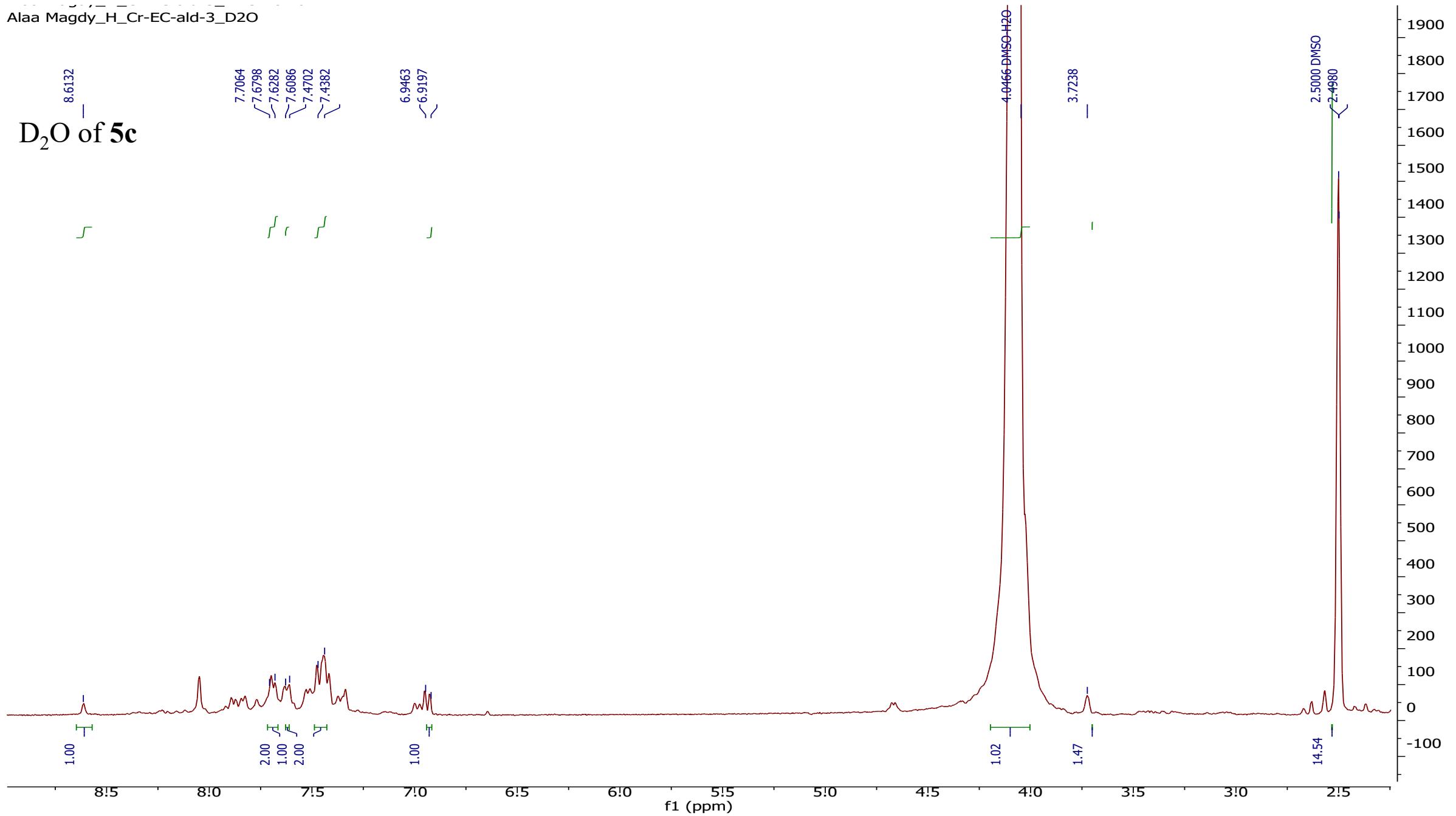
IR of compound 5c

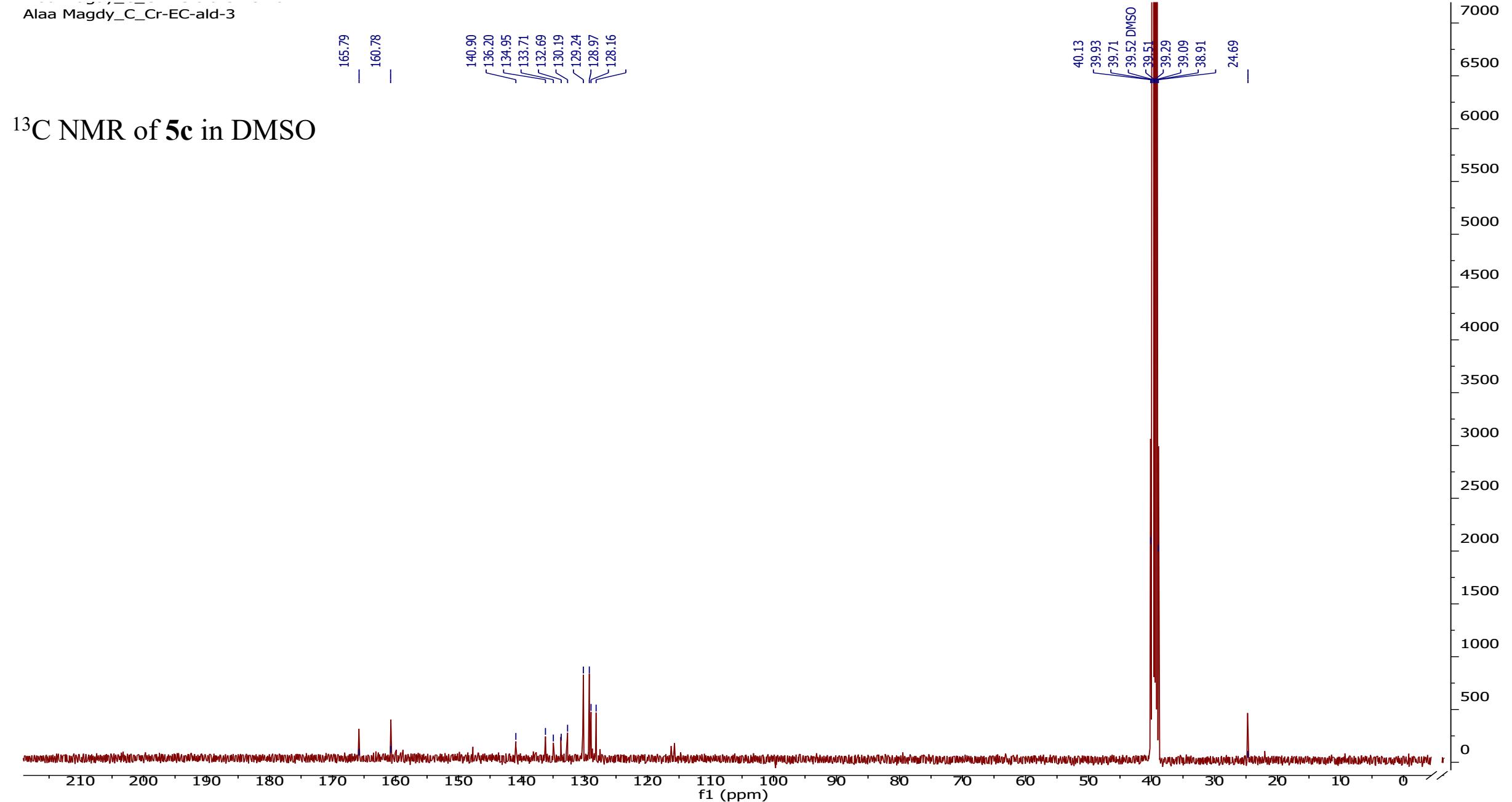


IR Lab





D₂O of 5c

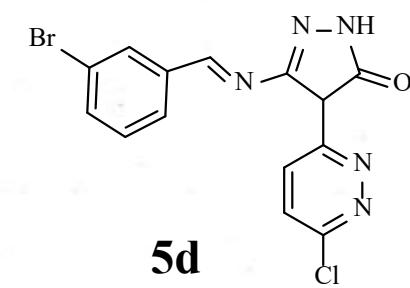
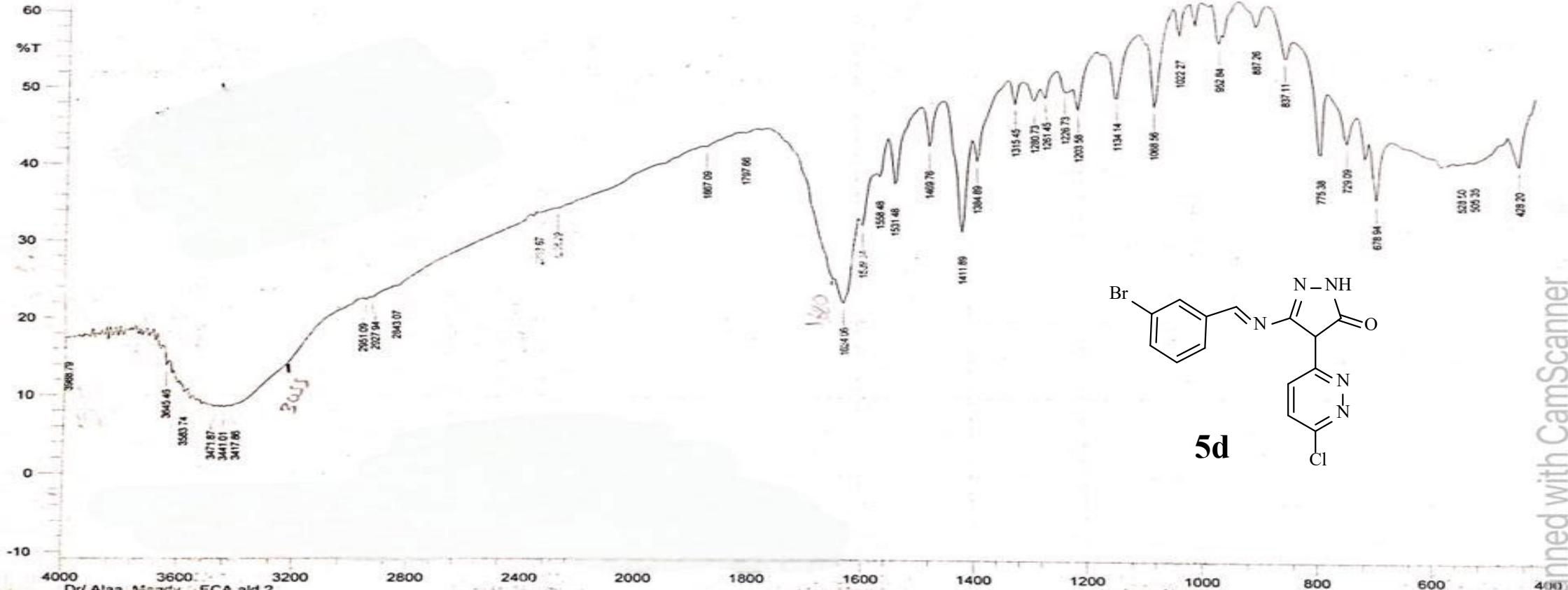
¹³C NMR of **5c** in DMSO

IR of compound 5d

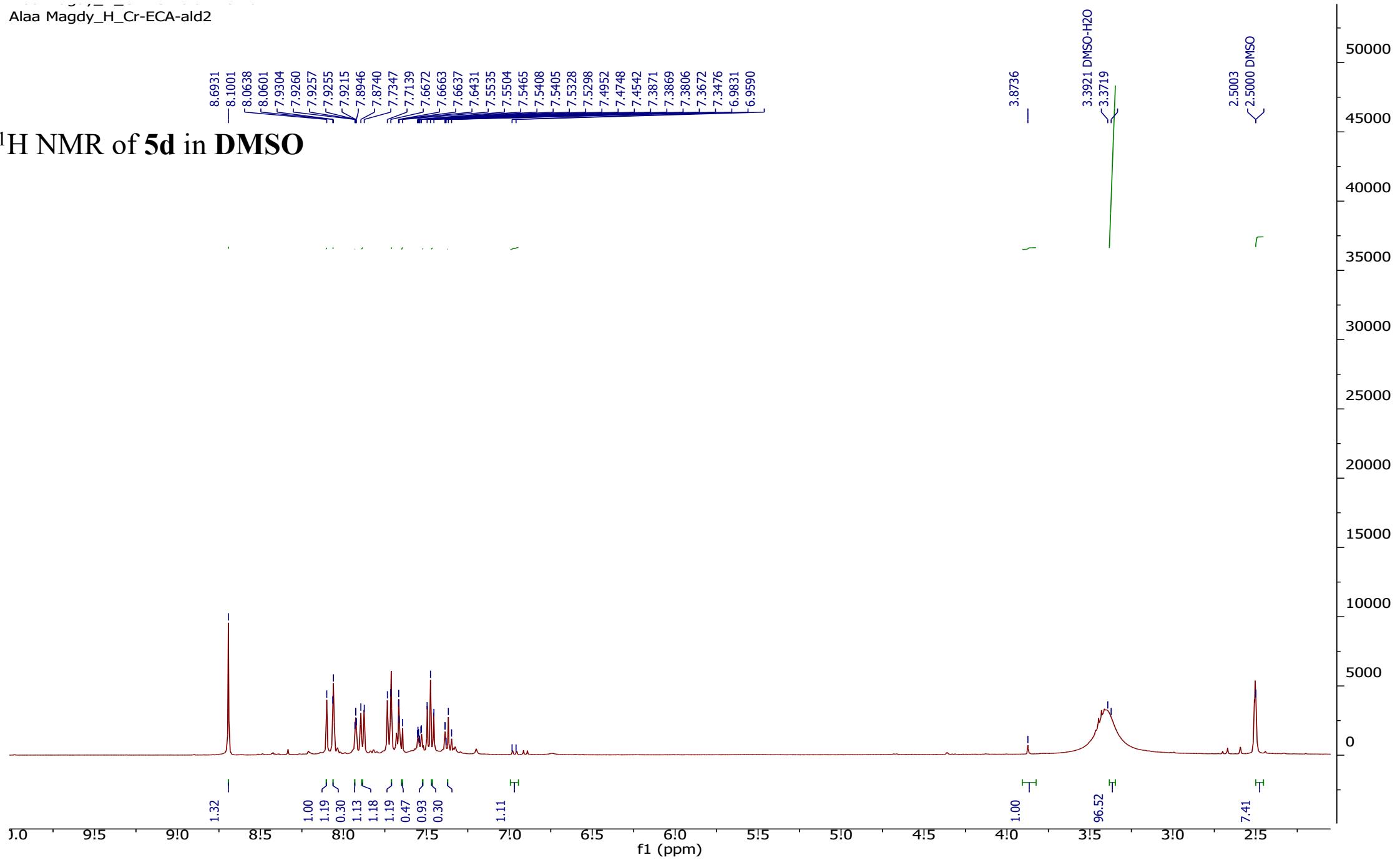


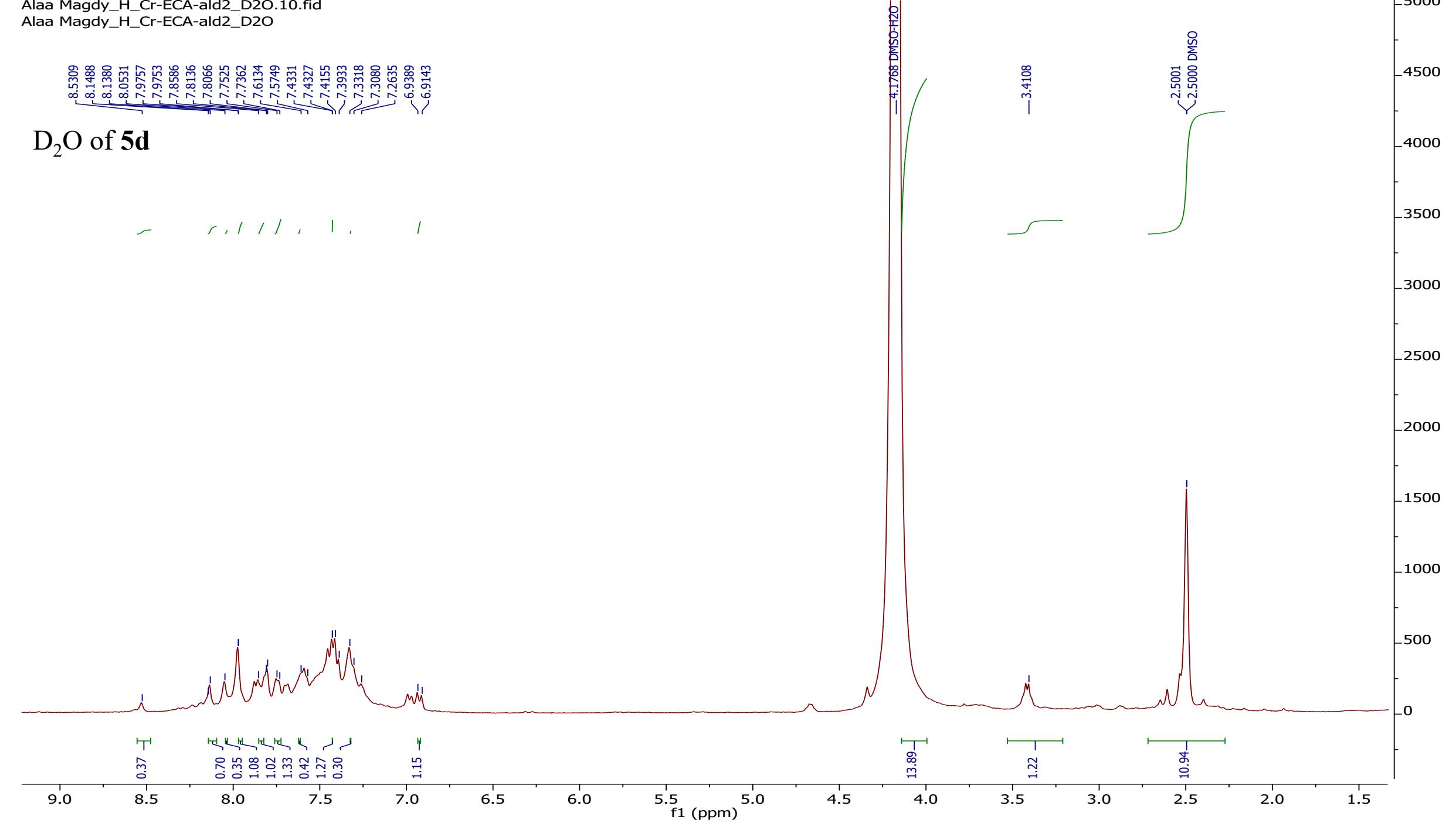
MAU
Microanalytical Unit-FOPCU
وحدة التحاليل الدقيقة
معمل الأشعة تحت الحمراء

SHIMADZU

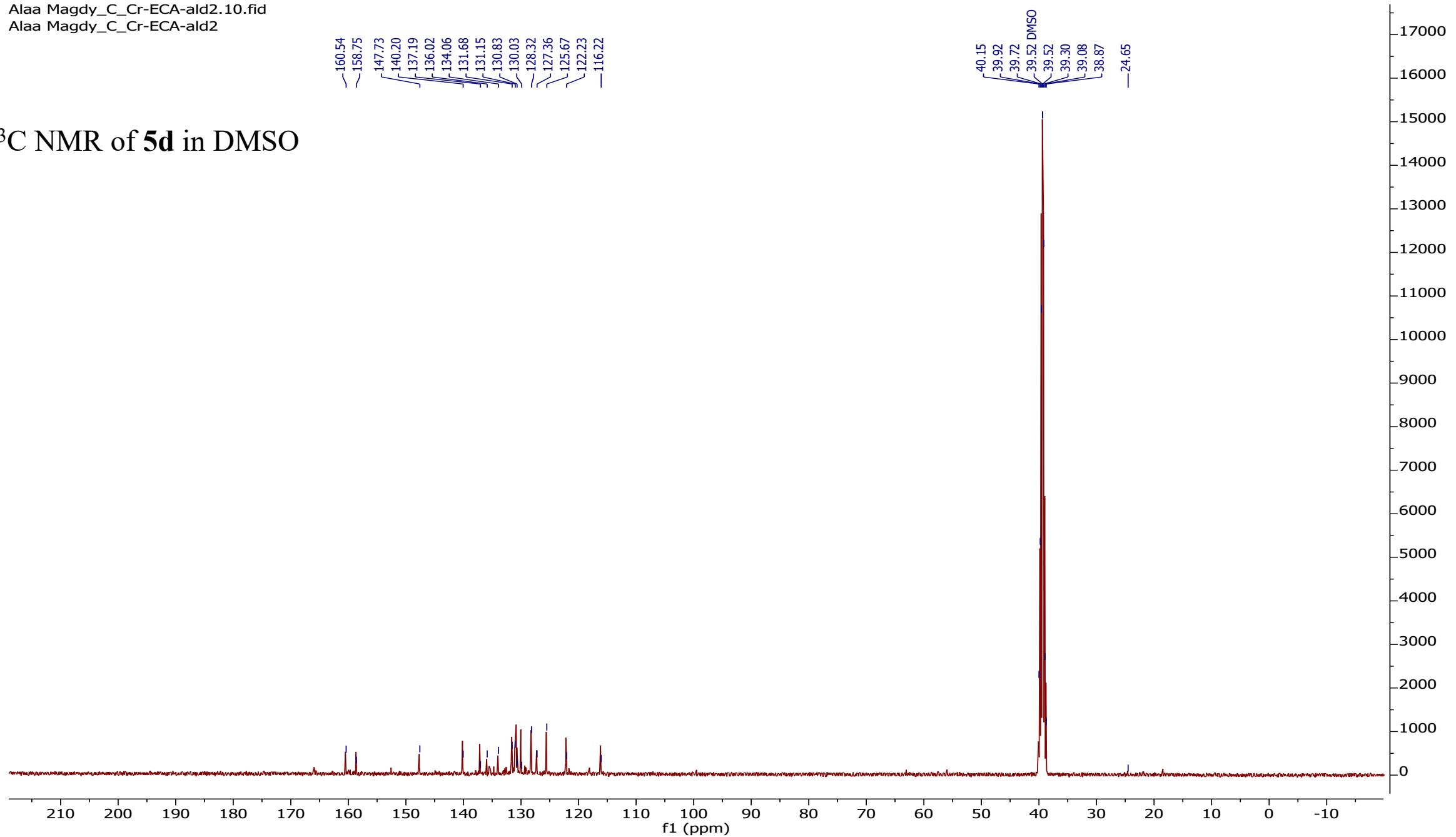


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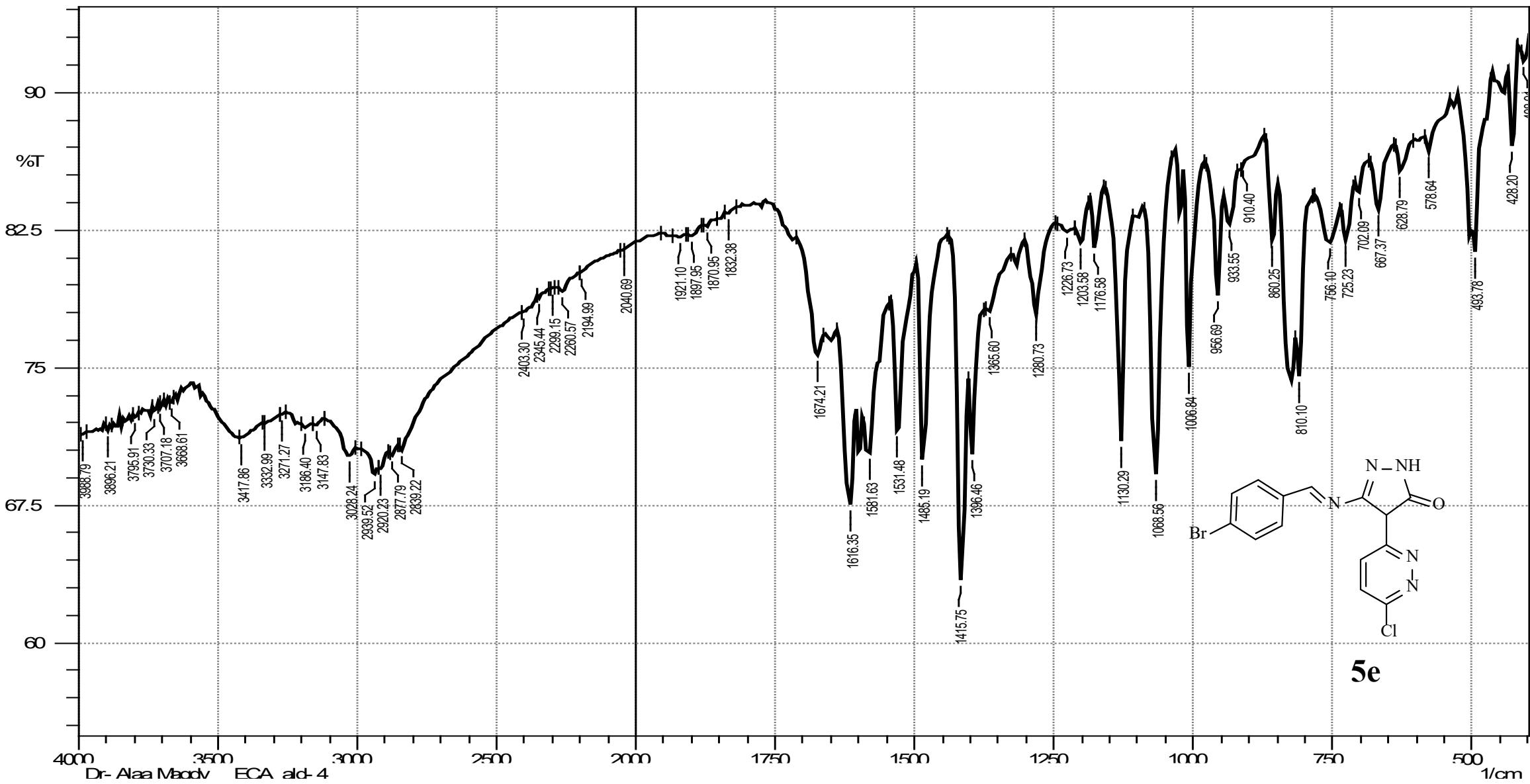
¹H NMR of 5d in DMSO

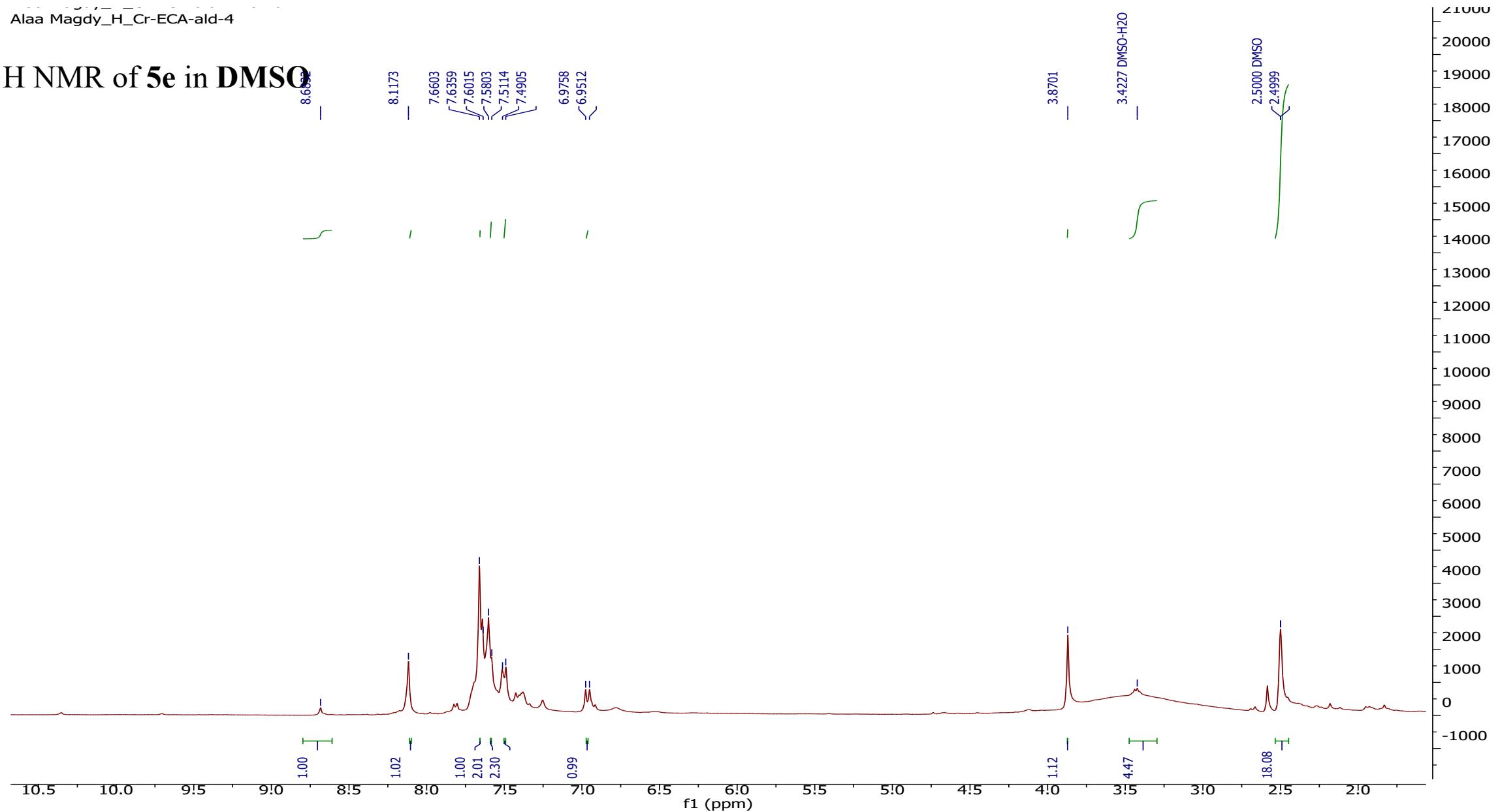


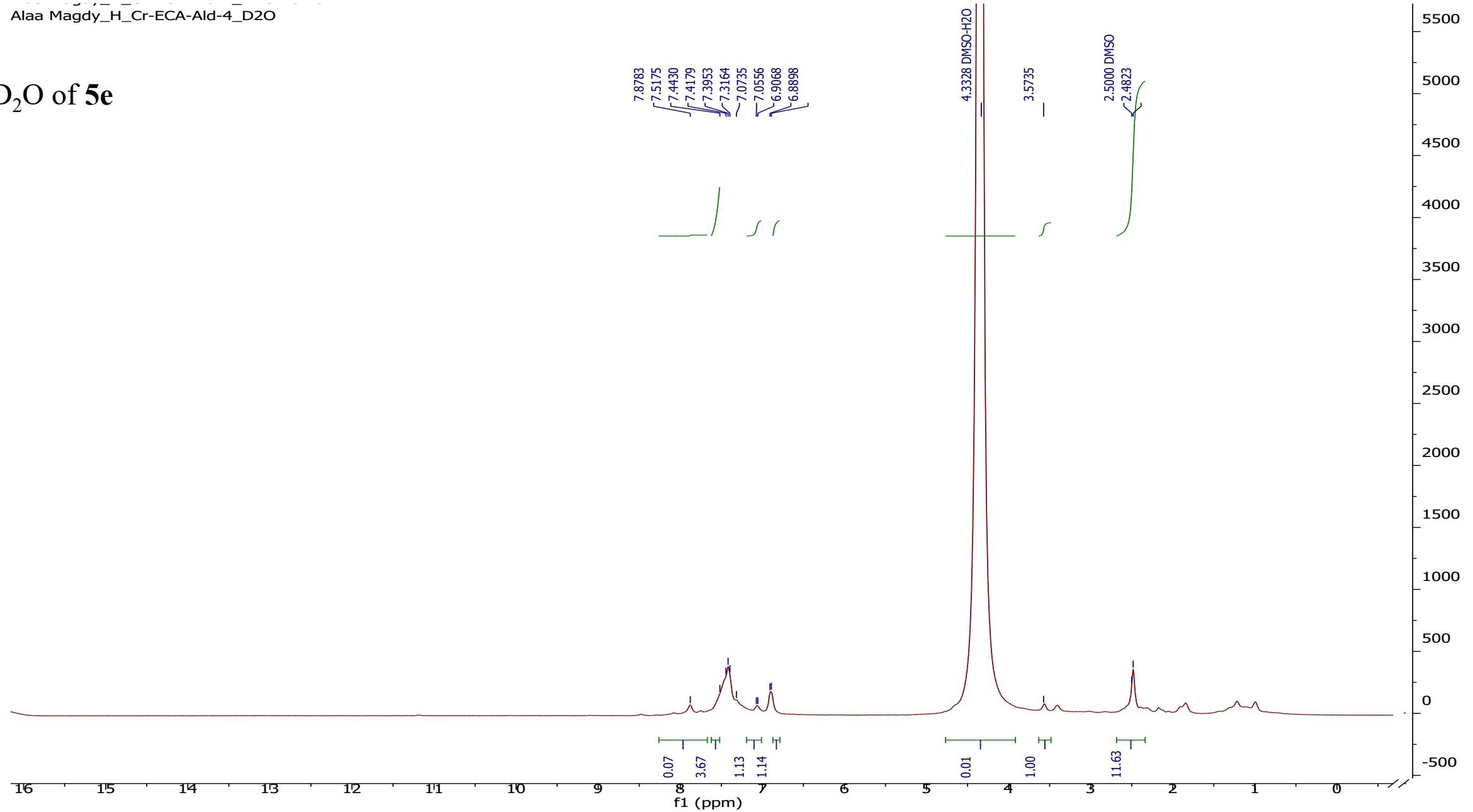
^{13}C NMR of **5d** in DMSO

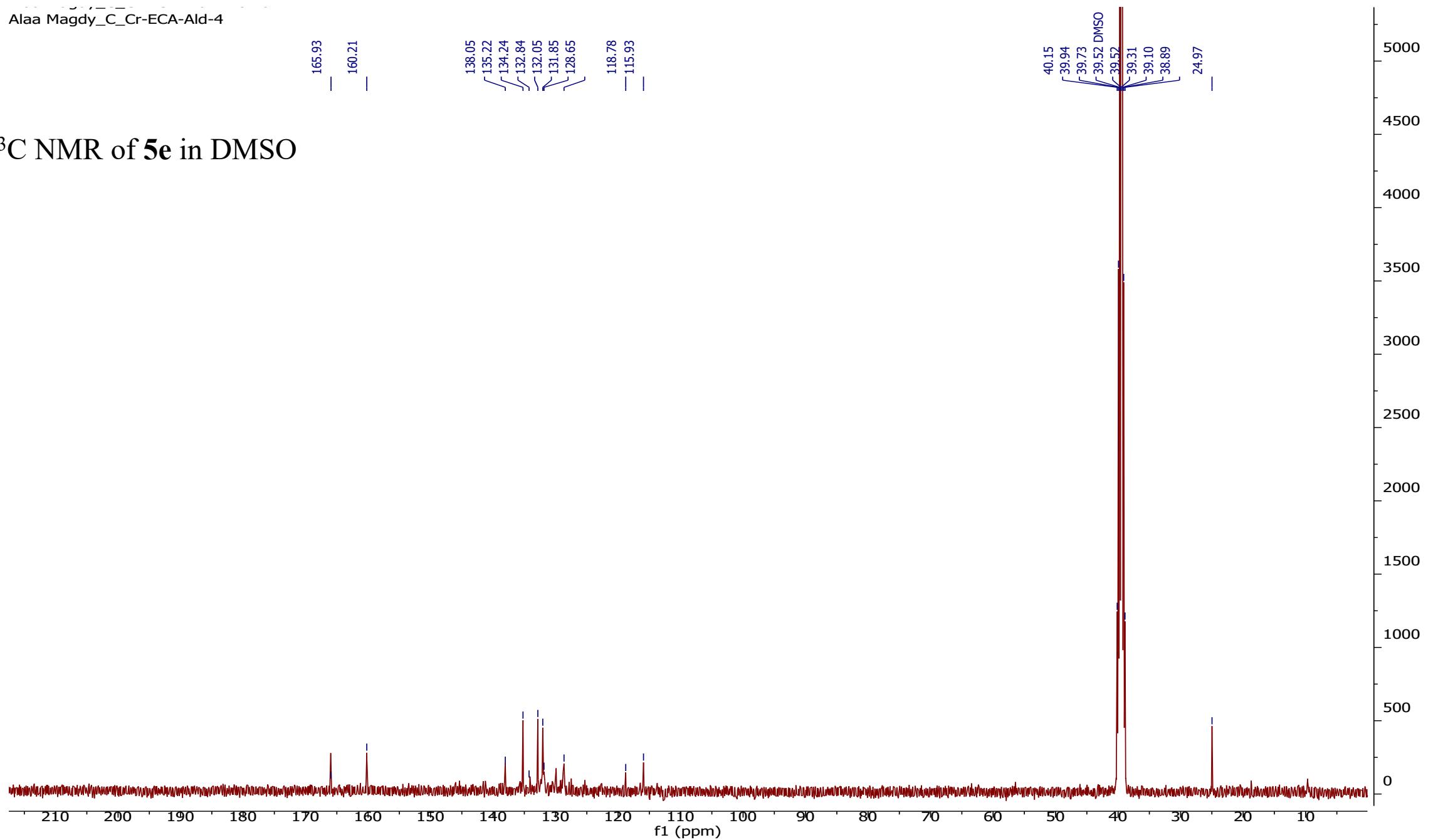


IR of compound 5e

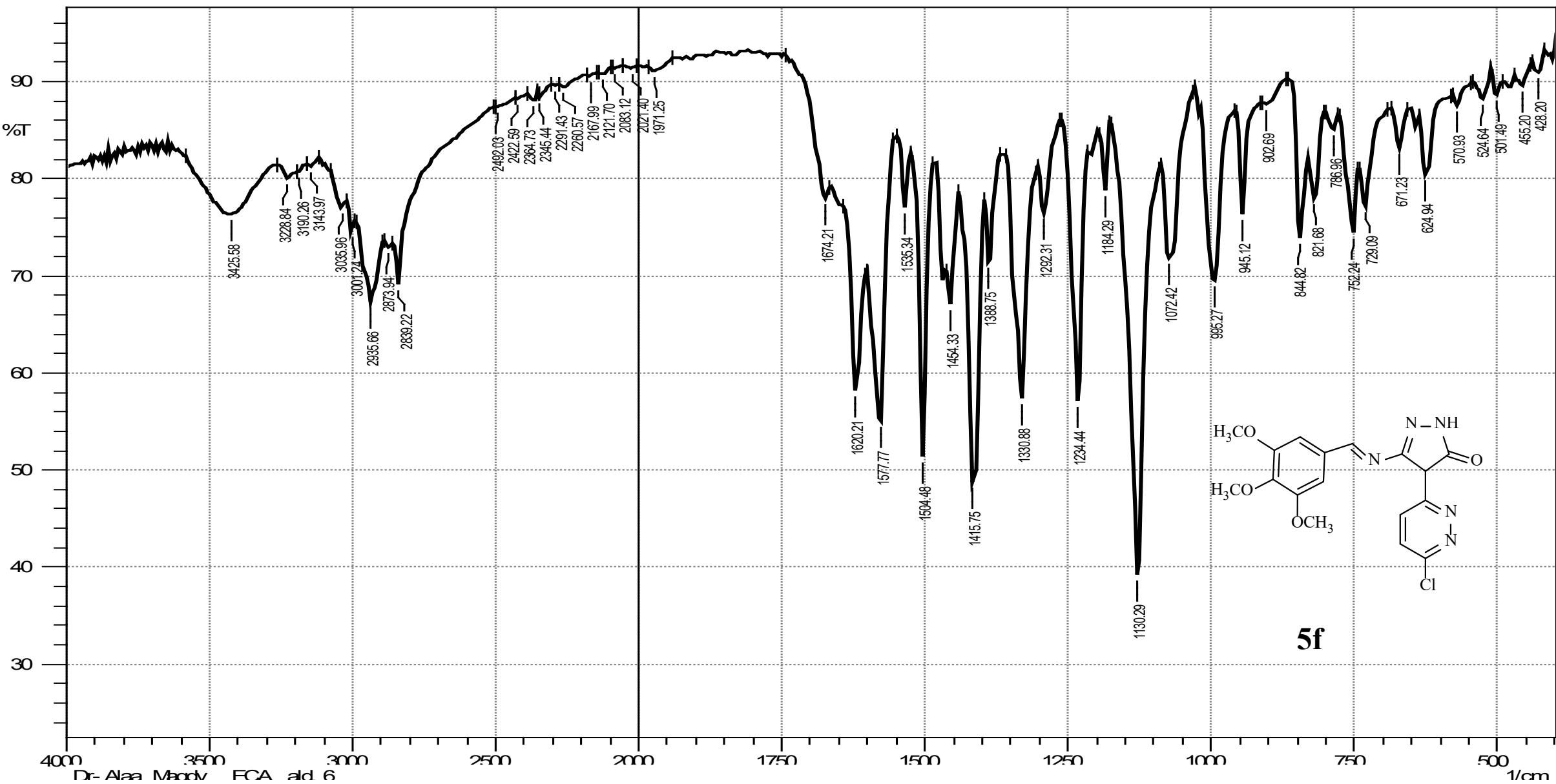


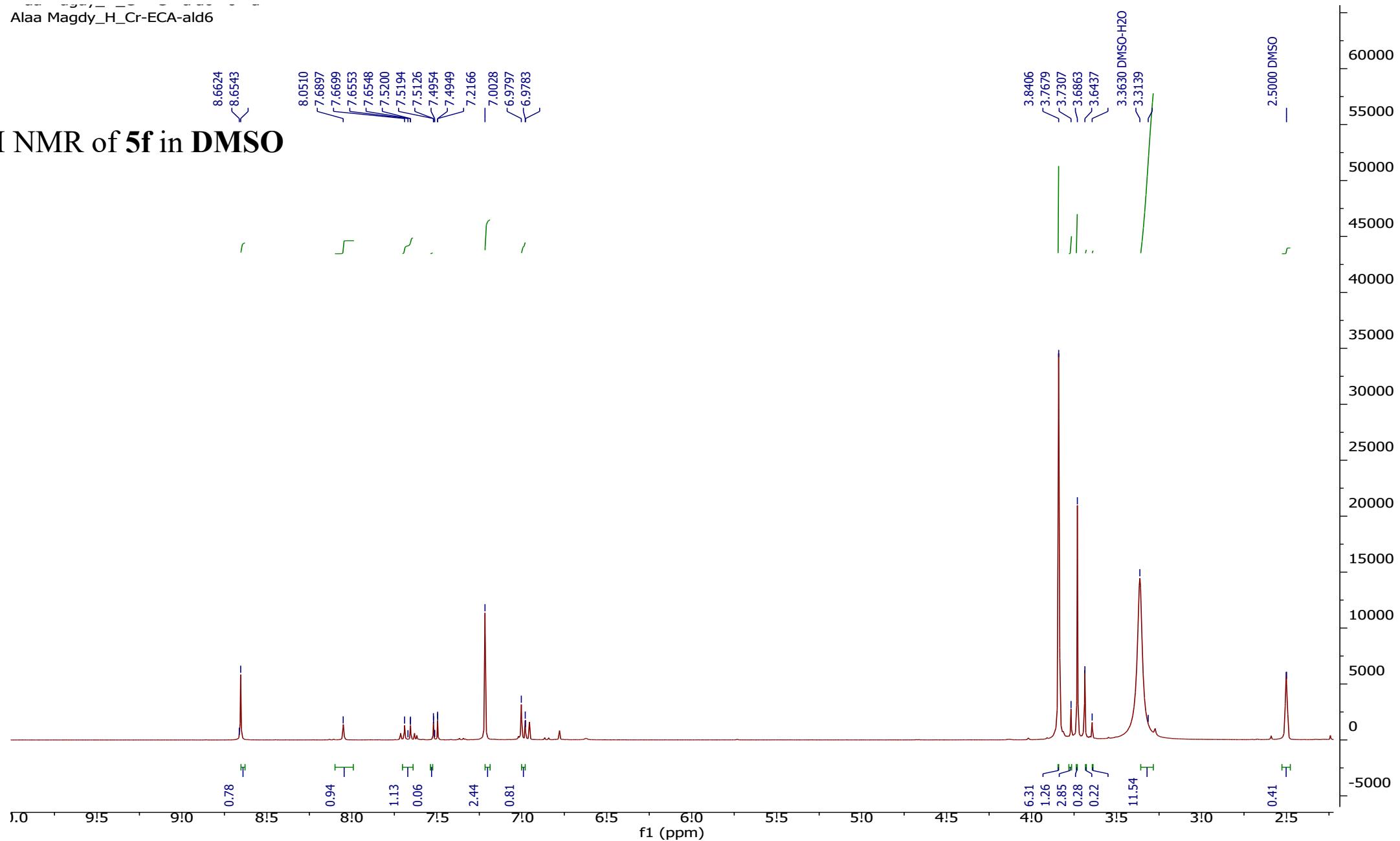
¹H NMR of 5e in DMSO

D₂O of 5e

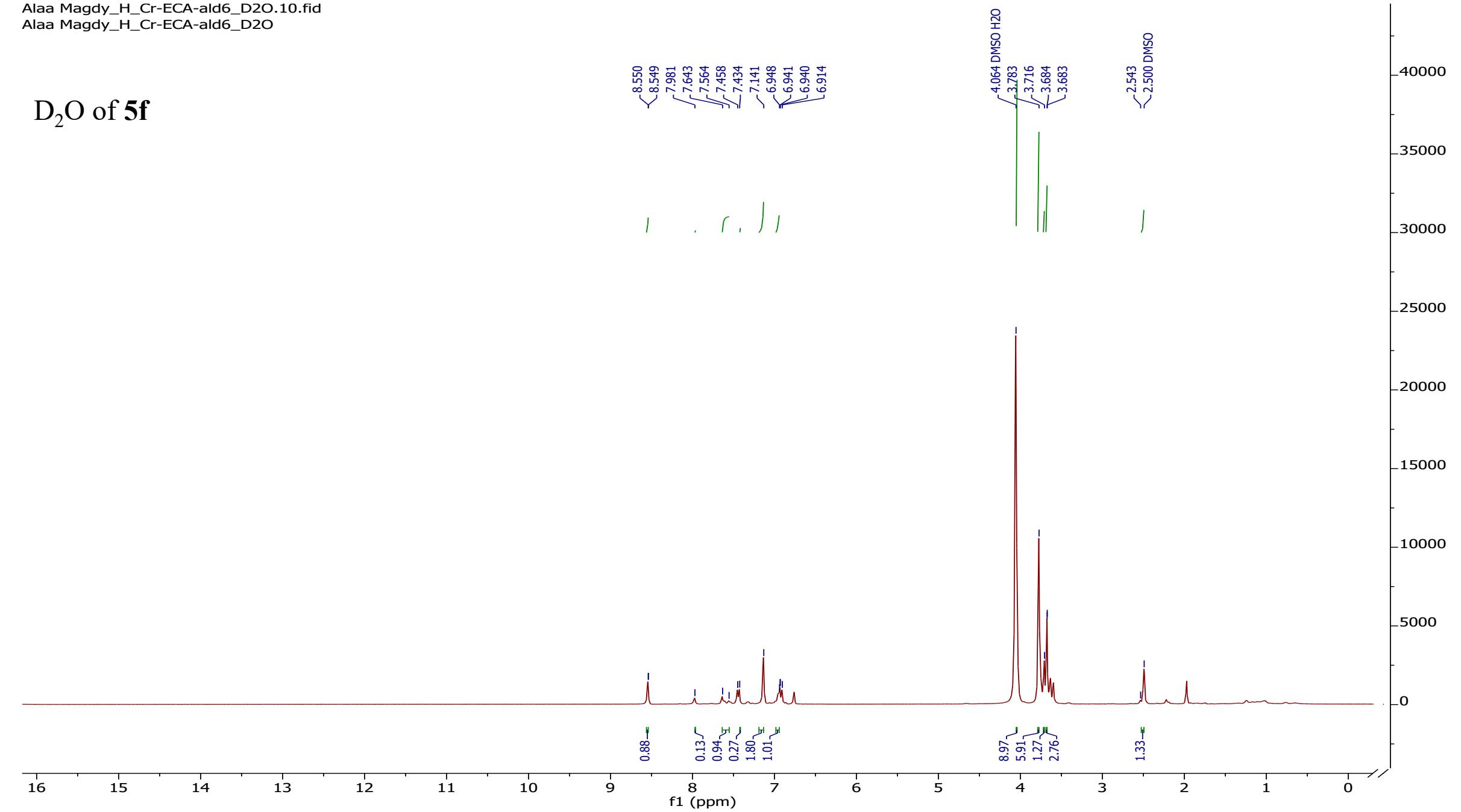
¹³C NMR of **5e** in DMSO

IR of compound 5f

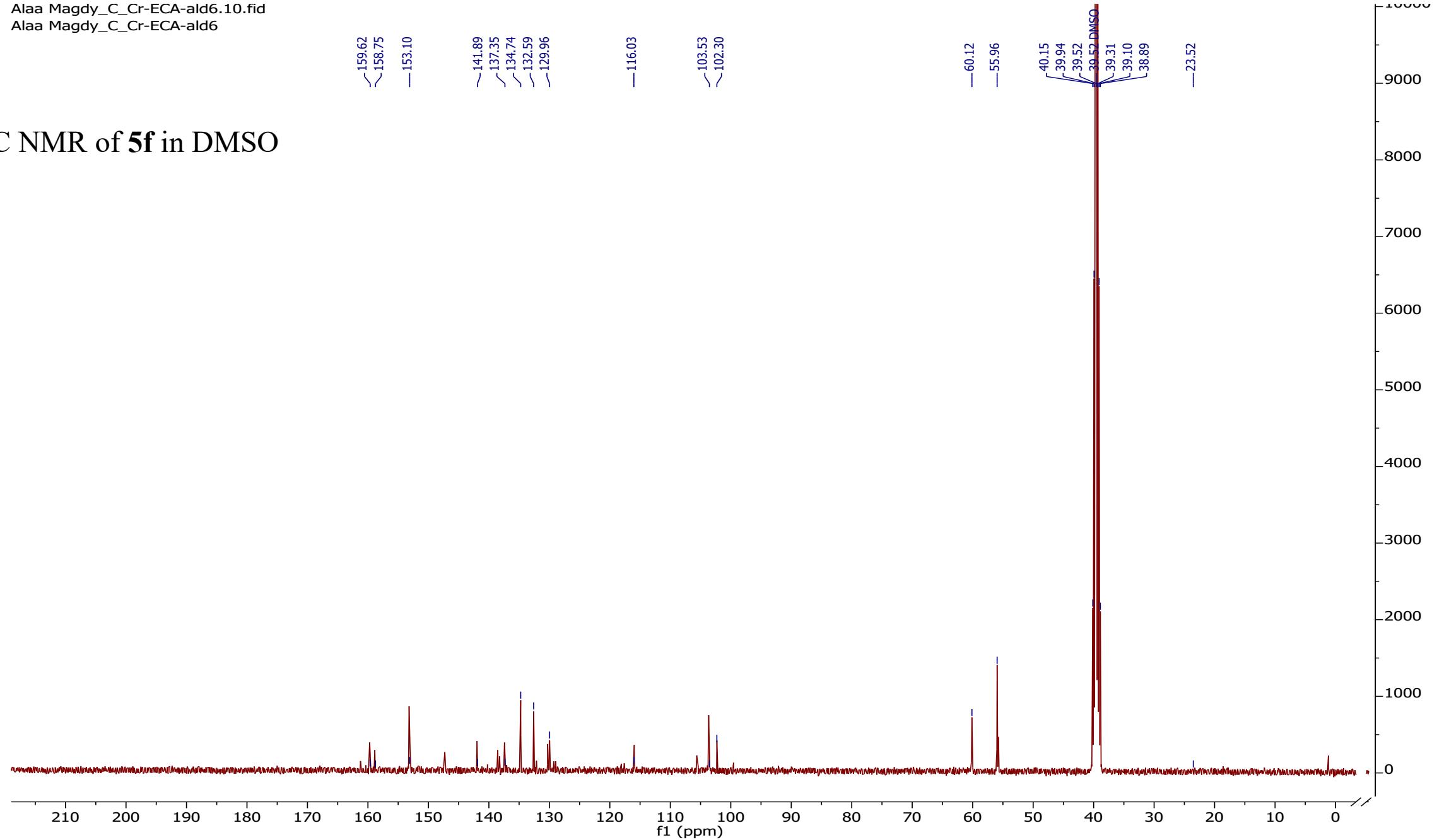


¹H NMR of 5f in DMSO

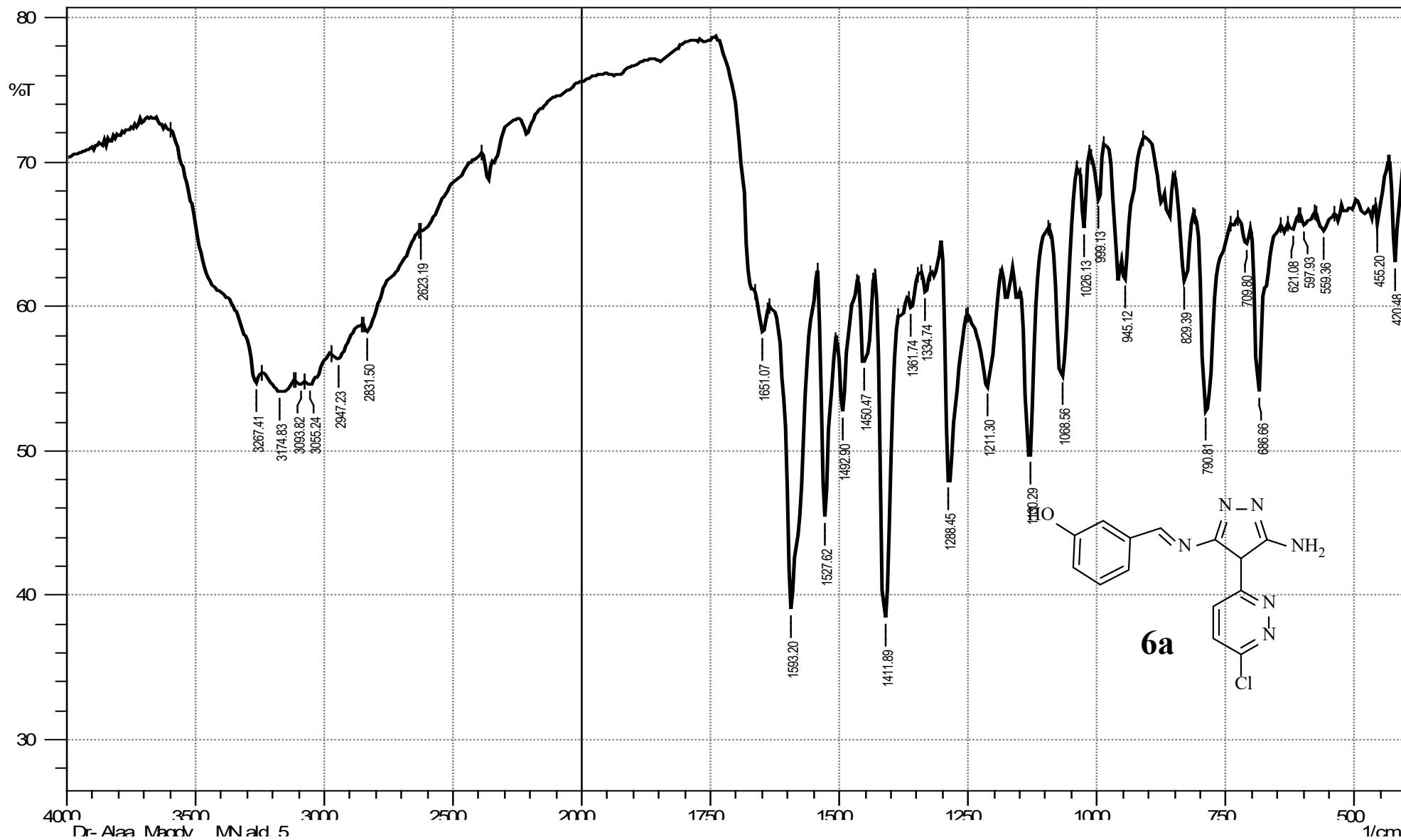
D₂O of 5f

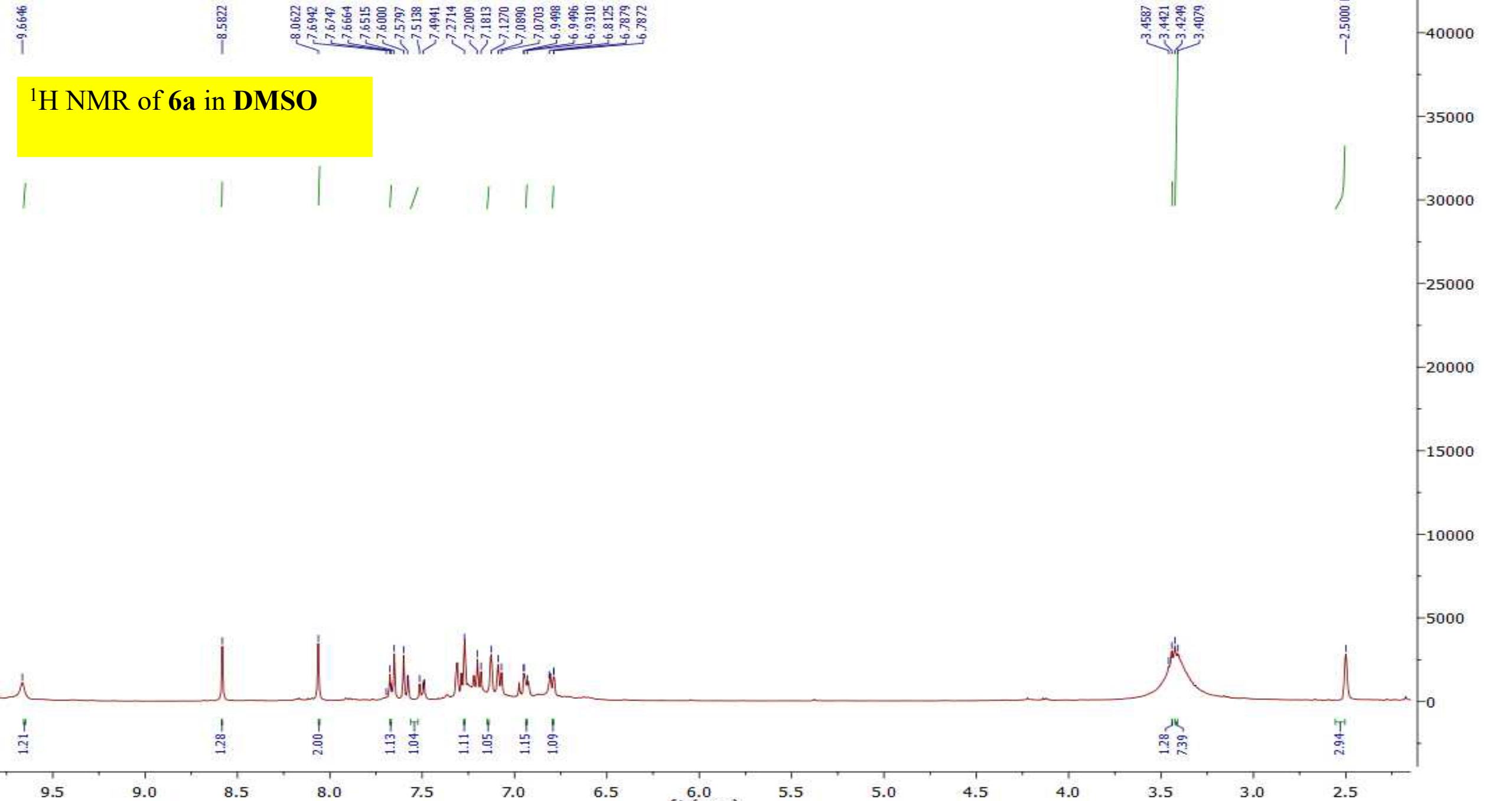


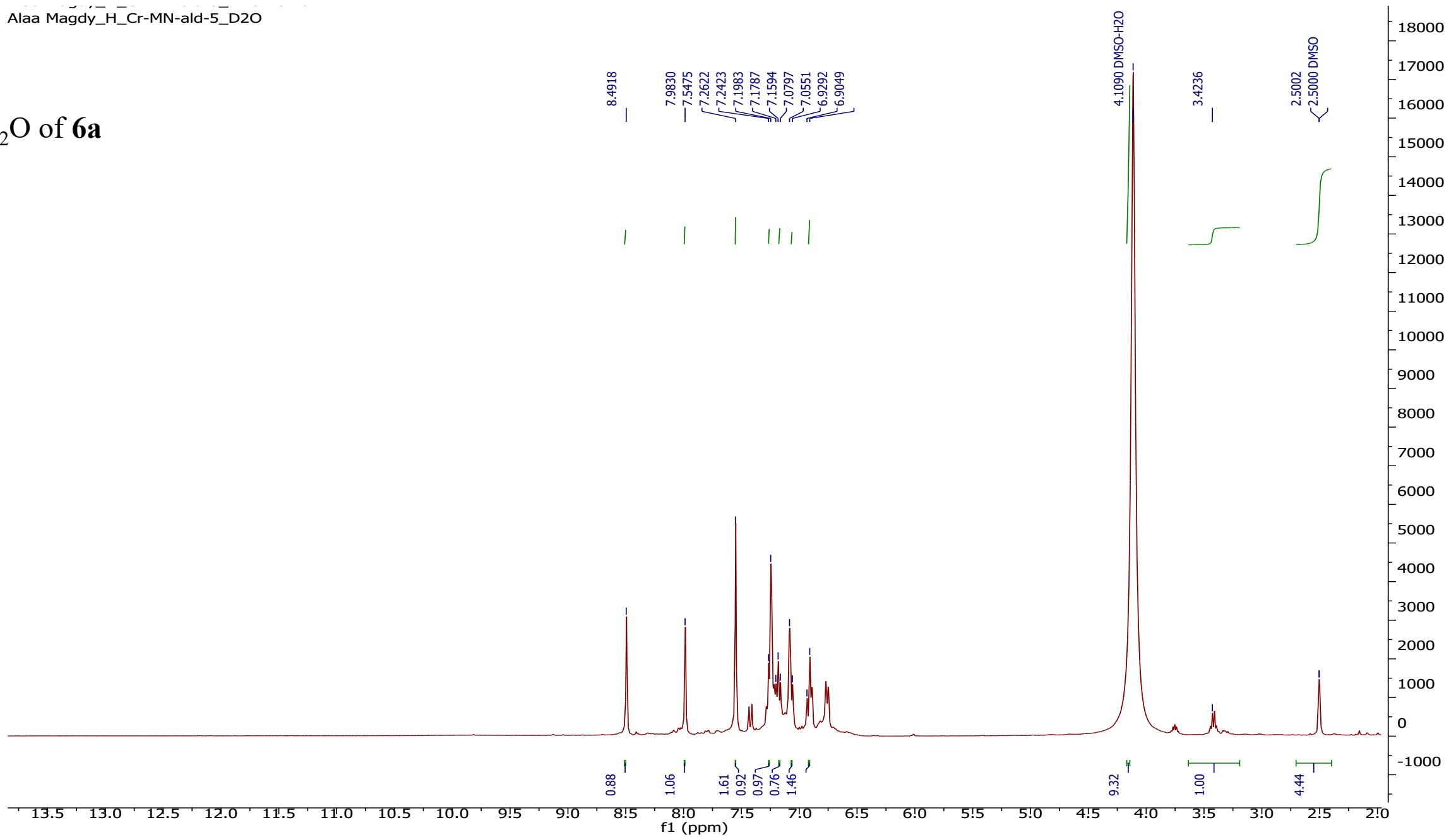
¹³C NMR of **5f** in DMSO

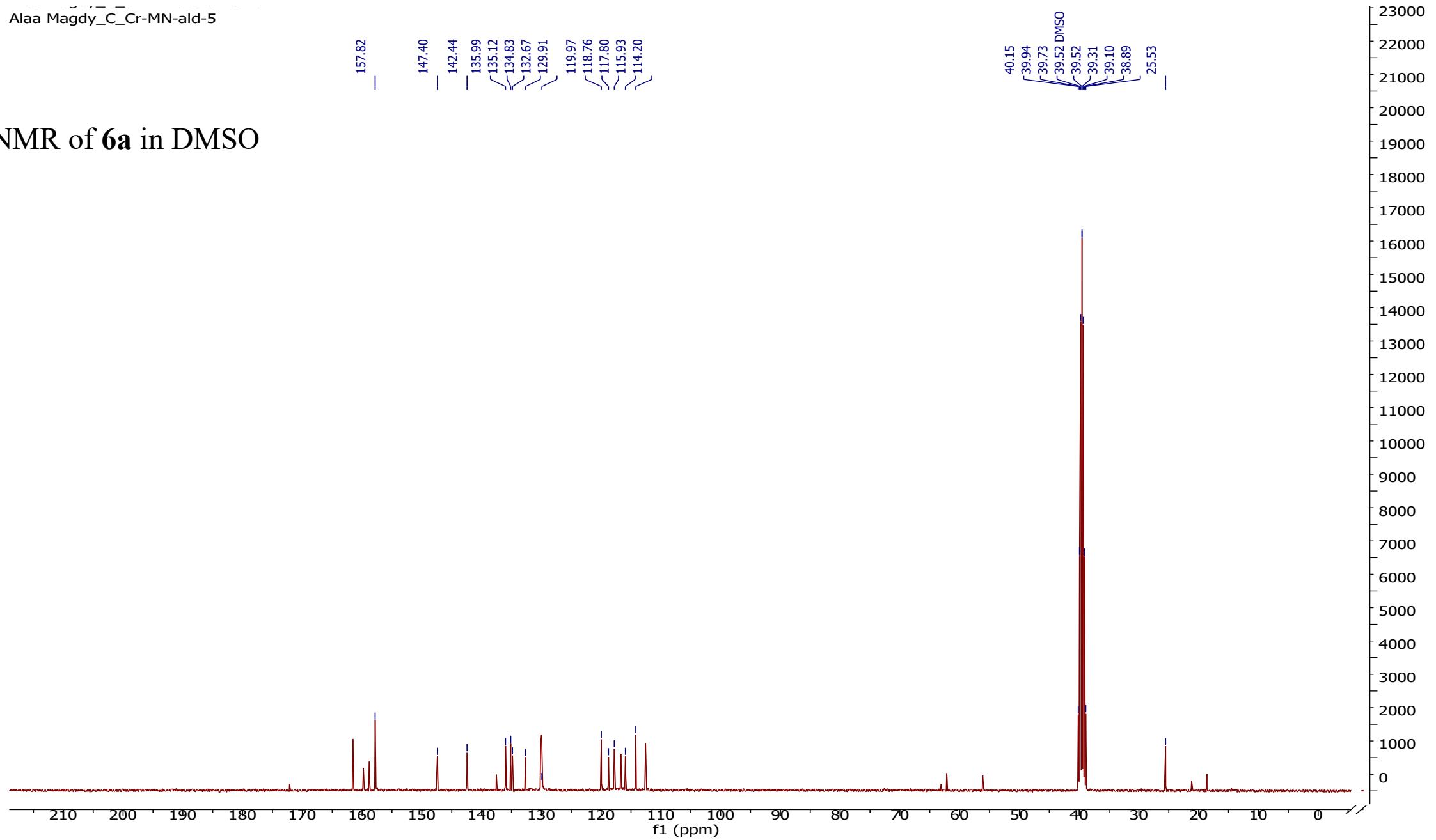


IR of compound 6a





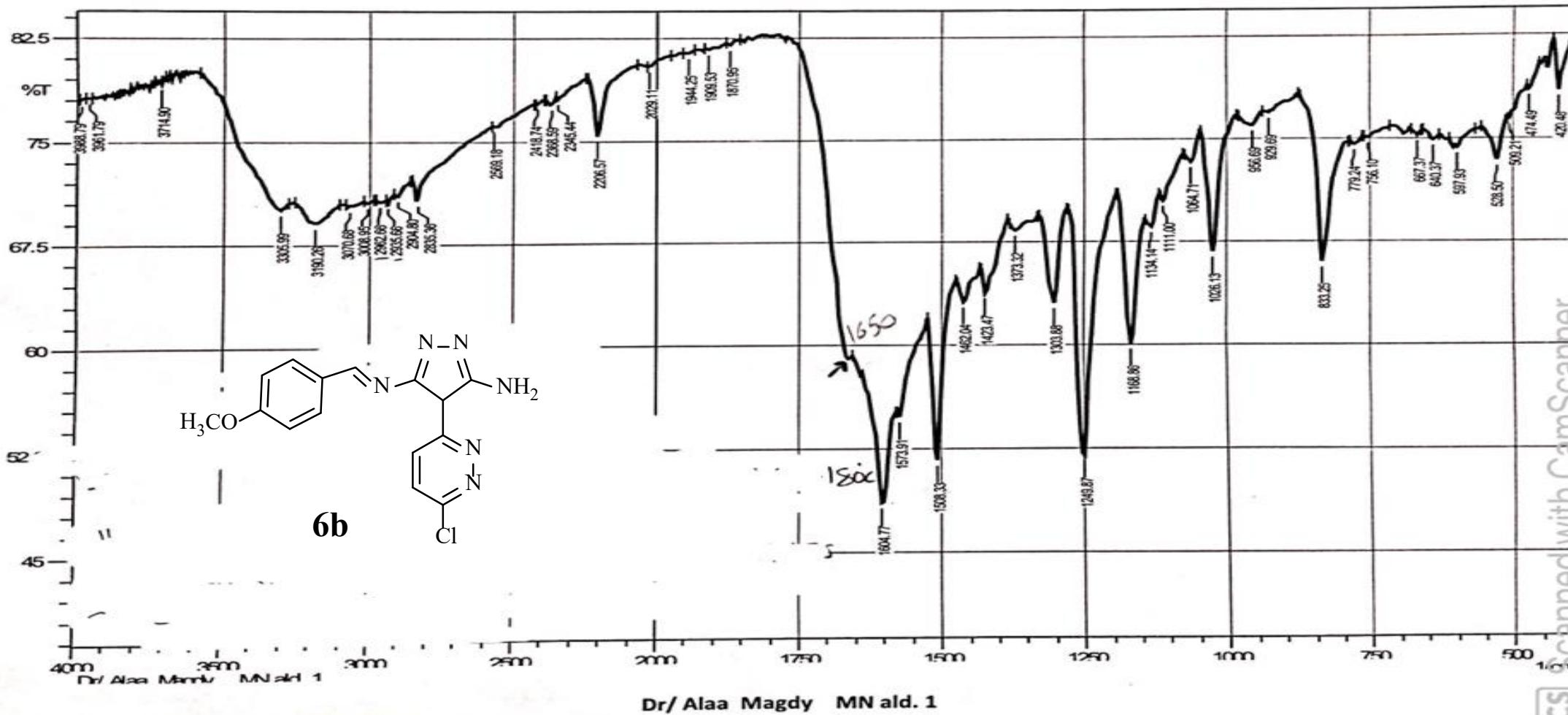
D₂O of 6a

¹³C NMR of **6a** in DMSO

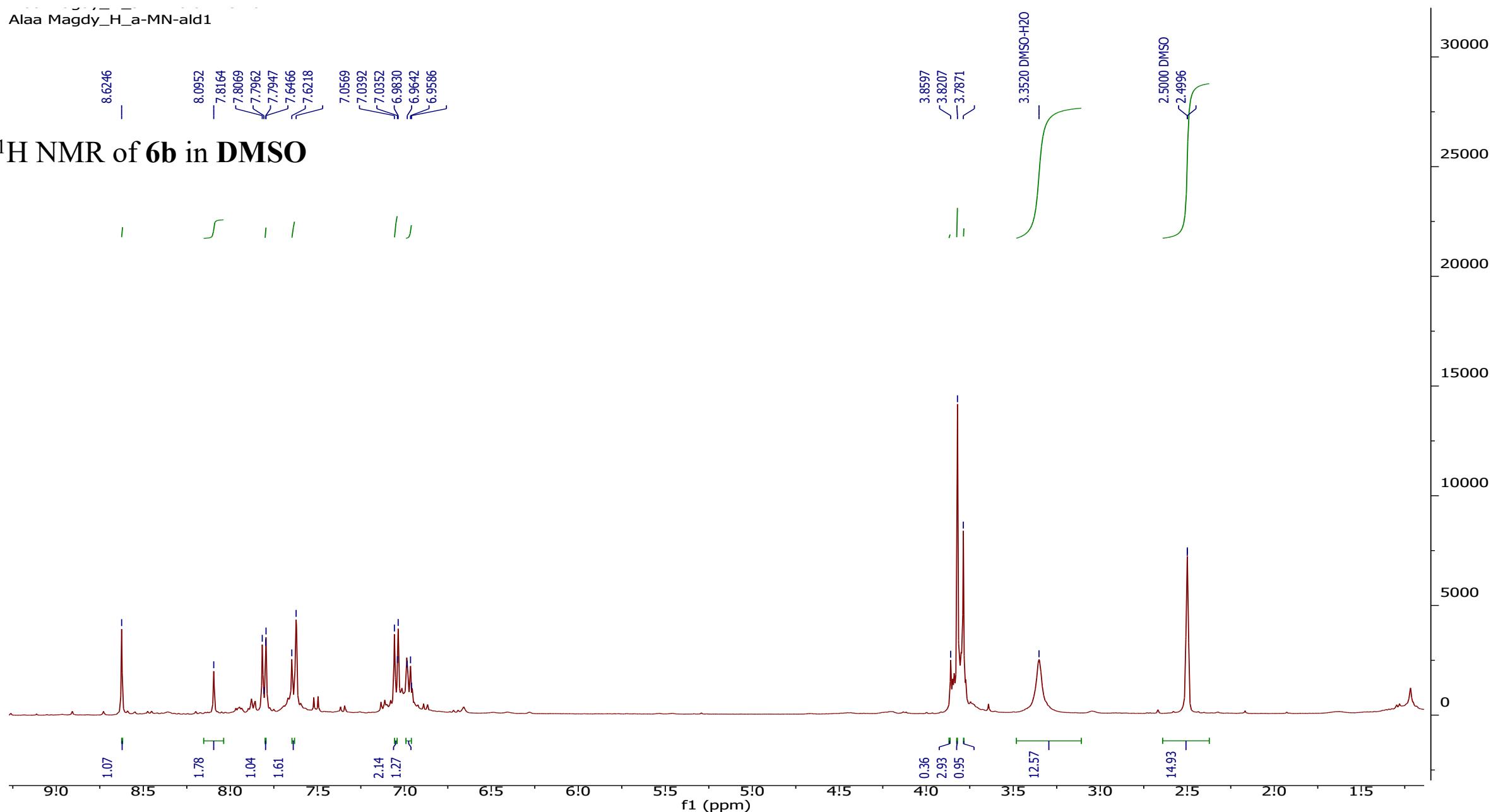
IR of compound 6b



MAW

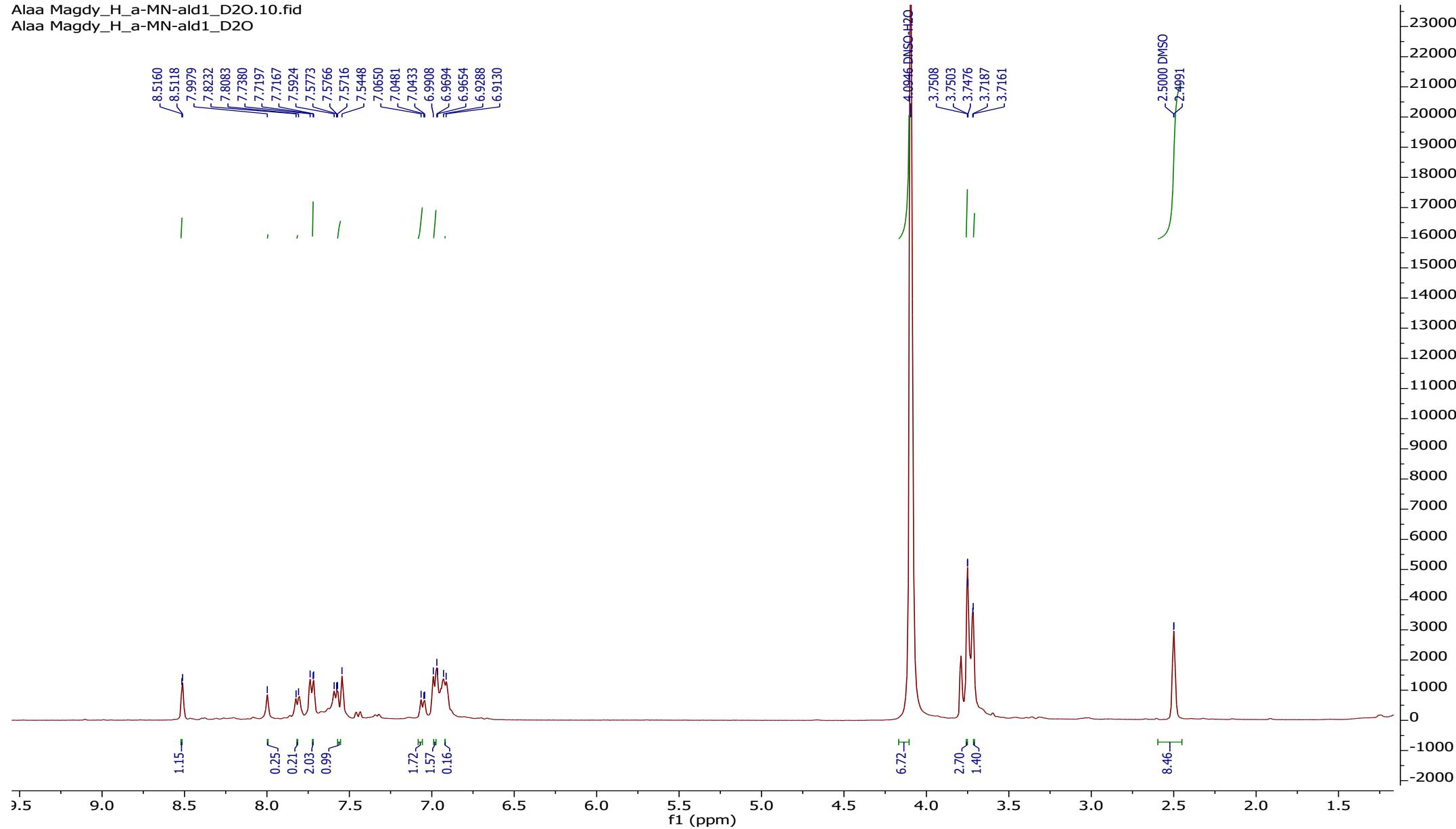


Scanned with CamScanner

¹H NMR of 6b in DMSO

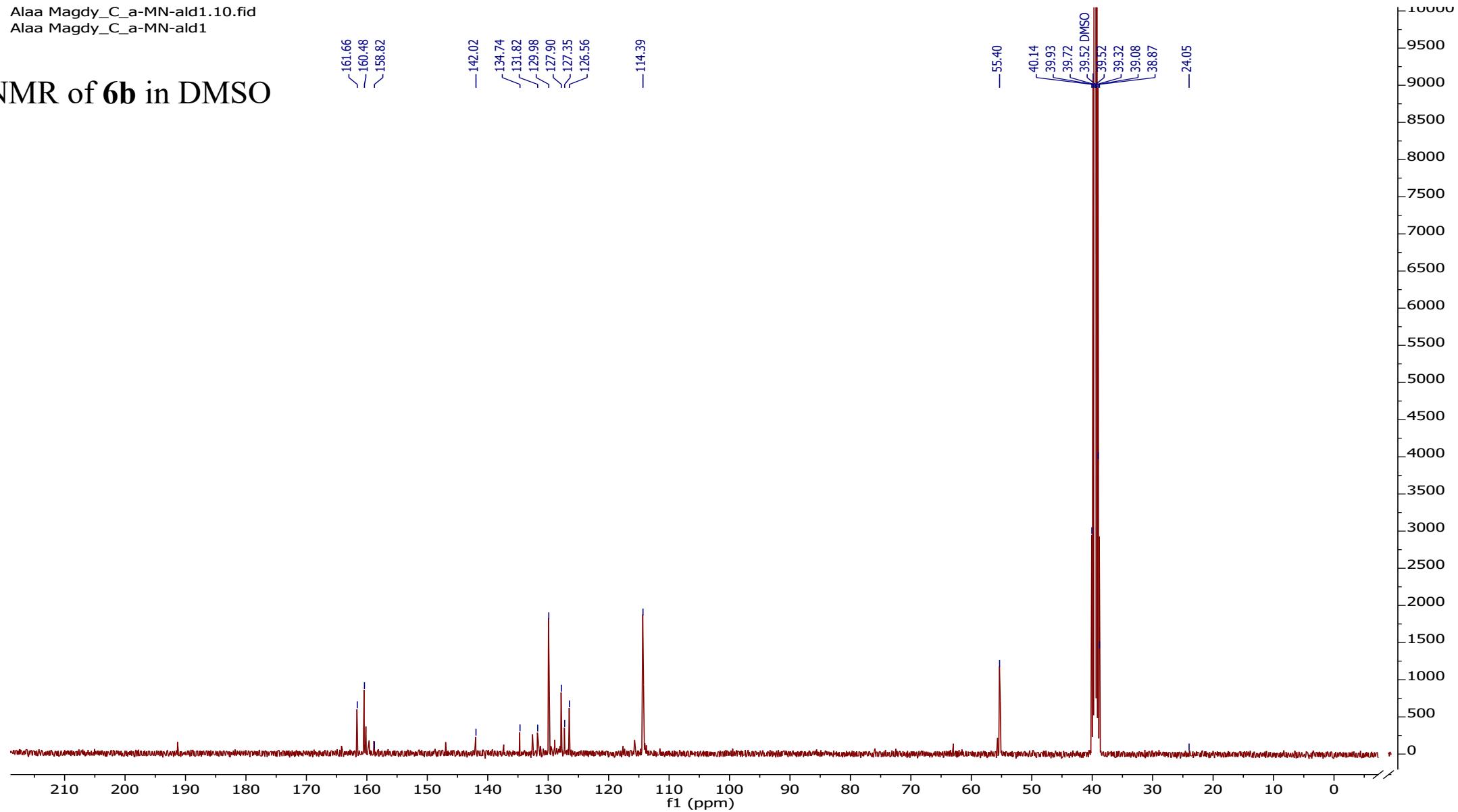
D₂O of 6b

Alaa Magdy_H_a-MN-ald1_D2O.10.fid
Alaa Magdy_H_a-MN-ald1_D2O

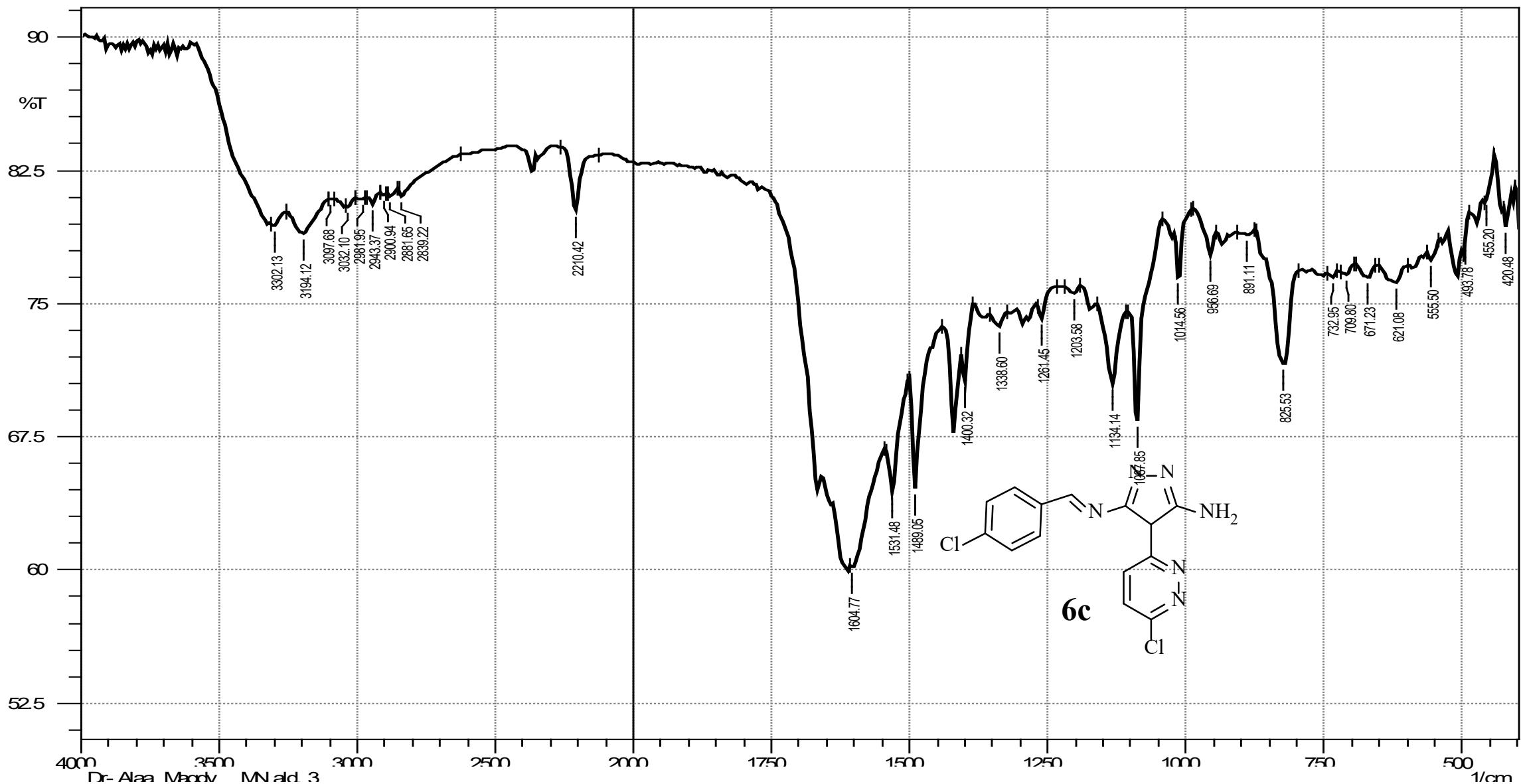


Alaa Magdy_C_a-MN-ald1.10.fid
Alaa Magdy_C_a-MN-ald1

¹³C NMR of **6b** in DMSO

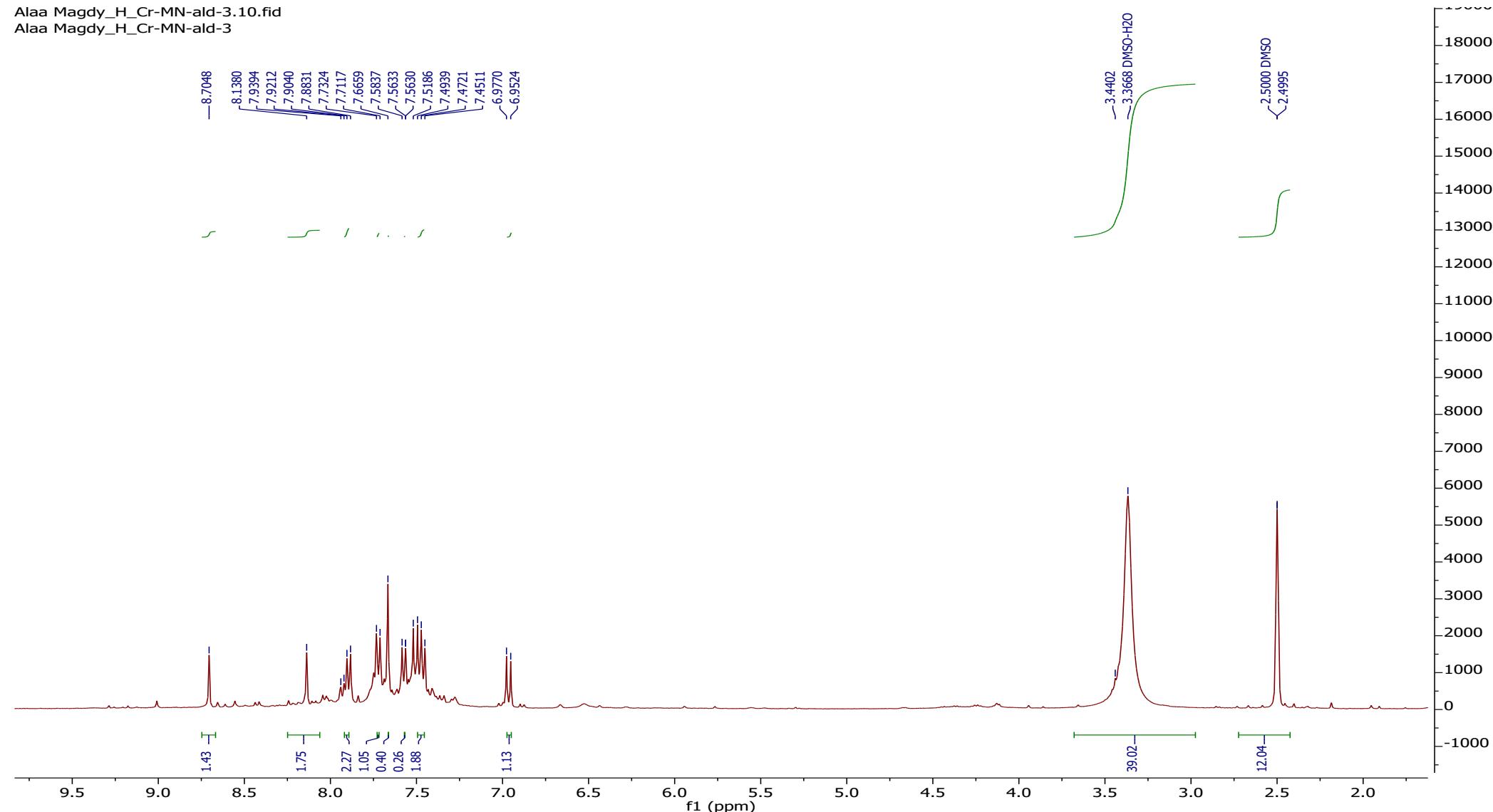


IR of compound **6c**

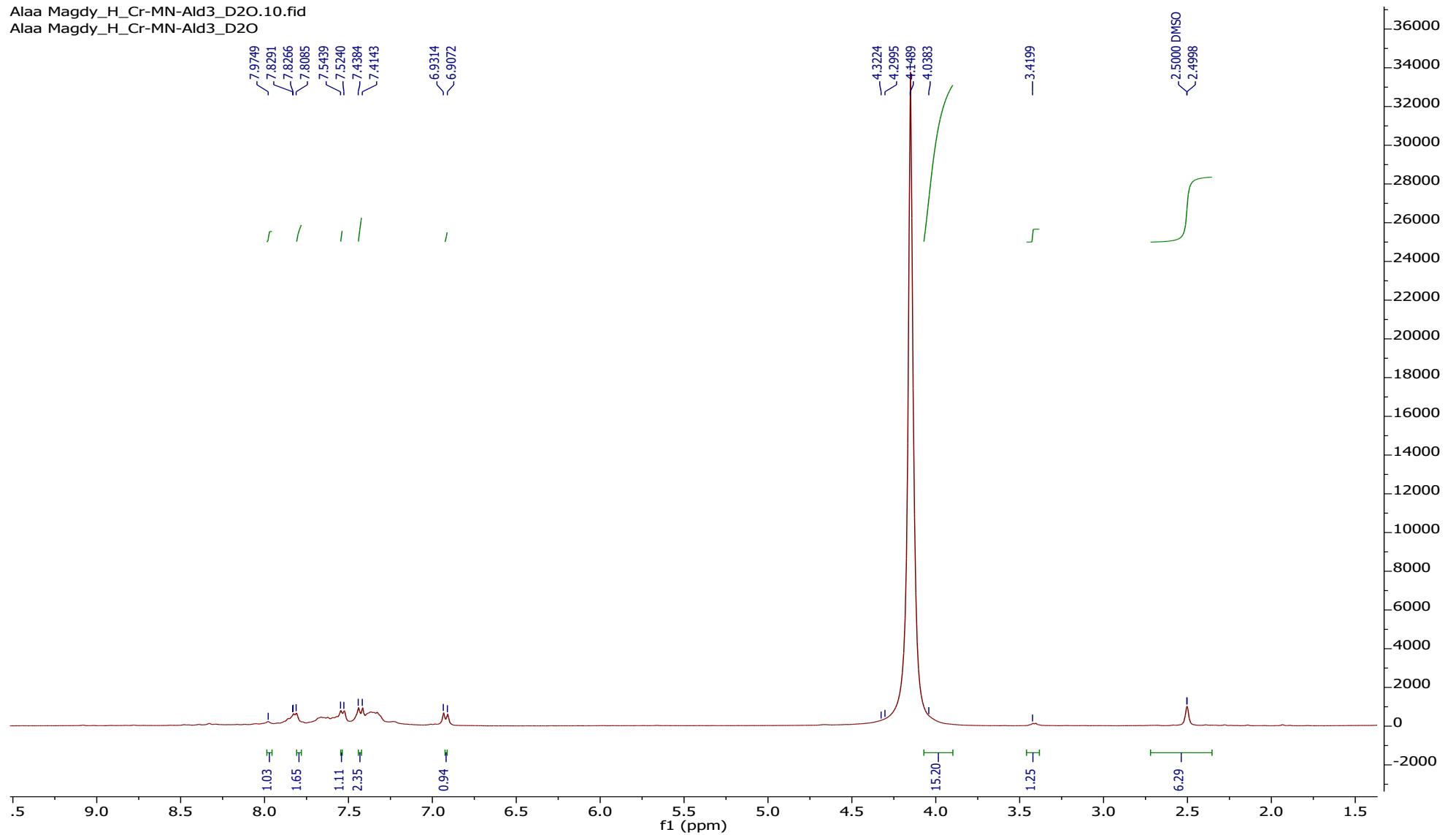


¹H NMR of 6c in DMSO

Alaa Magdy_H_Cr-MN-ald-3.10.fid
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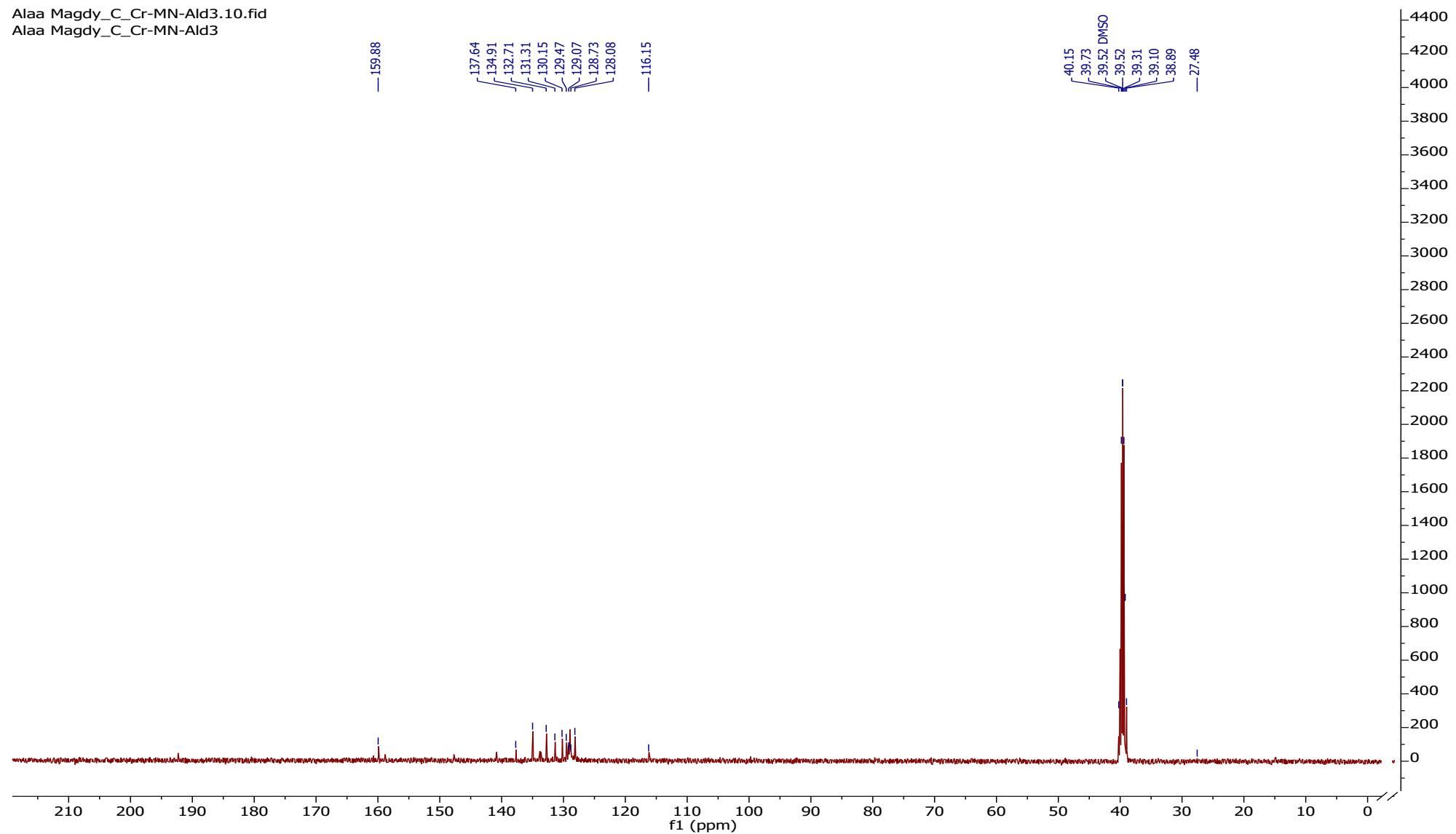


D₂O of 6c

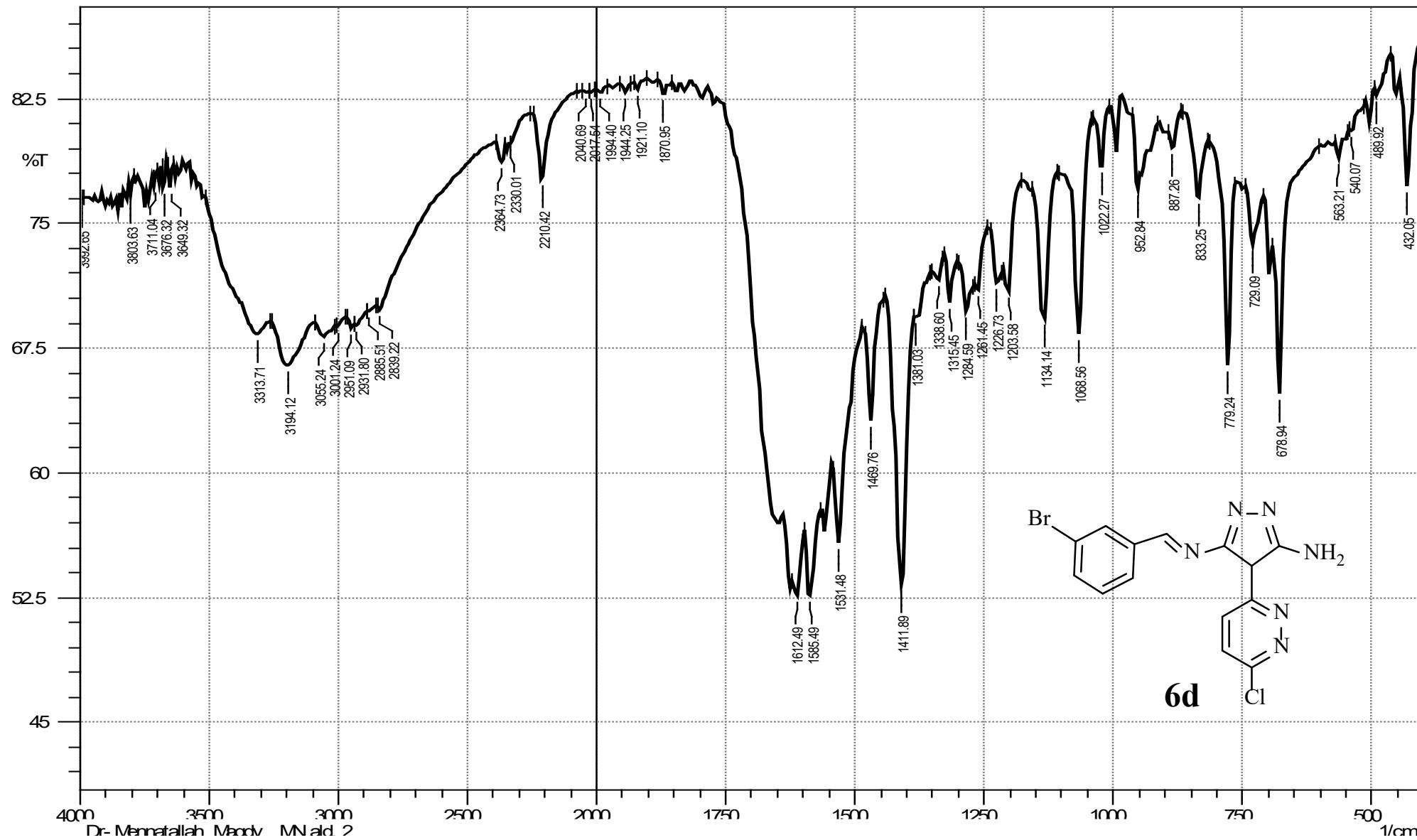


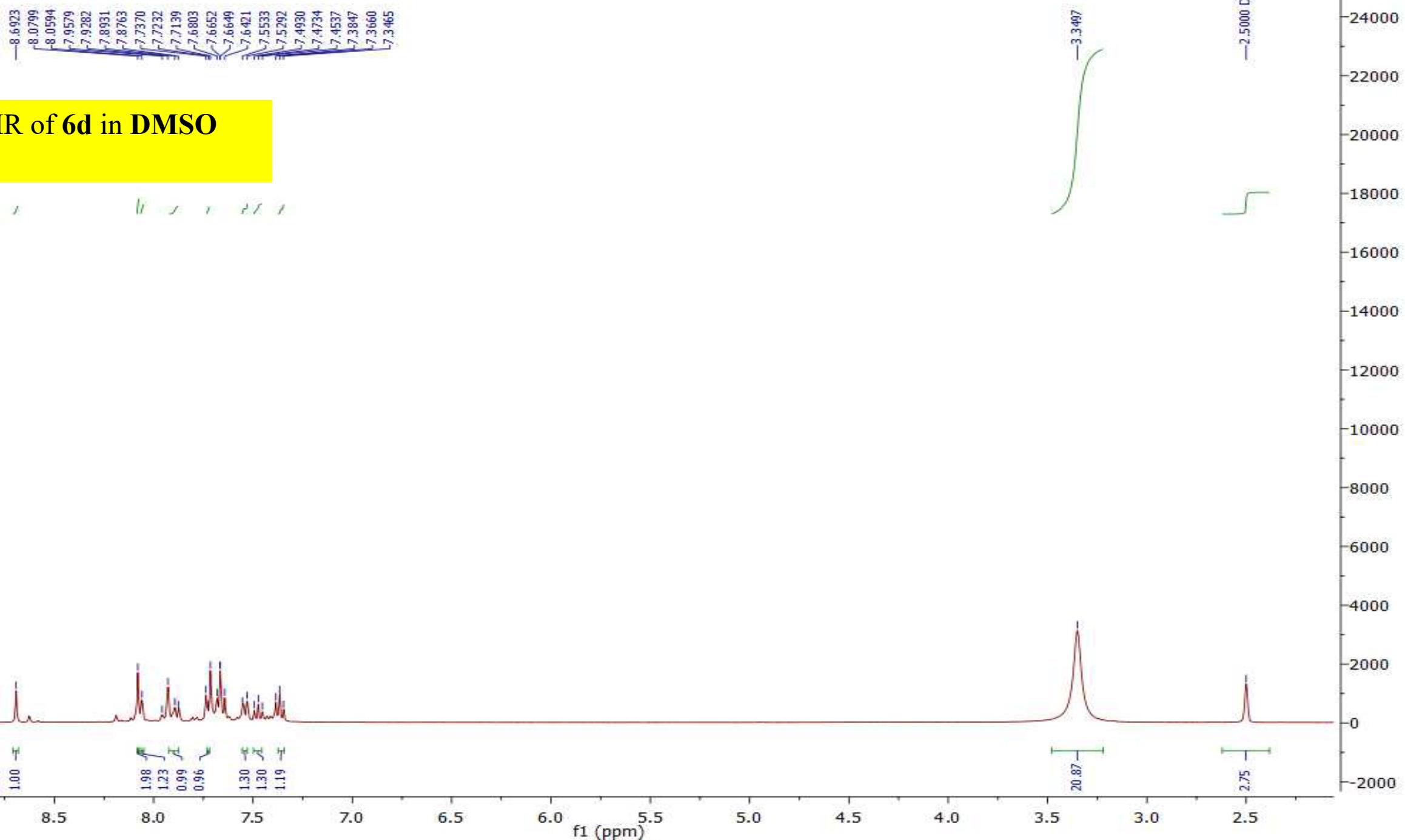
¹³C NMR of **6c** in DMSO

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Alaa Magdy_C_Cr-MN-Ald3

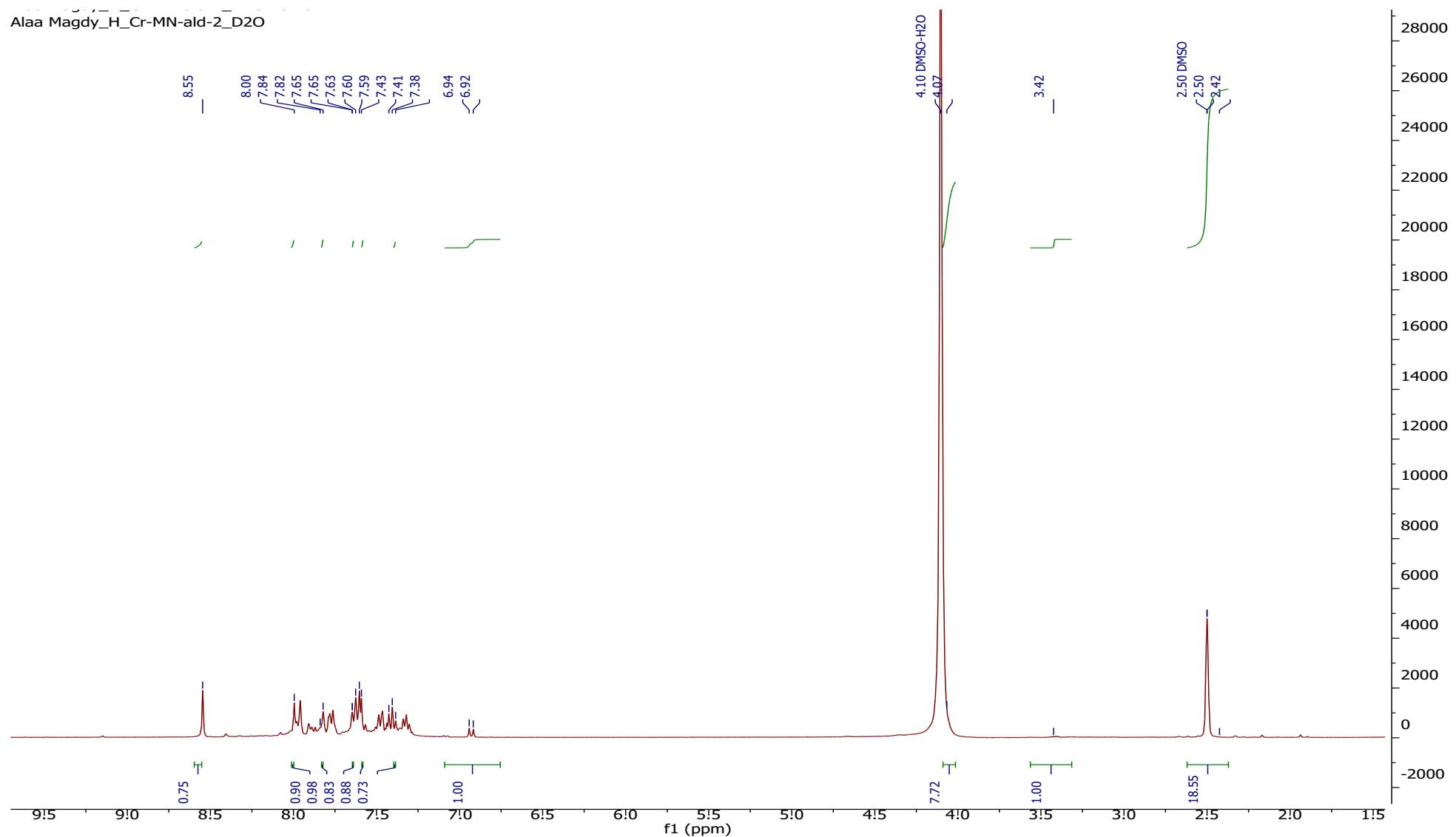


IR of compound 6d



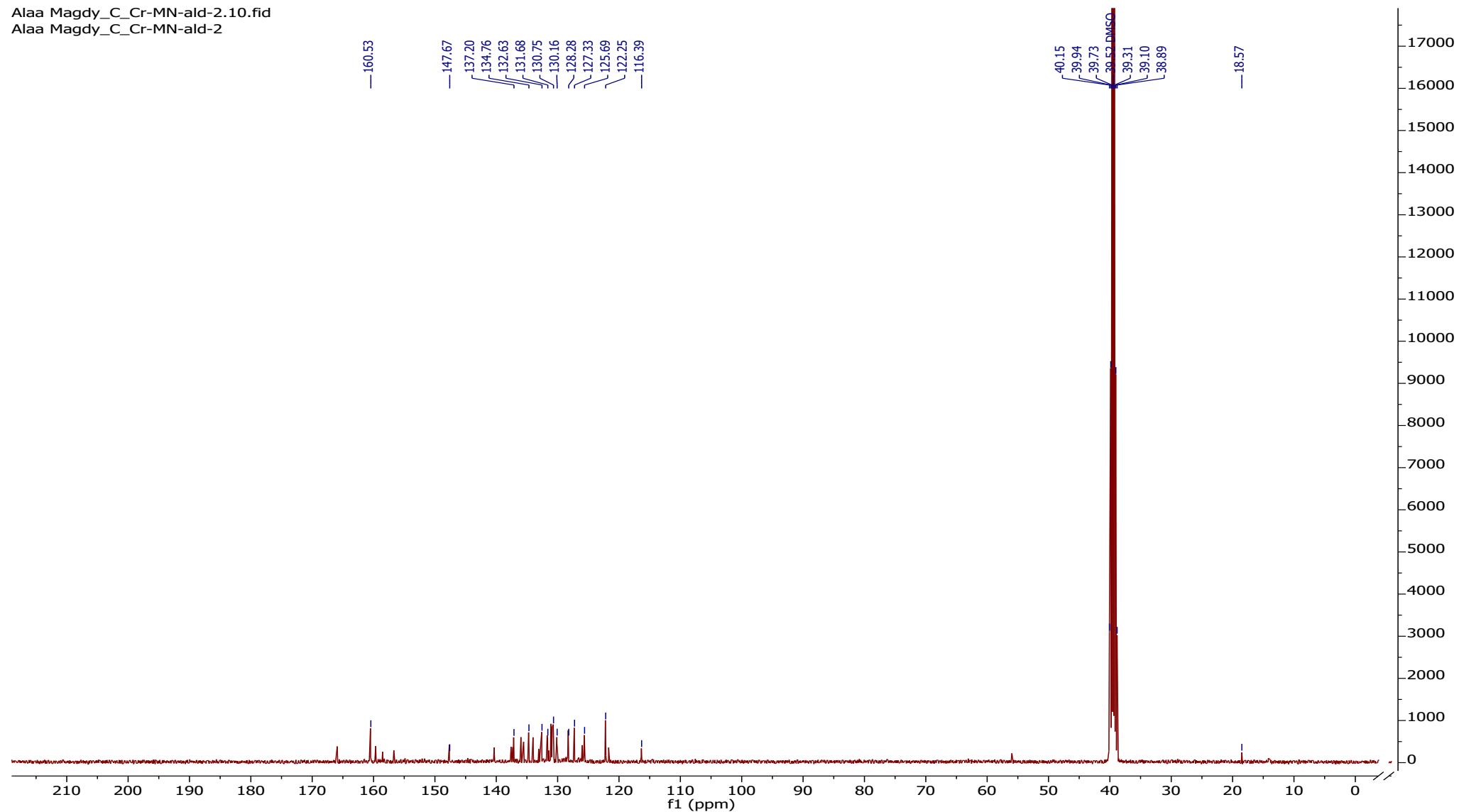


D₂O of 6d

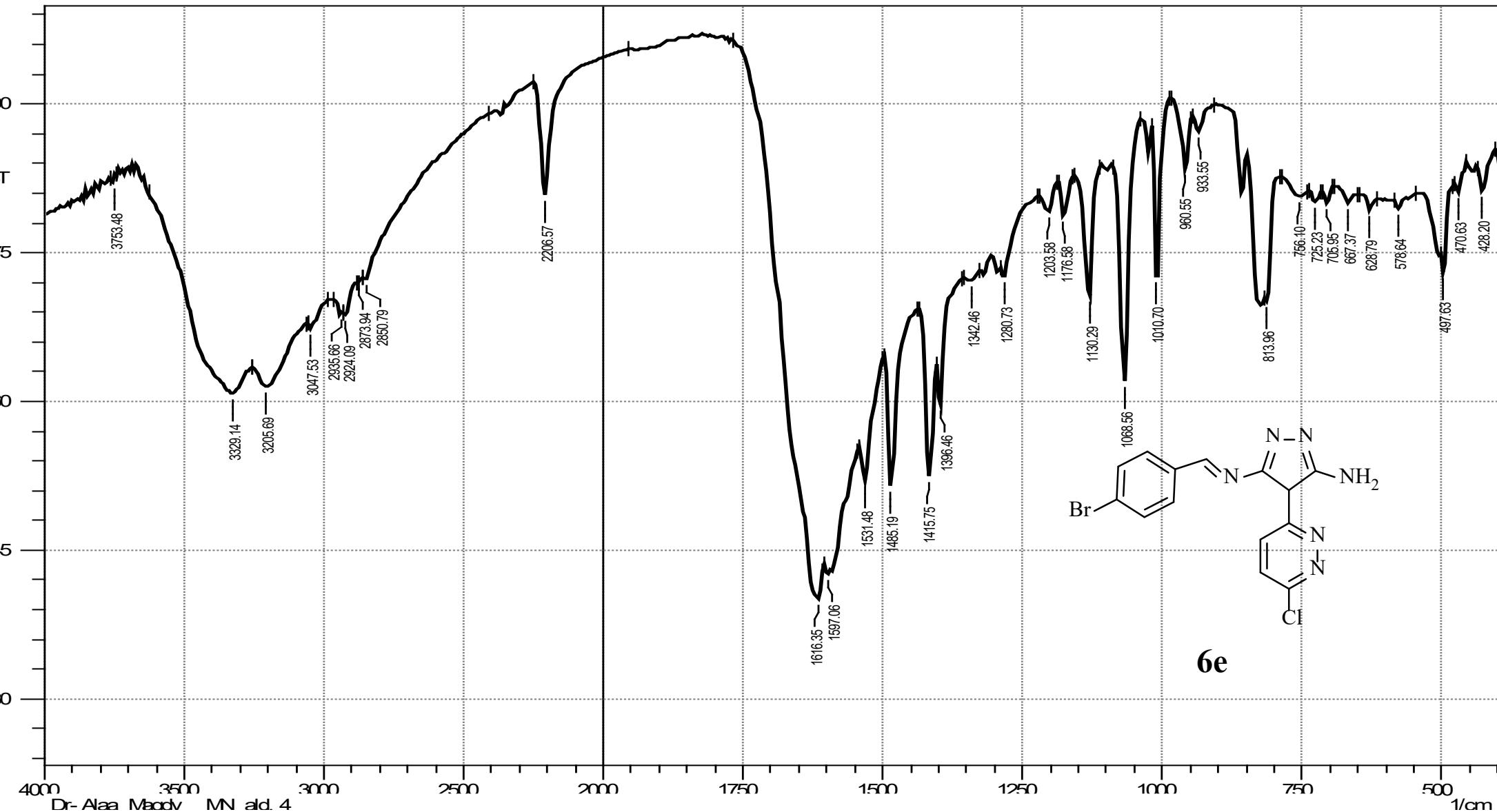


¹³C NMR of **6d** in DMSO

Alaa Magdy_C_Cr-MN-ald-2.10.fid
Alaa Magdy_C_Cr-MN-ald-2

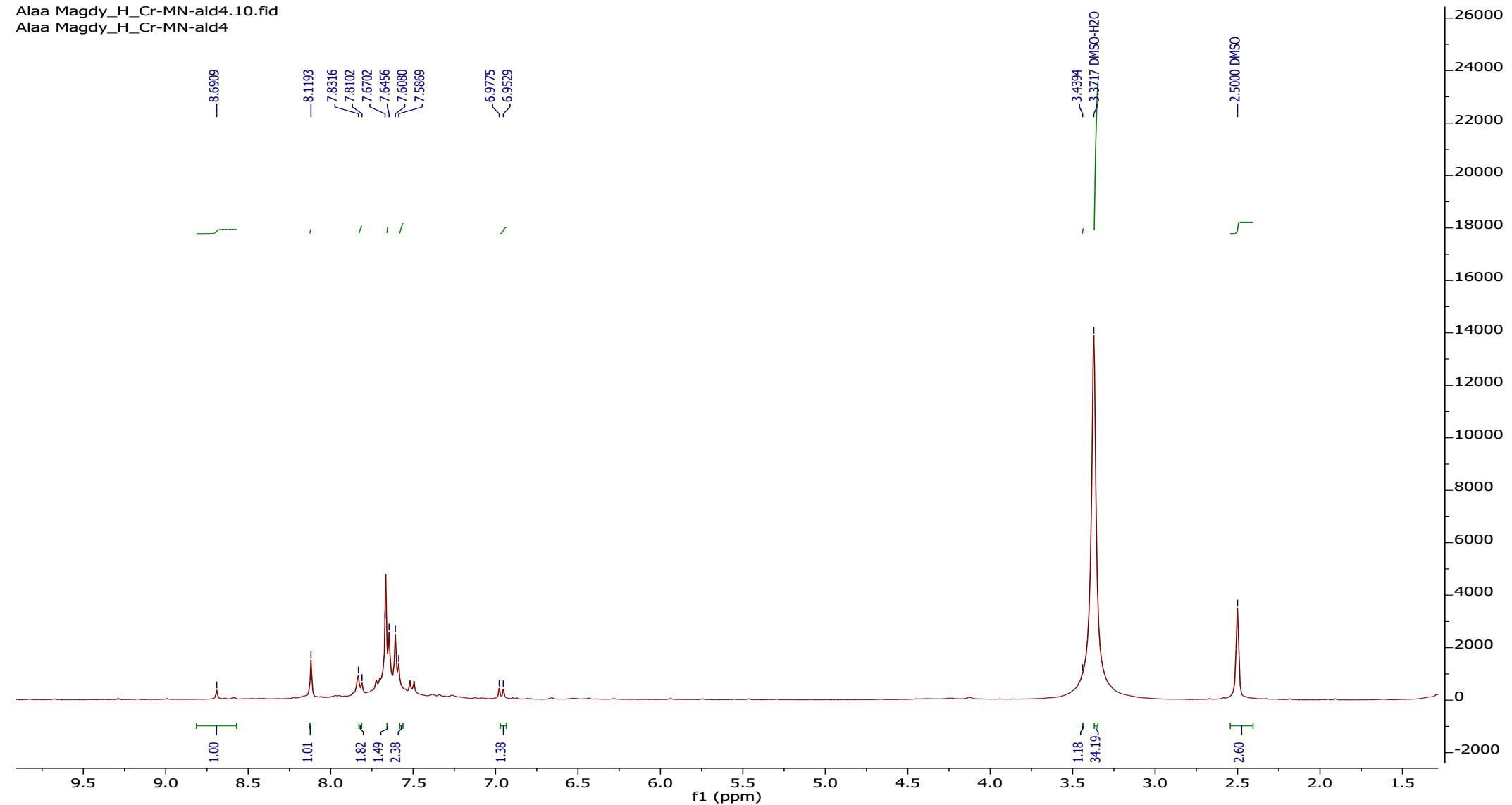


IR of compound 6e



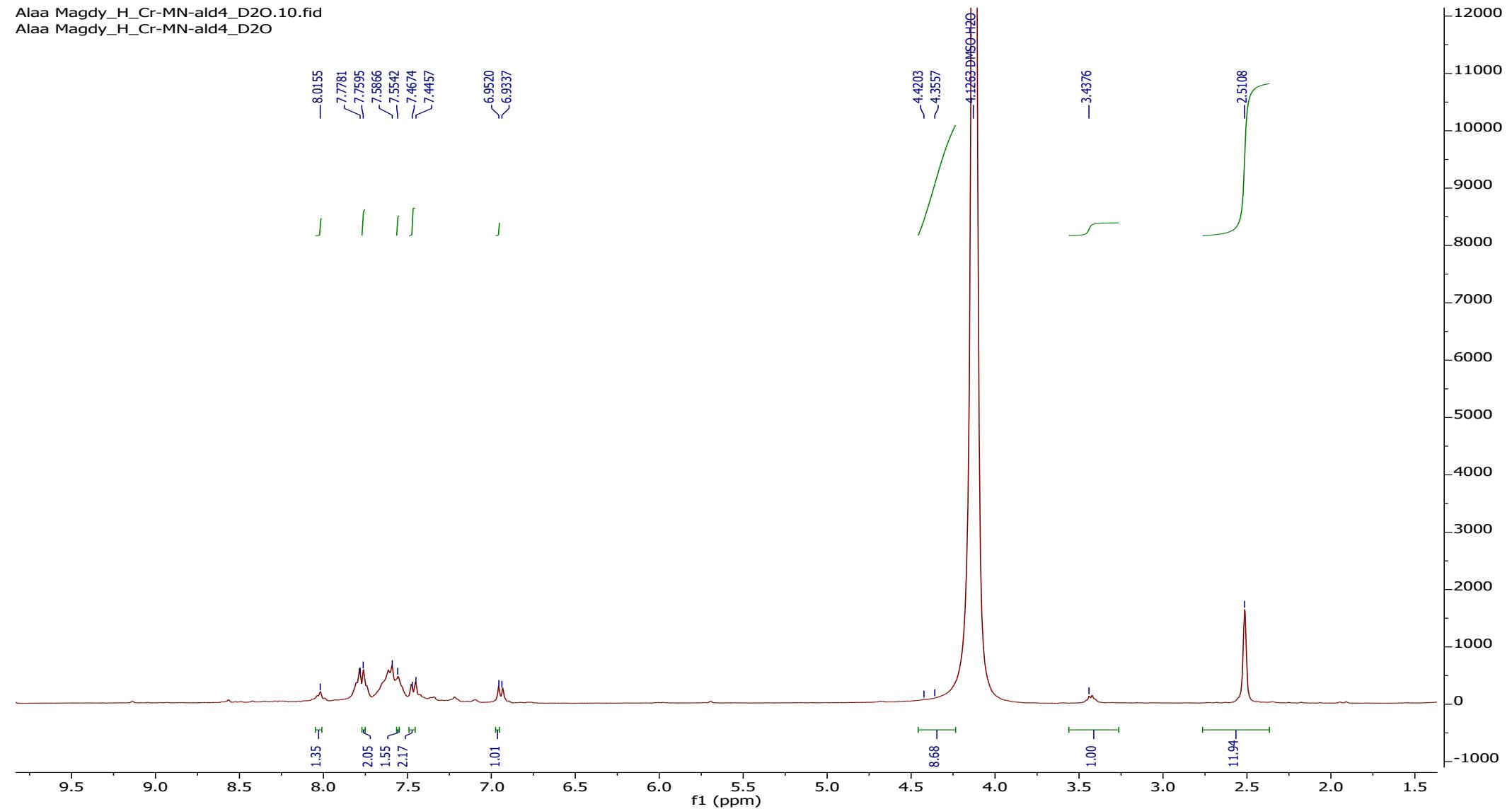
¹H NMR of 6e in DMSO

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Alaa Magdy_H_Cr-MN-ald4



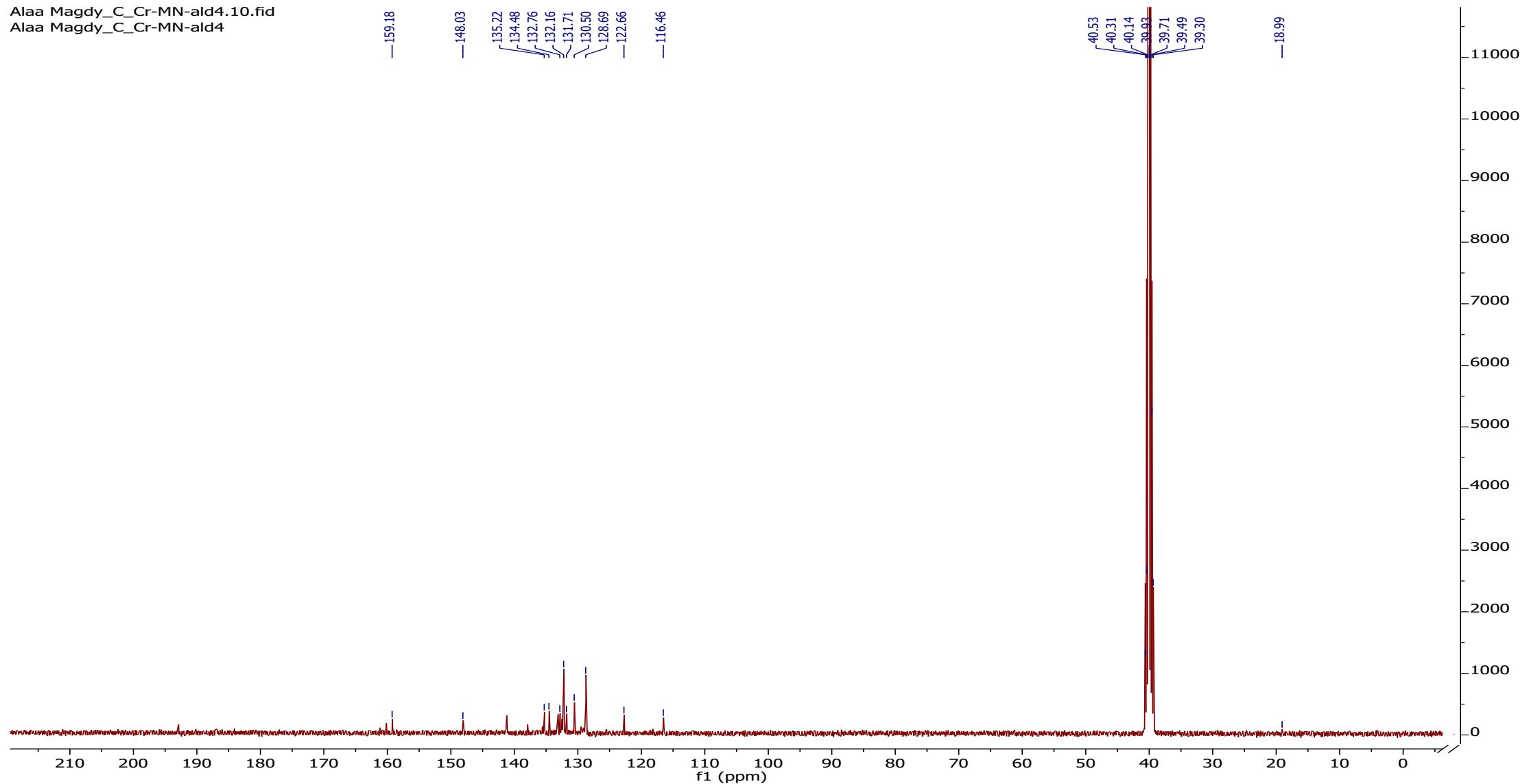
D₂O of 6e

Alaa Magdy_H_Cr-MN-ald4_D2O.10.fid
Alaa Magdy_H_Cr-MN-ald4_D2O

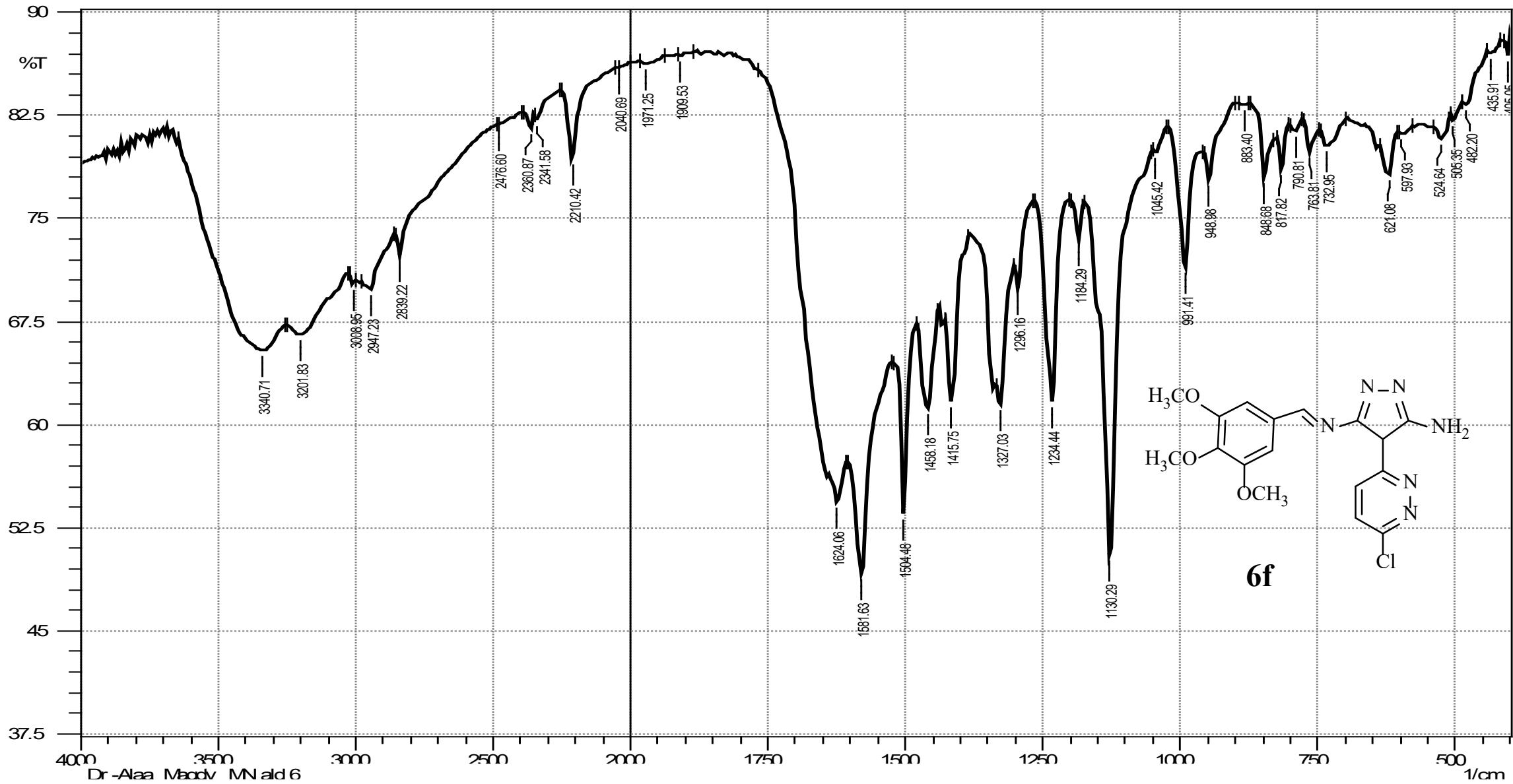


¹³C NMR of **6e** in DMSO

Alaa Magdy_C_Cr-MN-ald4.10.fid
Alaa Magdy_C_Cr-MN-ald4

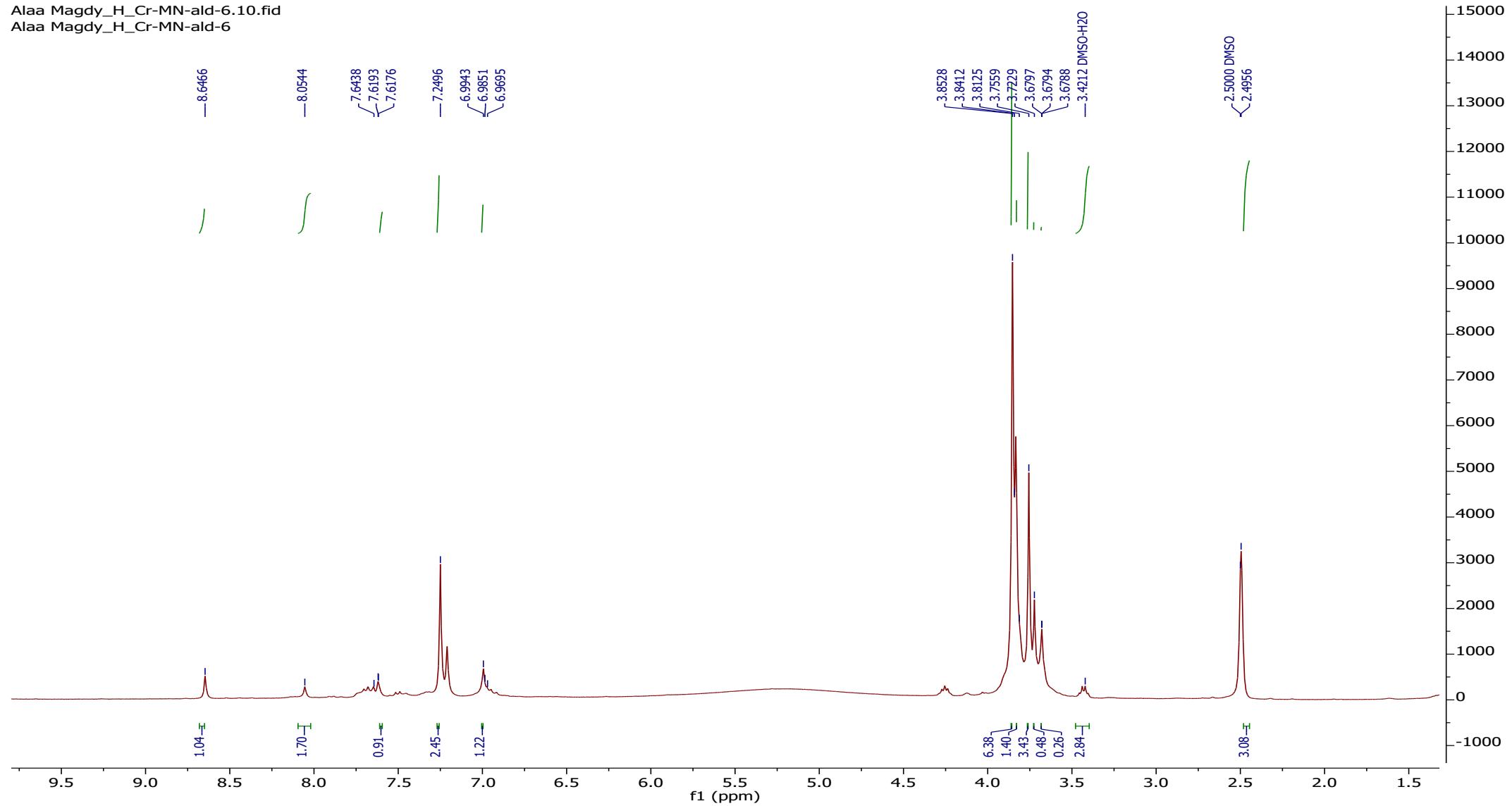


IR of compound 6f



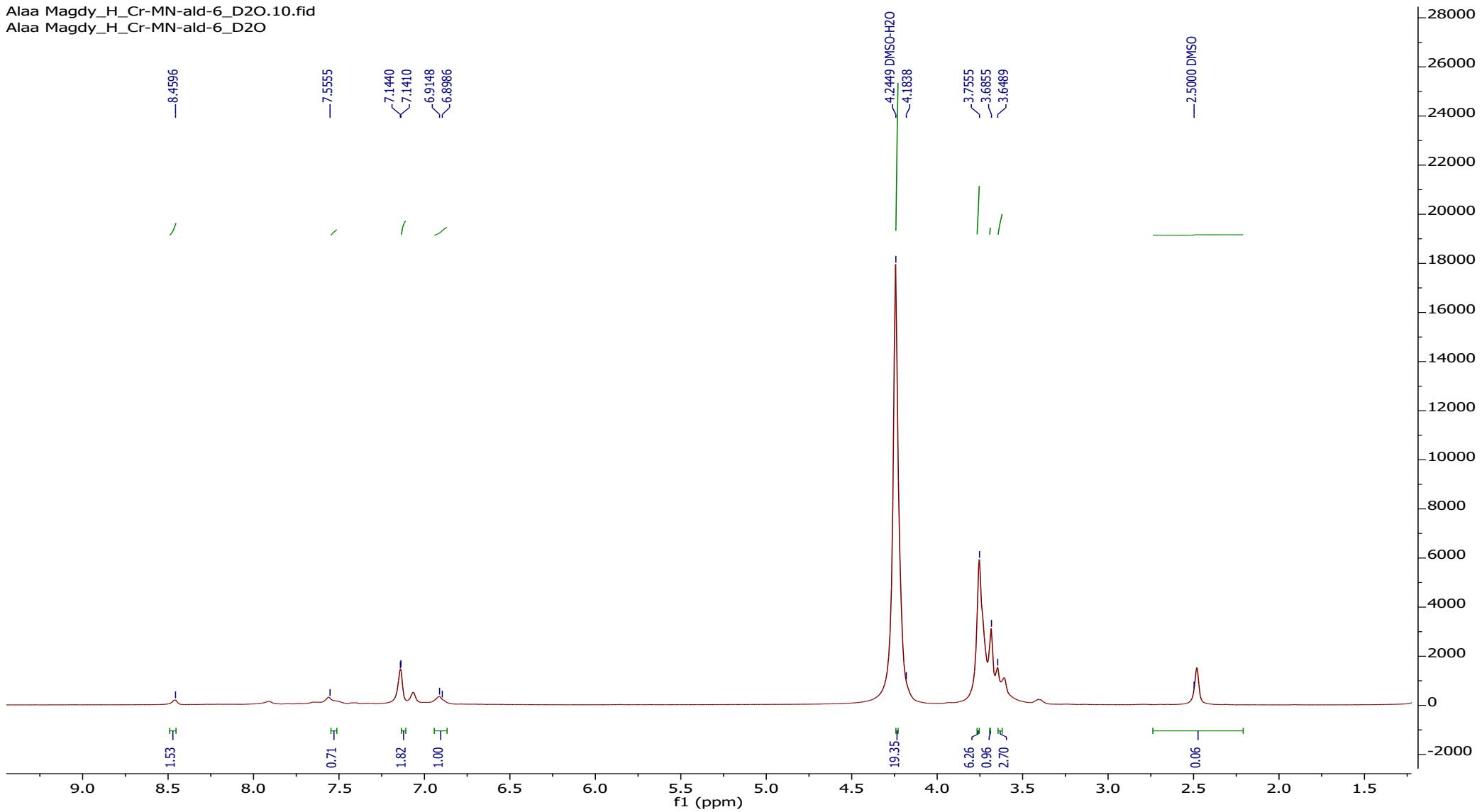
¹H NMR of 6f in DMSO

Alaa Magdy_H_Cr-MN-ald-6.10.fid
Alaa Magdy_H_Cr-MN-ald-6



D₂O of 6f

Alaa Magdy_H_Cr-MN-ald-6_D2O.10.fid
Alaa Magdy_H_Cr-MN-ald-6_D2O



¹³C NMR of **6f** in DMSO

