Supporting Information

Gold(I) and Gold(III) Carbene Complexes from the Marine Betaine Norzooanemonin: Inhibition of Thioredoxin Reductase, Antiproliferative and Antimicrobial Activity

CONTENTS

S1.1. General

Materials and Analytical Methods

All materials were purchased from Aldrich, Wako or TGI. (Me₂S)AuCl was prepared according to literature procedure.[1] Norzooanemonin, gold complexes (**3c**), and (**5c**) have been prepared according to our previous report.[2] All reactions were carried under air and technical grade solvent were used unless otherwise stated. K_2CO_3 and NaOH were used as received without further purification. ¹H and ¹³C NMR spectra were measured on the spectrometers Bruker AV 300 (300 MHz) spectrometers. The chemical shifts are given in parts per million (δ; ppm) relative to residual solvent peaks (δ; 3.31 (CD₃OD), 4.79 (D₂O), 7.26 (CDCl₃), 2.50 (DMSO-D₆). Coupling constants (*J*) are reported in Hertz (Hz), and splitting patterns are indicated as s (*singlet*), d (*doublet*), t (*triplet*), m (*multiplet*), sept (*septet*) and br (*broad*). All the spectra were measured at room temperature unless otherwise stated. Elemental analyses of all the complexes were carried out with a Vario Micro Cube System (Elementar Analysensysteme GmbH). High- and Low-resolution electrospray ionization (ESI) measurements were made with a Bruker MicroTOF II mass spectrometer.

S1.2. Experimental Procedures

S1.2.1 Synthesis of Compound 1b

Methyl trifluoromethanesulfonate (0.11 mL, 1 mmol, 1 eq.) was added to norzooanemonin **1a** (0.140 g, 1 mmol, 1 eq.) in dichloromethane and stirred for one hour at room temperature. The solvent was removed, and compound **1b** was washed with cold ethanol and hexane to be obtained as a white solid (0.120 g, 86%). ¹H NMR (CDCl₃, 300.1 MHz, 293 K): δ. 9.00 (1H, s, CH_{im}), 7.98 (1H, s, CH_{im}), 4.02 (3H, s, CH₃), 3.90 (3H, s, CH₃), 3.84 (3H, s, CH3). ¹³C{¹H} NMR (CDCl3, 75.5 MHz, 293 K): *δ.* 157.6 (*C*OO), 140.5 (N*C*HN), 129.0 (N*C*HC), 124.3 (N*C*COO), 120.3 (q, ¹ JC,F= 320 MHz, *C*F3), 52.7

(OCH₃), 36.5 (NCH₃), 36.3 (NCH₃). Elemental analysis calculated (%) for C₈H₁₁F₃N₂O₅S: C, 31.58, H, 3.64, N, 9.21; found: C, 31.02, H, 3.61, N, 9.10. ESI-MS *m/z* anion: calculated for [M]⁺ 155.08; Found [M]⁺ 155.14 m/z.

Figure S1. ¹H NMR spectrum of **1b** in CDCl₃ at room temperature.

Figure S2. ¹³C{¹H} NMR spectrum of **1b** in CDCl₃ at room temperature.

Figure S3. ¹³C dept NMR spectrum of 1b in CDCl₃ at room temperature.

S1.2.2. Synthesis of compound 1c

Triethyloxonium tetrafluoroborate (0.190 g, 1 mmol, 1. eq.) was added to norzooanemonin **1a** (0.140 g, 1 mmol, 1. eq.) in dichloromethane and stirred for four hours. Upon removing the solvent and washing with cold ethanol and diethyl ether, compound **1c** was obtained a white solid (0.130 g, 51%). ¹H NMR (CDCl3, 300.1 MHz, 293 K): *δ.* 8.84 (1H, s, CHim), 7.89 (1H, s, CH_{im}), 4.37 (2H, q, ³J_{HH}= 7.2 Hz, *CH*₂CH₃), 4.09 (3H, s, CH₃), 3.96 (3H, s, CH₃), 1.36 (3H, t, ³J_{HH}= 7.1 Hz, CH₂CH₃). ¹³C{¹H} NMR (CDCl₃, 75.5 MHz, 293 K): *δ.* 157.3 (*C*OO), 140.7 (N*C*HN), 128.8 (N*C*HC), 124.7

(NCCOO), 62.5 (CH₂CH₃), 36.7 (NCH₃), 36.5 (NCH₃), 13.9 (CH₃CH₂). Elemental analysis calculated (%) for C8H13F4N2O2: C, 37.53, H, 5.12, N, 10.94; found: C, 37.21, H, 5.07, N, 10.89. ESI-MS m*/z* anion: calculated for [M]⁺ 169.10; Found [M]⁺ 169.12 m/z.

Figure S4. ¹H NMR spectrum of 1c in CDCl₃ at room temperature.

Figure S5. ¹³C{¹H} NMR spectrum of 1c in CDCl₃ at room temperature.

S1.2.3. Synthesis of compound 2b

A mixture of compound **1b** (0.076 g, 0.25 mmol, 1 eq.) and K_2CO_3 (0.034 g, 0.25 mmol, 1 eq.) was stirred in methanol for three hours at room temperature. (Me₂S)AuCl (0.073 g, 0.25 mmol, 1 eq) was added to the mixture and stirred for 12 hours. The brownish mixture was filtered using a Whatman filter, and the solvent was removed under vacuum. Complex **2b** was obtained as a white powder after chloroform extraction and solvent removal under vacuum (0.048 g, 50%). ¹H NMR (CDCl₃, 300.1 MHz, 293 K): *δ.* 7.59 (1H,s, CHim), 4.09 (3H,s, CH3), 3.88 (3H,s, CH3), 3.87

(3H, s, CH3). ¹³C{¹H} NMR (CDCl3, 75.5 MHz, 293 K): *δ.* 176.7 (*C*Au), 158.7 (*C*OO), 128.3 (N*C*HC), 123.9 (NCCOO), 52.4 (OCH₃), 38.8 (NCH₃), 38.2 (NCH₃). Elemental analysis calculated (%) for C₇H₁₀AuClN₂O₂: C, 21.75, H, 2.61, N, 7.25; found: C, 21.88, H, 2.89, N, 7.18. ESI-MS m*/z* anion: calculated for [M + Na] + 409.01; Found [M + Na]⁺ 409.08 m/z and calculated for [M – CH₃]⁻ 370.99; Found [M – CH₃]⁻ 370.68 m/z.

Figure S7. ¹H NMR spectrum of 2b in CDCl₃ at room temperature.

Figure S8. ¹³C{¹H} NMR spectrum of **2b** in CDCl₃ at room temperature.

Figure S9. ¹³C dept NMR spectrum of 2b in CDCl₃ at room temperature.

S1.2.4. Synthesis of compound 2c

At 50 °C overnight, K_2CO_3 (0.034 g, 0.25 mmol, 1 eq.) was dissolved in ethanol. Compound 1c (0.064 g, 0.25 mmol, 1 eq.) and (Me₂S)AuCl (0.074 g, 0.25 mmol, 1 eq.) were added at room temperature and stirred overnight. The brownish mixture was filtered and the ethanol was removed under vacuum to yield a white powder. Extraction with chloroform and removal of the solvent yielded complex **2c** as a white powder (0.046 g, 46%).¹H NMR (CDCl3, 300.1 MHz, 293 K): *δ.* 7.59 (1H, s, CH_{im}), 4.35 (2H, q, ³J_{HH}= 7.2 Hz, *CH₂CH₃), 4.10 (3H, s, CH₃), 3.87 (3H,*

s, CH₃), 1.36 (3H, t, ³J_{HH}= 7.1 Hz, CH₂CH₃). ¹³C{¹H} NMR (CDCl₃, 75.5 MHz, 293 K): *δ.* 176,6 (CAu), 158.3 (*COO*), 128.1 (NCHC), 124.3 (NCCOO), 61.7 (CH₂CH₃), 38.7 (NCH₃), 38.3 (NCH₃), 14.2 (CH₃CH₂). Elemental analysis calculated (%) for $C_8H_{12}AuClN_2O_2$: C, 23.79, H, 3.02, N, 6.99; found: C, 23.89, H, 2.99, N, 6.89. ESI-MS m*/z* anion: calculated for [M + Na] ⁺ 423.02; Found [M + Na] ⁺423.00 m/z and calculated for [M – H] - 399.02; Found [M – H] - 398.92 m/z.

Figure S10. ¹H NMR spectrum of 2c in CDCl₃ at room temperature.

Figure S11. ¹³C{¹H} NMR spectrum of 2c in CDCl₃ at room temperature.

S1.2.5. Synthesis of compound 3b

Compounds **1b** (0.228 g, 0.70 mmol, 2 eq.), K_2CO_3 $(0.104 \text{ g}, 0.70 \text{ mmol}, 2 \text{ eq.})$, and $(Me_2S)AuCl$ (0.103 g, 0.35 mmol, 1 eq.) were stirred in acetone for 7 days, resulting in a brownish mixture which wasthen filtered using a Whatman filter. The solvent of the filtrate was removed under vacuum, following column chromato-

graphy (99% DCM: 1% MeOH), and complex **3b** was obtained as a white powder (0.015 g, 66%). ¹H NMR (CD2Cl2, 300.1 MHz, 293 K): *δ.* 7.77 (1H, s, CHim), 4.15 (3H, s, CH3), 3.95 (3H, s, CH3), 3.89 (3H, s, CH3). ¹³C{¹H} NMR (CD2Cl2, 75.5 MHz, 293 K): *δ.* 188.5 (*C*Au), 159.2 (*C*OO), 129.6 (N*C*HC), 124.9 (NCCOO), 52.8 (OCH₃), 38.9 (NCH₃), 38.4 (NCH₃). Elemental analysis calculated (%) for C₁₅H₂₀AuF₃N₄O₇S: C, 27.53, H, 3.08, N, 8.56; found: C, 27.72, H, 3.12, N, 8.48. ESI-MS m*/z* anion: calculated for [M]⁺ 505.11; Found [M]⁺ 505.20 m/z.

Figure S13. ¹H NMR spectrum of **3b** in CD₂Cl₂ at room temperature.

Figure S14. ¹³C{¹H} NMR spectrum of **3b** in CD₂Cl₂ at room temperature.

Figure S16. ¹⁹F NMR spectrum of 3b in CD₂Cl₂ at room temperature.

S1.2.6. Synthesis of compound 3c

Compound **1c** (1 eq, 0.128 g, 0.5 mmol, 1 eq.), K_2CO_3 (0.069 g, 0.5 mmol, 1 eq.), and (Me₂S)AuCl (0.057 g, 0.25 mmol, 0.5 eq.) were stirred in acetone for four days at room temperature to form a brownish mixture. The solution was filtered using a Whatman filter and the solvent of filtrate was removed under vacuum. After

purification by column chromatography (99% DCM: 1% MeOH), complex **3c** was obtained as a white powder (0.084 g, 54%). ¹H NMR (CDCl₃, 300.1 MHz, 293 K): *δ. 7.7*9 (1H, s, CH_{im}), 4.35 (2H, q, ³J_{HH}= 7.1 Hz, CH₂CH₃), 4.15 (3H, s, CH₃), 4.00 (3H, s, CH₃), 1.37 (3H, t, ³J_{HH}= 7.1 Hz, CH₂CH₃). ¹³C{¹H} NMR (CDCl₃, 75.5 MHz, 293 K): δ. 188.1 (CAu), 158.5 (COO), 129.5 (NCHC), 124.6 (NCCOO), 61.8 (CH₂CH₃), 38.7 (NCH₃), 38.0 (NCH₃), 14.1 (CH₃CH₂). Elemental analysis calculated (%) for C₁₆H₂₄AuBF₄N₄O₄: C, 30.99, H, 3.90, N, 9.03; found: C, 30.91, H, 3.89, N, 9.08. ESI-MS m*/z* anion: calculated for [M]⁺ 533.15; Found [M]⁺ 533.25 m/z.

Figure S17. ¹H NMR spectrum of **3c** in CDCl₃ at room temperature.

Figure S18. ¹³C{¹H} NMR spectrum of **3c** in CDCl₃ at room temperature.

Figure S19. ¹³C dept NMR spectrum of 3c in CDCl₃ at room temperature.

S1.2.7. Synthesis of compound 4a

A mixture of MeOH/DCM (1:1) was used to dissolve compound **2a** (0.039, 0.1 mmol, 1 eq.). Iodobenzene dichloride (0.038 g, 0.14 mmol, 1.4 eq.) was added, and the mixture was stirred for 24 hours at room temperature. The resulting light-yellow solution was vacuum evaporated and washed three times with hexane (20 mL) and diethyl ether (20 mL) to obtain complex **4a** as a white powder (0.038 g, 85%). ¹H NMR (CD₃OD, 300.1 MHz, 293 K): δ. 8.21 (1H, s, CH_{im}), 4.21 (3H, s, CH₃), 3.99 (3H, s, CH₃). ¹³C{¹H} NMR (CD₃OD, 75.5 MHz, 293 K): *δ*. 160.2 (*C*OO), 146,4 (*C*Au), 132.2 (N*C*HC), 128.5 (N*C*COO), 38.4 (N*C*H3), 37.7

(NCH₃). Elemental analysis calculated (%) for C₆H₈AuCl₃N₂O₂: C, 16.25, H, 1.82, N, 6.32; found: C, 16.17, H, 1.717, N, 5.78. ESI-MS *m/z* anion: calculated for [M + Na] + 464.92; Found [M + Na] ⁺ 464.86 m/z.

Figure S20. ¹H NMR spectrum of **4a** in CD₃OD at room temperature.

S1.2.8. Synthesis of compound 4b

Iodobenzene dichloride (PhICl₂, 0.14 mmol, 0.038 g, 1.4 eq.) was added to the methanol solution of compound **2b** (0.1 mmol, 0.039 g, 1 eq.). The mixture was stirred for 24 h at room temperature resulting in a light-yellow solution. The solvent was removed under vacuum. The residue was washed three times with hexane (20 mL), diethyl ether (20 mL), and cold chloroform (5 mL) to obtain complex **4b** as a white powder (0.040 g, 87%). ¹H NMR (CDCl3, 300.1 MHz, 293 K): *δ.* 7.76 (1H, s, CHim),

4.26 (3H, s, CH3), 4.03 (3H, s, CH3), 3.95 (3H, s, CH3). ¹³C{¹H} NMR (CDCl3, 75.5 MHz, 293 K): *δ.* 157.7 (*C*OO), 148.9 (*C*Au), 129.8 (N*C*HC), 126.4 (N*C*COO), 53.0 (O*C*H3), 38.6 (N*C*H3), 37.7 (N*C*H3). Elemental analysis calculated (%) for $C_7H_{10}AuCl_3N_2O_2$: C, 18.38, H, 2.20, N, 6.12; found: C, 18.27, H, 2.01, N, 5.93. ESI-MS m/z anion: calculated for [M + Na]⁺ 478.94; Found [M + Na]⁺478.84 m/z and calculated for [M $-$ CH₃]⁻ 370.99; Found [M $-$ CH₃]⁻ 370.68 m/z.

gure S23. ¹H NMR spectrum of 4**b** in CDCl₃ at room temperature.

Figure S24. ¹³C{¹H} NMR spectrum of **4b** in CDCl₃ at room temperature.

S1.2.9. Synthesis of compound 4c

Iodobenzene dichloride (PhICl₂, 0.038 g, 0.14 mmol, 1.4 eq.) was added to a methanol solution of compound **2c** (0.040 g, 0.1 mmol, 1 eq.). After stirring the mixture for 24 hours at room temperature, a light-yellow solution was obtained. The solvent was removed under vacuum. Three portions of hexane (20 mL) and cold diethyl ether (10 mL) were applied to the residue to obtain complex **4c** as a white powder (0.041 g, 89%).

¹H NMR (CDCl₃, 300.1 MHz, 293 K): *δ.* 7.78 (1H, s, CH_{imid}), 4.42 (2H, q, ³J_{HH}= 7.2 Hz, *CH₂CH₃), 4.25 (3H,* s, CH₃), 4.03 (3H, s, CH₃), 1.39 (3H, t, ³J_{HH}= 7.1 Hz, CH₂CH₃). ¹³C{¹H} NMR (CDCl₃, 75.5 MHz, 293 K): *δ.* 157.2 (*C*OO), 148,3 (*C*Au), 129.8 (N*C*HC), 126.6 (N*C*COO), 62.5 (*C*H2CH3), 38.6 (N*C*H3), 37.7 (N*C*H3), 14.2 (CH₂CH₃). Elemental analysis calculated (%) for C₈H₁₂AuCl₃N₂O₂: C, 20.38, H, 2.57, N, 5.94; found: C, 20.49, H, 2.34, N, 5.61. ESI-MS m*/z* anion: calculated for [M - H] - 468.95; Found [M - H] - 469.02 m/z.

Figure S26. ¹H NMR spectrum of 4c in CDCl₃ at room temperature.

Figure S27. ¹³C{¹H} NMR spectrum of 4c in CDCl₃ at room temperature.

Figure S28. ¹³C dept NMR spectrum of 4c in CDCl₃ at room temperature.

S1.2.10. Synthesis of compound 5a

Compound **3a** (0.051 g, 0.1 mmol, 1 eq.) was dissolved in a mixture of MeOH and DCM (1:1), then iodobenzene dichloride (0.027 g, 0.1 mmol, 1 eq.) was added and the mixture was stirred at room temperature for 24 hours. The solvent was removed in vacuum, and the residue was washed three times with hexane (20 mL) and diethyl ether

(20 mL) and cold chloroform (5 mL) to afford complex **5a** as a white powder (0.041 g, 70%). ¹H NMR (CD3OD, 300.1 MHz, 293 K): *δ.* 7.96 (1H, s, CHim), 4.27 (3H, s, CH3), 4.04 (3H, s, CH3). ¹³C{¹H} NMR (CD3OD, 75.5 MHz, 293 K): *δ.* 157.9 (*C*Au), 130.3 (N*C*HC), 37.9 (N*C*H3), 37.4 (N*C*H3). due to low solubility ¹³C resonances for (*C*OO) and (N*C*COO) were not observed. Elemental analysis calculated (%) for C12H16AuCl3F3N4O4: C, 24.70, H, 2.76, N, 9.60; found: C, 24.13, H, 2.48, N, 9.35. ESI-MS *m/z* anion: calculated for $[M]^+$ 547.02; Found $[M]^+$ 547.00 m/z.

Figure S29. ¹H NMR spectrum of **5a** in CD₃OD at room temperature.

Figure S30. ¹³C{¹H} NMR spectrum of 5a in CD₃OD at room temperature.

S1.2.11. Synthesis of compound 5b

The mixture of compound **3b** (0.131 g, 0.2 mmol, 1 eq.) and iodobenzene dichloride (PhICl $_2$, 0.055 g, 0.2 mmol, 1 eq.) in dichloromethane was stirred for 24 h at room temperature, resulting in a light-yellow solution. After removing the solvent, a yellowish solid was obtained. It was washed three times with hexane (20 mL)

followed by diethyl ether (20 mL) and once with cold methanol (5 mL) to obtain complex **5b** as a white powder. A flash chromatography for further purification (93% DCM: 7% MeOH) to afford **5b** (0.103 g, 71%). ¹H NMR (CDCl3, 300.1 MHz, 293 K): *δ.* 8.17 (1H, s, CHim), 4.24 (3H, s, CH3), 4.12 (3H, s, CH3), 3.92 (3H, s, CH3). ¹³C{¹H} NMR (CDCl3, 75.5 MHz, 293 K): *δ.* 158.4 (*C*OO), 158.3 (*C*Au), 131.6 (N*C*HC), 125.9 (NCCOO), 120.9 (q, ¹J_{C,F}= 320 MHz, CF₃), 52.9 (OCH₃), 38.6 (NCH₃), 37.4 (NCH₃). Elemental analysis calculated (%) for C₁₅H₂₀AuCl₂F₃N₄O₇S: C, 24.84, H, 2.78, N, 7.73; found: C, 24.37, H, 2.77, N, 7.42. ESI-MS m*/z* anion: calculated for [M]⁺ 575.05; Found [M]⁺ 575.04 m/z.

Figure S31. ¹H NMR spectrum of **5b** in CDCl₃ at room temperature.

Figure S32. ¹³C{¹H} NMR spectrum of 5b in CDCl₃ at room temperature.

Figure S33. ¹³C dept NMR spectrum of 5b in CDCl₃ at room temperature.

Figure S34. ¹⁹F NMR spectrum of **5b** in CDCl₃ at room temperature.

S1.2.12. Synthesis of compound 5c

The mixture of compound **3c** (0.62 g, 0.1 mmol, 1 eq.) and iodobenzene dichloride (PhICl $_2$, 0.027 g, 0.1 mmol, 1 eq.) in dichloromethane was stirred for 24 h at room temperature resulting in a lightyellow solution. The solvent was removed in vacuum to obtain a yellowish solid. The residue was washed three times with hexane (20 mL),

twice with diethyl ether (20 mL), and once with cold methanol (5 mL) to obtain complex **5c**. Further purificatin by flash chromatography using silica 60 (90% DCM: 10% MeOH) afforded **5c** as a white solid (0.049 g, 72%). ¹H NMR (CDCl₃, 300.1 MHz, 293 K): *δ*. 8.07 (1H, s, CH_{imid}), 4.38 (2H, q, ³J_{HH}= 7.1 Hz, *CH*₂CH₃), 4.23 (3H, s, CH₃), 4.11 (3H, s, CH₃), 1.38 (3H, t, ³J_{HH}= 7.1 Hz, CH₂*CH₃*). ¹³C{¹H} NMR (CDCl₃, 75.5 MHz, 293 K): *δ.* 158.2 (*C*OO), 157.9 (*C*Au), 131.2 (N*C*HC), 126.1 (N*C*COO), 62.3 (*C*H2CH3), 38.4 (N*C*H3), 37.2 (NCH₃), 14.1 (CH₃CH₂). ¹⁹F NMR (CDCl₃, 282.5 MHz, 293 K): *δ.* -152.7 (m,¹¹BF₄), -152.6 (q, ³J_{HH}= 1.3 MHz, ¹⁰BF₄). Elemental analysis calculated (%) for C₁₆H₂₄AuBCl₂F₄N₄O₄ (691.06): C, 27.81, H, 3.50, N, 8.11; found: C, 27.42, H, 3.75, N, 7.80. ESI-MS m*/z* anion: calculated for [M]⁺ 603.08; Found [M]⁺ 603.06 m/z.

Figure S35. ¹H NMR spectrum of 5c in CDCl₃ at room temperature.

Figure S36. ¹³C{¹H} NMR spectrum of **5c** in CDCl₃ at room temperature.

Figure S37. ¹³C dept NMR spectrum of 5c in CDCl₃ at room temperature.

Figure S38. ¹⁹F NMR spectrum of **5c** in CDCl₃ at room temperature.

S1.3. Single Crystal X-Ray Diffraction

Crystals were mounted on top of a human hair or on a Hampton Research CryoLoopTM with perfluorinated inert oil. Data were recorded on Rigaku XtaLAB Synergy S Single Source diffractometers equipped with a HyPix-6000HE detector and a PhotonJet microfocus source with Cu-Kα (**1c**, **3c**, **5c**) or Mo-Kα (**2b**, **1b**, **2c**, **4b**, **4c**, **4a**, **3b**, **5b**) radiation. Data reduction was performed with CrysalisPro[3] . Absorption correction was based on multi-scansfor **1c** and **2b**. For all otherstructures additionally face indexation and integration on a Gaussian grid was applied. The structure of compound **3b** was solved with the Patterson method with SHELXS-2013/1^[4] whereas all other structures were solved by intrinsic phasing with SHELXT-2018/2^[5] and refinements are based on F² using the program SHELXL-2018/3^[5] in OLEX^{2 [6]}. The hydrogen atoms of the OH groups in 4a have been refined freely. All other hydrogen atoms were placed in idealized positions and refined using a riding model.

1c was refined as a non-merohedral twin with the second component rotated by 179.8790° around [0.71 0.71 -0.01] (reciprocal) or [0.69 0.71 -0.14] (direct) and the relative volume of the smaller component refined to 42.8 %.

2b was refined as a non-merohedral twin with the minor component rotated by 179.8488° around [0.16 0.77 -0.61] (reciprocal) or [0.58 0.58 -0.58] (direct). The relative volume of the smaller component refined to 2.3 %.

For the structure of 2c the program ADDSYMM as part of the CHECKCIF^[7] routine suggests a different cell setup. A significant number of reflections could not be indexed with the cell suggested by ADDSYMM (97% indexation with the given cell vs. 73 % indexation with the suggested smaller cell volume). Additionally, the two molecules are not exactly overlapping. The torsion angles C6-O2-C7-C8 f e. are 156.22° and 171.91° for the two independent molecules.

4a was refined as a non-merohedral twin with component 2 rotated by 179.9923° around [0.39 0.84 0.37] (reciprocal) or [0.00 1.00 0.00] (direct). The relative volume of the smaller component refined to 48.9 %.

3b was refined as a pseudo merohedral twin (with twin matrix 1 0 0 / 0 -1 0 / -1 0 -1) and the relative volume of the smaller component refined to 33.7 %.

3c was refined as a non-merohedral 3-component twin with component 2 rotated by 6.0221° around [0.83 0.37 0.41] (reciprocal) or [1.00 0.05 0.02] (direct) and component 3 rotated by 1.8219° around [- 0.18 -0.97 0.15] (reciprocal) or [-0.10 -0.99 0.12] (direct). The relative volume of the smaller components refined to 28.9 % and 30.2 % respectively. Additionally, the BF₄ anion was found disordered and refined accordingly. This structure suffers from low bond length precision and exhibits high R values. Despite several attempts only weakly diffracting crystals of this compound were obtained. This structure only confirms the connectivity of the compound and any geometrical parameters should be interpreted with caution.

5b was refined as a non-merohedral twin with the second component rotated by -179.9751° around [0.62 -0.78 -0.07] (reciprocal) or [0.71 -0.71 -0.00] (direct). The relative volume of the smaller component refined to 27.9 %.

Two BF⁴ anions in **5c** were found disordered and refined accordingly. Chemically equivalent B-F and F-F distances were restrained to be equal.

Table S1. Crystal data and structure refinement of **1b**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.3238(12), N1−C2 1.3861(11), N1−C4 1.4657(12), N2−C1 1.3369(12), N2−C3 1.3712(11), O1−C6 1.3287(10), O1−C7 1.4460(12), O2−C6 1.2107(11), S1−O4 1.4470(7), S1−C8 1.8257(9), F1−C8 1.3424(12), N1−C1−N2 109.21(7), C3−C2−N1 107.11(7), C1−N1−C2 108.15(7), C1−N1−C4 123.40(8), C1−N2−C3 108.65(7), O2−C6−O1 125.34(8), C6−O1−C7 115.35(8).

Table S2. Crystal data and structure refinement of **1c**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.316(7), N1−C2 1.382(7), N1−C4 1.472(7), N2−C1 1.336(7), N2−C3 1.372(7), N2−C5 1.463(7), C2−C3 1.362(7), C2−C6 1.470(7), O1−C6 1.212(6), O2−C7 1.464(6), O2−C6 1.331(7), F1−B1 1.404(7), N1−C1−N2 109.7(5), C3−C2−N1 107.0(4), C1−N1−C2 108.2(4), C1−N1−C4 123.7(4), C1−N2−C3 108.1(4), O1−C6−O2 125.4(5), C6−O2−C7 116.2(4), O2−C6−C2 110.4(4).

Table S3. Crystal data and structure refinement of **2b**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.354(6), N1−C2 1.390(6), Au1−Cl1 2.2844(6), Au1−C1 1.987(2), N1−C4 1.466(6), N2−C1 1.362(5), N2−C3 1.366(5), N2−C5 1.467(5), C6−C2 1.469(6), C2−C3 1.356(6), O1−C6 1.341(6), O1−C7 1.451(6), O2−C6 1.203(6), N1−C1−N2 105.2(4), C3−C2−N1 107.2(4), C1−N1−C2 109.8(4), C1−N1−C4 123.8(4), C1−N2−C3 111.1(4), O1−C6−O2 124.6(4), C6−O1−C7 114.9(4), O1−C6−C2 110.5(4), N1−C1−Au1 129.3(3), C1−Au1−Cl1 178.05(13).

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.347(3), N1−C2 1.366(3), Au1−Cl1 2.2873(5), Au1−C1 1.984(2), N2−C3 1.392(3), N2−C5 1.459(3), C2−C3 1.365(3), C3−C6 1.464(3), O1−C6 1.205(3), O2−C7 1.453(3), O2−C6 1.341(3), N1−C1−N2 105.84(17), C3−C2−N1 106.89(18), C1−N1−C2 111.04(17), C1−N1−C4 124.31(17), C1−N2−C3 109.81(17), O1−C6−O2 124.5(2), C6−O2−C7 115.91(17), O2−C6−C3 109.73(18), N1−C1−Au1 126.52(15), C1−Au1−Cl1 178.42(6).

Table S5. Crystal data and structure refinement of **3b**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.357(11), N1−C4 1.471(11), Au1−C1 2.014(9), N2−C1 1.363(11), N2−C5 1.475(12), O1−C6 1.204(12), O2−C7 1.452(13), O2−C6 1.338(12), N1−C1−N2 104.4(7), C3−C2−N1 106.9(8), C1−N1−C2 110.2(7), C1−N1−C4 122.9(7), O1−C6−O2 125.0(9), C6−O2−C7 116.5(9), N1−C1−Au1 127.1(6), C1−Au1−C8 178.7(4).

Table S6. Crystal data and structure refinement of **3c**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.34(3), N1−C4 1.43(3), Au1−C1 2.05(2), N2−C1 1.33(3), N2−C5 1.48(4), O1−C6 1.20(3), O2−C6 1.34(3), O2−C7 1.47(3), N2−C1−N1 108(2), C3−C2−N1 107(2), C1−N1−C2 108(2), C1−N1−C4 126(2), O1−C6−O2 125(3), C6−O2−C7 114(2), N1−C1−Au1 124.1(18), C9−Au1−C1 177.5(9).

Table S7. Crystal data and structure refinement of **4a**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.337(3), N1−C2 1.388(3), Au1−Cl1 2.3192(6), Au1−C1 1.997(2), N2−C3 1.367(3), N2−C5 1.468(3), C2−C3 1.365(3), C2−C6 1.462(3), O1−C6 1.223(3), O2−C6 1.324(3), N1−C1−N2 107.70(17), C3−C2−N1 107.26(17), C1−N1−C2 108.65(17), C1−N1−C4 124.76(17), C1−N2−C3 109.70(18), O1−C6−O2 124.25(19), O2−C6−C2 111.99(18), N1−C1−Au1 126.48(15), C1−Au1−Cl1 176.95(6), Cl2−Au1−Cl1 93.17(2), Cl3−Au1−Cl2 174.44(2).

Table S8. Crystal data and structure refinement of **4b**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.3478(18), N1−C2 1.3692(19), Au1−Cl1 2.3129(4), Au1−C1 2.0022(14), N2−C3 1.3903(18), N2−C5 1.465(2), C2−C3 1.363(2), C3−C6 1.472(2), O1−C6 1.331(2), O1−C7 1.451(2), O2−C6 1.207(2), N2−C1−N1 107.68(12), C3−C2−N1 107.06(12), C1−N1−C2 109.51(12), C1−N1−C4 125.78(13), C1−N2−C3 108.84(12), O2−C6−O1 125.16(15), C6−O1−C7 115.40(15), O2−C6−C3 124.87(15), N1−C1−Au1 125.84(10), C1−Au1−Cl1 178.89(4), C1−Au1−Cl2 89.70(4), Cl3−Au1−Cl2 177.747(15).

Table S9. Crystal data and structure refinement of **4c**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.3512(16), N1−C2 1.3718(17), Au1−Cl1 2.3165(3), Au1−C1 1.9997(12), N2−C3 1.3937(16), N2−C5 1.4652(17), C2−C3 1.3614(18), C3−C6 1.4707(18), O1−C6 1.3341(17), O1−C7 1.4546(19), O2−C6 1.2084(18), N2−C1−N1 107.49(11), C3−C2−N1 107.05(11), C1−N1−C2 109.62(11), C1−N1−C4 126.04(11), C1−N2−C3 108.81(10), O2−C6−O1 125.72(13), C6−O1−C7 115.94(12), O2−C6−C3 124.65(13), N1−C1−Au1 127.20(9), C1−Au1−Cl1 176.53(4), Cl2−Au1−Cl1 91.157(13), Cl3−Au1−Cl2 177.234(14).

Table S10. Crystal data and structure refinement of **5b**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.343(12), N1−C4 1.464(12), Au1−C1 2.036(9), Au1−Cl1 2.275(2), N2−C1 1.347(11), N2−C5 1.470(11), O1−C6 1.196(11), O2−C6 1.336(11), O2−C7 1.455(11), N1−C1−N2 107.4(8), C3−C2−N1 107.0(8), C1−N1−C2 108.7(8), C1−N1−C4 123.6(8), O1−C6−O2 125.3(9), C6−O2−C7 113.9(8), N1−C1−Au1 125.1(7), Cl1#1−Au1−Cl1 180.0, C1−Au1−C1#1 180.0(5) (Symmetry transformations used to generate equivalent atoms: #1 -x+2,-y+2,-z+1).

Table S11. Crystal data and structure refinement of **5c**.

Thermal ellipsoids are presented at the 50 % level of probability. Bond distances and bond angles are reported in Å or degree (°), respectively. N1−C1 1.343(4), N1−C4 1.468(4), Au1−C1 2.033(3), N2−C1 1.352(4), N2−C5 1.475(4), O1−C6 1.211(4), O2−C6 1.336(4), O2−C7 1.457(4), N1−C1–N2 106.8(3), C3−C2−N1 107.1(3), C1−N1−C2 109.0(3), C1−N1−C4 124.3(2), O1−C6−O2 125.0(3), C6−O2−C7 115.7(3), N1−C1−Au1 126.7(2), Cl1^{#1}−Au1−Cl1 180.0, C1^{#1}−Au1−C1 180.0 (Symmetry transformations used to generate equivalent atoms: #1 -x+1,-y+2,-z+1).

S1.4. Stability Tests of Gold Complexes

The stability of the gold complexes **2a-c** to **5a-c** was investigated by ¹H NMR monitoring with respect to a conceivable Au-C bonds scission and the concomitant formation of protonated carbenes **1a-c**. A mixture of DMSO- d_6 and D₂O (9:1, w/w) was employed as an NMR solvent to simulate the conditions of the biological assays applied. ¹H NMR spectra of the pure ligand precursors **1a-c** displayed signals, which were significantly different from those of complexes **2a-c** to **5a-c**. All ¹H NMR samples did not show any diagnostic changes after 24 h. Additionally, ¹H NMR samples of equimolar mixture of ligands **1a-c** and complexes **2a-c** to **5a-c** gave rise to spectra, which resulted from the overlap of two independent species, i.e. ligand and respective complex.

Example 1

Figure S39. ¹H NMR spectra of **1b**, **2b** and an equimolar mixture of **1b** + **2b** at room temperature. All spectra were retained after 24 h without significant changes.

Example 2

Figure S40. ¹H NMR spectra of **1b**, **3b** and an equimolar mixture of **1b** + **3b** at room temperature. All spectra were retained after 24 h without significant changes.

Example 3

Figure S41. ¹H NMR spectra of **1b**, **4b** and an equimolar mixture of **1b** + **4b** at room temperature. All spectra were retained after 24 h without significant changes.

S2.1. Antibacterial Studies

Bacterial cultures were prepared by overnight aerobic incubation at 37 °C. Gram-negative strains were cultivated in Müller Hinton broth supplemented with 1% glucose at pH 7.2, while Gram-positive strains were grown in Trypticase soy yeast extract medium (TSY) with a composition of 30 g/l trypticase soy broth, 3 g/l yeast extract, and pH 7.2. The cultures were adjusted to an OD600 of 0.001, resulting in a final starting OD600 of 0.0005 for the subsequent tests. For testing, 25 μL of the prepared culture was combined with 25 μL of serially diluted test compounds in 384 well plates, following established protocols for different strains. Screening was performed at compound concentrations of 0.5, 5, and 50 µM, and for selected compounds, growth inhibition curves were recorded using DMSO stock solutions at final concentrations of 100, 50, 25, 12.5, 6,25, 3.125, 1.56, 0.78, 0.39, and 0.2 µM.

Positive control compounds, namely linezolid (for MRSA strains), ciprofloxacin (for *E. faecium* and *E. coli*), and amikacin (for *P. aeruginosa*), were included. The maximum DMSO concentration in the assay was 1%, which did not impact bacterial growth. After an 18-hour incubation at 37 °C under moist conditions, the optical density at 600 nm was measured using a Fusion Universal Microplate Analyser (Perkin–Elmer, Waltham, USA). The minimum inhibitory concentration (MIC) values, representing the lowest compound concentration that completely suppressed growth, were determined through curve fitting with Sigma Plot. The bacterial strains used included Gram-negative strains: *Escherichia coli* (DSM 1116), *A. baumannii* (DSM30007), *K. pneumoniae* (DSM111678), and *Pseudomonas aeruginosa* PA7 (DSM 24068); and Gram-positive strains: *E. faecium* (DSM20477) and *Staphylococcus aureus* MRSA (DSM 11822).

Table S12. Antibacterial activities of mono and dicarbene gold (I) and gold (III) complexes. Minimal inhibitory concentrations (MIC) are given in μM ± standard error of the mean. As positive control antibiotics, amikacin (P.a.), linezolid (MRSA) and ciprofloxacin (all other strains) have been used.

S2.2. Cytotoxicity and Anticancer Studies

Cell culture and cytotoxicity

Cell lines were sourced from DSMZ (German Collection of Microorganisms and Cell Cultures GmbH) and maintained through standard procedures in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal calf serum and 50 mg/L gentamycin. Antiproliferative effects were assessed using a well-established assay protocol. In brief, 100 μL volumes of A549 cells (13,243 cells per mL), HT-29 cells(14,657 cells per mL), MDA-MB-231 cells(6,613 cells per mL), or MCF-7 cells(6,917 cells per mL) were transferred into the wells of 96-well plates and incubated at 37° C/5% CO₂ for 72 hours (MCF-7, MDA-MB-231) or 48 hours (A549, HT-29).

Freshly prepared stock solutions of the compounds in DMSO were diluted with the respective cell culture medium to achieve graded concentrations (final concentration of DMSO: 0.1% v/v). After 72 hours (A549, HT-29) or 96 hours (MCF-7, MDA-MB-231) of exposure, the cell biomass was assessed through crystal violet staining, and the IC_{50} value was determined as the concentration causing 50% inhibition of cell proliferation compared to an untreated control. Results were calculated as the mean values of three independent experiments.

S2.3. Inhibition of bacterial TrxR from *E. coli*

The TrxR (*E. coli*) inhibition assay was performed according to previously published procedures.[8] The assay is partly based on the procedure developed by Lu et al^[9] and makes use of the reduction of DTNB (5,5′-dithiobis-(2-nitrobenzoic acid)). Solutions of *E. coli* TrxR (10.1 U/mL) and *E. coli* thioredoxin (Trx) (312 μg/mL) (both purchased from Sigma-Aldrich) were prepared in distilled water. Stock solutions of the test compounds (2 mM) were prepared in DMSO and serially diluted with TE buffer (Tris-HCl 50 mM, EDTA 1 mM, pH 7.5). 20 μL of these solutions or TE buffer without the test compounds (positive control) were mixed with the TrxR solution (10 μ L), the Trx solution (10 μ L) and 100 μ l of NADPH (200 μM) in TE buffer in a well on a 96-well plate. As a blank control, 200 μM NADPH in TE buffer (100 μL) mixed with a DMSO / buffer mixture (40 μL) was used (final concentrations of DMSO: 0.5% v/v). The plate was incubated for 75 min at 25°C with moderate shaking. After incubation, 100 μL of a reaction mixture (TE buffer containing 200 μM NADPH and 5 mM DTNB) was added to each well to initiate the reaction. After thorough mixing, the formation of 5-TNB was monitored by a microplate reader at 405 nm in 35 sintervals(10 measurements). The values were corrected by subtraction of the blank solution absorption values. The increase in concentration of 5-TNB followed a linear trend (r²≥ 0.990) and the enzymatic activities were calculated as the gradients (increase in absorbance per second) thereof. Absence of interference with the assay components was confirmed by a negative control experiment for each test compound, where the highest test compound concentration was used and the enzyme solution was replaced by TE buffer. The inhibition is presented as the mean IC_{50} values and standard deviations obtained in three independent experiments.

Table S13. Activity of TrxR from *E. coli* in presence of compounds at 0.5 µM (n = 2, n.d.= not determined)

Compound	2a	2 _b	2c	3a	3 _b	3 _c	4a	4b	4c	5a	5b	5c
TrxR activity, % of control	94.6	8.9	13.8	n.d.	106	115					86.1 36.5 28.5 91.6 90.8 93.5	

S2.4. Cellular uptake and metal quantification studies in A549 cells

Cellular uptake studies in A549 cells

For cellular accumulation studies, A549 lung carcinoma cells were seeded in 75 cm² cell culture flasks, incubated with 10 ml DMEM/10% FCS cell culture medium at 37 °C in a 5% CO₂ humidified atmosphere until they reached a confluence of 75-80%. The cells were treated with DMEM containing selected test compounds **3a** and **3c** dissolved in DMF (final DMF concentration: 0.1%; final test compound concentration: 2 μM). After 6 and 24 h of exposure, cells were isolated by removing the media followed by trypsinization. The trypsinized cells were washed with 9 ml PBS buffer (pH 7.4), scraped, and isolated by centrifugation (5 min, 3500 rpm). The obtained cell pellets were stored at -20 °C for further use. All experiments were carried out in triplicate.

Metal quantification in cell lysates by HRCS-AAS

Dry cell pellets were diluted with 400 μl of Milli-Q® water and lysed for 30 min in an ultrasonic bath (240 W, 65 Hz). For protein quantification. 20 μl of each sample was added to 200 μl of Bradford reagent (25 mg Serva Blue G (Sigma Aldrich) in 25 ml Ethanol 96%, 25 ml Milli-Q[®] water and 50 ml H₃PO₄ 86%, stored at -20 °C and freshly diluted 1+4 with Milli-Q[®] water prior to use) and incubated on a 96-well plate while shaking at room temperature for 30 min and measured by a Perkin Elmer Victor X4 plate reader at a wavelength of 595 nm. The measurements were performed in duplicate, and the mean was used to calculate the protein content. For a standard calibration curve, bovine serum albumin (Sigma Aldrich) was used in graded concentrations.

The gold quantification was performed using a high-resolution continuum source atomic absorption spectrometer ContrAA® 700 (Analytik Jena AG). A pure sample of the respective complex **5c** was used as standard, and calibration was done in a matrix-matched manner (meaning all samples and standards were adjusted to a protein concentration of 1 mg/ml by dilution with Milli-Q® water). The calibration and samples were measured in duplicate. The final cellular gold concentrations were calculated from data obtained in three independent experiments and are expressed as pmol of metal per mg of cell protein. Sample preparation: Triton-X 100 (1%, 20 μl) and ascorbic acid (1%, 20 μl) were added to each calibration standard or probe (200 μ). A volume of 20 μ l was injected into a coated standard graphite tube (Analytik Jena AG) and thermally processed as previously described.^[10] The gold amount was analyzed at a wavelength of 242.79 nm.

S3.1. References

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