Supplementary Information (SI) for RSC Medicinal Chemistry. This journal is © The Royal Society of Chemistry 2024

#### **Supporting Information**

#### Hydrophobic CPP/HDO Conjugates: A New Frontier in Oligonucleotide-

## Warheaded PROTAC Delivery

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Figure S1. Molecules synthesized in this study.

	Compounds	Sequence	
1	LCL-ER(dec)	Forward (LCL-ER(dec)-F): 5'-LCL-GTCAGGTCACAGTGACCTGA T-3' Reverse (ER(dec)-R): 3'-CAGTCCAGTGTCACTGGACTA-5'	
2	LCL-HDO	Forward (LCL-HDO-F): 5'-LCL-GTCAGGTCACAGTGACCTGATC AAAGTTAA-3' Reverse (HDO-R): 3'-CAGTCCAGTGTCACTGGACTAguuucaauu-	
3	LCL-HDOdna	5' Forward (LCL-HDO-F): 5'-LCL-GTCAGGTCACAGTGACCTGATC AAAGTTAA-3' Reverse (HDOdna-R): 3'-CAGTCCAGTGTCACTGGACTAGTTTCA ATT-5'	
4	LCL-HDOcl	Forward (LCL-HDO-F): 5'-LCL-GTCAGGTCACAGTGACCTGATC AAAGTTAA-3' Reverse (ER(dec)-R): 3'-CAGTCCAGTGTCACTGGACTA-5'	
5	CPP/HDO-PROTAC	Forward (LCL-HDO-F): 5'-LCL-GTCAGGTCACAGTGACCTGATC AAAGTTAA-3' Reverse (P4-HDO-R): 3'-CAGTCCAGTGTCACTGGACTAguuucaa uu-5'-P4	
6	FP-probe	Forward (5'-FAM-ER(dec)-F) 5'-FAM-GTCAGGTCACAGTGACCT GAT-3' Reverse (5'-FAM-ER(dec)-R): 3'-CAGTCCAGTGTCACTGGACTA- FAM-5'	
7	FAM-ER(dec)	Forward (5'-FAM-ER(dec)-F) 5'-FAM-GTCAGGTCACAGTGACCT GAT-3' Reverse (ER(dec)-R): 3'-CAGTCCAGTGTCACTGGACTA-5'	
8	FAM-HDOdna	Forward (5'-FAM-HDO-F) 5'-FAM-GTCAGGTCACAGTGACCTGA TCAAAGTTAA-3' Reverse (HDOdna-R): 3'- CAGTCCAGTGTCACTGGACTAGTTTC	
9	FAM-HDO-P4	Forward (5'-FAM-HDO-F) 5'-FAM-GTCAGGTCACAGTGACCTGA TCAAAGTTAA-3' Reverse (P4-HDO-R): 3'-CAGTCCAGTGTCACTGGACTAguuucaa uu-5'-P4	
10	LCL-ER(dec)-P4	Forward (LCL-ER(dec)-F): 5'-LCL-GTCAGGTCACAGTGACCTGA T-3' Reverse (P4-ER(dec)-R): 3'-CAGTCCAGTGTCACTGGACTA-5'-P4	

Table S1. Sequence of decoys and peptide used in this study.

LCL: LCL161, lowercase : RNA, FAM: fluorescein, P4:  $N_3$ -C6-LGAQSNF-NH<sub>2</sub>

# HPLC analysis of decoys

HPLC conditions

Column : CAPCELL PAK MG-II (C18, 4.6 x 250 mm, 5 µm; OSAKA soda).

Mobile phase : A = 0.1 M triethylammonium acetate (TEAA) buffer (pH7.0),  $B = CH_3CN$ .

Gradient : B% = 10-40 over 20 min, 40-100 over 5 min.

Flow rate : 1.0 mL/min.

Column temperature : 35°C.

Detection : 260 nm.





#### P4-HDO-R purity 97%

FAM-HDO-F purity 90%







FAM-ER(dec)-F purity 99%



FAM-ER(dec)-R purity 99%



## UPLC analysis of peptide

UPLC conditions

Column : ACQUITY UPLC® BEH C18 1.7 µm column (2.1 × 50 mm)

Mobile phase : A = 0.1% TFA/water, solvent, B: 0.1% TFA/CH<sub>3</sub>CN gradient: 10-90% gradient of solvent B over 4 min).

flow rate: 0.5 mL/min



Figure S2. HPLC trace of the synthesized decoys, peptide and chimeric molecules.

## MALDI TOF MS

LCL-HDO-F



Figure S3. MALDI TOF MS spectrum of decoys and chimeric molecules.

# HR-MS (ESI)

 $P4-N_3 \quad N_3-C6-LGAQSNF-NH_2$ 



m/z: calcd for C<sub>38</sub>H<sub>58</sub>N<sub>12</sub>O<sub>12</sub> [M+H]<sup>+</sup> : 874.4535, found : 874.4218



Figure S4. ESI TOF MS spectrum of peptide.

## Fluorescence polarization assay



Figure S5. The competitive binding assay of LCL-ER(dec), LCL-HDOcl, LCL-HDOdna and CPP-HDO-PROTAC to the ER $\alpha$ . Condition: 62.5  $\mu$ M-0.032 nM decoy in TE buffer (pH 7.5) containing 25 mM NaCl, measured at 25°C. The data represent means S.D. (n = 3).

# Biology Western blotting



**Figure S6.** Degradation of ERα by LCL-ER(dec) and LCL-ER(dec)-P4. MCF-7 cells were treated (for 24 h) with the indicated concentrations of LCL-ER(dec) and LCL-ER(dec)-P4.

#### **Cell proliferation assay**



**Figure S7.** Effects of PROTACs on the proliferation of ER $\alpha$ -positive breast cancer cells. Growth inhibition of ER $\alpha$ -positive breast cancer cells by PROTACs. MCF-7 cells were transfected with 1–10  $\mu$ M of the indicated PROTACs for 72 h, and cell proliferation was then evaluated using a cell viability assay. Data represent the mean ± standard deviation (n = 5).



**Figure S8.** Effects of PROTACs on the proliferation of ER $\alpha$ -positive breast cancer cells. Growth inhibition of ER $\alpha$ -positive breast cancer cells by PROTACs. MCF-7 cells were treated with 1–10  $\mu$ M of the indicated PROTACs for 72 h, and cell proliferation was then evaluated using a cell viability assay. All PRORACs were added without transfection reagents. Data represent the mean  $\pm$  standard deviation (n = 5).