Probing the Structural Requirements for Thiazole-Based Mimetics of Sunitinib as Potent VEGFR-2 Inhibitors

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Biological screening

In vitro antitumor testing against HepG2, HCT-116, MCF-7, HeP-2 and Hela cancer cell lines, and cytotoxicity against WI38 and WISH normal cell lines

Materials

Cell lines

Liver (HepG2), colon (HCT-116), breast (MCF-7), larynx (HeP-2) and cervix (Hela) cancer cell lines as well as lung fibroblast (WI38) and amnion epithelial (WISH) normal cell lines were obtained from American Type Culture Collection (ATCC) *via* Holding Company for Biological Products and Vaccines (VACSERA), Cairo, Egypt.

Chemical reagents

The reagents, Roswell Park Memonial Institute (RPMI-1640) medium, 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), DMSO, doxorubicin and sunitinib were obtained from Sigma-Aldrich co., St. Louis, USA. Fetal bovine serum was obtained from GIBCO, UK.

Procedure

Compounds **3-6** and **8-11** were screened for their *in vitro* antitumor activity against HepG2, HCT-116, MCF-7, HeP-2 and Hela cancer cell lines. In addition, compounds **3a,b**, **6g**, **8a** and **10c** were screened for their *in vitro* cytotoxicity against WI38 and WISH normal cell lines adopting MTT assay [1-3], and using doxorubicin, sunitinib as reference antitumor agent. Cells were cultured in Roswell Park Memorial Institute (RPMI) 1640 medium with 10% fetal bovine serum, penicillin (100 units/mL) and streptomycin (100 µg/mL) at 37 °C in an atmosphere of

5% CO₂ were added. Cells were placed in 96-multiwell microtiter plates (10⁴ cells/well) for 24 hours at 37 °C and in an atmosphere of 5% CO₂ before treatment with the compounds to allow attachment of the cells to the wall of the plate. The tested compounds were dissolved in DMSO and diluted with phosphate buffer saline (PBS) to obtain different concentrations. Tested compounds of different concentrations were added to each well and cells were incubated with the compounds for 48 hours at 37 °C and in atmosphere of 5% CO₂. All tests were performed in triplicates. The treated cells were washed with PBS and 100 μ L of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide solution (MTT) (5 mg/mL MTT stock in PBS diluted to 1 mg/mL with 10% RPMI-1640 medium) was added. The 96-multiwell plates were read by microarray reader Perkinelmer vector 3V multilabel counter model 1420 (Perkinelmer, Boston, MA) for optical density at 490 nm [1-3]. The relative percentage cell viability was calculated from the following equation:

% Cell viability =
$$\frac{A_{\text{treated cells}} - A_{\text{blank}}}{A_{\text{untreated cells}} - A_{\text{blank}}} \times 100$$

The relation between surviving fraction and drug concentration is plotted to get the survival curve for HepG2, HCT-116, MCF-7, HeP-2 and Hela cancer cell lines, and WI38 and WISH normal cell lines. The concentration required for 50% inhibition of cell viability ($IC_{50}/\mu M$) was obtained for each compound from the curve fitting using Sigma plot10.

In vitro kinase inhibition assay against VEGFR-2 for compounds 3a,b, 6g, 8a and 10c

In vitro kinase inhibition assay was carried out against VEGFR-2 using ELISA kit (Cat. # 40325, BPS Bioscience, USA). The procedure of the used kits was done according to the manufacturer's instructions. Briefly, compounds **3a,b**, **6g**, **8a** and **10c** and control drug, sunitinib, were made to different serial dilution. 5x kinase buffer 1, ATP and PTK substrate poly (Glu:Tyr 4:1) (10 mg/mL) were thawed. After that, the master mixture [N wells x (6 μ 5x kinase buffer 1 + 1 μ l ATP (500 μ M) + 1 μ l PTK substrate Poly (Glu:Tyr 4:1) (10 mg/mL)+ 17 μ l water)] was prepared followed by addition of 25 μ l to every well. Then, 5 μ l of inhibitor solution was added to each well and labeled as "test inhibitor". For the "positive control" and "blank wells", 5 μ l of the same solution without inhibitor (inhibitor buffer) was added. 3 mL of 1x kinase buffer 1 was prepared by mixing 600 μ l of 5x kinase buffer 1 was added. VEGFR-2

enzyme was thawed on ice. Upon first thaw, tube containing enzyme was spinned briefly to recover full content of the tube. The amount of VEGFR-2 required for the assay was calculated and the enzyme was diluted to 1 ng/µl with 1x kinase buffer 1. The reaction was initiated by adding 20 µl of diluted VEGFR-2 enzyme to the wells designated "positive control" and "test inhibitor control" and incubated at 30°C for 45 minutes. Kinase-Glo Max reagent was thawed. After the 45 minutes, 50 µl of Kinase-Glo Max reagent was added to each well. The plate was covered with aluminum foil and incubated at room temperature for 15 minutes. The luminescence was measured using the microplate reader. The values of % activity versus different concentrations of the test compound (with semi-log decrease in concentration) were then plotted using nonlinear regression analysis of sigmoidal dose-response curve, and the concentration that induces 50% inhibition of activity (IC₅₀) of VEGFR-2 were determined [4,5].

Cell cycle analysis and induction of apoptosis

Flow cytometry analysis of DNA content for cell cycle

HCT-116, MCF-7 and Hela cancer cells (2 x 10^5 cells/well) were treated with different concentrations of compound **10c**, and incubated for 24 hours at 37 °C in an atmosphere of 5% CO₂. The cells were washed twice with ice-cold PBS, collected by centrifugation, and fixed in ice-cold 70% ethanol, washed with PBS, and then they were incubated with propidium iodide (PI) staining solution that contains 50 µg/mL PI, 0.1 mg/mL RNase A and 0.05% Triton X-100. After incubation for 1 hour at room temperature, the cells were analyzed in the dark at 37 °C by flow cytometry (Becton Dickinson FACS, San Jose, CA) [6,7]. The cell cycle distributions were determined using Cell-Quest software.

Analysis of cellular apoptosis

Annexin V-fluorescein isothiocyanate (Annexin V-FITC) is a protein that possesses high affinity to phosphatidyl serine (PS), which can be detected by staining with annexin V-FITC and counter staining with propidium iodide (PI). Apoptosis was initially induced by incubation of cells in a density of 4 x 10⁶ cells/well in 100 μ L of complete growing medium with addition of the tested compound **10c** (100 μ L) in 96-microwell plates for 24 hours. Then, 0.5 x 10⁶ cells were collected by centrifugation and resuspended in 500 μ L of binding buffer. Annexin V-FITC (5 μ L) along with PI (5 μ L) were added, and incubated in the dark for 5 minutes at room temperature. Analysis of annexin-V-FITC binding was performed by flow cytometry (Becton Dickinson FACS, San Jose, CA) [6,7].

Caspase3/9 assay

The level of human caspase-3 was assessed using ELISA kit that was obtained from Invitrogen INC. Catalog # KHO1091 (Invitrogen, Inc., Waltham, Massachusetts, USA) in accordance with the manufacturer instructions. Also, human caspase-9 ELISA kit was obtained from DRG International, INC. Catalog # EIA-4860 (DRG International, Inc., Mountain Avenue Springfield, USA) in accordance with the manufacturer instructions. The tested compound **10c** (10μ M) was added to the cells and lysed with cell extraction buffer. After incubation at room temperature for 2 hours, wells were washed 4 times with the wash buffer, followed by addition of 100 mL of Invitrogen caspase-3 (active) or caspase-9 monoclonal antibody solution and incubated again at room temperature for another 1 hour. Wells were washed 4 times with the wash buffer, the anti-rabbit IgG horseradish peroxidase (HRP) solution (100 mL) was added, and the wells were incubated for 30 minutes at room temperature. After washing 4 times with the wash buffer, stabilized chromogen was added. Later on, the stop solution (100 mL) was added after incubation for 30 minutes. Absorbance was measured at 450 nm [8].

In vivo antitumor activity of compound 10c

In vivo antitumor screening was conducted after fulfillment of ethical statement and approval protocol for animal study from research ethical committee, Faculty of Pharmacy, Mansoura University. Evaluation of the *in vivo* antitumor activity of compound **10c** was achieved using EAC solid tumor animal model [9]. Tweny five swiss albino eight-week female mice (weighing 20-25 g) were obtained and divided into three groups (n = 5). Group 1: positive control-received equivalent volume of physiological saline; group 2: treated intraperitoneally with compound **10c** (10 mg/kg); group 3: treated intraperitoneally with doxorubicin (2 mg/kg). 7,12-Dimethylbenz[a]anthracene was used to induce breast cancer in mice. All groups were injected intraperitoneally one dose per day in a period of 20 days. Monitoring of the body weight and tumor volumes of mice was done every five days, the volume of solid tumor was monitored using a digital Vernier caliper and was calculated using the following equation:

Tumor volume (mm³) = Length(mm) × [Height(mm)]² × 0.52

Tumor growth inhibition (%TGI) was calculated after 5,10,15 and 2 days by the formula:

%TGI =
$$\frac{1 - \frac{(Tt/T0)}{(Ct/C0)}}{1 - (\frac{C0}{Ct})} \times 100$$

Where:

Tt = median tumor volume of treated group at time t.

T0 = median tumor volume of treated group at time 0.

Ct = median tumor volume of control group at time t.

C0 = median tumor volume of control group at time 0.

At the end of the experiment, mice were sacrificed and tumors were excised, fixed in 10 % formalin solution, and embedded in paraffin.

Molecular modeling study

The molecular modeling calculations were done applying swiss dock online server [10-12]. The 3D poses was visualized by UCSF chimera software [13-15] and the 2D poses were visualized by ProteinsPlus structure-based modeling support server [16-19]. The most stable conformer of the newly synthesized analogs **3a,b**, **6g**, **8a** and **10c** was docked into the binding pocket of VEGFR-2 (PDB code: 4agd) [20,21] which was obtained from the Protein Data Bank (PDB).

HPLC purity testing

The HPLC purity for compounds **3b**, **6g**, **8a** and **10c** was tested using Knauer high performance liquid chromatography apparatus with variable-wavelength UV-visualization detecdtor operatrd at 240 nm. The mixture was separated using a shim-pack cyano column (150 mm x 4.6 mm, 5 μ m particle size). The composition of the mobile phase was Methanol: phosphate buffer (50: 50, v/v) adjusted at pH 4, and programmed at flow rate of 1 mL/min with UV detection at 240 nm. Filtration of the mobile phase was carried out through 0.45 μ m millipore membrane filter. Separation was performed after sonication of the mobile phase for 10 min to remove air bubbles. All determinations were performed at ambient temperature.



Figure S1. Superimposition of the co-crystallized sunitinib (blue) and the re-docked sunitinib (green) in the VEGFR-2 active site (RMSD = 1.05 Å).





Figure S2. 3D and 2D Interaction pattern of sunitinib with the binding site of VEGFR-2.



Figure S3. 3D and 2D Interaction pattern of compound 3a with the binding site of VEGFR-2.





Figure S4. 3D and 2D Interaction pattern of compound 3b with the binding site of VEGFR-2.



Figure S5. 3D and 2D Interaction pattern of compound 6g with the binding site of VEGFR-2.





Figure S6. 3D and 2D Interaction pattern of compound 8a with the binding site of VEGFR-2.





Figure S7. 3D and 2D Interaction pattern of compound 10c with the binding site of VEGFR-2.

Group -	Body weight (g)					
	day 0	day 5	day 10	day 15	day 20	
Positive Control	18.20±1.068	23.00±0.707	28.20±0.860	32.60±0.509	37.00±0.707	
Doxorubicin	20.00±0.707	21.60±0.927	24.40±1.030*	26.40±0.979***	28.60±0.812****	
10c	19.20±0.734	23.00±1.140	27.00±0.948	30.20±0.583##	34.40±0.509*###	

Table S1. Body weight of mice for control, doxorubicin, and 10c-treated groups at 0, 5, 10, 15 and 20 days of treatment.

Positive control: mice with cancer implants and receiving no drug. **** p < 0.0001, *** p < 0.0005, * p < 0.05 vs. Positive control group. ### p < 0.0005, ## p < 0.01 vs. doxorubicin group.

Table S2. Tumor volume of the mammary gland of mice for control, doxorubicin, and **10c**-treated groups at 0, 5, 10, 15 and 20 days of treatment.

Group -	Tumor volume (mm ³)					
	day 0	day 5	day 10	day 15	day 20	
Positive Control	0.180±0.010	0.680±0.039	1.322±0.151	2.250±0.129	3.078±0.160	
Doxorubicin	0.186±0.010	$0.268 \pm 0.010^{****}$	0.368±0.014****	0.484±0.016****	0.600±0.012****	
10c	0.188±0.010	0.504±0.028**###	$0.680 \pm 0.050^{***}$	0.964±0.108****#	1.024±0.088****#	

Positive control: mice with cancer implants and receiving no drug.

**** p < 0.0001, *** p < 0.001, ** p < 0.005 vs. Positive control group.

p < 0.001, # p < 0.05 vs. doxorubicin group.

Group	%TGI				
	day 5	day 10	day 15	day 20	
Doxorubicin	84.16	84.58	86.07	86.18	
10c	39.49	58.75	64.11	72.38	

Table S3. % Tumor Growth Inhibition (%TGI) of breast solid tumor in mice after 5, 10, 15 and 20 days of treatment with doxorubicin and compound **10c**.

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Mansoura University Faculty of Pharmacy Research Ethics Committee

> السيد الأستاذ الدكتور / رئيس مجلس قسم الكيمياء الدوائية تحية طبية وبعدي

بناء على الخطاب الوارد من سيادتكم بشأن موافقة مجلس قسمكم الموقر في ٢٠٢/٨/١٠ على الطلب المقدم من أ.د / سعيد محمد بيومي - المشرف الرئيسي علي رسالة الدكتوراه الخاصة بالصيدلانية / الاء عبدالله عبدالحميد محمد – المدرس المساعد بقسم الكيمياء الدوائية لإضافة جزء خاص باجراء أبحاث علي الحيوان (الفئران المعملية) وكذلك تعديل عنوان الرسالة حتي يكون العنوان اكثر تحديدا ليتوافق مع ما تم تحضيره من مركبات وانجازه فعليا في خطة الرسالة.

عنوان الرسالة باللغة العربية والانجليزية قبل التعديل

تشييد ودراسات حاسوبية لأنظمة نيتروجينية غير متجانسة جديدة متوقع لها فعالية بيولوجية

Synthesis and *in silico* studies of certain new nitrogenous heterocyclic systems of expected biological activity

عنوان الرسالة باللغة العربية والانجليزية بعد التعديل

"تصميم وتشييد ودر اسات حاسوبية لبعض مشتقات الثيازول ومشتقات الثيازول المدمجة الجديدة كمضادات محتملة للسرطان"

"Design, Synthesis and in silico studies of some new thiazole and fused

thiazole derivatives as potential anticancer agents "

فان لجنة اخلاقيات البحث العلمي في اجتماعها بتاريخ ٢٠٢٢/٨/٢٤ قد احيطت علما بالتعديل الذي تم في عنوان الرسالة كما وافقت علي استمارة البحوث الحيوان المستوفاة بمعرفة وبتوقيع ١.د / المشرف الرئيسي وذلك بعد التأكد من توافق التجارب المقترحة مع معايير أخلاقيات البحث العلمي مع التزام المشرف الرئيسي بتقديم اسم الباحث الذي اجري جزء التجارب العملية علي الحيوان عند انهاء البحث.

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وتفضلوا سيادتكم بقبول وافر الاحترام والتقدير...

مقرر لجنة اخلاقيات البحث العلمي (المح^{24/21} مـ أ.م.د / امل احمد عطوه سلام

MANSOURA:35516 - Tel & Fax:050/2247496



alaa-16a #148 RT: 2.49 AV: 1 SB: 6 2.79 , 2.48-2.54 NL: 6.36E2 T: + cEl Full ms [40.00-1000.00]



Alaa Abdullah-2-HNMR-DMSO-AF



Alaa Abdullah-2-HNMR-DMSO-AF













alaa-16b#66 RT: 1.12 AV: 1 SB: 2 1.61 , 1.61 NL: 6.57E2 T: + cEl Full ms [40.00-1000.00]



Alaa Abdullah-2-HNMR-DMSO-AF













alaa-17 #108 RT: 1.82 AV: 1 SB: 6 2.79 , 2.48-2.54 NL: 4.19E2 T: + cEl Full ms [40.00-1000.00]





Alaa Abdullah-12-HNMR-DMSO-AF
















alaa-19a #239-242 AV: 4 SB: 2 2.71, 2.71 NL: 1.11E2 RT: 4.02-4.07

m/z











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alaa-19f #1 RT: 2.24-2.29 AV: 4 SB: 2 1.74, 1.67 33-138

m/z







alaa-19g #34 RT:0.59 AV:1 SB:18 0.54-0.74 ,0.50-0.57 NL:1.89E3 T:{0,0} + c El Full ms [40.00-1000.00]















Alaa abdallah-VII-Hnmr-ES





alaa-21a #64 RT: 1.09 AV: 1 SB: 2 0.97 , 1.15 NL: 3.72E3 T: {0,0} + c El Full ms [40.00-1000.00]




























alaa-22b #8 SB: 2 0.87, 0.89 NL: 4.64E2 RT: 0.15 AV:

m/z







Alaa Abdallah-14-carbon-WH





alaa-22c #131-133 RT: 2.21-2.24 AV: 3 SB: 6 2.80 , 2.48-2.54 NL: 1.24E2 T: + cEl Full ms [40.00-1000.00]





0.95

2.05

Alaa Abdullah-24-HNMR-DMSO-AF

1.00

4.15

8

8

1.00







alaa-23a #29-31 RT: 0.50-0.54 AV: 3 SB: 6 2.80 , 2.48-2.54 NL: 9.87E1 T: + cEl Full ms [40.00-1000.00]

m/z





















m/z





















SI		65536	
SF		400.2000000	MHZ
WDW		EM	
SSB	0		
LB		0.30	HZ
GB	0		
PC		1.00	








alaa-24 #185-187 RT: 3.11-3.15 AV: 3 SB: 6 2.80 , 2.48-2.54 NL: 2.07E2 T: + cEl Full ms [40.00-1000.00]

m/z



alaa abdallah 22-M c13



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Current Data Parameters alaa abdallah clbcl -M cl3 10 1 F2 - Acquisition Parameters 20210302 9.37 h spect Z108618 0945 (zgpg30 65536 DMSO 2100 4 24038.461 Hz 0.733596 Hz 1.3631488 sec 197.77 20.800 usec 6.50 usec 2.00000000 sec 0.03000000 sec 1 100.6404331 MHz 13C 10.00 usec 47.0000000 W 400.2016008 MHz 1H waltz16 90.00 usec 13.0000000 W 0.29249999 W 0.14713000 W F2 - Processing parameters 32768 100.6303700 MHz EM 1.00 Hz