Supplementary Information

Ov-rich γ-MnO² enhanced electrocatalytic three-electron oxygen reduction to hydroxyl radicals for sterilization in neutral media

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Electrochemical measurements.

All the electrochemical tests were conducted on CS2350M electrochemical workstation (Wuhan Corrtest Instrument Co., LTD). Before electrochemical measurements, 5 mg of as-prepared catalysts were dispersed in 1 mL mixture of DI water, ethanol, and 5 wt% Nafion solution (volume ratio is 1: 1: 0.0001) to form a uniform ink. All the electrochemical measurements were conducted in a three-electrode system, with a catalyst-modified RRDE as working electrode, saturated calomel electrode (SCE) as reference electrode, and carbon rod as counter electrode. 10 μL ink was dropped onto the glassy carbon disk electrode of RRDE, and rotary drying under the ambient environment. Cyclic voltammetry was conducted in N₂-saturated 3.5 wt% NaCl solution at a potential range of 0-1.2 V *vs.* RHE, with a scan rate of 500 mV/s for 50 cycles, to get a stable curve. The linear polarization curve (LSV) was measured at 0- 1.1 V *vs.* RHE with a scan rate of 20 mV/s and a rotate speed of 1600 rpm. The RRDE tests were performed in an O_2 -saturated solution with a scan rate of 20 mV/s and rotate speed of 1600 rpm, with the ring potential was fixed at 1.2 V *vs.* RHE. The selectivity of H_2O_2 (H_2O_2 %) via 2e⁻ ORR pathway and the number of transfer-electron (n) can be calculated by the following equation based on the disk current (I_D) and ring current (I_R) measured by RRDE.

$$
H_2O_2\% \, (\%) = \frac{200 \times I_R/N}{|I_D| + I_R/N} \tag{S1}
$$

$$
n = \frac{4 \times |I_D|}{|I_D| + I_R/N} \tag{S2}
$$

Here, N refers to the collection coefficient of RRDE, which depends on only the size of RRDE and is 37% in this system.

H2O² yield tests.

The tests of H_2O_2 yield were conducted in a traditional three-electrode system with catalyst-loaded carbon paper (mass loading: 0.2 mg cm^{-2}) as working electrode, saturated calomel electrode as reference electrode, and graphite rod as counter electrode. All the tests were performed in 35 mL O₂-satureted 3.5 wt% NaCl solution at 0.1 V (*vs.*) RHE) for 2 h in a separate chamber with Nafion 117 as the membrane. The concentration of H_2O_2 was measured using the cerium sulfate titration method based on

the color variation with Ce^{4+} concentration as described in Eq. S3. The Ce^{4+} concentration-absorbance standard curve was plotted by UV-visible adsorption peak at 318 nm for a range of Ce⁴⁺ solutions, and the final correction curve was $v = -0.015 +$ 5.21 x (y is UV-visible absorbance, x is the concentration of Ce^{4+}). The electrolyte after i-t test was mixed with $0.5 \text{ mM Ce(SO₄)₂$ solution at a volume ratio of 1: 49, and reacted for another 2 h, and the corresponding H_2O_2 concentration $(c_{H_2O_2})$ was calculated by Eq. S4. Finally, the H_2O_2 yield was calculated by Eq. S5.

$$
2Ce^{4+}(\text{yellow}) + H_2O_2 \rightarrow 2Ce^{3+}(\text{colorless}) + 2H^+ + O_2 \uparrow \tag{S3}
$$

$$
c_{H_2O_2}(mM) = \frac{(c_0 - x)/2}{1/50}
$$
 (S4)

Where the c_0 is the original Ce⁴⁺ standard solution with a concentration of 0.5 mM.

$$
H_2O_2 \text{ yield rate } (\text{mmol } g_{cat}^{-1} \text{ } h^{-1}) = \frac{c_{H_2O_2} \cdot v}{t \cdot m_{cat}} \tag{S5}
$$

Where v is the volume of electrolyte (35 mL), t is reaction time (2 h), and m_{cat} is the loading mass of as-prepared catalysts $(0.2 \,\mu g \text{ cm}^{-2})$.

Conditions of on-line SPE LC-MS/MS system.

Salicylic acid in samples was concentrated and detected by the on-line SPE-LC-MS/MS system, which was carried out on a 1290 II ultrahigh-performance liquid chromatography coupled with a 6470 triple quadrupole mass spectrometry system (Agilent, Santa Clara, CA, USA). A schematic of the online SPE-LC-MS/MS is presented in **Fig. S1.** The online SPE system consists of a high-performance (with a 900 μ L sample loop), a quaternary pump (P1), binary pump (P2) and a thermostatic column compartment equipped with a six-port valve. Chromatographic separation was performed on a Zorbax Extend C18 (3.5 μ m, 3.0 mm \times 150 mm) from Agilent (Santa Clara, CA, USA). A guard column Zorbax Eclipse XDB-C8 (5 μ m, 2.1 mm \times 12.5 mm) from Agilent was employed as a trap column for the online SPE. Details of on-line SPE-LC-MS/MS conditions are listed in **Table S1 and S2**.

DFT calculation.

All density functional theory (DFT) calculations carried out on the $MnO₂(O_v)$ and $Mn₃O₄(O_v)$ catalysts were performed via Vienna ab initio simulation package (VASP) with the projector augmented wave (PAW) method¹. The generalized gradient

approximation (GGA) with the Perdew-Burke-Ernzerhof (PBE) was adopted to describe the exchange-correlation functional². The DFT-D3 method is used to introduce van der Waals (vdW) interaction. Spin polarization was considered in the calculation³. The U parameter of the Mn atom is set to 4 eV^4 . The kinetic energy cutoff was set to 400 eV, and the force and energy convergence criteria were set to 0.03 eV/ \AA and 10⁻⁴ eV, respectively.

A slab model of MnO² with (300) crystal faces consisting of 16 Mn atoms and 32 O atoms was constructed using an optimized $MnO₂$ single cell. A slab model of $Mn₃O₄$ with (101) crystal faces consisting of 28 Mn atoms and 48 O atoms was constructed using an optimized Mn_3O_4 single cell. The vacuum layer of the slab model for MnO_2 and Mn_3O_4 is set to 20 Å. $MnO_2(O_v)$ and $Mn_3O_4(O_v)$ slab models with oxygen vacancies are constructed by removing one oxygen atom from the surface of the $MnO₂$ and $Mn₃O₄$ slab models. For geometric optimization, the bottom two atomic layers of all slab models are fixed, and the upper atomic layers remain relaxed.

The adsorption energy of O_2 molecules at active sites on the catalyst surface was calculated by the following equation:

$$
\Delta E_{*O_2} = E_{*O_2} - E_* - E_{O_2}
$$

Where * refers to O_v - γ -Mn O_2 or O_v -Mn₃ O_4 . E_{Q_2} refer to the total energy after the adsorption of O₂ on the catalyst surface. E_* is the total energy of the catalyst. E_{O_2} refer to the energy of O₂ molecules.

During the calculation of the reaction mechanism, Gibbs free energy (G) was obtained by the following equation:

$$
G = E + ZPE - TS
$$

Where E , ZPE and TS were total energy, zero-point energy and entropic contributions, respectively. ZPE and TS were processed by the vaspkit code at $298.15 K⁵$.

Antibacterial performance tests.

Pseudomonas aeruginosa (*P. aeruginosa*) and staphylococcus aureus (*S. aureus*) were used as the typical strain to invest the antibacterial performance of as-prepared catalysts. First of all, the typical strains were cultured in LB medium, after 12 h preservation at

37℃, the bacteria body was centrifugated at 4000 rpm for 5 min, and redispersed in 0.1 M phosphate buffer saline (PSB) to get a uniform solution with a concentration of $10⁷$ cfu/mL. The antibacterial test was carried in 35 mL above solution, with catalystmodified carbon paper (1×2 cm, with a loading mass of 0.2 mg/cm²) as working electrode, SCE as reference electrode, and carbon rod as counter electrode. Electrolytes were collected after 0, 10, 30, 60, 120, and 180 min chronoamperometry test with an applied potential at 0.1 V *vs.* RHE. Coating the above electrolytes onto nutrient agar medium plates, keeping at 37℃ for 24 h, counting the colony number of each plate and calculating the disinfection rate with the variation of time.

Figure S1 Schematic of the online SPE-LC-MS/MS.

Figure S2 SEM images of PBA precursor.

Figure S3 SEM images of ammonia-treated PBA.

Figure S4 SEM images of Mn₃O₄-x with different calcination temperature. (a) Mn3O4-700, (**b**) Mn3O4-800, (**c**) Mn3O4-900.

Figure S5 SEM images of MnO₂-700A

Figure S6 SEM images of MnO₂-800A

Figure S7 SEM images of MnO₂-900A

Figure S8 XRD patterns of as-prepared MnO₂-xA catalysts.

Figure S9 XPS patterns of as-prepared MnO₂-xA catalysts. (a) survey-scan spectra,

(**b**) O 1s spectra, (**c**) Mn 2p spectra.

Figure S10 RRDE polarization curves of Mn₃O₄-x.

Figure S11 Electrochemical performance of as-prepared MnO₂-xA catalysts. (a) RRDE polarization curves, (**b**) H2O² selectivity, (**c**) number of transfer electron, (**d**) Tafel curves.

Figure R12 The stability of as-prepared catalysts.

Figure 13 (**a**) UV-visible spectra of CeSO⁴ at various concentrations, (**b**) calibration curve of the absorbance and Ce^{4+} concentration at 318 nm.

Figure S14 The equation of the reaction of salicylic acid and hydroxyl radical.

Figure S15 (**a**) curves of LC-MS/MS for salicylic acid at different masses, (**b**) calibration curve of the peak area and mass of salicylic acid, (**c**) variation of salicylic acid concentration with different reaction times.

Figure S16 The configurations of (**a**) Mn₃O₄ (101), O_v-Mn₃O₄ (101), (**b**) γ -MnO₂ (300) and O_v - γ -MnO₂ (300) models.

Figure S17 The O position on adsorbed *OOH in Ov-γ-MnO² (300).

Figure S18 Geometry adsorption configurations of 3e⁻ ORR progress on O_v-γ-MnO₂

(300).

Figure S19 Geometry adsorption configurations of 3e⁻ ORR progress on O_v-Mn₃O₄

(101).

Figure S20 The projected density of state (PDOS) analysis. (a) O_v -Mn₃O₄ (101) and (**b**) Ov-γ-MnO² (300). (The two Mn atoms in the blue dotted circles are the active sites of as-prepared catalysts, all the PDOS and d-band center analysis are based on that.)

Figure S21 Digital pictures of colony plates with different reaction times without the addition of IPA. (**a**) *P. aeruginosa*, (**b**) *S. aureus*.

Figure S22 Variety of sterilization efficiencies for different reaction times.

Figure S23 Digital pictures of colony plates with different reaction times with the addition of IPA. (**a**) *P. aeruginosa*, (**b**) *S. aureus*.

Table S1. LC parameters and MS/MS parameters of the instrumental method for Salicylic acid analysis.

LC parameters					
Instrument	HPLC 1260 and UPLC 1290 (Agilent Technologies, USA)				
Analytical	Zorbax Extend C18, 3.5 μ m, 3.0 mm × 150 mm (Agilent				
column	Technologies, USA)				
Trapping column	Zorbax Eclipse XDB-C8, 5 μ m, 2.1 mm × 12.5 mm (Agilent				
		Technologies, USA)			
Injection volume			$100 \mu L$		
		(Needle rinsed once with 1:1 methanol: water before injection)			
Column					
temperature	30 °C				
Mobile phases	Flow rate: 0.3 mL min ⁻¹				
UP 1290	A1: Ultrapure water with 0.5% formic acid				
	B1: Methanol				
Mobile phases	A: Ultrapure water with 0.5% formic acid				
HP 1260	B : Methanol				
Gradient	Time / min	$A / \%$ $B / \%$		note	
UP 1290	0.0	80	20		equilibration
	2.0	80	20		
	7.0	50	50		
	10.5	$\boldsymbol{0}$	100		
HP 1260	Time /	Flow rate / mL	$A / \%$	$B / \%$	note
	min	min^{-1}			
	$\boldsymbol{0}$	0.5	95	$\boldsymbol{0}$	loading
	$\overline{2}$	0.5	95	$\boldsymbol{0}$	
	10.5	0.5	95	$\boldsymbol{0}$	equilibration
MS/MS parameters					

Instrument $\begin{vmatrix} 6470 \text{ triple quadrupole mass spectrometer (Agilent)} \end{vmatrix}$

	Technologies, USA)			
Ion source	Agilent Jet stream (Agilent Technologies, USA)			
Ionization	Electrospray ionization (ESI) in negative mode			
Gas temp	300 °C			
Gas flow	3 L/min			
Nebulizer	5 psi			
Sheath gas temp	350 °C			
Sheath gas flow	7 L/min			
Capillary voltage	3500 V			
Scan type	MS ₂ SIM			

Table S2. Retention times, molecular formula (precursor ion), monitored mass, fragmentor, collision energy.

Acronym	Retention			Molecular Monitored Fragmentor Collision	
analyte	Time/min	formula	${\rm mass / m/z}$ / V		energy / V
SA	9.85	$[C_7H_6O_3]$	-137	100	\mathcal{L}

Table S3 Binding energy and ∆E of as-prepared catalysts in Mn 3s XPS spectra.

	Binding energy /	ΔE /eV	
	eV		
Mn_3O_4-800	88.59 / 83.08	5.51	
$MnO2$ -700A	89.05/84.23	4.82	
$MnO2-800A$	88.76 / 83.99	4.77	
$MnO2$ -900A	88.78 / 83.94	4.84	

Table S4 Percentage of O with different configuration of as-prepared catalysts in O 1s spectra.

$MnO2$ -700A	38.8	19.4	12.2	29.6
$MnO2-800A$	39.1	17.6	22.2	21.1
$MnO2$ -900A	47.9	15.5	26.7	9.9

Table S5 The peak intensity of UV-visible spectrum and the corresponding Ce4+ concentration, the amount of H2O² and its yield rate after 2 h i-t tests.

	Peak	c_{Ce4+} / mM	n_{H2O2} /	$H2O2$ yield rate
	intensity		mmol	/ mmol g^{-1} h ⁻¹
Mn_3O_4-800	2.10	0.406	82.25	205.63
$MnO2$ -700A	2.12	0.401	78.75	196.88
$MnO2-800A$	1.25	0.243	224.88	562.19
$MnO2$ -900A	2.06	0.398	89.25	223.13

Table S6 The peak area of HPLC curve and the corresponding SA mass and ·OH concentration.

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