1	Supplementary Information
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3	A quenching electrochemiluminescence energy resonance
4	transfer system based on CdS and COFs for ultrasensitive
5	detection of CA242
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#### 26 Additional experimental section

27 CA242, anti-CA242, and other cancer antigens used in this work were bought from Shanghai Linc-Bio Science Co. Ltd. 2,5-dimethoxyterephaldehyde (DMTP) and 1,3,5-28 Tris(4-aminophenyl) benzene (TAPB) were obtained from Anhui Zesheng Technology 29 Co., Ltd., Cadmium nitrate tetrahydrate (Cd(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O), sodium suifide 9-hydrate 30 (Na<sub>2</sub>S·9H<sub>2</sub>O), Tris (2-carboxyethyl) phosphine hydrochloride (TCEP) (3-aminopropyl) 31 triethoxysilane were obtained from Shanghai Macklin Biochemical Technology Co., 32 33 Ltd. 1-(3-Dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC), Nhydroxysuccinimide (NHS) and Bovine serum albumin (BSA, 96-99%) were obtained 34 from Shanghai Aladdin Biochemical Technology Co.Ltd. Phosphate buffer solution 35 (PBS, 0.1 mol/L KH<sub>2</sub>PO<sub>4</sub>, and 0.1 mol/L Na<sub>2</sub>HPO<sub>4</sub>) was used during the 36 electrochemical measurements. 37

X-ray diffraction (XRD) patterns were performed with D8S-3 advance X-ray 38 diffractometer (Bruker AXS, Germany), transmission electronmicroscopy (TEM) 39 images were taken by Tecnai G2 F20S-TWIN transmission electron microscope (Field 40 Electron and Ion Co., United States). Scanning electron microscopy (SEM) images 41 were taken by Quanta FEG250 field emission environmental electron microscope 42 (Field Electron and Ion Co., United States). The F-4500 fluorescence 43 spectrophotometer (Japan) was used to complete fluorescence emission spectrum. The 44 electrochemical data were obtained by using CHI-660 electrochemical analyzer in 45 standard three-electrode system with a glassy carbon electrode (GCE, diameter 4 mm) 46 as the working electrode, The fluorescence emission spectrum was measured by F-380 47 fluorescence spectrophotometer. All the chemical reagents were analytical grade and 48 were directly used without further purification. 49

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## 54 Characterization of CdS NCs



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Fig. S1. SEM image of CdS NCs.

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## 58 ECL stability of CdS/GCE



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- 60 Fig. S2. The CdS/GCE continuous scanned for 8 cycles in PBS containing 40 mM  $H_2O_2$ .
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### 63 Electrochemical characterization of the immunosensor



65 **Fig. S3.** (B) CV of (a) bare GCE, (b)CdS/GCE, (c) Ab1/CdS/GCE, (d) 66 BSA/Ab1/CdS/GCE, (e) CA242/BSA/Ab1/CdS/GCE, (f) Ab2-

#### 67 COFs/CA242/BSA/Ab1/CdS/GCE.

### 68 Optimization experiments



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**Fig. S3.** The optimizations of  $H_2O_2$  concentration.

#### 71 Limit of detection

The limit of detection can be calculated by a most common way and the equation
as follow<sup>1</sup>:

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$$X_{L} = X_{b1} + kS_{b1}$$

where  $X_L$  is derived from the smallest measure,  $X_{b1}$  is the mean of the blank 75 measures, S<sub>b1</sub> is the standard deviation of the blank measures and k is a numerical factor 76 chosen according to the confidence level desired. A value of 3 for k in equation is 77 strongly recommended, and it usually corresponds to a confidence level of about 90% 78 in a practical sense. The values for  $X_L$  and  $S_{b1}$  cannot usually be determined from theory 79 80 but must be found experimentally by making a sufficiently large number of measurements. In this work, after making twenty ECL measurements of blank, the X<sub>b1</sub> 81 = 10798.6 and  $S_{b1}$  = 66.3 was calculated. According to  $X_L$  =  $X_{b1}$  +  $3S_{b1},$  the  $X_L$  = 82 10997.5 was obtained, and the fitting equation of this proposed biosensor is I = 8185.7583 - 752.41×lg c. Thus, the LOD of this proposed biosensor is  $c_L = 10^{(10997 - 8185.75)/(-752.41)}$ 84  $\approx 0.000183$  U/mL. 85

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Detection method	Linear	Limit of	Reference	
Detection method	range(U/mL)	Detection(U/mL)		
Electrochemical biosensor	0.01-100	20	2	
Electrochemical biosensor	0.001-10000	0.0015	3	
AlphaLISA	0.16-400	0.16	4	
Electrochemiluminescence	0.005.100	0.0010	5	
biosensor	0.003-100	0.0019		
Electrochemical biosensor	1-150	0.04	6	
Electrochemiluminescence	0.001.1000	0.000527	7	
biosensor	0.001-1000	0.000557	/	
Electrochemiluminescence	0.001.1000	0.000182	this would	
biosensor	0.001-1000	0.000185		

92 Table S1. Comparison of different methods for the detection of CA242

**Table S2.** The recovery analysis results (bovine serum sample).

Concentration of CA242	The addition	The detection	RSD	Recovery
in the sample (U/mL)	(U/mL)	(U/mL)	(%, n=3)	(%)
0.55	1	1.73, 1.56, 1.52	6.9	105.4
	2	2.33, 2.59, 2.46	5.3	95.6
	3	3.41, 3.71, 3.91	3.0	104.0

# **Table S3.** The recovery analysis results (human serum sample).

Concentration of CA242	The addition	The detection	RSD	Recovery
in the sample (U/mL)	(U/mL)	(U/mL)	(%, n=3)	(%)
0.55	1	1.53, 1.61, 1.48	4.3	99.3
	2	2.54, 2.29, 2.67	7.6	97.5
	3	3.60, 3.74, 3.81	3.0	105.7

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