

1 **Supporting information**

2

3 **Dual responsive Curcumin loaded N-Isopropylacrylamide/Acrylated linseed oil**
4 **copolymeric hydrogel films: Tuned LCST with value-added properties**

5 P.A Parvathy ^{a,b}, Victor A Ajisafe ^c, Ashok M Raichur ^c, Sushanta K. Sahoo* ^{a,b}

6 a. Materials Science and Technology Division, CSIR-National Institute for Interdisciplinary Science and
7 Technology, Thiruvananthapuram 695019, India

8 b. Academy of Scientific and Innovative Research (AcSIR), Ghaziabad- 201002, India

9 c. Department of Materials Engineering, Indian Institute of Science (IISc), Bangalore 560012, Karnataka, India.

10

11

12

13

14

15

16

17

18

19 **Author Information**

20 Corresponding Author

21 *E-mail: sushanta@niist.res.in; Tel +91-471-2515373 (S.K.S)

22 **ORCID**

23 Sushanta K Sahoo: 0000-0003-4845-1049

24 **Materials**

25 Linseed oil (L.O.) (α -linolenic acid 51.9-55.2 %, palmitic acid 7 %, stearic acid 3.4-4.6 %,
26 oleic acid 18.5-22.6 %, linolenic acid 14.2-17 %) was purchased from HIMEDIA. Acrylic
27 acid and CDCl_3 were procured from Sigma-Aldrich. Hydroquinone and N, N, N, N'-
28 Tetramethyl ethylenediamine (TEMED) were obtained from TCI chemicals. 30 % w/v
29 hydrogen peroxide (H_2O_2) was procured from Nice chemicals. Seralite SRC 120 was
30 obtained from SRL Pvt. Ltd, India. Glacial acetic acid, sodium carbonate, sodium bicarbonate
31 and sodium sulfate were procured from Merck.

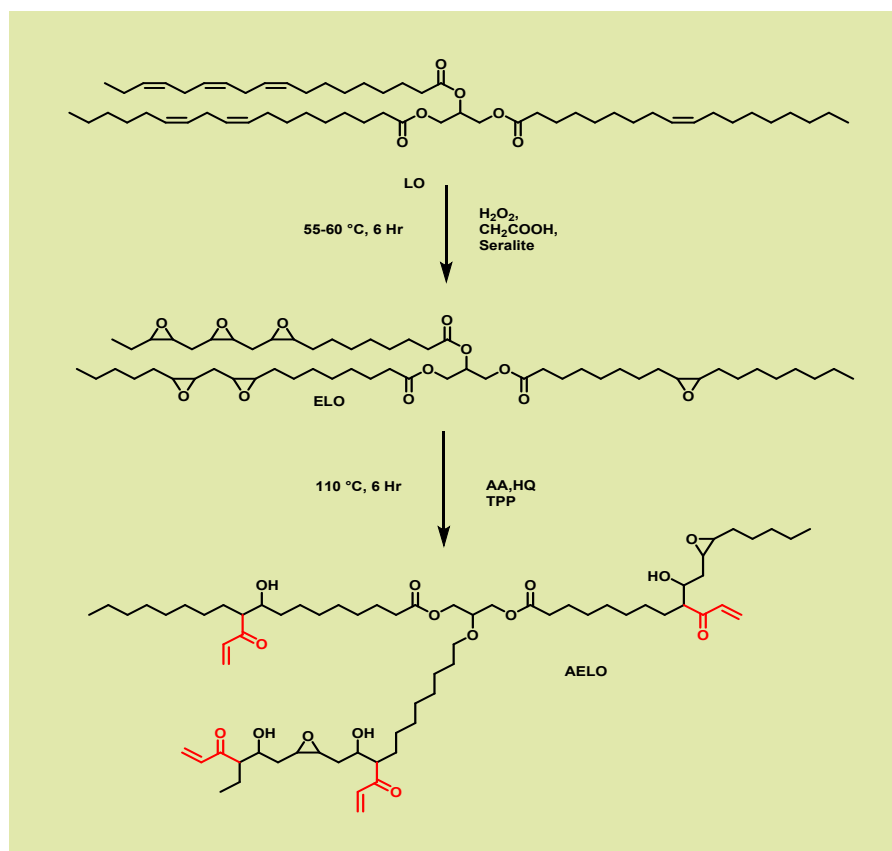
32 **Experimental section**

33 **Synthesis of Acrylated epoxidized linseed oil (AELO)**

34 Acrylated epoxidized linseed oil (AELO) was prepared via a two-step process reported by
35 our group (**Scheme S1**)¹. Initially, linseed oil was epoxidized employing Prilezhaev
36 epoxidation. 79 g of linseed oil was added to the 3-neck round bottom flask connected with
37 magnetic stirrer, followed by the addition of 30 g glacial acetic acid and seralite (19.75 g)
38 mixture and stirred for 30 min. After this, 113 g of H_2O_2 was added using a dropping funnel,
39 and then the reaction was carried out for 6 hr by maintaining the temperature between 55-60
40 °C. Afterward, the epoxidized layer was separated using filtration and washed with 2 wt.%
41 Na_2CO_3 solution and then distilled water. Epoxidized linseed oil (ELO) was then passed
42 through anhydrous sodium sulphate and dried using a vacuum oven.

43 For the acrylation process, 65 g of ELO and 0.5 wt. % hydroquinone were thoroughly mixed
44 in a two-neck round bottom flask. 19.58 g of acrylic acid was introduced dropwise to the
45 reaction media using a dropping funnel with continuous stirring. 1.23g (1.5 wt.% ELO+AA
46 mixture) of triphenylphosphine was added as a catalyst. The reaction continued for 6 hr at
47 110 °C. Acrylated epoxidized linseed oil (AELO) was washed with 2 wt.% NaHCO_3 solution

48 and passed over anhydrous Na_2SO_4 . Further drying of AELO was performed in a vacuum
49 oven.



50

51

Scheme S1. Acrylation of epoxidized linseed oil

52 Characterization

53 ^1H NMR spectrum was recorded using the 500 MHz Bruker Advance DPX spectrometer
54 using CDCl_3 as solvent.

55

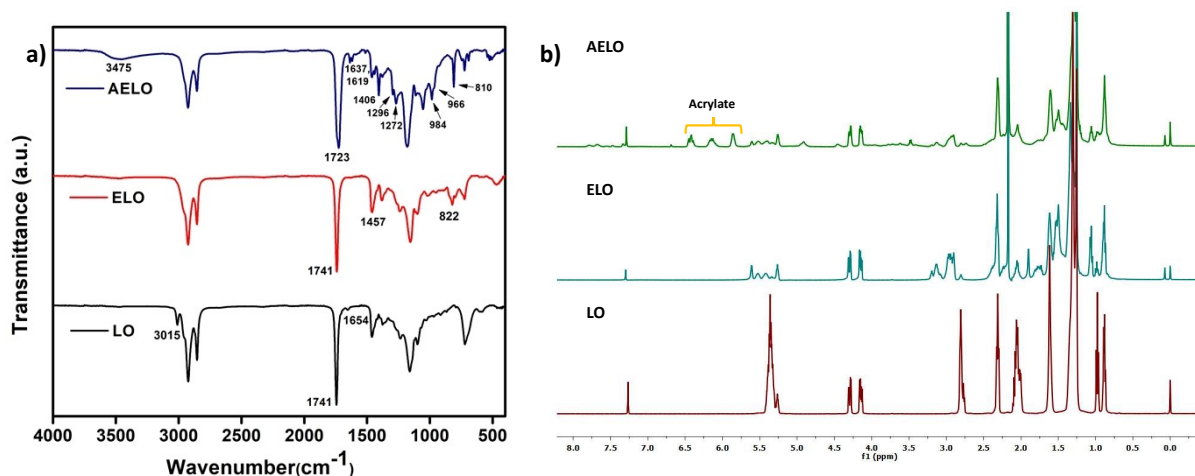
56 Results and discussion

57 Functional group analysis

58 ATR-FTIR and ^1H NMR (**Fig. S1**) are used to validate the formation of ELO and AELO. The
59 typical peak associated with C-H stretching from LO's $\text{C}=\text{C}-\text{H}$ at 3015 cm^{-1} is noticeably
60 diminished in ELO, as shown in **Fig. S1**¹. The peak at 1654 cm^{-1} attributed to $\text{C}=\text{C}$ is not

61 prominent in ELO, confirming that LO had undergone epoxidation. The vibration seen at 822
62 cm^{-1} repeatedly confirms the epoxidation of linseed oil. The acrylate moiety's double bond in
63 AELO is shown as a doublet at 1637 and 1619 cm^{-1} . The CH_2 scissoring vibration of $\text{CH}_2=\text{C}$
64 in the acrylate group is represented by the peak at 1406 cm^{-1} . The peaks at 984 and 966 cm^{-1}
65 and those at 1296 cm^{-1} and 1272 cm^{-1} are attributed to C-H scissoring vibration in acrylate
66 $\text{CH}=\text{C}$ and CH_2 rocking vibration of CH_2 in the acrylate groups, respectively^{2,3}.

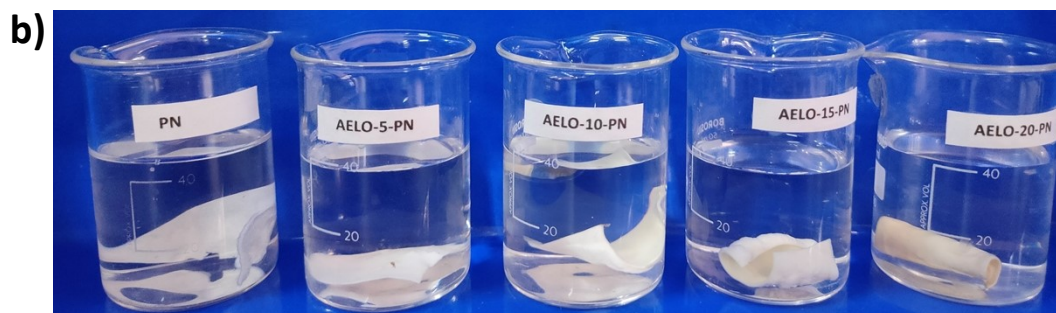
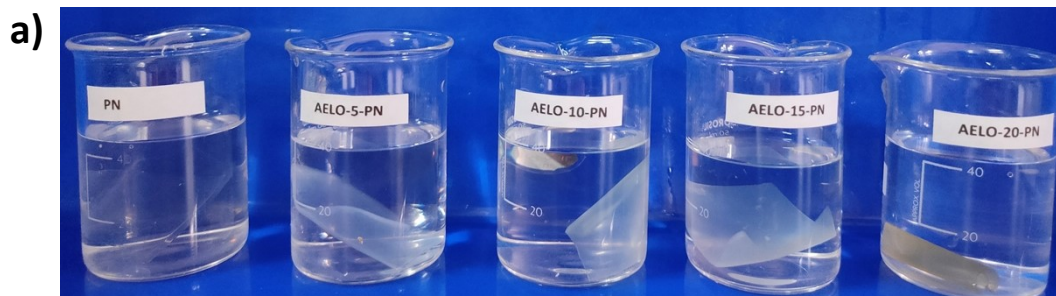
67 Epoxidation and acrylation are further confirmed with ^1H NMR. The olefinic hydrogen atoms
68 of double bond in LO. appeared at 5.3 ppm, which has significantly reduced in ELO. The
69 characteristic peaks of -CH- protons from epoxide moiety occurred at 2.9-3.12 ppm in ELO,
70 validating the epoxidation of LO. Three successive peaks at 5.8–6.5 ppm are seen after
71 acrylation, and these peaks correspond to the three acrylate group protons in AELO^{2,3}. Proton
72 peaks corresponding to the epoxy ring observed in the AELO spectrum with reduced
73 intensity suggest that complete acrylation is impossible. This might be due to the steric ratio
74 between the area of the epoxide peaks (^1H NMR) peak at 2.85-3.2 ppm and the area of the
75 unsaturation bond peak, which appeared at 5.27–5.45 ppm, is used to calculate the degree of
76 epoxidation, which was calculated to be 82 %. In a similar method, the degree of acrylation
77 was calculated by dividing the area of the acrylate group by the sum of the areas of epoxy and
78 acrylate, and it was calculated to be 56 %⁴.



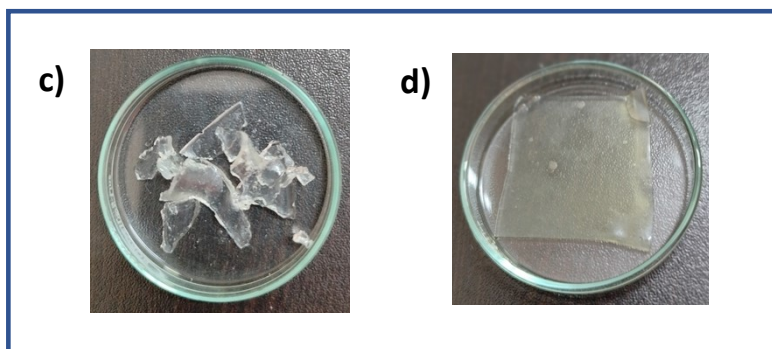
79

80 Fig. S1. a) ATR-FTIR comparison analysis and b) ^1H NMR analysis of LO., ELO and
81 AELO

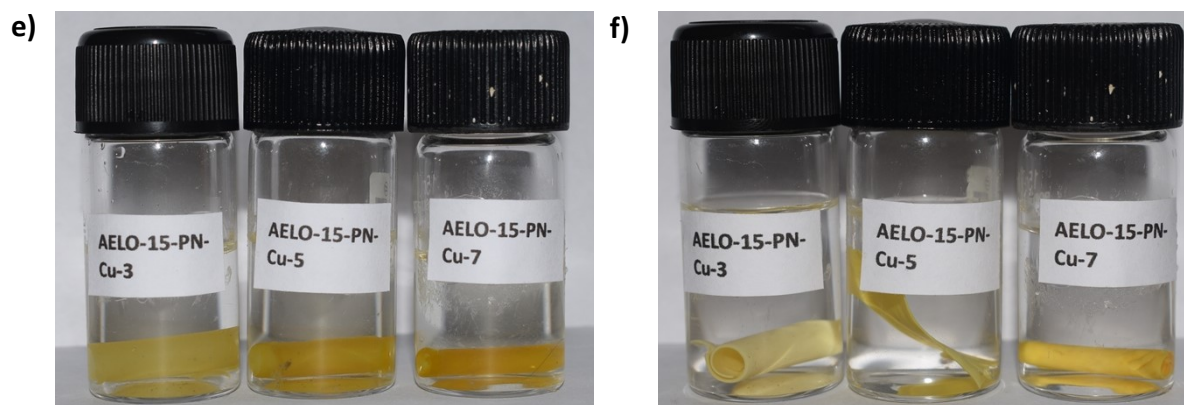
82 Swelling studies



83



84



85

86 **Fig. S2. (a) Images of the swelled samples at 27 °C and (b) at 40 °C (c) image of the PN.**
87 **after equilibrium swelling and (d) image of the AELO incorporated sample after**
88 **equilibrium swelling, (e) images of curcumin incorporated films at 27 °C and (f) at 40**
89 **°C**

90

91 **Table S1. Equilibrium swelling capacity of hydrogel films.**

Sample	Equilibrium swelling at 27 °C (g/g)
PN	9.87±0.114
AELO-5-PN	4.50±0.089
AELO-10-PN	3.03±0.121
AELO-15-PN	2.28±0.091
AELO-20-PN	1.36±0.098
AELO-15-PN-Cu-3	1.02±0.009
AELO-15-PN-Cu-5	0.82±0.009
AELO-15-PN-Cu-7	0.72±0.011

92

93

94

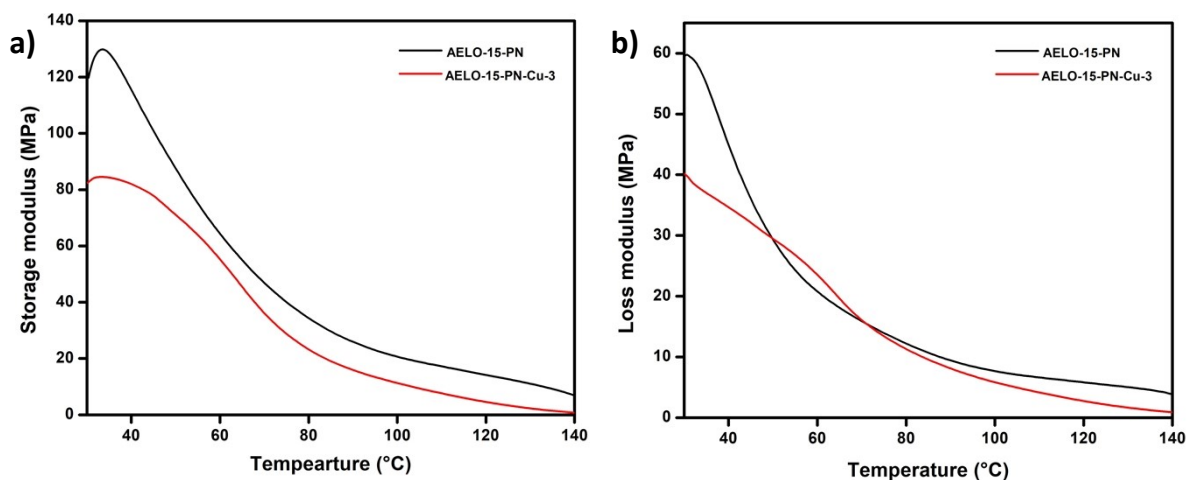
95

96

97

98

99 **DMA Analysis**



100

101 **Fig. S3. Comparison of (a) storage and (b) loss modulus of AELO-15-PN and AELO-15-**
 102 **PN-Cu-3**

103

104 **Colour changes with different pH**

105 **Table S2: Colour parameter analysis**

	Dry sample			pH 4			pH 7.4			pH 9.2		
	L	a	b	L	a	b	L	a	b	L	a	b
AELO-15-PN-Cu-3	81.82	-	44.79	82.68	-4.29	39.43	80.6	-2.23	36.79	77.46	2.4	40.53
		6.69										
AELO-15-PN-Cu-5	80.57	-5	64.52	80.81	-3.43	62.04	73.92	1.33	61.01	66.98	12.32	61.02
AELO-15-PN-Cu-3	74.44	6.4	74.81	73.9	4.05	73.63	66.34	14.14	58.73	57.37	24.58	53.82

106

107

108 **References**

109 1 R. Paul, B. John and S. K. Sahoo, *Biomacromolecules*, 2022, **23**, 816–828.

110 2 Y. Su, S. Zhang, Y. Chen, T. Yuan and Z. Yang, *Prog. Org. Coatings*, 2020, **148**,
111 105820.

112 3 S. K. Sahoo, V. Khandelwal and G. Manik, *Int. J. Adhes. Adhes.*, 2019, **89**, 174–177.

113 4 M. Jebrane, S. Cai, C. Sandström and N. Terziev, *Express Polym. Lett.*, 2017, **11**, 383–
114 395.

115