

Development of chiral modular bifunctional C_2 -symmetric bipyridine/phenanthroline-bipyrroloimidazolone ligands and application in noncovalent interaction-assisted enantioselective catalysis

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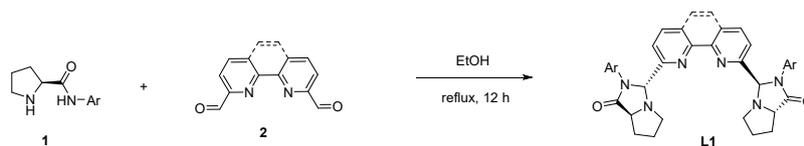
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1. General information

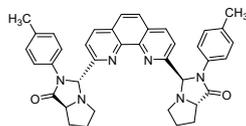
Reactions were monitored by thin layer chromatography using UV light to visualize the course of reaction. Purification of reaction products was carried out by flash chromatography. ^1H and ^{13}C NMR spectra were obtained using a Bruker DPX-400 spectrometer. ^1H NMR chemical shifts are reported in ppm (δ) relative to tetramethylsilane (TMS) with the solvent resonance employed as the internal standard. Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet), coupling constants (Hz) and integration. ^{13}C NMR chemical shifts are reported in ppm (δ) from tetramethylsilane (TMS) with the solvent resonance as the internal standard. Melting points were measured on an electrothermal digital melting point apparatus.

2. General procedure for preparation of chiral Bpy/Phen-BPI ligands L1



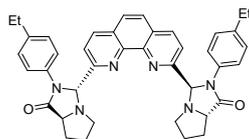
In a sealed tube equipped with a magnetic stirring bar, optically pure prolinamide **1** (2.4 mmol, 2.4 equiv) and bipyridine/phenanthroline-dicarbaldehyde **2** (1.0 mmol) were added. Then, ethanol (10.0 mL) was added and the reaction was heated with stirring at reflux for 12 h. After completion of the reaction, as indicated by TLC, the aftertreatment residue was purified by flash column chromatography to give the Bpy/Phen-BPI ligands **L1**.

3. Characterization data of ligands L

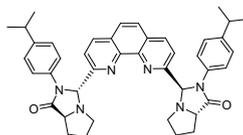


L1a: Yellow solid, yield 75%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.84-1.89 (m, 4H), 2.13 (s, 6H), 2.17-2.22 (m, 4H), 3.06-3.12 (m, 2H), 3.47-3.53 (m, 2H), 4.20-4.24 (m, 2H), 6.13 (s, 2H), 6.96 (d, $J = 8.4$ Hz, 4H), 7.45-7.49 (m, 6H), 7.62 (s, 2H), 8.09 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 19.7, 23.8, 27.0, 55.4, 63.9, 83.8, 118.5, 120.3, 125.4, 127.4, 128.4, 133.7, 133.8, 136.4, 144.6, 157.7, 173.8; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{38}\text{H}_{36}\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$:

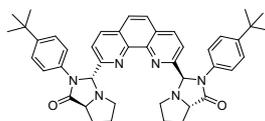
631.2792; Found: 631.2783.



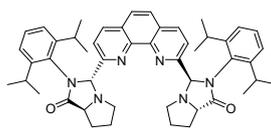
L1b: Yellow solid, yield 73%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.02-1.05 (m, 6H), 1.83-1.89 (m, 4H), 2.16-2.22 (m, 4H), 2.40-2.46 (m, 4H), 3.04-3.10 (m, 2H), 3.47-3.52 (m, 2H), 4.19-4.23 (m, 2H), 6.14 (s, 2H), 6.99 (d, $J = 8.8$ Hz, 4H), 7.47-7.50 (m, 6H), 7.60 (s, 2H), 8.08 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 15.4, 24.9, 28.1, 28.2, 56.5, 65.0, 85.0, 119.6, 121.5, 126.5, 128.4, 128.5, 135.0, 137.6, 141.3, 145.7, 158.9, 175.0; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{40}\text{H}_{40}\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 659.3105; Found: 659.3099.



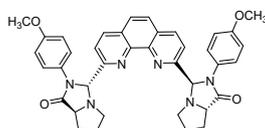
L1c: Yellow solid, yield 73%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.12-1.14 (m, 12H), 1.90-1.96 (m, 4H), 2.24-2.30 (m, 4H), 2.74-2.81 (m, 2H), 3.12-3.18 (m, 2H), 3.55-3.60 (m, 2H), 4.28-4.32 (m, 2H), 6.22 (s, 2H), 7.11 (d, $J = 8.4$ Hz, 4H), 7.55-7.60 (m, 6H), 7.65 (s, 2H), 8.14 (d, $J = 8.0$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 23.9, 24.9, 28.1, 33.5, 56.5, 65.0, 85.0, 119.6, 121.4, 126.5, 127.0, 128.5, 135.1, 137.6, 145.7, 145.8, 158.9, 175.0; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{42}\text{H}_{44}\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 687.3418; Found: 687.3423.



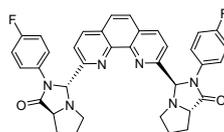
L1d: Yellow solid, yield 72%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.13 (s, 18H), 1.84-1.90 (m, 4H), 2.17-2.23 (m, 4H), 3.06-3.12 (m, 2H), 3.51-3.57 (m, 2H), 4.18-4.21 (m, 2H), 6.17 (s, 2H), 7.18-7.21 (m, 4H), 7.47-7.52 (m, 6H), 7.61 (s, 2H), 8.09 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 23.9, 27.1, 30.2, 33.3, 55.5, 63.9, 83.8, 118.5, 119.8, 124.9, 125.5, 127.5, 133.8, 136.6, 144.7, 147.0, 157.9, 174.0; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{44}\text{H}_{48}\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 715.3731; Found: 715.3722.



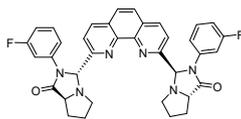
L1e: Yellow solid, yield 67%, >20:1 dr; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ : 0.83 (s, 3H), 0.85 (s, 3H), 1.16-1.19 (m, 12H), 1.48 (s, 3H), 1.49 (s, 3H), 1.89-1.94 (m, 4H), 2.18-2.26 (m, 4H), 2.28-2.36 (m, 2H), 3.08-3.14 (m, 4H), 3.39-3.44 (m, 2H), 4.49-4.53 (m, 2H), 5.58 (s, 2H), 6.74-6.76 (m, 2H), 7.16-7.19 (m, 4H), 7.38 (d, $J = 8.4$ Hz, 2H), 7.63 (s, 2H), 8.03 (d, $J = 8.4$ Hz, 2H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ : 22.4, 23.3, 24.9, 25.3, 25.6, 28.4, 28.9, 29.0, 57.3, 65.2, 88.0, 122.2, 123.8, 124.2, 126.5, 128.5, 129.3, 129.8, 136.5, 145.5, 146.4, 148.1, 158.6, 175.1; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{48}\text{H}_{56}\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 771.4357; Found: 771.4348.



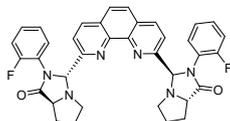
L1f: Yellow solid, yield 75%, >20:1 dr; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ : 1.84-1.88 (m, 4H), 2.17-2.22 (m, 4H), 3.06-3.12 (m, 2H), 3.46-3.51 (m, 2H), 3.59 (s, 6H), 4.22-4.25 (m, 2H), 6.08 (s, 2H), 6.68 (d, $J = 9.2$ Hz, 4H), 7.42-7.49 (m, 6H), 7.64 (s, 2H), 8.11 (d, $J = 8.4$ Hz, 2H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ : 22.9, 26.2, 53.3, 54.5, 63.0, 83.4, 112.2, 117.8, 121.6, 124.5, 126.5, 128.2, 135.6, 143.6, 155.1, 156.9, 172.7; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{38}\text{H}_{36}\text{N}_6\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 663.2690; Found: 663.2681.



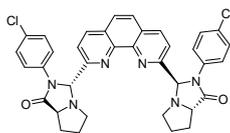
L1g: Yellow solid, yield 75%, >20:1 dr; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ : 1.85-1.91 (m, 4H), 2.17-2.24 (m, 4H), 3.03-3.09 (m, 2H), 3.45-3.50 (m, 2H), 4.25-4.29 (m, 2H), 6.07 (s, 2H), 6.82-6.86 (m, 4H), 7.48-7.54 (m, 6H), 7.65 (s, 2H), 8.13 (d, $J = 8.4$ Hz, 2H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ : 23.9, 27.1, 55.5, 63.9, 84.2, 114.7 (d, $J_{CF} = 22.3$ Hz), 118.8, 122.6 (d, $J_{CF} = 7.3$ Hz), 125.6, 127.6, 132.3 (d, $J_{CF} = 3.2$ Hz), 136.6, 144.6, 157.5, 159.8 (d, $J_{CF} = 244.2$ Hz), 174.0; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{F}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 639.2291; Found: 639.2287.



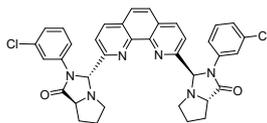
L1h: Yellow solid, yield 70%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.82-1.89 (m, 4H), 2.17-2.22 (m, 4H), 3.02-3.08 (m, 2H), 3.49-3.54 (m, 2H), 4.19-4.22 (m, 2H), 6.14 (s, 2H), 6.65-6.70 (m, 2H), 7.05-7.11 (m, 2H), 7.19-7.21 (m, 2H), 7.47 (d, $J = 8.0$ Hz, 2H), 7.64 (s, 2H), 7.69-7.73 (m, 2H), 8.13 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 22.9, 26.0, 54.5, 62.9, 82.6, 106.7 (d, $J_{CF} = 26.3$ Hz), 109.8 (d, $J_{CF} = 22.2$ Hz), 114.2 (d, $J_{CF} = 3.4$ Hz), 117.6, 124.6, 126.6, 128.1 (d, $J_{CF} = 10.2$ Hz), 135.7, 137.1, 137.2, 143.7, 156.3, 161.8 (d, $J_{CF} = 244.1$ Hz), 173.4; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{F}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 639.2291; Found: 639.2297.



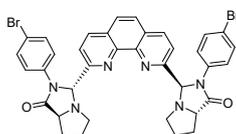
L1i: Yellow solid, yield 70%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.83-1.96 (m, 4H), 2.16-2.30 (m, 4H), 3.14-3.20 (m, 2H), 3.41-3.46 (m, 2H), 4.39-4.42 (m, 2H), 5.96 (s, 2H), 6.82-6.86 (m, 2H), 6.96-7.07 (m, 4H), 7.30-7.34 (m, 2H), 7.56-7.61 (m, 4H), 8.09 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 25.0, 28.1, 56.5, 64.3, 85.6, 116.4 (d, $J_{CF} = 20.3$ Hz), 120.5, 124.0 (d, $J_{CF} = 11.2$ Hz), 124.5, 126.5, 128.5, 128.8 (d, $J_{CF} = 8.2$ Hz), 129.8, 137.2, 145.7, 158.5, 158.7 (d, $J_{CF} = 249.0$ Hz), 175.5; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{F}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 639.2291; Found: 639.2285.



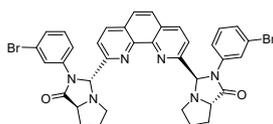
L1j: Yellow solid, yield 71%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.83-1.91 (m, 4H), 2.18-2.23 (m, 4H), 3.02-3.08 (m, 2H), 3.47-3.52 (m, 2H), 4.22-4.25 (m, 2H), 6.11 (s, 2H), 7.10-7.14 (m, 4H), 7.48 (d, $J = 8.4$ Hz, 2H), 7.54-7.58 (m, 4H), 7.66 (s, 2H), 8.13 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 23.0, 26.3, 54.7, 63.1, 82.9, 117.8, 120.5, 124.8, 126.7, 127.2, 128.4, 134.2, 135.9, 143.8, 156.5, 173.3; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{Cl}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 671.1700; Found: 671.1708.



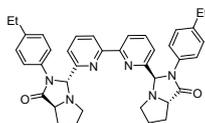
L1k: Yellow solid, yield 74%, >20:1 dr; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ : 1.84-1.90 (m, 4H), 2.18-2.23 (m, 4H), 3.02-3.08 (m, 2H), 3.49-3.54 (m, 2H), 4.21-4.25 (m, 2H), 6.12 (s, 2H), 6.93-6.96 (m, 2H), 7.03-7.07 (m, 2H), 7.30-7.33 (m, 2H), 7.48 (d, $J = 8.4$ Hz, 2H), 7.65 (s, 2H), 7.90-7.91 (m, 2H), 8.12 (d, $J = 8.0$ Hz, 2H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ : 23.9, 27.0, 55.5, 63.9, 83.6, 117.9, 118.6, 120.3, 124.1, 125.6, 127.5, 128.9, 133.6, 136.6, 137.7, 144.7, 157.2, 174.4; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{Cl}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 671.1700; Found: 671.1709.



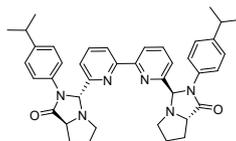
L1l: Yellow solid, yield 73%, >20:1 dr; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ : 1.86-1.92 (m, 4H), 2.18-2.24 (m, 4H), 3.03-3.09 (m, 2H), 3.48-3.53 (m, 2H), 4.22-4.26 (m, 2H), 6.11 (s, 2H), 7.25-7.29 (m, 4H), 7.48-7.54 (m, 6H), 7.66 (s, 2H), 8.14 (d, $J = 8.4$ Hz, 2H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ : 24.9, 28.2, 56.6, 65.0, 84.7, 118.1, 119.7, 122.7, 126.7, 128.6, 132.0, 136.6, 137.7, 145.7, 158.3, 175.2; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{Br}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 759.0689; Found: 759.0694.



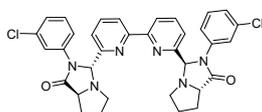
L1m: Yellow solid, yield 70%, >20:1 dr; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ : 1.85-1.91 (m, 4H), 2.18-2.23 (m, 4H), 3.02-3.08 (m, 2H), 3.48-3.53 (m, 2H), 4.23-4.26 (m, 2H), 6.11 (s, 2H), 6.97-7.01 (m, 2H), 7.09-7.12 (m, 2H), 7.34-7.37 (m, 2H), 7.48 (d, $J = 8.4$ Hz, 2H), 7.65 (s, 2H), 8.05-8.06 (m, 2H), 8.13 (d, $J = 8.4$ Hz, 2H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ : 23.2, 26.4, 54.8, 63.2, 82.9, 117.7, 118.0, 121.0, 122.4, 124.9, 126.3, 126.9, 128.5, 136.0, 137.2, 144.1, 156.5, 173.7; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{36}\text{H}_{30}\text{Br}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 759.0689; Found: 759.0695.



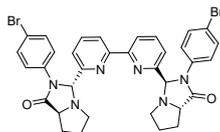
L1n: Light yellow solid, yield 71%, >20:1 dr; ¹H NMR (CDCl₃, 400 MHz) δ: 1.05-1.09 (m, 6H), 1.81-1.87 (m, 4H), 2.13-2.19 (m, 4H), 2.44-2.49 (m, 4H), 2.87-2.93 (m, 2H), 3.39-3.44 (m, 2H), 4.18-4.21 (m, 2H), 5.66 (s, 2H), 7.00 (d, *J* = 8.8 Hz, 4H), 7.19-7.27 (m, 6H), 7.67-7.71 (m, 2H), 8.20-8.22 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ: 15.5, 24.9, 27.7, 28.3, 56.4, 64.8, 84.6, 120.9, 121.0, 121.8, 128.4, 128.9, 135.1, 138.3, 141.4, 155.7, 157.2, 175.3; HRMS (ESI-TOF) *m/z*: Calcd. for C₃₈H₄₀N₆NaO₂ [M+Na]⁺: 635.3105; Found: 635.3112.



L1o: Light yellow solid, yield 70%, >20:1 dr; ¹H NMR (CDCl₃, 400 MHz) δ: 1.07 (s, 6H), 1.08 (s, 6H), 1.80-1.86 (m, 4H), 2.12-2.18 (m, 4H), 2.69-2.76 (m, 2H), 2.86-2.92 (m, 2H), 3.38-3.43 (m, 2H), 4.17-4.21 (m, 2H), 5.66 (s, 2H), 7.02 (d, *J* = 8.4 Hz, 4H), 7.20-7.28 (m, 6H), 7.67-7.71 (m, 2H), 8.20 (d, *J* = 7.6 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ: 23.9, 24.9, 27.7, 33.6, 56.4, 64.8, 84.6, 120.9, 121.0, 121.8, 127.0, 135.2, 138.3, 146.0, 155.7, 157.3, 175.3; HRMS (ESI-TOF) *m/z*: Calcd. for C₄₀H₄₄N₆NaO₂ [M+Na]⁺: 663.3418; Found: 663.3405.

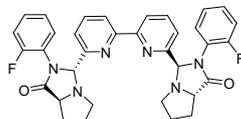


L1p: Light yellow solid, yield 67%, >20:1 dr; ¹H NMR (CDCl₃, 400 MHz) δ: 1.81-1.86 (m, 4H), 2.13-2.19 (m, 4H), 2.84-2.90 (m, 2H), 3.39-3.44 (m, 2H), 4.14-4.17 (m, 2H), 5.71 (s, 2H), 6.95-6.98 (m, 2H), 7.05-7.09 (m, 2H), 7.19-7.27 (m, 4H), 7.61-7.62 (m, 2H), 7.71-7.75 (m, 2H), 8.16 (d, *J* = 7.6 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ: 23.9, 26.7, 55.3, 63.8, 83.0, 117.9, 119.9, 120.1, 120.3, 124.0, 128.9, 133.6, 137.5, 137.9, 154.5, 155.6, 174.6; HRMS (ESI-TOF) *m/z*: Calcd. for C₃₄H₃₀Cl₂N₆NaO₂ [M+Na]⁺: 647.1700; Found: 647.1691.

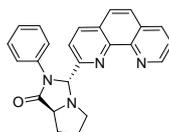


L1q: Light yellow solid, yield 68%, >20:1 dr; ¹H NMR (CDCl₃, 400 MHz) δ: 1.81-1.85 (m, 4H), 2.12-2.18 (m, 4H), 2.85-2.91 (m, 2H), 3.39-3.44 (m, 2H), 4.13-4.16 (m, 2H), 5.68 (s, 2H), 7.19-

7.32 (m, 10H), 7.69-7.73 (m, 2H), 8.12 (d, $J = 8.0$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 23.0, 25.8, 54.4, 62.9, 82.2, 116.2, 119.0, 119.2, 120.9, 130.1, 134.9, 136.6, 153.6, 154.8, 173.6; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{34}\text{H}_{30}\text{Br}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 735.0689; Found: 735.0685.

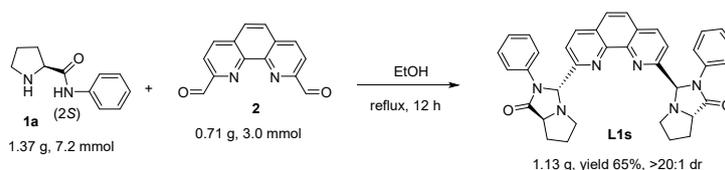


L1r: Light yellow solid, yield 63%, >20:1 dr; ^1H NMR (CDCl_3 , 400 MHz) δ : 1.75-1.81 (m, 2H), 1.87-1.92 (m, 2H), 2.01-2.08 (m, 2H), 2.14-2.20 (m, 2H), 3.02-3.08 (m, 2H), 3.34-3.39 (m, 2H), 4.27-4.30 (m, 2H), 5.79 (s, 2H), 6.91-6.95 (m, 2H), 7.03-7.08 (m, 4H), 7.13-7.18 (m, 2H), 7.34-7.36 (m, 2H), 7.73-7.77 (m, 2H), 8.07-8.09 (m, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ : 24.4, 27.4, 55.9, 64.3, 84.7, 116.0 (d, $J_{\text{CF}} = 20.3$ Hz), 120.4, 122.0, 123.7 (d, $J_{\text{CF}} = 12.0$ Hz), 124.3, 128.9, 129.2 (d, $J_{\text{CF}} = 8.1$ Hz), 138.2, 155.3, 156.9, 157.8 (d, $J_{\text{CF}} = 257.0$ Hz), 176.1; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{34}\text{H}_{30}\text{F}_2\text{N}_6\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 615.2292; Found: 615.2299.



L5a: Light yellow solid, yield 65%, >20:1 dr; ^1H NMR (CD_3OD , 400 MHz) δ : 1.72-1.85 (m, 2H), 2.07-2.14 (m, 2H), 3.10-3.16 (m, 1H), 3.27-3.33 (m, 1H), 4.19-4.23 (m, 1H), 6.24 (s, 1H), 6.88-6.92 (m, 1H), 7.09-7.13 (m, 2H), 7.50-7.59 (m, 6H), 8.11-8.16 (m, 2H), 8.94-8.95 (m, 1H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 24.3, 28.2, 56.2, 65.0, 85.4, 119.8, 122.2, 123.3, 125.6, 126.0, 126.7, 128.3, 128.6, 129.1, 136.6, 136.7, 137.8, 144.5, 145.0, 149.6, 158.8, 174.8; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{24}\text{H}_{20}\text{N}_4\text{NaO}$ $[\text{M}+\text{Na}]^+$: 403.1529; Found: 403.1534.

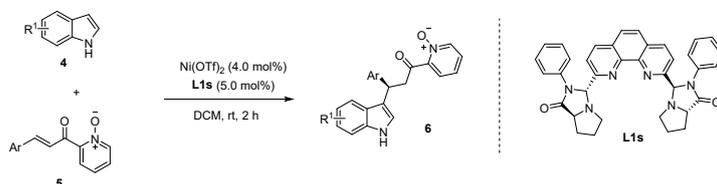
4. The gram scale synthesis of the Phen-BPI ligand L1s



In a sealed tube equipped with a magnetic stirring bar, phenanthroline-dicarbaldehyde **2** (0.71 g, 3.0 mmol) and optically pure prolinamide **1a** (1.37 g, 7.2 mmol) were added. Then, anhydrous ethanol (30.0 mL) was added and the reaction was heated with stirring at reflux for 12 h. After

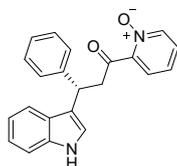
completion of the reaction, as indicated by TLC, the aftertreatment residue was purified by flash column chromatography to give the Phen-BPI ligand **L1s** (1.13 g, yield 65%, >20:1 dr).

5. Catalytic asymmetric synthesis of compounds **6**

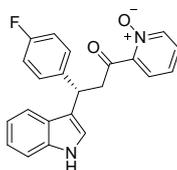


In a sealed tube equipped with a magnetic stirring bar, to the mixture of Ni(OTf)₂ (4.0 mol %), **L1s** (5.0 mol %) in 1.5 mL of DCM was added **4** (0.30 mmol), and **5** (0.20 mmol). The reaction mixture was stirred at room temperature for 2 h and was directly loaded onto a silica gel and purified by flash chromatography to give the desired product **6**, using hexane/EtOAc (8/1, v/v) as the eluent.

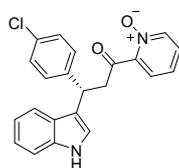
6. Characterization data of compounds **6**



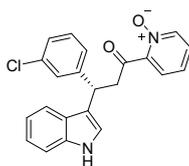
6aa: Product in accordance with literature characterization data⁸. 91%, 95% ee, $[\alpha]_D^{20} = -16.4$ (*c* 0.50, CHCl₃). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 15.65$ min; $\tau_{minor} = 19.01$ min). ¹H NMR (CDCl₃, 400 MHz) δ : 3.85-3.91 (m, 1H), 3.97-4.03 (m, 1H), 4.83-4.87 (m, 1H), 6.87-6.91 (m, 1H), 6.95-7.06 (m, 5H), 7.10-7.14 (m, 3H), 7.17-7.23 (m, 3H), 7.34 (d, *J* = 8.0 Hz, 1H), 8.04 (d, *J* = 6.8 Hz, 1H), 8.20 (br s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ : 37.5, 48.0, 110.1, 117.6, 118.3, 118.4, 120.6, 121.0, 124.7, 125.3, 125.4, 125.6, 126.5, 126.9, 127.3, 135.5, 139.0, 142.8, 146.0, 196.1.



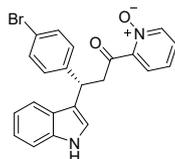
6ab: Product in accordance with literature characterization data⁸. 92%, 93% ee, $[\alpha]_{\text{D}}^{20} = -8.2$ (*c* 0.50, CHCl₃). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 12.66$ min; $\tau_{\text{minor}} = 14.84$ min). ¹H NMR (DMSO-*d*₆, 400 MHz) δ : 3.79-3.85 (m, 1H), 3.99-4.05 (m, 1H), 4.81-4.85 (m, 1H), 6.87-6.91 (m, 1H), 7.02-7.06 (m, 3H), 7.22-7.24 (m, 1H), 7.30-7.38 (m, 6H), 7.49-7.53 (m, 1H), 8.32 (d, $J = 6.4$ Hz, 1H), 10.91 (br s, 1H); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ : 37.4, 48.6, 111.9, 115.3 (d, $J_{\text{CF}} = 21.2$ Hz), 117.7, 118.9 (d, $J_{\text{CF}} = 23.3$ Hz), 121.6, 122.6, 126.3, 126.5 (d, $J_{\text{CF}} = 8.2$ Hz), 126.7, 128.9, 129.8 (d, $J_{\text{CF}} = 8.1$ Hz), 136.9, 140.6, 141.2, 141.3, 146.8, 161.2 (d, $J_{\text{CF}} = 240.3$ Hz), 197.6.



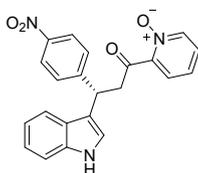
6ac: Product in accordance with literature characterization data⁸. 93%, 99% ee, $[\alpha]_{\text{D}}^{20} = -18.2$ (*c* 0.50, CHCl₃). The ee was determined by HPLC analysis using a Chiralpak IA column (70/30 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 14.69$ min; $\tau_{\text{minor}} = 13.44$ min). ¹H NMR (DMSO-*d*₆, 400 MHz) δ : 3.79-3.85 (m, 1H), 3.98-4.04 (m, 1H), 4.80-4.83 (m, 1H), 6.87-6.90 (m, 1H), 7.01-7.05 (m, 1H), 7.26-7.36 (m, 9H), 7.46-7.52 (m, 1H), 8.32 (d, $J = 6.0$ Hz, 1H), 10.91 (br s, 1H); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ : 37.5, 48.3, 111.9, 117.3, 118.9, 119.0, 121.6, 122.7, 126.3, 126.4, 126.6, 128.6, 128.9, 130.0, 131.0, 136.9, 140.6, 144.1, 146.7, 197.5.



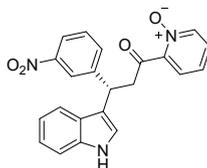
6ad: Product in accordance with literature characterization data⁸. 91%, 90% ee, $[\alpha]_{\text{D}}^{20} = -1.3$ (*c* 0.50, CHCl₃). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 11.24$ min; $\tau_{\text{minor}} = 15.82$ min). ¹H NMR (CDCl₃, 400 MHz) δ : 3.84-3.91 (m, 1H), 3.97-4.06 (m, 1H), 4.84-4.88 (m, 1H), 6.90-6.94 (m, 1H), 7.02-7.09 (m, 5H), 7.14-7.23 (m, 5H), 7.32 (d, $J = 8.0$ Hz, 1H), 8.07 (d, $J = 6.4$ Hz, 1H), 8.10 (br s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ : 37.0, 47.9, 110.1, 117.0, 118.2, 118.5, 120.6, 121.2, 124.8, 125.2, 125.5, 125.6, 125.7, 126.7, 127.0, 128.6, 133.1, 135.5, 139.2, 145.1, 145.8, 195.4.



6ae: Product in accordance with literature characterization data⁸. 90%, 94% ee, $[\alpha]_{\text{D}}^{20} = -17.1$ (*c* 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 12.64$ min; $\tau_{\text{minor}} = 15.32$ min). ¹H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 3.78-3.85 (m, 1H), 3.97-4.04 (m, 1H), 4.77-4.81 (m, 1H), 6.86-6.90 (m, 1H), 7.01-7.05 (m, 1H), 7.28-7.42 (m, 9H), 7.50-7.57 (m, 1H), 8.32 (d, $J = 6.4$ Hz, 1H), 10.90 (br s, 1H); ¹³C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 37.5, 48.2, 111.9, 117.3, 118.9, 119.0, 119.4, 121.6, 122.7, 126.3, 126.4, 126.6, 128.9, 130.4, 131.5, 136.9, 140.6, 144.6, 146.7, 197.4.

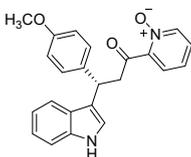


6af: Product in accordance with literature characterization data⁸. 88%, 93% ee, $[\alpha]_{\text{D}}^{20} = -20.3$ (*c* 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 22.05$ min; $\tau_{\text{minor}} = 29.97$ min). ¹H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 3.91-3.98 (m, 1H), 4.05-4.11 (m, 1H), 4.97-5.00 (m, 1H), 6.88-6.91 (m, 1H), 7.02-7.06 (m, 1H), 7.32-7.40 (m, 5H), 7.52-7.55 (m, 1H), 7.64 (d, $J = 8.4$ Hz, 2H), 8.09 (d, $J = 8.4$ Hz, 2H), 8.32 (d, $J = 6.4$ Hz, 1H), 10.99 (br s, 1H); ¹³C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 37.8, 47.9, 112.0, 116.6, 118.9, 119.0, 121.7, 123.0, 123.9, 126.4, 126.5, 126.6, 129.0, 129.4, 136.9, 140.6, 146.3, 146.6, 153.3, 197.0.

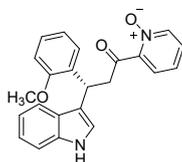


6ag: Product in accordance with literature characterization data⁸. 89%, 93% ee, $[\alpha]_{\text{D}}^{20} = +3.0$ (*c* 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 18.06$ min; $\tau_{\text{minor}} = 26.36$ min). ¹H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 3.90-3.97 (m, 1H), 3.99-4.05 (m, 1H), 4.97-5.01 (m, 1H), 6.85-

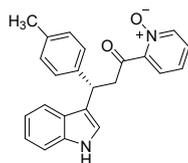
6.89 (m, 1H), 7.00-7.03 (m, 1H), 7.28-7.34 (m, 3H), 7.37-7.39 (m, 2H), 7.48-7.52 (m, 2H), 7.84 (d, $J = 7.6$ Hz, 1H), 7.97-7.99 (m, 1H), 8.16 (s, 1H), 8.29 (d, $J = 6.4$ Hz, 1H), 10.96 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 37.6, 48.0, 112.0, 116.8, 118.9, 119.0, 121.6, 121.7, 122.7, 123.0, 126.3, 126.4, 126.5, 129.0, 130.1, 135.1, 136.9, 140.6, 146.6, 147.5, 148.2, 197.2.



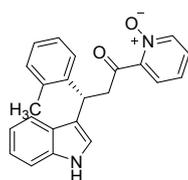
6ah: Product in accordance with literature characterization data⁸. 87%, 90% ee, $[\alpha]_{\text{D}}^{20} = -12.2$ (c 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IA column (70/30 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 16.42$ min; $\tau_{\text{minor}} = 14.71$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 3.66 (s, 3H), 3.74-3.80 (m, 1H), 3.97-4.03 (m, 1H), 4.73-4.77 (m, 1H), 6.77 (d, $J = 8.4$ Hz, 2H), 6.86-6.90 (m, 1H), 7.01-7.04 (m, 1H), 7.16-7.37 (m, 7H), 7.46-7.50 (m, 1H), 8.31 (d, $J = 6.4$ Hz, 1H), 10.86 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 37.5, 48.8, 55.4, 111.8, 114.0, 118.1, 118.7, 119.2, 121.5, 122.4, 126.2, 126.3, 126.8, 128.8, 129.0, 136.9, 137.0, 140.5, 146.9, 157.9, 197.9.



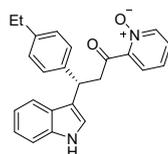
6ai: Product in accordance with literature characterization data⁸. 85%, 90% ee, $[\alpha]_{\text{D}}^{20} = -20.3$ (c 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IE column (70/30 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 14.64$ min; $\tau_{\text{minor}} = 15.77$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 3.54-3.60 (m, 1H), 3.71 (s, 3H), 3.99-4.07 (m, 1H), 5.19-5.23 (m, 1H), 6.73-6.77 (m, 1H), 6.84-6.89 (m, 2H), 6.97-7.01 (m, 1H), 7.07-7.11 (m, 3H), 7.18 (s, 1H), 7.26-7.29 (m, 2H), 7.33 (d, $J = 8.0$ Hz, 1H), 7.46-7.50 (m, 1H), 8.29 (d, $J = 6.4$ Hz, 1H), 10.79 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 30.4, 47.8, 55.9, 111.2, 111.8, 117.5, 118.7, 119.1, 120.7, 121.5, 122.9, 126.2, 126.3, 127.0, 127.7, 128.7, 128.8, 132.5, 136.8, 140.5, 146.9, 156.6, 198.0.



6aj: Product in accordance with literature characterization data⁸. 90%, 93% ee, $[\alpha]_{\text{D}}^{20} = -4.1$ (c 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IA column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 12.39$ min; $\tau_{\text{minor}} = 10.97$ min). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 2.21 (s, 3H), 3.73-3.80 (m, 1H), 3.97-4.03 (m, 1H), 4.72-4.76 (m, 1H), 6.85-6.89 (m, 1H), 7.00-7.03 (m, 3H), 7.16-7.20 (m, 3H), 7.25-7.36 (m, 4H), 7.49-7.53 (m, 1H), 8.31 (d, $J = 6.4$ Hz, 1H), 10.85 (br s, 1H); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 21.0, 37.8, 48.6, 111.8, 117.9, 118.7, 119.1, 121.5, 122.5, 126.2, 126.3, 126.7, 127.9, 128.8, 129.2, 135.4, 136.9, 140.5, 142.0, 146.8, 197.8.

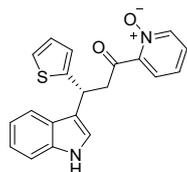


6ak: 87%, 90% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 12.05$ min; $\tau_{\text{minor}} = 15.18$ min). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 2.37 (s, 3H), 3.70-3.76 (m, 1H), 3.96-4.02 (m, 1H), 5.01-5.04 (m, 1H), 6.88-6.91 (m, 1H), 7.02-7.05 (m, 3H), 7.11-7.16 (m, 3H), 7.21-7.23 (m, 1H), 7.26-7.34 (m, 3H), 7.47-7.51 (m, 1H), 8.33 (d, $J = 6.4$ Hz, 1H), 10.87 (br s, 1H); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 19.6, 33.8, 48.3, 111.9, 117.3, 118.8, 118.9, 121.5, 123.3, 126.3, 126.4, 126.5, 126.8, 127.4, 128.9, 130.7, 135.5, 136.9, 140.5, 142.7, 146.8, 197.9; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{23}\text{H}_{20}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 379.1417; Found: 379.1414.

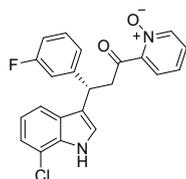


6al: 89%, 93% ee. The ee was determined by HPLC analysis using a Chiralpak IA column (70/30 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 11.86$ min; $\tau_{\text{minor}} = 10.33$ min). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 1.09-1.13 (m, 3H), 2.47-2.53 (m, 2H), 3.75-3.81 (m, 1H), 3.95-

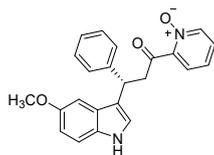
4.02 (m, 1H), 4.73-4.77 (m, 1H), 6.85-6.89 (m, 1H), 7.00-7.05 (m, 3H), 7.14-7.16 (m, 1H), 7.21 (d, $J = 8.0$ Hz, 2H), 7.26-7.31 (m, 3H), 7.35 (d, $J = 8.0$ Hz, 1H), 7.47-7.51 (m, 1H), 8.30 (d, $J = 6.4$ Hz, 1H), 10.85 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 16.0, 28.2, 37.9, 48.6, 111.8, 117.9, 118.7, 119.1, 121.5, 122.4, 126.2, 126.3, 126.8, 128.0, 128.1, 128.8, 136.9, 140.5, 141.8, 142.3, 146.9, 197.9; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{24}\text{H}_{22}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 393.1573; Found: 393.1573.



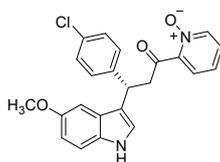
6am: Product in accordance with literature characterization data⁸. 90%, 93% ee, $[\alpha]_{\text{D}}^{20} = -19.6$ (c 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/ i -PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 18.48$ min; $\tau_{\text{minor}} = 23.41$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 3.89-3.95 (m, 1H), 4.03-4.09 (m, 1H), 5.11-5.14 (m, 1H), 6.87-7.07 (m, 4H), 7.22-7.35 (m, 5H), 7.43-7.51 (m, 2H), 8.31 (d, $J = 6.4$ Hz, 1H), 10.92 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 33.5, 49.5, 112.0, 117.6, 118.9, 119.1, 121.6, 122.7, 124.3, 124.4, 126.3, 126.5, 127.0, 129.0, 136.9, 140.6, 146.6, 149.5, 197.1.



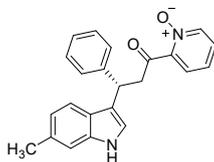
6ba: 87%, 94% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/ i -PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 11.98$ min; $\tau_{\text{minor}} = 9.26$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 3.83-3.89 (m, 1H), 4.03-4.10 (m, 1H), 4.84-4.88 (m, 1H), 6.90-6.98 (m, 2H), 7.12-7.45 (m, 8H), 7.52-7.56 (m, 1H), 8.33 (d, $J = 6.4$ Hz, 1H), 11.31 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 37.6, 48.1, 113.4 (d, $J_{\text{CF}} = 21.3$ Hz), 114.8 (d, $J_{\text{CF}} = 21.4$ Hz), 116.3, 118.2, 118.7, 120.0, 121.1, 124.2 (d, $J_{\text{CF}} = 9.1$ Hz), 126.3 (d, $J_{\text{CF}} = 4.4$ Hz), 128.7, 129.0, 130.5 (d, $J_{\text{CF}} = 8.3$ Hz), 133.6, 140.6, 146.7 (d, $J_{\text{CF}} = 7.0$ Hz), 147.9, 162.6 (d, $J_{\text{CF}} = 242.3$ Hz), 197.3; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{22}\text{H}_{16}\text{ClFN}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 417.0777; Found: 417.0782.



6ca: Product in accordance with literature characterization data⁸. 92%, 90% ee, $[\alpha]_{\text{D}}^{20} = -24.4$ (c 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 9.48$ min; $\tau_{\text{minor}} = 8.43$ min). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 3.68 (s, 3H), 3.78-3.84 (m, 1H), 4.02-4.08 (m, 1H), 4.75-4.79 (m, 1H), 6.69-6.72 (m, 1H), 6.86 (d, $J = 2.0$ Hz, 1H), 7.12-7.18 (m, 2H), 7.22-7.31 (m, 5H), 7.35 (d, $J = 7.6$ Hz, 2H), 7.47-7.51 (m, 1H), 8.32 (d, $J = 6.8$ Hz, 1H), 10.73 (br s, 1H); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 38.3, 48.5, 55.8, 101.2, 111.4, 112.5, 117.5, 123.3, 126.2, 126.3, 126.5, 127.1, 128.1, 128.6, 128.8, 132.0, 140.5, 145.0, 146.9, 153.3, 197.9.

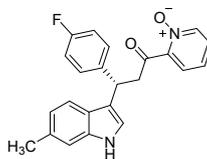


6cb: 93%, 92% ee. The ee was determined by HPLC analysis using a Chiralpak IA column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 11.31$ min; $\tau_{\text{minor}} = 9.50$ min). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 3.68 (s, 3H), 3.77-3.83 (m, 1H), 3.98-4.04 (m, 1H), 4.74-4.78 (m, 1H), 6.68-6.71 (m, 1H), 6.83 (s, 1H), 7.19-7.38 (m, 8H), 7.51-7.55 (m, 1H), 8.32 (d, $J = 6.4$ Hz, 1H), 10.74 (br s, 1H); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 37.5, 48.2, 55.8, 101.1, 111.4, 112.5, 117.1, 123.3, 126.3, 126.4, 127.0, 128.5, 128.9, 130.0, 130.9, 132.0, 140.6, 144.0, 146.7, 153.4, 197.6; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{23}\text{H}_{19}\text{ClN}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$: 429.0976; Found: 429.0971.

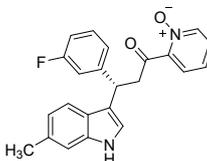


6da: 91%, 91% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{\text{major}} = 16.47$ min; $\tau_{\text{minor}} = 18.91$ min). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz) δ : 2.31 (s, 3H), 3.83-3.89 (m, 1H), 3.94-4.00 (m, 1H), 4.80-4.84 (m, 1H), 6.74 (d, $J = 8.0$ Hz, 1H), 6.91 (s, 1H), 6.99-7.07 (m, 5H), 7.12-7.20 (m, 5H), 8.05 (s, 1H), 8.07 (br s, 1H); ^{13}C NMR ($\text{DMSO-}d_6$, 100 MHz) δ : 21.8, 38.3, 48.6, 111.6, 117.6, 118.8, 120.5,

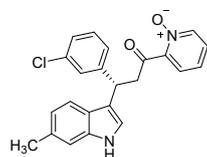
121.8, 124.7, 126.2, 126.3, 126.4, 128.1, 128.6, 128.8, 130.5, 137.3, 140.5, 145.1, 146.8, 197.8;
HRMS (ESI-TOF) m/z : Calcd. for $C_{23}H_{20}N_2NaO_2$ $[M+Na]^+$: 379.1417; Found: 379.1418.



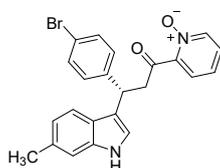
6db: 90%, 92% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 13.23$ min; $\tau_{minor} = 15.26$ min). 1H NMR (DMSO- d_6 , 400 MHz) δ : 2.33 (s, 3H), 3.77-3.83 (m, 1H), 3.95-4.01 (m, 1H), 4.76-4.80 (m, 1H), 6.71 (d, $J = 8.0$ Hz, 1H), 7.01-7.05 (m, 2H), 7.10 (s, 1H), 7.18-7.23 (m, 3H), 7.30-7.35 (m, 3H), 7.49-7.53 (m, 1H), 8.31 (d, $J = 6.4$ Hz, 1H), 10.72 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 21.8, 37.5, 48.6, 111.7, 115.3 (d, $J_{CF} = 21.3$ Hz), 117.5, 118.8, 120.6, 121.8, 124.6, 126.3, 126.4, 128.9, 129.8 (d, $J_{CF} = 8.1$ Hz), 130.6, 137.4, 140.6, 141.2 (d, $J_{CF} = 3.4$ Hz), 146.8, 161.8 (d, $J_{CF} = 240.3$ Hz), 197.7; HRMS (ESI-TOF) m/z : Calcd. for $C_{23}H_{19}FN_2NaO_2$ $[M+Na]^+$: 397.1323; Found: 397.1325.



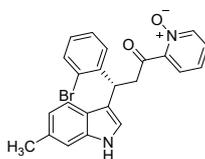
6dc: 92%, 90% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 11.24$ min; $\tau_{minor} = 15.17$ min). 1H NMR (DMSO- d_6 , 400 MHz) δ : 2.33 (s, 3H), 3.80-3.86 (m, 1H), 3.95-4.01 (m, 1H), 4.77-4.81 (m, 1H), 6.73 (d, $J = 8.0$ Hz, 1H), 6.91-6.96 (m, 1H), 7.10-7.27 (m, 7H), 7.32-7.35 (m, 1H), 7.51-7.55 (m, 1H), 8.32 (d, $J = 6.4$ Hz, 1H), 10.73 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 21.8, 37.8, 48.2, 111.7, 113.2 (d, $J_{CF} = 21.2$ Hz), 114.7 (d, $J_{CF} = 21.1$ Hz), 117.0, 118.7, 120.6, 122.0, 124.2, 124.6, 126.3, 126.4, 128.9, 130.4 (d, $J_{CF} = 8.4$ Hz), 130.6, 137.3, 140.6, 146.8, 148.2 (d, $J_{CF} = 7.3$ Hz), 162.8 (d, $J_{CF} = 242.0$ Hz), 197.5; HRMS (ESI-TOF) m/z : Calcd. for $C_{23}H_{19}FN_2NaO_2$ $[M+Na]^+$: 397.1323; Found: 397.1327.



6dd: 92%, 91% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 10.96$ min; $\tau_{minor} = 14.59$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 2.34 (s, 3H), 3.80-3.86 (m, 1H), 3.95-4.01 (m, 1H), 4.76-4.80 (m, 1H), 6.72 (d, $J = 8.0$ Hz, 1H), 7.10 (s, 1H), 7.17-7.19 (m, 1H), 7.23-7.27 (m, 4H), 7.31-7.36 (m, 3H), 7.51-7.55 (m, 1H), 8.32 (d, $J = 6.4$ Hz, 1H), 10.74 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 21.8, 37.8, 48.2, 111.7, 116.9, 118.7, 120.7, 122.0, 124.6, 126.3, 126.4, 126.5, 126.9, 127.9, 128.9, 130.5, 130.7, 133.3, 137.3, 140.6, 146.7, 147.8, 197.4; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{23}\text{H}_{19}\text{ClN}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 413.1027; Found: 413.1026.

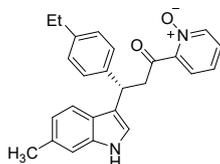


6de: 85%, 91% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 13.04$ min; $\tau_{minor} = 15.64$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 2.33 (s, 3H), 3.78-3.84 (m, 1H), 3.95-4.01 (m, 1H), 4.74-4.78 (m, 1H), 6.71 (d, $J = 8.0$ Hz, 1H), 7.10 (s, 1H), 7.19-7.22 (m, 2H), 7.25-7.33 (m, 4H), 7.39-7.41 (m, 2H), 7.50-7.54 (m, 1H), 8.31 (d, $J = 6.4$ Hz, 1H), 10.73 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 21.8, 37.6, 48.3, 111.7, 117.1, 118.7, 119.4, 120.6, 121.9, 124.6, 126.3, 126.4, 128.9, 130.4, 130.6, 131.4, 137.3, 140.6, 144.6, 146.7, 197.4; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{23}\text{H}_{19}\text{BrN}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 457.0522; Found: 457.0524.

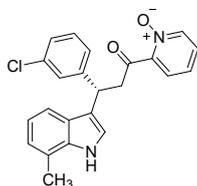


6df: 90%, 93% ee. The ee was determined by HPLC analysis using a Chiralpak IA column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 12.31$ min; $\tau_{minor} = 9.76$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 2.34 (s, 3H), 3.62-3.68 (m, 1H), 4.02-4.07 (m, 1H), 5.23-5.26 (m, 1H), 6.73 (d, $J = 8.4$ Hz, 1H), 7.06-7.15 (m, 3H), 7.20-7.37 (m, 5H), 7.52-7.59 (m, 2H), 8.33 (d, $J = 6.4$ Hz, 1H), 10.75 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 21.8, 37.2, 47.9, 111.8, 116.5, 118.7, 120.7, 122.6, 124.1, 124.7, 126.4, 126.5, 128.3, 128.6, 129.1, 129.9, 130.7, 133.0,

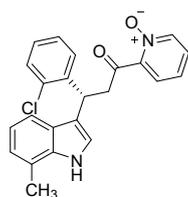
137.3, 140.7, 143.5, 146.6, 197.0; HRMS (ESI-TOF) m/z : Calcd. for $C_{23}H_{19}BrN_2NaO_2$ $[M+Na]^+$: 457.0522; Found: 457.0525.



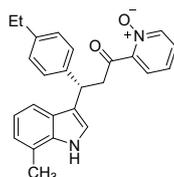
6dg: 88%, 91% ee. The ee was determined by HPLC analysis using a Chiralpak IA column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 10.01$ min; $\tau_{minor} = 8.61$ min). 1H NMR (DMSO- d_6 , 400 MHz) δ : 1.06-1.10 (m, 3H), 2.30 (s, 3H), 2.44-2.50 (m, 2H), 3.71-3.77 (m, 1H), 3.90-3.96 (m, 1H), 4.66-4.70 (m, 1H), 6.66-6.68 (m, 1H), 7.00 (d, $J = 8.0$ Hz, 2H), 7.06 (s, 1H), 7.10-7.12 (m, 2H), 7.16-7.20 (m, 3H), 7.23-7.27 (m, 1H), 7.44-7.48 (m, 1H), 8.27 (d, $J = 6.0$ Hz, 1H), 10.66 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 16.1, 21.8, 28.2, 38.0, 48.6, 111.6, 117.8, 118.9, 120.5, 121.7, 124.7, 126.2, 126.3, 128.0, 128.1, 128.8, 130.5, 137.3, 140.5, 141.7, 142.3, 146.9, 197.9; HRMS (ESI-TOF) m/z : Calcd. for $C_{25}H_{24}N_2NaO_2$ $[M+Na]^+$: 407.1730; Found: 407.1735.



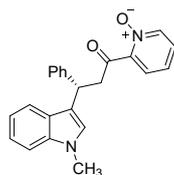
6ea: 91%, 93% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 11.73$ min; $\tau_{minor} = 17.14$ min). 1H NMR (DMSO- d_6 , 400 MHz) δ : 2.42 (s, 3H), 3.83-3.89 (m, 1H), 4.00-4.06 (m, 1H), 4.80-4.84 (m, 1H), 6.79-6.84 (m, 2H), 7.16-7.28 (m, 4H), 7.32-7.37 (m, 4H), 7.50-7.54 (m, 1H), 8.33 (d, $J = 6.4$ Hz, 1H), 10.89 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 17.2, 37.8, 48.2, 116.6, 117.5, 119.1, 121.0, 122.1, 122.5, 126.3, 126.4, 126.5, 126.6, 126.9, 128.0, 128.9, 130.5, 133.3, 136.3, 140.6, 146.7, 147.9, 197.4; HRMS (ESI-TOF) m/z : Calcd. for $C_{23}H_{19}ClN_2NaO_2$ $[M+Na]^+$: 413.1027; Found: 413.1024.



6eb: 90%, 93% ee. The ee was determined by HPLC analysis using a Chiralpak IC column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 15.55$ min; $\tau_{minor} = 18.83$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 2.42 (s, 3H), 3.68-3.74 (m, 1H), 4.09-4.15 (m, 1H), 5.29-5.33 (m, 1H), 6.80-6.84 (m, 2H), 7.16-7.21 (m, 3H), 7.26-7.30 (m, 2H), 7.33-7.41 (m, 3H), 7.52-7.56 (m, 1H), 8.34 (d, $J = 6.4$ Hz, 1H), 10.89 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 17.2, 34.3, 47.7, 116.4, 117.0, 119.2, 121.1, 122.1, 123.0, 126.3, 126.4, 126.5, 127.7, 128.3, 129.0, 129.7, 129.8, 132.9, 136.4, 140.7, 142.0, 146.6, 197.1; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{23}\text{H}_{19}\text{ClN}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 413.1027; Found: 413.1023.



6ec: 88%, 91% ee. The ee was determined by HPLC analysis using a Chiralpak IA column (60/40 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 8.53$ min; $\tau_{minor} = 6.85$ min). ^1H NMR (DMSO- d_6 , 400 MHz) δ : 1.09-1.13 (m, 3H), 2.41 (s, 3H), 2.47-2.53 (m, 2H), 3.75-3.81 (m, 1H), 3.95-4.01 (m, 1H), 4.71-4.75 (m, 1H), 6.76-6.82 (m, 2H), 7.03 (d, $J = 8.0$ Hz, 2H), 7.14-7.31 (m, 6H), 7.47-7.51 (m, 1H), 8.31 (d, $J = 6.4$ Hz, 1H), 10.81 (br s, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 16.1, 17.2, 28.2, 38.0, 48.6, 116.8, 118.4, 119.0, 120.9, 122.0, 122.1, 126.2, 126.3, 126.5, 128.0, 128.8, 136.4, 140.5, 141.7, 142.3, 146.9, 197.9; HRMS (ESI-TOF) m/z : Calcd. for $\text{C}_{25}\text{H}_{24}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 407.1730; Found: 407.1735.

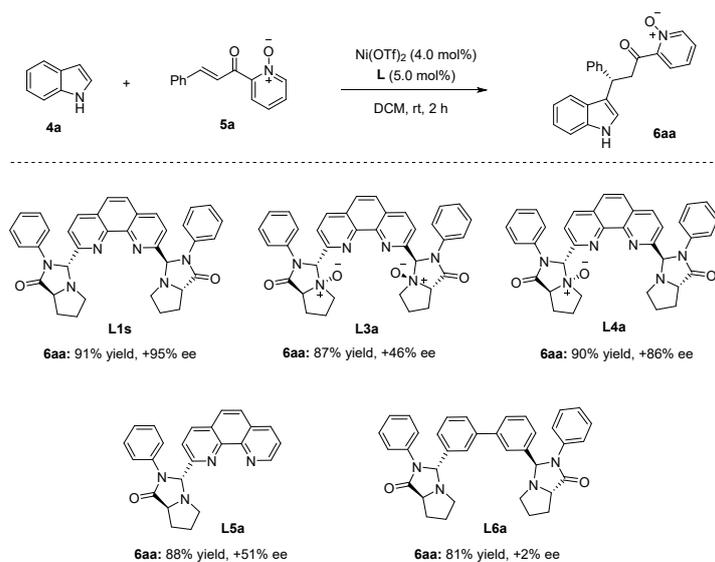


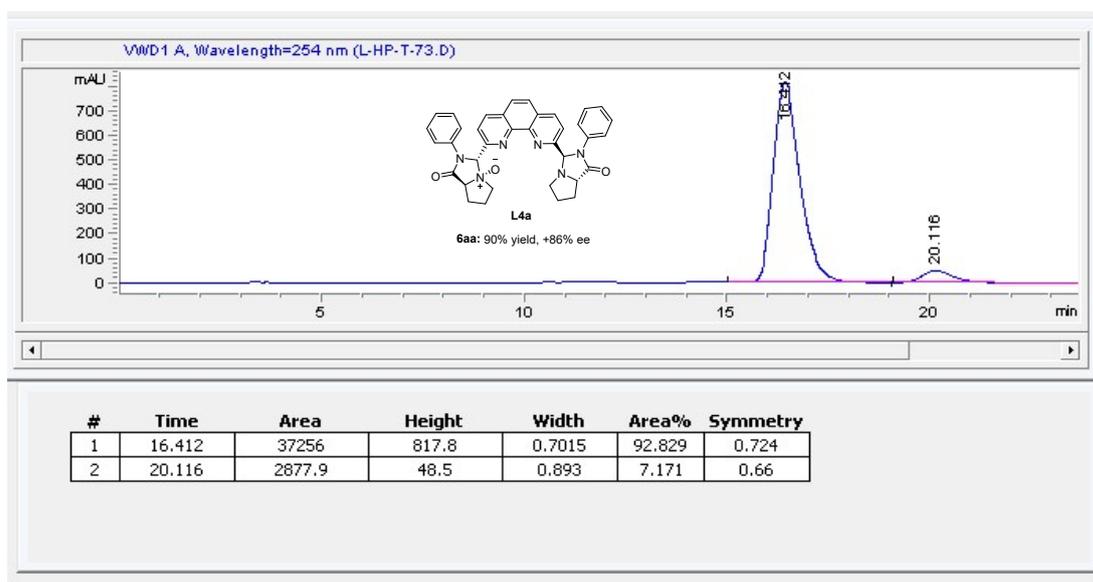
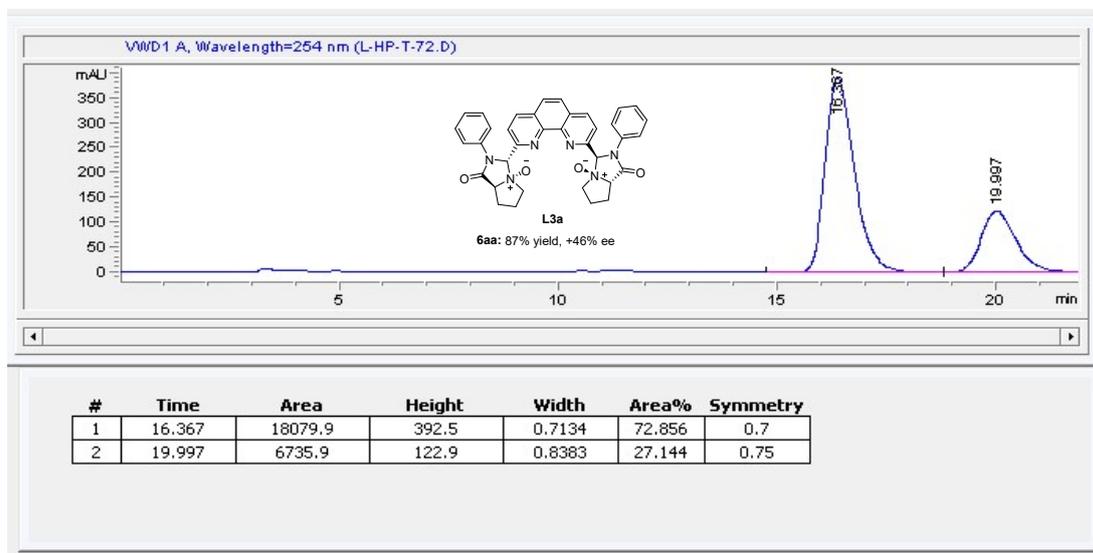
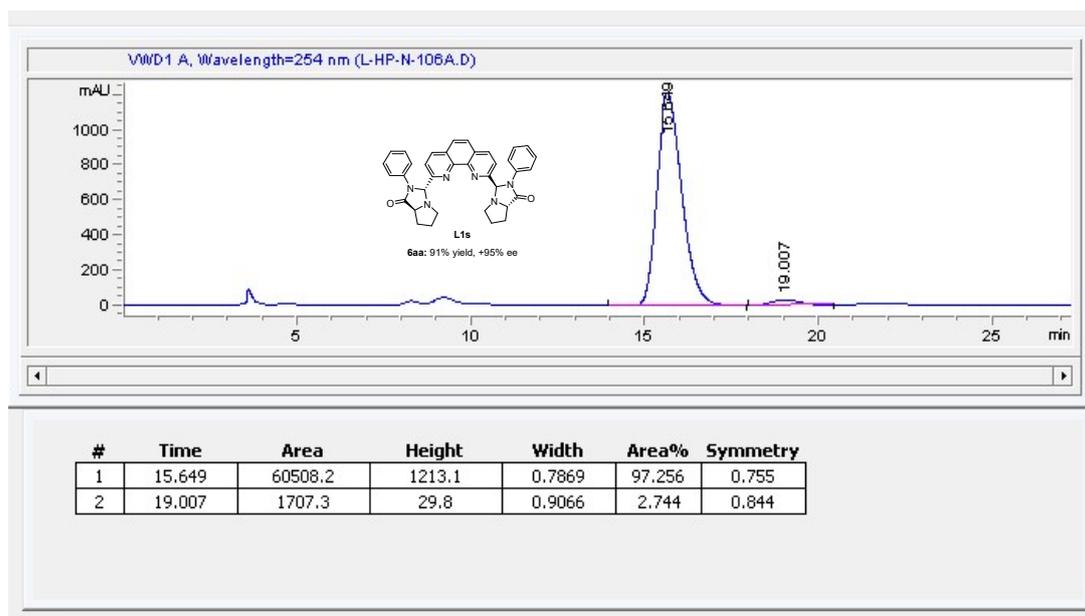
6fa: Product in accordance with literature characterization data⁸. 65% yield, 82% ee, $[\alpha]_{\text{D}}^{20} = +2.0$ (c 0.50, CHCl_3). The ee was determined by HPLC analysis using a Chiralpak IA column

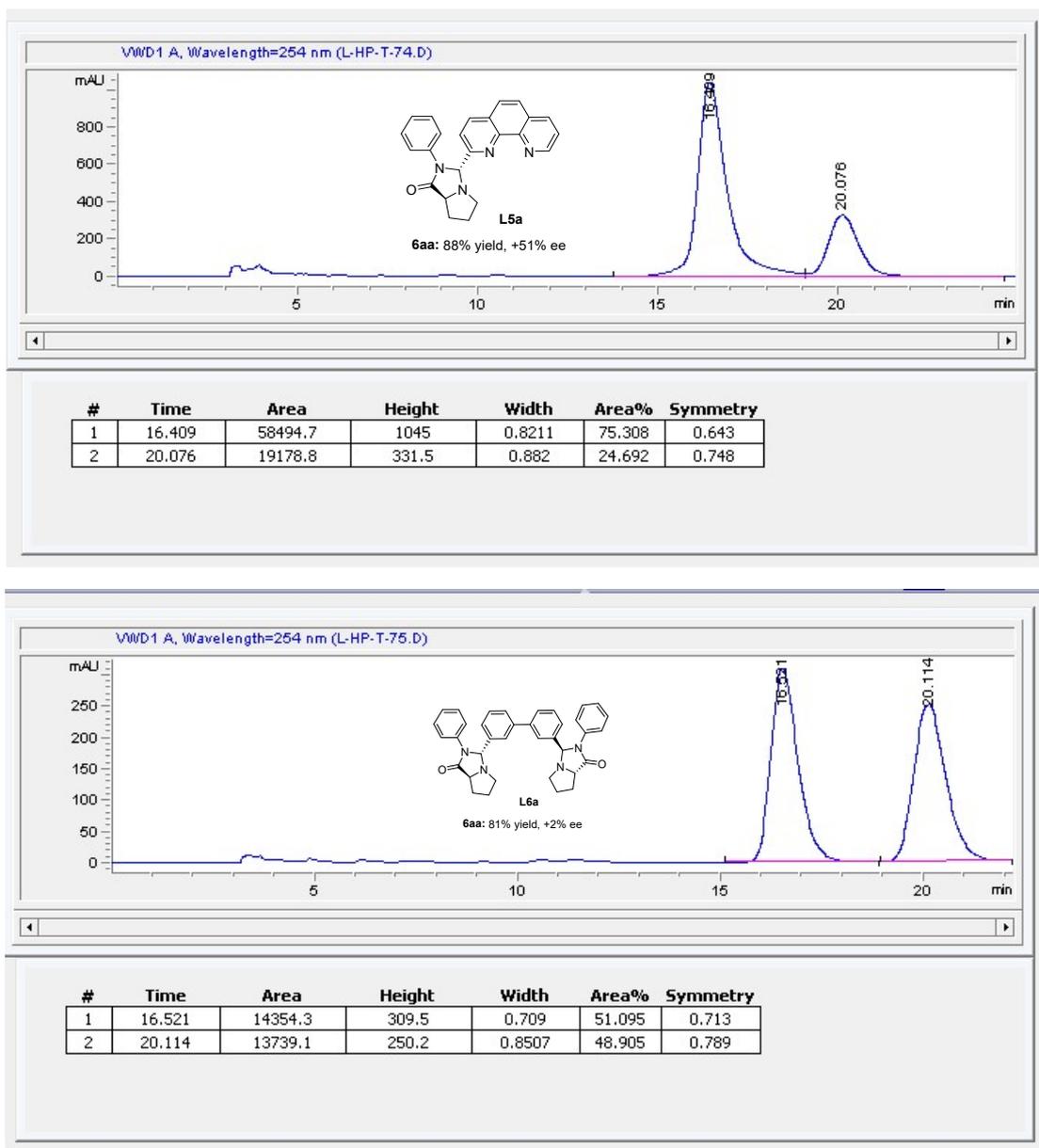
(70/30 hexane/*i*-PrOH; flow rate: 1.0 mL/min; $\lambda = 254$ nm; $\tau_{major} = 10.44$ min; $\tau_{minor} = 9.83$ min).
 ^1H NMR (DMSO- d_6 , 400 MHz) δ : 3.62 (s, 3H), 3.86-3.92 (m, 1H), 3.97-4.03 (m, 1H), 4.82-4.86 (m, 1H), 6.85 (s, 1H), 6.88-6.92 (m, 1H), 7.00-7.07 (m, 4H), 7.11-7.17 (m, 4H), 7.23-7.25 (m, 2H), 7.35 (d, $J = 7.6$ Hz, 1H), 8.06 (d, $J = 6.4$ Hz, 1H); ^{13}C NMR (DMSO- d_6 , 100 MHz) δ : 31.6, 37.5, 48.1, 108.0, 116.3, 117.8, 118.5, 120.6, 124.5, 125.2, 125.3, 125.5, 126.0, 126.5, 126.8, 127.3, 136.2, 139.1, 143.0, 146.0, 196.1.

7. Control experiments and HPLC spectra for compound 6aa

In a sealed tube equipped with a magnetic stirring bar, to the mixture of $\text{Ni}(\text{OTf})_2$ (4.0 mol %), 5.0 mol % of **L** in 1.5 mL of DCM was added **4a** (0.30 mmol), and **5a** (0.20 mmol). The reaction mixture was stirred at room temperature for 2 h and was directly loaded onto a silica gel and purified by flash chromatography to give the desired product **6aa**, using hexane/EtOAc (10/1, v/v) as the eluent.







8. References

- (a) P. K. Singh and V. K. Singh, *Org. Lett.*, 2008, **10**, 4121-4124; (b) J. George and B. V. S. Reddy, *Org. Biomol. Chem.*, 2012, **10**, 4731-4738; (c) X. Liang, Y. Gui, K. Li, J. Li, Z. Zha, L. Shi and Z. Wang, *Chem. Commun.*, 2020, **56**, 11118-11121.

9. X-ray crystal data for compounds L1p, L1r and L1a-Ni(OTf)₂·3H₂O complex

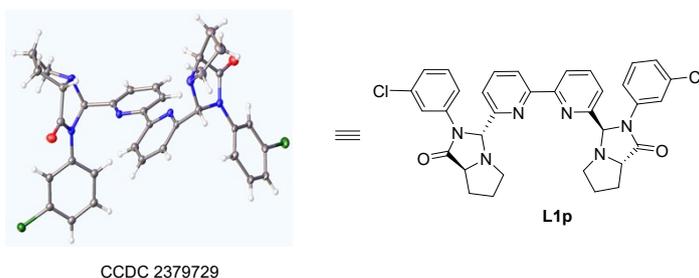


Table S1 Crystal data and structure refinement for L1p

| | |
|---|---|
| Identification code | L1p |
| Empirical formula | C ₃₄ H ₃₀ Cl ₂ N ₆ O ₂ |
| Formula weight | 625.54 |
| Temperature/K | 169.99(10) |
| Crystal system | monoclinic |
| Space group | P2 ₁ |
| a/Å, b/Å, c/Å | 13.2041(3), 8.8612(3), 14.7734(3) |
| α/°, β/°, γ/° | 90, 92.657(2), 90 |
| Volume/Å ³ | 1726.69(8) |
| Z | 2 |
| ρ _{calc} /cm ³ | 1.203 |
| μ/mm ⁻¹ | 1.993 |
| F(000) | 652.0 |
| Radiation | Cu Kα (λ = 1.54184) |
| Crystal size/mm ³ | 0.15 × 0.12 × 0.11 |
| 2θ range for data collection/° | 5.988 to 147.192 |
| Index ranges | -13 ≤ h ≤ 16, -10 ≤ k ≤ 10, -18 ≤ l ≤ 17 |
| Reflections collected | 25280 |
| Independent reflections | 6690 [R _{int} = 0.0479, R _{sigma} = 0.0370] |
| Data/restraints/parameters | 6690/451/461 |
| Goodness-of-fit on F ² | 1.057 |
| Final R indexes [I ≥ 2σ (I)] | R ₁ = 0.0696, wR ₂ = 0.1974 |
| Final R indexes [all data] | R ₁ = 0.0716, wR ₂ = 0.1991 |
| Largest diff. peak/hole / e Å ⁻³ | 0.76/-0.47 |
| Flack parameter | 0.033(12) |

Crystal Data for C₃₄H₃₀Cl₂N₆O₂ (*M* = 625.54 g/mol): monoclinic, space group P2₁ (no. 4), *a* = 13.2041(3) Å, *b* = 8.8612(3) Å, *c* = 14.7734(3) Å, β = 92.657(2)°, *V* = 1726.69(8) Å³, *Z* = 2, *T* = 169.99(10) K, μ(Cu Kα) = 1.993 mm⁻¹, *D*_{calc} = 1.203 g/cm³, 25280 reflections measured (5.988° ≤ 2θ ≤ 147.192°), 6690 unique (*R*_{int} = 0.0479, *R*_{sigma} = 0.0370) which were used in all calculations. The final *R*₁ was 0.0696 (*I* > 2σ(*I*)) and *wR*₂ was 0.1991 (all data).

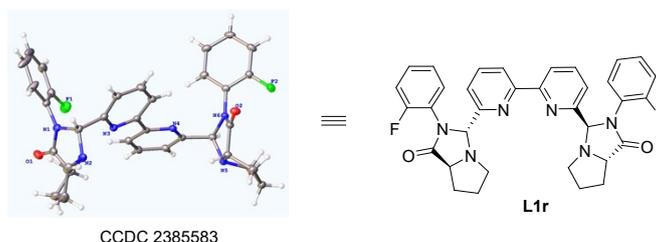
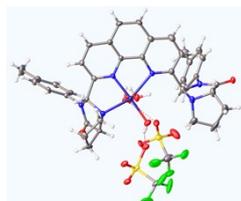


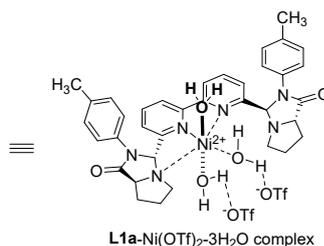
Table S2 Crystal data and structure refinement for L1r

| | |
|---|--|
| Identification code | L1r |
| Empirical formula | C ₃₄ H ₃₂ F ₂ N ₆ O ₃ |
| Formula weight | 610.65 |
| Temperature/K | 149.98(10) |
| Crystal system | orthorhombic |
| Space group | P2 ₁ 2 ₁ 2 ₁ |
| a/Å, b/Å, c/Å | 8.7651(2), 13.5600(3), 24.7099(7) |
| α/°, β/°, γ/° | 90, 90, 90 |
| Volume/Å ³ | 2936.88(14) |
| Z | 4 |
| ρ _{calc} /cm ³ | 1.381 |
| μ/mm ⁻¹ | 0.820 |
| F(000) | 1280.0 |
| Radiation | Cu Kα (λ = 1.54184) |
| Crystal size/mm ³ | 0.14 × 0.12 × 0.1 |
| 2θ range for data collection/° | 7.154 to 146.576 |
| Index ranges | -10 ≤ h ≤ 10, -14 ≤ k ≤ 16, -29 ≤ l ≤ 30 |
| Reflections collected | 24274 |
| Independent reflections | 5730 [R _{int} = 0.0405, R _{sigma} = 0.0326] |
| Data/restraints/parameters | 5730/3/412 |
| Goodness-of-fit on F ² | 1.035 |
| Final R indexes [I ≥ 2σ(I)] | R ₁ = 0.0472, wR ₂ = 0.1223 |
| Final R indexes [all data] | R ₁ = 0.0589, wR ₂ = 0.1308 |
| Largest diff. peak/hole / e Å ⁻³ | 0.46/-0.24 |
| Flack parameter | -0.09(9)/-0.05(7) |

Crystal Data for C₃₄H₃₂F₂N₆O₃ (*M* = 610.65 g/mol): orthorhombic, space group P2₁2₁2₁ (no. 19), *a* = 8.7651(2) Å, *b* = 13.5600(3) Å, *c* = 24.7099(7) Å, *V* = 2936.88(14) Å³, *Z* = 4, *T* = 149.98(10) K, μ(Cu Kα) = 0.820 mm⁻¹, *D*_{calc} = 1.381 g/cm³, 24274 reflections measured (7.154° ≤ 2θ ≤ 146.576°), 5730 unique (*R*_{int} = 0.0405, *R*_{sigma} = 0.0326) which were used in all calculations. The final *R*₁ was 0.0472 (*I* > 2σ(*I*)) and *wR*₂ was 0.1308 (all data).



CCDC: 2387992



L1a-Ni(OTf)₂-3H₂O complex

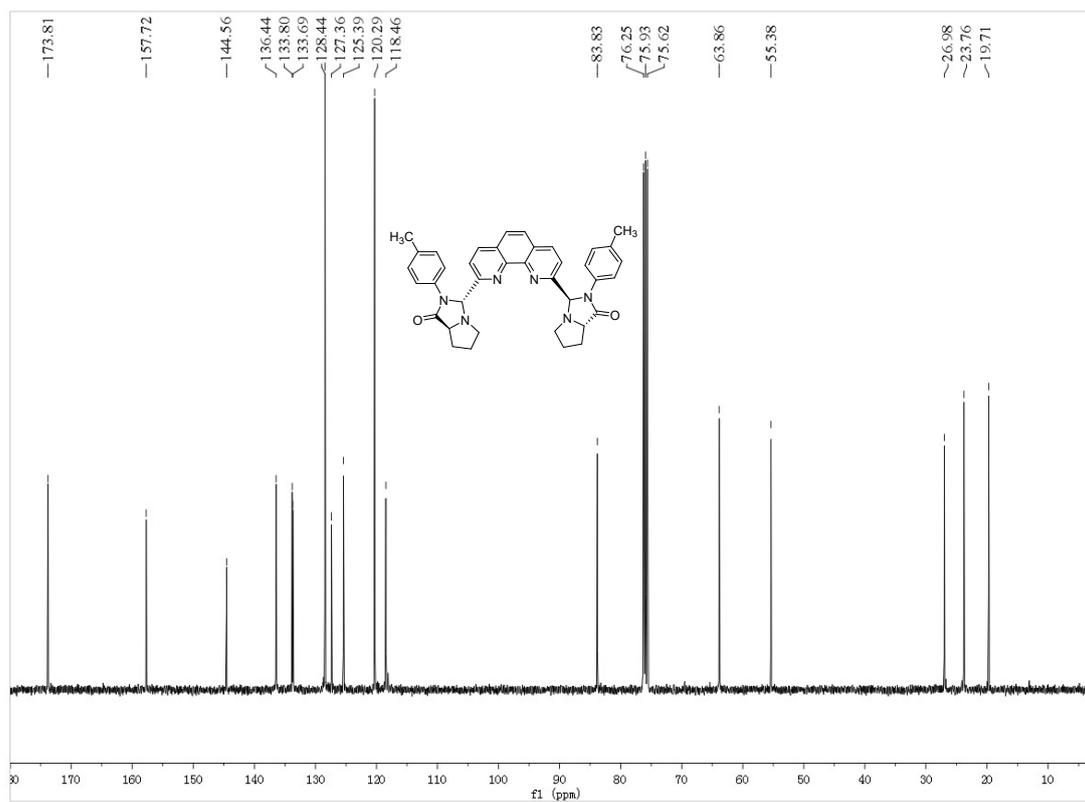
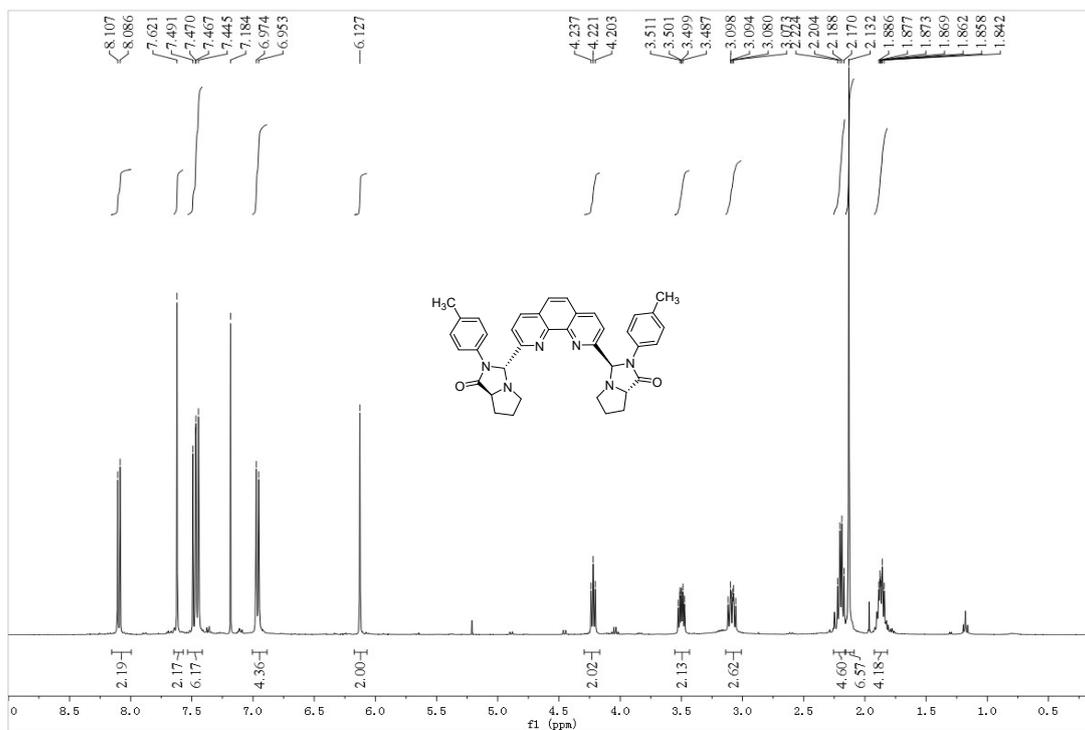
Table S3 Crystal data and structure refinement for L1a-Ni(OTf)₂-3H₂O complex

| | |
|---|--|
| Identification code | L1a-Ni(OTf)₂-3H₂O complex |
| Empirical formula | C ₄₀ H ₄₂ F ₆ N ₆ NiO ₁₁ S ₂ |
| Formula weight | 1019.62 |
| Temperature/K | 169.99(10) |
| Crystal system | orthorhombic |
| Space group | P2 ₁ 2 ₁ 2 ₁ |
| a/Å, b/Å, c/Å | 14.2901(4), 22.2090(8), 15.6839(4) |
| α/°, β/°, γ/° | 90, 90, 90 |
| Volume/Å ³ | 4977.6(3) |
| Z | 4 |
| ρ _{calc} /cm ³ | 1.361 |
| μ/mm ⁻¹ | 2.065 |
| F(000) | 2104.0 |
| Radiation | Cu Kα (λ = 1.54184) |
| Crystal size/mm ³ | 0.15 × 0.13 × 0.1 |
| 2θ range for data collection/° | 6.9 to 147.906 |
| Index ranges | -17 ≤ h ≤ 14, -22 ≤ k ≤ 27, -18 ≤ l ≤ 19 |
| Reflections collected | 27893 |
| Independent reflections | 9876 [R _{int} = 0.0507, R _{sigma} = 0.0504] |
| Data/restraints/parameters | 9876/405/741 |
| Goodness-of-fit on F ² | 1.016 |
| Final R indexes [I >= 2σ(I)] | R ₁ = 0.0616, wR ₂ = 0.1675 |
| Final R indexes [all data] | R ₁ = 0.0666, wR ₂ = 0.1730 |
| Largest diff. peak/hole / e Å ⁻³ | 0.53/-0.99 |
| Flack parameter | 0.011(16)/0.02(3) |

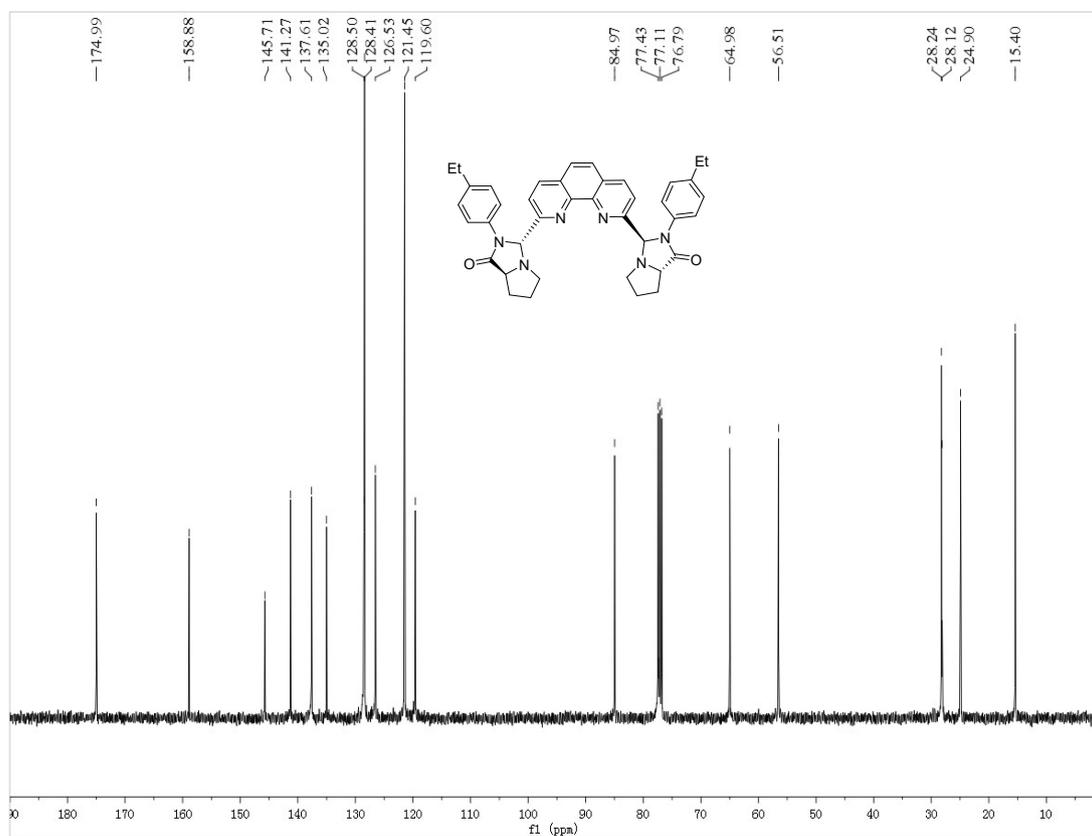
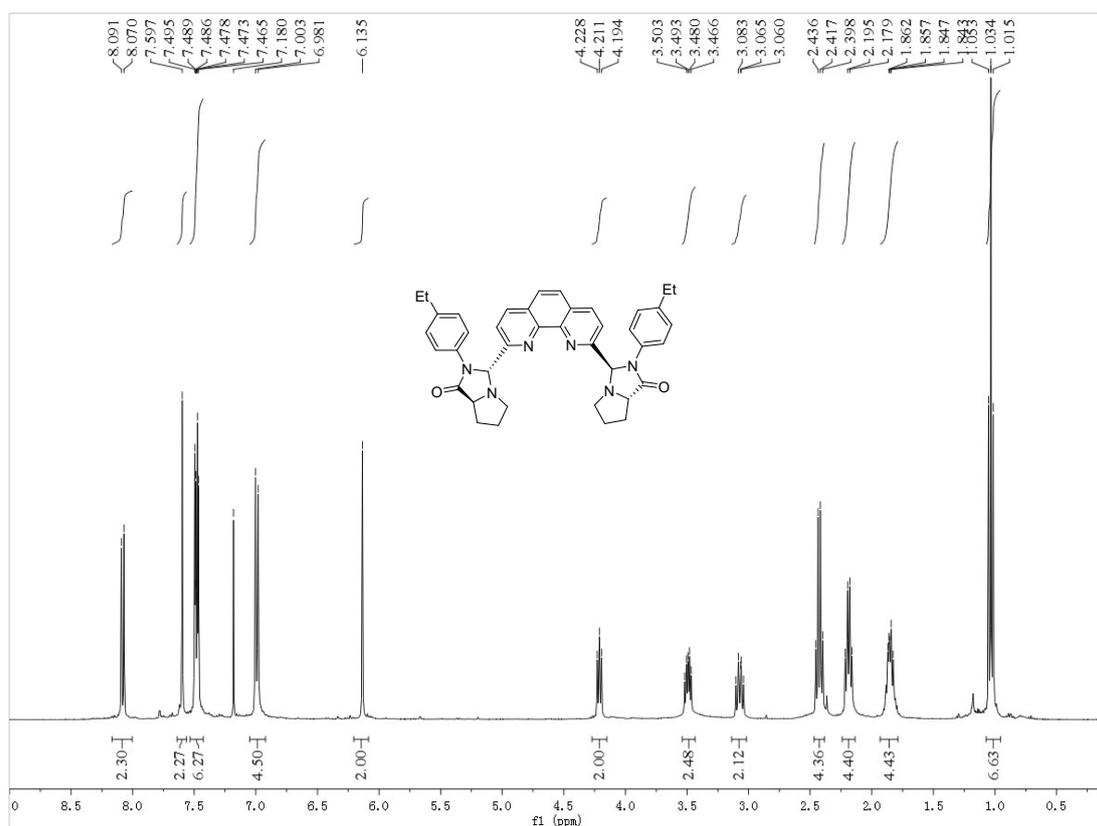
Crystal Data for C₄₀H₄₂F₆N₆NiO₁₁S₂ (*M* = 1019.62 g/mol): orthorhombic, space group P2₁2₁2₁ (no. 19), *a* = 14.2901(4) Å, *b* = 22.2090(8) Å, *c* = 15.6839(4) Å, *V* = 4977.6(3) Å³, *Z* = 4, *T* = 169.99(10) K, μ(Cu Kα) = 2.065 mm⁻¹, *D*_{calc} = 1.361 g/cm³, 27893 reflections measured (6.9° ≤ 2θ ≤ 147.906°), 9876 unique (*R*_{int} = 0.0507, *R*_{sigma} = 0.0504) which were used in all calculations. The final *R*₁ was 0.0616 (*I* > 2σ(*I*)) and *wR*₂ was 0.1730 (all data).

10. The copies of ^1H NMR, ^{13}C NMR and HPLC spectra for compounds L and 6

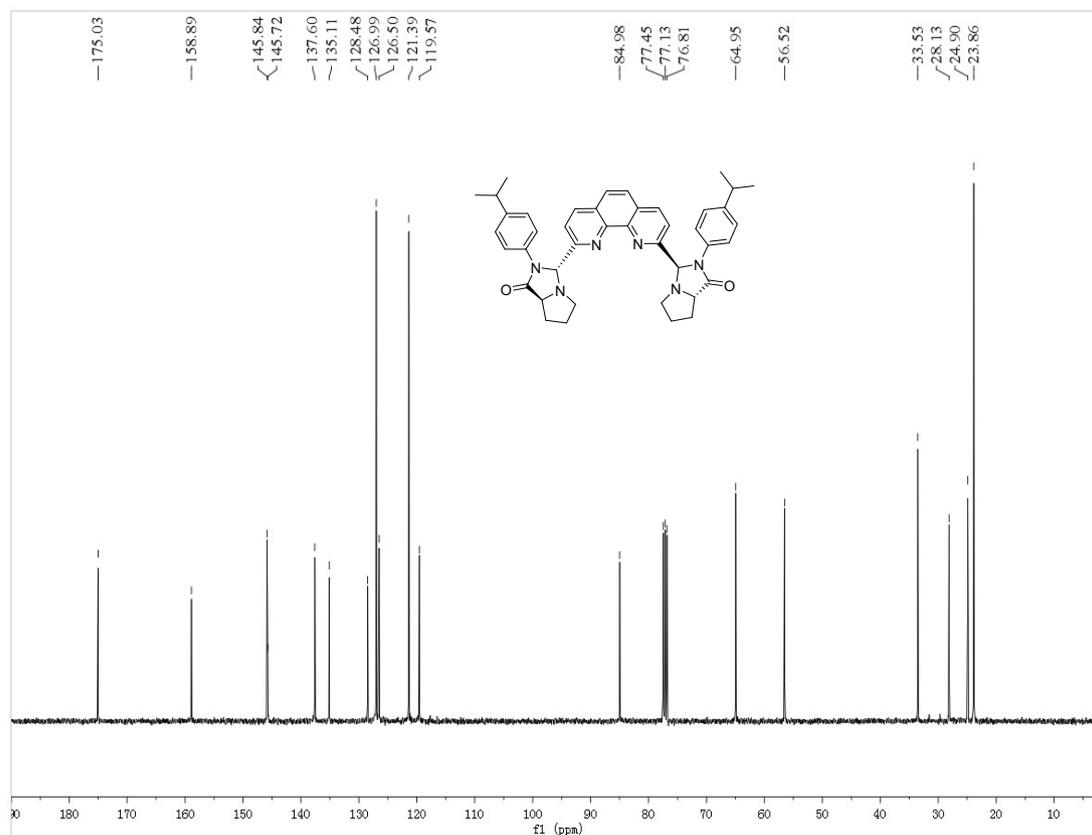
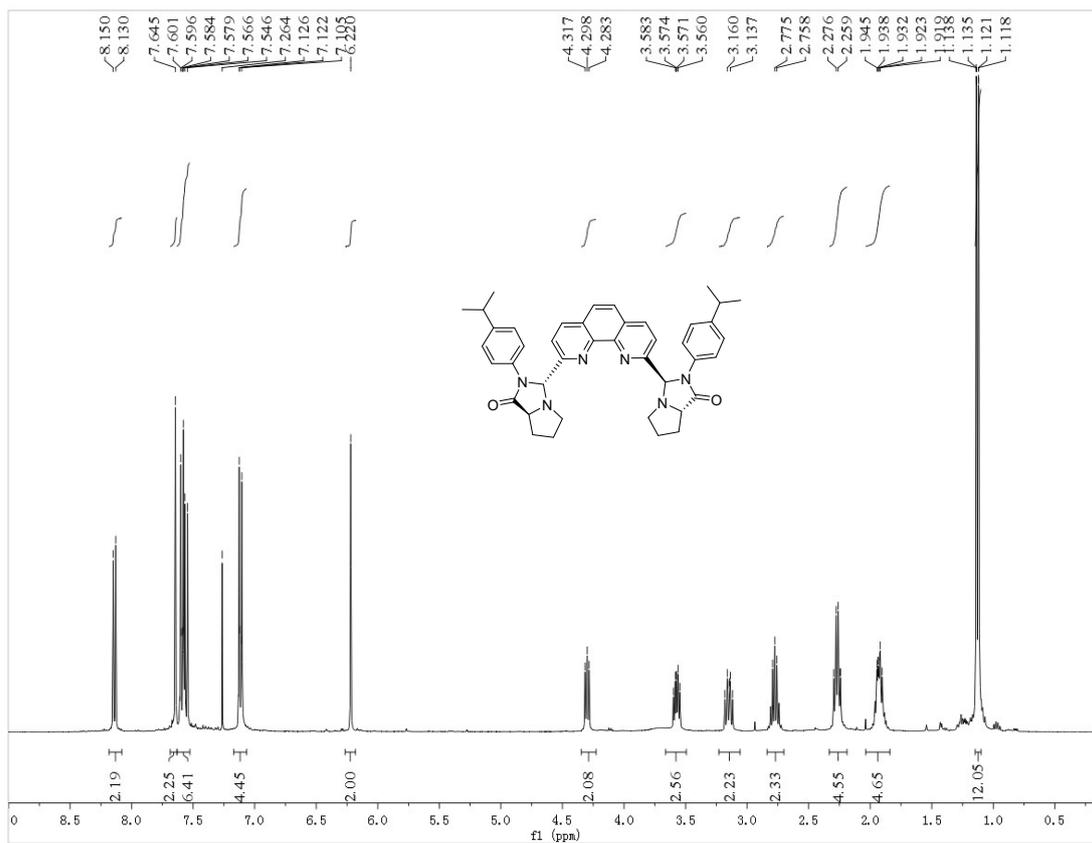
^1H and ^{13}C NMR of L1a



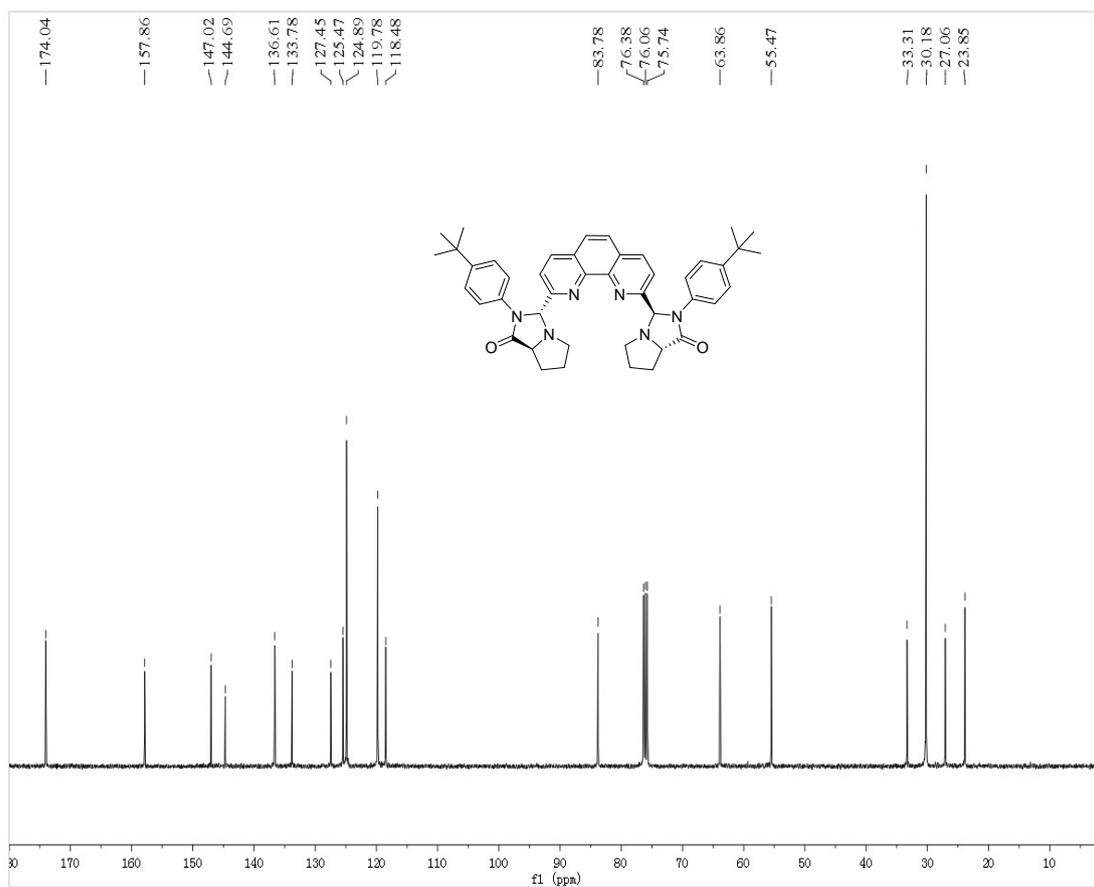
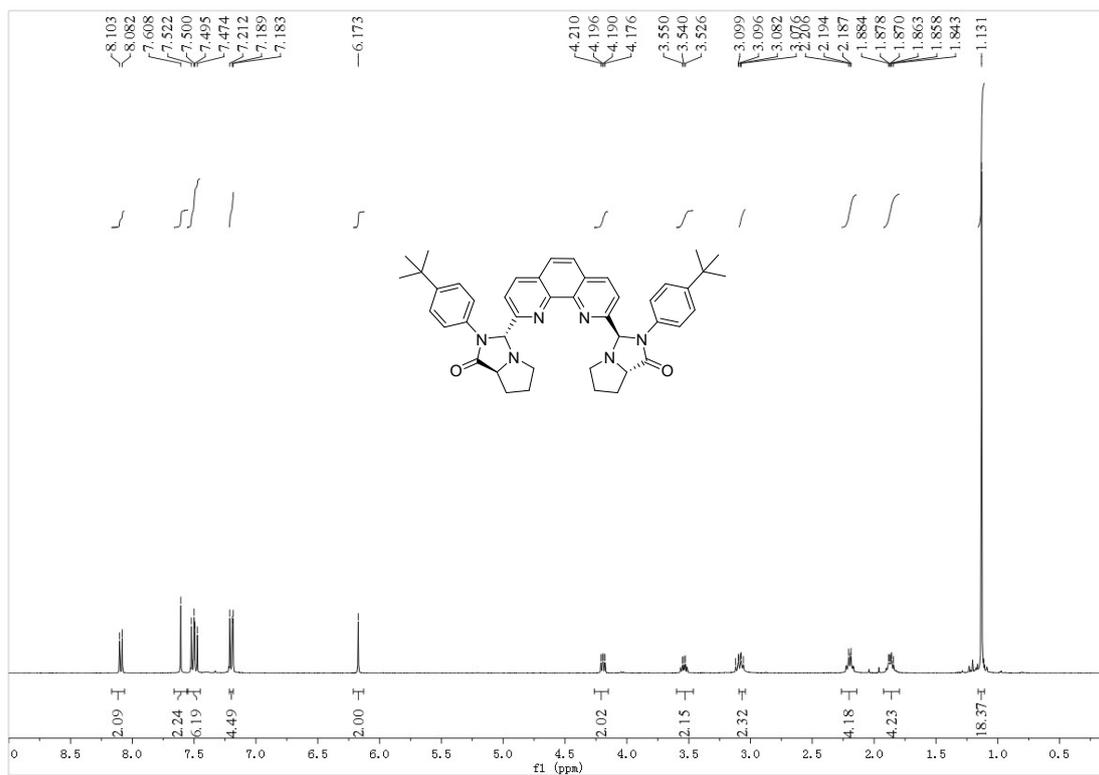
¹H and ¹³C NMR of L1b



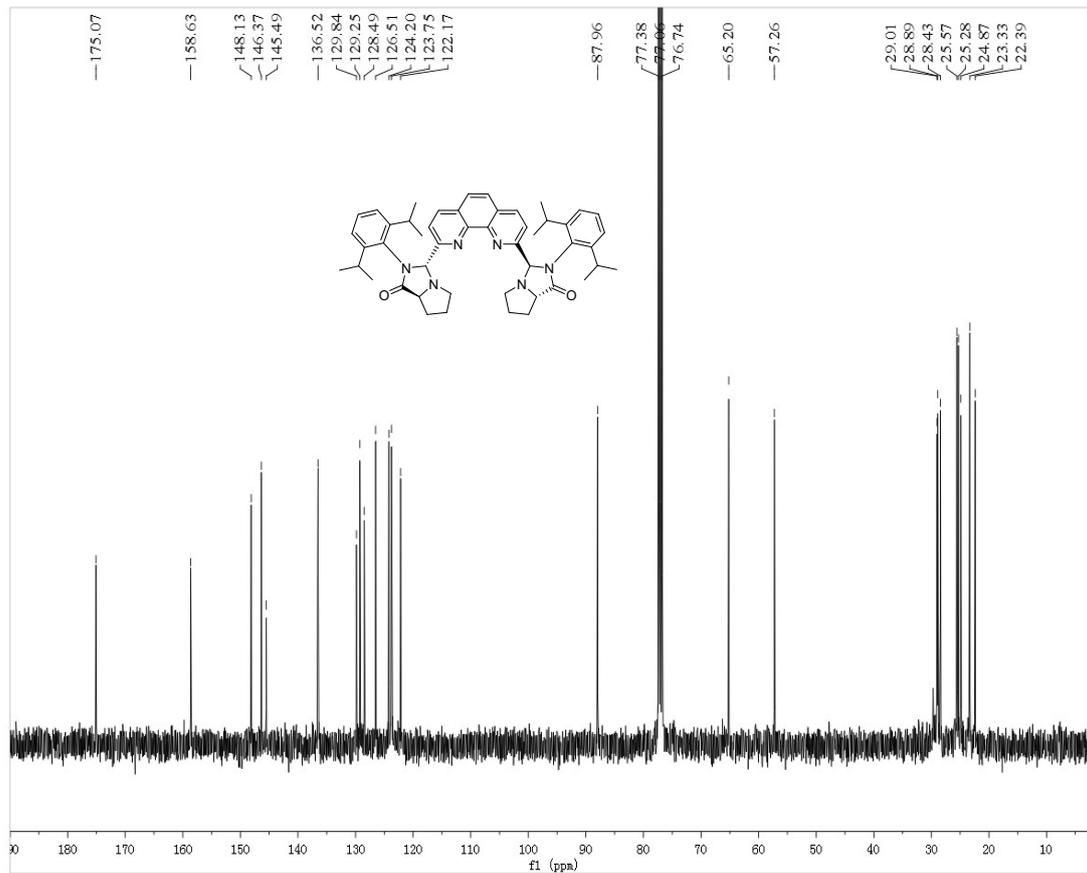
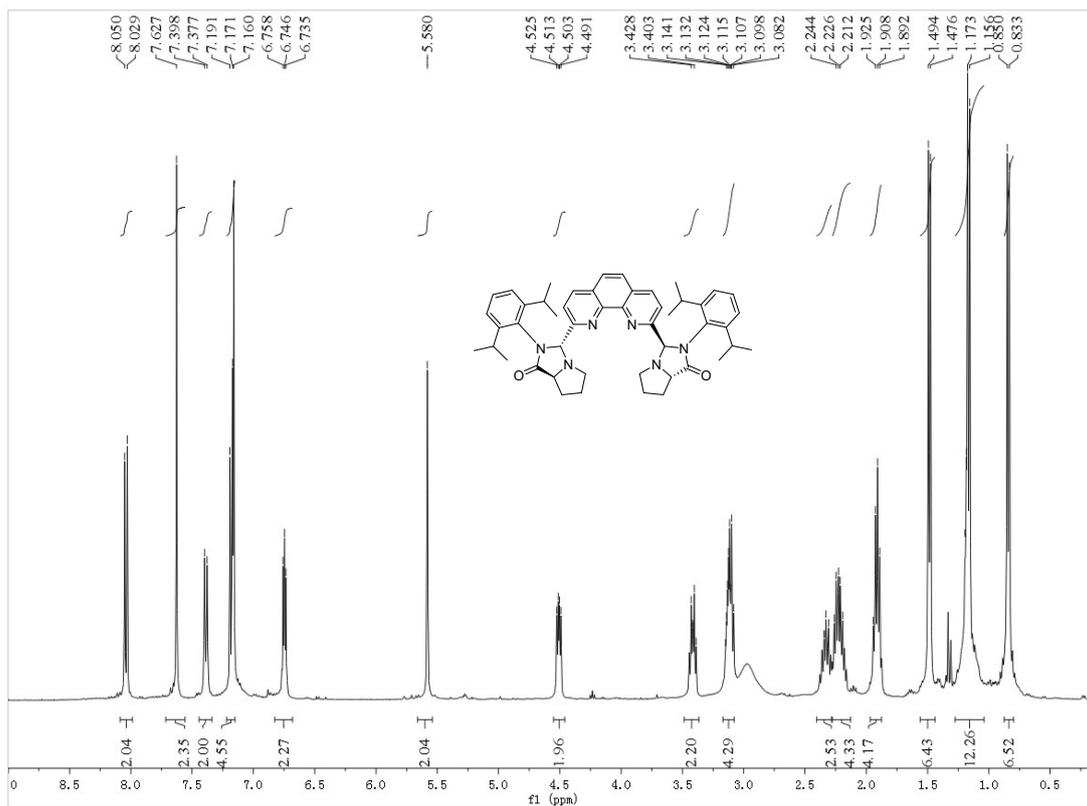
¹H and ¹³C NMR of L1c



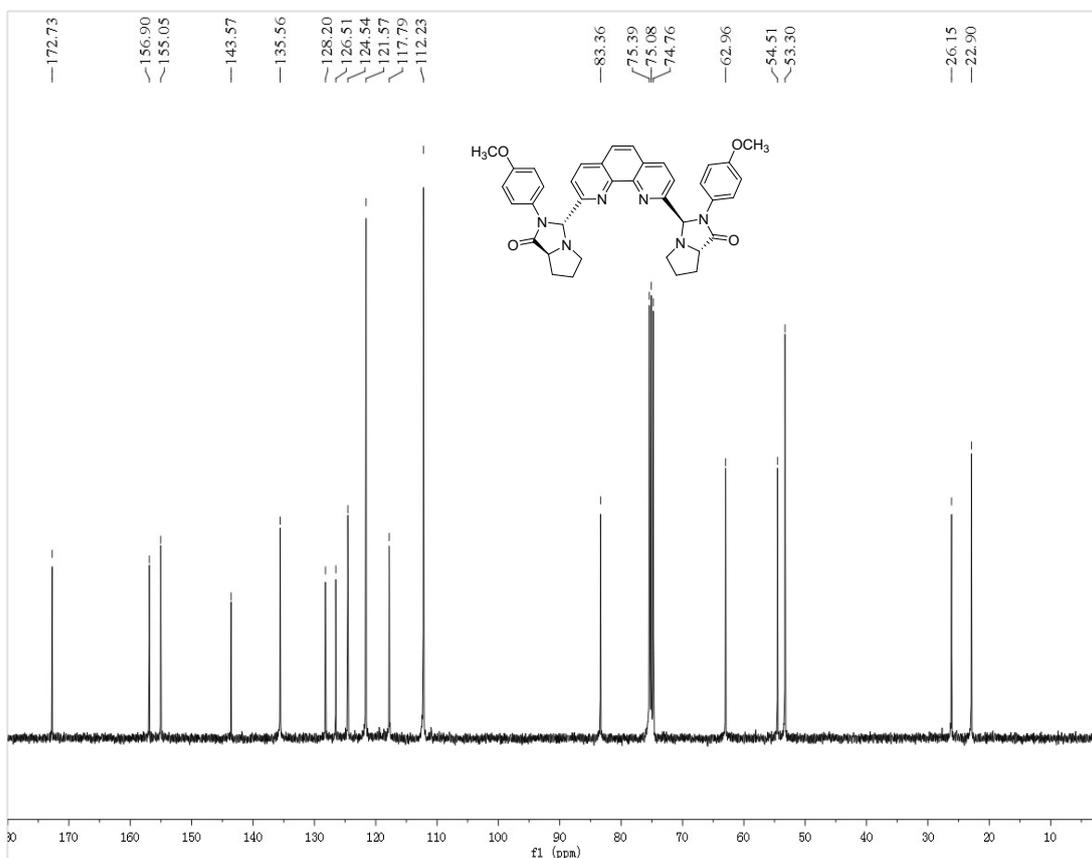
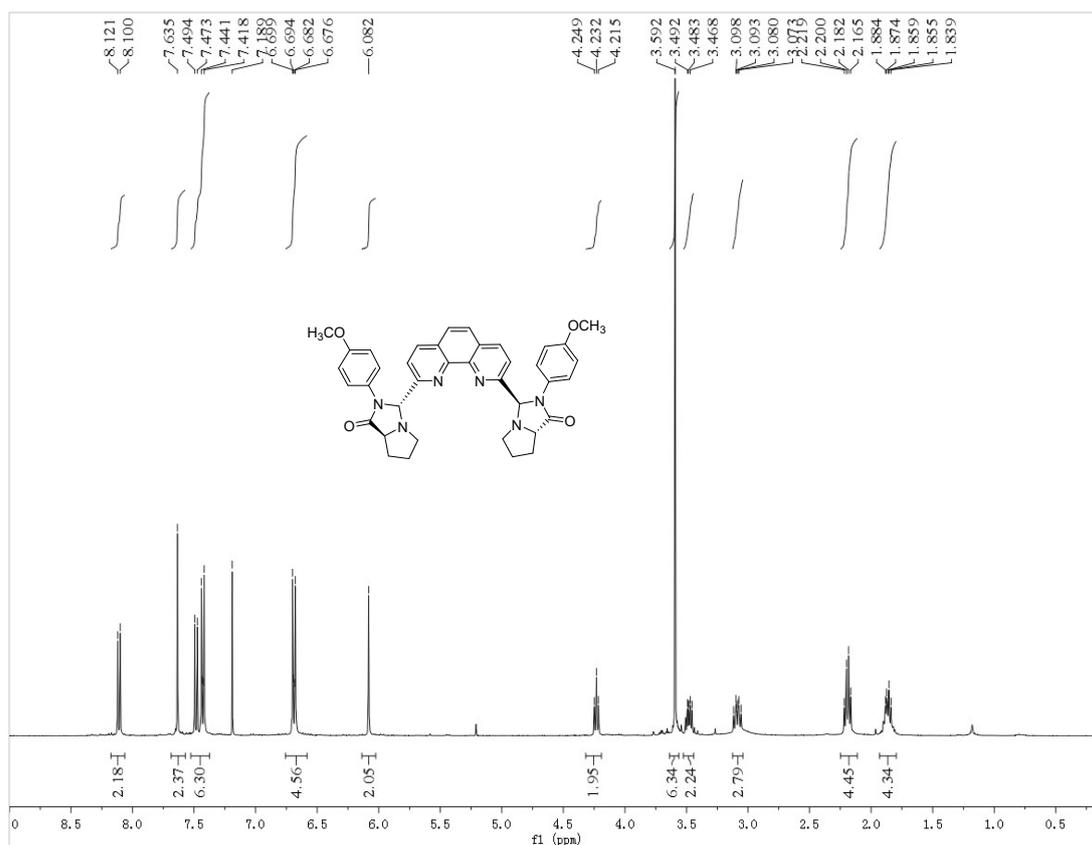
¹H and ¹³C NMR of L1d



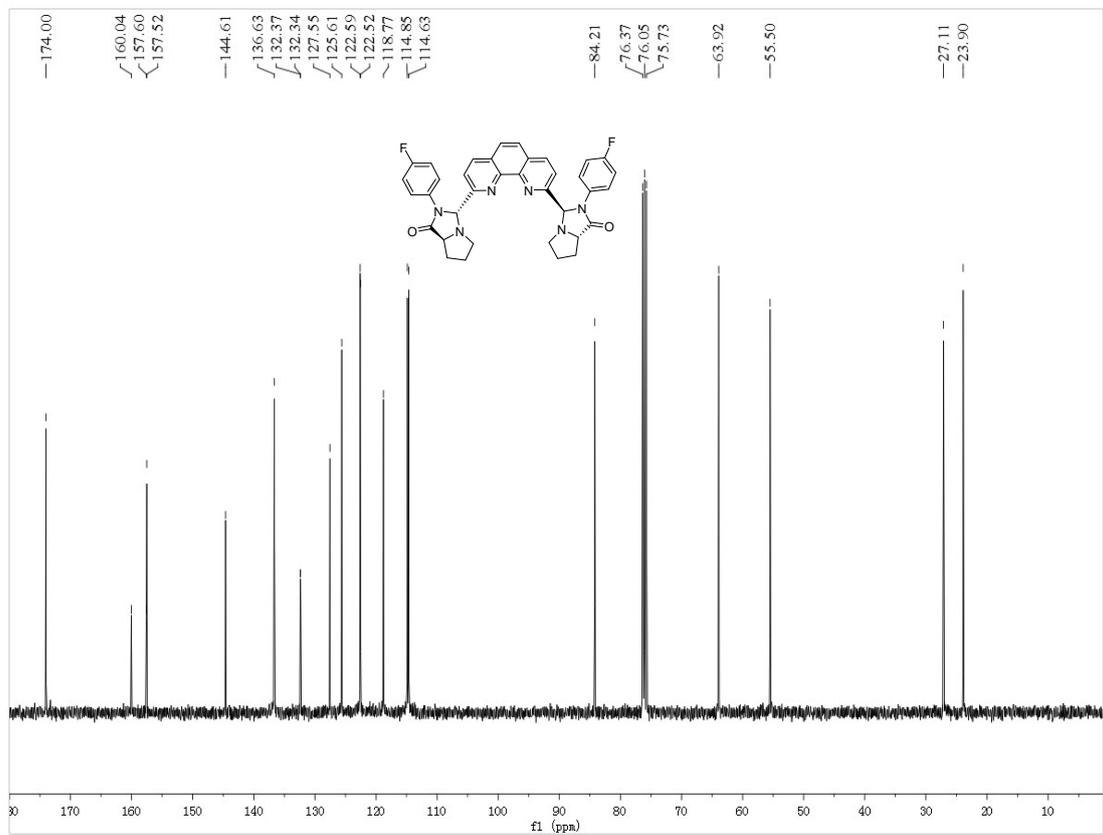
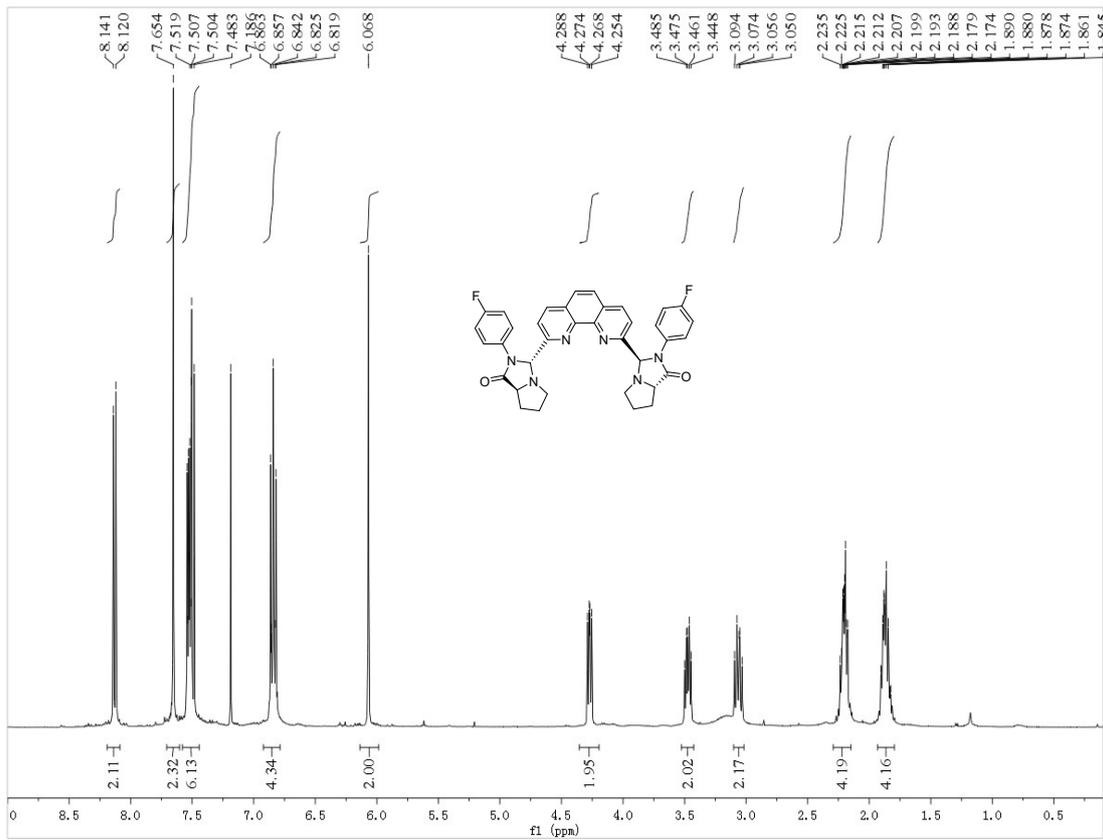
¹H and ¹³C NMR of L1e



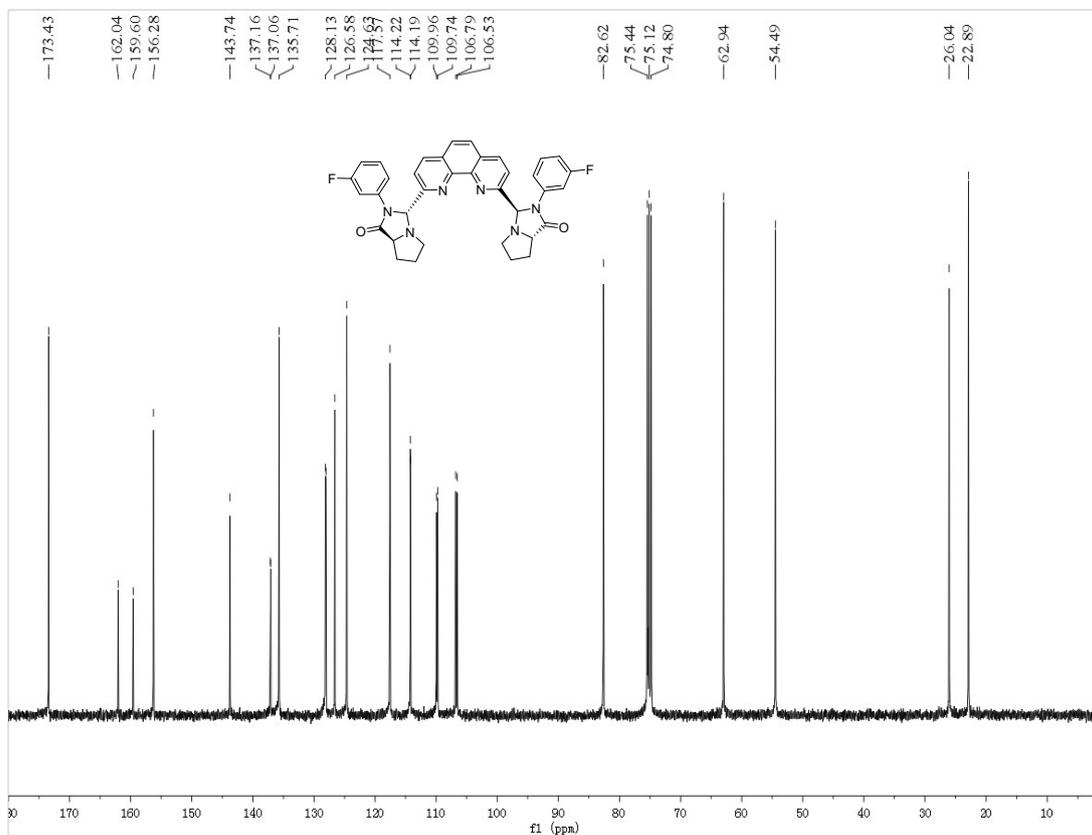
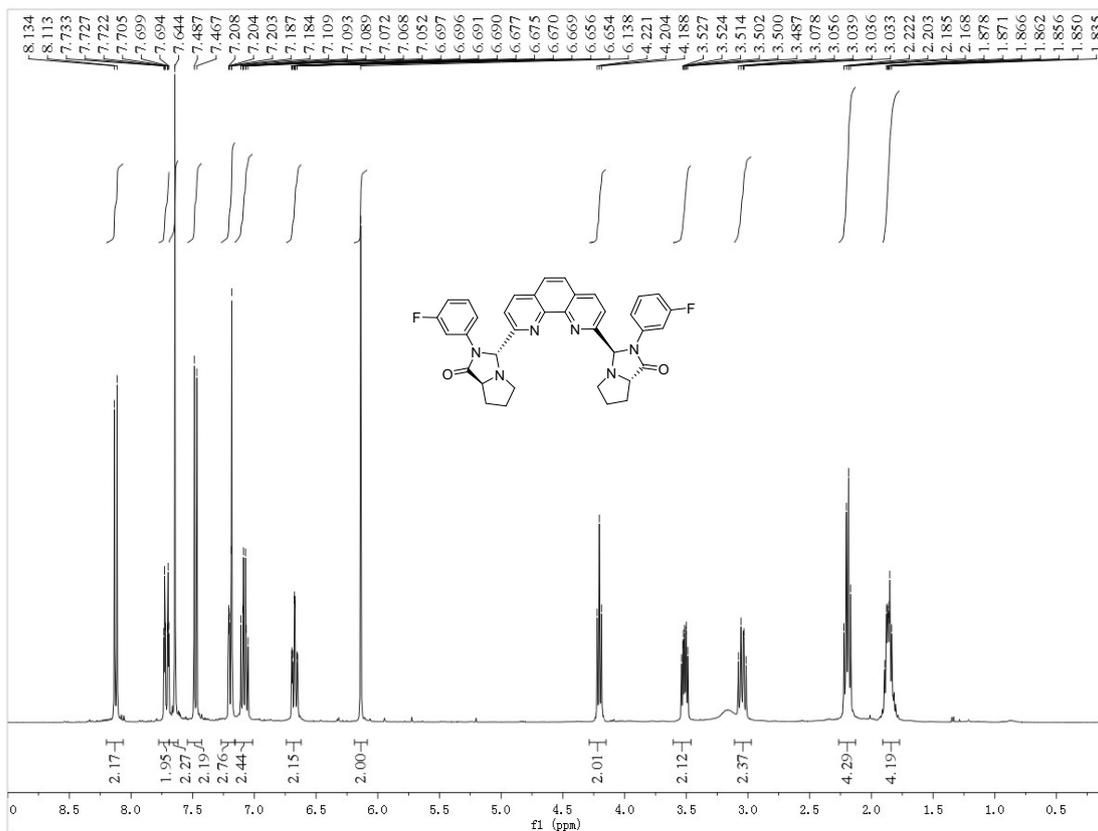
¹H and ¹³C NMR of L1f



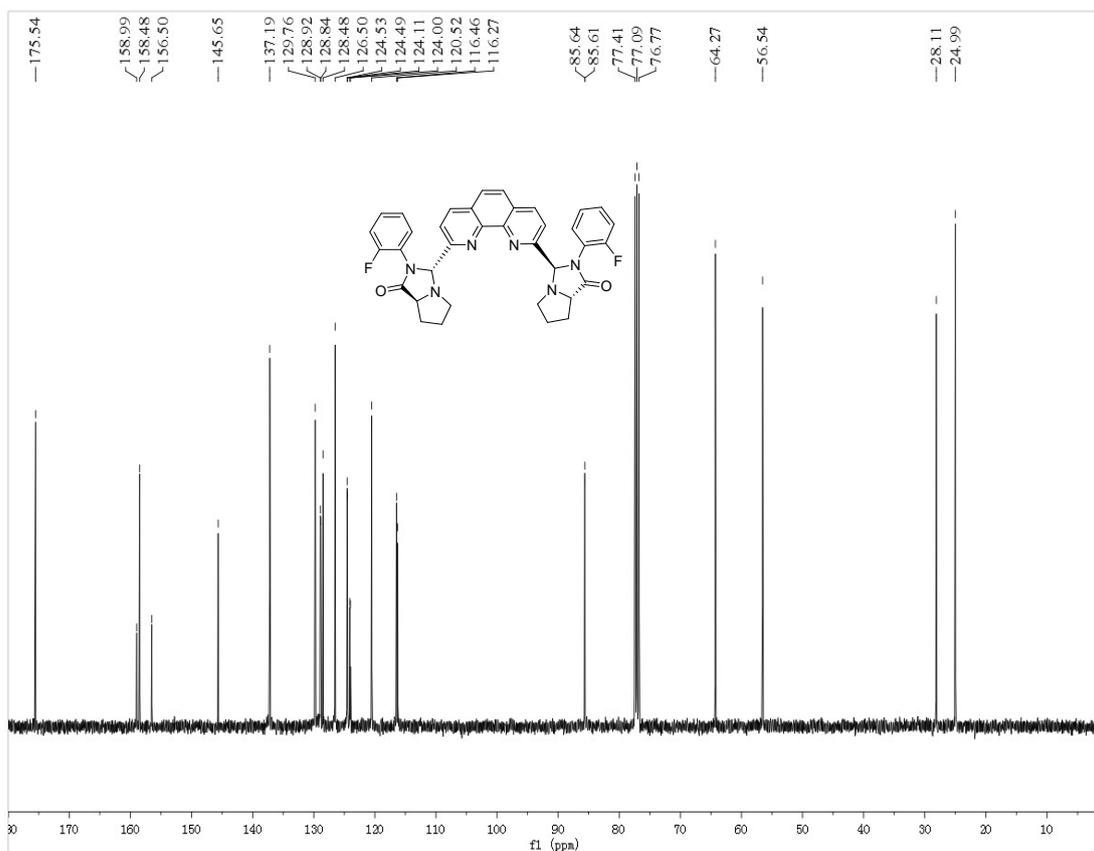
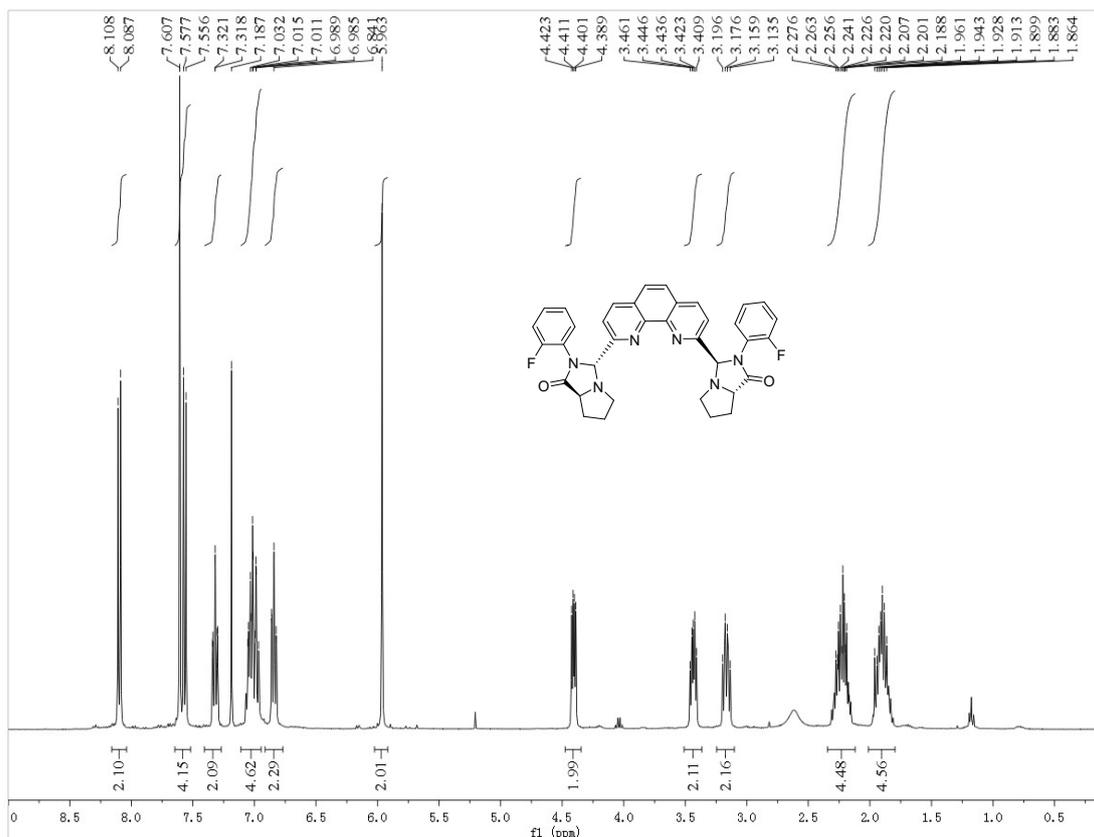
¹H and ¹³C NMR of L1g



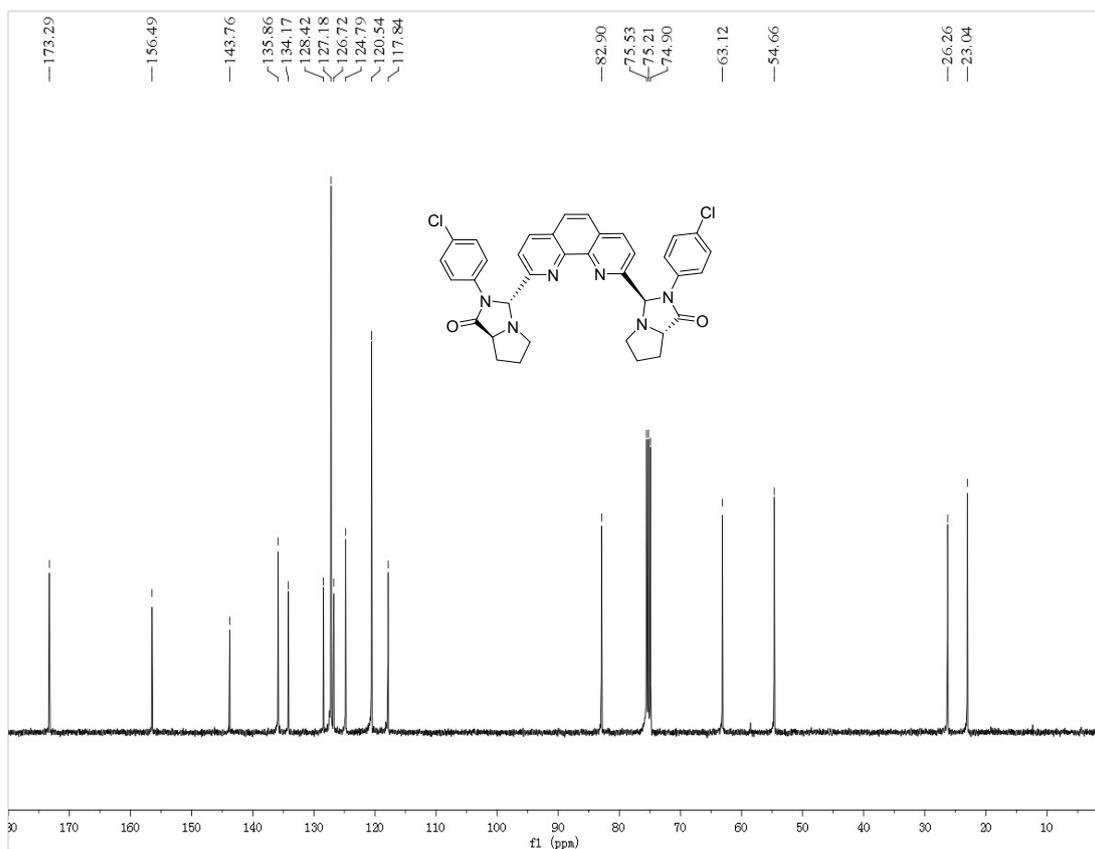
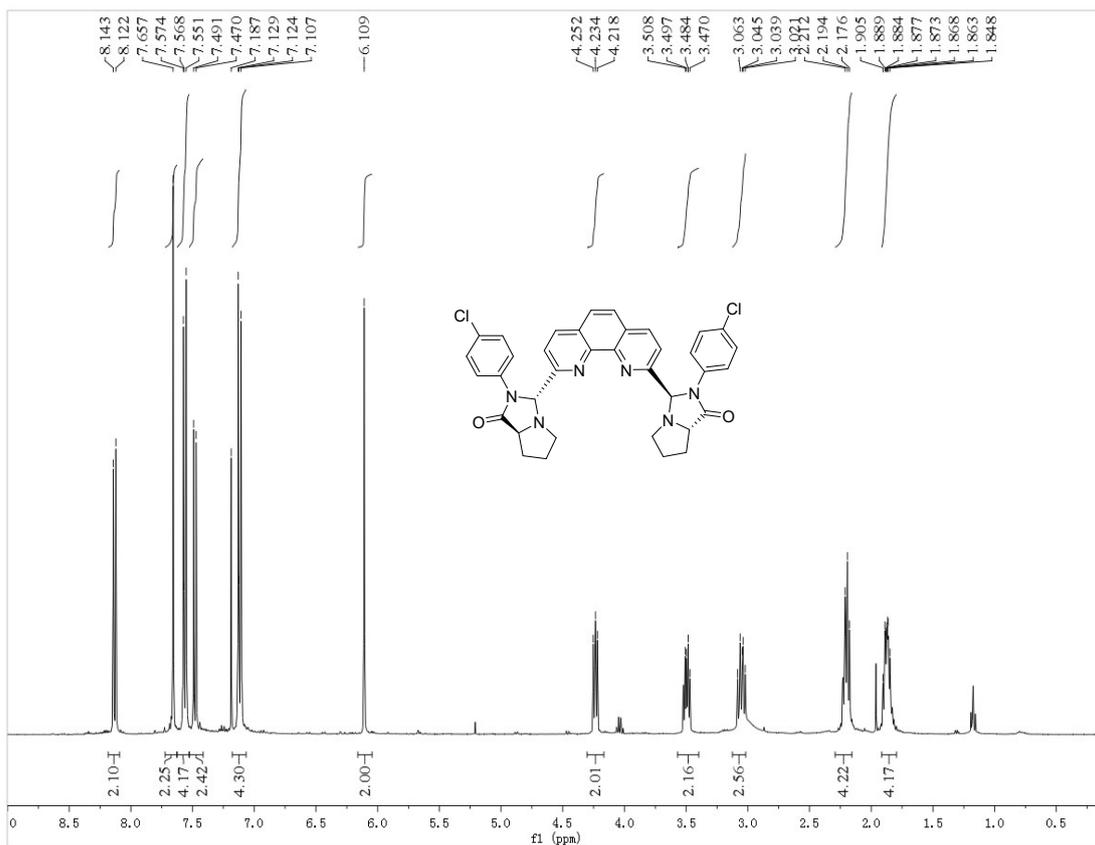
^1H and ^{13}C NMR of L1h



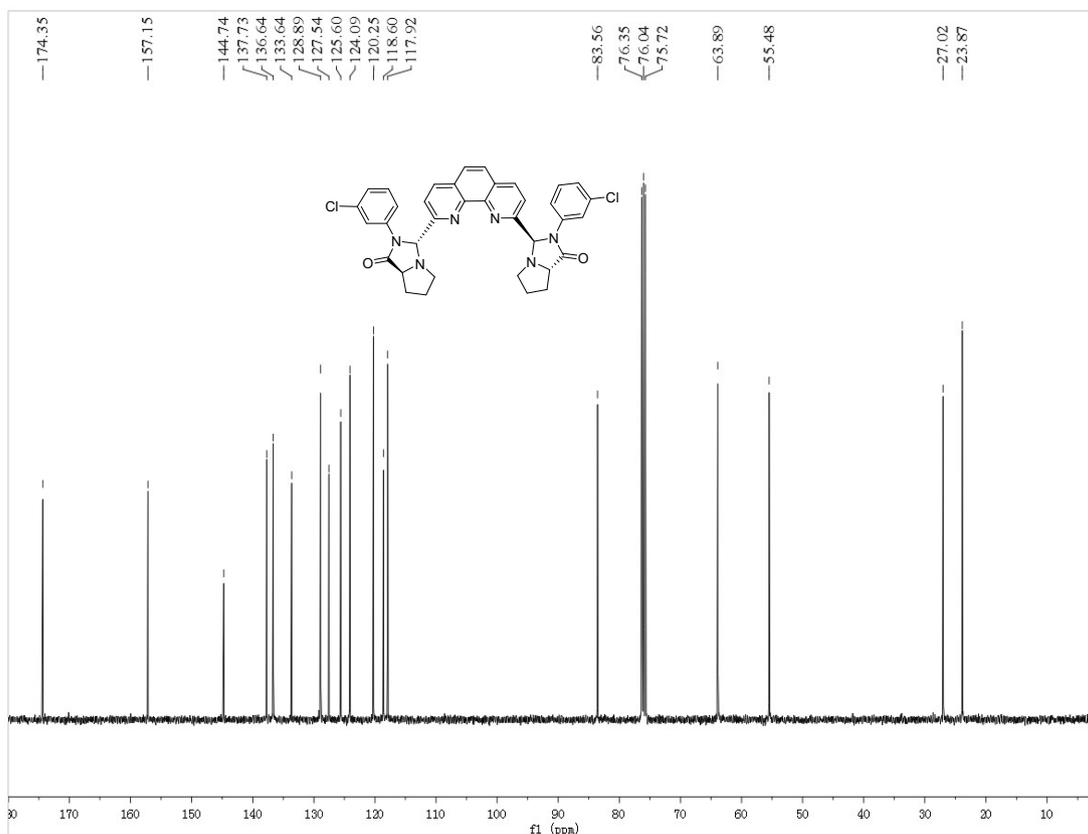
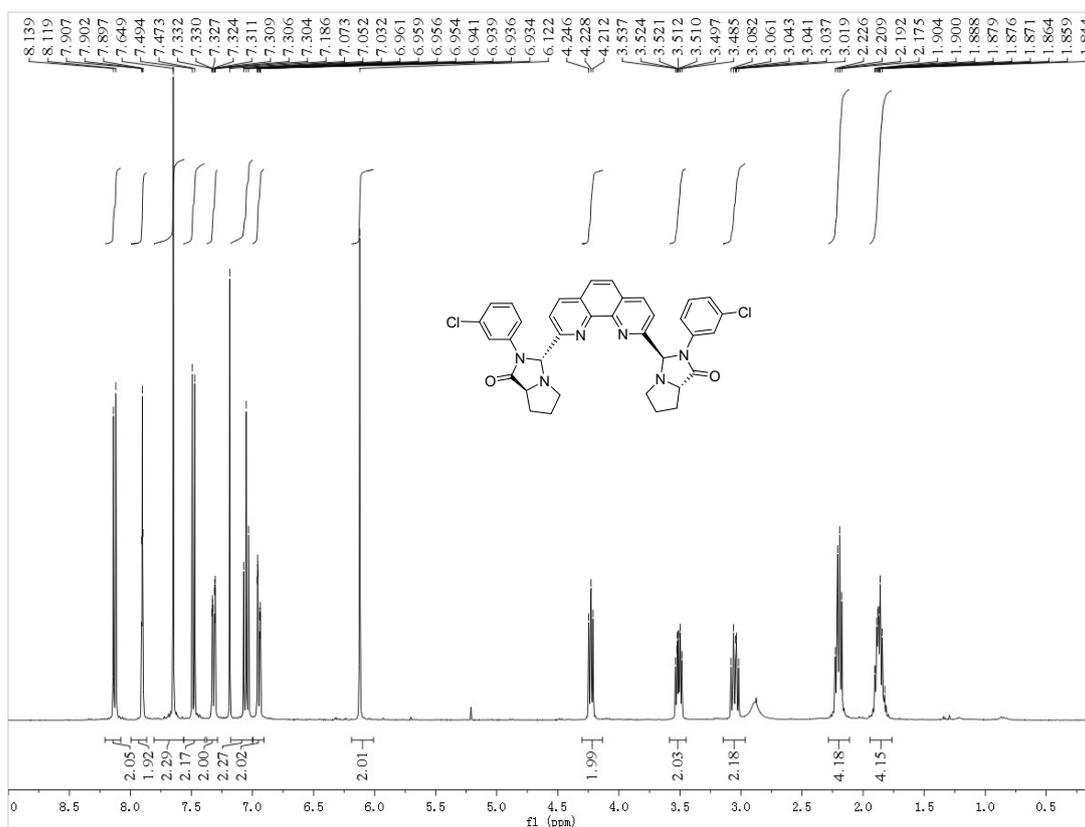
¹H and ¹³C NMR of L1i



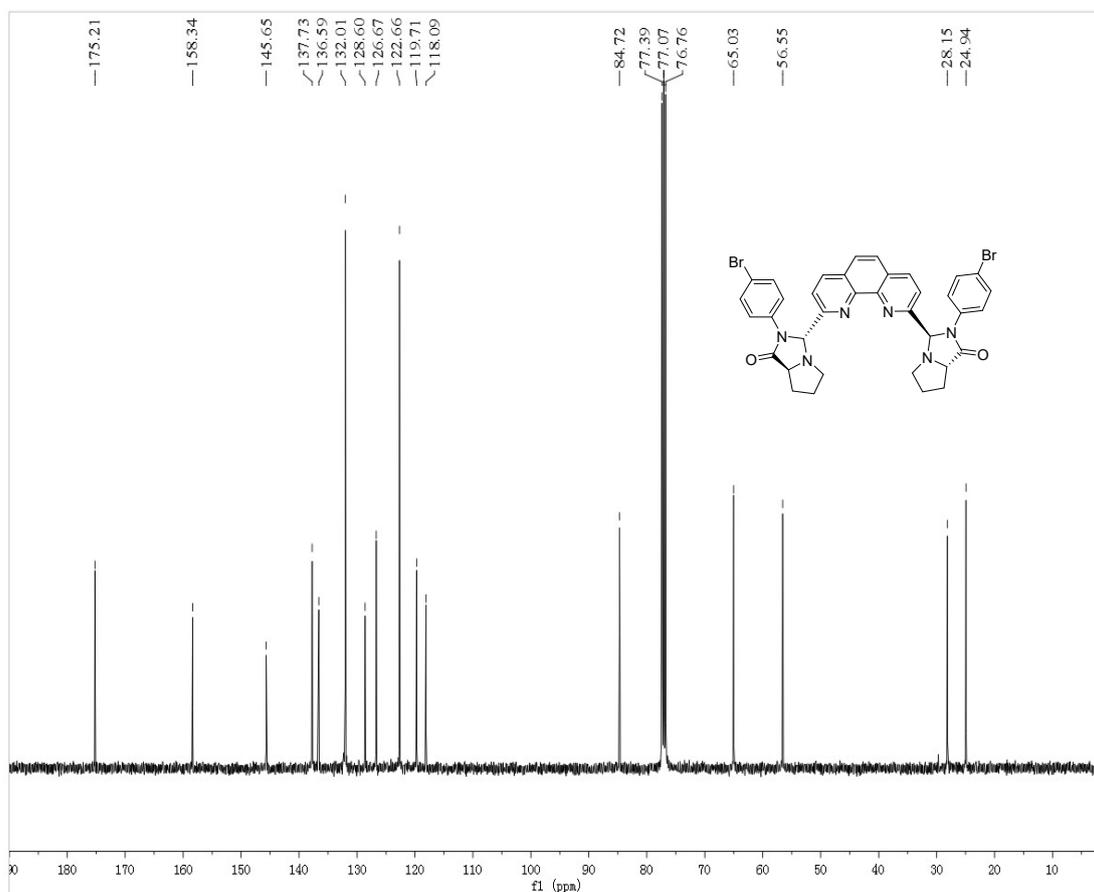
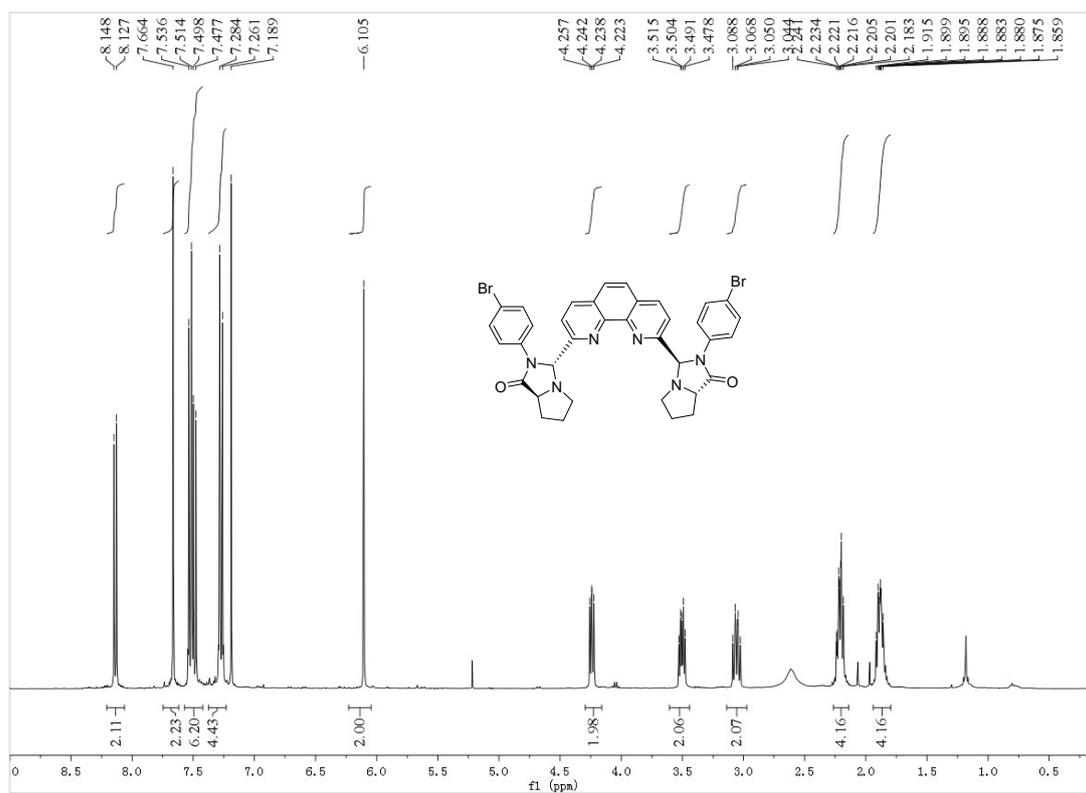
¹H and ¹³C NMR of L1j



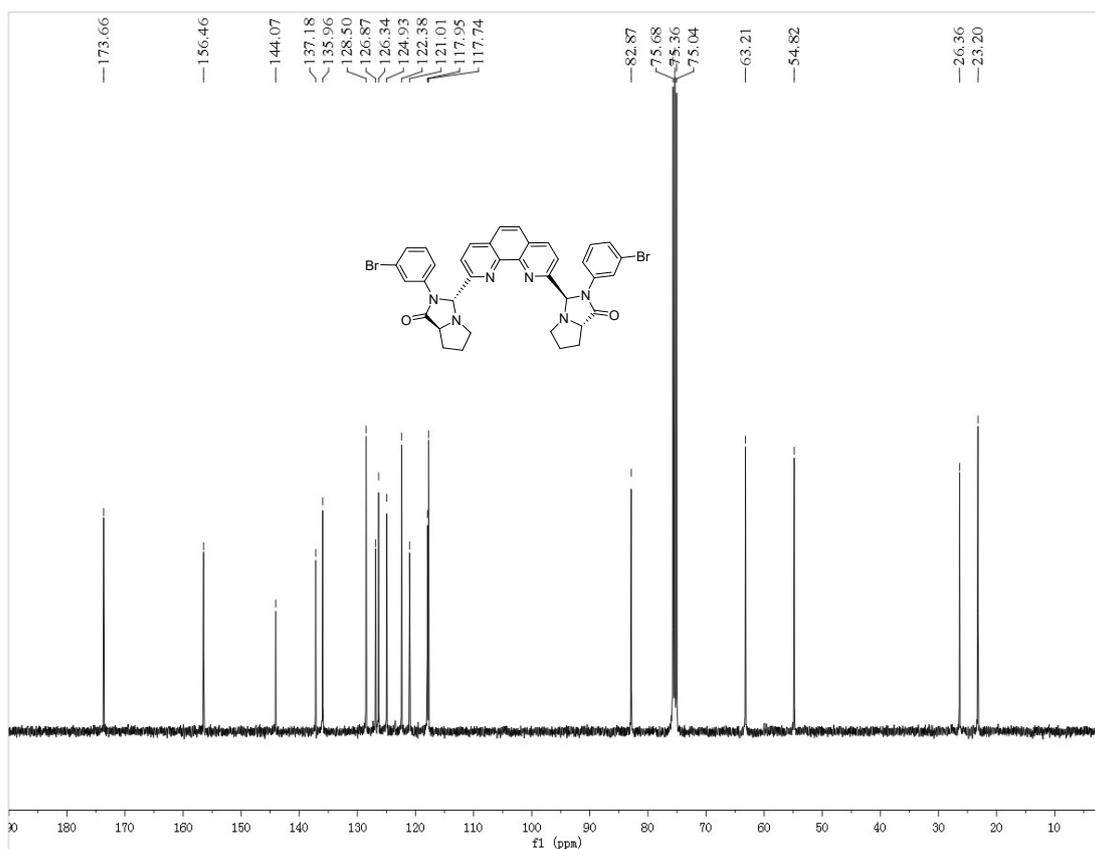
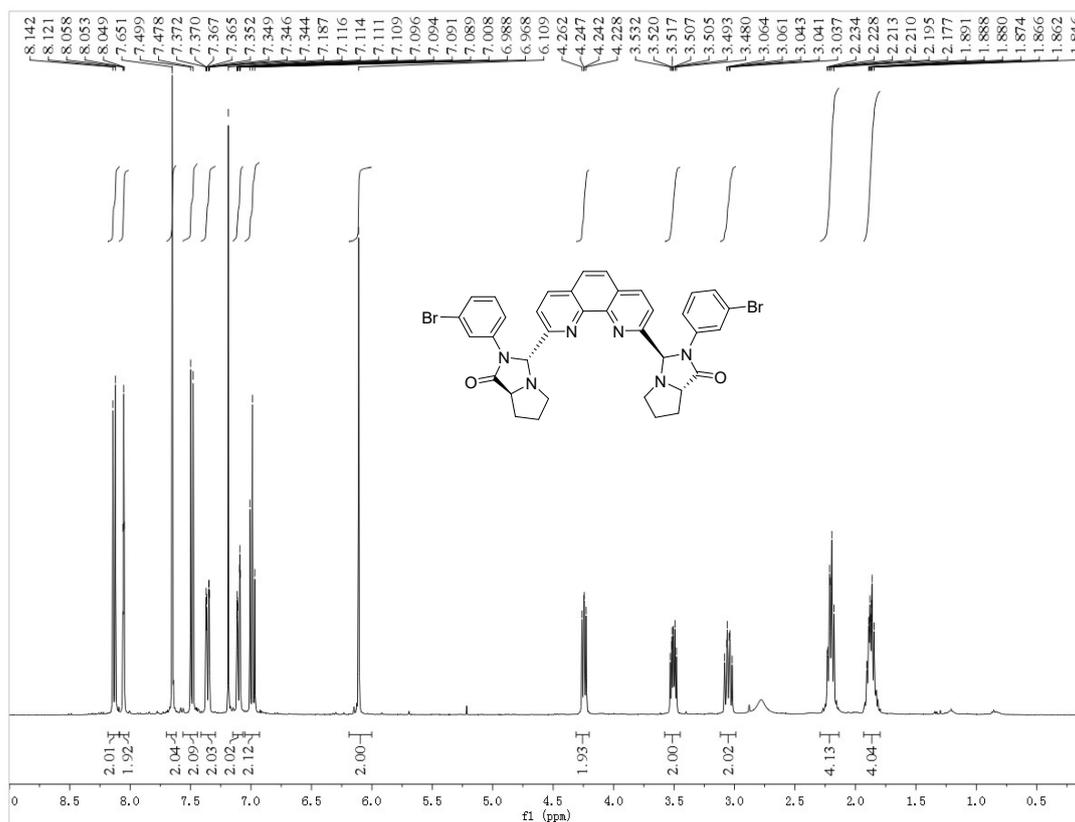
¹H and ¹³C NMR of L1k



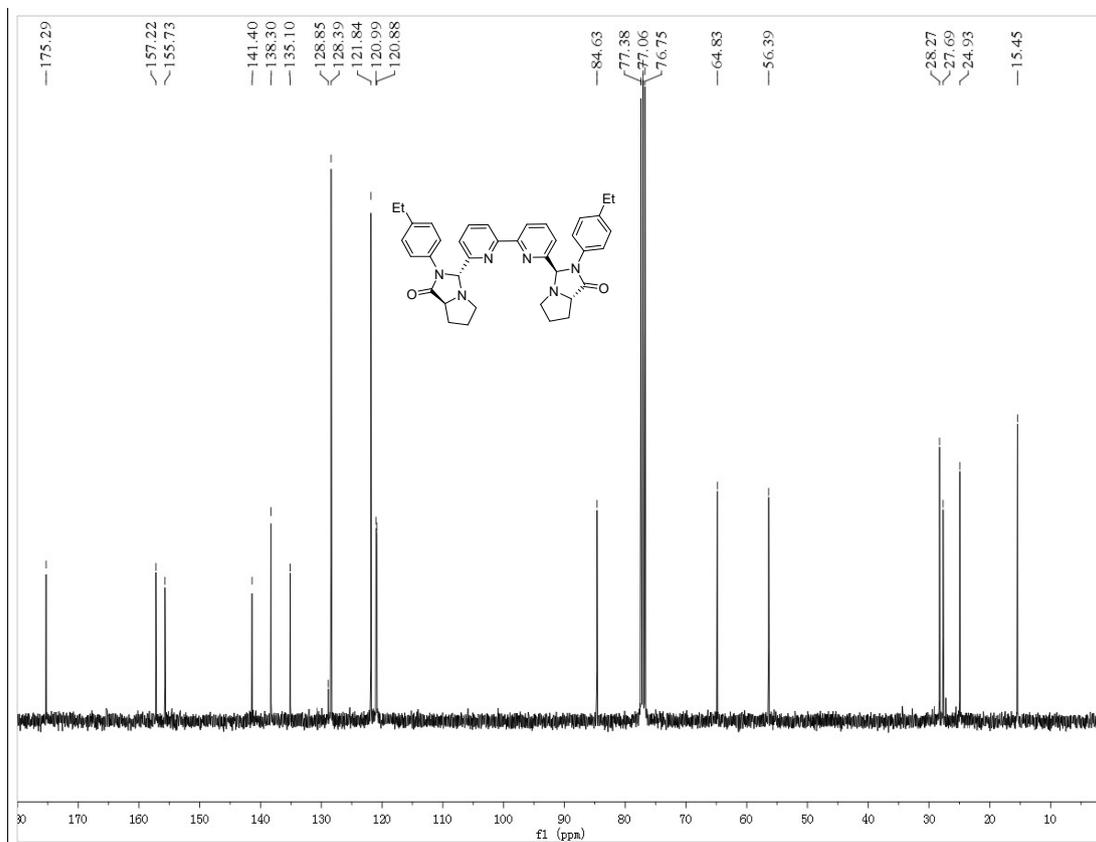
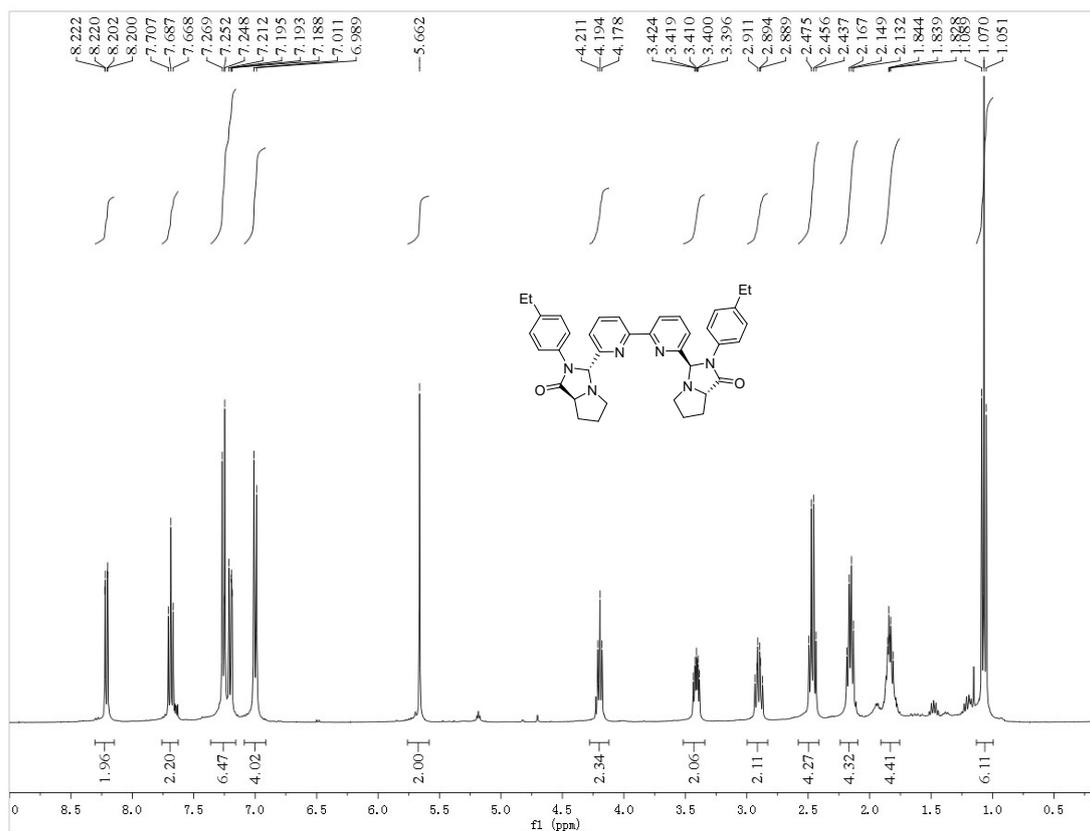
^1H and ^{13}C NMR of L11



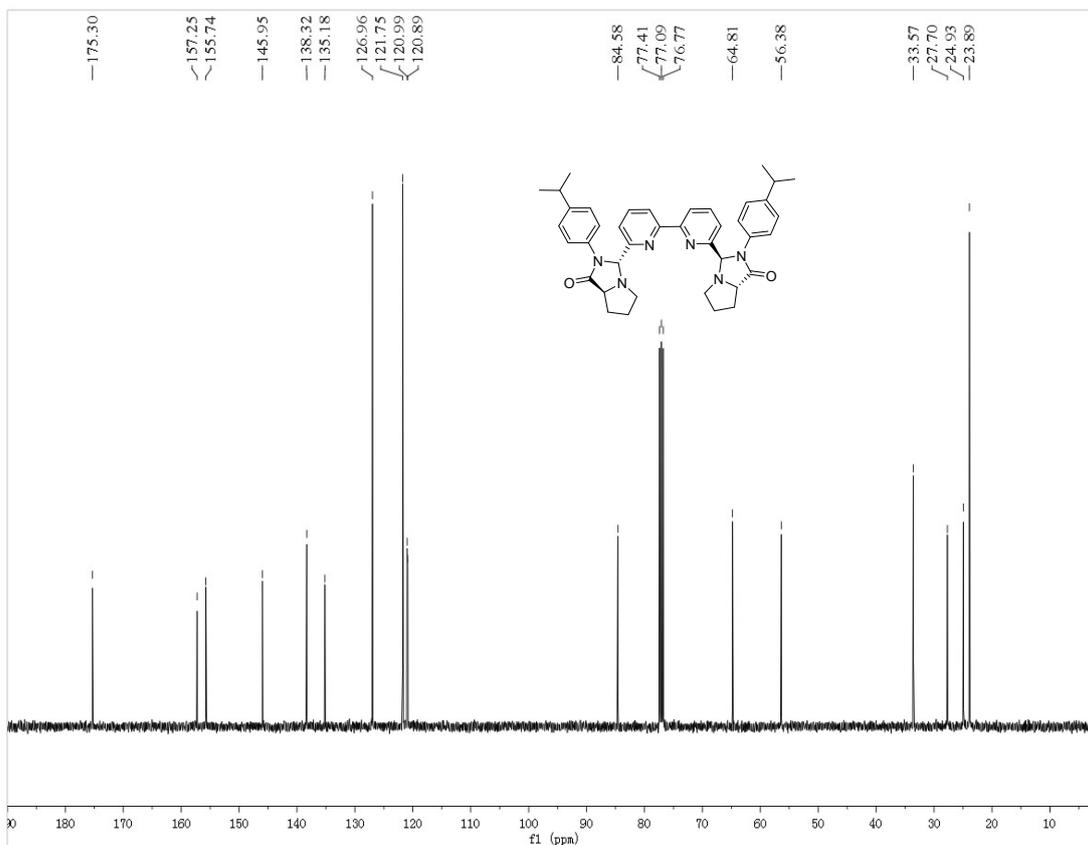
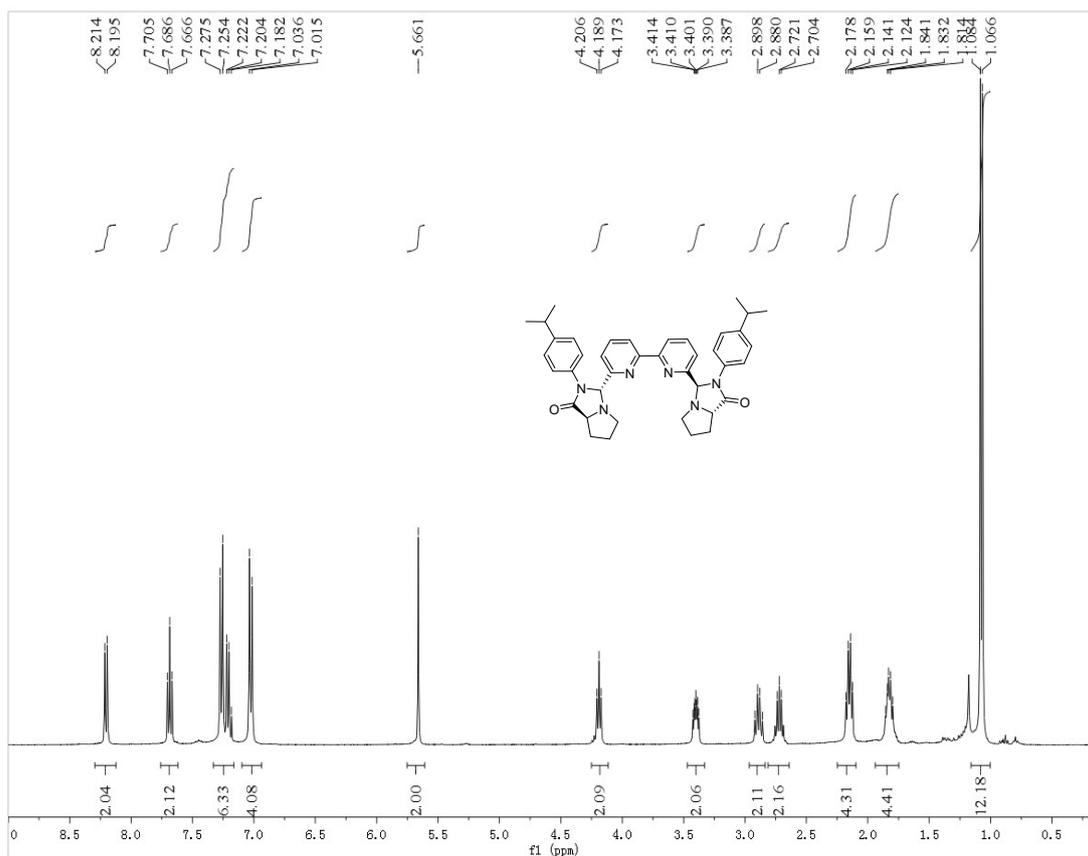
¹H and ¹³C NMR of L1m



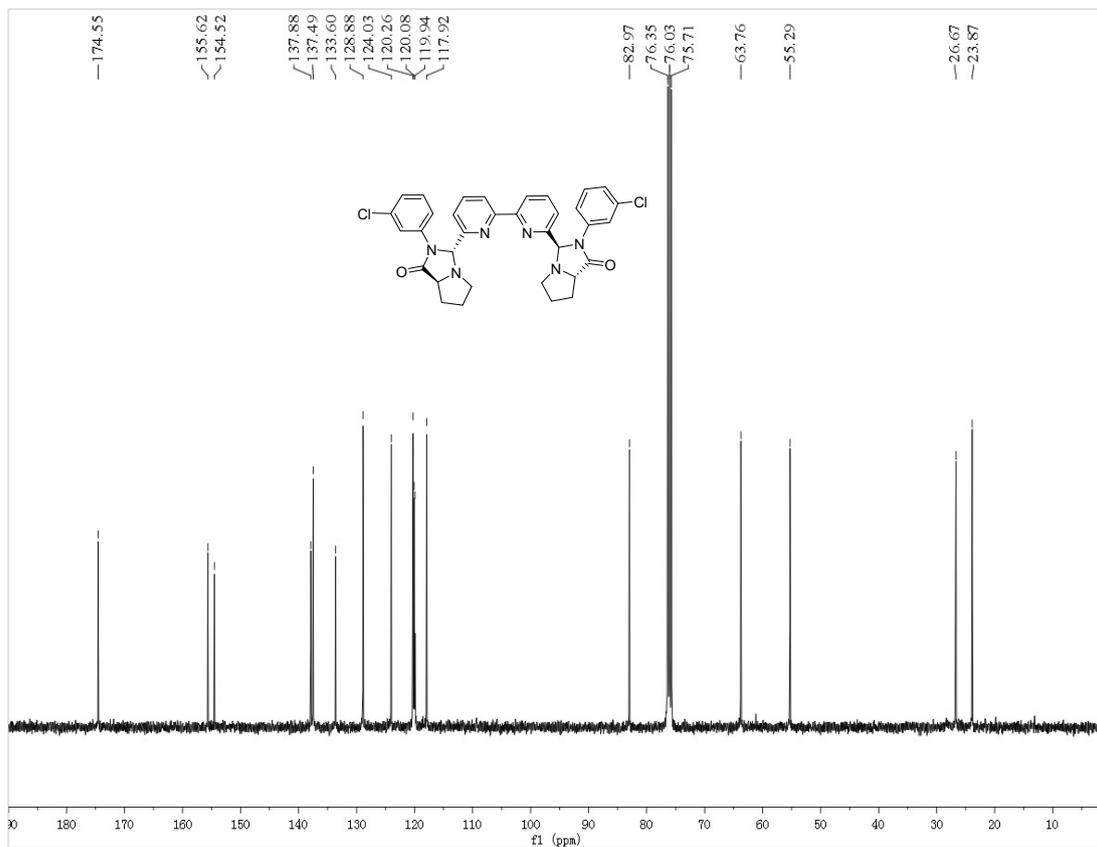
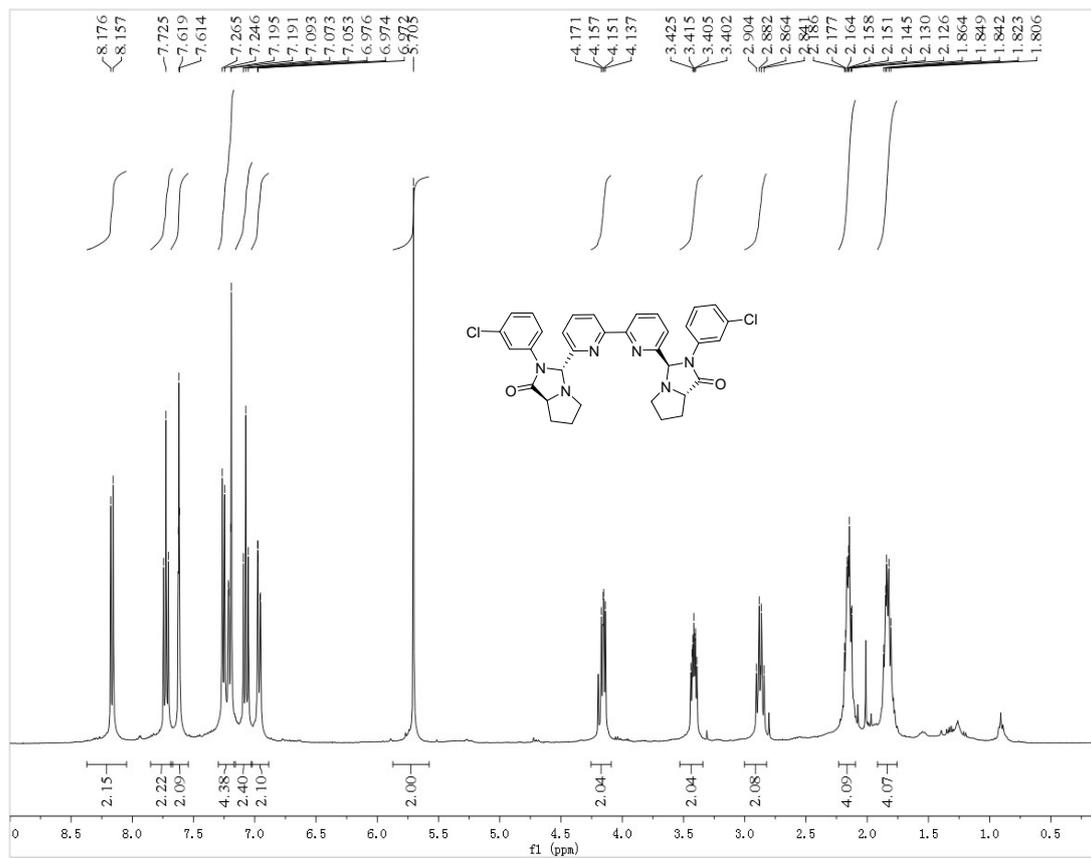
¹H and ¹³C NMR of L1n



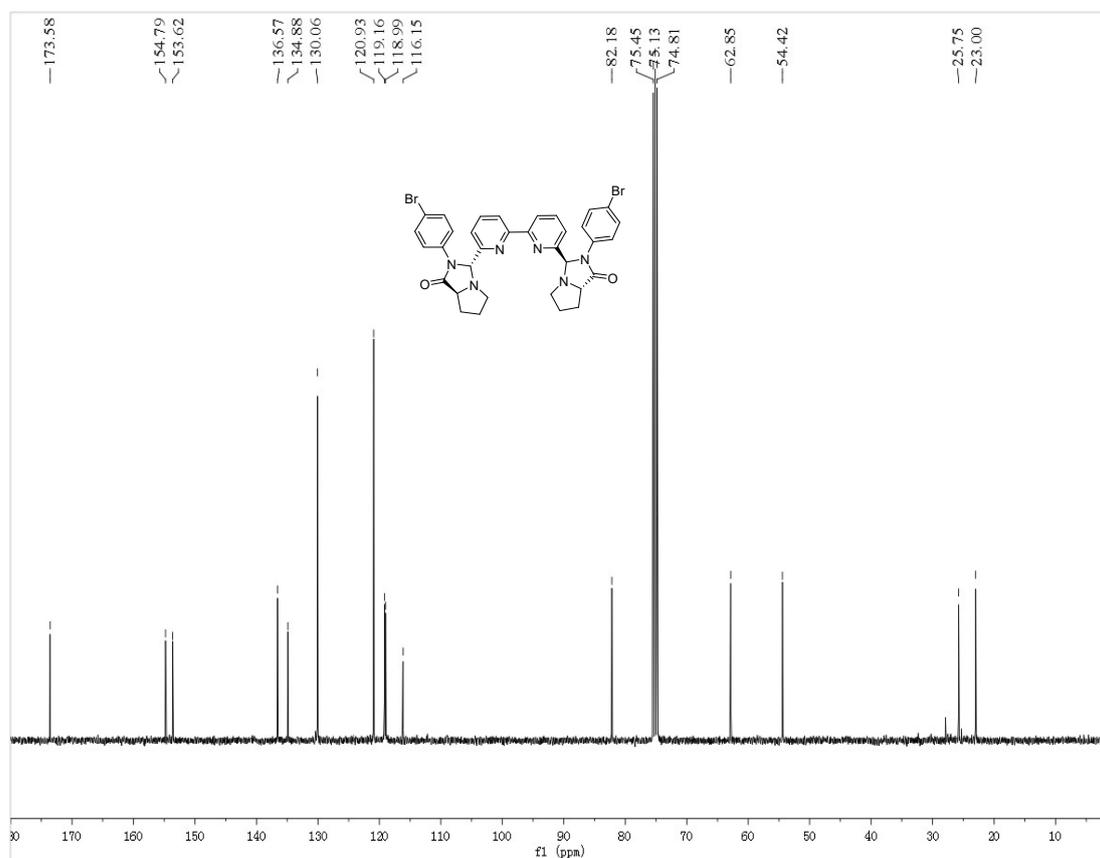
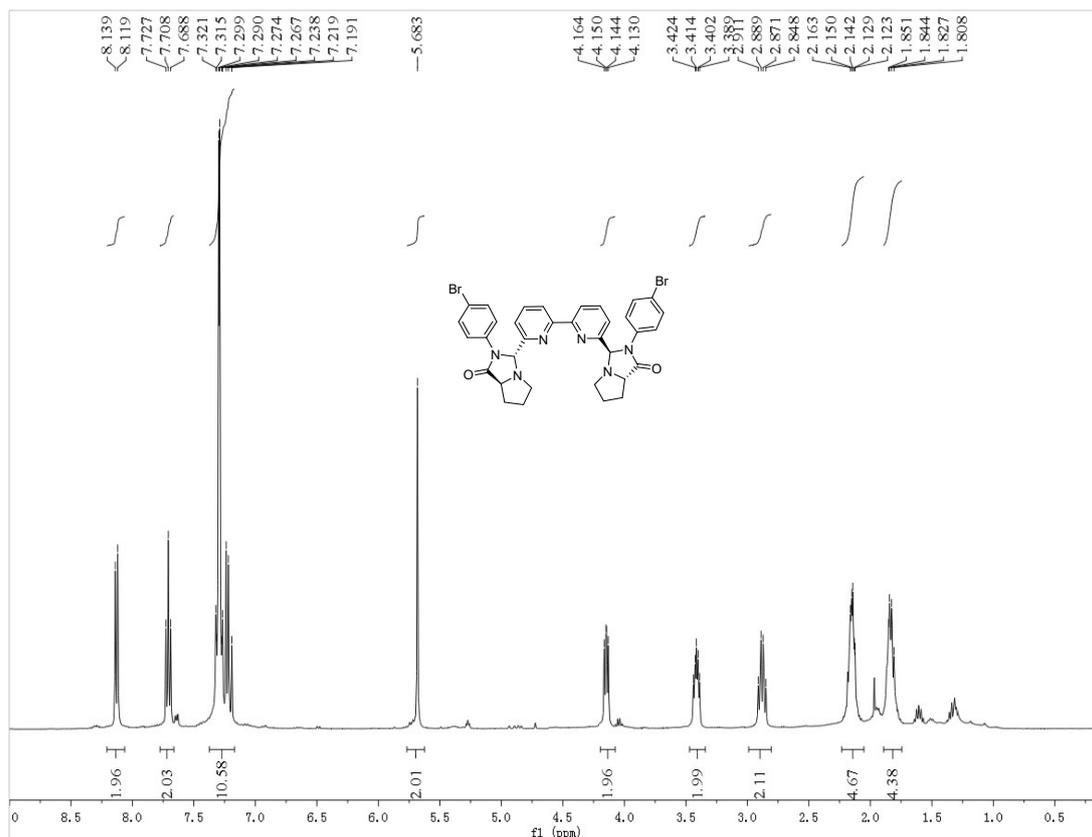
¹H and ¹³C NMR of L1o



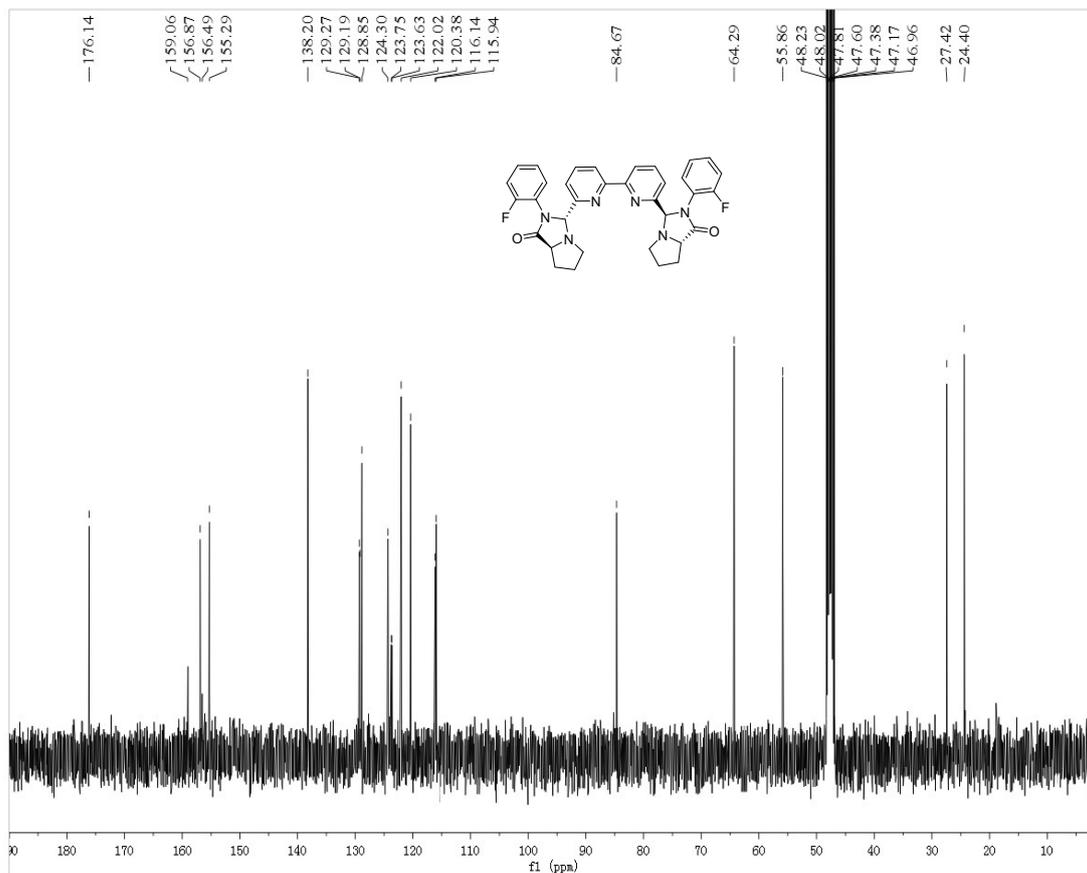
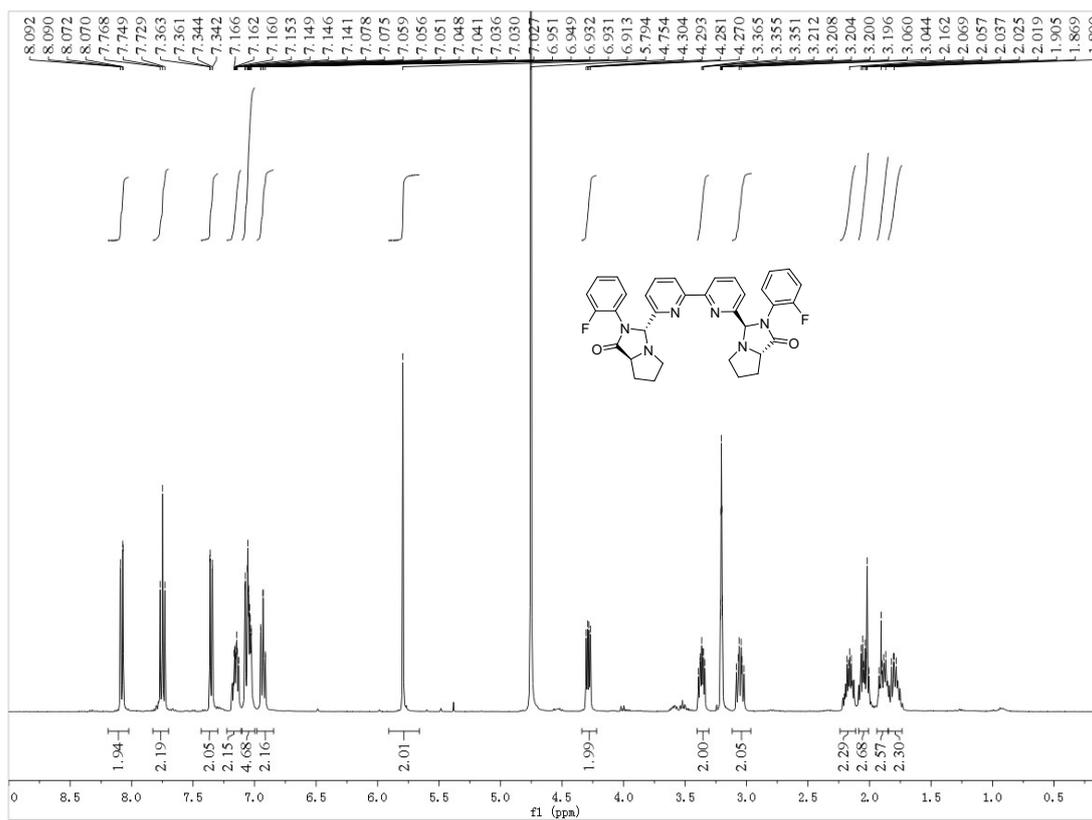
¹H and ¹³C NMR of L1p



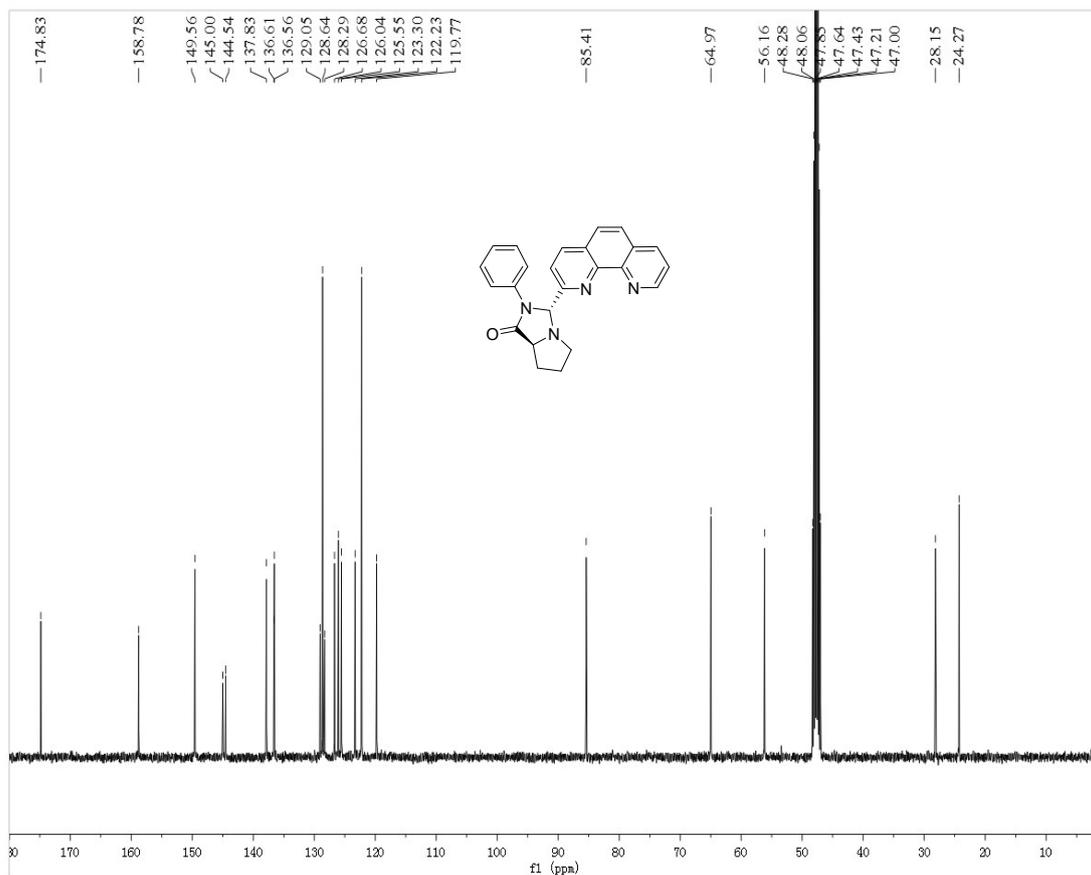
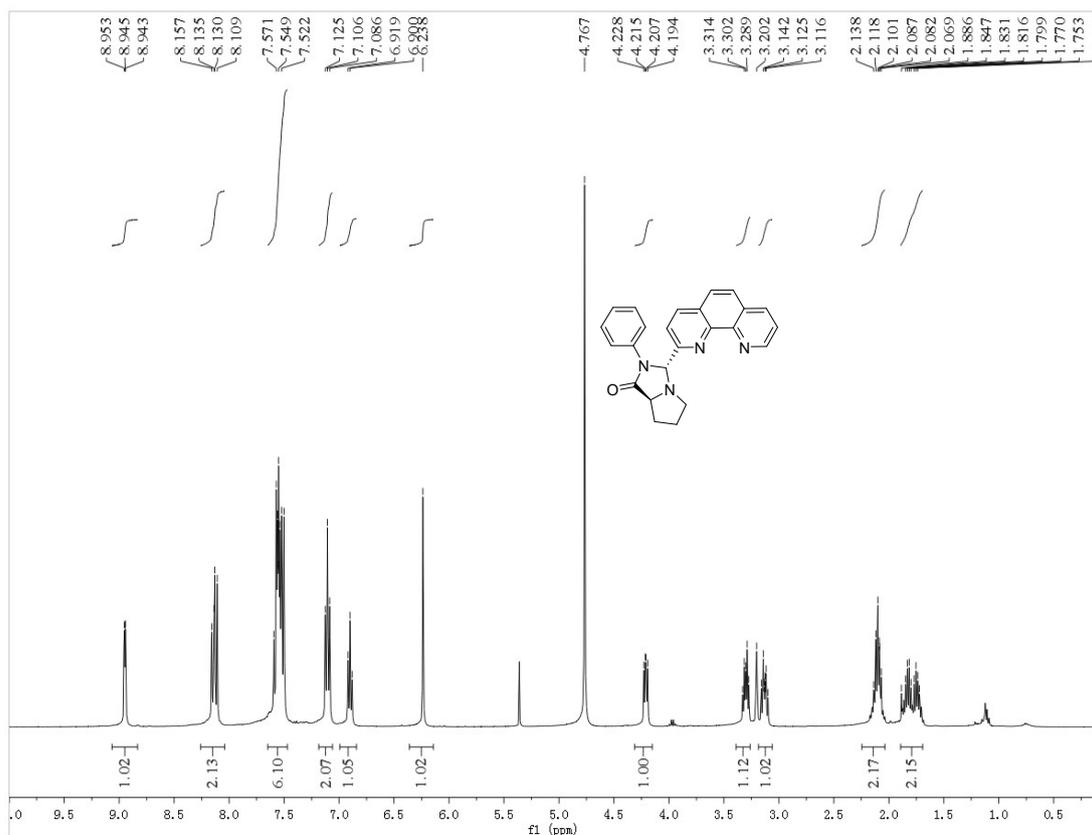
¹H and ¹³C NMR of L1q



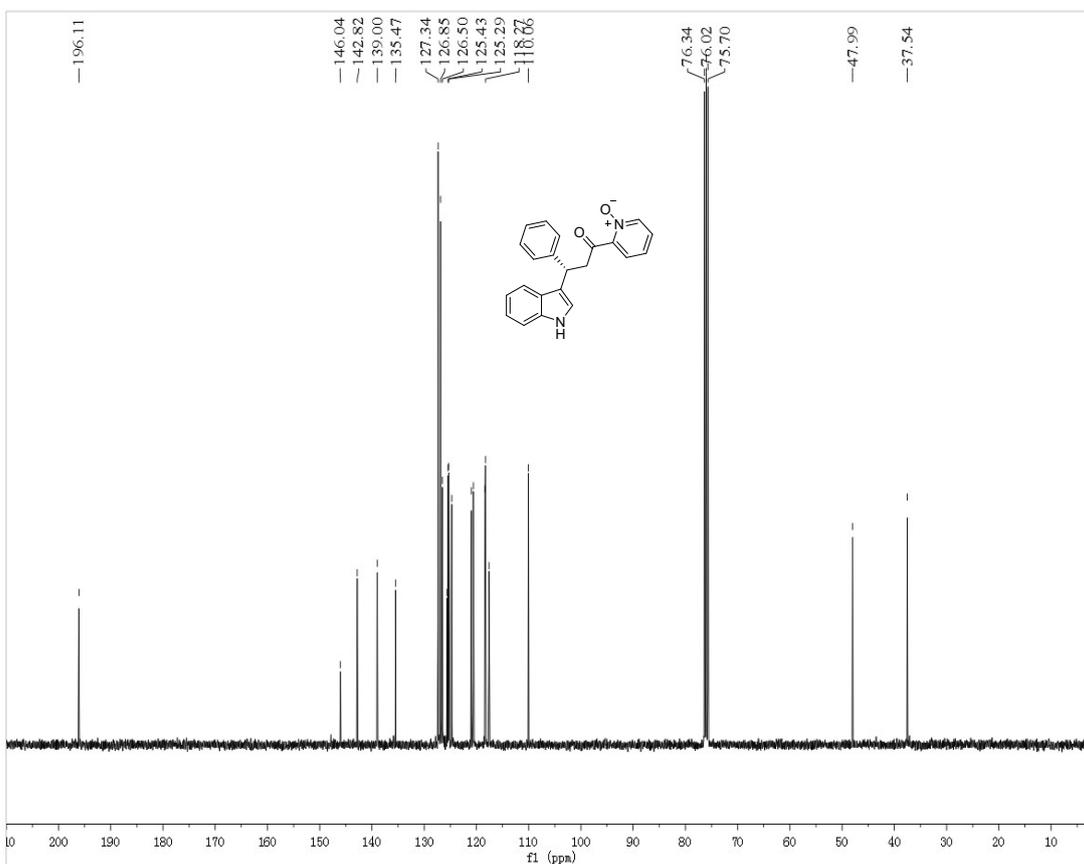
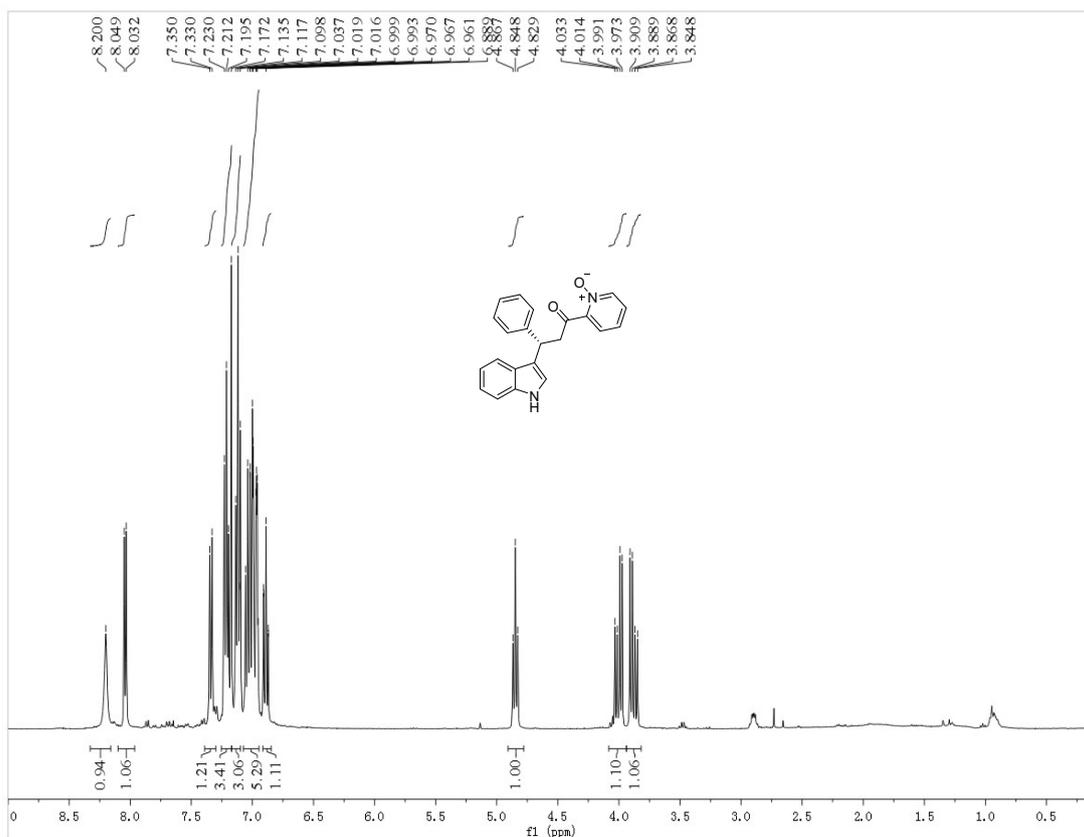
¹H and ¹³C NMR of L1r



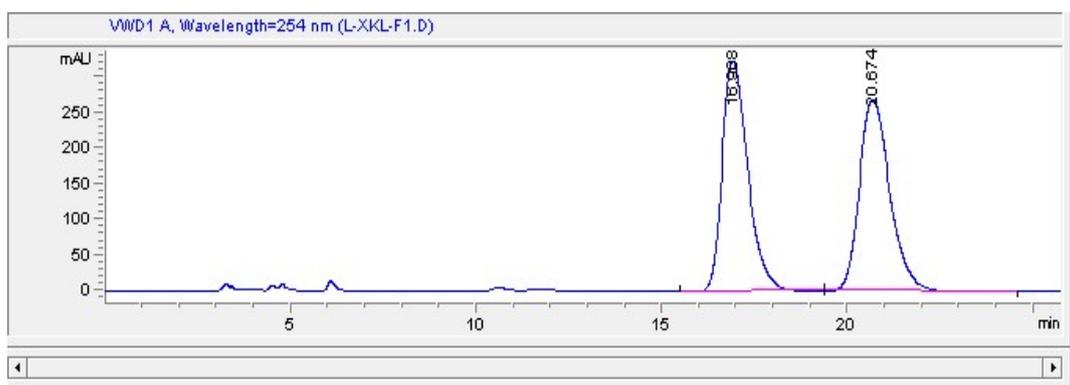
¹H and ¹³C NMR of L5a



¹H and ¹³C NMR of 6aa



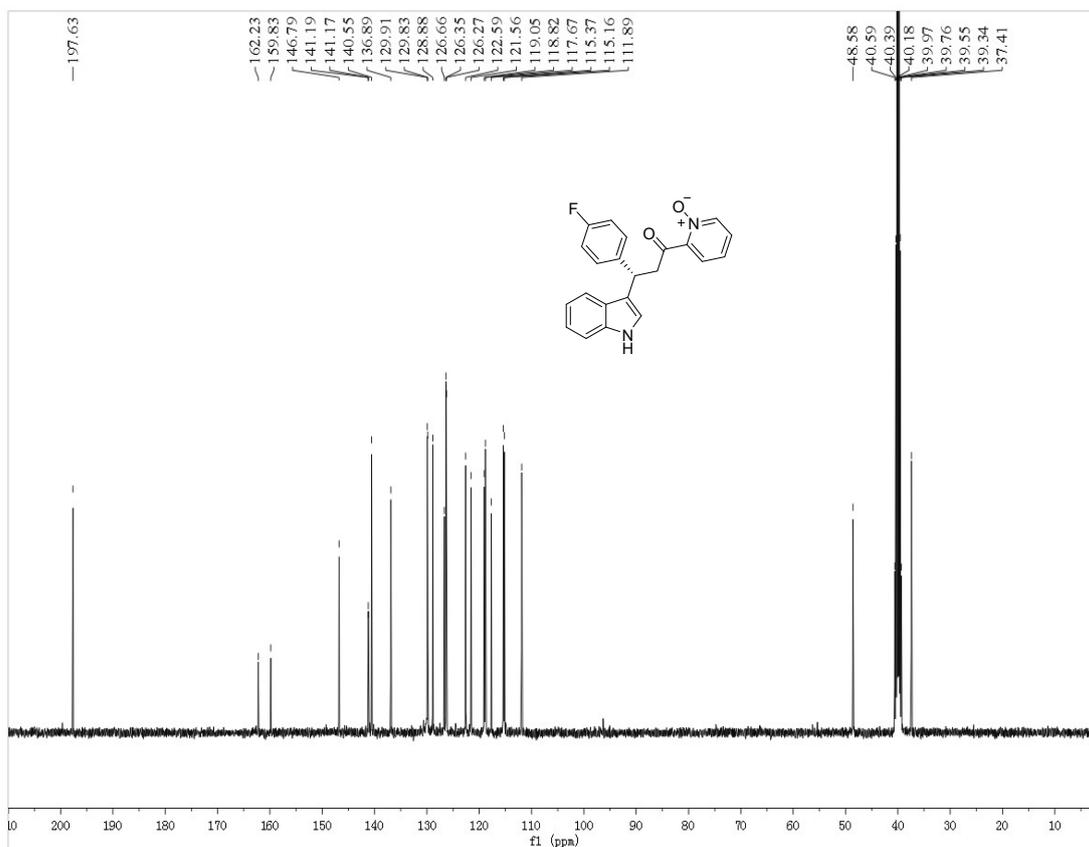
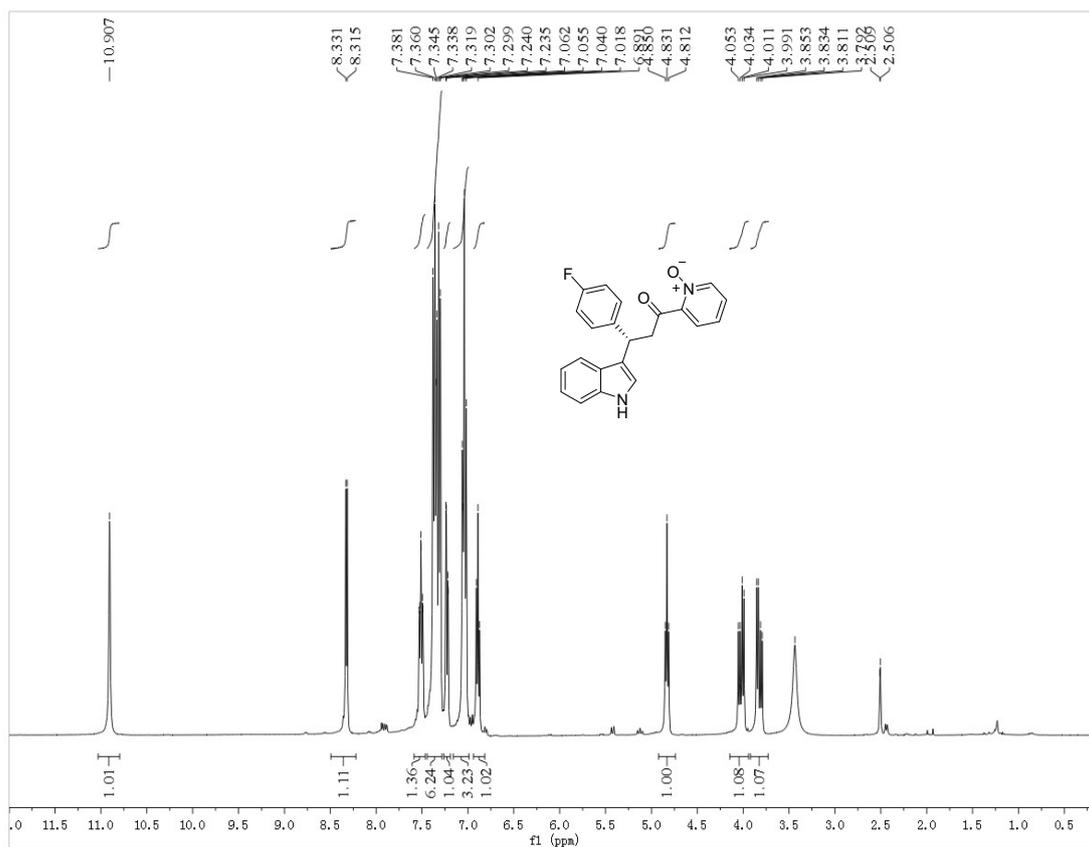
HPLC of 6aa



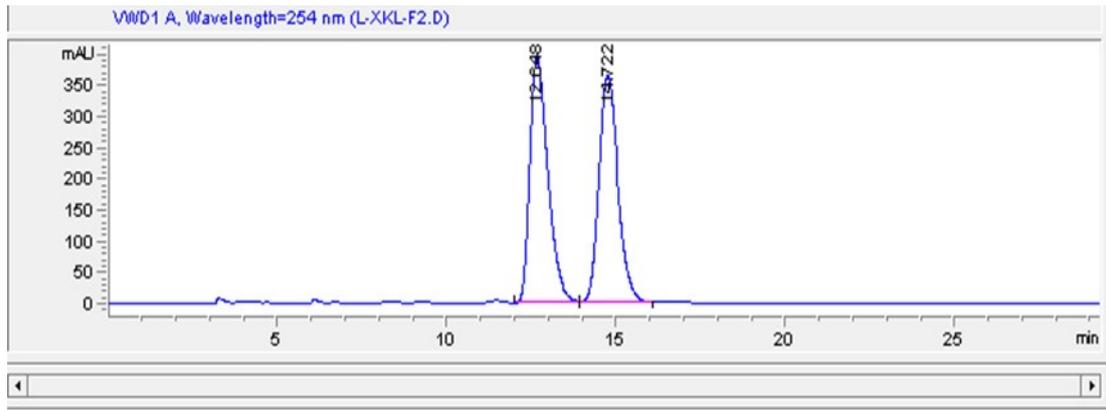
| # | Time | Area | Height | Width | Area% | Symmetry |
|---|--------|---------|--------|--------|--------|----------|
| 1 | 16.908 | 15469.6 | 322.8 | 0.7311 | 50.175 | 0.684 |
| 2 | 20.674 | 15361.4 | 269 | 0.8823 | 49.825 | 0.746 |



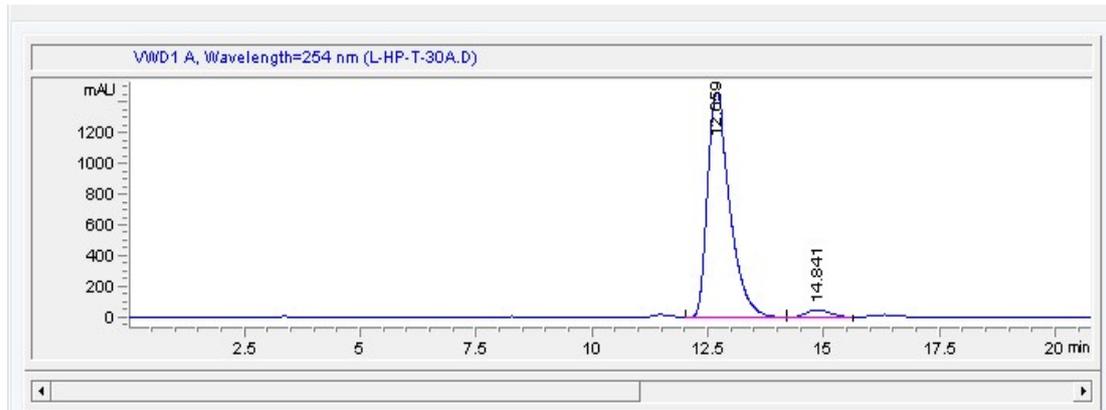
¹H and ¹³C NMR of 6ab



HPLC of 6ab

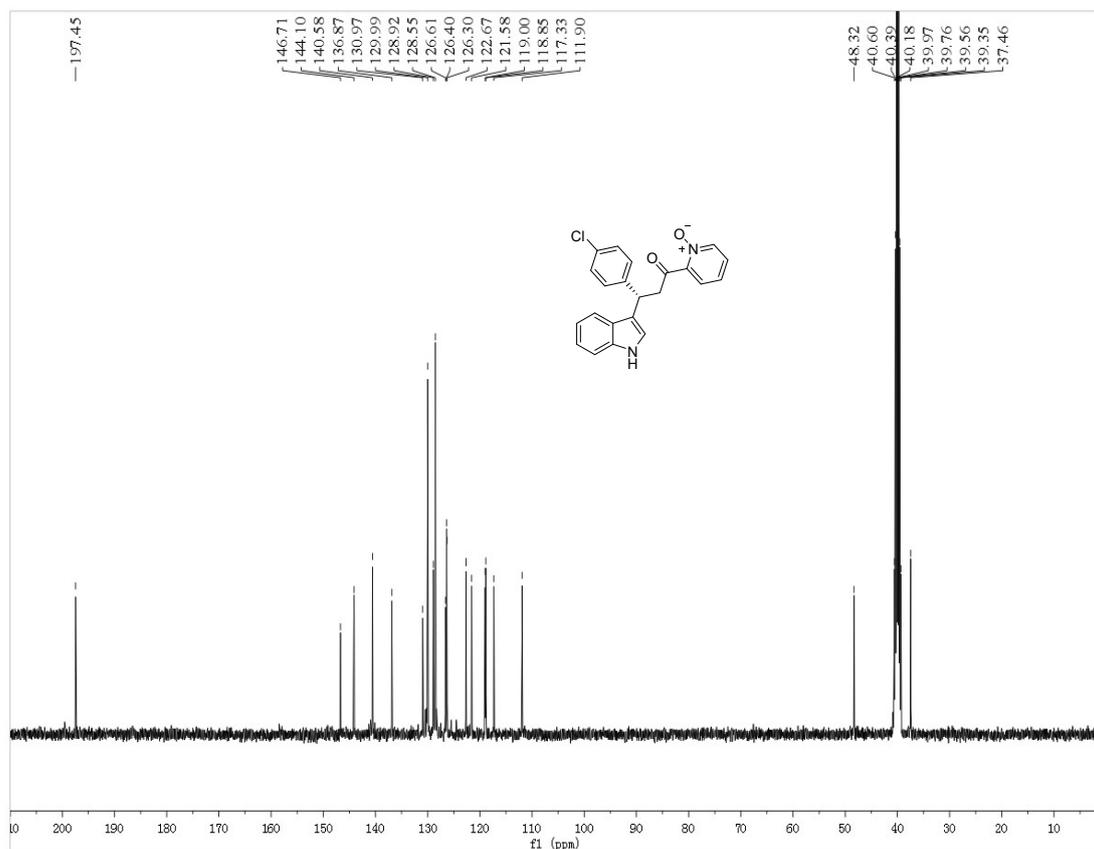
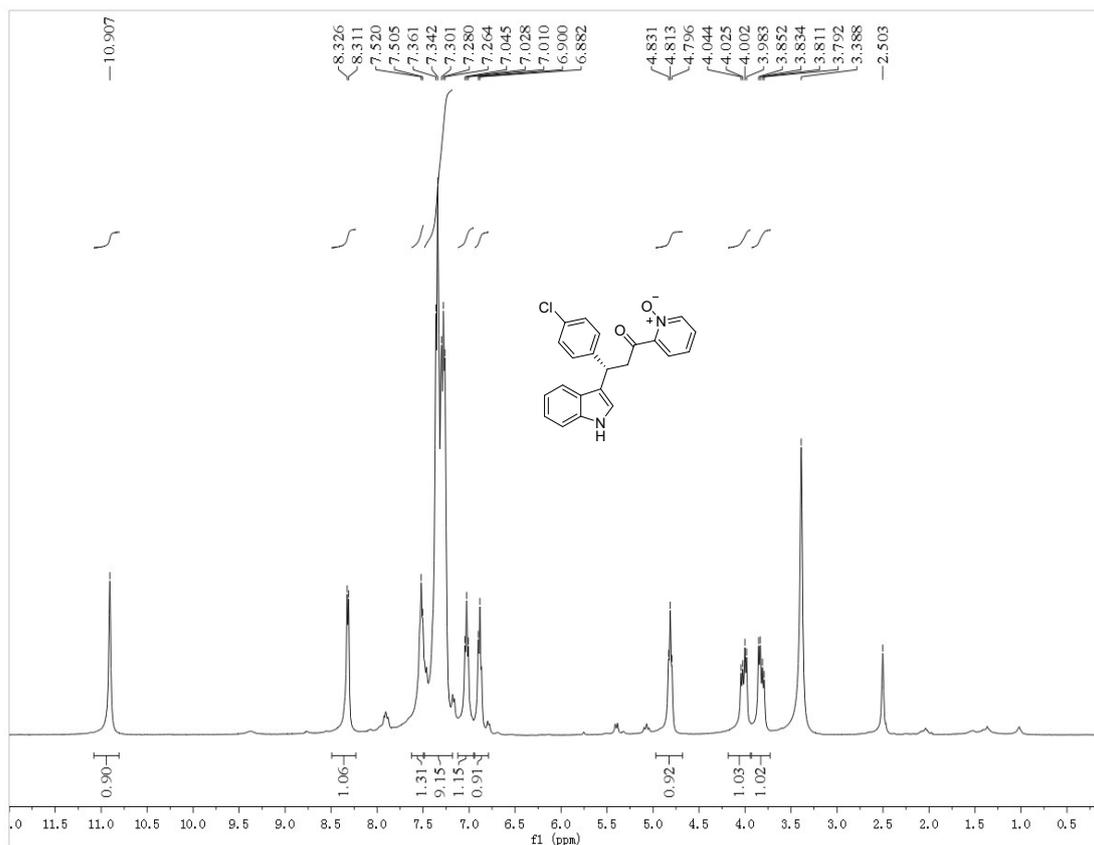


| # | Time | Area | Height | Width | Area% | Symmetry |
|---|--------|---------|--------|--------|--------|----------|
| 1 | 12.648 | 14400.3 | 396.7 | 0.5545 | 49.968 | 0.617 |
| 2 | 14.722 | 14418.9 | 365.4 | 0.6134 | 50.032 | 0.759 |

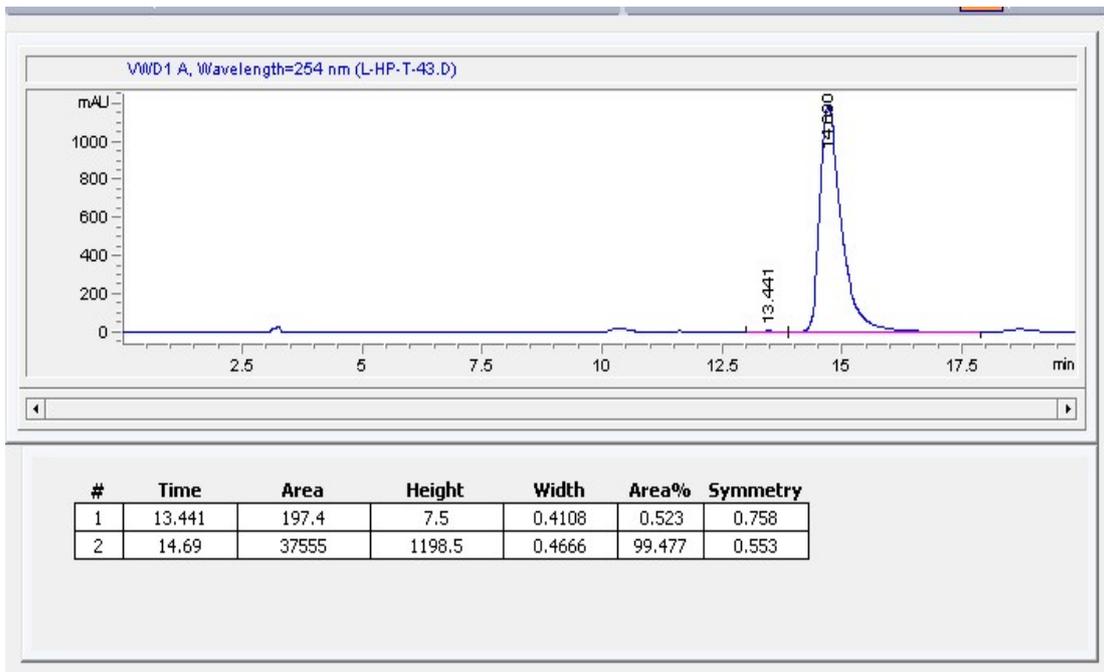
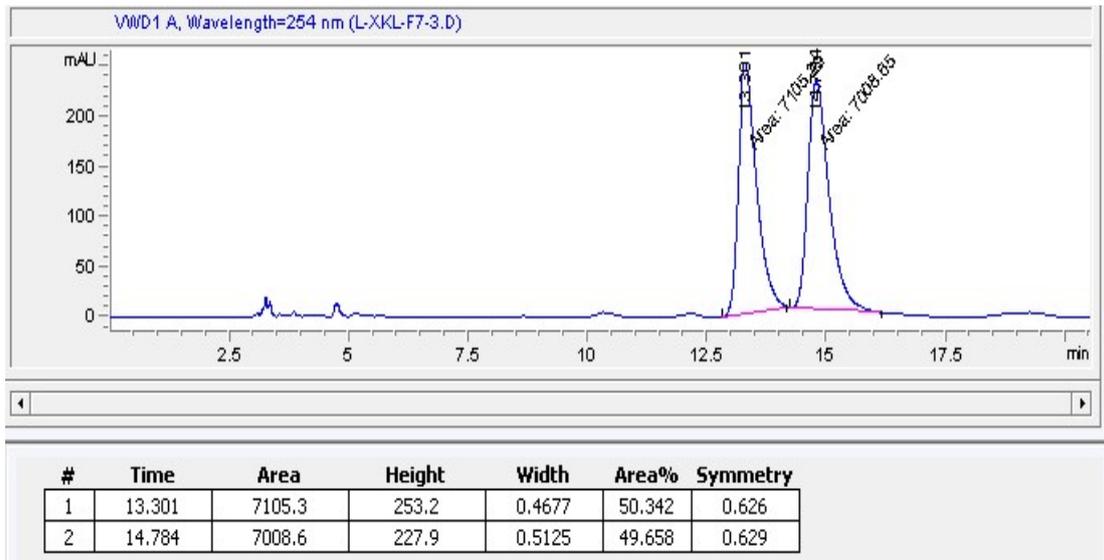


| # | Time | Area | Height | Width | Area% | Symmetry |
|---|--------|---------|--------|--------|--------|----------|
| 1 | 12.659 | 50583.1 | 1469.2 | 0.5257 | 96.257 | 0.603 |
| 2 | 14.841 | 1967.1 | 51.6 | 0.5889 | 3.743 | 0.866 |

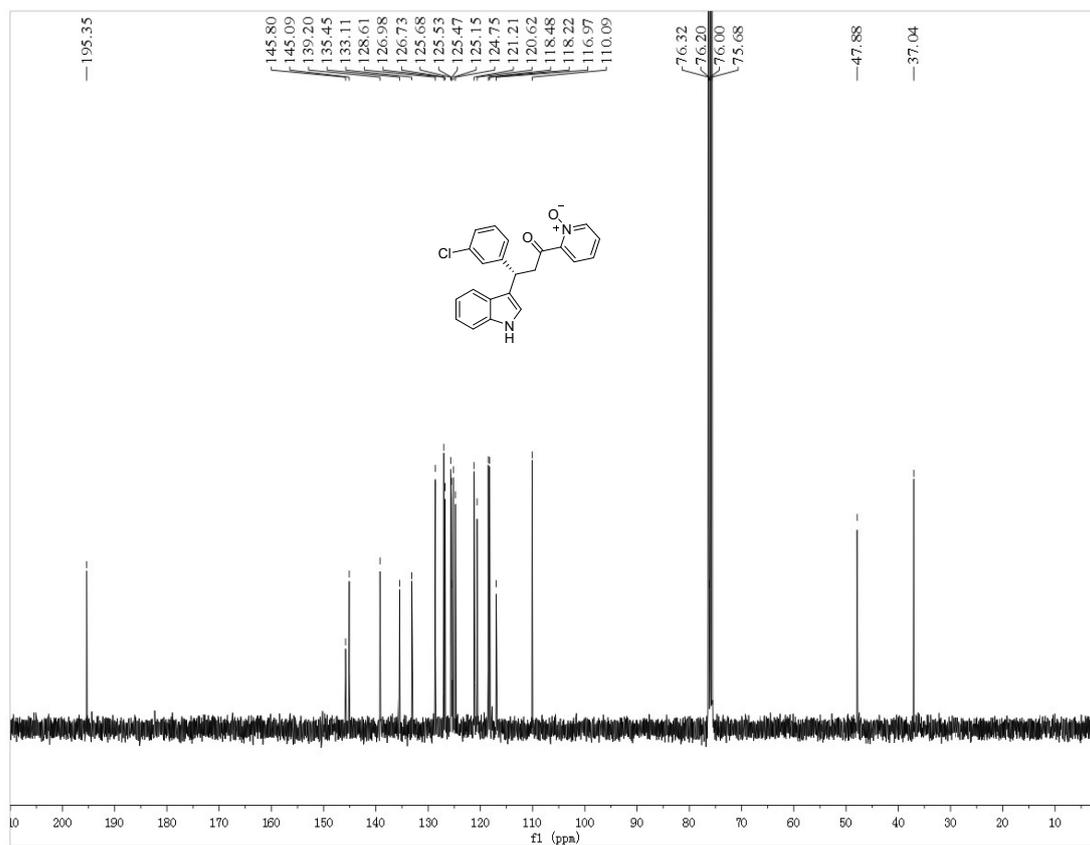
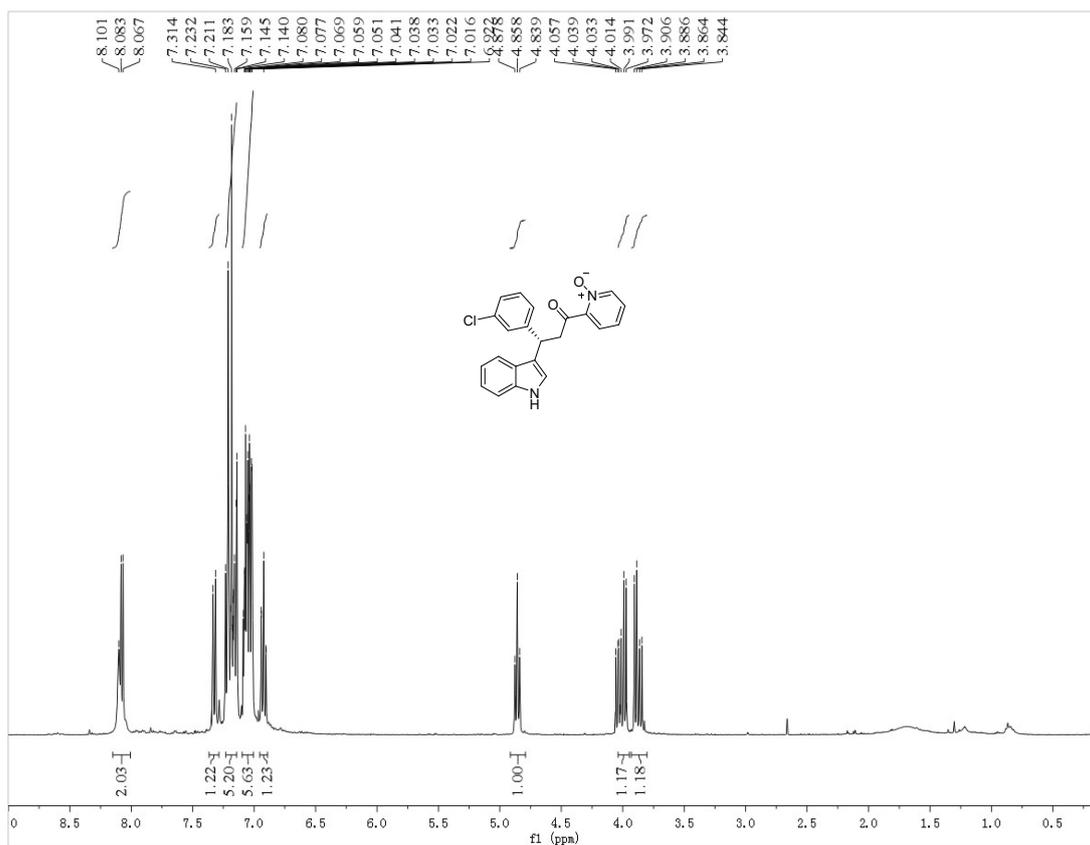
^1H and ^{13}C NMR of 6ac



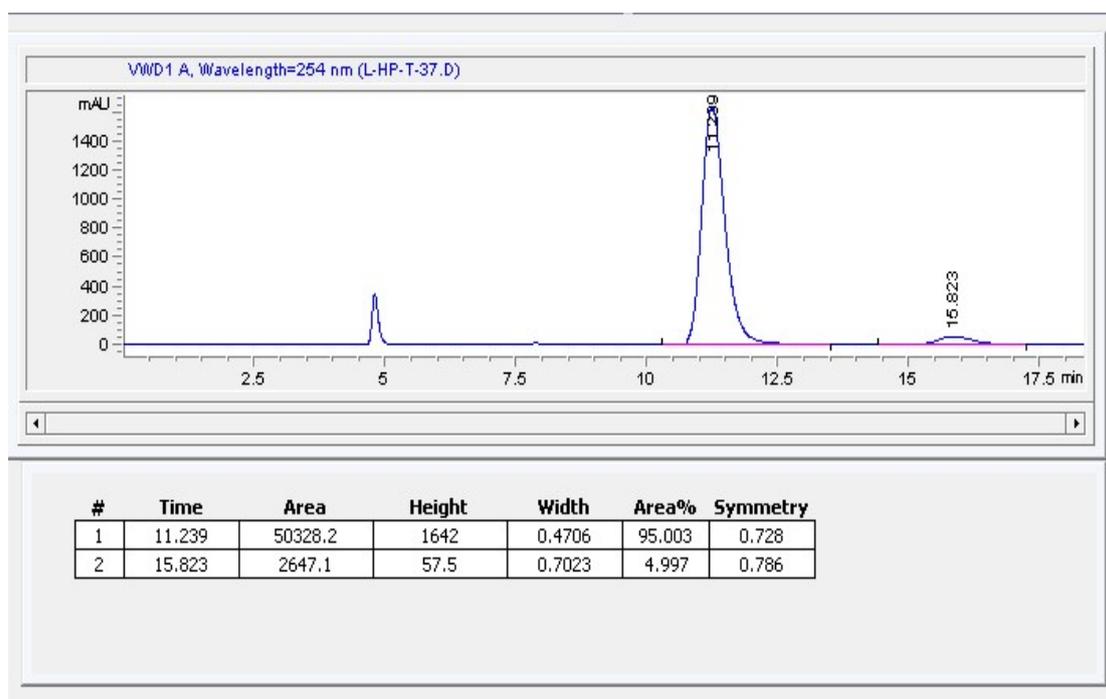
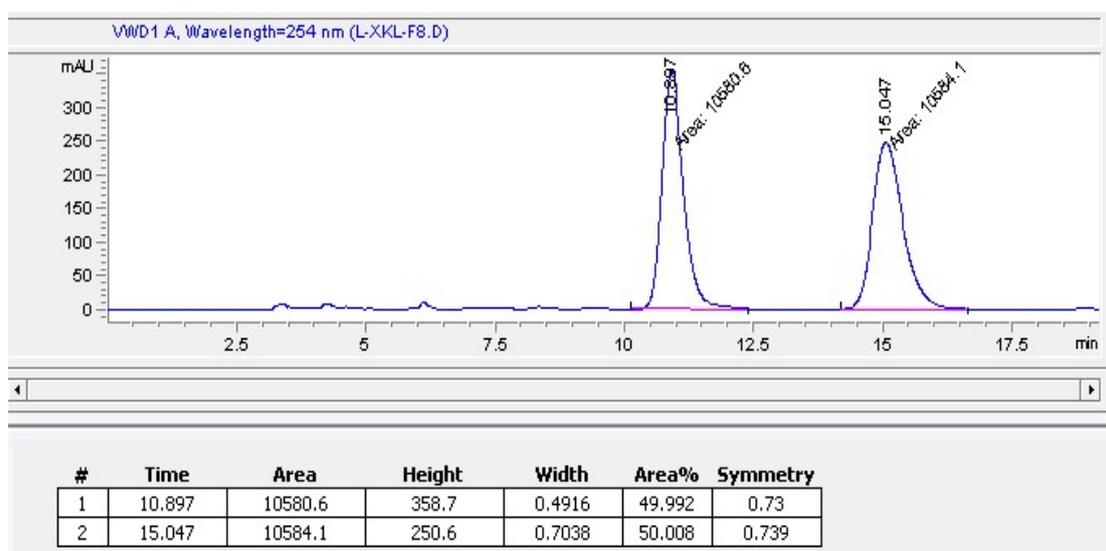
HPLC of 6ac



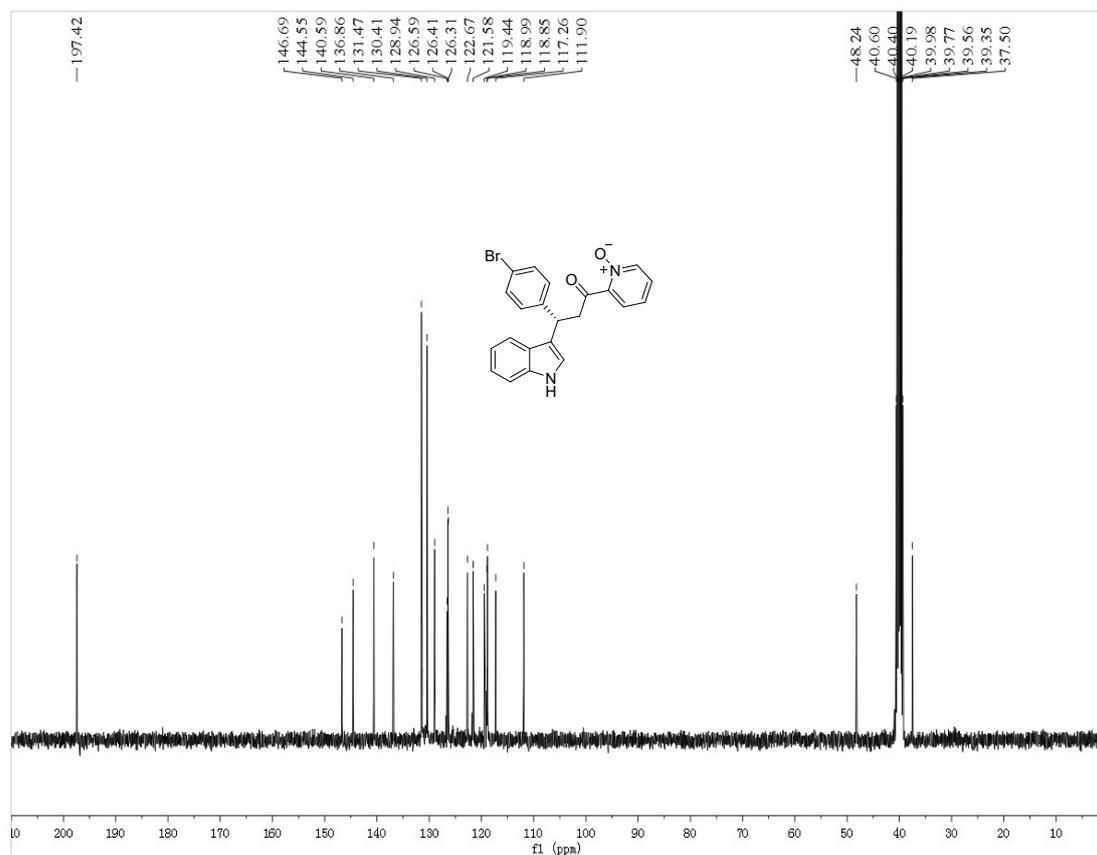
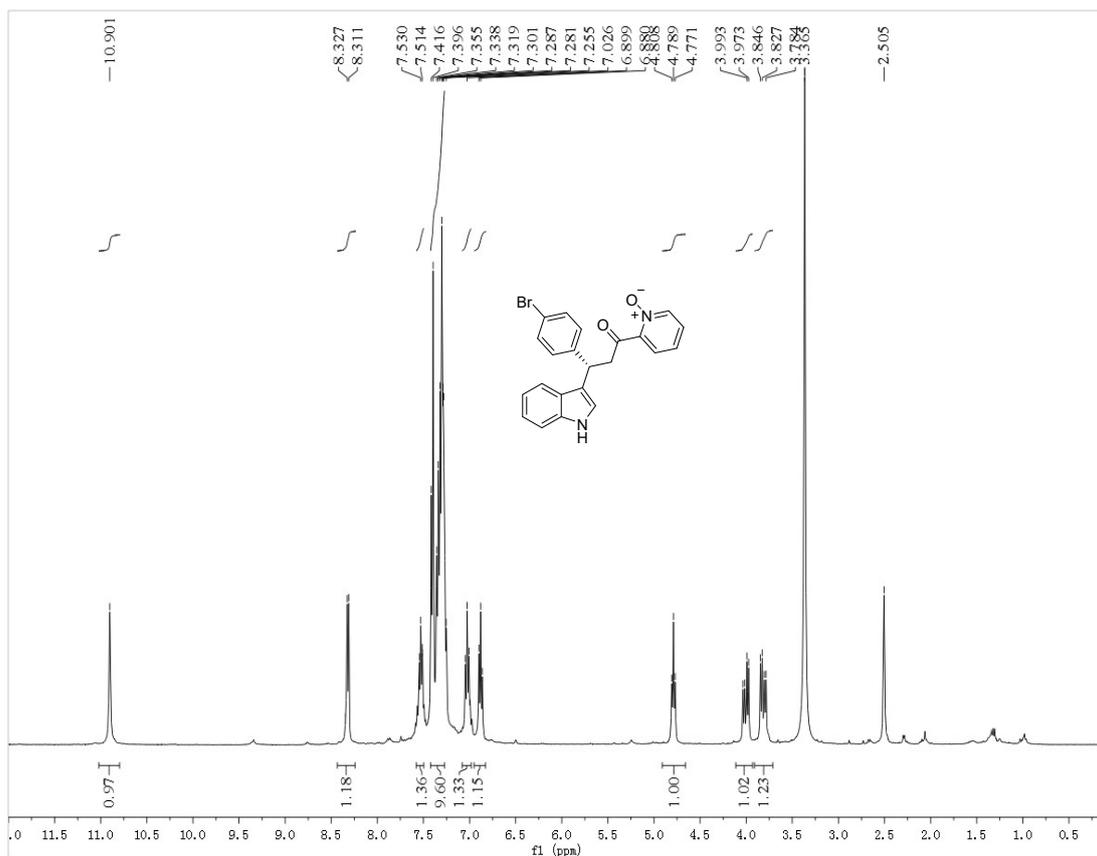
¹H and ¹³C NMR of 6ad



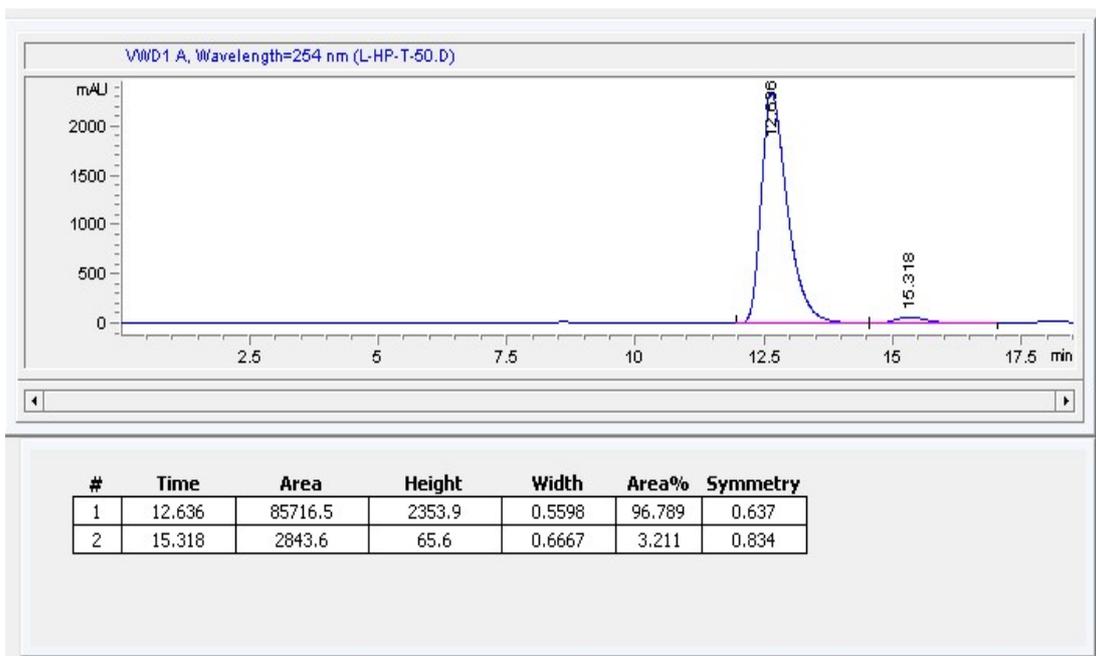
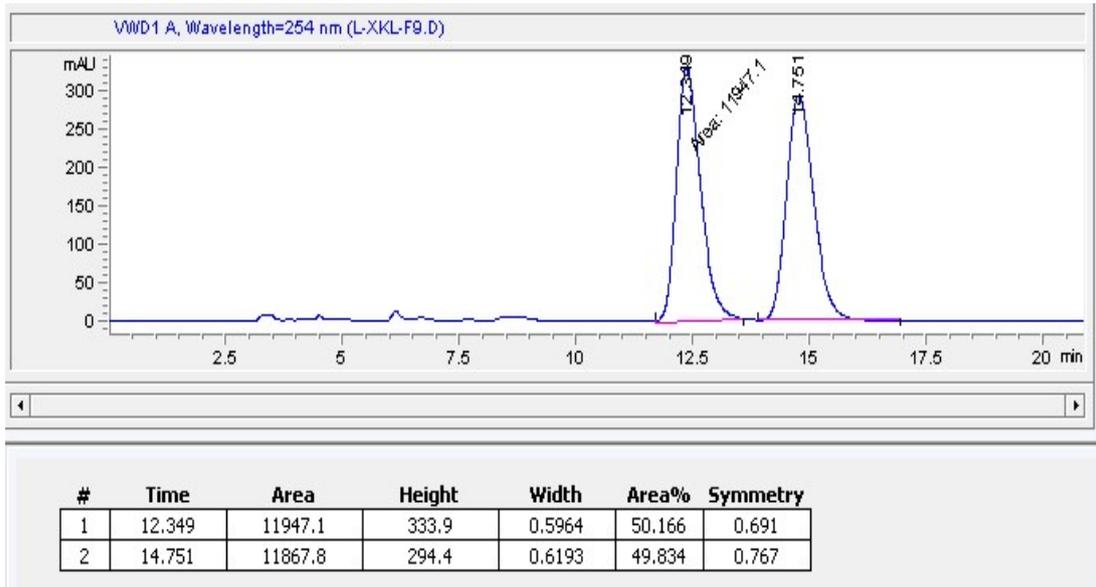
HPLC of 6ad



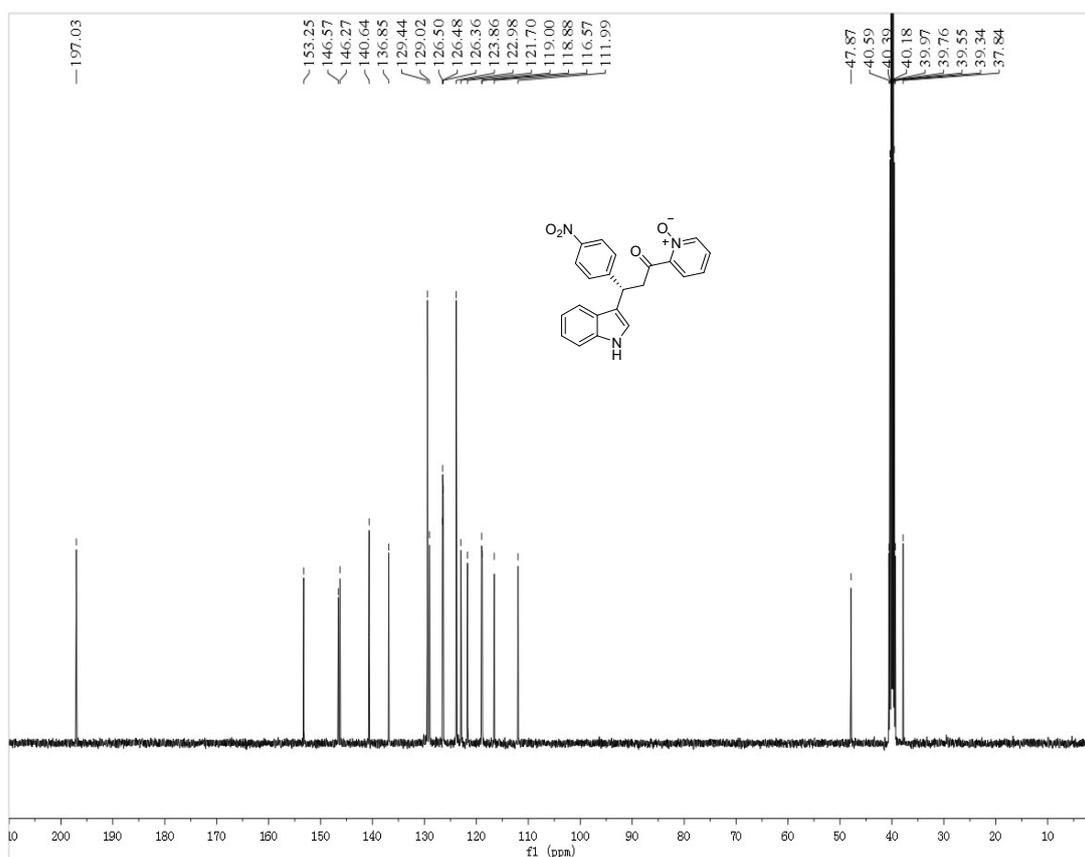
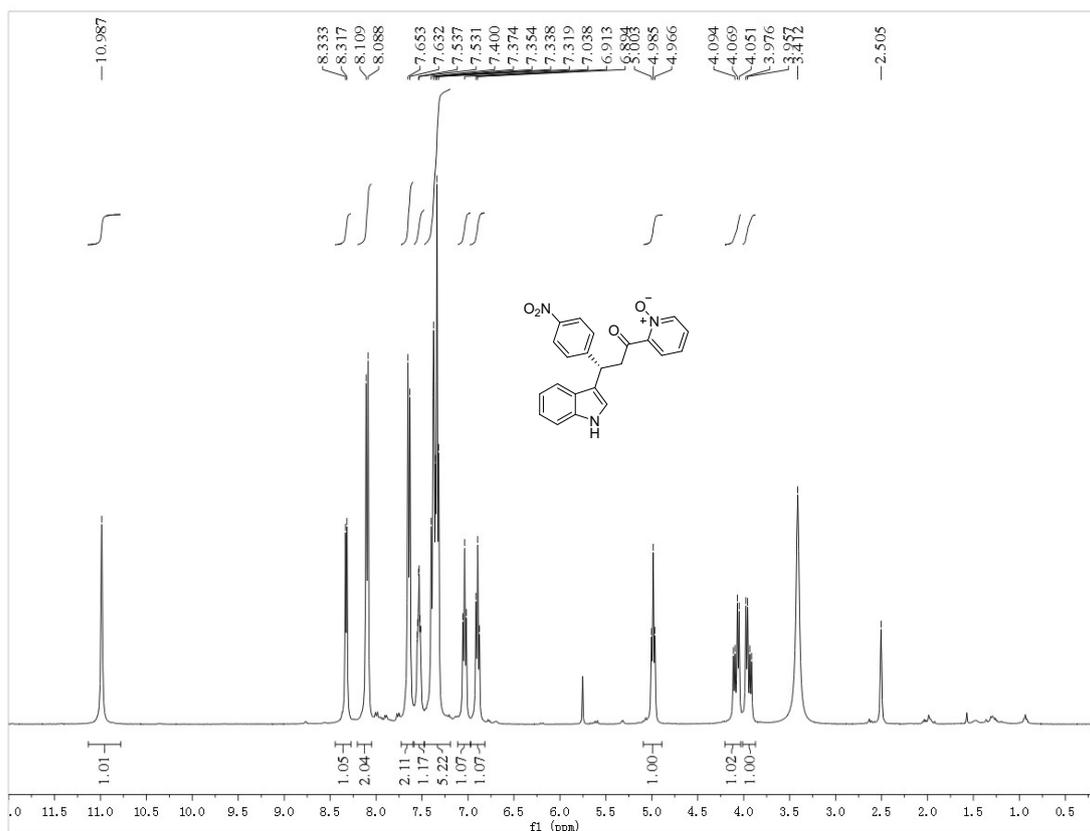
¹H and ¹³C NMR of 6ae



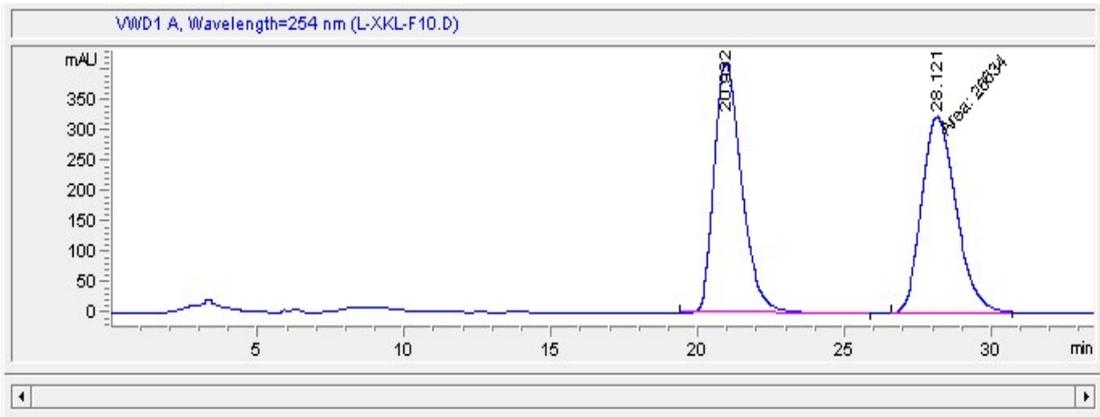
HPLC of 6ae



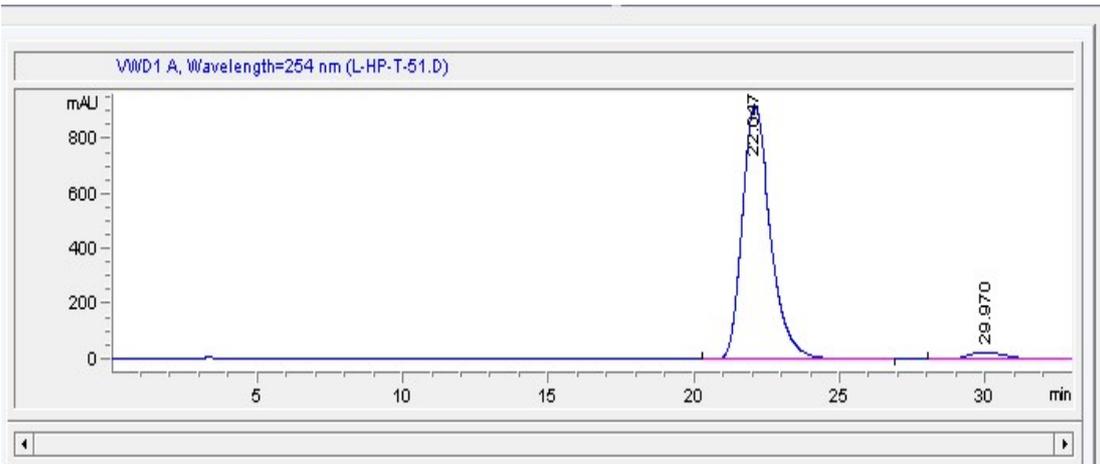
¹H and ¹³C NMR of 6af



HPLC of 6af

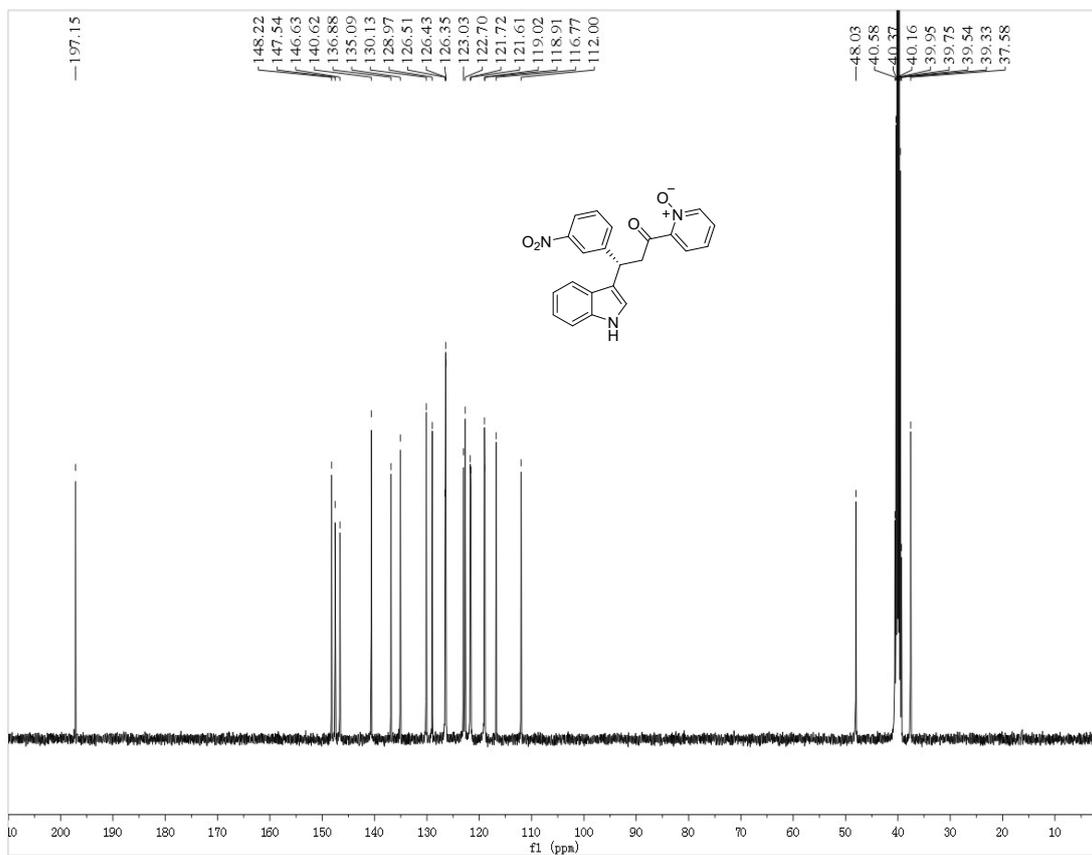
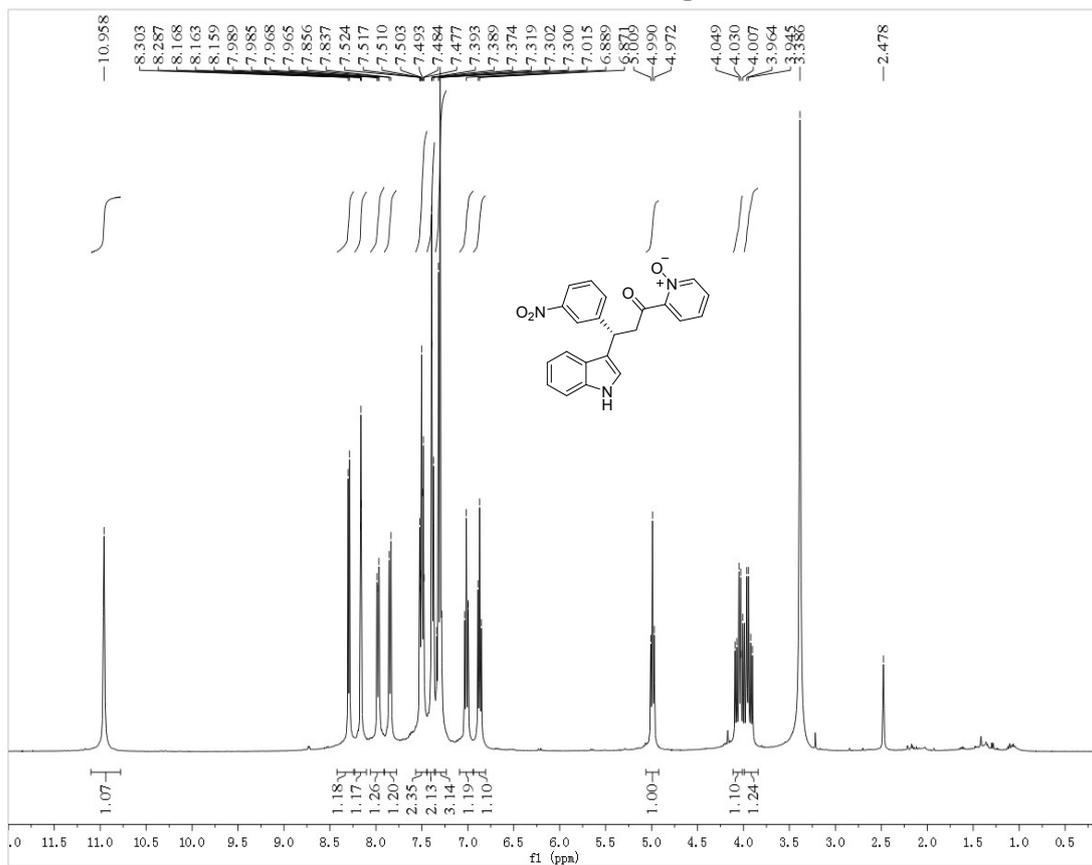


| # | Time | Area | Height | Width | Area% | Symmetry |
|---|--------|---------|--------|--------|--------|----------|
| 1 | 20.932 | 26457.1 | 412.8 | 0.9808 | 49.833 | 0.686 |
| 2 | 28.121 | 26634 | 324.6 | 1.3674 | 50.167 | 0.742 |

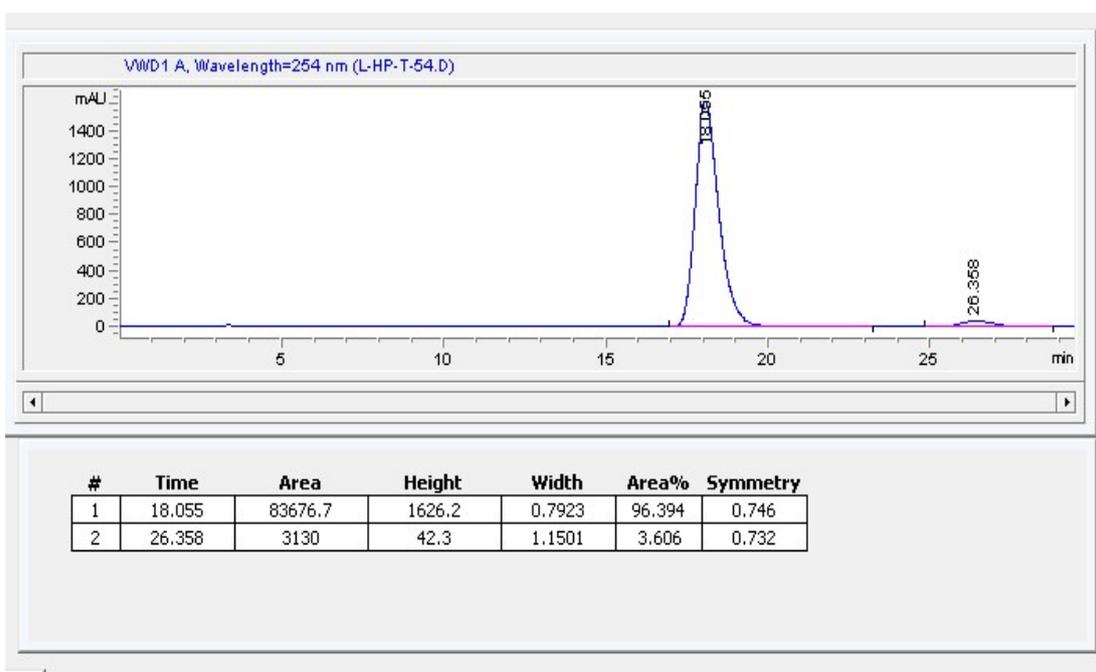
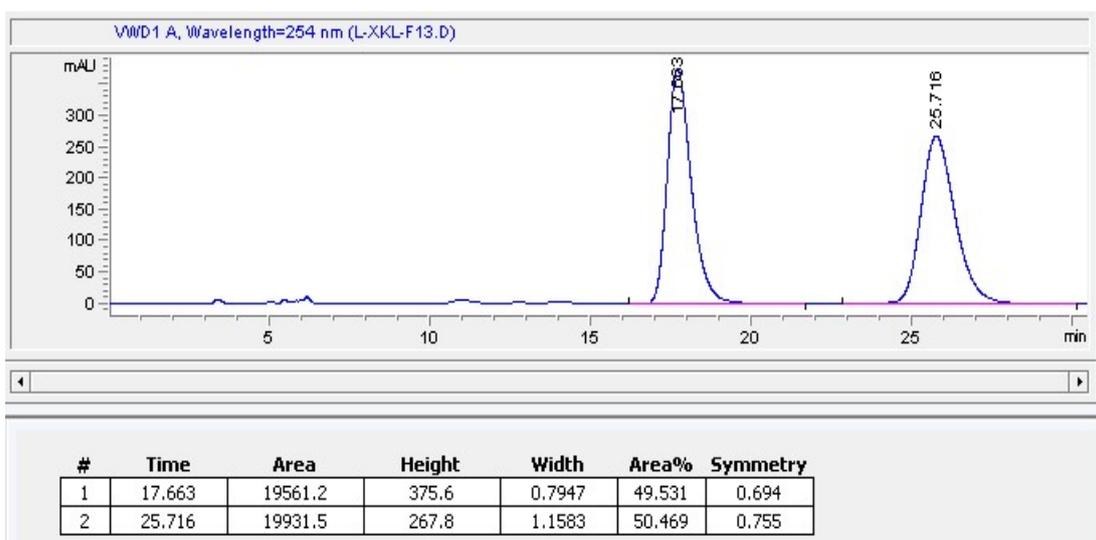


| # | Time | Area | Height | Width | Area% | Symmetry |
|---|--------|---------|--------|--------|--------|----------|
| 1 | 22.047 | 62850.2 | 921.9 | 1.0514 | 96.277 | 0.71 |
| 2 | 29.97 | 2430.7 | 27 | 1.3485 | 3.723 | 0.796 |

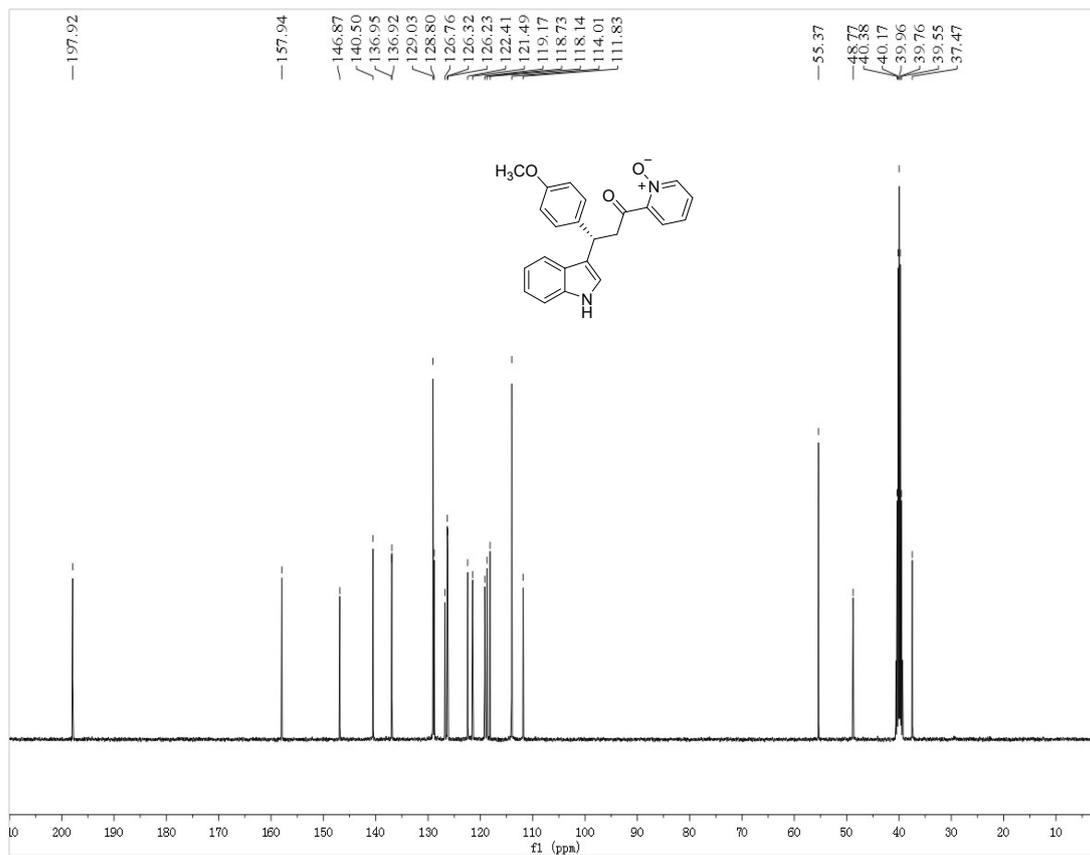
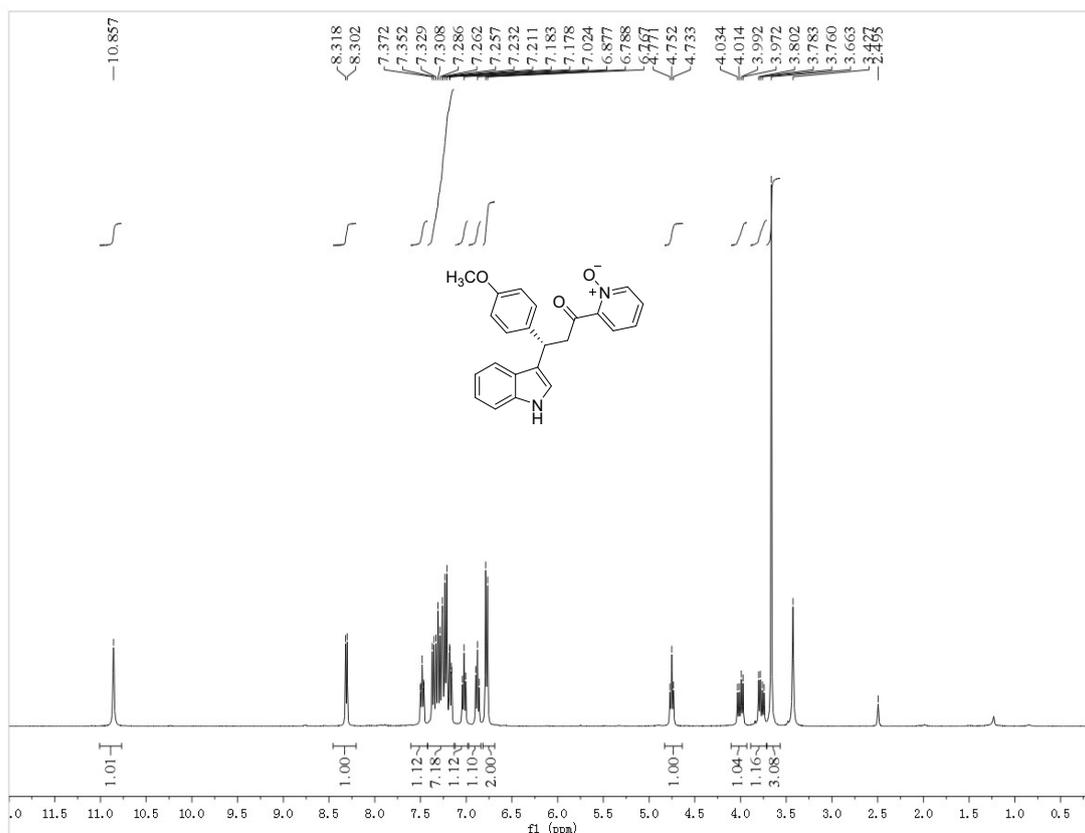
¹H and ¹³C NMR of 6ag



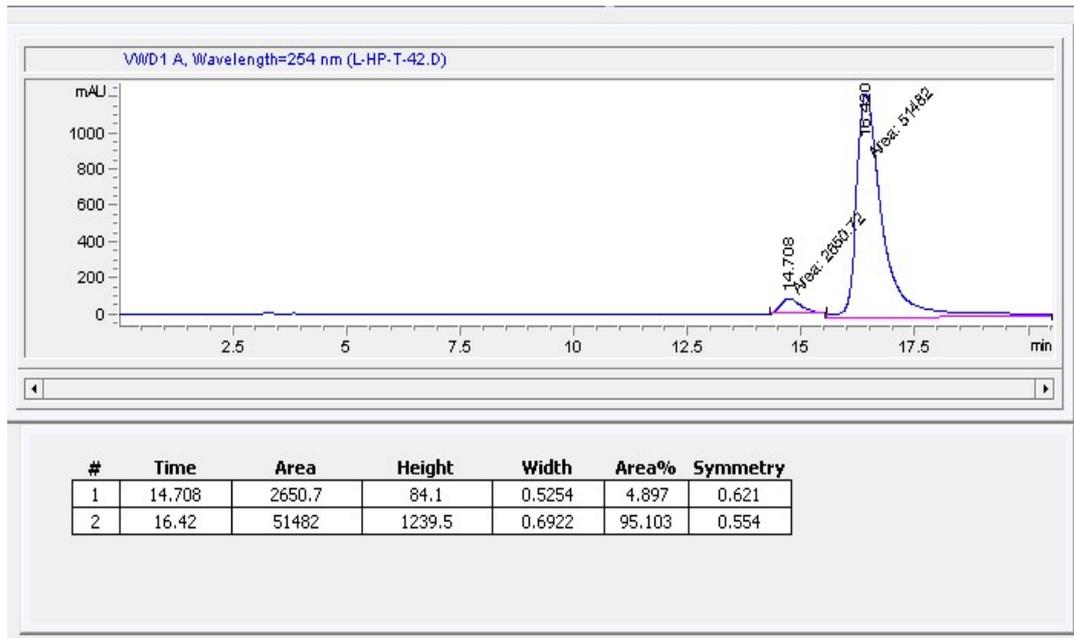
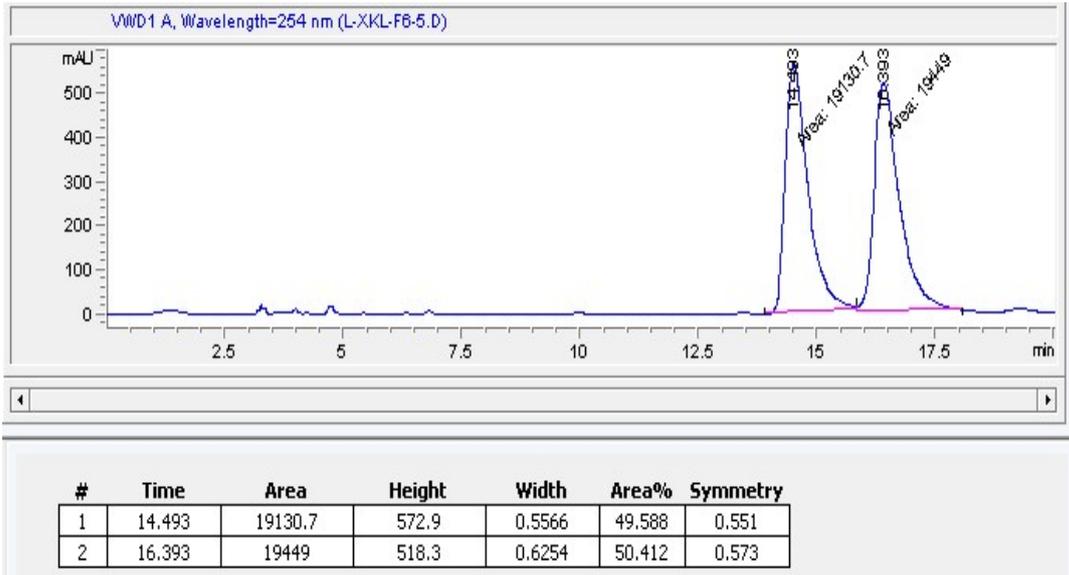
HPLC of 6ag



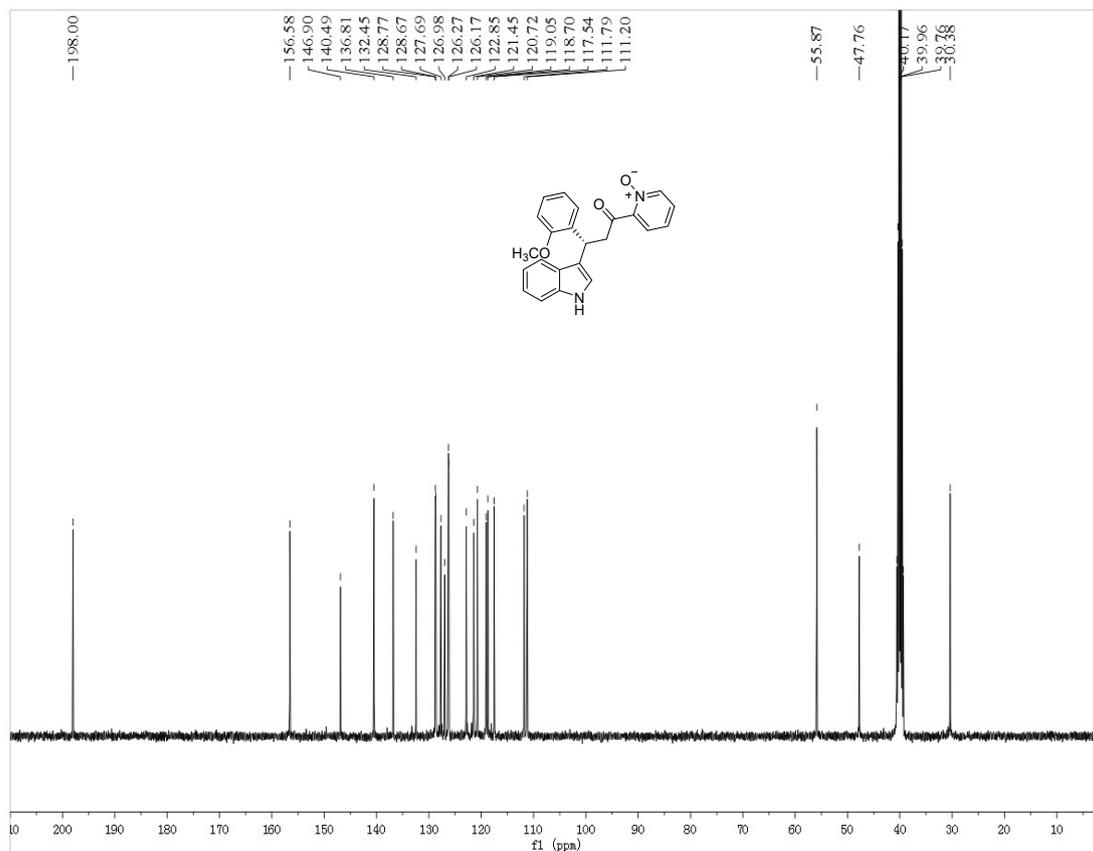
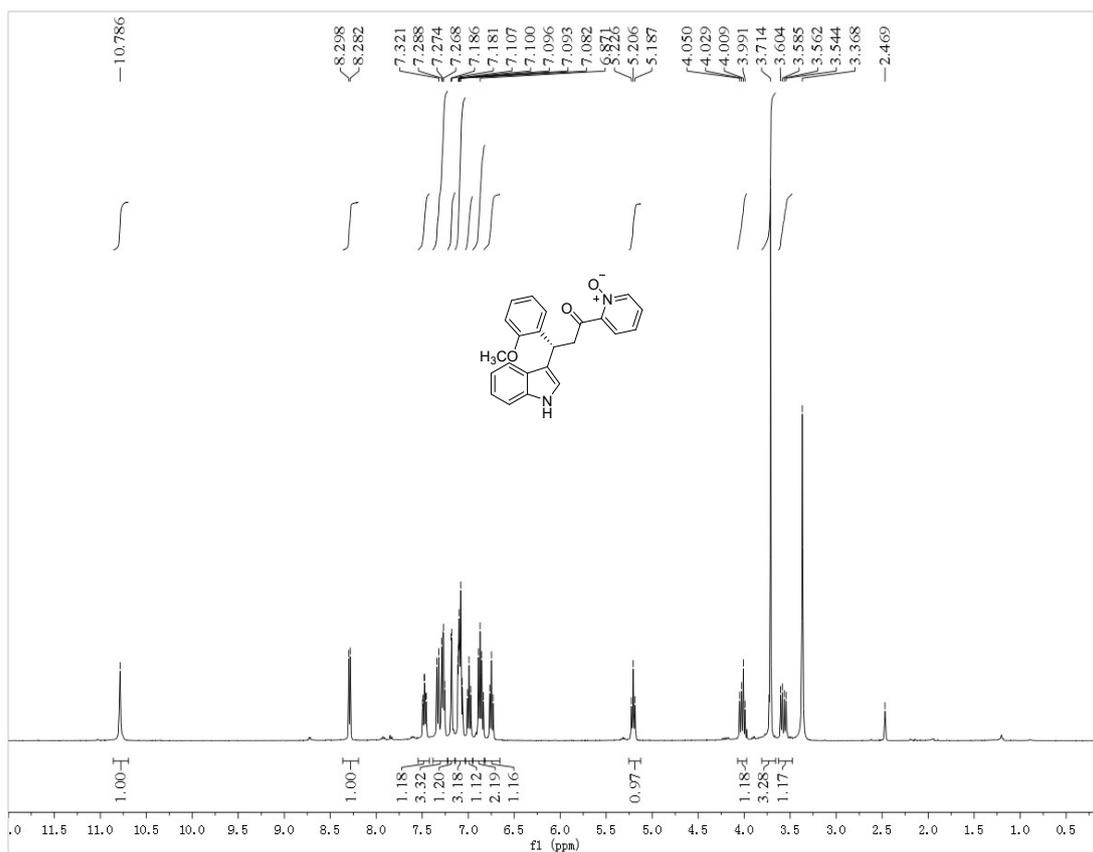
¹H and ¹³C NMR of 6ah



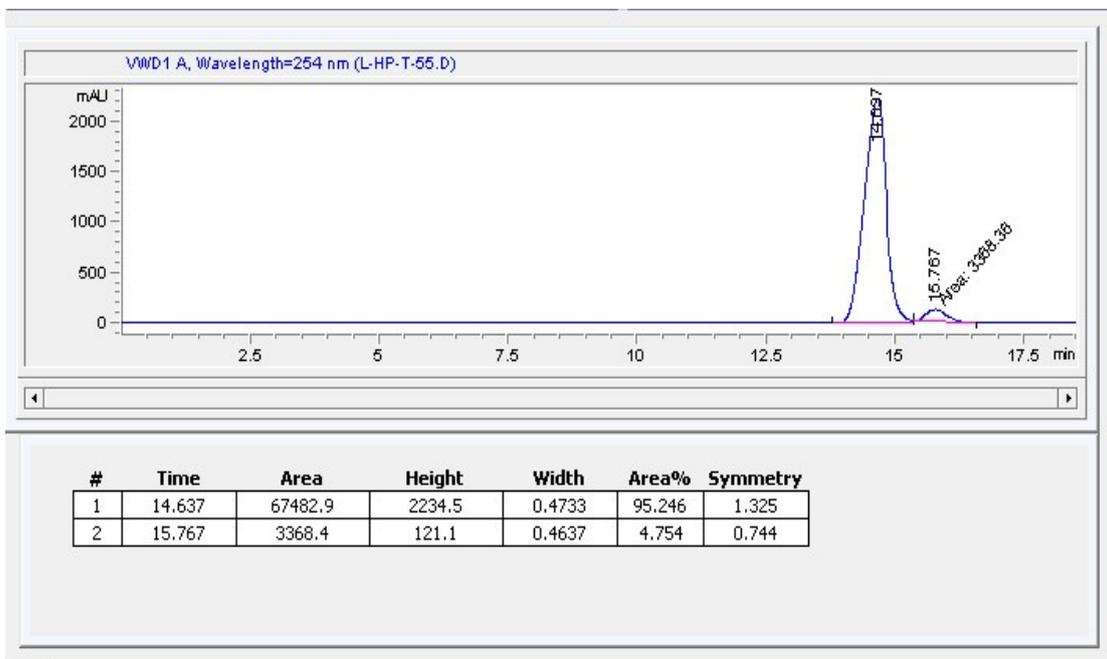
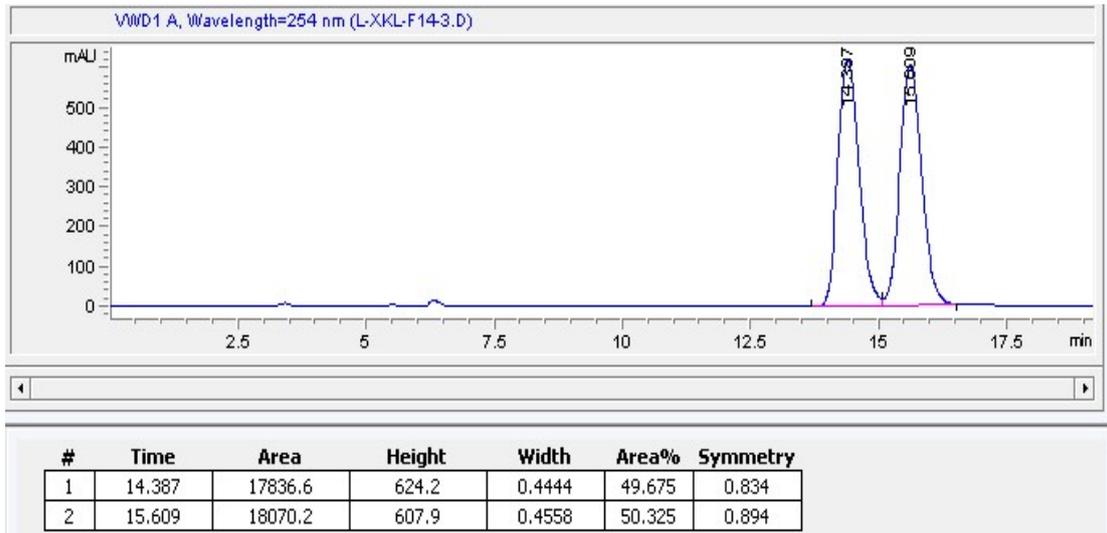
HPLC of 6ah



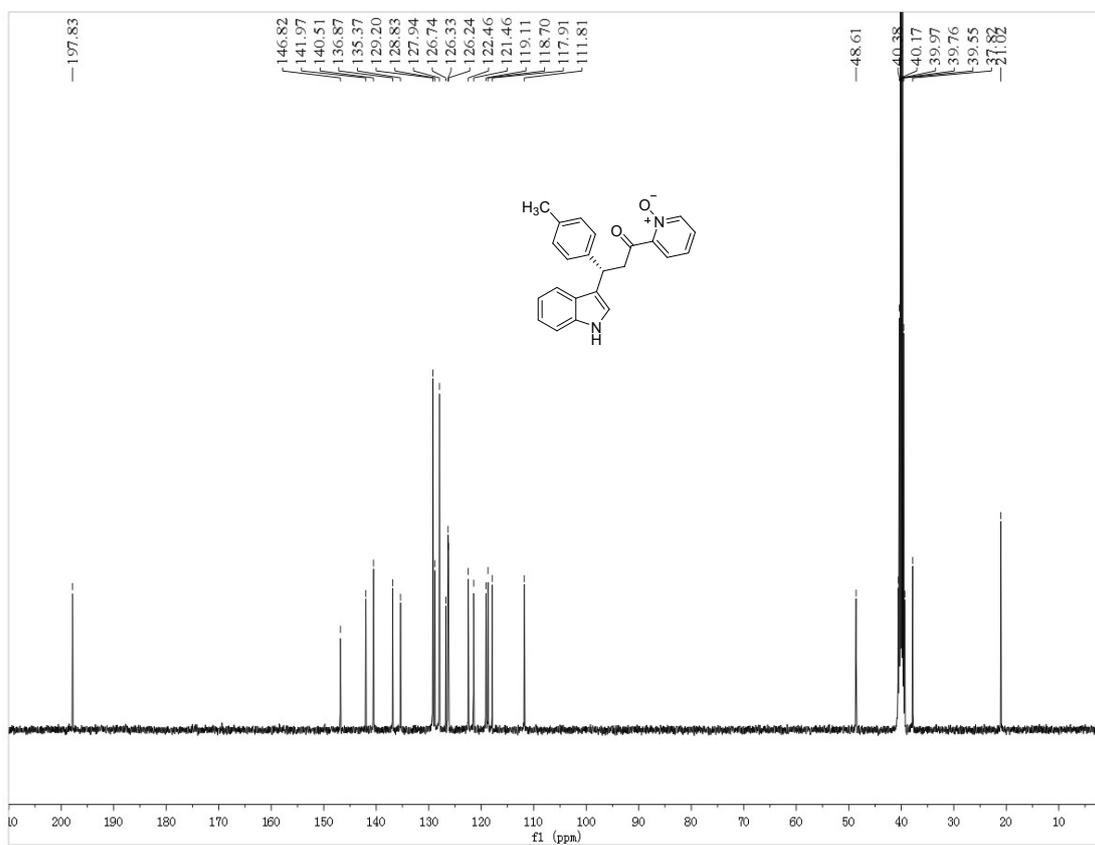
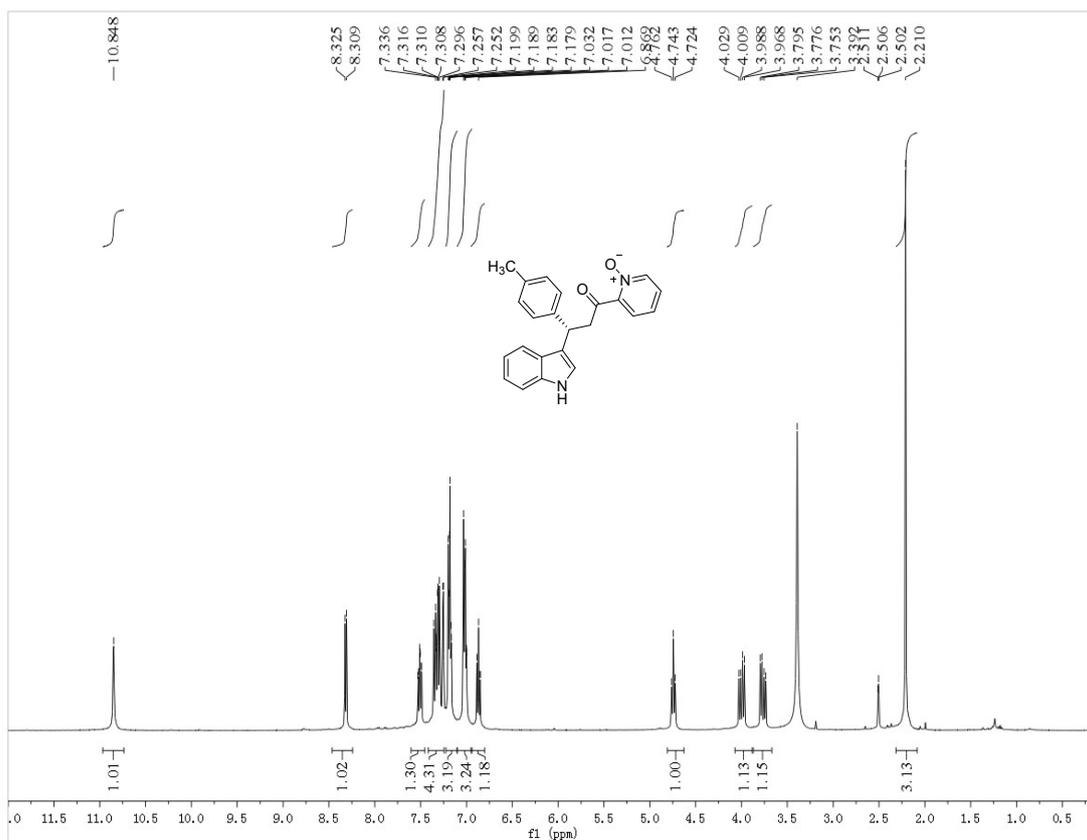
¹H and ¹³C NMR of 6ai



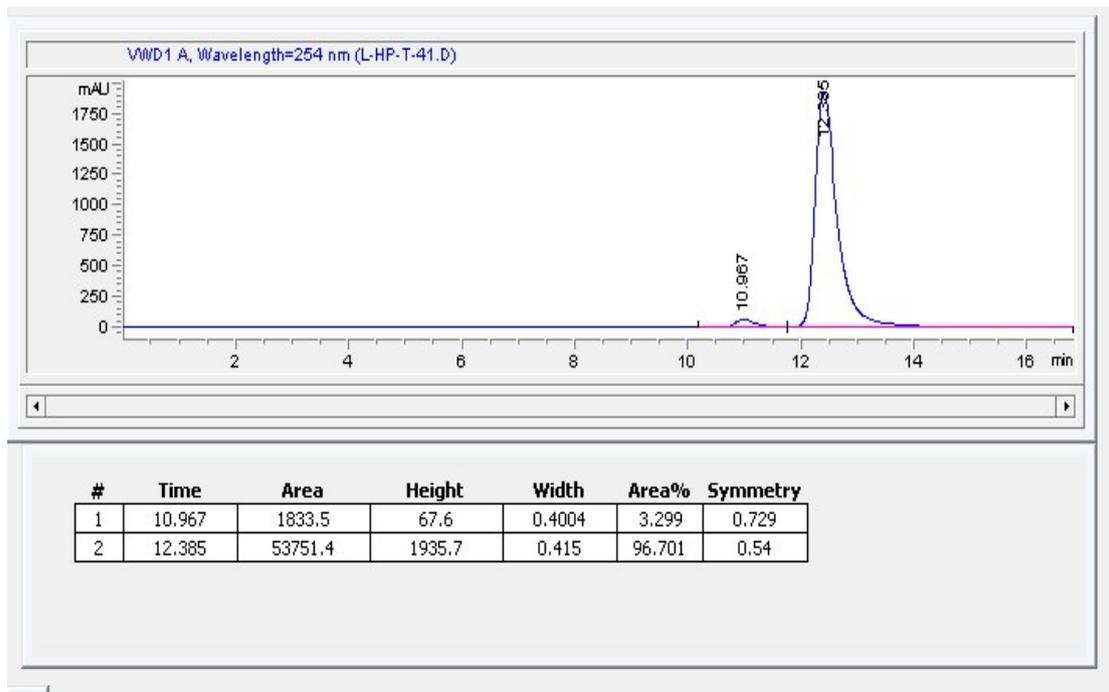
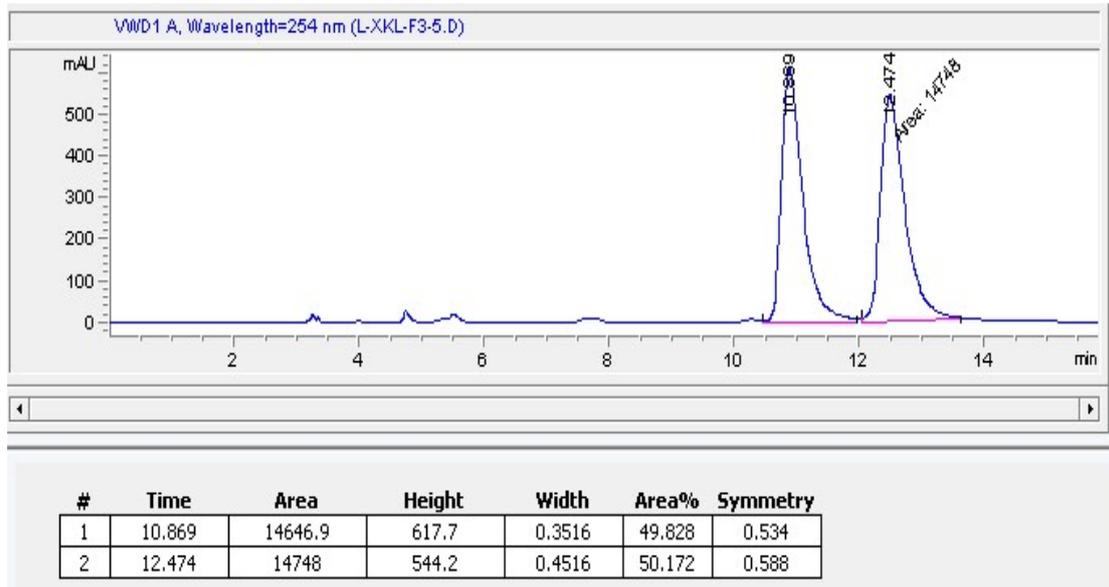
HPLC of 6ai



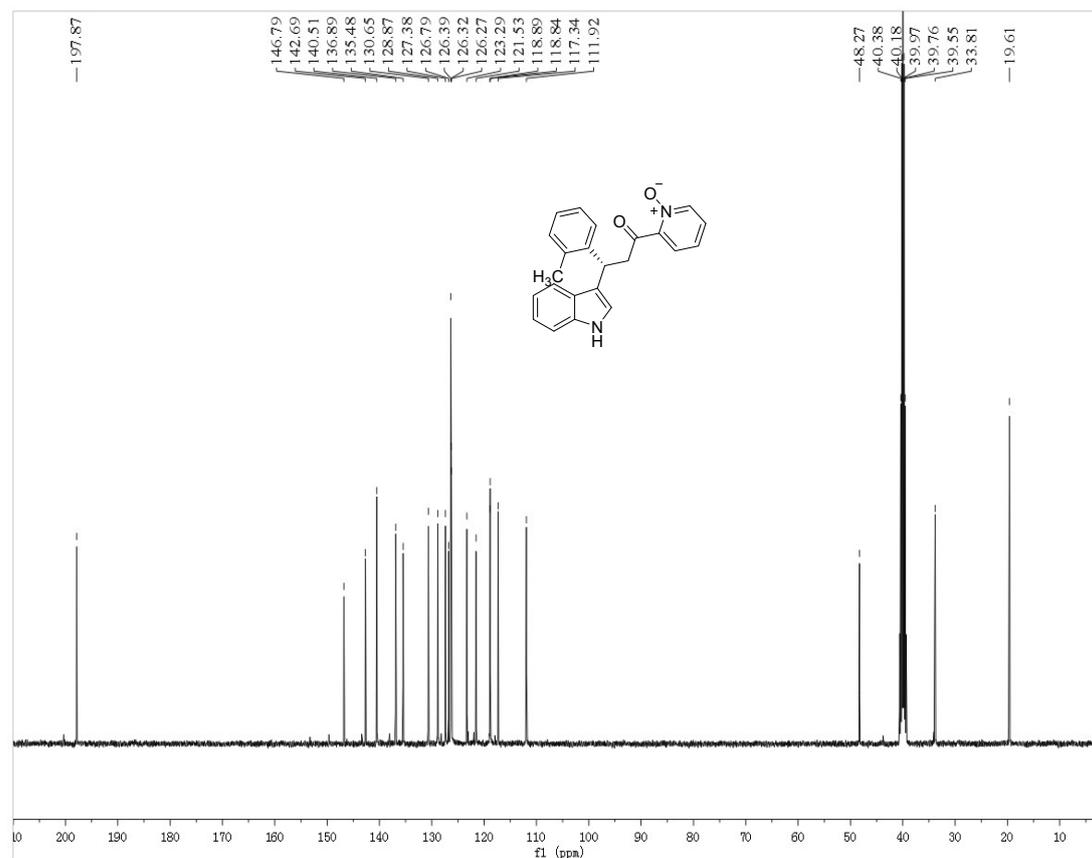
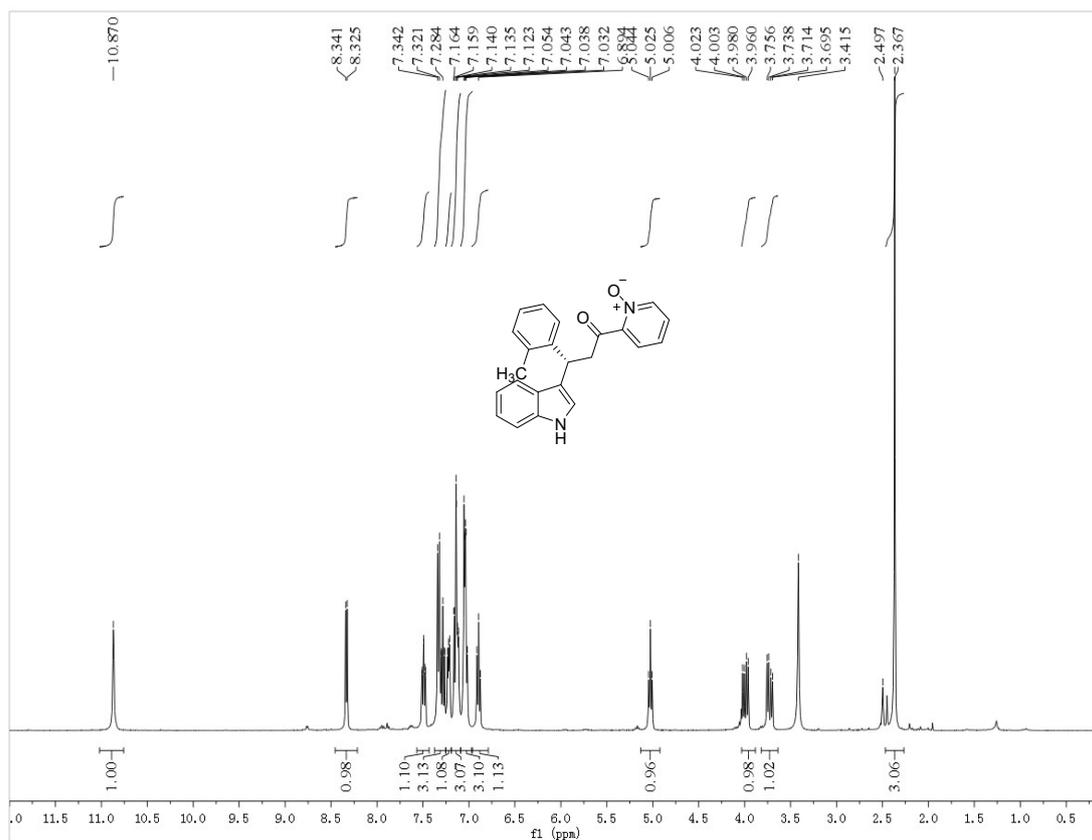
^1H and ^{13}C NMR of 6aj



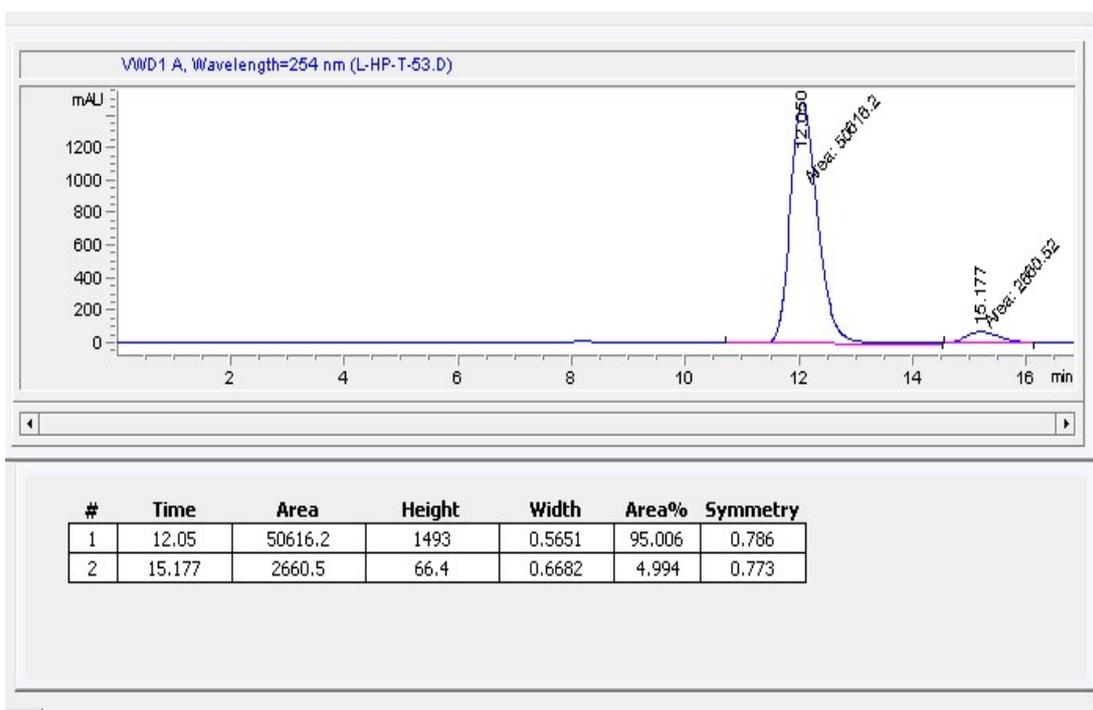
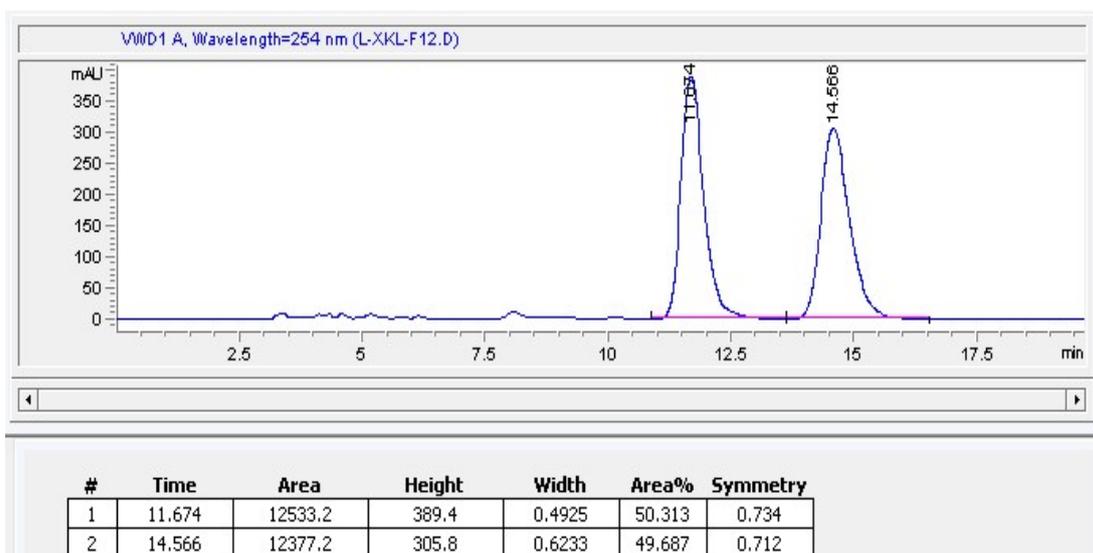
HPLC of 6aj



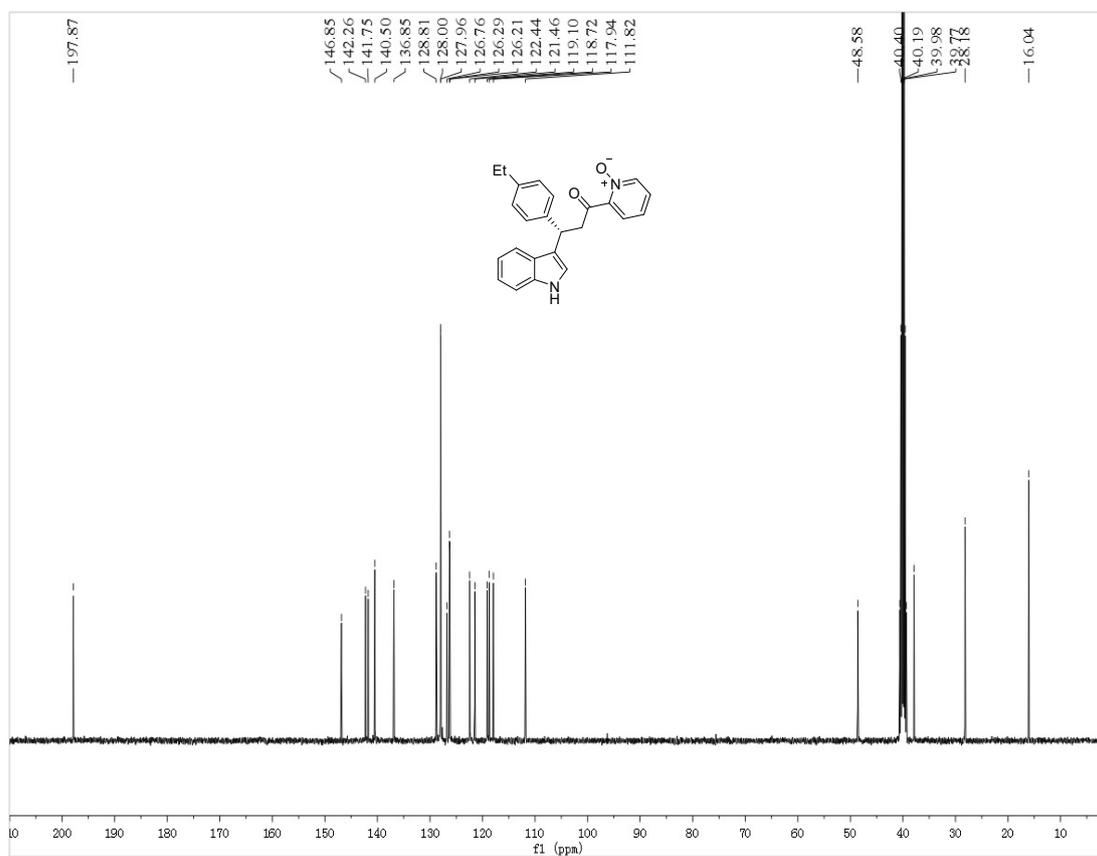
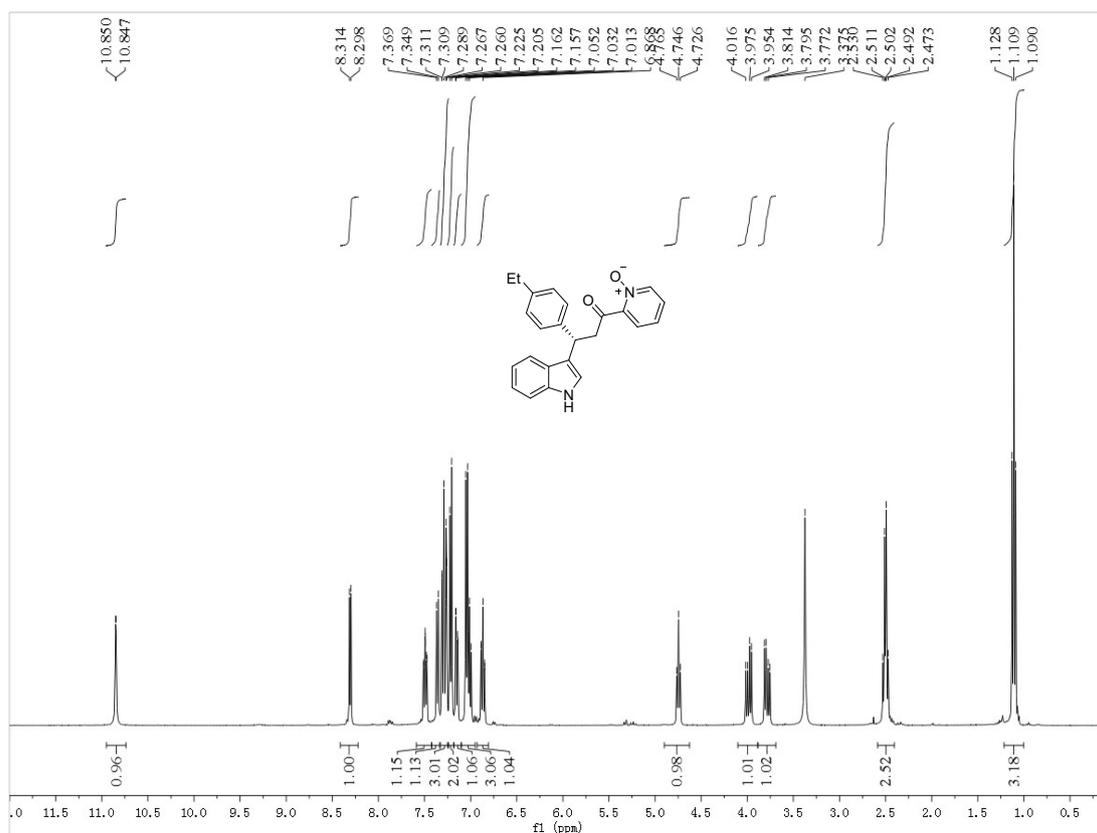
¹H and ¹³C NMR of 6ak



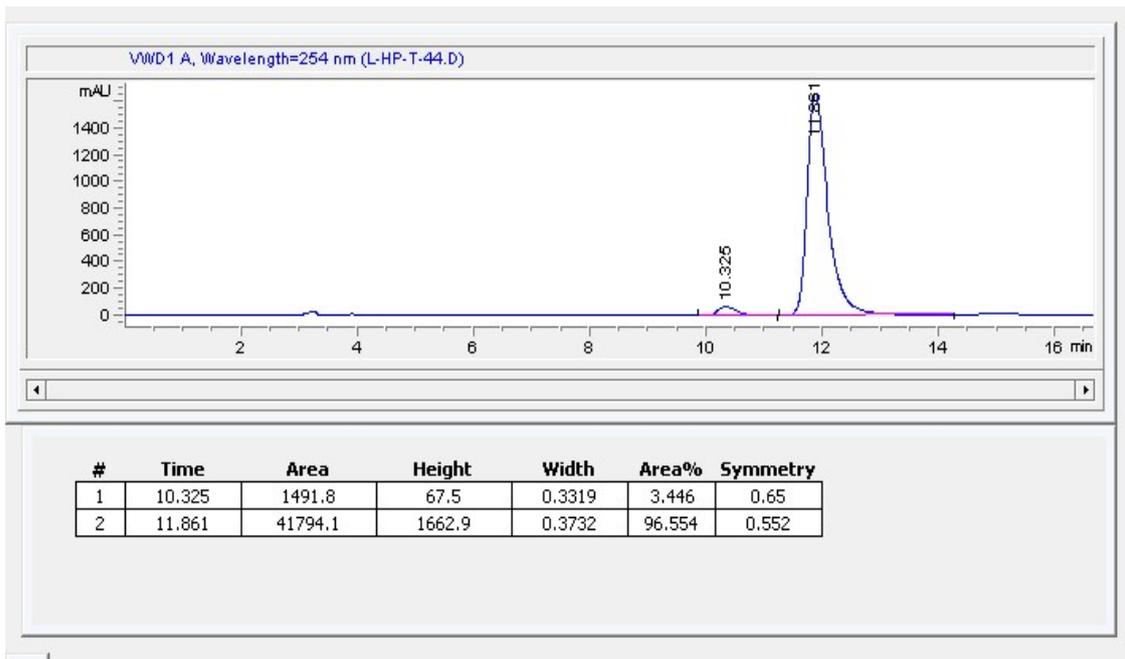
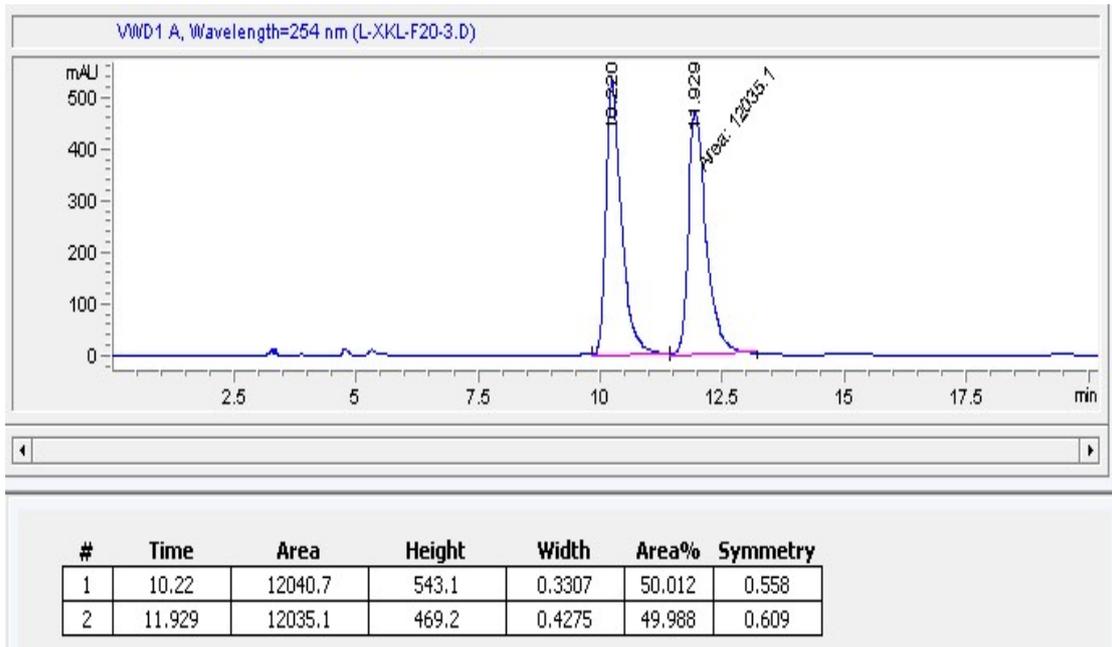
HPLC of 6ak



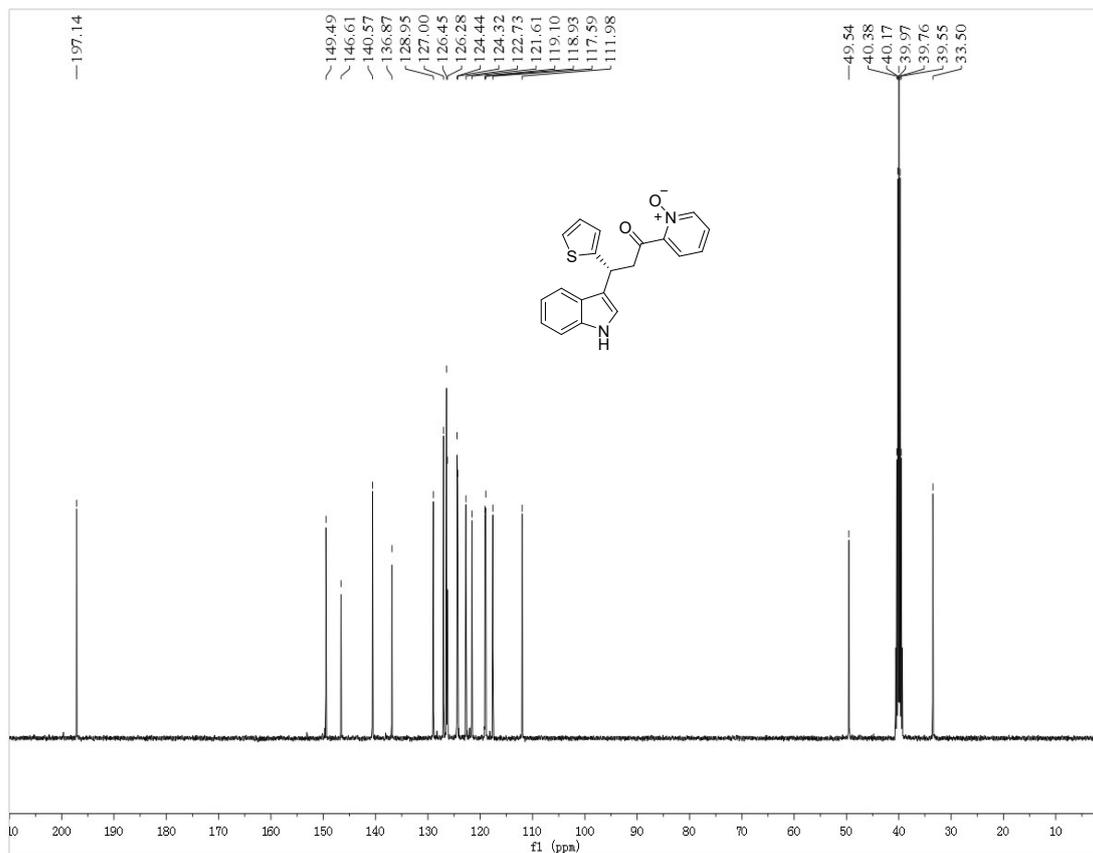
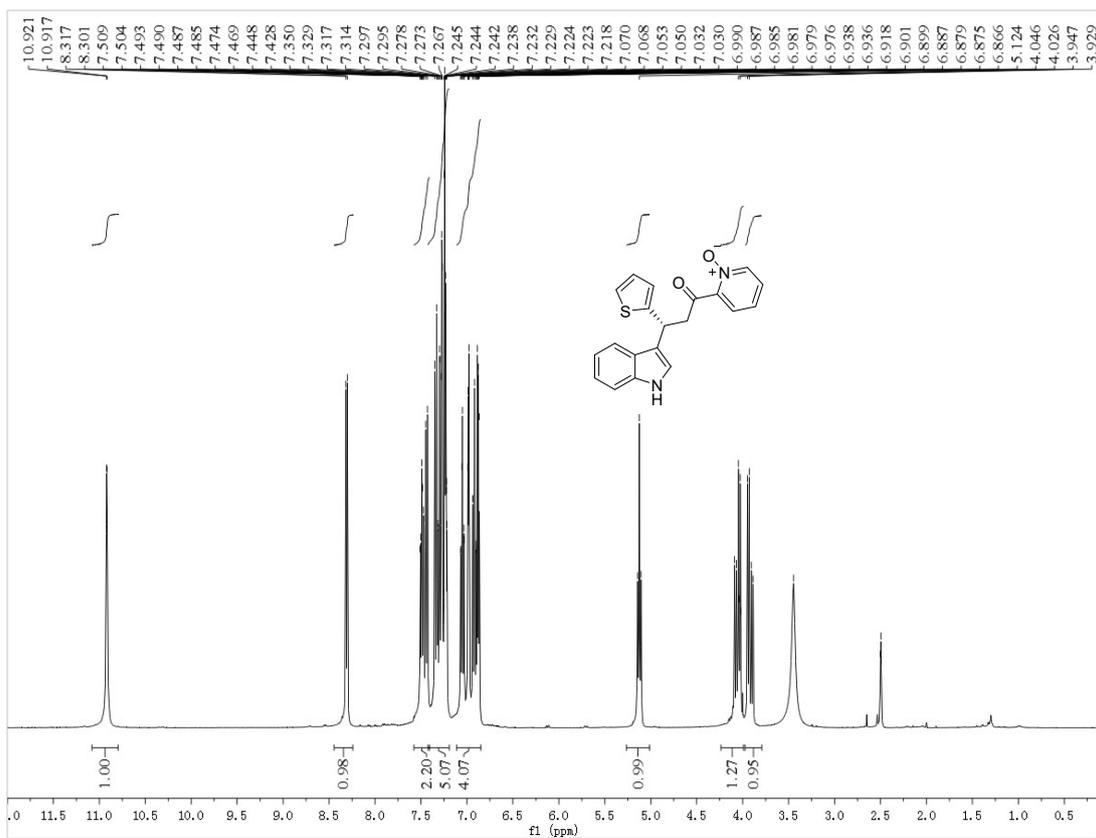
^1H and ^{13}C NMR of 6al



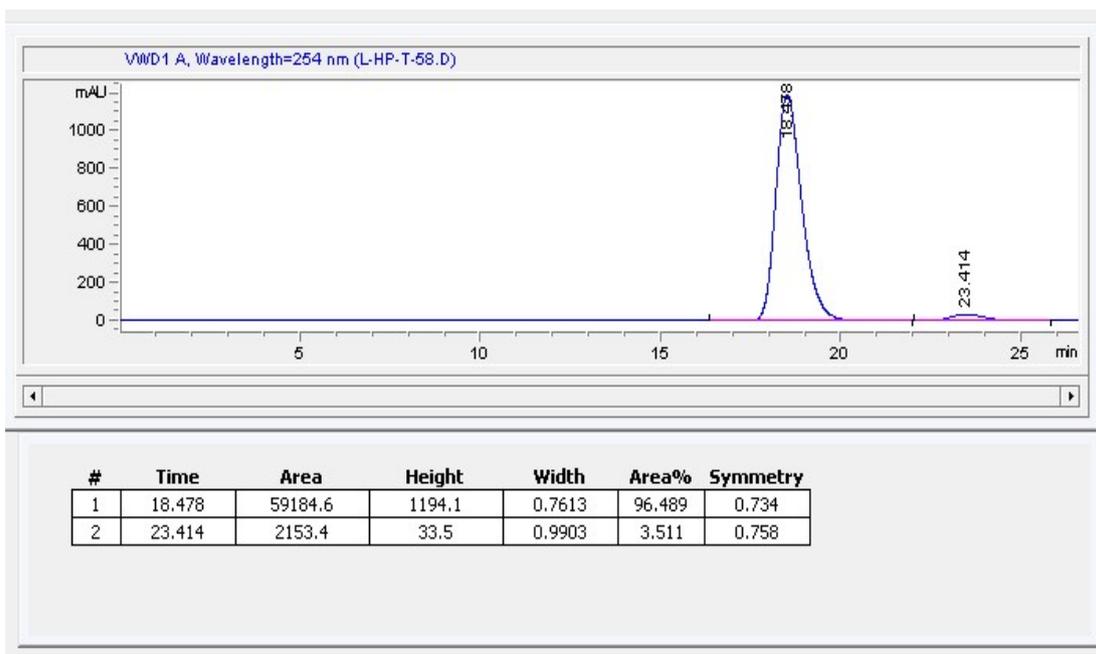
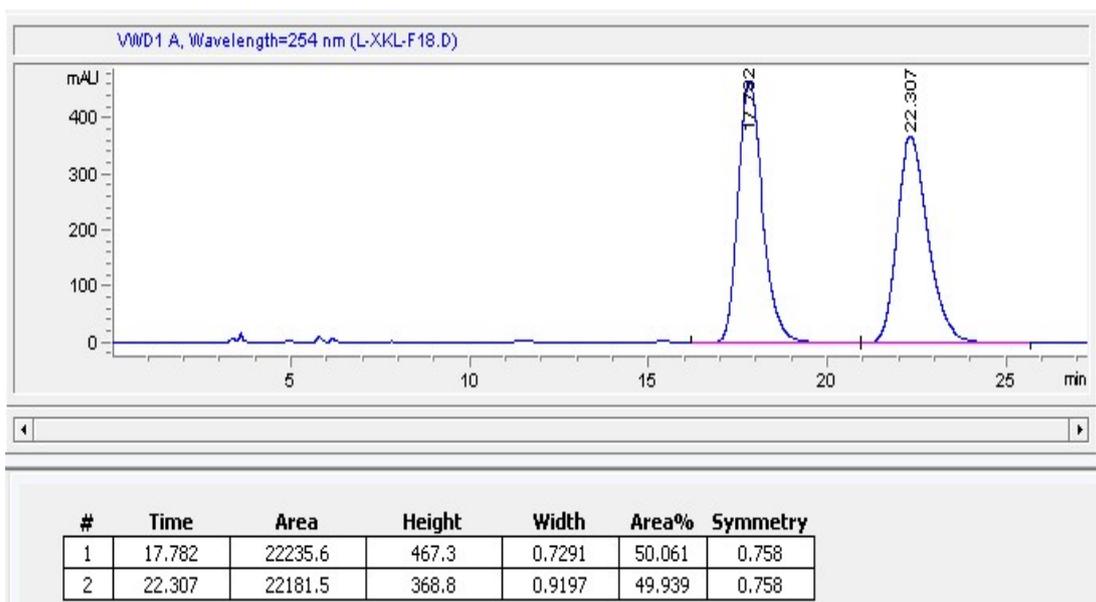
HPLC of 6al



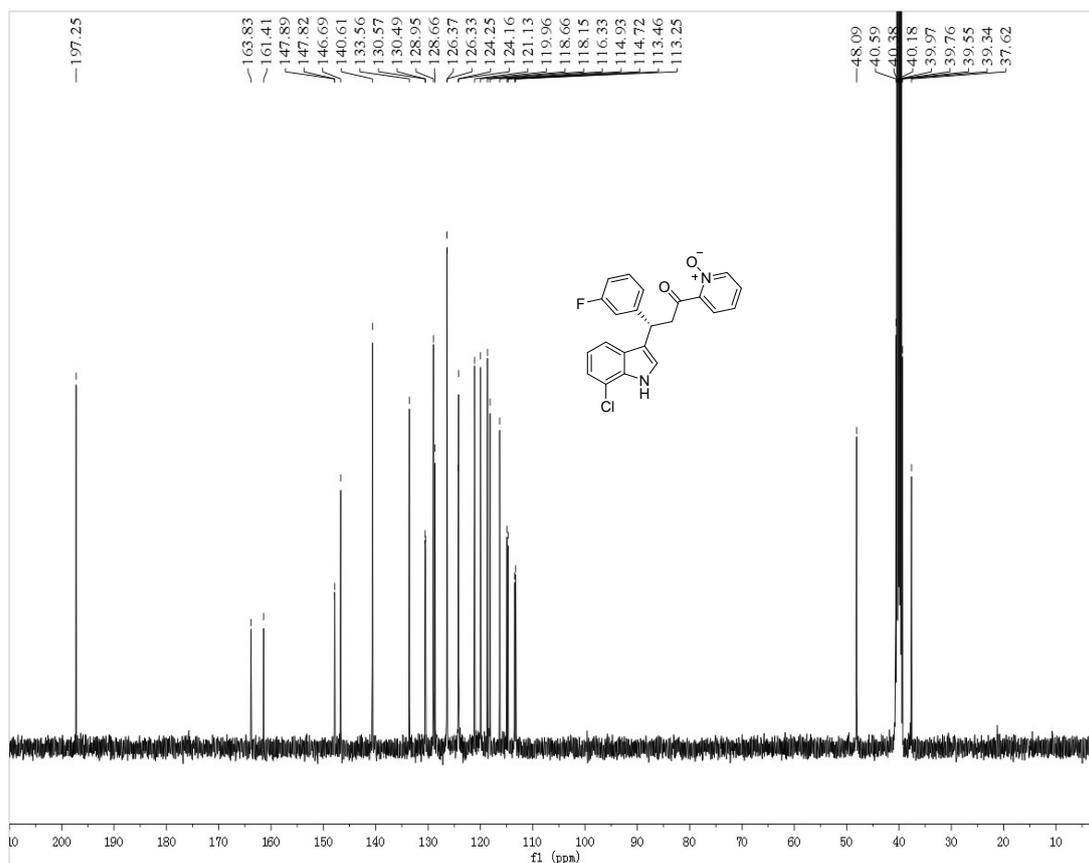
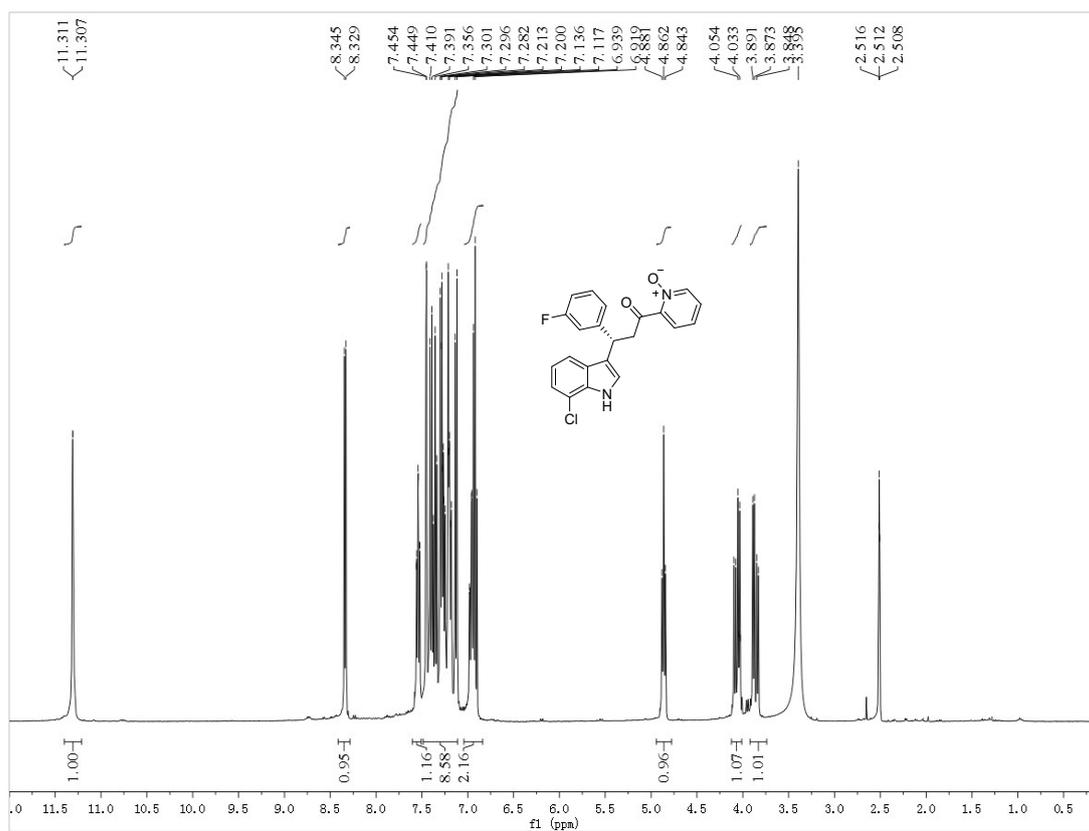
¹H and ¹³C NMR of 6am



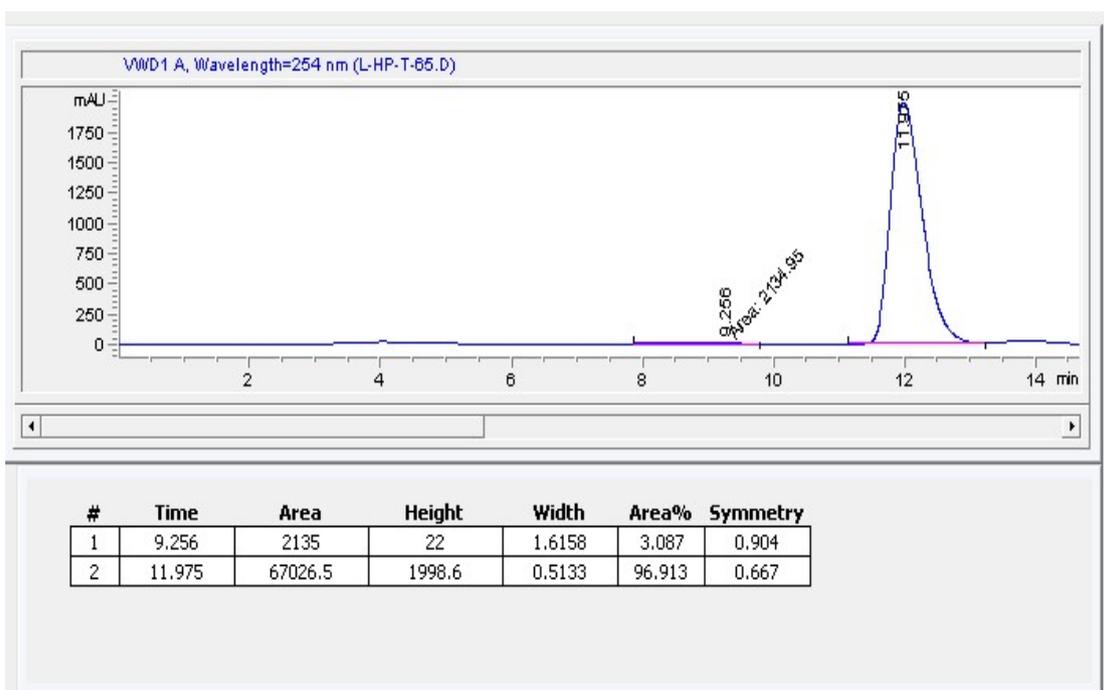
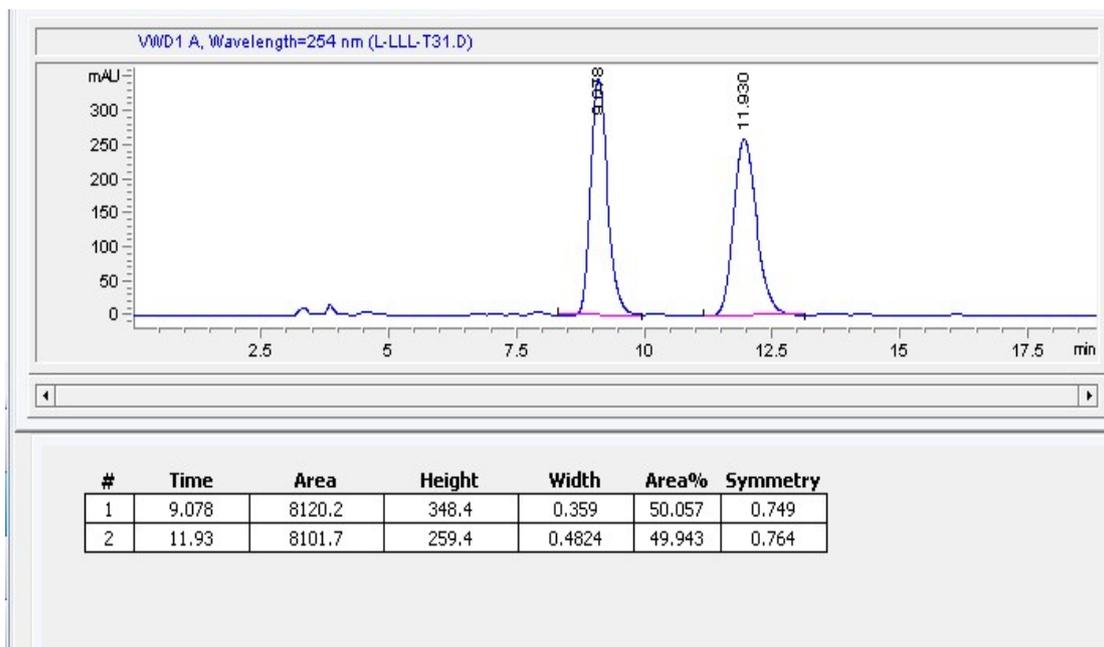
HPLC of 6am



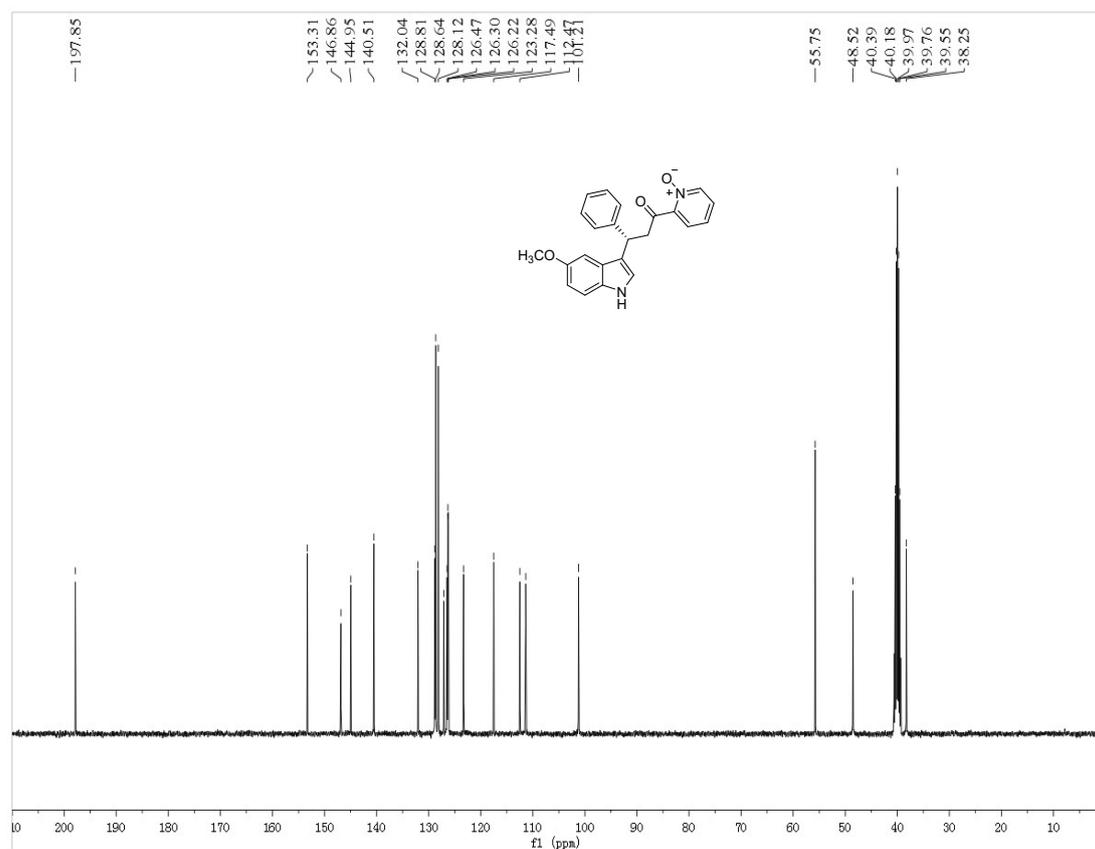
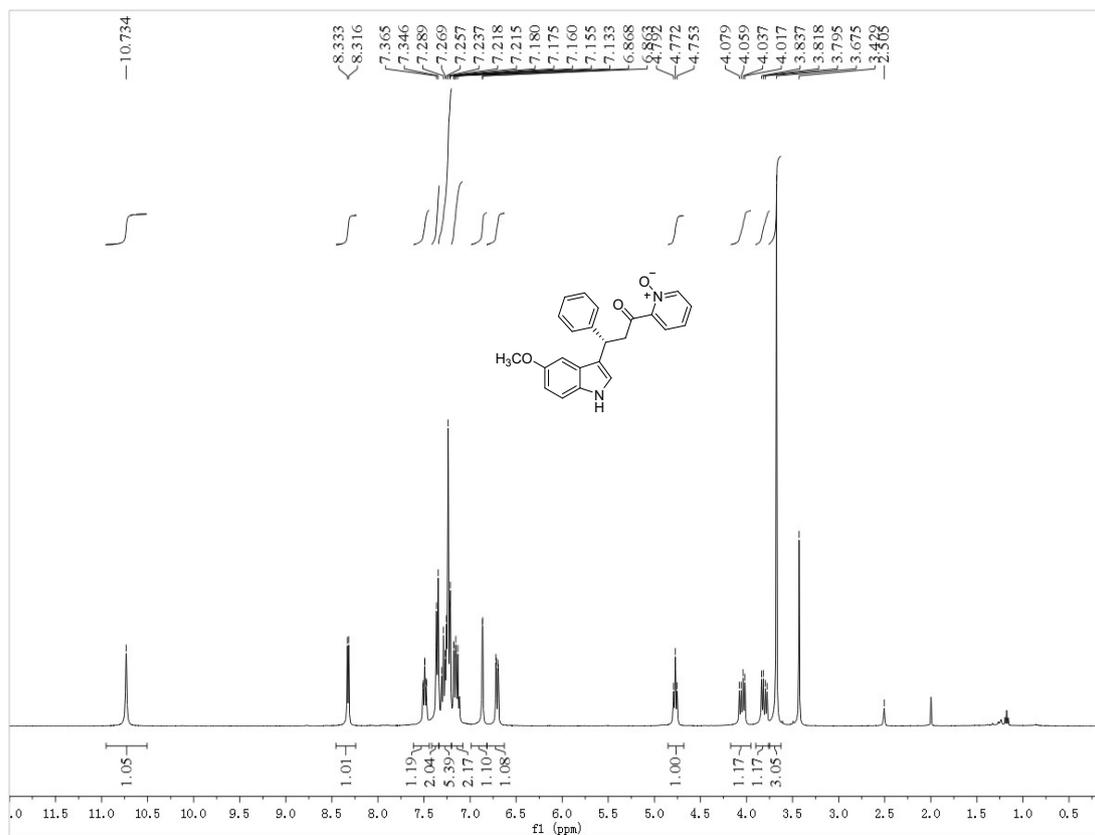
¹H and ¹³C NMR of 6ba



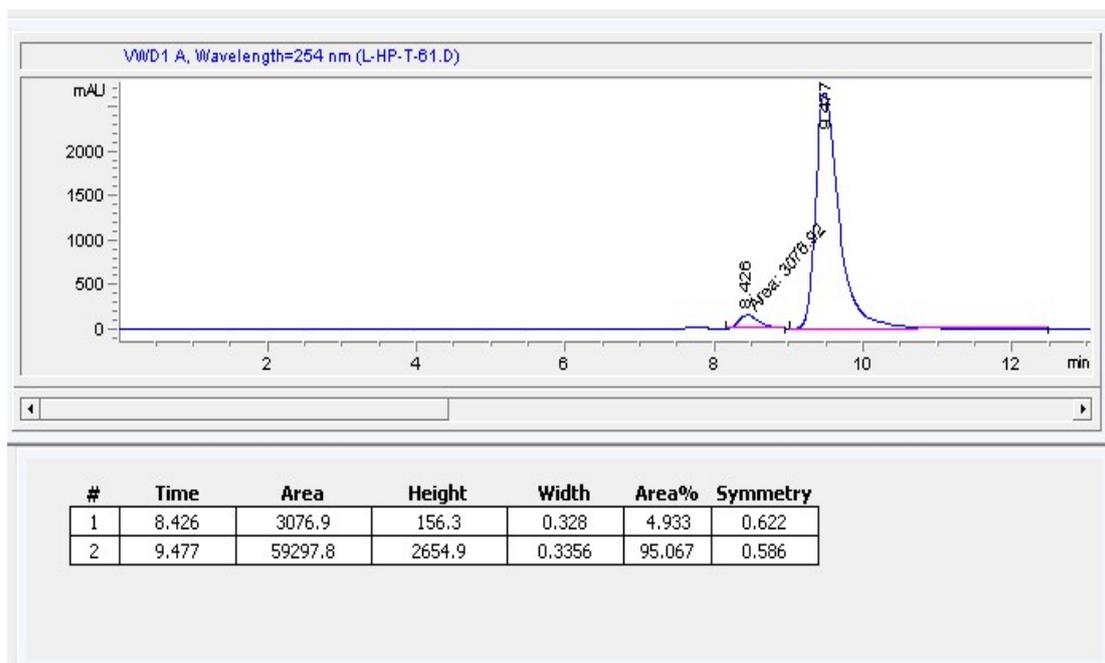
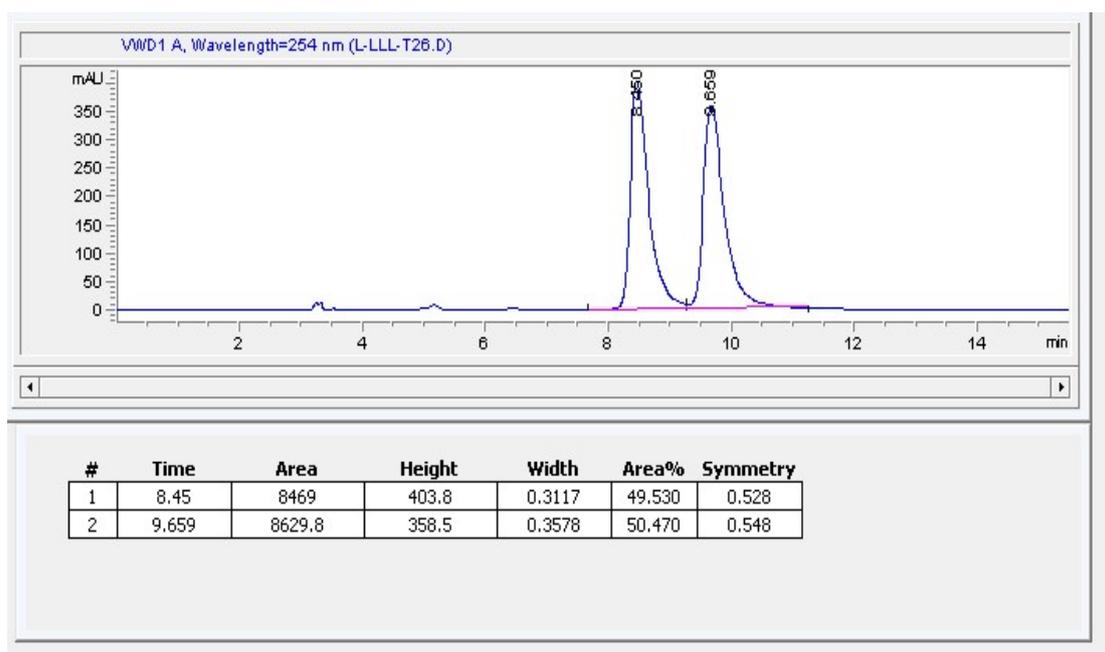
HPLC of 6ba



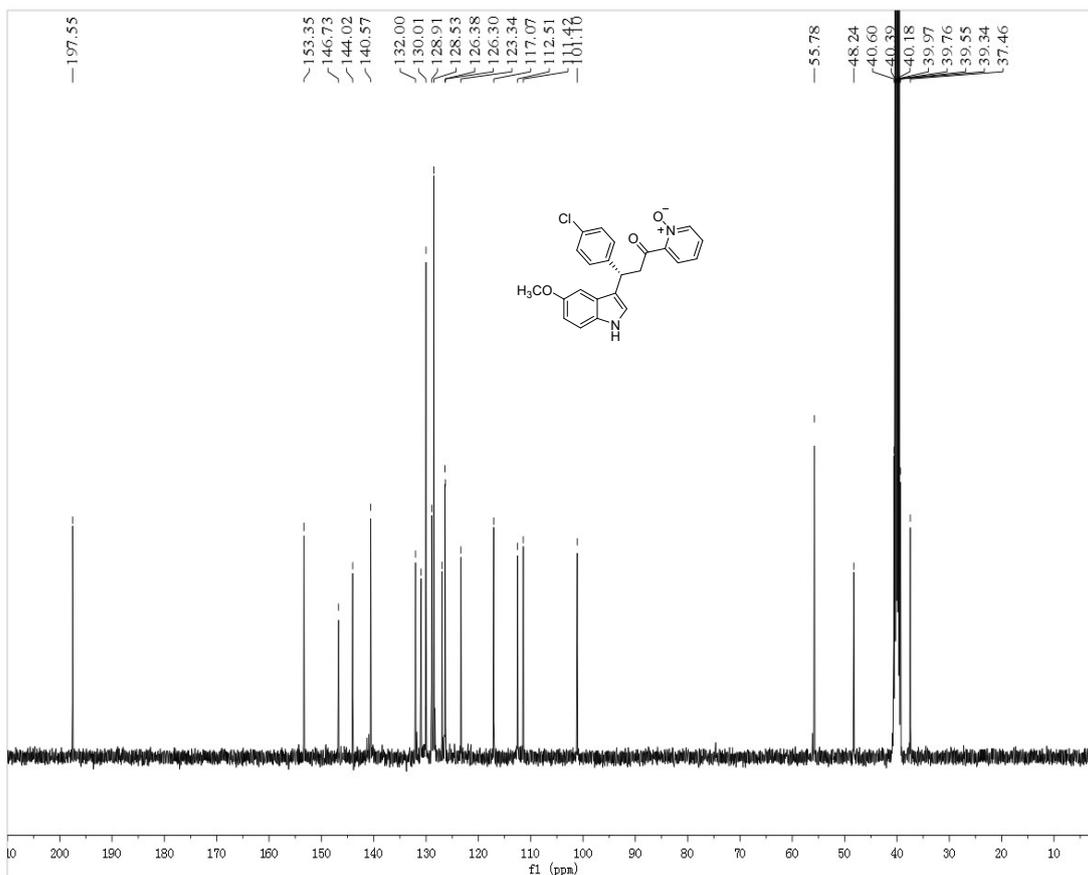
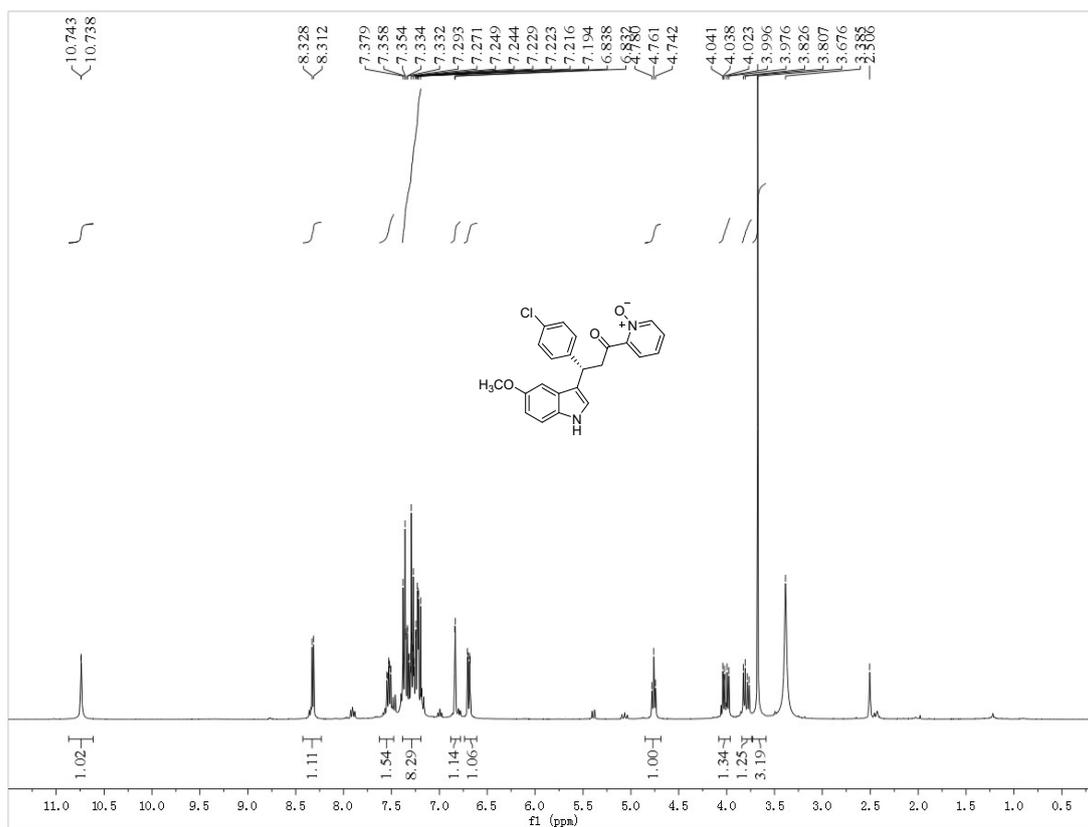
¹H and ¹³C NMR of 6ca



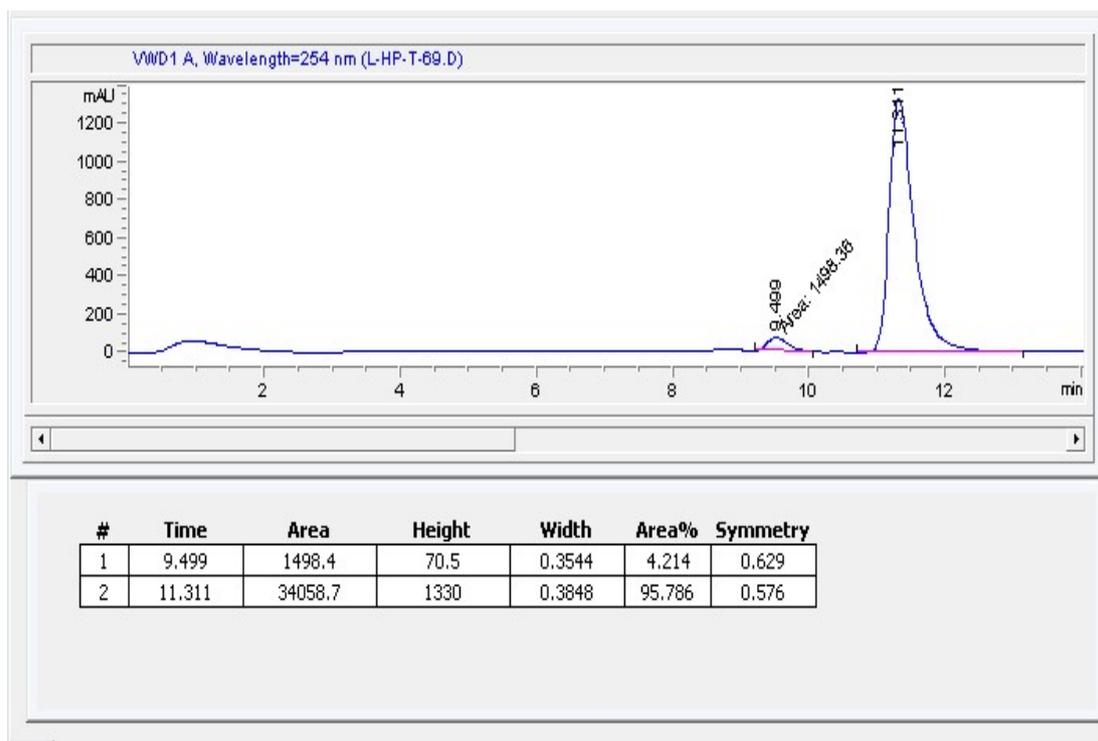
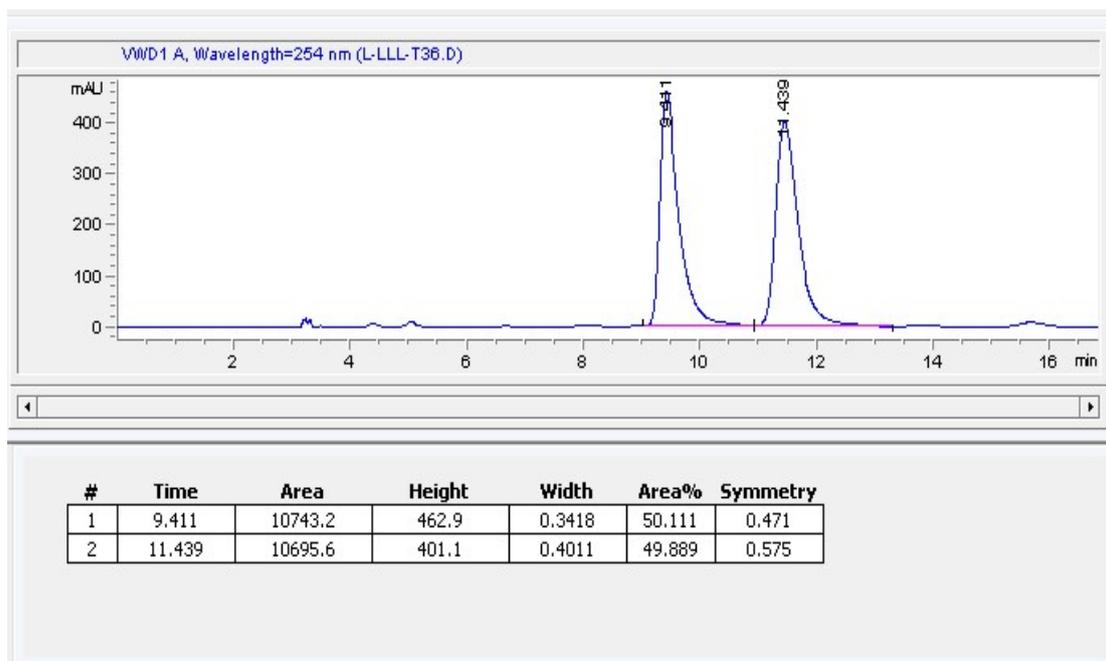
HPLC of 6ca



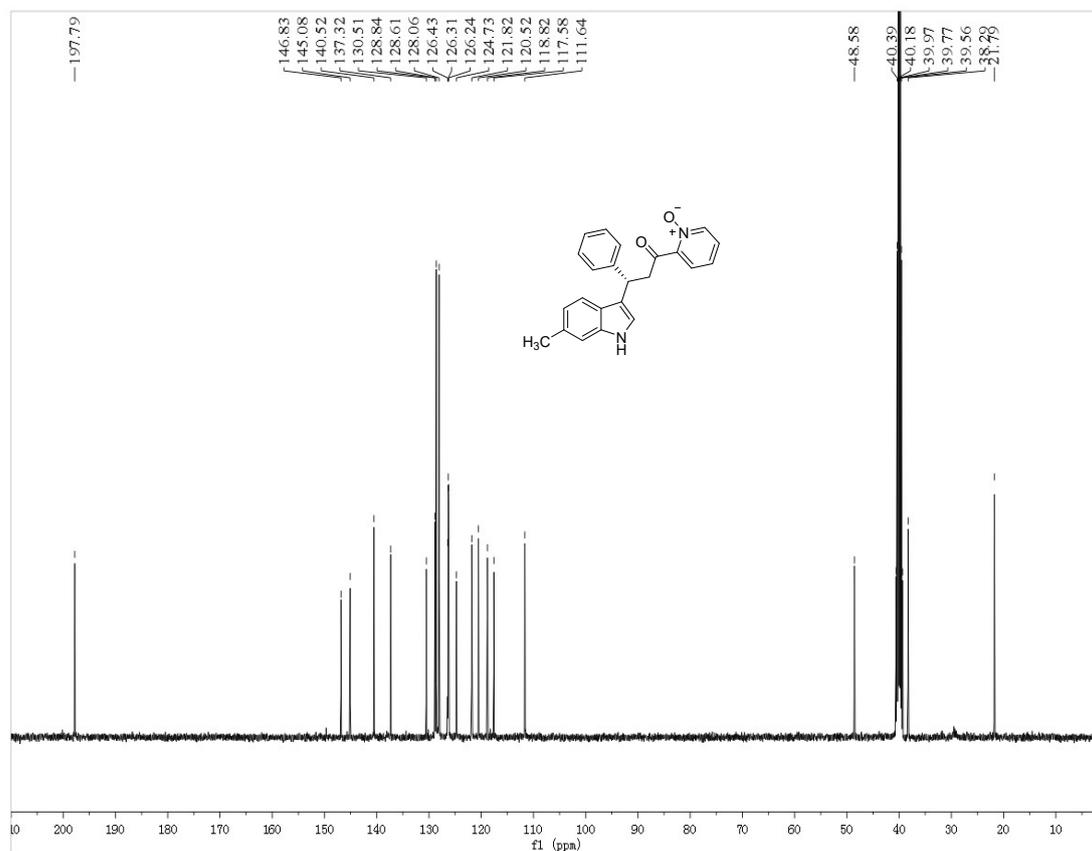
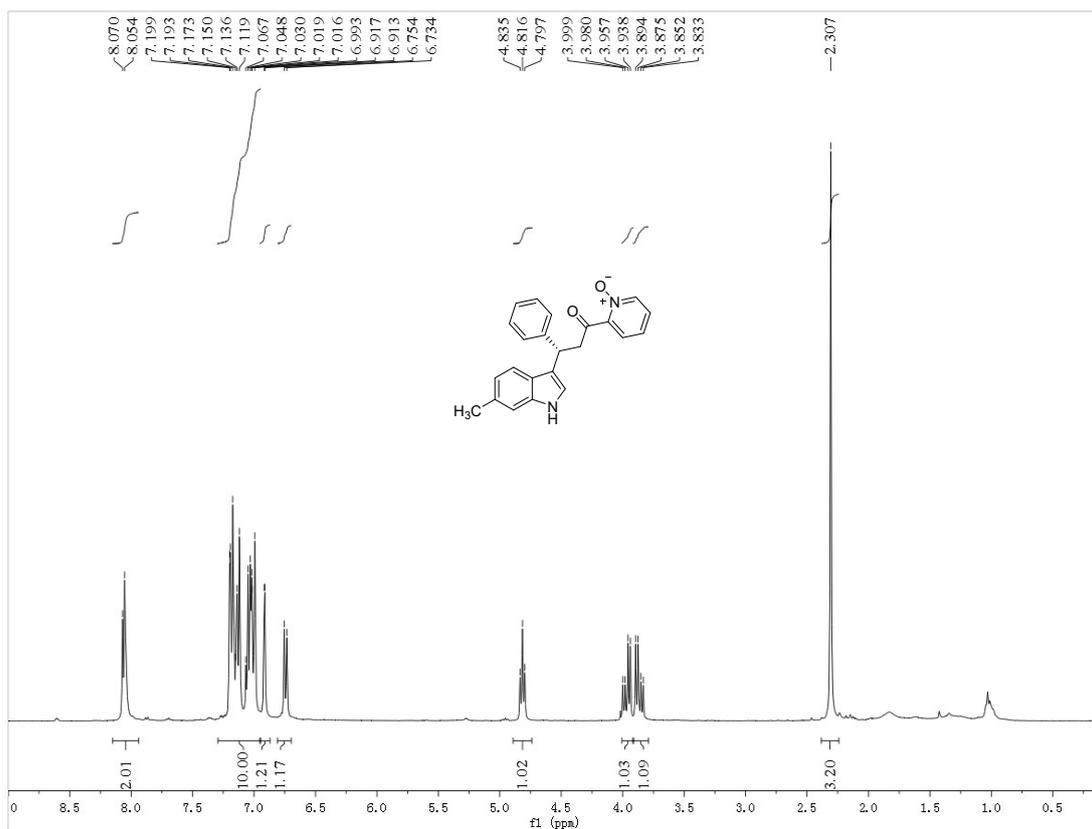
¹H and ¹³C NMR of 6cb



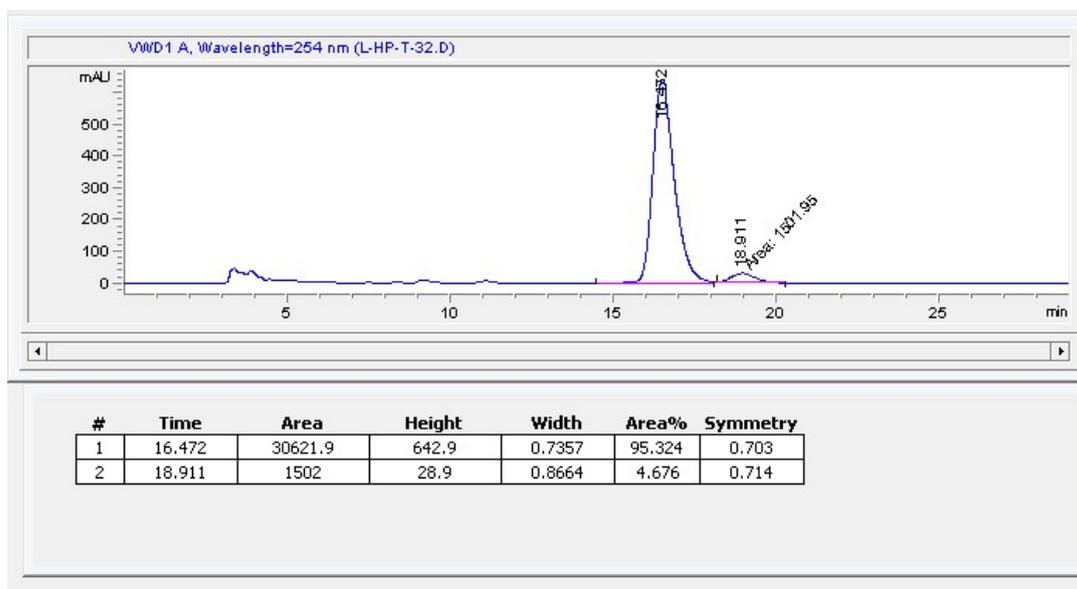
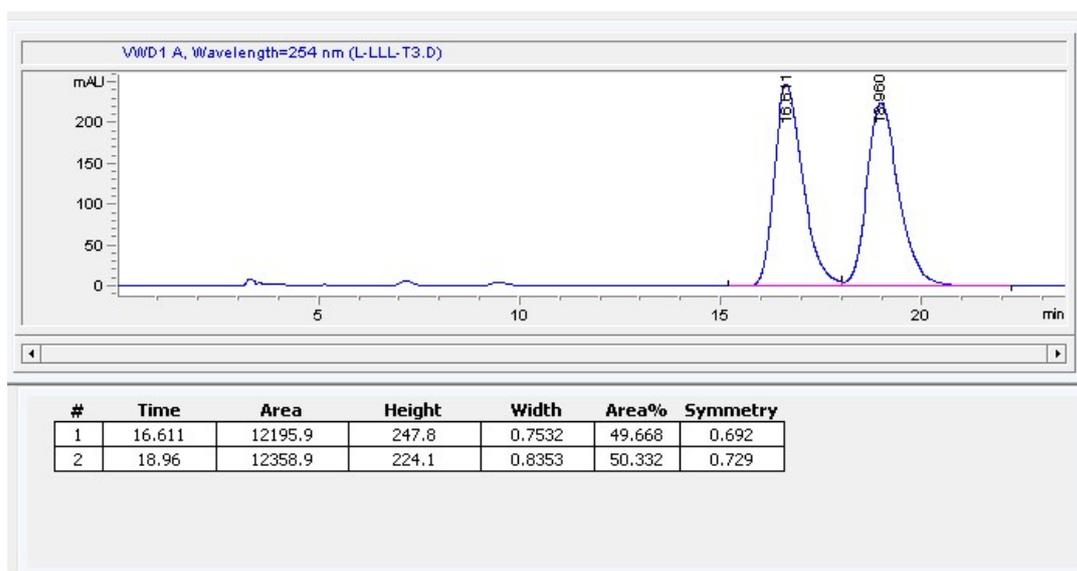
HPLC of 6cb



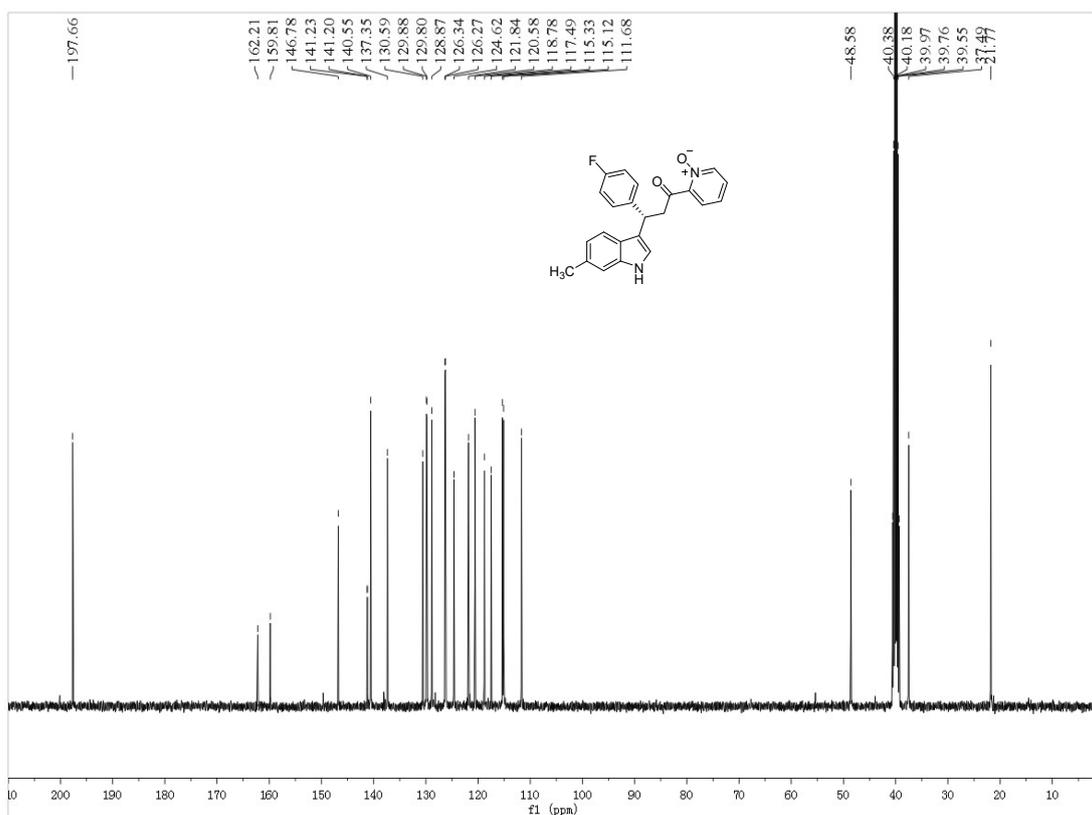
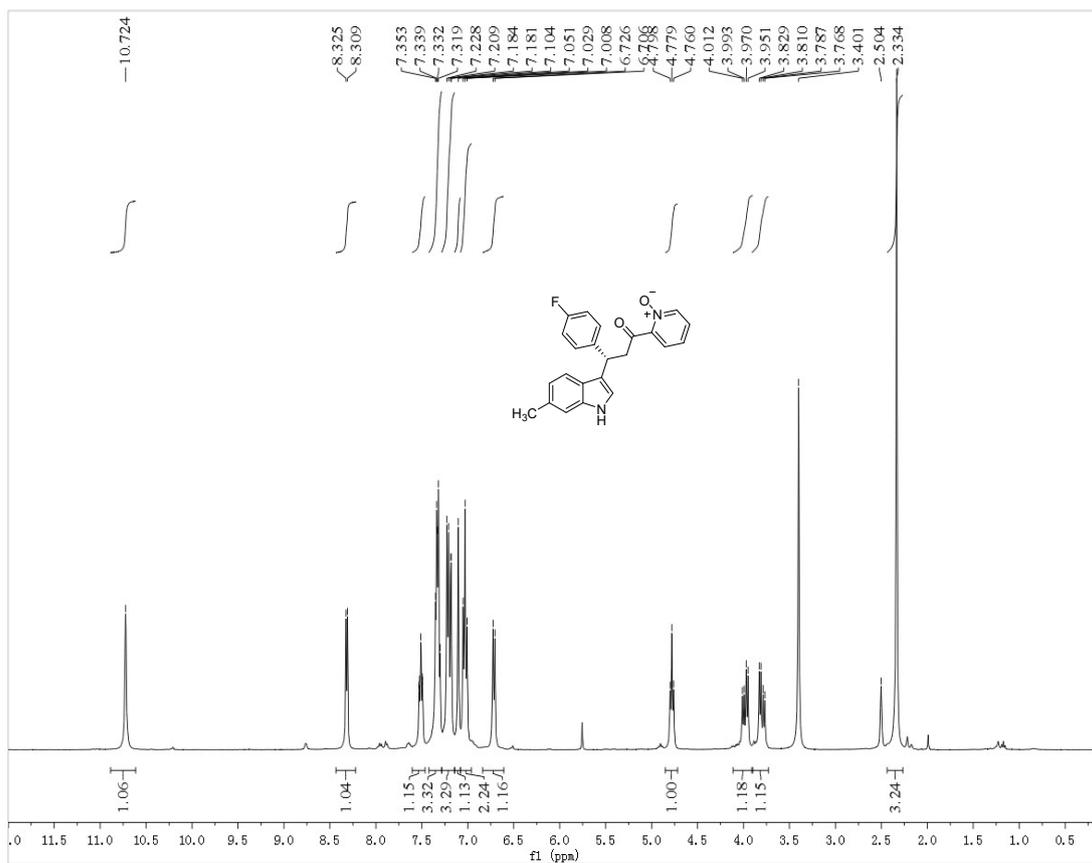
¹H and ¹³C NMR of 6da



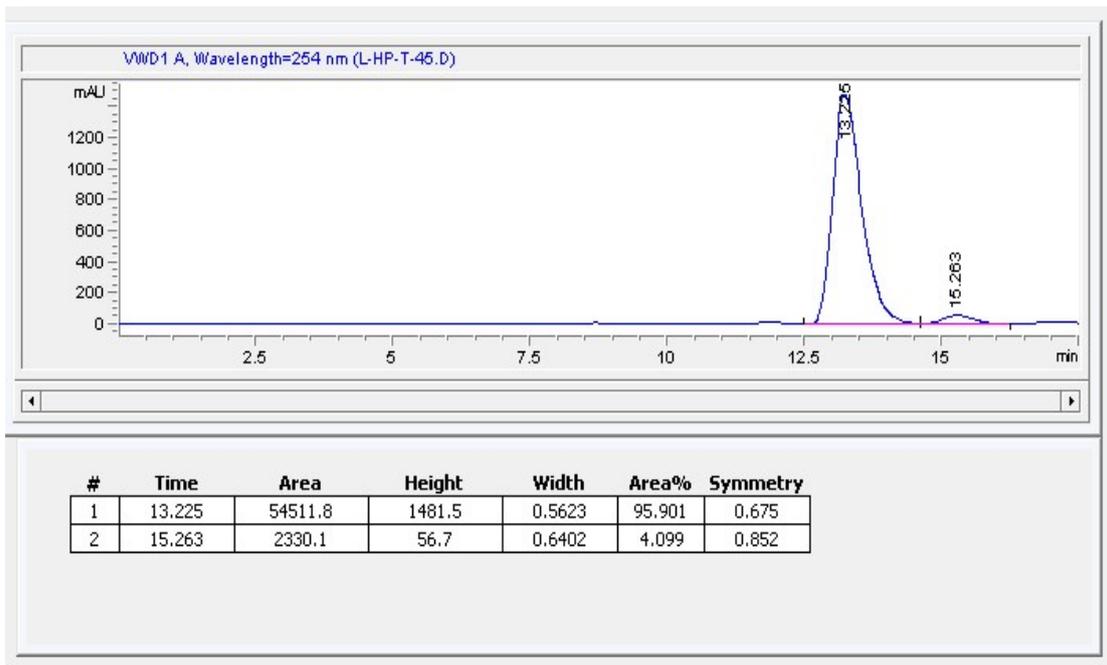
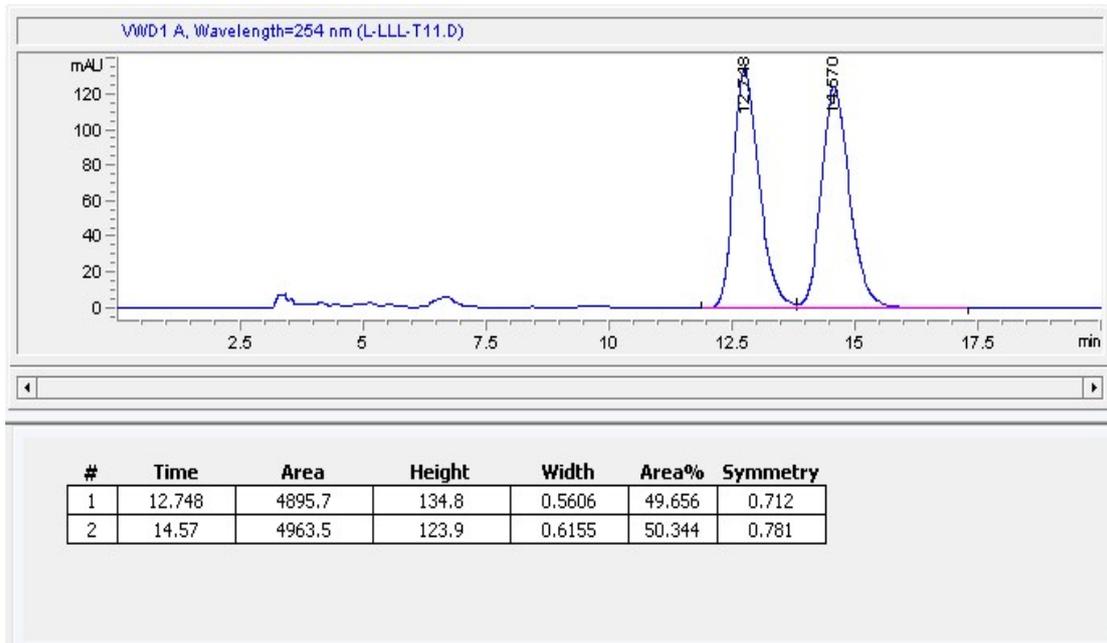
HPLC of 6da



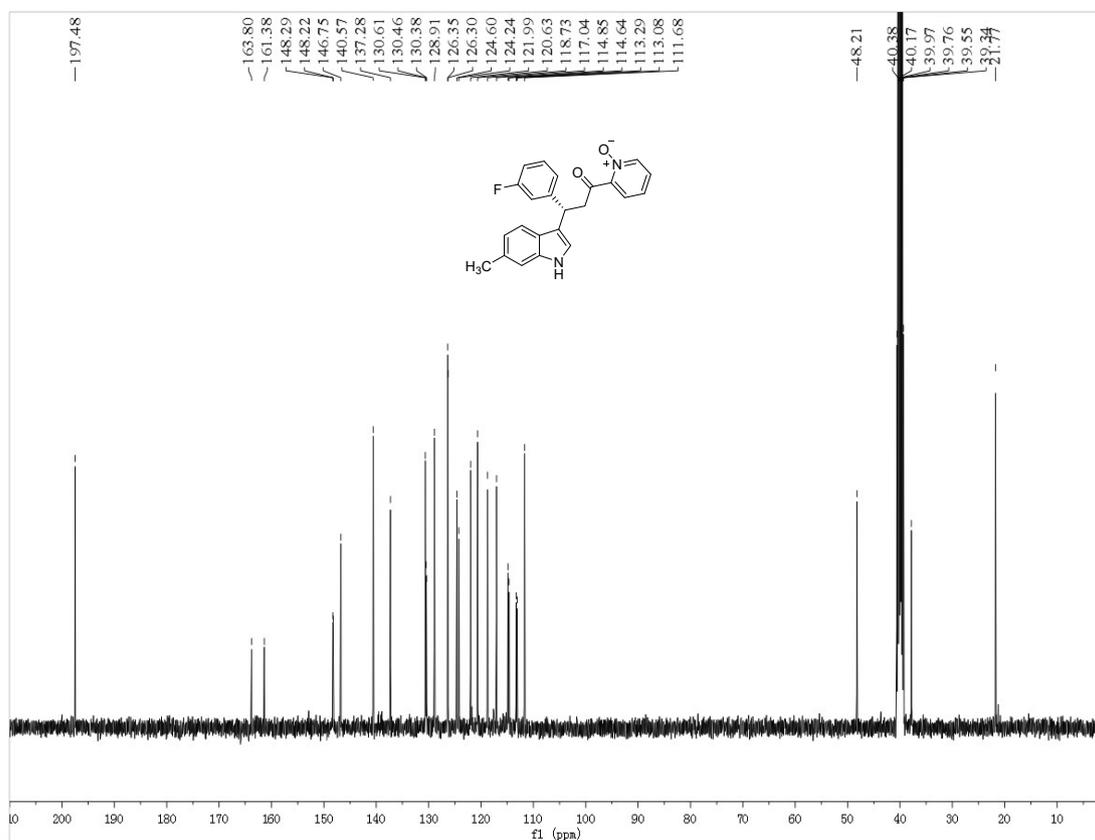
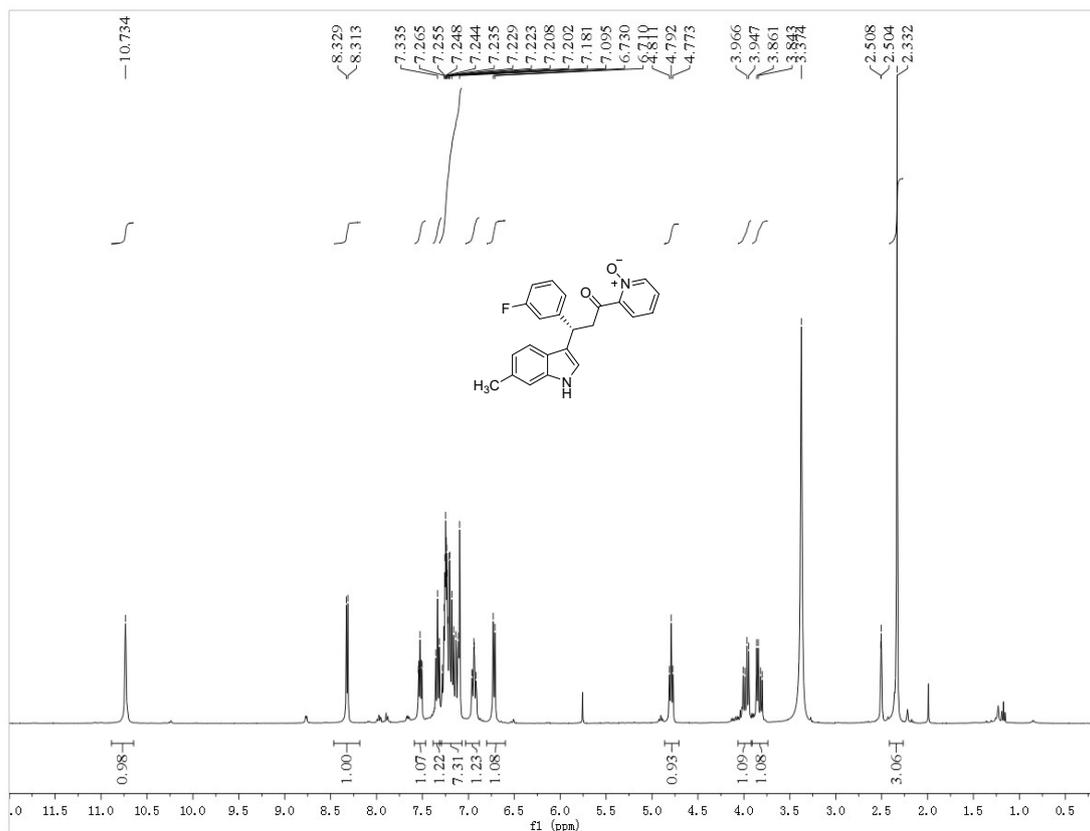
¹H and ¹³C NMR of 6db



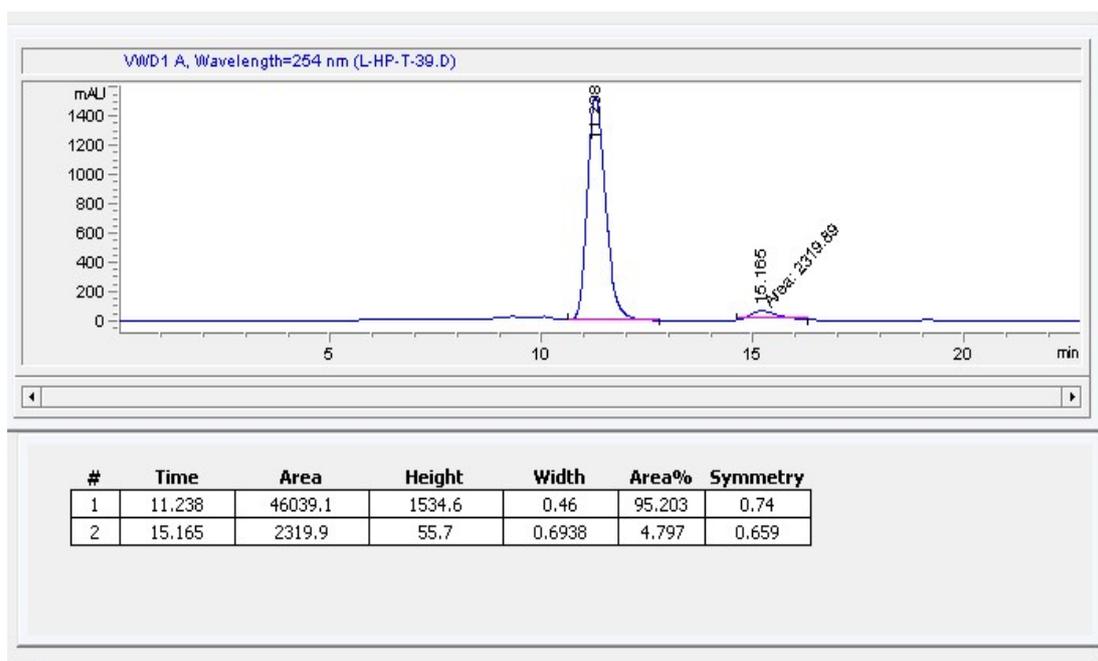
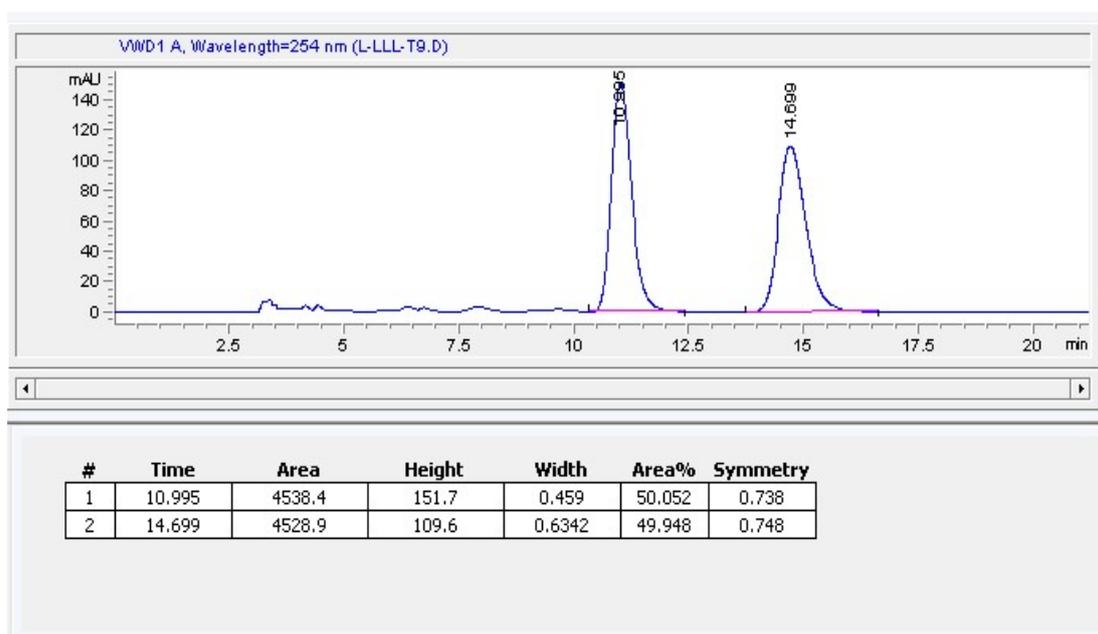
HPLC of 6db



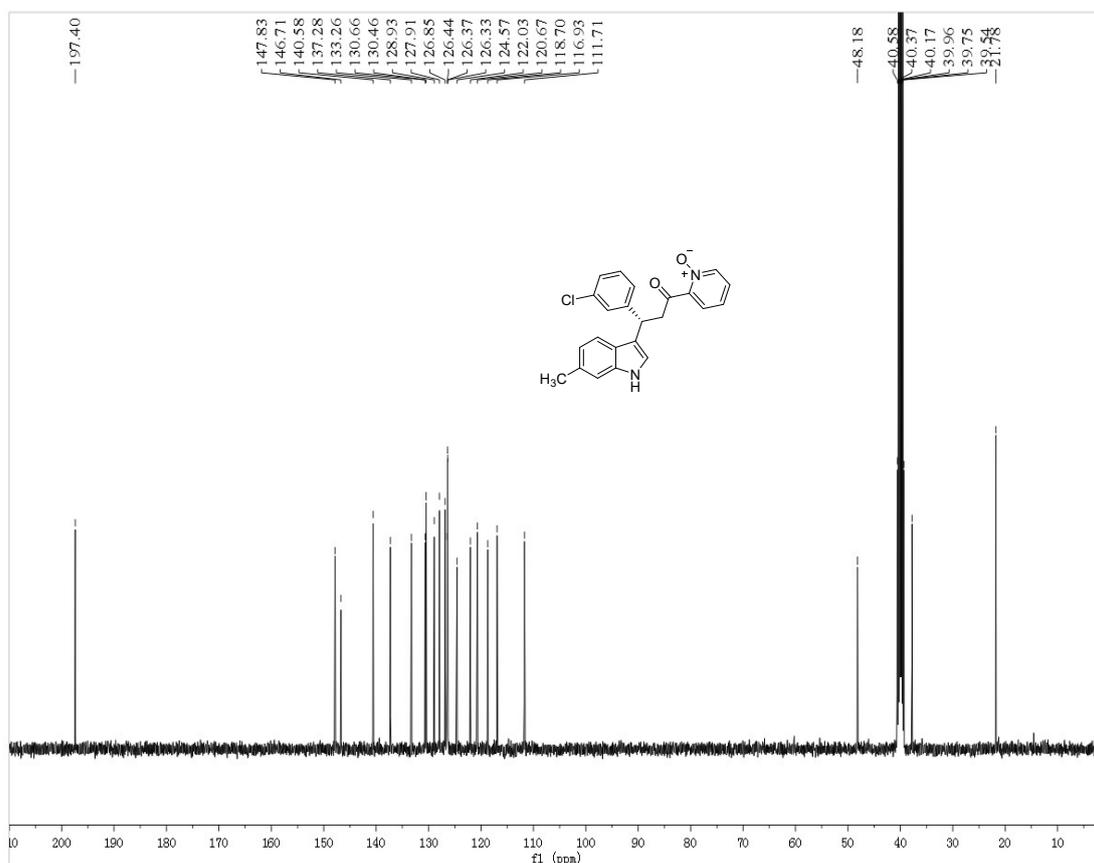
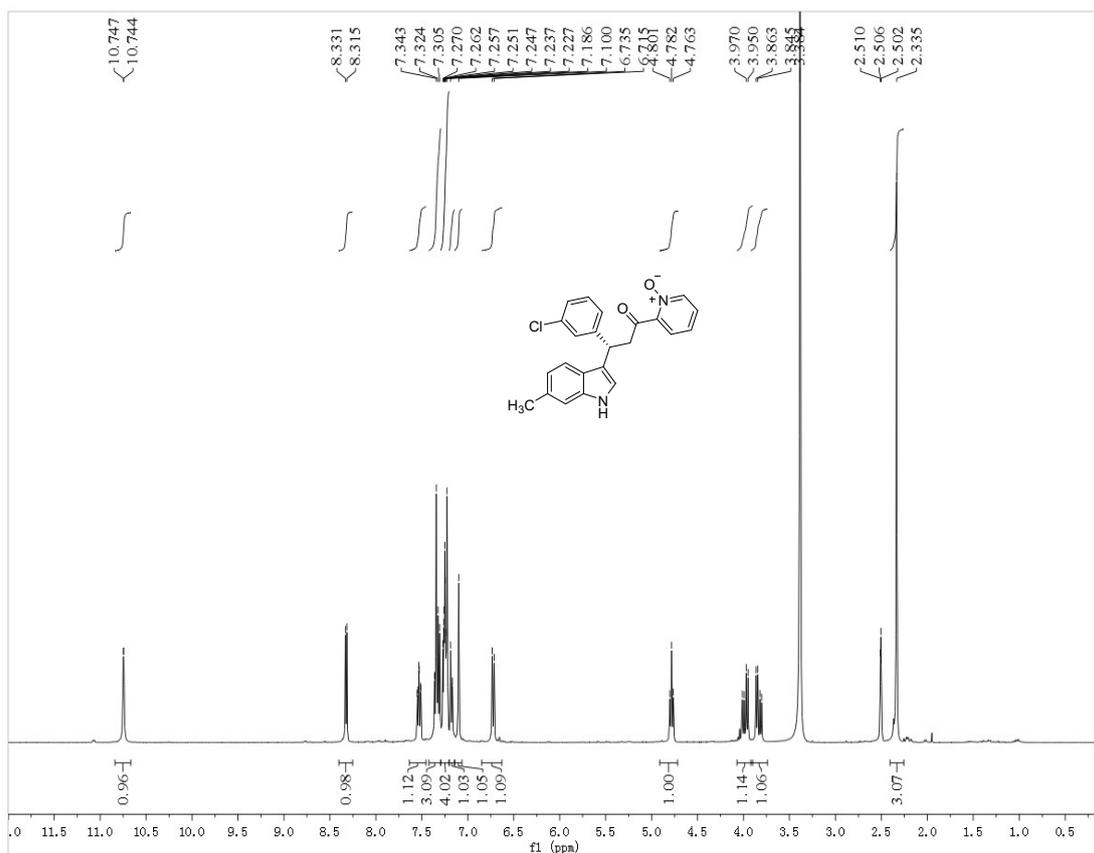
¹H and ¹³C NMR of 6dc



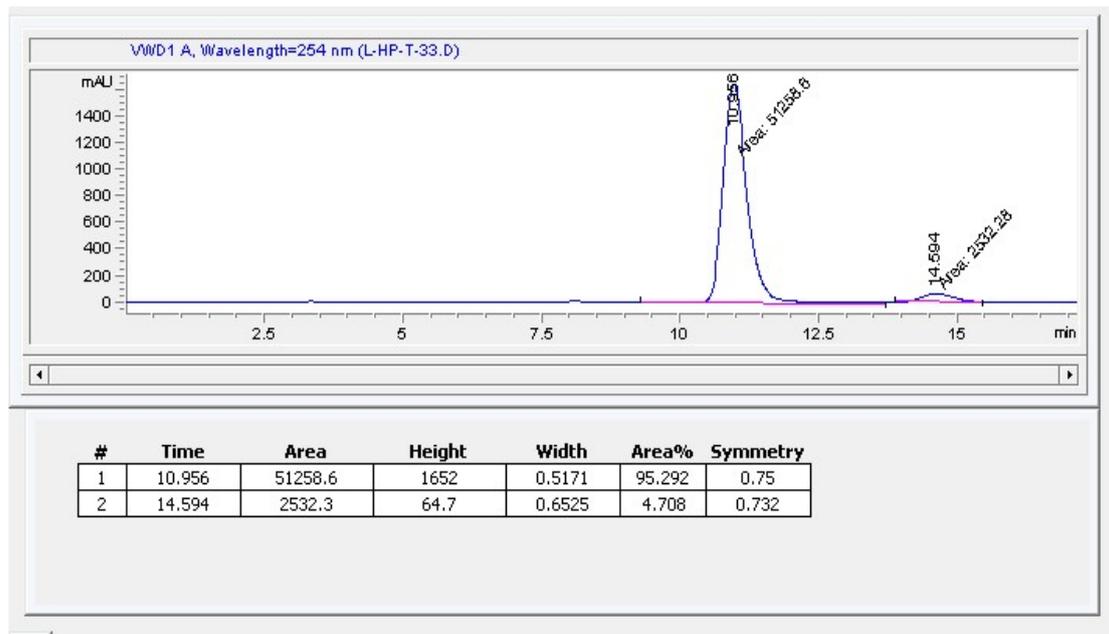
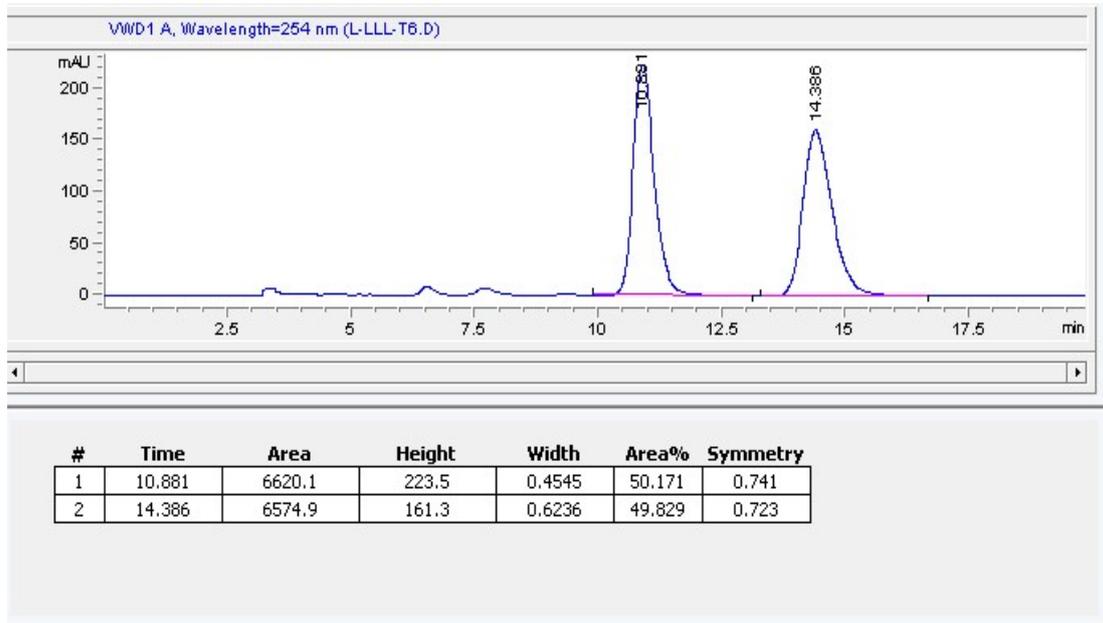
HPLC of 6dc



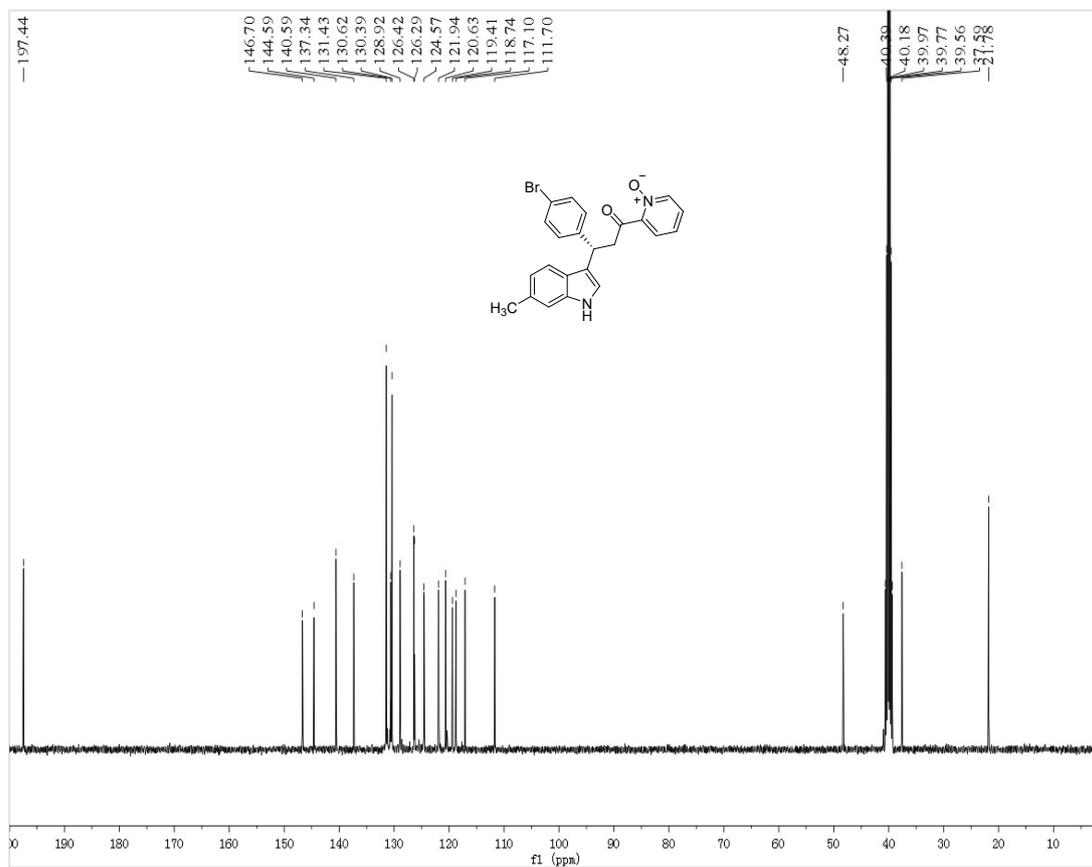
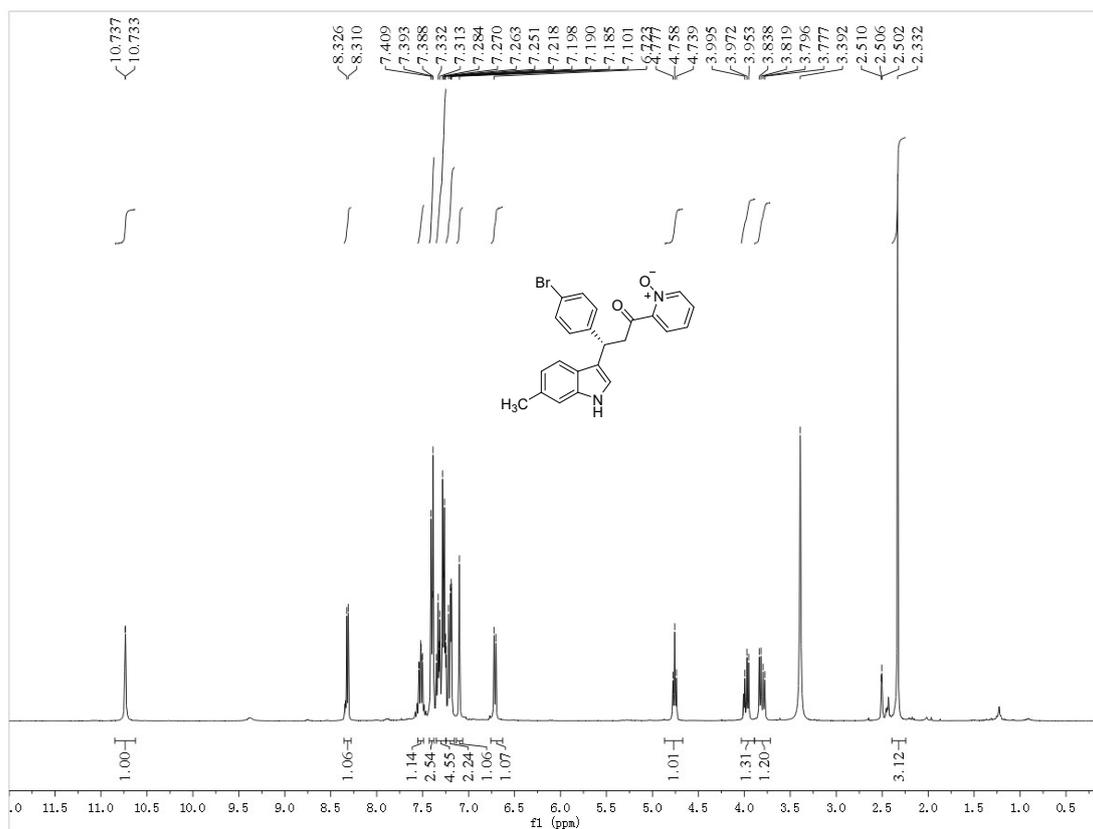
¹H and ¹³C NMR of 6dd



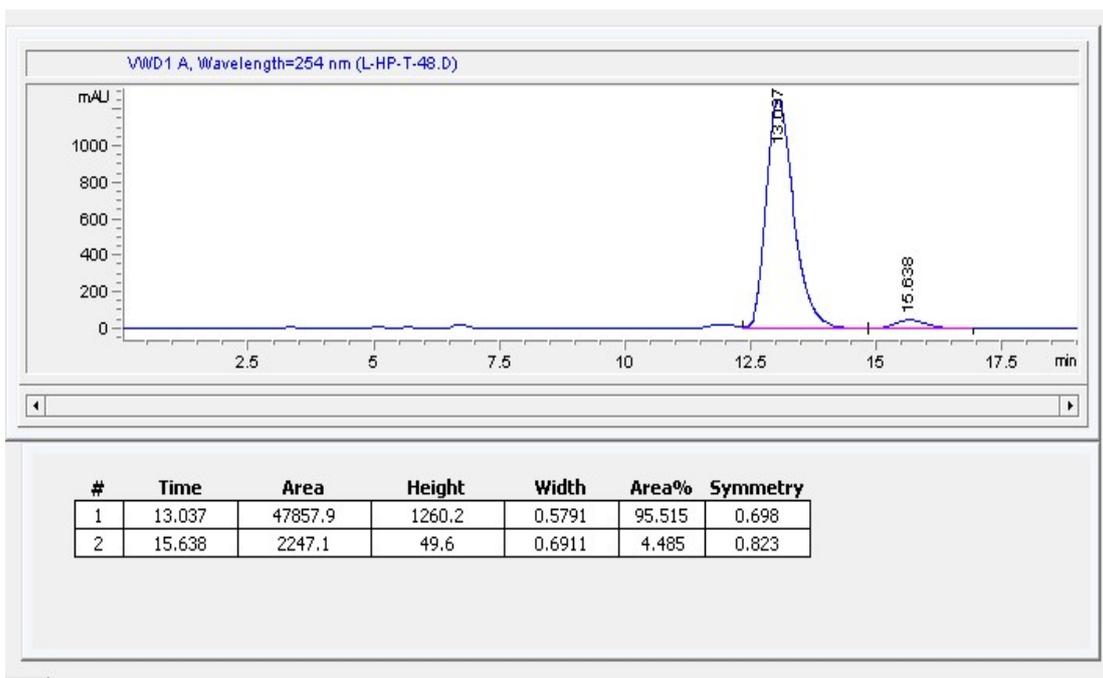
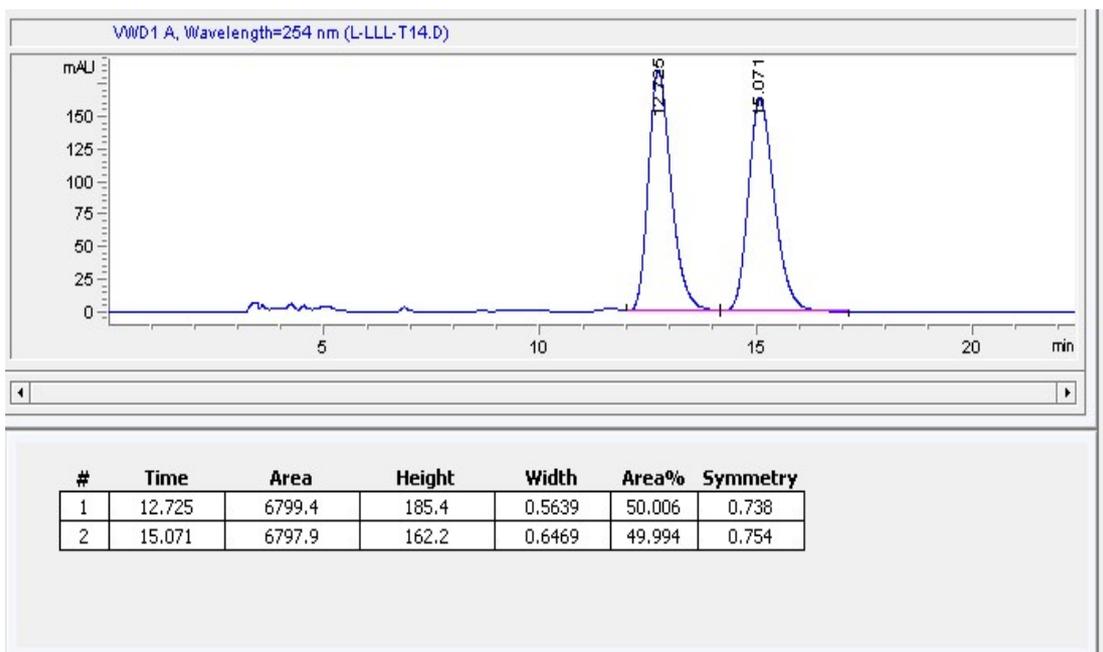
HPLC of 6dd



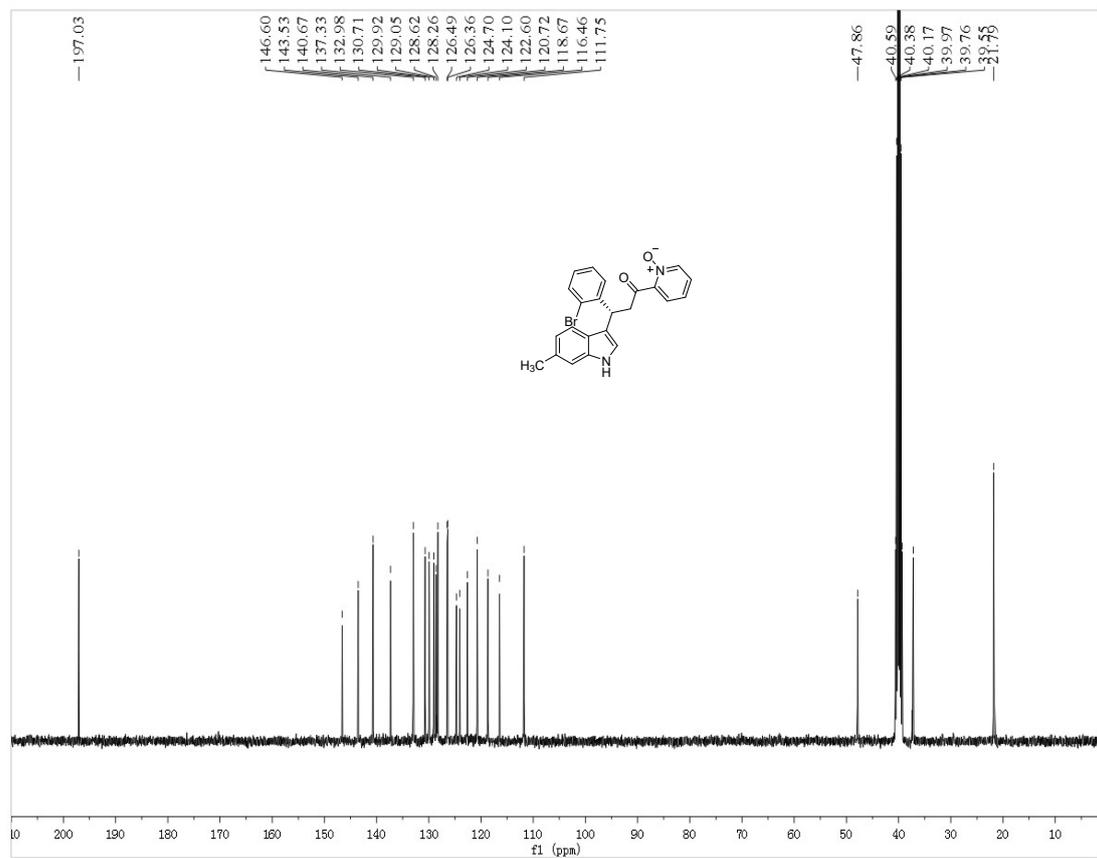
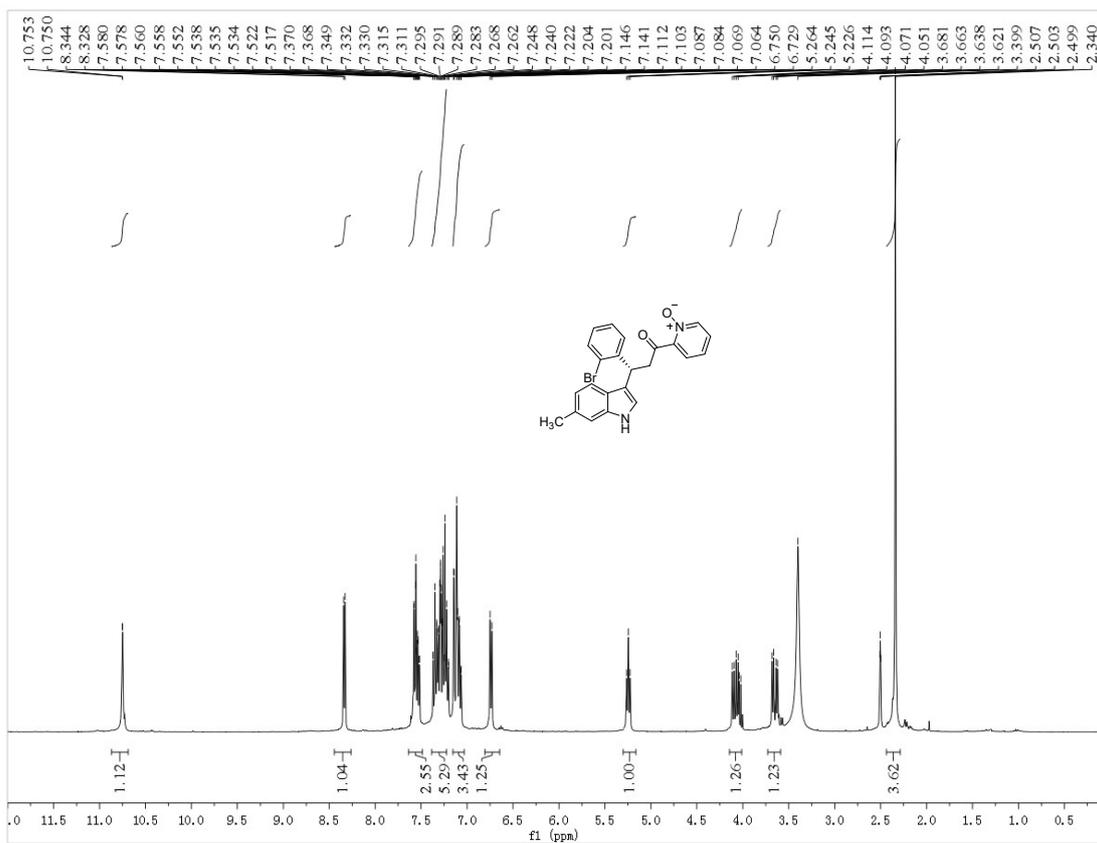
¹H and ¹³C NMR of 6de



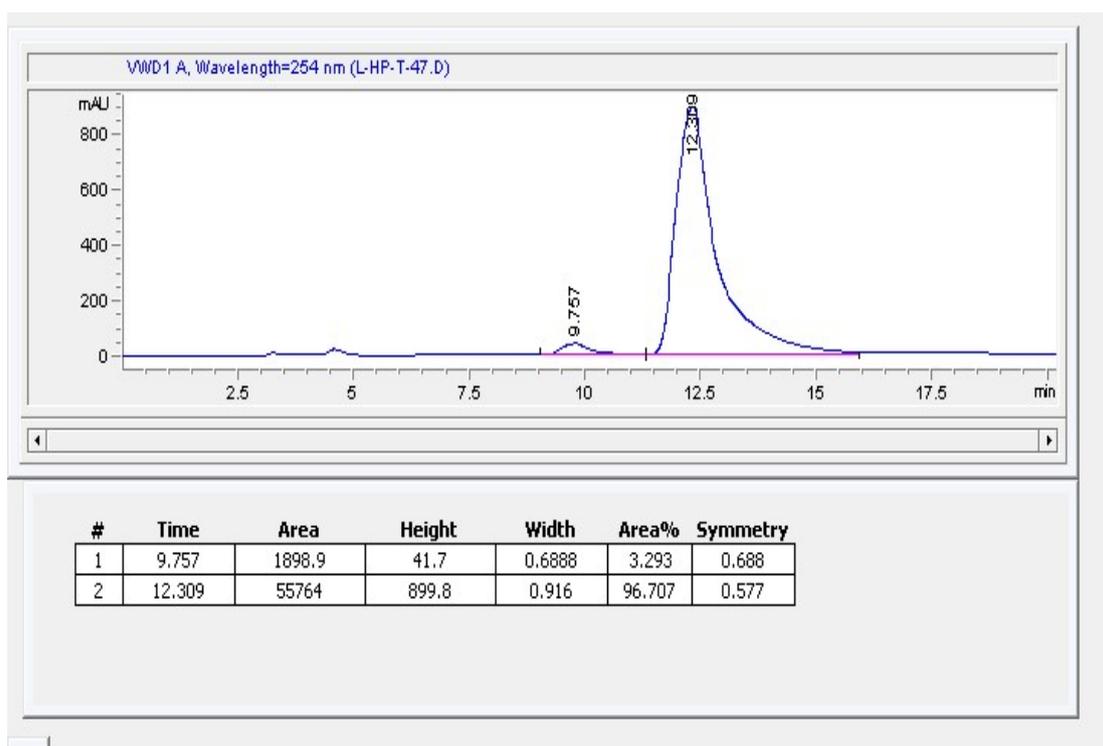
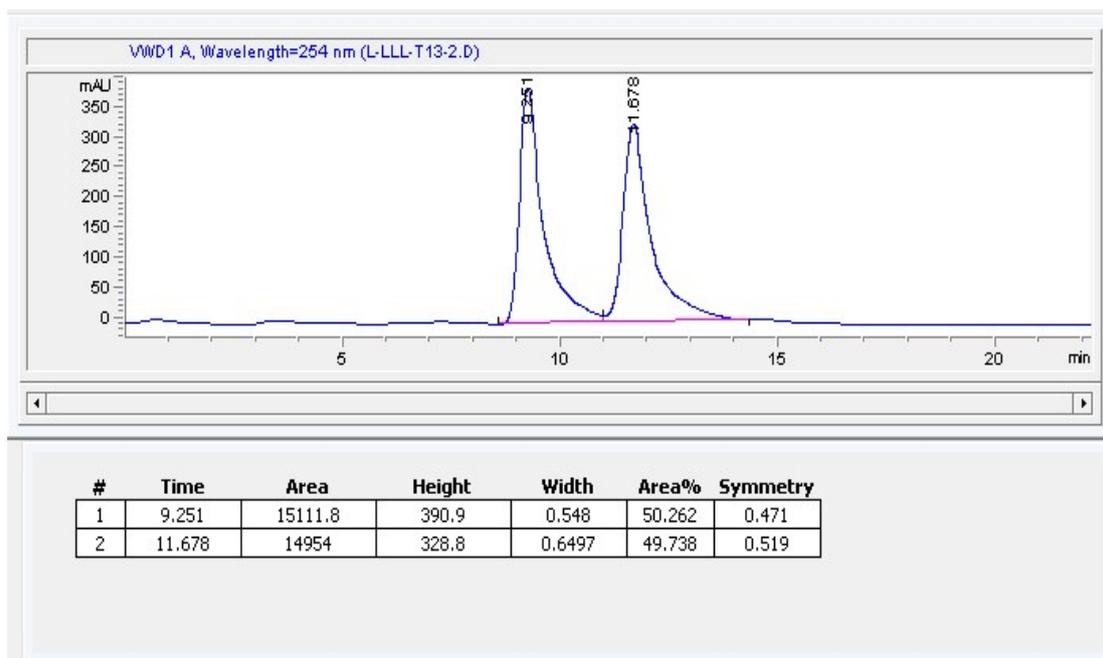
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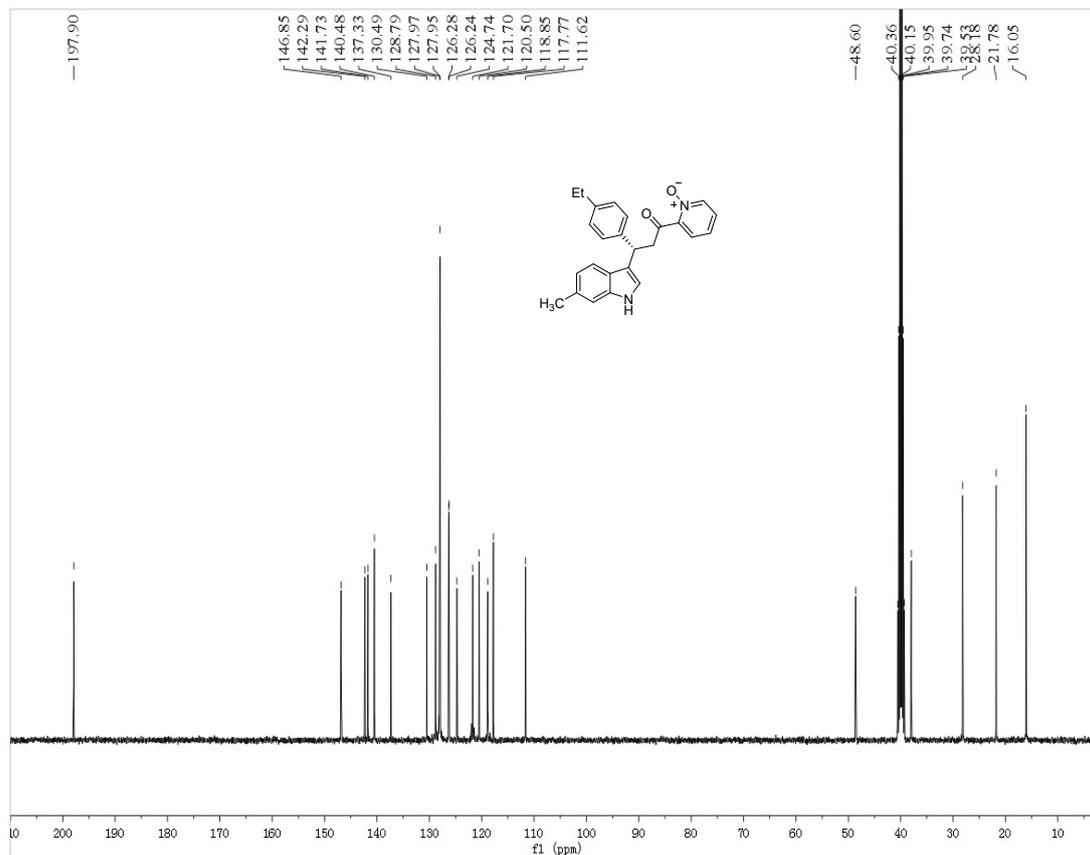
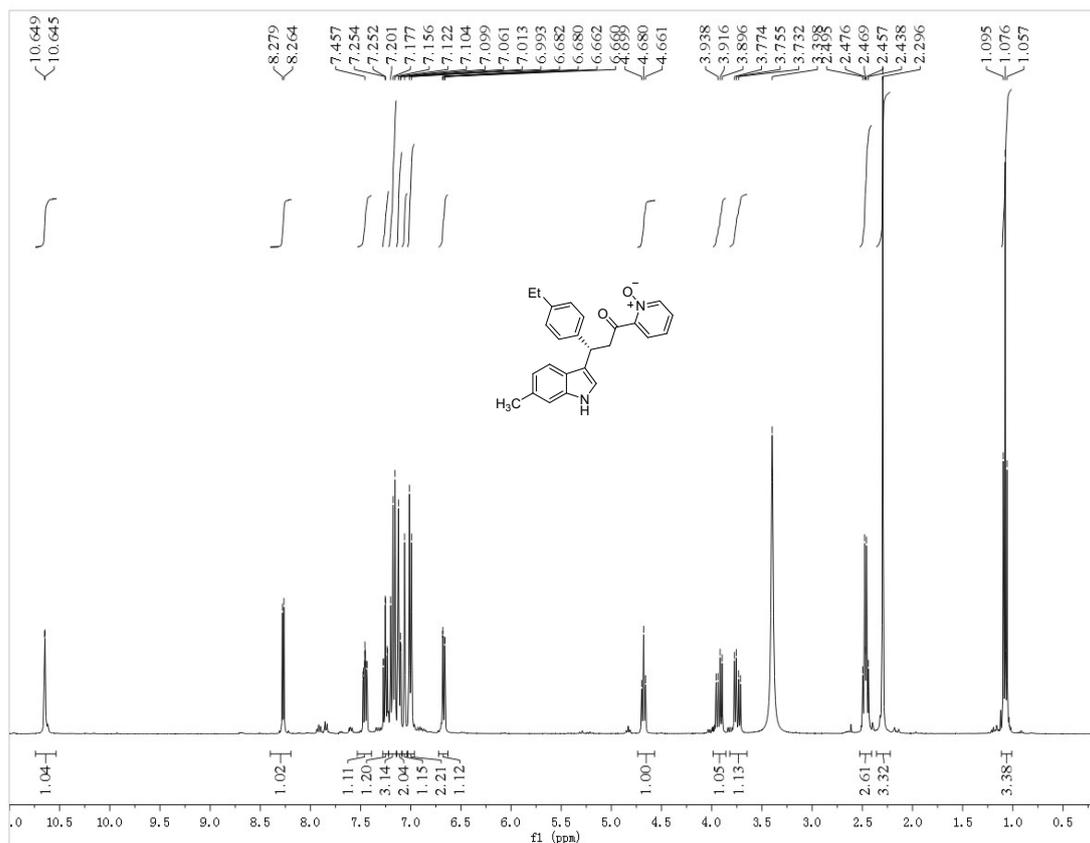
¹H and ¹³C NMR of 6df



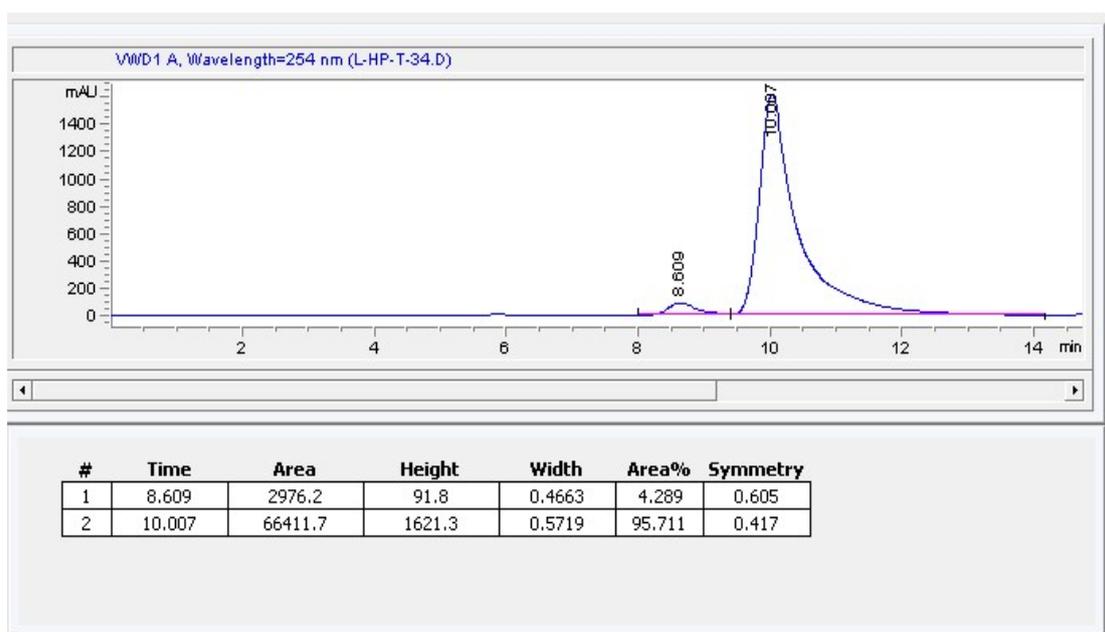
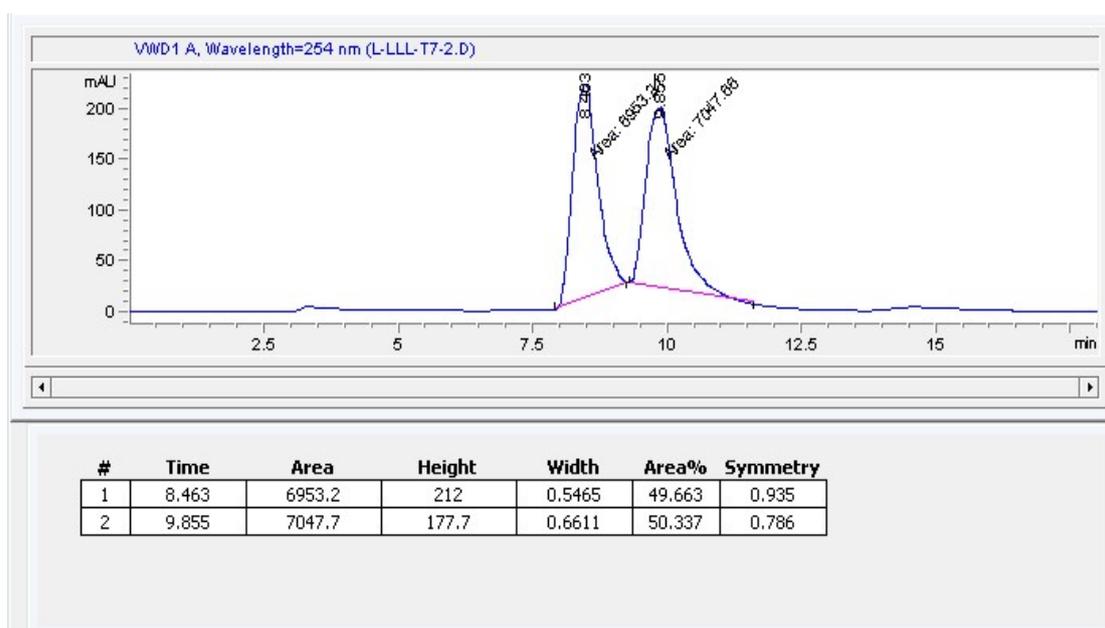
HPLC of 6df



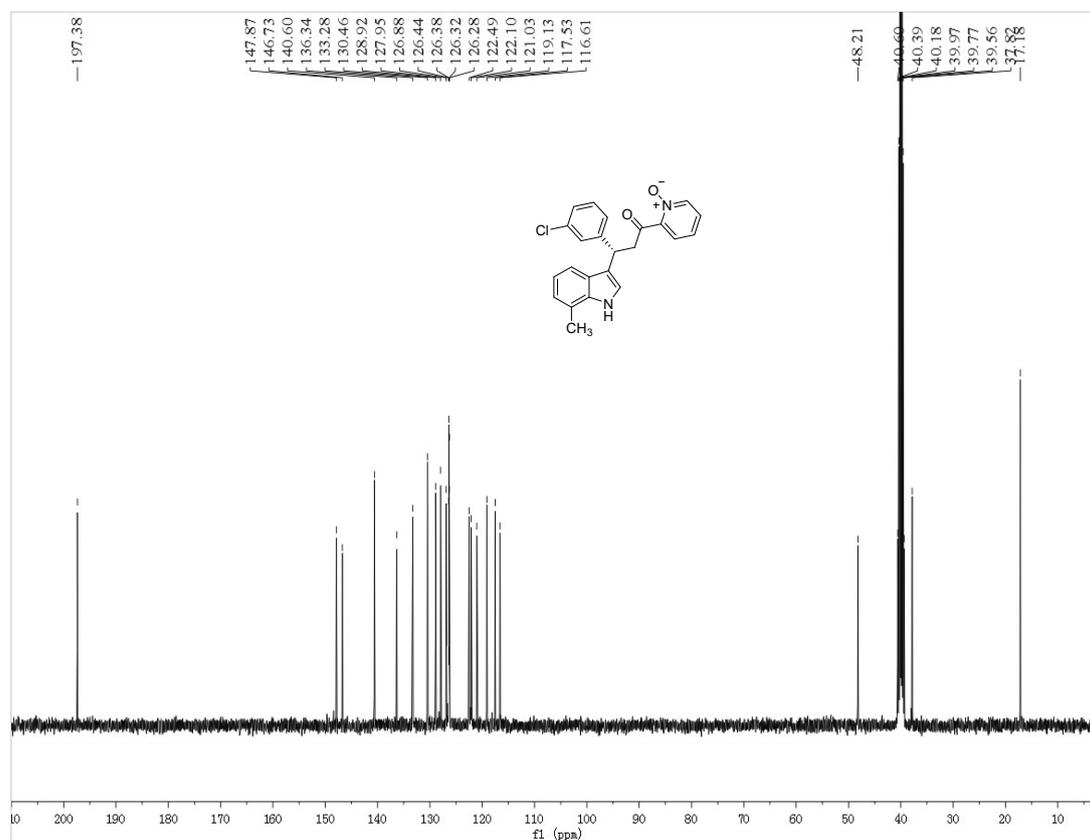
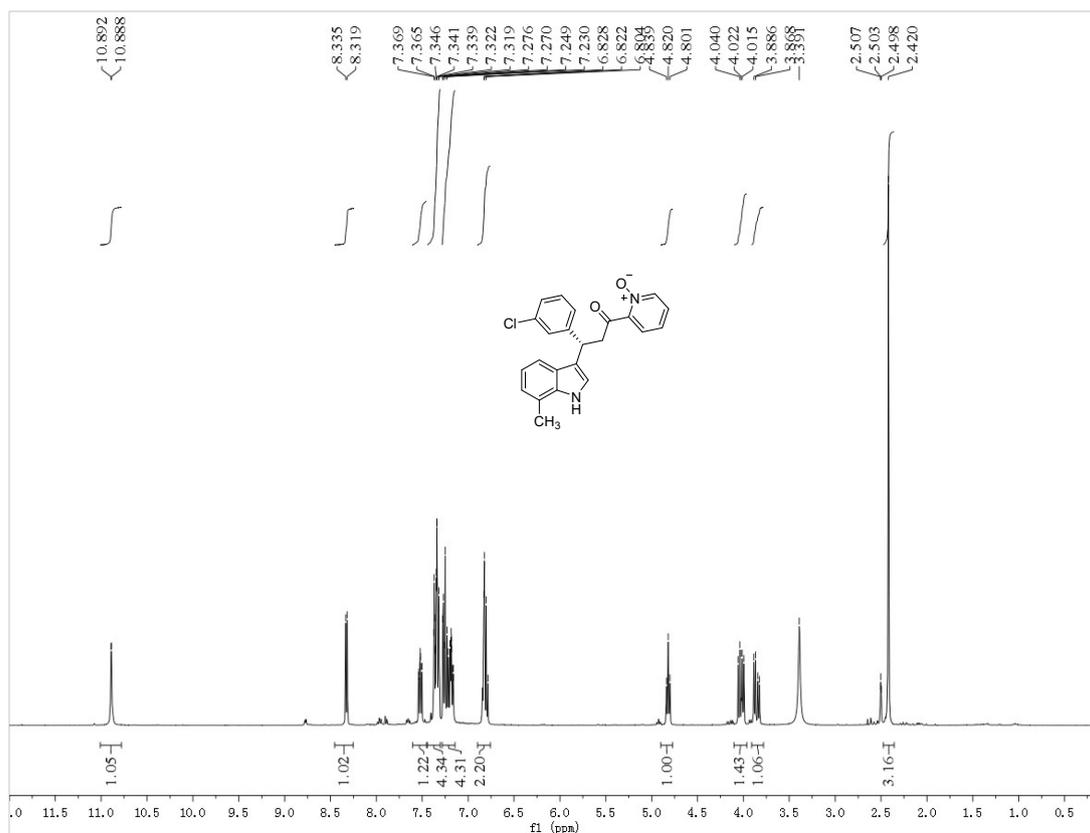
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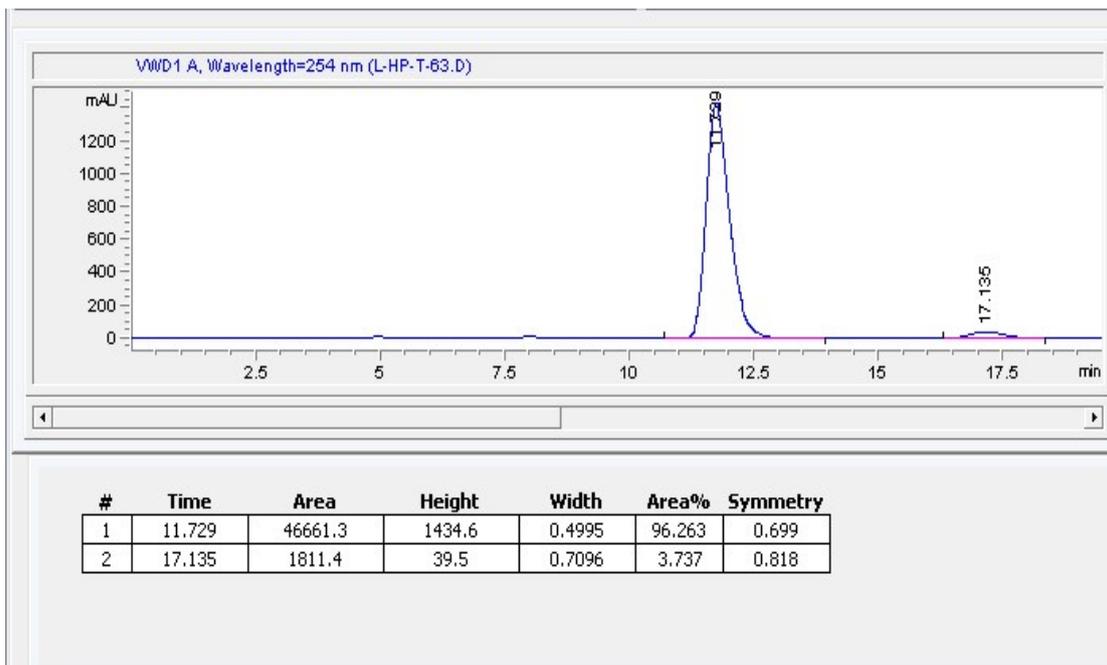
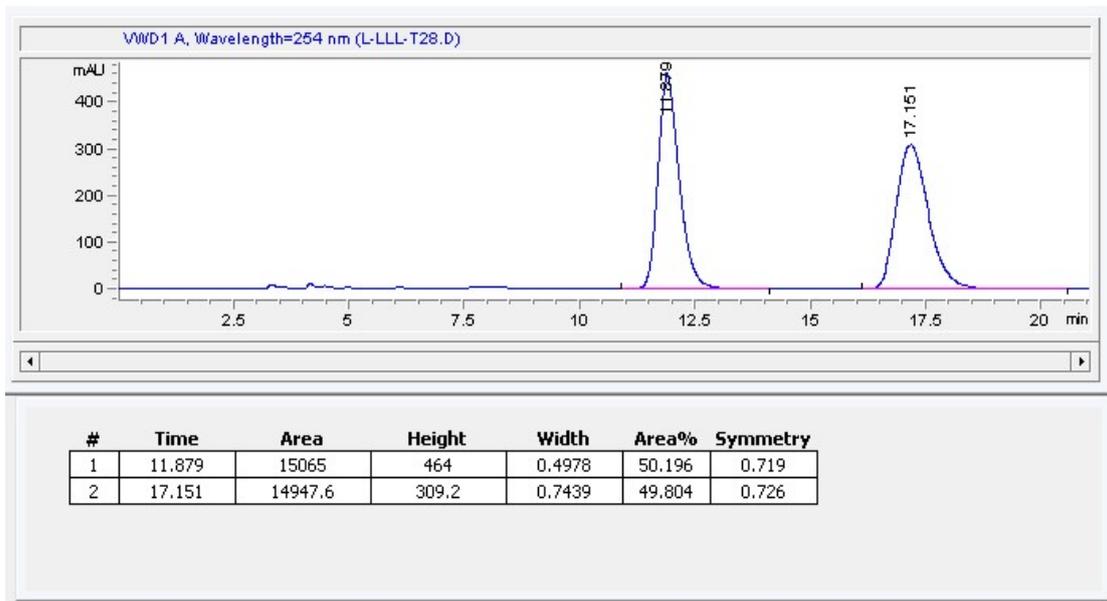
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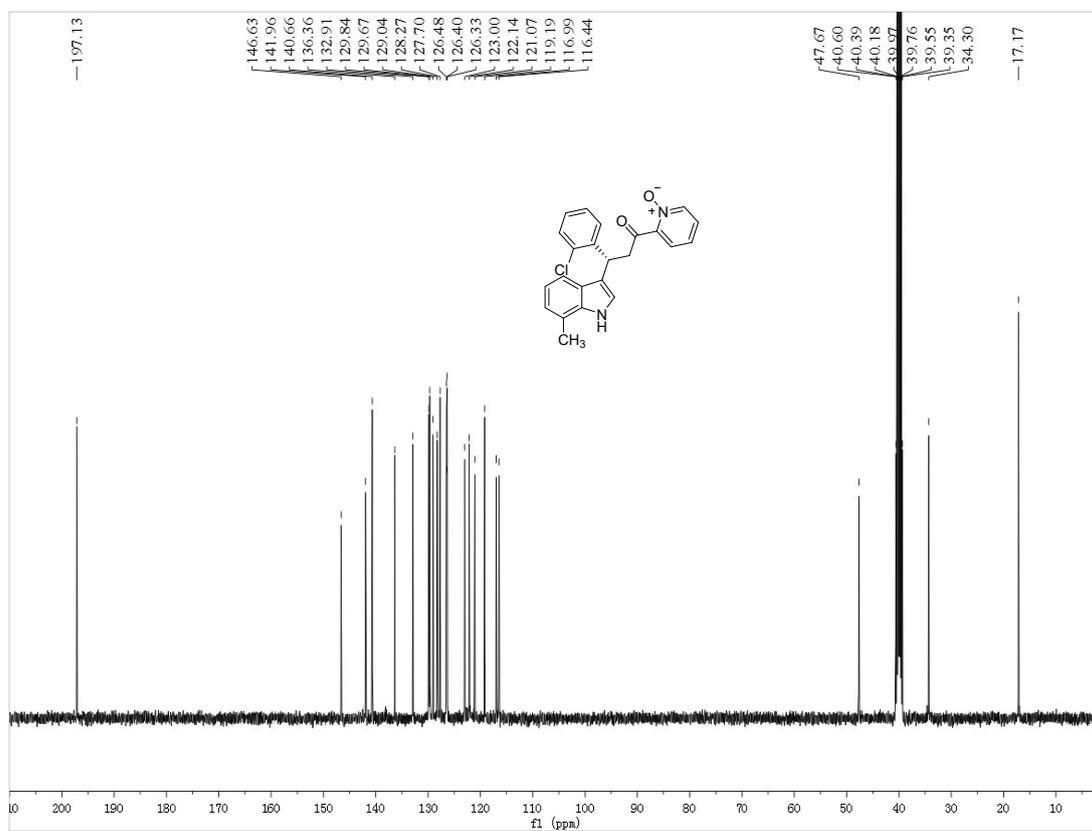
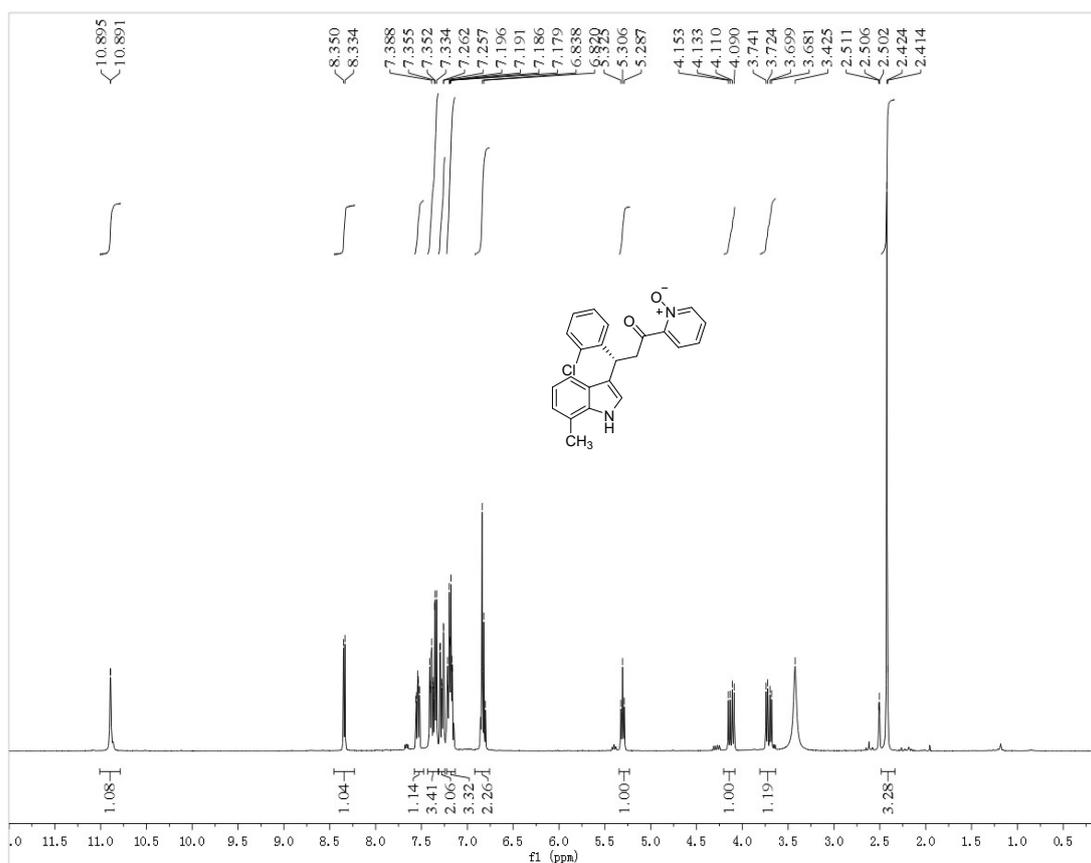
¹H and ¹³C NMR of 6ea



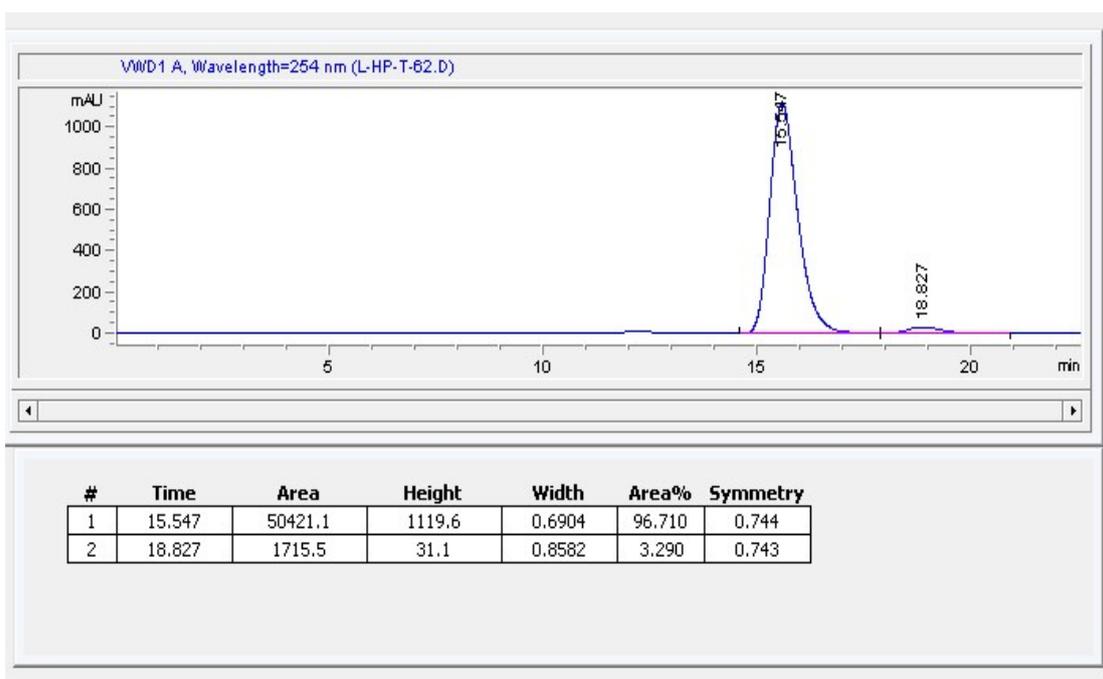
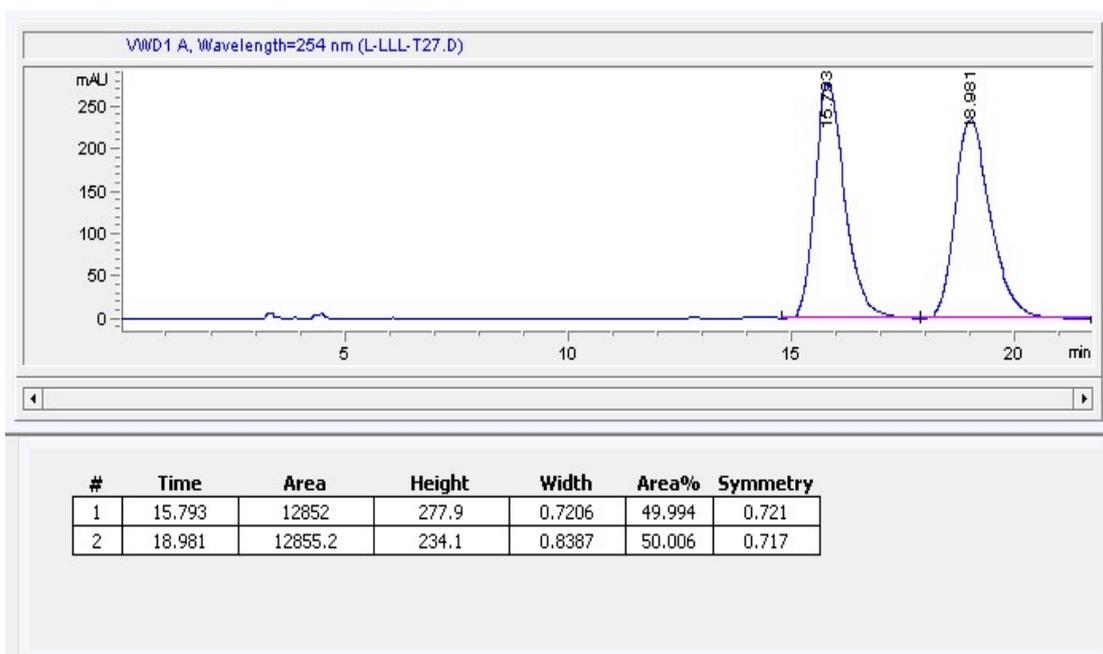
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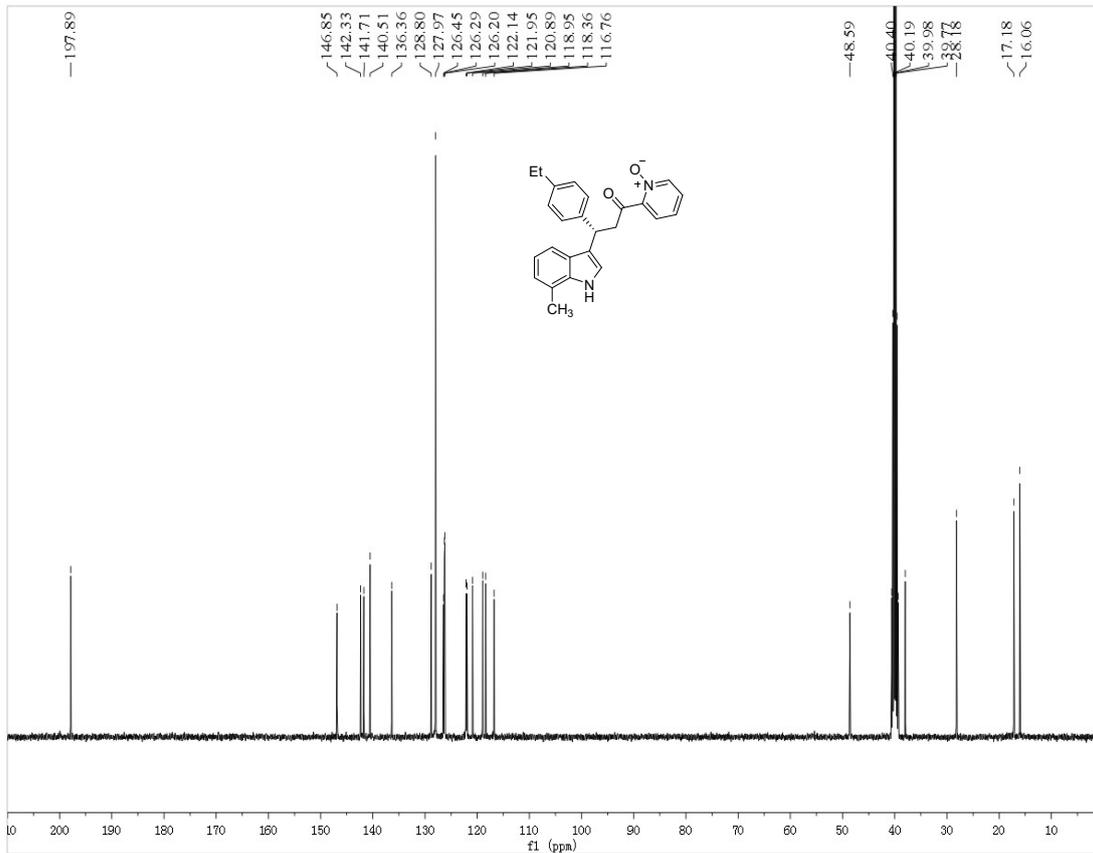
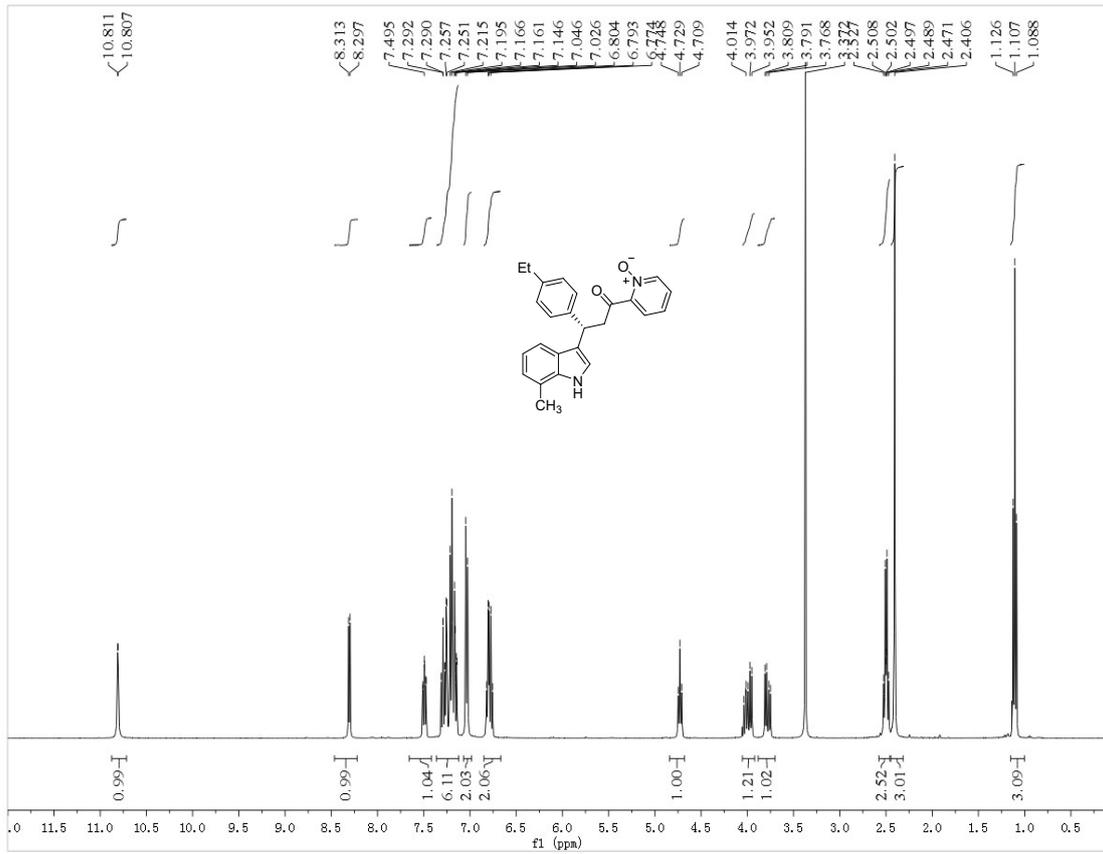
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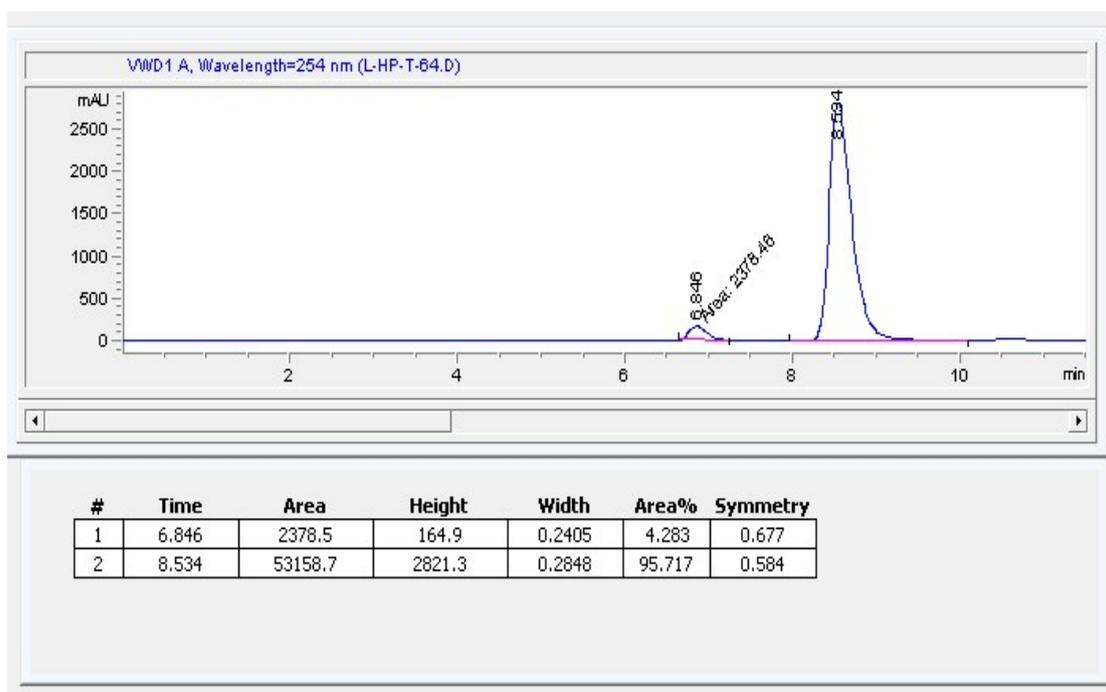
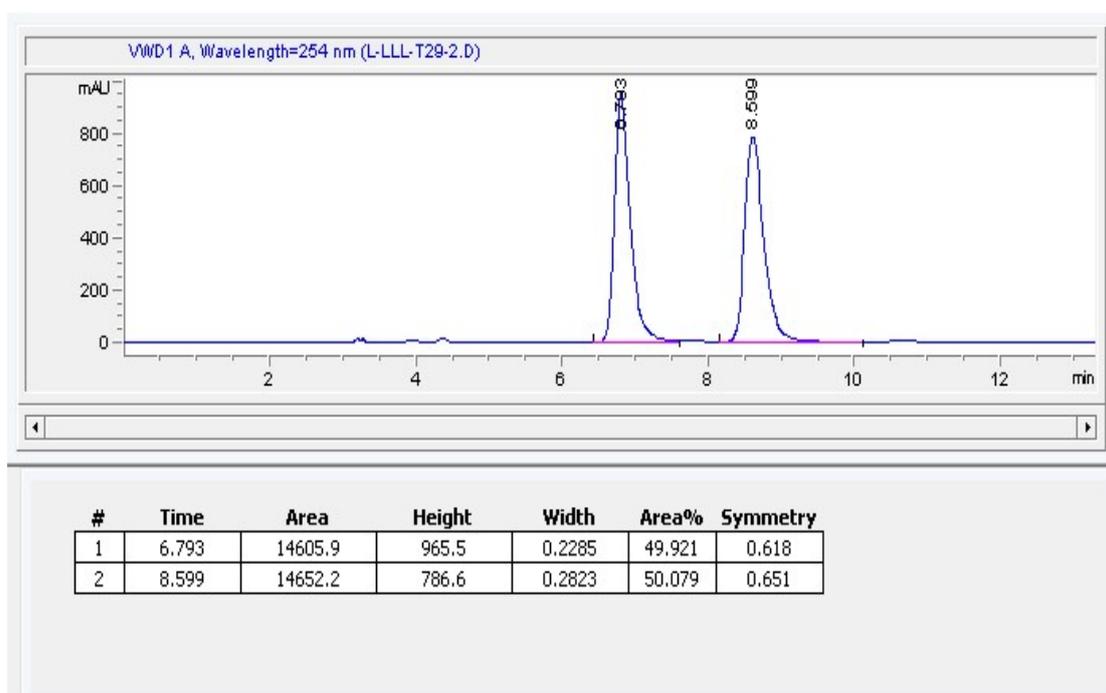
HPLC of 6eb



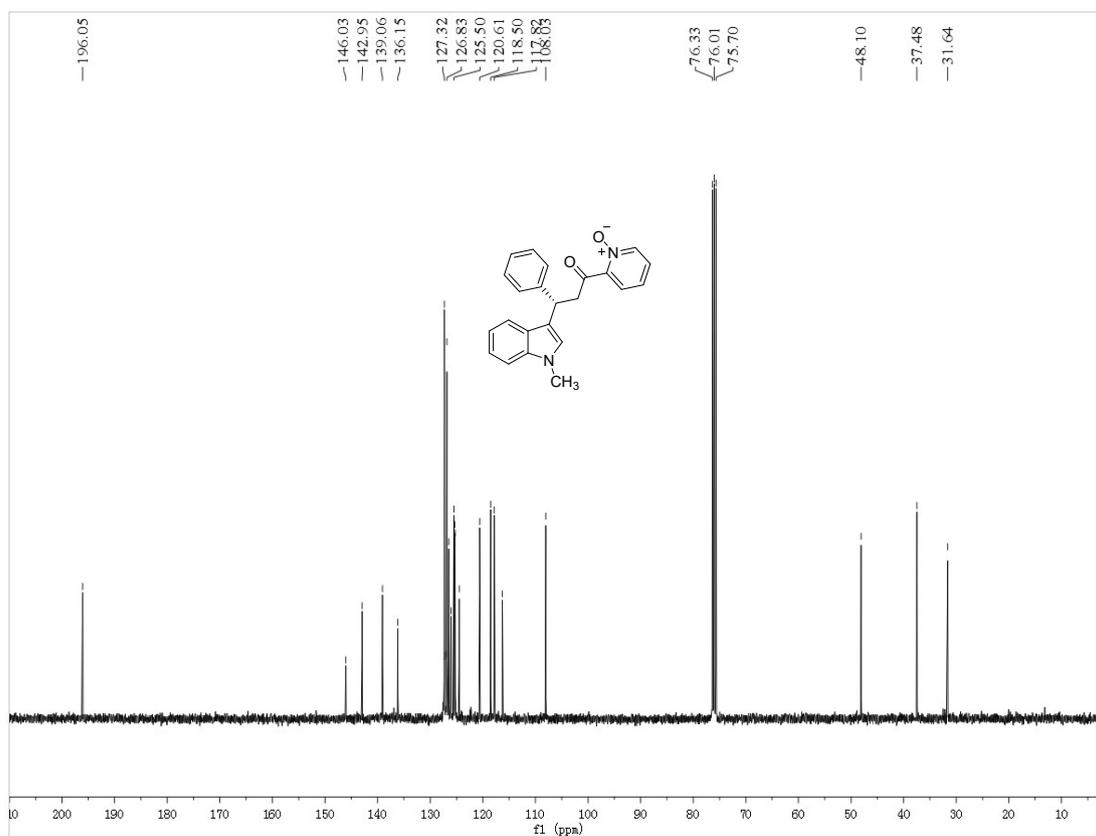
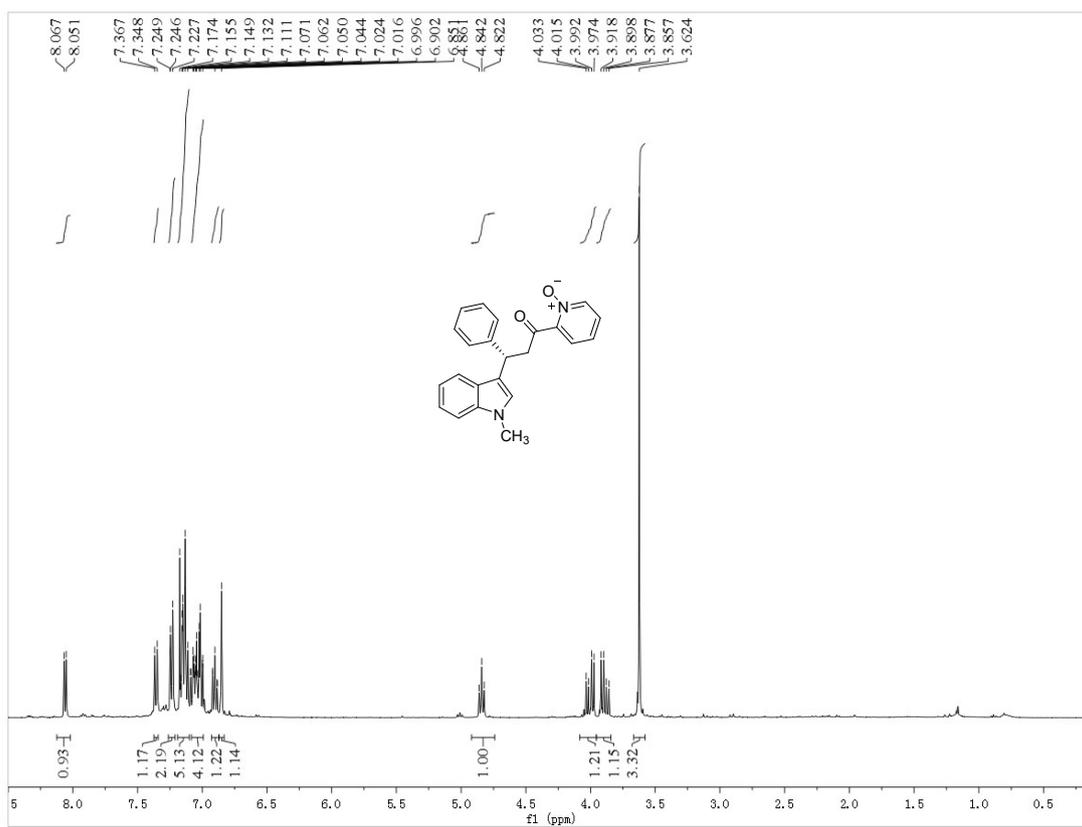
¹H and ¹³C NMR of 6ec



HPLC of 6ec



¹H and ¹³C NMR of 6fa



HPLC of 6fa

