

Supplemental Information

Rapid detection and visible light driven photocatalytic degradation of chloramphenicol in aqueous medium using $\text{CoAl}_2\text{O}_4/\text{rGO}$ nanocomposite

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Figure S21	Schematic illustration of CAP real sample analysis.
Table 1	Real sample analysis of CAP in water
Table 2.	Comparison with literature reports for CAP detection

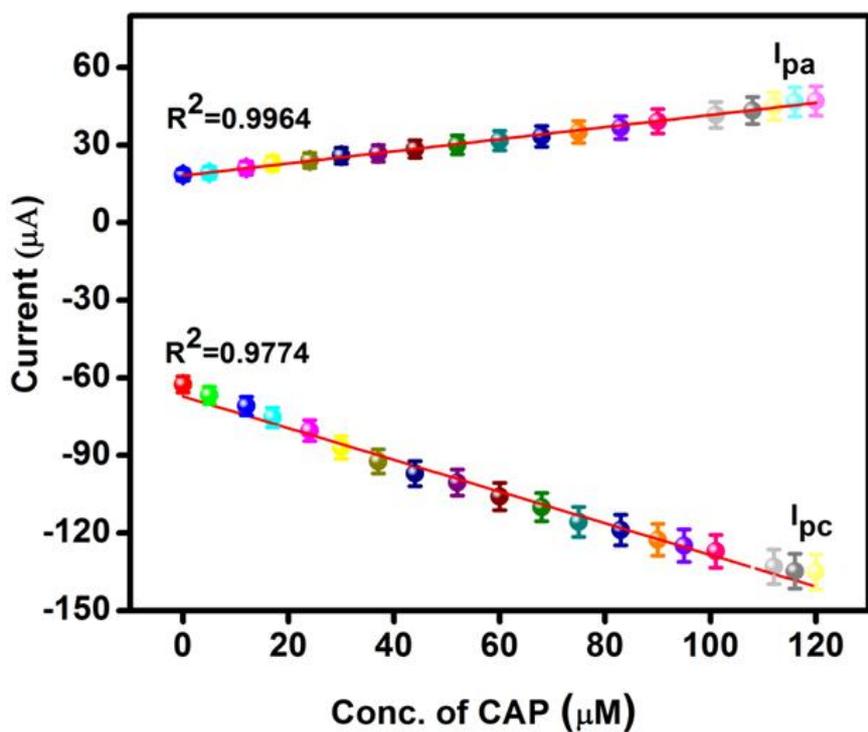


Figure S1. Linear calibration plot of CV showing the linear change in peak current of $\text{CoAl}_2\text{O}_4\text{-rGO/GCE}$ respectively with increasing concentration of CAP.

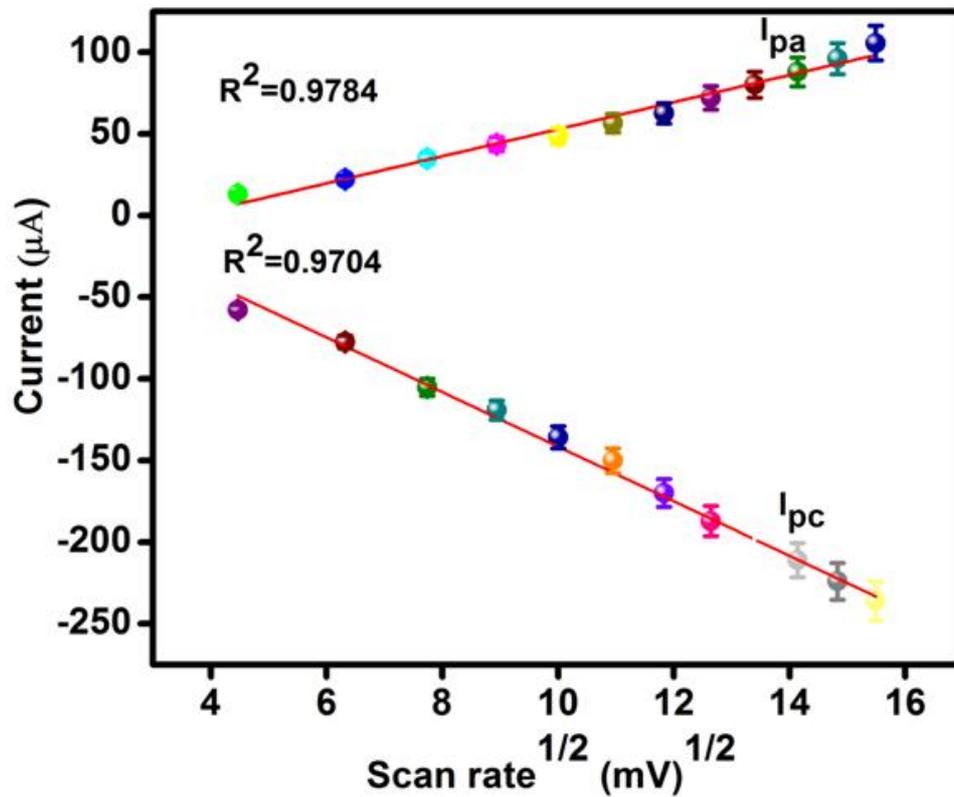


Figure S2. Plot of square root scan rate vs current with error bars

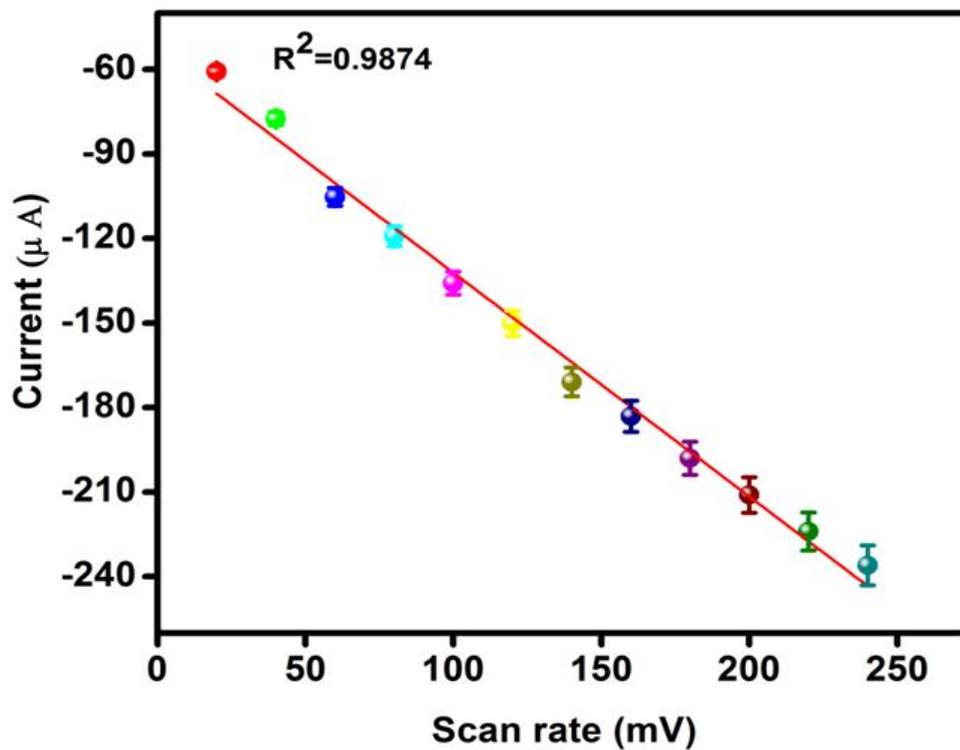


Figure S3. Plot of scan rate vs current

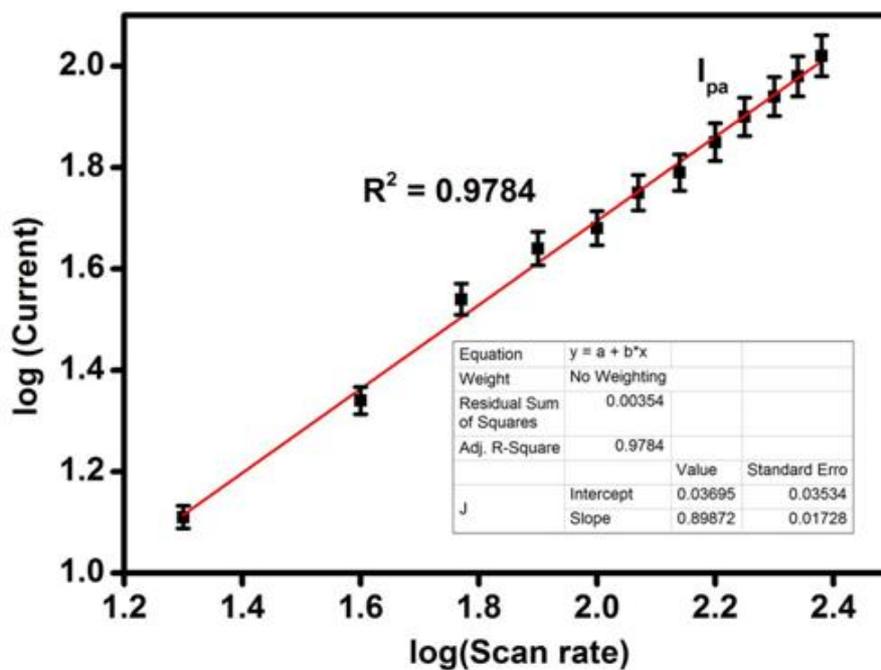


Figure S4. Linear regression plot between log current versus log of scan rate.

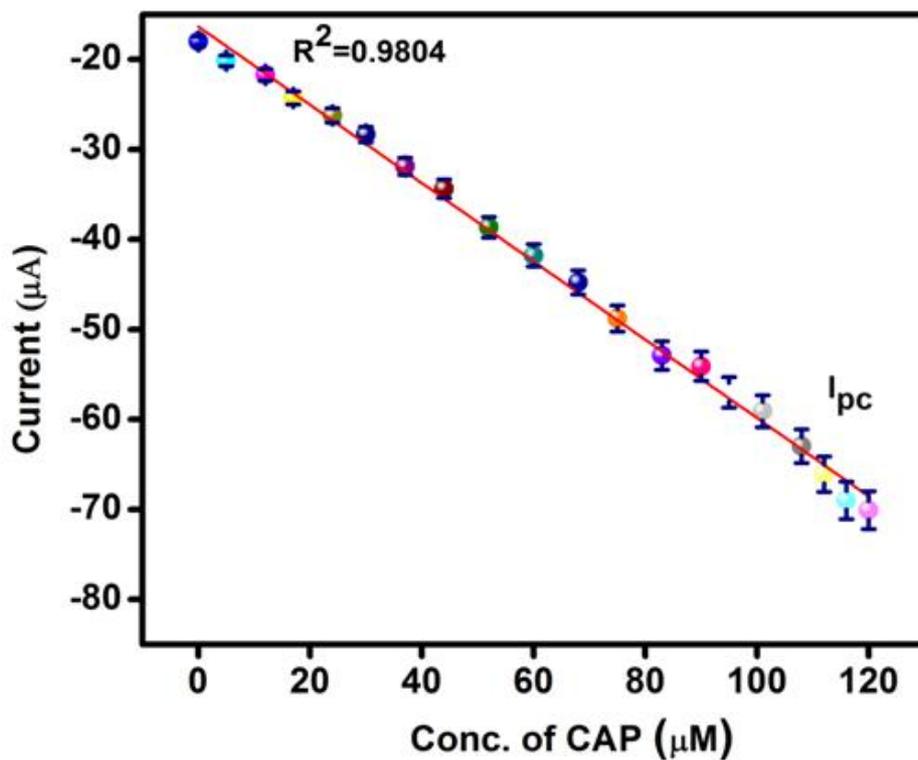


Figure S5. Linear calibration plot of LSV showing the linear change in peak current with a concentration of CAP.

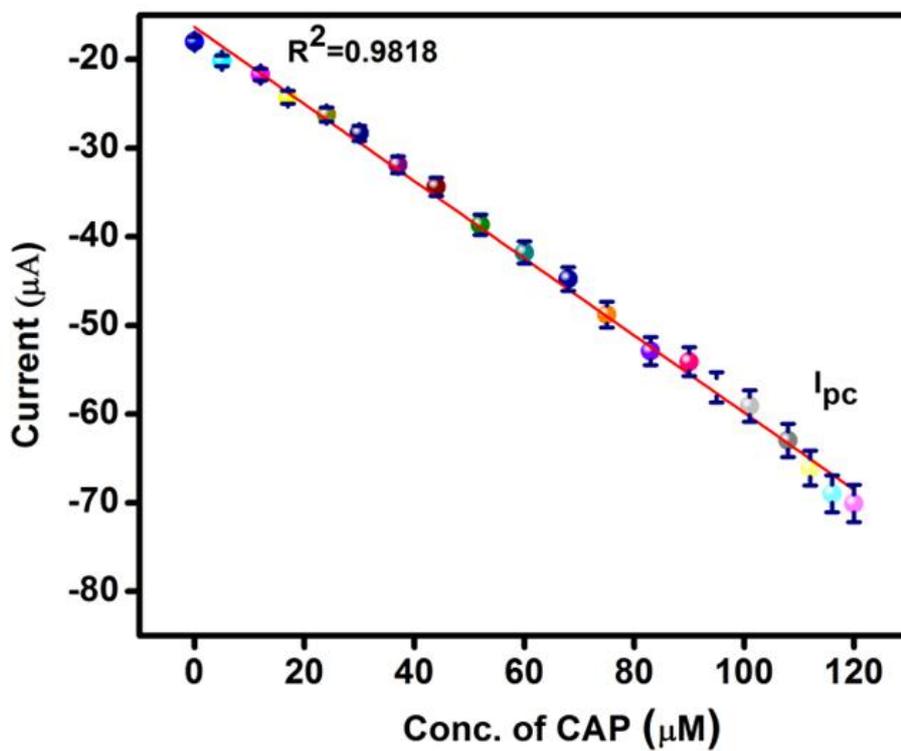


Figure S6. Linear calibration plot of DPV showing the linear change in peak current with a concentration of CAP.

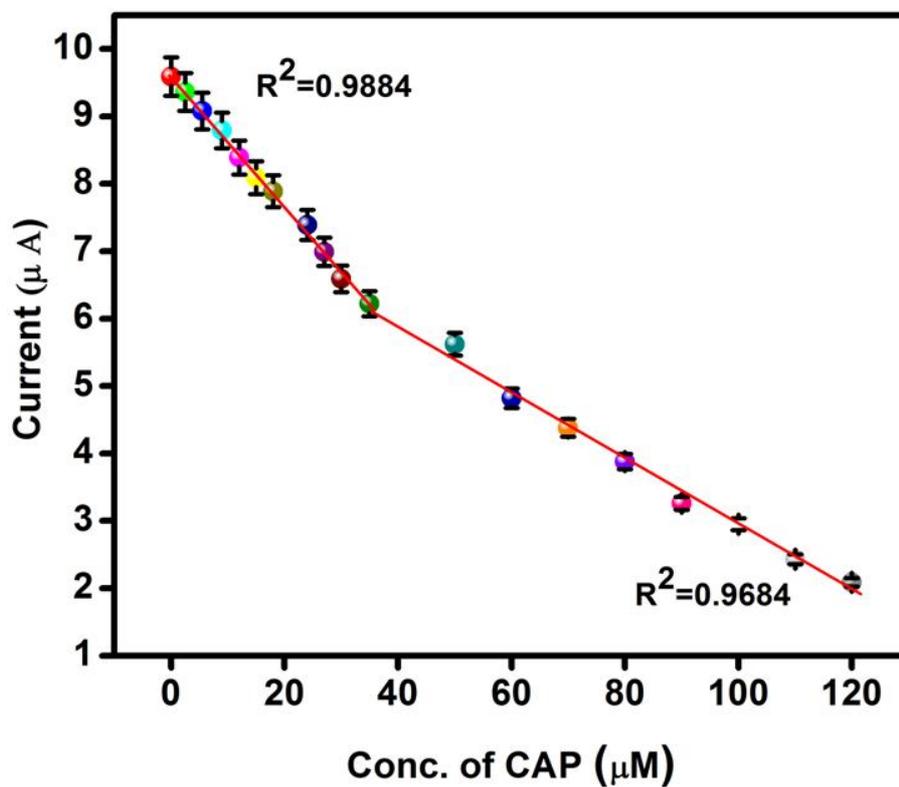


Figure S7. Linear calibration plot of the amperometry study after successive addition of CAP.

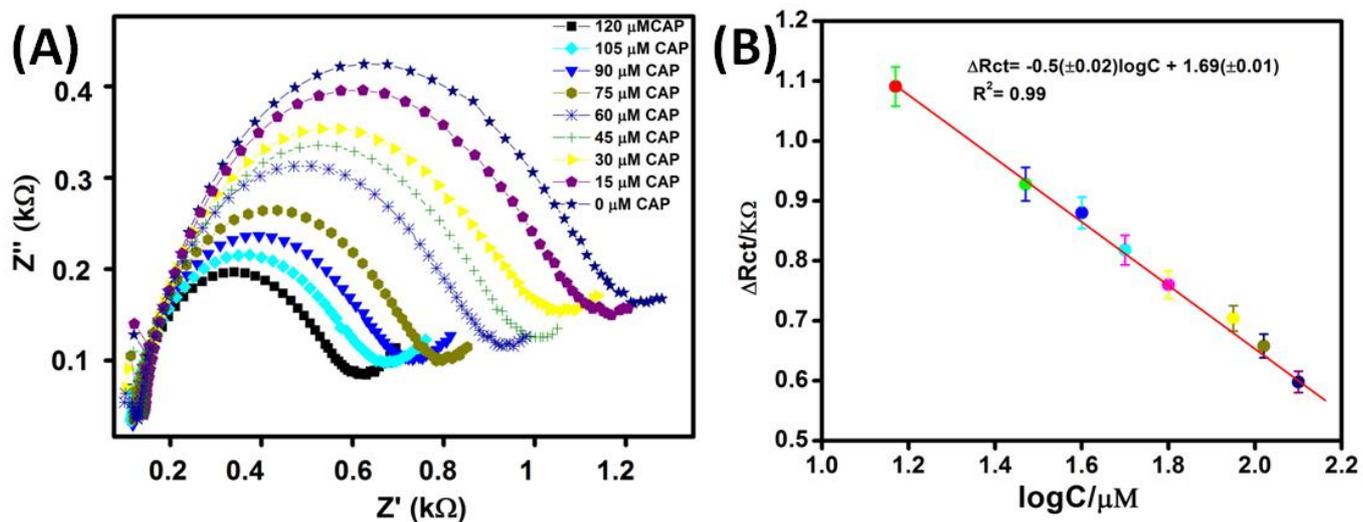


Figure S8. (A) EIS measurements for various concentrations of CAP at rGO-CoAl₂O₄/GCE. (B) The calibration curve for the determination of CAP.

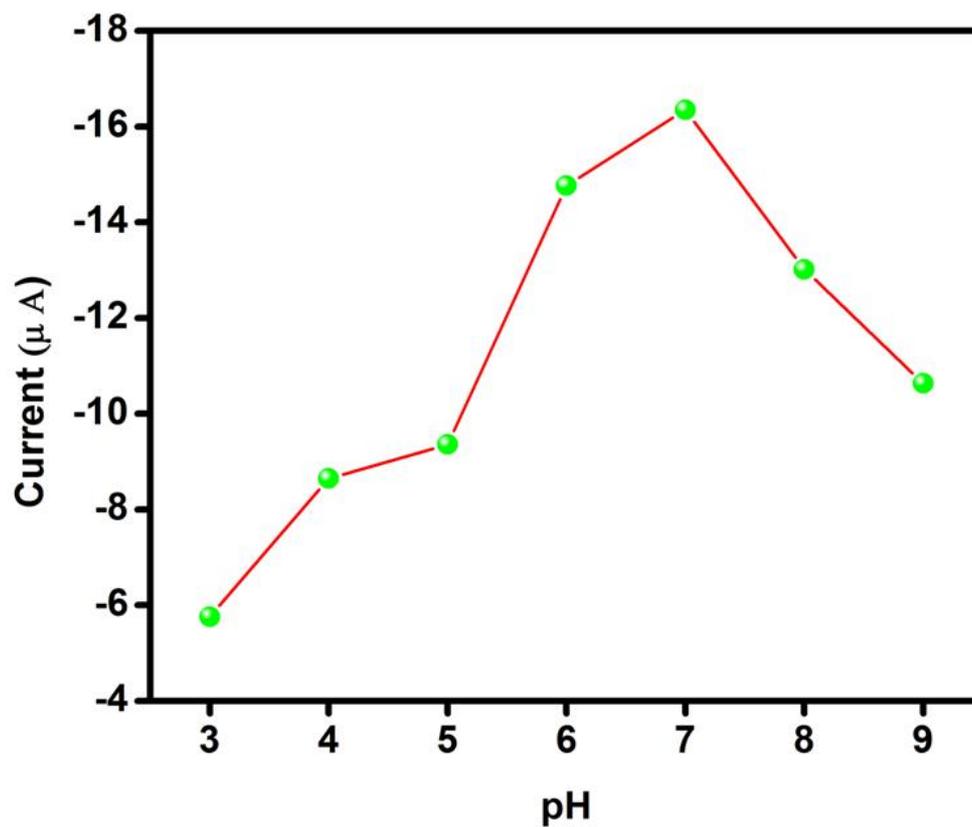


Figure S9. Plot of pH vs. current in 0.1 M PBS containing 120 μM solution of CAP at the CoAl₂O₄-rGO.

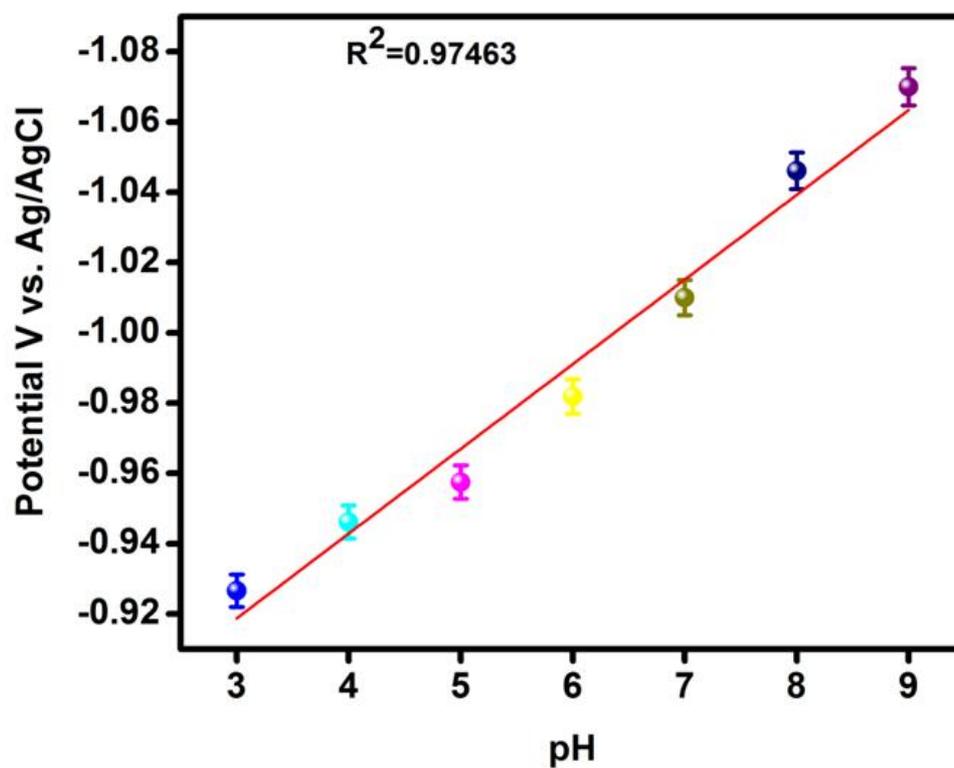


Figure S10. Correlation of pH with Ep.

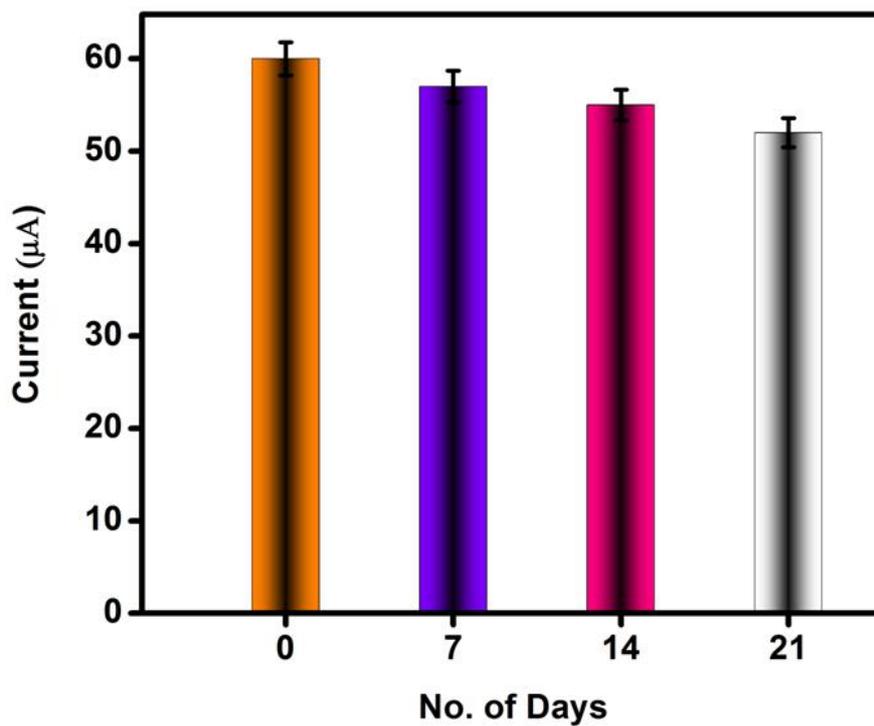


Figure S11. Bar graph showing the stability of response of $\text{CoAl}_2\text{O}_4\text{-rGO}$ for CAP detection with number of days.

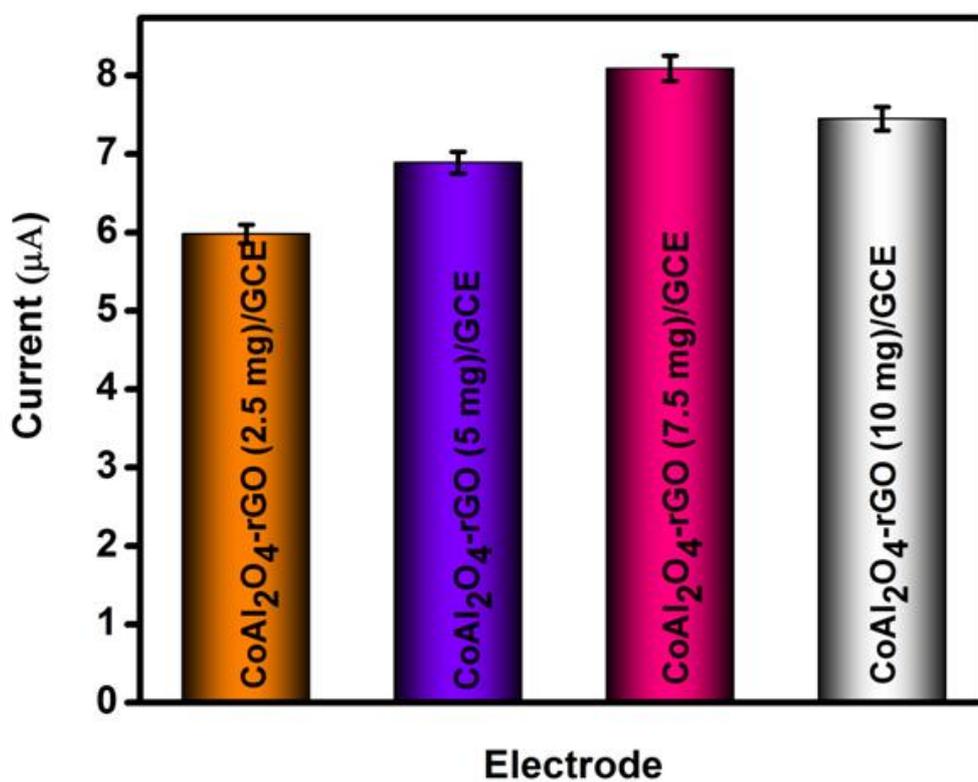


Figure S12. Reduction peak currents of 2.5, 5, 7.5 and 10 mg $\text{CoAl}_2\text{O}_4\text{-rGO}/\text{GCE}$ of $120 \mu\text{M}$ CAP in 0.1 M PBS pH 7.00 at scan rate 100 mV s^{-1}

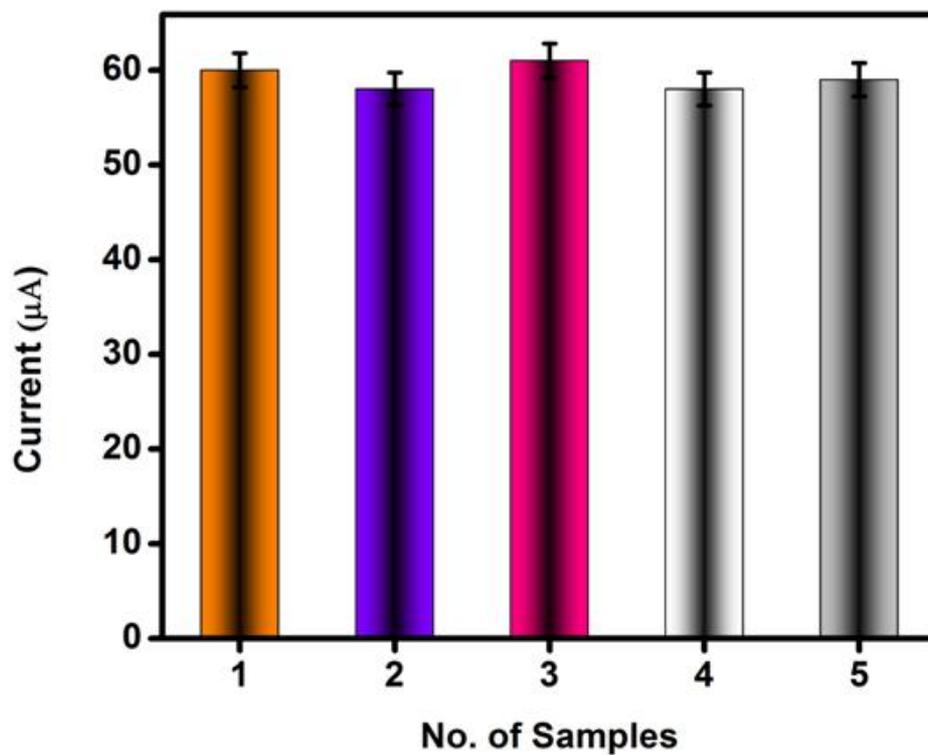


Figure S13. Bar graph showing the stability of response of $\text{CoAl}_2\text{O}_4\text{-rGO}$ for CAP detection with number of different samples.

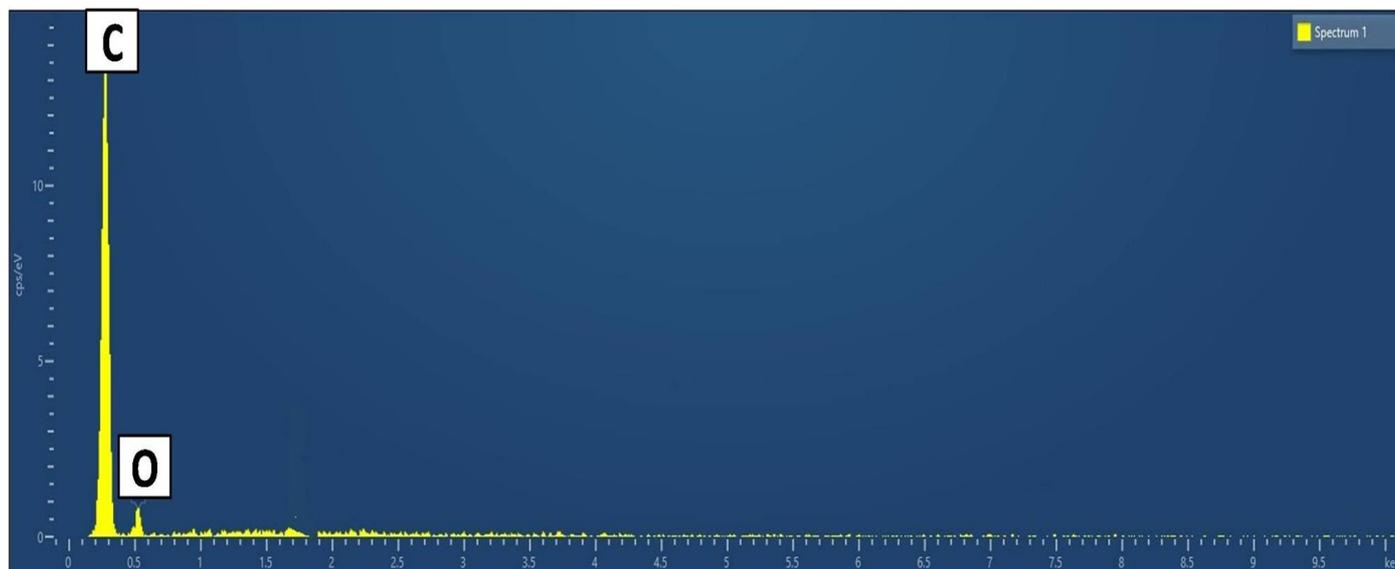


Figure S14. EDAX analysis of rGO.

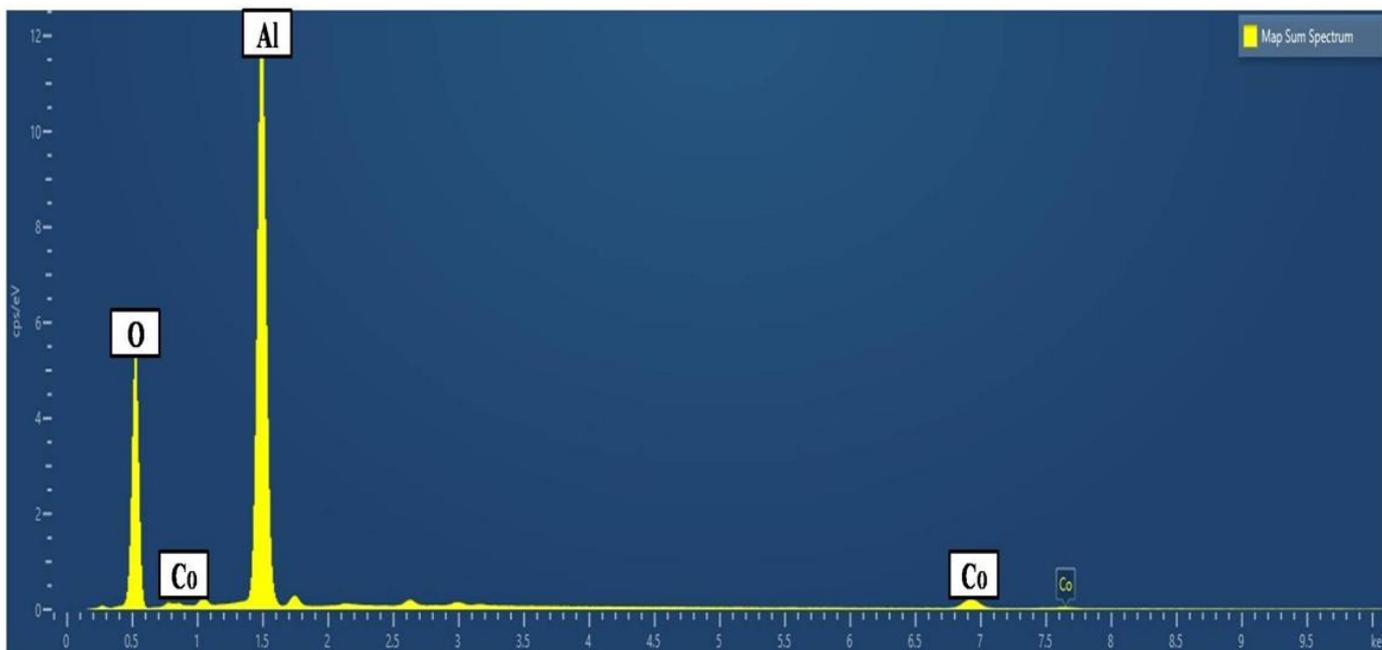


Figure S15. EDAX analysis of CoAl₂O₄.

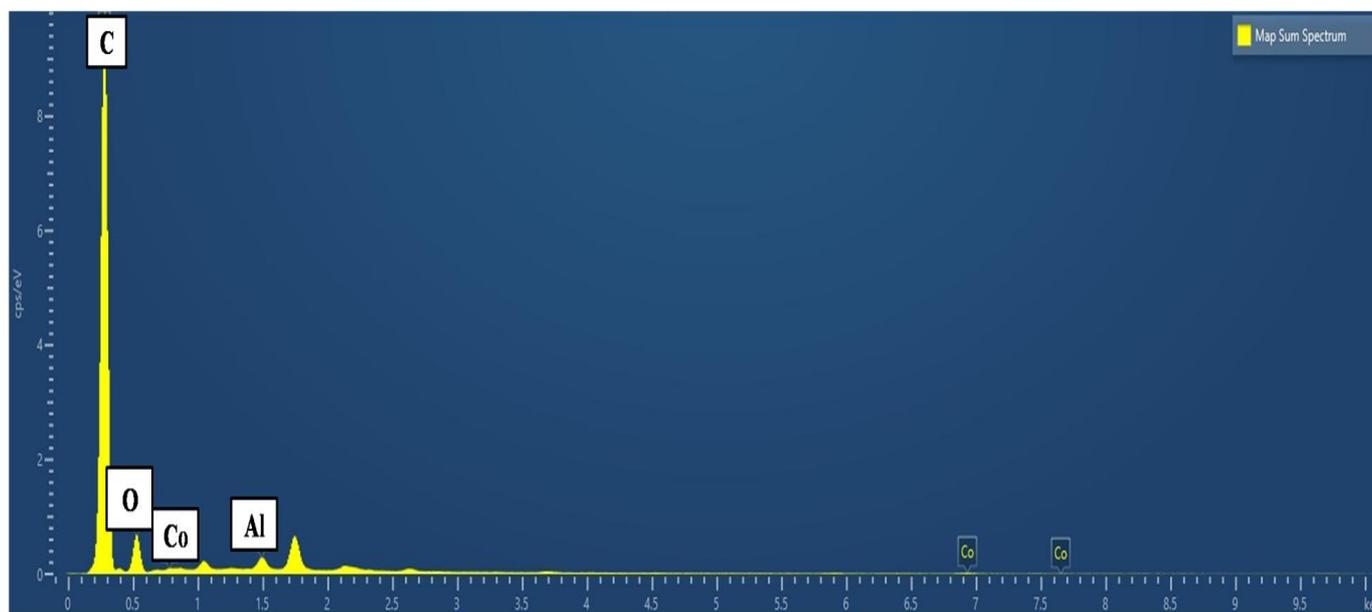


Figure S16. EDAX analysis of rGO@CoAl₂O₄.

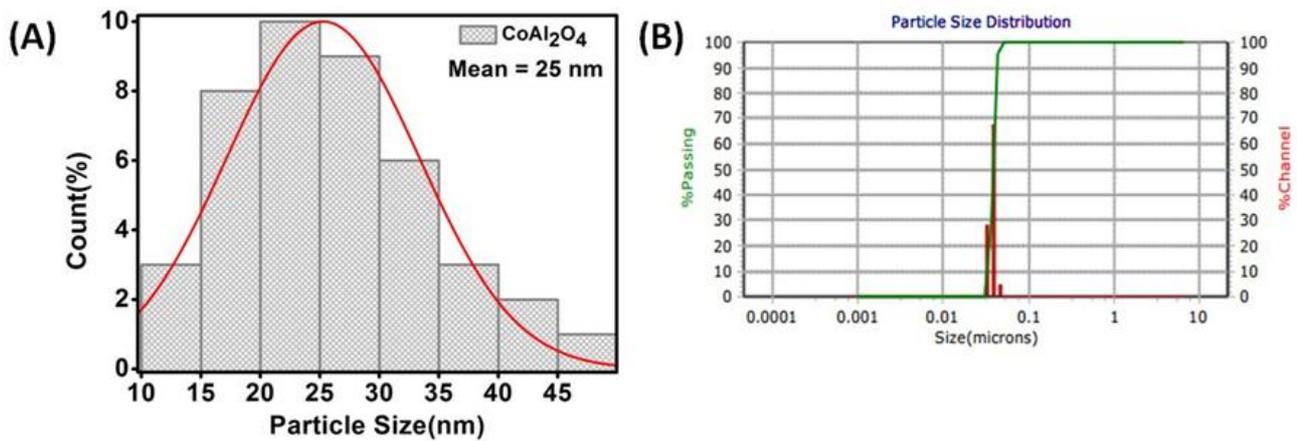


Figure S17. (A) The particle size distribution histogram of CoAl_2O_4 nanoparticles, (B) DLS histogram of CoAl_2O_4 nanoparticles

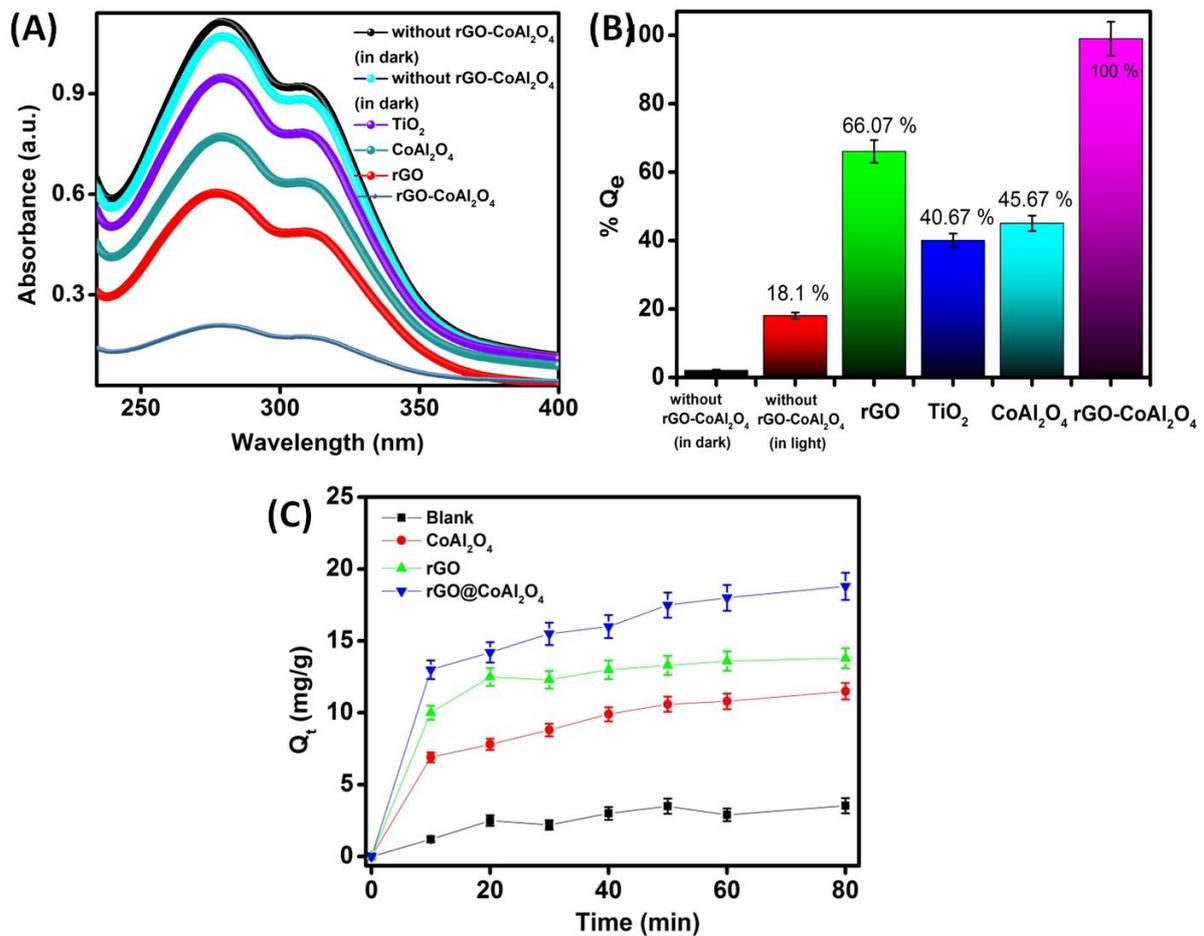


Figure S18. (A) Absorption spectra of CAP photocatalytic degradation under the influence of various catalysts, (B) Photocatalytic degradation efficiencies of various catalysts and (C) adsorption capacities of various catalysts.

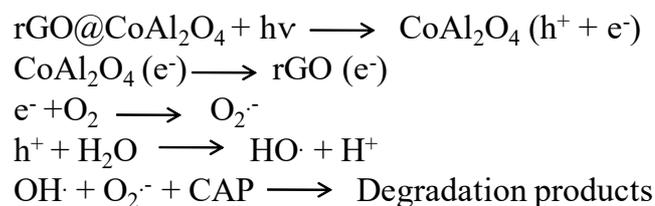


Figure S19. Reactions involving degradation of CAP using rGO@CoAl₂O₄.

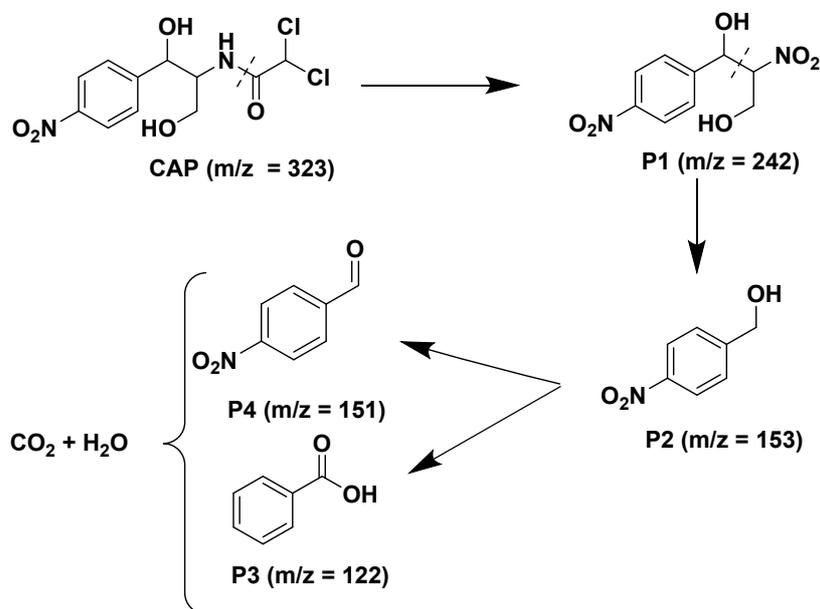


Figure S20. Degradation products of CAP using CoAl₂O₄/rGO nanocomposite.

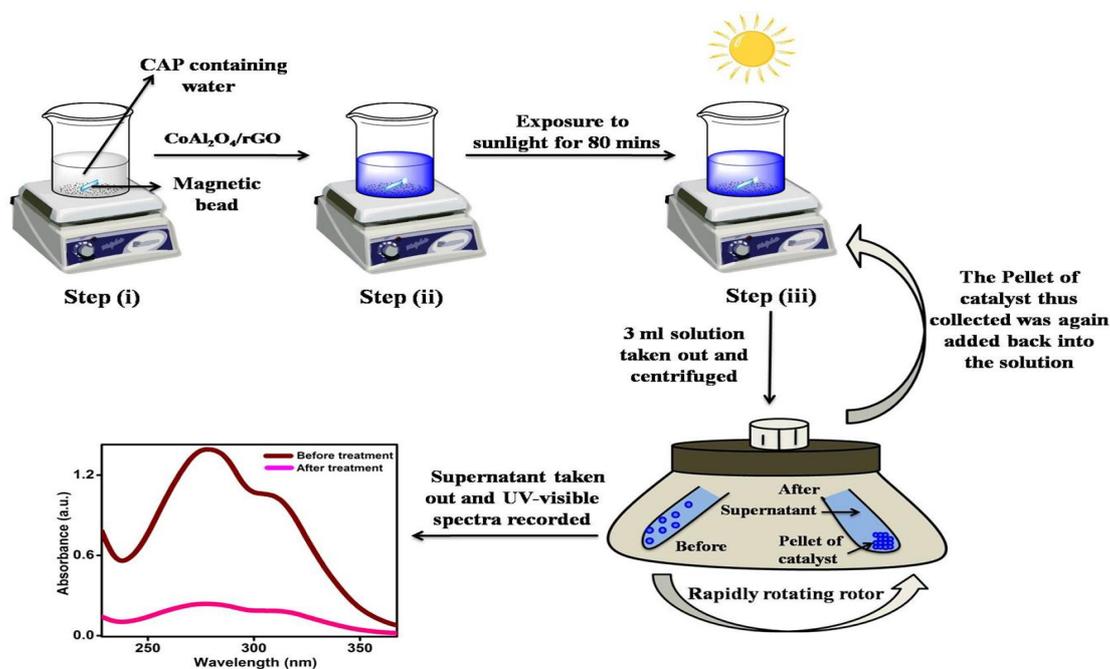


Figure S21: Schematic illustration of CAP real sample analysis.

Real sample analysis of CAP in water

Table 1.

Sample	Added/ 10^{-6} mol/L	Found/ 10^{-6} mol/L	RSD (%)	Degradation (%)
River water	10.00	9.18	0.83	91.80
	25.00	24.19	2.07	96.76
Tap water	10.00	9.89	0.75	98.90
	50.00	50.55	2.08	101.10
Pharmaceutical Sewage wastewater	10.00	10.36	0.43	103.6
	25.0	25.19	1.05	100.76

Table 2.

Comparison with literature reports for CAP detection

S.No.	Probe	Detection method	Application	Detection Limit	Linear range	References
1.	Mn ₃ O ₄ nanoparticles for detection of chloramphenicol	Electrochemical sensor	Milk samples	0.008 μ M	0.007-0.013 μ M	1
2.	GCE modified with CS-MWCNTs electro-polymerization for detection of chloramphenicol	Electrochemical sensor	Milk samples	3.3×10^{-2} μ M	0.1-1000 μ M	2
3.	MIP/Uio-66@CDs/GCE for detection of chloramphenicol	Electrochemical Impedance Spectroscopy (EIS)	Water samples	61×10^{-9} μ M	40×10^{-9} μ M - 61×10^{-9} μ M	3
4.	CdS _{0.75} Se _{0.25} @oligo peptide quantum dots for detection of chloramphenicol	Fluorescent spectroscopy	Milk samples	0.89 μ g/L	3.13 to 500 μ g/L	4
5.	MIP-functionalized rGO for adsorption and detection of chloramphenicol	Electrochemical detection	Honey samples	0.204 μ M	0.1 μ M – 1.2 μ M	5

6.	Sn/rGO/SPCE (Screen printed electrode) for detection of chloramphenicol	Electrochemical detection	Milk, honey samples	0.2 μ M	0.5–30 μ M	6
7.	Ag nanoparticles for detection of chloramphenicol	Surface-enhanced Raman scattering (SERS)	Food samples	10–5 μ g/mL	10 ² to 10 ⁻⁵ μ g/mL	7
8.	AuNPs/MoS ₂ /TiO ₂ for detection of Chloramphenicol	Photochemical aptasensor	Milk samples	0.5 pM	0.3-0.5 pM	8
9.	CoAl₂O₄-rGO for removal as well as detection of chloramphenicol	Electrochemical detection	Water samples	13.5 nM	3.4-13.5 nM	This work

References

1. R. Nehru, S. Gnanakrishnan, C.-W., Chen, C.-D, Dong, *ACS Appl. Nano Mater.*,2023, **6**, 1235–1249.
2. L. Geng, J. Huang, H. Zhai, Z. Shen, J. Han, Y. Yu, H. Fang, F. Li, X. Sun, Y. Guo, *Microchemical Journal*,2022, **182**, 107887.
3. L. Bu, X. Chen, Q. Song, D. Jiang, X. Shan, W, Wang, Z. Chen, *Microchemical Journal*,2022, **179**, 107459.
4. X. Xu, Y. Yang, H. Jin, B. Pang, R. Yang, L. Yan, C. Jiang, D. Shao, J. Shi, *ACS Sustainable Chem. Eng.*,2020, **8**, 6806–6814.
5. T. N. T. Nguyen, N. Thi Pham, D.-H . Ngo, S. Kumar, X. T. Cao,*ACS Omega*,2023, **8** , 25385–25391.
6. S. Bunnasit, K. Thamsirianunt, R. Rakthabut, K. Jeamjumnunja, C. Prasittichai, W. Siriwatcharapiboon, *ACS Appl. Nano Mater.*,2024, **7**, 267–278.
7. H. Li, W. Geng, M. M. Hassan, M. Zuo, W. Wei, X. Wu, Q. Ouyang, Q. Chen, Q. *Food Control*, 2021, **128**, 108186.
8. C. Zhao, T. Jing, M. Dong, D. Pan, J, Guo, J. Tian, M. Wu, N. Naik, M. Huang, Z. Guo, *Langmuir*, 2022, **38**, 2276–2286.