

Supporting Information

Hydrogel based on Fe(II)-GMP demonstrates Tunable Emission, Self-healing Mechanical Strength and Fenton Chemistry mediated Notable Antibacterial Properties

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Materials and methods

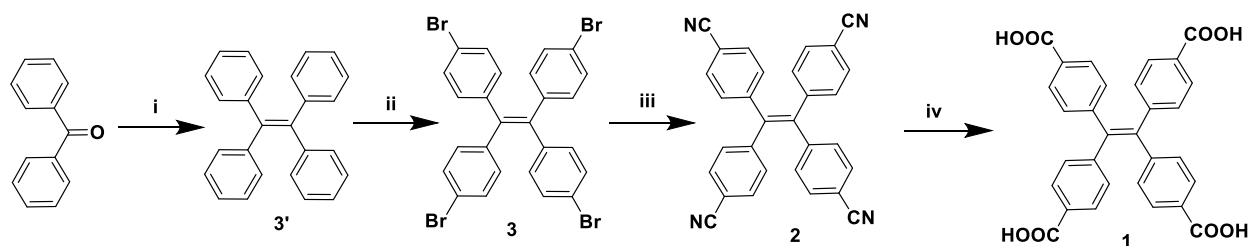
Materials

All reagents, both chemical and biological, employed in this study were sourced from reputable manufacturers and maintained the highest quality standards. Sigma supplied 5'-GMP and $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$, while spectrochem chemicals provided MgCl_2 and CaCl_2 . Biological materials, including bacterial growth media such as Luria Bertani broth and Nutrient agar, were obtained from HIMEDIA. Nuclear Magnetic Resonance spectra ($^1\text{H-NMR}$) were recorded using a FT-NMR Bruker 400 MHz NMR spectrometer. UV-visible spectra were recorded using an Agilent Cary 60 spectrophotometer. Circular Dichroism (CD) spectra were obtained using a JASCO J-1500 Circular Dichroism Spectrometer in Easton, MD, USA. Fluorescence spectra were recorded with an Edinburgh Instruments Spectrofluorometer FS5. X-ray diffraction (XRD) studies were conducted using a Bruker D8 Advance Powder X-ray Diffractometer. Scanning Electron Microscopy (FESEM) images were acquired using a JEOL JEM 2100 scanning electron microscope from Tokyo, Japan.

Synthesis of Tetra(4-carboxylphenyl) ethylene (1)

The molecule TPE, **1** was synthesized by following the previously reported procedure^{S1} and characterized by $^1\text{H-NMR}$ spectroscopy.

Scheme 1



Reagents & Conditions: (i) TiCl_4 , Zn Powder, Dry THF, 0 -70°C, overnight; (ii) Br_2 , Glacial Acetic acid, DCM, 0 °C, 3h; (iii) CuCN , DMF, reflux, 2 days; (iv) KOH , ethylene glycol, 200 °C, 3 days.

$^1\text{H-NMR}$ ($\text{DMSO-}d_6$) of **3**, δ (ppm): 12.93 (br s, 4H), 7.73 (d, 2H), 7.13 (d, 2H).

NMR characterization of 1_{GMP} hydrogel

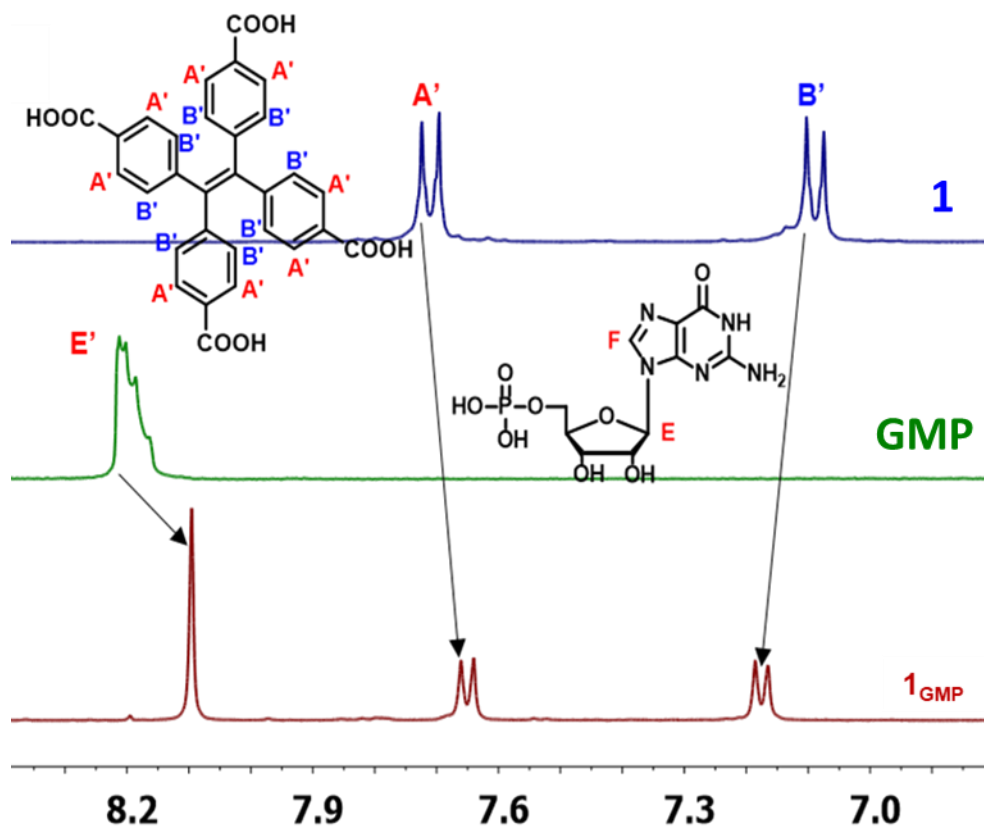


Fig. S1 $^1\text{H-NMR}$ spectra for the **1** in DMSO-d_6 , $5'$ -GMP in D_2O and 1_{GMP} in D_2O , showing the shifts in peak position of the aromatic protons.

Characterization and properties of metallogels, M-1_{GMP}

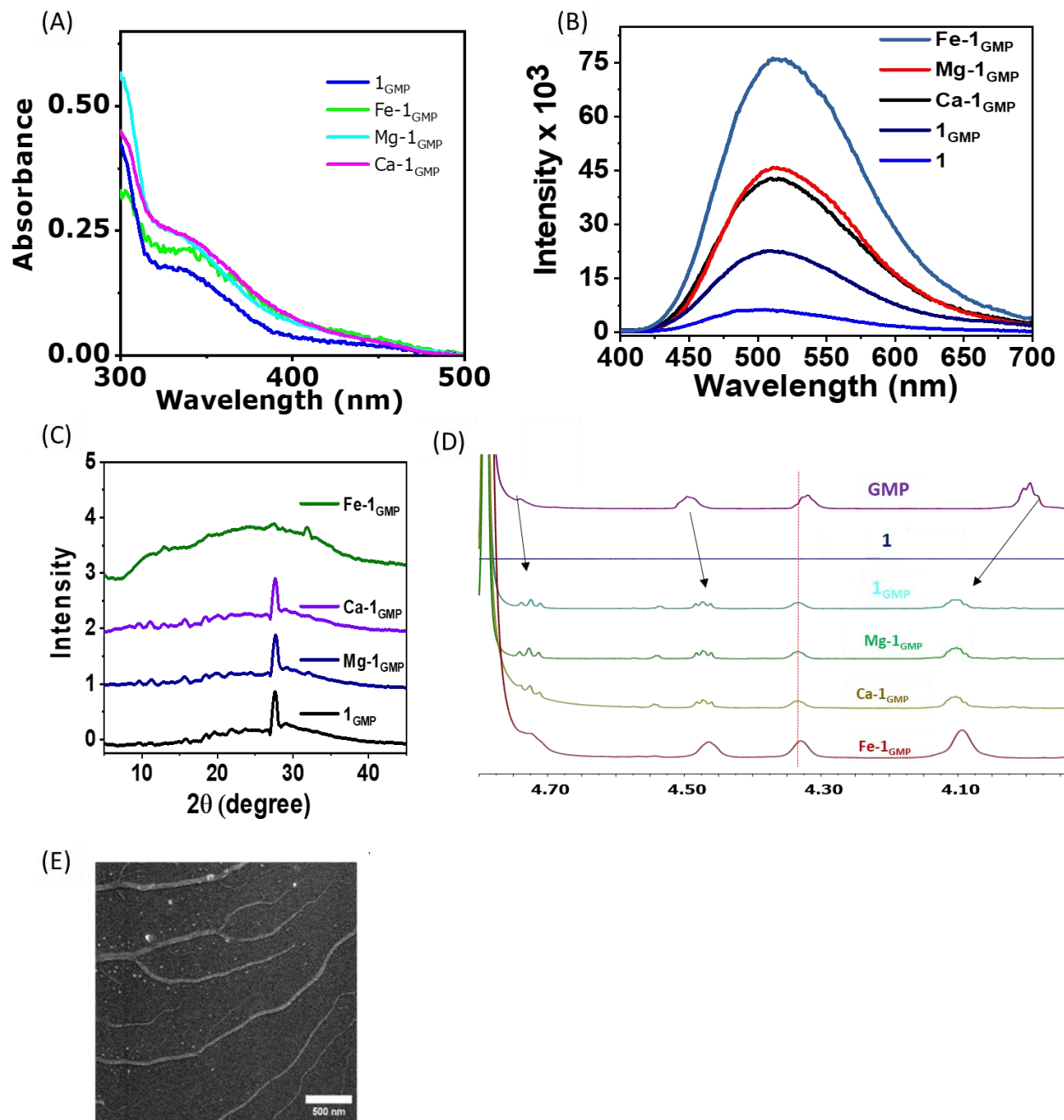


Fig. S2 (A) Comparative UV-vis spectra of the metallogels, **M-1_{GMP}** with **1_{GMP}**. (B) Fluorescence spectra showing enhanced emission of metallogels, **M-1_{GMP}**. (C) PXRD data showing absence of peak at $2\theta = 27.5^\circ$ corresponding to G4-quadruplex. (D) ^1H NMR spectra of the metallogels, **M-1_{GMP}** showing shift of the pentose sugar protons. (E) FE-SEM image of **Fe-1_{GMP}** showing fibrous morphology.

Gelation studies

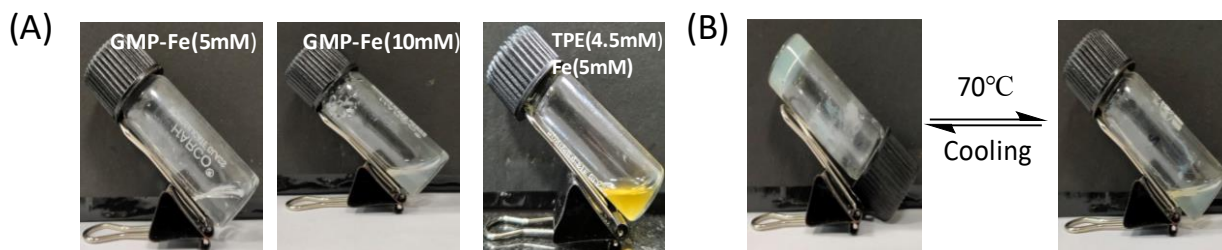


Fig. S3 (A) Control gelation experiment of G-quartet of GMP with TPE (1) and Fe. (B) Reversible hydrogel formation of **Fe-1_{GMP}** by subsequent heat-cool cycles.

Fenton reaction in metallo gels, M-1_{GMP}

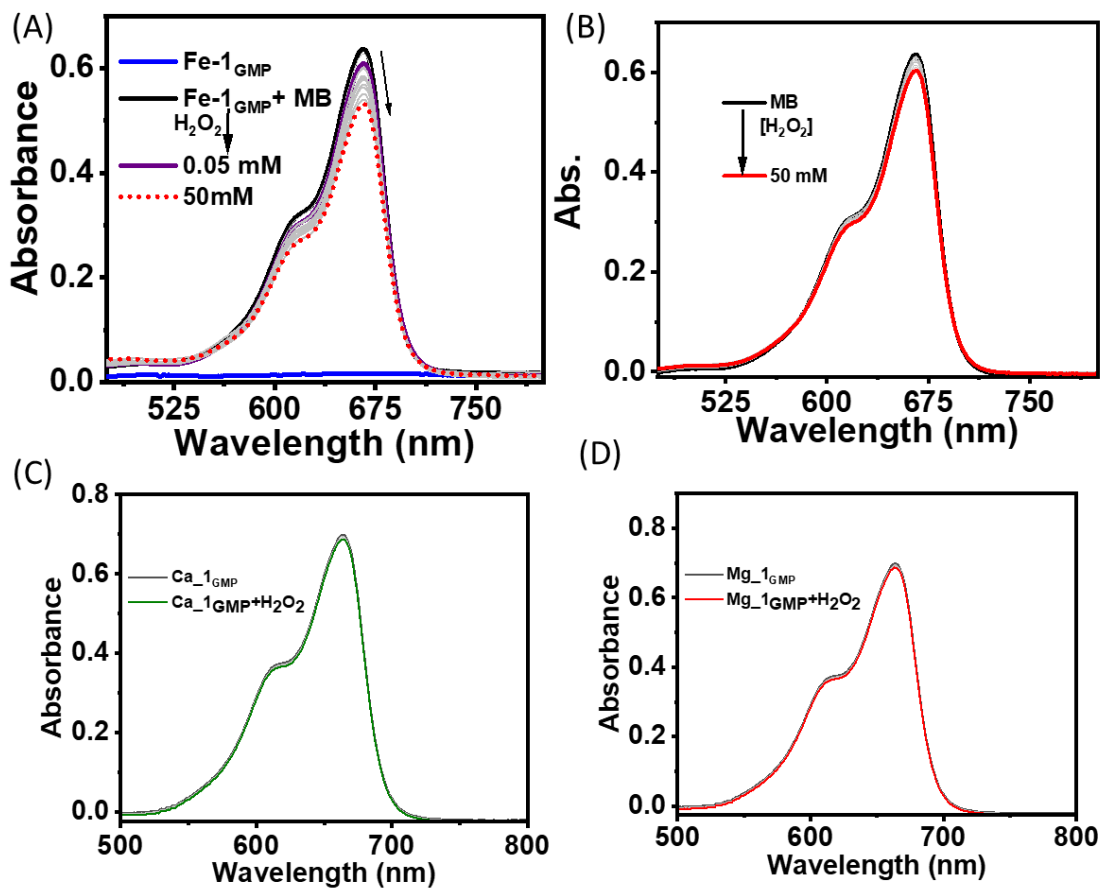


Fig. S4 (A) Concentration dependent degradation of MB dye with addition of H₂O₂ in presence of **Fe-1_{GMP}**. (B) Degradation MB in presence of H₂O₂ only. (C-D) Control experiments showing no ROS formation in the hydrogels of **Ca-1_{GMP}** & **Mg-1_{GMP}**.

Rheological studies of Fe-1_{GMP} V/S Fe-1_{GMP}+AA

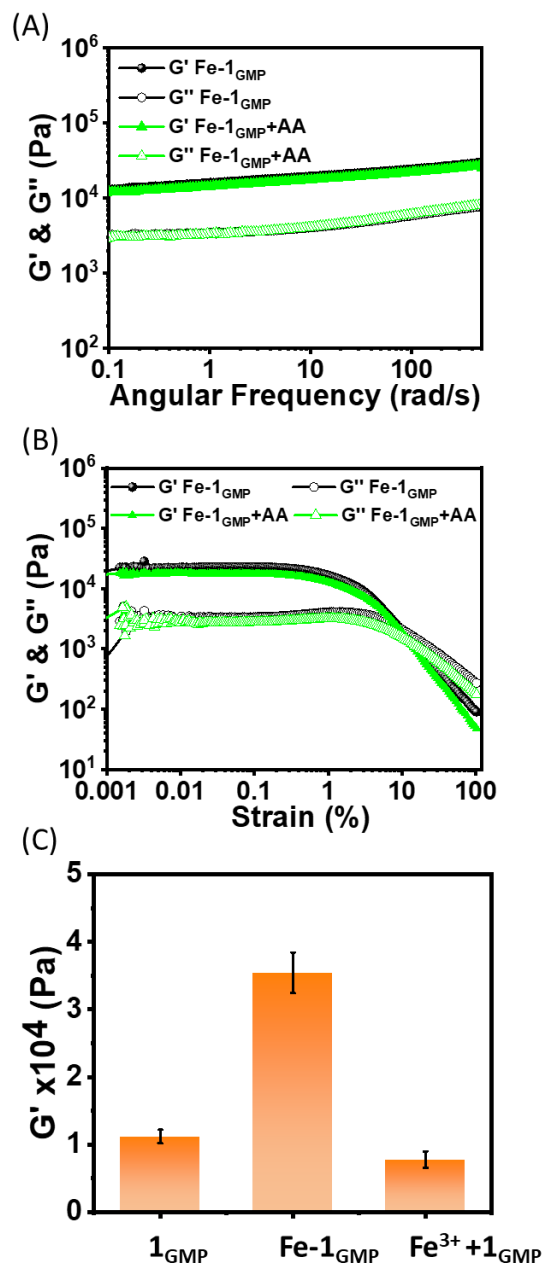


Fig. S5 (A-B) Frequency sweep and amplitude sweep rheological study of Fe-1_{GMP} after loading AA. (C) Comparison of the storage moduli of 1_{GMP}, Fe-1_{GMP}+AA and Fe³⁺ loaded 1_{GMP}.

Antimicrobial and Biocompatibility studies

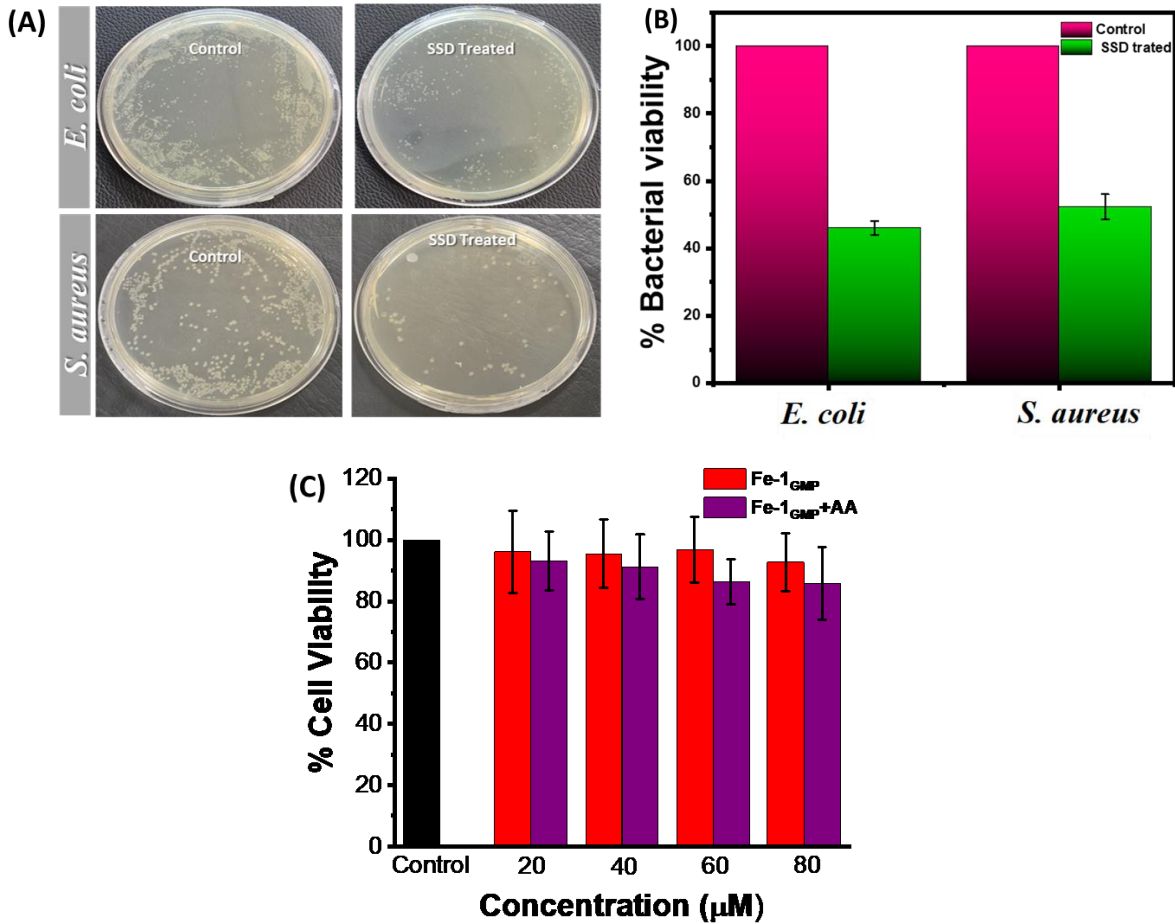


Fig. S6 (A) Images of bacterial colonies formed treating silver sulfadiazine (SSD) at its MIC, *E. coli* 23.7 μg/mL (upper panel) *S. aureus* 47.5 μg/mL (lower panel) and (B) corresponding histogram of comparison of survival percentage of bacterial cells. Control is without SSD administration. (C) MTT assay for Fe-1_{GMP} and Fe-1_{GMP}+AA loaded hydrogels in L929 cell line.

Reference:

S1. A. Bhunia, V. Vasylyeva, C. Janiak, *Chem. Commun.*, 2013, **49**, 3961-3963.