

## Supporting Information

### Three-layered nanoplates and amorphous/crystalline interface synergism boost CO<sub>2</sub> photoreduction on bismuth-oxychloride nanospheres

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## 1. Experimental Section

### 1.1 Chemicals

Ethylene glycol (EG, 99.9 %) and ethanol (99.9 %) were purchased from Xilong Chemical Industry Incorporated Co. Ltd. Poly (sodium 4-styrene sulfonate) (PS, average Mw ~200,000, 30 wt % in H<sub>2</sub>O) were obtained from Sigma-Aldrich. Bismuth nitrate pentahydrate (Bi(NO<sub>3</sub>)<sub>3</sub>·5H<sub>2</sub>O 99 %) were purchased from Macklin. Sodium chloride (NaCl, NaBr, NaI, 99.9 %), sodium sulfate (Na<sub>2</sub>SO<sub>4</sub>, 99.9 %), H<sub>2</sub>AuCl<sub>4</sub>, Na<sub>2</sub>PdCl<sub>4</sub>, and H<sub>2</sub>PtCl<sub>6</sub>, were purchased from Sinopharm Chemical Reagents Co. Ltd. Deionized water was prepared with a Milli-Q water purification system. All chemicals were analytical grade and used without further purification. The indium-doped tin oxide (ITO) glass substrates were obtained from China Southern Glass Co., Ltd., Shenzhen. Then they were well cleaned by ultrasonication in a solution containing deionized water, absolute ethanol, and isopropanol for 15 min sequentially.

### 1.2 Synthesis of BiOCl-NS and other related photocatalysts

For a typical synthesis of BiOCl-NS, 3 mL of 0.25 M Bi(NO<sub>3</sub>)<sub>3</sub> aqueous solution and 9 mL deionized water were successively injected into 15 mL of EG, in a 50 mL plastic tube under vigorous magnetic stirring. Subsequently, 0.45 mL of PS was added dropwise into the above colourless solution. After stirring for 10 min, 2.5 mL of 3 M NaCl aqueous solution was introduced dropwise and then kept stirring for 30 min till forming a uniform colloid solution. The product was further collected through high rpm centrifugation and was washed repeatedly with water and ethanol. The product was dried at 50 °C overnight and stored for characterization and performance study. The final volume ratio of water: EG in the reaction mixture solution is 1:1, and the concentrations of PS, Bi(NO<sub>3</sub>)<sub>3</sub>, and NaCl were 25, 25, and 250 mM, respectively. Further experiments were conducted to expose the influences of reaction time on the formation of BiOCl-NS. Moreover, BiOBr-NS and BiOI-NS were also synthesized by changing the halogen precursor to NaBr and NaI. To synthesize BiOCl-MS, similar conditions (as BiOCl-

NS) were applied except for inserting PS in the reaction vessel. While EG and PS were absent from the system during the synthesis of BiOCl-NP. Furthermore, Au, Pd, or Pt-doped BiOCl-NS were synthesized by using the photo-deposition method (irradiated with Xe light for 30 minutes) with the aid of precursors H<sub>2</sub>AuCl<sub>4</sub>, Na<sub>2</sub>PdCl<sub>4</sub>, H<sub>2</sub>PtCl<sub>6</sub> respectively.

### 1.3 Sample characterizations

The products were characterized by transmission electron microscopy (TEM, JEM-1400), field-emission scanning electron microscopy (FESEM, Sigma 500), energy dispersive X-ray spectroscopy (EDS), and high-resolution TEM (HRTEM, JEM-2100F). X-ray diffraction (XRD) was recorded on a Bruker D8 Focus X-ray diffractometer with Cu K $\alpha$  radiation ( $\lambda=0.15418$  nm) by depositing the sample on an amorphous silicon substrate. X-ray photoelectron spectroscopy (XPS) was performed on a Thermo ESCALAB 250Xi X-ray photoelectron spectrometer with an Al K $\alpha$  excitation source, and the binding energies of all elements were calibrated with the C1s peak at 284.8 eV. Nitrogen (N<sub>2</sub>) adsorption-desorption isotherms were measured on a Micromeritics TriStar II 3020 instrument at 77 K. Before measurement, the samples were degassed and dried at 200 °C under vacuum for 4 h. The specific surface area was calculated according to the Brunauer-Emmett-Teller (BET) method. The diffuse reflectance spectra (DRS) of BiOCl samples were measured using a UV-Vis-NIR spectrophotometer (Shimadzu UV-3101PC) in the wavelength range of 200 to 600 nm. Photoluminescence (PL) spectra and PL lifetime were recorded on Edinburgh FLS920 Multifunction Steady State and Transient State Fluorescence. Electron paramagnetic resonance (EPR) was measured by Bruker EPR EMXplus. In situ diffuse reflection infrared Fourier transform spectroscopic (in-situ DRIFTS) measurements were conducted using diffuse reflectance infrared Fourier transform spectroscopy (Nicolet iS50, Thermo). The sample was placed in the in-situ reactor equipped with a temperature controller for heating. Firstly, the sample was heated to 100 °C for 2 h to remove the adsorbed water molecules. After being cooled down to room temperature, the background IR spectra were recorded as a reference.

Then CO<sub>2</sub> continuous flow (10 mL/min) was passed through the in-situ pool at room temperature, and after adsorption equilibrium, the sample was irradiated, and IR spectra were recorded simultaneously with a certain time interval.

#### **1.4 Electrochemical procedures**

Electrochemical measurements were measured on a Solartron Analytical electrochemical analyzer (ModuLab XM) in a standard three-electrode system by utilizing Pt foil as the counter electrode and Ag/AgCl (KCl, 3 M) as a reference electrode. The working electrodes were prepared by spreading DMF-dispersed slurries of samples (20 mg in a 0.1 mL DMF) onto the well-cleaned ITO glass substrates. Subsequently, the sample-coated substrates were dried in air for 10 min and then annealed at 100 °C for 30 min for electrochemical impedance spectroscopy (EIS) analysis and photocurrent test. In the photoelectrochemical measurements, all three electrodes were placed in a quartz cell containing 0.5 M Na<sub>2</sub>SO<sub>4</sub> aqueous solution as the electrolyte which was bubbled by N<sub>2</sub> thoroughly to remove oxygen before the measurement. The transient photocurrent responses were conducted at a bias potential of 0.6 V under a 60 s on/off chopped illumination. The distance between the Xenon lamp and the working electrode was about 2 cm. EIS analysis was measured in the three-electrode system and recorded over a frequency range from 0.01 Hz to 1 MHz with a current voltage amplitude of 0.01 V. Mott-Schotky measurements were performed at various frequencies 1000 Hz, 1500 Hz, and 2000 Hz.

#### **1.5 Photocatalytic CO<sub>2</sub> reduction**

A Pyrex photoreactor was used to execute photocatalytic CO<sub>2</sub> reduction experiments. A simulated light source with a 300 W Xe lamp (Solaredge 700, 100 mW cm<sup>-2</sup>, 25 °C) was used. The desired photocatalyst (10 mg) was homogeneously dispersed in water (10 ml), and the reactor was evacuated and then purged with CO<sub>2</sub> under continuous stirring several times to remove air completely. Finally, CO<sub>2</sub> gas was filled and the reactor was irradiated at given time intervals. The evolved product (CO) was quantified through periodic headspace gas analysis

(500  $\mu\text{L}$ ) using gas chromatography (GC7820A, Agilent).  $\text{CH}_4$  was measured using a flame ionization detector (FID), and  $\text{CO}$  was converted to  $\text{CH}_4$  using a methanation reactor and analysed using the FID. To test the reusability of BiOCl-SP, the first cycle (8h) was completed following the procedure described above. For the second cycle, the reactor was evacuated, refilled with  $\text{CO}_2$ , and run again for 8 h, and the procedure was uninterruptedly performed for four consecutive cycles (approximately 32 h in total).

The selectivity of photocatalytic  $\text{CO}_2$  Reduction for  $\text{CO}$  can be calculated by the following equation:

$$\text{Selectivity (\%)} = \frac{R_{\text{CO}} \times 2}{(R_{\text{H}_2} \times 2 + R_{\text{CO}} \times 2 + R_{\text{CH}_4} \times 8)} \times 100\%$$

Where  $R_{\text{H}_2}$ ,  $R_{\text{CO}}$  and  $R_{\text{CH}_4}$  are the production rates of photocatalytic reactions of  $\text{H}_2$ ,  $\text{CO}$  and  $\text{CH}_4$ .

The corresponding apparent quantum yield (AQY) of the BiOCl-NS for the product generation was calculated through the following equation:

$$\text{AQY} = \frac{N_e}{N_p} \times 100\%$$

$$N_e = [2 \times (n_{\text{H}_2} + n_{\text{CO}}) + 8 \times n_{\text{CH}_4}] \times N_A$$

$$N_p = \frac{IAt\lambda}{hc}$$

Where  $N_e$  is the total number of reactive transfer electrons, and  $N_p$  is the number of incident photons. where  $n_{\text{H}_2}$ ,  $n_{\text{CO}}$  and  $n_{\text{CH}_4}$  is the  $\text{H}_2$ ,  $\text{CO}$  and  $\text{CH}_4$  production amount (15, 8 and 85  $\mu\text{mol}$ ) per hour under different band-pass filters,  $N_A$  is the Avogadro constant ( $6.02 \times 10^{23} \text{ mol}^{-1}$ ),  $h$  is the Plank constant ( $6.626 \times 10^{-34} \text{ J s}^{-1}$ ),  $c$  is vacuum light velocity ( $3 \times 10^8 \text{ m s}^{-1}$ ),  $\lambda$  is the monochromatic light wavelength (420 nm),  $t$  is the light irradiation time (1 h = 3600 s),  $I$  is the incident monochromatic light intensity ( $30 \text{ W/m}^2$ ),  $A$  is the irradiation area ( $5.7 \text{ cm}^2$ ).

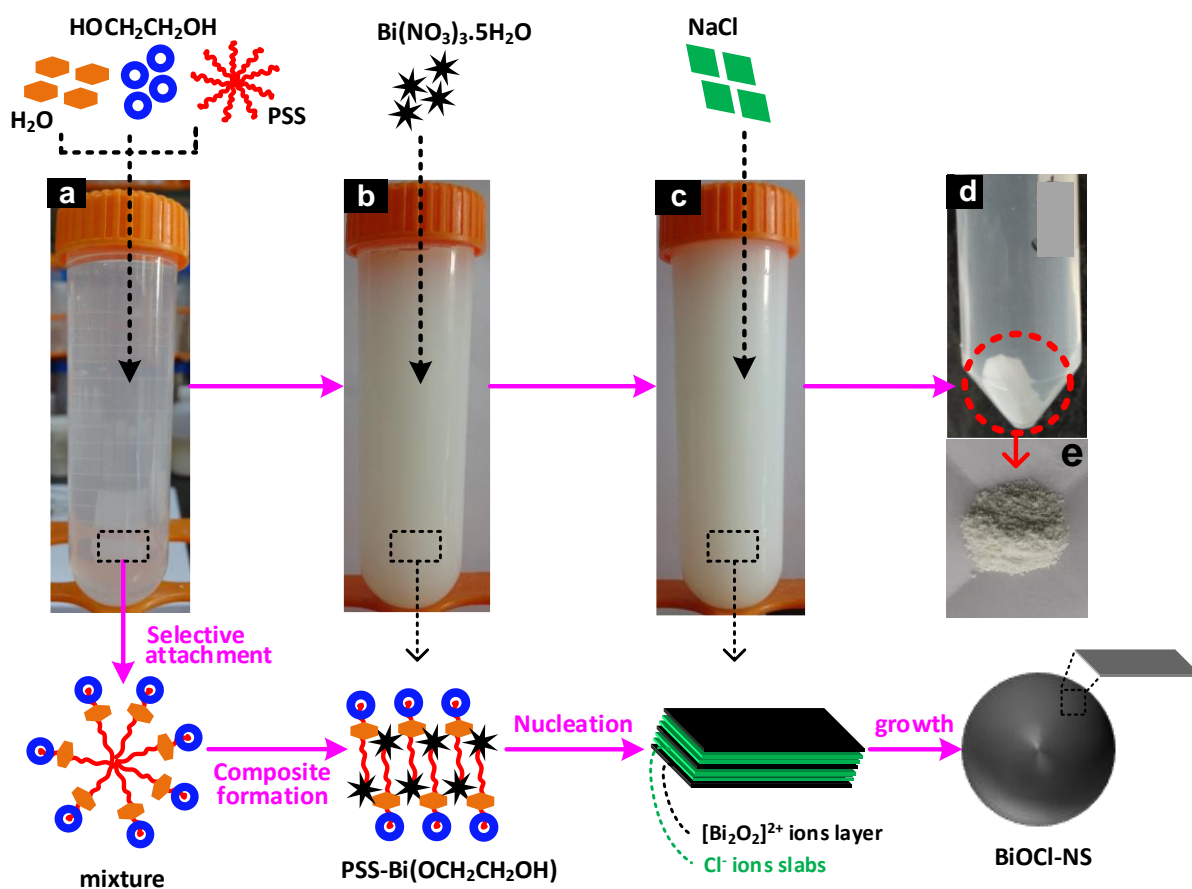
The turnover number (TON) of BiOCl-NS was estimated from the following formula;

$$\text{TON} = (\text{Number of moles of CO evolution}) / (\text{Number of moles of BiOCl-NS taken})$$

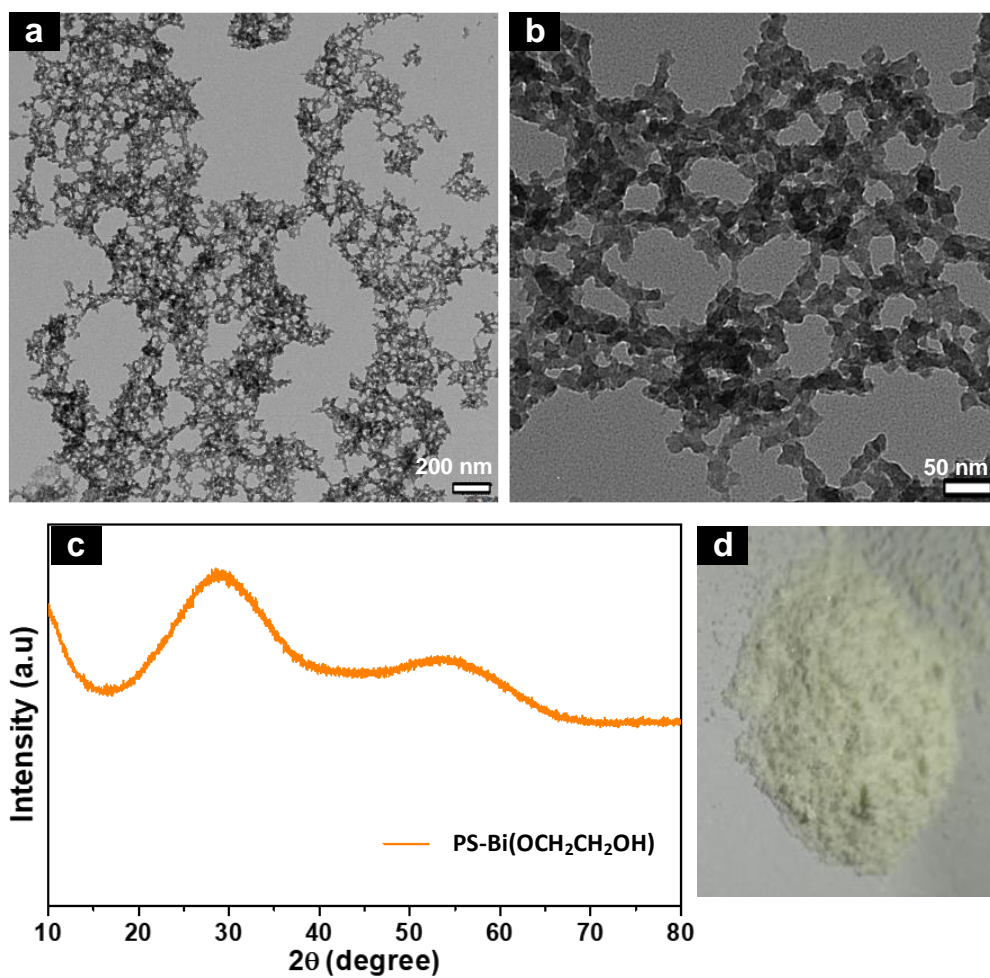
## **1.6 DFT simulations**

All DFT calculations in this work were performed by the Vienna Ab-initio Simulation Package (VASP). The electron-ion interaction was described by the projector augmented wave (PAW) pseudopotential method, and exchange-correlation energy was described by Perdew-Burke-Ernzerhof (PBE) in generalized gradient approximation (GGA). The Monkhorst-Pack grid mesh-based Brillouin zone k-points are set as  $2 \times 2 \times 1$  for all surface structures with the cutoff energy of 450 eV and the equilibrium was reached when the forces on the relaxed atoms became less than  $0.05 \text{ eV}/\text{\AA}$ . Furthermore, the (110) facet was selected as a typical catalysis active surface of BiOCl. To avoid interlayer interference, a  $20 \text{ \AA}$  vacuum layer along the Z direction is employed.

## 2. Supplementary Figs. (S1-S20)



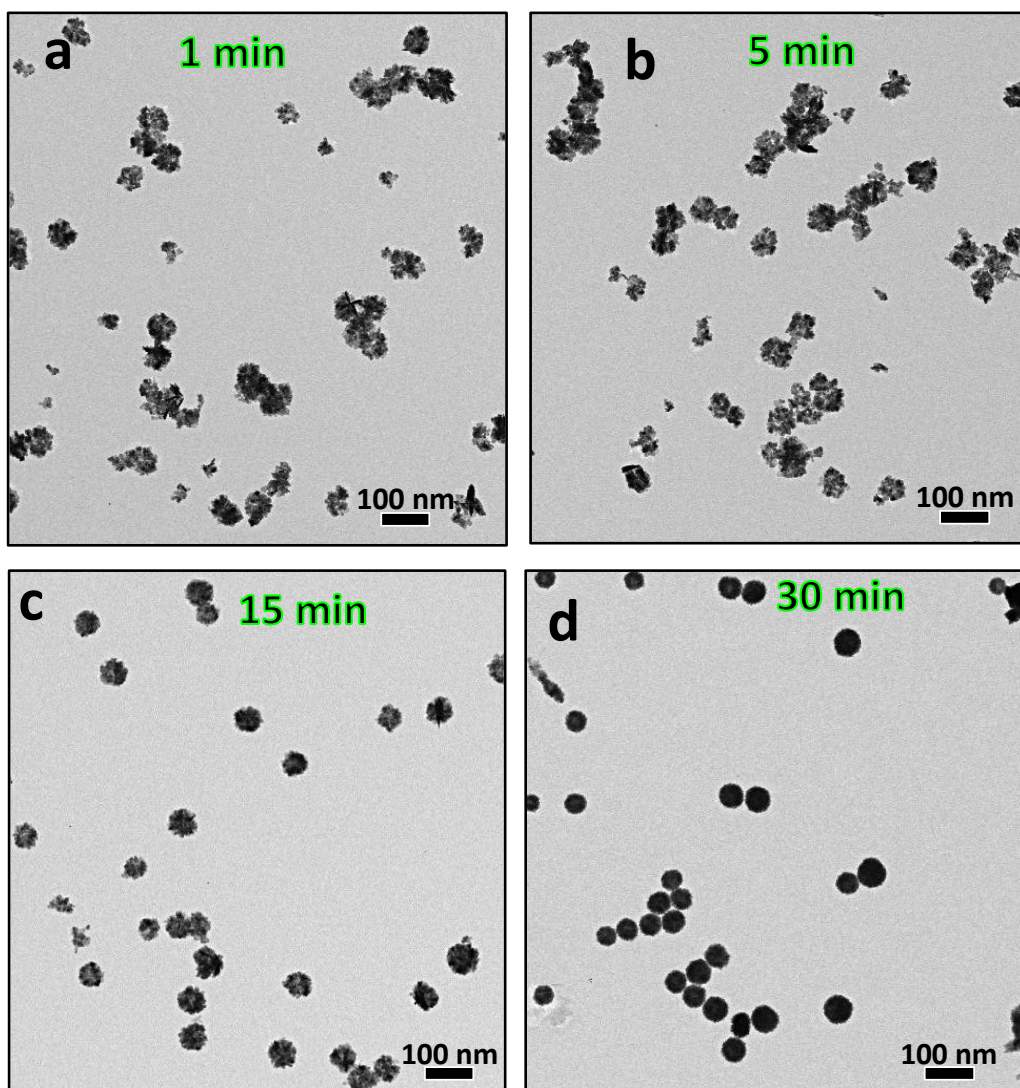
**Fig. S1** The formation process of BiOCl-NS with real-time photographs, displaying the synthesis propagation with typical complex generation after the addition of EG, Water, PS, and Bi precursor.



**Fig. S2** (a, b) TEM images of complex (PS-Bi(OCH<sub>2</sub>CH<sub>2</sub>OH)) formed during the synthesis of BiOCl-NS at RT. (c) Corresponding powder and XRD pattern of composite.

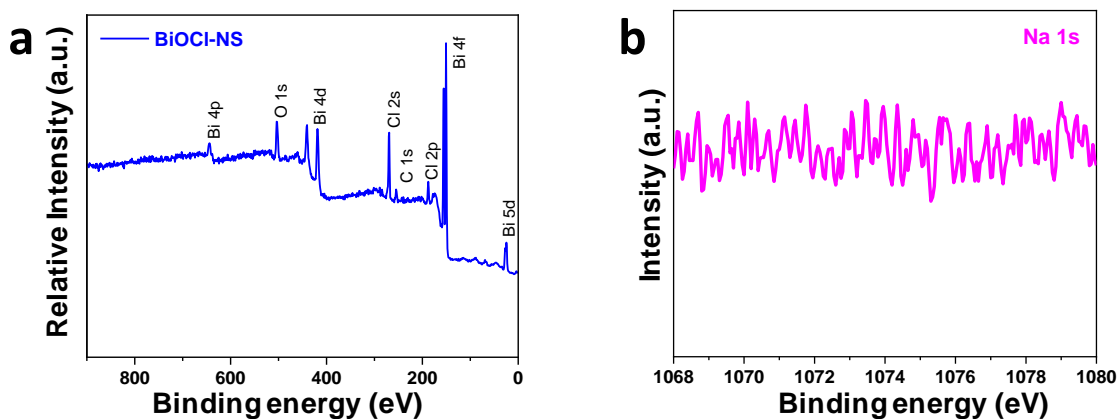
The temporary composite thus restricts the sudden contact between bismuth precursors and halogen precursors. This leads to the controlled, non-agglomerated formation of 3D-structured BiOCl-NS.





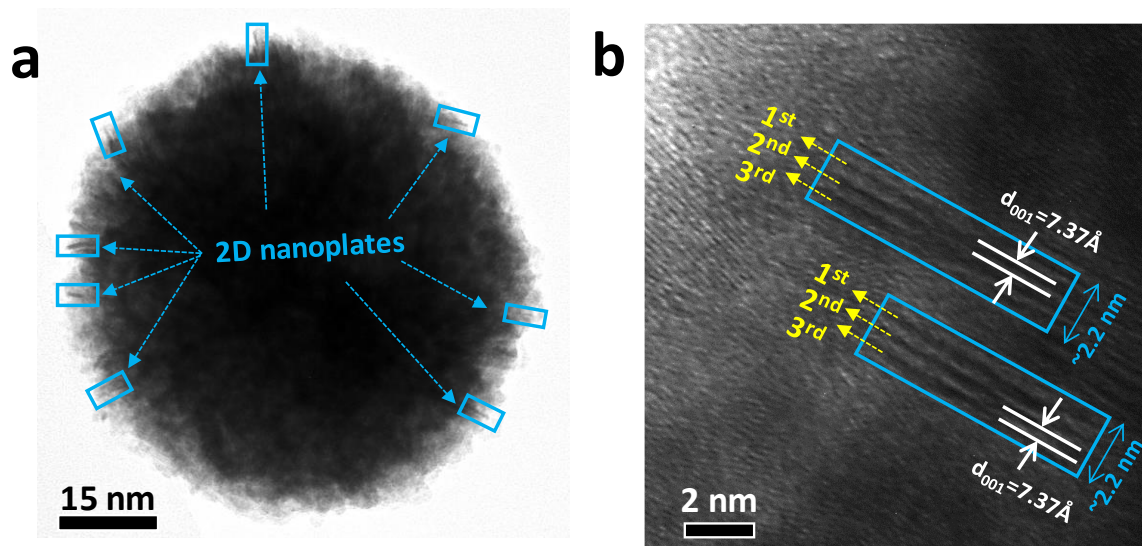
**Fig. S3** (a-d) Time-dependent structure evolution of BiOCl-NS.

The structural transition from the relatively open morphology to regularly-confined morphology. That confirms that innovatively controlling the synthesis route, can inhibit the conventional habitual growth to micron size.

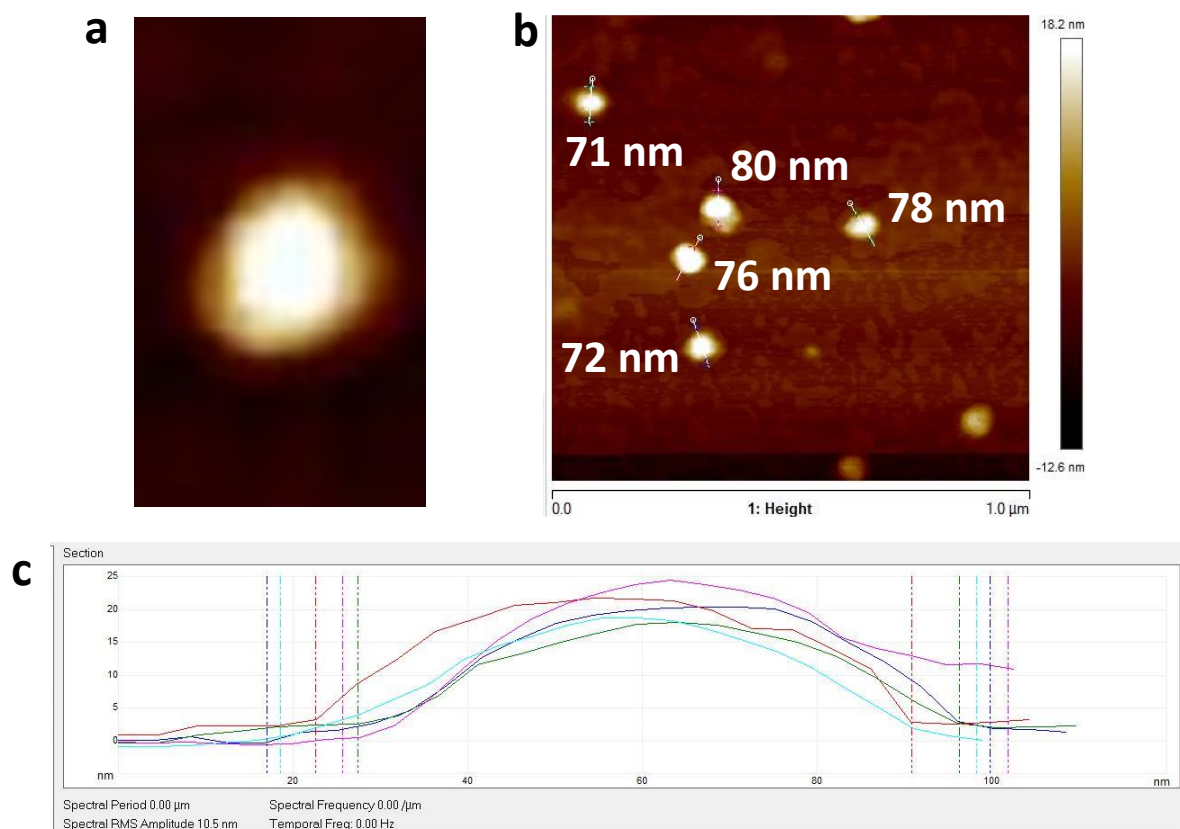


**Fig. S4** (a) XPS survey spectrum, (b) high-resolution XPS spectra of element Na 1s, ruling out the existence of PS residues on the surface of BiOCl-NS.

The absence of any Na 1s signal further validates that PS has just participated temporarily during the formation process, and it has not been detected in BiOCl-NS, confirming the purity of the product.

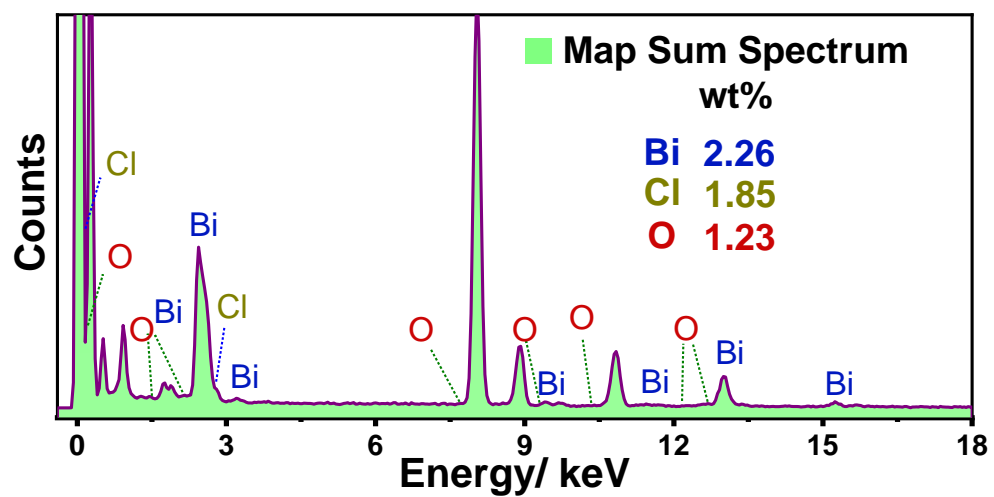


**Fig. S5** HRTEM images of BiOCl-NS (a) single nanoparticle, (b) three-layered nanoplates configuration.

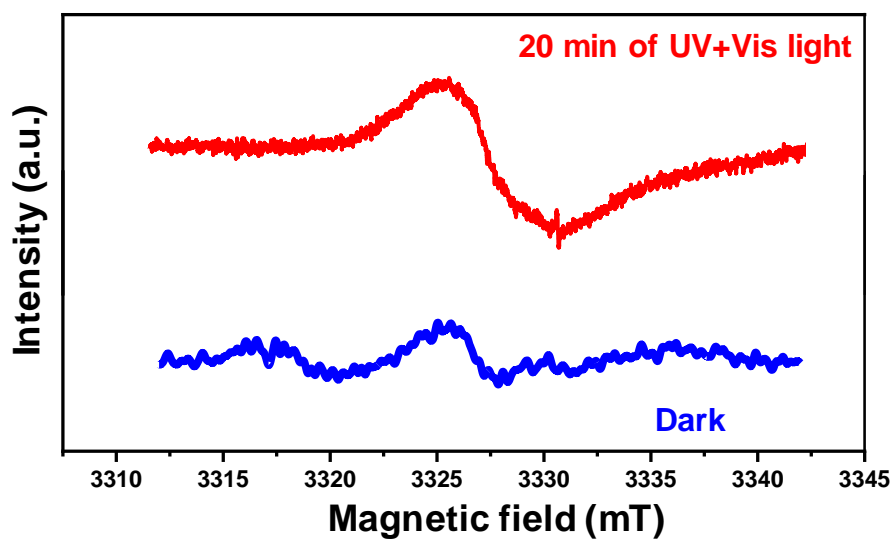


**Fig. S6** (a, b) AFM images and (c) corresponding height profile.

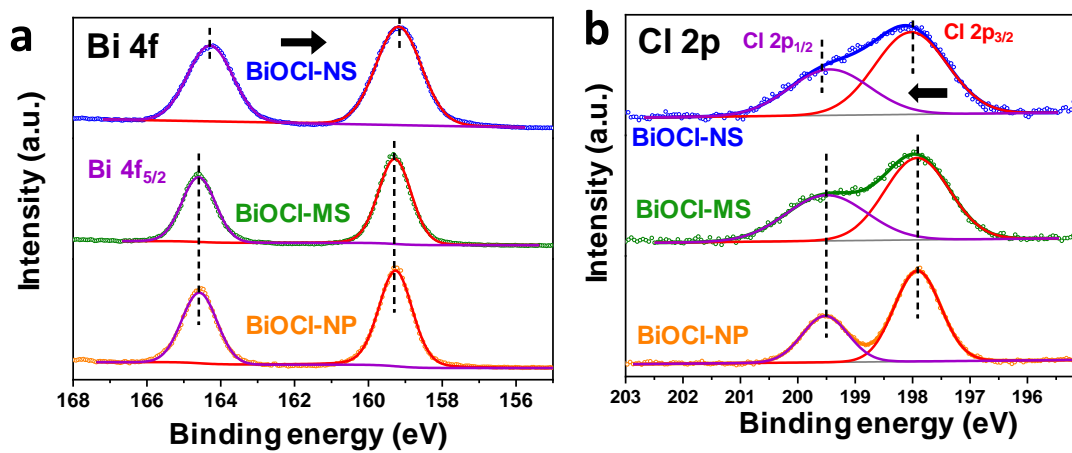
AFM image of a single particle of BiOCl-NS showing a distinct morphology, and size distribution of various nanoparticles. The profile is based on the individual particles' distance from one point to another, further confirming the diameter distribution of BiOCl-NS.



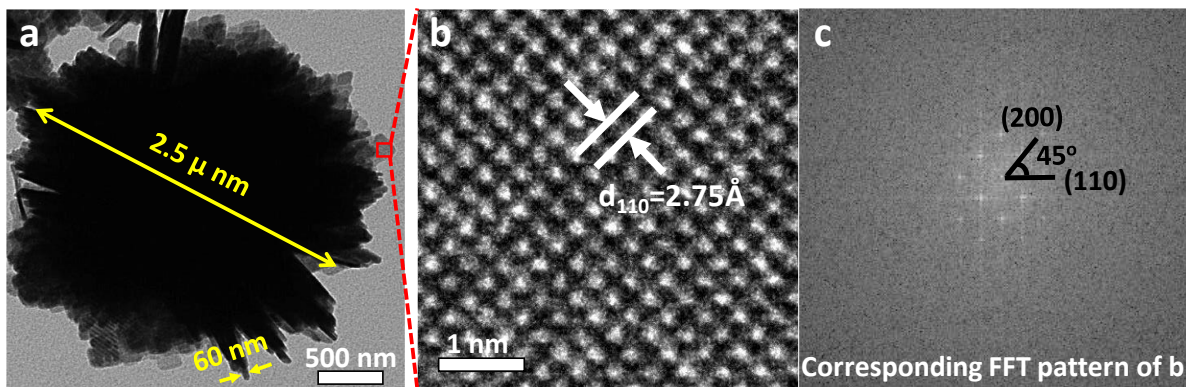
**Fig. S7** EDS spectra highlight a relatively lower weight percentage of element O in BiOCl-NS.



**Fig. S8** EPR spectra of BiOCl-NS after 10 min of light irradiation in comparison to dark conditions, demonstrating the existence of dynamic or light-induced defects.

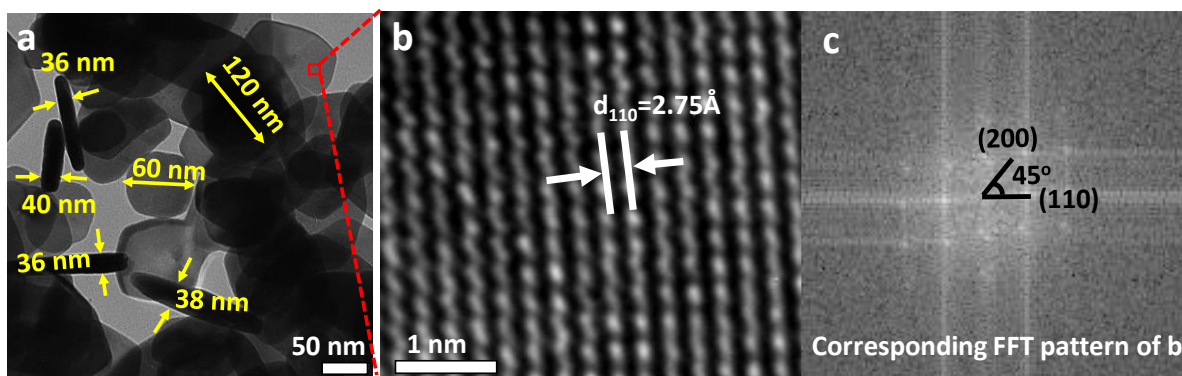


**Fig. S9** Comparative illustration of high-resolution *ex-situ* XPS spectra of (a) Bi 4f and (b) Cl 2p.

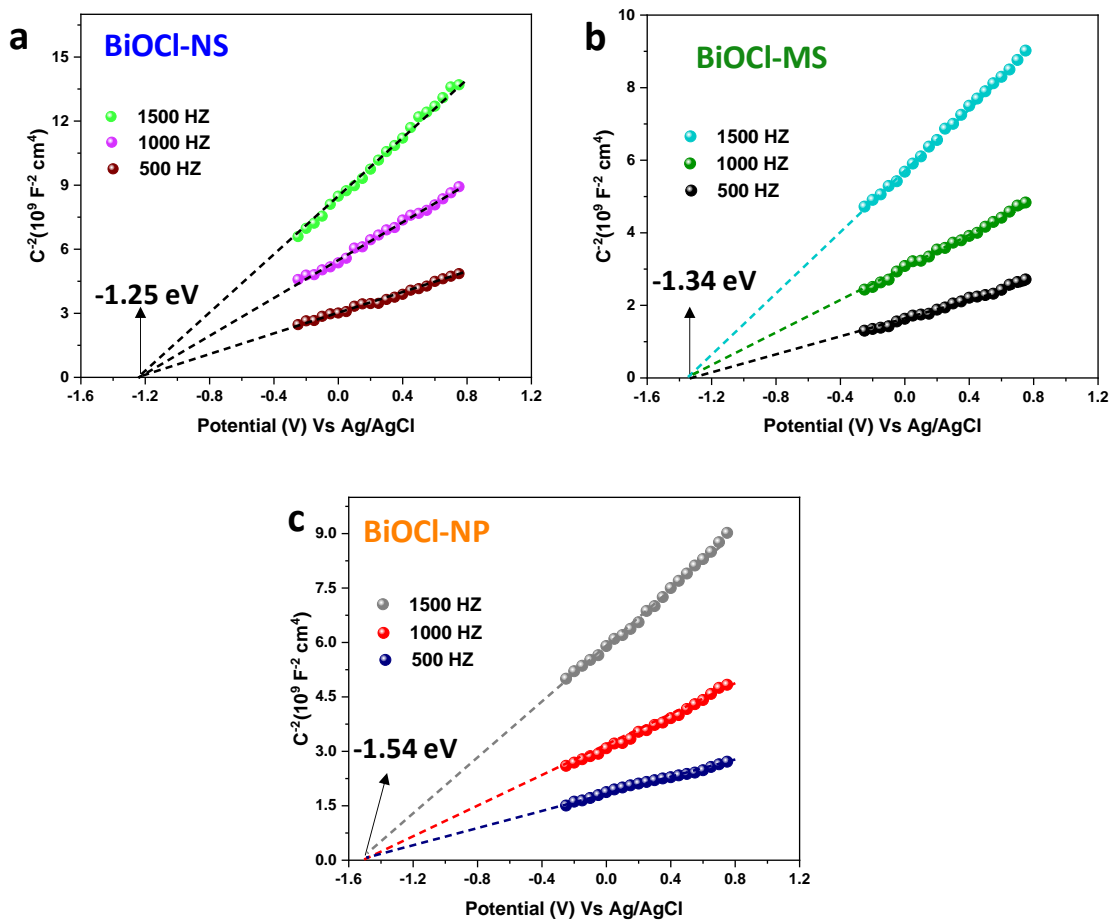


**Fig. S10** (a) TEM image of BiOCl-MS displaying about  $\sim 2.5 \mu\text{m}$  diameter and thickness of about  $\sim 60 \text{ nm}$ . (b) Lattice distribution and (c) corresponding FFT pattern.



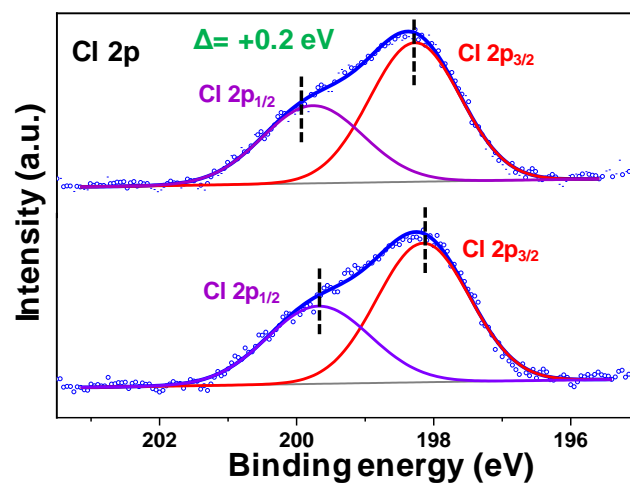


**Fig. S11** (a) TEM image of BiOCl-NP demonstrating nanoplates randomly distributed horizontally and vertically with planar size range from 60-120 nm, and thickness ranging from 36-40 nm. (b) Lattice distribution and (c) corresponding FFT pattern.

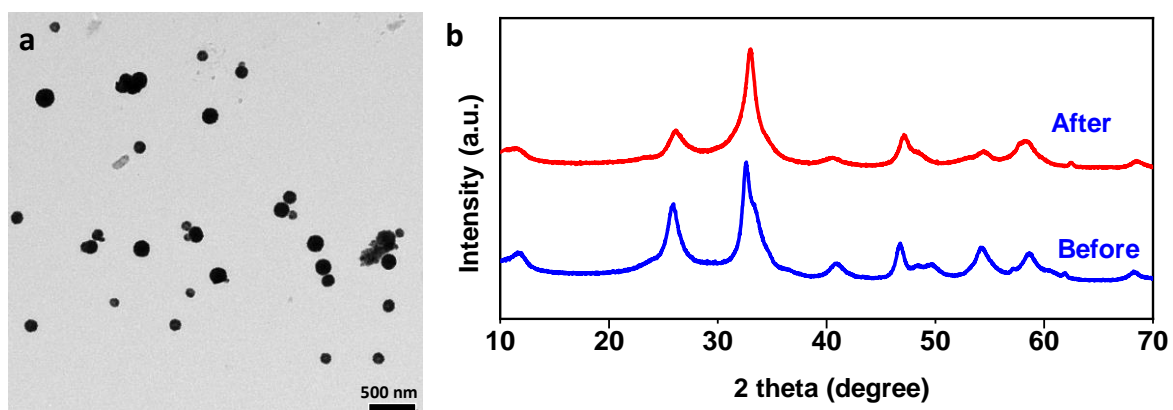


**Fig. S12** Mott Schottky analysis of (a) BiOCl-NS, (b) BiOCl-MS, and (c) BiOCl-NP measured at different frequencies. (d) Band alignment showing corresponding CB and VB positions of BiOCl-NS, BiOCl-MS, and BiOCl-NP.

The flat band ( $E_{FB}$ ) for BiOCl-NS, BiOCl-MS and BiOCl-NP was found to be -1.25 V, -1.34 V, and -1.54 V against Ag/AgCl. As is known, BiOCl is an indirect semiconductor whose flat band is very close to VB, considering this,  $E_{CB}$  was calculated against NHE using the following formula [ $E_{NHE} = E_{Ag/AgCl} + E^{\circ}_{Ag/AgCl} + 0.059 \cdot pH$ ]. Where, the value of  $E_{Ag/AgCl}$  is -1.251 V for BiOCl-NS and -1.34 V for BiOCl-MS, and -1.54 V for BiOCl-NP,  $E^{\circ}_{Ag/AgCl} = 0.199$  V, and  $pH = 7$ . Accordingly, the  $E_{CB}$  values were calculated to be -0.63 V, -0.72 V, and -0.92 Vs NHE for BiOCl-NP, BiOCl-MS and BiOCl-NS, respectively. Further, by using the equation  $E_{VB} = E_g - E_{CB}$ , the values of the valence band potential ( $E_{VB}$ ) turned out to be 2.74 V, 2.54 V, and 2.02 V for BiOCl-NP, BiOCl-MS and BiOCl-NS, respectively.

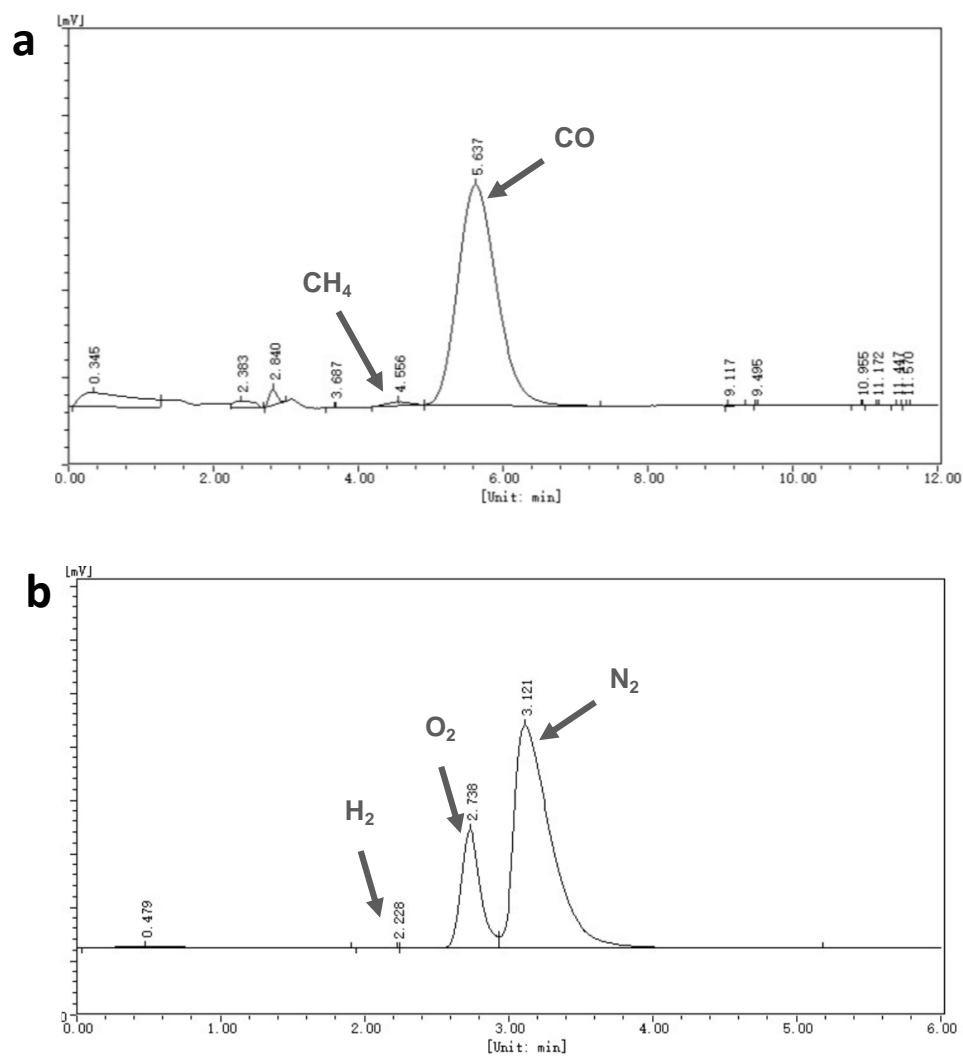


**Fig. S13** *In-situ* XPS spectra of Cl 2p under simulated light irradiation in comparison to the dark conditions.



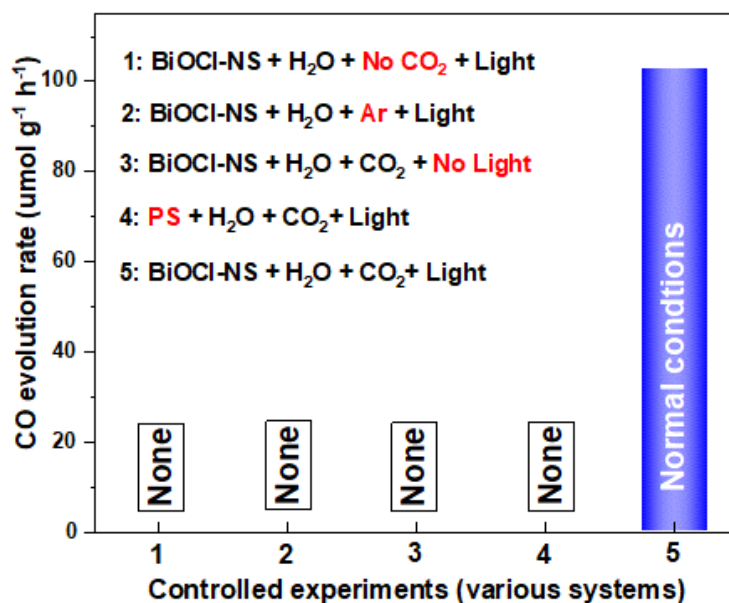
**Fig. S14** The characteristics of BiOCl-NS after reusability, (a) TEM and (b) XRD pattern.

The results confirm the morphological and phase purity after using BiOCl-NS for consecutive photocatalytic cycles.



**Fig. S15** The GC graph from (a) FID detector and (b) TCD detector.

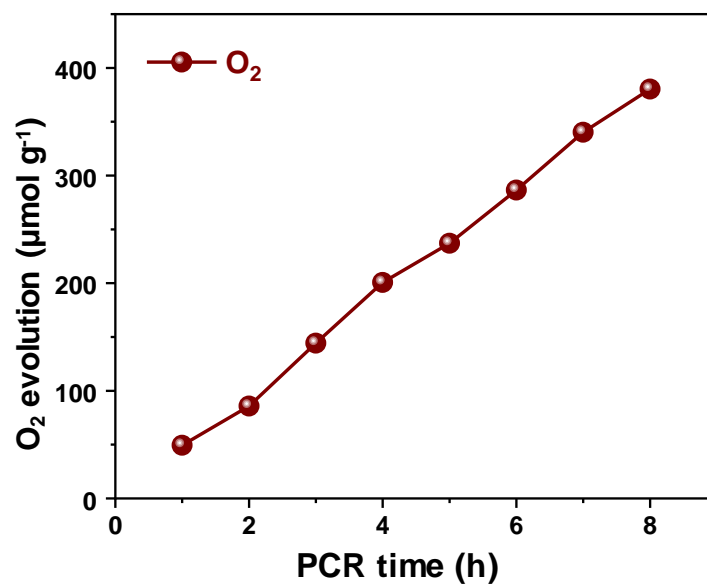
The results highlight the negligible detection of other evolved gases, particularly CH<sub>4</sub> or H<sub>2</sub>, ensuring high selectivity of BiOCl-NS towards CO.



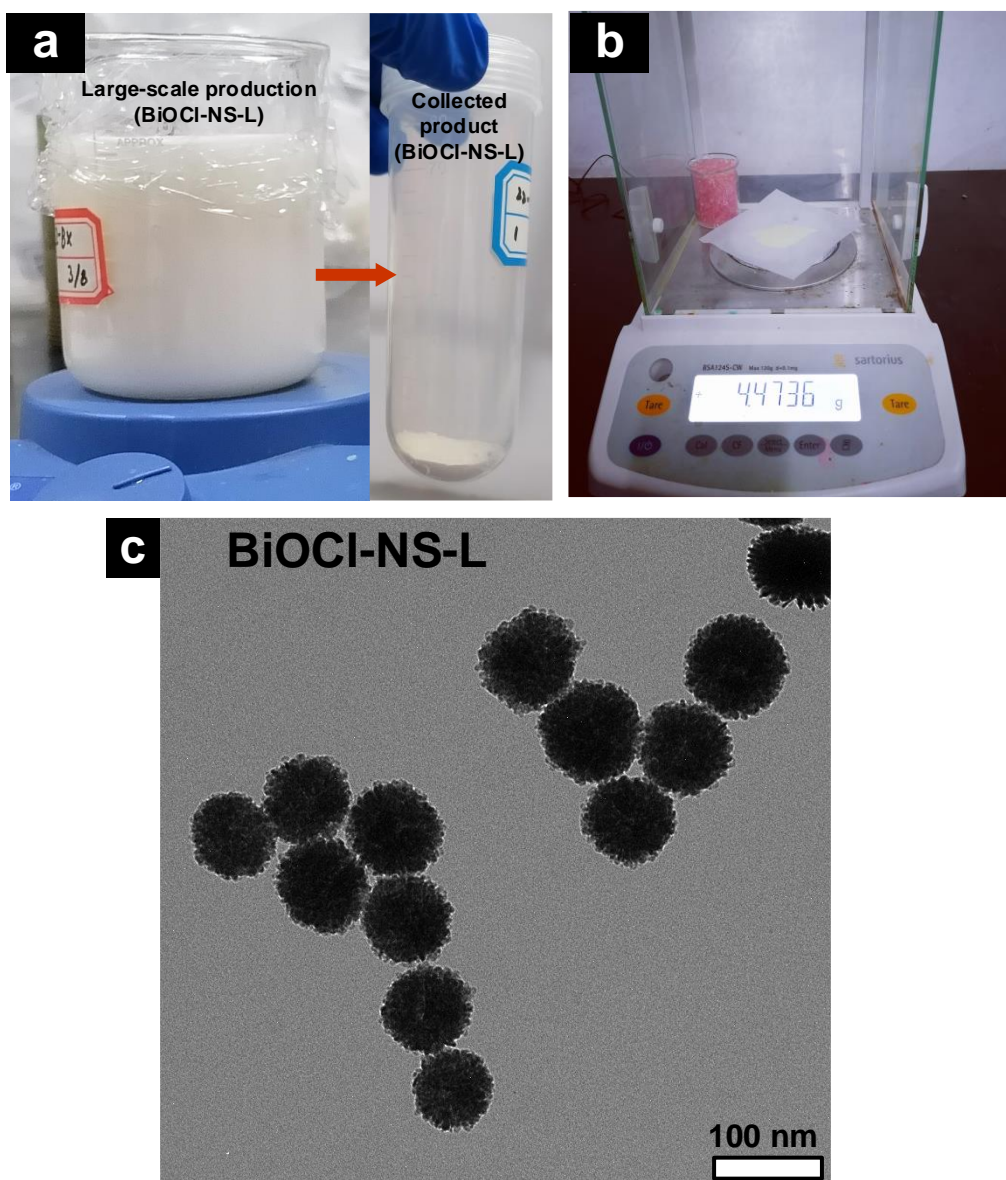
**Fig. S16** The results of the control experiments under various conditions to verify the origin of the produced products.

#### **Additional discussion:**

Several control experiments were designed and conducted to determine the origin of evolved CO. For instance, system-1 was employed without feedstock CO<sub>2</sub>, and in consequence, no CO was detected as a product, validating that feedstock CO<sub>2</sub> is necessary and is the origin of produced CO. This result also confirms that BiOCl-NS does not self-degrade during PCR. System 2, using argon (Ar) instead of CO<sub>2</sub>, also fails to produce any CO, again highlighting that CO<sub>2</sub> is required to produce CO. Likewise, system 3 certifies the importance of light irradiation, as no CO was detected when the light was absent. Other than that, PS residues were not detected on the surface of BiOCl-NS, and hence unable to participate in PCR, to further endorse this, system-4 was processed, wherein PS was used as a photocatalyst instead of BiOCl-NS. The result confirms that no CO was produced, confirming that PS cannot participate during PCR. Finally, CO in high yield was detected when normal conditions were employed in system-5, namely, BiOCl-NS as photocatalysts, H<sub>2</sub>O as a source of the proton, CO<sub>2</sub> as a feedstock, and light irradiation.

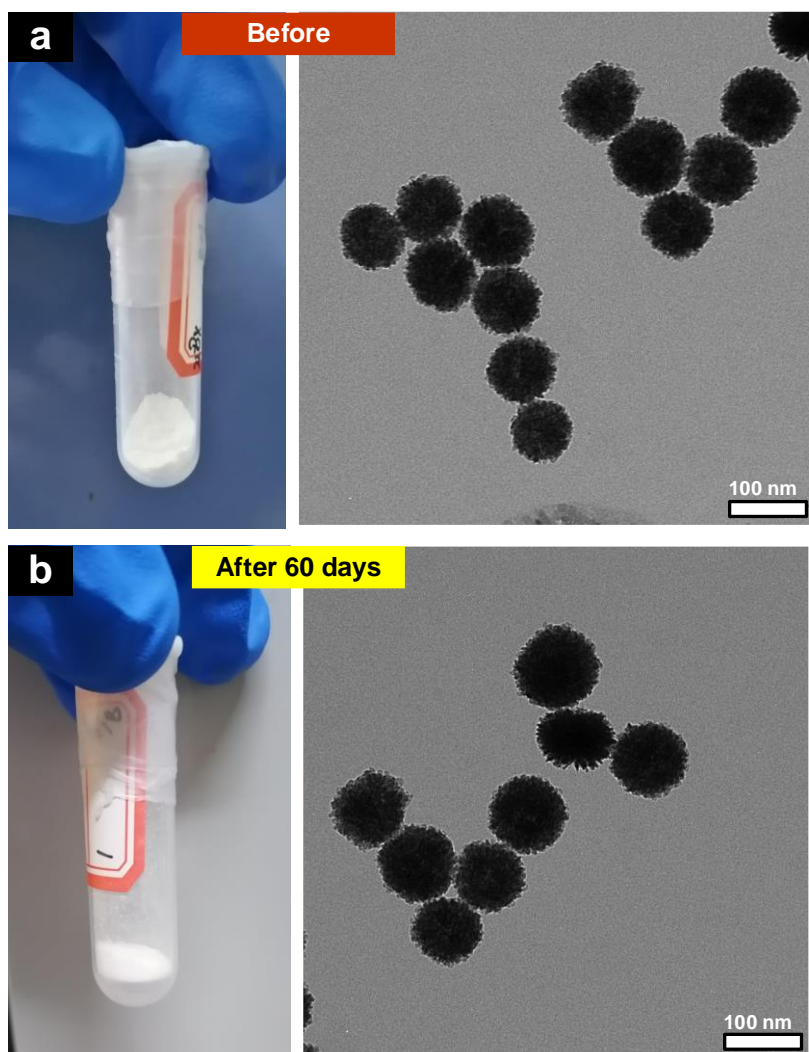


**Fig. S17** The result shows the oxidation of water at the VB end of BiOCl-NS, due to which an uninterrupted supply of protons became available during PCR.

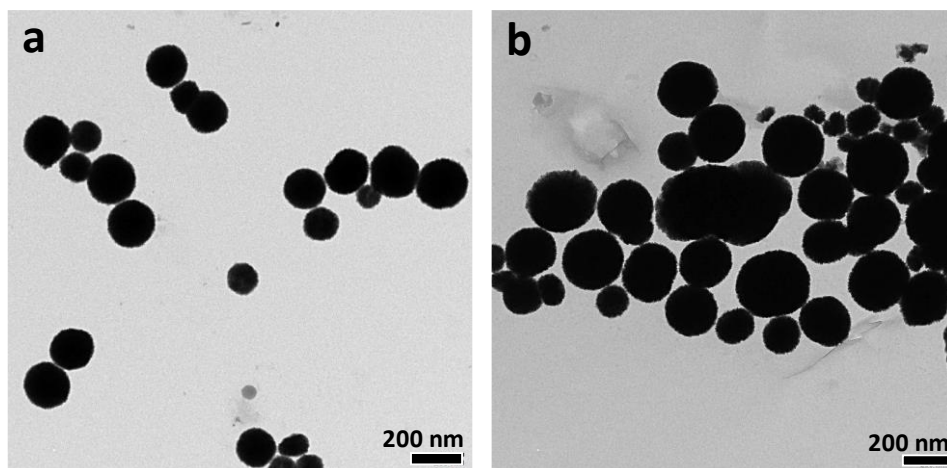


**Fig. S18** (a) Large-scale synthesis of BiOCl nanospheres (named BiOCl-NS-L), and corresponding collected product. (b) gram-scale weightage, and (c) corresponding TEM image of BiOCl-NS-L displaying similar morphology as obtained in small-scale.

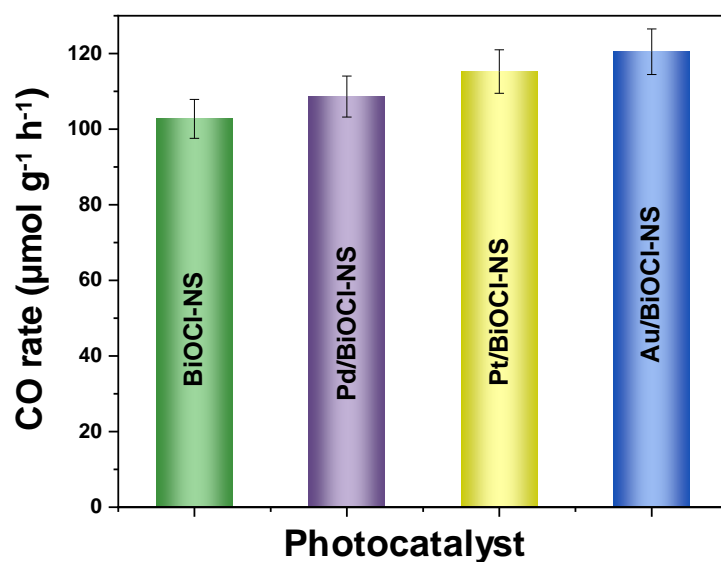




**Fig. S19** Digital photograph and corresponding TEM images of BiOCl-NS (a) before and (b) after storing for months.



**Fig. S20** TEM images of (a) BiOBr-NS, and (c) BiOI-NS, synthesized by changing the halogen precursors with NaBr and NaI, respectively.



**Fig. S21** The preliminary results highlight the further PCR optimization by introducing Nobel metals (Au, Pd, Pt) in BiOCl-NS, a valuable indicator for considering BiOCl-NS as a potential support material for diverse applications.

### 3. Supplementary Tables (S1-S4)

**Table S1.** Comparison of BiOCl-NS with 3D BiOX reported in the literature.

| 3D BiOX   | Morphology             | Synthesis                | Approximate dia (nm) | Uniform/Non-uniform | Ref.             |
|---|------------------------|--------------------------|----------------------|---------------------|------------------|
| BiOCl   | Flower-like            | Solvothermal             | ~2 $\mu\text{m}$     | Non-uniform         | [1]              |
| BiOCl   | Mesopores flowers-like | Hydrothermal             | >10 $\mu\text{m}$    | Non-uniform         | [2]              |
| BiOCl   | Flower-like            | Solvothermal             | ~1 $\mu\text{m}$     | Mildly-uniform      | [3]              |
| Bi-BiOCl  | Flower-like            | Mild-temperature         | ~2 $\mu\text{m}$     | Non-uniform         | [4]              |
| H-BiOCl   | Flower-like            | Hydrothermal             | ~2 $\mu\text{m}$     | Non-uniform         | [5]              |
| BiOCl   | Flower-like            | Solvothermal             | ~2 $\mu\text{m}$     | Non-uniform         | [6]              |
| BiOCl/TiO <sub>2</sub>                                      | Hierarchical           | Solvothermal             | ~2 $\mu\text{m}$     | Non-uniform         | [7]              |
| BiOCl/CN  | Aggregated sheet-like  | Mild-Temperature         | ~3 $\mu\text{m}$     | Non-uniform         | [8]              |
| P/Bi-BiOBr  | Hierarchical           | Solvothermal             | ~2 $\mu\text{m}$     | Mildly-uniform      | [9]              |
| BiVO <sub>4</sub> /BiOBr                                    | Hierarchical           | Solvothermal             | ~3 $\mu\text{m}$     | Mildly-uniform      | [10]             |
| BiOCl <sub>x</sub> Br <sub>1-x</sub>                        | Hierarchical           | colloidal                | ~3 $\mu\text{m}$     | Non-uniform         | [11]             |
| BiOBr   | Flower-like            | hydrothermal             | ~2.5 $\mu\text{m}$   | Non-uniform         | [12]             |
| BiOI  | Hierarchical           | Chemical-precipitation   | ~2.5 $\mu\text{m}$   | Non-uniform         | [13]             |
| Pd/BiOI/MnOx  | Hollow spheres         | In-situ colloidal        | >2 $\mu\text{m}$     | Non-uniform         | [14]             |
| BiOBr/Bi <sub>2</sub> SiO <sub>5</sub>                      | Flower-like            | Solvothermal             | ~2 $\mu\text{m}$     | Non-uniform         | [15]             |
| BiOBr/Cu <sub>2-x</sub> S                                   | Flower-like            | Solvothermal             | ~1.2 $\mu\text{m}$   | Non-uniform         | [16]             |
| BiOBr <sub>0.75</sub> I <sub>0.25</sub> /BiOIO <sub>3</sub> | Hierarchical           | Solvothermal-assisted    | >2 $\mu\text{m}$     | Non-uniform         | [17]             |
| BiOBr <sub>x</sub> Cl <sub>1-x</sub>                        | Buttercup-like         | Colloidal                | 8-10 $\mu\text{m}$   | Non-uniform         | [18]             |
| BiOI  | Hierarchical           | Solvothermal             | 1-3 $\mu\text{m}$    | Non-uniform         | [19]             |
| BiOCl <sub>x</sub> Br <sub>1-x</sub>                        | Flower-like            | Solvothermal             | ~1.5 $\mu\text{m}$   | Non-uniform         | [20]             |
| BiOCl <sub>0.6</sub> Br <sub>0.4</sub>                      | Hierarchical           | Solvothermal             | ~1.3 $\mu\text{m}$   | Non-uniform         | [21]             |
| Bi <sub>5</sub> O <sub>7</sub> I                            | Porous Hierarchical    | Solvothermal-calcination | ~2 $\mu\text{m}$     | Non-uniform         | [22]             |
| Anti-restack BiOCl  | Hierarchitecture       | Solvothermal             | 1–3 $\mu\text{m}$    | Non-uniform         | [23]             |
| BiOI  | Flower-like            | Precipitation            | 2–5 $\mu\text{m}$    | Non-uniform         | [24]             |
| Bi-BiOCl <sub>x</sub>                                       | Hierarchical           | Solvothermal             | 1–3 $\mu\text{m}$    | Non-uniform         | [25]             |
| BiOCl-NS  | Self-organized spheres | Room-temperature         | ~76 nm               | Uniform             | <b>This work</b> |

**Table S2.** The PCR over BiOCl-NS as compared with other recent state-of-the-art photocatalysts. The catalytic system does not involve any sensitizer, co-catalyst, scavengers, sacrificial agents, and organic solvents.

| Photocatalyst  | Catalyst dosage (mg) | Light source                 | CO evolving rate ( $\mu\text{mol g}^{-1} \text{h}^{-1}$ ) | CO <sub>2</sub> source                              | Ref.             |
|--|----------------------|------------------------------|---|---|------------------|
| BiOIO <sub>3</sub>   | 20                   | 300 W Xe (UV-vis)            | 17.33   | H <sub>2</sub> SO <sub>4</sub> + NaHCO <sub>3</sub> | [26]             |
| Br-grafted Bi <sub>2</sub> O <sub>2</sub> (OH)(NO <sub>3</sub> ) NSs   | 20                   | 300 W Xe (UV-vis)            | 8.12  | H <sub>2</sub> SO <sub>4</sub> + NaHCO <sub>3</sub> | [27]             |
| UN-BiOIO <sub>3</sub>  | 50                   | 300 W Xe (UV-vis)            | 5.42  | H <sub>2</sub> SO <sub>4</sub> + NaHCO <sub>3</sub> | [28]             |
| Sr <sub>2</sub> Bi <sub>2</sub> Nb <sub>2</sub> TiO <sub>12</sub> -OVs | 10                   | 300 W Xe (UV-vis)            | 11.7  | H <sub>2</sub> SO <sub>4</sub> + NaHCO <sub>3</sub> | [29]             |
| BiOBr  | 50                   | Xe (0.2 W cm <sup>-2</sup> ) | 4.45  | H <sub>2</sub> SO <sub>4</sub> + NaHCO <sub>3</sub> | [30]             |
| BiOI   | 50                   | 300 W Xe (UV-vis)            | 5.18  | H <sub>2</sub> SO <sub>4</sub> + NaHCO <sub>3</sub> | [31]             |
| Bi <sub>12</sub> O <sub>17</sub> Cl <sub>2</sub> NTs                   | 30                   | 300 W Xe (UV-vis)            | 48.6  | high-purity CO <sub>2</sub>                         | [32]             |
| BiOBr-OVs Atomic layers  | 10                   | 300 W Xe (UV-vis)            | 87.4  | high-purity CO <sub>2</sub>                         | [33]             |
| BiOBr <sub>x</sub> Cl <sub>1-x</sub>                                   | 10                   | Xe (0.2 W cm <sup>-2</sup> ) | 15.86   | high-purity CO <sub>2</sub>                         | [18]             |
| BiOCl  | 100                  | 500 W Xe                     | 1.01  | high-purity CO <sub>2</sub>                         | [34]             |
| Co-Bi <sub>3</sub> O <sub>4</sub> Br atomic layer                      | 30                   | 300 W Xe (UV-vis)            | 107.1   | high-purity CO <sub>2</sub>                         | [35]             |
| BiOCl with surface pits  | 10                   | 300 W Xe (UV-vis)            | 89  | high-purity CO <sub>2</sub>                         | [36]             |
| BiOCl-large surface area   | 10                   | 300 W Xe (UV-vis)            | 11.42   | high-purity CO <sub>2</sub>                         | [36]             |
| BiOCl-abundant OVs   | 10                   | 300 W Xe (UV-vis)            | 23.45   | high-purity CO <sub>2</sub>                         | [36]             |
| BiOCl-ultrathin thickness  | 10                   | 300 W Xe (UV-vis)            | 14.33   | high-purity CO <sub>2</sub>                         | [36]             |
| BiOCl-NS   | 10                   | 300 W Xe (UV-vis)            | 102.72  | high-purity CO <sub>2</sub>                         | <b>This work</b> |

**Table S3.** The PCR activity over BiOCl-NS in comparison with other recent state-of-the-art photocatalysts. The catalytic system is additive engaged e.g., it involves sensitizers, co-catalyst, scavengers, sacrificial agents, and organic solvents.

| Photocatalyst  | Light source                             | CO evolving ( $\mu\text{mol}\cdot\text{g}^{-1}\cdot\text{h}^{-1}$ ) | Co-Catalyst              | Hole scavenger                                     | Reaction medium                            | Ref.             |
|--|--|---|--------------------------|--|--|------------------|
| g-C <sub>3</sub> N <sub>4</sub>                      | 300 W Xe lamp (>400 nm)                  | 17  | Co-porphyrin             | TEOA   | CH <sub>3</sub> CN                         | [37]             |
| CdS  | 300 W Xe lamp (400 nm)                   | 50.4  | Co-ZIF-9                 | TEOA   | CH <sub>3</sub> CN + H <sub>2</sub> O      | [38]             |
| MOF-525  | 300 W Xe lamp (400 nm)                   | 200.6   | Single-atom Co           | TEOA   | CH <sub>3</sub> CN                         | [39]             |
| o-PCN  | 300 W Xe lamp (>400 nm)                  | 286   | Pt                       | Na <sub>2</sub> S+ Na <sub>2</sub> SO <sub>3</sub> | H <sub>2</sub> O                           | [40]             |
| UiO-66/C <sub>3</sub> N <sub>4</sub>                 | 300 W Xe lamp (>400 nm)                  | 59.4  | -                        | TEOA   | CH <sub>3</sub> CN                         | [41]             |
| Co-tuned Au nanoclusters                             | 300 W Xe lamp (420 nm)                   | 3.451   | -                        | TEOA   | H <sub>2</sub> O                           | [42]             |
| Ni-doped CdS quantum dots                            | 300 W Xe lamp (>400 nm)                  | 9.5   | dinuclear cobalt complex | TEOA   | H <sub>2</sub> O                           | [43]             |
| CsPbBr <sub>3</sub> /GO                              | 100 W Xe lamp (150 mW cm <sup>-2</sup> ) | 23.7  | -                        | -  | Ethyl acetate                              | [44]             |
| CsPbBr <sub>3</sub> /g-C <sub>3</sub> N <sub>4</sub> | 300 W Xe lamp (420 nm)                   | 149   | -                        | -  | Acetonitrile/water and ethyl acetate/water | [45]             |
| Cs <sub>2</sub> AgBiBr <sub>6</sub>                  | AM 1.5G, 150 mW cm <sup>-2</sup>         | 105   | -                        | -  | Ethyl acetate                              | [46]             |
| BiOCl-NS   | 300 W Xe (UV-vis)                        | 102.72  |                          |  | H <sub>2</sub> O                           | <b>This work</b> |

**Table S4.** The BiOCl-NS stability in PCR compared with some state-of-the-art photocatalysts.

| Photocatalyst   | Light source | Stability (h) | CO evolving ( $\mu\text{mol g}^{-1} \text{h}^{-1}$ ) | Reaction medium          | Ref.             |
|---|--------------|---------------|--|--------------------------|------------------|
| BiOCl@Bi <sub>2</sub> O <sub>3</sub>  | >420 nm      | 36            | 30   | H <sub>2</sub> O         | [47]             |
| V <sub>Bi</sub> -BiOBr NSs  | UV-Vis       | 20            | 20.1   | H <sub>2</sub> O         | [48]             |
| Br-grafted  | UV-Vis       | 12            | 8.12   | H <sub>2</sub> O (vapor) | [27]             |
| Bi <sub>2</sub> O <sub>2</sub> (OH)(NO <sub>3</sub> ) NSs<br>BiOIO <sub>3</sub> -OV | UV-Vis       | 12            | 17.33  | H <sub>2</sub> O (vapor) | [26]             |
| Sr <sub>2</sub> Bi <sub>2</sub> Nb <sub>2</sub> TiO <sub>12</sub> -OV               | UV-Vis       | 4             | 17.11  | H <sub>2</sub> O (vapor) | [29]             |
| Bi <sub>4</sub> Ti <sub>3</sub> O <sub>12</sub> -UOV                                | UV-Vis       | 4             | 11.7   | H <sub>2</sub> O (vapor) | [49]             |
| BiOBr-Ovs Atomic Layers   | >420 nm      | 60            | 87.4   | H <sub>2</sub> O         | [33]             |
| Bi <sub>12</sub> O <sub>17</sub> Cl <sub>2</sub> NTs                                | UV-Vis       | 12            | 48.6   | H <sub>2</sub> O         | [32]             |
| Partially oxidized SnS <sub>2</sub> atomic layers                                   | >420 nm      | 12.28         | 20   | H <sub>2</sub> O (vapor) | [50]             |
| BP@g-C <sub>3</sub> N <sub>4</sub>  | UV-Vis       | 8             | 6.54   | H <sub>2</sub> O (vapor) | [51]             |
| SiC-NW/C  | UV-Vis       | 9             | 5.87   | H <sub>2</sub> O (vapor) | [52]             |
| Ni-SA-5/ZrO <sub>2</sub>  | UV-Vis       | 25            | 11.8   | H <sub>2</sub> O (vapor) | [53]             |
| BiOCl-NS  | UV-Vis       | 32            | 102.72   | H <sub>2</sub> O         | <b>This work</b> |

**Table S5.** The AQE of BiOCl-NS in PCR compared with some state-of-the-art BiOX photocatalysts.

| Photocatalyst   | AQE (%) | References       |
|---|---------|------------------|
| S-BiOCl   | 0.28    | [54]             |
| C <sub>3</sub> N <sub>4</sub> /bismuthene/BiOCl                 | 0.42    | [55]             |
| BiOBr-5   | 0.33    | [56]             |
| Bi <sub>4</sub> O <sub>5</sub> Br <sub>x</sub> I <sub>2-x</sub> | 0.37    | [57]             |
| Bi <sub>4</sub> O <sub>5</sub> I-Fe30                           | 0.12    | [58]             |
| Pt <sub>1</sub> /BOB-V <sub>O</sub>                             | 0.20    | [59]             |
| BiOIO <sub>3</sub> -b <sub>3</sub>                              | 0.10    | [28]             |
| BiOCl-NS  | 0.51    | <b>This work</b> |



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