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Supporting Information

A single-gold-atom addition regulates sharp redshift in

fluorescence of atomically precise nanoclusters

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1. Experimental Procedures

1.1 Materials

All the solvents and reagents used in this study were commercially available and used as received without further purification. Dichloromethane (DCM, HPLC grade), Dichloromethane (CDCl₃, HPLC grade), methanol (MeOH, HPLC grade), toluene (HPLC grade), *n*-hexane (*n*-Hex, HPLC grade), 4-tert-butylphenylthiophenol (TBBT, ≥98%), triphenylphosphine (TPP, \geq 98%), HAuCl₄.4H₂O (\geq 99.99%, metals basis) and sodium borohydride (NaBH4, ≥98%) were all purchased from Shanghai Aladdin Bio-Chem Technology CO., Ltd. The ultrapure water (\geq 18.2 M Ω) used in this study was purified on a Millipore system (Millipore, USA). All glassware was washed with aqua regia and rinsed with ultrapure water prior to use.

1.2 Synthesis and purification of the Au³⁷ nanocluster.

Au₃₇ has two synthetic strategies, which are method I and method II.

In **method I**, the Au₃₇ nanocluster was synthesized using a one-pot and size-focusing synthesis strategy, which is a similar method reported for the synthesis of Au₃₆,^[1] but with extra phosphine ligands. Typically, $HAuCl_4·4H_2O$ (82.4 mg, 0.20 mmol) was dissolved in 5 mL water, and TOAB (150 mg, 0.28 mmol) was dissolved in 15 mL dichloromethane. These two solutions were combined in a 25 mL tri-neck round bottom flask. The solution was vigorously stirred (~1100 rpm) with a magnetic stir bar to facilitate the phase transfer of Au(III) salt into the organic phase. After ~15 min, phase transfer was completed and the aqueous was then removed. After that, 4-tertbutylphenylthiophenol (150 μ L, 0.90 mmol) and PPh₃ (15 mg, 0.057 mmol) were added into the above solution order, and the color of the solution changed to colorless. After \sim 30 min, 3 mL aqueous solution of NaBH₄ (77 mg, 2.03 mmol) was added slowly. Then, the reaction was allowed to proceed for \sim 12 h. The product was washed several times with CH₃OH to remove the excess ^tBuPhSH, PPh_{3,} and by-products. Then, the crude product was extracted with CH_2Cl_2 twice. The crude product was then re-dissolved in 3 mL of toluene and reacted with 100 μL of *^t*BuPhSH under 80 °C. After reacting for 12 h, the product was washed with methanol several times and further purified by thinlayer chromatography (TLC, DCM: n-hexane= $1:2$). The raw product containing Au₃₇ was loaded on TLC plates for purification with a yield of about 2.95% (1.80 mg) based on the gold.

In **method II**, Pure Au³⁶ (20 mg) was used as a precursor by reacting with pre-prepared gold-phosphine complexes (Ph₃P)AuCl (1 mg, \sim 1 equivalent) and the reaction was allowed to proceed overnight. (Ph₃P)AuCl was prepared by mixing HAuCl₄·4H₂O (20.6) mg, 0.05 mmol) with PPh₃ (26.2 mg, 0.10 mmol) in ethanol solution (HAuCl₄ + H₂O + 2 $PPh_3 \rightarrow (Ph_3P)AuCl + Ph_3PO + 3 HCl$ and washed three times with ethanol to remove the excess impurity. The purification of the targeted Au_{37} nanocluster was performed by thin layer chromatography with DCM/n-hexane (v: $v = 1 : 2$) as developing solvent. The raw product containing Au_{37} was loaded on TLC plates for purification with a yield of about 5.49% (1.03 mg) based on Au_{36} .

1.3 X-ray Crystallographic Determination.

Single crystal X-ray diffraction (SCXRD) of it was carried out on a Stoe Stadivari diffractometer at 293 K, using a Cu Kα radiation (λ = 1.54186 Å) for Au₃₇. The structure

was solved by direct methods and refined with full-matrix least-squares on F² using the SHELXL-2014/7 (Sheldrick, 2014) suite of programs. All the refinement parameters are summarized in Table S2.

1.4 Computational method and details

Both DFT calculations and TDDFT calculations were done using ORCA5.0.2 software^[2]. The geometric optimization of the ground and excited states of the gold clusters was carried out using the PBE0 exchange-correlation functional^[3] and the def2-SV(P) basis set. To improve the computational efficiency, both DFT and TDDFT calculations were performed using the RI (Resolution of Identity) approximation with the auxiliary basis set def2/J[4] . To facilitate the calculation, we simplified the thiol ligand Bu*^t*PhSH and phosphine ligand Ph₃P to CH₃SH and $(CH_3)_3P$. All structures were optimized in the gas phase. The convergence criteria for geometric optimization are set as 1.0×10^{-6} Hartree for energy change and 3.0×10-4Hartree/Å for gradient change, respectively. The geometric optimization of the S_0 is performed by using DFT method, and the structural optimization of S_1 is performed using TDDFT method. Vertical excitation energy and Vertical emission energy are calculated at the optimized ground state S_0 and excitedstate S_1 structures, respectively.

Based on the results of optimized S_0 and S_1 by ORCA5.0.2, the wave function information of hole and electron pairs were obtained by calculating TDDFT single point energy at PBE0/def2-SVP level with Gaussian 09 program^[5], and then electron-hole^[6] and inter fragment charge transfer (IFCT) $[7]$ analysis were performed using Multiwfn 3.8 (dev) program^[8].

1.5 Characterization.

UV-vis absorption spectra were recorded on an Agilent 8453 spectrophotometer. Fluorescence spectra were obtained using an F-7000 fluorescence spectrophotometer. X-ray photoelectron spectroscopy (XPS) measurements were performed on a thermal ESCALAB 250, equipped with a monochromated Al Kα (1486.8 eV) 150 W X-ray source, 0.5 mm circularspotsize, and a flood gun (to counter charging effects). The analysis chamber base pressure was lower than 1×10−9 mbar, and data were collected with FAT = 20 eV. Electrospray ionization mass spectra (ESI-MS) were recorded using a Waters Xevo G2-XS mass spectrometer. The source temperature was maintained at 80 °C. The sample was directly infused into the chamber at 5 μL/min. ESI sample was prepared by dissolving it in dichloromethane/methanol (0.1 mg/mL). NIR emission was taken in an edinburgh instruments FS5 Spectrophotometer. QY of the nanoclusters in this work was estimated using Au_{25} as a reference.^[9]

Figure S1. Near-infrared fluorescence emission wavelength measurements of Au₁₈(S $c - C_6H_{11}$ ₁₄, Au₂₅(PET)₁₈, Au₃₆(TBBT)₂₄ and Au₃₈(PET)₂₄ nanoclusters.

Figure S2. ESI-MS of [(Ph₃P)AuCl], interpolation is experimental and theoretical simulation. The m/z peak at 459.3 is $[(Ph_3P)Au]^+$, which is formed by the $[(Ph_3P)AuCl]$ molecule losing a Cl atom.

Figure S3. Temperature-dependent UV-vis absorption spectra of (a) Au₃₆ and (b) Au₃₇ nanoclusters.

Figure S4. The ³¹P NMR spectra of (a) TPP, (b) reaction process and (c) Au₃₇ nanocluster.

Figure S5. Full X-ray photoelectron spectroscopy (XPS) of (a) Au₃₆ and (b) Au₃₇ nanoclusters.

Figure S6. Unit cells of Au₃₇ nanocluster (color coded: Au = blue; S = yellow; P = green; $C = gray; H = white$).

Figure S7. Kernel Au-Au bond length statistics of (a) Au₃₆ and (b) Au₃₇ nanoclusters.

Figure S8. The calculated absorption spectra of Au₃₆ and Au₃₇ at TD DFT are compared with the experimental spectra (PBE0/def2-SV(P)).

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Figure S11. Au₃₇ nanocluster protected by different phosphine ligands.

Figure S12. Intensity spectra of near infrared emission peaks of Au₃₇ nanocluster in different solvents.

Figure S13. Intensity spectra of near infrared emission peaks of Au₃₇ nanocluster for phosphine ligands with different substituents.

Figure S14. Possible steps of structural transformation from Au₃₆ to Au₃₇ nanoclusters.

Figure S15. Fragment Definition of Au₃₆ and Au₃₇.

Figure S16. Contribution of phosphine ligands to emission.

Figure S17.The charge transfer heat map by IFCT Analysis.

Table S2 Crystal data and structure refinement for Au37(TBBT)21(TPP)2.

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