#### **Electronic Supplementary Information**

# Metallopeptide Nanoreservoirs for Concurrent Imaging and Detoxification of Lead (Pb) from human Retinal Pigment Epithelial (hRPE1) Cells

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1.0 Materials and Methods (General): - The solvents and reagents employed in this investigation were obtained from diverse commercial suppliers as per specific requirements. L-Phenylalanine and L-Tyrosine amino acids were procured from Spectrochem in Mumbai, India. 2,6-Pyridinedicarboxylic acid, employed as a linker, was sourced from SRL Pvt. Ltd. in India. Peptide coupling agents, including N, N'dicyclohexylcarbodiimide (DCC), N-hydroxybenzotriazole (HOBt), and 1-ethyl-3-(3dimethylaminopropyl) carbodiimide (EDC. HCl), were obtained from Avra Synthesis Pvt. Ltd. in Hyderabad, India. The protecting reagent, Di-tert-butyl dicarbonate (Boc anhydride), and the deprotecting reagent, Trifluoroacetic acid (TFA), were sourced from Spectrochem Pvt. Ltd. Organic bases, such as triethylamine (Et<sub>3</sub>N), diisopropylethylamine (DIPEA), and 4-Dimethylamino pyridine (DMAP), were supplied by S D Fine-Chem Ltd. Strong anion exchange resin (Dowex 1-X8) was acquired from HiMedia Laboratories Pvt. Ltd. in India, and strong cation exchange resin (Amberlite IR-120 Na form) was purchased from Spectrochem Pvt. Ltd. in Mumbai, India. Silica gel with mesh sizes of 60-120 and 100-200, along with precoated aluminum sheets for thin-layer chromatography (TLC Silica gel 60 F254), were procured from Merck Chemicals in India. Solvents, including methanol (MeOH), ethanol (EtOH), dichloromethane (DCM), ethyl acetate (EtOAc), acetone (Ac<sub>2</sub>O), chloroform (CHCl<sub>3</sub>), dimethylsulfoxide (DMSO), diethyl ether (Et<sub>2</sub>O), acetonitrile (ACN), pyridine, N, Ndimethylformamide (DMF), and tetrahydrofuran (THF), were purchased from S D Fine Chemicals Ltd. in India. Furthermore, HPLC-grade solvents and deuterated solvents for nuclear magnetic resonance (NMR) spectra were sourced from Merck Pvt. Ltd. Metal salts, specifically those of mercury, zinc, cadmium, and lead, were obtained from Himedia Laboratories Pvt. Ltd. in Mumbai, India. All other requisite reagents were secured from HiMedia.

**2.0 Peptide Synthesis-** The synthesis of tripeptide(Tyr-Phe-Phe) and the  $C_2$ -Symmetric pyridine-bis- Tyr-Phe-Phe metallopeptide conjugate MPC was performed by conventional solution phase synthesis via established lab protocol mentioned in

research paper published our group. The purity and identity of these compounds were confirmed before their utilization. All experiments were conducted under ambient room temperature conditions.

**3.0 Fluorescence spectroscopy-** Fluorescence intensity was assessed at room temperature using an excitation wavelength ( $\lambda_{ex}$ ) of 260 nm. The interaction between Tyr-Phe-Phe and MPC-1 with relevant metal ions was individually investigated to discern alterations in their behaviour. Emission spectra were captured within the range of 280 nm to 500 nm. A stock solution of each metal ion salt at a concentration of 15 mM in water was prepared. For each metal ion, the ethanolic solution was titrated with the peptide (20  $\mu$ M) up to a maximum concentration of 80  $\mu$ M. Fluorescence spectra were recorded using a Varian Luminescence Cary Eclipse spectrophotometer equipped with a 10 nm quartz cell, while maintaining a controlled temperature of 25 ± 0.1°C. High-performance liquid chromatography (HPLC)-grade water and ethanol were employed in these research endeavours.

**4.0 UV studies**- UV-Vis absorption spectra were collected using a Lab India UV-VIS Spectrophotometer 3000+ equipped with a 10 mm quartz cell, and the measurements were maintained at a constant temperature of  $25\pm0.1$  °C. To initiate the experiment, a stock solution with a concentration of 1 mM for each type of metal ion was meticulously prepared in water. Subsequently, solutions containing 10 µM of both tripeptide and pyridine-bis- Tyr-Phe-Phe Metallopeptide Conjugate (MPC) in ethanol were titrated with each specific metal ion until reaching a final concentration of 200 µM, utilizing the previously mentioned stock solution.

**5.0 FT-IR Study-** To assess the functional group composition of both the peptides, with and without the presence of metal ions, Fourier-transform infrared (FT-IR) spectra were acquired utilizing a Bruker Alfa II attenuated total reflection (ATR) instrument. The spectral data encompassed the range of 4000-500 cm<sup>-1</sup>, with a spectral resolution of 4

cm<sup>-1</sup>, employing 2 sample gain and 32 sample/background scans. Data processing was carried out using OPUS 7.0 software, incorporating noise removal procedures.

A 200 µM solution of MPC, both in the absence and in the presence of metal ions at a 1:3 ratio, was applied and dried onto a ZnSe substrate. Subsequently, the spectral region corresponding to Amide I, ranging from 1700 to 1600 cm<sup>-1</sup>, underwent deconvolution analysis using Origin software. This deconvolution process was guided by the count of initial peak values derived from the second derivative spectrum. A quantitative evaluation of the area linked to each spectral component was carried out to determine its influence on the secondary structural features.

**6.0 Circular Dichroism spectroscopy:**- CD spectra were collected between 195 nm to 270 nm and each spectrum was the average of 3-5 scans. All experiments were carried out at  $25\pm0.1$  °C. Spectra were recorded at the final concentration of MPC-1 alone and MPC-1+Pb<sup>2+</sup> at 200  $\mu$ M, on JASCO J-815 CD SPECTROMETER by using a quartz cuvette with a path length of 1mm. To avoid any instrumental baseline drift between any measurements, the background value was subtracted for each sample measurement with ethanol and ethanol water wherever needed.

7.0 Atomic Force Microscopy (AFM)- The test sample(s) at appropriate concentration were meticulously prepared in a controlled, dust-free environment and deposited onto a freshly cleaved muscovite mica surface. Subsequently, they were subjected to drying under a 60W light bulb for 30 minutes, followed by vacuum drying followed by imaging using an atomic force microscope (AFM) of the INNOVA model, an ICON analytical equipment by Bruker. The AFM operated in both contact and tapping modes, utilizing a cantilever (NSC 12(c) from MikroMasch) featuring a Silicon Nitride Tip managed by NanoDrive<sup>™</sup> version 8 software. The cantilever had a resonant frequency of approximately 260 kHz. Imaging was conducted under ambient conditions at a temperature of 25±0.1 °C, with a scan speed ranging from 2 to 1.5 lines per second. Subsequent data analysis was performed employing the Nanoscope Analysis software.

**8.0 Scanning Electron Microscopy (SEM)-** Scanning Electron Microscopy (SEM) images were acquired using a FEI QUANTA 200 microscope equipped with a tungsten filament gun. The microscope was set to a working distance of 4.0 mm and operated at a voltage of 7.99 kV. A minute volume (10  $\mu$ L) of, MPC-1 at 200  $\mu$ M concentrations was delicately deposited onto the designated surface(s) and allowed to undergo overnight air drying at room temperature. Subsequently, it underwent an additional phase of drying under high vacuum conditions lasting 30 minutes. The specimen was then subjected to a 1-minute gold coating process and subsequently observed using SEM.

**9.0 Transmission Electron Microscopy (TEM):-** In accordance with established protocols for Transmission Electron Microscopy (TEM) sample preparation, a freshly prepared 10  $\mu$ L solution of 200  $\mu$ M concetration was meticulously dispensed onto a copper grid that had been coated with a carbon layer featuring a mesh size of 200. Any surplus sample was delicately eliminated from the grid, and the residual specimen was permitted to undergo an air-drying process at room temperature for duration of 6 hours. The samples were subsequently subjected to examination utilizing a FEI Titan G2 60-300 Transmission Electron Microscope (TEM).

**10.0 Confocal microscopy-** Cellular specimens affixed to glass substrates were subjected to comprehensive examination through the utilization of a Leica TCS SP8 confocal scanning laser microscope. The selective excitation of distinct fluorophores was achieved through laser emissions of specific wavelengths: 405 nm for 4',6-diamidino-2-phenylindole (DAPI) and 488 nm for phalloidin. Subsequently, image analysis was meticulously conducted employing the Fiji ImageJ software, facilitating the precise removal of background signal artifacts and the subsequent quantification of cellular intensity and area measurements. A systematic quantification protocol was applied to 30 to 40 individual cells. The resulting dataset, comprising only validated and rigorously assessed data points, was then subjected to graphic representation using the

GraphPad Prism software, aligning with the established practices of highly reputable journals within the biological sciences.

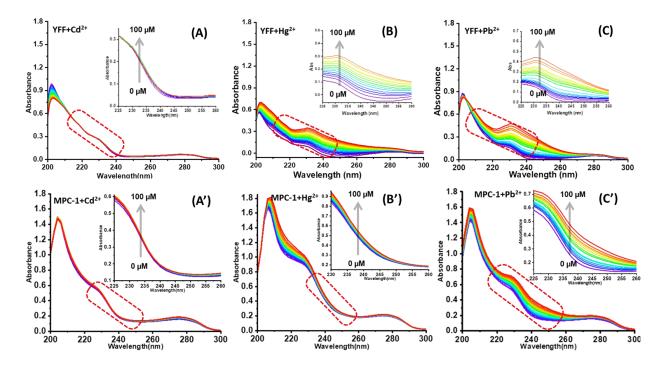
**11.0 Cell Culture and Treatment:-** For the cellular uptake experiment, the RPE1 cells were cultured in DMEM containing 10% fetal bovine serum and antibiotic at 37 °C with 5% CO<sub>2</sub> in a humidified incubator. Approximately  $1\times10^5$  cell counts per well were seeded on a glass coverslip in a 6-well plate overnight. Before treatment, the seeded cells were washed with 1× PBS buffer three times. After washing, the cells were treated with Pb<sup>2+</sup> salt solution (500nm) and Pb<sup>2+</sup> ions +peptide (1:1) for 24h. The treated cells were fixed for 15 min at 37 °C with 4% paraformaldehyde and rinsed three times with 1×-PBS. The cells were then permeabilized with 0.1% Triton-X100 and stained with 0.1% Alexa Fluor<sup>TM</sup> 488 phalloidin to visualize the actin filaments. Then the cells were washed three times with 1× PBS and mounted onto the slides with Mowiol and DAPI to stain the nucleus.

**12.0 Computational studies/ Details:** Calculations were executed utilizing the ORCA quantum chemical program package. Geometries were optimized using the GGA density functional BP86 and def2-SVP basis sets. Non-covalent interactions were considered through dispersion corrections (D3) with Becke-Johnson (BJ) damping. Subsequent numerical frequency calculations were performed on optimized geometries to ensure they represent stationary points with no imaginary frequencies. Binding energies were determined through single-point calculations using the BP86/def2-TZVP/C-PCM(EtOH) method applied to BP86/def2-SVP geometries(references cited).

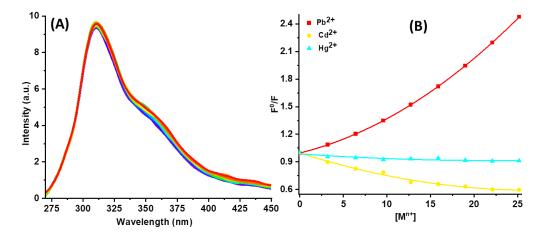
**13.0 MTT Assay:** An MTT assay was performed to evaluate the cytotoxicity of the synthesized peptide, lead(II) ions (Pb(II)), and the peptide-Pb(II) conjugate on human retinal pigment epithelial (RPE-1) cells. RPE-1 cells were seeded in a 96-well plate at a density of 20,000 cells per well and incubated for 24 hours at 37°C to allow for cell attachment and recovery. Subsequently, the cells in different wells were treated with

the peptide, Pb(II), and the peptide-Pb(II) conjugate at a concentration of 500 nM, while untreated cells served as a control group. Following an additional 24-hour incubation at 37°C, a solution of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) was added to each well, and the plates were incubated for 4 hours to allow viable cells to metabolize the MTT and form insoluble formazan crystals. The MTT solution was then removed, and dimethyl sulfoxide (DMSO) was added to solubilize the formazan crystals. After a 15-minute incubation in the dark, the absorbance of the resulting coloured solution was measured at 570 nm using a multi-well microplate reader, with higher absorbance values indicating greater cell viability.

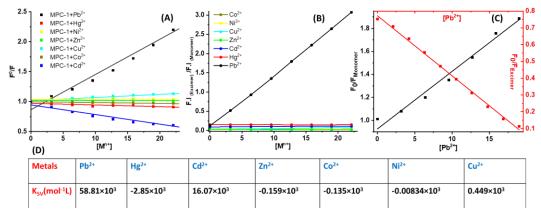
#### 14.0 Figures:



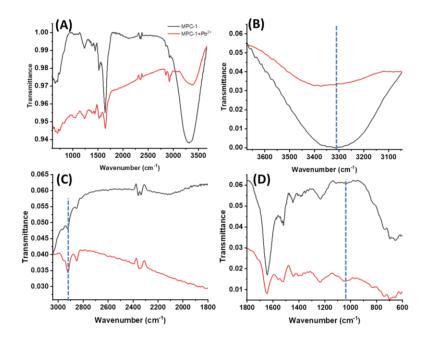
**Figure S1:** UV-Vis titration spectra illustrating the impact of the gradual addition of  $Cd^{2+}$  ions (A, A'),  $Hg^{2+}$  ions (B, B'), and  $Pb^{2+}$  ions (C, C') (1 mM) on tripeptide and MPC respectively. The spectra show the increase in absorbance intensity of the aromatic sidechain, acid and amide region.



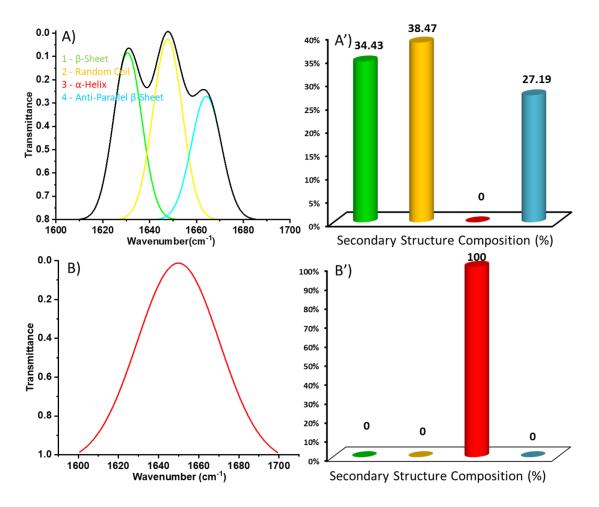
**Figure S2:** (A) Fluorescence emission spectra of MPC-1 with the gradual addition of  $Hg^{2+}$  ions ( $\lambda_{ex} = 260$  nm). (B) Stern-Volmer plots illustrate the competitive complexation of MPC-1 with Pb<sup>2+,</sup> Cd<sup>2+,</sup> and Hg<sup>2+</sup> ions.



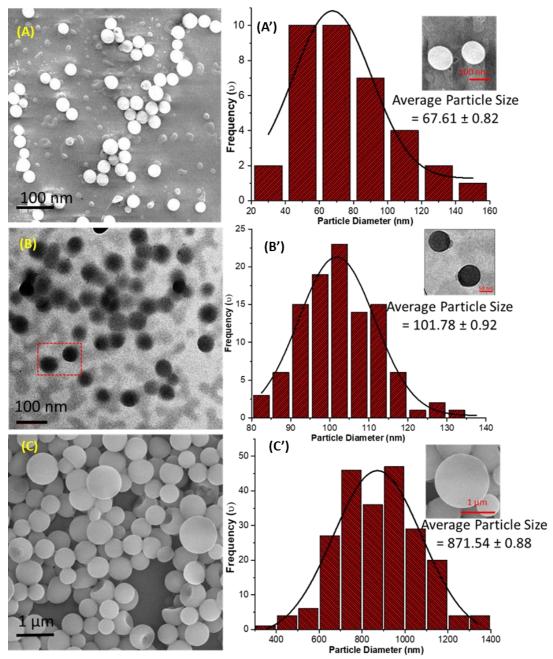
**Figure S3:** (A) Stern–Volmer plots of MPC-1 with the gradual addition of Co<sup>2+</sup>, Ni<sup>2+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Cd<sup>2+</sup>, Hg<sup>2+</sup>, and Pb<sup>2+</sup>. (B) A ratiometric plot of MPC-1 for Excimer to Monomer formation (Excimer  $\lambda_{em} = 450$ nm/Monomer  $\lambda_{em} = 360$ nm) in the presence of various metal ions, utilizing an excitation wavelength of  $\lambda_{ex} = 260$ nm. (C) Stern–Volmer plots of monomer (**Left Axis**) and excimer (**Right axis**) formation with MPC-1 on gradual addition of Pb<sup>2+</sup> ions.



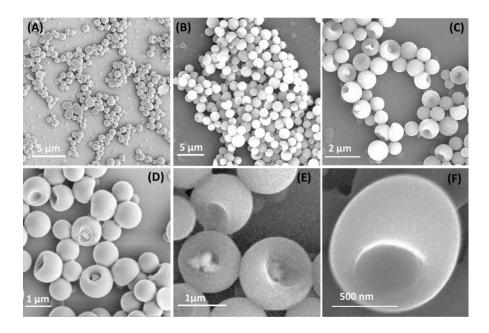
**Table S4: (A)** FT-IR spectra of MPC-1 alone (**Black**) and with Pb<sup>2+</sup> ion (**Red**); (**B**) Amide A and B regions ranging from 3600–3000 cm<sup>-1</sup>, representing N–H bond stretching; (**C**) Amide I and II regions ranging from 1800 to 1500 cm<sup>-1</sup>; and (**D**) Fingerprint region ranging from 1400 to 600 cm<sup>-1</sup>. These spectra depict clear differences in the free H-bonding and N–H vibrations in the amide regions A, as well as differences in the nature of H-bonding interactions after the addition of Pb<sup>2+</sup> ion. Conversely, differences in the out-of-plane (Ar C–H) vibration band were observed in the fingerprint region (D) (1400 cm<sup>-1</sup>to 600 cm<sup>-1</sup>). Additionally, the amide regions I and II, which provide insights into the differences in the amide backbone, showed quit similar band positions.



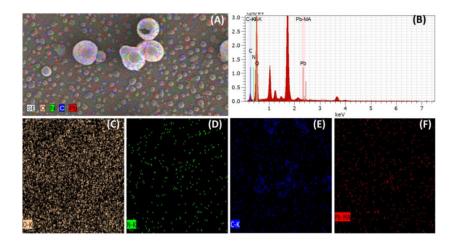
**Figure S5: (A)** FT-IR spectrum of MPC-1 (solid line) and its deconvolution (coloured line), A') Corresponding secondary structure contribution in self-assembly, **(B)** FT-IR spectrum of MPC-1 with lead (solid line) and its deconvolution (dashed line), B') Corresponding secondary structure contribution in self-assembly. Deconvolution was done using multiple Gaussian peaks fit in the amide I region, ranging from 1,600 to 1,700 cm<sup>-1</sup>.



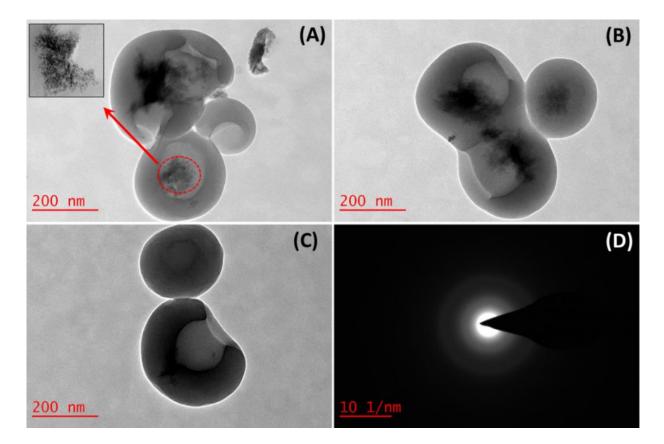
**Figure S6:** Quantitative morphological analysis of MPC-1 before and after the addition of Pb<sup>2+</sup> ions **(A)** SEM images of MPC-1 showing nanovesicle-like self-assembly and (A') its corresponding particle (diameter of vesicles) size distribution histogram suggest the average size of particles around 67 nm which is well corresponded with **(C)** TEM images of MPC-1 and (C') represent its particle size distribution histogram depict the average particles size around 101 nm, (D)& (D') shows SEM image and its particle size distribution after the addition of Pb<sup>2+</sup> ions suggest the unusual enlargement of vesicles size from 100 nm to 871 nm and aggregation behaviour might be due to encapsulation or complexation of pb<sup>2+</sup> ions into nanovesicles.



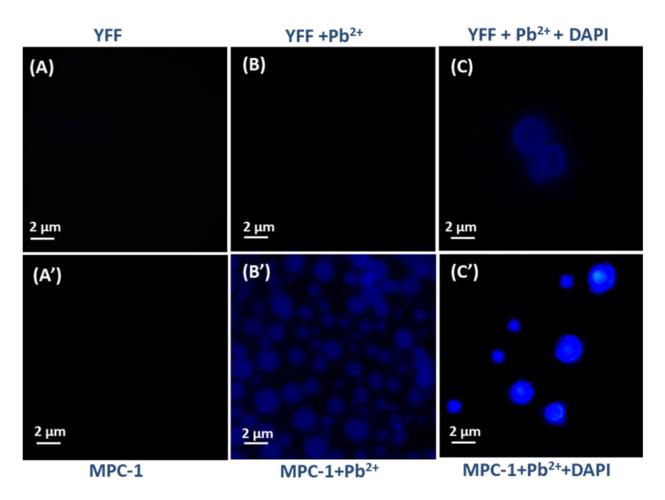
**Figure S7:** Insight into the Morphological Characteristics of the MPC1-Pb<sup>2+</sup> Complex via SEM Analysis. Detailed SEM images delineating the hierarchical assembly of coagulated spherical/pot-like structures, indicative of peptide-mediated self-assembly processes nano-manipulated by Pb<sup>2+</sup> ions. Notably, an ordered arrangement is observed, with a distinct inner view highlighting the cohesive chain-like structures of MPC1-Pb complexes. Further SEM observations reveal the intricate pot-like architecture filled with internal material, underscoring the complexity and organization inherent in peptide-driven assembly mechanisms.



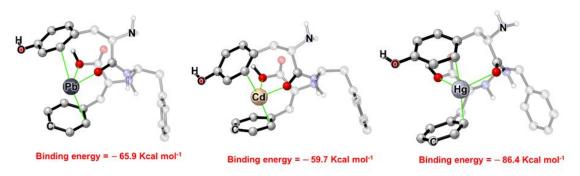
**Figure S8:** Characterization of self-assembled structures through EDS and colour mapping analysis of SEM images. The elemental composition revealed by EDS indicates the presence of core components such as Oxygen, Nitrogen, and Carbon from the MPC molecule. Additionally, the structures exhibit regions highlighted in red, indicative of the presence of Pb<sup>2+</sup> ions. This observation directly correlates with the nanomanipulated embedding of lead within MPC-1 self-assembled vesicles.



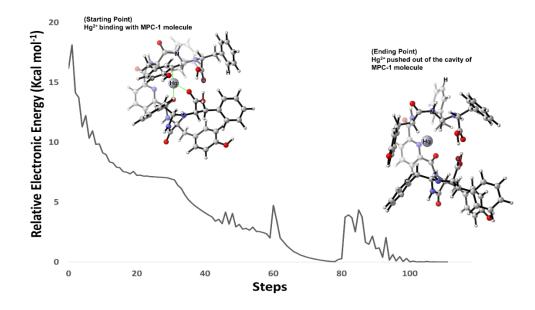
**Figure S9:** TEM image analysis unveils the intricate self-assembly dynamics of MPC1-Pb<sup>2+</sup> structures. (A) Highlights the pot/vesicle-like assembly, revealing the fusion of vesicles embedded by lead or the initial stages of pot-like structure formation. Notably, TEM images (B) exhibit varying colour contrasts within the soft structures, indicative of potential heavy metal ion presence, specifically lead. (D) Presents the SAED pattern of the image (B), affirming the amorphous nature of lead-embedded structures, and providing further evidence for the incorporation of lead within these assembled nanostructures.



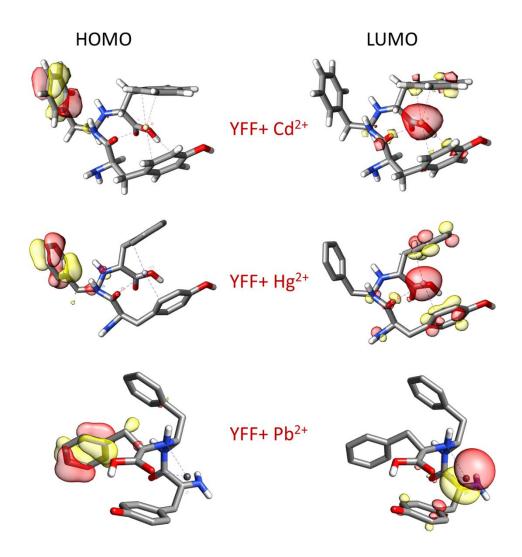
**Figure S10:** Fluorescence microscopy was utilized to probe the morphology of YFF and MPC-1 in the presence of Pb<sup>2+</sup> ions and DAPI. Initial imaging (A) & (A') without staining revealed no discernible morphology for either YFF or MPC1 under 358 nm excitation observed through a blue filter ( $\lambda_{em}$ =450nm) in the dark field, indicating the absence of inherent fluorophores necessary for proper imaging. Upon introduction of Pb<sup>2+</sup> ions (B) & (B'), YFF+Pb<sup>2+</sup> showed no noticeable morphological changes, whereas MPC-1&Pb<sup>2+</sup> exhibited distinct blue emission, suggesting intrinsic blue light emission capability within the MPC-Pb complex. Subsequent staining with DAPI for a 12-hour incubation period resulted in minute signals for YFF-Pb complexes, while MPC1-Pb showed increased emission intensity (C) & (C'), particularly concentrated around the Pot/Sphere-like structure, potentially indicative of MPC-Pb-DAPI interactions.



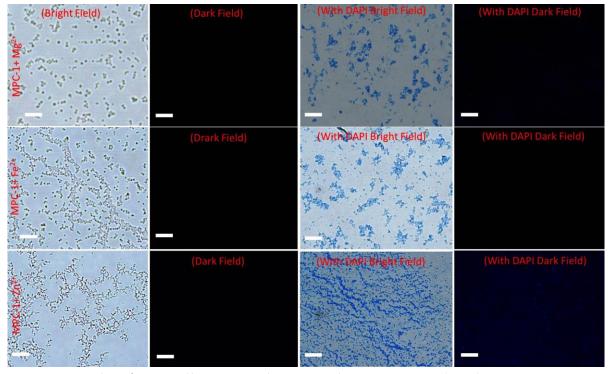
**Figure S11:** Depicts the active binding site of YFF's cavity and its interaction with different metals (Pb<sup>2+</sup>, Cd<sup>2+</sup>, Hg<sup>2+</sup>) along with their binding energies.



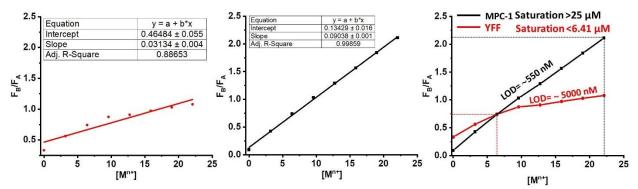
**Figure S12:** Energy convergence plot depicting the interaction between  $Hg^{2+}$  ions and the MPC-1 molecule. The plot illustrates the placement of  $Hg^{2+}$  within the cavity of the MPC-1 molecule, and as the convergence point approaches, it indicates the gradual release of  $Hg^{2+}$  from the cavity.



**Figure S13:** Computational model illustrating the optimized geometry of the YFF molecule, highlighting the configuration of its binding cavities/sites. The image depicts the interactions of Pb<sup>2+,</sup> Cd<sup>2+,</sup> and Hg<sup>2+</sup> ions within the specified cavities.



**Figure S14:** This figure illustrates the optical microscopy (OM) images captured in both bright field and dark field in the presence of three biologically relevant metal ions:  $Mg^{2^+}$ ,  $Fe^{2^+}$ , and  $Zn^{2^+}$ . The top panel displays OM images of MPC-1 with  $Mg^{2^+}$ ; the middle panel shows OM images of MPC-1 with  $Fe^{2^+}$ ; and the bottom panel presents OM images of MPC-1 with  $Zn^{2^+}$ , both without and with DAPI. These images demonstrate the specificity of MPC-1 for lead ions in complex biological environments. Scale bar 5µm.



**Figure S15:** Ratiometric plot of (A) YFF with Pb(II) and (B) MPC-1 with Pb(II) used for calculations of LOD and LOQ which is further merged in figure (c) for better comparison.

S1. No.	Probe	LOD	Methods/Techniques	Selectivity for Pb <sup>2+</sup> tested in the presence of M <sup>n+</sup> ions	Real-world Practical Applications	Reference no. in the main text
1.	Gallic acid-Au NPs	5.0 μΜ	Based on Ratiometric Uv-Vis Spectroscopic Absorbance	Ca <sup>2+</sup> , Cu <sup>2+</sup> , Cd <sup>2+</sup> , Hg <sup>2+</sup> , Mg <sup>2+</sup> , Ni <sup>2+</sup> , and Zn <sup>2+</sup>	Naked-Eye Detection from Aqueous Media	67
2.	Glutathione-Capped Quantum Dots	40 nm	Based on Fluorescence Quenching of GSH-capped QDs	Na <sup>+</sup> , Ba <sup>2+</sup> , Ca <sup>2+</sup> , Mg <sup>2+</sup> , Co <sup>2+</sup> , Ni <sup>2+</sup> , Li <sup>+</sup> , K <sup>+</sup>	Not Available	68
3.	Peptide-Au NPs	242 nM	Based on Uv-Vis Spectroscopic Absorbance and Colorimetric response	No selectivity	Not Available	58
4.	2- Mercaptoethanol/S <sub>2</sub> O <sub>3</sub> 2-Au NPs	0.5 nM	Based on the Leaching of Gold Nanoparticles	No selectivity	Not Available	69
5.	Au NPs, DNAzyme and substrate	0.5 μΜ	Based on quenching and ratiometric absorbance of DNAzyme using unmodified gold nanoparticle probes	Towards $(Zn^{2+}, Co^{2+}, Mg^{2+}, Ca^{2+}, Mn^{2+}, Cd^{2+}, Ni^{2+}, and Cu^{2+}$	Not Available	70
6.	DNA-Au NPs, DNAzyme and substrate	0.5 μΜ	Based on Uv-Vis Spectroscopic Absorbance and Colorimetric response	NA/NA	Not Available	71
7.	Graphene Quantum Dots/L-Cysteine	70 nM.	Quenching of Graphene Quantum Dots/L-Cysteine Coreactant Electrochemiluminescence System	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Not Available	72
8.	1,8-naphthalimide dye with cellulose nanocrystals (CNCs)	150 nM	Fluorescence enhancement of cellulose nanocrystals CNCs	Ba <sup>2+</sup> , Cu <sup>2+</sup> , Zn <sup>2+</sup> , Fe <sup>2+</sup> , Co <sup>2+</sup> , Ni <sup>2+</sup> , and Mg <sup>2+</sup>	Not Available	73

9.	A group of Cysteine-rich cyclic peptides	NA	Not mentioned	Zn <sup>2+</sup> and Ca <sup>2+</sup>	Detoxification in bacterial (DH5a cells) and human cell culture (HT- 29 cells)	8
10.		0.49 μM	Based on the fluorescent enhancement response	$\begin{array}{c} Cr^{3+,}\ Mn^{2+}\ ,\ Fe^{3+,}\ ,\ Co^{2+}\ ,\\ Ni^{2+}\ ,\ Cu2^{2+}\ ,\ Zn^{2+}\ ,\ Hg^{2+}\ ,\\ Bi^{3+,}\ \ Ag^{+,}\ \ Cd^{2+}\ ,\ Th^{4+}\ ,\\ Ce^{4+}\ ,\ Nd^{3+,}\ and\ U^{6+} \end{array}$	Not Available	74
11		NA	Based on Ratiometric Fluorescence Spectroscopic response.	K <sup>+</sup> , Na <sup>+</sup> , Ag <sup>+</sup> , Ba <sup>2+</sup> , Ca <sup>2+</sup> , Cd <sup>2+</sup> , Co <sup>2+</sup> , Cu <sup>2+</sup> , Fe <sup>2+</sup> , Hg <sup>2+</sup> , Mg <sup>2+</sup>	Not Available	75
12.		160 nM	Based on the excited state intramolecular proton transfer (ESIPT)	Fe <sup>2+</sup> , Co <sup>2+</sup> , Ni <sup>2+</sup> ,Cu <sup>2+</sup> , Zn <sup>2+</sup> , Cd <sup>2+</sup> , Hg <sup>2+</sup> , Mg etc	Not Available	76
13.		6.3 nM	Based on the Ratiometric fluorescent detection	Fe <sup>2+</sup> , Co <sup>2+</sup> , Ni <sup>2+</sup> , Cu <sup>2+</sup> , Zn <sup>2+</sup> , Cd <sup>2+</sup> , Hg <sup>2+</sup> , Mg+, Mn <sup>2+</sup> etc.	Ratiometric detection of Pb <sup>2+</sup> in tap water, groundwater, and live cell.	77
14.	N N	3.4 µM	Based on ratiometric fluorescent response.	K <sup>+</sup> , Na <sup>+</sup> , Ca <sup>2+</sup> , Ag <sup>+</sup> , Mg <sup>2+</sup> , Hg <sup>2+</sup> , Co <sup>2+</sup> , Fe <sup>2+</sup> , Mn <sup>2+</sup> , Pd <sup>2+</sup> , Cd <sup>2+</sup> , Ni <sup>2+</sup> , Zn <sup>2+</sup>	Cu <sup>2+</sup> and Pb <sup>2+</sup> were detected in a test strip	78
15.		NA	Based on Ratiometric Fluorescence Spectroscopic response.	Ca <sup>2+</sup> , Cd <sup>2+</sup> , or Zn <sup>2+</sup>	Not Available	79

16.		210 nM	Based on chromogenic and fluorogenic 'turn-on' spectral responses	Mn <sup>2+</sup> , Fe <sup>2+</sup> , Co <sup>2+</sup> , Ni <sup>2+</sup> , Cu <sup>2+</sup> , Zn <sup>2+</sup> , Cd <sup>2+</sup> ,Hg <sup>2+</sup> , Ag <sup>+</sup> etc	Not Available	80
17.		3.4 nM	Based on the Ratiometric response of Intermolecular fluorescence resonance energy transfer (FRET) processes from the porphyrin donor to the phthalocyanine acceptor.	Ag <sup>+</sup> , Al <sup>3+</sup> , Ba <sup>2+</sup> , Ca <sup>2+</sup> , Cd <sup>2+</sup> , Co <sup>2+</sup> , Cr <sup>3+</sup> , Cu <sup>2+</sup> , Fe <sup>2+</sup> , Fe <sup>3+</sup> , Hg <sup>2+</sup> , K <sup>+</sup> , Mg <sup>2+</sup> , Mn <sup>2+</sup> , Na <sup>+</sup> , , NH <sup>4+</sup> , Ni <sup>2+</sup> , and Zn <sup>2+</sup>	Not Available	81
18.		4.1 nM	Based on the Ratiometric response of intramolecular fluorescence resonance energy transfer (FRET) process	Ag <sup>+</sup> , Al <sup>3+</sup> , Ba <sup>2+</sup> , Ca <sup>2+</sup> , Co <sup>2+</sup> , Cr <sup>3+</sup> , Cu <sup>2+</sup> , Fe <sup>2+</sup> , Fe <sup>3+</sup> , K <sup>+</sup> , Mg <sup>2+</sup> , Mn <sup>2+</sup> , Na <sup>+</sup> , NH <sub>4</sub> <sup>+</sup> and Ni <sup>2+</sup>	Not Available	82
19.		NA	Enhancement of the monomer emission of pyrene	Ag+, Co <sup>2+</sup> , Ni <sup>2+</sup> , Hg <sup>2+</sup> , Zn <sup>2+</sup> , Cd <sup>2+</sup> , Cu <sup>2+</sup>	Not Available	83
20.	Pyridine-bis-YFF peptide conjugate	~550 nM	Based on Ratiometric Monomer- Excimer formation	Fe <sup>2+</sup> ,Co <sup>2+</sup> , Ni <sup>2+</sup> , Cu <sup>2+</sup> , Zn <sup>2+</sup> , Cd <sup>2+</sup> , Hg <sup>2+</sup> , Mg <sup>2+</sup>	In vivo detoxification in RPE-1 cell lines	Current work

**Table S1:** Offers a detailed review of current technologies, encompassing their detection limits, selectivity, and practical applications. This comprehensive assessment highlights the novelty and superiority of our proposed metallopeptide nanoreservoirs.

# 15.0 Cartesian coordinates for the optimized Geometries

DFT optimization method: BP86-D3/def-SVP

### MPC-1 molecule

С	10.882182618014	-6.935027692862	18.684288341374
Η	11.073546942116	-7.942547205117	19.092886865569
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С	10.345777257789	-6.030880571280	19.818032700603
Η	10.132866062743	-5.026640202713	19.395692368652
Н	9.388071226404	-6.460566798872	20.176332880251
С	11.350827440181	-5.929817186414	20.943676160511
С	11.405106629101	-6.909977424762	21.954993971828
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Ν	13.345854707396	-6.870204626197	18.366024215502
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Η	13.018038456934	-8.438816454598	15.772773201694
С	16.300381896639	-10.493445545134	16.299329533521
С	14.207853449944	-9.579811816721	17.187479183105
Η	17.216067169253	-11.085748134059	16.449072914997
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Н	16.695928154674	-11.524209938505	25.150517992060
H	16.723553814647	-6.764479177501	23.603533326531
H	12.877216924901	-8.050825720775	20.023333417914
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H	17.084469185667	-9.271534572125	20.900378384308
H	12.210271227282	-9.318354232238	17.966779100794
H	13.288074684806	-10.235600602134	19.013646072599
H	14.176101249832	-6.408989423095	17.982665373478
H	14.178101249832	-7.820409382261	21.728518117441
H	14.493303327747	-8.591139862505	19.076832369458
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C	12.523688957438	-12.648794650401	16.593868967957
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Н	9.979809681388	-15.333181085701	15.350269745299
Н	13.247055759861	-11.895424085912	16.934230679840
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Η	14.116003576733	-11.998286514816	20.181147462194
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Η	4.598946565084	-12.650264679247	20.986573448021
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### MPC-1 molecule + Pb<sup>2+</sup>

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Ο	10.808894000000	-9.334531000000	18.286956000000
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Pb	11.612738000000	-11.450480000000	17.821808000000

## YFF molecule

С	-3.096373000000	-1.792173000000	1.898545000000
Η	-4.093559000000	-1.448818000000	2.241613000000
С	-2.573660000000	-2.963327000000	2.766865000000
Ο	-1.607338000000	-3.618809000000	2.359423000000
С	-2.119486000000	-0.595728000000	2.024495000000
Η	-1.118352000000	-0.936900000000	1.688186000000
Η	-2.465408000000	0.182300000000	1.314372000000
С	-2.059399000000	-0.076746000000	3.439550000000
С	-3.061537000000	0.786520000000	3.930158000000
С	-1.089190000000	-0.544735000000	4.350049000000
С	-3.123073000000	1.140400000000	5.285478000000
С	-1.135483000000	-0.198541000000	5.706652000000
С	-2.171448000000	0.625361000000	6.191059000000
Η	-3.823052000000	1.179503000000	3.236776000000
Η	-0.302347000000	-1.226136000000	3.990687000000
Η	-3.918746000000	1.805119000000	5.653651000000

Н	-0.393890000000	-0.587800000000	6.419726000000
0	-2.230914000000	0.866654000000	7.535092000000
Η	-3.175544000000	0.972434000000	7.783164000000
Ν	-3.192614000000	-3.261274000000	3.949259000000
С	-4.247989000000	-2.542230000000	4.688273000000
Ν	-4.191063000000	-3.000681000000	6.066448000000
С	-4.103892000000	-2.025000000000	7.128218000000
С	-5.342037000000	-1.124821000000	7.244754000000
Ο	-5.039528000000	0.145324000000	7.653748000000
0	-6.490194000000	-1.469538000000	7.034459000000
Η	-5.900321000000	0.605641000000	7.778127000000
С	-5.658796000000	-2.755637000000	4.061319000000
С	-6.134387000000	-6.525035000000	3.579076000000
С	-6.953253000000	-6.847813000000	4.675477000000
С	-5.738248000000	-5.196733000000	3.360648000000
Η	-7.264330000000	-7.890098000000	4.844634000000
Η	-5.094093000000	-4.949711000000	2.502797000000
С	-7.377092000000	-5.833334000000	5.549005000000
С	-6.156740000000	-4.168162000000	4.231965000000
Η	-8.024698000000	-6.077214000000	6.405418000000
С	-6.981053000000	-4.502653000000	5.330027000000
Η	-7.309142000000	-3.701634000000	6.011747000000
Η	-5.801818000000	-7.315102000000	2.888246000000
С	-3.905675000000	-2.764710000000	8.481297000000
С	-2.742781000000	-1.078755000000	9.977851000000
С	-3.886996000000	-1.846683000000	9.680159000000
С	-2.751781000000	-0.165036000000	11.042464000000
Η	-1.850806000000	0.429507000000	11.257601000000
С	-5.039141000000	-1.681836000000	10.475765000000
С	-3.906396000000	-0.006783000000	11.828453000000
Η	-5.939879000000	-2.275222000000	10.249975000000
Η	-3.912142000000	0.708540000000	12.665151000000
С	-5.050544000000	-0.770309000000	11.544122000000
Η	-5.957084000000	-0.657633000000	12.158713000000
Η	-1.841495000000	-1.182089000000	9.355615000000
Η	-4.017676000000	-1.456242000000	4.659178000000
Η	-2.958701000000	-3.333356000000	8.387396000000
Η	-4.726499000000	-3.506195000000	8.585339000000
Η	-5.614251000000	-2.487167000000	2.986985000000
Η	-6.351305000000	-2.045535000000	4.554240000000
Η	-2.755323000000	-4.016613000000	4.486150000000
Η	-3.225844000000	-1.374396000000	6.954775000000
Η	-4.919670000000	-3.702462000000	6.253445000000

Ν	-3.205526000000	-2.192930000000	0.494781000000
Η	-3.963227000000	-2.884170000000	0.401306000000
Η	-2.343968000000	-2.721808000000	0.280418000000

### YFF molecule + Pb<sup>2+</sup>

С	-3.573231348781	-0.974476675882	3.078453061435
Η	-4.351352577125	-0.329266792869	3.551125172304
С	-3.346540573274	-2.132698115971	4.064512903884
Ο	-2.257113798274	-2.230552636720	4.730666238230
С	-2.298141811785	-0.087180479855	2.917051437350
Η	-1.431691559848	-0.722288353340	2.647205060312
Η	-2.496204905928	0.579884674545	2.053759189270
С	-1.989825177321	0.749108507673	4.138005974082
С	-2.816175437859	1.841730127175	4.495473013198
С	-0.905995618705	0.455582611042	4.995532822111
С	-2.625725141879	2.565884937057	5.688886710031
С	-0.703780276198	1.153730465396	6.210340963165
С	-1.568910763803	2.223608716193	6.570854380089
Η	-3.635339625038	2.149651540734	3.825479219219
Η	-0.183041256300	-0.321228774553	4.696669981056
Η	-3.268091321244	3.433972916171	5.910453941815
Η	0.191729700020	0.978998311338	6.828607604352
Ο	-1.315503051458	2.856270311078	7.727050166895
Η	-1.871578751503	3.656134315879	7.825077587736
Ν	-4.304830636409	-3.056246268948	4.213042010937
С	-5.783659896372	-2.849059582399	4.158881261993
Ν	-6.268891034014	-2.680120694584	5.529898998126
С	-5.426134020752	-1.966895342488	6.434805695024
С	-5.285276182316	-0.499494009267	6.020241555417
Ο	-4.067485662519	0.012446362720	6.491109806468
Ο	-6.032701205404	0.170590711363	5.364224470104
Η	-3.901850836646	0.933858526561	6.122508912741
С	-6.472673882712	-4.055514568983	3.493650770332
С	-4.748907329761	-7.186188412768	4.888193687912
С	-5.651017231916	-7.583731319486	5.890261101012
С	-5.033210722745	-6.067919009240	4.085602188772
Η	-5.436250123547	-8.469408089779	6.506608425149
Η	-4.344266959365	-5.793187096356	3.270038185172
С	-6.847398331053	-6.870579825557	6.073868804753
С	-6.223952961094	-5.328578121370	4.273302359382
Η	-7.576307118444	-7.200823526821	6.829275195814
С	-7.131926227933	-5.751414608638	5.271306697301

Η	-8.094686973009	-5.226557891382	5.390648631317
Η	-3.830866837125	-7.767143747006	4.712290888037
С	-5.801514057612	-2.115794000306	7.943501211217
С	-3.760688505230	-3.292492286903	8.888978622137
С	-4.543118628608	-2.109935026431	8.789566770341
С	-2.532906367218	-3.295753900731	9.583528799872
Η	-1.958855229960	-4.230233800546	9.675696587414
С	-4.058447661519	-0.931796038000	9.406520630462
С	-2.055895897518	-2.109621272574	10.179814280159
Η	-4.655882376844	-0.008836928753	9.358909040913
Η	-1.106299102046	-2.112050407856	10.735720019345
С	-2.822321003159	-0.929167229918	10.090073805308
Η	-2.469764592074	-0.005932918447	10.574437052162
Η	-4.131247462855	-4.228183944726	8.438975097939
Η	-5.956253993225	-1.941239089605	3.553181428209
Η	-6.321227197974	-3.088722083820	8.056599850488
Η	-6.515515617357	-1.328979336826	8.255253213606
Η	-6.083295709998	-4.121639447682	2.459344577954
Η	-7.555235227889	-3.829147116710	3.429341049267
Η	-4.027917571620	-3.887507672210	4.760028579575
Η	-4.368346987016	-2.366171940668	6.397323266502
Η	-6.565673660171	-3.586219416839	5.922934059113
Ν	-4.166715952047	-1.506714942350	1.866336640184
Η	-4.521771191679	-0.758047199475	1.259933640028
Η	-3.494661428430	-2.059061788301	1.318268207639
Pb	-2.092668108511	-1.434297300985	6.976427497864

# YFF molecule + Cd<sup>2+</sup>

С	-3.573537000000	-1.026254000000	3.254802000000
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0	-2.318251000000	-2.413726000000	4.856017000000
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Η	-1.427401000000	-0.902669000000	2.775011000000
Η	-2.418176000000	0.479873000000	2.236165000000
С	-1.842841000000	0.577959000000	4.302071000000
С	-2.538642000000	1.751867000000	4.672025000000
С	-0.803266000000	0.150539000000	5.169111000000
С	-2.284062000000	2.433544000000	5.876116000000
С	-0.555081000000	0.797192000000	6.419022000000
С	-1.307802000000	1.954136000000	6.781228000000
Η	-3.305806000000	2.160967000000	3.994499000000

тт	0 10010500000	0.44510400000	4.04070000000
Н	-0.133195000000	-0.665196000000	4.849799000000
Н	-2.826822000000	3.36640500000	6.100796000000
H	0.319839000000	0.52814000000	7.032817000000
0	-1.015584000000	2.52037800000	7.960792000000
Н	-1.483237000000	3.373122000000	8.076277000000
Ν	-4.356274000000	-3.178711000000	4.209358000000
С	-5.841327000000	-2.926903000000	4.190542000000
Ν	-6.305575000000	-2.838341000000	5.568682000000
С	-5.444166000000	-2.145540000000	6.485363000000
С	-5.437875000000	-0.640580000000	6.205663000000
Ο	-4.190838000000	-0.083195000000	6.587683000000
Ο	-6.281786000000	0.047562000000	5.708044000000
Η	-4.197322000000	0.881416000000	6.370047000000
С	-6.570733000000	-4.057819000000	3.445546000000
С	-4.978723000000	-7.366509000000	4.556729000000
С	-5.888339000000	-7.804109000000	5.534681000000
С	-5.216572000000	-6.171761000000	3.856096000000
Η	-5.709492000000	-8.747476000000	6.071723000000
Η	-4.52202000000	-5.859917000000	3.058876000000
С	-7.043948000000	-7.050877000000	5.799332000000
С	-6.367130000000	-5.395164000000	4.124282000000
Η	-7.777509000000	-7.406718000000	6.538427000000
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Η	-8.214431000000	-5.294381000000	5.280318000000
Η	-4.090855000000	-7.971965000000	4.320366000000
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С	-3.233026000000	-2.653444000000	8.527292000000
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С	-2.014824000000	-2.116671000000	9.051008000000
Η	-1.077931000000	-2.693558000000	8.956350000000
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С	-2.051801000000	-0.948161000000	9.860916000000
Η	-5.440874000000	-0.394866000000	9.854077000000
Η	-1.120295000000	-0.537740000000	10.277423000000
С	-3.289495000000	-0.368223000000	10.166320000000
Η	-3.332915000000	0.507016000000	10.831613000000
Η	-3.208203000000	-3.618272000000	7.991821000000
Η	-5.984155000000	-1.972842000000	3.650160000000
Η	-5.850110000000	-3.538586000000	8.083628000000
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Η	-6.189970000000	-4.063950000000	2.406005000000
Η	-7.644922000000	-3.789505000000	3.406001000000
Η	-4.103087000000	-4.034995000000	4.728787000000

Η	-4.384081000000	-2.457533000000	6.342888000000
Η	-6.582670000000	-3.765515000000	5.924356000000
Ν	-4.237733000000	-1.441998000000	2.037001000000
Η	-4.551045000000	-0.642022000000	1.475485000000
Η	-3.636508000000	-2.029653000000	1.445587000000
Cd	-2.115386000000	-1.213210000000	6.734576000000

# YFF molecule + Hg<sup>2+</sup>

С	-3.556596711357	-1.090485035099	3.145569686626
Η	-4.337980479704	-0.481009941811	3.653055699670
С	-3.324590759228	-2.350375169523	4.012209300142
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С	-2.247041245780	-0.219331843237	3.102781238395
Η	-1.389260738837	-0.876650058772	2.863866684718
Η	-2.381821092188	0.487918697985	2.260812639056
С	-1.972807750582	0.585283874534	4.343151966866
С	-2.670001189821	1.784605347738	4.596916184810
С	-0.946692291101	0.221989578375	5.294076558983
С	-2.359788934198	2.615328282793	5.686019538021
С	-0.597268023400	1.096424901444	6.384483116650
С	-1.317346071008	2.289479533140	6.595439689049
Η	-3.449404611500	2.115581898879	3.892342126201
Η	-0.220405976797	-0.555754359071	4.995744280892
Η	-2.906695550831	3.565064462153	5.810666385984
Η	0.262751181493	0.868175314255	7.032622309344
Ο	-0.958369564131	3.054615419102	7.636091752795
Η	-1.423941610746	3.915481063260	7.631376351145
Ν	-4.335314904453	-3.235016620107	4.112681582491
С	-5.800172289388	-2.931425722297	4.104860943676
Ν	-6.251974579676	-2.738599030996	5.475947052515
С	-5.390819314799	-1.987669835041	6.348791554097
С	-5.450083418779	-0.484516051190	6.056380532830
0	-4.507892279037	0.188366424863	6.814363941169
0	-6.123912125238	0.072855023792	5.226048705823
Η	-4.549652357381	1.136849809983	6.548975554674
С	-6.580058459514	-4.081930106344	3.442046827607
С	-5.050559212705	-7.376201427402	4.678012175726
С	-5.943456164863	-7.736963857494	5.701662900471
С	-5.275485055850	-6.212005521596	3.924426172163
Η	-5.777121759759	-8.657974915939	6.279944632197
Η	-4.598225216747	-5.962077368930	3.091535127073
С	-7.070116624215	-6.937020723265	5.959120748211

С	-6.395380960306	-5.387457458736	4.185065598496
Η	-7.791566069816	-7.234159845495	6.735143193884
С	-7.294834182415	-5.771267290869	5.206371958026
Η	-8.205114377809	-5.174577695878	5.383668450390
Η	-4.187705031442	-8.019233697050	4.447944851825
С	-5.575976893184	-2.322637203833	7.852103674980
С	-3.086394381335	-2.643400622448	8.388232606918
С	-4.355337878356	-2.016802612669	8.689731119133
С	-1.949297011535	-2.436140100630	9.256150777328
Η	-1.025723912523	-3.009614941747	9.075625008118
С	-4.412219994848	-1.160732229721	9.795690044745
С	-2.046684856990	-1.559173882279	10.342094882603
Η	-5.364904586529	-0.673274179507	10.052043083213
Η	-1.182384531146	-1.393858237530	11.001387780210
С	-3.279333582964	-0.933519172341	10.608175264454
Η	-3.370602145407	-0.265530656121	11.478678403194
Η	-3.077331893374	-3.534852346089	7.733229633889
Η	-5.931603481463	-2.009527674453	3.510272592154
Η	-5.767429137036	-3.416783926864	7.911169362211
Η	-6.470213148715	-1.817095781610	8.264413641214
Η	-6.226787676705	-4.159594460221	2.395491292962
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Η	-6.530014852987	-3.638768311314	5.891851775074
Ν	-4.102153096525	-1.493626172016	1.875520725106
Η	-4.491703515098	-0.722345924090	1.323250687195
Η	-3.457506565171	-2.045447338420	1.297654760612
Hg	-2.115400184192	-1.243306881380	6.723168934174

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