

Supporting Information

Biomass-derived substrates hydrogenation over Rhodium Nanoparticles Supported on Functionalized Mesoporous Silica

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A. Infrared spectra

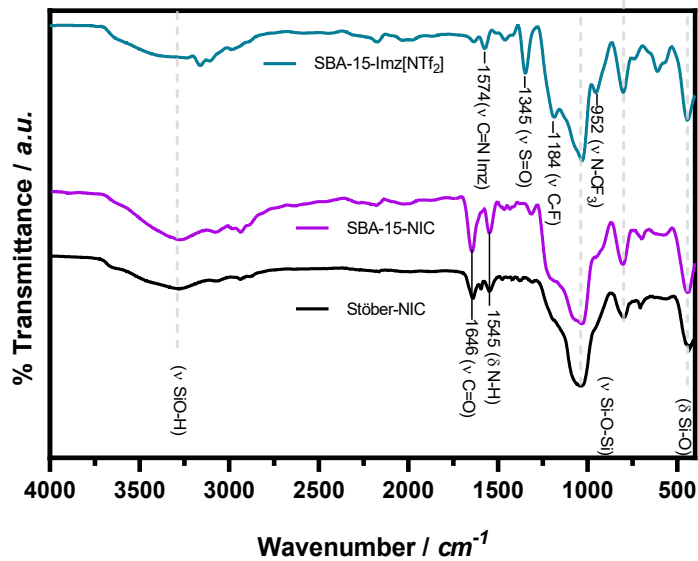


Figure S1. FTIR (ATR) spectra of functionalized SBA-15 based materials and Stöber silica.

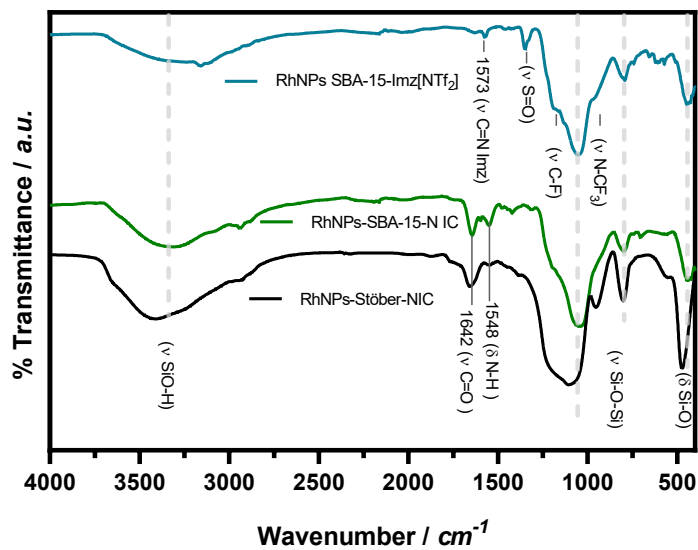


Figure S2. FTIR (ATR) of RhNPs supported on functionalized SBA-15 and Stöber silica

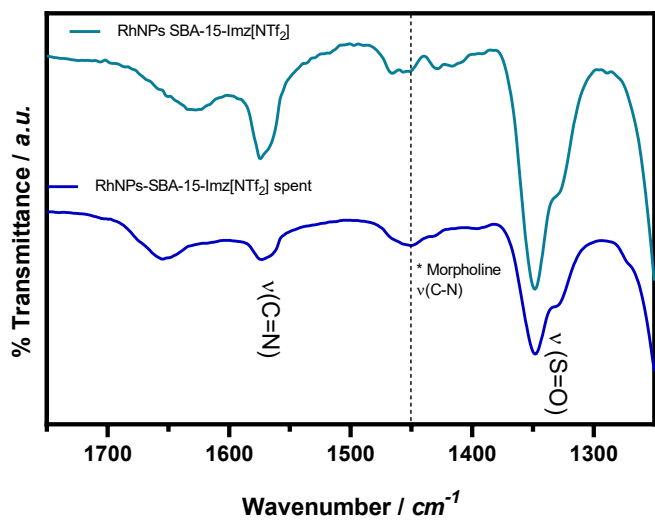


Figure S3. FTIR (ATR) of RhNPs@SBA-15-Imz [NTf₂] before and after phenol reductive amination in 1750-1250 cm^{-1} region.

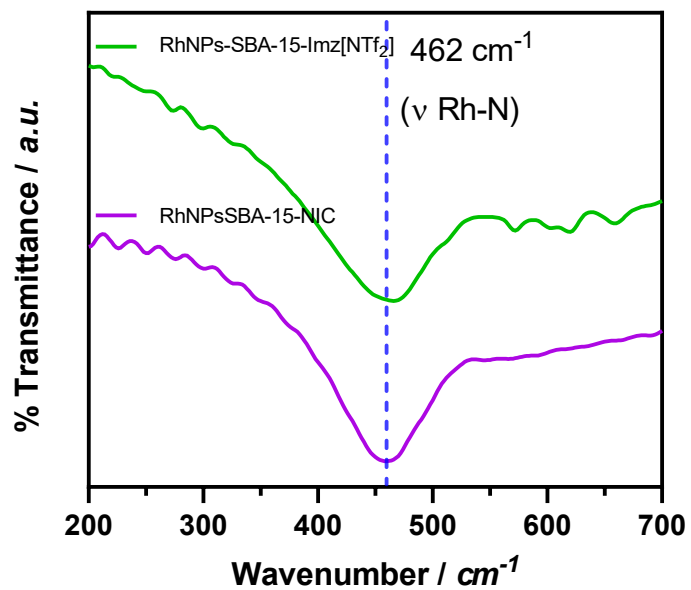


Figure S4. Far infrared spectra of RhNPs supported on functionalized SBA-15

B. TGA thermograms

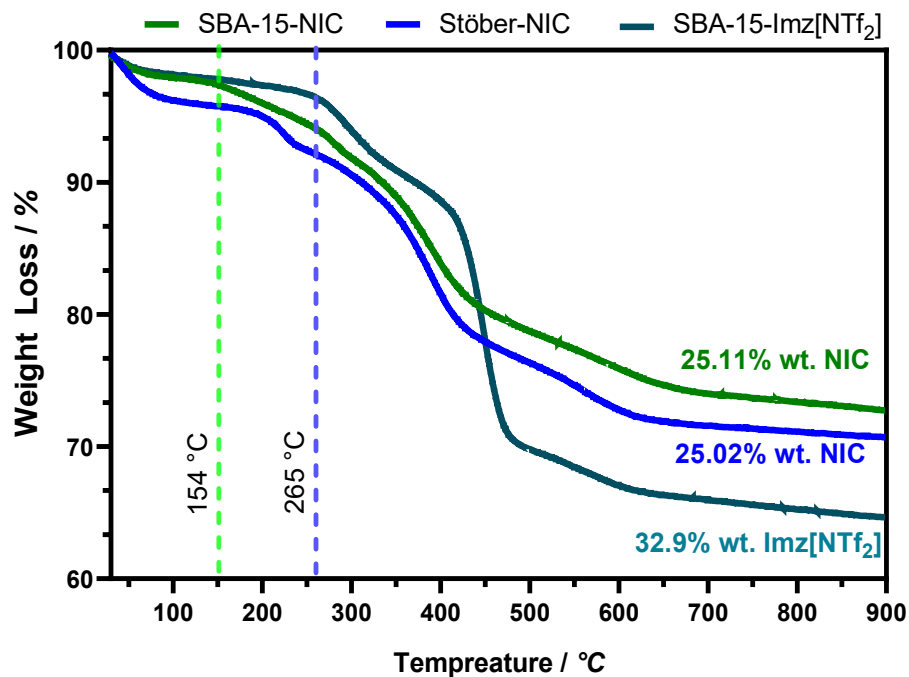


Figure S5. TGA curve of functionalized SBA-15 and Stöber silica

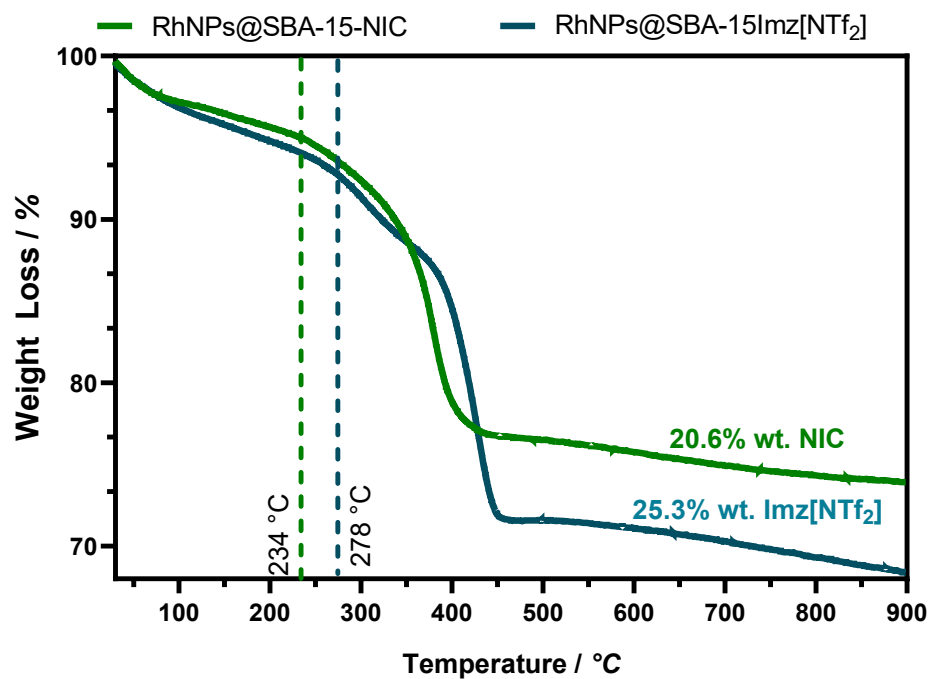
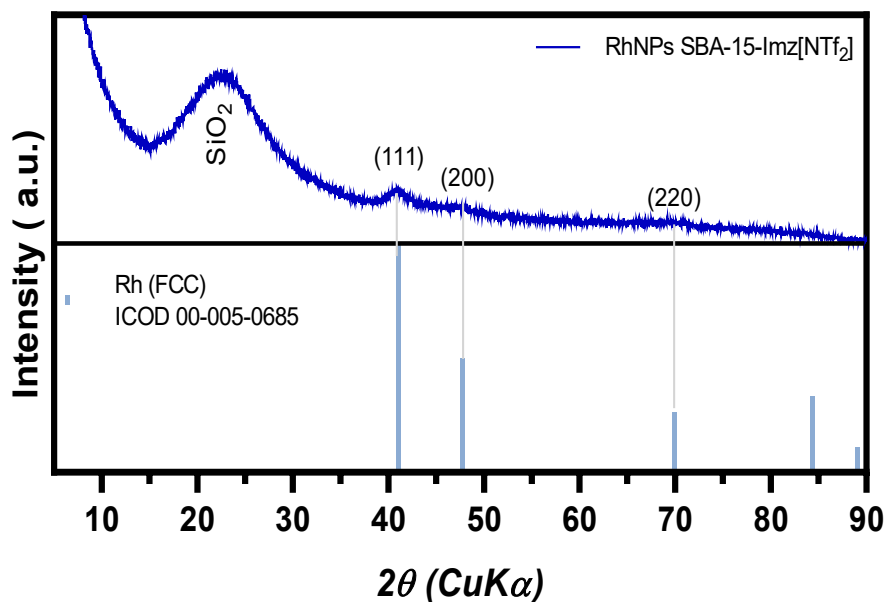


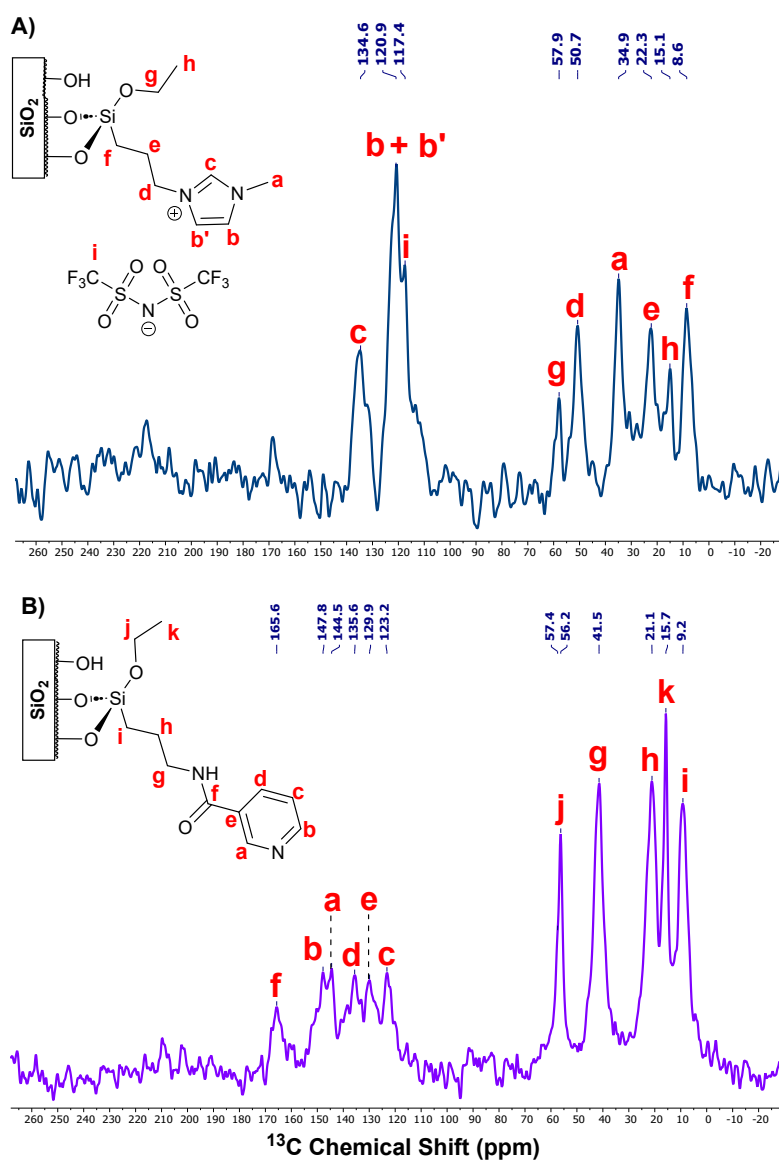
Figure S6. TGA curve of RhNPs supported in functionalized SBA-15.

C. PXRD data

Figure S7. PXRD of RhNPs@SBA-15-Imz[NTf₂].D. ²⁹Si NMR deconvolution dataTable S1. Relative concentration of Tⁿ and Qⁿ sites obtained from ²⁹Si UDEFT spectra.

Material	²⁹ Si-UDEFT-MAS ^a						φ ratio	% of silane Grafting ^b	% of Silanols (Si-OH) ^b	Grafting density (mmol g ⁻¹) ^c	BET Surface area (m ² g ⁻¹)	Surface coverage (molecules nm ⁻²)
	T ¹	T ²	T ³	Q ²	Q ³	Q ⁴						
SBA-15-Imz [NTf ₂]	0.5	5.2	5.7	6.6	16.9	65.2	0.49	11.4	23.5	1.1	279	2.4
SBA-15-NIC	1.4	7.6	3.9	1.8	22.2	63.1	0.53	12.9	24.0	2.1	328	3.9

a) Percentage Area after deconvolution of ²⁹Si NMR spectrum. b) Determined based on the relative concentrations of Tⁿ and Qⁿ. c) Determined by E.A and TGA.

E. ^{13}C CP-MAS SpectraFigure S8. Solid ^{13}C CP-MAS NMR spectrum of SBA-15-Imz[NTf₂] (A), and SBA-15-NIC (B)

F. Nitrogen sorption/desorption isotherms at 77 K of MMS

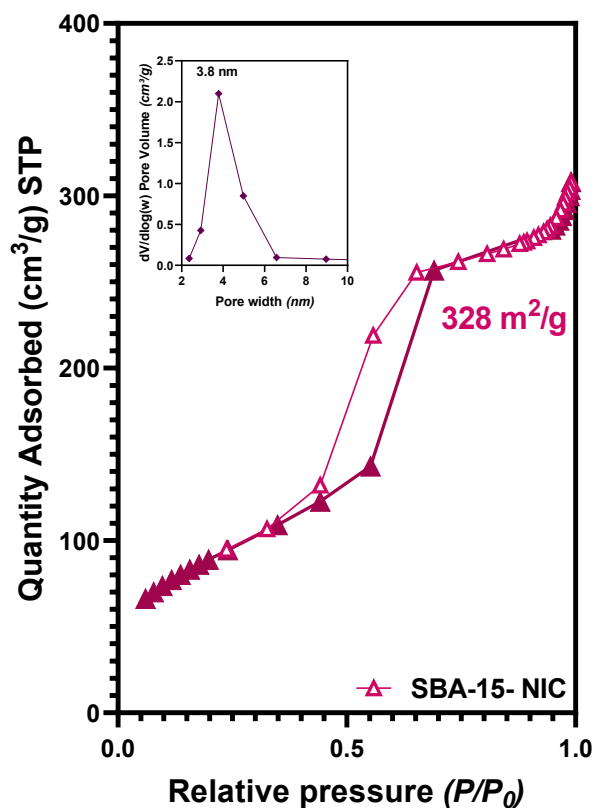


Figure S9. Adsorption-desorption isotherms of functionalized SBA-15 with pore size.

Table S2 BET isotherms analysis of functionalized silicas.

Entry	Material	S_{BET} ($\text{m}^2 \text{g}^{-1}$) ^a	Pore width ^b (nm)
1	SBA-15	902	5.7
2	SBA-15 Imz[NTf₂]	279	5.4
3	SBA-15 NIC	328	3.8
4	RhNPs@ SBA-15 Imz[NTf₂]	211	5.4

^a Calculated using Brunauer–Emmett–Teller (BET) model on the adsorption branch in the range of relative pressure (P/P₀) from 0.06 to 0.196.

^b Calculated using the BJH model for cylindrical pores

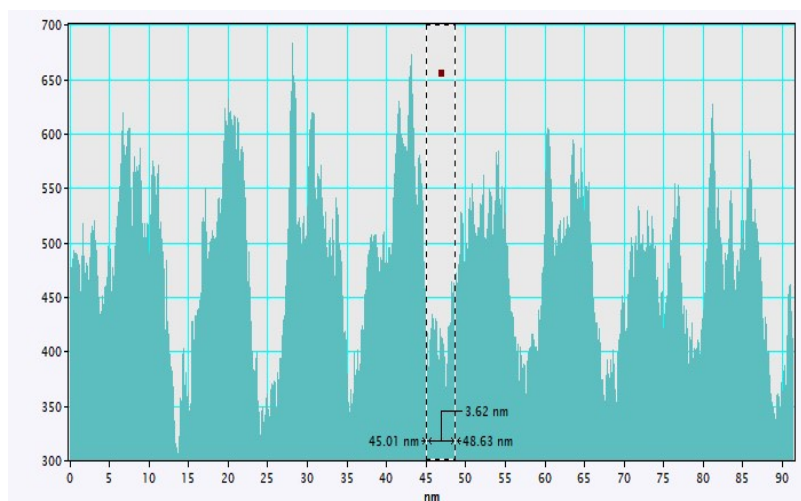
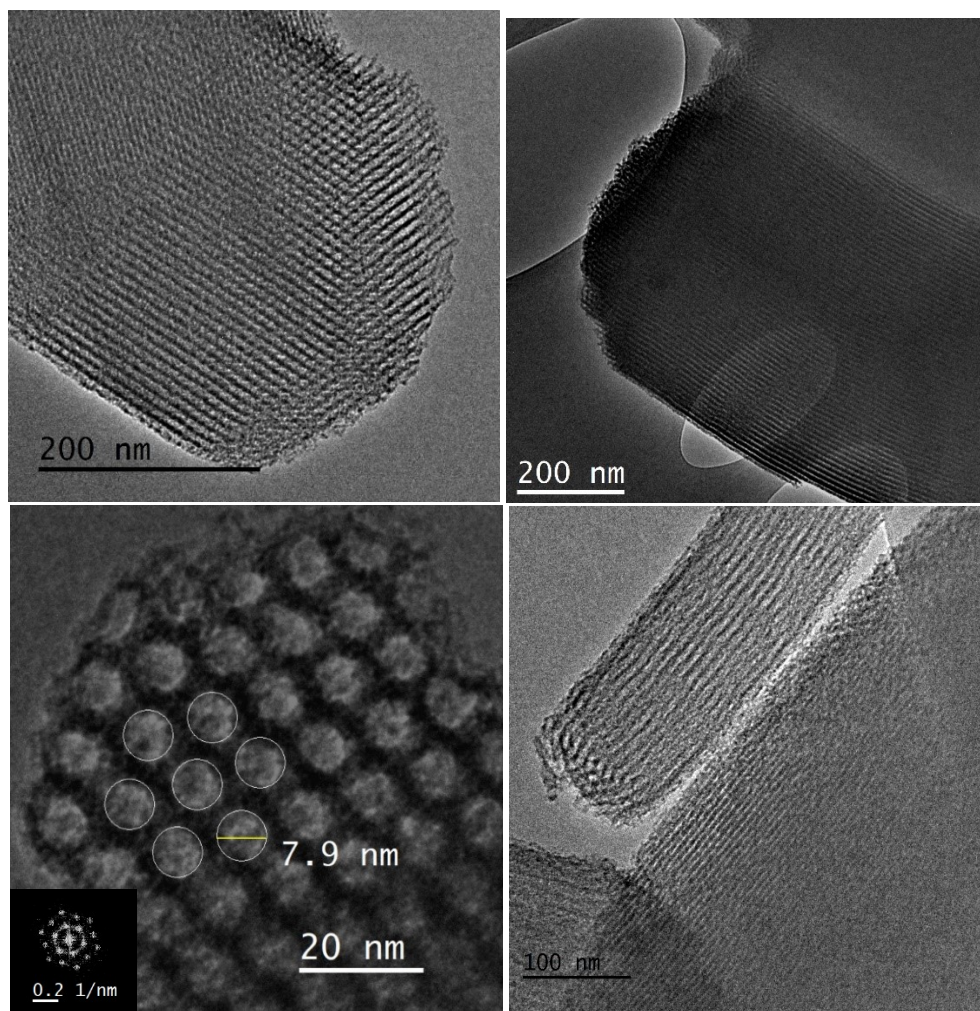
G. TEM and SEM micrographs with EDX analysis of pristine SBA-15

Figure S10. TEM micrographs of pristine calcinated SBA-15 where it showed a ca. 7.9 nm of pore diameter and 3.6 nm of walls width.

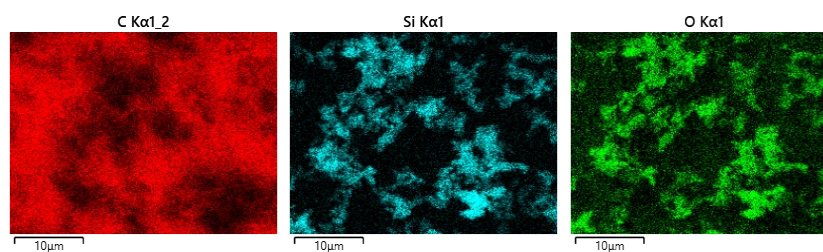
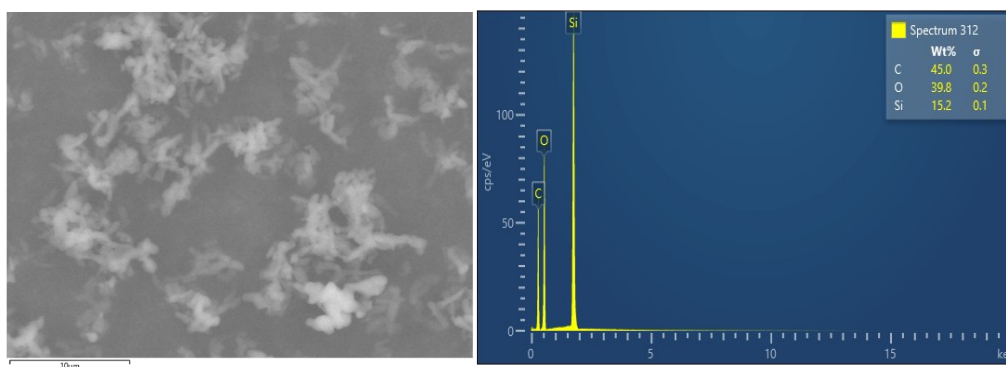
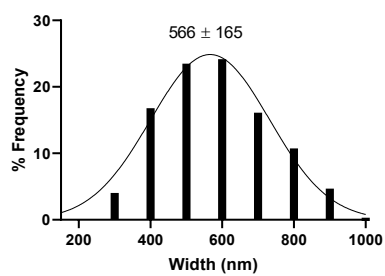
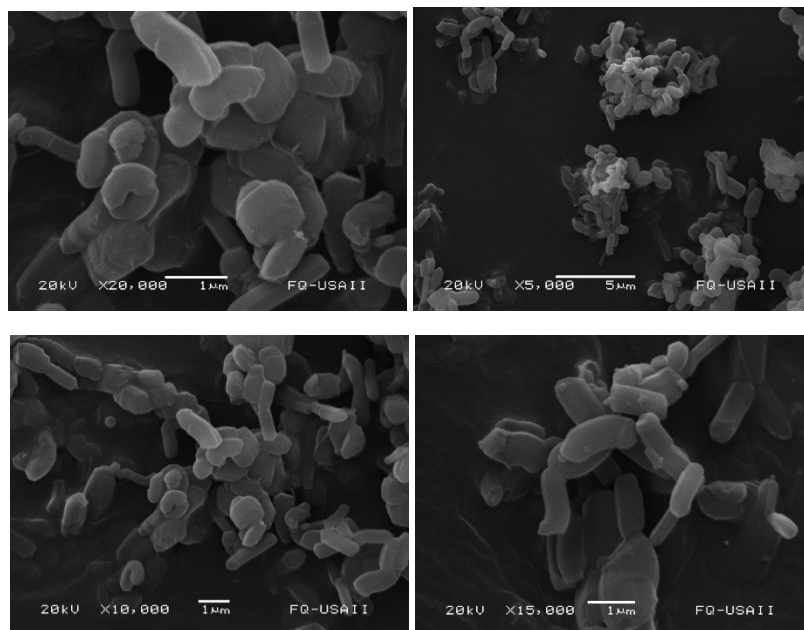


Figure S11. SEM micrographs of pristine calcinated SBA-15 with particles size distribution (566 ± 165) with EDX and element mapping.

H. TEM micrographs with EDX analysis of RhNPs@MMS

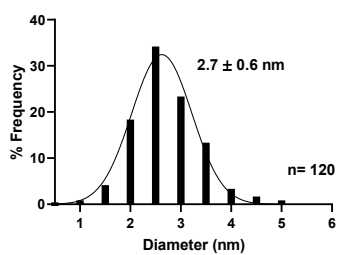
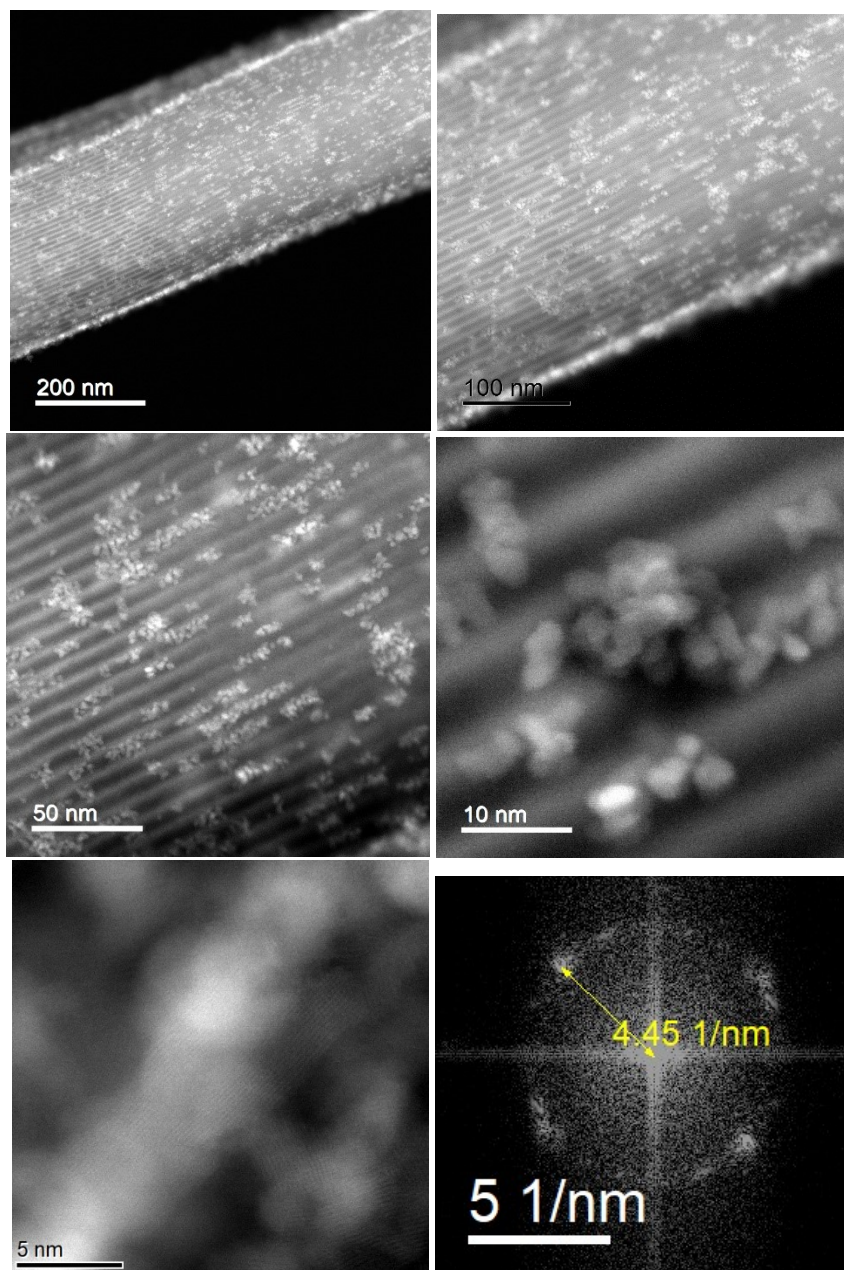


Figure S12. HAADF-STEM images of RhNPs@SBA-15-Imz[NTf₂] with FFT. Mean diameter: 2.7 ± 0.6 nm.

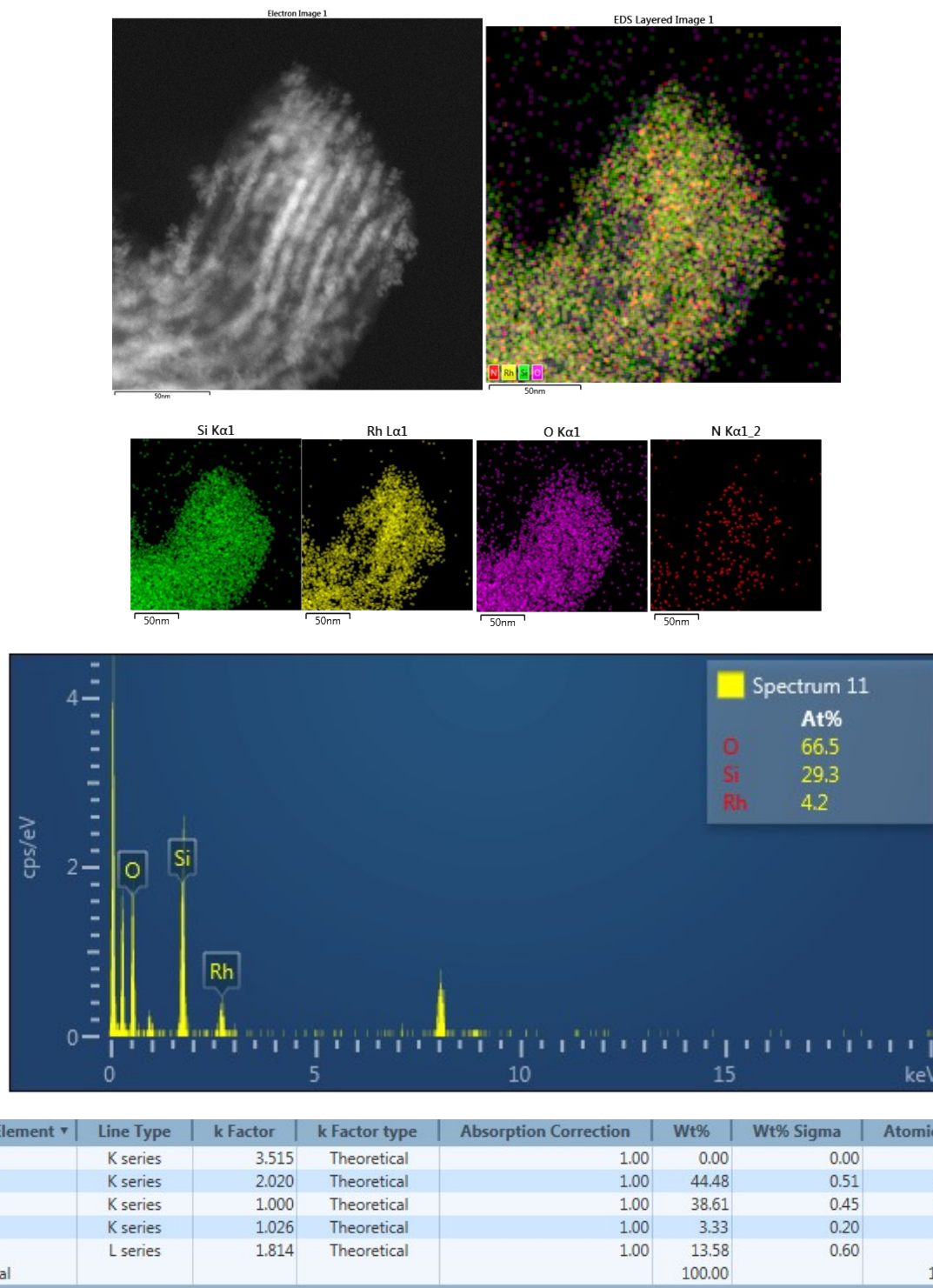


Figure S13. Element mapping and EDX quantification of RhNPs@SBA-15-Imz[NTf₂]

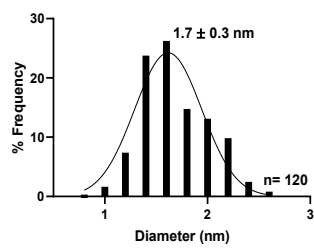
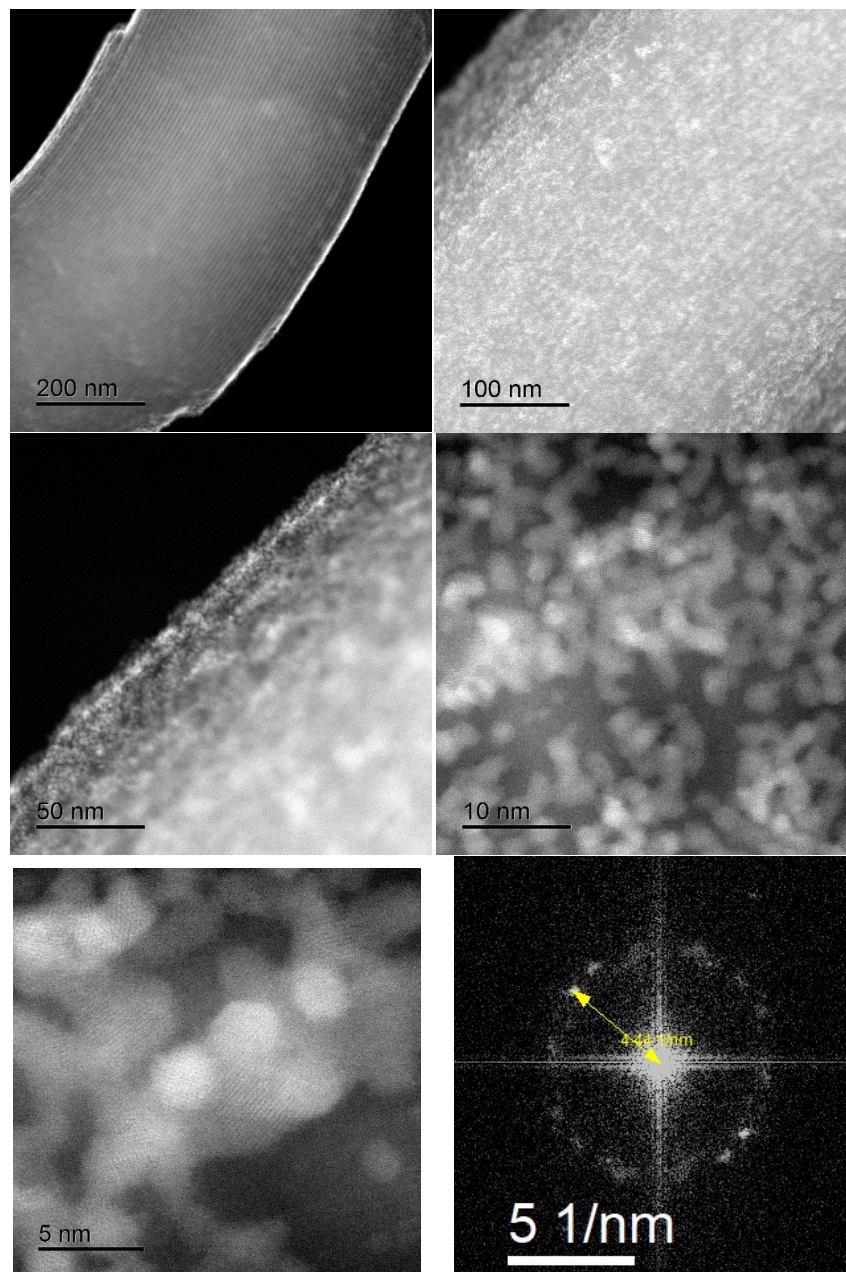


Figure S14. HAADF-STEM images of RhNPs@SBA-15NIC with FFT. Mean diameter: 1.7 ± 0.3 nm.

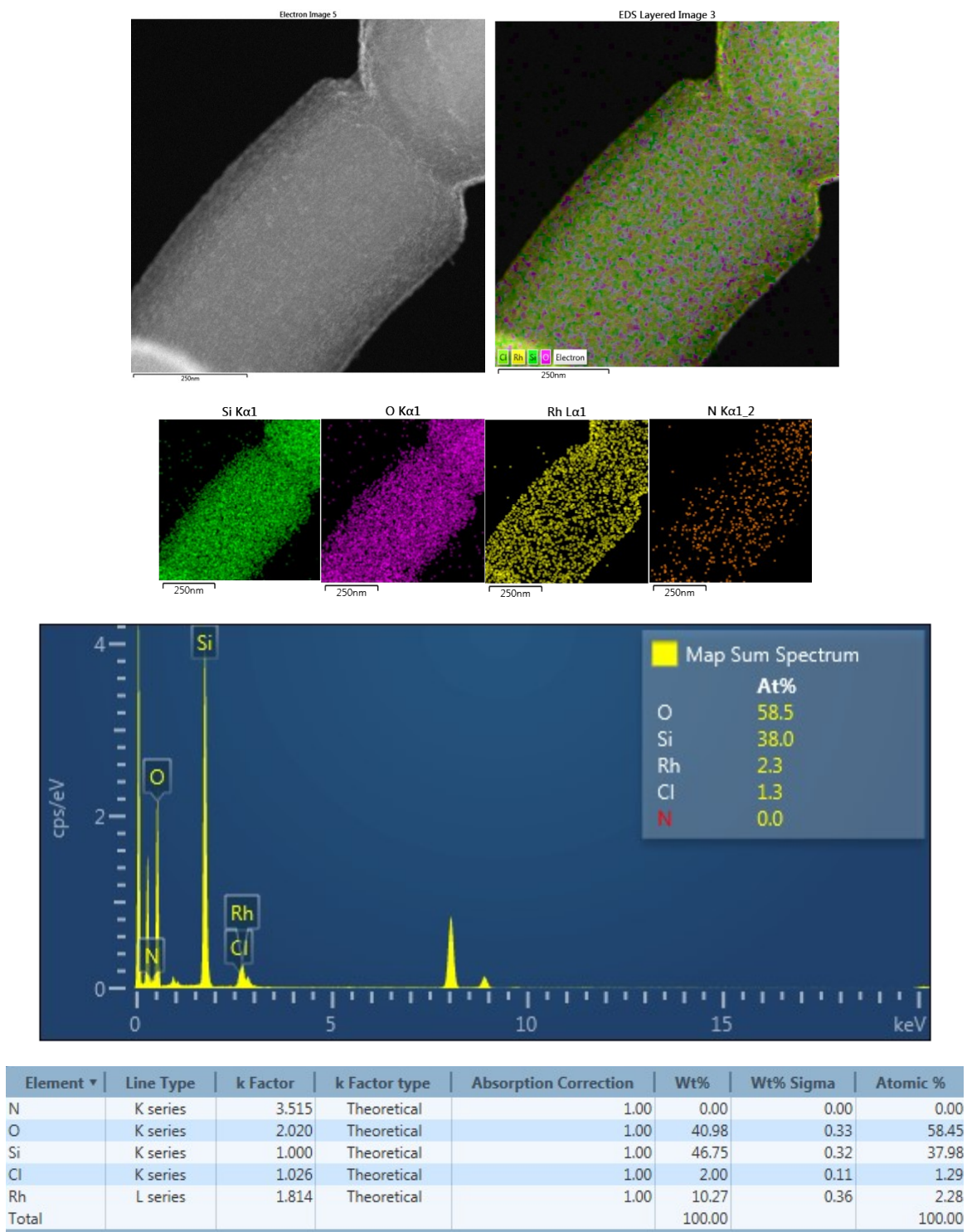


Figure S15. Element mapping and EDX quantification of RhNPs@SBA-15-NIC

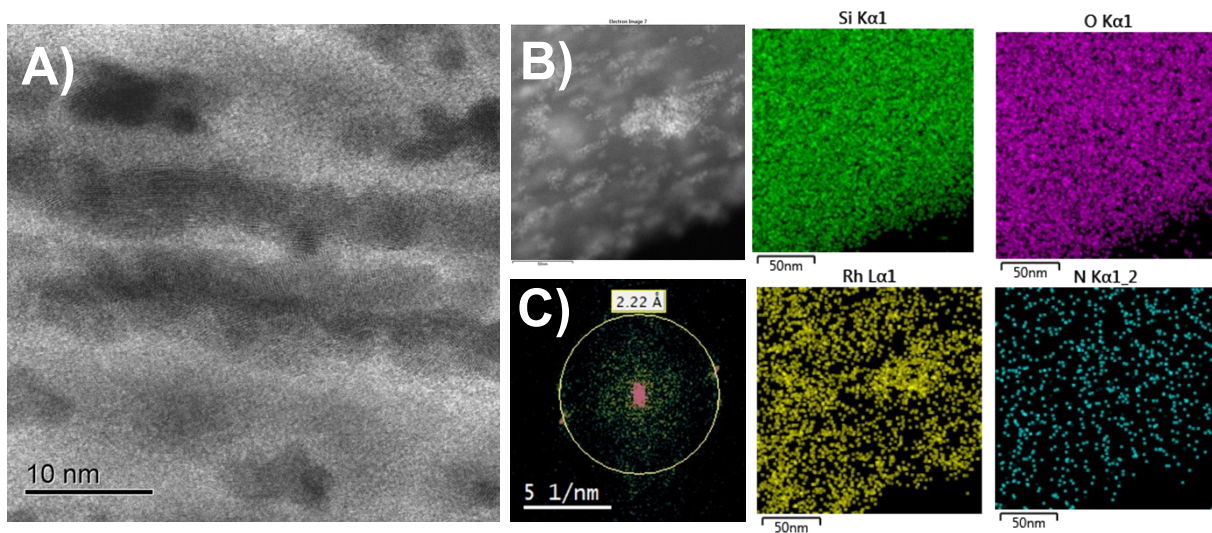


Figure S16. STEM image of RhNPs in spent catalyst (A) with EDX elemental mapping (B) and FFT of RhNPs (C).

I. XPS spectra of RhNPs@MMS

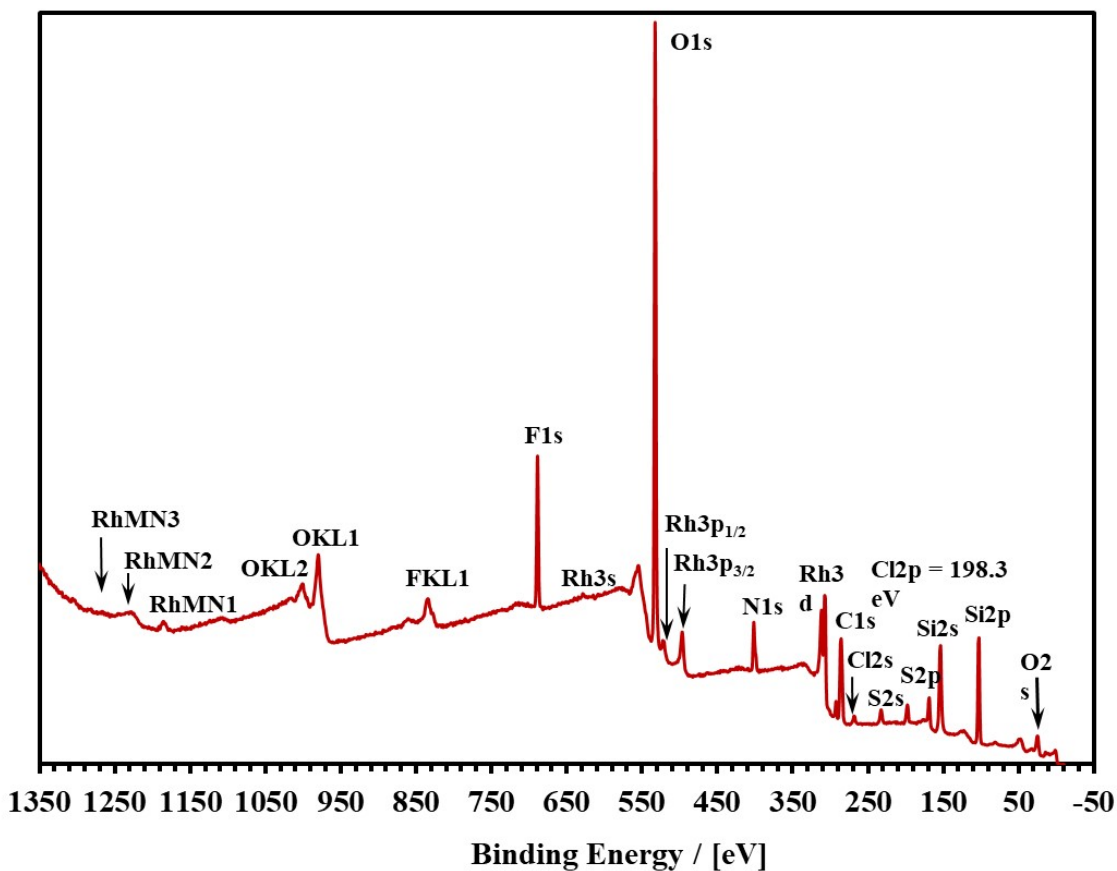


Figure S17. XPS survey of RhNPs@SBA-15Imz[NTf2]

Table S3 Element distribution on surface of RhNPs@SBA-15Imz[NTf2] by XPS

Name	Peak BE	Atomic %	Wt. %
O1s	532.95	40.07	31.8
Rh3d	307.93	2.24	11.4
F1s	689.03	8.26	7.8
Si2p	103.38	18.56	25.9
C1s	286.13	20.86	12.4
N1s	401.82	6.17	4.3
S2p	169.23	2.59	4.1
Cl2p	198.3	1.25	2.2

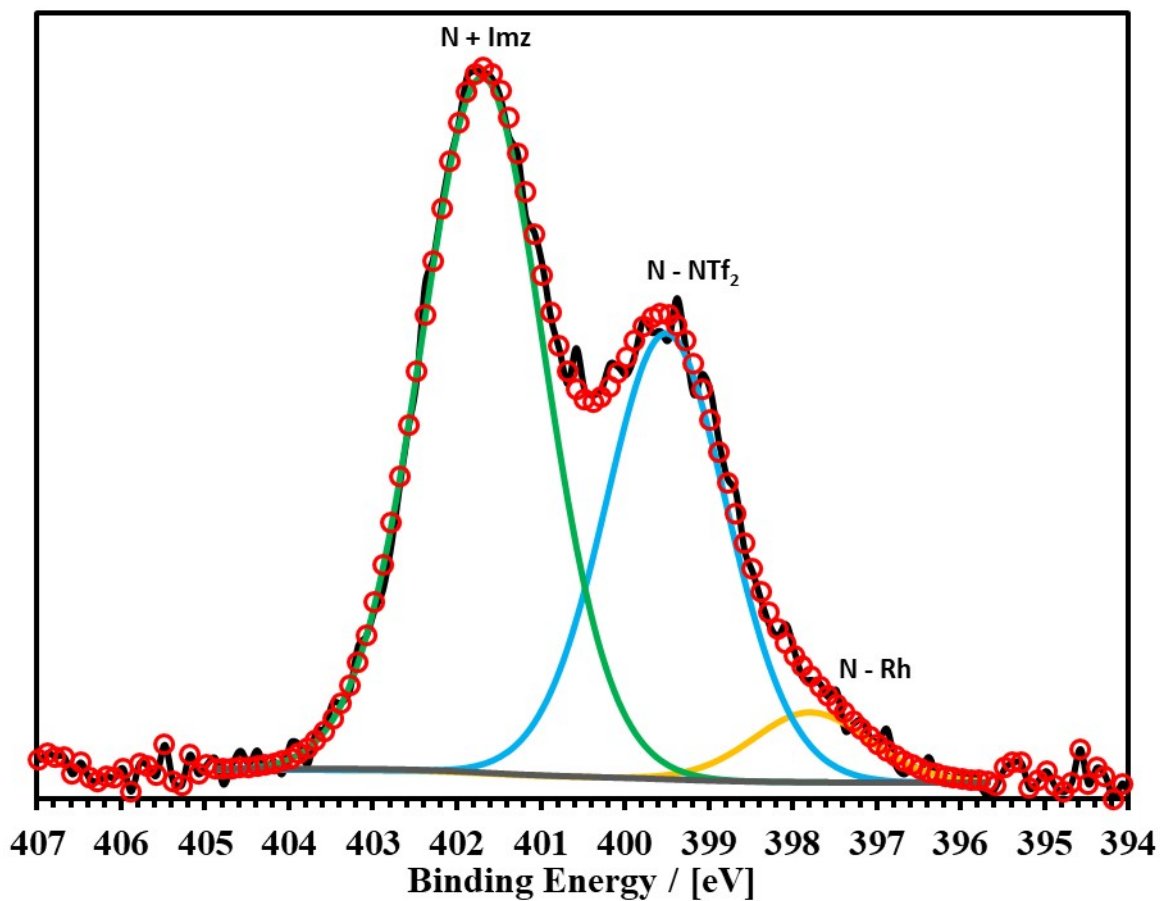


Figure S18. High resolution -XPS of N 1s in RhNPs@SBA-15Imz[NTf₂]

Table S4 Nitrogen species distribution on surface of RhNPs@SBA-15Imz[NTf₂] by XPS

Name	Peak BE	Atomic %
N [*] -Rh	397.83	5.42
N ⁻ (NTf ₂)	399.09	36.65
N ⁺ (Imz)	401.28	57.93

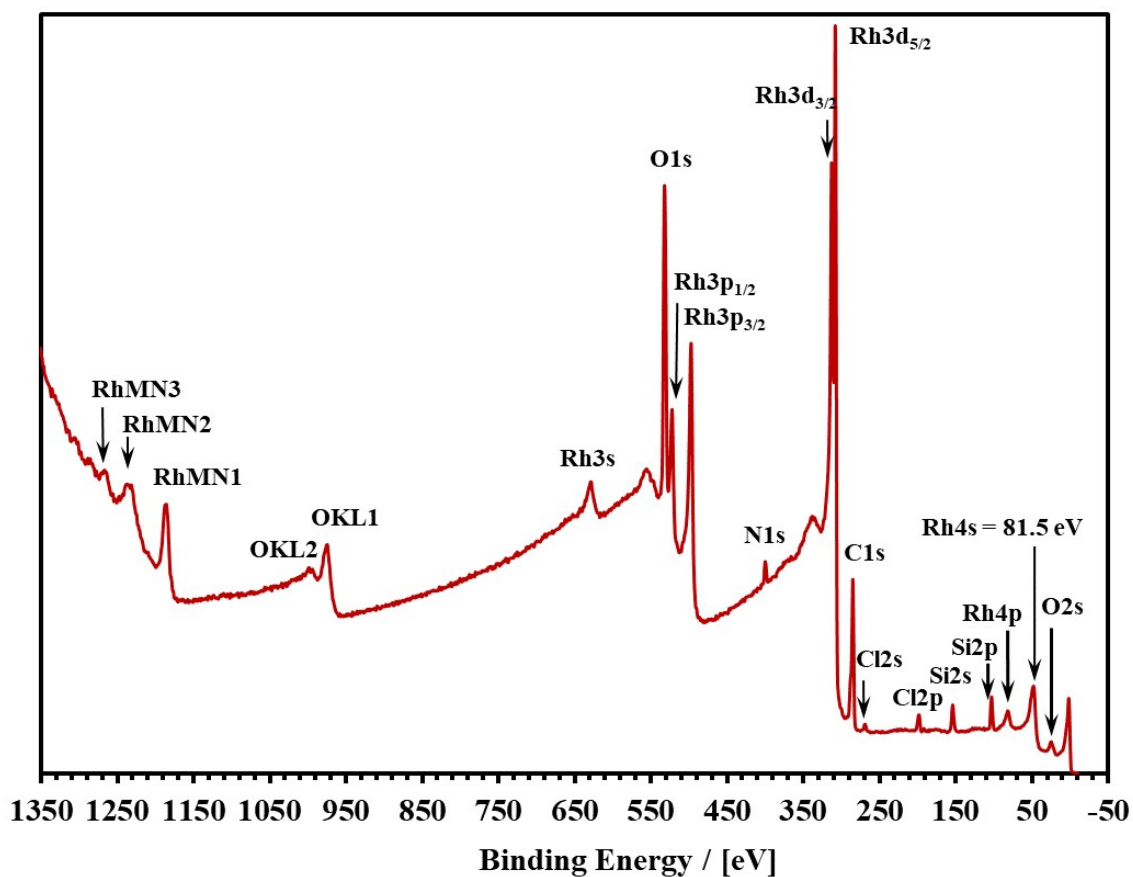


Figure S19. XPS survey of RhNPs@SBA-15NIC

Table S5 Element distribution on surface of RhNPs@SBA-15NIC by XPS

Name	Peak BE	Atomic %	Wt. %
Rh3d	308.14	14.99	53.78
O1s	531.81	36.12	20.15
C1s	285.2	36.18	15.15
Si2p	103.17	6.87	6.73
N1s	399.91	4.03	1.97
Cl2p	198.56	1.81	2.24

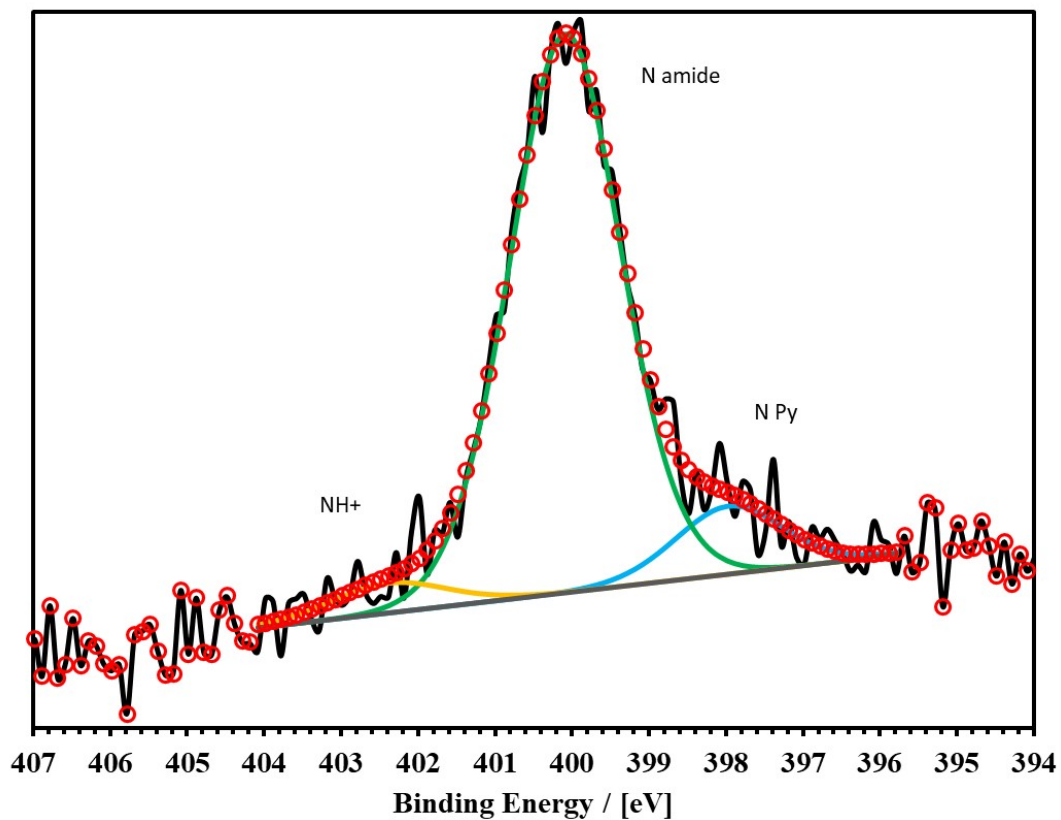


Figure S20. High resolution -XPS of N 1s in RhNPs@SBA-15NIC

Table S6 Nitrogen species distribution on surface of RhNPs@SBA-15NIC by XPS

Name	Peak BE	Atomic %
N Pyridine	397.96	10.1
N amide	400.09	85.21
NH +	402.39	4.69

J. Calculation of specific surface area and metal Dispersion.

For the calculation of the dispersion (D) of the RhNPs, we assume nanoparticles as spheres; let NS be the total number of metal atoms present on the surface and NT the total number of metal atoms (surface and bulk). The metal dispersion D is given by

$$D = NS/NT \text{ (eq. 1)}$$

Taking in account the relationship between specific surface area (S_{sp}) and dispersion (D) is

$$S_{sp} = a_m (N_A/M)D \text{ (eq. 2)}$$

Where a_m is the surface area occupied by an atom (Rh) on a polycrystalline surface, N_A is Avogadro's number ($6.022 \times 10^{23} \text{ mol}^{-1}$) and M is the atomic mass of the metal.

And the definition of v_m (the volume occupied by an atom (Rh) in the bulk of metal) given by:

$$v_m = M/\rho N_A \text{ (eq. 3)}$$

where ρ the mass density

It possible to rewrite the D in terms of a_m , v_m and d_{NP} .^{1,2} and rewrites as follows:

$$D = [6(v_m/a_m)]/d_{NP} \text{ (eq. 4)}$$

Where d_{NP} is the mean diameter of nanoparticles.

For Rh (FCC) $a_m = 7.58 \text{ \AA}^2$, $v_m = 13.78 \text{ \AA}^3$, $\rho = 12.4 \text{ g cm}^{-3}$,

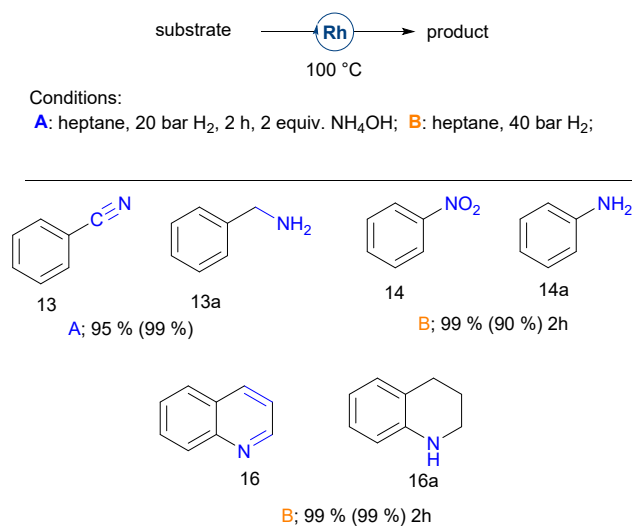
$$D(\text{RhNPs@SBA-15Imz[NTf}_2]) = [6*(13.78 \text{ \AA}^3/7.58 \text{ \AA}^2)]/ 27 \text{ \AA} = \mathbf{0.404 (40.4 \%)}$$

$$S_{sp}(\text{RhNPs@SBA-15Imz[NTf}_2]) = 7.58 \times 10^{-20} \text{ m}^2 (N_A/102.905) = 443.6 * D = \mathbf{179 \text{ m}^2 \text{ g}^{-1}}$$

$$D(\text{RhNPs@SBA-15NIC}) = [6*(13.78 \text{ \AA}^3/7.58 \text{ \AA}^2)]/ 17 \text{ \AA} = \mathbf{0.642 (64.2 \%)}$$

$$S_{sp}(\text{RhNPs@SBA-15Imz[NTf}_2]) = 7.58 \times 10^{-20} \text{ m}^2 (N_A/102.905 \text{ g mol}^{-1}) = 443.6 * D = \mathbf{285 \text{ m}^2 \text{ g}^{-1}}$$

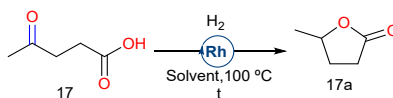
K. Selected catalytic Reactions.



Reaction conditions: 2 mmol of substrate, 5 mg RhNPs@SBA-15-NIC
 0.25 mol% , 100 °C. Conversion % (Selectivity %)

Figure S21. RhNPs@SBA-15NIC hydrogenation of different functional groups.

Table S7. Rh-catalyzed hydrogenation of Levulinic Acid.



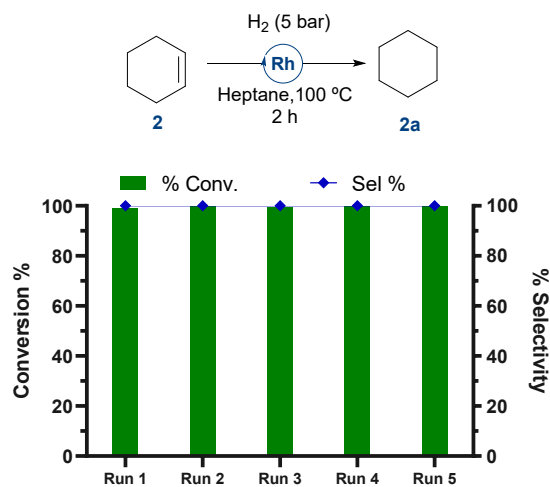
Entry	Catalyst	Solvent	Time (h)	H ₂ (bar)	Conv.% ^a	Yield 17a% ^a	TON ^b
1	RhNPs@SBA-15-NIC	Dioxane	2h	5	<1	n.d.	n.d.
2		2-MeTHF		5	n.d.	n.d.	
3		Heptane		5	n.d.	n.d.	
4		Heptane	4	40	<1	n.d.	n.d.
5		Neat		40	15	12	96
6	RhNPs@SBA-15-Imz[NTf ₂]	Dioxane	2h	5	<1	n.d.	n.d.
7		2-MeTHF		18	9	72	
8		Heptane		7	n.d.	n.d.	
9		Heptane	4	40	<1	n.d.	n.d.
10		Neat		40	42	42	336

Reaction conditions: Levulinic acid (203 μL, 2 mmol), 2 mL of solvent, 5 mg of catalyst, 100 °C.

Table S8. Rh-catalyzed hydrogenation of Furfural.

Entry	Catalyst	Solvent	Time (h)	H ₂ (bar)	Conv.% ^a	Yield 17a% ^a	TON ^b	
1	RhNPs@SBA-15-Imz[NTf ₂]	Heptane	4	5	<1	n.d.	n.d.	
2				40	<1	n.d.	n.d.	
3		EtOH	4	5	25	77	154	
4				40	30	57	137	
5		RhNPs@SBA-15-NIC	MeOH	2	40	30	65	156
6			Dioxane	4		19	44	67
7			Neat	6		32	78	200
8						47	62	233
9			Heptane	4		<1	n.d.	n.d.
10			MeOH	2		32	60	154
11		Dioxane	4	27	16	37		
12		Neat	4	44	56	197		

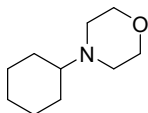
Reaction conditions: Furfural (166 μ L, 2 mmol), 2 mL of solvent, 5 mg of catalyst, 100 $^{\circ}$ C.



Scheme S1. Recycling experiments of RhNPs@SBA-15-Imz[NTf₂] on cyclohexene hydrogenation.

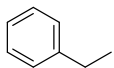
L. NMR Data of catalytic products

4-cyclohexylmorpholine: ³



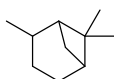
¹H NMR (600 MHz, CDCl₃) δ 3.79 – 3.71 (m, 4H), 2.67 – 2.55 (m, 4H), 2.29 – 2.17 (m, 1H), 1.96 – 1.74 (m, 4H), 1.37 – 1.13 (m, 6H). ¹³C NMR (151 MHz, CDCl₃) δ 66.90, 63.93, 49.44, 28.45, 26.09, 25.66.

Ethylbenzene:



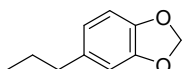
¹H NMR (600 MHz, CDCl₃) δ: 7.35 – 7.25 (m, 3H, Ar-H), 7.26 – 7.15 (m, 1H, Ar-H), 2.67 (q, *J* = 7.6 Hz, 2H, -CH₂-), 1.26 (t, *J* = 7.6 Hz, 3H, -CH₃).

Pinane: ⁴



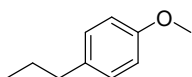
Mixture of Cis and Trans, ¹H NMR (600 MHz, CDCl₃) δ: 2.38 (q, *J* = 7.2 Hz, 2H), 2.20 (q, *J* = 8.1 Hz, 2H), 2.16 – 1.64 (m, 13H), 1.56 – 1.43 (m, 2H), 1.26 (d, *J* = 4.1 Hz, 9H), 1.08 (d, *J* = 12.0 Hz, 13H), 0.96 – 0.87 (m, 5H). ¹³C NMR (151 MHz, CDCl₃) δ: 48.24, 47.79, 41.51, 41.03, 39.56, 38.90, 36.11, 35.57, 34.08, 29.48, 28.38, 26.91, 26.65, 24.72, 24.07, 23.97, 23.27, 23.14, 22.96, 21.65, 20.13.

5-propylbenzo[d][1,3]dioxole:



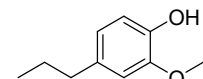
¹H NMR (600 MHz, CDCl₃) δ: 6.83 – 6.69 (m, 2H), 6.69 – 6.61 (m, 1H), 5.93 (d, *J* = 1.1 Hz, 2H), 2.54 (t, *J* = 7.7 Hz, 2H), 1.67 – 1.60 (m, 2H), 0.97 (td, *J* = 7.3, 2.6 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ: 147.59, 145.57, 136.65, 121.21, 109.00, 108.10, 100.77, 37.90, 24.93, 13.80.

1-methoxy-4-propylbenzene:



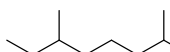
¹H NMR (600 MHz, CDCl₃) δ: 7.18 – 7.10 (m, 2H), 6.94 – 6.84 (m, 2H), 3.83 (s, 4H), 2.63 – 2.54 (m, 2H), 1.67 (h, *J* = 7.4 Hz, 2H), 0.99 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ: 157.78, 134.91, 129.42, 113.75, 55.31, 37.27, 24.92, 13.89.

2-methoxy-4-propylphenol:

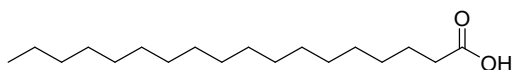


¹H NMR (600 MHz, CDCl₃) δ: 6.96 (d, *J* = 7.9 Hz, 1H), 6.82 – 6.74 (m, 2H), 5.91 (s, 1H), 3.90 (s, 3H), 2.65 – 2.60 (m, 2H), 1.73 (h, *J* = 7.4 Hz, 2H), 1.05 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ: 146.38, 143.57, 134.58, 120.93, 114.23, 111.10, 55.70, 37.70, 24.82, 13.71.

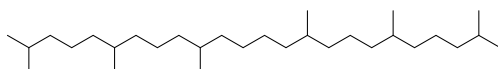
2,6-dimethyloctane:



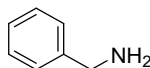
¹H NMR (600 MHz, CDCl₃) δ: 1.58 – 1.48 (m, 1H), 1.42 – 1.19 (m, 5H), 1.19 – 1.03 (m, 4H), 0.90 – 0.83 (m, 12H). ¹³C NMR (151 MHz, CDCl₃) δ: 39.57, 37.07, 34.62, 29.71, 28.17, 25.02, 22.88, 22.79, 19.39, 11.58.

Stearic acid:⁵

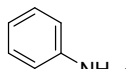
¹H NMR (600 MHz, CDCl₃) δ: 11.69 (s, 1H), 2.34 (t, *J* = 7.6 Hz, 2H), 1.63 (p, *J* = 7.5 Hz, 2H), 1.26 (s, 26H), 0.88 (t, *J* = 7.0 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ: 180.62, 34.26, 32.09, 29.86, 29.85, 29.83, 29.80, 29.75, 29.59, 29.52, 29.40, 29.22, 24.83, 22.85, 14.26.

Squalane:⁵

¹H NMR (600 MHz, CDCl₃) δ: 1.65 – 1.51 (m, 3H), 1.52 – 1.24 (m, 25H), 1.25 – 1.06 (m, 12H), 0.99-0.82 (m, 24H). ¹³C NMR (151 MHz, CDCl₃) δ: 39.73, 37.82, 37.79, 37.77, 37.75, 37.66, 33.13, 33.10, 28.30, 27.80, 27.79, 25.17, 25.16, 24.82, 23.04, 22.94, 20.09, 20.06, 20.02, 20.00.

Benzylamine:

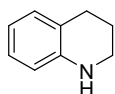
¹H NMR (600 MHz, CDCl₃) δ: 7.31 – 7.17 (m, 5H), 3.77 (s, 2H), 1.39 (s, 2H). ¹³C NMR (151 MHz, CDCl₃) δ: 143.07, 128.13, 126.70, 126.35, 46.10.

Aniline:

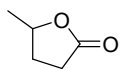
¹H NMR (600 MHz, CDCl₃) δ: 7.36 – 7.23 (m, 2H), 6.89 (t, *J* = 7.4 Hz, 1H), 6.82 – 6.68 (m, 2H), 3.68 (s, 2H). ¹³C NMR (151 MHz, CDCl₃) δ: 146.42, 129.17, 118.31, 114.99.

Methylcyclohexane:⁶

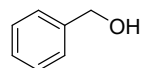
¹H NMR (600 MHz, CDCl₃) δ: 1.78 – 1.57 (m, 6H), 1.41 – 1.30 (m, 1H), 1.30 – 1.19 (m, 2H), 1.20 – 1.08 (m, 1H), 0.88 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ: 35.66, 32.97, 26.67, 26.55, 23.05.

1,2,3,4-tetrahydroquinoline:⁷

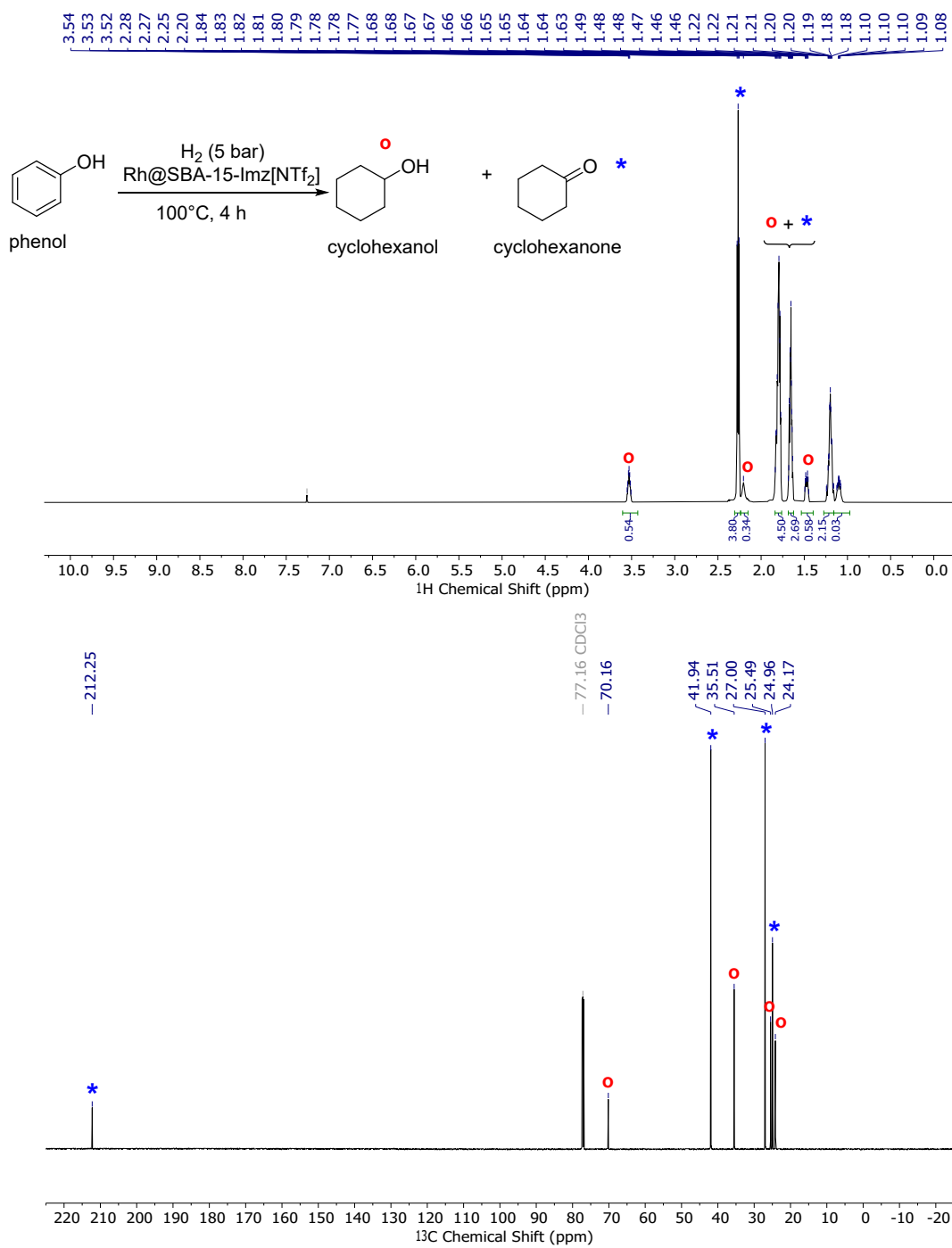
¹H NMR (600 MHz, CDCl₃) δ: 7.01 – 6.95 (m, 2H), 6.68 – 6.59 (m, 1H), 6.53 – 6.45 (m, 1H), 3.35 – 3.26 (m, 3H), 2.79 (t, *J* = 6.4 Hz, 2H), 2.00 – 1.92 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ: 144.88, 129.61, 126.82, 121.56, 117.05, 114.30, 42.08, 27.07, 22.28.

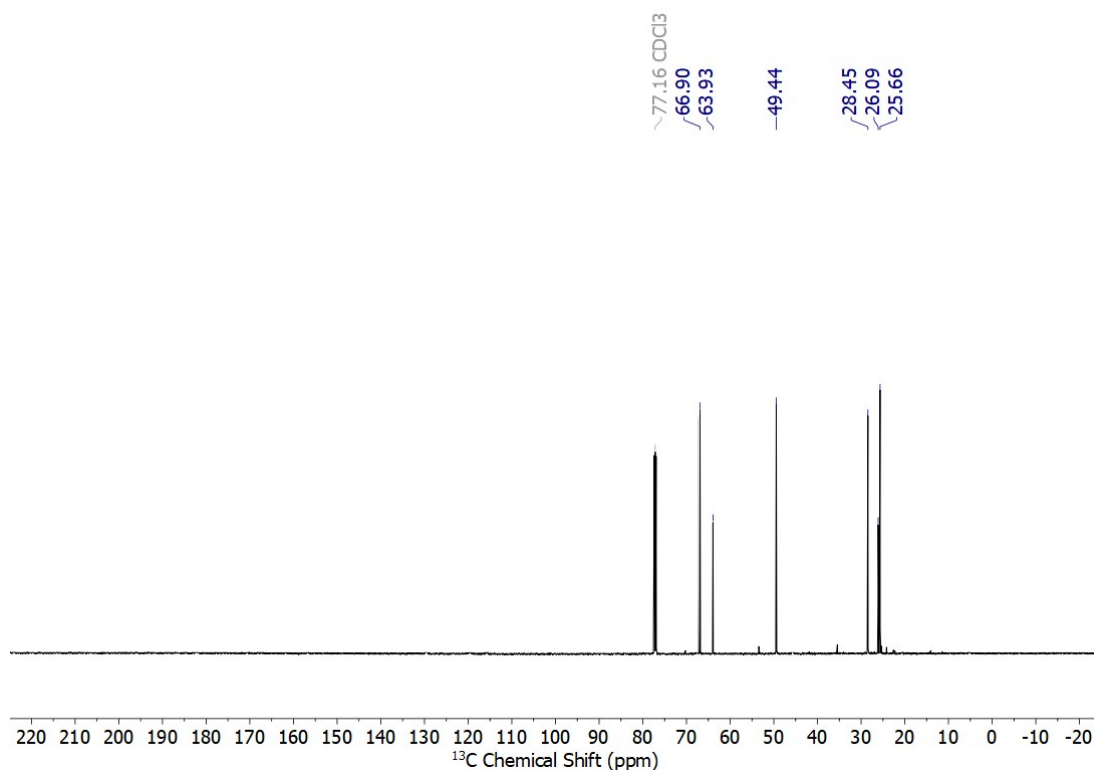
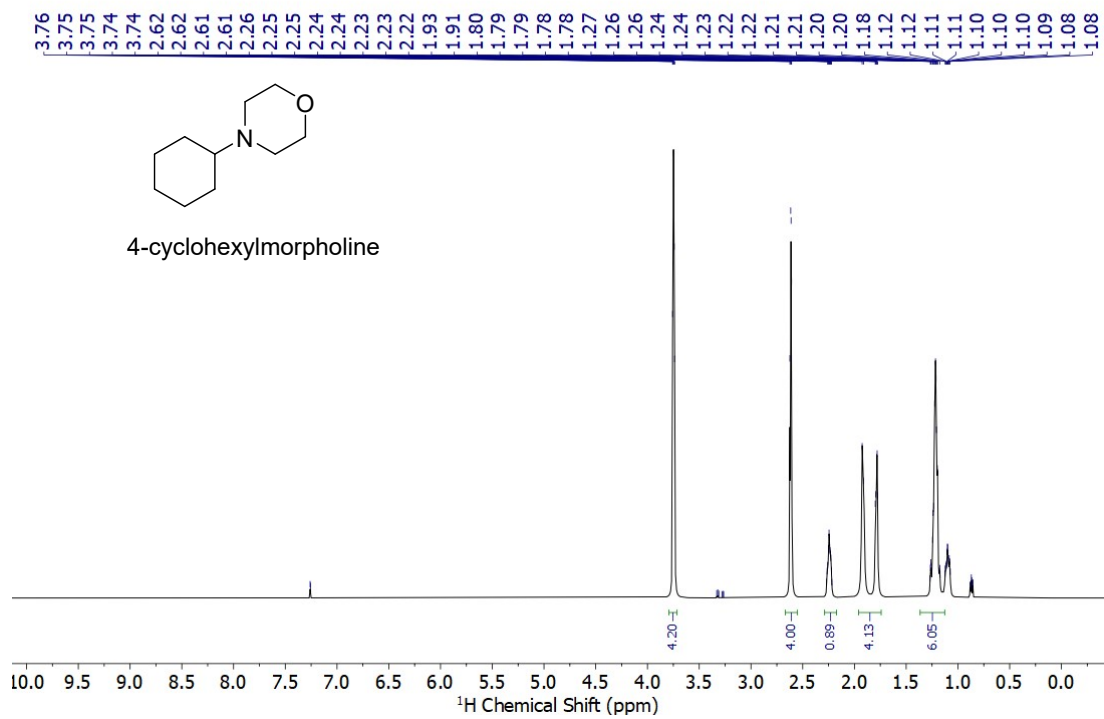
Gamma-valerolactone:⁵

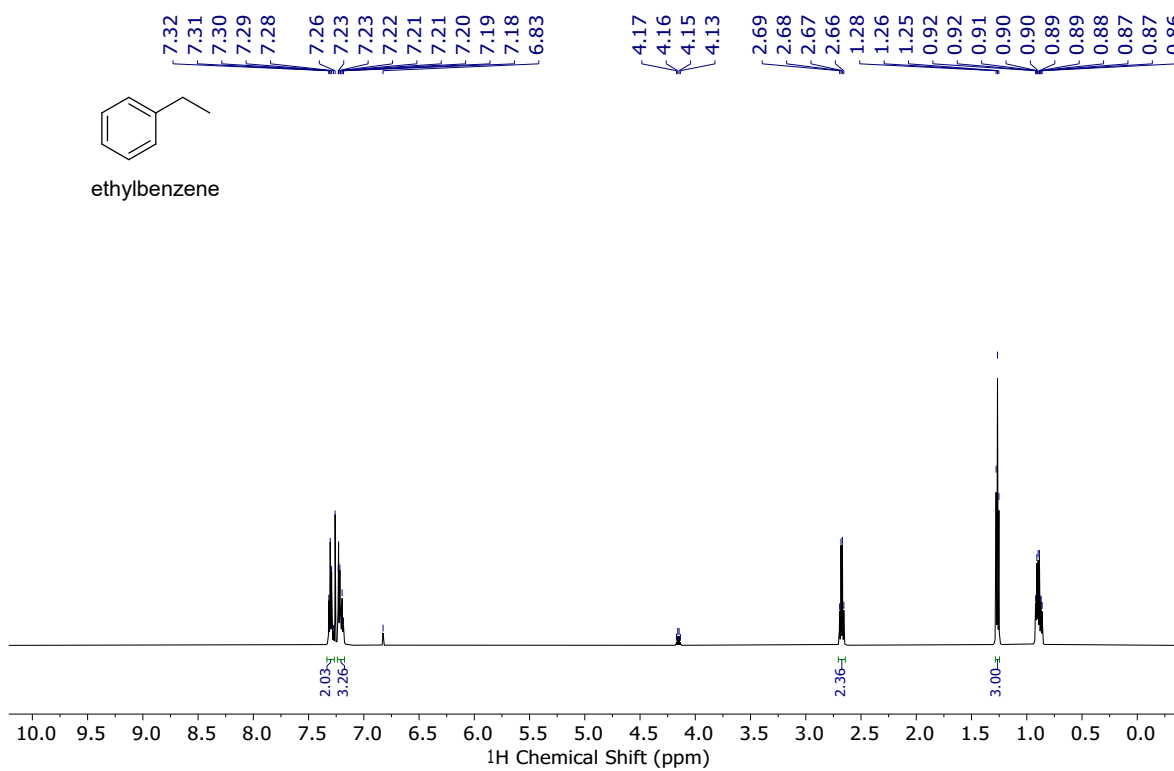
¹H NMR (600 MHz, CDCl₃) δ: 4.50 – 4.39 (m, 1H), 2.32 (dd, *J* = 10.2, 7.2 Hz, 2H), 2.23 – 2.10 (m, 1H), 1.69 – 1.58 (m, 1H), 1.20 – 1.16 (m, 3H). ¹³C NMR (151 MHz, CDCl₃) δ: 176.89, 76.84, 29.19, 28.60, 20.53.

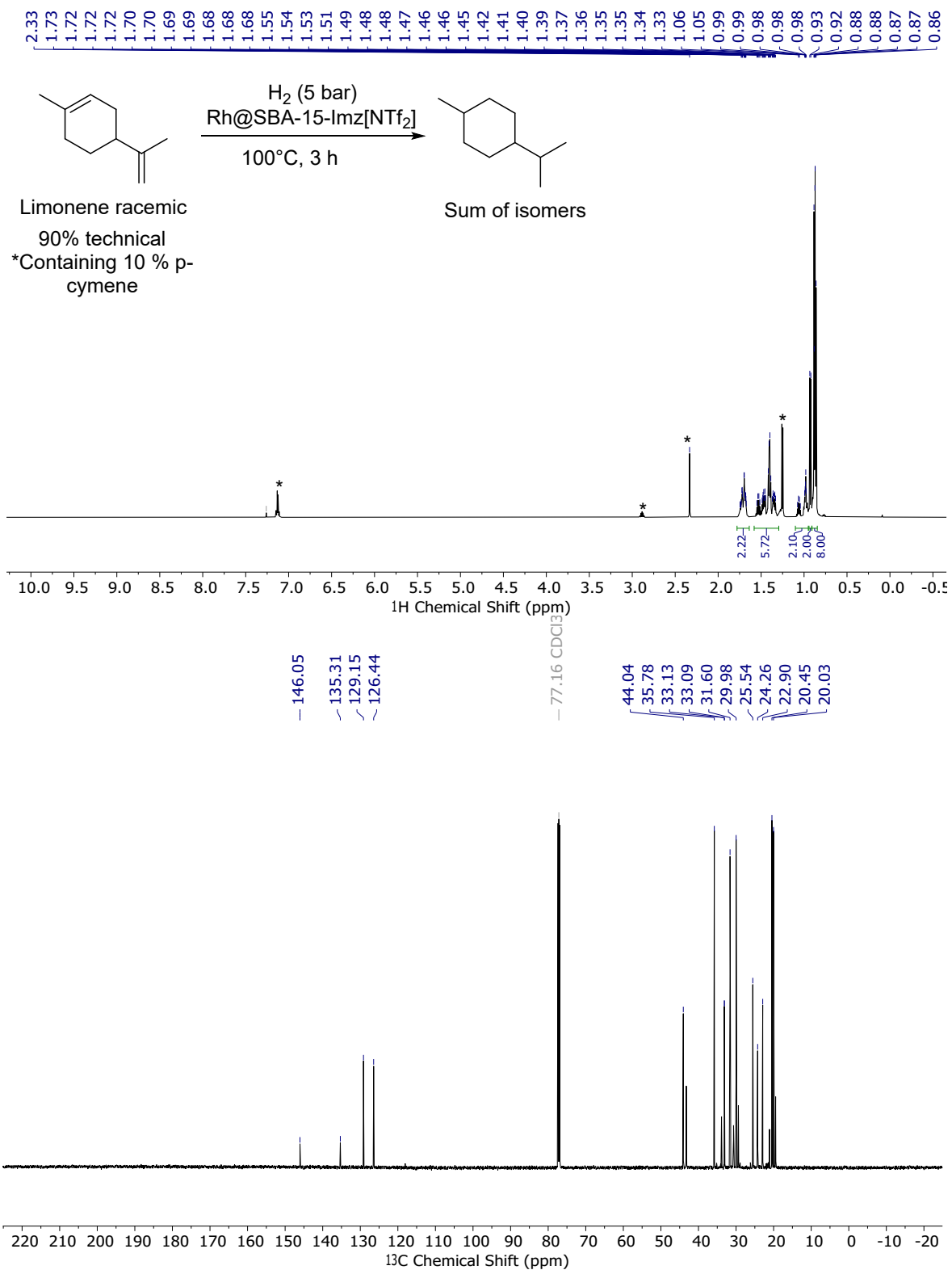


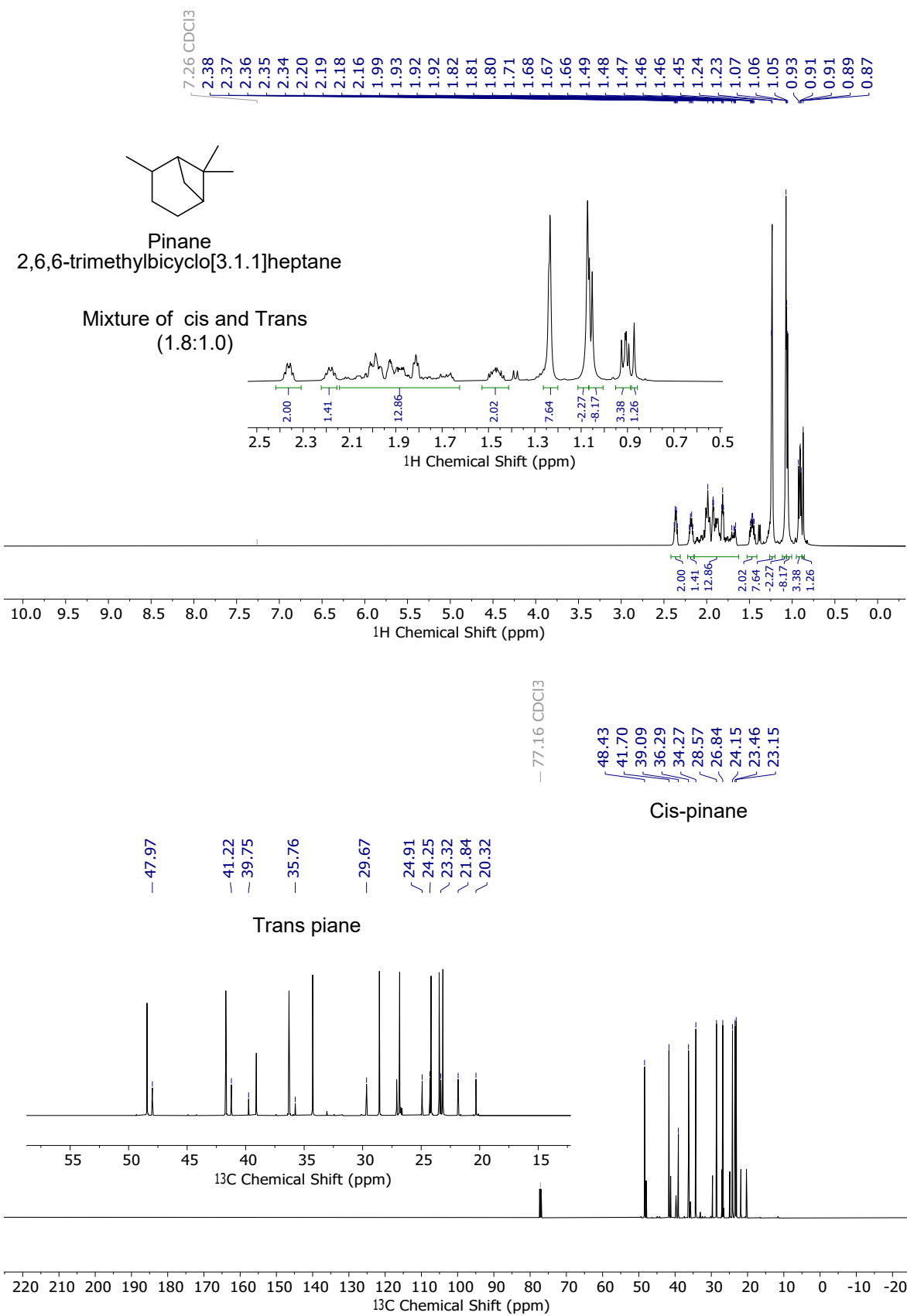
phenylmethanol ¹H NMR (600 MHz, CDCl₃) δ: 7.51 – 7.29 (m, 5H), 4.59 (s, 2H), 4.18 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ: 140.79, 128.25, 127.23, 126.85, 64.44.

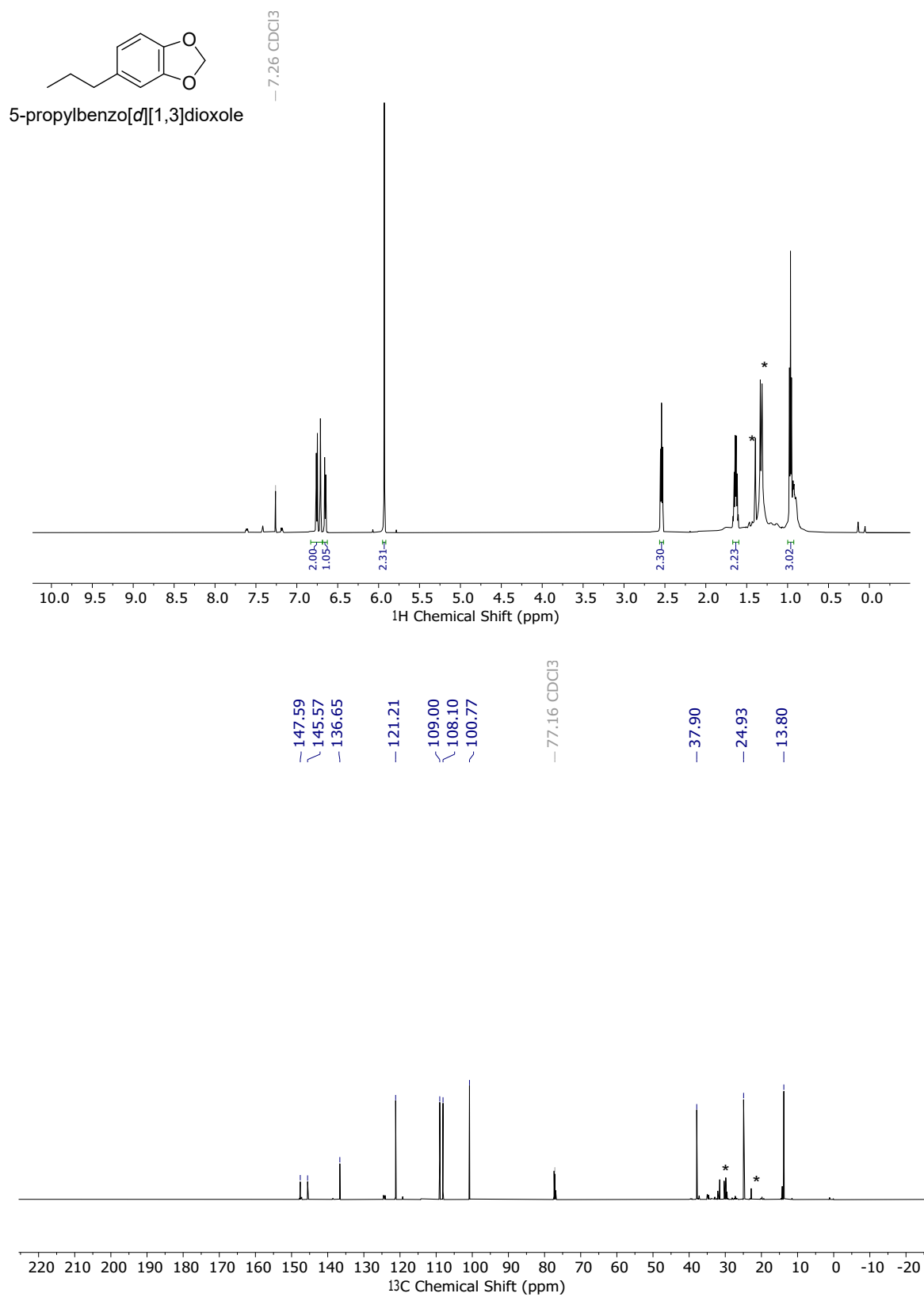


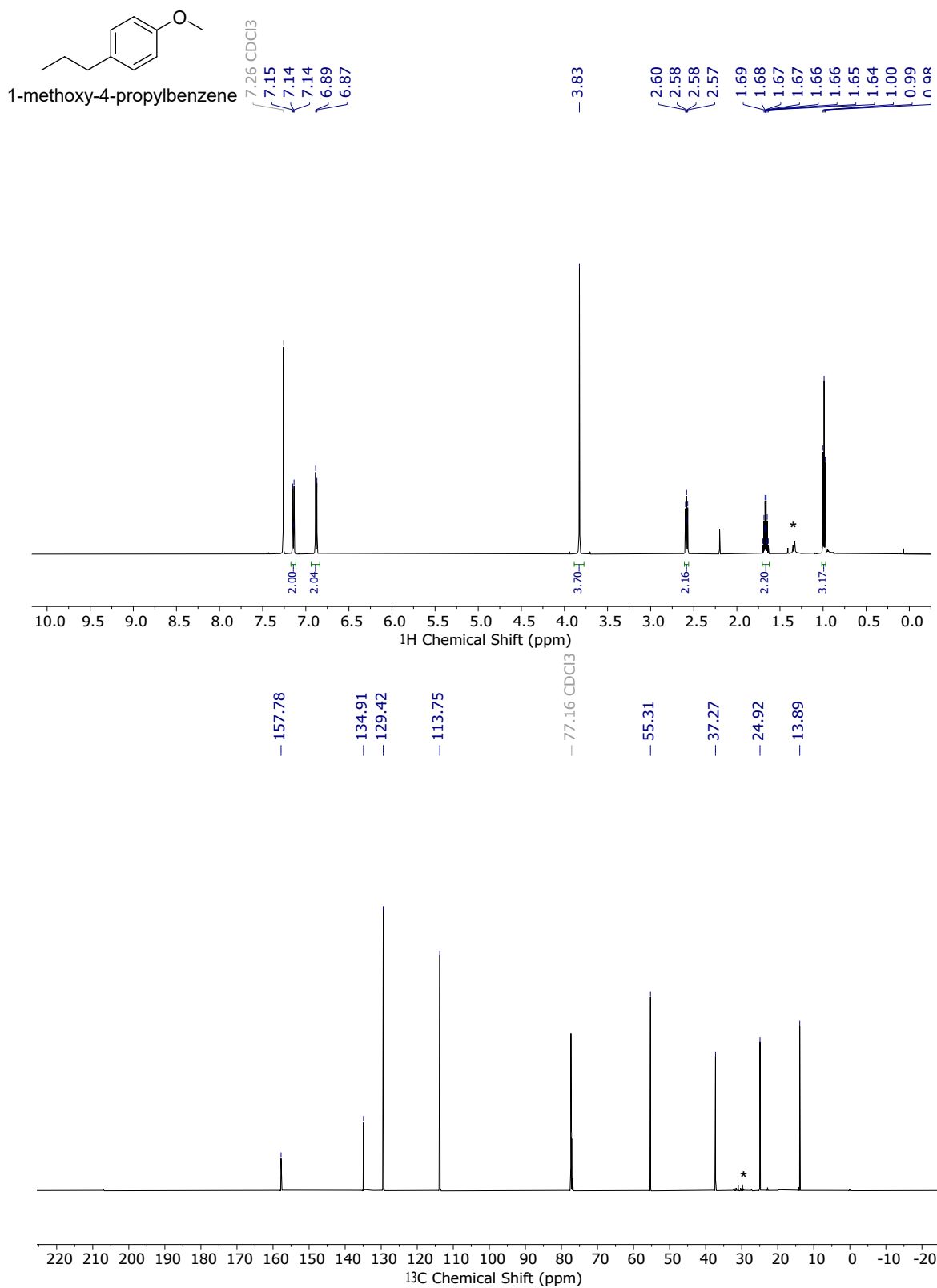


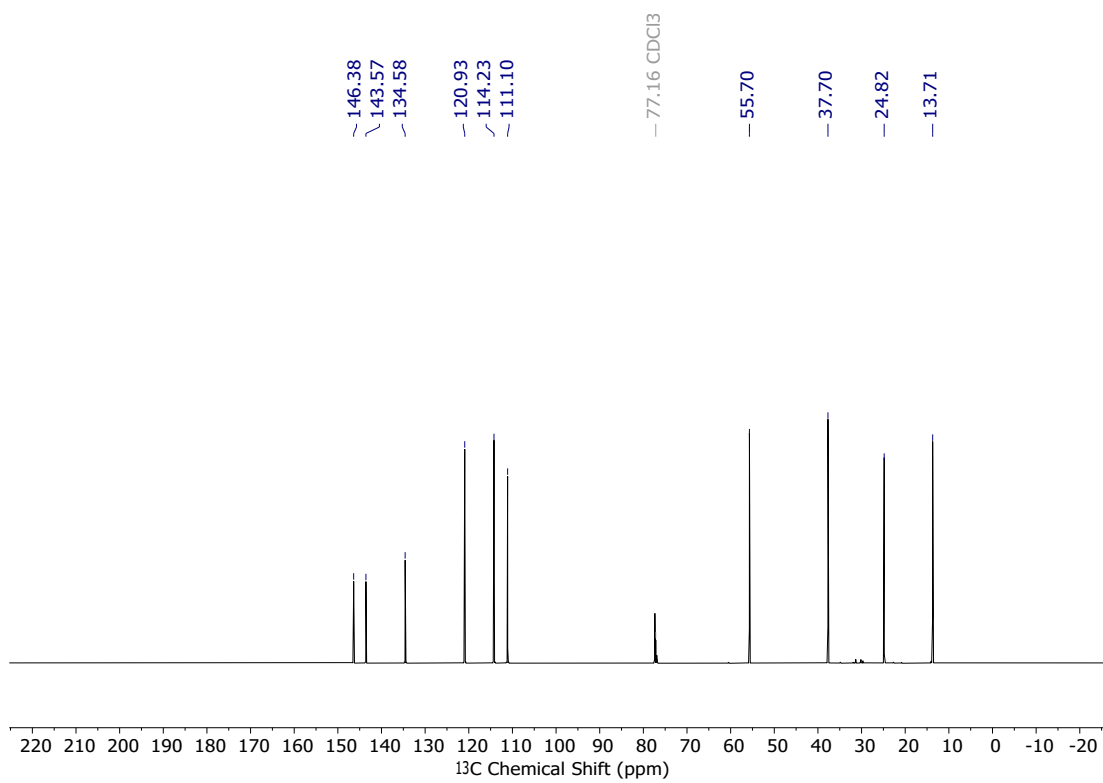
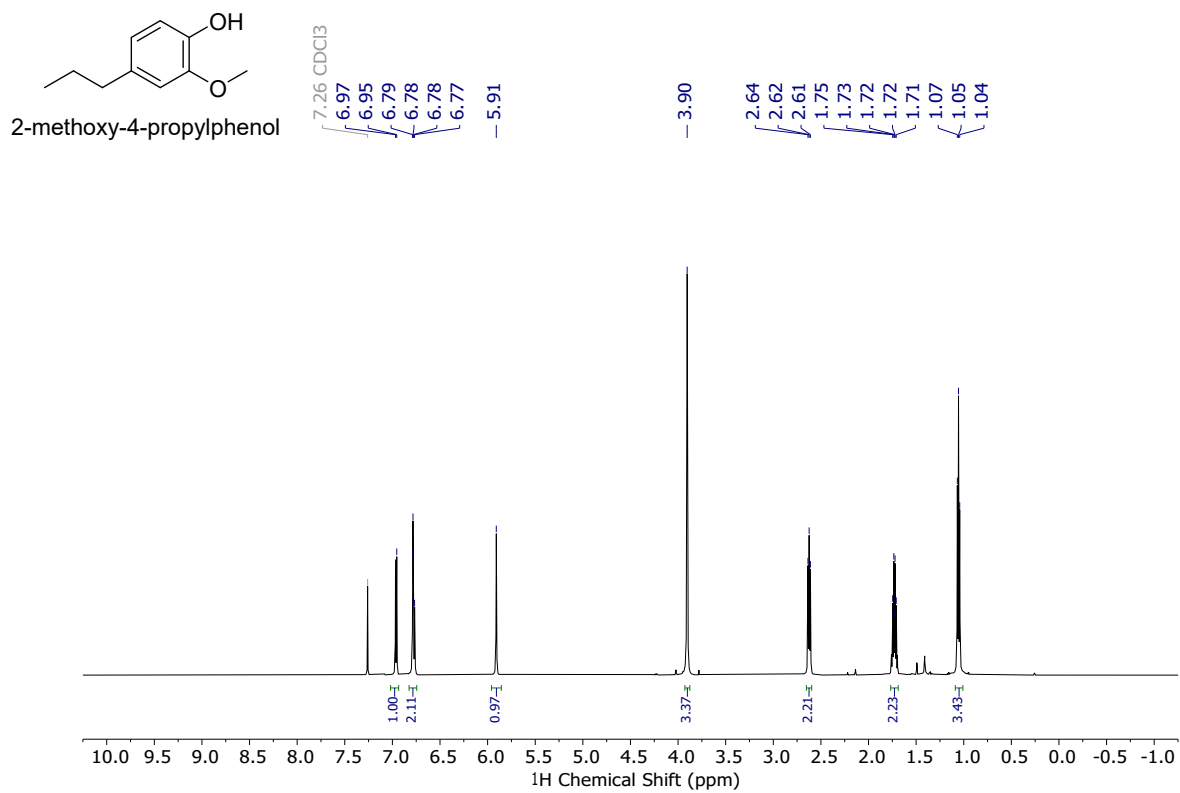


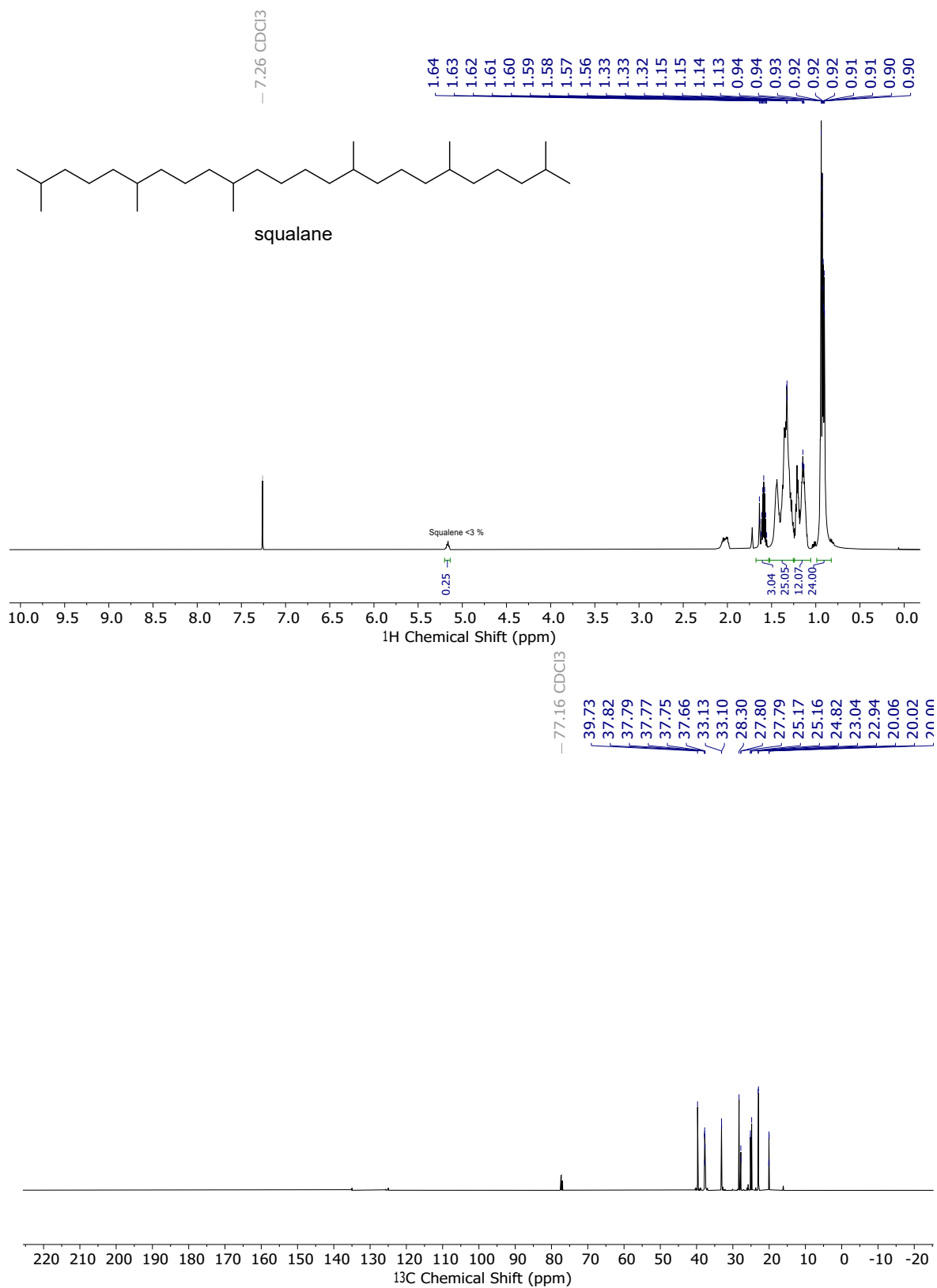


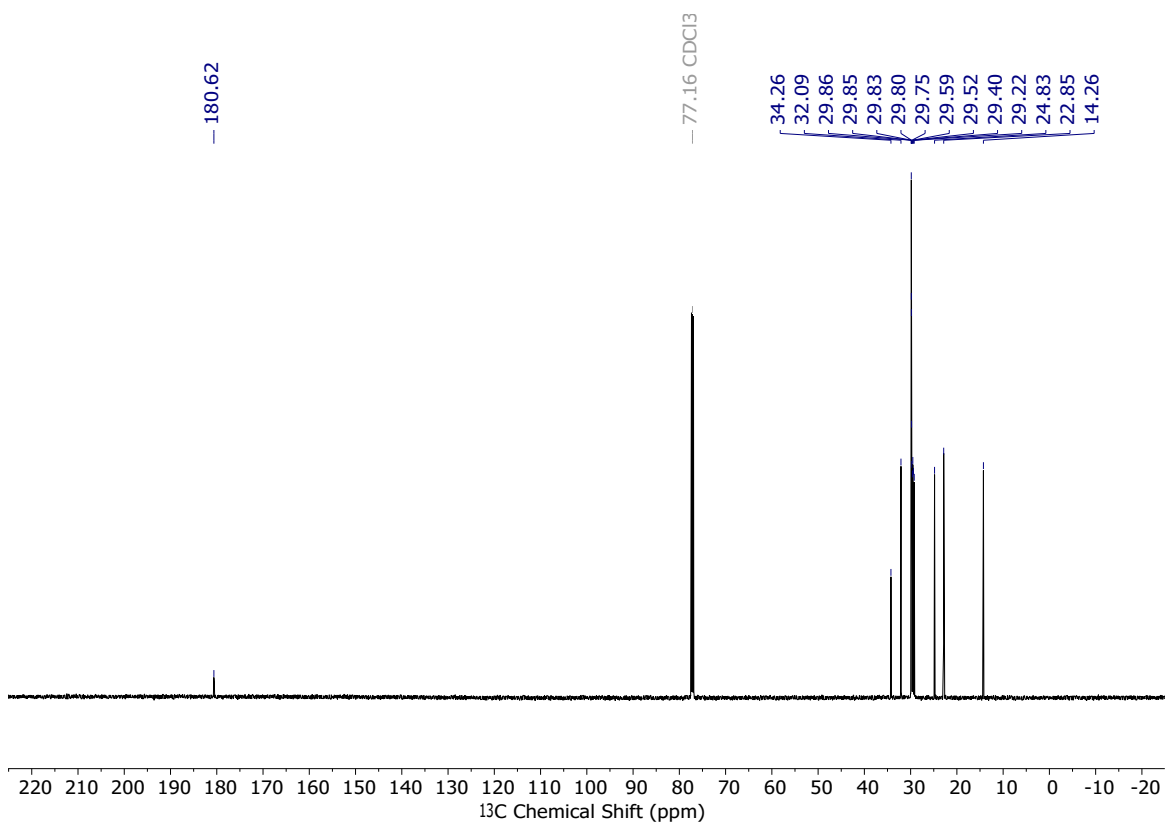
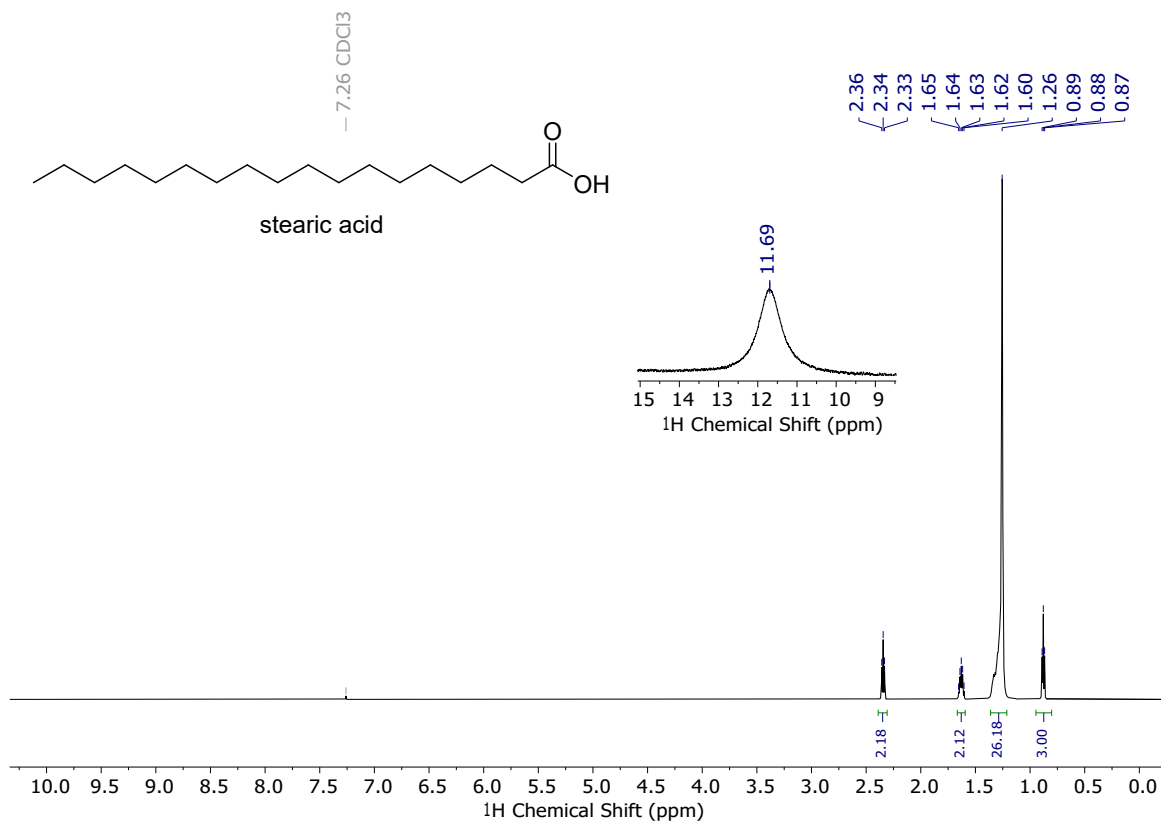


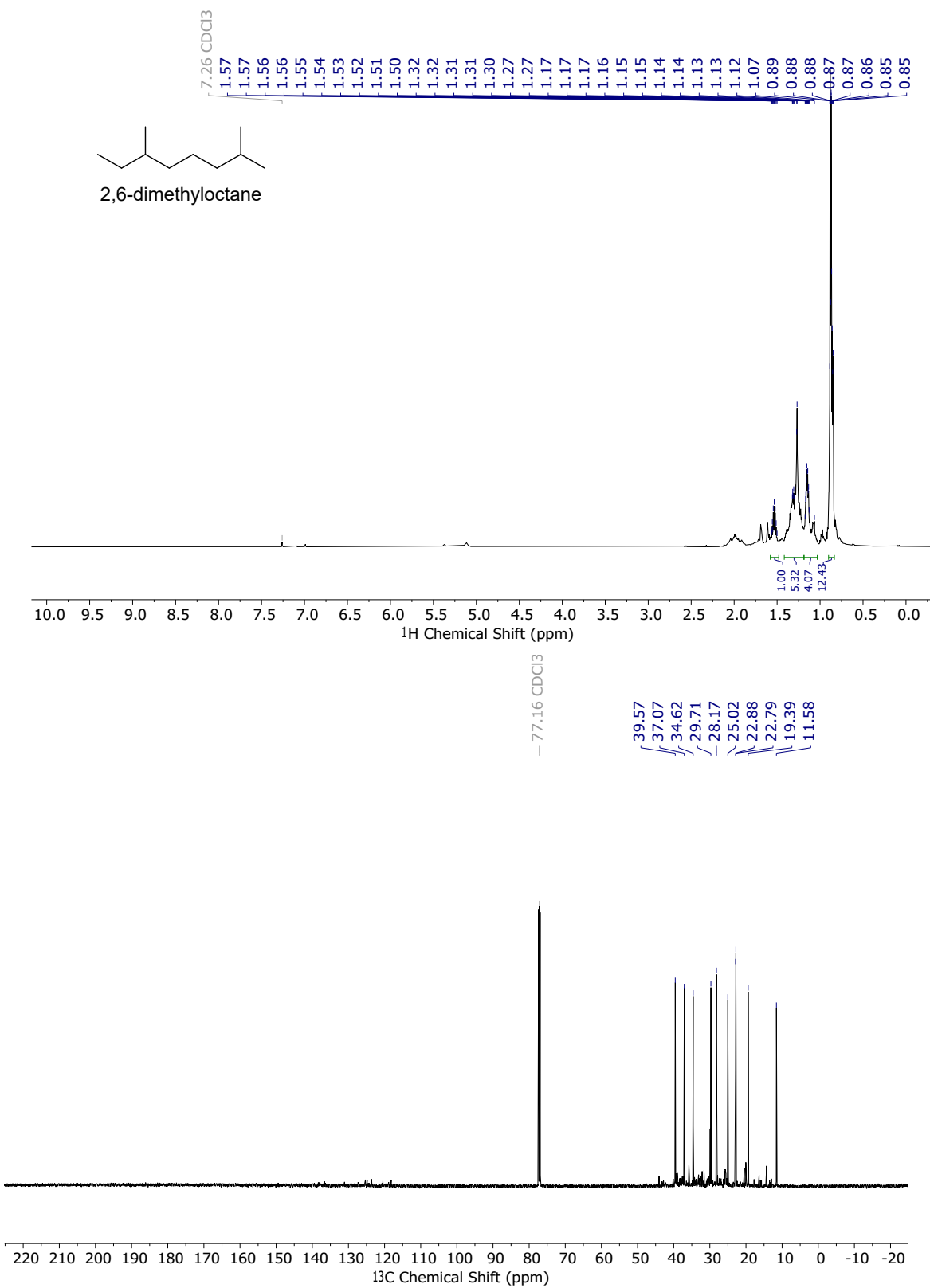


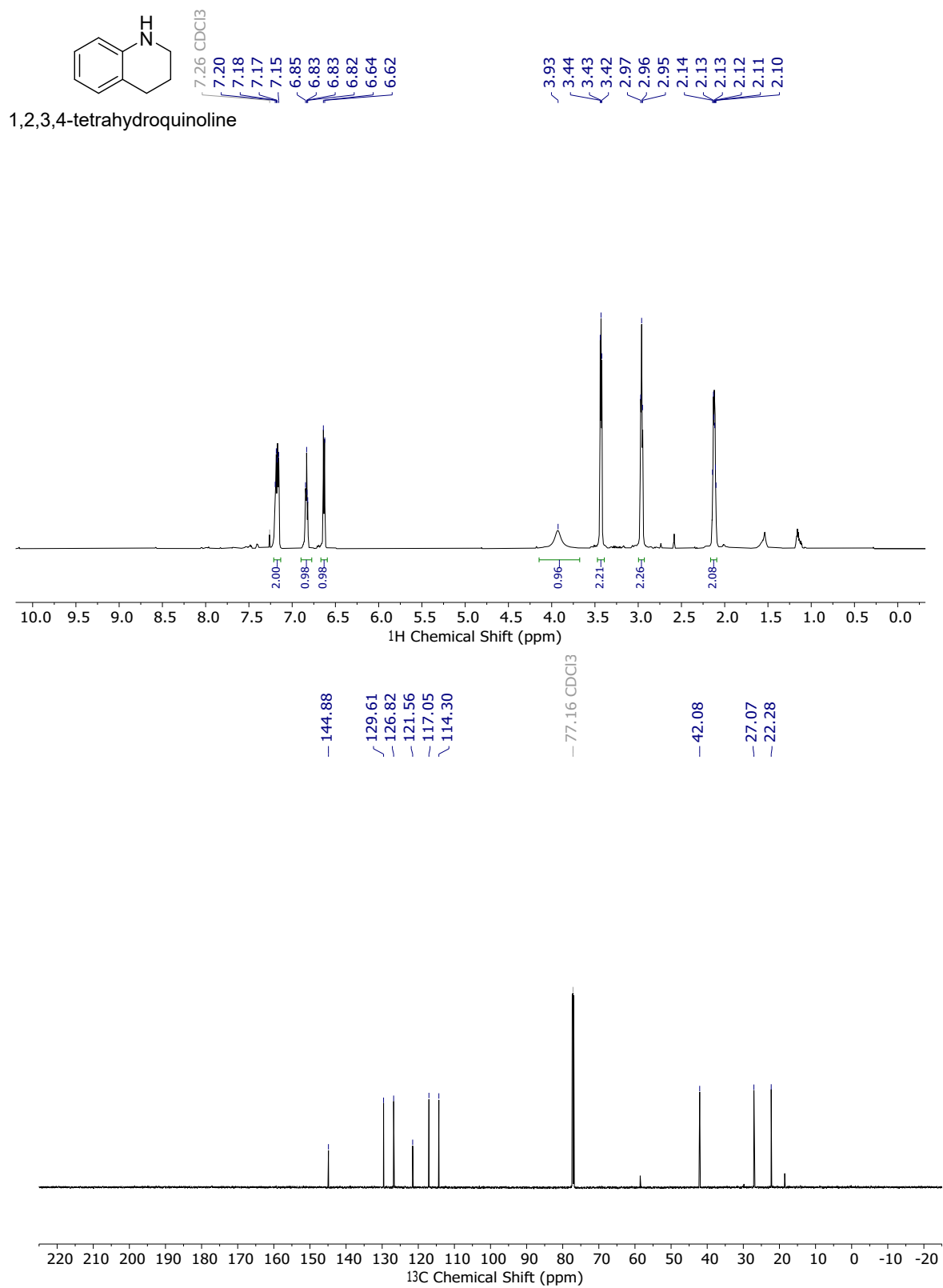


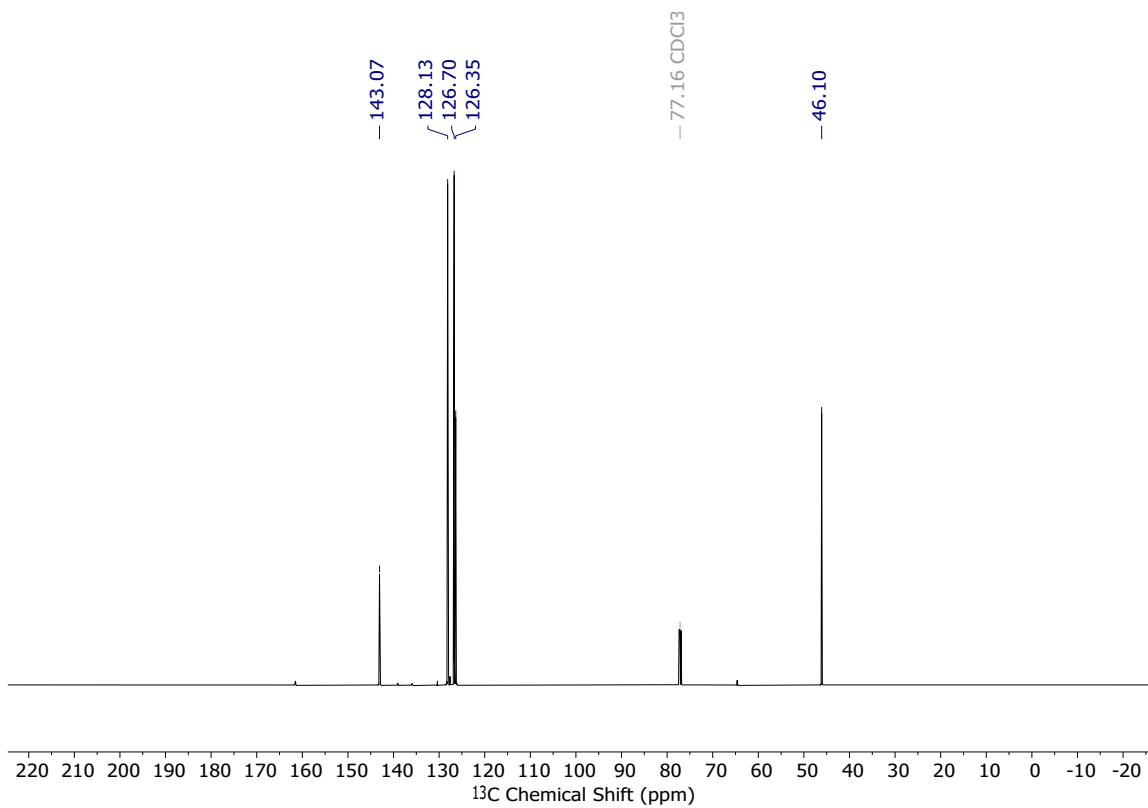
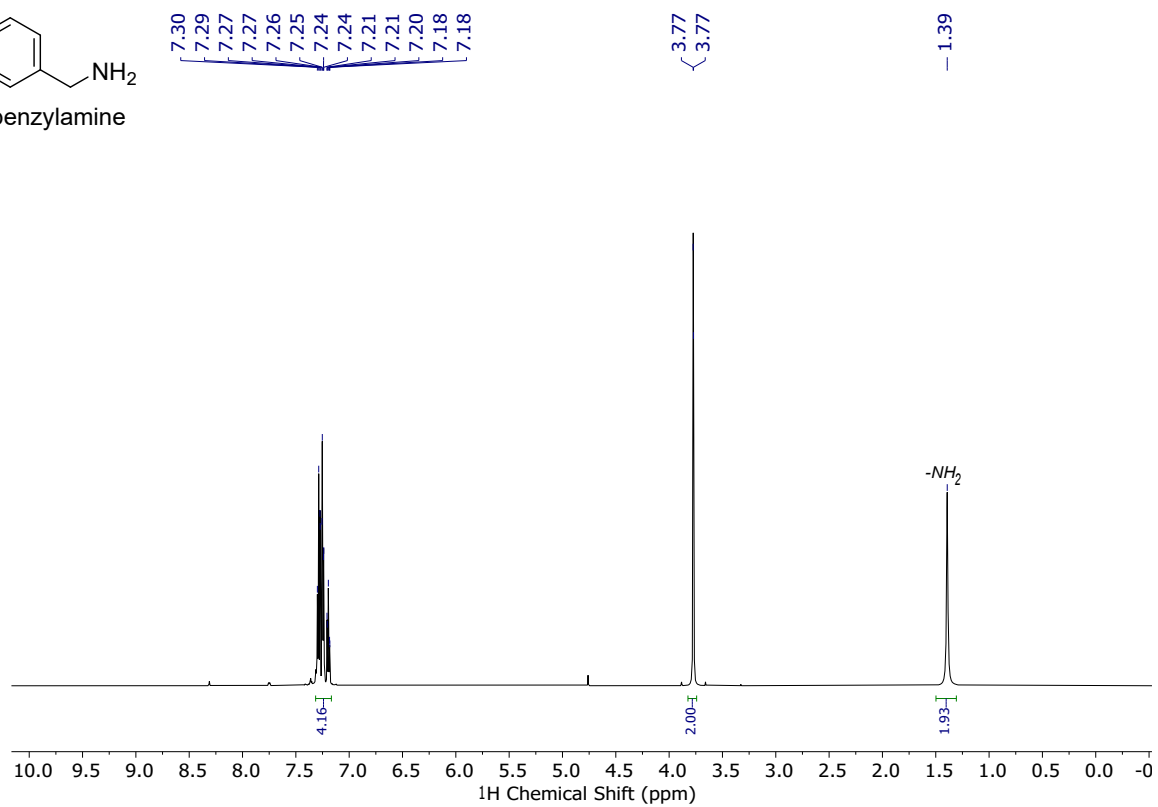
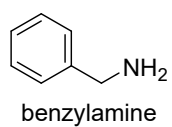


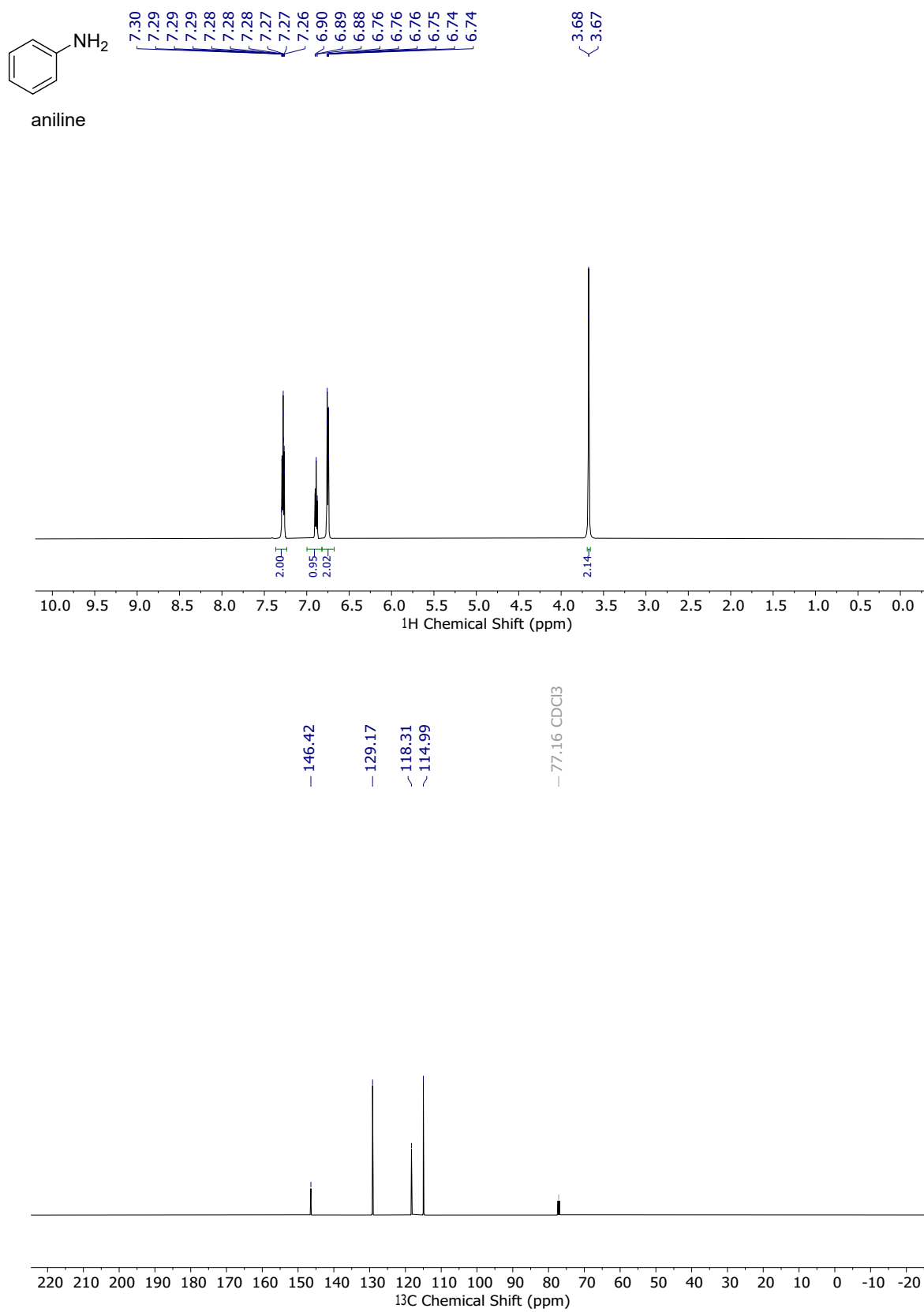


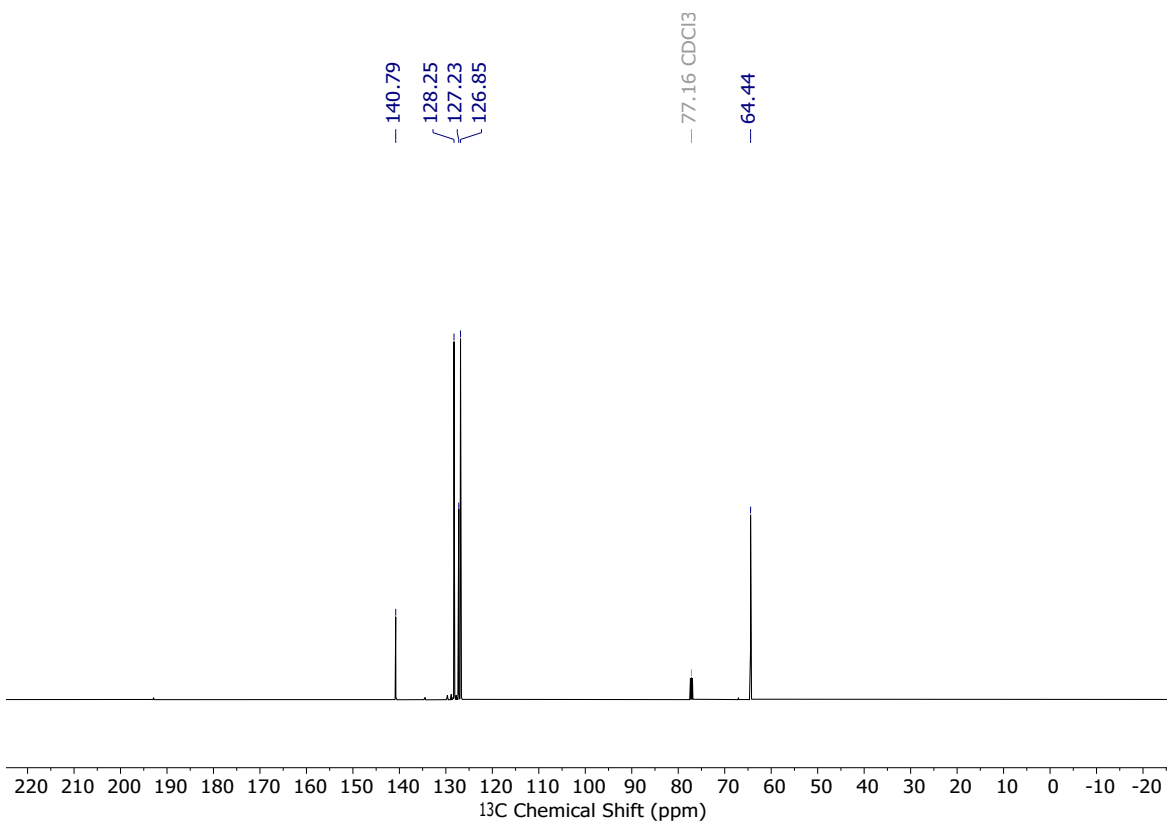
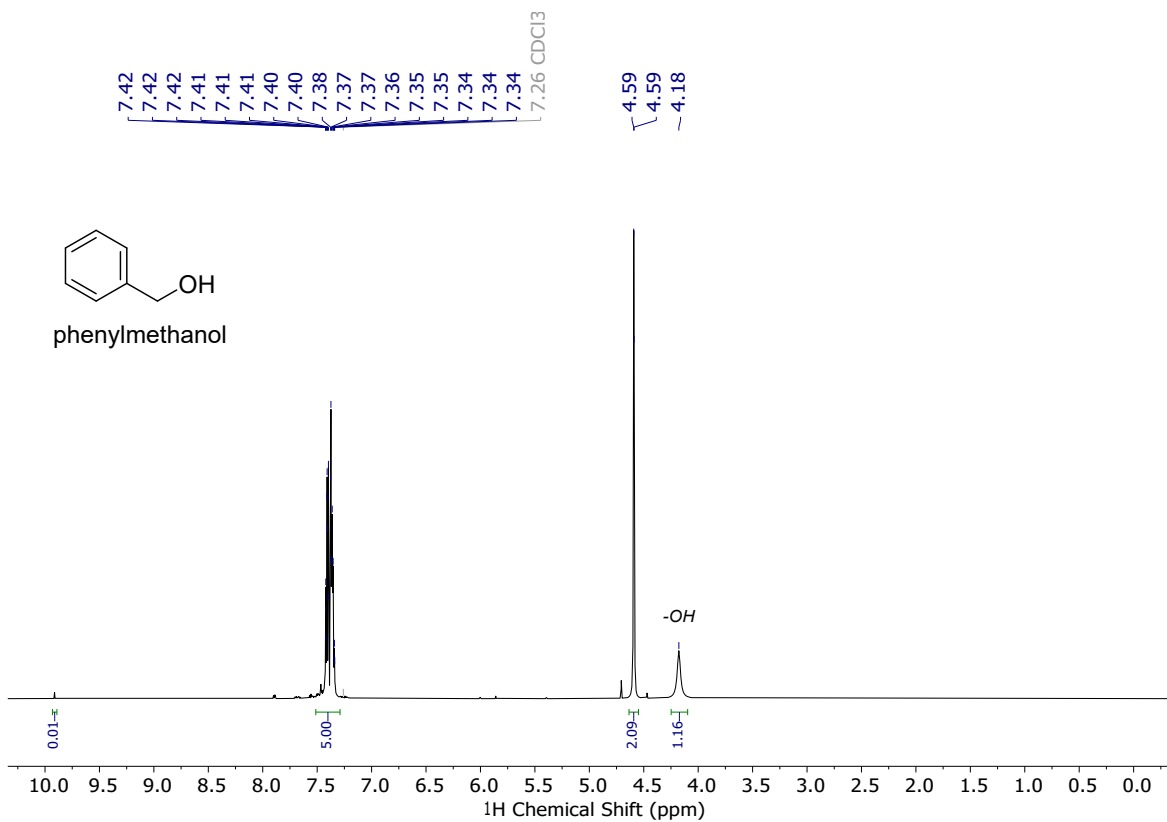


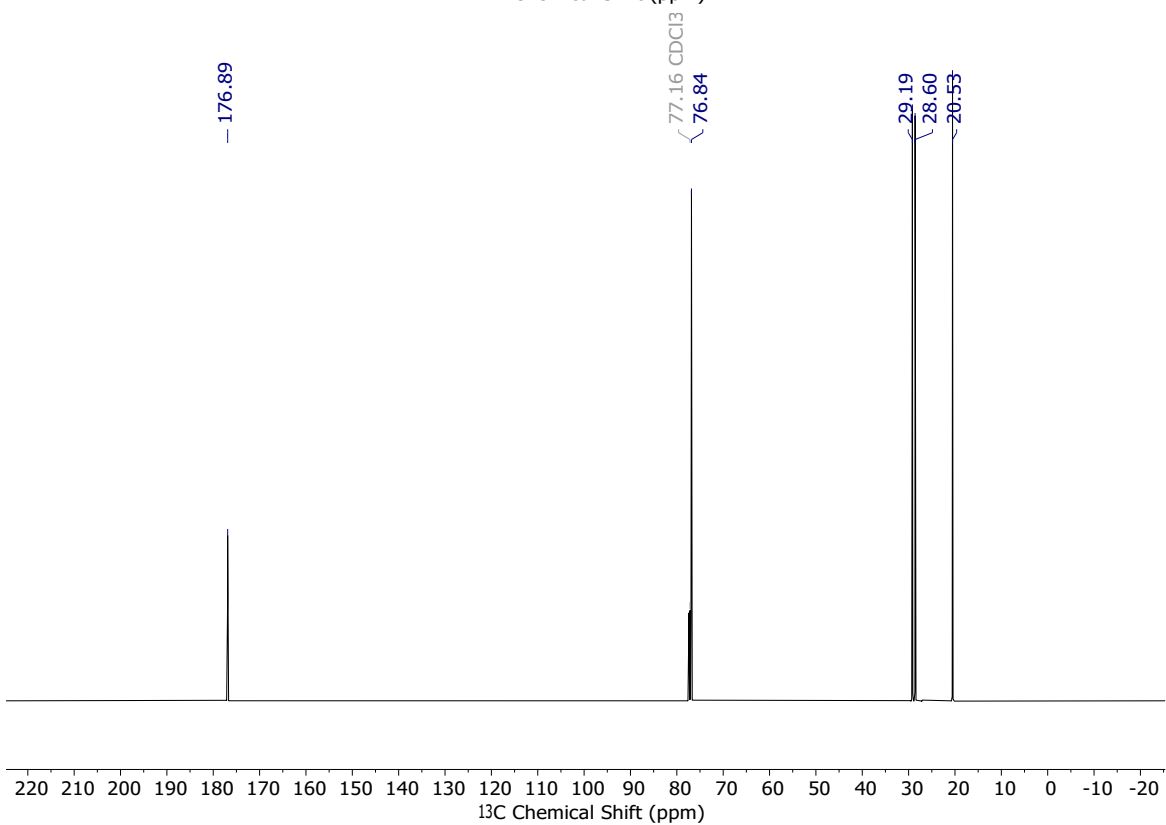
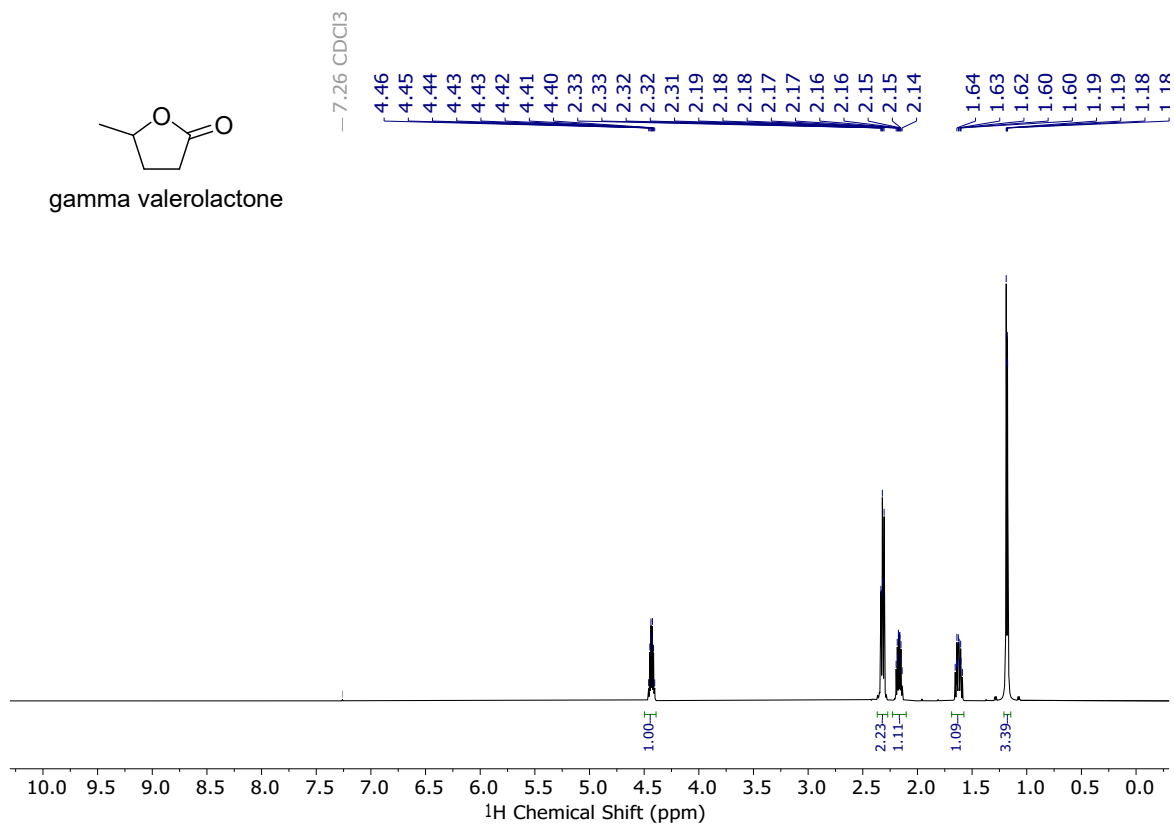
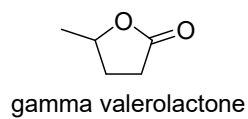


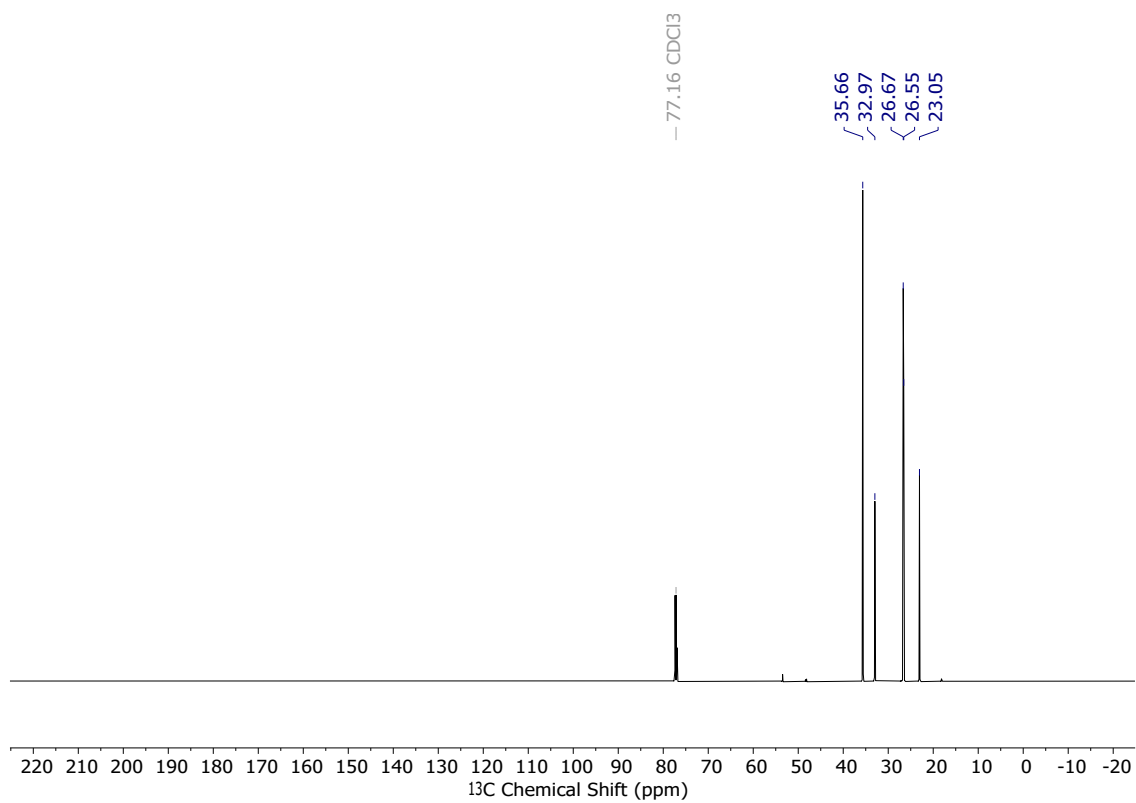
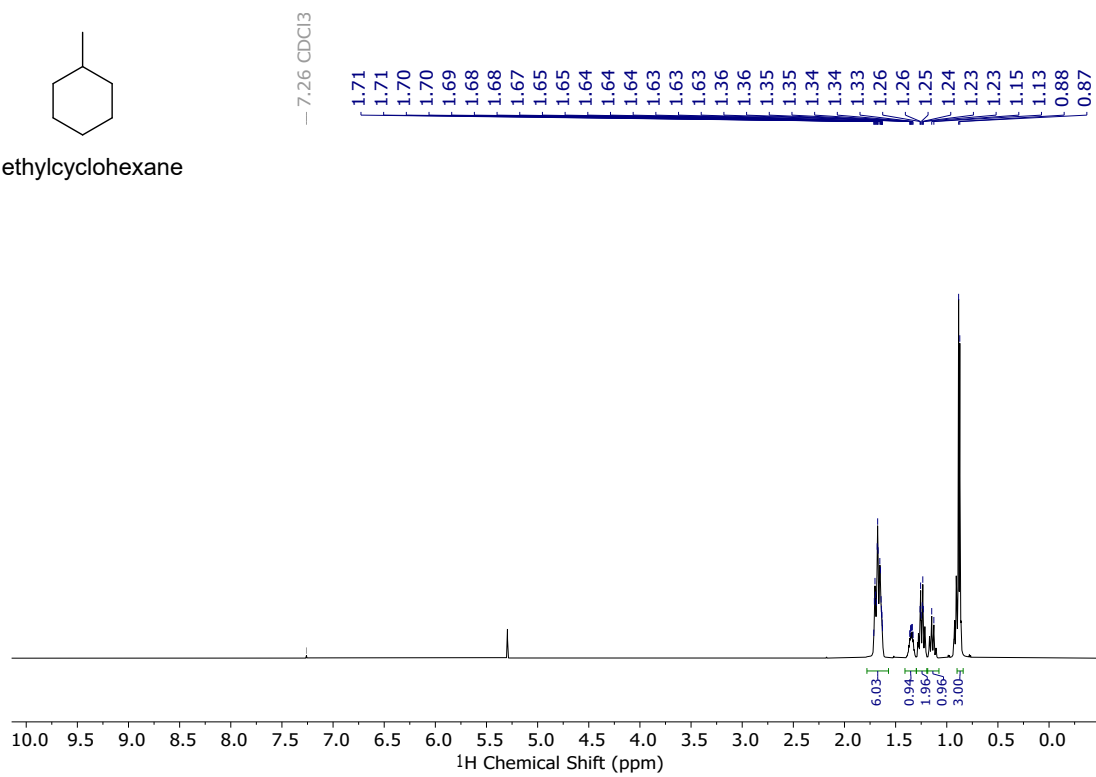
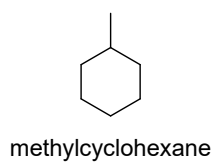












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