

Supplementary Material

# Immobilization of Horseradish Peroxidase on UIO- 66-NH<sub>2</sub> for Colorimetric and Fluorometric Sensing of Nitrite

Zuyao Fu <sup>a</sup>, Lingfeng Yang <sup>a</sup>, Zhaoyang Ding <sup>a,b,c \*</sup>, Jing Xie <sup>a,b \*</sup>

*a* College of Food Science and Technology, Shanghai Ocean University, Shanghai 201306, China

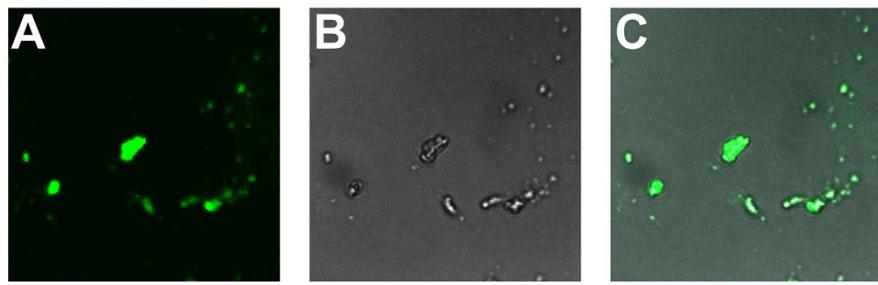
*b* Shanghai Engineering Research Center of Aquatic-Product Processing & Preservation, Shanghai 201306, China

*c* Marine Biomedical Science and Technology Innovation Platform of Lin-gang Special Area, Shanghai 201306, China

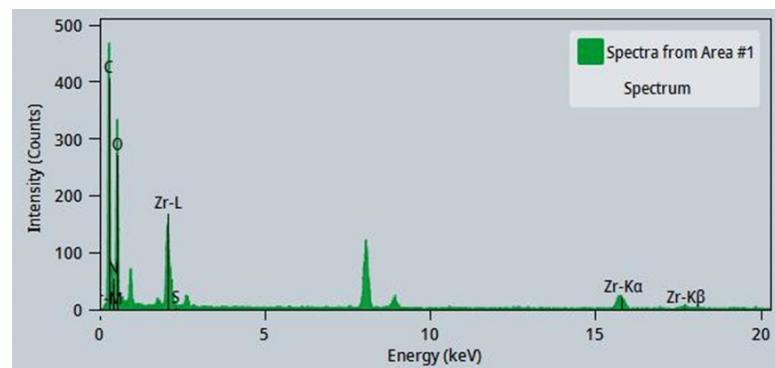
\* Corresponding authors. E-mail addresses: zyding@shou.edu.cn (Z. Ding); jxie@shou.edu.cn (J. Xie).

## **Characterizations**

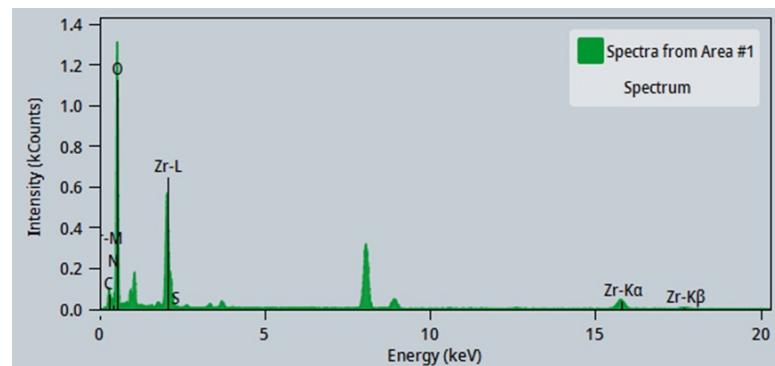
The morphology of the materials was observed using a Talos F200X transmission electron microscope (TEM) and an SU5000 scanning electron microscope (SEM). X-ray diffraction (XRD) measurements were obtained using an Ultima IV diffractometer. Fourier transform infrared spectroscopy spectra (FTIR) were recorded with a Nicolet iS5 spectrometer. Particle size and zeta potential were measured using a Zetasizer Pro Nanoparticle Size and Zeta Potential Analyzer. X-ray photoelectron spectroscopy (XPS) analysis was performed with an EscaLab 250Xi spectrometer. CLSM images were taken on Leica SP5. Ultraviolet-visible absorption spectra were acquired using a U-3900 spectrophotometer. Fluorescence signals were recorded using an F-7000 spectrophotometer.



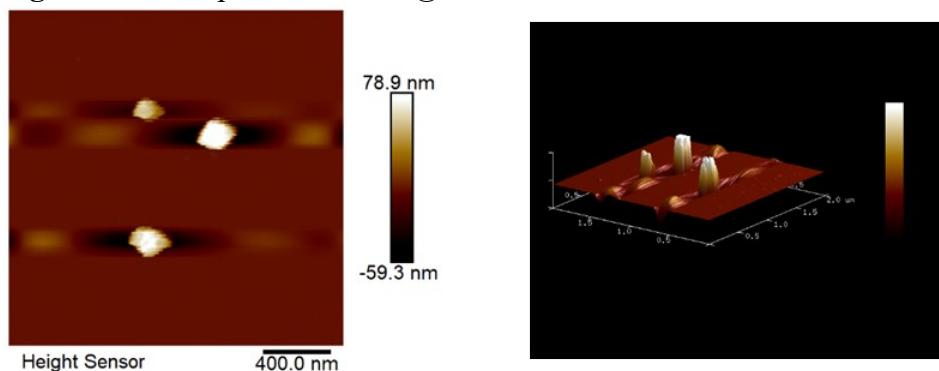
**Figure S1.** CLSM images of (A) fluorescent, (B) optical, and (C) overlap of optical and fluorescent images of HRP@UiO-66-NH<sub>2</sub>.



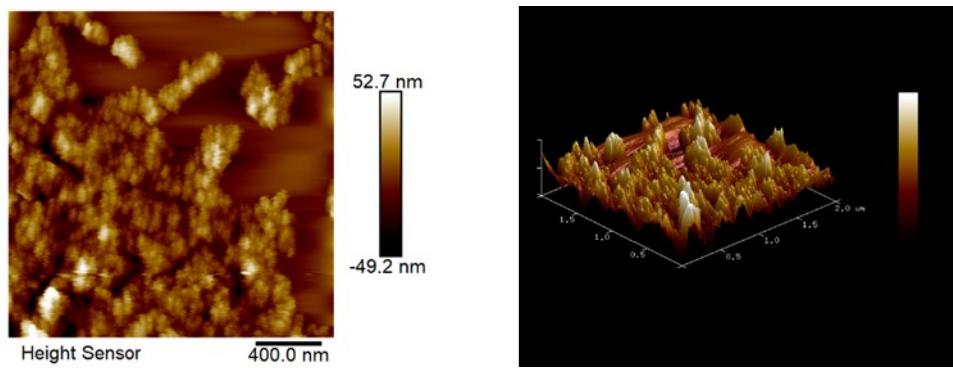
**Figure S2.** EDS pattern of UiO-66-NH<sub>2</sub>.



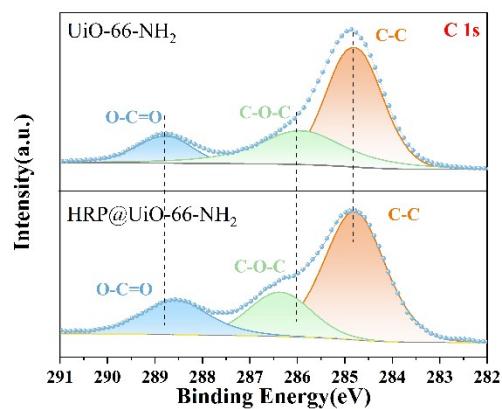
**Figure S3.** EDS pattern of HRP@UiO-66-NH<sub>2</sub>



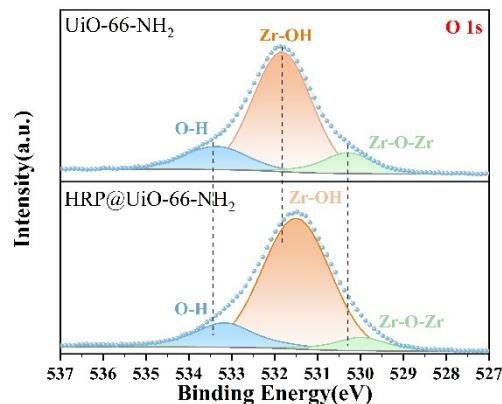
**Figure S4.** AFM pattern of UiO-66-NH<sub>2</sub>.



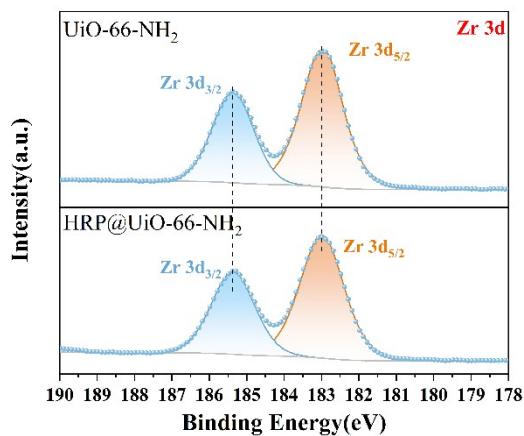
**Figure S5.** AFM pattern of  $\text{UiO-66-NH}_2$ .



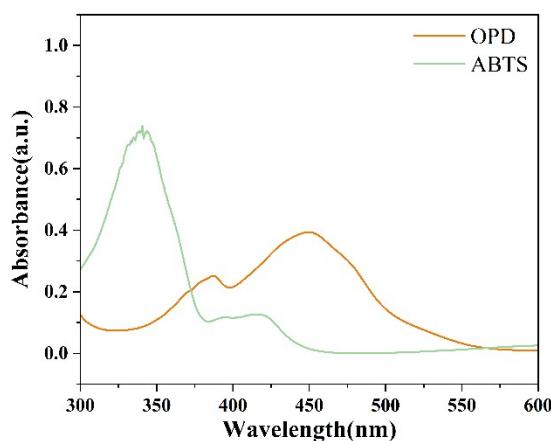
**Figure S6.** C 1s high resolution XPS spectra of  $\text{UiO-66-NH}_2$  (top) and  $\text{HRP@UiO-66-NH}_2$  (below).



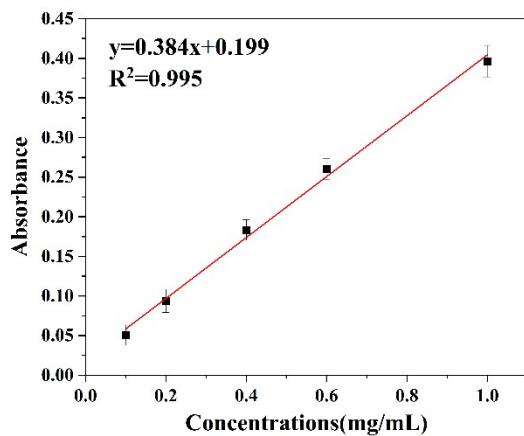
**Figure S7.** O 1s high resolution XPS spectra of  $\text{UiO-66-NH}_2$  (top) and  $\text{HRP@UiO-66-NH}_2$  (below).



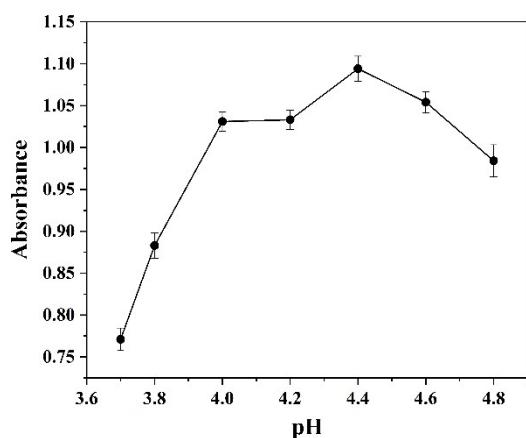
**Figure S8.** Zr 3d high resolution XPS spectra of  $\text{UiO-66-NH}_2$  (top) and  $\text{HRP@UiO-66-NH}_2$  (below).



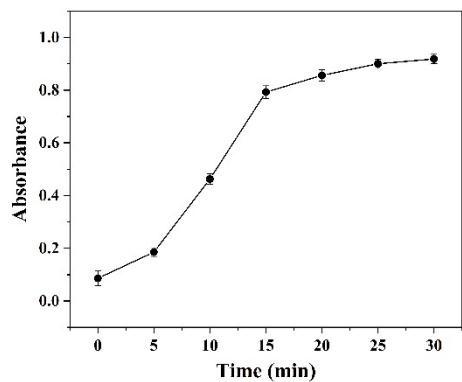
**Figure S9.** UV-vis absorption patterns of  $\text{HRP@UiO-66-NH}_2 + \text{OPD} + \text{H}_2\text{O}_2$  and  $\text{HRP@UiO-66-NH}_2 + \text{ABTS} + \text{H}_2\text{O}_2$ .



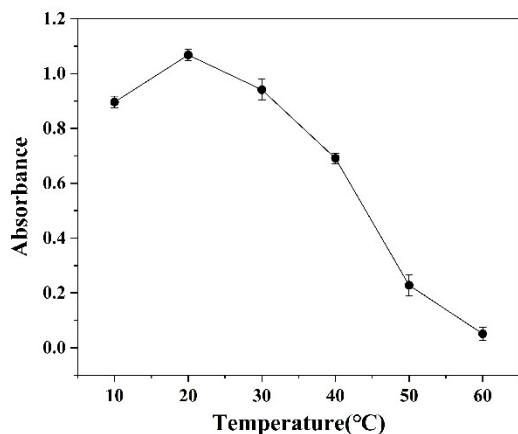
**Figure S10.** The standard curve of absorbance and BSA concentration.



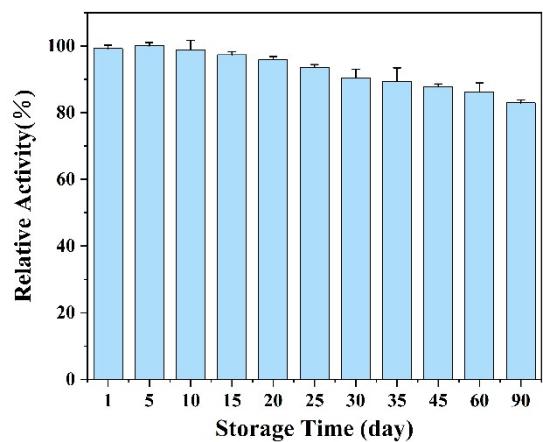
**Figure S11.** Effect of buffer pH on the absorbance of HRP@UiO-66-NH<sub>2</sub> + TMB + H<sub>2</sub>O<sub>2</sub>.



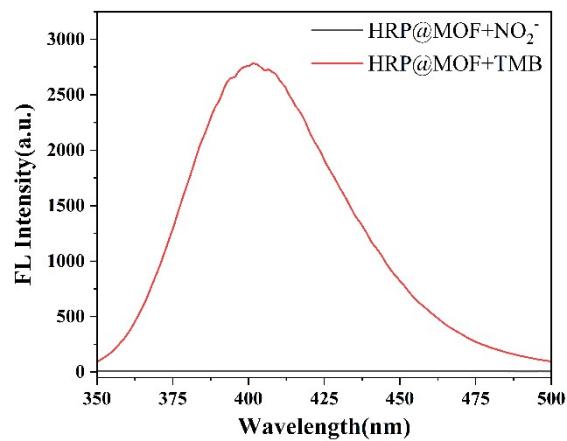
**Figure S12.** The absorbance of HRP@UiO-66-NH<sub>2</sub> + TMB + H<sub>2</sub>O<sub>2</sub> at different time.



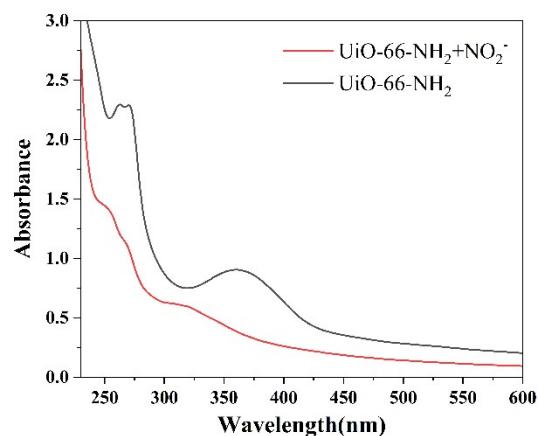
**Figure S13.** Effect of temperature on the absorbance of HRP@UiO-66-NH<sub>2</sub> + TMB + H<sub>2</sub>O<sub>2</sub>.



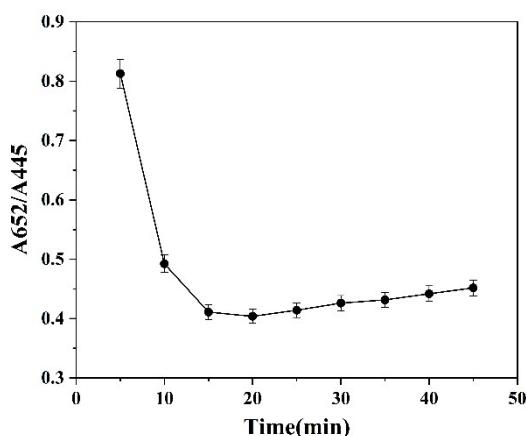
**Figure S14.** Oxidase-mimetic activity stability of HRP@UiO-66-NH<sub>2</sub> during storage



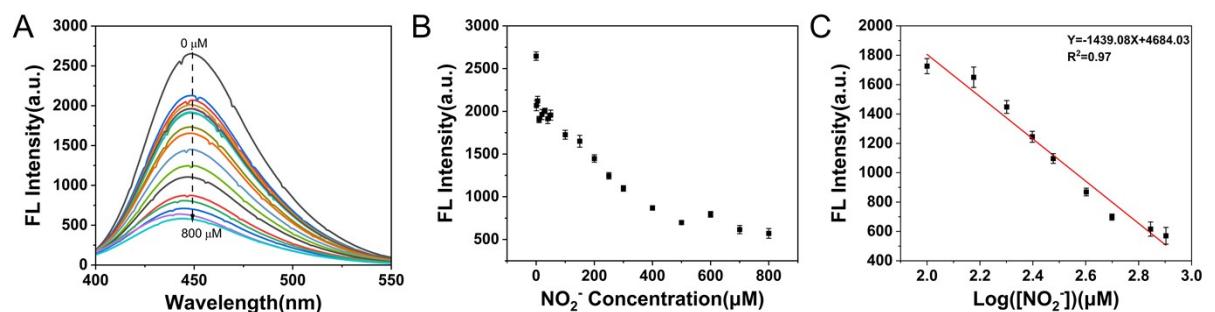
**Figure S15.** Fluorescence spectra of HRP@UiO-66-NH<sub>2</sub> in the presence and absence of NO<sub>2</sub><sup>-</sup>.



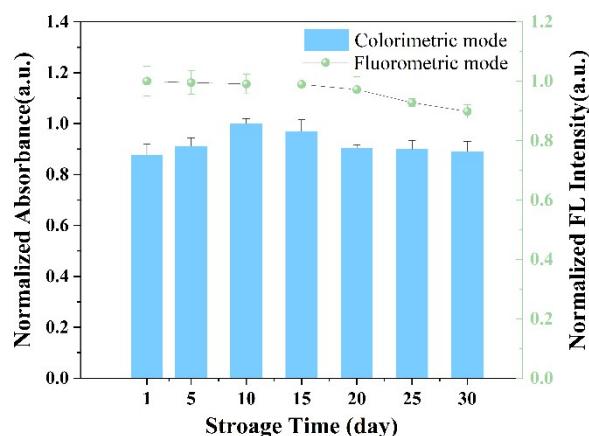
**Figure S16.** UV-vis absorption spectra of HRP@UiO-66-NH<sub>2</sub> in the presence and absence of NO<sub>2</sub><sup>-</sup>.



**Figure S17.** Effect of time on the colorimetric signal A652/A445 along with the diazotization time of TMB<sup>+</sup> and NO<sub>2</sub><sup>-</sup>.



**Figure S18.** The response of UiO-66-NH<sub>2</sub> toward nitrite.



**Figure S19.** The stability of colorimetric/fluorometric dual-mode sensors.

Table S1 The calculated catalytic kinetic parameters for free enzymes and HRP@UiO-66-NH<sub>2</sub>

substrate	enzyme probe	K <sub>m</sub> (mM)	V <sub>max</sub> (M s <sup>-1</sup> )	K <sub>cat</sub> (s <sup>-1</sup> )
TMB	HRP@UiO-66-NH <sub>2</sub>	0.23	15.36 × 10 <sup>-8</sup>	6.14 × 10 <sup>3</sup>
	Free HRP	0.434	10.00 × 10 <sup>-8</sup>	4.00 × 10 <sup>3</sup>
H <sub>2</sub> O <sub>2</sub>	HRP@UiO-66-NH <sub>2</sub>	1.00	2.36 × 10 <sup>-8</sup>	0.94 × 10 <sup>3</sup>
	Free HRP	3.70	8.71 × 10 <sup>-8</sup>	3.48 × 10 <sup>3</sup>

Table S2 Comparison of the reusability presented in this work with that reported previously.

Enzyme@(material)	Reusability	Residual activity	Ref
HRP@UiO-66-NH <sub>2</sub>	10	75.76%	This work
HRP@(amidoximated acrylic polymer)	10	45.0%	1
HRP@(functionalized reduced graphene oxide)	10	70%	2
HRP@(electrospun magnetic nanofibers)	5	52%	3
HRP@(functionalized reduced graphene oxide-SiO <sub>2</sub> )	10	70%	4
HRP@(cationic microporous starch)	10	66%	5
HRP@CN/Cu <sub>3</sub> (PO <sub>4</sub> ) <sub>2</sub>	5	83.6%	6
HRP@(PDA/expanded polystyrene foam)	15	60%	7

Table S3 Performance comparison of our colorimetric/fluorometric method with previously reported approaches for nitrite detection

Sensor Material	Year	Detection mode	Detecting range	Detection limit	Ref.
UiO-66-NH <sub>2</sub>	2020	Fluorometric	0–10 mM	77 μM	8
UiO-66-NH <sub>2</sub> -Cit	2020	Fluorometric	0–800 μM	–	9
Tb-MOF	2021	Fluorometric	4–200 μM	1.25 μM	10
Eu <sup>3+</sup> @UiO-66-(COOH) <sub>2</sub>	2021	Fluorometric	0–60 μM	0.69 μM	11
MnFeO	2021	Colorimetric	3.3~133.3 μM	0.2 μM	12
MnCDs	2022	Colorimetric	2–150 μM	0.16 μM	13

			Fluorometric	2 to 150 $\mu\text{M}$	1.07 $\mu\text{M}$	
CeO <sub>2</sub>	2023	Colorimetric	2–100 $\mu\text{M}$	0.29 $\mu\text{M}$	14	
Ru@UiO-66-NH <sub>2</sub>	2024	Fluorometric	10.0 - 15.0 $\mu\text{M}$	0.3 $\mu\text{M}$	15	
P–N–C nonmetal nanozymes	2024	Electrochemical	1–800 $\mu\text{M}$	0.21 $\mu\text{M}$		16
		Colorimetric	1 to 700 $\mu\text{M}$	0.24 $\mu\text{M}$		

Table S4 Detection results by our colorimetric/ fluorometric sensing platform of nitrite in real samples.

Detection mode	Sample	Spiked ( $\mu\text{M}$ )	Detected ( $\mu\text{M}$ )	RSD (%, n=3)	Recovery (%)
colorimetric		0	0.20	0.13	NA
	lake water	50	50.68	1.86	101.96
		100	89.31	0.92	89.11
		0	8.32	2.36	NA
	sausages	50	58.61	1.83	100.58
		100	107.93	4.61	99.61
		0	10.36	0.82	NA
	pickled fish	50	55.60	0.37	90.48
		100	105.75	1.64	95.39
		0	1.92	0.38	NA
Fluorometric	pickle brine	50	52.37	4.93	100.9
		100	101.49	1.73	99.57
		0	0.22	2.37	NA
	lake water	50	51.36	2.66	102.28
		100	103.84	1.87	103.62
		0	8.57	2.47	NA
	sausages	50	53.28	0.63	89.24
		100	109.83	1.79	101.26
		0	10.52	1.21	NA
	pickled fish	50	60.88	3.07	100.72
		100	104.39	0.93	93.87
		0	2.81	4.71	NA
	pickle brine	50	49.29	0.89	92.96
		100	103.82	5.32	101.01

NA = not applicable

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