

Supporting Information

Synthesis of 2*H*-imidazoles via Copper-Catalyzed Homo/Cross-Coupling of Oxime Acetates

Min Liu^a, Bifu Liu^{a*}, Hongyan Chen^a, Qian Wang^a, Lixin Liu^a, Kejun Feng^a, Zijia Wang^a and Qiang Li^b

A School of Chemistry and Materials Engineering, Huizhou University, Huizhou, 516007, Guangdong, China

b Shandong Provincial Key Laboratory of Chemical Energy Storage and Novel Cell Technology, School of Chemistry and Chemical Engineering, Liaocheng University, No. 1, Hunan Street, Liaocheng, Shandong 252000, P. R. China

E-Mail: liubf@hzu.edu.cn

Contents:

1. General information	Page 2
2. Typical procedure for preparation of oxime acetates	Page 2
3. Typical procedure for the synthesis of 2 <i>H</i> -imidazoles	Page 2-3
4. Typical procedure for the scale-up synthesis of 2 <i>H</i> -imidazoles	Page 3
5. Typical procedure for the scale-up synthesis of 5aa	Page 3
6. Condition screening	Page 3
7. Characterization and analytical data of products	Page 3-9
8. References	Page 9
9. ¹ H and ¹³ C-NMR spectrum of products	Page 10-42

General information

All reactions were carried out under air atmosphere using standard Schlenk technique in the parallel synthesizer. All reagent/reactant were commercially available unless other noted, DCE (99.5%, Extra Dry, with molecular sieves) and the related dry solvents screened were purchased from Energy Chemical (Note: the solvents used above could be altered to normal solvent with the additive of 4A Ms.). Column chromatography was performed using Silica Gel 60 (particle size and 37-54 μm). The pure products were obtained by column chromatography using ethyl acetate/petroleum ether as the eluent and characterized by NMR spectroscopy using CDCl_3 as the deuterium reagent. GC analysis was performed on GC 7820A (Shimadzu). GC-MS results were recorded on GC-MS QP2030 (Shimadzu). The ^1H NMR and ^{13}C NMR data were acquired on a Bruker ADVANCE III spectrometer (400 MHz for ^1H NMR spectroscopy and 100 MHz for ^{13}C NMR spectroscopy). HRMS analysis was conducted at the College of Chemistry and Materials Engineering, Wenzhou University, Wenzhou, 325000, China.

Typical procedure for preparation of oxime acetates

General Procedure for oximes: To a solution of aromatic ketones (2 mmol) in the mixture of $\text{C}_2\text{H}_5\text{OH}/\text{H}_2\text{O}$ ($v/v = 1:1$) was added hydroxylamine hydrochloride (2.2 mmol), NaOAc (3 mmol) in one portion, and the reaction mixture was stirred at 100 $^\circ\text{C}$ for 6-8 h. Upon completion of the reaction as indicated by TLC, the reaction mixture was diluted with water, extracted with ethyl acetate, and dried over anhydrous Na_2SO_4 . The solvent was removed and concentrated under reduced pressure to give oximes.

General Procedure for oxime acetates: The mixture of ketoximes (2.0 mmol), anhydride (4.0 mmol, 2.0 equiv) was stirred in CH_2Cl_2 (10 mL) at room temperature for 24 h. Upon completion of the reaction as indicated by TLC, the reaction mixture was diluted with EtOAc (25 mL), washed with H_2O (20 mL) and aq. NaHCO_3 (10 mL). Neutralization with NaHCO_3 and dried over anhydrous Na_2SO_4 and evaporated in vacuo. The crude residue was purified by column chromatography using silica gel with hexane as the eluent to give oxime acetates.

Typical procedure for the synthesis of 2*H*-imidazoles

Process a: the synthesis of 2-methyl-2,4-diphenyl-2*H*-imidazole

An oven-dried Schlenk tube containing a stir bar was charged with **1a** 1-phenylethan-1-one O-acetyl oxime (0.3 mmol, 53.1 mg) and CuCl (0.03 mmol, 2.97 mg), for which then were dissolved in 2 mL anhydrous DCE. The mixture was stirred at 120 $^\circ\text{C}$ for 4 h under air. After reaction, the mixture was diluted with 3-5 mL EA, then the volatiles was removed and the residues were passed through a short silica chromatography column (particle size 37-54 μm , petroleum ether/ethyl acetate as eluent) to afford analytically pure product **3a**.

Process b: the synthesis of 2,2,4-triphenyl-2*H*-imidazole

An oven-dried Schlenk tube containing a stir bar was charged with **1a** 1-phenylethan-1-one O-acetyl oxime (0.15 mmol, 26.5 mg), **4a** diphenylmethanone O-acetyl oxime (0.15 mmol, 35.8 mg) and CuCl (0.03 mmol, 2.97 mg), for which then were dissolved in 2 mL anhydrous DCE. The mixture was stirred at 120 $^\circ\text{C}$ for 8 h under air. After reaction, the mixture was diluted with 3-5 mL EA, then the volatiles was removed and the residues were passed through a short silica

chromatography column (particle size 37–54 μm , petroleum ether/ethyl acetate as eluent) to afford analytically pure product **4aa**.

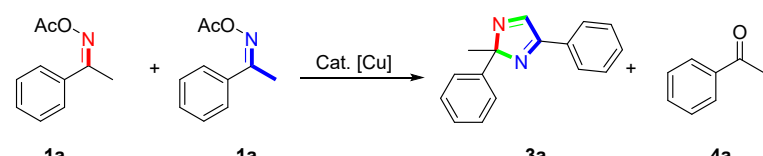
Typical procedure for the scale-up synthesis of 2*H*-imidazoles

An oven-dried 100 mL Schlenk tube containing a stir bar was charged with **1a** 1-phenylethan-1-one O-acetyl oxime (6 mmol, 1062 mg) and CuCl (0.6 mmol, 59.4 mg), for which then were dissolved in 20 mL anhydrous DCE. The mixture was stirred at 120 $^{\circ}\text{C}$ for 12 h under air. After reaction, the volatiles was removed and the residues were passed through a short silica chromatography column (particle size 37–54 μm , petroleum ether/ethyl acetate as eluent) to afford analytically pure product **3a**, 582.7mg, 83% yield.

Typical procedure for the scale-up synthesis of **5aa**

To a solution of 2,2,4-triphenyl-2*H*-imidazole **4aa** (1.0 mmol), dry CH_3CN (2 mL) and *m*-CPBA (344 mg, 2.0 equiv) were added to a 10 mL screw-capped tube at 80 $^{\circ}\text{C}$. Upon completion of the reaction as indicated by TLC, the solvent was removed and concentrated under reduced pressure to give crude raffinate. The crude raffinate was purified by column chromatography using silica gel with eluent (petroleum ether: EtOAc = 5 : 1) to afford **5aa** (188 mg, 60% yield).

Condition screening

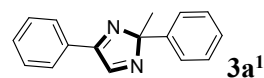


The reaction scheme shows two molecules of 1a (1-phenylethan-1-one O-acetyl oxime) reacting with catalytic copper (Cat. [Cu]) to produce 3a (2,2-diphenyl-2*H*-imidazole) and 4a (acetophenone).

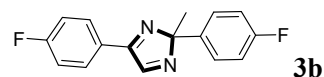
Entry	Acids/ 2 eq.	Yield 3a /%	Yield 4a /%
1	CF_3COOH	trace	95
2	CH_3COOH	trace	94
3	PTSA	trace	98
4	Citric acid	trace	98
5	H_3PO_4	trace	87
6	NH_4Cl	6	93

^a Reaction conditions: **1a** 0.15 mmol, **2a** 0.15 mmol, cat. 0.03 mmol, additives 0.3 mmol, solvent 2.0 mL, under air, 4 h. GC yield using tridecane as the internal standard based on 0.15 mmol of **1a**.

Characterization and analytical data of products

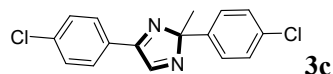


Yield 93%. ^1H NMR(400 MHz, CDCl_3) δ 8.48 (s, 1H), 8.07 (d, J = 6.8 Hz, 2H), 7.75 (d, J = 8.0 Hz, 2H), 7.53-7.51 (m, 3H), 7.38-7.27 (m, 3H), 1.87 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 163.7, 154.3, 140.0, 131.4, 131.0, 129.0, 128.3, 128.3, 127.6, 126.9, 109.1, 27.0.

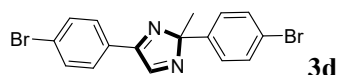


Yield 87%. ^1H NMR(400 MHz, CDCl_3) δ 8.43 (s, 1H), 8.06 (t, J = 6.4 Hz, 2H), 7.68 (t, J = 6.4 Hz,

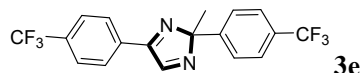
2H), 7.19 (t, $J = 8.0$ Hz, 2H), 7.02 (t, $J = 8.0$ Hz, 2H), 1.80 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 166.0 (d, $J_{\text{F-C}} = 251.3$ Hz), 163.4 (d, $J_{\text{F-C}} = 244.8$ Hz), 162.6, 154.1, 135.9 (d, $J_{\text{F-C}} = 3.2$ Hz), 130.5 (d, $J_{\text{F-C}} = 8.8$ Hz), 128.7 (d, $J_{\text{F-C}} = 8.0$ Hz), 127.2 (d, $J_{\text{F-C}} = 3.3$ Hz), 116.4 (d, $J_{\text{F-C}} = 142.0$ Hz), 116.2 (d, $J_{\text{F-C}} = 98.8$ Hz), 108.7, 27.2; ^{19}F NMR(377 MHz, CDCl_3), δ -107.4, -114.6. HRMS (ESI): calcd for $\text{C}_{16}\text{H}_{12}\text{F}_2\text{N}_2$: $[\text{M}+\text{H}^+]$ 271.1041, found 271.1047.



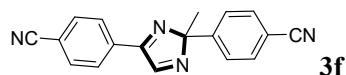
Yield 84%. ^1H NMR(400 MHz, CDCl_3) δ 8.42 (s, 1H), 7.97 (d, $J = 8.0$ Hz, 2H), 7.65 (d, $J = 8.0$ Hz, 2H), 7.47 (d, $J = 8.0$ Hz, 2H), 7.30 (d, $J = 8.0$ Hz, 2H), 1.79 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 162.8, 154.1, 138.4, 137.7, 133.5, 129.5, 129.3, 129.2, 128.3, 128.3, 108.7, 27.0. HRMS (ESI): calcd for $\text{C}_{16}\text{H}_{12}\text{Cl}_2\text{N}_2$: $[\text{M}+\text{H}^+]$ 303.0450, found 303.0456.



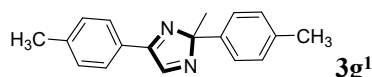
Yield 82%. ^1H NMR(400 MHz, CDCl_3) δ 8.43 (s, 1H), 7.91 (d, $J = 8.4$ Hz, 2H), 7.64 (d, $J = 8.0$ Hz, 2H), 7.59 (d, $J = 8.0$ Hz, 2H), 7.46 (d, $J = 8.4$ Hz, 2H), 1.79 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 163.0, 154.1, 138.9, 132.3, 131.4, 129.7, 129.7, 128.7, 126.2, 121.8, 108.9, 27.0. HRMS (ESI): calcd for $\text{C}_{16}\text{H}_{12}\text{Br}_2\text{N}_2$: $[\text{M}+\text{H}^+]$ 390.9440, found 390.9444.



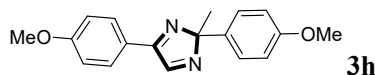
Yield 62%. ^1H NMR(400 MHz, CDCl_3) δ 8.52 (s, 1H), 8.19 (d, $J = 8.0$ Hz, 2H), 7.89 (d, $J = 8.0$ Hz, 2H), 7.79 (d, $J = 8.0$ Hz, 2H), 7.62 (d, $J = 8.4$ Hz, 2H), 1.85 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 163.0, 154.2, 143.7, 134.0, 133.4 (d, $J_{\text{F-C}} = 32.6$ Hz), 130.2 (d, $J_{\text{F-C}} = 32.2$ Hz), 128.7, 127.5, 126.1 (q, $J_{\text{F-C}} = 3.7$ Hz), 125.4 (d, $J_{\text{F-C}} = 36.8$ Hz), 125.3 (q, $J_{\text{F-C}} = 3.7$ Hz), 122.7 (d, $J_{\text{F-C}} = 37.4$ Hz), 109.2, 27.2; ^{19}F NMR(377 MHz, CDCl_3), δ -62.6, -63.0. HRMS (ESI): calcd for $\text{C}_{18}\text{H}_{12}\text{F}_6\text{N}_2$: $[\text{M}+\text{H}^+]$ 371.0977, found 371.0982.



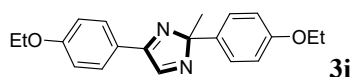
Yield 76%. ^1H NMR(400 MHz, CDCl_3) δ 8.51 (s, 1H), 8.17 (d, $J = 7.6$ Hz, 2H), 7.86 (d, $J = 8.0$ Hz, 2H), 7.82 (d, $J = 8.0$ Hz, 2H), 7.63 (d, $J = 7.6$ Hz, 2H), 1.81 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 162.8, 154.1, 144.7, 134.5, 132.8, 132.1, 128.9, 127.9, 118.5, 117.9, 115.1, 111.7, 109.2, 27.1. HRMS (ESI): calcd for $\text{C}_{18}\text{H}_{12}\text{N}_4$: $[\text{M}+\text{H}^+]$ 285.1135, found 285.1141.



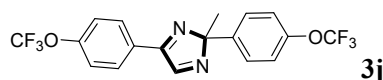
Yield 91%. ^1H NMR(400 MHz, CDCl_3) δ 8.46 (s, 1H), 7.96 (d, $J = 8.0$ Hz, 2H), 7.61 (d, $J = 7.6$ Hz, 2H), 7.32 (d, $J = 8.0$ Hz, 2H), 7.17 (d, $J = 8.0$ Hz, 2H), 2.43 (s, 3H), 2.33 (s, 3H), 1.84 (s, 3H); ^{13}C NMR(100 MHz, CDCl_3), δ 163.6, 154.3, 141.9, 137.4, 137.3, 129.8, 129.0, 128.4, 128.3, 126.8, 1008.9, 27.0, 21.6, 21.1.



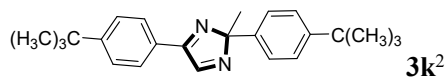
Yield 67%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.42 (s, 1H), 8.00 (d, $J = 8.0$ Hz, 2H), 7.74 (d, $J = 8.0$ Hz, 2H), 7.00 (d, $J = 8.0$ Hz, 2H), 6.87 (d, $J = 8.0$ Hz, 2H), 3.85 (s, 3H), 3.77 (s, 3H), 1.81 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3), δ 162.9, 162.0, 158.9, 154.1, 132.4, 129.9, 127.9, 123.7, 114.3, 113.5, 108.4, 55.3, 55.1, 26.9. HRMS (ESI): calcd for $\text{C}_{18}\text{H}_{18}\text{N}_2\text{O}_2$: $[\text{M}+\text{H}^+]$ 295.1441, found 295.1447.



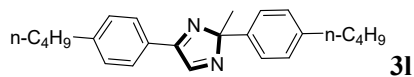
Yield 72%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.41 (s, 1H), 8.00 (d, $J = 8.4$ Hz, 2H), 7.61 (d, $J = 8.8$ Hz, 2H), 7.00 (d, $J = 8.8$ Hz, 2H), 6.86 (d, $J = 8.4$ Hz, 2H), 4.10-4.05 (dd, $J_1 = 6.8$ Hz, $J_2 = 14$ Hz, 2H), 4.02-3.97 (dd, $J_1 = 6.8$ Hz, $J_2 = 14$ Hz, 2H), 1.81 (s, 3H), 1.43 (t, $J = 7.2$ Hz, 3H), 1.38 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3), δ 162.9, 161.4, 158.2, 154.1, 132.2, 129.9, 127.9, 123.5, 114.8, 114.1, 108.4, 63.5, 63.3, 26.9, 14.7, 14.6. HRMS (ESI): calcd for $\text{C}_{20}\text{H}_{22}\text{N}_2\text{O}_2$: $[\text{M}+\text{H}^+]$ 323.1754, found 323.1757.



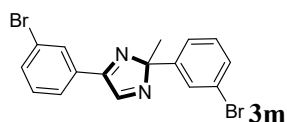
Yield 73%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.46 (s, 1H), 8.1 (d, $J = 8.0$ Hz, 2H), 7.77 (d, $J = 8.0$ Hz, 2H), 7.37 (d, $J = 8.0$ Hz, 2H), 7.20 (d, $J = 8.04$ Hz, 2H), 1.82 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3), δ 162.7, 154.1, 151.5, 148.7, 138.7, 130.1, 129.4, 128.5, 121.7 (d, $J_{\text{F-C}} = 265$ Hz), 121.6 (d, $J_{\text{F-C}} = 247$ Hz), 119.1, 119.1, 108.9, 27.2. $^{19}\text{F NMR}$ (377 MHz, CDCl_3), δ -57.7, -57.9. HRMS (ESI): calcd for $\text{C}_{18}\text{H}_{12}\text{F}_6\text{N}_2\text{O}_2$: $[\text{M}+\text{H}^+]$ 403.0876, found 403.0881.



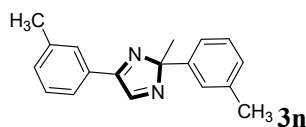
Yield 60%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.47 (s, 1H), 8.01 (d, $J = 8.0$ Hz, 2H), 7.65 (d, $J = 8.0$ Hz, 2H), 7.54 (d, $J = 8.0$ Hz, 2H), 7.38 (d, $J = 8.0$, 2H), 1.86 (s, 3H), 1.38 (s, 9H), 1.32 (s, 9H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3), δ 163.5, 154.9, 154.3, 150.4, 137.1, 128.3, 128.1, 126.5, 125.9, 125.2, 108.8, 34.9, 34.4, 31.3, 31.1, 26.8.



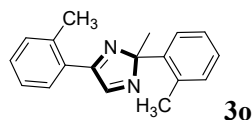
Yield 53%. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.45 (s, 1H), 7.97 (d, $J = 8.0$ Hz, 2H), 7.61 (d, $J = 7.6$ Hz, 2H), 7.32 (d, $J = 7.6$ Hz, 2H), 7.16 (d, $J = 7.6$, 2H), 2.69 (t, $J = 7.6$, 2H), 2.59 (t, $J = 7.6$, 2H), 1.84 (s, 3H), 1.68-1.54 (m, 4H), 1.42-1.30 (m, 4H), 0.97-0.90 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3), δ 163.5, 154.3, 146.8, 142.3, 137.3, 129.1, 128.5, 128.3, 128.3, 126.7, 108.9, 35.5, 35.2, 33.5, 33.3, 26.9, 22.3, 22.2, 13.88, 13.87. HRMS (ESI): calcd for $\text{C}_{24}\text{H}_{30}\text{N}_2$: $[\text{M}+\text{H}^+]$ 347.2482, found 347.2488.



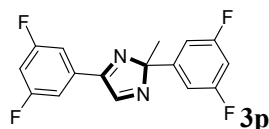
Yield 62%. ¹H NMR(400 MHz, CDCl₃) δ 8.44 (s, 1H), 8.22 (s, 1H), 7.95 (d, *J* = 7.6 Hz, 2H), 7.86 (s, 1H), 7.67 (t, *J* = 7.2 Hz, 2H), 7.42-7.36 (m, 2H), 7.23 (t, *J* = 8.0 Hz, 2H), 1.80 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 162.7, 154.2, 141.9, 134.5, 132.7, 131.2, 130.8, 130.5, 130.1, 129.9, 126.8, 125.7, 123.3, 122.3, 108.7, 27.1. HRMS (ESI): calcd for C₁₆H₁₂Br₂N₂: [M+H⁺] 390.9440, found 390.9445.



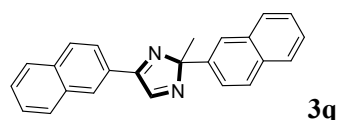
Yield 80%. ¹H NMR(400 MHz, CDCl₃) δ 8.51 (s, 1H), 7.95 (s, 1H), 7.87 (d, *J* = 7.2 Hz, 1H), 7.57 (d, *J* = 8.4 Hz, 2H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.39 (d, *J* = 7.6 Hz, 1H), 7.30 (d, *J* = 6.8 Hz, 1H), 7.15 (d, *J* = 7.6 Hz, 1H), 2.48 (s, 3H), 2.40 (s, 3H), 1.89 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 163.9, 154.4, 139.9, 138.9, 137.9, 132.2, 130.9, 128.8, 128.8, 128.4, 128.2, 127.5, 125.4, 124.0, 123.9, 26.9, 21.4, 21.3. HRMS (ESI): calcd for C₁₈H₁₈N₂: [M+H⁺] 263.1543, found 263.1547.



Yield 94%. ¹H NMR(400 MHz, CDCl₃) δ 8.31 (s, 1H), 7.78 (d, *J* = 7.6 Hz, 1H), 7.68 (d, *J* = 7.2 Hz, 1H), 7.36-7.27 (m, 3H), 7.22-7.16 (m, 2H), 7.13 (t, *J* = 7.2 Hz, 1H), 2.88 (s, 3H), 2.61 (s, 3H), 1.83 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 163.9, 155.6, 138.9, 138.6, 137.1, 132.3, 131.7, 130.7, 130.4, 129.6, 128.4, 127.8, 126.1, 125.8, 110.3, 25.5, 22.2, 21.7. HRMS (ESI): calcd for C₁₈H₁₈N₂: [M+H⁺] 263.1543, found 263.1547

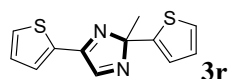


Yield 65%. ¹H NMR(400 MHz, CDCl₃) δ 8.41 (s, 1H), 7.59 (d, *J* = 5.6 Hz, 2H), 7.27 (d, *J* = 6.0 Hz, 2H), 7.03 (t, *J* = 8.8 Hz, 1H), 6.77 (t, *J* = 8.8 Hz, 1H), 1.80 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 164.6 (d, *J*_{F-C} = 12.3 Hz), 164.0 (d, *J*_{F-C} = 185.0 Hz), 163.9 (d, *J*_{F-C} = 184.7 Hz), 162.4 (t, *J*_{F-C} = 3.3 Hz), 161.2 (d, *J*_{F-C} = 12.6 Hz), 154.1, 143.0 (t, *J*_{F-C} = 9.3 Hz), 133.6 (t, *J*_{F-C} = 9.3 Hz), 111.5 (d, *J*_{F-C} = 26.5 Hz), 111.4 (d, *J*_{F-C} = 11.3 Hz), 110.5 (d, *J*_{F-C} = 26.4 Hz), 110.4 (d, *J*_{F-C} = 11.9 Hz), 108.7, 107.1 (t, *J*_{F-C} = 25.1 Hz), 103.2 (t, *J*_{F-C} = 25.1 Hz), 27.1; ¹⁹F NMR(470 MHz, CDCl₃), δ -107.6, -109.3. HRMS (ESI): calcd for C₁₆H₁₀F₄N₂: [M+H⁺] 307.0853, found 307.0859.

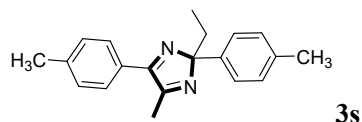


Yield 60%. ¹H NMR(400 MHz, CDCl₃) δ 9.58 (d, *J* = 8.8 Hz, 1H), 8.99 (d, *J* = 8.0 Hz, 1H), 8.56 (s, 1H), 8.03-7.89 (m, 5H), 7.84 (d, *J* = 8.0 Hz, 1H), 7.70-7.53 (m, 5H), 7.44 (t, *J* = 7.6 Hz, 1H), 2.18 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 163.4, 155.99, 137.0, 134.63, 133.99, 131.69, 131.59, 131.3, 129.0, 128.8, 128.7, 128.3, 127.8, 127.6, 126.6, 125.9, 125.8, 125.7, 125.4, 125.2, 124.9,

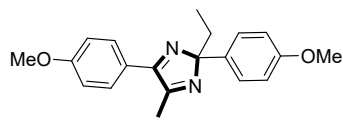
110.7, 26.5. HRMS (ESI): calcd for C₂₄H₁₈N₂: [M+H⁺] 335.1543, found 335.1547



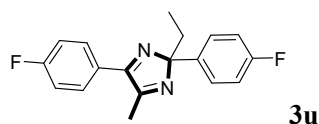
Yield 35%. ¹H NMR(400 MHz, CDCl₃) δ 8.32 (s, 1H), 7.71 (d, *J* = 7.6 Hz, 1H), 7.58 (d, *J* = 6.8 Hz, 1H), 7.22-7.17 (m, 3H), 6.98 (t, *J* = 4.0 Hz, 1H), 1.91 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 158.6, 154.4, 141.0, 134.6, 131.0, 130.6, 128.0, 126.7, 125.0, 124.8, 106.9, 27.1. HRMS (ESI): calcd for C₁₂H₁₀N₂S₂: [M+H⁺] 247.0358, found 247.0362.



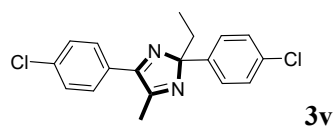
Yield 87%. ¹H NMR(400 MHz, CDCl₃) δ 7.71 (d, *J* = 8.0 Hz, 2H), 7.62 (d, *J* = 8.0 Hz, 2H), 7.27 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 7.6 Hz, 2H), 2.51 (s, 3H), 2.41 (s, 3H), 2.32 (s, 3H), 2.24-2.20 (m, 2H), 0.75 (t, *J* = 7.2 Hz, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 165.6, 163.9, 140.7, 137.6, 137.0, 129.4, 129.3, 128.8, 128.4, 127.5, 106.9, 33.9, 21.4, 21.0, 17.5, 8.5. HRMS (ESI): calcd for C₂₀H₂₂N₂: [M+H⁺] 291.1856, found 291.1860



Yield 62%. ¹H NMR(400 MHz, CDCl₃) δ 7.82 (d, *J* = 8.8 Hz, 2H), 7.67 (d, *J* = 8.8 Hz, 2H), 6.99 (d, *J* = 8.8 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 3.85 (s, 3H), 3.78 (s, 3H), 2.52 (s, 3H), 2.23-2.15 (m, 2H), 0.76 (t, *J* = 7.2 Hz, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 164.9, 163.7, 161.3, 158.7, 132.8, 130.0, 128.6, 124.7, 114.0, 113.4, 106.0, 55.3, 55.1, 33.9, 17.7, 8.5. HRMS (ESI): calcd for C₂₀H₂₂N₂O₂: [M+H⁺] 323.1754, found 323.1758.

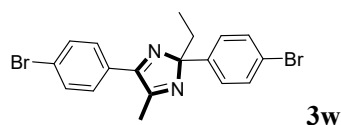


Yield 85%. ¹H NMR(400 MHz, CDCl₃) δ 7.83 (t, *J* = 7.2 Hz, 2H), 7.71 (d, *J* = 7.2 Hz, 2H), 7.16 (t, *J* = 8.4 Hz, 2H), 7.01 (t, *J* = 8.4 Hz, 2H), 2.52 (s, 3H), 2.20-2.13 (m, 2H), 0.74 (t, *J* = 7.2 Hz, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 165.4 (d, *J*_{F-C} = 211.3 Hz), 164.7 (d, *J*_{F-C} = 104.1 Hz), 162.9, 160.8, 136.1, 130.5 (d, *J*_{F-C} = 8.6 Hz), 129.3 (d, *J*_{F-C} = 7.9 Hz), 128.3 (d, *J*_{F-C} = 3.3 Hz), 115.9 (d, *J*_{F-C} = 21.7 Hz), 114.9 (d, *J*_{F-C} = 21.1 Hz), 106.4, 34.1, 17.5, 8.5; ¹⁹F NMR(377 MHz, CDCl₃), δ -109.3, -115.1; HRMS (ESI): calcd for C₁₈H₁₆F₂N₂: [M+H⁺] 299.1354, found 299.1357.

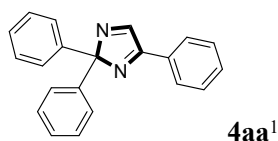


Yield 83%. ¹H NMR(400 MHz, CDCl₃) δ 7.68 (d, *J* = 8.4 Hz, 2H), 7.59 (d, *J* = 8.4 Hz, 2H), 7.37 (d, *J* = 8.4 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 2H), 2.42 (s, 3H), 2.11-2.06 (m, 2H), 0.65 (t, *J* = 7.2 Hz, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 164.8, 163.7, 138.6, 136.8, 133.3, 130.5, 129.7, 129.0, 129.0,

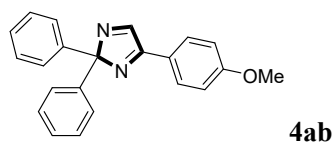
128.2, 106.6, 34.1, 17.5, 8.5. HRMS (ESI): calcd for C₁₈H₁₆Cl₂N₂: [M+H⁺] 331.0763, found 331.0769.



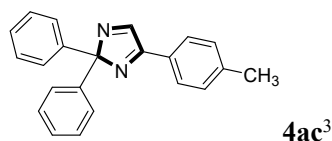
Yield 80%. ¹H NMR(400 MHz, CDCl₃) δ 7.70 (d, *J* = 8.4 Hz, 2H), 7.63-7.59 (m, 4H), 7.46 (d, *J* = 8.4 Hz, 2H), 2.51 (s, 3H), 2.17-2.14 (m, 2H), 0.74 (t, *J* = 7.6 Hz, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 164.9, 163.6, 139.1, 131.9, 131.1, 130.9, 129.9, 129.4, 125.2, 121.5, 106.6, 34.0, 17.4, 8.5. HRMS (ESI): calcd for C₁₈H₁₆Br₂N₂: [M+H⁺] 418.9752, found 417.9758.



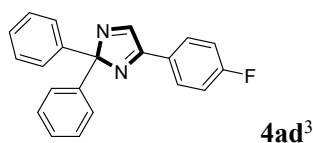
Yield 77%. ¹H NMR(400 MHz, CDCl₃) δ 8.54 (s, 1H), 8.06 (d, *J* = 4.8 Hz, 2H), 7.69 (d, *J* = 6.4 Hz, 4H), 7.45 (s, 3H), 7.28-7.21 (m, 6H); ¹³C NMR(100 MHz, CDCl₃), δ 164.3, 163.3, 155.1, 148.7, 140.9, 133.6, 131.6, 131.5, 131.0, 129.6, 129.1, 128.6, 128.4, 127.8, 126.5, 112.7.



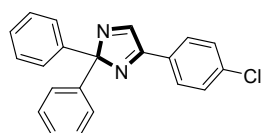
Yield 60%. ¹H NMR(400 MHz, CDCl₃) δ 8.53 (s, 1H), 8.04 (d, *J* = 8.8 Hz, 2H), 7.67 (d, *J* = 7.2 Hz, 4H), 7.30-7.20 (m, 6H), 6.98 (d, *J* = 8.8 Hz, 2H), 3.82 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 163.5, 162.2, 155.0, 141.0, 130.2, 128.2, 127.6, 127.6, 123.6, 114.4, 112.3, 55.3. HRMS (ESI): calcd for C₂₂H₁₈N₂O: [M+H⁺] 327.1491, found 327.1495.



Yield 72%. ¹H NMR(400 MHz, CDCl₃) δ 8.54 (s, 1H), 7.98 (d, *J* = 7.6 Hz, 2H), 7.68 (d, *J* = 7.2 Hz, 4H), 7.30-7.26 (m, 6H), 7.23 (t, *J* = 7.2 Hz, 2H), 2.38 (s, 3H); ¹³C NMR(100 MHz, CDCl₃), δ 164.2, 155.2, 142.1, 141.0, 129.8, 128.6, 128.3, 127.8, 127.7, 112.6, 21.7.

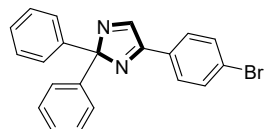


Yield 73%. ¹H NMR(400 MHz, CDCl₃) δ 8.42 (s, 1H), 7.98-7.95 (m, 2H), 7.58 (d, *J* = 7.2 Hz, 4H), 7.20-7.10 (m, 6H), 7.2307 (t, *J* = 8.4 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 166.1 (t, *J*_{F-C} = 251.3 Hz), 163.1, 154.7, 140.8, 130.8, 130.7, 128.4, 127.8, 127.7, 116.3 (d, *J*_{F-C} = 27.5 Hz), 112.8.



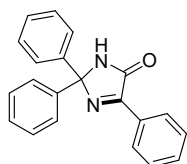
4ae³

Yield 71%. ¹H NMR(400 MHz, CDCl₃) δ 8.38 (s, 1H), 7.87 (d, *J* = 8.4 Hz, 2H), 7.57 (d, *J* = 7.2 Hz, 4H), 7.32 (d, *J* = 8.8 Hz, 2H), 7.17 (t, *J* = 7.2 Hz, 4H), 7.10 (t, *J* = 7.2 Hz, 2H); ¹³C NMR(100 MHz, CDCl₃), δ 163.1, 154.5, 140.6, 137.7, 129.7, 129.3, 128.3, 127.7, 127.6, 112.7.



4af³

Yield 70%. ¹H NMR(400 MHz, CDCl₃) δ 8.50 (s, 1H), 7.92 (d, *J* = 8.4 Hz, 2H), 7.67 (d, *J* = 7.2 Hz, 4H), 7.60 (d, *J* = 8.4 Hz, 2H), 7.28 (t, *J* = 7.6 Hz, 4H), 7.21 (t, *J* = 7.2 Hz, 2H); ¹³C NMR(100 MHz, CDCl₃), δ 163.2, 154.4, 140.5, 132.2, 129.9, 129.7, 128.3, 127.8, 127.6, 112.8.



5aa³

Yield 60%, ¹H NMR (500 MHz, CDCl₃) δ 9.83 (s, 1H), 8.54 (d, *J* = 7.0 Hz, 2H), 7.56-7.53 (m, 1H), 7.50-7.49 (m, 6H), 7.36-7.31 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 166.2, 160.9, 141.3, 131.9, 130.3, 128.8, 128.6, 128.6, 128.3, 126.8, 87.5.

Reference

1. Z. Zhu, X. Tang, J. Li, X. Li, W. Wu, G. Deng and H. Jiang, *Org. Lett.*, 2017, **19**, 1370-1373;
2. F. Geng, S. Wu, X. Gan, W. Hou, J. Dong and Y. Zhou, *Org. Biomol. Chem.*, 2022, **20**, 5416-5422;
3. Z. Zhu, H. Lin, B. Liang, J. Huang, W. Liang, L. Chen, Y. Huang, X. Chen and Y. Li, *Chem. Commun.*, 2020, **56**, 5621-5624

Copies of ^1H NMR and ^{13}C NMR spectroscopies

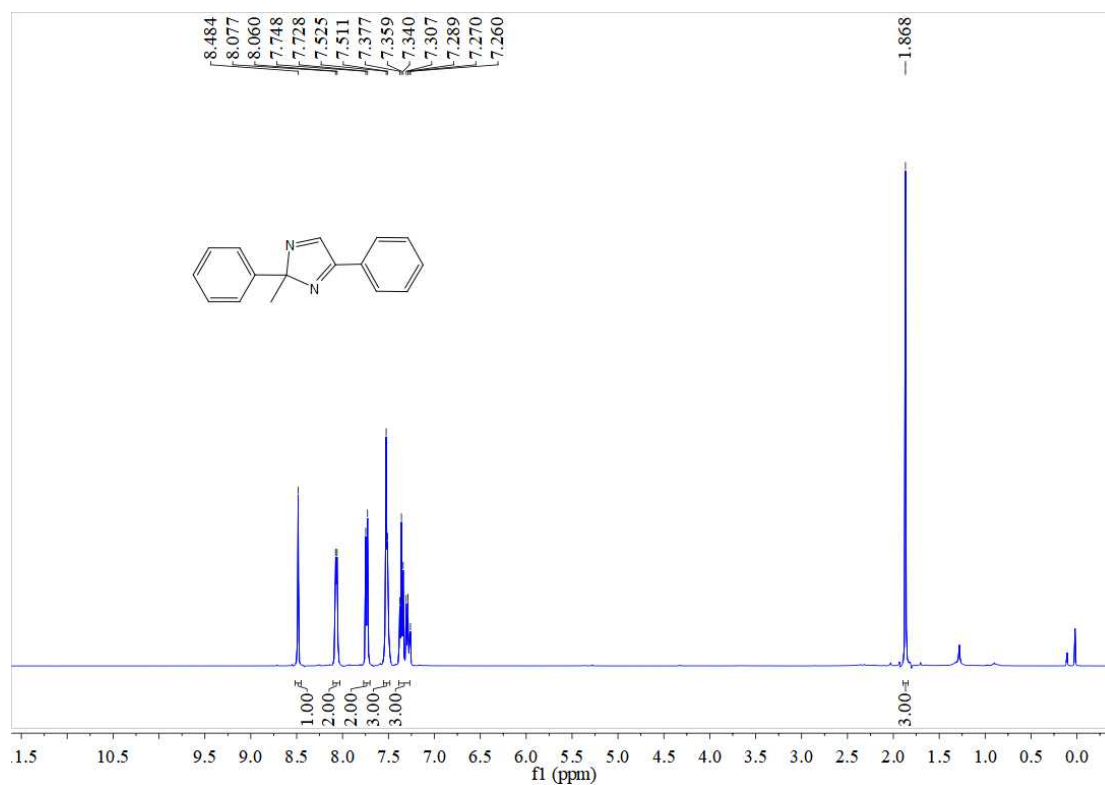


Figure S1 ^1H NMR of **3a**

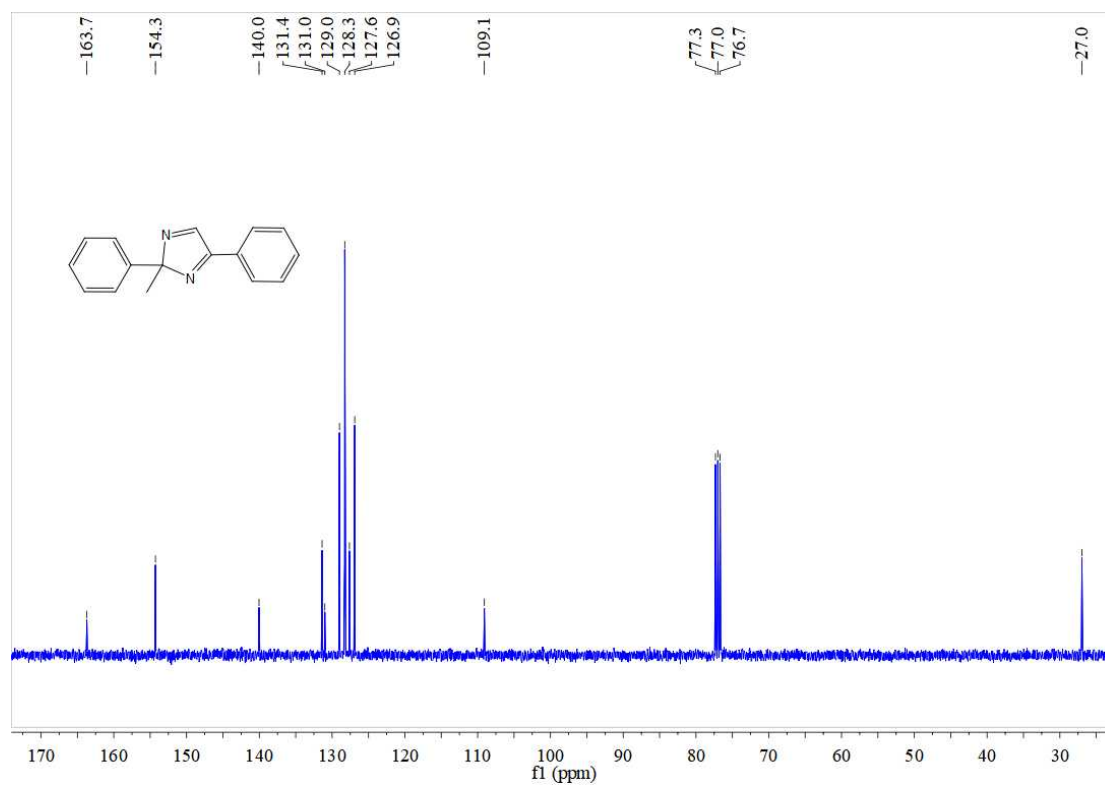


Figure S2 ^{13}C NMR of **3a**

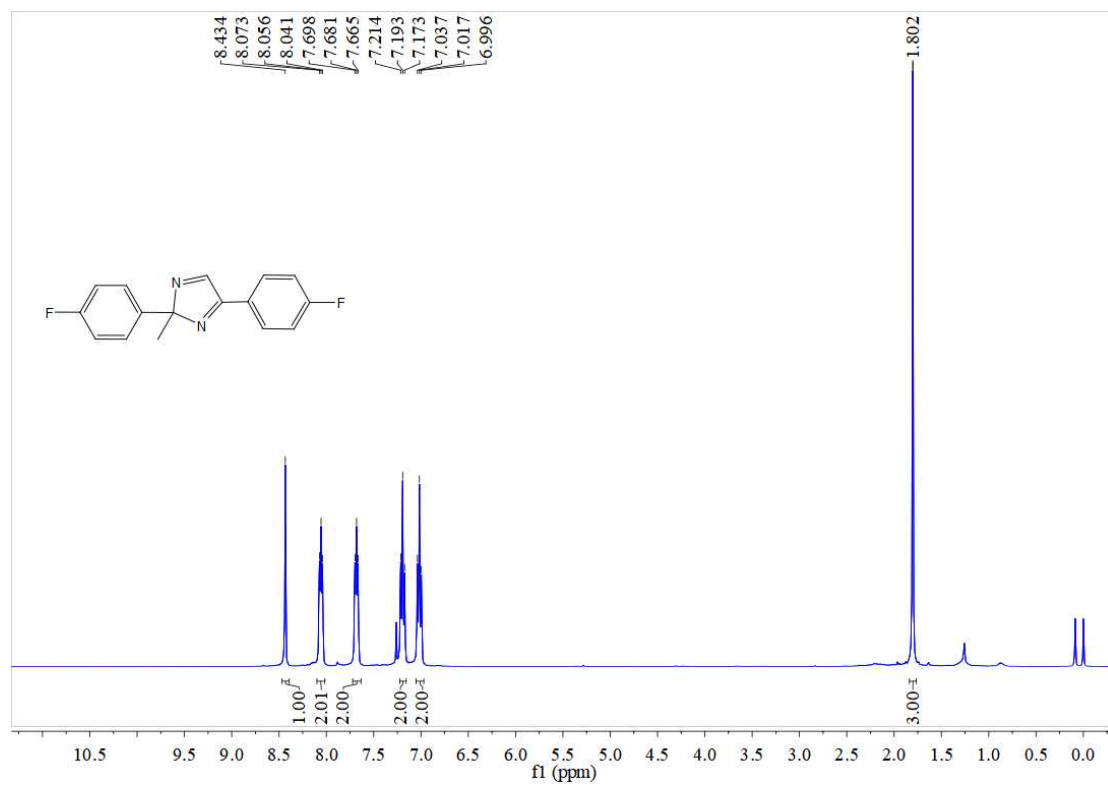


Figure S3 ¹H NMR of **3b**

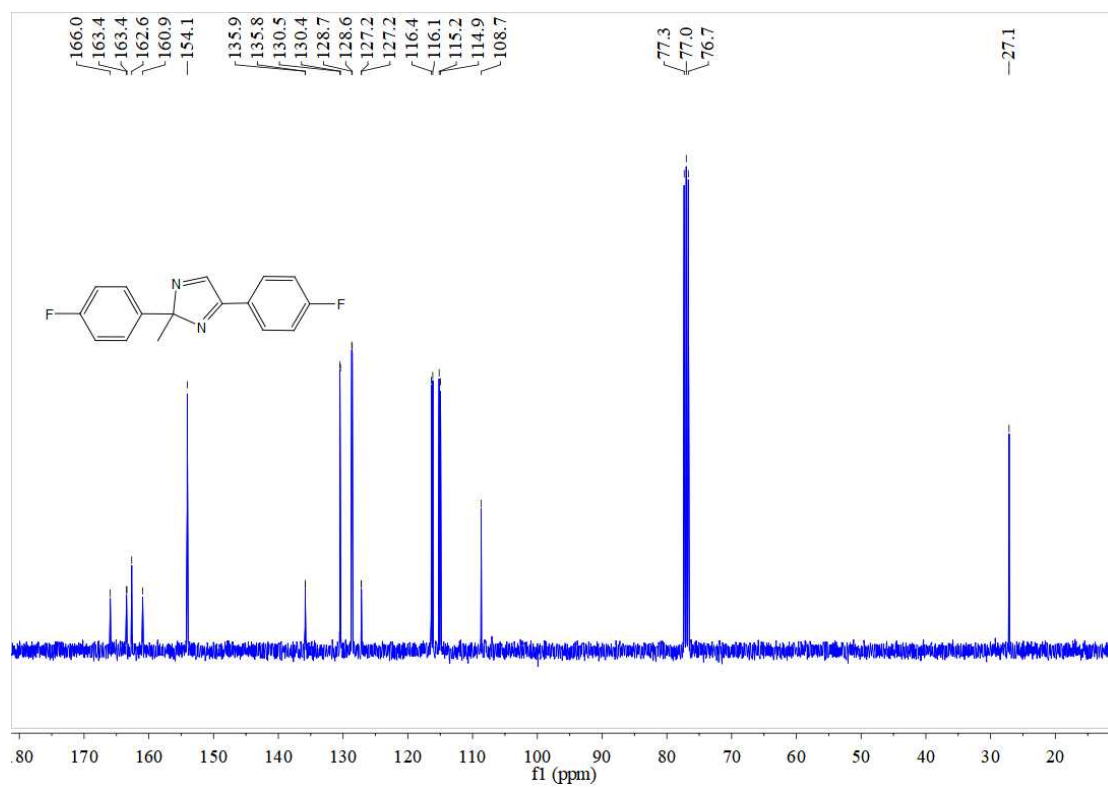


Figure S4 ¹³C NMR of **3b**

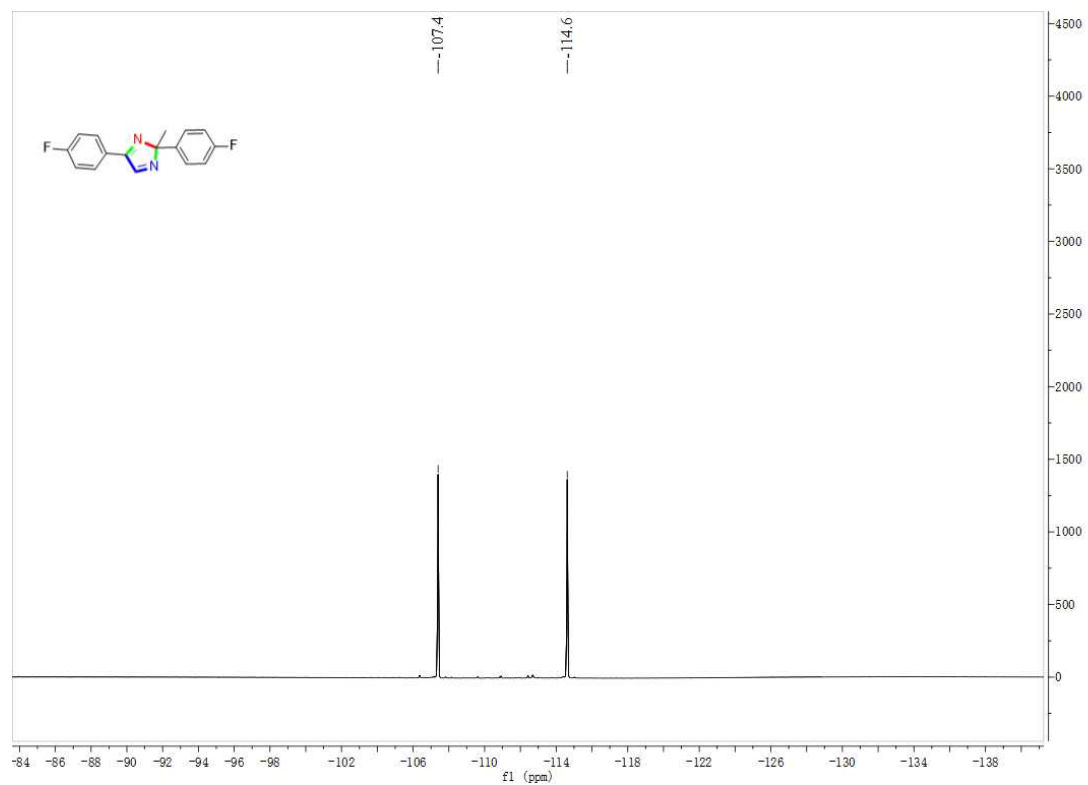


Figure S5 ^{19}F NMR of **3b**

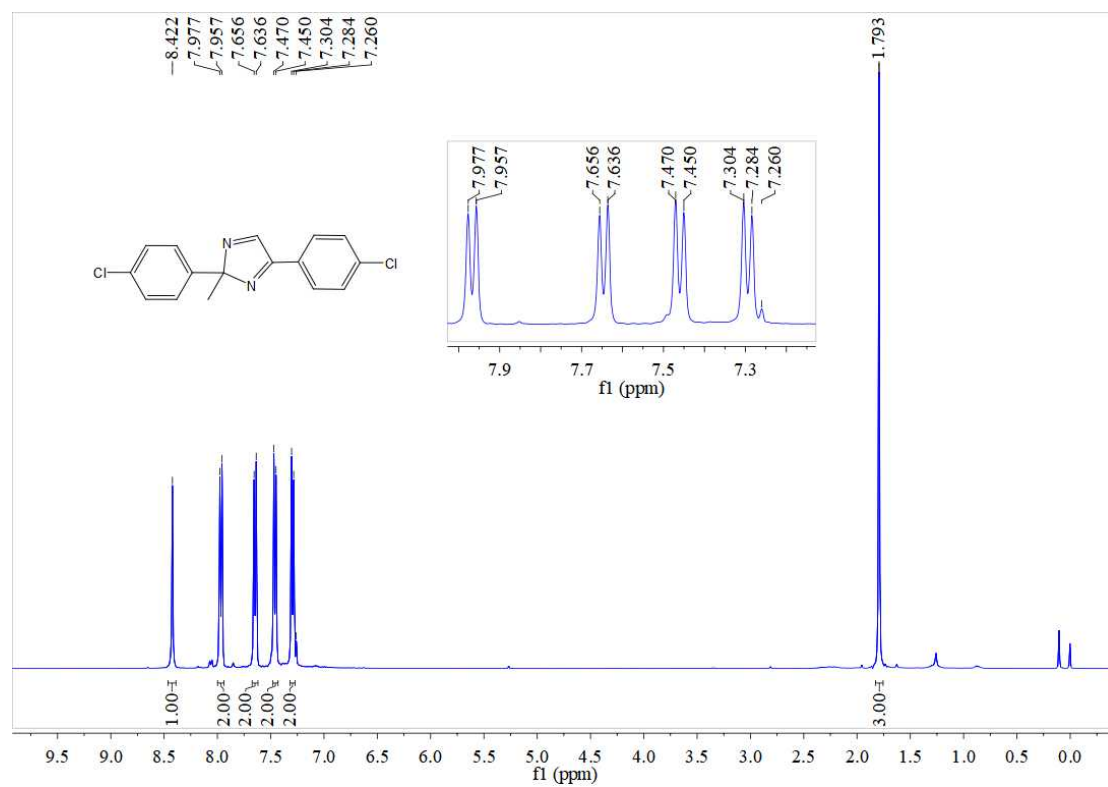


Figure S6 ^1H NMR of **3c**

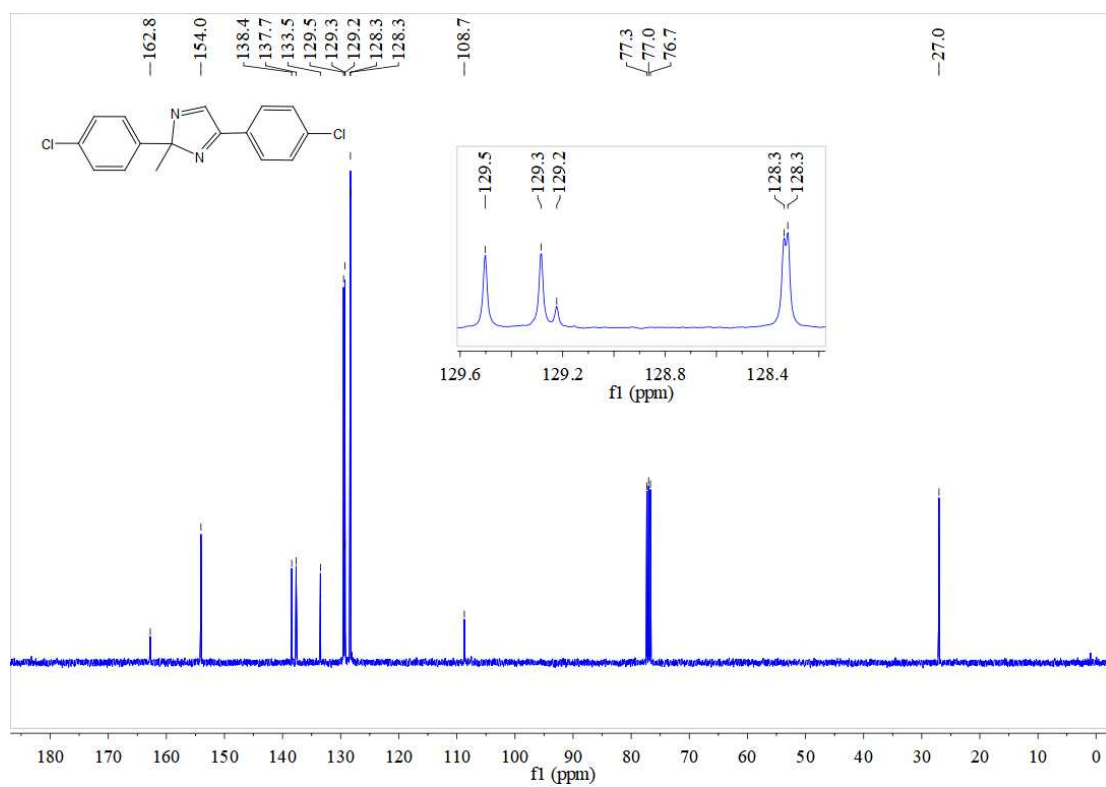


Figure S7 ¹³C NMR of **3c**

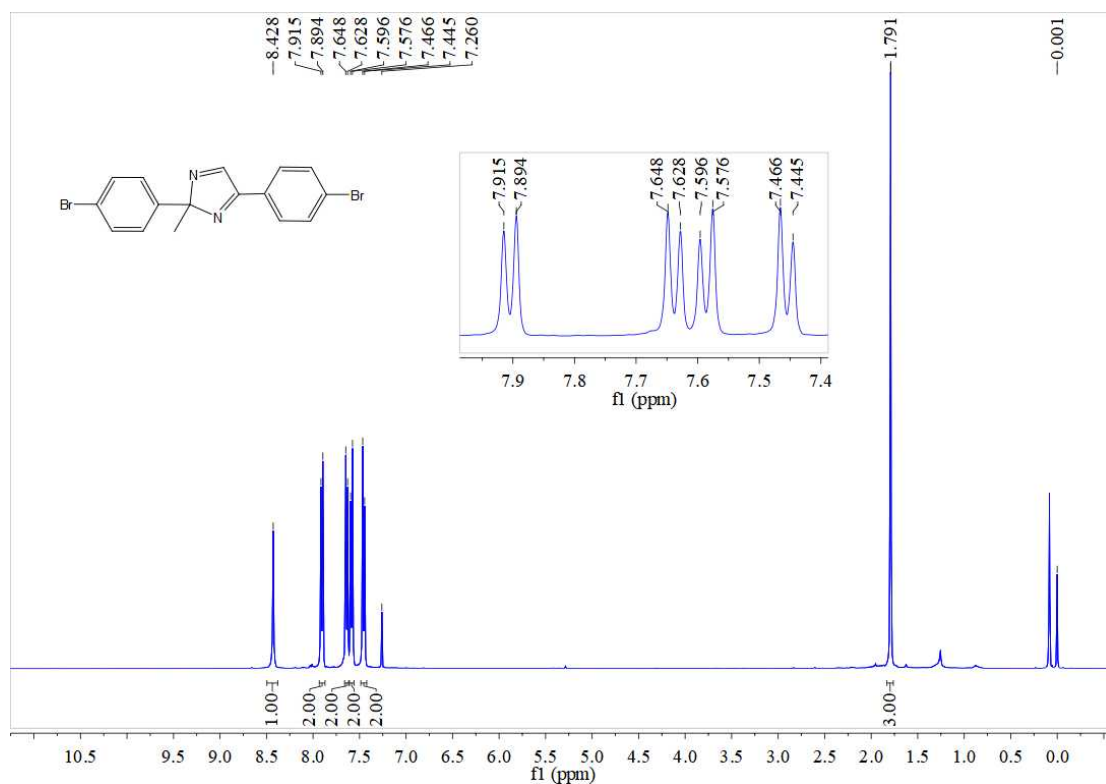


Figure S8 ¹H NMR of **3d**

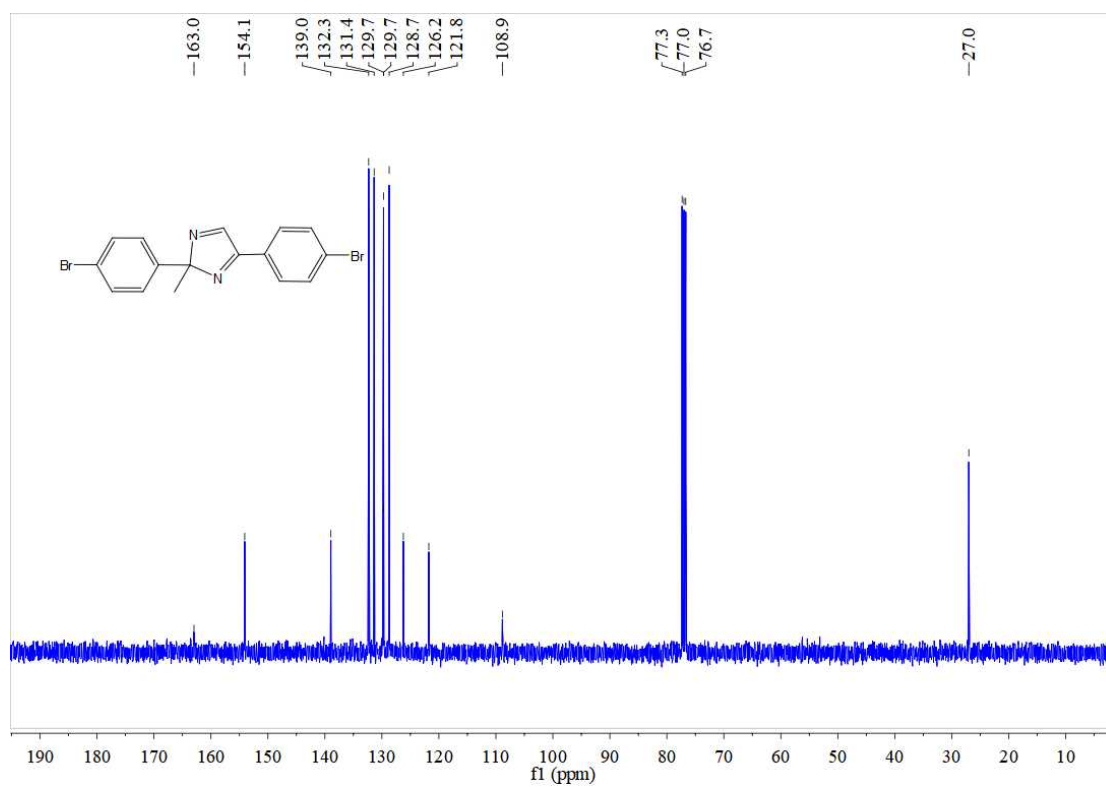


Figure S9 ¹³C NMR of **3d**

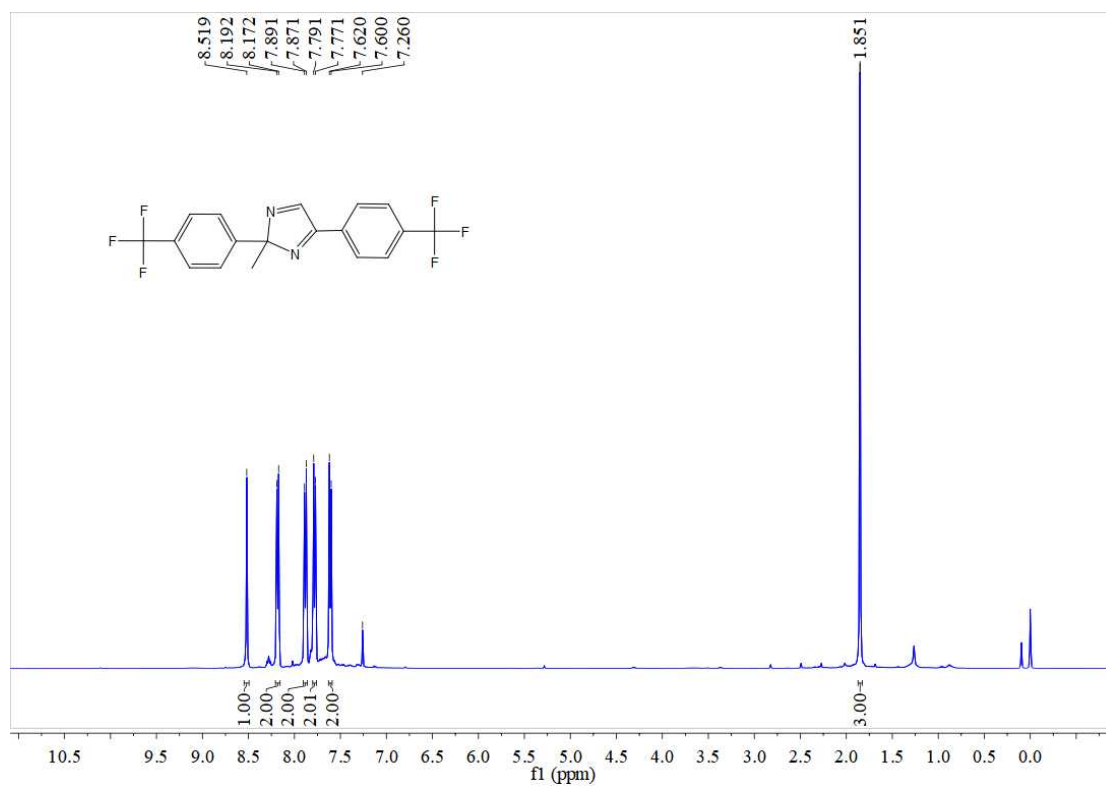


Figure S10 ¹H NMR of **3e**

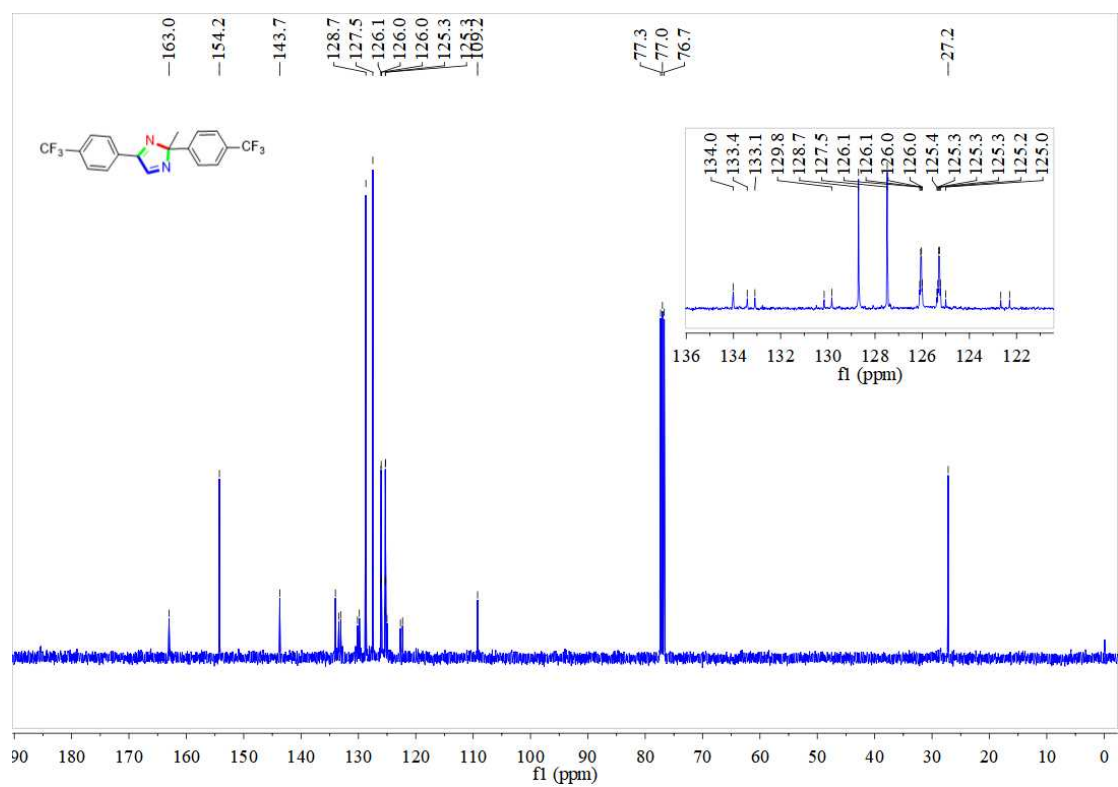


Figure S12 ¹³C NMR of **3e**

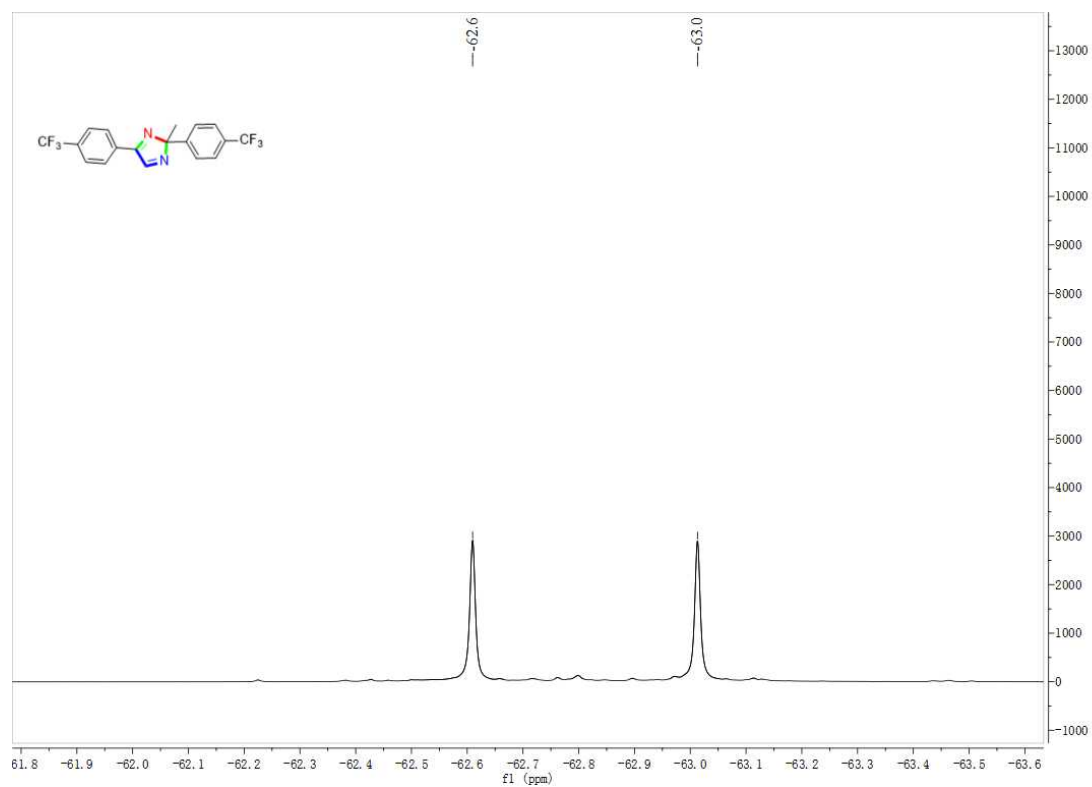


Figure S13 ¹⁹F NMR of **3e**

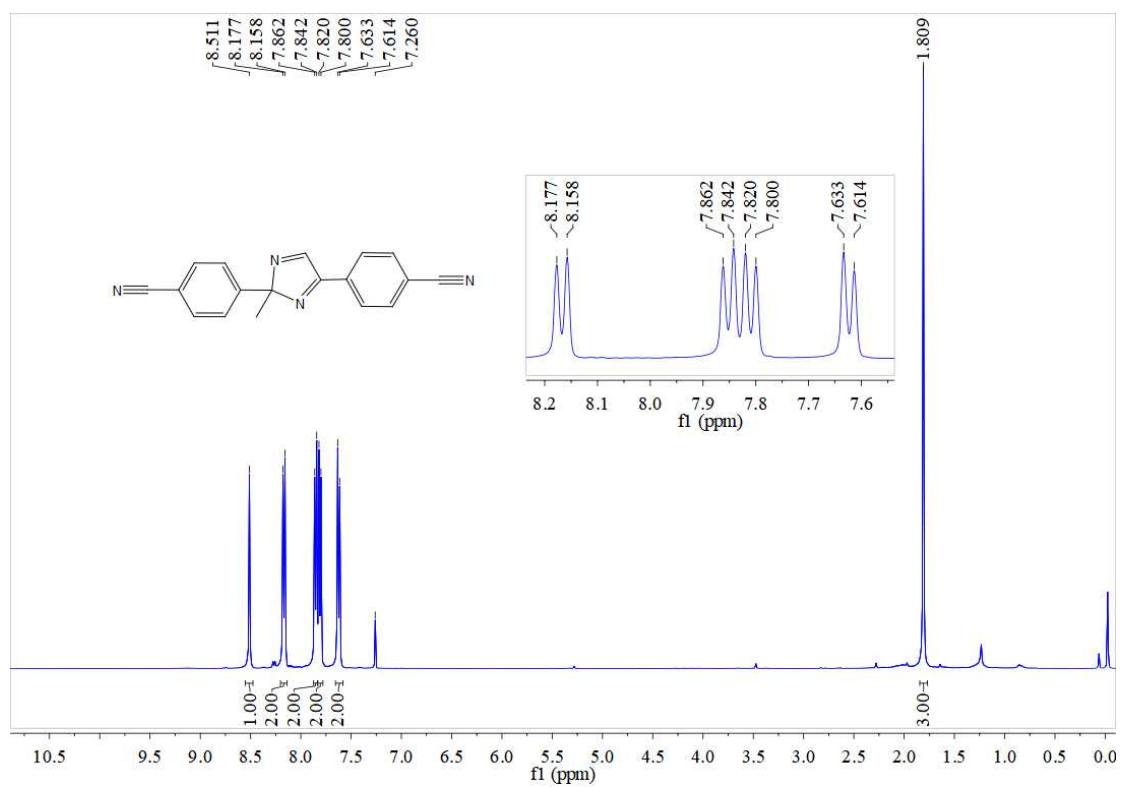


Figure S14 ¹H NMR of 3f

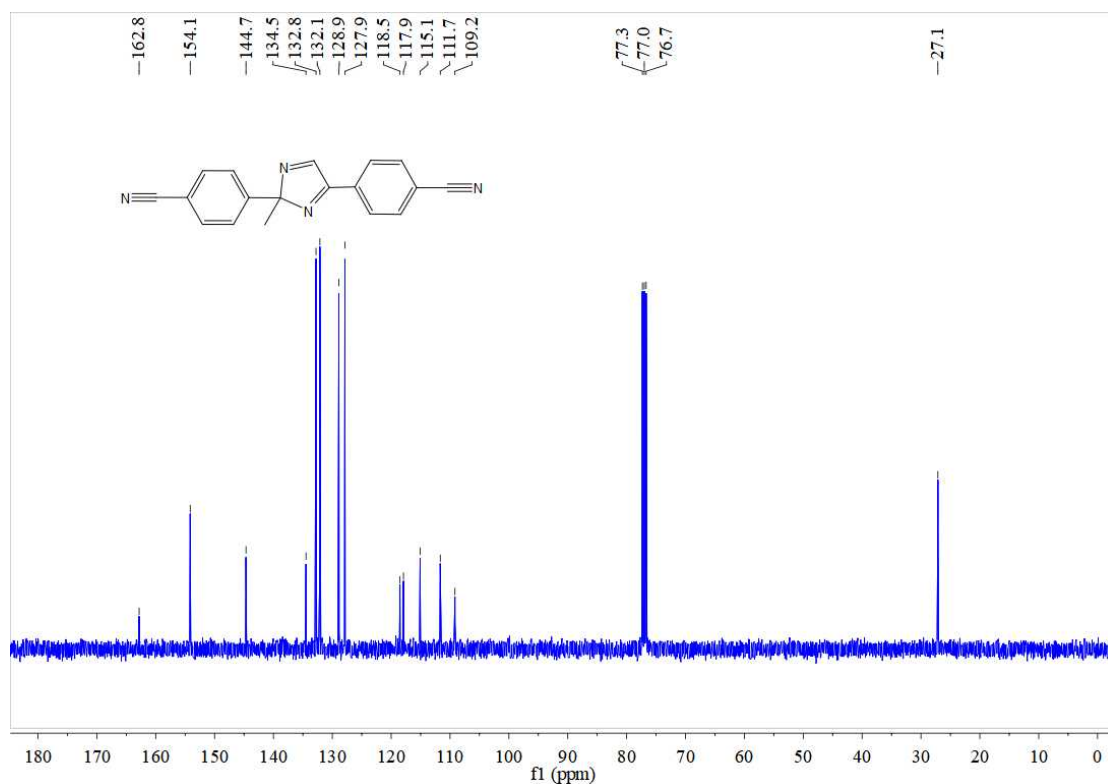


Figure S15 ¹³C NMR of 3f

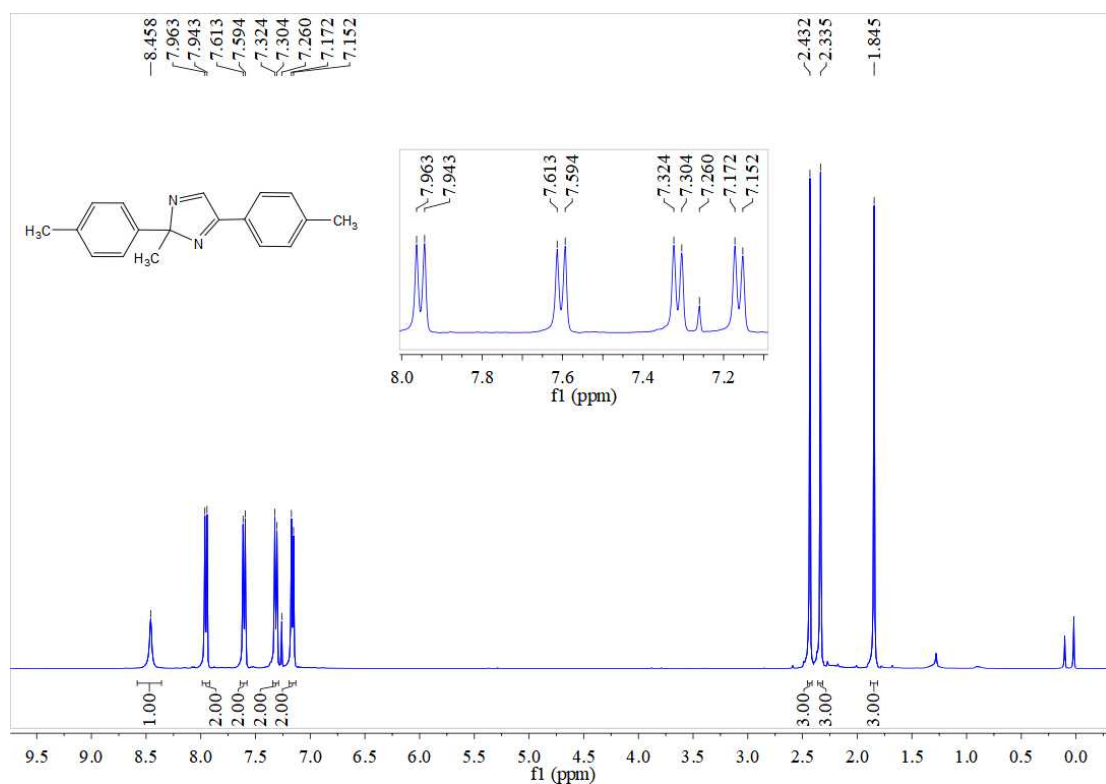


Figure S16 ¹H NMR of 3g

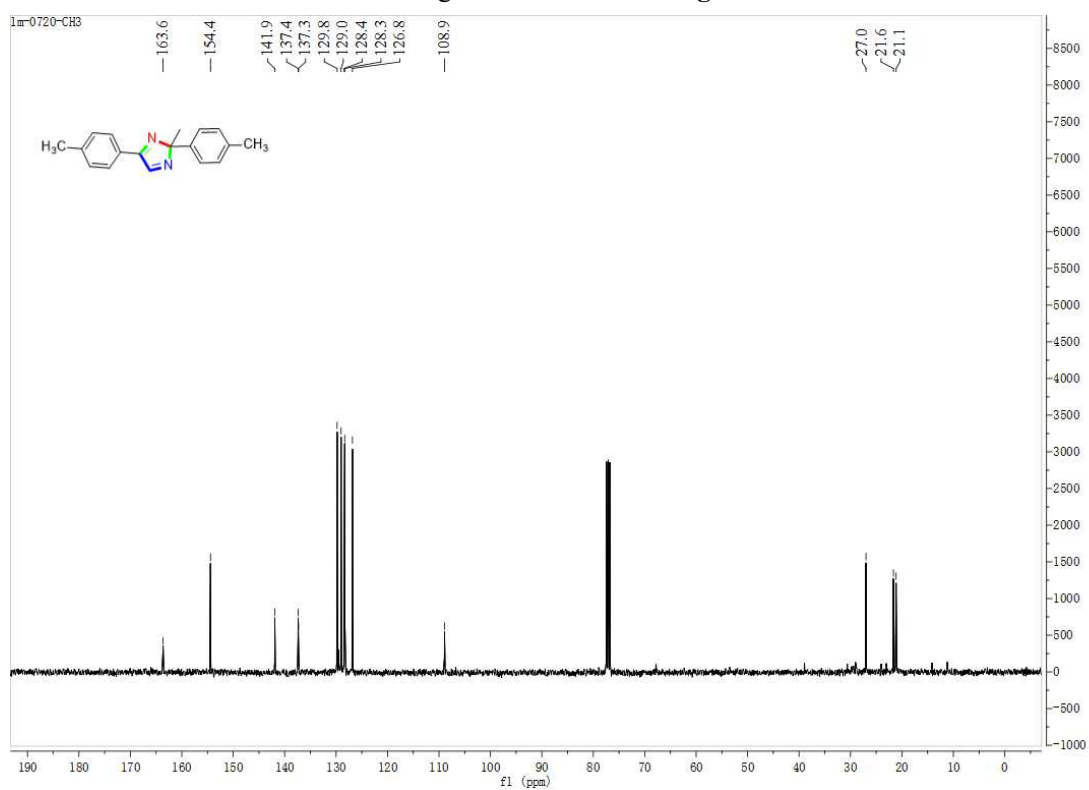


Figure S17 ¹³C NMR of 3g

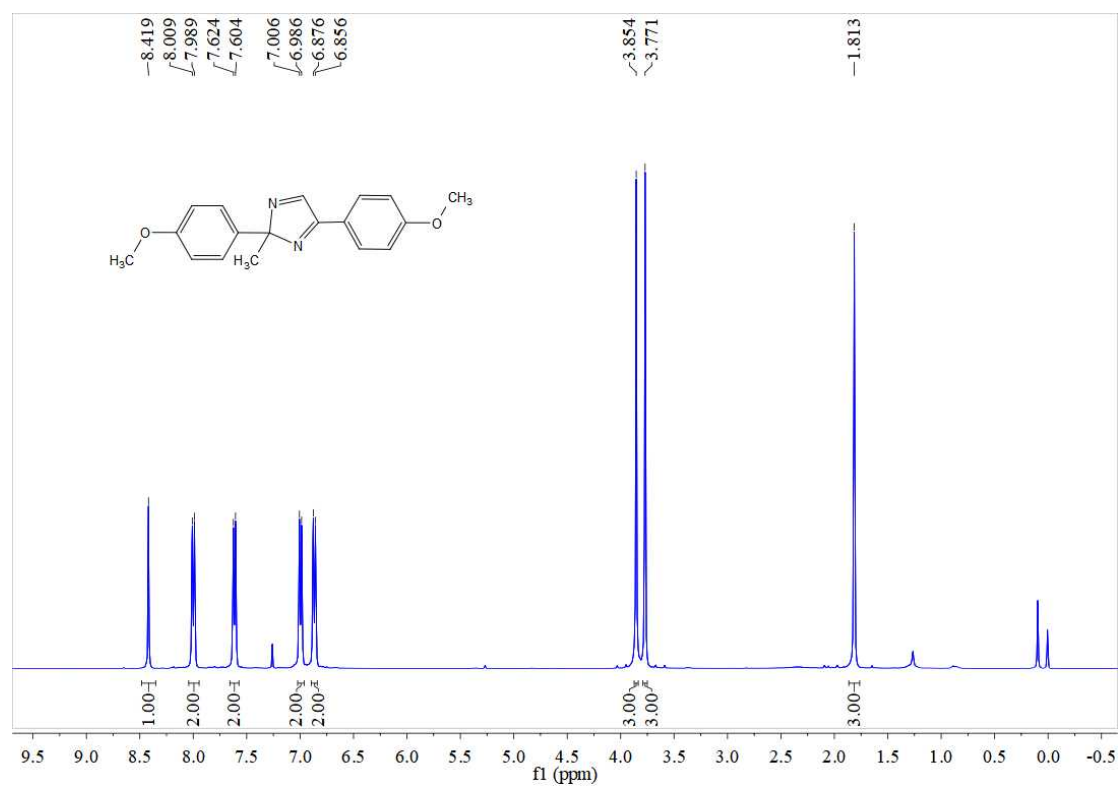


Figure S18 ¹H NMR of 3h

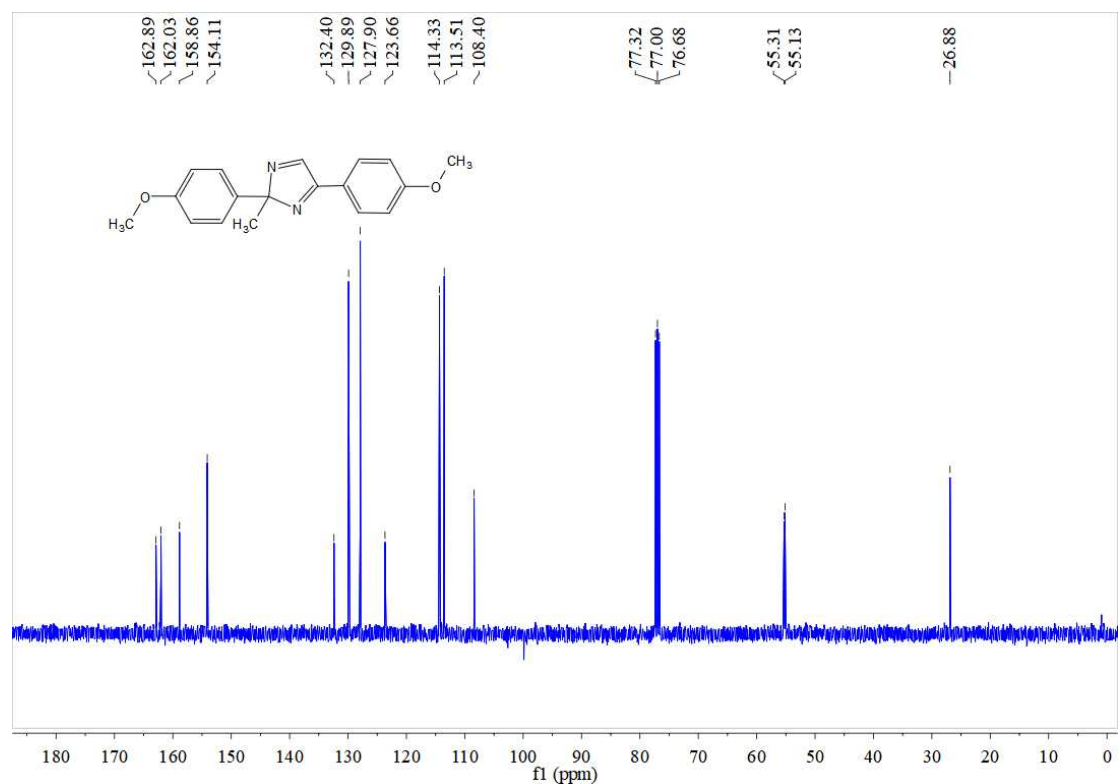


Figure S19 ¹³C NMR of 3h

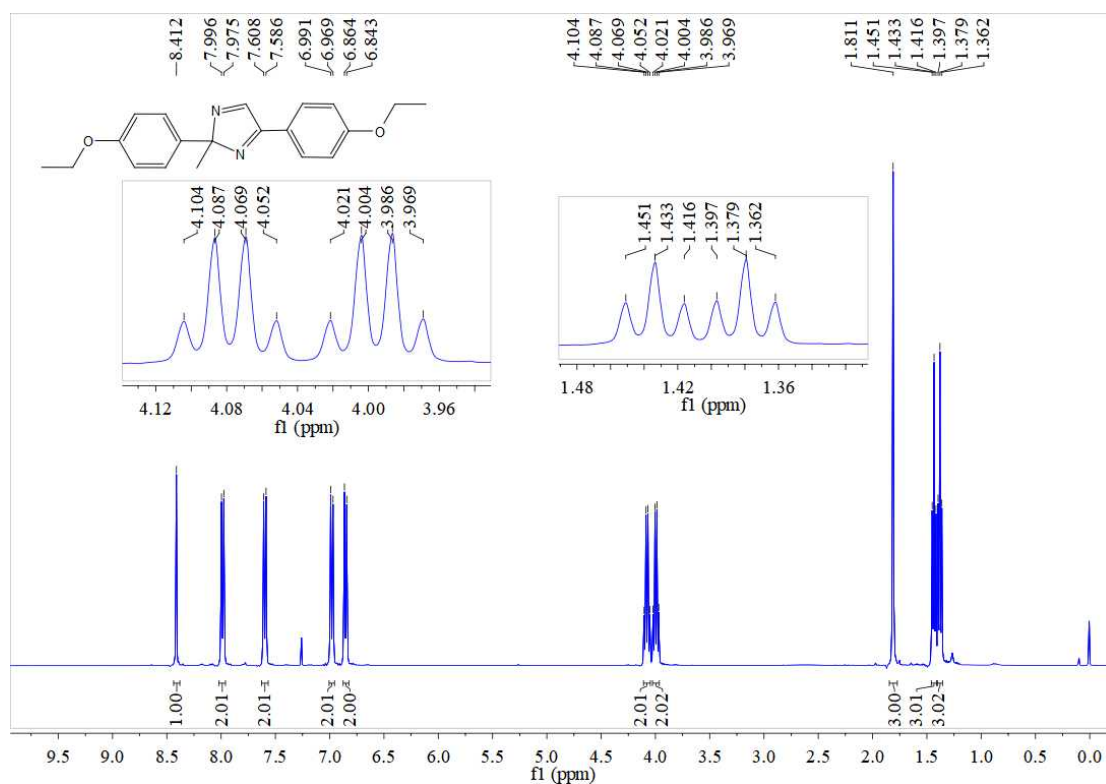


Figure S20 ¹H NMR of **3i**

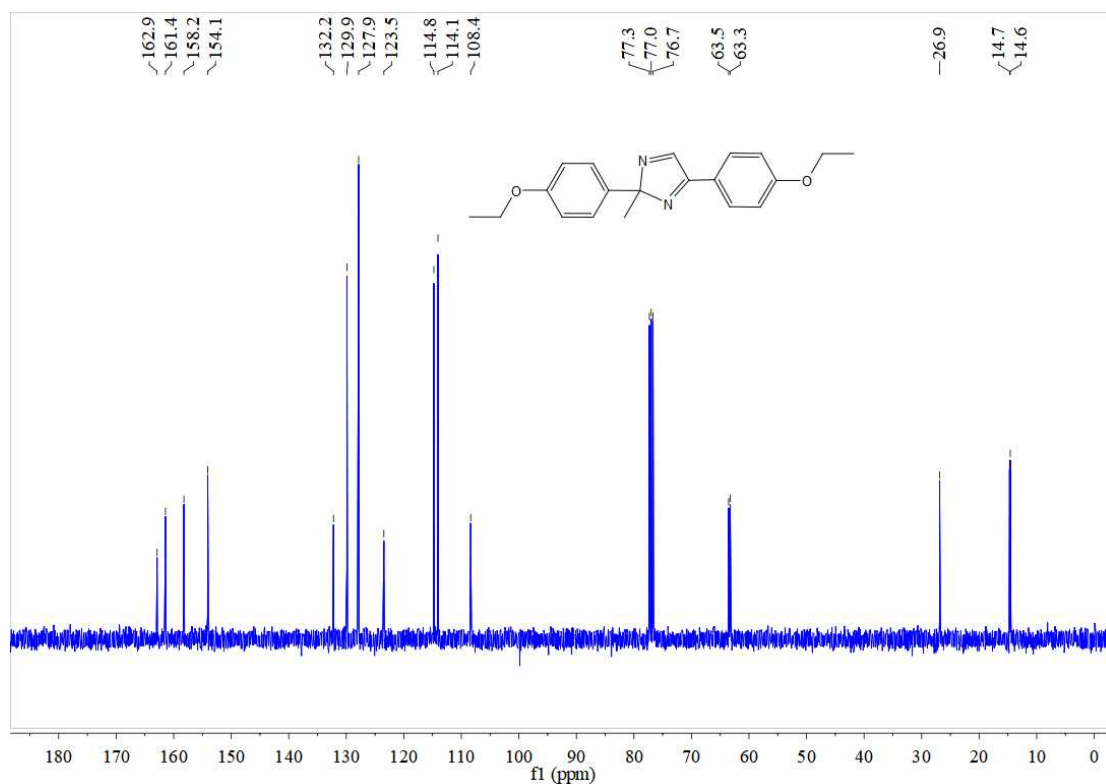


Figure S21 ¹³C NMR of **3i**

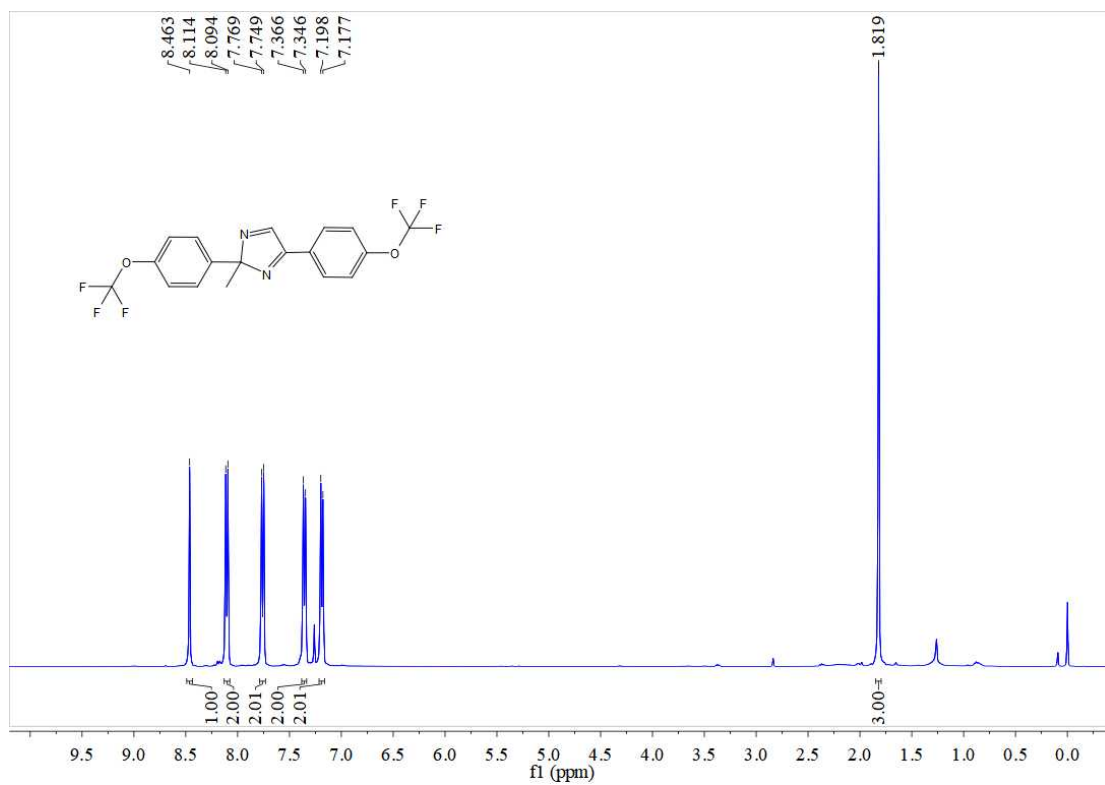


Figure S22 ¹H NMR of **3j**

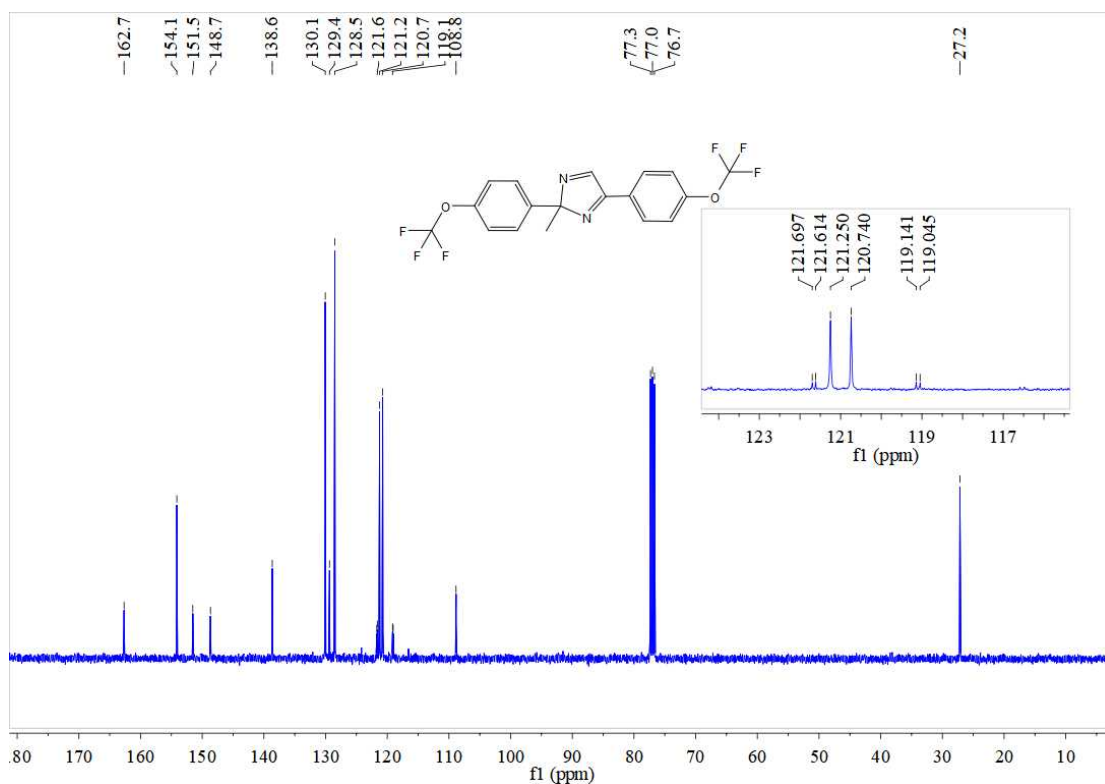


Figure S23 ¹³C NMR of **3j**

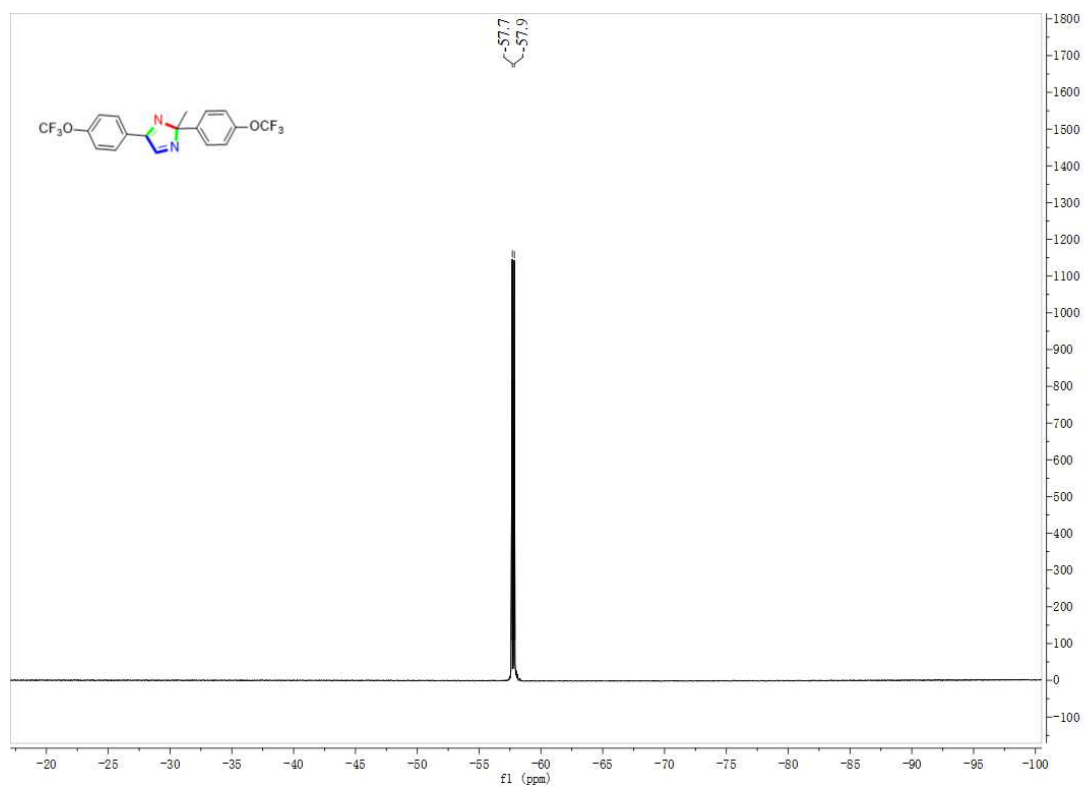


Figure S24 ^{19}F NMR of **3j**

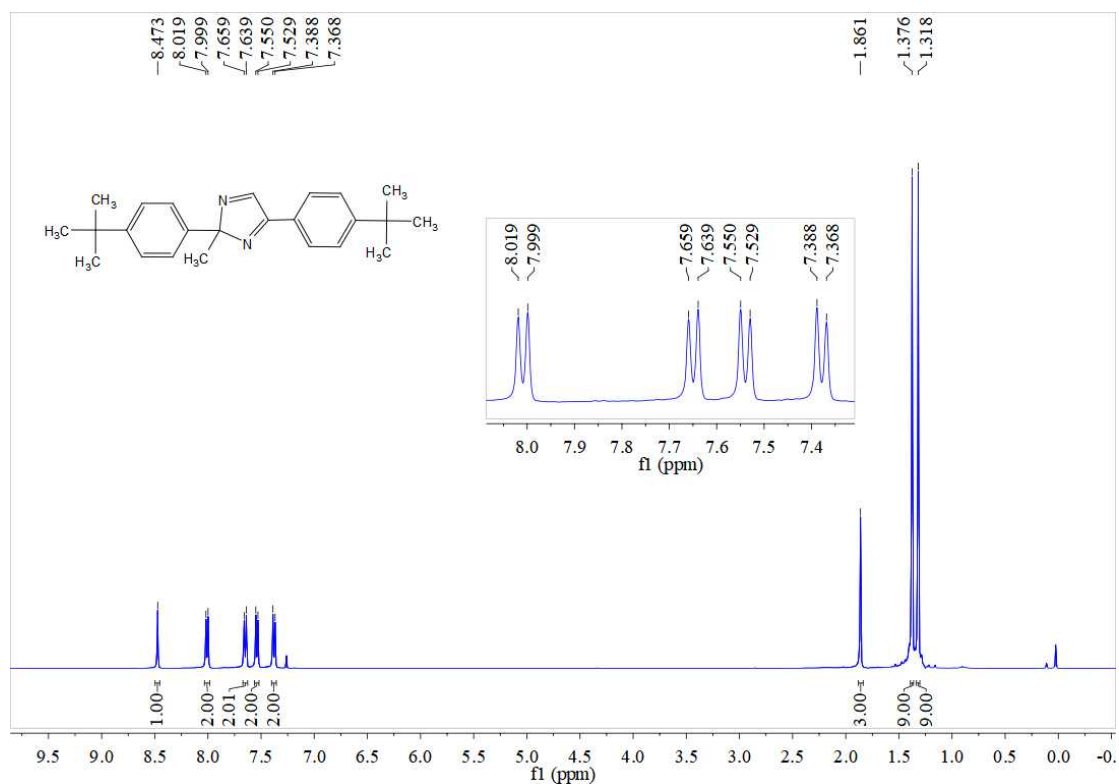


Figure S25 ^1H NMR of **3k**

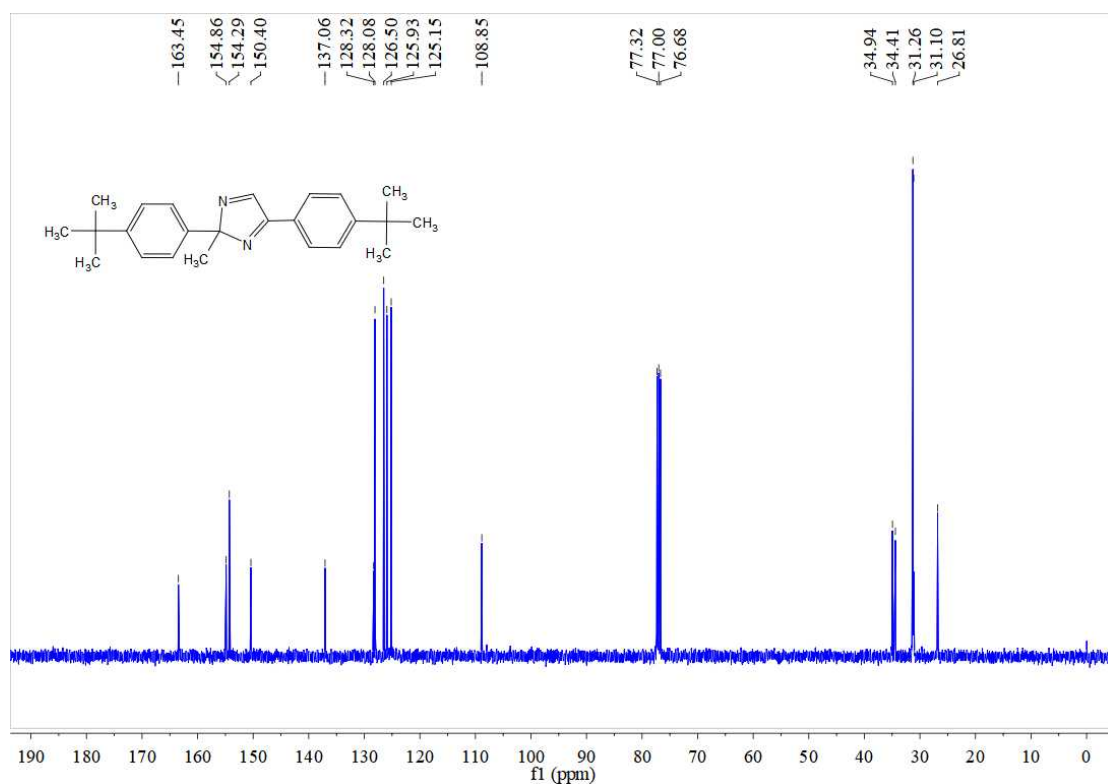


Figure S26 ¹³C NMR of **3k**

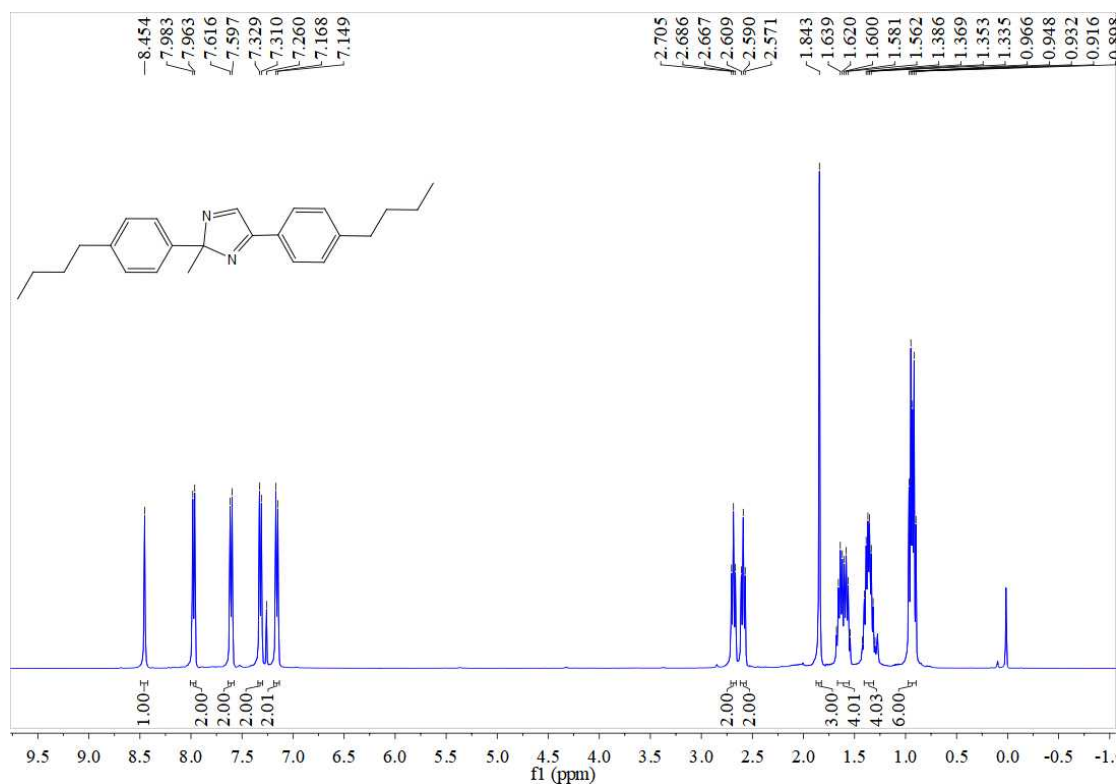


Figure S27 ¹H NMR of **3l**

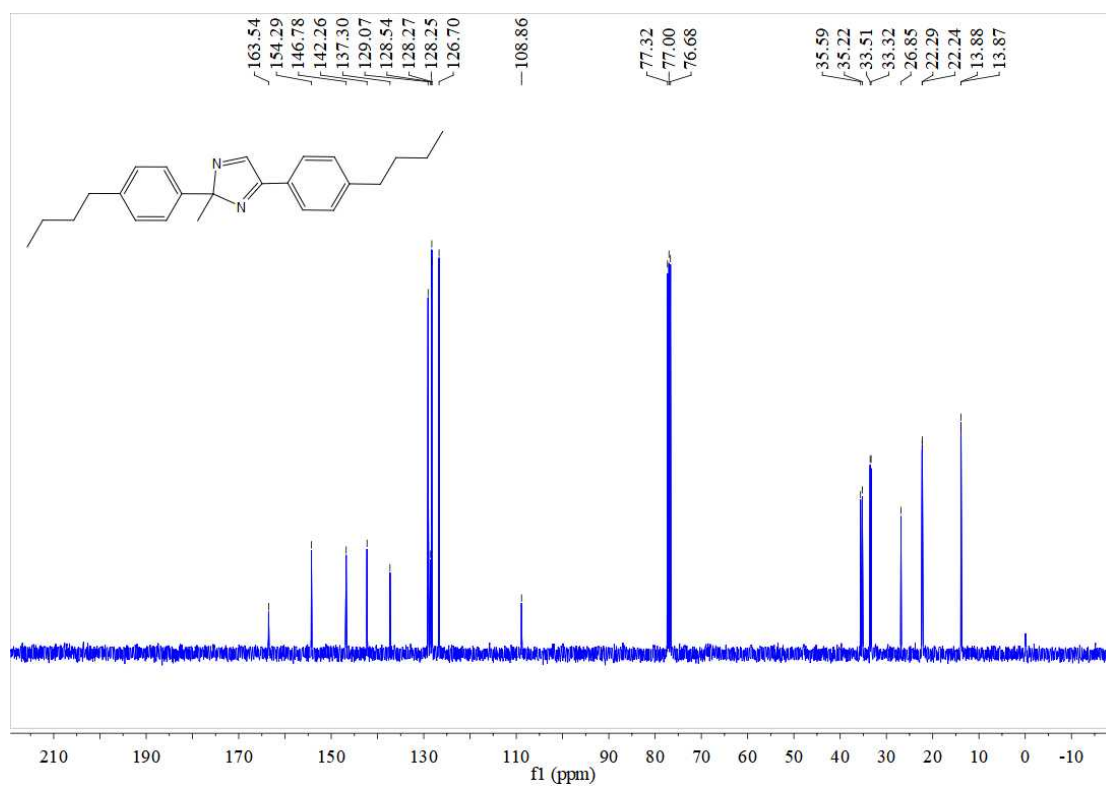


Figure S28 ¹³C NMR of **31**

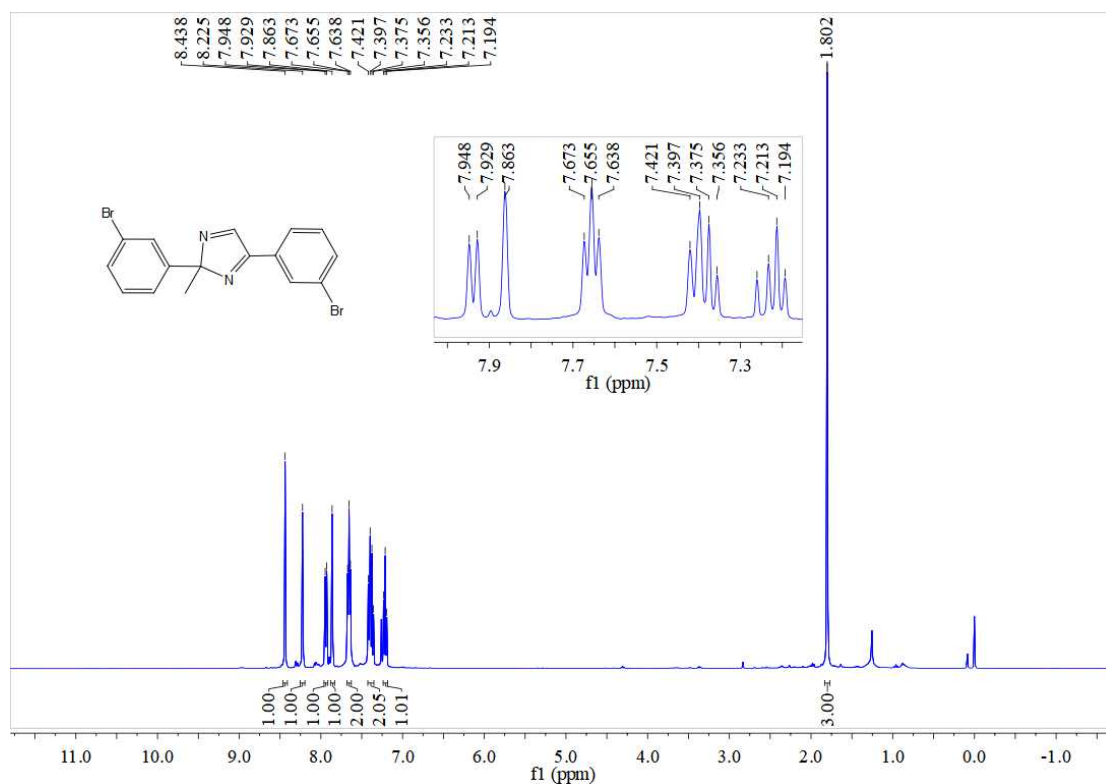


Figure S29 ¹H NMR of **3m**

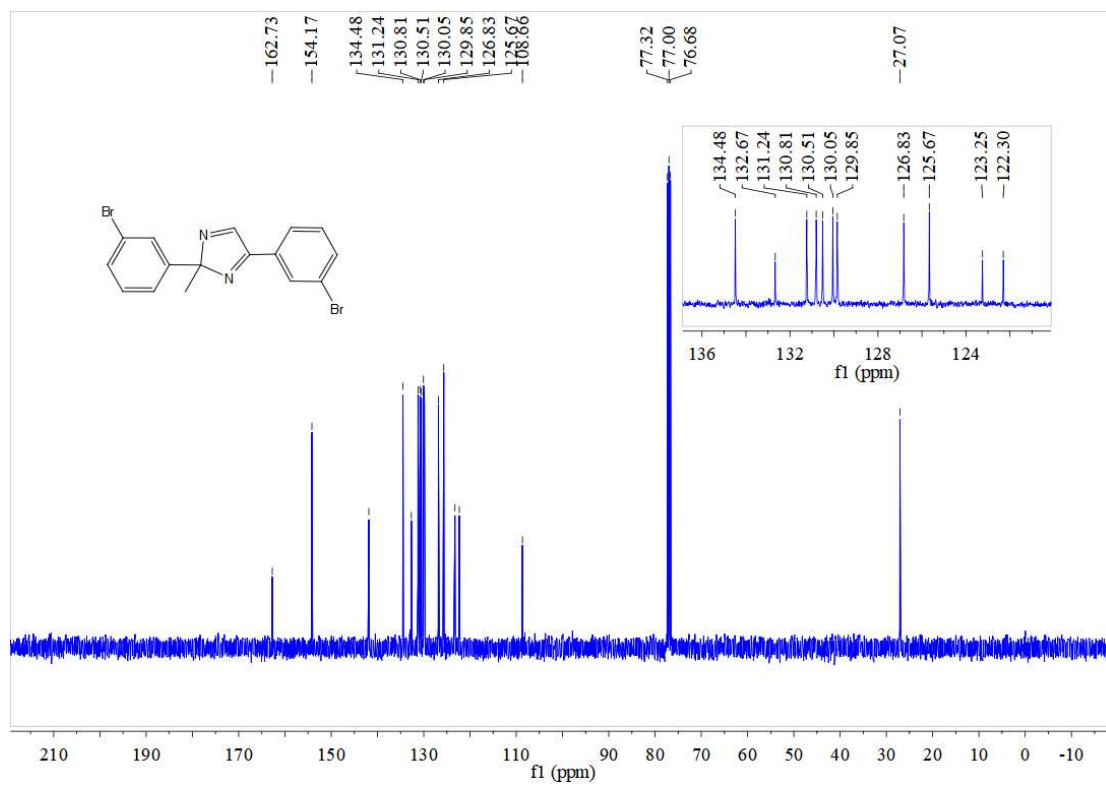


Figure S30 ¹³C NMR of **3m**

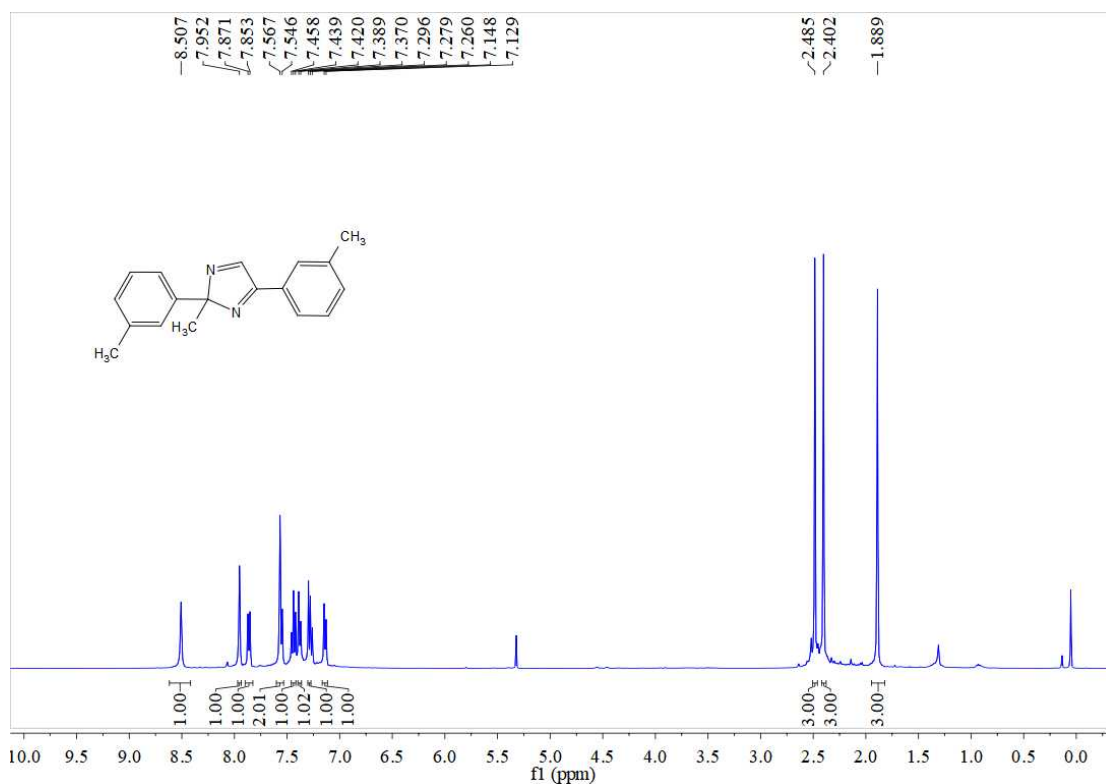


Figure S31 ¹H NMR of **3n**

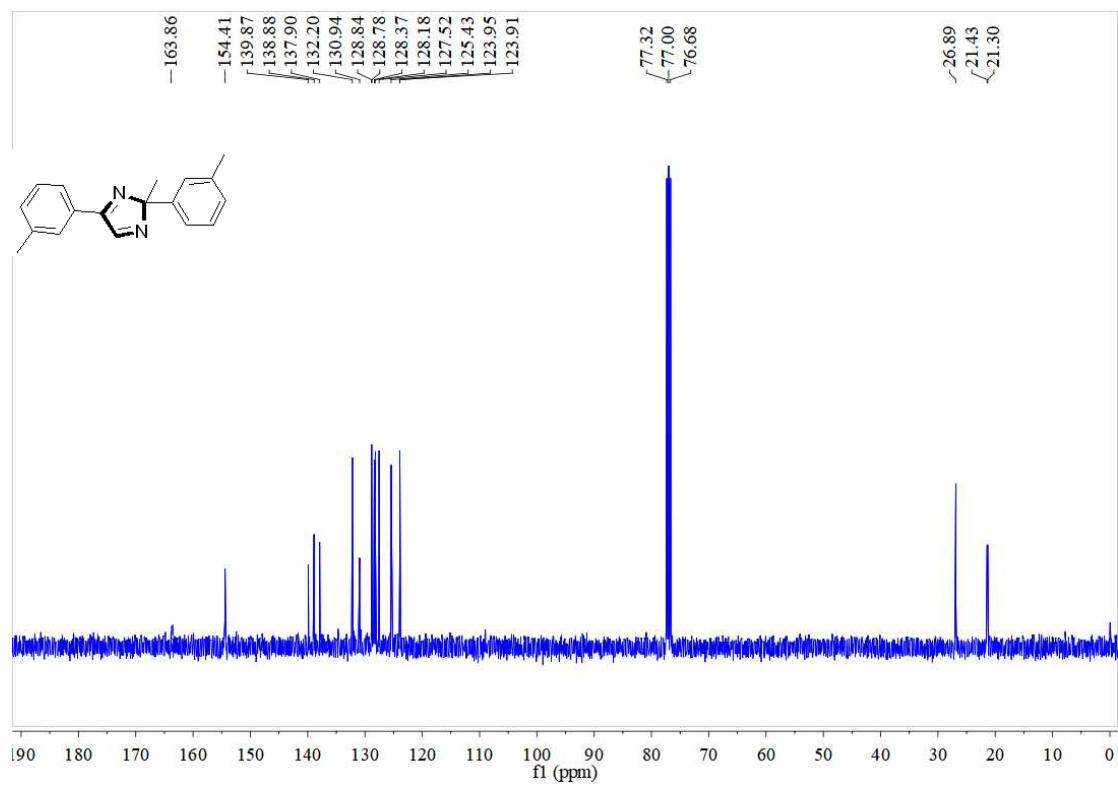


Figure S32 ^{13}C NMR of **3n**

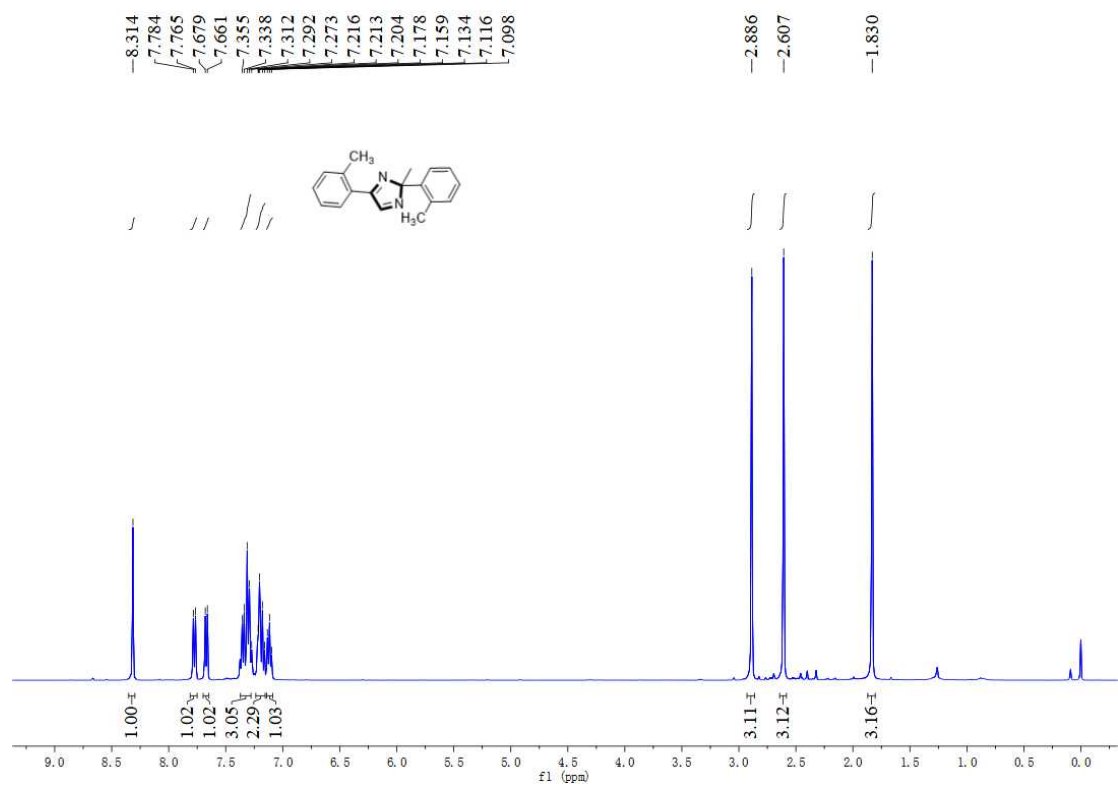


Figure S33 ^1H NMR of **3o**

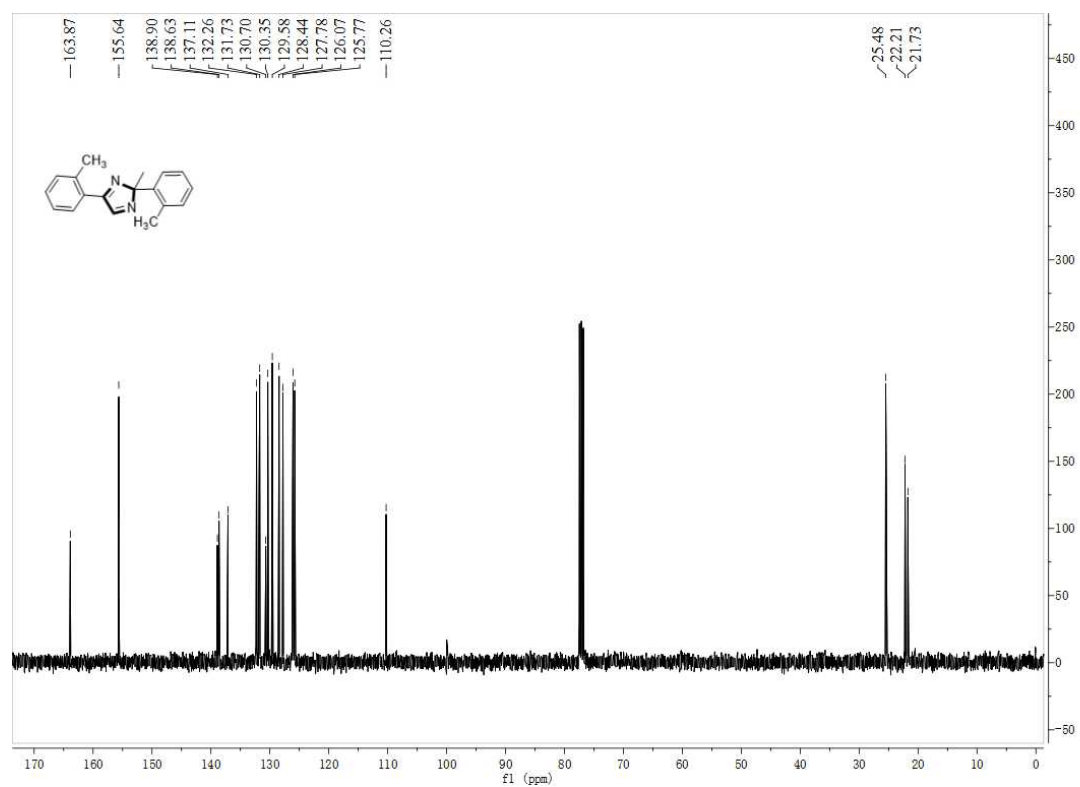


Figure S34 ¹³C NMR of **3o**

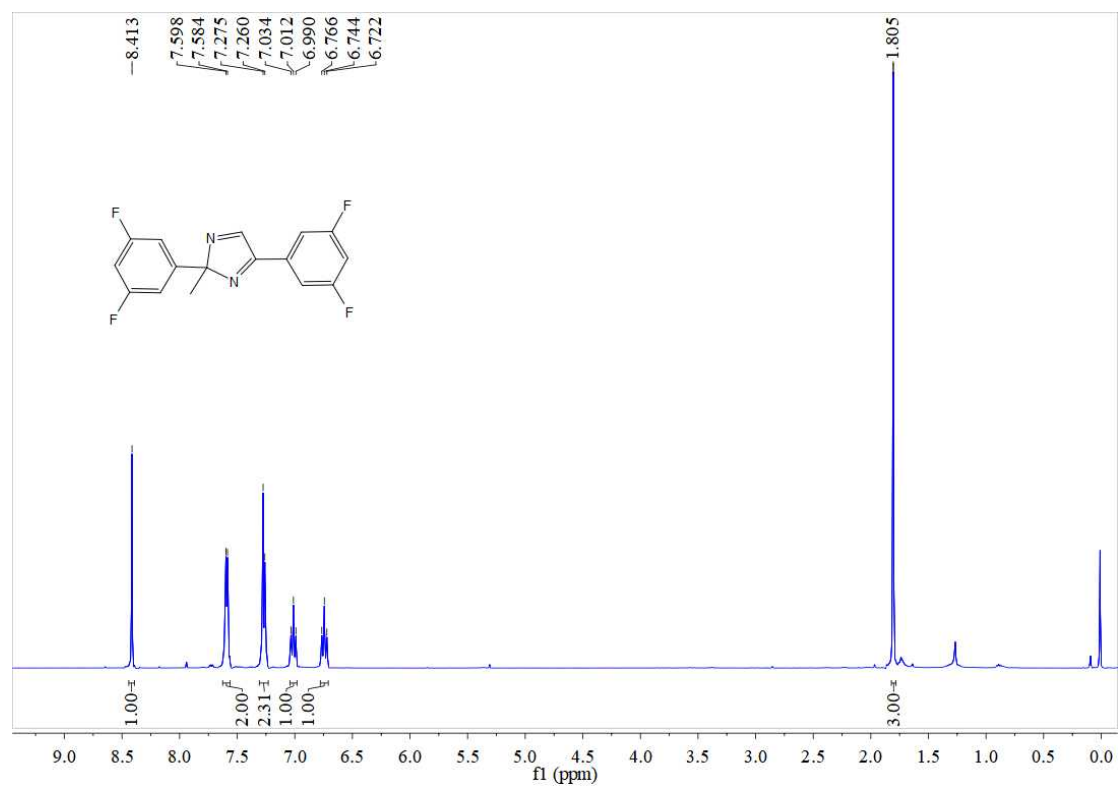


Figure S35 ¹H NMR of **3p**

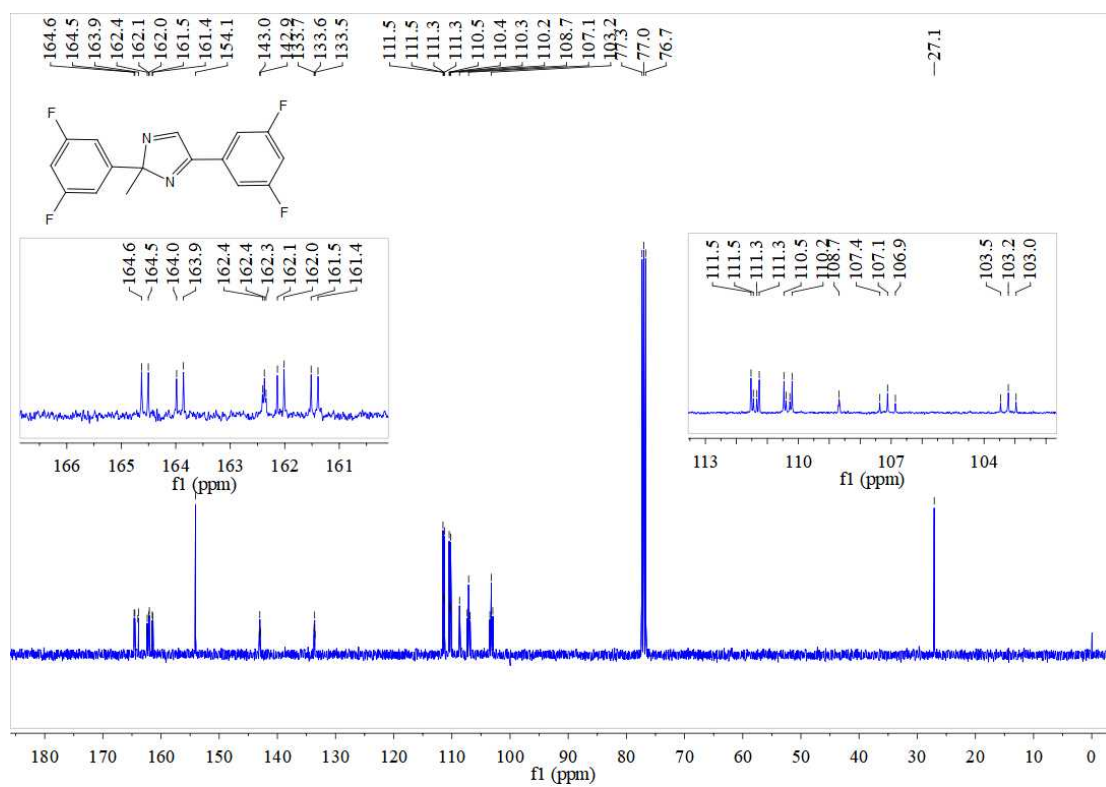


Figure S36 ¹³C NMR of **3p**

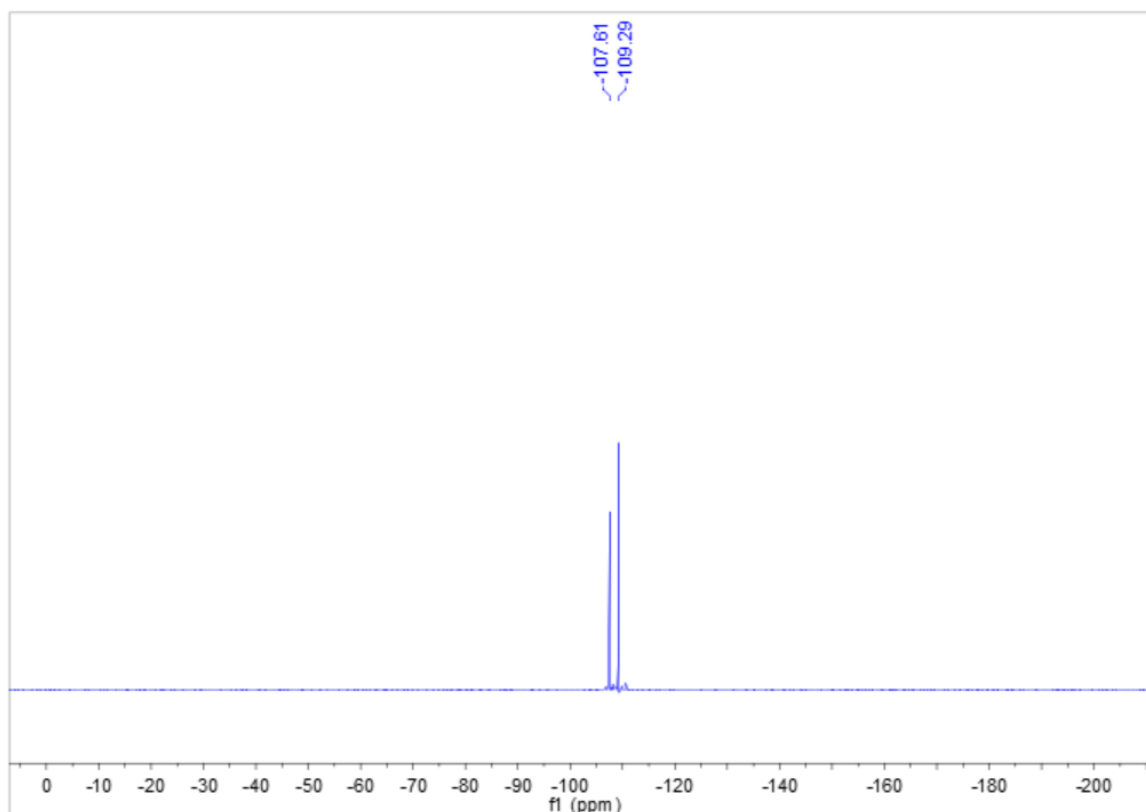


Figure S37 ¹⁹F NMR of **3p**

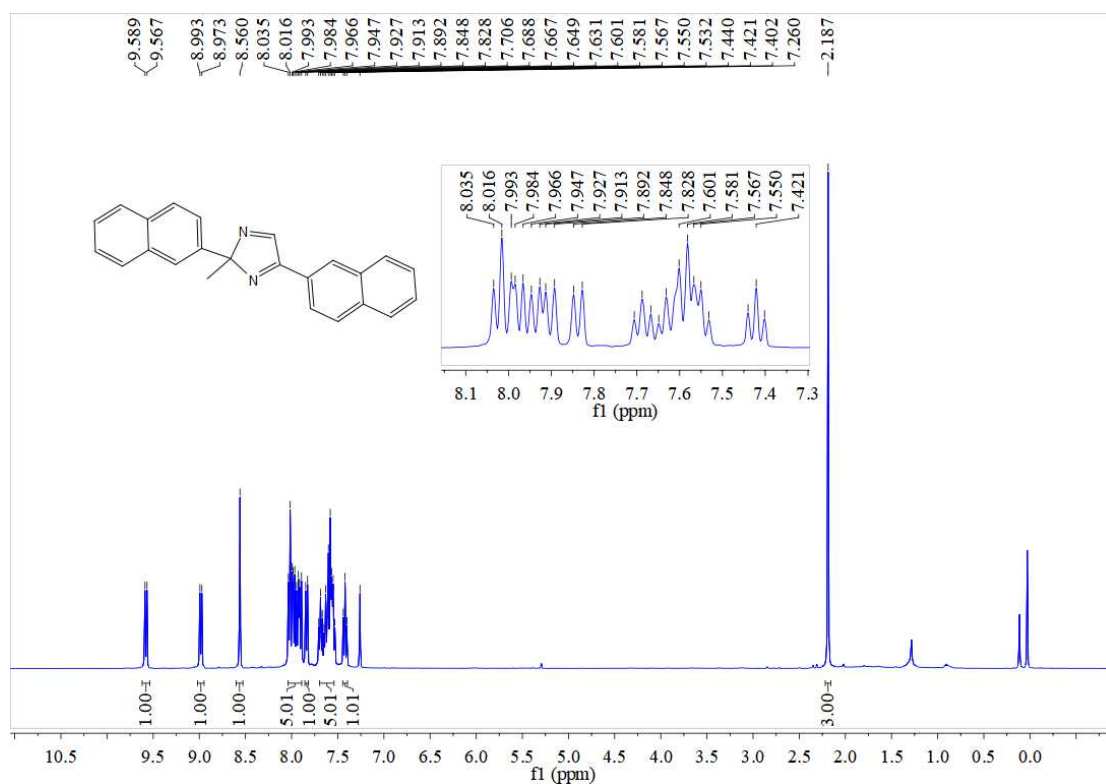


Figure S38 ¹H NMR of 3q

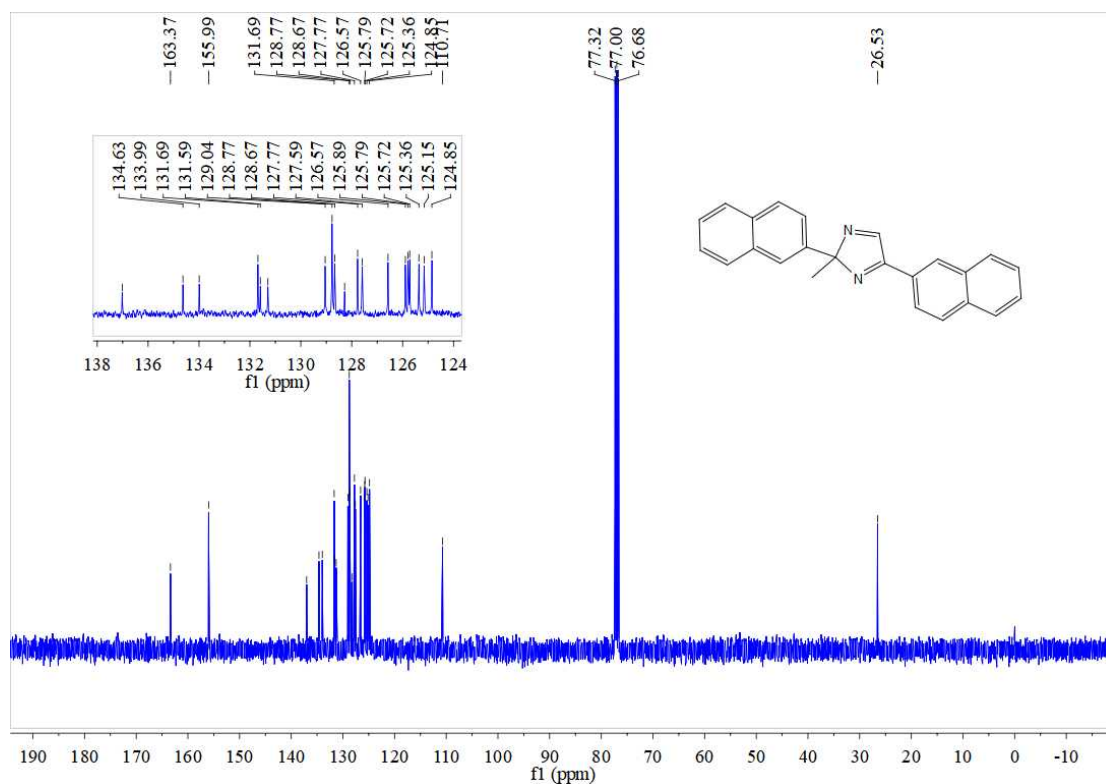


Figure S39 ¹³C NMR of 3q

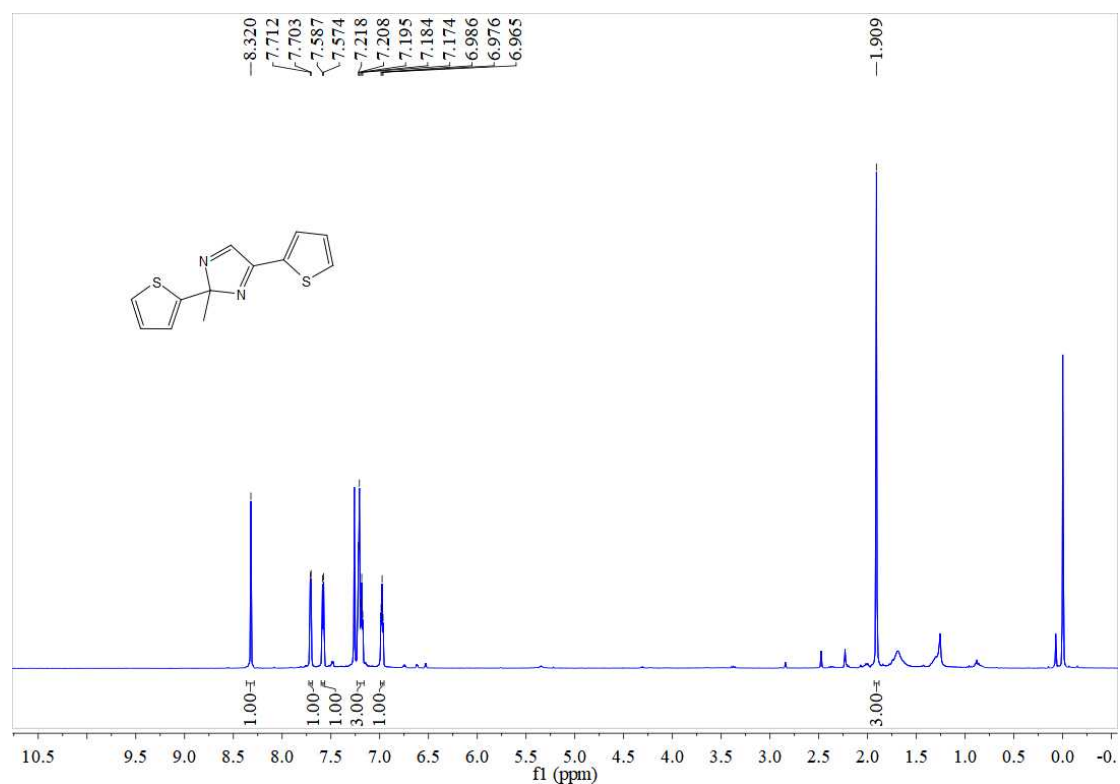


Figure S40 ¹H NMR of **3r**

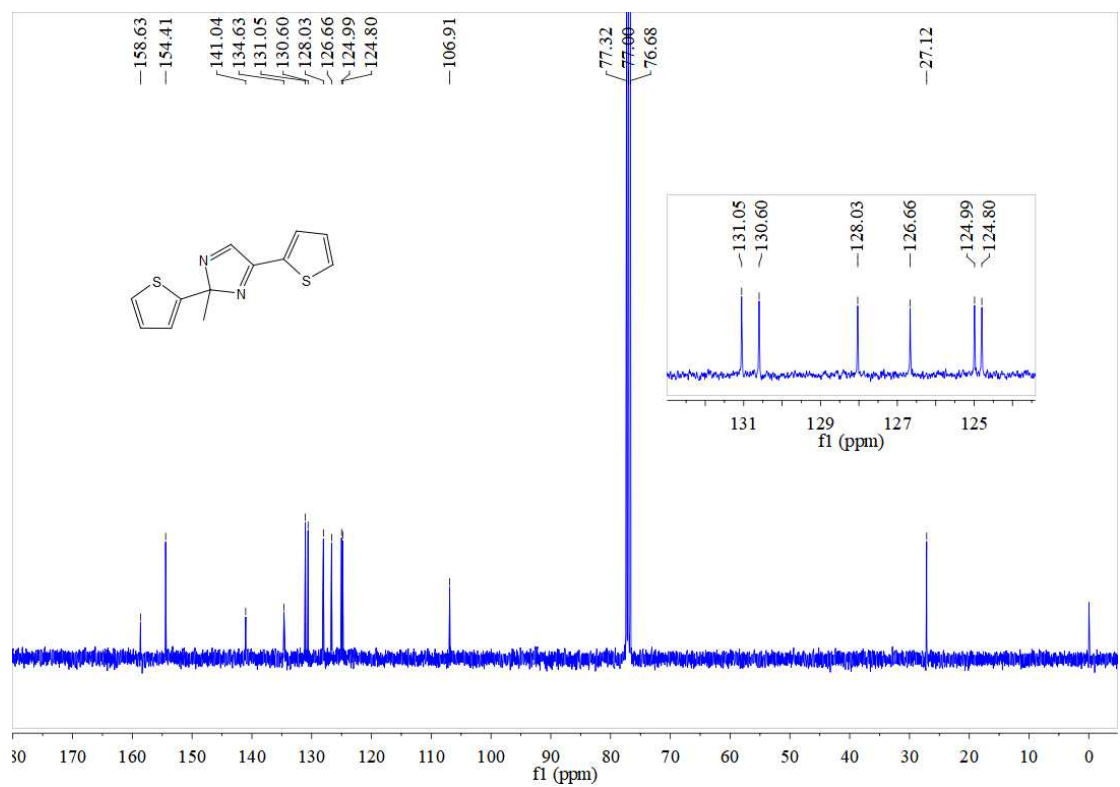


Figure S41 ¹³C NMR of **3r**

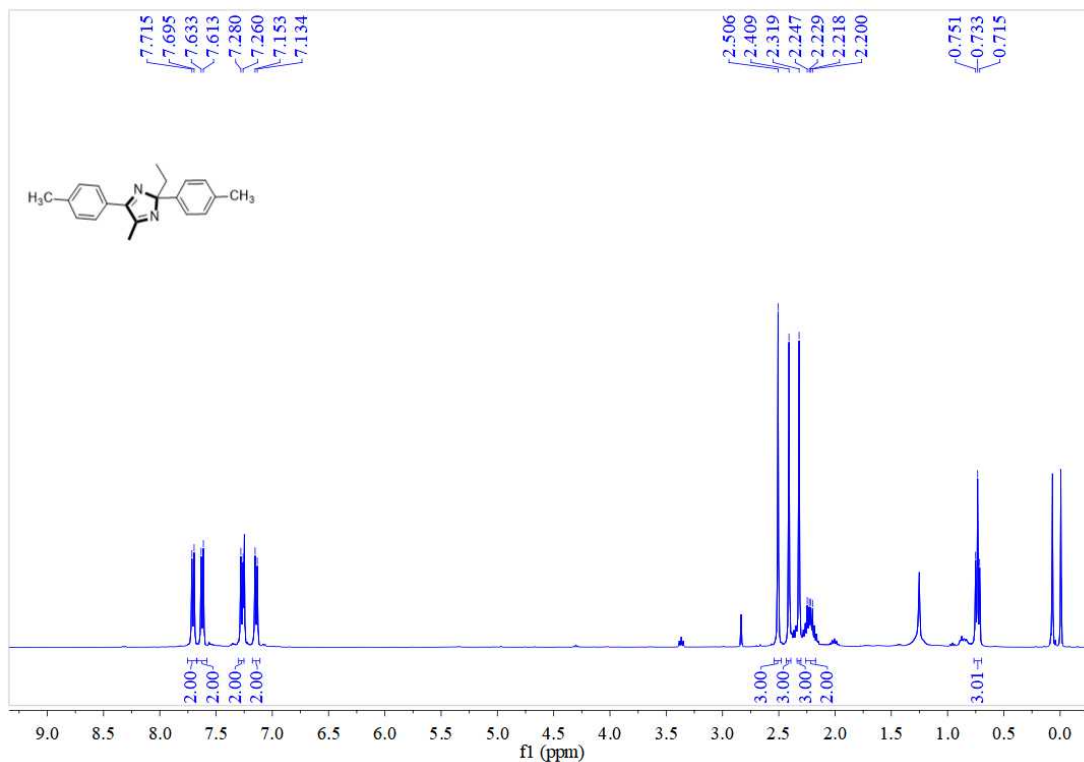


Figure S42 ^1H NMR of **3s**

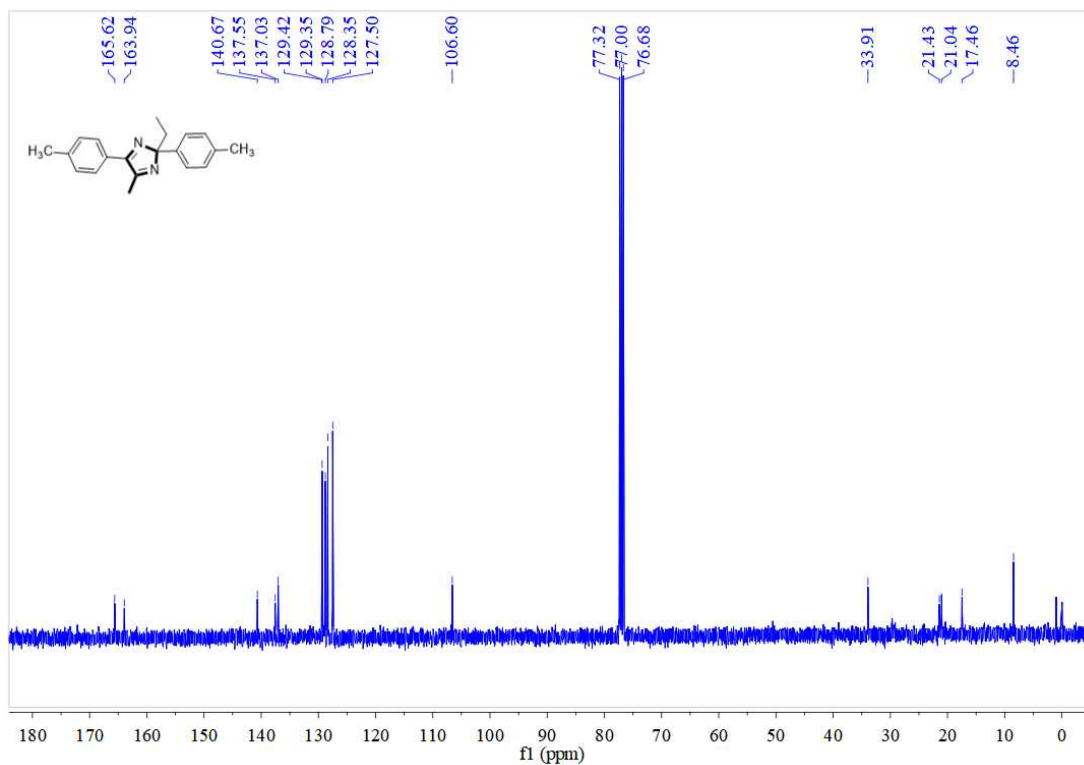


Figure S43 ^{13}C NMR of **3s**

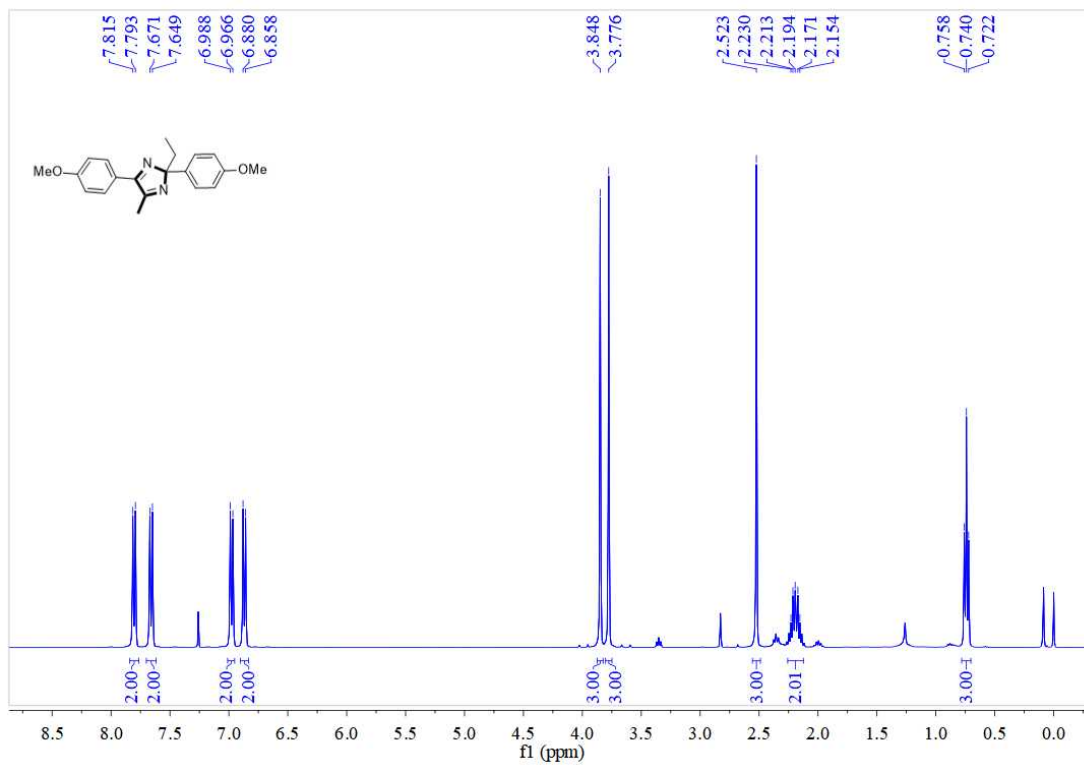


Figure S44 ^1H NMR of **3t**

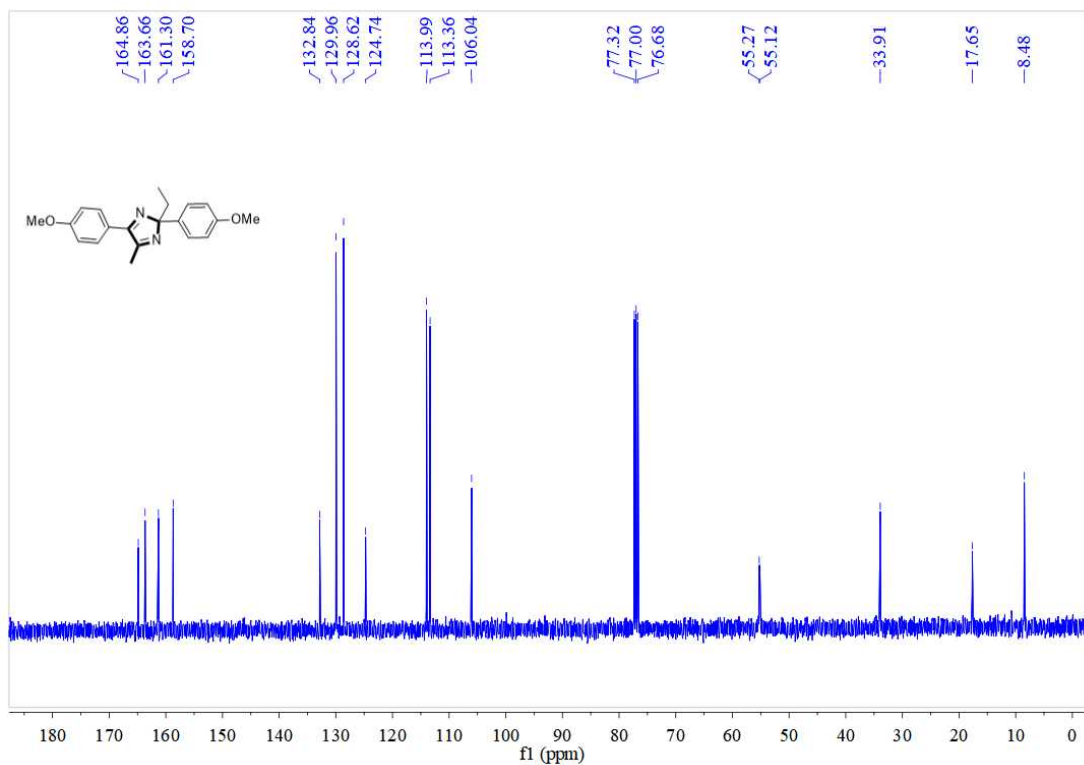


Figure S45 ^{13}C NMR of **3t**

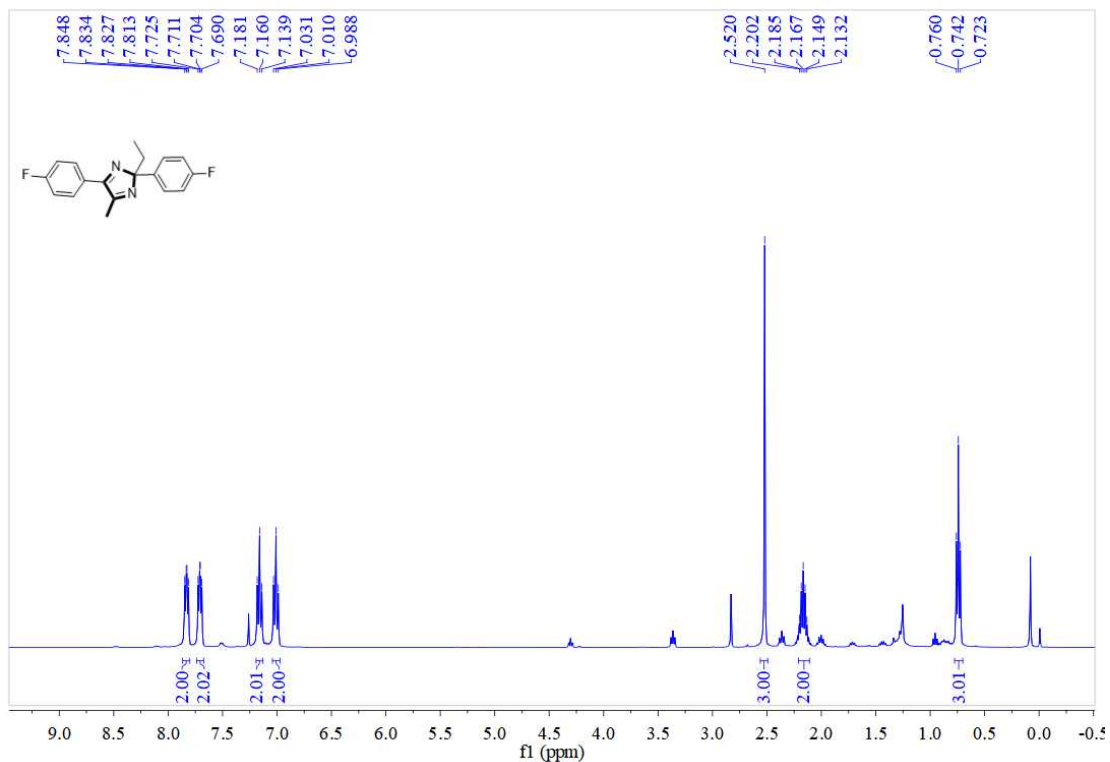


Figure S46 ¹H NMR of **3u**

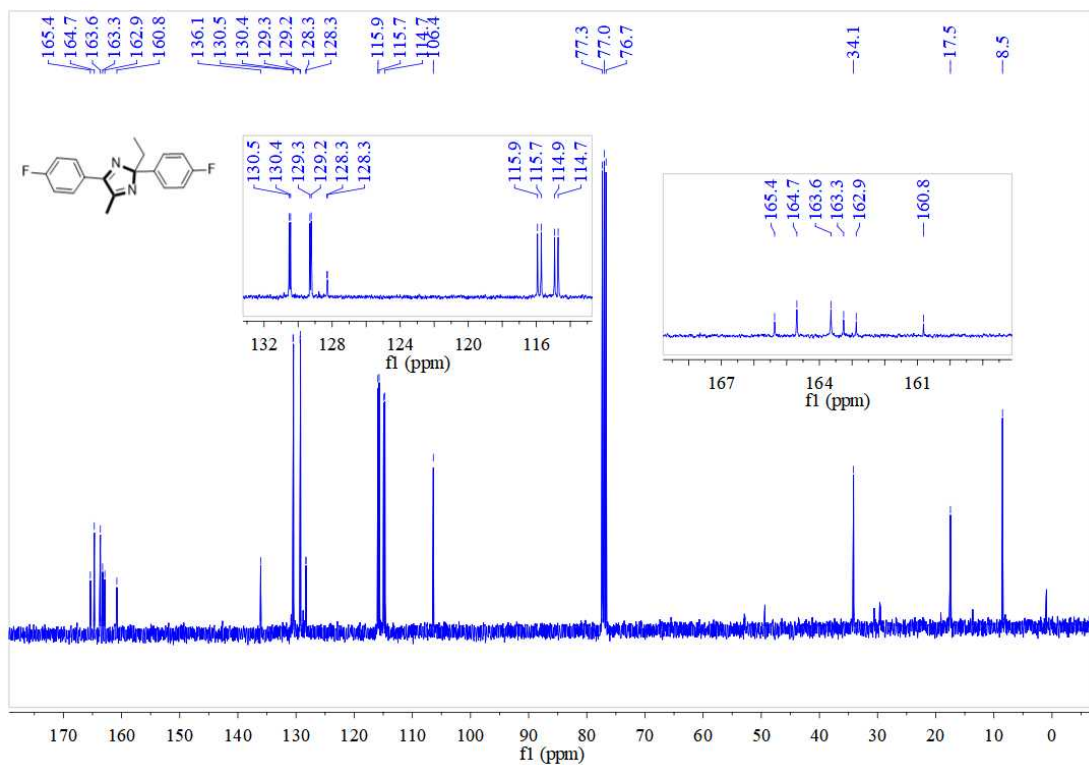


Figure S47 ¹³C NMR of **3u**

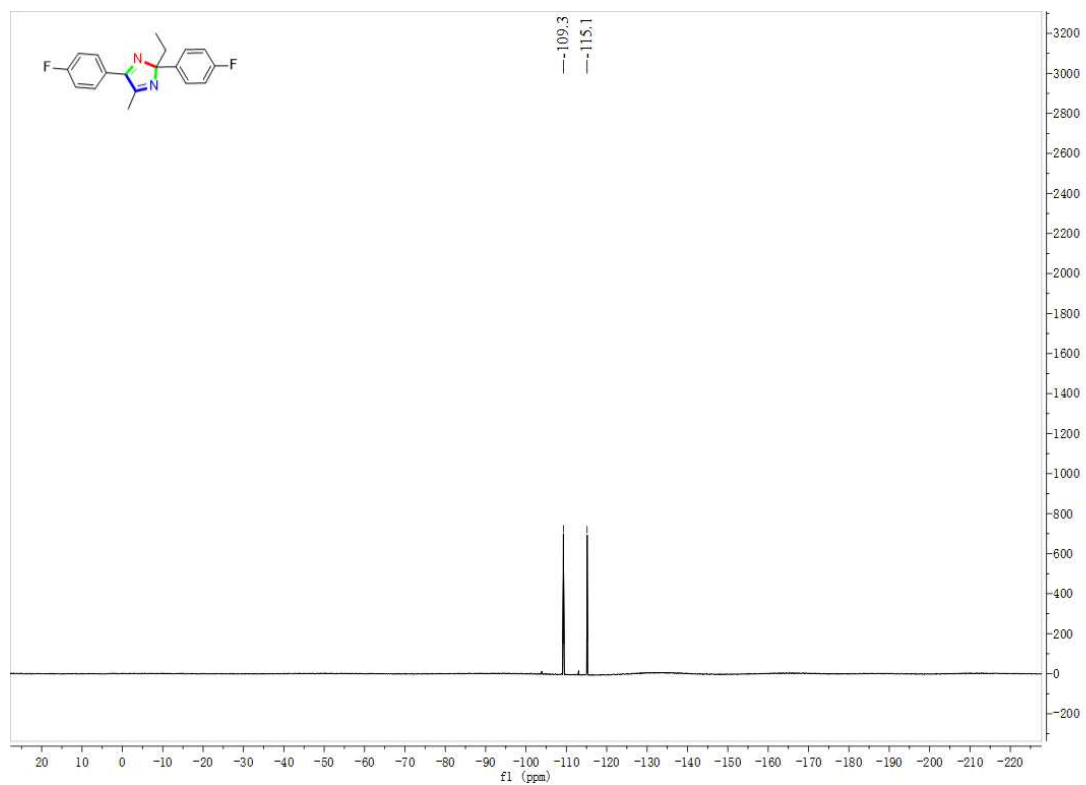


Figure S48 ^{19}F NMR of **3u**

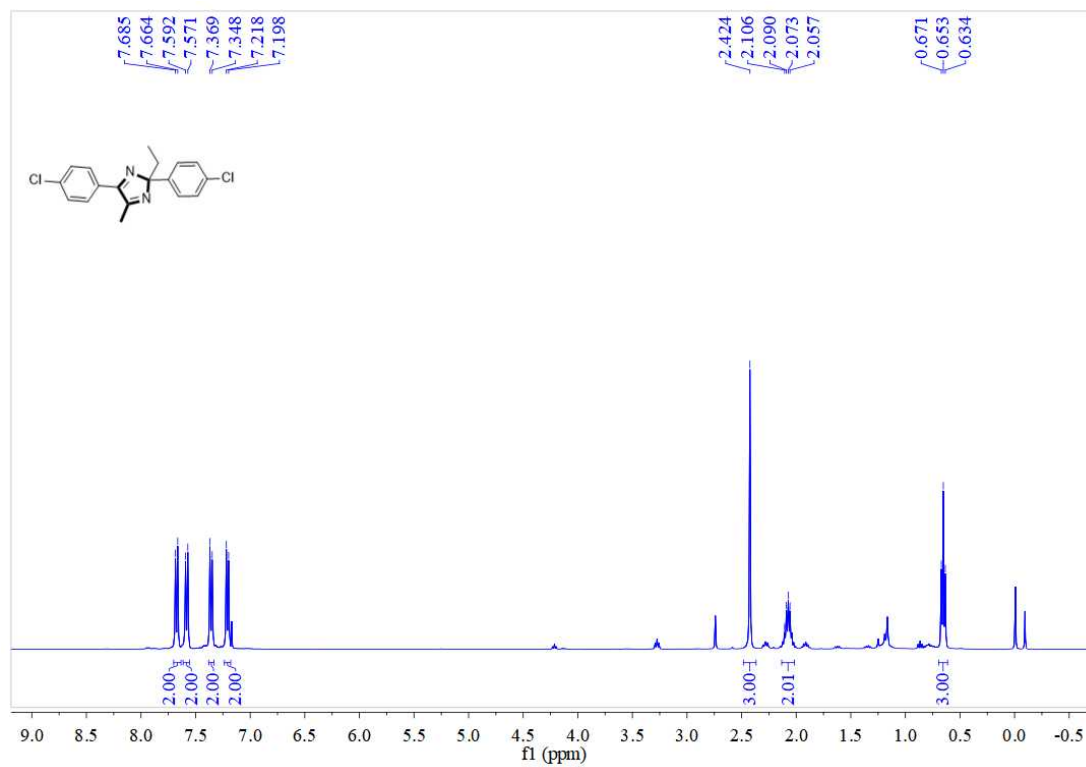


Figure S49 ^1H NMR of **3v**

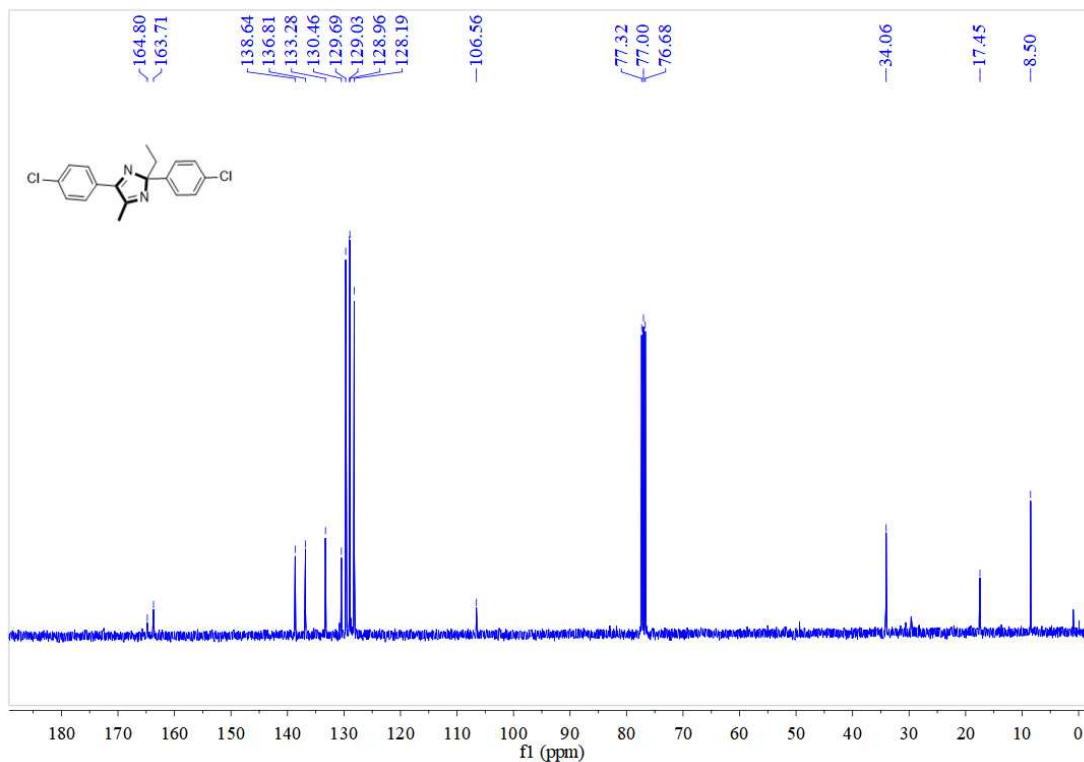


Figure S50 ¹³C NMR of **3v**

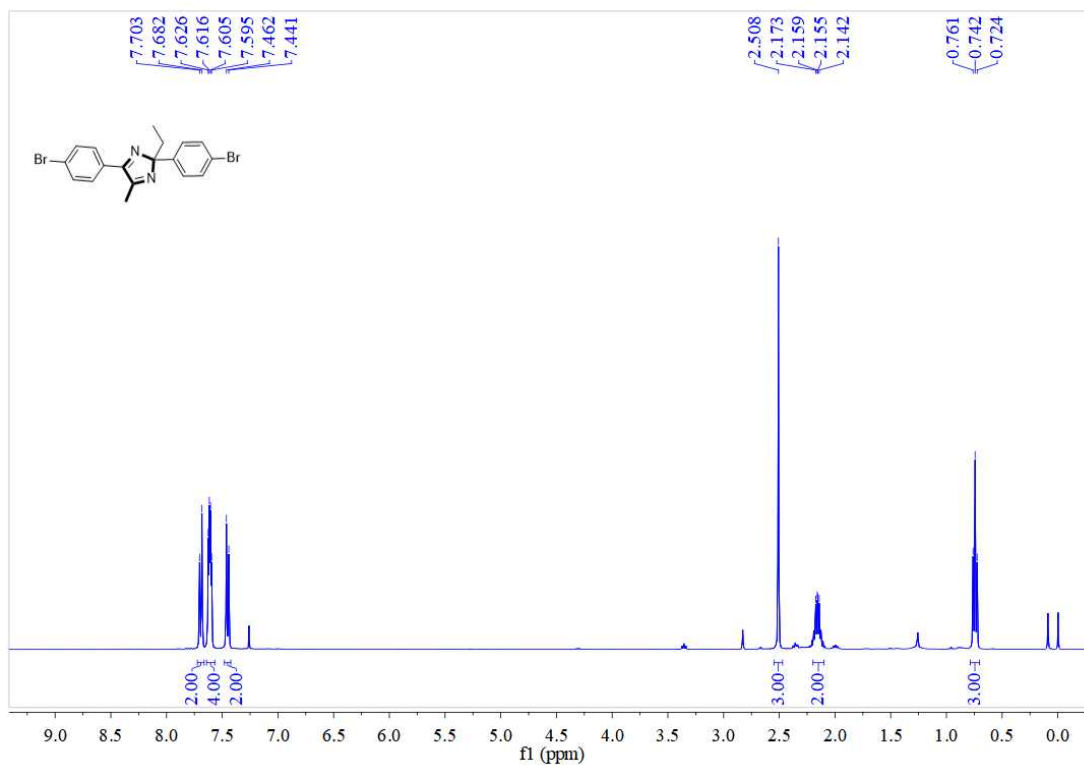


Figure S51 ¹H NMR of **3w**

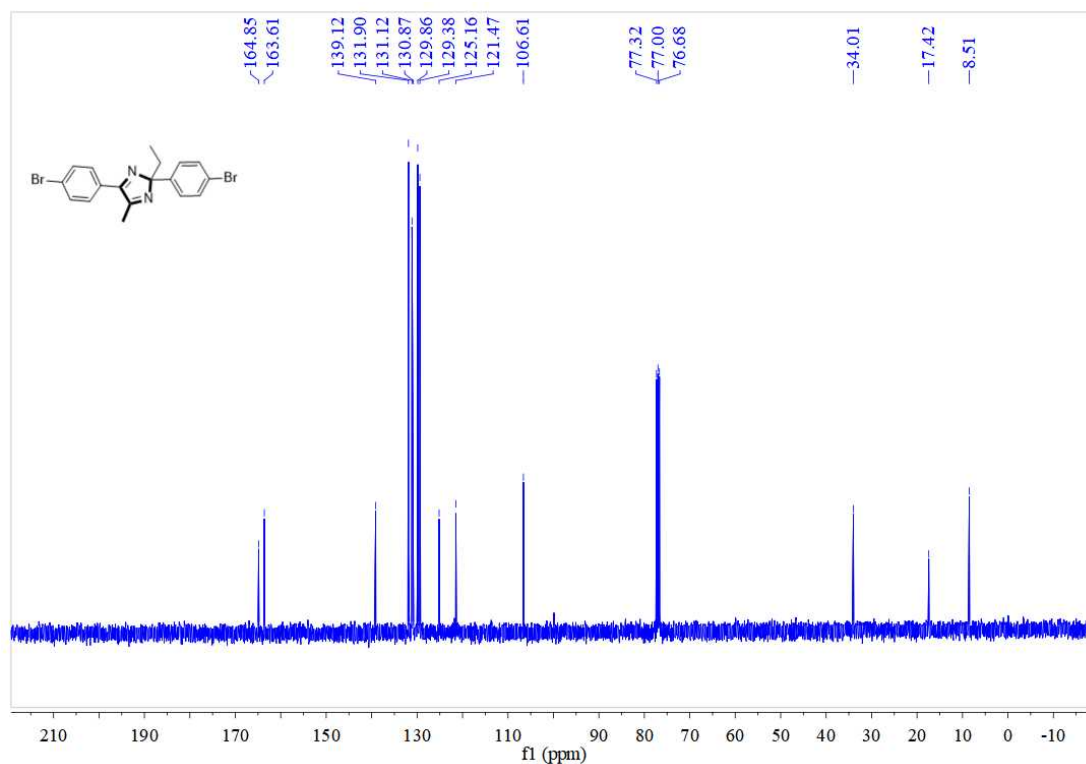


Figure S52 ¹³C NMR of **3w**

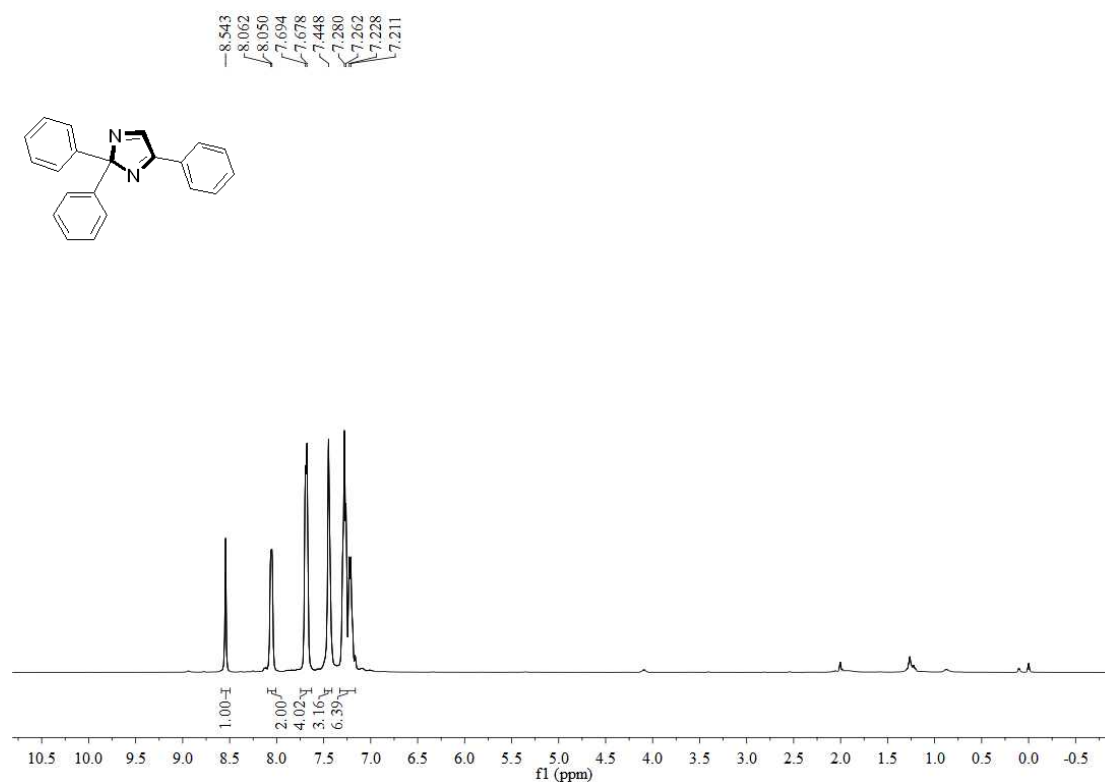


Figure S53 ¹H NMR of **4aa**

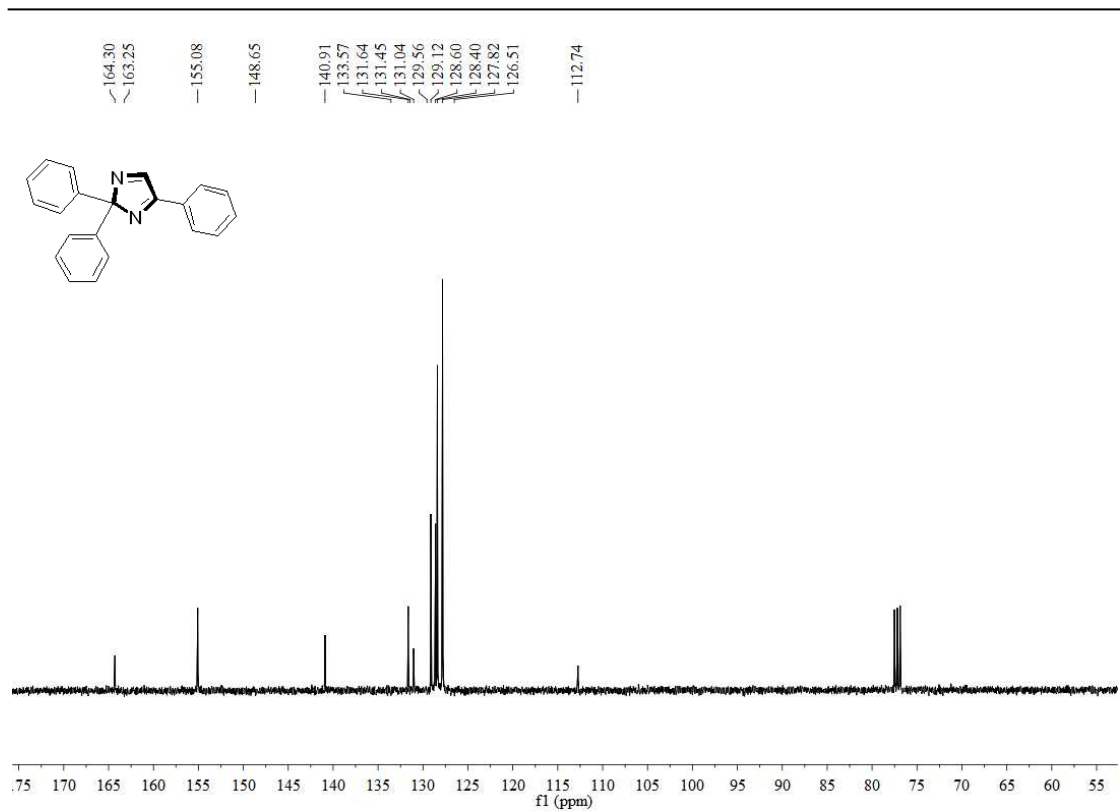


Figure S54 ¹³C NMR of **4aa**

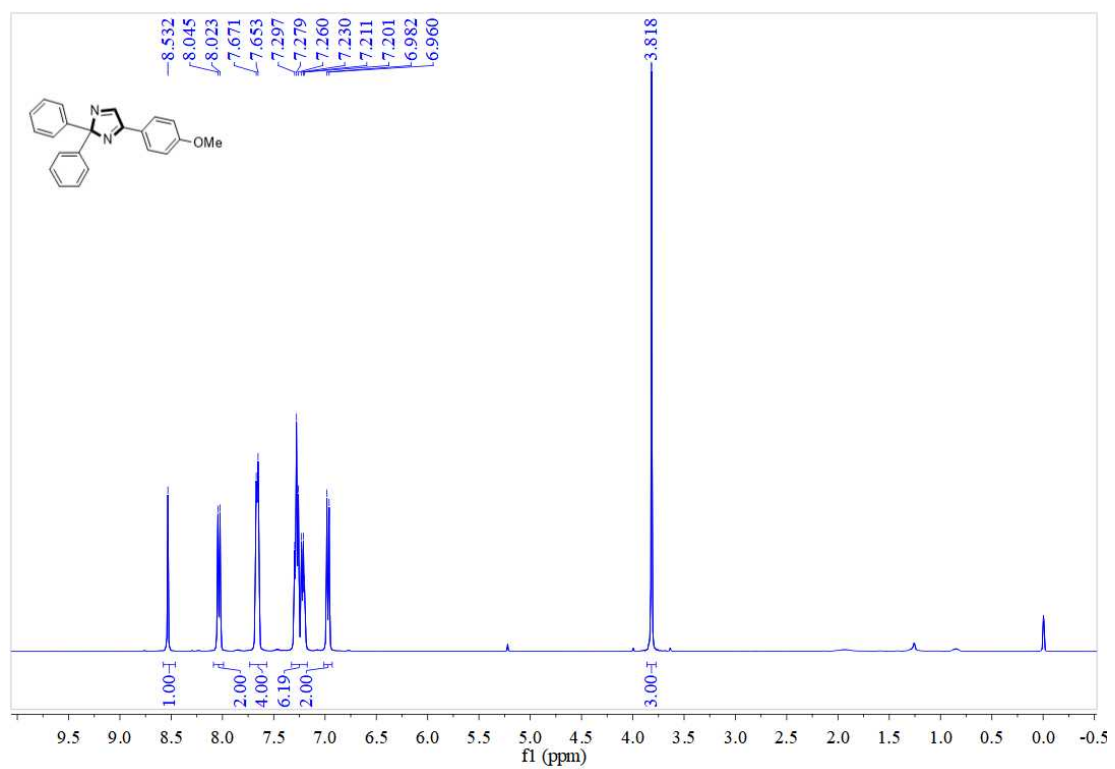


Figure S55 ¹H NMR of **4ab**

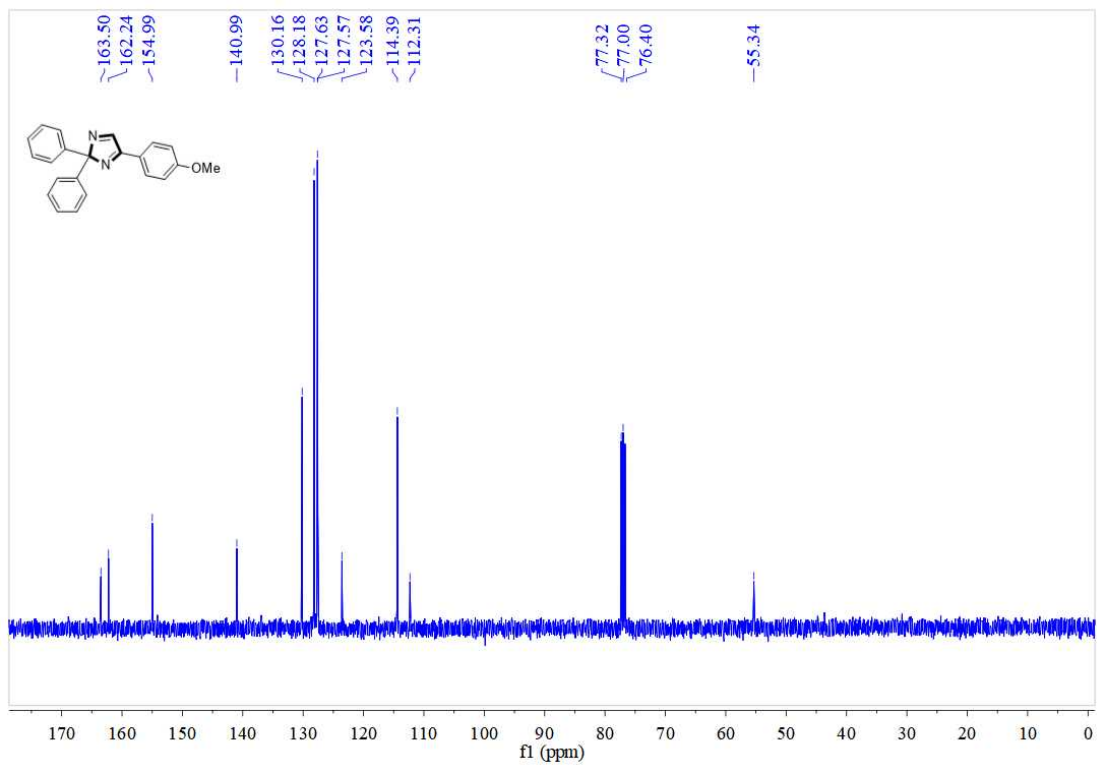


Figure S56 ¹³C NMR of **4ab**

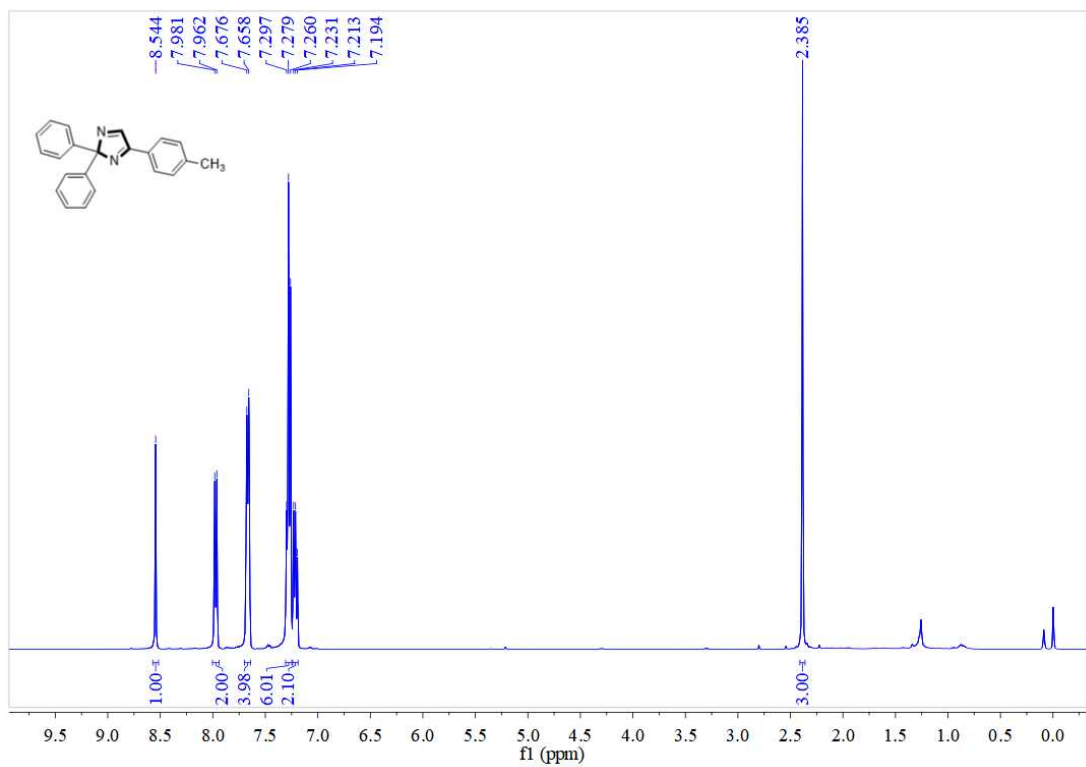


Figure S57 ¹H NMR of **4ac**

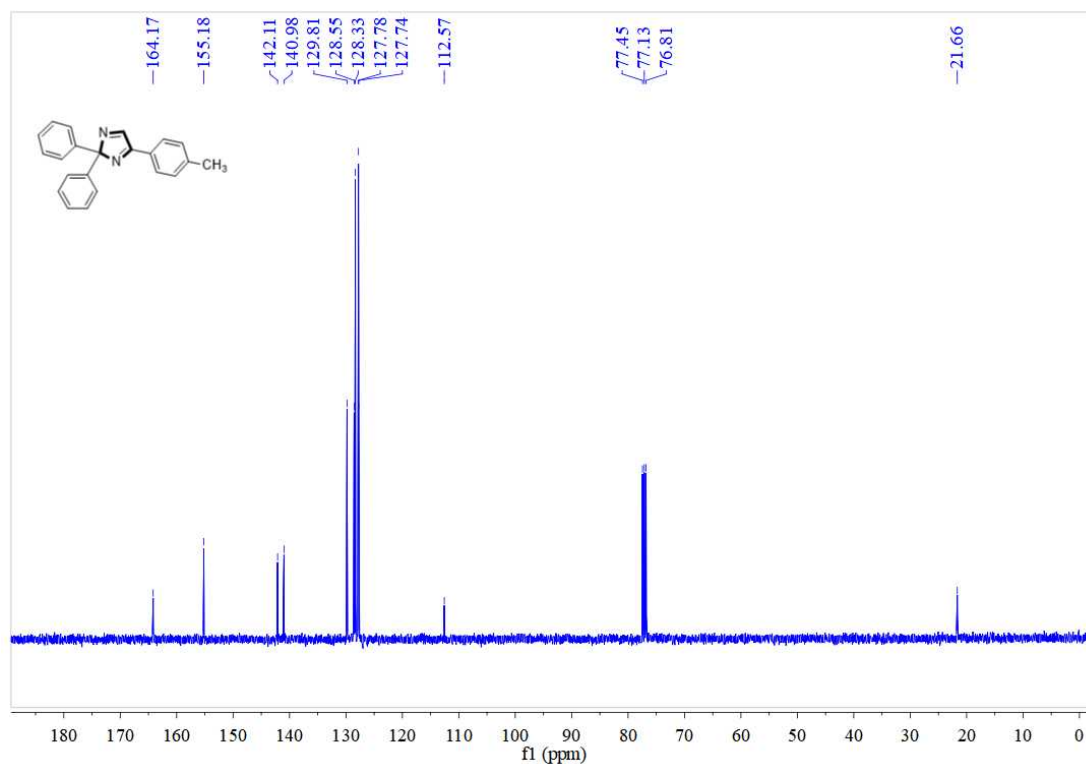


Figure S58 ¹³C NMR of **4ac**

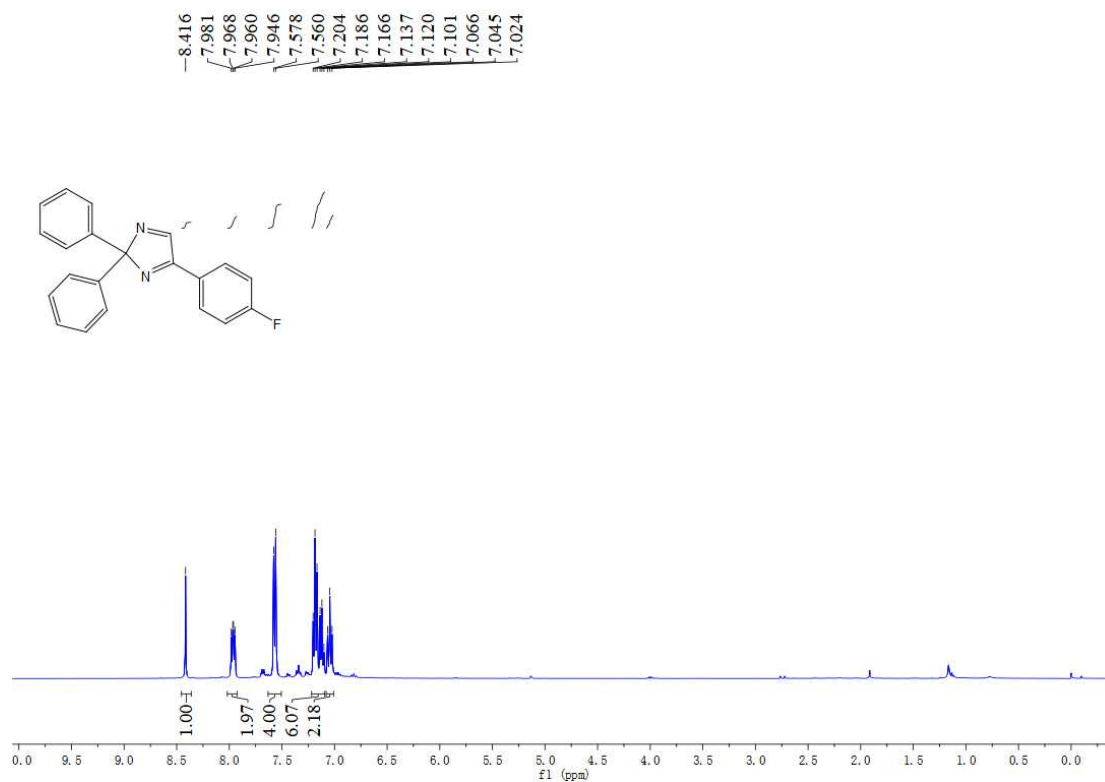


Figure S59 ¹H NMR of **4ad**

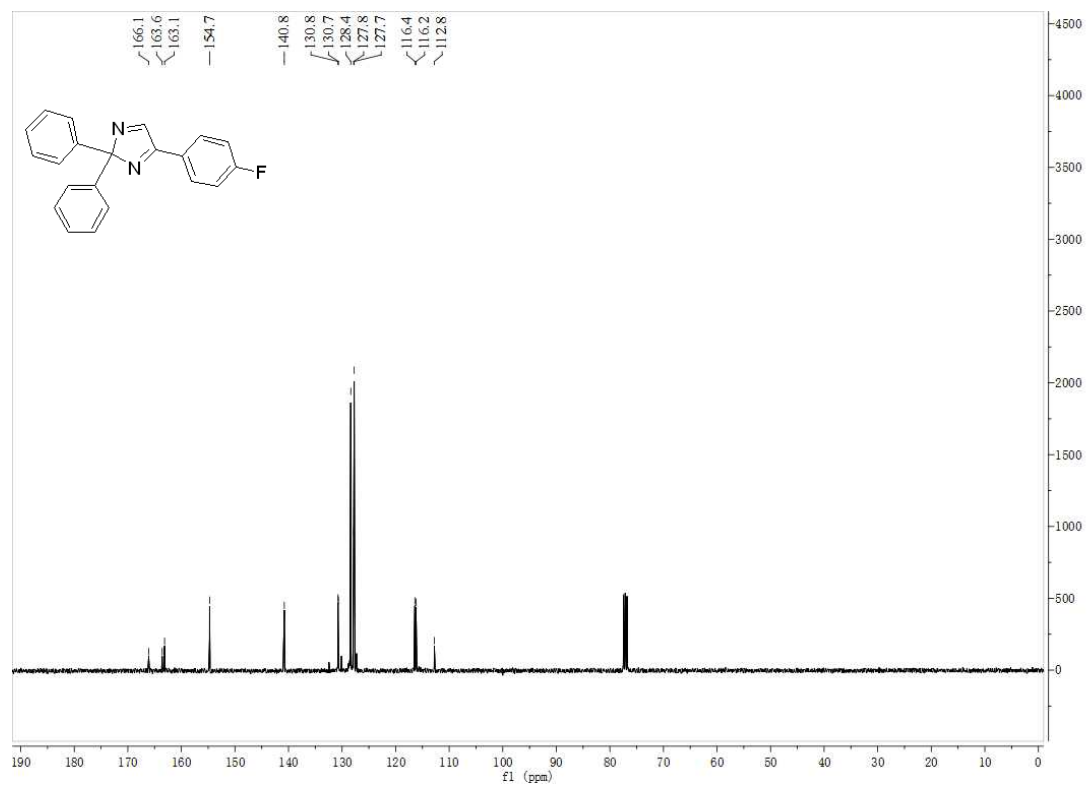


Figure S60 ^{13}C NMR of **4ad**

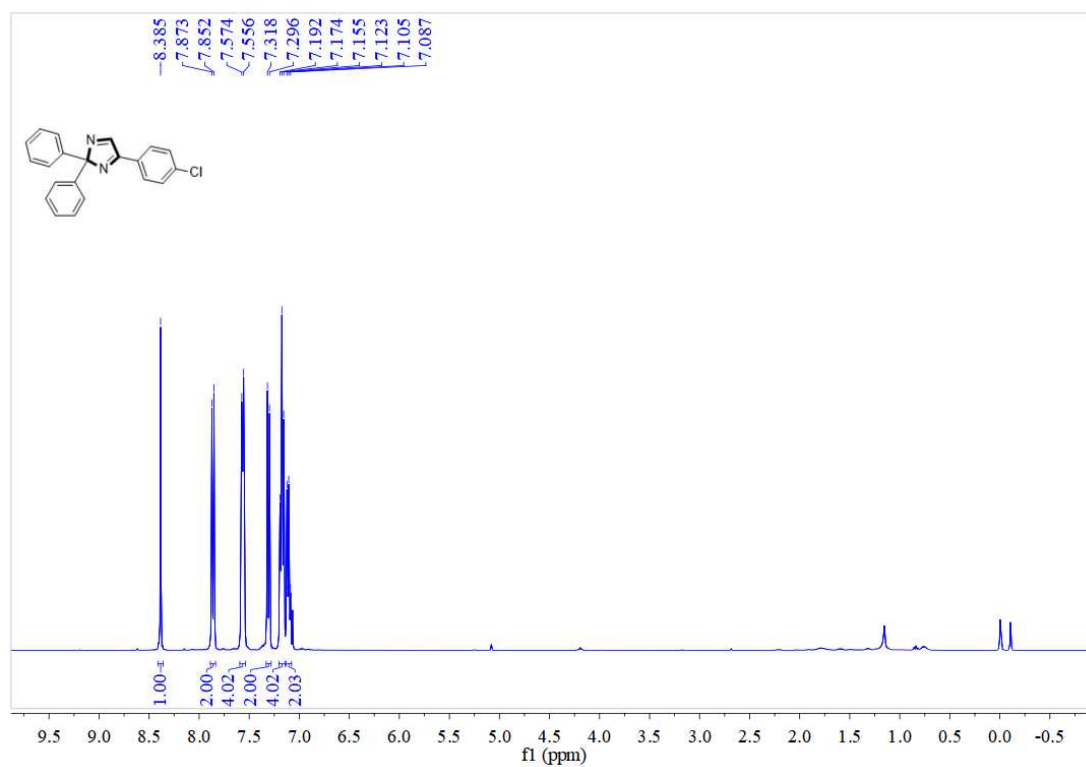


Figure S61 ^1H NMR of **4ae**

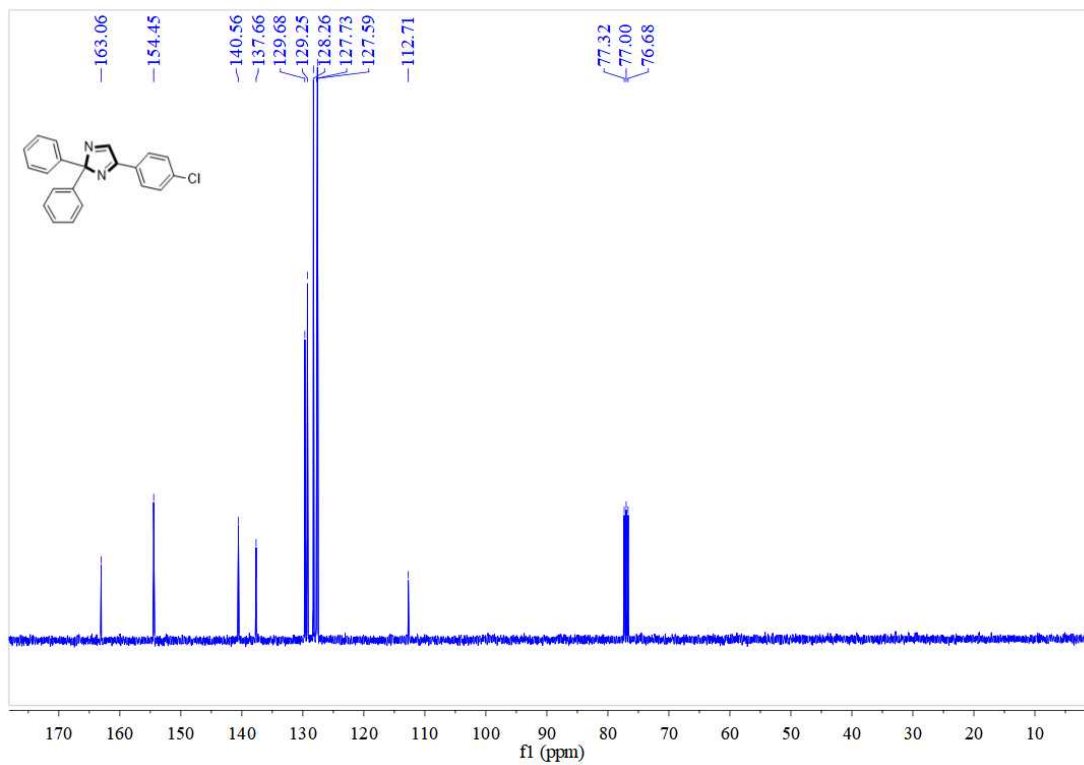


Figure S62 ¹³C NMR of **4ae**

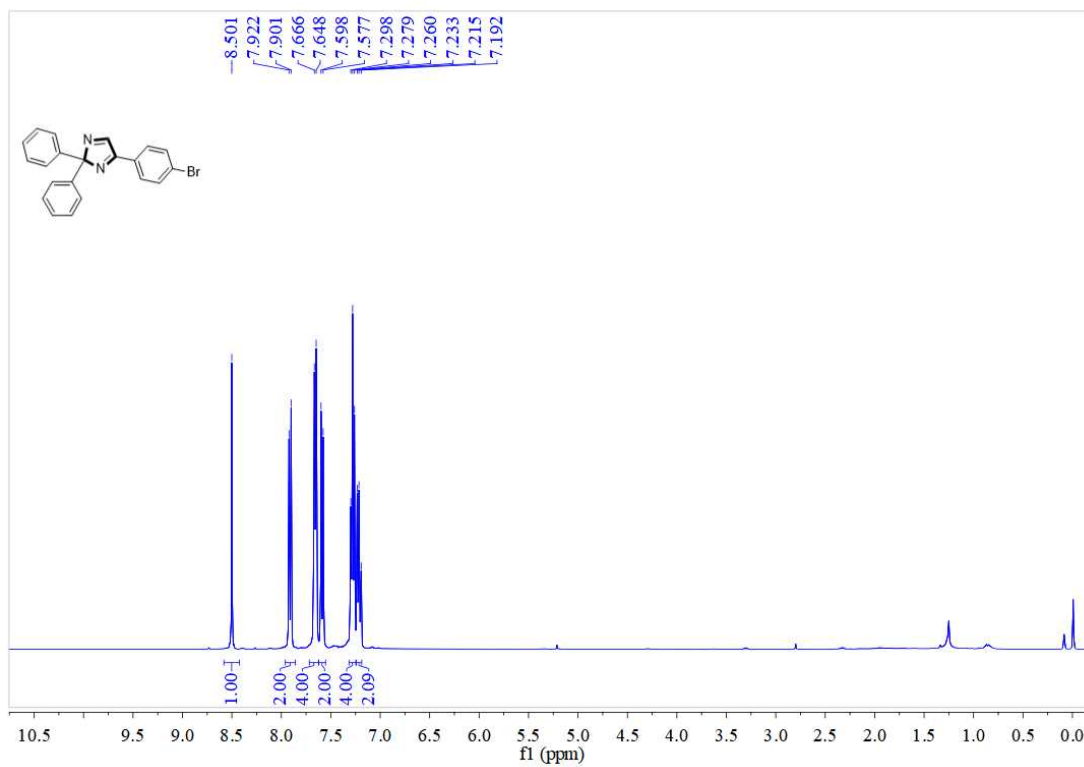


Figure S63 ¹H NMR of **4ae**

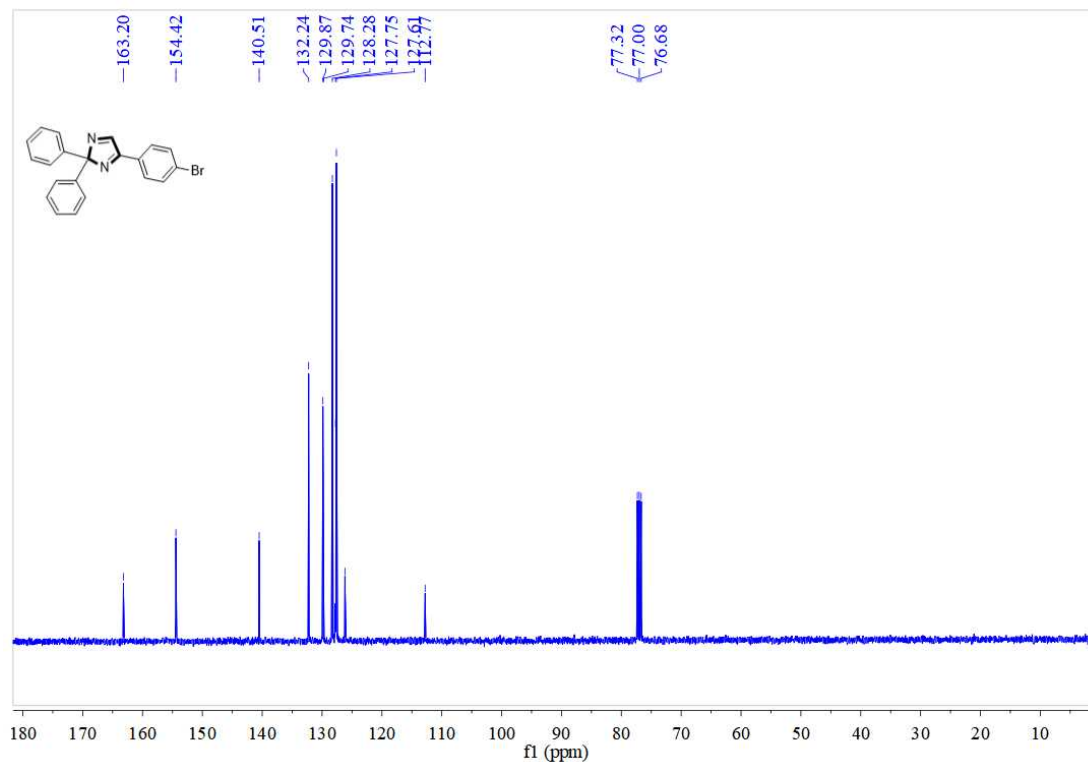


Figure S64 ¹³C NMR of **4ae**

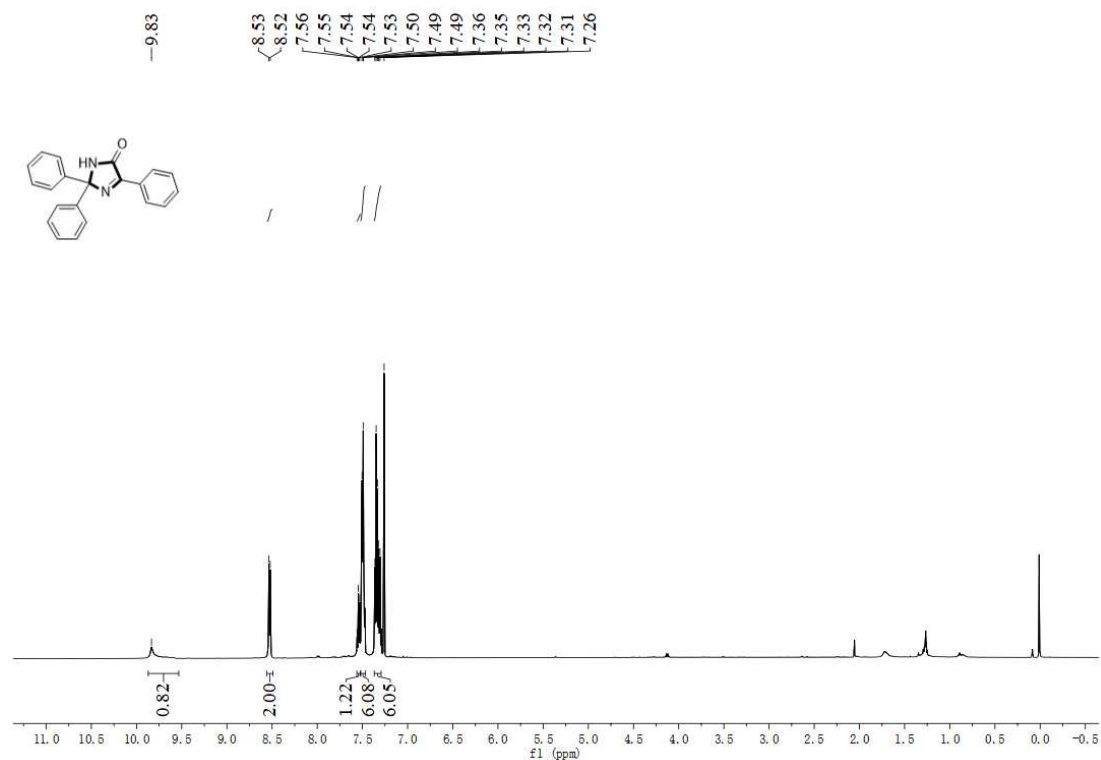


Figure S65 ¹H NMR of **5aa**

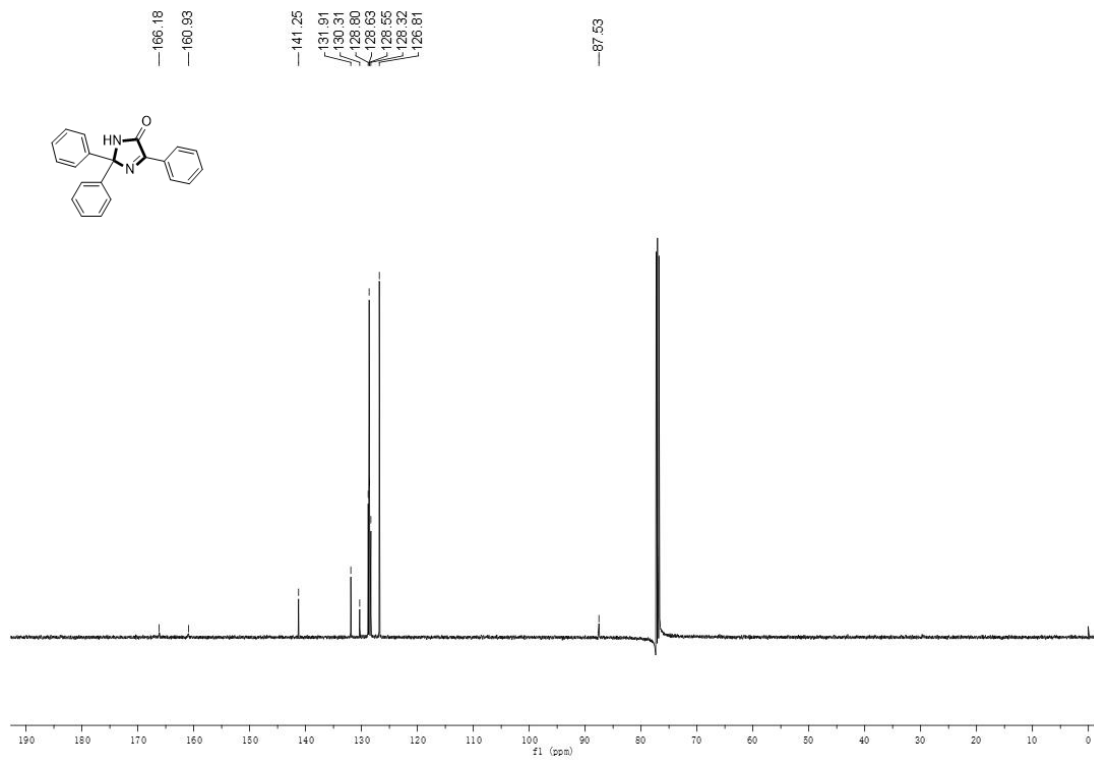


Figure S66 ¹³C NMR of **5aa**