Supporting Information

Enhancing Molecular Diversity in Peptoid Oligomers Using Amino Acid Synthons

Peter T. Smith, Jennifer L. Franco, and Kent Kirshenbaum*

Department of Chemistry, New York University, New York, New York 10003 (USA)

Contents

- 1. Summarized Mass Spectrometry Characterization Data
- 2. Analytical RP-HPLC Spectra and MS Spectra
- 3. Example Peptoids Using Unprotected Tyr, Cys, and His Submonomers
- 4. Intermolecular, On-Resin DKP Formation
- 5. Circular Dichroism of Compounds 1,4, and 5

1. Summaraized MS Characterization Data

Table S1. Calculated and observed electrospray ionization mass spectrometry data for synthesized peptoid oligomers 1-16. Data was obtained using a Waters AutoPurification HPLC/MS system.

Peptoid	Molecular Formula	Calculated m/z	Observed m/z
1	C25H32N4O6	[<i>M</i> + H ⁺]: 485.24	[<i>M</i> + H ⁺]: 485.4
2	C29H39N5O6	[<i>M</i> + H ⁺]: 554.30	[<i>M</i> + H ⁺]: 554.5
3	C41H61N7O9	[<i>M</i> + H ⁺]: 796.46	[<i>M</i> + H ⁺]: 796.8
4	C29H40N4O6	[<i>M</i> + H ⁺]: 541.30	[<i>M</i> + H ⁺]: 541.5
5	C41H62N6O9	[<i>M</i> + H ⁺]: 783.47	[<i>M</i> + H ⁺]: 783.7
6	C31H42N4O5S	[<i>M</i> + H ⁺]: 583.30	[<i>M</i> + H ⁺]: 583.6
7	C68H89N11O11	[<i>M</i> + H ⁺]: 1236.68	$[M + H^+]: 1237.0$ $[M + 2H^+]/2: 619.4$
8	$C_{58}H_{78}N_{14}O_{14}$	[<i>M</i> + H ⁺]: 1195.59	$[M + H^+]: 1195.9$ $[M + 2H^+]/2: 598.6$
9	C66H91IN12O11	[<i>M</i> + H ⁺]: 1355.61	$[M + H^+]: 1356.1$ $[M + 2H^+]/2: 678.7$
10	C ₆₃ H ₈₅ BrF ₃ N ₁₁ O ₁₄ S	[<i>M</i> + H ⁺]: 1388.52	$[M + 2H^{+}](2:676.7)$ $[M + H^{+}]:1390.7$ $[M + 2H^{+}]/2:695.9$
11	C67H88N12O11	[<i>M</i> + H ⁺]: 1237.68	$[M + 2H^{+}]: 1238.1$ $[M + 2H^{+}]/2: 610.7$
12	C61H87F3N10O12	[<i>M</i> + H ⁺]: 1209.66	$[M + 2H^{+}]: 1209.1$ $[M + 2H^{+}]/2: 605.6$
13	C72H91N11O14S	[<i>M</i> + H ⁺]: 1366.66	[M + 2H]/2.003.0 $[M + H^+]: 1367.0$
14	C68H100N10O11S	[<i>M</i> + H ⁺]: 1265.74	$[M + 2H^{+}]/2: 684.1$ $[M + H^{+}]: 1266.2$
15	C52H85N9O12S	$[M + H^+]$: 1060 61	$\frac{[M + 2H^+]/2:633.7}{[M + H^+]:1061.0}$
10			$\frac{[M + 2H^+]/2:531.2}{[M + H^+]:1239.1}$
16	C62H95N9O13S2	$[M + H^+]: 1238.66$	$[M + 2H^+]/2:620.1$

High-resolution Mass Spectrometry (HRMS) of Peptoids 1, 4, and 5

HRMS spectra were obtained for compounds 1,4, and 5 using a Waters G3 QTof MS system (ZORBAX StableBond 300, C₁₈, 4.6 x 50 mm).







2. Analytical RP-HPLC Spectra and HPLC/MS Spectra

Analytical RP-HPLC and ESI-MS data for peptoids 1-16. ESI-MS spectra were obtained using a Waters AutoPurification HPLC/MS system with a semi-preparative C_{18} column. Analytical HPLC spectra were obtained from a linear gradient of 5-95% ACN/H₂O (0.1% TFA) over 10 min using a C_{18} column (Waters XBridge BEH300 4.6x50 mm) with a flow rate of 0.7 mL/min at UV detection 220 nm. For compound 5, an Agilent HPLC 1260 Infinity with an Eclipse Plus C_{18} column (3.5 uM, 4.6 x 100 mm) was used in place of the Waters instrument.

The overall purified percent yield values were calculated by dividing the mass of isolated pure product by the theoretical mass determined from the reaction scale using 100 mg of resin with 0.2 mmol/g loading. The average yields per number of steps was determined using the overall purified yield and the number of reaction steps required for each oligomer synthesis, including the initial resin deprotection step and the final resin cleavage step.

Amine submonomer 1: 2-Methoxyethylamine Amine submonomer 2: Benzylamine Amine submonomer 3: L-Phenylalanine Theoretical yield: 9.69 mg Crude Mass: 9.3 mg Crude Mass: 7.3 mg Average Yield over 8 Steps: 97% Calculated $m/z [M + H^+]$: 485.24 2.0



Amine submonomer 1: 2-Methoxyethylamine Amine submonomer 2: Benzylamine Amine submonomer 3: L-Phenylalanine Amino acid side chain coupling partner: Morpholine Theoretical Yield: 11.07 mg Crude Mass: 11.0 mg Crude Yield: 99% Pure Mass: 7.6 mg **Overall Purified Yield: 69%** Average Yield Over 9 Steps: 96% Calculated $m/z [M + H^+]$: 554.30 2.0 1.5 Q 1.0 2 0.5 0.0 0.0 1.0 2.0 3.0 4.0 5.0 6.0 7.0 8.0 9.0 10.0 min ^{554.5} M + H⁺ 6.72e7 ר100 555.5

56.5 2M + H+ M + Nat 1108.0 577.5 1109.2 1942.5/ 634.5 850.0 1276.1,1297. 169.7 235.5 1500 1600 1700 1800 1900 200 300 400 500 600 900 1000 1100 1200 1300 1400 700 8Ó0





Amine submonomer 1: 2-MethoxyethylamineAmine submonomer 2: BenzylamineAmine submonomer 3: L-PhenylalanineAmino acid side chain coupling partner: 1-ButanolAmine submonomer 4: 2-MethoxyethylamineAmine submonomer 5: IsopentylamineTheoretical Yield: 15.65 mgCrude Mass: 15.5 mgCrude Mass: 6.9 mgOverall Purified Yield: 44%Average Yield Over 13 Steps: 94%Calculated m/z $[M + H^+]$: 783.47



Amine submonomer 1: 2-Methoxyethylamine Amine submonomer 2: Benzylamine Amine submonomer 3: L-Phenylalanine Amino acid side chain coupling partner: Cyclohexanethiol Theoretical Yield: 11.65 mg Crude Mass: 11.2 mg Crude Yield: 96% Pure Mass: 5.5 mg **Overall Purified Yield: 47%** Average Yield Over 9 Steps: 92% Calculated $m/z [M + H^+]$: 583.30 1.0 0.8 0.6 AU 0.4 6 0.2 0.0 0.0 1.0 2.0 3.0 4.0 5.0 6.0 7.0 8.0 9.0 10.0 min ^{583.6} M + H⁺ 5.23e7 100-584.6 85.6 .605.6 M + Na⁺ 606.5 451.4

.334.0 1338.3 1300 1166.1 752.5 785.9 894.7 251.1 292.3 402.6 611.1 Ĥ. 1506.7 1617.0 1733.91798.2 1889.5 553.6 ⊢ m/z 800 300 400 700 900 1100 1500 200 500 600 1000 1200 1400 1600 1700 1800 1900



Amine submonomer 1: L-Phenylalanine

Amino acid side chain coupling partner: Propargylamine

Amine submonomer 2: Aniline

Amine submonomer 3: L-Glutamine

Amino acid side chain coupling partner: β-Alanine *tert*-butyl ester Amine submonomer 4: L-Serine

Amino acid side chain coupling partner: (*S*)-1-Phenylethylamine Amine submonomer 5: *N*-Boc-ethylenediamine

Amine submonomer 6: L-Proline

Amino acid side chain coupling partner: N-Boc-ethylenediamine

Theoretical Yield: 23.9 mg

Crude Mass: 15.0 mg

Pure Mass: 2.3 mg

Average Yield Over 18 Steps: 88%





Crude Yield: 63%

Overall Purified Yield: 10%

Amine submonomer 1: Glycine Amino acid side chain coupling partner: Isopentylamine Amine submonomer 2: (S)-1-Phenylethylamine Amine submonomer 3: L-Isoleucine Amino acid side chain coupling partner: N-Boc-ethylenediamine Amine submonomer 4: L-Phenylalanine Amino acid side chain coupling partner: Cyclopropylmethanamine Amine submonomer 5: 2-Methoxyethylamine Amine submonomer 6: D-Phenylalanine Amino acid side chain coupling partner: 4-Iodoaniline Theoretical Yield: 27.1 mg Crude Mass: 5.7 mg Crude Yield: 21% Pure Mass: 1.9 mg **Overall Purified Yield: 7%** Average Yield Over 18 Steps: 86% Calculated m/z [*M* + H⁺]: 1355.61 1.0 0.8 0.6 ₹ _{0.4} 0.2 -0.0 -0.2 0.0 1.0 2.0 3.0 4.0 5.0 6.0 7.0 8.0 9.0 10.0 min 1.64e7 678.7 M + 2H+ 100-679.2 79.6 М 1357 (358.0 1380.0 522.3 555.4 1147.4_1176.8 1325.6 1384.6 1508.7 1545.9 649 4760.3 857.8 918.9 943.2 1059.2 476 1660.6 - m/z 1800 1900 200 800 1100 1200 1300 1400 1500 1600 1700 300 400 500 600 700 ann 1000

















3. Example Peptoids using Unprotected Tyr, Cys, and His Submonomers

4. Intermolecular, On-Resin DKP Formation

Figure S1. Proposed formation of diketopiperazines during the activation of a carboxylic acid peptoid side chain. An example of a putative DKP product formed between two L-phenylalanine carboxylic acid termini is shown. The side product was isolated by preparative HPLC and observed by LCMS. The extent of DKP formation appears to increase when the nucleophilicity of the desired synthon decreases. Notably this DKP formation results in termination of chain elongation.



5. Circular Dichroism of Compounds 1, 4 and 5.

All CD experiments were conducted using a Jasco J-1500 circular dichroism spectrometer.

The sign for the ellipticity of compound 5 is opposite of that for compounds 1 and 4, which we attribute to a distinct conformational influence resulting from the presence of this monomer at non-terminal residues. Variations in circular dichroism may also arise due to partial racemization of the side chains under basic conditions. The influence of stereochemistry on conformation will be the subject of additional study.



Figure S2. Circular Dichroism (CD) spectra of peptoids 1,4, and 5. Samples were prepared at 0.625 mM (~0.3-0.5 mg/mL) peptoid in acetonitrile and filtered (0.2 micron). 5 accumulations per sample were obtained at room temperature in a quartz cuvette (1mm pathlength). Solvent baseline was subtracted from the spectra and no smoothing functions were applied.