

Supplementary Information

Foldameric Receptors with Domain-Swapping Cavities Capable of Selectively Binding and Transporting Monosaccharides

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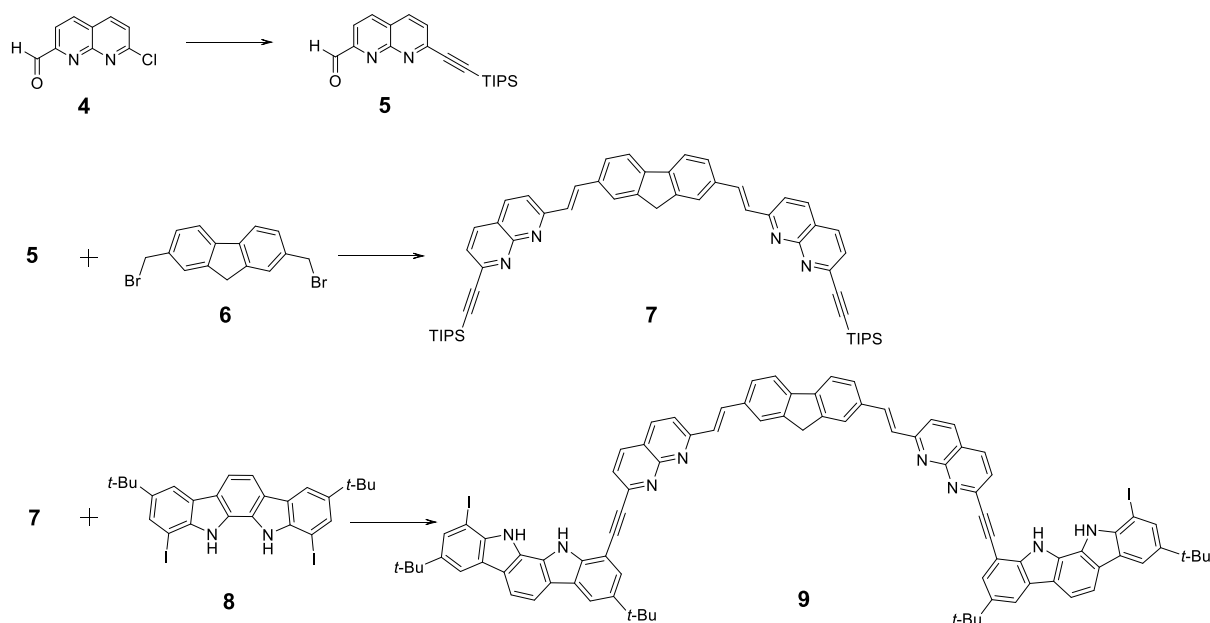
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General method: All chemicals were purchased from commercial suppliers and used without further purification unless otherwise specified. Dichloromethane (CH₂Cl₂) were purified by drying over calcium hydride (CaH₂), followed by distillation. Ethyl acetate (EtOAc) and acetone were distilled. Thin layer chromatography (TLC) was performed on Merck (silica gel 60, F-254, 0.25 mm). Silica gel 60 (230-400 mesh, Merck) was used for column chromatography. Melting points were determined with a Barnstead Electrothermal (IA9100) apparatus. NMR spectra were measured by using Bruker DRX 400, Avance II instruments. Abbreviations used when describing NMR spectra are as follows: s, singlet; d, doublet; dd, doublet of doublet; t, triplet; m, multiplet. Coupling constants (*J*) are given in Hz. Chemical shifts are reported using residual protonated solvent peaks (for ¹H NMR spectra, DMSO-*d*₆ 2.50 ppm; Acetone-*d*₆ 2.05 ppm; CD₂Cl₂ 5.32 ppm; CDCl₃ 7.32 ppm and for ¹³C NMR spectra, DMSO-*d*₆ 39.5 ppm; Acetone-*d*₆ 29.8 ppm; CD₂Cl₂ 53.8 ppm CDCl₃ 77.2 ppm). NMR data processing was achieved with Topspin 4.1.1 software. ESI-HRMS spectrometric measurements were obtained from the National Instrument Center for Environmental Mangement at Seoul National University. MALDI-TOF-HRMS spectra were recorded on a Bruker Autoflex Max MALDI-TOF/TOF MS (Bruker Daltonics, Bremen, Germany). Energy-minimized structures were obtained using *MacroModel 9.1 program* with MMFFs force field in CHCl₃ phase.

1. Syntheses and characterisation of new compounds



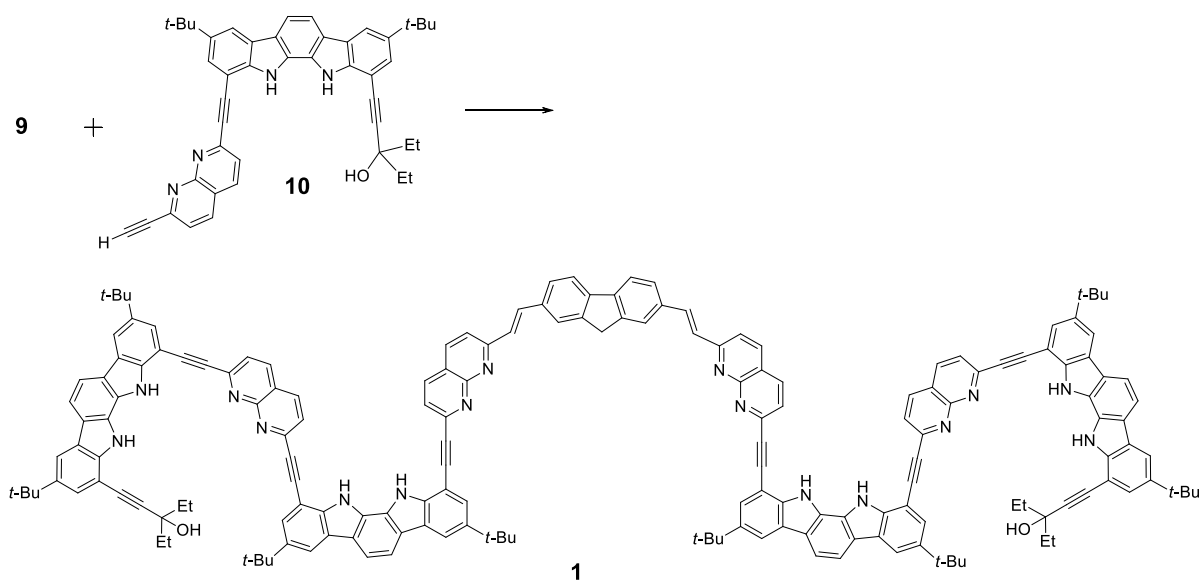
5: A Schlenk flask containing **4**^{S1} (220 mg, 1.14 mmol), CuI (6.5 mg, 0.034 mmol) and Pd(PPh₃)₂Cl₂ (24 mg, 0.034 mmol) was evacuated under vacuum and back-filled with nitrogen. Anhydrous, degassed tetrahydrofuran (THF) (4.0mL), triethylamine (Et₃N) (0.6 mL), and (triisopropylsilyl)acetylene (0.5 mL, 2.28 mmol) were sequentially added and the solution was stirred at 55 °C for 3.5 h. The reaction mixture was cooled to room temperature, filtered through

Celite with CH₂Cl₂ and concentrated. The residue was dissolved in CH₂Cl₂, washed with brine, and dried over anhydrous Na₂SO₄. The residue was concentrated and purified by flash column chromatography (silica gel, hexanes:CH₂Cl₂:ethyl acetate (EtOAc)= 1:1:0.2 (v/v/v)) to give compound **5** (200 mg, 52 %) as a grey solid; mp > 160 °C (dec); ¹H NMR (400 MHz, Acetone-*d*₆, 25 °C, ppm) δ 10.18 (d, *J* = 0.9 Hz, 1H), 8.68 (dd, *J* = 8.3 Hz, 0.9 Hz, 1H), 8.58 (d, *J* = 8.4 Hz, 1H), 8.09 (d, *J* = 8.3 Hz, 1H), 7.87 (d, *J* = 8.4 Hz, 1H), 1.23–1.08 (m, 21H); ¹³C NMR (100 MHz, Acetone-*d*₆, 25 °C, ppm) δ 194.1, 156.5, 156.4, 148.2, 140.2, 139.0, 128.1, 125.6, 118.9, 107.2, 94.9, 18.9, 11.9; ESI-HRMS, *m/z* calcd for C₂₀H₂₆N₂OSi [M+H]⁺ 339.1887 found 339.1890

7: A round bottom flask containing **6**^{S2} (76 mg, 0.216 mmol), triphenylphosphin (141 mg, 0.54 mmol) and NaH (24 mg, 0.45 mmol) was evacuated under vacuum and back-filled with nitrogen. Anhydrous dimethylformamide (DMF) was added and the solution as stirred at 40 °C. After 30 min, **5** (146 mg, 0.43 mmol) was added and stirred for 4h. The reaction mixture was cooled to room temperature, diluted through ethyl acetate, and washed with brine. After dried over anhydrous Na₂SO₄, the residue was concentrated and purified by flash column chromatography (silica gel, hexanes:CH₂Cl₂:tetrahydrofuran (THF) = 4:4:1 (v/v/v)) to give compound **7** (80 mg, 44 %) as a yellow solid; mp > 225 °C (dec); ¹H NMR (400 MHz, Acetone-*d*₆, 25 °C, ppm) δ 8.39 (d, *J* = 8.3 Hz, 1H), 8.38 (d, *J* = 8.3 Hz, 1H), 8.12 (d, *J* = 16.1 Hz, 1H), 8.00 (s, 1H), 7.97 (d, *J* = 8.1 Hz, 1H), 7.87 (d, *J* = 8.1 Hz, 1H), 7.82 (d, *J* = 8.2 Hz, 1H), 7.67 (d, *J* = 8.1 Hz, 1H), 7.57 (d, *J* = 16.2 Hz, 1H), 4.05 (s, 1H), 1.23–1.20 (m, 21H); ¹³C NMR (100 MHz, CDCl₃, 25 °C, ppm) δ 159.8, 156.1, 147.1, 144.6, 142.5, 137.1, 136.9, 136.6, 135.5, 127.3, 127.2, 125.4, 124.1, 121.9, 121.3, 120.6, 106.4, 94.7, 18.7, 11.5; ESI-HRMS, *m/z* calcd for C₅₅H₆₂N₄Si₂ [M+H]⁺ 835.4586 found 835.4593

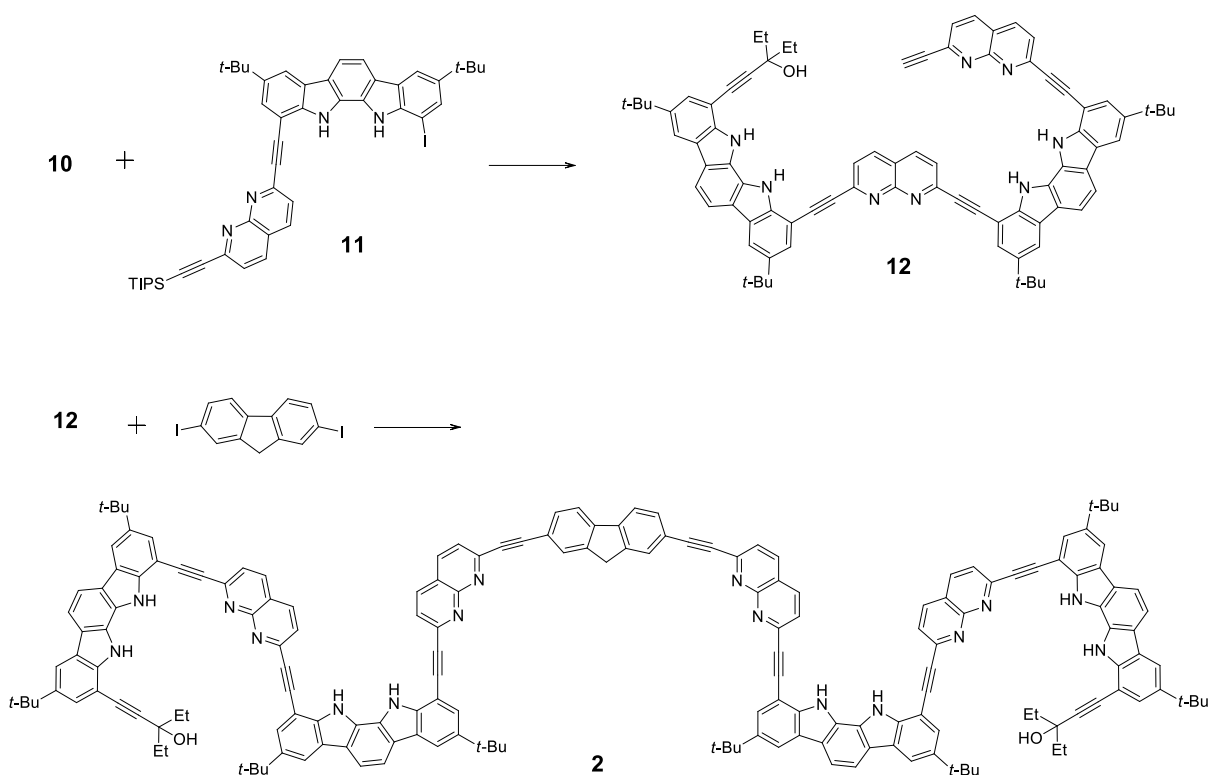
9: A Schlenk flask containing **7** (53 mg, 0.063 mmol), **8**^{S3} (305 mg, 0.51 mmol), CuI (1.2 mg, 0.006 mmol), and Pd(PPh₃)₂Cl₂ (4.4 mg, 0.006 mmol) was evacuated under vacuum and back-filled with nitrogen. Anhydrous, degassed dimethylformamide (DMF) (1.2 mL), triethylamine (Et₃N) (0.2 mL), and tetrabutylammonium fluoride (0.14 mL, 1.0 M solution in THF) were sequentially added, and the solution was stirred at 45 °C for 2.5 h. The mixture was cooled to room temperature, filtered through Celite with THF, and concentrated. The residue was dissolved in ethyl acetate, washed with brine, and dried over anhydrous Na₂SO₄. After concentrated, the residue was purified by flash column chromatography (silica gel, hexanes : THF : MeOH : TEA = 10:10:1:1 (v/v/v/v)) to give compound **9** (59 mg, 62 %) as a yellow

solid; mp > 240 °C (dec); ¹H NMR (400 MHz, DMSO-*d*₆, 25 °C, ppm) δ 11.49 (s, NH, 1H), 11.13 (s, NH, 1H), 8.56 (d, *J* = 8.2 Hz, 1H), 8.53 (d, *J* = 8.3 Hz, 1H), 8.40 (d, *J* = 1.4 Hz, 1H), 8.24 (s, 1H), 8.12 (d, *J* = 16.1 Hz, 1H), 8.09–7.99 (m, 6H), 7.86 (d, *J* = 8.3 Hz, 1H), 7.83 (d, *J* = 1.3 Hz, 1H), 7.82 (d, *J* = 1.3 Hz, 1H), 7.66 (d, *J* = 16.2 Hz, 1H), 4.13 (s, 1H), 1.49 (s, *t*-Bu, 9H), 1.43 (s, *t*-Bu, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆, 25 °C, ppm) δ 159.5, 155.6, 146.0, 144.4, 143.9, 142.2, 141.8, 139.1, 137.8, 135.1, 130.7, 128.8, 127.8, 127.2, 126.3, 125.9, 125.5, 124.0, 123.8, 121.2, 121.1, 120.4, 118.9, 116.3, 112.7, 112.4, 103.0, 93.2, 88.1, 83.7, 76.7, 72.2, 60.2, 34.6, 34.4, 31.8, 31.7; MALDI-TOF-HRMS, *m/z* calcd for C₈₉H₇₂I₂N₈ [M+H]⁺ 1507.4042 found 1507.4029



1: A Schlenk flask containing **9** (80 mg, 0.053 mmol), **10**^{S4} (76 mg, 0.12 mmol), CuI (1.0 mg, 0.0053 mmol), and Pd(PPh₃)₂Cl₂ (3.7 mg, 0.0053 mmol) was evacuated under vacuum and back-filled with nitrogen. Anhydrous, degassed dimethylformamide (DMF) (1.0 mL), and triethylamine (Et₃N) (0.1 mL) were sequentially added, and the solution was stirred at 45 °C for 2.5 h. The mixture was cooled to room temperature, filtered through Celite with CH₂Cl₂, and concentrated. The residue was dissolved in ethyl acetate, washed with brine, and dried over anhydrous Na₂SO₄. After concentrated, the residue was purified by flash column chromatography (silica gel, hexanes : THF : MeOH : TEA = 10:10:1:1 (v/v/v/v)) to give compound **1** (71 mg, 52 %) as a yellow solid; mp > 275 °C (dec); ¹H NMR (400 MHz, DMSO-*d*₆, 25 °C, ppm) δ 11.63 (s, 1H), 11.45 (s, 1H), 11.38 (s, 1H), 10.74 (s, 1H), 8.57 (d, *J* = 8.4 Hz, 1H), 8.45 (s, 1H), 8.43 (s, 1H), 8.35 (d, *J* = 1.4 Hz, 1H), 8.35 (d, *J* = 8.3 Hz, 1H), 8.18 (d, *J* = 1.4 Hz, 1H), 8.14 (s, 4H), 8.01 (d, *J* = 8.3 Hz, 1H), 7.96 (d, *J* = 8.4 Hz, 1H), 7.88 (d, *J* = 8.3

Hz, 1H), 7.94 (d, $J = 1.5$ Hz, 1H), 7.82 (s, 1H), 7.80 (d, $J = 7.5$ Hz, 1H), 7.79 (d, $J = 8.3$ Hz, 1H), 7.76 (d, $J = 8.2$ Hz, 1H), 7.68 (d, $J = 16.1$ Hz, 1H), 7.66 (d, $J = 1.4$ Hz, 1H), 7.63 (s, 1H), 7.45 (d, $J = 7.5$ Hz, 1H), 7.43 (d, $J = 1.6$ Hz, 1H), 7.31 (d, $J = 8.3$ Hz, 1H), 7.21 (d, $J = 16.2$ Hz, 1H), 5.18 (s, OH), 3.70 (s, 1H), 1.75–1.62 (m, 4H), 1.50 (s, 18H, *t*-Bu), 1.44 (s, 9H, *t*-Bu), 1.38 (s, 9H, *t*-Bu), 0.98 (t, $J = 7.3$ Hz, 6H); ^{13}C NMR (100 MHz, DMSO- d_6 , 25 °C, ppm) δ 158.9, 155.3, 155.2, 146.7, 145.7, 143.9, 142.2, 142.1, 141.8, 141.7, 141.5, 138.0, 137.9, 137.8, 137.1, 134.7, 127.1, 126.3, 126.0, 125.9, 125.9, 125.8, 125.7, 124.2, 124.1, 123.7, 121.2, 120.8, 120.7, 120.6, 120.4, 119.1, 118.8, 116.7, 112.5, 112.3, 112.1, 105.0, 103.2, 102.9, 97.5, 93.2, 93.1, 92.9, 89.4, 89.0, 87.9, 79.7, 70.9, 34.5, 34.4, 34.3, 33.8, 31.7, 31.6, 29.0, 20.7, 8.7; ESI-HRMS, m/z calcd for $\text{C}_{179}\text{H}_{154}\text{N}_{16}\text{O}_2$ [$\text{M}+2\text{H}$] $^{2+}$ 1280.6293 found 1280.6277



12: A Schlenk flask containing **10** (300 mg, 0.46 mmol), **11**^{SS} (380 mg, 0.46 mmol), CuI (4.4 mg, 0.023 mmol), and Pd(PPh₃)₂Cl₂ (16 mg, 0.023 mmol) was evacuated under vacuum and back-filled with nitrogen. Anhydrous, degassed dimethylformamide (DMF) (3.0 mL), and triethylamine (Et₃N) (0.26 mL) were sequentially added, and the solution was stirred at 45 °C for 3 h. The mixture was cooled to room temperature, filtered through Celite with CH₂Cl₂, and concentrated. The residue was dissolved in ethyl acetate, washed with brine, and dried over anhydrous Na₂SO₄. After concentrated, the residue was dissolved in THF (6 mL) and added dropwise TBAF (0.6 mL, 1.0 M solution in THF). After 30 min stirring in room temperature,

the residue was concentrated and purified by flash column chromatography (silica gel, CH₂Cl₂ : MeOH = 50:1 (v/v)) to give compound **12** (450 mg, 81 %) as a yellow solid; mp > 214 °C (dec); ¹H NMR (400 MHz, DMSO-*d*₆, 25 °C, ppm) δ 11.56 (s, 1H), 11.53 (s, 1H), 11.46 (s, 1H), 10.88 (s, 1H), 8.70 (d, *J* = 8.4 Hz, 1H), 8.69 (d, *J* = 8.3 Hz, 1H), 8.44 (s, 1H), 8.43 (s, , 2H), 8.39 (d, *J* = 8.4 Hz, 1H), 8.24 (s, 1H), 8.18 (d, *J* = 8.2 Hz, 1H), 8.15 (d, *J* = 8.1 Hz, 1H), 8.13 (d, *J* = 8.0 Hz, 1H), 8.12 (s, 2H), 8.08 (d, *J* = 8.2 Hz, 1H), 8.05 (d, *J* = 8.2 Hz, 1H), 8.00 (d, *J* = 8.3 Hz, 1H), 7.83 (s, 1H), 7.81 (s, 2H), 7.47 (d, *J* = 8.1 Hz, 1H), 7.45 (d, *J* = 1.4 Hz, 1H), 5.26 (s, 1H), 4.58 (s, 1H), 1.77–1.67 (m, 4H), 1.50 (s, 9H, *t*-Bu), 1.48 (s, 18H, *t*-Bu), 1.42 (s, 9H, *t*-Bu), 1.01 (t, *J* = 7.5 Hz, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆, 25 °C, ppm) δ 155.4, 154.9, 146.8, 146.7, 146.6, 145.7, 142.2, 142.1, 141.8, 138.2, 138.0, 137.9, 137.9, 137.8, 137.2, 126.4, 126.2, 126.1, 126.0, 125.9, 125.7, 125.3, 124.2, 124.1, 123.7, 121.5, 121.4, 120.8, 120.7, 120.5, 119.0, 119.0, 116.8, 112.5, 112.4, 112.2, 105.0, 102.9, 102.8, 97.6, 93.0, 93.0, 92.8, 89.3, 89.1, 88.8, 83.0, 82.6, 79.6, 70.9, 37.6, 34.6, 34.4, 33.8, 31.7, 17.8, 12.0, 8.7; ESI-HRMS, *m/z* calcd for C₈₃H₇₂N₈O [M+H]⁺ 1197.5902 found 1197.5916

2: A Schlenk flask containing **12** (292 mg, 0.244 mmol), 2,7-diiodo-9H-fluorene (49 mg, 0.116 mmol), CuI (2.2 mg, 0.0116 mmol), and Pd(PPh₃)₂Cl₂ (8.1 mg, 0.0116 mmol) was evacuated under vacuum and back-filled with nitrogen. Anhydrous, degassed dimethylformamide (DMF) (1.5 mL), and triethylamine (Et₃N) (0.2 mL) were sequentially added, and the solution was stirred at 45 °C for 3.5 h. The mixture was cooled to room temperature, filtered through Celite with CH₂Cl₂, and concentrated. The residue was dissolved in ethyl acetate, washed with brine, and dried over anhydrous Na₂SO₄. After concentrated, the residue was purified by flash column chromatography (silica gel, hexanes : CH₂Cl₂ : THF = 4:4:1 (v/v/v)) to give compound **2** (160 mg, 54 %) as a yellow solid; mp > 265 °C (dec); ¹H NMR (400 MHz, DMSO-*d*₆, 25 °C, ppm) δ 11.56 (s, 1H), 11.44 (s, 2H), 10.78 (s, 1H), 8.61 (d, *J* = 8.4 Hz, 1H), 8.47 (d, *J* = 8.4 Hz, 1H), 8.43 (s, 1H), 8.42 (s, 1H), 8.38 (d, *J* = 1.5 Hz, 1H), 8.21 (d, *J* = 8.4 Hz, 1H), 8.18 (d, *J* = 1.6 Hz, 1H), 8.11 (d, *J* = 8.4 Hz, 1H), 8.04 (d, *J* = 8.3 Hz, 1H), 7.98 (d, *J* = 8.3 Hz, 1H), 7.98 (d, *J* = 8.3 Hz, 1H), 7.89 (d, *J* = 8.3 Hz, 1H), 7.86 (d, *J* = 8.3 Hz, 1H), 7.83 (d, *J* = 7.5 Hz, 1H), 7.83 (d, *J* = 1.5 Hz, 1H), 7.81 (d, *J* = 1.6 Hz, 1H), 7.72 (d, *J* = 1.6 Hz, 1H), 7.66 (s, 1H), 7.44 (d, *J* = 1.6 Hz, 1H), 7.42 (d, *J* = 7.5 Hz, 1H), 7.29 (d, *J* = 8.3 Hz, 1H), 5.21 (s, OH), 3.78 (s, 1H), 1.76–1.64 (m, 4H), 1.49 (s, 18H, *t*-Bu), 1.43 (s, 9H, *t*-Bu), 1.37 (s, 9H, *t*-Bu), 1.00 (t, *J* = 7.3 Hz, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆, 25 °C, ppm) δ 155.8, 155.5, 147.3, 147.2, 146.9, 146.7, 144.2, 142.7, 142.6, 142.4, 142.2, 142.0, 138.6, 138.4, 138.4, 138.3,

138.2, 137.9, 137.6, 131.4, 129.0, 126.9, 126.5, 126.4, 126.4, 126.2, 125.5, 124.7, 124.6, 124.6, 124.2, 121.8, 121.4, 121.3, 121.2, 121.1, 120.9, 120.0, 119.5, 119.4, 117.2, 113.1, 112.8, 112.6, 105.4, 103.4, 103.4, 103.3, 98.0, 93.6, 93.4, 92.1, 90.1, 90.0, 89.5, 89.3, 80.1, 71.4, 35.0, 34.9, 34.8, 34.3, 33.7, 32.2, 32.1, 23.1, 15.2, 9.2; ESI-HRMS, m/z calcd for $C_{179}H_{150}N_{16}O_2 [M+2H]^{2+}$ 1278.6137 found 1278.6130

2. 1H NMR studies

2.1 1H NMR spectra of receptors **1** and **2** in the presence of various guests

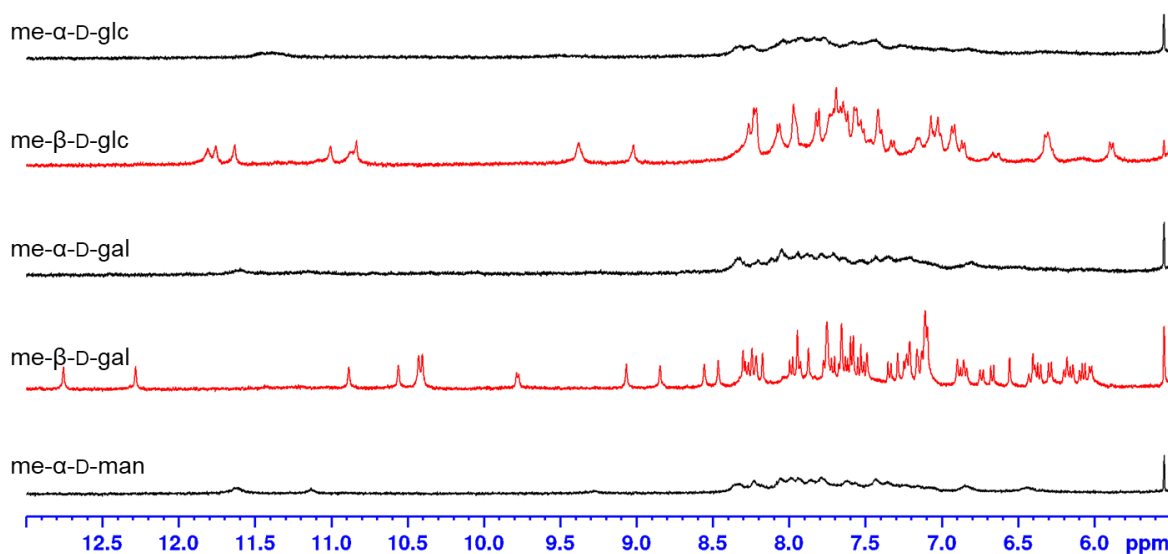


Fig. S1 Partial 1H NMR spectra (400 MHz, 25 $^\circ C$) of receptor **1** in the presence of various guests (2.0 equiv of each guest) in 5% (v/v) $DMSO-d_6/CD_2Cl_2$ (containing ca. 0.05 % water).

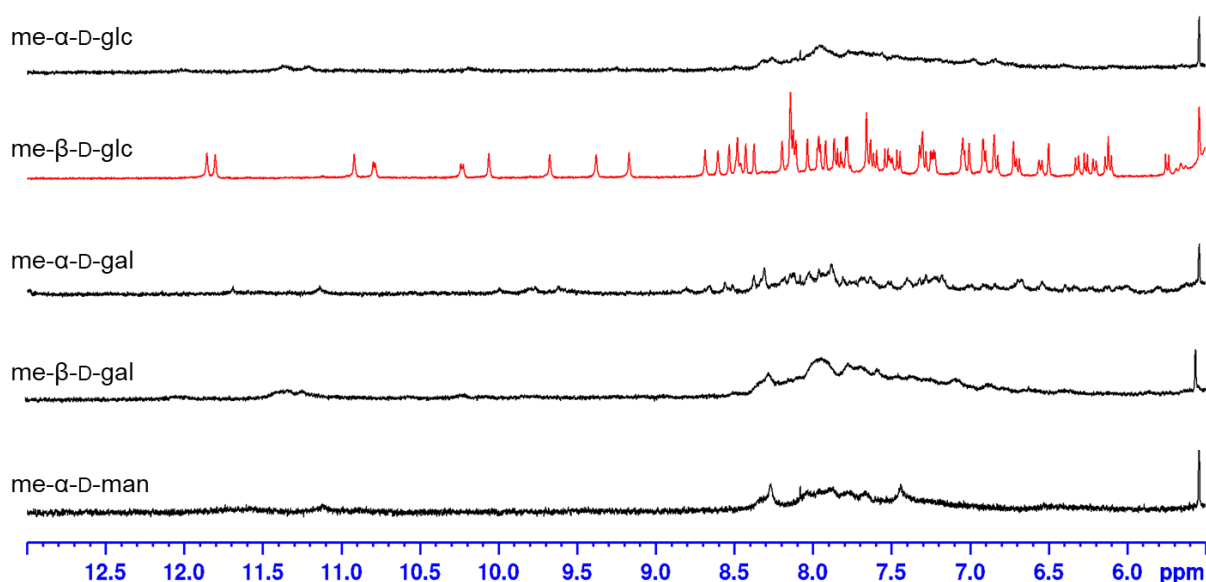


Fig. S2 Partial 1H NMR spectra (400 MHz, 25 $^\circ C$) of receptor **2** in the presence of various guests (2.0 equiv of each guest) in 5% (v/v) $DMSO-d_6/CD_2Cl_2$ (containing ca. 0.05 % water).

2.2 ^1H NMR spectra of **1**, $(1\text{-MM})_2\supset(\text{me-}\beta\text{-D-gal}\cdot 2\text{H}_2\text{O})_2$, and $(1\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$

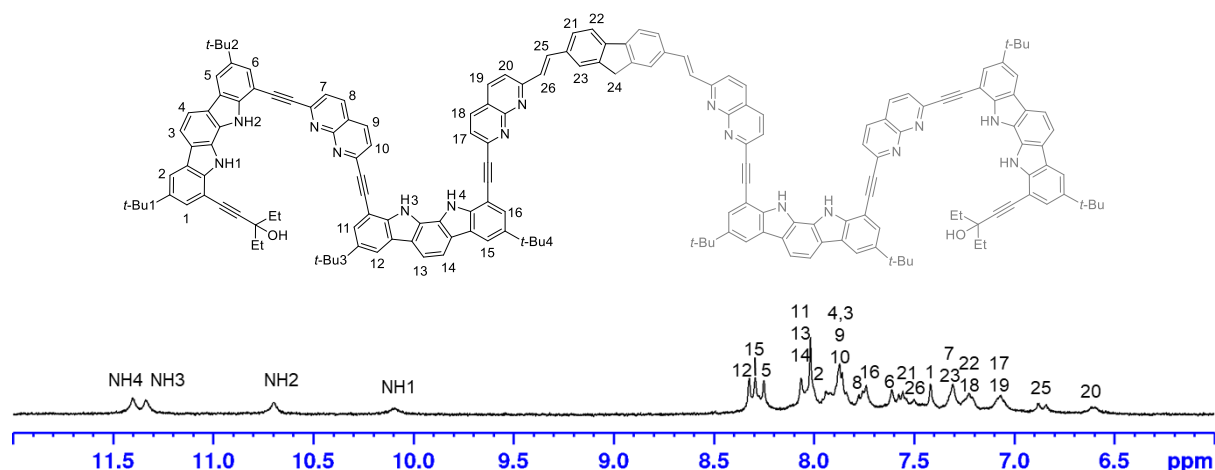


Fig. S3 Partial ^1H NMR spectrum (400 MHz, 25 °C) of **1** (2 mM) in 5% (v/v) $\text{DMSO-}d_6/\text{CD}_2\text{Cl}_2$.

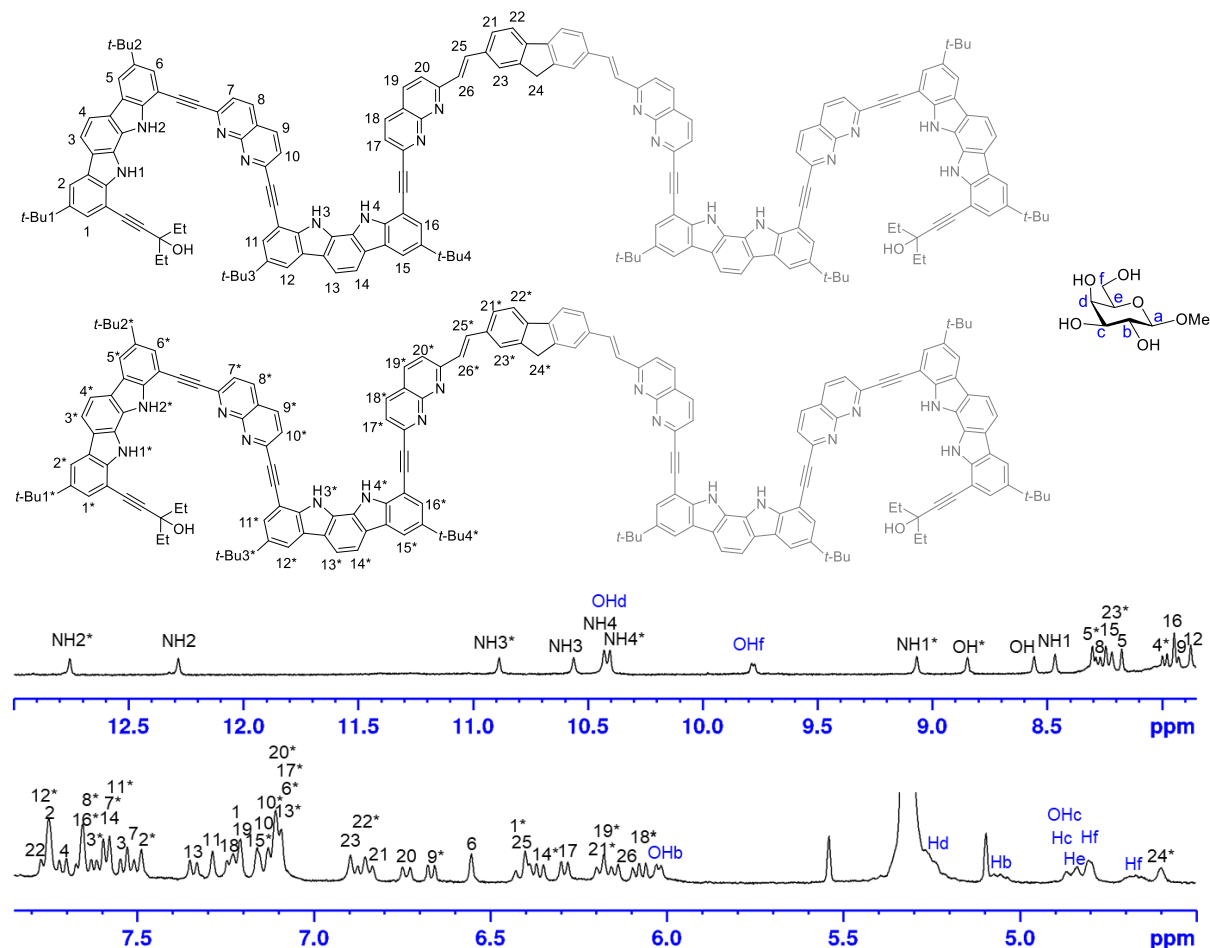


Fig. S4 Partial ^1H NMR spectrum (400 MHz, 25 °C) of $(1\text{-MM})_2\supset(\text{me-}\beta\text{-D-gal}\cdot 2\text{H}_2\text{O})_2$ (2 mM) in 5% (v/v) $\text{DMSO-}d_6/\text{CD}_2\text{Cl}_2$. Me- β -D-gal signals are depicted in blue.

Table S1 ^1H NMR chemical shifts (400 MHz, 25 °C) of $(1-MM)_2\supset(\text{me-}\beta\text{-D-gal}\cdot 2\text{H}_2\text{O})_2$ in 5% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$ and their differences between free **1**.

$(1-MM)_2\supset(\text{me-}\beta\text{-D-gal}\cdot 2\text{H}_2\text{O})_2$			$(1-MM)_2\supset(\text{me-}\beta\text{-D-gal}\cdot 2\text{H}_2\text{O})_2$		
atom	δ (ppm)	δ (free) – δ (complex)	atom	δ (ppm)	δ (free) – δ (complex)
1	7.24 (d, $J = 1.4$ Hz)	0.18	1*	6.43 (d, $J = 1.5$ Hz)	0.99
2	7.76 (d, $J = 1.5$ Hz)	0.16	2*	7.51 (d, $J = 1.5$ Hz)	0.41
3	7.55 (d, $J = 8.1$ Hz)	0.32	3*	7.66 (d, $J = 8.0$ Hz)	0.21
4	7.73 (d, $J = 7.9$ Hz)	0.14	4*	8.00 (d, $J = 8.0$ Hz)	-0.13
5	8.19 (d, $J = 1.4$ Hz)	0.06	5*	8.31 (d, $J = 1.6$ Hz)	-0.06
6	6.56 (d, $J = 1.4$ Hz)	1.05	6*	7.10 (s)	0.51
7	7.54 (d, $J = 8.2$ Hz)	-0.24	7*	7.61 (d, $J = 7.9$ Hz)	-0.31
8	8.29 (d, $J = 8.1$ Hz)	-0.53	8*	7.68 (d, $J = 7.9$ Hz)	0.08
9	7.96 (d, $J = 8.1$ Hz)	-0.11	9*	6.67 (d, $J = 8.0$ Hz)	1.18
10	7.15 (d, $J = 7.9$ Hz)	0.70	10*	7.14 (d, $J = 8.2$ Hz)	0.71
11	7.31 (s)	0.71	11*	7.60 (s)	0.42
12	7.89 (d, $J = 1.4$ Hz)	0.43	12*	7.77 (d, $J = 1.6$ Hz)	0.55
13	7.36 (d, $J = 8.1$ Hz)	0.66	13*	7.11 (d, $J = 8.0$ Hz)	0.91
14	7.62 (d, $J = 8.1$ Hz)	0.40	14*	6.37 (d, $J = 8.0$ Hz)	1.65
15	8.26 (d, $J = 1.5$ Hz)	0.03	15*	7.18 (d, $J = 1.4$ Hz)	1.11
16	7.97 (d, $J = 1.5$ Hz)	-0.23	16*	7.68 (d, $J = 1.7$ Hz)	0.06
17	6.31 (d, $J = 7.9$ Hz)	0.75	17*	7.11 (d, $J = 7.8$ Hz)	-0.05
18	7.25 (d, $J = 7.9$ Hz)	-0.03	18*	6.09 (d, $J = 8.0$ Hz)	1.13
19	7.23 (d, $J = 8.2$ Hz)	-0.17	19*	6.19 (d, $J = 8.0$ Hz)	0.87
20	6.76 (d, $J = 8.6$ Hz)	-0.14	20*	7.11 (d, $J = 8.2$ Hz)	-0.49
21	6.86 (d, $J = 7.8$ Hz)	0.70	21*	6.21 (d, $J = 7.4$ Hz)	1.35
22	7.78 (d, $J = 8.0$ Hz)	-0.56	22*	6.89 (d, $J = 7.9$ Hz)	0.33
23	6.91 (s)	0.39	23*	8.25 (s)	-0.95
24	3.84 (s)	-0.36	24*	4.63 (s)	-1.15
25	6.42 (d, $J = 16.5$ Hz)	0.44	26	6.13 (d, $J = 16.5$ Hz)	1.40
OMe	2.41 (s)		Ha	2.93 (d, $J = 8.0$ Hz)	
OHb	6.08 (d, $J = 5.6$ Hz)		Hb	5.08	
OHc	4.85		Hc	4.88	
OHd	10.48 (d, $J = 6.2$ Hz)		Hd	5.27	
OHf	9.83 (d, $J = 5.0$ Hz)		He	4.85	
Hf	4.84		Hf	4.68	

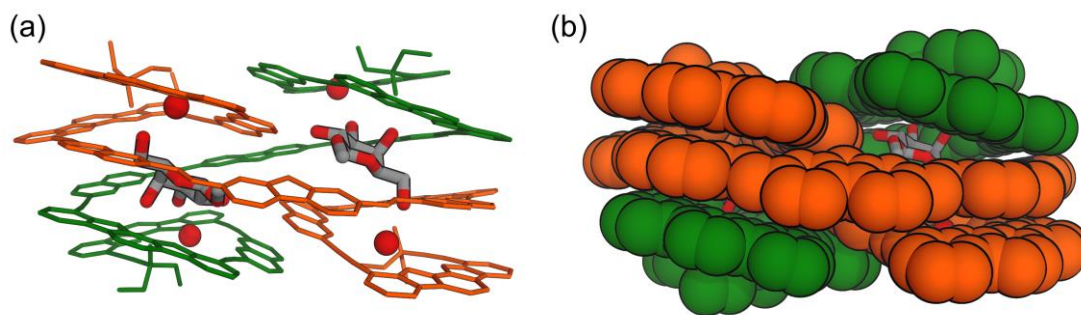


Fig. S5 (a) Tube and (b) space-filling representations of energy-minimized structure of $(1-MM)_2 \supset (me-\beta-D-gal \cdot 2H_2O)_2$ (MacroModel 9.1, MMFFs force field, $CHCl_3$ phase).

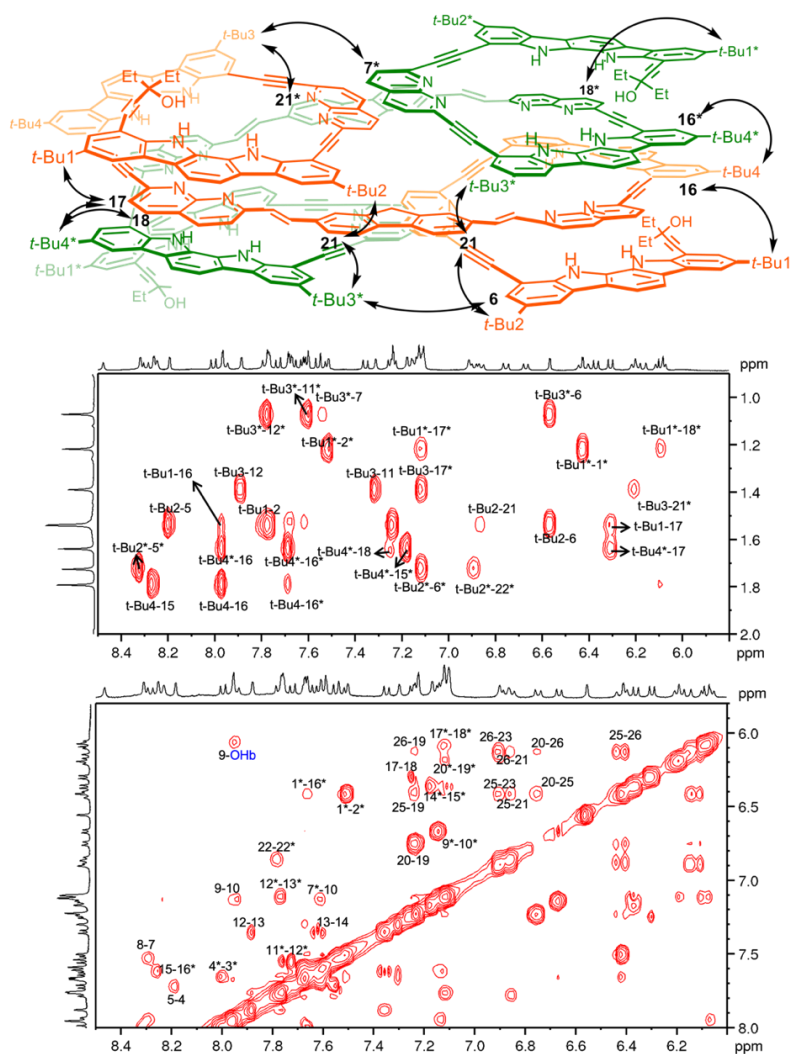


Fig. S6 Partial $^1H-^1H$ ROESY spectrum (400 MHz, 25 °C, mixing time: 400 ms) of $(1-MM)_2 \supset (me-\beta-D-gal \cdot 2H_2O)_2$ (4.0 mM) in 2% (v/v) $DMSO-d_6/CD_2Cl_2$.

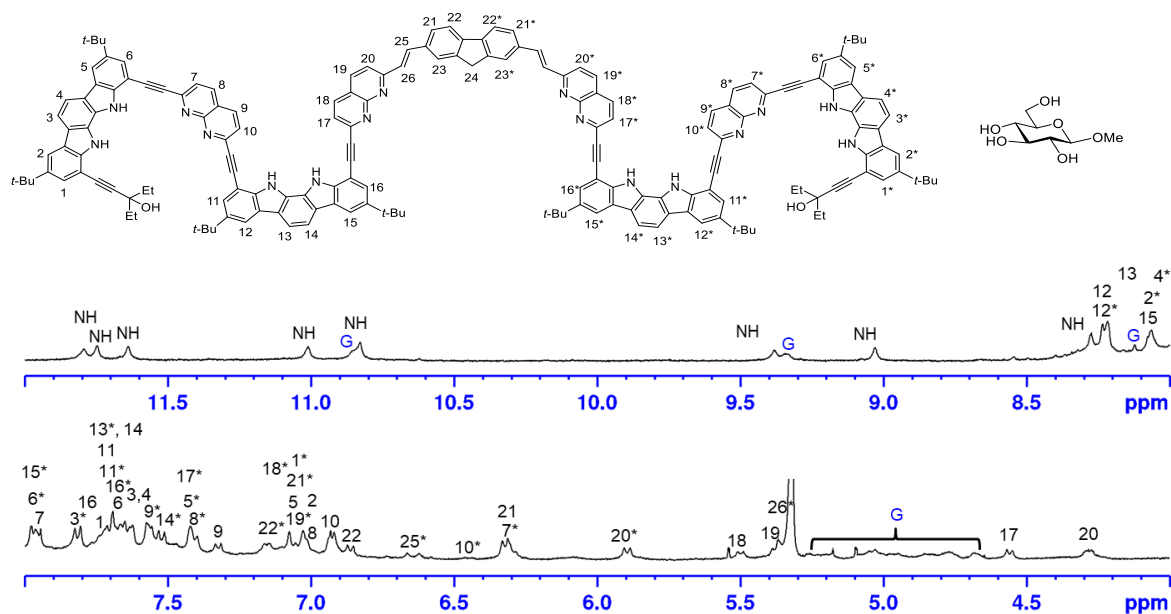


Fig. S7 Partial ^1H NMR spectrum (400 MHz, 25 °C) of $(1\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$ (2 mM) in 5% (v/v) $\text{DMSO-}d_6/\text{CD}_2\text{Cl}_2$. Me- β -D-glc signals are depicted in blue G.

Table S2 ^1H NMR chemical shifts (400 MHz, 25 °C) of $(1\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$ in 5% (v/v) $\text{DMSO-}d_6/\text{CD}_2\text{Cl}_2$ and their differences between free **1**.

$(1\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$			$(1\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$		
atom	δ (ppm)	δ (free) – δ (complex)	atom	δ (ppm)	δ (free) – δ (complex)
1	7.65 (s)	-0.23	1*	7.05 (s)	0.37
2	7.00 (s)	0.92	2*	8.05 (s)	-0.13
3	7.60	0.27	3*	7.82 (d, $J = 8.1$ Hz)	0.05
4	7.60	0.27	4*	8.03	-0.16
5	7.10 (s)	1.15	5*	7.40 (s)	0.85
6	7.70 (s)	-0.09	6*	7.95 (s)	-0.34
7	7.95	-0.65	7*	6.35 (d, $J = 8.2$ Hz)	0.95
8	7.05 (d, $J = 8.1$ Hz)	0.71	8*	7.38 (d, $J = 8.0$ Hz)	0.38
9	7.33 (d, $J = 8.0$ Hz)	0.52	9*	7.55 (d, $J = 8.1$ Hz)	0.3
10	6.95 (d, $J = 8.1$ Hz)	0.9	10*	6.45	1.4
11	7.70	0.32	11*	7.70	0.32
12	8.25 (s)	0.07	12*	8.25 (s)	0.07
13	8.15	-0.13	13*	7.75	0.27
14	7.75	0.27	14*	7.55 (d, $J = 8.1$ Hz)	0.47
15	8.05 (s)	0.24	15*	7.95 (s)	0.34
16	7.80 (s)	-0.06	16*	7.70 (s)	0.04
17	4.55 (d, $J = 8.0$ Hz)	2.51	17*	7.43	-0.37
18	5.50 (d, $J = 8.1$ Hz)	1.72	18*	7.10	0.12
19	5.38 (d, $J = 8.1$ Hz)	1.68	19*	7.10 (d, $J = 8.2$ Hz)	-0.04
20	4.28 (d, $J = 8.0$ Hz)	2.34	20*	5.90 (d, $J = 8.1$ Hz)	0.72
21	6.30 (d, $J = 7.9$ Hz)	1.26	21*	7.05	0.51
22	6.87 (d, $J = 7.9$ Hz)	0.35	22*	7.15	0.07
24	3.09 (s)	0.39	24*	3.07 (s)	0.41
25*	6.65 (d, $J = 16.0$ Hz)	0.21	26*	5.35	2.18

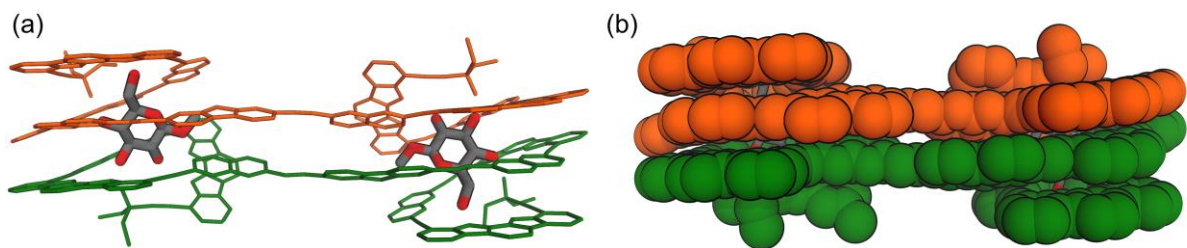


Fig. S8 (a) Tube and (b) space-filling representations of energy-minimized structure of $(1-MP)_2 \supset (me-\beta-D-glc)_2$ (MacroModel 9.1, MMFFs force field, $CHCl_3$ phase).

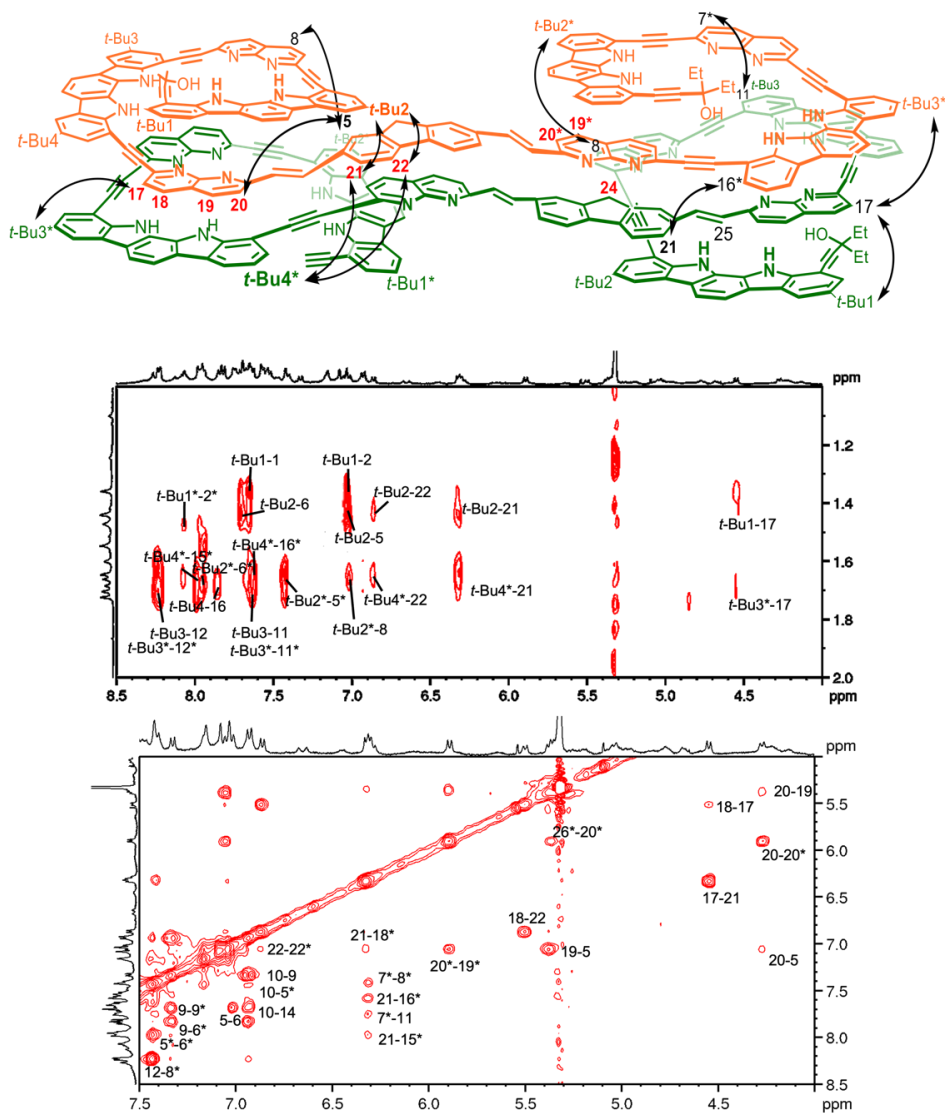


Fig. S9 Partial 1H - 1H ROESY spectrum (400 MHz, 25 °C, mixing time: 400 ms) of $(1-MP)_2 \supset (me-\beta-D-glc)_2$ (4.0 mM) in 3% (v/v) $DMSO-d_6/CD_2Cl_2$.

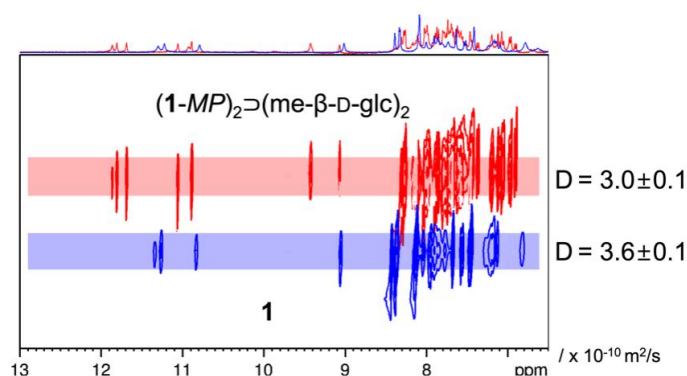


Fig. S10 Partial DOSY spectra of **1** in the absence and presence of me- β -D-glc (2 mM) in 2% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$.

2.3 ^1H NMR spectra of **2**, and $(2\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$

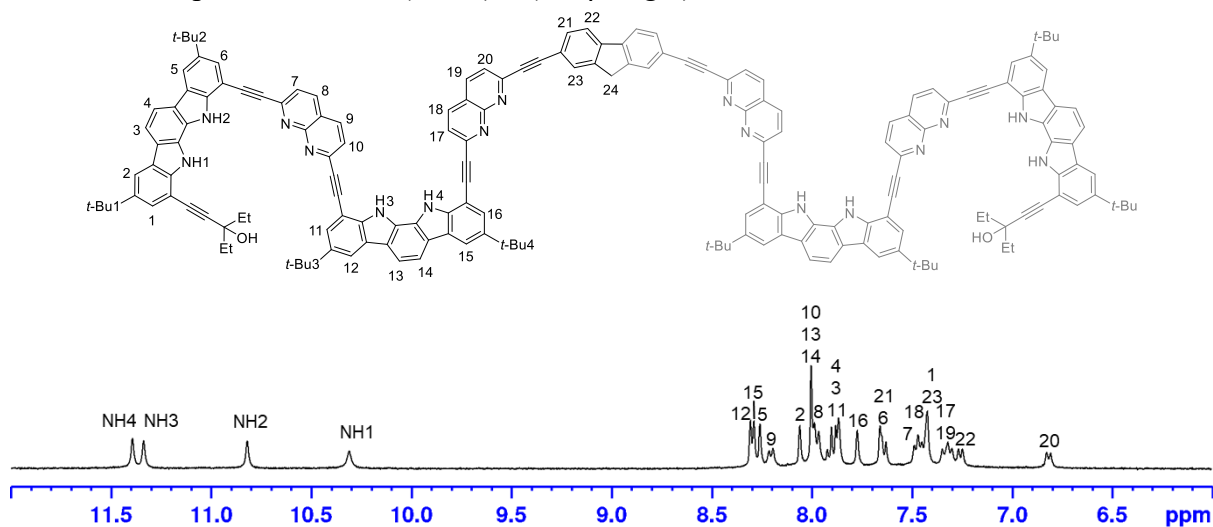


Fig. S11 Partial ^1H NMR spectrum (400 MHz, 25 °C) of **2** (2 mM) in 5% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$.

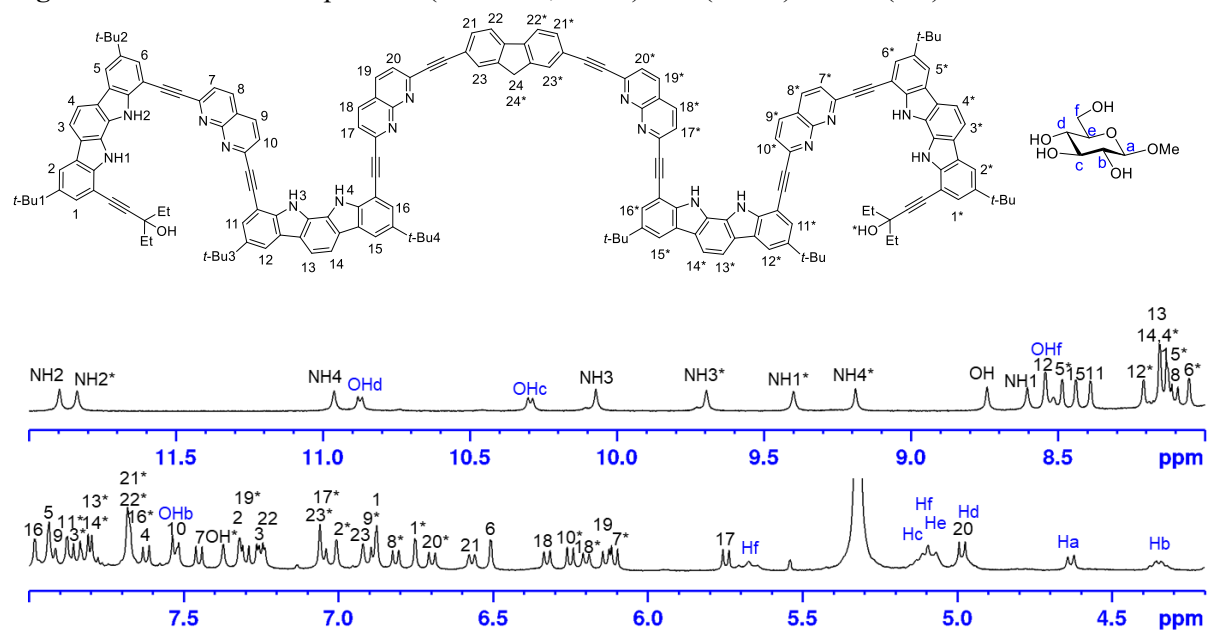


Fig. S12 Partial ^1H NMR spectrum (400 MHz, 25 °C) of $(2\text{-MP})_2\supset(\text{me-}\beta\text{-D-glc})_2$ (2 mM) in 5% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$. Me- β -D-glc signals are depicted in blue.

Table S3 ¹H NMR chemical shifts (400 MHz, 25 °C) of (2-MP)₂⊃(me-β-D-glc)₂ (2 mM) in 5% (v/v) DMSO-*d*₆/CD₂Cl₂ and their differences between free **2**.

(2-MP) ₂ ⊃(me-β-D-glc) ₂			(2-MP) ₂ ⊃(me-β-D-glc) ₂		
atom	δ (ppm)	δ (free) - δ (complex)	atom	δ (ppm)	δ (free) - δ (complex)
1	6.88 (s)	0.54	1*	6.75 (s)	0.67
2	7.32 (s)	0.74	2*	7.00 (s)	1.06
3	7.25 (d, <i>J</i> = 8.2 Hz)	0.62	3*	7.84 (d, <i>J</i> = 8.2 Hz)	0.03
4	7.62 (d, <i>J</i> = 8.1 Hz)	0.29	4*	8.15 (s)	-0.24
5	7.93 (s)	0.33	5*	8.48 (s)	-0.22
6	6.51 (s)	1.15	6*	8.05 (s)	-0.39
7	7.45 (d, <i>J</i> = 8.1 Hz)	0.03	7*	6.11 (d, <i>J</i> = 8.0 Hz)	1.37
8	8.10 (d, <i>J</i> = 8.1 Hz)	-0.12	8*	6.81 (d, <i>J</i> = 8.1 Hz)	1.17
9	7.92 (d, <i>J</i> = 8.2 Hz)	0.28	9*	6.88 (d, <i>J</i> = 8.0 Hz)	1.32
10	7.53 (d, <i>J</i> = 8.1 Hz)	0.47	10*	6.25 (d, <i>J</i> = 8.1 Hz)	1.75
11	8.39 (s)	-0.53	11*	7.88 (d, <i>J</i> = 1.5 Hz)	-0.02
12	8.54 (s)	-0.24	12*	8.21 (s)	0.09
13	8.15 (s)	-0.15	13*	7.78 (d, <i>J</i> = 8.2 Hz)	0.22
14	8.15 (s)	-0.15	14*	7.82 (d, <i>J</i> = 8.1 Hz)	0.18
15	8.44 (s)	-0.15	15*	8.13 (s)	0.16
16	7.98 (s)	-0.21	16*	7.68 (s)	0.09
17	5.75 (d, <i>J</i> = 8.0 Hz)	1.56	17*	7.05 (d, <i>J</i> = 8.0 Hz)	0.26
18	6.32 (d, <i>J</i> = 8.0 Hz)	1.14	18*	6.20 (d, <i>J</i> = 8.1 Hz)	1.26
19	6.13 (d, <i>J</i> = 8.1 Hz)	1.17	19*	7.30 (d, <i>J</i> = 8.0 Hz)	0.00
20	4.98 (d, <i>J</i> = 8.0 Hz)	1.84	20*	6.70 (d, <i>J</i> = 8.0 Hz)	0.12
21	6.57 (d, <i>J</i> = 7.4 Hz)	1.06	21*	7.68 (br)	-0.05
22	7.25 (d, <i>J</i> = 7.4 Hz)	0.00	22*	7.68 (br)	-0.43
23	6.92 (s)	0.50	23*	7.06 (s)	0.36
24	3.12 (s)	0.53	24*	3.07 (s)	0.56
OMe	0.95 (s)		Ha	4.63 (d, <i>J</i> = 8.0 Hz)	
OHb	7.53 (d, <i>J</i> = 6.0 Hz)		Hb	4.34	
OHc	10.29 (d, <i>J</i> = 6.2 Hz)		Hc	5.13	
OHd	10.87 (d, <i>J</i> = 5.8 Hz)		Hd	4.96	
OHf	8.51 (s)		He	5.07	
Hf	5.67		Hf	5.09	

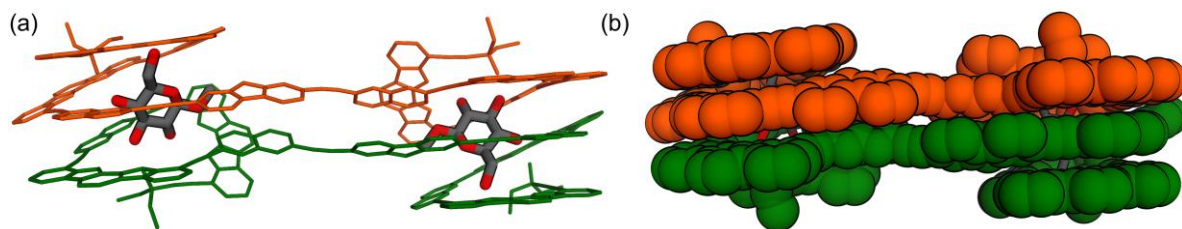


Fig. S13 (a) Tube and (b) space-filling representations of energy-minimized structure of $(2-MP)_2 \supset (\text{me-}\beta\text{-D-glc})_2$ (MacroModel 9.1, MMFFs force field, CHCl_3 phase).

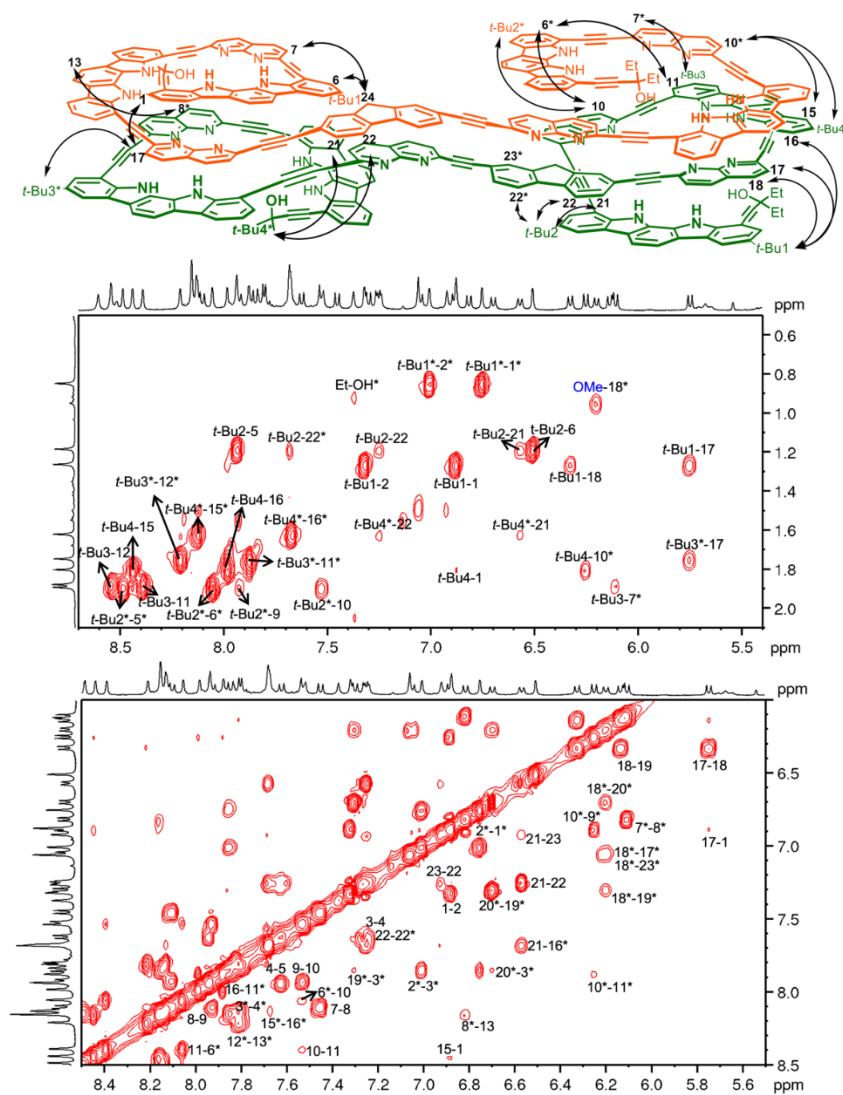


Fig. S14 Partial ^1H - ^1H ROESY spectrum (400 MHz, 25 °C, mixing time: 400 ms) of $(2-MP)_2 \supset (\text{me-}\beta\text{-D-glc})_2$ (4.0 mM) in CD_2Cl_2 .

2.4 ^1H NMR spectra in the U-tube transport experiments

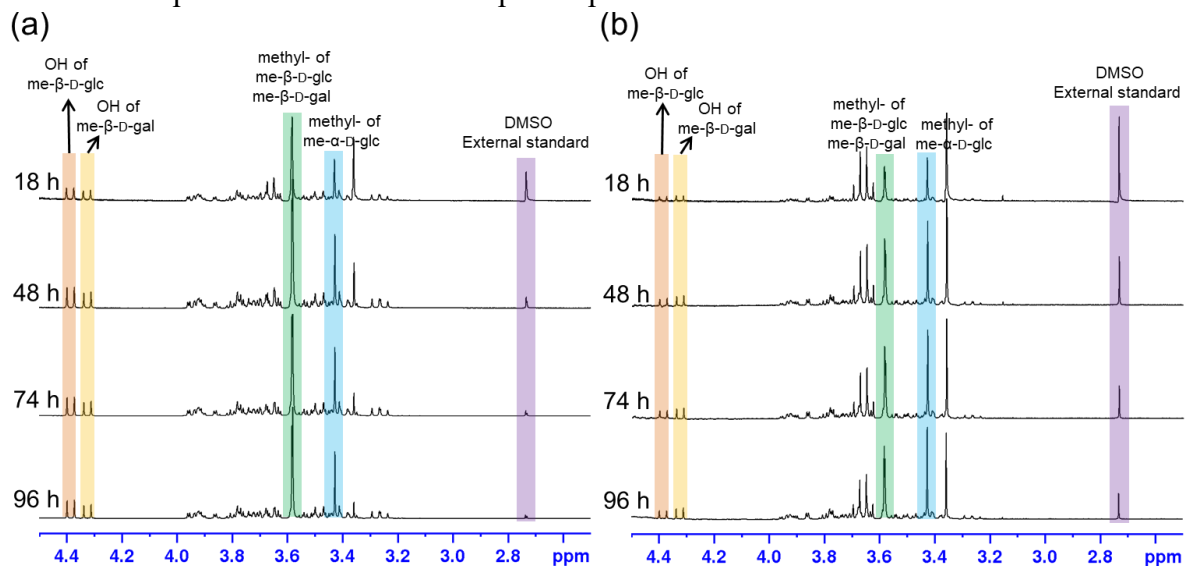


Fig. S15 Time-dependent ^1H NMR spectra (400 MHz, D_2O , 25 °C) of the receiving phase in U-tube transport experiments: (a) in the presence of receptor **2** (3 mM) or (b) in its absence.

3. CD studies

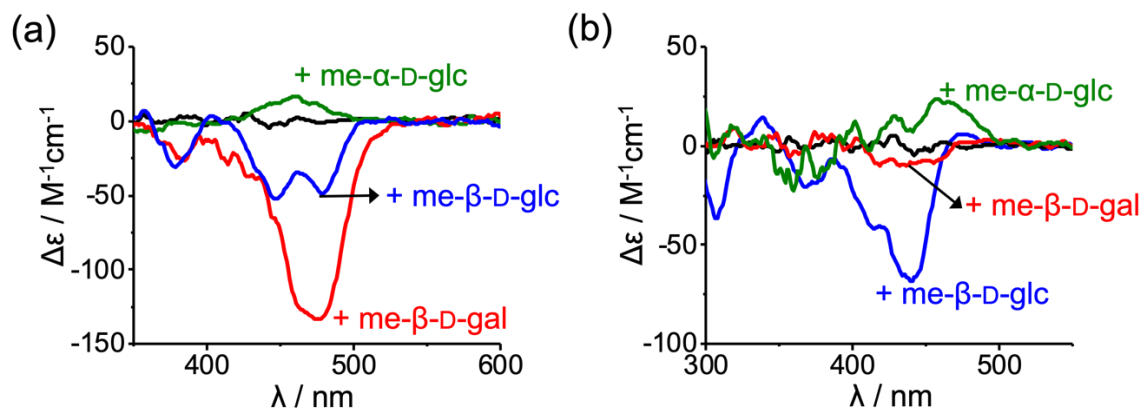


Fig. S16 CD spectra of (a) **1** and (b) **2** (2.00×10^{-5} M) in the absence and presence of guests (200 equiv) in 5% (v/v) $\text{DMSO}/\text{CH}_2\text{Cl}_2$ at 25 °C.

4. Binding studies

4.1 ^1H NMR titrations

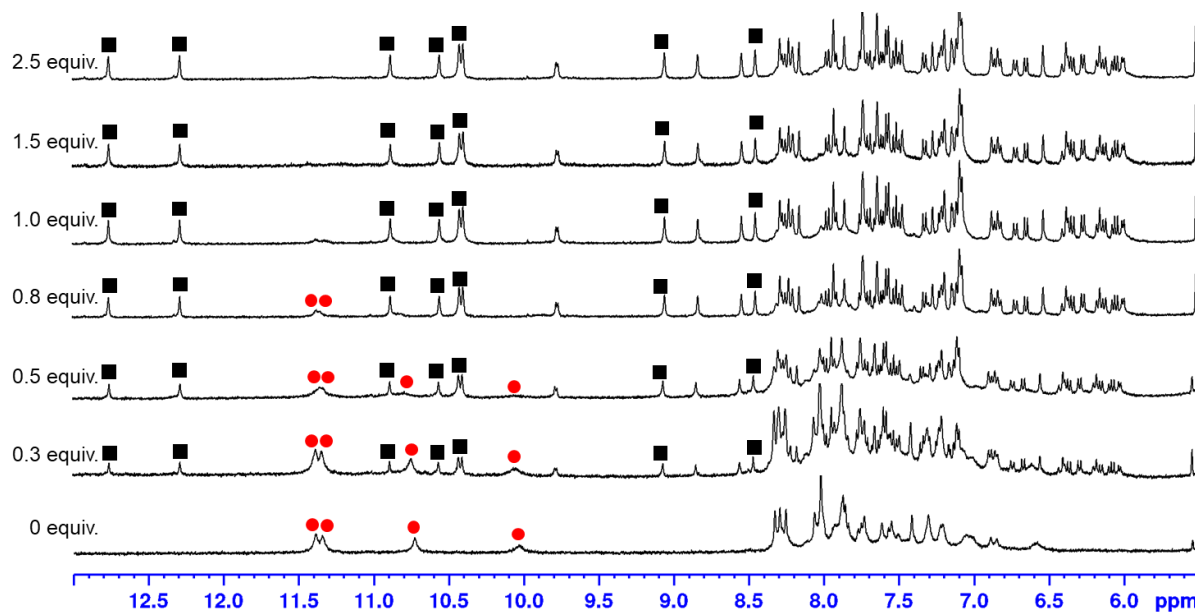


Fig. S17 Partial ^1H NMR spectral changes of **1** (1.50 mM, 25 ± 1 °C) with increasing the amount of me- β -D-gal in 5% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$ (containing ca. 0.05 % water). The NH signals of free **1** and its 2:2 complex are marked with red circles and black squares, respectively.

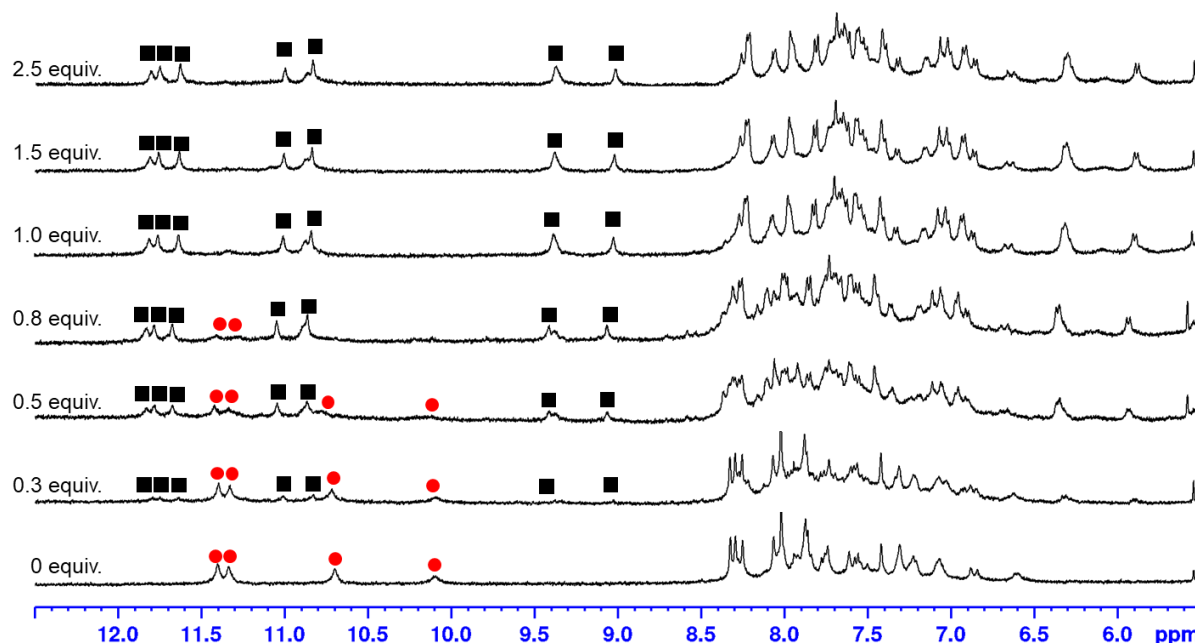


Fig. S18 Partial ^1H NMR spectral changes of **1** (1.50 mM, 25 ± 1 °C) with increasing the amount of me- β -D-gal in 5% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$ (containing ca. 0.05 % water). The NH signals of free **1** and its 2:2 complex are marked with red circles and black squares, respectively.

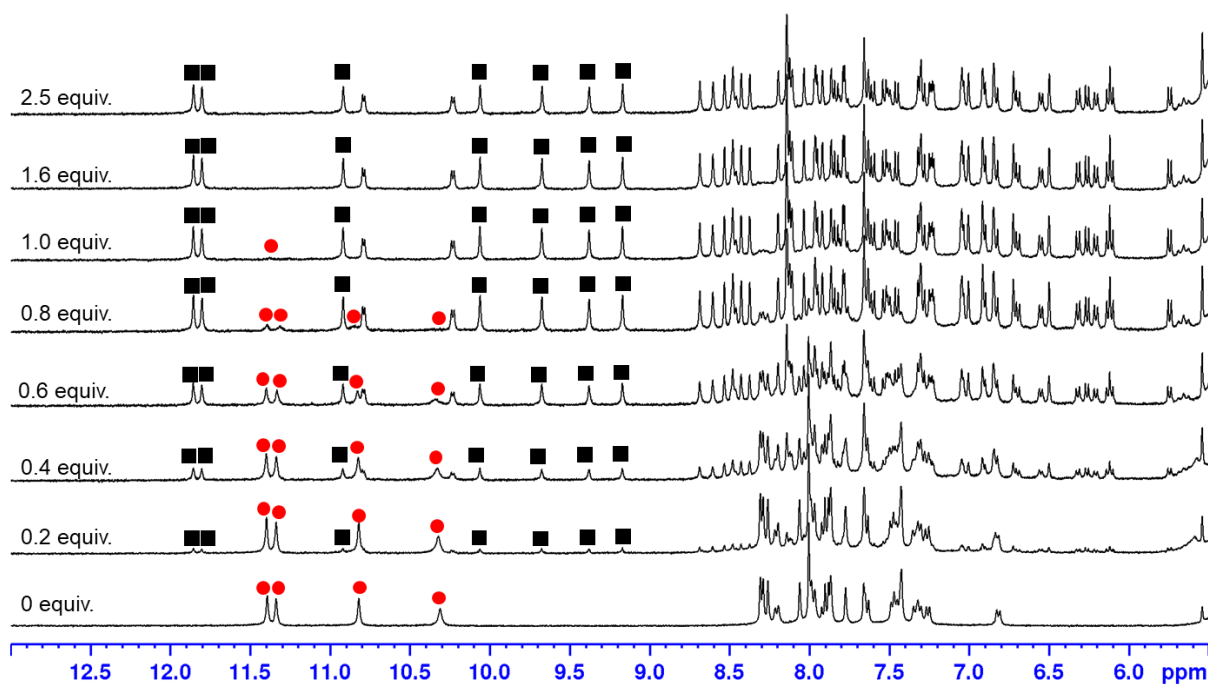


Fig. S19 Partial ^1H NMR spectral changes of **2** (1.50 mM, 25 ± 1 °C) with increasing the amount of me- β -D-glc in 5% (v/v) DMSO- d_6 /CD $_2$ Cl $_2$ (containing ca. 0.05 % water). The NH signals of free **2** and its 2:2 complex are marked with red circles and black squares, respectively.

4.2 Isothermal titration calorimetry (ITC) experiments

The Hypcal software program analyzed the ITC experimental data using a 2:2 binding model, yielding values for ΔH and K . The Gibbs free energy change (ΔG) was subsequently derived from K , allowing for the calculation of the entropy change (ΔS) based on the relationship between ΔH and ΔG (common thermodynamic equation).

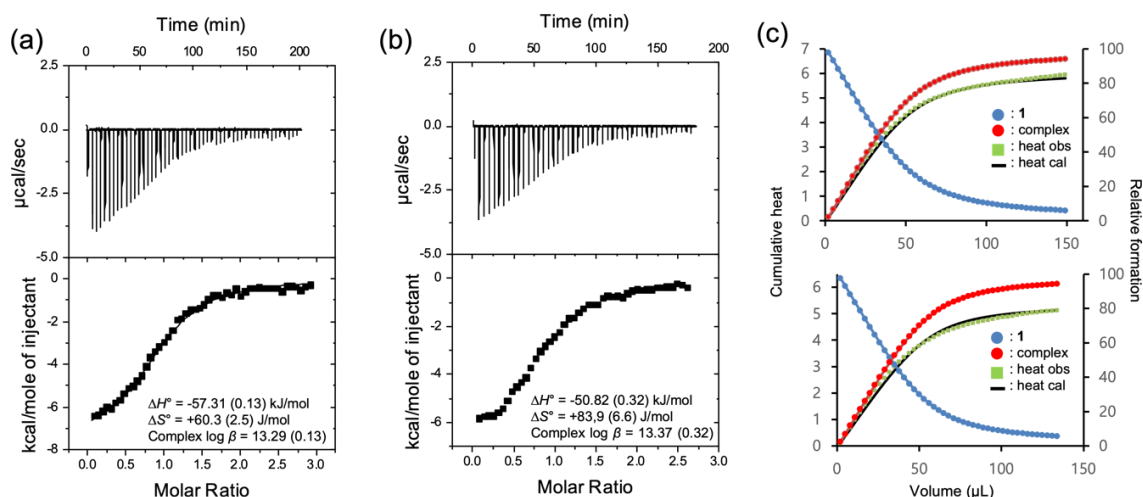


Fig. S20 (a,b) Experimental ITC measurements and binding isotherms of **1** (1.50×10^{-4} M) with me- β -D-glc (4.00×10^{-3} M) in 5 % (v/v) DMSO/ CH_2Cl_2 (containing ca. 0.05 % water) at 22 ± 1 °C. (c) Relative distributions of unbound **1** and its complex, along with experimental and theoretical cumulative heat profiles. In (c), the top represents (a), and the bottom (b).

Table S4 Thermodynamic parameters between **1** and me- β -D-glc in 5 % (v/v) DMSO/ CH_2Cl_2 at 22 °C.

	L	ΔG° (kJ·mol ⁻¹)	ΔH° (kJ·mol ⁻¹)	$T\Delta S^\circ$ (kJ·mol ⁻¹)
(a)	13.3	-75.1	-57.3	+17.8
(b)	13.4	-75.6	-52.8	+22.8

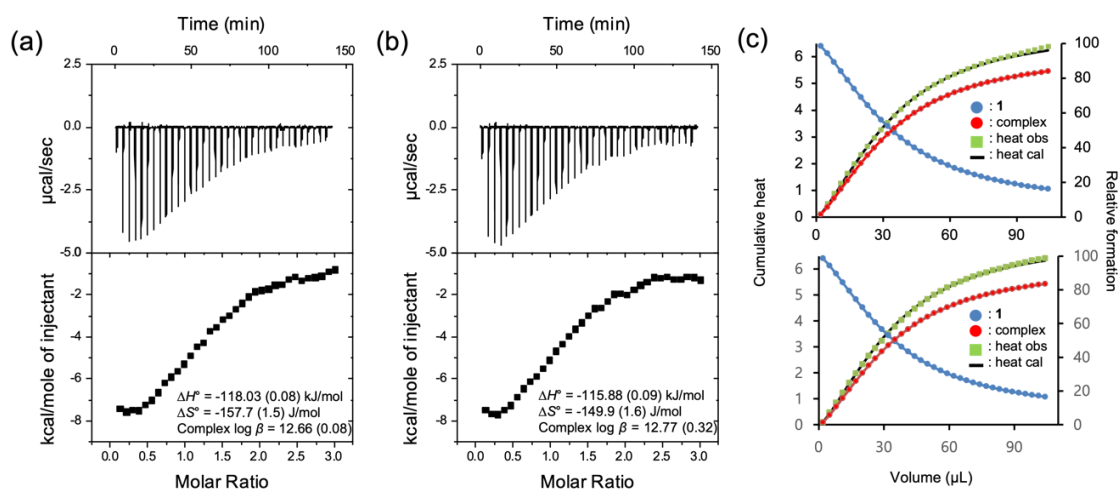


Fig. S21 (a,b) Experimental ITC measurements and binding isotherms of **1** (9.00×10^{-5} M) with me- β -D-gal (4.00×10^{-3} M) in 5 % (v/v) DMSO/ CH_2Cl_2 (containing ca. 0.05 % water) at 22 ± 1 °C. (c) Relative distributions of unbound **1** and its complex, along with experimental and theoretical cumulative heat profiles. In (c), the top represents (a), and the bottom (b).

Table S5 Thermodynamic parameters between **1** and me- β -D-gal in 5 % DMSO/ CH_2Cl_2 at 22 °C.

	L	ΔG° (kJ·mol ⁻¹)	ΔH° (kJ·mol ⁻¹)	$T\Delta S^\circ$ (kJ·mol ⁻¹)
(a)	12.7	-71.5	-118.0	-46.5
(b)	12.8	-71.7	-115.9	-44.2

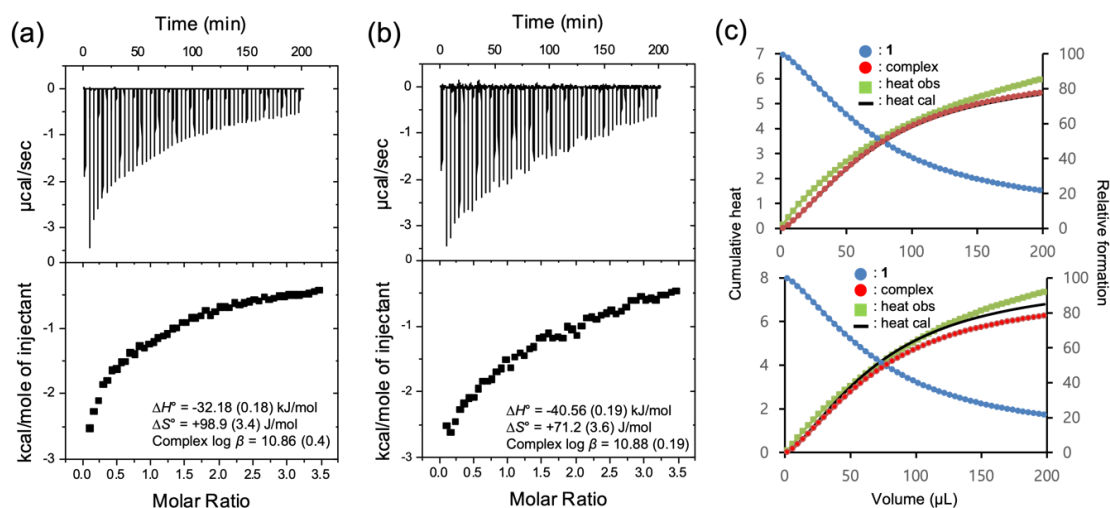


Fig. S22 (a,b) Experimental ITC measurements and binding isotherms of **1** (3.00×10^{-4} M) with me- α -D-glc (7.00×10^{-3} M) in 5 % (v/v) DMSO/CH₂Cl₂ (containing ca. 0.05 % water) at 22 ± 1 °C. (c) Relative distributions of unbound **1** and its complex, along with experimental and theoretical cumulative heat profiles. In (c), the top represents (a), and the bottom (b).

Table S6 Thermodynamic parameters between **1** and me- α -D-glc in 5 % (v/v) DMSO/CH₂Cl₂ at 22 °C.

	L	ΔG° (kJ·mol ⁻¹)	ΔH° (kJ·mol ⁻¹)	$T\Delta S^\circ$ (kJ·mol ⁻¹)
(a)	10.9	-61.2	-32.2	+29.0
(b)	10.9	-61.6	-40.6	+21.0

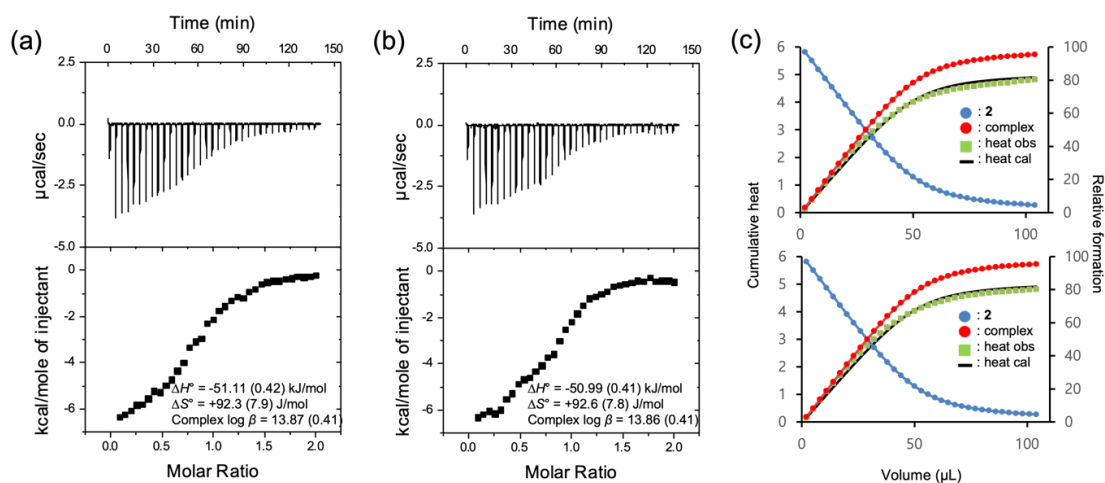


Fig. S23 (a,b) Experimental ITC measurements and binding isotherms of **2** (1.50×10^{-4} M) with me- β -D-glc (4.00×10^{-3} M) in 5 % (v/v) DMSO/CH₂Cl₂ (containing ca. 0.05 % water) at 22 ± 1 °C. (c) Relative distribution of unbound **2** and its complex, along with experimental and theoretical cumulative heat profiles. In (c), the top represents (a), and the bottom (b).

Table S7 Thermodynamic parameters between **2** and me- β -D-glc in 5 % DMSO/CH₂Cl₂ at 22 °C.

	L	ΔG° (kJ·mol ⁻¹)	ΔH° (kJ·mol ⁻¹)	$T\Delta S^\circ$ (kJ·mol ⁻¹)
(a)	13.9	-78.3	-51.1	+27.2
(b)	13.9	-78.3	-51.0	+27.3

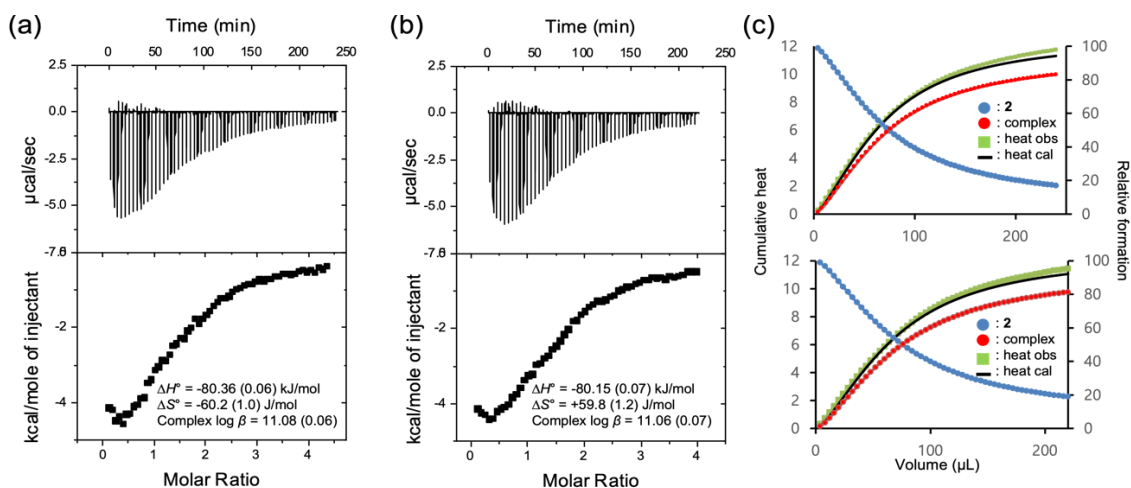


Fig. S24 (a,b) Experimental ITC measurements and binding isotherms of **2** (2.50×10^{-4} M) with me- β -D-gal (6.00×10^{-3} M) in 5 % (v/v) DMSO/CH₂Cl₂ (containing ca. 0.05 % water) at 22 ± 1 °C. (c) Relative distribution of unbound **2** and its complex, along with experimental and theoretical cumulative heat profiles. In (c), the top represents (a), and the bottom (b).

Table S8 Thermodynamic parameters between **2** and me- β -D-gal in 5 % DMSO/CH₂Cl₂ at 22 ± 1 °C.

	L	ΔG° (kJ·mol ⁻¹)	ΔH° (kJ·mol ⁻¹)	$T\Delta S^\circ$ (kJ·mol ⁻¹)
(a)	11.1	-62.6	-80.4	-17.8
(b)	11.1	-62.6	-80.2	-17.6

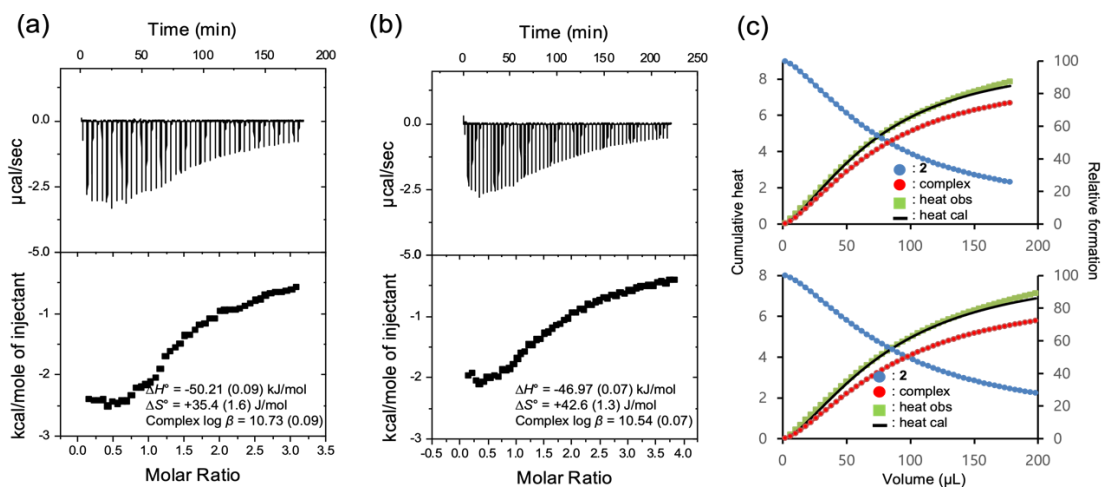


Fig. S25 (a,b) Experimental ITC measurements and binding isotherms of **2** (3.00×10^{-4} M) with me- α -D-glc (7.00×10^{-3} M) in 5 % (v/v) DMSO/CH₂Cl₂ (containing ca. 0.05 % water) at 22 ± 1 °C. (c) Relative distribution of unbound **2** and its complex, along with experimental and theoretical cumulative heat profiles. In (c), the top represents (a), and the bottom (b).

Table S9. Thermodynamic parameters between **2** and me- α -D-glc in 5 % DMSO/CH₂Cl₂ at 22 ± 1 °C.

	L	ΔG° (kJ·mol ⁻¹)	ΔH° (kJ·mol ⁻¹)	$T\Delta S^\circ$ (kJ·mol ⁻¹)
(a)	10.7	-60.6	-50.2	+10.4
(b)	10.5	-59.6	-47.0	+12.6

5. Mass spectra

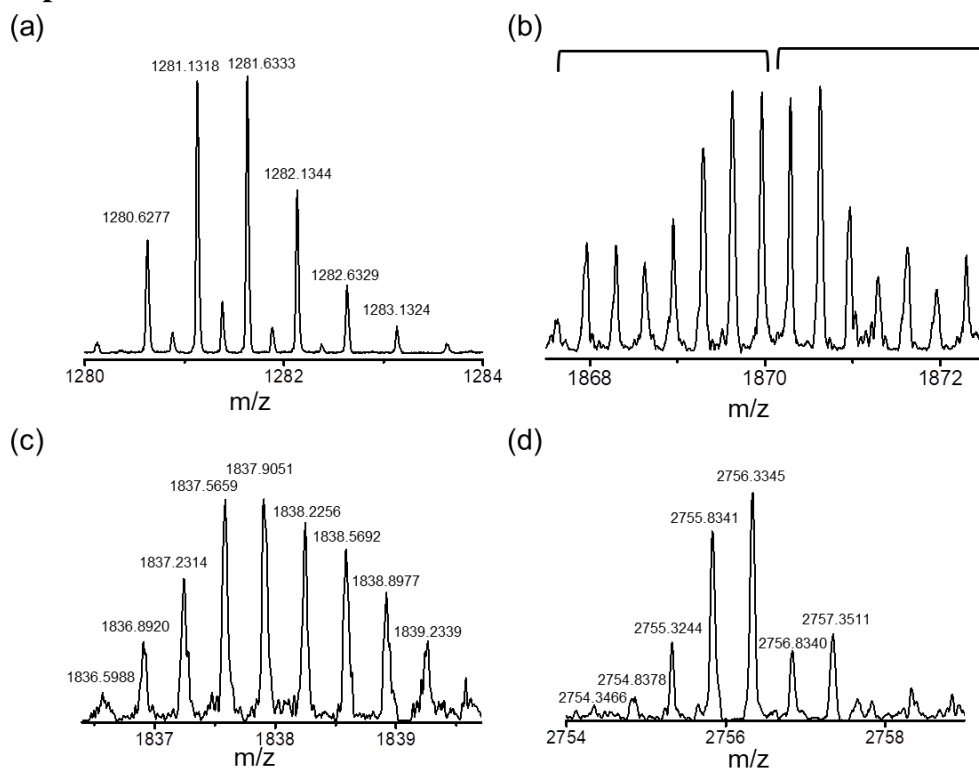


Fig. S26 ESI-MS isotopic distribution of (a) $[1+2H]^{2+}$, (b) $[(1-MM)_2\supset(m\text{-}\beta\text{-D-gal}\cdot 2H_2O)_2+2H+Na]^{3+}$ and $[(1-MM)_2\supset(m\text{-}\beta\text{-D-gal}\cdot 2H_2O)_2+H+2Na-H_2O]^{3+}$, (c) $[(1-MP)_2\supset(m\text{-}\beta\text{-D-glc})_2+3H]^{3+}$, and (d) $[(1-MP)_2\supset(m\text{-}\beta\text{-D-glc})_2+2H]^{2+}$.

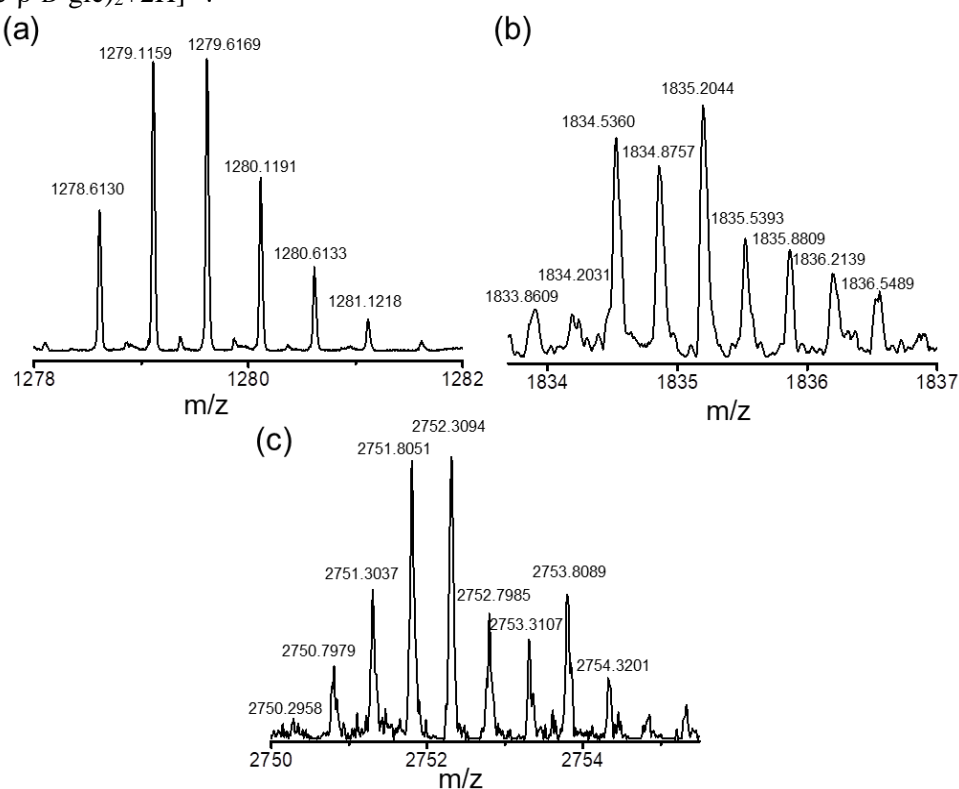


Fig. S27 ESI-MS isotopic distribution of (a) $[2+2H]^{2+}$, (b) $[(2-MP)_2\supset(m\text{-}\beta\text{-D-glc})_2+3H]^{3+}$, and (c) $[(2-MP)_2\supset(m\text{-}\beta\text{-D-glc})_2+2H]^{2+}$.

6. ^1H , ^{13}C NMR spectra of new compounds

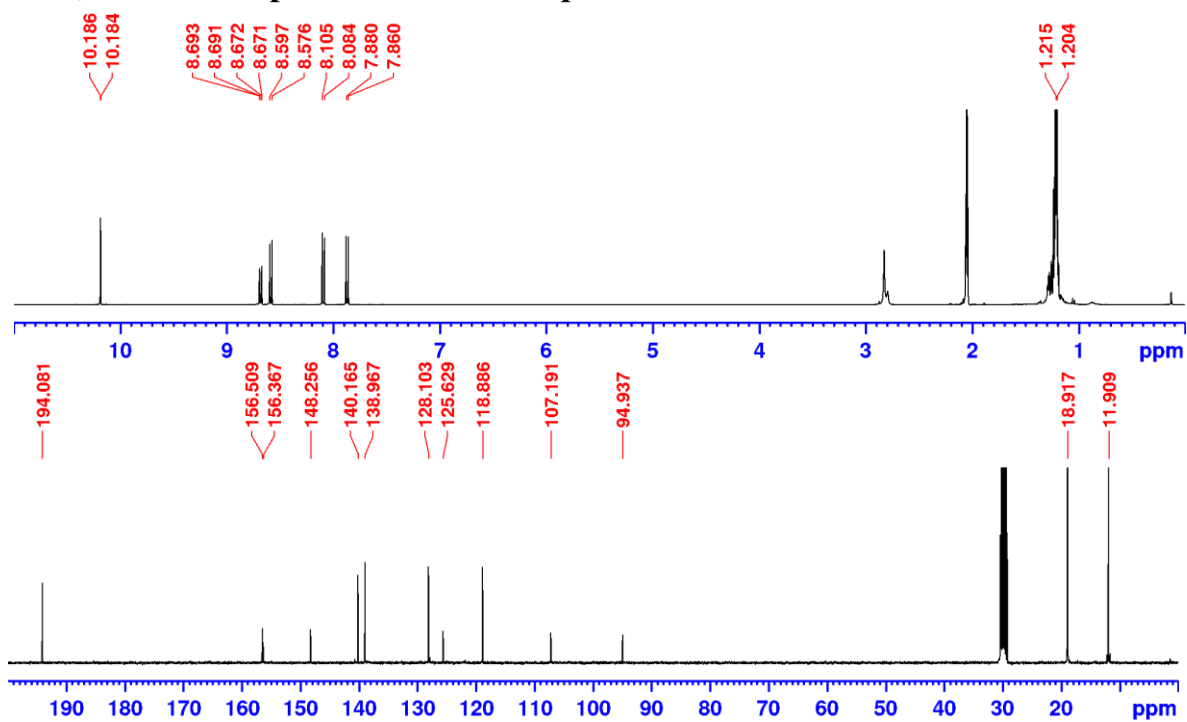


Fig. S28 ^1H and ^{13}C spectra of compound 5 (Acetone- d_6 , 25 $^\circ\text{C}$).

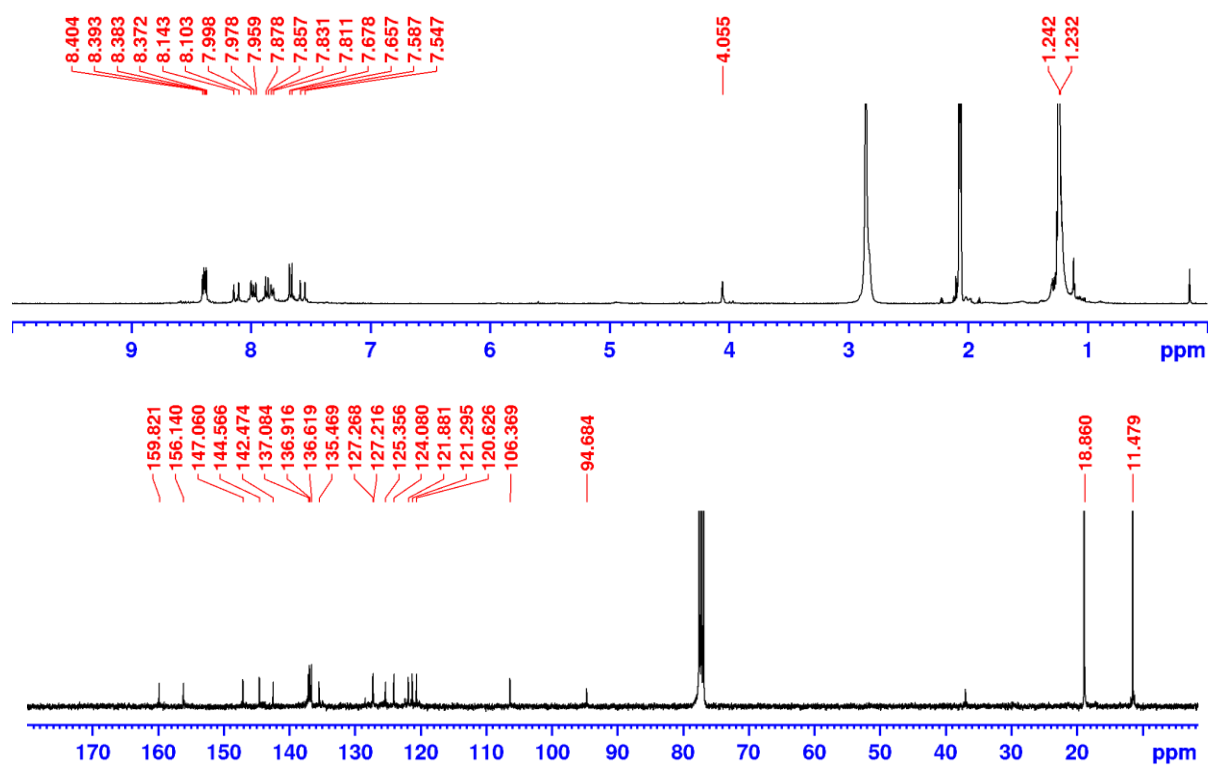


Fig. S29 ^1H (Acetone- d_6) and ^{13}C (CDCl_3) spectra of compound 7 (25 $^\circ\text{C}$).

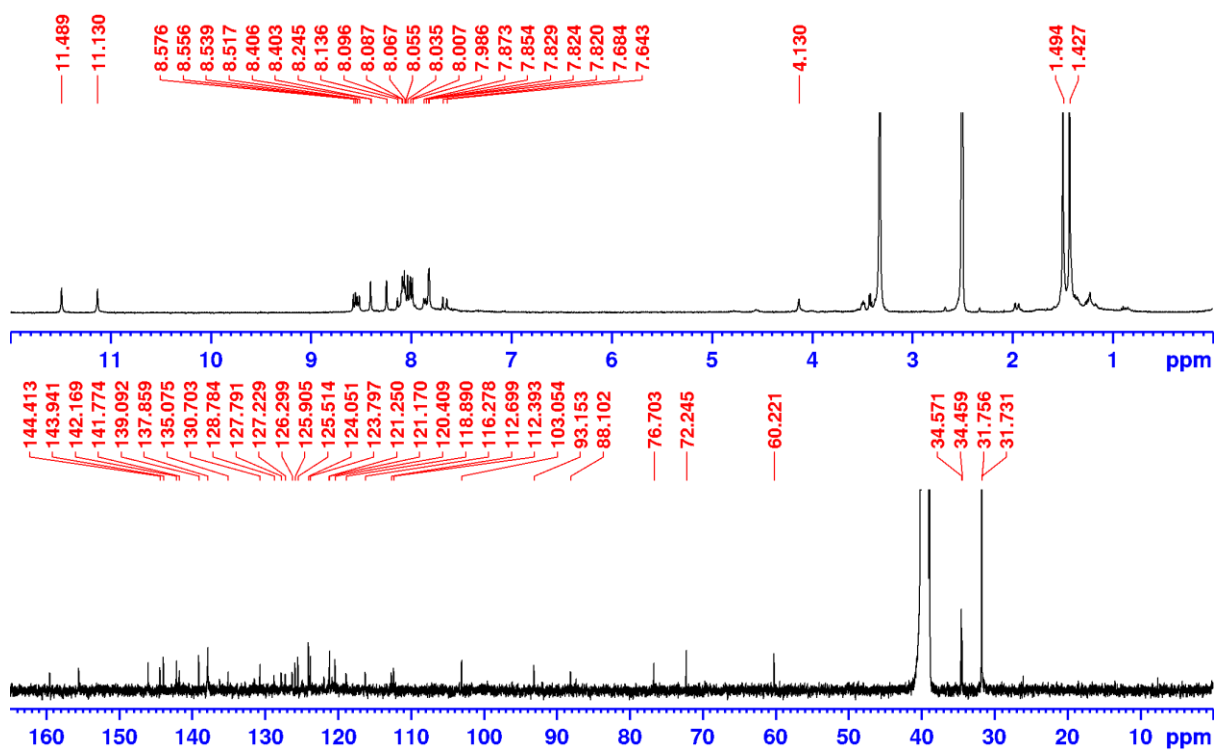


Fig. S30 ¹H and ¹³C spectra of compound 9 (DMSO-*d*₆, 25 °C).

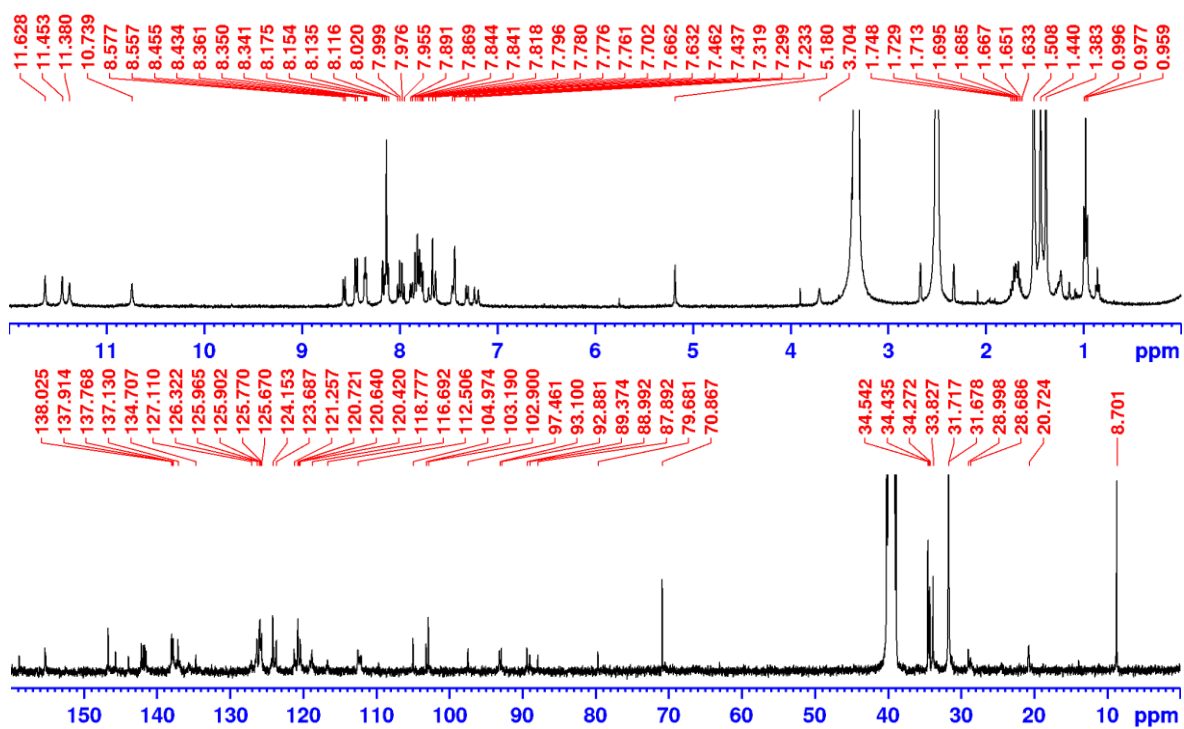


Fig. S31 ¹H and ¹³C spectra of compound 1 (DMSO-*d*₆, 25 °C).

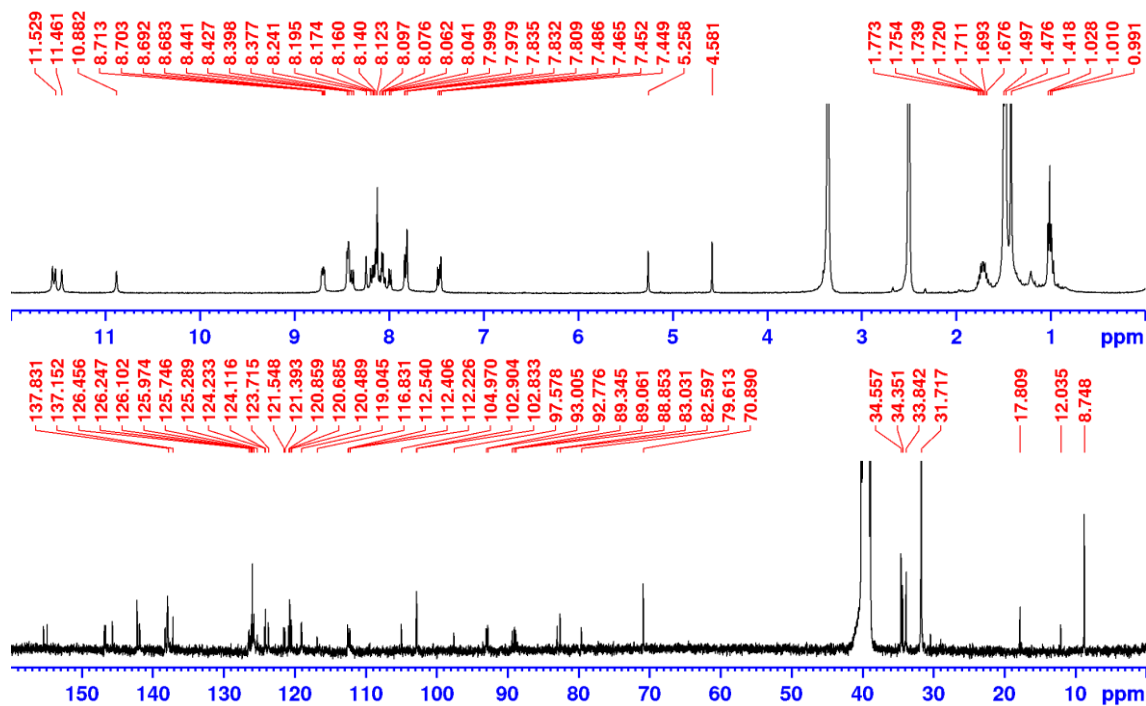


Fig. S32 ¹H and ¹³C spectra of compound 12 (DMSO-*d*₆, 25 °C).

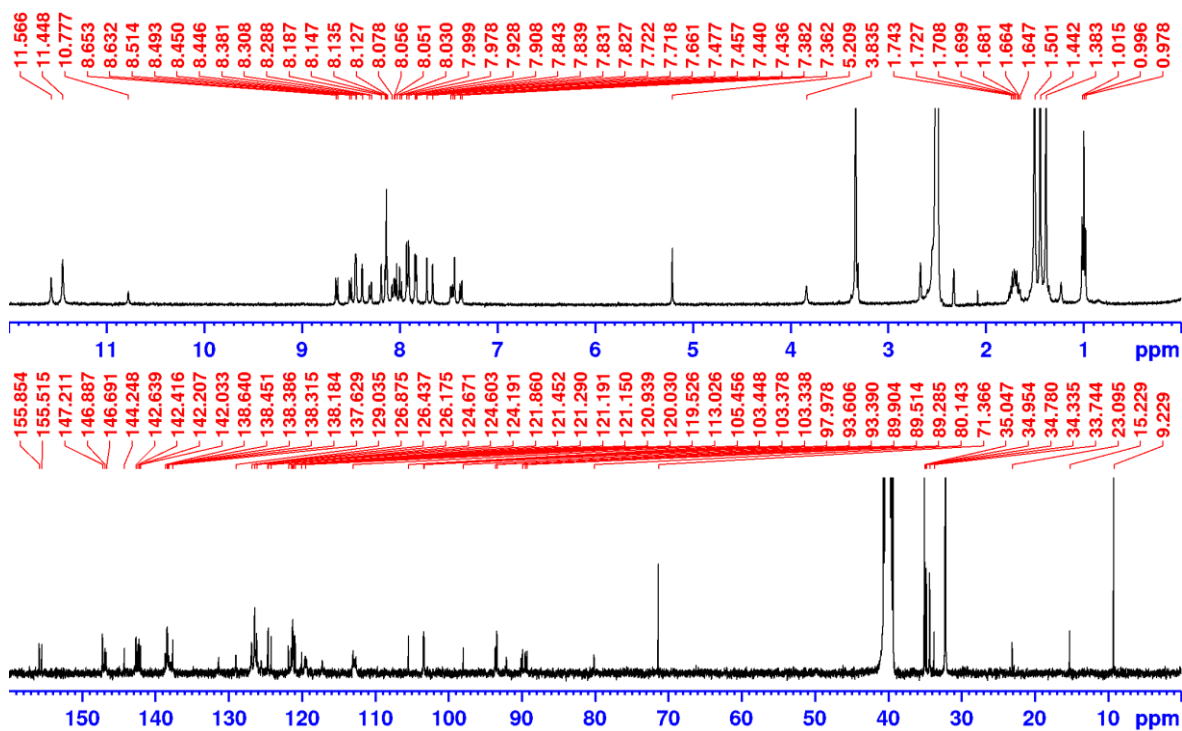


Fig. S33 ¹H and ¹³C spectra of compound 2 (DMSO-*d*₆, 25 °C).

7. References

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