Supplementary Information (SI) for Polymer Chemistry. This journal is © The Royal Society of Chemistry 2024

# Constructing phase diagrams of block copolymers with A-block-(B-stat-C) architecture

# Supporting Information

Britta Weidinger<sup>§</sup>, Nadine von Coelln<sup>§</sup>, Guohui Yang, Hermann Nirschl, Irene Wacker, Rasmus R. Schröder, Petra Tegeder, Eva Blasco\* <sup>§</sup> equal contribution

#### Contents

1.	Synthesis and chemical characterization of the block copolymers	2					
1.	1 P(MMA-stat-HEMA) macroCTAs	2					
Ex	emplary synthesis of P(MMA-stat-HEMA) macroCTAs	2					
N	MR spectra of P(MMA- <i>stat</i> -HEMA) macroCTAs	4					
G	PC chromatograms of P(MMA- <i>stat</i> -HEMA) macroCTAs						
1.	2 PS-b-P(MMA-stat-HEMA) BCPs	16					
Ex	emplary synthesis of PS- <i>b</i> -P(MMA- <i>stat</i> -HEMA) BCPs	16					
N	MR spectra of PS- <i>b</i> -P(MMA- <i>stat</i> -HEMA) BCPs						
G	PC chromatograms of PS- <i>b</i> -P(MMA- <i>stat</i> -HEMA) BCPs						
1.	3 Functionalized BCPs	40					
Ex	emplary synthesis of functionalized BCPs	40					
N	MR spectra of functionalized BCPs	41					
2.	Morphology characterization	50					
SE	M images and SAXS spectra	50					
A	FM and IR-SNOM phase images	63					
Cι	uts along different planes of cylindrical and gyroidal morphologies	64					
D	Domain spacings						

## 1. Synthesis and chemical characterization of the block copolymers

### 1.1 P(MMA-stat-HEMA) macroCTAs

#### Exemplary synthesis of P(MMA-stat-HEMA) macroCTAs

In a Schlenk tube, MMA (6.40 g, 64.0 mmol, 819 eq.), HEMA (2.08 g, 16.0 mmol, 204 eq.) and CPDB (17.3 mg, 78.1 µmol, 1.0 eq.) were dissolved in 1,4-dioxane (8 mL). AIBN solution (323 mg, 9.78 µmol, 0.125 eq., 10 mg in 2.010 mg 1,4-dioxane) was added. The solution was degassed via freeze-pump-thaw (3 x 8 min), followed by backfilling with nitrogen. The reaction mixture was stirred at 90 °C for 2.5 h, cooled in liquid nitrogen and opened to the atmosphere. After dilution with DCM, the solution was precipitated into diethyl ether twice. After centrifugation and decanting of the supernatant, copolymer macroCTA-38 was received as a light pink solid.

<sup>1</sup>H NMR 600 MHz (CD<sub>2</sub>Cl<sub>2</sub>):  $\delta$  [ppm] = 7.80 (m), 7.47 (m), 7.31 (m), 4.0 (bs), 3.74 (bs), 3.50 (s), 2.00 - 0.61 (m).

		т [b]	M (GPC)	Ð	Precursor for
Polymer		. [.,]	win(GrC)	D	
macroCTA-	304:76:1:0.125	4	22454	1.11	S <sub>0.54</sub> -MH <sub>0.46</sub> -45
22					
macroCTA-	441:110:1:0.125	4	28749	1.12	S <sub>0.52</sub> -MH <sub>0.48</sub> -67
29					
macroCTA-	304:76:1:0.125	2	20667	1.13	S <sub>0.75</sub> -MH <sub>0.25</sub> -66
21					
macroCTA-	819:204:1:0.125	5	53925	1.05	S <sub>0.49</sub> -MH <sub>0.51</sub> -92
54					
macroCTA-	304:76:1:0.125	2.5	23083	1.08	S <sub>0.45</sub> -MH <sub>0.55</sub> -42
23					S <sub>0.66</sub> -MH <sub>0.34</sub> -70
macroCTA-	819:204:1:0.125	2.5	48065	1.12	S <sub>0.50</sub> -MH <sub>0.50</sub> -91
48					
macroCTA-	819:204:1:0.125	2.5	38850	1.17	S <sub>0.46</sub> -MH <sub>0.54</sub> -81
39					S <sub>0.42</sub> -MH <sub>0.58</sub> -89
macroCTA-	819:204:1:0.125	3.2	43205	1.15	S <sub>0.28</sub> -MH <sub>0.72</sub> -67
43					S <sub>0.27</sub> -MH <sub>0.73</sub> -70
macroCTA-	819:204:1:0.125	2.5	37641	1.10	S <sub>0.37</sub> -MH <sub>0.63</sub> -65
38					S <sub>0.53</sub> -MH <sub>0.47</sub> -94
macroCTA-	819:204:1:0.125	2.6	33587	1.13	S <sub>0.42</sub> -MH <sub>0.58</sub> -57
34					S <sub>0.09</sub> -MH <sub>0.91</sub> -39
					S <sub>0.31</sub> -MH <sub>0.69</sub> -48
					S <sub>0.16</sub> -MH <sub>0.84</sub> -45
					S <sub>0.12</sub> -MH <sub>0.88</sub> -40
					S <sub>0.16</sub> -MH <sub>0.84</sub> -43
macroCTA-	304:76:1:0.125	2	18985	1.10	S <sub>0.39</sub> -MH <sub>0.61</sub> -35
19					S <sub>0.28</sub> -MH <sub>0.72</sub> -29
					S <sub>0.50</sub> -MH <sub>0.50</sub> -45
					S <sub>0.56</sub> -MH <sub>0.44</sub> -52
macroCTA-9	224:56:1:0.125	1	9303	1.12	S <sub>0.68</sub> -MH <sub>0.32</sub> -33

Table S1: Monomer ratio, reaction time, molecular weight, and dispersity of the synthesized MacroCTAs-XX, where xx corresponds to the molecular weight in kDa.

NMR spectra of P(MMA-stat-HEMA) macroCTAs



Figure S2: <sup>1</sup>H NMR spectrum of macroCTA-29 in CDCl<sub>3</sub>.



re S4: <sup>1</sup>H NMR spectrum of macroCTA-54 in CDCl<sub>3</sub>.



re S6: <sup>1</sup>H NMR spectrum of macroCTA-48 in CDCl<sub>3</sub>.







re S10: <sup>1</sup>H NMR spectrum of macroCTA-34 in CDCl<sub>3</sub>.



e S12: <sup>1</sup>H NMR spectrum of macroCTA-9 in CDCl<sub>3</sub>.



Figure S13: GPC chromatogram (RI detector) of macroCTA-22.



Figure S14: GPC chromatogram (RI detector) of macroCTA-29.



Figure S15: GPC chromatogram (RI detector) of macroCTA-21.



Figure S16: GPC chromatogram (RI detector) of macroCTA-54.



Figure S17: GPC chromatogram (RI detector) of macroCTA-23.



Figure S18: GPC chromatogram (RI detector) of macroCTA-48.



Figure S19: GPC chromatogram (RI detector) of macroCTA-39.



Figure S20: GPC chromatogram (RI detector) of macroCTA-43.



Figure S21: GPC chromatogram (RI detector) of macroCTA-38.



Figure S22: GPC chromatogram (RI detector) of macroCTA-34.



Figure S23: GPC chromatogram (RI detector) of macroCTA-19.



Figure S24: GPC chromatogram (RI detector) of macroCTA-9.

#### 1.2 PS-b-P(MMA-stat-HEMA) BCPs

#### Exemplary synthesis of PS-b-P(MMA-stat-HEMA) BCPs

In a Schlenk tube, 560 mg (MW = 37641, 0.0149 mmol, 1 eq.) of macroCTA-*38* were dissolved in 18 mL 1,4-dioxane. Styrene (18.3 g, 176 mmol, 11000 eq.) and AIBN stock solution (79 mg, 0.0032 mmol, 7.5 mg in 1.125 g 1,4-dioxane, 0.2 eq.) were added. The solution was degassed via freeze-pump-thaw (3 x 8 min), followed by backfilling with nitrogen. The reaction mixture was stirred at 90 °C for 2 h, cooled in liquid nitrogen and opened to the atmosphere. The solvent and most of the remaining styrene were removed in vacuo, the viscous mixture was dissolved in small amounts of DCM and precipitated into diethyl ether twice. After centrifugation and decanting of the supernatant,  $S_{0.53}$ -MH<sub>0.47</sub>-94 was received as a light pink solid.

<sup>1</sup>H NMR 600 MHz (CD<sub>2</sub>Cl<sub>2</sub>): δ [ppm] = 7.80 (m), 7.47 (m), 7.31 (m), 7.28 - 6.31 (m), 4.07 (bs), 3.80 (bs), 3.56 (s), 2.14 - 0.63 (m).

Polymer	MacroCTA	Styrene:Macro	Т	M <sub>n</sub> (GPC)	Ð	Funct. polymer
		CTA:AIBN	[min]			
S <sub>0.75</sub> -MH <sub>0.25</sub> -66	macroCTA-	5000:1:0.2	120	65769	1.20	S <sub>0.75</sub> -MH* <sub>0.25</sub> -66
	21					
S <sub>0.68</sub> -MH <sub>0.32</sub> -33	macroCTA-9	5000:1:0.2	80	32524	1.06	S <sub>0.68</sub> -MH* <sub>0.32</sub> -33
S <sub>0.66</sub> -MH <sub>0.34</sub> -70	macroCTA-	8000:1:0.2	120	69535	1.10	S <sub>0.66</sub> -MH* <sub>0.34</sub> -70
	23					
S <sub>0.56</sub> -MH <sub>0.44</sub> -52	macroCTA-	10000:1:0.2	60	52082	1.07	S <sub>0.56</sub> -MH* <sub>0.44</sub> -52
	19					
S <sub>0.54</sub> -MH <sub>0.46</sub> -45	macroCTA-	3720:1:0.2	60	45272	1.18	-
	22					
S <sub>0.53</sub> -MH <sub>0.47</sub> -94	macroCTA-	11000:1:0.2	120	93566	1.10	S <sub>0.53</sub> -MH* <sub>0.47</sub> -94
	38					
S <sub>0.52</sub> -MH <sub>0.48</sub> -67	macroCTA-	7000:1:0.2	120	66991	1.10	S <sub>0.52</sub> -MH* <sub>0.48</sub> -67
	22					
So 50-MH0 50-91	macroCTA-	6150:1:0.2	90	91421	1.14	So 50-MH*0 50-91
- 0.50 0.50 -	18			-		- 0.50 - 0.50 -
	40					
So 50-MH0 50-45	macroCTA-	6150:1:0.2	60	45136		So 50-MH*0 50-45
30.50 1011 0.50 7.3	10	0100.1.0.2	00	13130		0.50 1011 0.50 70
	17					
So 40-MH92	macroCTA-	6150.1.0.2	100	56600	1 29	
50.49 10110.51 52		0150.1.0.2	100	50000	1.55	
	54					
S	macroCTA	6150.1.0.2	90	81005	1 1 /	S
J0.46 IVI 0.54 OL		0130.1.0.2	50	01003	1.14	J0.46 IVI 0.54-01
	39					
<u> </u>	magraCTA	2720.1.0.2	40	42004	1 1 1	C NALI50%* 47
J <sub>0.45</sub> -IVI⊓ <sub>0.55</sub> -4∠	macrociA-	3720.1.0.2	40	42004	1.11	J <sub>0.45</sub> -IVI∏ <sup>2000</sup> 0.55-4∠

	23					
S <sub>0.42</sub> -MH <sub>0.58</sub> -89	macroCTA-	6150:1:0.2	90	89172	1.16	S <sub>0.42</sub> -MH* <sub>0.58</sub> -89
	39					
S <sub>0.42</sub> -MH <sub>0.58</sub> -57	macroCTA-	10000:1:0.2	120	56861	1.11	S <sub>0.42</sub> -MH* <sub>0.58</sub> -57
	34					
S <sub>0.39</sub> -MH <sub>0.61</sub> -35	macroCTA-	3720:1:0.2	40	35156	1.09	-
	19					
	10					
S0 37-MH0 63-65	macroCTA-	6150:1:0.2	90	65036	1.09	S <sub>0 37</sub> -MH* <sub>0 63</sub> -65
0.57 0.05	28					0.05
	50					
So 21-MHo co-48	macroCTA-	5000:1:0.2	90	47659	1.09	So 21-MH*2 co-48
0.31 0.69	24					0.31 0.89 .0
	54					
SMH67	macro(TA-	3720.1.0 2	90	67188	1 1 3	SMH*67
50.28 101 10.72 07	42	5720.1.0.2	50	07100	1.15	50.28 1011 0.72 07
	43					
S	macro(TA-	2000.1.0.2	40	29291	1 09	S79
50.28 10110.72 25	10	2000.1.0.2	40	25251	1.05	50.28 1011 0.72 2.5
	19					
S _MH _70	macroCTA	2720.1.0.2	60	70101	1 1 2	S _N/H* _70
3 <sub>0.27</sub> -10111 <sub>0.73</sub> -70		3720.1.0.2	00	70191	1.15	S <sub>0.27</sub> -1011 0.73-70
	43					
<u> </u>	magraCTA	12200.1.0.2	120	45070	1 22	
30.16-IVIN0.84-43	macrocra-	13200.1.0.2	120	45079	1.25	-
	34					
<u> </u>	macroCTA	2500.1.0.2	20	42624	1 1 2	C MU* 40
S <sub>0.16</sub> -IVI⊓ <sub>0.84</sub> -43	macrocia-	3500:1:0.2	30	42021	1.15	S <sub>0.16</sub> -IVI - 0.84-43
	34					
		2400 4 0 0	20	20050	4 4 2	
S <sub>0.12</sub> -IVIH <sub>0.88</sub> -40	macroCIA-	2400:1:0.2	28	39858	1.13	-
	34					
S <sub>0.09</sub> -MH <sub>0.91</sub> -39	macroCTA-	2500:1:0.2	23	38743	1.13	-
	34					



Figure S25: <sup>1</sup>H NMR spectrum of S<sub>0.53</sub>-MH<sub>0.47</sub>-94 in CDCl<sub>3</sub> with peak assignment.



Figure S26: <sup>1</sup>H NMR spectrum of S<sub>0.75</sub>-MH<sub>0.25</sub>-66 in CDCl<sub>3</sub>.



Figure S28: <sup>1</sup>H NMR spectrum of S<sub>0.66</sub>-MH<sub>0.34</sub>-70 in CDCl<sub>3</sub>.



Figure S30: <sup>1</sup>H NMR spectrum of  $S_{0.54}$ -MH<sub>0.46</sub>-45 in CDCl<sub>3</sub>.



Figure S31: <sup>1</sup>H NMR spectrum of  $S_{0.52}$ -MH<sub>0.48</sub>-67 in CDCl<sub>3</sub>.



Figure S33: <sup>1</sup>H NMR spectrum of  $S_{0.50}$ -MH<sub>0.50</sub>-45 in CDCl<sub>3</sub>.



Figure S35: <sup>1</sup>H NMR spectrum of  $S_{0.46}$ -MH<sub>0.54</sub>-81 in CDCl<sub>3</sub>.



Figure S38: <sup>1</sup>H NMR spectrum of  $S_{0.42}$ -MH<sub>0.58</sub>-57 in CDCl<sub>3</sub>.



Figure S39: <sup>1</sup>H NMR spectrum of  $S_{0.39}$ -MH<sub>0.61</sub>-35 in CDCl<sub>3</sub>.



Figure S40: <sup>1</sup>H NMR spectrum of S<sub>0.37</sub>-MH<sub>0.63</sub>-65 in CDCl<sub>3</sub>.



Figure S41: <sup>1</sup>H NMR spectrum of  $S_{0.31}$ -MH<sub>0.69</sub>-48 in CDCl<sub>3</sub>.



Figure S43: <sup>1</sup>H NMR spectrum of S<sub>0.28</sub>-MH<sub>0.72</sub>-29 in CDCl<sub>3</sub>.



e S45: <sup>1</sup>H NMR spectrum of S<sub>0.16</sub>-MH<sub>0.84</sub>-45 in CDCl3.



Figure S47: 1H NMR spectrum of  $S_{0.12}$ -MH<sub>0.88</sub>-40 in CDCl3.



Figure S48: <sup>1</sup>H NMR spectrum of  $S_{0.09}$ -MH<sub>0.91</sub>-39 in CDCl<sub>3</sub>.

GPC chromatograms of PS-b-P(MMA-stat-HEMA) BCPs



Figure S49: GPC chromatogram (RI detector) of  $S_{0.75}$ -MH<sub>0.25</sub>-66.



Figure S50: GPC chromatogram (RI detector) of  $S_{0.68}\text{-}\mathsf{MH}_{0.32}\text{-}33.$ 



Figure S51: GPC chromatogram (RI detector) of  $S_{0.66}$ -MH<sub>0.34</sub>-70.



Figure S52: GPC chromatogram (RI detector) of  $S_{0.56}$ -MH<sub>0.44</sub>-52.



Figure S53: GPC chromatogram (RI detector) of  $S_{0.54}$ -MH<sub>0.46</sub>-45.



Figure S54: GPC chromatogram (RI detector) of  $S_{0.53}$ -MH<sub>0.47</sub>-94.



Figure S55: GPC chromatogram (RI detector) of  $S_{0.52}$ -MH<sub>0.48</sub>-67.



Figure S56: GPC chromatogram (RI detector) of  $S_{0.50}$ -MH<sub>0.50</sub>-91.



Figure S57: GPC chromatogram (RI detector) of  $S_{0.50}$ -MH<sub>0.50</sub>-45.



Figure S58: GPC chromatogram (RI detector) of  $S_{0.49}$ -MH<sub>0.51</sub>-92.



Figure S59: GPC chromatogram (RI detector) of  $S_{0.46}$ -MH<sub>0.54</sub>-81.



Figure S60: GPC chromatogram (RI detector) of  $S_{0.45}$ -MH<sub>0.55</sub>-42.



Figure S61: GPC chromatogram (RI detector) of  $S_{0.42}$ -MH<sub>0.58</sub>-89.



Figure S62: GPC chromatogram (RI detector) of  $S_{0.42}$ -MH<sub>0.58</sub>-57.



Figure S63: GPC chromatogram (RI detector) of S<sub>0.39</sub>-MH<sub>0.61</sub>-35.



Figure S64: GPC chromatogram (RI detector) of  $S_{0.37}$ -MH<sub>0.63</sub>-65.



Figure S65: GPC chromatogram (RI detector) of S<sub>0.31</sub>-MH<sub>0.69</sub>-48.



Figure S66: GPC chromatogram (RI detector) of  $S_{0.28}$ -MH<sub>0.72</sub>-67.



Figure S67: GPC chromatogram (RI detector) of S<sub>0.28</sub>-MH<sub>0.72</sub>-29.



Figure S68: GPC chromatogram (RI detector) of  $S_{0.27}$ -MH<sub>0.73</sub>-70.



Figure S69: GPC chromatogram (RI detector) of  $S_{0.16}$ -MH<sub>0.84</sub>-45.



Figure S70: GPC chromatogram (RI detector) of  $S_{0.16}$ -MH<sub>0.84</sub>-43.



Figure S71: GPC chromatogram (RI detector) of S<sub>0.12</sub>-MH<sub>0.88</sub>-40.



Figure S72: GPC chromatogram (RI detector) of S<sub>0.09</sub>-MH<sub>0.91</sub>-39.

#### 1.3 Functionalized BCPs

#### Exemplary synthesis of functionalized BCPs

S<sub>0.53</sub>-MH<sub>0.47</sub>-94 (250 mg, MW = 93566, 2.67 µmol, 207 µmol OH-groups) was added to a Schlenk flask. After flushing with nitrogen, dry DCM (19 mL) was added. After dissolving, NEt<sub>3</sub> (592 µL, 430 mg, 4.24 mmol, 20 eq.) was added, the solution was cooled to 0 °C and methacryloyl chloride (348 µL, 389 mg, 3.74 mmol, 18 eq.) was added slowly under stirring. After stirring at 0°C to RT for 16-72 h, most of the DCM was removed in vacuo. MeCN (65 mL) was added and the mixture was poured into aqueous NaHCO<sub>3</sub> (5%, 130 mL). After extracting with DCM (3 x 65 mL) the united organic fractions were dried over Na<sub>2</sub>SO<sub>4</sub>, the solvent was removed in vacuo and the washing step was repeated if NEt<sub>3</sub>HCl was still present. After precipitation in Et<sub>2</sub>O, S<sub>0.53</sub>-MH\*<sub>0.47</sub>-94 was obtained as a solid.

<sup>1</sup>H NMR 600 MHz ( $CD_2Cl_2$ ):  $\delta$  [ppm] = 7.80 (m), 7.47 (m), 7.31 (m), 7.28 - 6.31 (m), 6.12 (bs), 5.63 (bs), 4.32 (bs), 4.17 (bs), 3.56 (s), 2.14 - 0.63 (m).



Figure S73: <sup>1</sup>H NMR spectrum of S<sub>0.53</sub>-MH\*<sub>0.47</sub>-94 in CD<sub>2</sub>Cl<sub>2</sub> with peak assignment.



Figure S74: <sup>1</sup>H NMR spectrum of  $S_{0.75}$ -MH\*<sub>0.25</sub>-66 in CDCl<sub>3</sub>.



Figure S76: <sup>1</sup>H NMR spectrum of  $S_{0.66}$ -MH\*<sub>0.34</sub>-70 in CDCl<sub>3</sub>.



Figure S78: <sup>1</sup>H NMR spectrum of  $S_{0.52}$ -MH\*<sub>0.48</sub>-67 in CDCl<sub>3</sub>.



Figure S80: <sup>1</sup>H NMR spectrum of  $S_{0.50}$ -MH\*<sub>0.50</sub>-45 in CDCl<sub>3</sub>.



Figure S81: <sup>1</sup>H NMR spectrum of  $S_{0.46}$ -MH\*<sub>0.54</sub>-81 in CDCl<sub>3</sub>.



Figure S82: <sup>1</sup>H NMR spectrum of  $S_{0.45}$ -MH\*<sub>0.55</sub>-42 in CDCl<sub>3</sub>.



Figure S84: <sup>1</sup>H NMR spectrum of  $S_{0.42}$ -MH\*<sub>0.58</sub>-57 in CDCl<sub>3</sub>.



Figure S86: <sup>1</sup>H NMR spectrum of  $S_{0.31}$ -MH\*<sub>0.69</sub>-48 in CDCl<sub>3</sub>.



Figure S87: <sup>1</sup>H NMR spectrum of S<sub>0.28</sub>-MH\*<sub>0.72</sub>-67 in CD<sub>2</sub>Cl<sub>2</sub>.



Figure S88: <sup>1</sup>H NMR spectrum of S<sub>0.28</sub>-MH\*<sub>0.72</sub>-29 in CDCl<sub>3</sub>.



Figure S90: <sup>1</sup>H NMR spectrum of S<sub>0.16</sub>-MH<sub>0.84</sub>-43 in CDCl<sub>3</sub>.

## 2. Morphology characterization

SEM images and SAXS spectra





Figure S91: SEM images and SAXS spectra for  $S_{0.75}$ - $MH_{0.25}$ -66 and  $S_{0.75}$ - $MH_{0.25}$ -66. Scale bars = 200 nm.



Figure S92: SEM image and SAXS spectra for S0.68-MH0.32-33 and S0.68-MH\*0.32-33. Scale bar = 200 nm.



Figure S93: SEM images and SAXS spectra for  $S_{0.66}$ -MH $_{0.34}$ -70 and  $S_{0.66}$ -MH $_{0.34}$ -70. Scale bars = 200 nm.



Figure S94: SEM images and SAXS spectra for  $S_{0.56}$ -MH $_{0.44}$ -52 and  $S_{0.56}$ -MH $_{0.44}$ -52. Scale bars = 200 nm.



Figure S95: SEM image and SAXS spectrum for  $S_{0.54}$ -MH<sub>0.46</sub>-45. Scale bar = 200 nm.







Figure S97: SEM images and SAXS spectrum for  $S_{0.52}$ -MH<sub>0.48</sub>-67 and  $S_{0.52}$ -MH\*<sub>0.48</sub>-67. Scale bars = 200 nm.



Figure S98: SEM images and SAXS spectra for  $S_{0.50}$ -MH $^{*}_{0.50}$ -91 and  $S_{0.50}$ -MH $^{A*}_{0.50}$ -91. Scale bars = 200 nm.



Figure S99: SEM images and SAXS spectra for  $S_{0.50}$ -MH $_{0.50}$ -45 and  $S_{0.50}$ -MH $_{0.50}$ -45. Scale bars = 200 nm.



Figure S100: SEM image for  $S_{0.49}$ -MH<sub>0.51</sub>-92. Scale bar = 200 nm.



Figure S101: SEM images for S0.46-MH0.54-81 and S0.46-MH\*0.54-81. Scale bars = 200 nm.



Figure S102: SEM images and SAXS spectra for S0.45-MH0.55-42 and S0.45-MH $^{50\%}$ \*0.55-42. Scale bars = 200 nm.



Figure S103: SEM images and SAXS spectra for  $S_{0.42}$ -MH<sub>0.58</sub>-89 and  $S_{0.42}$ -MH\*<sub>0.58</sub>-89. Scale bars = 200 nm.



Figure S104: SEM images and SAXS spectra for  $S_{0.42}$ -MH<sub>0.58</sub>-57 and  $S_{0.42}$ -MH\*<sub>0.58</sub>-57. Scale bars = 200 nm.



Figure S105: SEM image and SAXS spectrum for  $S_{0.39}$ -MH<sub>0.61</sub>-35. Scale bar = 200 nm.



Figure S106: SEM images and SAXS spectra for  $S_{0.37}$ -MH $_{0.63}$ -65 and  $S_{0.37}$ -MH $_{0.63}$ -65. Scale bars = 200 nm.



Figure S107: SEM images for  $S_{0.31}$ -MH<sub>0.69</sub>-48 and  $S_{0.31}$ -MH\*<sub>0.69</sub>-48. Scale bars = 200 nm.



Figure S108: SEM image and SAXS spectra for  $S_{0.28}$ -MH<sub>0.72</sub>-67 and  $S_{0.28}$ -MH\*<sub>0.72</sub>-67. Scale bar = 200 nm.



Figure S109: SEM image and SAXS spectra for  $S_{0.28}$ -MH $_{0.72}$ -29 and  $S_{0.28}$ -MH $_{0.72}$ -29. Scale bar = 200 nm.



Figure S110: SEM images and SAXS spectrum for  $S_{0.27}$ -MH $_{0.73}$ -70 and  $S_{0.27}$ -MH $_{0.73}$ -70. Scale bars = 200 nm.



Figure S111: SAXS spectra for S $_{0.16}$ -MH $_{0.84}$ -43 and S $_{0.09}$ -MH $_{0.91}$ -39.

## AFM and IR-SNOM phase images



Figure S112: AFM height images (top) and corresponding IR-SNOM near-field optical phase images (bottom), mapped at an independently addressable absorption band of the PMMA block at 1152 cm<sup>-1</sup>.

# Cuts along different planes of cylindrical and gyroidal morphologies



Figure S113: 3D modeled, rendered images cut along different planes of the ordered cylindrical (top and middle row) and gyroidal (bottom row) morphologies.

#### **Domain spacings**



Figure S114: Double logarithmic plot of the domain spacing  $d_{SAXS}$  against the molecular weight for the cylindrical samples. The dotted lines are the power law fits with the parameters stated below in Table S3. Blue = precursor BCP and red = functionalized BCP.



Figure S115: Double logarithmic plot of the domain spacing against the molecular weight for the lamellar morphology determined from  $d_{SEM}$ . The dark grey lines are the power law fits to  $d=a^*MW^b$ .

Morphology	Fit to power law	d ∼ MW <sup>ь</sup>	R <sup>2</sup>
Lamella (d <sub>saxs</sub> )	0.036 +/-0.012 * MW 0.63 +/- 0.03	0.63	0.98
Lamella functionalized (d <sub>SAXS</sub> )	0.040 +/- 0.035 * MW 0.59 +/- 0.08	0.59	0.95
Lamella (d <sub>sem</sub> )	0.141 +/- 0.011 * MW 0.50 +/- 0.07	0.50	0.80
Lamella functionalized (d <sub>SEM</sub> )	0.009 +/-0.012 * MW 0.73 +/- 0.12	0.73	0.93
Cylinder	0.043 +/- 0.001 * MW 0.62 +/- 0.20	0.62	0.85
Cylinder functionalized	0.187 +/- 0.490 * MW 0.47+/- 0.32	0.47	0.43*

## Table S3: Fit data for the power law for the different polymer types