# *Supporting Information for*

# **Composition-dependent emission colors for biomass-based main-chain luminescent liquid crystalline copolyesters with excellent tensile properties**

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#### **1. Experimental**

#### **1.1. Materials**

1,6-Hexanediol was purchased from Aladdin Reagent Co., Ltd. 4- Methoxyphenylacetonitrile, 4-methoxybenzaldehyde and tetraisopropyl orthotitanate Ti(OiPr)<sup>4</sup> were obtained from TCI. Diethyl [2,2'-bifuran]-5,5'-dicarboxylate (BFDCE) and (*Z*)-2,3-bis(4-ethoxyphenyl) acrylonitrile (*Z*-CS) were synthesized according to previous literatures.1-3

### **1.2. Characterizations**

<sup>1</sup>H nuclear magnetic resonance (NMR) spectra were measured on a Bruker AVANCE III-400 NMR instrument at room temperature. Fourier transform-infrared (FT-IR) spectra were taken on a Bruker Equinox 55 spectrometer in the attenuated total reflectance (ATR) mode. The number-average molar masses  $\binom{M_n}{n}$  and the polydispersity  $\binom{M_w}{M_n}$  were measured on the Waters 1515 GPC instrument at 40 °C using chloroform as the eluent at a flow rate of 0.5 mL min<sup>-1</sup>. The molecular weight calibration curves were obtained with polystyrene standards.  $UV-vis$  absorption spectra and fluorescence spectra were recorded using a Shimadzu UV-2550 diode-array spectrophotometer and EDINBURGH FS5 spectrofluorometer at room temperature, respectively. The solutions were tested in 1 cm quartz cuvettes, while the films prepared by spin coating the corresponding chloroform solutions with a concentration of 30 mg mL<sup>-1</sup> onto the quartz plate were directly tested. Polarized optical microscopic (POM) observations were carried out using a Zeiss Axio Scope P1 polarizing optical microscope with a hot stage (Linkam THMS 600) at different temperature. Thermogravimetric analyses (TGA) were performed on a Perkin Elmer Diamond

TGA8000 under an argon flow rate of 20 mL min−1 . Differential scanning calorimetry (DSC) was run on a DSC2A-01634 instrument at a heating or cooling rate of 10 °C min−1 under an argon atmosphere. X-ray diffraction (XRD) patterns for the aspolymerized samples were recorded on an X-ray diffraction meter (D/max 2500 PC, Rigaku) with a Cu-K $\alpha$  radiation source with a wavelength of 0.154 nm. The mechanical strength measurements were carried out on an UTM2103 testing machine with a transducer-coupled 1 kN load cell. The films with a thickness of  $100 \sim 200 \mu m$  were prepared by compression molding for testing, where an appropriate amount of the polymer sample was loaded between two iron plates that were preheated at 180 ℃ for 5 min.

**1.3. Syntheses of** *Z***-CS, BFDCE, and PBFCS**



**Scheme S1**. Synthetic routes for *Z*-CS (a), BFDCE (b), and PBFCS (c).

The syntheses of *Z*-CS and copolyesters were carried out according to the synthetic route illustrated in **Scheme S1**.

(*Z*)-2,3-Bis(4-hydroxyphenyl)acrylonitrile (CS-OH). 4-Methoxybenzaldehyde (0.88 g, 5.0 mmol) and sodium hydroxide (0.40 g, 10.0 mmol) were added to a three-necked flask containing 100 mL of ethanol. The flask was equipped with a reflux condenser, and the resulting mixture was stirred at 60 ℃ until it completely dissolved. Subsequently, 4-methoxyphenylacetonitrile (0.82 g, 5.0 mmol) was added to the solution and stirred at 80 ℃ for 4 h. The mixture was extracted using ethyl acetate and deionized water. The aqueous phase was then acidized by 36.5 wt % hydrochloric acid. After filtration, the residue was washed with deionized water for three times. The obtained product was dried in an oven at 60 ℃ for 24 h to afford *Z*-CS as a pale-yellow crystal (0.96 g, 66.3%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  = 13.23 (s, 2H), 8.30 (s, 1H), 8.08 (m, 6H), 7.94 (d, *J* = 8.3 Hz, 2H).

(*Z*)-2,3-Bis(4-ethoxyphenyl)acrylonitrile (*Z*-CS). CS-OH (1.47 g, 5.0 mmol) was added to a three-necked flask equipped with a reflux condenser containing 100 mL ethanol. Then, 1 mL of concentrated sulfuric acid was slowly added to the reaction flask. After stirring at 80 ℃ for 4 h, the mixture was cooled, extracted, and dried to obtain the desired product as a yellow solid  $(1.26 \text{ g}, 72.1\%)$ . <sup>1</sup>H NMR  $(400 \text{ MHz},$ CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 8.22 (s, 1H), 7.98 (dd,  $J$  = 8.9, 5.6 Hz, 6H), 7.86 (d,  $J$  = 8.3 Hz, 2H), 4.25 (q, *J* = 7.1 Hz, 4H), 1.25 (t, *J* = 7.1 Hz, 6H).

The synthetic procedures of PBF and the copolyesters are similar. Take the synthesis of P79.5 as an example. BFDCE (1.00 g, 3.60 mmol), *Z*-CS (0.32 g, 0.90 mmol), 1,6 hexanediol (0.53 g, 4.50 mmol), and a solution of  $Ti(OiPr)_4$  with a concentration of 0.3 wt % relative to the diester in ethylene glycol dimethyl ether (10 wt %) were added to a 50 mL three-necked flask equipped with an mechanical stirrer. The flask was evacuated to a low vacuum (below 100 Pa) and then filled with argon three times to reduce the generation of by-products. The mixture was then heated at 160  $^{\circ}$ C for 2 h, followed by 180 °C for 2 h under  $N_2$ . The reaction mixture was further heated at 235 °C for 3 h under reduced pressure (less than 100 Pa). After cooling to room temperature, the product was dissolved in a mixture solution of chloroform and trifluoroacetic acid (1:1) and then the solution was poured into ethanol (20 times). The precipitate was collected by centrifugation, and dried at 80 °C in vacuum for 48 h to afford P79.5 as a white block (1.12 g, 79.4%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 8.05 (m, 4H), 7.96 (d, 2H), 7.76 (d, 2H), 7.66 (s, 1H), 7.22 (d, 8H), 6.88 (d, 8H), 4.34 (q, 20H),  $1.82 - 1.77$  (m, 24H),  $1.52 - 1.48$  (m, 17H).

P86.7. Yield: 76.0%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 8.12 (m, 4H), 7.96 (d, 2H), 7.76 (d, 2H), 7.66 (s, 1H), 6.88 (d, 13H), 4.33 (t, 30H), 1.82-1.77 (m,  $37H$ ),  $1.54-1.23$  (m,  $23H$ ).

P88.6. Yield: 85.1%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 8.12 (m, 4H), 7.96 (d, 2H), 7.77 (d, 2H), 7.66 (s, 1H), 7.22 (d, 15H), 6.88 (d, 15H), 4.33 (t, 39H),  $1.82 - 1.77$  (m, 33H),  $1.52 - 1.48$  (m, 36H).

P90.5. Yield: 91.3%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 8.12 (m, 7.0 Hz, 4H), 7.96 (d, 2H), 7.76 (d, 2H), 7.66 (s, 1H), 7.23 (d, 19H), 6.88 (d, 19H), 4.33 (q, 48H), 1.82–1.77 (m, 46H), 1.53–1.48 (m, 39H).

P96.0. Yield: 87.4%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 8.17 (m, 4H), 7.96 (d, 2H), 7.77 (d, 2H), 7.66 (s, 1H), 7.23 (d, 48H), 6.88 (d, 48H), 4.33 (q, 118H),  $1.79$  (m, 92H),  $1.52-1.48$  (m, 116H).

PBF. Yield: 86.7%. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, TMS, ppm):  $\delta$  = 7.22 (d, 2H), 6.88  $(d, 2H)$ , 4.33 (t, 4H), 1.79 (t, 4H), 1.52–1.48 (m, 4H).

#### **1.4. Preparation of samples for latent fingerprints**

Firstly, a fingertip was pressed onto ethanol-cleaned glass slides. Secondly, a PBFCS solution in chloroform was gradually added into silica powder, which was continuously stirred until a uniform mixture formed. Finally, after chloroform was evaporated completely, a small amount of the prepared powder was then gently applied to the fingerprinted surface of the glass slides, and the excess powder was removed to obtain the latent fingerprints.

## **2. Supporting data**



**Figure S1.** <sup>1</sup>H NMR spectrum of *Z*-CS in DMSO- $d_6$  at room temperature.



Figure S2.<sup>1</sup>H NMR spectrum of BFDCE in CDCl<sub>3</sub> at room temperature.



Figure S3.<sup>1</sup>H NMR spectrum of PBF in CDCl<sub>3</sub> at room temperature.



**Figure S5.** <sup>1</sup>H NMR spectrum of P90.5 in CDCl<sub>3</sub> at room temperature.





**Figure S7.** <sup>1</sup>H NMR spectrum of P86.7 in CDCl<sub>3</sub> at room temperature.



**Figure S8.** <sup>1</sup>H NMR spectrum of P80.5 in CDCl<sub>3</sub> at room temperature.



**Figure S9.** ATR-FTIR spectra of PBF (a), P96.0 (b), P90.5 (c), P88.6 (d), P86.7 (e) and P79.5 (f) in films.



**Figure S10.** Fluorescence spectra of PBF in DCM/ethanol mixtures with different ethanol volume fraction  $({}^{f}e)$  at room temperature.



Figure S11. Emission spectra and excitation spectra of *Z*-CS and BFDCE in CHCl<sub>3</sub> (20 mg/mL) (a) and in solid state (b).



**Figure S12.** Plot of the maximum emission wavelength of PBFCS versus  $\phi_{\text{BFDCE}}$ .



**Figure S13.** TGA curves of BFDCE, *Z*-CS, PBF and PBFCS at a heating rate of 10 °C min<sup>-1</sup> under nitrogen atmosphere.



**Figure S14.** DSC curves of BFDCE (a) and *Z*-CS (b) at a rate of  $10^{\circ}$ C min<sup>-1</sup> under nitrogen atmosphere.



**Figure S15.** DSC curves of PBF and PBFCS on the second heating scans (a) and first cooling (b) at a rate of  $10^{\circ}$ C min<sup>-1</sup> under nitrogen atmosphere.



**Figure S16.** Glass transition temperature of the copolyesters as a function of BFDCE content and the fitting lines from Fox (black line), Fried (red line), and Gordon-Taylor (blue line) equations.

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 $+ w_{CS}$ 

 $T_{g,~PCS}$ 

 $T_{g,PBF}$ 

The experimental data can be fitted well by the following equations,

$$
Fox \, Equ.: \frac{1}{T_{g, copolymer}} = \frac{w_{BFDCE}}{T_{g, PBF}} + \frac{w_{CS}}{T_{g, PCS}}
$$
\n
$$
(1)
$$
\n
$$
w_{CS} \times ln^{[0]}_{\infty}(\frac{T_{g, PCS}}{T_{g, PBF}})
$$
\n
$$
\ln(T_{g, copolymer}/T_{g, PBF}) = \frac{T_{g, PCS}}{T_{g, PBF}}
$$

Fried Equ.:

(2)

$$
T_{g, copolymer} = \frac{w_{BFDCE} \times T_{g,PBF} + k \times w_{CS} \times T_{g, PCS}}{w_{BFDCE} + k \times w_{CS}}
$$

(3)

where  $W_{BFDCE}$  and  $W_{CS}$  are the weight fractions of BFDCE and *Z*-CS, respectively; PBF and PCS represent the corresponding homopolyesters obtained from BFDCE and *<sup>Z</sup>*-CS, respectively; *<sup>k</sup>* is an adjustable parameter.

 $W_{BFDCE}$   $\times \frac{1}{7}$ 

**Table S1**. The fitting parameters of Fox, Fried and Gordon-Taylor equations.

Equation name Fitted equation $\overline{f}$ g, PCS (K)	
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**Figure S17.** XRD patterns and analysis data for PBF (a), P90.5 (b), P88.6 (c), P86.7 (d), and P80.5 (e).



**Figure S18.** Stress-strain curves (a) and the partial curves (b) of PBFCS.

**Table S2**. Young's modulus (*E*), tensile strength at break (<sup> $\sigma$ </sup>b), elongation at break ( ${}^{\epsilon}$ b) of common engineering plastics and general-purpose plastics.<sup>a</sup>

sample	conditions	E(MPa)	$\sigma_b$ (MPa)	$\varepsilon_b$ (%)		
<b>Engineering plastics</b>						
Nylon 6,6	23 °C	3300	82.7-90.3	$30 - 60$		
	50% relative humidity (23 $\degree$ C)		62.1-77.2	$200 - 300$		
Polycarbonate	ASTM: D638	2380	62.1	110		
Polyoxymethylene	296 K, ASTM D638	2825	60.6	40-75		
Poly(butylene terephthalate)	ASTM D638	2600	55	200-300		
<b>General-purpose plastics</b>						
Polyethylene, high-density	$25^{\circ}$ C	60-290	10-60	18-40		
Polyethylene, low-density	D 638	100-310	$9 - 15$	100-800		
Polystyrene	Unoriented	3200-3400	$30 - 60$	$1 - 4$		
Poly(ethylene terephthalate)		1700	50	180		
Acrylonitrile-butadiene elastomers	ACN (20-40%)	$2.9 - 3.6$	13.4-17.9	387-466		
Polypropylene, isotactic	APAO, Eastoflex <sup>TM</sup> P1010 and P1023	1.38	7	100-200		
Poly(vinyl chloride)	With 10% dioctyl phthalate	2964	55.5	104		



<sup>a</sup> Source: Data collected in Mark, J. E. *Polymer Data Handbook*. Oxford University Press. **1998**.

**Figure S19.** UV-vis absorption spectra of P96.5 (a), P88.6 (c), P86.7 (e), and P79.5 (g) in CHCl<sub>3</sub> upon 365 nm UV irradiation, and P96.5 (b), P88.6 (d), P86.7 (f), and P79.5 (h) in CHCl<sub>3</sub> then by 254 nm UV irradiation for different times at room temperature.



Figure S20.<sup>1</sup>H NMR spectra of P79.5 in CDCl<sub>3</sub> before and after irradiation with 365 nm UV light  $(20 \text{ mW cm}^{-2})$  for 30 min, and subsequent 254 nm UV light  $(20 \text{ mW cm}^{-2})$  for 60 min.



**Figure S21.** Fluorescence spectra of P96.5 (a), P88.6 (c), P86.7 (e), and P79.5 (g) in films upon 365 nm UV irradiation (20 mW cm<sup>-2</sup>), and P96.5 (b), P88.6 (d), P86.7 (f), and P79.5 (h) in films then by 254 nm UV irradiation (20 mW cm<sup>-2</sup>) for different times at room temperature.



**Figure S22.** FT-IR spectra of P79.5 before and after 365 nm UV irradiation.

#### **References**

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