Supplementary Information

Structure-Property Relationships to Direct the Dynamic Properties of Acylsemicarbazide-Based Materials

Stefan J.D. Maessen,^a Siebe Lekanne Deprez,^b Pascal Vermeeren,^b Bart W.L. van den Bersselaar,^a Martin Lutz,^c Johan P.A Heuts,^a Célia Fonseca Guerra^b and Anja R.A. Palmans*a

- a. Department of Chemical Engineering & Chemistry and Institute for Complex Molecular Systems, Eindhoven University of Technology, P.O. Box 513, 5600 MB Eindhoven, The Netherlands.
- b. Department of Chemistry and Pharmaceutical Sciences, Amsterdam Institute of Molecular and Life Sciences (AIMMS), Vrije Universiteit Amsterdam, De Boelelaan 1108, 1081 HZ Amsterdam, The Netherlands.
- c. Structural Biochemistry, Bijvoet Centre for Biomolecular Research, Faculty of Science, Utrecht University, Universiteitsweg 99, 3584 CG Utrecht, The Netherlands.

*corresponding author:

a.palmans@tue.nl

Contents

1. Experimental section

1.1 Materials and Methods

Materials

Unless stated otherwise, all reagents and chemicals were obtained from commercial sources at the highest purity available and used without further purification. Polydimethylsiloxane, hydride terminated, viscosity 7-10 cSt and polydimethylsiloxane, monohydride terminated, viscosity 5-9 cSt were obtained from ABCR. All solvents were obtained from Biosolve except for acetonitrile which was obtained from Actu-All Chemicals. Dry solvents were obtained from an MBraun solvent purification system (MB SPS-800). Dry unstabilized THF from the solvent purification system was stored under argon and further dried with activated 4 Å molecular sieves, and filtered through a Whatman 2 um PTFE syringe filter before use. Deuterated chloroform (CDC l_3) was purchased from Cambridge Isotope Laboratories and kept dry with 4 Å molecular sieves. Deuterated DMSO (DMSO- d_6) and deuterated THF (THF- d_8) were purchased from Deutero GmbH and used as received.

General Methods

Reactions were followed by thin-layer chromatography (TLC) 60-F254 silica gel plates from Merck and visualized by 254 nm UV light or by staining with a ceric ammonium molybdate solution. Automated column chromatography was performed on a Grace Reveleris X2 automated column machine with Reveleris Silica Flash Cartridges and on a Biotage Isolera 1 with Biotage SNAP KP-SIL columns.

NMR spectroscopy

¹H, ¹³C NMR spectra were recorded on a 400 MHz Bruker Avance III Spectrometer (¹H using 400 MHz, ¹³C using 100 MHz). Peak multiplicities are abbreviated as s: singlet, d: doublet, t: triplet, q: quartet, p: pentet, m: multiplet, dd: doublet of doublets, td: triplet of doublets, tt: triplet of triplets. Proton and carbon chemical shifts are reported in parts per million (ppm) downfield from tetramethylsilane (TMS) using the deuterated solvent resonance frequency as internal standard (7.26 ppm for CDCl₃ and 2.50 ppm for DMSO- d_6).

Matrix-assisted laser desorption/ionization time-of-flight

Matrix-assisted laser desorption/ionization time-of-flight followed by mass spectrometry (MALDI-ToF-MS) mass analysis measurements were performed on a Bruker Autoflex Speed using α-cyano-4-hydroxycinnamic acid (CHCA) and *trans*-2-[3-(4-tert-butylphenyl)-2-methyl-2 propenylidene]malononitrile (DCTB) as matrices. Samples for MALDI-ToF-MS were dissolved with a concentration of 1 mg mL $^{-1}$ in either chloroform, DCM or THF.

Fourier transform infrared spectroscopy

Fourier transform infrared spectroscopy (FTIR) spectra were recorded on a Shimadzu IRTracer-100 spectrometer using the MIRacle 10 single reflection ATR accessory.

Growth of single crystals

Crystals suitable for single-crystal X-ray diffraction were prepared via a vapor diffusion method. Saturated solutions of PhASCPh and PhASCBz in methanol were prepared and diluted twofold. An open GCMS vial was charged with 0.2 mL of this half-saturated solution, which was then placed in a large closed vial containing 2 mL of countersolvent (diethyl ether for PhASCPh, methyl *tert*-butyl ether for PhASCBz). After several days, the formation of crystals was observed.

X-Ray crystal structure determination of PhASCPh

 $C_{14}H_{13}N_3O_2$, Fw = 255.27, colorless needle, 0.33 \times 0.15 \times 0.05 mm³, orthorhombic, Pbca (no. 61), a = 9.9581(3), b = 9.1704(3), c = 27.6575(7) Å, V = 2525.70(12) Å³, Z = 8, D_x = 1.343 $g/cm³$, $\mu = 0.09$ mm⁻¹. The diffraction experiment was performed on a Bruker Kappa ApexII diffractometer with sealed tube and Triumph monochromator ($\lambda = 0.71073$ Å) at a temperature of 150(2) K up to a resolution of (sin θ/λ)_{max} = 0.65 Å⁻¹. The intensities were integrated with the Eval15 software¹. A multi-scan absorption correction and scaling was performed with SADABS² (correction range 0.65-0.75). A total of 41429 reflections was measured, 2887 reflections were unique ($R_{int} = 0.062$), 2119 reflections were observed [$|>2\sigma(1)|$. The structure was solved with Patterson superposition methods using SHELXT.³ Structure refinement was performed with SHELXL-2018⁴ on F^2 of all reflections. Non-hydrogen atoms were refined freely with anisotropic displacement parameters. All hydrogen atoms were located in difference Fourier maps. N-H hydrogen atoms were refined freely with isotropic displacement parameters, C-H hydrogen atoms were refined with a riding model. 184 Parameters were refined with no restraints. R1/wR2 $[1 > 2\sigma(1)]$: 0.0453 / 0.1023. R1/wR2 [all refl.]: 0.0683 / 0.1130. S = 1.062. Residual electron density between -0.19 and 0.23 e/\AA ³. Geometry calculations and checking for higher symmetry was performed with the PLATON program.⁵

X-Ray crystal structure determination of PhASCBz

 $C_{15}H_{13}N_3O_3$, Fw = 283.28, colorless plate, 0.37 \times 0.35 \times 0.08 mm³, monoclinic, P2₁/c (no. 14), a = 9.7633(3), b = 27.5437(7), c = 10.4855(3) Å, β = 98.644(2) °, V = 2787.70(13) Å³, Z = 8, D_x = 1.350 g/cm³, μ = 0.10 mm⁻¹. The diffraction experiment was performed on a Bruker Kappa ApexII diffractometer with sealed tube and Triumph monochromator ($\lambda = 0.71073$ Å) at a temperature of 150(2) K up to a resolution of (sin θ/λ)_{max} = 0.65 Å⁻¹. The intensities were integrated with the Eval15 software¹. A multi-scan absorption correction and scaling was performed with SADABS² (correction range 0.68-0.75). A total of 53558 reflections was

measured, 6410 reflections were unique ($R_{int} = 0.034$), 5233 reflections were observed $[1>2_σ(1)]$. The structure was solved with Patterson superposition methods using SHELXT.³ Structure refinement was performed with SHELXL-2018⁴ on F^2 of all reflections. Non-hydrogen atoms were refined freely with anisotropic displacement parameters. All hydrogen atoms were located in difference Fourier maps. N-H hydrogen atoms were refined freely with isotropic displacement parameters, C-H hydrogen atoms were refined with a riding model. 403 Parameters were refined with no restraints. R1/wR2 $\left|1 > 2\sigma(1)\right|$: 0.0402 / 0.1033. R1/wR2 [all refl.]: 0.0503 / 0.1084. S = 1.060. Residual electron density between -0.33 and 0.43 e/ A^3 . Geometry calculations and checking for higher symmetry was performed with the PLATON program.⁵

Compression molding

Compression molding was performed in a Fontijne LabEcon 300 hot press. The pDMS-ASC were placed in a mold and sandwiched between 0.05 mm thick Teflon sheets and 4 mm thick stainless steel sheets, which was placed under 100 kN of force at 140 °C for 60 minutes. The samples were cooled rapidly and inspected. In all cases, more polymer was added and the samples were molded again under 100 kN of force at 140 °C for 30 minutes. The pDMS-ASC polymers were pressed into 0.7-1 mm thick films, from which rectangles were cut for DMTA experiments and 8 mm disks were punched for creep experiments.

Size Exclusion Chromatography (SEC)

SEC was performed on Varian/Polymer Laboratories PL-SEC 50 equipment equipped with refractive index (RI) and UV (254 nm) detectors. The system was equipped with a combination of Agilent PLgel 5-um mixed-C column and Agilent PLgel 5-um mixed-D column. The mobile phase was THF with BHT stabilizer and run at a flow rate of 1 mL min⁻¹ at 40 °C. Polystyrene was used as calibration standard.

Thermogravimetric analysis (TGA)

The thermal stabilities of the pDMS-ASC materials were determined using a TGA 550 (TA instruments) under N_2 flow. Temperature ramps were performed by ramping from 20 °C to 40 °C at 20 °C min-1 , keeping the sample isothermal at 40 °C for 5 minutes, followed by a ramp to 800 °C at 10 °C min⁻¹. Temperature calibration was performed using the Curie points of high purity ferromagnetic standards.

Differential scanning calorimetry (DSC)

DSC was performed in a Q2000 DSC (TA instruments) calibrated with an indium standard. Measurements were performed using standard aluminum pans between -40 °C and 160 °C. Heating and cooling runs were performed at 10 $^{\circ}$ C min⁻¹. The first cooling and second heating runs were reported for all materials.

Variable temperature infrared spectroscopy (VTIR)

VTIR spectra were obtained at the Bruker Tensor 27. A sample was heated to 160 or 200 °C and cooled back to 25 °C, both with a heating rate of 3 °C min⁻¹. At 200 °C, the sample was first equilibrated for 5 min before cooling. IR spectra were collected every 5 °C.

Dynamic mechanical thermal analysis (DMTA)

DMTA was performed on rectangular shaped compression molded samples using the film tension setup in a Discovery DMA 850 (TA instruments). Samples were cooled to -150 °C and left to equilibrate for 2 hours. Then, a temperature ramp was performed at 3 $^{\circ}$ C min⁻¹ from -

150 °C to 200 °C. A preload force of 1.0 N and a force track of 125% was used. The storage and loss modulus were recorded as a function of temperature.

Medium- and wide-angle x-ray scattering (MAXS and WAXS)

Bulk small-angle X-ray scattering (SAXS) was performed on a Ganesha instrument from SAXSLab. The flight tube and sample holder are all under vacuum in a single housing, with a GeniX-Cu ultra-low divergence X-ray generator. The source produces X-rays with a wavelength (λ) of 0.154 nm and flux of 1 × 108 ph s⁻¹. Scattered X-rays were captured on a 2dimensional Pilatus 300K detector with 487 × 619 pixel resolution. The sample-to-detector distance was 0.084 m (WAXS mode) or 0.48 m (MAXS mode). The instrument was calibrated with diffraction patterns from silver behenate.

Probing dynamics in bulk experiments

Around 50 mg of the pDMS-ASC polymers were dissolved in 20 mL of THF (using a heat gun for ASC-HDI and ASC-PDI, at room temperature for ASC-BDI). Around 50 mg of the corresponding monoASCs (H-monoASC for ASC-HDI, P-monoASC for ASC-PDI and BmonoASC for ASC-BDI) were added to the polymer and dissolved. The mixtures were divided equally over 10 vials (2 mL per vial) and the solvent was allowed to evaporate over the course of 24 hours, after which the mixtures were dried in a vacuum oven at 50 °C overnight. For the measurements, an oven was preheated at 120 °C and 7 vials were put in. The vials were taken out at time intervals of 1, 2, 4, 8, 16, 24 and 48 hours. The mixtures were then dissolved in 4 mL of THF (using a heat gun for ASC-HDI and ASC-PDI, at room temperature for ASC-BDI) and SEC measurements were performed $(\sim 2 \text{ mg} \text{ mL}^{-1}$ in THF). For ASC-PDI, the samples were additionally diluted two-fold due to aggregation during the SEC measurements (~1 mg) mL⁻¹ in THF). The SEC chromatograms were baseline corrected at a retention time of 12 minutes, and normalized on the highest intensity polymer peak above 2000 Da.

Creep-recovery measurements

Creep recovery measurements were performed using a Discovery HR 20 (TA instruments) with environmental temperature control (ETC) setup and 8 mm parallel-plate geometry. All experiments were performed with an axial force range of 3 ± 0.1 N. Samples were equilibrated at the measurement temperature for 3 minutes, after which they were subjected to an oscillatory time sweep for 2 minutes under 1.0% oscillatory strain and at an angular frequency of 1 Hz. Creep measurements were performed between 100 and 140 °C. 20 kPa of shear stress was applied to the sample for 1800 seconds, with a recovery period of 5400 seconds. The slope of the creep compliance was determined between 1400 and 1800 s.

Tensile tests

Tensile tests were performed on rectangular shaped samples using the film tension setup in a Discovery DMA 850 (TA instruments). Samples were compression molded to a thickness between 0.3 and 0.4 mm for these measurements. A sample with an initial length of 5.0 mm \pm 0.2 mm and a width of 5.3 mm was clamped in the setup. A rate of 0.1 mm/min (~2%/min) was used to put the sample in tension. The stress-strain curves were obtained at room temperature.

Frequency sweep experiments

Frequency sweep experiments from 10^2 rad/s to 10^{-3} rad/s were performed using a Discovery HR 20 (TA instruments) with environmental temperature control (ETC) setup and 8 mm parallel-plate geometry. The oscillation amplitude was set to 1% strain.

1.2 Synthetic procedures

Synthesis of *tert***-butyl hydrazide:** Methyl pivalate (5 mL, 38 mmol, 1 eq) was added to a 25 mL round-bottom flask equipped with an oil bath and reflux cooler. Hydrazine monohydrate (10 mL, 206 mmol, 5.5 eq) was to the flask and the mixture was stirred at 80 °C overnight. The liquid was dried in under high vacuum for 6 hours, after which a white crystalline powder was obtained which was used without further purification $(3.5 g, 80\%)$. ¹H NMR $(400 MHz, DMSO$ *d*6) *δ* = 8.78 (s, 1H), 3.99 (s, 2H), 1.08 (s, 9H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 177.31, 37.59, 27.79.

Synthesis of ASC model compounds: *General procedure:* The hydrazide (1 eq) was dissolved in acetonitrile (dried on 3 Å molecular sieves) in a dried 50 mL round-bottom flask under argon atmosphere. The isocyanate (1 eq) was added to the suspension directly and the mixture was left to stir overnight. The next day, the suspension was filtered using filter paper in a funnel and washed with 2x 10 mL of acetonitrile and with 2x 10 mL of diethyl ether. The solid was collected in a vial and dried in a vacuum oven at 50 $^{\circ}$ C overnight, yielding the pure product.

MeASCPh: 300 mg (4.05 mmol) of acethydrazide in 10 mL of acetonitrile, 0.440 mL (4.05 mmol) of phenyl isocyanate, 10 mL of acetonitrile. Product obtained as a white solid (573 mg, 73%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 9.63 (d, *J* = 2.1 Hz, 1H), 8.70 (s, 1H), 7.96 (d, *J* = 2.1 Hz, 1H), 7.44 (d, *J* = 7.3 Hz, 2H), 7.25 (t, *J* = 7.2 Hz, 2H), 6.95 (tt, *J* = 7.0 Hz, 1.2 Hz, 1H), 1.87 (s, 3H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 169.29, 155.43, 139.66, 128.65, 121.89, 118.50, 20.64. MALDI-ToF-MS: m/z calc. for C₉H₁₁N₃O₂: 193.09 Da; found 194.12 Da [M+H]⁺, 216.09 Da [M+Na]⁺.

BuASCPh: 300 mg (2.94 mmol) of butyric acid hydrazide in 10 mL of acetonitrile, 0.321 mL (2.94 mmol) of phenyl isocyanate. Product obtained as a white solid (427 mg, 66%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 9.59 (s, 1H), 8.68 (s, 1H), 7.96 (s, 1H), 7.44 (d, *J* = 7.4 Hz, 2H), 7.24 (t, *J* = 7.6, 2H), 6.94 (tt, *J* = 7.4, 1.2 Hz, 1H), 2.12 (t, *J* = 7.3 Hz, 2H), 1.55 (h, *J* = 7.4 Hz, 2H), 0.89 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 172.10, 155.41, 139.68, 128.65, 121.82, 118.36, 35.11, 18.38, 13.58. MALDI-ToF-MS: m/z calc. for $C_{11}H_{15}N_3O_2$: 221.12 Da; found 222.15 Da [M+H]⁺, 244.12 Da [M+Na]⁺.

BnASCPh: 500 mg (3.33 mmol) of phenylacetic hydrazide in 10 mL of acetonitrile, 0.362 mL (3.3 mmol) of phenyl isocyanate. Product obtained as a white solid (820 mg, 91%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 9.95 (s, 1H), 8.72 (s, 1H), 8.08 (s, 1H), 7.44 (d, *J* = 7.3 Hz, 2H), 7.35 – 7.20 (m, 7H), 6.95 (t, *J* = 7.4 Hz, 1H), 3.50 (s, 2H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 170.07, 155.31, 139.61, 135.73, 129.12, 128.67, 128.23, 126.48, 121.89, 118.42, 40.14. MALDI-ToF-MS: m/z calc. for C₁₅H₁₅N₃O₂: 269.12 Da; found 270.14 Da [M+H]⁺, 292.11 Da [M+Na]⁺, 308.08 Da [M+K]⁺.

PhASCPh: 500 mg (3.67 mmol) of benzhydrazide in 10 mL of acetonitrile, 0.400 mL (3.67 mmol) of phenyl isocyanate. Product obtained as a white solid (832 mg, 89%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 10.30 (s, 1H), 8.86 (s, 1H), 8.18 (s, 1H), 7.93 (d, *J* = 6.9 Hz, 2H), 7.58 (tt, *J* = 7.6 Hz, 1.2 Hz, 1H), 7.54 – 7.43 (m, 4H), 7.26 (t, *J* = 7.3 Hz, 2H), 6.96 (tt, *J* = 7.3, 1.2 Hz, 1H). ¹³C NMR (100 MHz, DMSO-d₆) δ = 166.42, 155.65, 139.69, 132.58, 131.78, 128.65,

128.39, 127.56, 121.90, 118.53. MALDI-ToF-MS: m/z calc. for C₁₄H₁₄N₃O₂: 255.10 Da; found 256.10 Da [M+H]⁺, 278.08 Da [M+Na]⁺, 294.06 Da [M+K]⁺.

*t***-BuASCPh:** 500 mg (3.67 mmol) of *tert*-butyl hydrazide in 15 mL of acetonitrile, 0.470 mL (3.67 mmol) of phenyl isocyanate. Product was hot-filtrated with acetonitrile and the filtrate was recrystallized from acetonitrile due to some diphenyl urea impurities. Obtained as a colorless crystals (280 mg, 28%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 9.36 (d, *J* = 2.1 Hz, 1H), 8.67 (s, 1H), 7.82 (s, 1H), 7.43 (d, *J* = 7.4 Hz, 2H), 7.26 (t, *J* = 7.3 Hz, 2H), 6.95 (t, *J* = 7.4 Hz, 1H), 1.16 (s, 9H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 177.97, 155.98, 140.18, 129.16, 122.21, 118.61, 37.86, 27.63. MALDI-ToF-MS: m/z calc. for C₁₂H₁₇N₃O₂: 235.13 Da; found 236.17 Da [M+H]⁺, 258.14 Da [M+Na]⁺, 274.11 Da [M+K]⁺.

PhASCBu: 300 mg (2.20 mmol) of benzhydrazide in 20 mL of acetonitrile, 0.245 mL (2.20 mmol) of butyl isocyanate. Product obtained as a white solid (78 mg, 15%). Low yield obtained due to gelation of the reaction mixture leading to loss of stirring during the reaction. ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 10.10 (s, 1H), 7.89 (d, *J* = 7.2 Hz, 2H), 7.78 (s, 1H), 7.56 (t, *J* = 7.3 Hz, 1H), 7.48 (t, *J* = 7.5 Hz, 2H), 6.45 (t, *J* = 5.7 Hz, 1H), 3.02 (q, *J* = 6.6 Hz, 2H), 1.38 (p, *J* = 7.0 Hz, 2H), 1.27 (h, *J* = 7.2 Hz, 2H), 0.87 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 166.28, 158.33, 132.76, 131.58, 128.26, 127.53, 38.86, 31.99, 19.42, 13.72. MALDI-ToF-MS: m/z calc. for C₁₂H₁₇N₃O₂: 235.13 Da; found 236.18 Da [M+H]⁺, 258.14 Da [M+Na]⁺, 274.29 Da [M+K]⁺.

PhASCBn: 500 mg (3.67 mmol) of benzhydrazide in 10 mL of acetonitrile, 0.454 mL (3.67 mmol) of benzyl isocyanate. Product obtained as a white solid (678 mg, 69%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 10.18 (s, 1H), 7.98 (s, 1H), 7.91 (d, *J* = 7.0 Hz, 2H), 7.56 (tt, *J* = 7.6 Hz, 1.2 Hz, 1H), 7.48 (t, *J* = 7.4 Hz, 2H), 7.35 – 7.16 (m, 5H), 7.08 (s, 1H), 4.25 (d, *J* = 6.1 Hz, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ = 166.41, 158.50, 140.65, 132.74, 131.64, 128.27, 128.10, 127.61, 126.94, 126.49, 42.63. MALDI-ToF-MS: m/z calc. for $C_{15}H_{15}N_3O_2$: 269.12 Da; found 270.14 Da [M+H]⁺, 292.11 Da [M+Na]⁺, 308.08 Da [M+K]⁺.

PhASCBz: 369 mg (2.79 mmol) of benzhydrazide in 10 mL of acetonitrile, 369 mg (2.71 mmol) of benzoyl isocyanate. Product obtained as a white solid (680 mg, 89%). ¹H NMR (400 MHz, DMSO-*d*6) *δ* = 11.11 (s, 1H), 10.67 (s, 1H), 10.20 (s, 1H), 8.03 (d, *J* = 7.2 Hz, 2H), 7.93 (d, *J* = 7.0 Hz, 2H), 7.69 – 7.49 (m, 6H). ¹³C NMR (100 MHz, DMSO-*d*6) *δ* = 168.07, 165.55, 153.77, 133.05, 132.23, 132.19, 131.98, 128.61, 128.51, 128.34, 127.56. MALDI-ToF-MS: m/z calc. for $C_{15}H_{13}N_3O_3$: 283.10 Da; found 284.11 Da [M+H]⁺, 306.09 Da [M+Na]⁺, 322.06 Da [M+K]⁺.

Synthesis of methyl ester-terminated pDMS: Polydimethylsiloxane, hydride terminated, viscosity 7-10 cSt (40.0 g, 1 eq) was mixed with methyl undec-10-enoate (14.0 g, 70.6 mmol, \approx 2.4 eq) in bulk in a dried 100 mL round-bottom flask under argon atmosphere. The mixture was cooled with a room-temperature water bath, and Karstedt's catalyst (2 wt% in xylene) was added (5-6 drops), after which the mixture was stirred for 24 hours. The mixture was transferred to a separatory funnel and extracted with 3x 50 mL of acetonitrile to remove excess methyl undec-10-enoate. After this, the polymer oil was dissolved in 100 mL of DCM and filtered over a plug of activated charcoal in a glass filter to remove the platinum catalyst. The charcoal plug was washed with 3x 50 mL of DCM. The solvent was removed in *vacuo* after which the product was obtained as a colorless oil (50.0 g, 99%). ¹H NMR (400 MHz, CDCl₃) δ = 3.65 (s, 6H), 2.29 (t, *J* = 7.5 Hz, 4H), 1.62 (p, *J* = 7.3 Hz, 4H), 1.39 – 1.17 (m, 28H), 0.57 – 0.47 (m, 4H), 0.06 (m, 96H). ¹³C NMR (100 MHz, CDCl₃) δ = 174.43, 51.52, 34.26, 33.60, 29.69, 29.65, 29.52, 29.42, 29.32, 25.12, 23.38, 18.42, 1.31, 1.20, 1.18. SEC (RI detector): M_n : 1.7 kg mol⁻¹, M_w : 1.9 kg mol⁻¹, PDI: 1.1.

Synthesis of acyl hydrazide terminated pDMS: Methyl ester-terminated pDMS (45.0 g) was dissolved in 150 mL of isopropanol in a 250 mL round-bottom flask equipped with an oil bath and reflux cooler. Hydrazine monohydrate (22.0 g, 0.44 mol) was added to the solution, after which the mixture was left to stir at 50 °C for 72 hours. The isopropanol was removed in *vacuo*, and the mixture was dissolved in 150 mL of DCM. The mixture was transferred to a separatory funnel and extracted with 3x 200 mL of demineralized water to remove excess hydrazine. After this, the DCM layer was dried over $Na₂SO₄$, filtered using gravity and the filter washed with 50 mL of DCM. The solvent was removed in *vacuo* after which the product was obtained as a colorless viscous oil (43.0 g, 96%). ¹H NMR (400 MHz, CDCl3) *δ* = 6.77 (s, 2H), 3.90 (s, 4H), 2.14 (t, *J* = 7.6 Hz, 4H), 1.62 (p, *J* = 7.2 Hz, 4H), 1.37 – 1.17 (m, 28H), 0.52 (t, *J* = 7.5 Hz, 4H), 0.24 – -0.17 (m, 96H). ¹³C NMR (100 MHz, CDCl3) *δ* = 174.24, 34.70, 33.57, 29.68, 29.64, 29.50, 29.45, 29.43, 25.66, 23.34, 18.38, 1.29, 1.17, 1.15. SEC (UV detector): M_n: 2.0 kg mol⁻ 1 , M_w: 2.8 kg mol⁻¹, PDI: 1.4. SEC (RI detector): M_n: 1.4 kg mol⁻¹, M_w: 1.9 kg mol⁻¹, PDI: 1.3.

$$
C1 \longrightarrow C1 \longrightarrow N H_3 \longrightarrow N H_2N \longrightarrow N H_2 \longrightarrow C1 \longrightarrow 0 \longrightarrow C2 \longrightarrow N^2 C^2
$$

Synthesis of isophthalamide: Ammonium hydroxide (50 mL, 25% in water) was added to a dried 100 mL round-bottom flask that was cooled with an ice bath and equipped with a dropping funnel under argon. Isophthaloyl chloride (5.0 g, 24.6 mmol) was dissolved in 20 mL of dry THF and transferred to the dropping funnel. The THF solution was added dropwise to the flask over 5 minutes, after which the mixture was stirred for an additional 15 minutes. The mixture was allowed to warm to room temperature after which the precipitate was filtered using a Büchner filter and washed with 3x 100 mL of demineralized water. over a plug of activated charcoal in a glass filter to remove the platinum catalyst. The charcoal plug was washed with 3x 50 mL of DCM. The solid dried in a vacuum oven at 60 °C overnight after which the product was obtained as a white powder (3.8 g, 95%). ¹H NMR (400 MHz, DMSO-*d*₆) δ = 8.39 (t, *J* = 1.8 Hz, 1H), 8.07 (s, 2H), 8.01 (dd, *J* = 7.7, 1.8 Hz, 2H), 7.53 (t, *J* = 7.7 Hz, 1H), 7.49 (s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ = 167.41, 134.23, 129.88, 128.06, 126.59.

Synthesis of isophthaloyl diisocyanate (BDI): Isophthalamide (2.0 g, 12.2 mmol, 1 eq) was added to a dried 500 mL flat-bottom flask equipped with an oil bath. The flask was purged several times by alternating high vacuum and argon. 1,2-Dichloroethane (90 mL, dried on 4 Å molecular sieves and filtered through a Whatman 2 µm PTFE syringe filter) was added to the flask. Oxalyl chloride (3.6 mL, 42.0 mmol, 3.4 eq) was added to the mixture, after which the mixture was left to stir at 80 °C for 24 hours. Then, a clear solution was obtained. The temperature was reduced to 60 °C and the solvent was removed in *vacuo* via the Schlenk line with a cold trap in between. The flask was refilled with argon, and the stopper was exchanged for a sublimation cooler. The product was purified via sublimation for between 80-100 °C at high vacuum for 4 hours, forming colorless crystal needles on the sublimation cooler. The flask was evacuated and transferred to a glove-box, in which the flask was opened and the pure product was obtained as colorless crystals (1.6 g, 61%). The product was stored in the glovebox due its extreme water sensitivity. ¹H NMR (400 MHz, CDCl3) *δ* = 8.73 (td, *J* = 1.7, 0.6 Hz, 1H), 8.34 (dd, *J* = 7.8, 1.8 Hz, 2H), 7.64 (td, *J* = 7.8, 0.6 Hz, 2H). ¹³C NMR (100 MHz, CDCl3) *δ* = 164.40, 136.21, 132.67, 132.43, 132.09, 129.76.

Synthesis of ASC-HDI: Hydrazide terminated pDMS (2.026 g) was dissolved in 15 mL of dry THF in a 100 mL round-bottom flask equipped with an oil bath under argon. The mixture was heated to 50 °C while stirring vigorously. Hexamethylene diisocyanate (HDI, 179 mg, 1.06 mmol) was dissolved in a vial in 5 mL of dry THF and added to the solution. Within 30 minutes, a gel was formed. Part of the solvent was removed in vacuo and the gel was further dried in a vacuum oven at 60 °C overnight after which the product was obtained as a translucent, opaque plastic (2.2 g, quantitative yield). ¹H NMR (400 MHz, THF-*d*₈) *δ* = 9.54 (m, 1H), 7.75 (m, 1H), 6.36 (m, 1H), 3.18 (m, 4H), 2.17 (m, 4H), 1.60 (m, 4H), 1.32 (m, 36H), 0.70 – 0.45 (m, 4H), $0.33 - 0.16$ (m, 108H). Solubility too poor for ¹³C NMR. SEC (UV detector): M_n: 13.9 kg mol⁻¹, M_w : 24.2 kg mol⁻¹, PDI: 1.7. SEC (RI detector): M_n : 20.9 kg mol⁻¹, M_w : 28.7 kg mol⁻¹, PDI: 1.4.

Synthesis of ASC-PDI: Hydrazide terminated pDMS (2.0 g) was dissolved in 20 mL of dry THF in a 100 mL round-bottom flask equipped with an oil bath under argon. The mixture was heated to 50 °C while stirring vigorously. 1,3-Phenylene diisocyanate (PDI, 191 mg, 1.19 mmol) was dissolved in 4 mL of dry THF after which it was added to the solution. Within 60 minutes, a gel was formed. Part of the solvent was removed in vacuo and the gel was further dried in a vacuum oven at 50 °C overnight after which the product was obtained as a slightly yellow, translucent, opaque plastic (2.2 g, quantitative yield). ¹H NMR (400 MHz, THF-*d*8) *δ* = 9.48 (s, 1H), 8.46 (s, 1H), 7.86 (s, 1H), 7.00 (m, 3H), 2.17 (m, 4H), 1.59 (m, 4H), 1.49 – 1.10 (m, 29H), 0.58 (t, *J* = 7.7 Hz, 4H), 0.33 – -0.17 (m, 92H). Solubility too poor for ¹³C NMR. SEC (UV detector): M_n: 7.7 kg mol⁻¹, M_w: 16.5 kg mol⁻¹, PDI: 2.1. SEC (RI detector): M_n: 7.8 kg mol⁻¹, M_w: 16.5 kg mol-1 , PDI: 2.1.

Synthesis of ASC-BDI: Hydrazide terminated pDMS (2.0 g) was dissolved in 10 mL of dry THF in a 50 mL round-bottom flask in a glovebox. The mixture was stirred vigorously. Isophthaloyl diisocyanate (BDI, 272 mg, 1.26 mmol) was dissolved in 5 mL of dry THF after which it was added to the solution. The mixture was left to stir for 1 hour, after which it was precipitated in 150 mL of cold methanol to remove cyclic oligomers. The precipitate was centrifuged, and the isolated solid was dried in a vacuum oven at 50 °C for 4 hours. The

polymer was redissolved in 10 mL of THF and the precipitation procedure was repeated once more. The polymer was dried in a vacuum oven at 50 °C overnight after which the product was obtained as a translucent, opaque plastic (1.6 g, 70%). ¹H NMR (400 MHz, THF-*d*8) *δ* = 10.15 (s, 3H), 9.23 (s, 2H), 8.58 (s, 1H), 8.19 (d, *J* = 7.8 Hz, 2H), 7.65 (t, *J* = 7.8 Hz, 1H), 2.31 – 2.06 (m, 4H), 1.63 (m, 4H), 1.33 (m, 28H), 0.68 – 0.46 (m, 4H), 0.30 – -0.13 (m, 97H). Solubility too poor for ¹³C NMR. SEC (UV detector): M_n: 13.5 kg mol⁻¹, M_w: 31.3 kg mol⁻¹, PDI: 2.3. SEC (RI detector): M_n: 15.9 kg mol⁻¹, M_w: 31.7 kg mol⁻¹, PDI: 2.0.

Synthesis of monofunctional methyl ester-terminated pDMS: Polydimethylsiloxane, monohydride terminated, viscosity 5-9 cSt (10.0 g, 1 eq) was combined with methyl undec-10 enoate (2.35 g, 11.9 mmol, ~1.4 eq) in bulk in a dried 50 mL round-bottom flask under argon atmosphere. The mixture was cooled with a room-temperature water bath, and Karstedt's catalyst (2 wt% in xylene) was added (3-4 drops), after which the mixture was left to stir for 24 hours. The bulk mixture was transferred to a separatory funnel and extracted with 3x 25 mL of acetonitrile to remove excess methyl undec-10-enoate. After this, the polymer oil was dissolved in 30 mL of DCM and filtered over a plug of activated charcoal in a glass filter to remove the platinum catalyst. The charcoal plug was washed with 3x 15 mL of DCM. The solvent was removed in *vacuo* after which the product was obtained as a colorless oil (10.4 g, 89%). ¹H NMR (400 MHz, CDCl3) *δ* = 3.66 (s, 3H), 2.30 (t, *J* = 7.5 Hz, 2H), 1.61 (p, *J* = 7.2 Hz, 2H), 1.29 (m, 19H), 0.93 – 0.84 (t, 3H), 0.60 – 0.46 (m, 4H), 0.25 – -0.16 (m, 78H). ¹³C NMR (100 MHz, CDCl3) *δ* = 174.45, 51.54, 34.27, 33.62, 29.71, 29.66, 29.54, 29.44, 29.34, 26.53, 25.62, 25.14, 23.39, 18.43, 18.12, 13.96, 1.32, 1.20, 0.34, 0.32. SEC (RI detector): M_n: 1.3 kg mol⁻¹, M_w: 1.5 kg mol-1 , PDI: 1.1.

$$
\sim\!\!<\!\!\cdot\! s_{i1}^{10} \cdot s_{i2}^{11} \cdot s_{i1}^{10} \cdot \sqrt{s_{i2}^{10}} \cdot \sqrt{s_{i3}^{10} \cdot s_{i1}^{10}} \cdot \sqrt{s_{i3}^{10} \cdot s_{i1}^{10}} \cdot \sqrt{s_{i3}^{10} \cdot s_{i1}^{10}} \cdot \sqrt{s_{i3}^{10} \cdot s_{i2}^{10}} \cdot \sqrt{s_{i3}
$$

Synthesis of monofunctional acyl hydrazide terminated pDMS: Monofunctional methyl ester-terminated pDMS (10.0 g) was dissolved in 10 mL of isopropanol in a 50 mL roundbottom flask equipped with an oil bath and reflux cooler. Hydrazine monohydrate (4.0 g, 79.9 mmol) was added to the solution, after which the mixture was left to stir at 50 °C for 24 hours. The isopropanol was removed in *vacuo*, and the mixture was dissolved in 100 mL of DCM. The mixture was transferred to a separatory funnel and extracted with 3x 100 mL of demineralized water to remove excess hydrazine. After this, the DCM layer was dried over $Na₂SO₄$, filtered using gravity and the filter washed with 50 mL of DCM. The solvent was removed in *vacuo* after which the product was obtained as a colorless viscous oil (9.0 g, 90%). ¹H NMR (400 MHz, CDCl3) *δ* = 6.64 (s, 1H), 3.88 (s, 2H), 2.20 – 2.08 (m, 2H), 1.63 (p, *J* = 7.3 Hz, 2H), 1.29 (m, 18H), 0.93 – 0.83 (m, 3H), 0.59 – 0.45 (m, 4H), 0.25 – -0.16 (m, 81H). ¹³C NMR (100 MHz, CDCl3) *δ* = 174.21, 34.73, 33.63, 33.60, 29.70, 29.67, 29.52, 29.47, 29.45, 26.50, 25.66, 25.59, 23.37, 18.41, 18.09, 13.94. SEC (UV detector): M_n : 1.7 kg mol⁻¹, M_w : 2.2 kg mol⁻¹, PDI: 1.3. SEC (RI detector): M_n : 1.3 kg mol⁻¹, M_w : 1.6 kg mol⁻¹, PDI: 1.3.

$$
\text{Tr}_{\text{S}_1}^{\text{I}}\text{Tr}_{\text{S}_1}^{\text{I}}\text{Tr}_{\text{S}_1}^{\text{I}}\text{Tr}_{\text{S}_1}^{\text{II}}\text{Tr}_{\text{S}_1}^{\text{II}}+\text{C}_2^{\text{C}_2N}\text{Tr}_{\text{S}_2}^{\text{II}}\text{Tr}_{\text{S}_1}^{\text{II}}\text{Tr}_{\text{S}_1}^{\text{II}}\text{Tr}_{\text{S}_1}^{\text{II}}\text{Tr}_{\text{S}_1}^{\text{II}}\text{Tr}_{\text{S}_1}^{\text{II}}\text{Tr}_{\text{S}_2}^{\text{II}}\text{Tr}_{\text{
$$

Synthesis of MonoASC with butyl isocyanate (H-monoASC): Monofunctional hydrazideterminated pDMS (1.0 g, 1 eq) was dissolved in 10 mL of dry THF in a 50 mL round-bottom flask equipped with an oil bath and reflux cooler. Phenyl isocyanate (0.164 mL, 1.48 mmol, 2 eq) was added to the solution, after which the mixture was left to stir for 24 hours. Ammonium hydroxide (1 mL, 25% in water) was added to the mixture to quench excess isocyanate. Then, 50 mL of chloroform and 50 mL of demineralized water were added to the mixture, which was transferred to a separatory funnel and extracted with 3x 50 mL of demineralized water. After this, the organic layer was dried over $Na₂SO₄$, filtered using gravity and the solvent was removed in *vacuo*. The product was further purified using flash column chromatography (liquid injection from DCM), with an eluent system of 10 column volumes of 100% DCM, followed by 20% acetone in DCM. The product fractions were combined and the solvent was removed in *vacuo* after which the product was obtained as colorless waxy solid (603 mg, 56%). ¹H NMR (400 MHz, CDCl3) *δ* = 8.28 (s, 1H), 7.65 (s, 1H), 5.63 (s, 1H), 3.18 (q, *J* = 6.6 Hz, 2H), 2.23 (t, *J* = 7.7 Hz, 2H), 1.64 (m, 2H), 1.54 – 1.41 (m, 2H), 1.41 – 1.17 (m, 20H), 1.02 – 0.78 (m, 6H), 0.61 – 0.42 (m, 4H), 0.26 – -0.18 (m, 73H). ¹³C NMR (100 MHz, CDCl3) *δ* = 172.99, 158.21, 40.05, 34.29, 33.65, 32.23, 29.78, 29.72, 29.58, 29.54, 29.47, 26.51, 25.59, 25.57, 23.40, 20.11, 18.43, 18.10, 13.95, 13.89, 1.31, 1.20, 0.32. SEC (UV detector): M_n : 1.4 kg mol⁻¹, M_w : 1.5 kg mol⁻¹, PDI: 1.1. SEC (RI detector): M_n : 1.2 kg mol⁻¹, M_w : 1.4 kg mol⁻¹, PDI: 1.1.

Synthesis of MonoASC with phenyl isocyanate (P-monoASC): Monofunctional hydrazideterminated pDMS (1.0 g, 1 eq) was dissolved in 10 mL of dry THF in a 50 mL round-bottom flask equipped with an oil bath and reflux cooler. Phenyl isocyanate (0.161 mL, 1.48 mmol, 2 eq) was added to the solution, after which the mixture was left to stir for 24 hours. Ammonium hydroxide (1 mL, 25% in water) was added to the mixture to quench excess isocyanate. 50 mL of chloroform and 50 mL of demineralized water were added to the mixture. Then, the mixture was transferred to a separatory funnel and extracted with 3x 50 mL of demineralized water. After this, the organic layer was dried over $Na₂SO₄$, filtered using gravity and the solvent was removed in *vacuo*. The product was further purified using flash column chromatography (liquid injection from DCM), with an eluent system of 10 column volumes of 100% DCM, followed by 5% acetone in DCM. The product fractions were combined and the solvent was removed in *vacuo* after which the product was obtained as colorless waxy solid (631 mg, 58%). ¹H NMR (400 MHz, CDCl3) *δ* = 8.78 (s, 1H), 8.24 (s, 1H), 8.17 (s, 1H), 7.28 (d, *J* = 7.6 Hz, 3H), 7.19 (t, *J* = 7.7 Hz, 2H), 6.98 (t, *J* = 7.4 Hz, 1H), 2.26 (t, *J* = 7.7 Hz, 2H), 1.69 – 1.52 (m, 2H), 1.43 – 1.12 (m, 18H), $0.97 - 0.81$ (m, 3H), 0.53 (m, 4H), $0.26 - 0.17$ (m, 81H). ¹³C NMR (100 MHz, CDCl3) *δ* = 173.98, 155.67, 138.26, 129.03, 123.43, 119.52, 34.23, 33.67, 29.79, 29.70, 29.58, 29.50, 29.45, 26.51, 25.59, 25.49, 23.41, 18.44, 18.10, 13.95, 1.32, 1.20, 0.33. SEC (UV detector): M_n : 1.4 kg mol⁻¹, M_w : 1.6 kg mol⁻¹, PDI: 1.1. SEC (RI detector): M_n : 1.4 kg mol⁻¹, M_w : 1.5 kg mol-1 , PDI: 1.1.

Synthesis of MonoASC with benzoyl isocyanate (B-monoASC): Monofunctional hydrazide-terminated pDMS (0.5 g, 1 eq) was dissolved in 10 mL of dry THF in a 50 mL roundbottom flask equipped with an oil bath and reflux cooler. Benzoyl isocyanate (65 mg, 0.44 mmol, 1.2 eq) was added to the solution, after which the mixture was left to stir for 24 hours. Then, 50 mL of chloroform and 50 mL of demineralized water were added to the mixture, which was transferred to a separatory funnel and extracted with 3x 50 mL of demineralized water. After this, the organic layer was dried over $Na₂SO₄$, filtered using gravity and the solvent was removed in *vacuo*. The product was further purified using flash column chromatography (liquid injection from DCM), with an eluent system of 15 column volumes of 100% DCM, followed by

5% acetone in DCM. The product fractions were combined and the solvent was removed in *vacuo* after which the product was obtained as colorless waxy solid (366 mg, 66%). ¹H NMR (400 MHz, CDCl3) *δ* = 10.33 (d, *J* = 3.1 Hz, 1H), 8.85 (s, 1H), 7.87 (d, *J* = 7.1 Hz, 2H), 7.61 (t, *J* = 7.4 Hz, 1H), 7.56 (s, 1H), 7.49 (t, *J* = 7.7 Hz, 2H), 2.28 (t, *J* = 7.6 Hz, 2H), 1.69 (p, *J* = 7.6 Hz, 2H), 1.43 – 1.15 (m, 18H), 0.95 – 0.82 (m, 3H), 0.61 – 0.44 (m, 4H), 0.25 – -0.17 (m, 77H). ¹³C NMR (100 MHz, CDCl₃) δ = 171.71, 167.52, 153.62, 133.61, 129.14, 127.88, 34.38, 33.63, 29.75, 29.68, 29.55, 29.50, 29.42, 26.50, 25.59, 25.47, 23.39, 18.42, 18.09, 13.95, 1.32, 1.20, 0.33. SEC (UV detector): M_n : 1.5 kg mol⁻¹, M_w : 1.6 kg mol⁻¹, PDI: 1.1. SEC (RI detector): M_n : 1.4 kg mol⁻¹, M_w : 1.6 kg mol⁻¹, PDI: 1.1.

2. Crystal structures of ASC model compounds

Figure S1: Molecular structure of PhASCPh in the crystal. Displacement ellipsoids are drawn at the 50% probability level, hydrogen atoms with arbitrary radii.

Figure S2: Topology of the hydrogen bonding network in the crystal structure of PhASCPh. Molecules are simplified to spheres (nodes). The connecting lines represent N-H…O hydrogen bonds. The hydrogen bonded two-dimensional sheets are oriented parallel to the

crystallographic a,b-plane. The simplification and the drawing were created with the ToposPro software⁶.

Table 31. Selected to Sign angles Fight the Grystal Structure of Final Summa					
LC2-C1-N1-C7	151.63(16)	C7-N2-N3-C8	61.83(18)		
LC1-N1-C7-N2	171.06(13)	N2-N3-C8-C9	$-169.31(13)$		
l N1-C7-N2-N3	-167.70(13)	N3-C8-C9-C10	11.9(2)		

Table S1: Selected torsion angles [°] in the crystal structure of PhASCPh.

Table S2: Hydrogen bonding geometries in the crystal structure of PhASCPh.

	$D-H[\AA]$	HA _[A]	DA [A]	$D-HA$ ^{[o}]
$N1-H1NO1$ ⁱ	0.87(2)	1.97(2)	2.7919(17)	158.1(16)
$N2-H2NO1$ ⁱ	0.878(19)	2.190(18)	2.9480(18)	144.4(14)
N3-H3NO2 ⁱⁱ	0.886(18)	1.965(19)	2.8361(17)	167.5(16)

Symmetry codes *i*: ½-x, y+½, z; *ii*: x-½, y, ½-z.

2.2 PhASCBz

Figure S3: Two independent molecules of PhASCBz in the asymmetric unit of the crystal structure. Projection along the crystallographic c-axis. Displacement ellipsoids are drawn at the 50% probability level, hydrogen atoms with arbitrary radii.

Figure S4: Overlay of the two independent molecules in the crystal structure of PhASCBz.

Figure S5: Topology of the hydrogen bonding network in the crystal structure of PhASCBz. Molecules are simplified to spheres (nodes). The connecting lines represent N-H…O hydrogen bonds. The hydrogen bonded one-dimensional chains are running along the crystallographic a-axis. The two independent molecules take part in the same chain. The simplification and the drawing were created with the ToposPro software 6 .

	$x = 1$	$x = 2$			
$C2x-C1x-C7x-N1x$	5.65(19)	$-20.1(2)$			
$C1x-C7x-N1x-C8x$	$-178.61(11)$	178.17(12)			
$C7x-N1x-C8x-N2x$	$-9.51(18)$	$-1.9(2)$			
$N1x-C8x-N2x-N3x$	173.53(10)	$-175.06(11)$			
$C8x-N2x-N3x-C9x$	85.95(15)	68.96(15)			
$N2x-N3x-C9x-C10x$	$-175.04(11)$	$-162.43(10)$			
$N3x-C9x-C10x-C11x$	32.08(18)	$-172.40(11)$			

Table S3: Selected torsion angles [°] in the two independent molecules in the crystal structure of PhASCBz.

Symmetry codes *i*: x+1, y, z, z; *ii*: 1-x, 1-y, -z.

CCDC 2377270 and 2377271 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

3. NMR kinetics experiments

Dissociation kinetics of ASC model compounds were measured by first making a 50 mM stock solution of benzylamine in DMSO- d_6 with added dimethyl sulfone (DMSO₂) as internal standard. This stock solution was added to the model compounds to a concentration of 50 mM, making a 1:1 ratio of model compound to benzylamine. Subsequently, 0.6 mL of the mixture was transferred to an NMR tube and submerged in an oil bath at the desired temperature. At certain time intervals, an NMR tube was quenched in ice water. Then, a 1 H NMR spectrum was recorded and the same sample was used for the next time interval. Dissociation rates were determined by setting the integral of $DMSO₂$ to 100, and following the decrease in intensity of an ASC model compound peak over time. The peaks used for integration is mentioned below the respective model compound in Table S5. As the system follows pseudo first-order kinetics, k_d was determined from the slope of $\ln\left(\frac{c}{a}\right)$ $\frac{c}{c_0}$) as a function of time (eqn(S1)). The activation energy, E_{act} , was determined from the slope of k_d as a function of inverse temperature (eqn(S2)).

$$
\ln\left(\frac{c}{c_0}\right) = -k_d t \tag{S1}
$$

$$
k_d = Ae^{-\frac{E_{act}}{RT}}
$$
 (S2)

Table S5: Model compound kinetics data. Chemical shift *δ* of the peak tracked to follow conversion is indicated below the corresponding ASC model compound (in ppm).

Model	Temperature	Time	Peak signal	ln(c/c ₀)	k_{d}	$E_{\rm act}$
compound	$(^{\circ}C)$	(min)	$(DMSO2 =$	(\cdot)	(min^{-1})	$(kJ \text{ mol}^{-1})$
			100)			
MeASCPh	130	$\overline{0}$	46.01	$\overline{0}$	0.0179	112 ± 7
δ = 1.87		10	38.41	0.181		
		20	32.04	0.362		
		30	26.9	0.537		
		40	22.59	0.711		
	120	$\overline{0}$	53.68	$\overline{0}$	0.00962	
		15	46.34	0.147		
		30	39.92	0.296		
		45	34.76	0.435		
		60	30.32	0.571		
	110	$\overline{0}$	46.61	Ω	0.00364	
		15	44.15	0.0542		
		30	41.86	0.107		
		45	39.49	0.166		
		60	37.48	0.218		
	100	0	46.4	0	0.00127	
		30	44.47	0.0425		
		60	43.24	0.0705		
		90	41.29	0.117		
		120	39.83	0.153		
BuASCPh	130	$\overline{0}$	28.35	$\overline{0}$	0.0223	112 ± 6
$\delta = 2.12$		10	22.64	0.225		
		20	17.91	0.459		
		30	14.45	0.674		
		40	11.74	0.882		

Figure S6: Example ¹H NMR spectra in DMSO- d_6 of PhASCPh during kinetic experiment at 120 °C.

Figure S7: Kinetic measurements of model ASC at various temperatures. A) MeASCPh, B) BuASCPh, C) BnASCPh, D) *t*-BuASCPh, E) PhASCBu, F) PhASCBn and G) PhASCBz. The lines represent linear fits through the origin of the corresponding data points.

Figure S8: Arrhenius plots of ASCs with various hydrazide (A) and isocyanate (B) substituents. The lines represent linear fits of the corresponding data points.

4. Supporting figures

Figure S9: Crystal structure of PhASCPh. Hydrogen bonds visualized in green using the CrystalExplorer⁷ software.

Figure S10: TGA curves of ASC-HDI (A), ASC-PDI (B) and ASC-BDI (C). Heating rate is 10 $^{\circ}$ C min⁻¹.

Figure S11: Wavenumbers of N-H stretch maximum of pDMS-ASC materials as a function of temperature.

Figure S12: Variable temperature infrared spectroscopy spectrum of ASC-BDI during first heating (A) and first cooling (B) with intervals of 5 °C.

Figure S13: Variable temperature infrared spectroscopy spectrum of ASC-PDI up to 160 °C during first heating (A) and first cooling (B) with intervals of 5 °C.

Figure S14: Variable temperature infrared spectroscopy spectrum of ASC-PDI up to 200 °C during first heating (A) and first cooling (B) with intervals of 5 °C.

Detailed analysis of VTIR of ASC-HDI

Before heating, ASC-HDI shows two well-defined N-H stretch peaks at 3217 and 3344 cm⁻¹, C=O stretches at 1656 and 1588 cm⁻¹, and an N-H bending peak at 1549 cm⁻¹ (Figure S14). Upon heating, the N-H stretches broaden and eventually fuse into one feature. The carbonyl peaks shift to higher wavenumbers at elevated temperatures and the relative intensities of the peaks change drastically. Upon cooling, all shifted peaks return to their values before heating, but there are significant differences in intensity remaining. The intensity differences before and after heating suggest that kinetic trapping of the material takes place during compression molding. The unchanged peak positions, however, suggest that the trapped state is very similar to the thermodynamically stable state. To check whether the morphology was stable after heating once, a second VTIR heating run was performed (Figure S14C and D). This run showed similar behavior to the first run, with now only minor intensity differences before and after heating.

Figure S15: Variable temperature infrared spectroscopy spectrum of ASC-HDI during first heating (A), first cooling (B), second heating (C) and second cooling (D) with intervals of 5 °C.

To test the stability of the hydrogen bonding patterns of the pDMS-ASC materials, isothermal IR spectra were taken at 140 °C (Figure S15). Here, for ASC-HDI there is some gradual change in the N-H and C=O stretch regions over a duration of 4 hours, indicating some change in hydrogen-bonding patterns. The other pDMS-ASC materials showed little to no change in hydrogen-bonding patterns during this time period.

 $\boldsymbol{\mathsf{A}}$

Figure S16: Isothermal IR spectra of ASC-HDI (A), ASC-PDI (B) and ASC-BDI (C) at 140 °C for 4 hours with intervals of 10 minutes.

Figure S17: A) Creep compliance of ASC-HDI at various temperatures. B) Recoverable compliance of ASC-HDI at various temperatures. Stress is 20 kPa.

28

Figure S18: A) Creep compliance of ASC-PDI at various temperatures. B) Recoverable compliance of ASC-PDI at various temperatures. Stress is 40 kPa.

Figure S19: A) Creep compliance of ASC-BDI at various temperatures. B) Recoverable compliance of ASC-BDI at various temperatures. Stress is 20 kPa.

Figure S20: DSC thermograms of the first cooling and second heating of ASC-HDI (A), ASC-PDI (B) and ASC-BDI (C). Heating rate and cooling rates are 10 °C min⁻¹. Exo up.

Figure S22: Frequency sweep data at different temperatures under 1% strain of ASC-HDI (A), ASC-PDI (B) and ASC-BDI (C).

5. Detailed DFT analysis of model compound crystal structures

This section contains analyses of hydrogen bonding motives present in crystal structures obtained for PhASCPh and PhASCBz (see Scheme S1).

Scheme S1: Structures of ASC motives considered in this study.

First, the PhASCPh motif is studied through comparing the experimentally determined crystal structure with the computationally optimized crystal structure (section 5.1). Hydrogen-bond patterns are examined, and the effect of hydrogen bonds on both experimental and theoretical IR spectra is established. Similar analyses for PhASCBz crystals are discussed in section 5.2.

5.1 PhASCPh

Crystal structure

Experimentally determined crystal structures for PhASCPh $(R_1, R_2$ = Phenyl; CIF ID: I1330a) were taken as a starting point for periodic DFT calculations at a ZORA-BLYP-D3(BJ)/TZP level of theory (see Computational Details). The structures are reported in Figure S23 and several points can be observed.

First, the computational and experimental structures show a high degree of similarity, particularly in their structural patterns (Figure S23). Bond angles and lengths are identical with a margin of 3 degrees and 0.02 Å, respectively. The most notable differences occur in C–H and N–H bond lengths, which are elongated by a maximum of 0.15 Å in the theoretical structure. However, as will be discussed in later sections, these variations only affect the magnitude of interaction strengths while the trends remain consistent. Consequently, the computationally determined crystal structure is considered valid and hence used for further, detailed analyses. Data derived from the experimental structure are always highlighted in grey.

Figure S23: Experimental and optimized crystal structure of PhASCPh (A) including a schematic representation (B) and structural HB patterns, being "bifurcated HB" and "single HB" (C). Values in parenthesis depict experimental bond lengths in Å. Computed at a ZORA-BLYP-D3(BJ)/TZP level of theory.

Second, the crystal structure exhibits multiple hydrogen bond (HB) patterns that form chains in different directions. The schematic representation (Figure S23B and C) illustrates these patterns, being the "bifurcated HB" and "single HB" patterns. Bifurcated HBs occur between urea scaffolds (part of the ASC motif, shown in purple) with hydrogen bond lengths (r_0, n) of 2.74 and 2.88 Å. Single HBs (shown in blue) form between the remaining atoms of the ASC motif with a length of 2.80 Å. These hydrogen-bonding patterns create chains of bifurcated HBs in one direction, which are connected perpendicularly by single HBs. Both the single HBs, *i.e.*, in and out of the screen, and the bifurcated HB chains, *i.e.*, up and down, alternate directions (Figure S23).

Interaction decomposition

Both hydrogen-bond patterns are analyzed further to determine their strength and compared to other intermolecular interactions present, such as the interaction between the phenyl substituents. To accomplish this, the interacting PhASCPh dimer is fragmented and bonds between R₁/R₂ and ASC motifs are replaced by hydrogen atoms⁸: *r* (**H–**C_{ph}) = 1.083 Å, *r* (**H–** C=O) = 1.08 Å), and *r* (**H–**NH) = 1.01 Å (see Scheme S2). The fragmentation defines two fragments, A and B, with various ASC and substituents combinations, allowing us to isolate the hydrogen bond interaction (ASC_A-ASC_B) from interactions between substituents.

Scheme S2: Fragmentation scheme for studying the interaction between two R₁ASCR₂ monomers forming a hydrogen-bonded dimer.

The decomposition results are shown in Table S6. The interaction energy (ΔE_{int}) summed over all fragmentations closely approximates the interaction energy of the complete dimer, with a maximum difference of 1.6 kcal mol⁻¹, validating the employed fragmentation scheme. The decomposition reveals that the hydrogen bond component is the most important interaction for both the bifurcated and single HB dimer, accounting for half of the total interaction energy in both HB patterns. The energy decomposition analysis (EDA, see Theoretical Methods) shows strong electrostatic interactions (ΔV_{elstat}) between the negatively charged HB acceptor (C=O^{δ –}) and positively charged HB donor ($N-H^{\delta+}$), alongside significant donor-acceptor interactions (ΔE_{oi}), where the lone pair on the C=O donates electrons to the σ^* orbital of the H–N.^{9,10} The other half of the interaction in the PhASCPh dimer stems from contributions of phenyl–phenyl $(\pi-\pi$ stacking) and mixed ASC–phenyl interactions, each contributing a small amount to the overall interaction.

		Bifurcated Hydrogen Bond				
System	Structure	ΔE_{int}	ΔV_{elstat}	$\Delta E_{\rm Pauli}$	$\Delta E_{\text{o}i}$	$\Delta E_{\rm disp}$
$[PhASCPh]_A -$		-20.2	-24.3	36.0	-13.6	-18.3
[PhASCPh] _B		-20.3	-17.1	20.8	-7.3	-16.7
$ASCA$ –		-10.1	-19.9	25.1	-10.9	-4.5
ASC _B		-10.4	-13.1	11.9	-5.1	-4.0
$Ph_A -$		-2.5	-2.0	5.3	-0.8	-5.0
Ph_B		-2.7	-1.5	4.1	-0.7	-4.6
$Ph_A -$		-2.1	-1.0	2.1	-0.7	-2.6
Ph _B		-2.2	-0.7	1.5	-0.5	-2.5
$[Ph+Ph]_A-$		-1.6	-0.7	0.8	-0.5	-1.2
ASC _B		-1.5	-0.6	0.7	-0.4	-1.1
$ASCA$ –		-5.5	-3.7	8.0	-2.1	-7.6
$[Ph+Ph]_B$		-5.7	-2.9	5.5	-1.6	-6.8
Sum ^[b]		-21.8	-27.2	41.3	-14.9	-20.9
		-22.5	-18.9	23.7	-8.3	-19.0
		Single Hydrogen Bond				
$[PhASCPh]_A -$		-15.3	-18.4	24.6	-11.2	-10.3
$[PhASCPh]_{B}$		-14.3	-12.3	12.8	-5.5	-9.3
$ASCA$ -		-7.2	-11.8	13.6	-7.0	-2.0
ASC _B		-6.3	-8.2	7.1	-3.3	-1.9
$Ph_A -$	\mathcal{X}^{\bullet}	-1.6	-1.0	3.1	-0.8	-2.9
Ph _B	y-L	-1.8	-0.5	1.7	-0.5	-2.5
$Ph_A -$		-0.2	0.0	0.1	0.0	-0.3
Ph_B		-0.2	0.0	0.1	0.0	-0.3
$[Ph+Ph]_A$ –		-2.5	-3.3	4.5	-1.7	-2.0
ASC _B		-2.6	-2.1	2.3	-1.0	-1.9
$ASC_A -$		-2.5	-1.7	3.9	-1.5	-3.2
$[Ph+Ph]_B$		-2.7	-0.9	2.0	-0.9	-2.9
Sum ^[b]		-14.0 -13.6	-17.8 -11.8	25.1 13.1	-11.0 -5.7	-10.3 -9.3

Table S6: Gas-phase decomposition of the interaction energy $\Delta E_{\text{int}}^{[a]}$ (in kcal mol⁻¹) of PhASCPh dimers decomposed into components defined in Scheme S2.

[a] Computed at a ZORA-BLYP-D3(BJ)/TZ2P (ADF) // ZORA-BLYP-D3(BJ)/TZP (BAND) level of theory with values obtained using the experimental crystal in gray.
[b] The sum of the total interaction energy and decomposition terms of the substituent-only, ASC-only, and mixed fragmentations.

Cooperativity analysis

When studying the interactions for the two distinctive patterns found in PhASCPh crystals, it is found that the hydrogen bond accounts for 50% of the total dimer interaction through favorable orbital interactions and electrostatics. Previous work on hydrogen bonds in supramolecular systems has shown that highly structured, chain-like hydrogen bonds exhibit cooperativity: the phenomenon that the hydrogen-bond strength of chains with n monomers is substantially higher than (n−1) times the hydrogen-bond strength of a dimer. This means that adding monomers to the chain becomes more favorable.^{9,11,12}

Given that the structural patterns in the PhASCPh crystal are chain-like (bifurcated and single HB, see Figure S23), the degree of cooperativity was examined by elongating the chain for both patterns, from a dimer (1+1) to a pentamer (4+1). The results presented in Table S7, indicate significant cooperativity for both patterns. As the chain lengthens, the interaction energy (ΔE_{int}) becomes more stabilizing, meaning that there is a stronger interaction between a longer chain and a monomer. For example, the bifurcated chain's interaction strength increases from -20.2 (1+1) to -24.5 (4+1) kcal mol⁻¹. The increase or the longer chains is smaller and goes to an asymptotic value. Additionally, charge transfer between a longer chain and an additional monomer increases, as shown by the VDD charges (ΔQ_n^{VDD} , see Theoretical Methods), which originates from stronger orbital interactions: +54 to +63 milli-electrons for the bifurcated HB pattern and +39 to +45 millielectrons for the single HB pattern. Note that the VDD charges obtained using the experimental crystal are smaller because the hydrogen bonds are longer, and the donor– acceptor interactions are weaker.

Bifurcated HB Single HB $n=2$ $\overline{1}$ $n=2$ *r*O-N = 2.74 (2.97) & 2.88 (2.95) Å *r*O-N = 2.80 (2.84) Å

Table S7: Cooperativity analysis of hydrogen-bonded PhASCPh chains. The interaction energy, ΔE_{int} (in kcal mol⁻¹) and associated VDD charges (in milli-electrons) are tabulated.^[a]

[a] computed at a ZORA-BLYP-D3(BJ)/TZ2P level of theory with values obtained with experimental structures in gray.

[b] The elongation direction is from bottom to top. Note that trends remain the same regardless of the elongation direction (except that the sign of charge transfer is inverted).

IR studies

Finally, the experimental and theoretical IR spectra of PhASCPh are reported to further understand the impact of the hydrogen bonds on the IR spectra (see Figure S24). These include spectra for the PhASCPh crystal (both experimental and theoretical) and the PhASCPh monomeric structure (theoretical). Peaks from bonds involving in hydrogen bonding are highlighted, including three N–H stretch vibrations between 3200 and 3500 cm– 1 and two C=O stretch vibrations around 1670 cm $^{-1}$.

Comparing the monomer IR spectrum (no hydrogen bonding) with the crystal IR spectrum (involving hydrogen bonding) reveals that the stretch vibrations corresponding to the orange and purple N–H bonds (See Figure S24 for the color scheme) undergo a redshift of 268 and 158 cm⁻¹, respectively. This originates from the hydrogen bond-induced weakening of the N–H bonds. Upon hydrogen-bond formation, electrons are transferred from the HB acceptor to the anti-bonding σ^* of the N–H bond, which induces and elongation of the N–H by 0.01 Å, as observed in the crystal geometry. The N–H stretch vibration denoted with the color blue in Figure S24, on the other hand, remains nearly unchanged, because the hydrogen bond comprising this N–H bond is significantly longer and weaker with respect to the hydrogen bonds with the prior discussed orange and purple N–H bonds (Figure S24).

Figure S24: IR spectra comparison of PhASCPh between the experimental crystal (A) with computed spectra of the crystal (B) and monomer (C). Highlighted peaks are associated with stretching modes.

5.2 PhASCBz

Crystal Structure

A procedure similar to that used for PhASCPh was applied to obtain the theoretical crystal structure of PhASCBz (R_1 = Phenyl, R_2 = Benzoyl; CIF ID: I1349a). DFT calculations were conducted at the ZORA-BLYP-D3(BJ)/TZP level of theory, starting from the experimental structure. Unlike PhASCPh, the PhASCBz monomer exhibits multiple conformers within the crystal structure that are not energetically equivalent: monomers in the repeating unit cell adopt distinct geometries with an energy difference of 15 kcal mol⁻¹ (see main text Figure 2). Furthermore, the hydrogen-bonding patterns form a stairs-like structure in both the theoretical and experimental structures (Figure S25).

Three distinct hydrogen-bond patterns contribute to this structure. The "Double HB" pattern (Figure S25, purple) comprises two hydrogen bonds oriented in opposite directions, connecting two ASC motifs to form each "step" of the stairway. These steps are linked perpendicularly by the " σ HB" pattern (blue), where a hydrogen bond forms between the C=O group of one step and the H–N group of an adjacent step, both lying in the same plane. The third pattern, " π HB" (green), involves hydrogen bonds between orthogonal C=O and H–N groups. This occurs when the H–N group of one step points towards a C=O group of an adjacent stairway step involved in a double HB, locking two stairs together. Thus, the double HB and σ HB patterns contribute to forming stairways and two stairways bind together through π HBs. Note that within a double stairway, one has the σ HBs in one direction and the other in the opposite direction (blue arrows in the schematic representation, Figure S25A). All these patterns combined describe a repeating, three-dimensional network of "double stairways."

Figure S25: Experimental and optimized crystal structure of PhASCBz including a schematic representation (A) and structural HB patterns, being "double HB", " σ HB", and " π HB" (B). Values in parenthesis depict experimental bond lengths in Å. Optimized at a ZORA-BLYP-D3(BJ)/TZP level of theory.

Interaction decomposition

The contributions stemming from the hydrogen-bond interaction for each pattern can be isolated through the same fragmentation scheme used for PhASCPh (see Scheme S2). Dimers of PhASCBz were made for each of the three hydrogen-bond patterns and the results are tabulated in Table S8. The interaction energy (ΔE_{int}) summed over all fragmentations has a maximum difference of 1.4 kcal mol⁻¹ compared to the interaction energy of the complete dimer which validates this approach.

The hydrogen-bond component in the double HB and σ HB dimers is strong, accounting for more than half of the total interaction energy. In the π HB dimer, this interaction accounts for less than half of the total interaction energy. The origin of this difference lies in the nature of the hydrogen bonds. In the double HB and s HB dimers, strong electrostatic interactions (ΔV_{elstat}) are present between the negatively charged HB acceptor $(C=O^{δ-})$ and positively charged HB donor $(N-H^{δ+})$, alongside significant donor-acceptor interactions (ΔE_{o}), where the lone pair on the C=O donates electrons to the σ^* orbital of the H–N. The hydrogen bond in the π HB dimer deviates because the involved lone pair is less favorable aligned with the σ* orbital of the H–N, resulting in less donor-acceptor interactions. The remaining interaction energy in the PhASCBz dimers arises from phenyl–benzoyl and mixed ASC–phenyl interactions, each making a relatively smaller contribution to the overall interaction.

[a] Computed at a ZORA-BLYP-D3(BJ)/TZ2P (ADF) // ZORA-BLYP-D3(BJ)/TZP (BAND) level of theory with values obtained using the experimental crystal in gray.

[b] The sum of the total interaction energy and decomposition terms of the substituent-only, ASC-only, and mixed fragmentations.

Cooperativity analysis

Since the stair-like chains are composed of steps connected by σ HB, the degree of cooperativity was assessed using two approaches. In the first approach, ASC-wise, the chain is elongated by adding ASC motives one at a time (Table S9), alternating between double and σ HBs. The second approach called "stepwise" forms the stairs using steps, *i.e.*, two ASC motives linked together by a double HB. The results show that both approaches indicate a lack of cooperativity, as the interaction energy (ΔE_{int}) and charge transfer ($\Delta Q_{\text{n}}^{\text{VDD}}$) do not significantly increase as the stairs become longer. Instead, an oscillating pattern emerges in the ASC-wise approach with interaction energies alternating between –14.3 and -28.3 kcal mol⁻¹, corresponding to the addition of a monomer via a σ HB or double HB. In addition, the stepwise approach shows a constant value of around -16.0 kcal mol⁻¹, and the charge transfer remains the same (+33 milli-electrons). The lack of cooperativity is due to charge not being able to be transferred in one direction; the opposing charge flow in double HBs cancels out, and the distance between adjacent σ HBs is too large to facilitate significant charge transfer.

Table S9: Cooperativity analysis of two hydrogen-bonded PhASCBz chains termed ASCwise and stepwise. The interaction energy, ΔE_{int} (in kcal mol⁻¹) and associated VDD charges (in milli-electrons) are tabulated. [a]

[a] computed at a ZORA-BLYP-D3(BJ)/TZ2P level of theory with values obtained with experimental structures in gray.

[b] The elongation direction is from top to bottom. Note that trends remain the same regardless of the elongation direction (except that the sign of charge transfer is inverted).

IR studies

The experimental IR spectrum of the PhASCBz crystal was measured, and the spectra of the optimized crystal and monomeric structures were calculated (see Figure S26). Highlighted in the spectra are the stretch vibrations of bonds involved in hydrogen bonding, including three N–H stretches between 3200 and 3500 cm–1 and three C=O stretches between 1600 and 1670 cm⁻¹. Comparing the monomeric spectrum (no hydrogen bonds) with the theoretical crystal spectrum (involving hydrogen bonds) reveals redshifts attributed to hydrogen bonding. Specifically, the N–H bond denoted with orange shows two distinct shifts: 133 cm⁻¹ for the σ HB with the yellow C=O bond and 201 cm⁻¹ for the π HB with the green C=O bond. The purple N–H bond, which participates in a bifurcated HB, exhibits a shift of 326 cm⁻¹. Meanwhile, the blue N-H bond, involved in an intramolecular hydrogen bond with the dark blue $C=O$ bond, shifts by 45 cm^{-1} . These redshifts correlate with elongations of the N–H bonds by 0.02 Å (purple) and 0.01 Å (orange) reflecting the bondweakening effects of hydrogen bonding. The blue N–H bond remains the same length up to two decimals, showing that the strength of the internal hydrogen bond changes less significantly upon complexation.

Figure S26: IR spectra comparison of PhASCBz between the experimental crystal (A) with computed spectra of the crystal (B) and monomer (C). Highlighted peaks are associated with stretching modes.

5.3 Theoretical methods

Computational details

Calculations were carried out using the periodic DFT module BAND13,14 and the DFT module ADF15-17 of the Amsterdam Modeling Suite 2023.104. The exchange-correlation function used is BLYP-D3(BJ), that is, a BLYP level of the generalized gradient approximation (GGA); exchange functional developed by Becke (B^{18}) , and the GGA correlation functional developed by Lee, Yang, and Parr (LYP¹⁹). The DFT-D3(BJ) method developed by Grimme and coworkers,²⁰ which contains the damping function proposed by Becke and Johnson,²¹ is used to describe non-local dispersion interactions. An uncontracted relativistically optimized Slater type orbitals (STOs) basis set was used containing diffuse function with either one set of diffuse functions (TZP, BAND calculations), or two sets of diffuse functions (TZ2P, ADF calculations). The smaller basis set for BAND calculations were chosen because of the unit cell's size (~260 atoms). However, the TZP performs sufficiently well for hydrogen bonds in large supramolecular systems.²² The 1*s* orbitals on carbon, nitrogen, and oxygen atoms were frozen within each basis set. Hydrogen-bonded systems have shown that the employed level of theory, *i.e.*, BLYP-D3(BJ)/TZ2P, gives a basis set superposition error (BSSE) of only a few tenths of a kcal mol $^{-1}$ and hence this will not affect the computed trends in hydrogen-bond strength.²³ Moreover, the employed level of theory has shown to give accurate results for describing hydrogen-bonded systems.²⁴ The zeroth-order regular approximation (ZORA²⁵) method was applied to account for relativistic effects. The molecular density was fitted by the systematically improvable Zlm fitting scheme; a numerical quality of "normal" was used for BAND calculations and "VeryGood" for ADF calculations.²⁶ The kspace grid for BAND calculations was set to "normal", that is, 3 k-space points for lattice vectors between 10 and 20 Bohr and 1 point for lattice vectors greater than 20 Bohr. The computational settings are summarized in Table S8.

Both the unit cell and molecular geometry were optimized with periodic DFT and verified through frequency analyses with zero imaginary frequencies for the equilibrium geometries.27, 28 Geometries were visualized using the CYLview2.0 software.²⁹ Most of the calculations were automated using the application programming interface PLAMS.³⁰

Energy decomposition analysis

To understand the different components that determine the hydrogen-bond strength, ∆*E* can be partitioned as formulated by eqn(S3) according to the activation strain model (ASM) of reactivity and bonding.³¹⁻³⁴ In this model, ΔE comprises two components, ΔE_{strain} and ΔE_{int}. Here, the strain energy (ΔE_{strain}) is the energy required to deform the equilibrium geometry of each fragment, to the geometry it acquires when it interacts in the hydrogen-bonded chain. The interaction energy (ΔE_{int}) accounts for the stabilizing interaction between the two deformed fragments.

$$
\Delta E = \Delta E_{\text{strain}} + \Delta E_{\text{int}} \tag{S3}
$$

In the framework of the Kohn-Sham molecular orbital model using quantitative canonical energy decomposition analysis (EDA),^{35, 36} the latter term, ΔE_{int} , can be further decomposed into electrostatic interaction (ΔV_{elstat}), Pauli repulsion (ΔE_{Pauli}), orbital interaction (ΔE_{o}), and an additional term that accounts for dispersion interactions ΔE_{disp} (eqn(S4)).

$$
\Delta E_{\text{int}} = \Delta V_{\text{elstat}} + \Delta E_{\text{Pauli}} + \Delta E_{\text{oi}} + \Delta E_{\text{disp}} \tag{S4}
$$

The term ΔV_{elstat} represents the usually attractive classical Coulomb interaction between the unperturbed charge distributions of the deformed fragments ΔE_{Pauli} comprises destabilizing interactions between occupied orbitals of the fragments and is responsible for steric repulsion. The orbital interaction energy $(\Delta E_{\text{o}i})$ accounts for charge transfer and polarization effects, including interactions between the highest occupied and lowest unoccupied MOs (HOMO–LUMO). Finally, ΔE_{disp} accounts for dispersion corrections as introduced by Grimme *et al*. 20

Voronoi deformation density charge analysis

For computing the charge transfer between molecular fragment that form a bond, the Voronoi deformation density (VDD) method was used.³⁷ In this method, Voronoi cells are considered for describing the spatial extent of an atom, which is defined as the compartment of space bounded by the bond midplanes on and perpendicular to all bond axes between one nucleus and its neighboring nuclei. Here, the charge rearrangement ΔQ^{VDD}_{A} compared to the initial density of the polyatomic fragments (ρ_i) is measured and gives insight into the change in electronic density upon interacting (eqn(S5)).

$$
\Delta Q_A^{VDD} = -\int_{Voronoi\ cell\ of\ A\ in\ molecule} \left[\rho_{molecule}(r) - \sum_{subsystems,i} \rho_i(r) \right] dr \tag{S5}
$$

Thus, Δ Q_A^{VDD} is an indication for the number of electrons that flows into (Δ Q_A^{VDD} < 0) or out of (ΔQ_A^{VDD} > 0) the Voronoi cell of a nucleus A as the result of the interaction between the two molecular fragments.

Theoretical IR spectra

Calculating the IR frequencies and normal modes with ADF yields peak heights (intensity) and positions (frequency in wavenumbers cm^{-1}) while BAND only positions; in this case, each vibration has an intensity of 1. The theoretical spectra presented in this work have been created by extracting the frequencies and peaks of ADF/BAND calculations and plotting them with a Lorentzian curve (width of 1) on a grid between 4000- and 400 cm^{-1} (18000 points with a spacing of 0.2). The resulting spectra were scaled to the maximum peak height found in the experimental section. The settings for the spacing and Lorentzian curve width were chosen to resemble the experimental spectrum as closely as possible.

The frequency of a vibration is given by the following formula which can be derived from the quantum harmonic oscillator model:

$$
\widetilde{w} = \frac{1}{2\pi\widetilde{c}} \sqrt{\frac{k}{u}}
$$
 (S6)

With \widetilde{w} being the wavenumber in cm⁻¹, \widetilde{c} the speed of light in cm s⁻¹, u the reduced mass, and k the force constant associated with a quantum harmonic oscillator. Upon forming hydrogen bonds comprised of C=O and H–N, the individual bonds are weakened, lowering their force constants and shifting the peak to the right on a spectrum plotted from high to low wavenumbers (redshift). Upon increasing the temperature, the reverse happens: hydrogen bonds are weakened, and the vibrations of the individual bonds shift to the left (blueshift).

6. Molecular characterization data

Figure S27: ¹H NMR spectrum of *tert*-butyl hydrazide in DMSO-*d*6.

Figure S28: ¹³C NMR spectrum of *tert*-butyl hydrazide in DMSO-*d*6.

Figure S29: ¹H NMR spectrum of MeASCPh in DMSO-*d*6.

Figure S30: ¹³C NMR spectrum of MeASCPh in DMSO-*d*6.

Figure S31: MALDI-ToF-MS spectra of MeASCPh (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S32: ¹H NMR spectrum of BuASCPh in DMSO-*d*6.

Figure S33*:* ¹³C NMR spectrum of BuASCPh in DMSO-*d*6.

Figure S34: MALDI-ToF-MS spectra of BuASCPh (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S35*:* ¹H NMR spectrum of BnASCPh in DMSO-d6.

Figure S36: ¹³C NMR spectrum of BnASCPh in DMSO-*d*6.

Figure S37: MALDI-ToF-MS spectra of BnASCPh (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S39: ¹³C NMR spectrum of PhASCPh in DMSO-*d*6.

Figure S40: MALDI-ToF-MS spectra of PhASCPh (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S41: ¹H NMR spectrum of *t-*BuASCPh in DMSO-*d*6.

Figure S42: ¹³C NMR spectrum of *t-*BuASCPh in DMSO-*d*6.

Figure S43: MALDI-ToF-MS spectra of PhASCPh (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S44: ¹H NMR spectrum of PhASCBu in DMSO-*d*6.

Figure S45: ¹³C NMR spectrum of PhASCBu in DMSO-*d*6.

Figure S46: MALDI-ToF-MS spectra of PhASCBu (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S48: ¹³C NMR spectrum of PhASCBn in DMSO-*d*6.

Figure S49: MALDI-ToF-MS spectra of PhASCBn (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S50: ¹H NMR spectrum of PhASCBz in DMSO-*d*6.

Figure S51: ¹³C NMR spectrum of PhASCBz in DMSO-*d*6.

Figure S52: MALDI-ToF-MS spectra of PhASCBz (black: CHCA, blue: DCTB maxtrix). Known matrix peaks indicated with x.

Figure S54: ¹³C NMR spectrum of methyl ester terminated pDMS in CDCl₃.

Total Average(RI)

Chromatogram Detector 2 Ch1
 $\frac{\# \qquad \text{Mn}}{1111114}$ 1938 1.1303 $\frac{1938}{1938}$ $\frac{1,13035}{1,13035}$ $\frac{1714}{1714}$

Figure S55: SEC chromatogram (RI detector) of methyl ester terminated pDMS.

Figure S56: ¹H NMR spectrum of acyl hydrazide terminated pDMS in CDCl₃.

Figure S57: ¹³C NMR spectrum of acyl hydrazide terminated pDMS in CDCl₃.

Figure S58: SEC chromatogram (left: UV detector, right: RI detector) of acyl hydrazide terminated pDMS. Note: There are interactions of the molecule with the column which gives rise to the tailing in the SEC trace.

Figure S59: ¹H NMR spectrum of isophthalamide in DMSO-*d*6.

Figure S60: ¹³C NMR spectrum of isophthalamide in DMSO-*d*6.

Figure S61: ¹H NMR spectrum of isophthaloyl diisocyanate in CDCl₃.

Figure S62: ¹³C NMR spectrum of isophthaloyl diisocyanate in CDCl₃.

Figure S63: ¹H NMR spectrum of ASC-HDI in THF-*d*8.

1,37509

Figure S68: SEC (left: UV detector, right: RI detector) chromatogram of ASC-BDI.

Figure S69: ¹H NMR spectrum of monofunctional methyl ester terminated pDMS in CDCl₃.

Figure S70: ¹³C NMR spectrum of monofunctional methyl ester terminated pDMS in $C\overline{D}Cl₃$.

Total Average(RI) 10
Chromatogram Detector 2 Ch1

Mn Mw $\frac{\text{Mw/Min}}{1,13858}$
1,13858 1462 $\overline{1}$ 1284 1284 1462

Figure S71: SEC chromatogram (RI detector) of monofunctional methyl ester terminated pDMS.

Figure S72: ¹H NMR spectrum of monofunctional acyl hydrazide terminated pDMS in CDCl₃.

Figure S73: ¹³C NMR spectrum of monofunctional acyl hydrazide terminated pDMS in CDCl₃.

Figure S74: SEC chromatogram (left: UV detector, right: RI detector) of monofunctional acyl hydrazide terminated pDMS. Note: There are interactions of the molecule with the column which gives rise to the tailing in the SEC trace.

Figure S75: ¹H NMR spectrum of H-monoASC in CDCl₃.

Figure S76: ¹³C NMR spectrum of H-monoASC in CDCl₃.

Figure S77: SEC chromatogram (left: UV detector, right: RI detector) of H-monoASC. Note: There are interactions of the molecule with the column which gives rise to the tailing in the SEC trace.

Figure S78: ¹H NMR spectrum of P-monoASC in CDCl₃.

Figure S79: ¹³C NMR spectrum of P-monoASC in CDCl₃.

Figure S81: ¹H NMR spectrum of B-monoASC in CDCl₃.

Figure S82: ¹³C NMR spectrum of B-monoASC in CDCl₃.

Figure S83: SEC chromatogram (left: UV detector, right: RI detector) of B-monoASC.

7. Cartesian Coordinates

Table S11: Cartesian coordinates (in Å) of the experimentally-measured crystal structures PhASCPh and PhASCBz.

PhASCPh (experimental)

Table S12. Cartesian coordinates (in Å) and bonding energies (in kcal mol⁻¹) for the stationary points of the optimized (crystal) structures PhASCPh and PhASCBz, computed with periodic DFT (BAND engine) at ZORA-BLYP-D3(BJ)/TZP.

PhASCPh (monomer)

 $E = -4683.5$ kcal mol⁻¹ *H* = –4519.8 kcal mol–¹ *G* = –4559.8 kcal mol–¹ $N_{image} = 0$

PhASCPh (theoretical)

 $E = -37752.1$ kcal mol⁻¹

 $N_{imag} = 0$

PhASCBz (monomer)

 \vec{E} = –5039.5 kcal mol⁻¹ *H* = –4868.8 kcal mol–¹ *G* = –4910.4 kcal mol–¹

 $N_{imag} = 0$

PhASCBz (theoretical)

 \vec{E} = –40579.0 kcal mol⁻¹

 $N_{imag} = 0$

8. References

1 A. M. M. Schreurs, X. Xian, L. M. J. Kroon-Batenburg. "*EVAL15*: a diffraction data integration method based on *ab initio* predicted profiles". *J. Appl. Cryst.,* **2010**, 43, 70-82.

2 L. Krause, R. Herbst-Irmer, G.M. Sheldrick, D. Stalke. "Comparison of silver and molybdenum microfocus X-ray sources for single-crystal structure determination". *J. Appl. Cryst.*, **2015**, 48, 3-10. **3** G. M. Sheldrick. "*SHELXT* - Integrated space-group and crystal-structure determination". *Acta Cryst.*, **2015**, A71, 3-8.

4 G. M. Sheldrick. "Crystal structure refinement with *SHELXL*". *Acta Cryst.*, **2015**, C71, 3-8.

5 A. L. Spek. "Structure validation in chemical crystallography". *Acta Cryst.*, **2009**, D65, 148-155. **6** V. A. Blatov, A. P. Shevchenko, D. M. Proserpio. "Applied Topological Analysis of Crystal Structures with the Program Package ToposPro". *Cryst. Growth Des.*, **2014**, 14, 3576-3586.

7 Spackman, P. R., Turner, M. J., McKinnon, J. J., Wolff, S. K., Grimwood, D. J., Jayatilaka, D. & Spackman, M. A. *J. Appl. Cryst.*, **2021**, 54, **3**, 1006–1011.

8 C. Nieuwland, S. Lekanne Deprez, C. de Vries, C. Fonseca Guerra, Chalcogen Atom Size Dictates Stability of Benzene-1,3,5-triamide Polymers: Overlooked Role of Geometrical Fit for Enhanced Hydrogen Bonding. *Chem. Eur. J.*, **2023**, 29 (34).

9 S. C. C. Van Der Lubbe, C. Fonseca Guerra, The Nature of Hydrogen Bonds: A Delineation of the Role of Different Energy Components on Hydrogen Bond Strengths and Lengths. *Chem. Asian J.*, **2019**, 14 (16), 2760-2769.

10 C. Nieuwland, C. Fonseca Guerra, Chalcogen Atom Size: A Key Parameter in Modulating Carbonyl Compound Properties. *Chem. Eur. J.*, **2024**, 30 (24).

11 L. De Azevedo Santos, D. Cesario, P. Vermeeren, S. C. C. Van Der Lubbe, F. Nunzi, C. Fonseca Guerra, σ‐Electrons Responsible for Cooperativity and Ring Equalization in Hydrogen‐Bonded Supramolecular Polymers. *ChemPlusChem*, **2022**, 87 (2).

12 C. Fonseca Guerra, H. Zijlstra, G. Paragi, F. M. Bickelhaupt, Telomere Structure and Stability: Covalency in Hydrogen Bonds, Not Resonance Assistance, Causes Cooperativity in Guanine Quartets. *Chem. Eur. J.*, **2011**, 17 (45), 12612-12622.

13 BAND 2023.1; SCM Theoretical Chemistry, Vrije Universiteit: Amsterdam, the Netherlands, **2023**. http://www.scm.com (accessed January 11, 2024).

14 G. Te Velde, E. J. Baerends, Precise density-functional method for periodic structures. *Phys. Rev. B*, **1991**, 44 (15), 7888-7903.

15 C. Fonseca Guerra, J. G. Snijders, G. te Velde, E. J. Baerends, Towards an order-N DFT method. *Theor. Chem. Acc.*, **1998**, 99 (6), 391-403.

16 G. te Velde, F. M. Bickelhaupt, E.J. Baerends, C. Fonseca Guerra, S. J. A. van Gisbergen, J. G. Snijders, T. Ziegler, Chemistry with ADF. *J. Comput. Chem.*, **2001**, 22 (9), 931-967.

17 ADF2023.104; SCM Theoretical Chemistry, Vrije Universiteit: Amsterdam, the Netherlands, **2023**. http://www.scm.com (accessed January 11, 2024).

18 A. D. Becke, Density-functional exchange-energy approximation with correct asymptotic behavior. *Phys. Rev. A*, **1988**, 38 (6), 3098-3100.

19 C. Lee, W. Yang, R. G. Parr, Development of the Colle-Salvetti correlation-energy formula into a functional of the electron density. *Phys. Rev. B*, **1988**, 37 (2), 785-789.

20 S. Grimme, J. Antony, S. Ehrlich, H. Krieg, A consistent and accurate ab initio parametrization of density functional dispersion correction (DFT-D) for the 94 elements H-Pu. *J. Chem. Phys.*, **2010**, 132 (15).

21 S. Grimme, S. Ehrlich, L. Goerigk, Effect of the damping function in dispersion corrected density functional theory. *J. Comput. Chem.*, **2011**, 32 (7), 1456-1465.

22 C. Nieuwland, F. Zaccaria, C. Fonseca Guerra, Understanding alkali metal cation affinities of multilayer guanine quadruplex DNA. *Phys. Chem. Chem. Phys*., **2020**, 22 (37), 21108-21118.

23 C. Fonseca Guerra, F. M. Bickelhaupt, J. G. Snijders, E. J. Baerends, Hydrogen Bonding in DNA Base Pairs:  Reconciliation of Theory and Experiment. *J. Am. Chem. Soc.*, **2000**, 122 (17), 4117- 4128.

24 P. Vermeeren, L. P. Wolters, G. Paragi, C. Fonseca Guerra, Cooperative Self‐Assembly in Linear Chains Based on Halogen Bonds. *ChemPlusChem*, **2021**, 86 (6), 812-819.

25 E. van Lenthe, E. J. Baerends, J. G. Snijders, Relativistic total energy using regular approximations. *J. Chem. Phys.*, **1994**, 101 (11), 9783-9792.

26 M. Franchini, P. H. T. Philipsen, E. van Lenthe, L. Visscher, Accurate Coulomb Potentials for Periodic and Molecular Systems through Density Fitting. *J. Chem. Theory Comput.*, **2014**, 10 (5), 1994-2004.

27 A. Bérces, R. M. Dickson, L. Fan, H. Jacobsen, D. Swerhone, T. Ziegler, An implementation of the coupled perturbed Kohn-Sham equations: perturbation due to nuclear displacements. *Comput. Phys. Commun.*, **1997**, 100, 247-262.

28 H. Jacobsen, A. Bérces, D. P. Swerhone, T. Ziegler, Analytic second derivatives of molecular energies: a density functional implementation. *Comput. Phys. Commun.*, **1997**, 100 (3), 263-276.

29 CYLview2.0; Université de Sherbrooke, 2020. http://www.cylview.org (accessed July 11, 2024).

30 PLAMS; SCM, Theoretical Chemistry, Vrije Universiteit: Amsterdam, the Netherlands, 2023. https://www.scm.com <https://github.com/SCM-NV/PLAMS> (accessed July 11, 2024).

31 W. -J. Van Zeist, F. M. Bickelhaupt, The activation strain model of chemical reactivity. *Org. Biomol. Chem.*, **2010**, 8 (14), 3118.

32 P. Vermeeren, P.; S. C. C. Van Der Lubbe, C. Fonseca Guerra, F. M. Bickelhaupt, T. A. Hamlin, Understanding chemical reactivity using the activation strain model. *Nat. Protoc.*, **2020**, 15 (2), 649- 667.

33 P. Vermeeren, P.; T. A. Hamlin, F. M. Bickelhaupt, Chemical reactivity from an activation strain perspective. *Chem. Commun.*, **2021**, 57 (48), 5880-5896.

34 F. M. Bickelhaupt, Understanding reactivity with Kohn–Sham molecular orbital theory: E2–SN2 mechanistic spectrum and other concepts. *J. Comput. Chem.*, **1999**, 20 (1), 114-128.

35 F. M. Bickelhaupt, E. J. Baerends, Kohn-Sham Density Functional Theory: Predicting and Understanding Chemistry. In Reviews in Computational Chemistry, John Wiley & Sons, Inc., **2007**; pp 1-86.

36 T. A. Hamlin, P. Vermeeren, C. Fonseca Guerra, F. M. Bickelhaupt, 8 Energy decomposition analysis in the context of quantitative molecular orbital theory. In Complementary Bonding Analysis, G. Simon, Ed.; De Gruyter, **2021**; pp 199-212.

37 C. Fonseca Guerra, J. W. Handgraaf, E. J. Baerends, F. M. Bickelhaupt, Voronoi deformation density (VDD) charges: Assessment of the Mulliken, Bader, Hirshfeld, Weinhold, and VDD methods for charge analysis. *J. Comput. Chem.*, **2004**, 25 (2), 189-210.