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¹School of Biomedical Sciences and Engineering

Supporting Information

for

Living Cascade Enyne Metathesis Polymerization of an Allylic Acetate Monomer and Post-Polymerization Modification via Tsuji-Trost Reaction

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General methods: All reagents are commercially available and used without further purification unless otherwise noted. All polymerization reactions were carried out under nitrogen atmosphere. Dry, degassed tetrahydrofuran (THF) and dichloromethane (DCM) were obtained from a Vigor solvent purification system. Greagent gel (100-200 mesh) was used for flash column chromatography. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance 400 MHz magnetic resonance spectrometer and calibrated using residual undeuterated solvent as an internal reference (CHCl₃ @ 7.26 ppm for ¹H NMR; 77.16 ppm for 13 C NMR; DMSO @ 2.50 ppm for 1 H NMR; 39.53 ppm for 13 C NMR; CH₂Cl₂ @ 5.30 ppm for ¹H NMR). The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. High Resolution Mass spectra were recorded on Thermo scientific Q Exactive Ultimate 3000 UPLC. Polymer samples were analyzed using a Waters 2414 Refractive Index Detector size exclusion chromatography (SEC) system with Waters (WAT045870) columns eluting THF (0.3 mL /min) and Tosoh EcoSEC HLC 8420GPC system with TSKgel SuperHM-N columns eluting CHCl₃ containing 0.25% Et₃N at a flow rate of 0.6 mL/min. All number-average molecular weights and polydispersities were calculated from refractive index chromatography using polyacrylate or polystyrene standards. Melting points were measured on RY-1G Digital Melting Point Apparatus. Thermogravimetric analysis (TGA) of the polymers was conducted using a METTLER TOLEDO Thermal Analysis System TGA2 under nitrogen atmosphere at a heating speed of 10 °C per minute from 30°C to 800 °C.

The monomer synthesis and experiment method.



2: A dry DCM (80 mL) solution of PMB (5.80 g, 42 mmol, 1.4 eq) and 1 (6.422 g, 30 mmol, 1.0 eq) was purged with nitrogen. Then a solution of the Pd₂(dba)₃ (686 mg, 0.75 mmol, 0.025 eq)

and PPh₃ (786 mg, 3 mmol, 0.1 eq) in dry DCM (4 mL) was added at 0 °C under nitrogen. The reaction was stirred at RT for 4 h before it was quenched with sat. aq. NaHCO₃. Then the reaction was extracted with DCM (3×20 mL), dried over Na₂SO₄. After evaporation, the residue was purified by column chromatography on silica gel (petroleum ether/EtOAc = 8:1) to afford the desired product as a yellowish oil (5.62 g, 80%).

HRMS (ESI) [M+H]⁺ calcd. for C₁₃H₁₄O₄, 233.0819, found, 233.0819.

¹**H NMR** (400 MHz, CDCl₃): δ 7.31–7.26 (m, 2H), 6.92–6.87 (m, 2H), 6.85 (dd, *J* = 10.3, 3.4 Hz, 1H), 6.11 (d, *J* = 10.3 Hz, 1H), 5.26 (dd, *J* = 3.4, 0.8 Hz, 1H), 4.77 (d, *J* = 11.4 Hz, 1H), 4.58 (d, *J* = 11.4 Hz, 1H), 4.46 (d, *J* = 16.9 Hz, 1H), 4.10 (d, *J* = 16.9 Hz, 1H), 3.79 (s, 3H).

¹³**C NMR** (100 MHz, CDCl₃): δ 194.69, 159.63, 144.54, 129.96, 128.94, 127.87, 114.02, 91.80, 70.40, 66.30, 55.30.



and the reaction mixture was stirred for 2 h at -78 °C. The resulting solution was quenched with saturated NaHCO₃ (20 mL), extracted (3x20 mL) with DCM, dried over Na₂SO₄, and concentrated under reduced pressure. After evaporation, the residue was purified by column chromatography on silica gel (petroleum ether/EtOAc = 2:1) to afford the desired product as a colorless oil (3.97 g, 70%).

HRMS (ESI) [M] calcd. for C₁₃H₁₆O₄, 235.0976, found, 235.0975.

¹**H NMR** (400 MHz, DMSO): δ 7.30–7.23 (m, 2H), 6.95–6.88 (m, 2H), 5.91 (dq, *J* = 10.3, 1.4 Hz, 1H), 5.67 (ddd, *J* = 10.3, 2.6, 2.0 Hz, 1H), 5.08 (d, *J* = 6.1 Hz, 1H), 4.97 (dt, *J* =

2.8, 1.4 Hz, 1H), 4.61 (d, *J* = 11.4 Hz, 1H), 4.42 (d, *J* = 11.4 Hz, 1H), 4.10 (dtq, *J* = 9.7, 6.0, 1.8 Hz, 1H), 3.75 (s, 3H), 3.69–3.61 (m, 1H), 3.52–3.45 (m, 1H). ¹³C NMR (100 MHz, DMSO): δ 159.20, 135.21, 130.54, 129.88, 126.18, 114.11, 93.03, 69.20, 62.76, 62.23, 55.52.

4: To a solution of 3 (1.89 g, 8 mmol, 1.0 eq), 2,4,6trimethyl-N-2-propyn-1-ylbenzenesulfonamide (1.90 g, 8 mmol, 1.0 eq) and PPh₃ (3.36 g, 12.8 mmol, 1.6 eq) in dry THF (100 mL, 0.08 M) was added DEAD (2.09 g, 1.84 mL,

12 mmol, 1.5 eq) at 0 °C. The mixture was then stirred for 24 h (TLC monitoring) while warmed to room temperature. After evaporation, the residue was purified by column chromatography on silica gel (petroleum ether/EtOAc = 6:1) to afford the desired product as a white solid (3.06 g, 84%).

Melting point: 72–74 °C.

HRMS (ESI) [M] calcd. for C₂₅H₂₉NO₅S, 456.1839, found, 456.1833.

¹**H NMR** (400 MHz, CDCl₃): δ 7.29–7.23 (m, 2H), 6.95 (s, 2H), 6.91–6.83 (m, 2H), 6.05 (ddd, *J* = 10.1, 3.2, 1.6 Hz, 1H), 5.89 (ddt, *J* = 10.1, 5.0, 1.3 Hz, 1H), 5.12–5.01 (m, 1H), 4.68 (d, *J* = 11.4 Hz, 1H), 4.48 (d, *J* = 11.4 Hz, 1H), 4.21 (dd, *J* = 12.7, 4.4 Hz, 1H), 4.18–4.10 (m, 1H), 4.07 (dd, *J* = 7.3, 1.9 Hz, 1H), 4.06–4.00 (m, 1H), 4.00–3.92 (m, 1H), 3.80 (s, 3H), 2.62 (s, 6H), 2.30 (s, 3H), 2.09 (t, *J* = 2.4 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃): δ 159.37, 142.95, 140.49, 132.47, 132.07, 131.20, 129.74, 129.66, 126.13, 113.90, 91.76, 80.17, 71.52, 69.40, 62.19, 55.30, 48.27, 33.23, 22.81, 21.00.



5: A solution of **4** (2.9 g, 6.4 mmol, 1.0 eq) in CH₃CN (60 mL) and H₂O (15 mL) was cooled to 0 °C. Ceric Ammonium Nitrate (8.77 g, 16 mmol, 2.5 eq) was added and the reaction mixture was stirred for 25 min at 0 °C. The resulting solution was quenched with saturated

NaHCO₃ (60 mL), extracted (3x20 mL) with EtOAc, dried over Na₂SO₄, and concentrated under reduced pressure. After evaporation, the residue was purified by column chromatography on silica gel (petroleum ether/EtOAc = 2:1) to afford the desired product as a mixture of diastereomers as a white solid (1.93 g, 90%).

Melting point: 116–118 °C.

¹**H NMR** (400 MHz, CDCl₃): δ 6.98 (s, 2H), 6.17–5.89 (m, 2H), 5.54–5.26 (m, 1H), 4.36–3.94 (m, 5H), 2.98–2.74 (m, 1H), 2.64 (s, 6H), 2.33 (s, 3H), 2.22–2.09 (m, 1H). **HRMS** (ESI) [M] calcd. for C₁₇H₂₁NO₄S, 334.1119, found, 334.1121.



M1: A solution of 5 (1.91 g, 5.7 mmol, 1.0 eq) and DMAP (0.033 g, 0.27 mmol, 0.05 eq) in DCM (90 mL) was cooled to -15 °C. Et₃N (1.16 g, 11.4 mmol, 2 eq) and Ac₂O (1.16 g, 11.4 mmol, 2 eq) was added and the reaction mixture was stirred for 4 h at -15 °C.

The resulting solution was quenched with H_2O (20 mL), extracted (3x20 mL) with DCM, dried over Na₂SO₄, and concentrated under reduced pressure. After evaporation, the residue was recrystallized twice by (petroleum ether/EtOAc = 4:1) to afford the product as a white solid (1.02 g, 50%).

Melting point: 118–120 °C.

HRMS (ESI) [M] calcd. for C₁₉H₂₃NO₅S, 378.1370, found, 378.1366.

¹**H NMR** (400 MHz, CDCl₃): δ 6.96 (s, 2H), 6.31 (d, *J* = 1.9 Hz, 1H), 6.08 (d, *J* = 2.3 Hz, 2H), 4.20–4.12 (m, 2H), 4.09 (d, *J* = 2.5 Hz, 1H), 4.08–4.06 (m, 1H), 4.03 (dd, *J* = 18.4, 2.5 Hz, 1H), 2.62 (s, 6H), 2.30 (s, 3H), 2.09 (t, *J* = 2.4 Hz, 1H), 2.07 (s, 3H).

¹³**C NMR** (100 MHz, CDCl₃): δ 169.90, 143.10, 140.50, 132.29, 132.12, 129.06, 127.45, 86.96, 79.70, 71.88, 64.10, 47.65, 33.39, 22.81, 21.10, 21.01.

General procedure for the polymerization of M1: Monomer M1 (25-75 eq) was added to a 10 mL vial equipped with a stir bar, placed under a nitrogen atmosphere, and was dissolved in degassed THF at RT. In a separate 3 mL vial, a solution of Grubbs 3rd generation (Grubbs 3) initiator (1 eq) in degassed THF was prepared. An appropriate volume of Grubbs 3 solution was rapidly added to the stirred reaction mixture containing monomer solution using a microliter syringe to make a total concentration of 0.08 M for the monomer. After the indicated time, the polymerization was quenched by addition of mixed solution of excess ethyl vinyl ether (0.1mL) and triethylamine (0.05 mL). The reaction mixture was stirred for 1 minute, and the reaction mixture was concentrated under vacuum. A small portion of the crude sample was analyzed by SEC and ¹H NMR. The remaining material was precipitated from DCM into 20-fold volume of diethyl ether. The precipitated polymer was then characterized using ¹H NMR.

P1: ¹**H NMR** (400 MHz, CD₂Cl₂): δ 7.02–7.00 (m, 2H), 6.56–6.14 (m, 1H), 6.14–5.74 (m, 2H), 5.61–5.39 (m, 1H), 4.91–4.65 (m, 1H), 4.48–4.22 (m, 1H), 4.17-3.87 (m, 1H), 3.74–3.67 (m, 1H), 3.50–3.15 (m, 1H), 2.72–2.58 (m, 6H), 2.41–2.27 (m, 3H), 2.15–1.94 (m, 3H).

Kinetic Studies on the polymerization of M1 targeting DP75: A solution of Grubbs 3 in degassed THF (2.4 mg/620 μ L) was prepared. Monomer M1 (30.2 mg, 0.08 mmol, 75 eq) was dissolved in 800 μ L THF. Then 200 μ L of Grubbs 3 was added. The reaction mixture was stirred at 0 °C and aliquots (100 μ l each) were taken and quenched by mixed solution of excess ethyl vinyl ether (50 μ L) and triethylamine (20 μ L) at 5 min, 10 min, 20 min, 30 min, 40 min, 60 min, 80 min and 100 min. Each aliquot was concentrated rapidly and analyzed by SEC and ¹H NMR.



Fig. S1 SEC traces for different time points of the P1

General procedure of the diblock copolymer P_{Bn} -*b*-P1 by sequential monomer addition: Monomer M_{Bn} and M1 were respectively added to two different 5 mL vials equipped with a stir bar, placed under a nitrogen atmosphere, and was dissolved in degassed THF at RT. In a separate 4 mL vial, a stock solution of G3 in degassed THF was prepared. An appropriate volume of G3 solution was rapidly added to the stirred solution that contained M_{Bn} using a microliter syringe to make a concentration of 0.08 M for the monomer. The reaction mixture was stirred before the M1 solution was rapidly added to the reaction solution using a microliter syringe to make a concentration of 0.08 M. Then the mixture was stirred at RT. After the indicated time, the polymerization was quenched by addition of mixed solution of excess ethyl vinyl ether (0.1mL) and triethylamine (0.05 mL). The reaction mixture was stirred for 1 minute, and the reaction mixture was concentrated under vacuum. The polymer was precipitated from DCM into 20-fold volume of diethyl ether. The precipitated polymer was then characterized using ¹H NMR and SEC.

General procedure of the copolymerization CP1: Monomer M1 and M_{Bn} (25, 50 eq respectively) were added to a 10 mL vial equipped with a stir bar, placed under a nitrogen atmosphere, and was dissolved in degassed THF at RT. In a separate 4 mL vial, a solution of Grubbs 3 in degassed THF was prepared. An appropriate volume of G3 solution was rapidly transferred to the stirred vial containing monomer solution using a microliter syringe to make a total monomer concentration of 0.1 M. Then the mixture was stirred at RT. After the indicated time, the polymerization was quenched by addition of mixed solution of excess ethyl vinyl ether (0.1mL) and triethylamine (0.05 mL). The reaction mixture was stirred for 1 minute, and the reaction mixture was concentrated under vacuum. The polymer was precipitated from DCM into 20-fold volume of diethyl ether. The precipitated polymer was then characterized using ¹H NMR and SEC.

General procedures of post-polymerization modifications: A dry DCM (1 mL) solution of **P1** (0.08 mmol monomer unit of **P1**, 30 mg, 1 eq), PPh₃ (0.04 mmol, 10.4mg, 0.5 eq), NaOAc (0.08 mmol, 6.5mg, 1 eq) and an appropriate nucleophile (such as PMB; 0.08 mmol, 3–5 eq) was added with nitrogen condition. Then a solution of the Pd₂(dba)₃ (0.012 mmol, 10.9mg, 0.15 eq) was added at 0 °C under nitrogen. The reaction was stirred at room temperature for 8 h, then the reaction was extracted with DCM/H₂O, The resulting DCM solution was subsequently filtered with a filter membrane of 0.22 μ m, The filtrate was slowly dropped into the diethyl ether solution, and subsequently centrifuged at 4000 r/min for 5min, the polymer solid obtained by centrifugation was dissolved in a small amount of DCM, then slowly dropped into n-hexane solution, centrifuged again at 4000 r/min for 5 min, The resulting polymer solid was dried in a vacuum drying chamber to obtain the target polymer. For the purification of **P2c**, precipitation used *n*-hexane instead, which was subsequently centrifuged at 4000 r/min for 5 min and dried in a vacuum drying chamber.

P2a: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 7.28–7.16 (m, 2H), 7.04–6.93 (m, 2H), 6.93–6.77 (m, 2H), 6.54–6.34 (m, 1H), 5.86 (d, *J* = 9.4 Hz, 1H), 5.47–5.36 (m, 1H), 5.03–4.76 (m, 2H), 4.54–4.42 (m, 1H), 4.41–4,24 (m, 2H), 4.05–3.91 (m, 1H), 3.80 (s, 3H), 3.66–3.53 (m, 1H), 3.42–3.16 (m, 1H), 2.69–2.58 (m, 6H), 2.33–2.23 (m, 3H).

P2b: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 7.02–6.86 (m, 2H), 6.50–6.3 (m, 1H), 5.81 (d, *J* = 11.0 Hz, 1H), 5.38–5.22 (m, 1H), 5.03–4.76 (m, 2H), 4.55–4.41 (m, 1H), 4.41–4.24 (m, 1H), 3.65–3.15 (m, 4H), 2.59 (m, 6H), 2.42–2.31 (m, 2H), 2.31–2.17 (m, 3H), 2.00–1.93 (m, 1H).

P2c: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 7.07–6.88 (m, 2H), 6.55–6.30 (m, 1H), 6.17–5.71 (m, 2H), 5.42-5.26 (m, 1H), 5.17–4.88 (m, 1.4H), 4.87–4.67 (m, 1.8H), 4.48–4.20 (m, 1H), 4.06–4.85 (m, 1H), 3.74–3.16 (m, 4H), 2.75–2.50 (m, 6H), 2.39–2.21 (m, 3H), 2.11–2.02 (m, 2.2H), 1.56–1.31 (m, 11H).

P2d: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 7.06–6.95 (m, 2H), 6.53–6.33 (m 1H), 6.25–5.72 (m, 3H), 5.45–5.25 (m, 1H), 5.00–4.62 (m, 2H), 4.50–4.24 (m, 1H), 4.07–3.84 (m, 1H), 3.78–2.76 (m, 6H), 2.70–2.52 (m, 6H), 2.38–2.21 (m, 3.6H), 1.85-1.74 (m, 0.6H), 1.56–1.42 (m, 1H), 1.34–1.20 (m, 2H), 1.19–1.03 (m, 0.5H), 0.56–0.39 (m, 0.6H).

P2e: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 8.15–7.95 (m, 0.7H), 7.81–7.61 (m, 1.3H),

7.53–7.42 (m, 0.7H), 7.08–6.88 (m, 2H), 6.59–6.25 (m, 1H), 6.32–5.38 (m, 3H),

5.09-4.74 (m, 3H), 4.45-4.16 (m, 1H), 4.13-3.85 (m, 1H), 3.69-3.34 (m, 2H),

2.74-2.53 (m, 6H), 2.39-2.16 (m, 3H), 2.08-2.01 (m, 1H).

P2f: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 7.85–7.49 (m, 1.4H), 7.43–7.21 (m, 1.4H), 7.09– 6.89 (m, 2H), 6.57–5.40 (m, 3H), 5.33–5.13 (m, 1H), 4.94–4.64 (m, 1H), 4.48–4.11 (m, 1H), 4.04–3.07 (m, 5H), 2.77–2.54 (m, 6H), 2.54–2.36 (m, 2H), 2.33 (7.36-7.23, 3H), 2.14–1.94 (m, 1.5H).

P2g: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 7.68–6.70 (m, 4.6H), 6.53–5.38 (m, 4H), 4.90–4.56 (m, 1H), 4.44–4.10 (m, 1H), 4.08–3.75 (m, 1H), 3.66–3.06 (m, 2H), 2.7–2.36 (m, 6H), 2.36–2.14 (m, 3H), 2.36–2.14 (m, 0.3H).

P2h: ¹**H NMR** (400 MHz, CD₂Cl₂) δ 8.00–7.84 (m, 2H), 7.68–7.56 (m, 1H), 7.55–7.41 (m, 2H), 7.01–6.82 (m, 2H), 6.70–5.49 (m, 4H), 4.81–4.67 (m, 1H), 4.54–4.23 (m, 1H), 4.10–3.85 (m, 1H), 3.72–3.27 (m, 2H), 2.79–2.44 (m, 6H), 2.35–2.09 (m, 3H), 2.08–2.00 (m, 0.3H).

Studies toward the degradation of representative polymers under different acidic conditions

Degradation studies of **P1** ($M_n = 15.8$ kDa) was examined under different acidic conditions in tetrahydrofuran using acetic acid or trifluoroacetic acid.

Procedure for degradation with acetic acid: In a 10 mL vial equipped with a stir bar, 15 mg of polymer **P1/P2a/P2f** (0.04 mmol monomer unit for each polymer) was dissolved in THF (6.5 mL). In a separate 10 mL vial, add acetic acid (55 μ L) to THF (6 mL), then dilute acetic acid solution (1 mL, 4 eq/monomer unit of **P1/P2a/P2f**) was rapidly added to the stirred mixture **P1/P2a/P2f** solution using a pipette to make a total concentration of 2 mg/mL for the **P1/P2a/P2f** polymer. The degradations were followed by size exclusion chromatography after concentration to monitor the decrease in molecular weight at 10 min, 20 min, 40 min, 1 hour, 2 hours, 4 hours, 12 hours, and 24 hours. Each aliquot was quenched with 0.05 mL triethylamine, concentrated under vacuum, and analyzed by SEC.

Procedure for degradation with trifluoroacetic acid: In a 10 mL vial equipped with a stir bar, 15 mg of polymer **P1** (0.04 mmol monomer unit for each polymer) was dissolved in THF (6.5mL). In a separate 10 mL vial, add trifluoroacetic acid (71 μ L) to TH F (6 mL), then dilute trifluoroacetic acid solution (1 mL, 4 eq/monomer unit of **P1**) was rapidly added

to the stirred mixture **P1** solution using a pipette to make a total concentration of 2mg/mL for the **P1** polymer. The degradations were followed by size exclusion chromatography after concentration to follow the decrease in molecular weight at 2min, 5min, 10 min, 20 min, 40 min, 80min, and 4 hours. Each aliquot was quenched with 0.05 mL triethylamine, concentrated under vacuum, and analyzed by SEC.



Fig. S2 (A) SEC trace of P1 polymer at different times under acetic acid conditions. (B) M_n of the P1 polymer at different times under acetate conditions. (C) SEC trace of P1 polymer at different times under trifluoroacetic acid. (D) M_n of polymer P1 at different times under trifluoroacetic acid.



Fig. S3a M_n of P2b, P2e, P2g, and P2h at different times under acetic acid conditions.



Fig. S3b SEC trace of P2f at different times under acetic acid conditions.



Fig. S3c SEC trace of P2b at different times under acetic acid conditions.



Fig. S3d SEC trace of P2e at different times under acetic acid conditions.



Fig. S3e SEC trace of P2g at different times under acetic acid conditions.



Fig. S3f SEC trace of P2h polymer at different times under acetic acid conditions.



Fig. S4 Comparison of ¹H NMR of polymer P1, P_{Bn}, CP1 and P_{Bn}-b-P1



Fig. S5 Stability testing for the P1. (A) P1 was stored under -20°C sealed conditions. (B) P1 was exposed to air at room temperature.



Fig. S6 TGA curves of the polymers before and after post-polymerization modification with different nucleophiles.



Fig. S7 SEC data of before and after post-polymerization modification of P1 ($M_n = 13.7 \text{ kDa}$; PDI = 1.12)



Fig. S8 Comparison of ¹H NMR and SEC of polymer **CP1** ($M_n = 15.9$ kDa, PDI=1.26): before and after PPM with 1-pyrenemethanol



Fig. S9 ¹H NMR spectrum of **P1** after 0.5 hour in deuterated chloroform and tentative assignment of resonance peaks for plausible products



Fig. S10 ¹H NMR spectrum of **P2a** after 1 hour in deuterated chloroform and tentative assignment of resonance peaks for plausible products



Fig. S11 Comparison of ¹H NMR spectra of P_{int4} (A) and P2a (B)

¹H NMR and ¹³C NMR spectra

¹H NMR and ¹³C NMR spectra of **2**



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

¹H NMR and ¹³C NMR spectra of **3**



¹H NMR and ¹³C NMR spectra of 4



¹H NMR spectrum of **5**



¹H NMR and ¹³C NMR spectra of **M1**



¹H NMR spectrum of polymer **P1**



¹H NMR spectrum of polymer **P2a**



¹H NMR spectrum of polymer **P2b**



¹H NMR spectrum of polymer **P2c**



¹H NMR spectrum of polymer **P2d**



¹H NMR spectrum of 5-norbornene-2-methanol



¹H NMR spectrum of polymer **P2e**



¹H NMR spectrum of polymer **P2f**



¹H NMR spectrum of polymer **P2g**



¹H NMR spectrum of polymer **P2h**

