1	SUPPLEMENTARY MATERIAL
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3	The potential of MOF accelerator in
4	electrochemiluminescence system for sensitivity detection
5	of menthol enantiomers
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16 3. References

1 1. Experiment section

2	1.1 Reagents and Apparatus. The reagents were obtained from commercial
3	sources and used directly without further purification. The reagents are
4	analytical grade. CdCl ₂ ·2.5H ₂ O, Na ₂ S·9H ₂ O, Zn(NO ₃) ₂ ·6H ₂ O, Triethylamine
5	(TEA), 2-Methylimidazole (Hmim), L-aspartic acid (L-Asp), L-/D-menthol (L-
6	/D-Men), L-/D-tyrosine (L-/D-Tyr), L-/D-arginine (L-/D-Arg), L-/D-tryptophan
7	(L-/D-Trp), L-/D-glutamic acid (L-/D-Glu), L-/D-proline (L-/D-Pro), L-/D-
8	threonine (L-/D-Thr), L-/D-histidine (L-/D-His), L-/D-serine (L-/D-Ser), L-/D-
9	methionine (L-/D-Met), R-/S-mandelic acid (R-/S-Man), L-/D-phenylalanine
10	(L-/D-Phe), L-/D-valine (L-/D-Val), L-/D-alanine (L-/D-Ala), L-/D-carnitine
11	(L-/D-Car), L-/D-penicillamine (L-/D-PA), R-/S-naproxen (R-/S-Nap),
12	potassium dihydrogen phosphate (KH2PO4), disodium hydrogen phosphate
13	(Na ₂ HPO ₄) and potassium chloride (KCl) were purchased from Sigma Aldrich
14	(Shanghai, China). Methanol and ethanol were obtained from Tianjin Fuyu Fine
15	Chemical Co, Ltd. All aqueous solutions were prepared with ultrapure water
16	(DW, 18.25 MΩ.cm).

17 Scanning electron microscopy (SEM) images were characterized by a FEI QUANTA FEG250 transmission electron microscope at 15.0 kV. Powder X-ray 18 patterns (PXRD) of samples were measured on a Bruker SMART APEX 19 charge coupled device (CCD)-based diffractometer using Cu K α radiation (λ = 20 1.5418 Å) at room temperature (298 K). Fourier transform infrared (FT-IR) 21 22 spectroscopy (KBr pellets) of materials was recorded on a Nicolet iS50 spectrometer. Electrochemical impedance analysis (EIS) was carried out in 0.1 23 M KCl aqueous solution containing $Fe(CN)_6^{3-/4-}$ as a redox marker with 24

scanning frequencies ranging from 1 to 10⁵ Hz. ECL measurements were
performed on an HYZ-3002 ECL analyzer obtained from Zhengzhou Shirui
Instrument Technology Co., Ltd (Xi'an, China), and the experimental
parameters were listed as follows: a photomultiplier voltage of 500 V, a scan
rate of 200 mV s⁻¹, and a potential ranging from -1.6 to 0 V (vs Ag/AgCl).

6 1.2 Characterisation of Circular dichroism spectra. To further validate the 7 adsorption of Men isomers by cyclodextrins, we continued to characterise the 8 experiments with Circular dichroism (CD). Two portions of β -CD (10 mM) were 9 taken and dissolved in water, followed by the addition of an equal amount of 10 aqueous L-/D-Men (10 mM) solution dissolved in water, respectively, and after 11 twenty minutes of adsorption, the β -CD was rotary distilled and washed with 12 EtOH to remove any remnants adsorption. After drying, equal amounts of the 13 above solid and β -CD (1 mg/mL) were dissolved in water and their CD signals 14 were measured.

1.3 The calculation of ECL efficiency. The calculation of ECL efficiency. The
ECL value refers to the ECL efficiency of Ru(bpy)₃Cl₂/K₂S₂O₈ using the
following formula¹⁻³:

 $\Phi_{ECL} = \frac{\left(\frac{\int ECL \, dt}{\int current \, dt}\right)^{x}}{\left(\frac{\int ECL \, dt}{\int current \, dt}\right)^{st}} \times 100\%$

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where "ECL intensity" and "current" represent the integration of ECL intensityand electrochemical current value from the cumulative ECL spectrum,

respectively, and "st" refers to the standard Ru(bpy)₃Cl₂/K₂S₂O₈. Specifically,
 Φ_{st} is the ECL efficiency of 1 mM [Ru(bpy)₃]²⁺ in 0.1 M K₂S₂O₈/PBS, which is
 be regarded as a standard, taken as 5.0%. According to calculations, the
 calculated ECL efficiency of the β-CD/L-CdS QDs/L-His-ZIF-8 system was
 34.7%.

1.4 The calculation of detection limit. An ECL measurement for blank samples
was implemented with ten parallel tests, which exhibited an average ECL
intensity (*I*_B) of 17416 with a standard deviation (*S*_B) of 28.2. k was a numerical
factor chosen according to the signal-to-noise ratio value and a value of 3 for it
in the equation was strongly recommended. Therefore, the smallest detectable
signal (*I*_L) could be calculated as follows:

$$I_{\rm L} = I_{\rm B} - \mathbf{k}^* S_{\rm B}$$

13 After that, by substituting $I_{\rm L}$ (17430.2) into the standard curve equation, the 14 lowest detection limit of both L-Men and D-Men can be calculated, which was 15 0.046 μ M.

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1 **2.** Figure and Table



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4 Figure S1. UV-vis absorption (blue) and FL emission spectrum (orange) of L-



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Figure S2. (A) Cyclic voltammograms and (B) Nyquist plots of (curve a) bare
GCE, (curve b) β-CD/GCE, (curve c) β-CD/L-CdS QDs/GCE and (curve d) βCD/L-CdS QDs/L-His-ZIF-8/GCE in 0.1 M KCl containing 5 mM
[Fe(CN)₆]^{4-/3-}.



2 Figure S3. Circular dichroism of β -CD (grey), β -CD + L-Men (blue) and β -CD





Figure S4. Influence of (A) the concentration of β-CD, (B) the concentration of
L-His-ZIF-8 and (C) pH on the ECL signals. Error bars are RSD (n = 3).





Figure S5. Comparison of enantioselectivity of enantiomers (0.1 mM) on the βCD/L-CdS QDs/L-His-ZIF-8/GCE + different amino acid enantiomers in PBS
(pH 7.4). Among them, the blue bar chart shows L-enantiomers, and the gray
bar chart shows D-enantiomers (except for Nap and Man, where the blue and
gray bar charts correspond to R and S enantiomers, respectively). Error bars are
RSD (n = 3).





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Figure S6. The linear relationship between ECL signals and L-Men%. Inset:
ECL of 0.1 mM Men mixture containing different content of L-Men in the
mixture (0, 30, 50, 70 and 100%, corresponding to curves a to e) on β-CD/L-CdS
QDs/L-His-ZIF-8/GCE. Error bars are RSD (n = 3).

1	Table S1. Comparison of the performance of the proposed and referenced ECL chiral
2	sensors for enantiomers detection.

Anaiyte	Methods	Linear range	LOD	Reference
Glu	ECL	0.005-5.0 mM	1.6 µM	4
Pro	ECL	0.001-1 mM	0.33 μΜ	5
РА	ECL	2-12 mM	0.3 mM	6
Pen	ECL	0.1-5.0 mM	33 µM	7
DAAO	ECL	0-10.0 mM	0.03 mM	8
Men	ECL	0.050-100 μM	0.046 µM	This work

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