

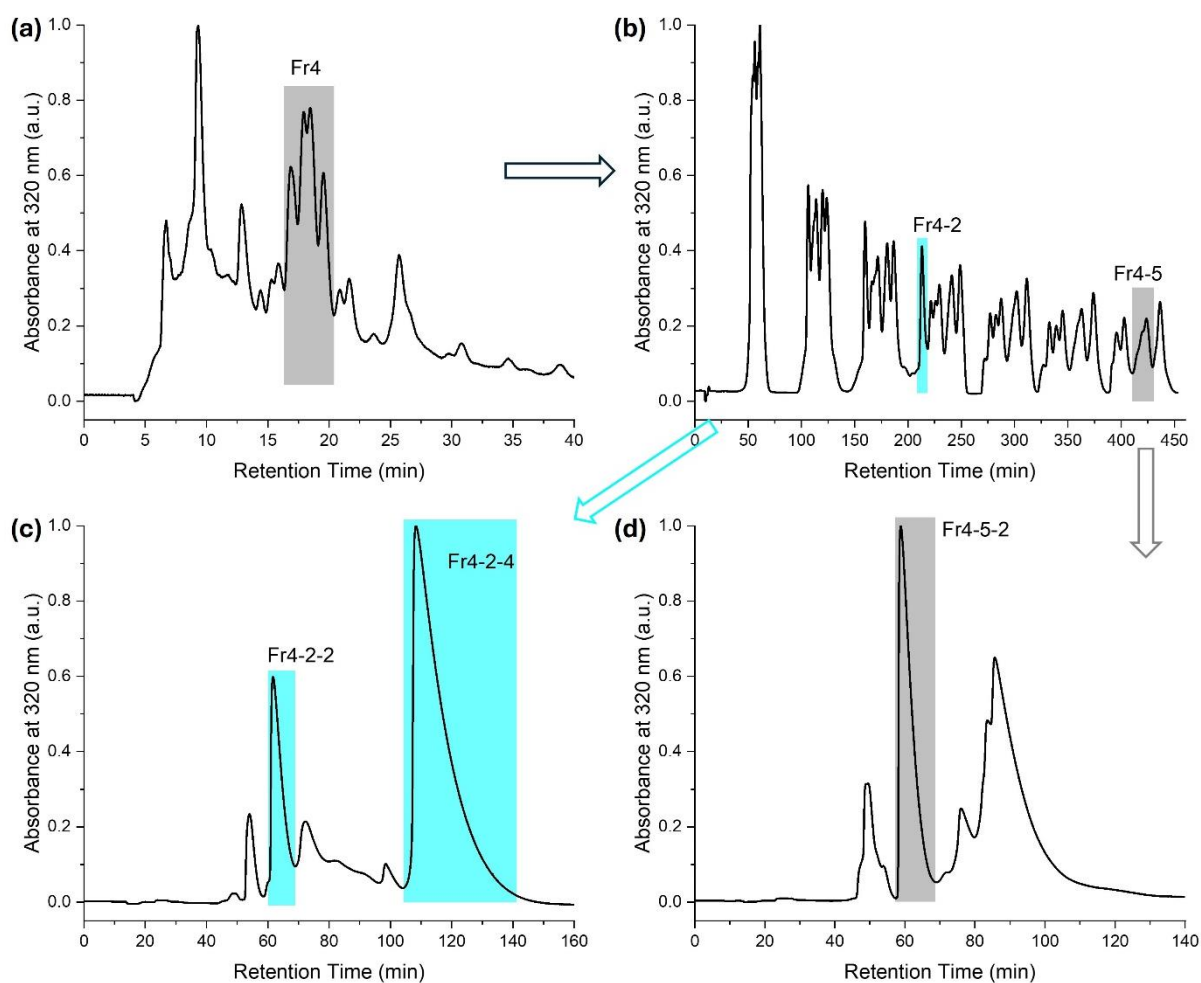
## **An Interplay between Metal-Fullerene and Metal-Metal Bonding in Molecular Magnetism of Erbium Metallofullerenes**

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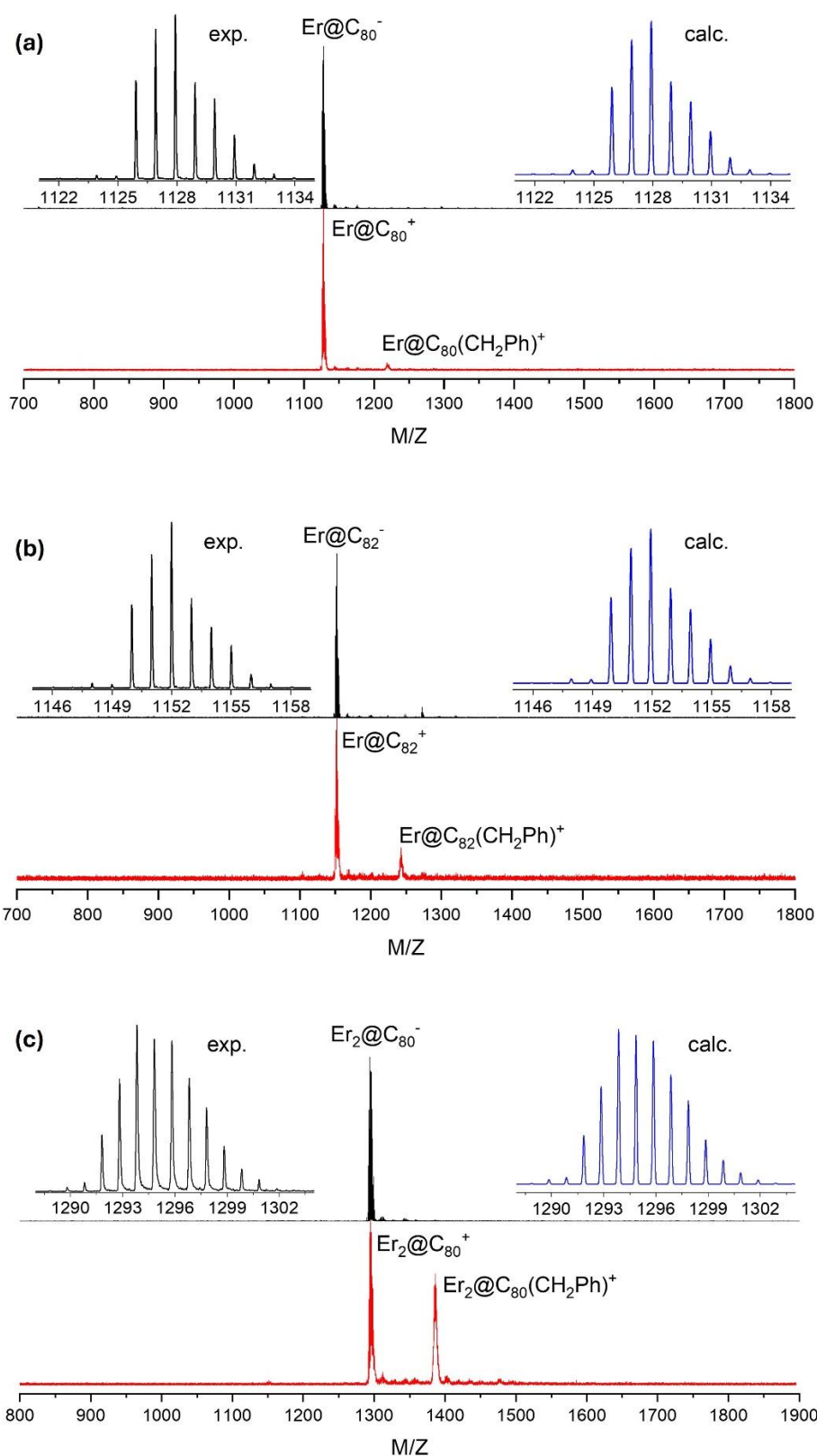
### **Supporting Information**

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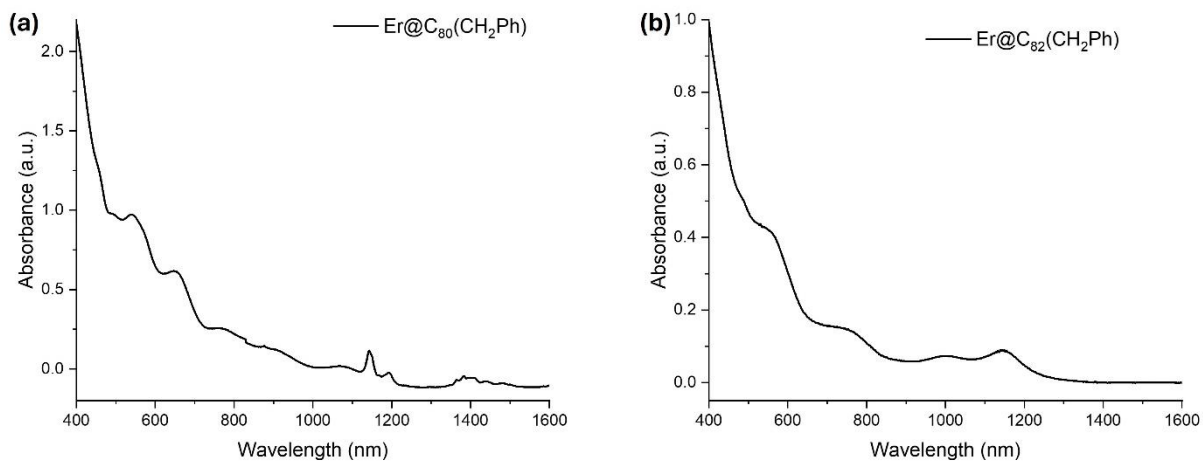
### HPLC separation of Er-EMFs



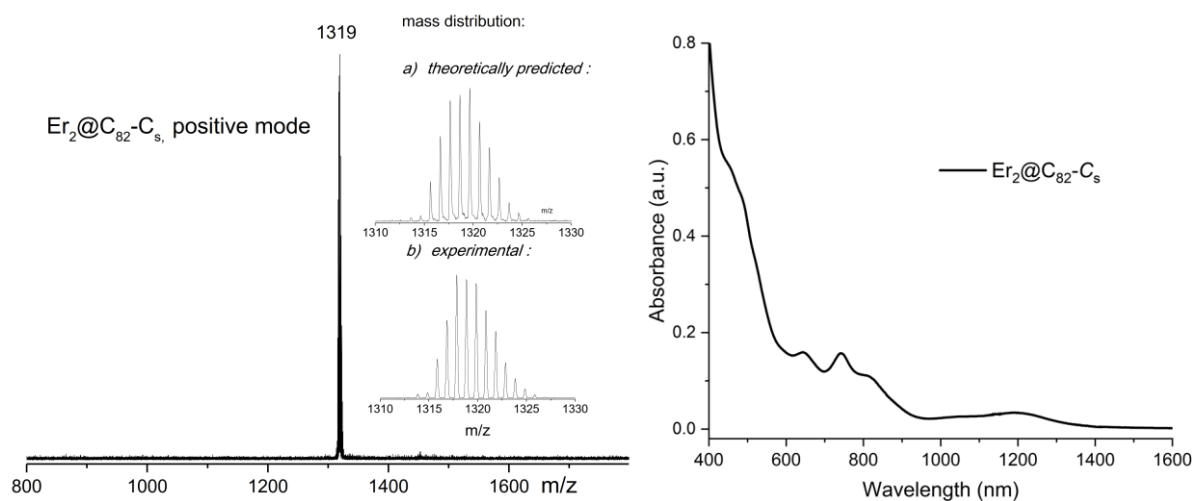
**Figure S1.** Separation of  $\text{Er}@C_{80}(\text{CH}_2\text{Ph})$ ,  $\text{Er}@C_{82}(\text{CH}_2\text{Ph})$ , and  $\text{Er}_2@C_{80}(\text{CH}_2\text{Ph})$ . Three steps HPLC separation were required to obtain the pure compounds. **(a)** HPLC profile of the mixture of benzyl-derivatized Er-EMFs. The highlighted fraction (Fr4) containing  $\text{Er}@C_{80}(\text{CH}_2\text{Ph})$ ,  $\text{Er}@C_{82}(\text{CH}_2\text{Ph})$ , and  $\text{Er}_2@C_{80}(\text{CH}_2\text{Ph})$  was collected for further separation. HPLC conditions: linear combination of two  $4.6 \times 250$  mm Buckyprep columns; flow rate 1.6 mL/min; injection volume  $800 \mu\text{L}$ ; toluene as eluent;  $40^\circ\text{C}$ . **(b)** Recycling HPLC separation of the Fr4. The highlighted fractions (Fr4-2 and Fr4-5) containing  $\text{Er}@C_{80}(\text{CH}_2\text{Ph})$ ,  $\text{Er}@C_{82}(\text{CH}_2\text{Ph})$ , and  $\text{Er}_2@C_{80}(\text{CH}_2\text{Ph})$  were collected for further separation ( $10 \times 250$  mm Buckyprep column; flow rate 1.5 mL/min; injection volume 4.5 mL; toluene as eluent). **(c)** HPLC separation of the Fr4-2. Pure  $\text{Er}@C_{80}(\text{CH}_2\text{Ph})$  and  $\text{Er}@C_{82}(\text{CH}_2\text{Ph})$  were obtained as Fr4-2-4 and Fr4-2-2, respectively. ( $10 \times 250$  mm Buckyprep-D column; flow rate 1.0 mL/min; injection volume 4.5 mL; toluene as eluent). **(d)** HPLC separation of the Fr4-5. Pure  $\text{Er}_2@C_{80}(\text{CH}_2\text{Ph})$  was obtained as Fr4-5-2. ( $10 \times 250$  mm Buckyprep-D column; flow rate 1.0 mL/min; injection volume 4.5 mL; toluene as eluent).



**Figure S2.** Matrix-assisted laser desorption/ionization time-of-flight (MALDI TOF) mass-spectra of **(a)** Er@C<sub>80</sub>(CH<sub>2</sub>Ph), **(b)** Er@C<sub>82</sub>(CH<sub>2</sub>Ph), and **(c)** Er<sub>2</sub>@C<sub>80</sub>(CH<sub>2</sub>Ph). Reflection negative (top) and linear positive (bottom) ionization modes, 1,1,4,4-tetraphenyl-1,3-butadiene was used as matrix. Resolution in positive mode is not high enough for analysis of isotopic distribution. In the negative ion mode, strong fragmentation does not allow for detection of molecular peak, but spectral resolution is sufficient to prove correct isotopic distribution of the Er-EMF<sup>-</sup> fragments.

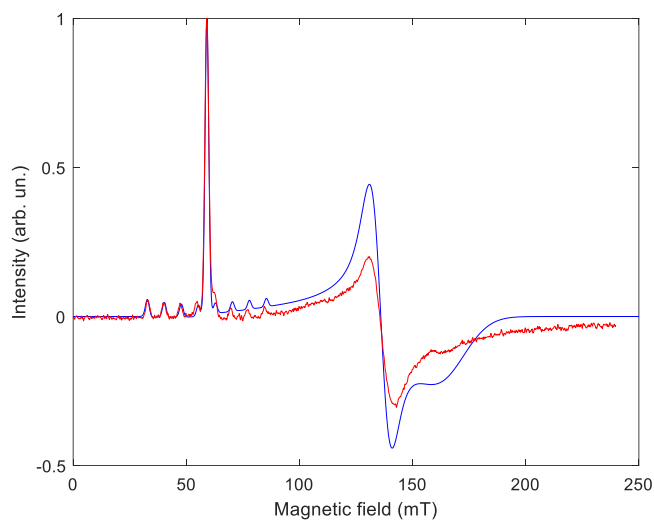


**Figure S3.** UV-vis-NIR absorption spectra of **(a)** Er@C<sub>80</sub>(CH<sub>2</sub>Ph) and **(b)** Er@C<sub>82</sub>(CH<sub>2</sub>Ph) measured in carbon disulfide.

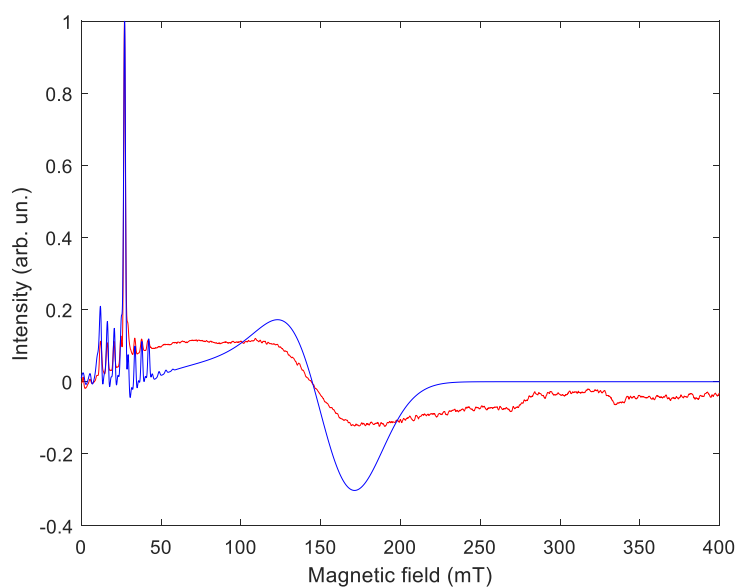


**Figure S4.** Mass-spectrum (left) and vis-NIR absorption spectrum (right) of Er<sub>2</sub>@C<sub>s</sub>(6)-C<sub>82</sub>.

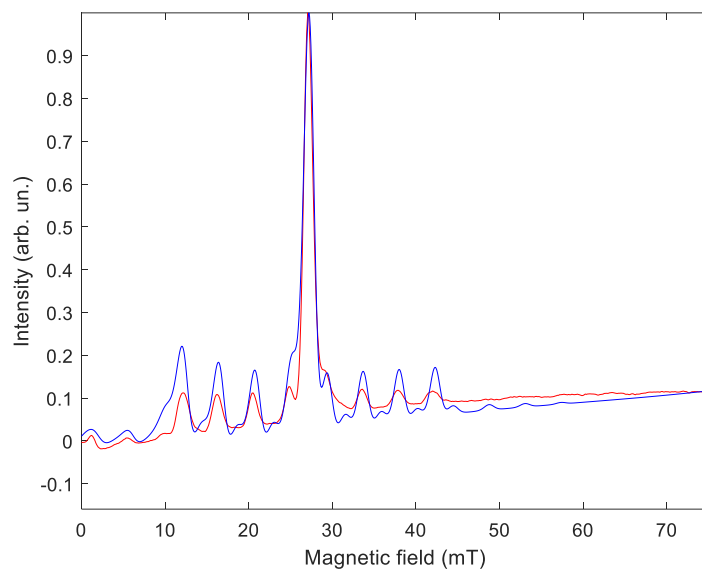
### Simulations of EPR spectra



**Figure S5.** Experimental (red) and simulated (blue) EPR spectra of Er@C<sub>80</sub>(CH<sub>2</sub>Ph);  $a_x$  and  $a_y$  are not considered since there is no observable hyperfine splitting in the range of  $g_x$ ,  $g_y$ . Parameters of simulated spectrum:  $g = (4.2, 5.05, 11.625)$ ;  $a_z = 1200$  MHz, HStrain = (1500, 600, 300)

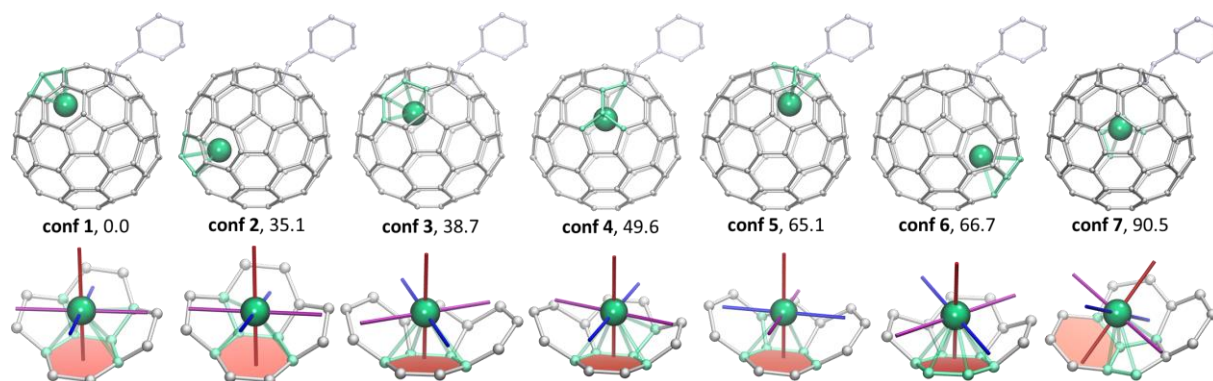


**Figure S6a.** Experimental (red) and simulated (blue) EPR spectra of Er<sub>2</sub>@C<sub>80</sub>(CH<sub>2</sub>Ph). Parameters of simulated spectrum:  $g = (4.3, 4.3, 25.00)$ ;  $a_z = 1500$  MHz, HStrain = (3000, 3000, 100).

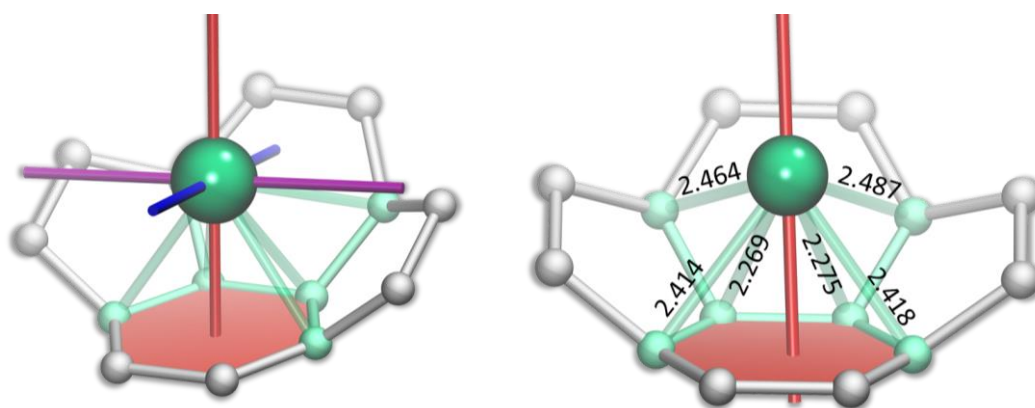


**Figure S6b.** Experimental (red) and simulated (blue) EPR spectra of  $\text{Er}_2@C_{80}(\text{CH}_2\text{Ph})$  in the low-field range.

## Computational studies of Er@C<sub>80</sub>(CH<sub>2</sub>Ph) conformers



**Figure S7a.** Seven conformers of Er@C<sub>80</sub>(CH<sub>2</sub>Ph) with their relative energies (kJ mol<sup>-1</sup>) and metal-cage coordination with orientation of principal axes for KD1 pseudospin g-tensor in each of them. Principal values of g-tensors are listed in Table S1.

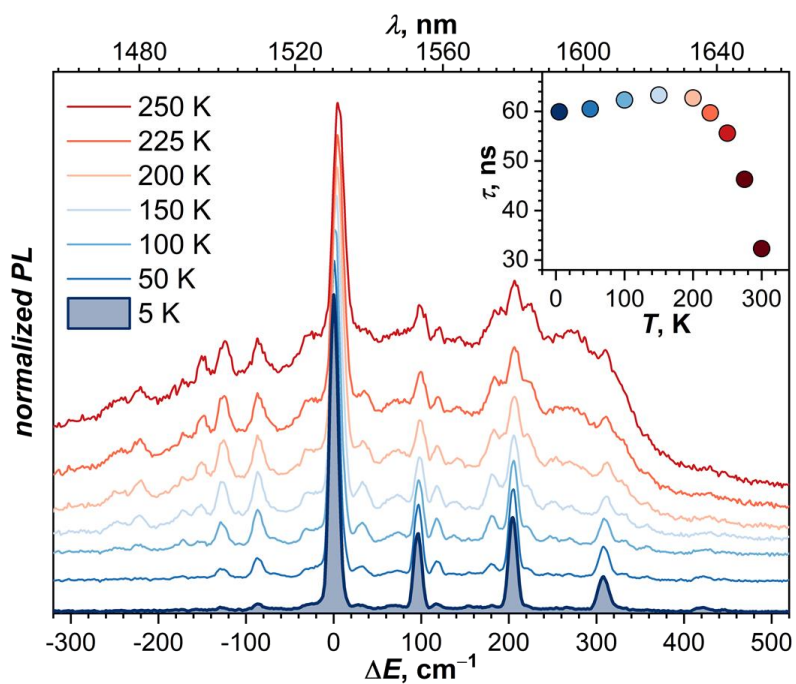


**Figure S7b.** Metal-cage coordination in **conf 1** with orientation of principal axes for KD1 pseudospin g-tensor (left, the same as Fig. 4c in the main text) and selected Er–C distances (Å) in DFT-optimized structure (right).

**Table S1.** Energies (cm<sup>-1</sup>) of LF-split Kramers doublets (KDs) and KD1 g-tensors in conformers of Er@C<sub>80</sub>(CH<sub>2</sub>Ph) computed with CASSCF/RASSI approach.

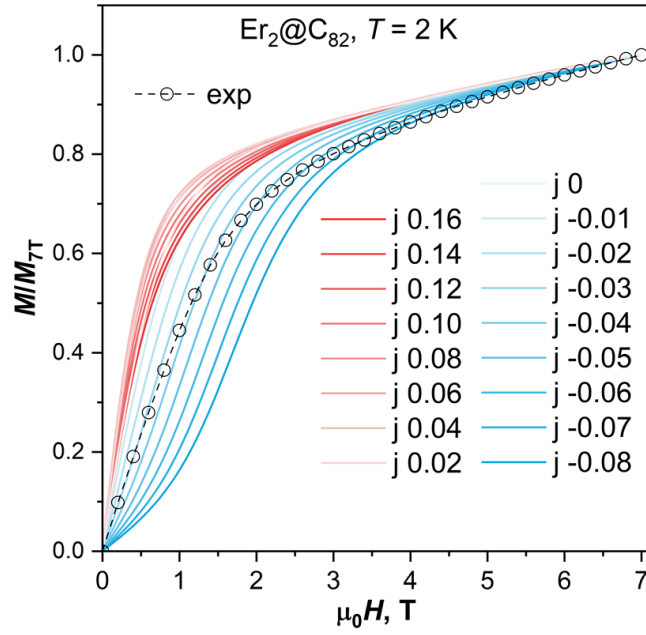
	conf 1	conf 2	conf 3	conf 4	conf 5	conf 6	conf 7
KD1	0 2.4, 6.8, 9.7	0 1.5, 2.9, 11.3	0 0.2, 6.7, 9.1	0 2.5, 4.1, 11.9	0 0.5, 4.6, 11.6	0 0.8, 6.4, 8.4	0 1.0, 2.6, 13.4
KD2	47	47	39	62	33	20	57
KD3	105	76	80	91	78	76	114
KD4	190	137	181	150	154	188	194
KD5	220	196	232	208	184	222	208
KD6	245	239	254	252	243	286	277
KD7	384	319	406	361	374	347	384
KD8	456	382	474	409	422	418	485

### Variable-temperature photoluminescence spectra of Er@C<sub>80</sub>(CH<sub>2</sub>Ph)



**Figure S8.** Photoluminescence spectra of Er@C<sub>80</sub>(CH<sub>2</sub>Ph) measured at different temperatures in the range of the  $^4I_{13/2} \rightarrow ^4I_{15/2}$  transitions under laser excitation at 488 nm. The inset shows temperature dependence of the PL decay time of Er@C<sub>80</sub>(CH<sub>2</sub>Ph). Each spectrum is normalized to the intensity of the most intensive peak at 1530 nm. Higher-temperature spectra show many features of comparable intensity, but the spectra become simpler at lower temperature.





**Figure S9.** Magnetization curve of powder  $\text{Er}_2@C_{82}$  measured by SQUID magnetometry at 2 K and compared to the curves calculated for different values of  $j_{\text{Er-Er}}$  (in  $\text{cm}^{-1}$ ). The Hamiltonian used in simulations is:  $\hat{H}_{\text{spin}} = \hat{H}_{\text{LF}'} + \hat{H}_{\text{LF}''} - 2j_{\text{Er-Er}}\hat{J}_{\text{Er}'}\hat{J}_{\text{Er}''} + \hat{H}_{\text{ZEE}}$ . For the sake of comparison, each curve is normalized to the magnetization at 7 T.