# **Supporting Information**

# TFPN-mediated racemization/epimerization-free amide and peptide bond formation

Jinhua Yang,<sup>a</sup>\* Dou Zhang,<sup>a</sup> Yajin Chang,<sup>a</sup> Bo Zhang,<sup>a</sup> Peng Shen,<sup>a</sup> Chunyu Han<sup>a</sup>\* and Junfeng Zhao<sup>b</sup>\*

<sup>a</sup>Hubei Key Laboratory of Pollutant Analysis & Reuse Technology, College of Chemistry and Chemical Engineering, Hubei Normal University, Huangshi 435002, P. R. China

<sup>b</sup>Affiliated Cancer Hospital, Guangdong Provincial Key Laboratory of Major Obstetric Diseases,

School of Pharmaceutical Sciences, Guangzhou Medical University, Guangzhou 511436, P. R. China

E-mail: jhyang@hunu.edu.cn; cyhan@hbnu.edu.cn; zhaojf@gzhmu.edu.cn

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# **General information**

Unless otherwise stated, all components as well as reagents and solvents were bought from commercial suppliers (Leyan, Energy Chemical) and used without further purification. TLC analysis was performed using commercially prepared silica gel plates, and visualization was affected at ultraviolet light (254 nm).  ${}^{1}H/{}^{13}C{H}$  NMR spectra were recorded on Bruker Avance 300 MHz and Bruker AMX 300 MHz spectrometer at 300/75 MHz, respectively, in CDCl<sub>3</sub> unless otherwise stated, using either TMS or the undeuterated solvent residual signal as the reference. CDCl<sub>3</sub> referenced at  $\delta$  7.26 and 77.00 ppm, DMSO- $d_6$  referenced at  $\delta$  2.50 and 39.8 ppm. Data for <sup>1</sup>H is reported as follows: chemical shift ( $\delta$  ppm), integration, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), broad peaks (br), coupling constant (Hz) and assignment. Data for <sup>13</sup>C{H} NMR are reported in terms of chemical shift ( $\delta$  ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant (Hz) and no special nomenclature is used for equivalent carbons. Flash column chromatography purification of compounds was carried out by gradient elution using ethyl acetate (EA) in light petroleum ether (PE). HRMS (ESI) spectra were obtained by the electrospray ionization time-of-flight (ESI-TOF) mass spectrometry. Analytical HPLC was performed on an UltiMate 3000 HPLC system with appropriate columns and elution conditions.

# Synthesis of acyl fluorides

In a round bottom flask, 4-methoxybenzoic acid (1a, 0.20 mmol), TFPN (2a, 0.24 mmol) was dissolved in DMF (1 mL), then DIPEA (0.24 mmol) was added to the solution and reaction mixture was stirred at room temperature until 4-methoxybenzoic acid was fully consumed. After the reaction was finished, water (10 mL) was added followed by EtOAc (3 x 10 mL). Combined organic layers were washed with brine (1 x 20 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford the 4-methoxybenzoyl fluoride (4a) in 93% yield.

### 3,4-dicyano-2,5,6-trifluorophenyl 4-methoxybenzoate (3a)



White solid, 92% yield,  $R_f = 0.3$  (PE/EA = 4:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.14 (d, J = 8.9 Hz, 2H), 7.03 (d, J = 8.9 Hz, 2H), 3.93 (s, 3H). <sup>13</sup>C{H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  165.4, 161.1, 154.0 (dt, J = 264.5, 3.4 Hz), 149.7 (ddd, J = 265.5, 14.0, 3.7 Hz), 148.4 (ddd, J = 266.9, 13.7, 4.9 Hz), 134.6 (tt, J = 16.0, 2.9 Hz), 133.4, 117.7, 114.5, 108.8 (dt, J = 10.3, 2.9 Hz), 102.7 (dt, J = 16.3, 2.6 Hz), 100.8 (ddd, J = 17.9, 4.6, 1.8 Hz), 55.8. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -113.15 – -117.83 (d, J = 8.8 Hz), -127.02 (dd, J = 21.0, 11.0 Hz), -131.00 (dd, J = 21.0, 8.4 Hz). (s, 1F). HRMS m/z (ESI) calcd for  $C_{16}H_8F_3N_2O_3^+$  [M+H]<sup>+</sup>: 333.0482, found: 333.0486.

### 4-methoxybenzoyl fluoride (4a)<sup>1</sup>

Colorless oil, 93% yield,  $R_f = 0.5$  (PE/EA = 10:1). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ 8.00 (d, *J* = 9.0 Hz, 2H), 7.15 (d, *J* = 9.0 Hz, 2H), 3.89 (s, 3H); <sup>13</sup>C{H} NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  165.5, 157.1 (d, *J* = 338.9 Hz), 134.0 (d, *J* = 4.1 Hz), 115.9 (d, *J* = 61.5 Hz), 115.2, 56.2. <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  16.0 (s, 1F).

## 4-ethylbenzoyl fluoride (4b)<sup>2</sup>



Colorless oil, 95% yield,  $R_f = 0.5$  (PE/EA = 10:1). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ 7.96 (d, *J* = 8.3 Hz, 2H), 7.48 (d, *J* = 7.9 Hz, 1H), 2.73 (q, *J* = 7.6 Hz, 2H), 1.21 (t, *J* = 7.6 Hz, 3H). <sup>13</sup>C{H} NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  157.3 (d, *J* = 342.2 Hz), 153.2, 131.72 (d, *J* = 4.0 Hz), 129.2, 121.7 (d, *J* = 60.6 Hz), 28.6, 15.2. <sup>19</sup>F NMR (376 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  18.0 (s, 1F). (a) <sup>1</sup>H NMR study of acyl fluoride formation and (b) Proposed mechanism for the deoxyfluorination intermediate formation





Acyl fluoride formation reaction monitored by <sup>1</sup>H NMR



(b) Proposed mechanism for the deoxyfluorination intermediate formation



# General procedure for the challenge amide formation



To a mixture of compound 1 (0.30 mmol) and TFPN (0.36 mmol) in DMF (1 mL) was added the DIPEA (0.36 mmol). The reaction solution was stirred at room temperature for 10 min. Then amine 5 (0.36 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluoride intermediate was fully consumed. After these reactions were finished, water (20 mL) was added followed by EtOAc (3 x 10 mL), and the layers were separated. Combined organic layers were washed with brine (1 x 20 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford 6 and 6-1.

### 4-(N,N-dipropylsulfamoyl)-N-phenylbenzamide (6a)<sup>3</sup>



White solid, 99% yield,  $R_f = 0.3$  (PE/EA = 4:1). <sup>1</sup>H NMR 8.67 (s, 1H), 7.89 (d, J = 8.1 Hz, 2H), 7.69 (t, J = 8.8 Hz, 4H), 7.35 (t, J = 7.8 Hz, 2H), 7.15 (t, J = 7.4 Hz, 1H), 3.05 (t, J = 7.7 Hz, 4H), 1.52 (h, J = 7.5 Hz, 4H), 0.85 (t, J = 7.4 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  165.0, 142.4, 138.9, 137.9, 129.0, 128.1, 127.1, 124.8, 120.4, 50.0, 21.9, 11.1.

N-(4-chlorophenyl)-4-(N,N-dipropylsulfamoyl)benzamide (6b)<sup>4</sup>



Yellow solid, 98% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.59 (s, 1H), 7.89 (d, J = 8.1 Hz, 2H), 7.69 (d, J = 8.1 Hz, 4H), 7.35 (d, J = 8.6 Hz, 2H), 7.27 (s, 1H), 3.22 – 2.86 (m, 4H), 1.53 (q, J = 7.5 Hz, 4H), 0.86 (t, J = 7.4 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  164.9, 142.5, 138.7, 136.5, 129.8, 129.1, 128.0, 127.2, 121.5, 50.0, 21.9, 11.1.

N-(4-bromophenyl)-4-(N,N-dipropylsulfamoyl)benzamide (6c)<sup>5</sup>



Yellow solid, 98% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.70 (s, 1H), 7.87 (d, J = 8.1 Hz, 2H), 7.64 (dd, J = 8.4, 2.0 Hz, 4H), 7.46 (d, J = 8.5 Hz, 2H), 3.05 (t, J = 7.6 Hz, 4H), 1.52 (h, J = 7.2 Hz, 4H), 0.85 (t, J = 7.4 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  164.9, 142.4, 138.7, 137.1, 132.0, 128.1, 127.1, 121.8, 117.4, 50.0, 21.9, 11.2.

N-(3-chlorophenyl)-4-(N,N-dipropylsulfamoyl)benzamide (6d)<sup>6</sup>



White solid, 90% yield,  $R_f = 0.25$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.83 (s, 1H), 7.86 (t, *J* = 8.1 Hz, 3H), 7.64 (d, *J* = 8.1 Hz, 3H), 7.28 (t, *J* = 8.0 Hz, 1H), 7.13

(d, *J* = 7.0 Hz, 1H), 3.06 (t, *J* = 7.6 Hz, 4H), 1.54 (h, *J* = 7.4 Hz, 4H), 0.87 (t, *J* = 7.4 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>) δ 165.1, 142.4, 139.2, 138.7, 134.6, 130.0, 128.1, 127.1, 124.7, 120.3, 118.3, 50.0, 22.0, 11.2.

### N-(3,5-bis(trifluoromethyl)phenyl)-4-(N,N-dipropylsulfamoyl)benzamide (6e)



White solid, 99% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  9.32 (s, 1H), 8.33 (s, 2H), 7.86 (d, J = 8.2 Hz, 2H), 7.65 (s, 1H), 7.59 (d, J = 8.3 Hz, 2H), 3.06 (d, J = 7.5 Hz, 5H), 1.55 (h, J = 7.4 Hz, 4H), 0.86 (t, J = 7.4 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  165.7, 142.2, 139.8, 138.6, 132.3 (q, J = 33.3 Hz), 128.3, 127.2, 123.2 (d, J = 272.8 Hz), 119.9 (d, J = 3.2 Hz), 117.8, 50.2, 22.0, 11.1. HRMS m/z (ESI) calcd for C<sub>21</sub>H<sub>23</sub>F<sub>6</sub>N<sub>2</sub>O<sub>3</sub>S<sup>+</sup> [M+H]<sup>+</sup>: 497.1328, found: 497.1317.

### 4-(*N*,*N*-dipropylsulfamoyl)-*N*-(o-tolyl)benzamide (6f)



White solid, 99% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.00 (d, J = 8.2 Hz, 2H), 7.95 – 7.83 (m, 3H), 7.79 (s, 1H), 7.34 – 7.23 (m, 2H), 7.18 (t, J = 7.3 Hz, 1H), 3.22 – 3.00 (m, 4H), 2.36 (s, 3H), 1.62 – 1.48 (m, 4H), 0.89 (t, J = 7.4 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  164.4, 143.2, 138.4, 135.2, 130.7, 129.8, 127.8, 127.5, 127.0, 126.0, 123.5, 50.0, 22.0, 17.9, 11.2. HRMS m/z (ESI) calcd for C<sub>20</sub>H<sub>27</sub>N<sub>2</sub>O<sub>3</sub>S<sup>+</sup> [M+H]<sup>+</sup>: 375.1737, found: 375.1729.

### 4-(*N*,*N*-dipropylsulfamoyl)-*N*-mesitylbenzamide (6g)



White solid, 90% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.99 (d, J = 8.1 Hz, 2H), 7.85 (d, J = 8.0 Hz, 2H), 7.62 (s, 1H), 6.94 (s, 2H), 3.20 – 3.01 (m, 4H), 2.30 (s, 3H), 2.23 (s, 6H), 1.55 (p, J = 7.4 Hz, 4H), 0.88 (t, J = 7.4 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  164.9, 142.9, 138.1, 137.5, 135.2, 130.8, 129.1, 128.0, 127.3, 50.0, 22.0, 21.0, 18.4, 11.2. HRMS m/z (ESI) calcd for C<sub>22</sub>H<sub>31</sub>N<sub>2</sub>O<sub>3</sub>S<sup>+</sup> [M+H]<sup>+</sup>: 403.2050, found: 403.2037.

N-(tert-butyl)-4-(N,N-dipropylsulfamoyl)benzamide (6h)<sup>7</sup>



White solid, 99% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.79 (d, J = 8.3 Hz, 2H), 7.71 (d, J = 7.8 Hz, 2H), 6.25 (s, 1H), 3.11 – 2.92 (m, 4H), 1.60 – 1.37 (m, 13H), 0.83 (t, J = 7.3 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  165.7, 142.1, 139.5, 127.6, 127.0, 52.0, 49.9, 28.7, 21.9, 11.1.

### N-benzyl-4-(N,N-dipropylsulfamoyl)-N-methylbenzamide (6i)



White solid, 99% yield,  $R_f = 0.4$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.92 – 7.71 (m, 2H), 7.63 – 7.50 (m, 2H), 7.47 – 7.22 (m, 4H), 7.19 – 7.06 (m, 1H), 4.61 (s, 2H), 3.17 – 2.76 (m, 7H), 1.69 – 1.44 (m, 4H), 1.00 – 0.75 (m, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  170.7, 170.0, 141.2, 140.0, 139.9, 136.5, 135.9, 130.5, 129.0, 128.8, 128.2, 127.9, 127.7, 127.6, 127.4, 127.3, 126.6, 55.0, 50.8, 50.1, 49.9, 36.9, 33.4, 22.1,

21.9, 11.2. HRMS m/z (ESI) calcd for  $C_{21}H_{29}N_2O_3S^+$   $[M\text{+}H]^+\!\!:$  389.1893, found: 389.1880

N-(tert-butyl)adamantane-1-carboxamide (6j) 8

White solid, 92% yield,  $R_f = 0.4$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  5.36 (s, 1H), 2.07 – 1.96 (m, 3H), 1.85 – 1.75 (m, 6H), 1.75 – 1.60 (m, 6H), 1.31 (s, 9H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  177.4, 50.5, 40.8, 39.3, 36.5, 28.8, 28.2.

## General procedure for the synthesis of peptides

$$1_{PG} \xrightarrow{H} \underbrace{O}_{\overline{R}} \xrightarrow{O}_{IOH} \underbrace{I}_{IOH} \xrightarrow{TFPN}_{DIPEA} \left[ 1_{PG} \xrightarrow{H} \underbrace{O}_{\overline{R}} \xrightarrow{I}_{\overline{R}} \right] \xrightarrow{H-L-AA-OPG^{2}(5)}_{IO \min - 1 \text{ h, r.t.}} 1_{PG} \xrightarrow{H} \underbrace{O}_{\overline{R}} \xrightarrow{I}_{O} \xrightarrow{I}_{O}$$

To a mixture of compound 1 (0.30 mmol) and TFPN (0.36 mmol) in DMF (1 mL) was added the DIPEA (0.36 mmol). The reaction solution was stirred at room temperature for 10 min. Then amine 5 (0.36 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluoride intermediate was fully consumed. After these reactions were finished, water (20 mL) was added followed by EtOAc (3 x 10 mL), and the layers were separated. Combined organic layers were washed with brine (1 x 20 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford 7.

## tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-alanyl-L-leucinate (7a) <sup>9</sup>



Yellow solid, 95% yield,  $R_f = 0.55$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.68 (d, J = 7.5 Hz, 2H), 7.51 (d, J = 7.4 Hz, 2H), 7.32 (t, J = 7.4 Hz, 2H), 7.23 (t, J = 7.4 Hz, 2H), 6.41 (d, J = 8.2 Hz, 1H), 5.49 (d, J = 7.8 Hz, 1H), 4.48 – 4.36 (m, 1H), 4.36 – 4.17 (m, 3H), 4.13 (t, J = 7.1 Hz, 1H), 1.62 – 1.48 (m, 2H), 1.47 – 1.25 (m, 13H), 0.83 (d, J = 5.7 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.9, 171.8, 155.9, 143.8, 141.3, 127.7, 127.1, 125.1, 120.0, 82.0, 67.1, 51.5, 50.4, 47.1, 41.7, 28.0, 24.9, 22.8, 22.1, 18.9.

### tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)glycyl-L-leucinate (7b) <sup>9</sup>



White solid, 97% yield,  $R_f = 0.55$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 7.5 Hz, 2H), 7.60 (d, J = 7.4 Hz, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.32 (d, J = 7.4 Hz, 2H), 6.73 (d, J = 8.3 Hz, 1H), 5.80 (t, J = 5.5 Hz, 1H), 4.56 (td, J = 8.5, 4.9 Hz, 1H), 4.40 (d, J = 7.1 Hz, 2H), 4.23 (t, J = 7.1 Hz, 1H), 3.96 (d, J = 5.5 Hz, 2H), 1.77 – 1.51 (m, 3H), 1.47 (s, 9H), 0.94 (dd, J = 6.3, 2.4 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.2, 168.8, 156.7, 143.8, 141.3, 127.7, 127.1, 125.1, 120.0, 82.1, 67.3, 51.4, 47.0, 44.4, 41.7, 28.0, 24.9, 22.8, 22.0.

(9H-fluoren-9-yl)methyl (S)-2-(((S)-1-(tert-butoxy)-4-methyl-1-oxopentan-2yl)carbamoyl)pyrrolidine-1-carboxylate (7c) <sup>8</sup>



White solid, 93% yield,  $R_f = 0.5$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d, J = 7.5 Hz, 2H), 7.51 (d, J = 8.0 Hz, 2H), 7.35 – 7.13 (m, 4H), 6.99 – 6.18 (m, 1H), 4.56 – 4.22 (m, 4H), 4.23 – 4.08 (m, 1H), 3.62 – 3.26 (m, 2H), 2.33 – 2.05 (m, 1H), 2.02 – 1.74 (m, 3H), 1.61 – 1.47 (m, 2H), 1.46 – 1.19 (m, 10H), 0.81 (d, J = 5.8 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.8, 171.2, 156.0, 143.8, 141.3, 127.7, 127.1, 125.1, 120.0, 81.7, 67.8, 61.1, 60.4, 51.6, 47.2, 47.0, 42.0, 41.6, 31.4, 28.3, 28.0, 25.0, 24.7, 23.6, 22.8, 22.2.

tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-valyl-L-leucinate (7d)<sup>8</sup>



White solid, 91% yield,  $R_f = 0.4$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (d, J = 7.5 Hz, 2H), 7.50 (d, J = 7.5 Hz, 2H), 7.29 (t, J = 7.5 Hz, 2H), 7.20 (t, J = 7.7 Hz, 2H), 6.45 (d, J = 8.2 Hz, 1H), 5.57 (d, J = 9.0 Hz, 1H), 4.48 – 4.29 (m, 2H), 4.23 (dd, J = 10.5, 7.0 Hz, 1H), 4.12 (t, J = 7.1 Hz, 1H), 4.06 – 3.96 (m, 1H), 2.10 – 1.93 (m,

1H), 1.65 – 1.45 (m, 2H), 1.45 – 1.23 (m, 10H), 0.95 – 0.69 (m, 12H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>) δ 171.9, 171.0, 156.4, 143.9, 143.8, 141.3, 127.7, 127.1, 125.2, 125.1, 120.0, 119.9, 81.9, 67.1, 60.2, 51.5, 47.1, 41.6, 31.5, 28.0, 24.9, 22.7, 22.1, 19.2, 18.0.

tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-isoleucyl-L-leucinate (7e)<sup>8</sup>



Yellow solid, 92% yield,  $R_f = 0.3$  (PE/EA = 3:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.65 (d, J = 7.5 Hz, 2H), 7.49 (d, J = 7.4 Hz, 2H), 7.28 (t, J = 7.5 Hz, 2H), 7.19 (t, J = 7.3 Hz, 2H), 6.47 (d, J = 8.1 Hz, 1H), 5.59 (d, J = 9.0 Hz, 1H), 4.48 – 4.28 (m, 2H), 4.27 – 4.18 (m, 1H), 4.16 – 4.06 (m, 1H), 4.03 (t, J = 8.1 Hz, 1H), 1.84 – 1.68 (m, 1H), 1.64 – 1.22 (m, 13H), 1.15 – 0.99 (m, 1H), 0.95 – 0.67 (m, 12H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.8, 171.0, 156.3, 143.9, 143.8, 141.3, 127.7, 127.1, 125.2, 125.1, 120.0, 119.9, 81.8, 67.1, 59.5, 51.5, 47.1, 41.6, 37.8, 28.0, 24.9, 22.7, 22.1, 15.4, 11.4.

tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-leucyl-L-leucinate (7f) 8



White solid, 99% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.75 (d, J = 7.4 Hz, 2H), 7.58 (d, J = 7.2 Hz, 2H), 7.39 (t, J = 7.3 Hz, 2H), 7.31 (t, J = 7.3 Hz, 2H), 6.46 (d, J = 7.8 Hz, 1H), 5.42 (d, J = 8.4 Hz, 1H), 4.53 – 4.43 (m, 1H), 4.44 – 4.29 (m, 2H), 4.29 – 4.13 (m, 2H), 1.77 – 1.49 (m, 6H), 1.45 (s, 9H), 0.94 (d, J = 4.5 Hz, 6H), 0.90 (d, J = 5.4 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.8, 171.8, 156.2, 143.9, 143.7, 141.3, 127.7, 127.1, 125.1, 120.0, 120.0, 81.9, 67.1, 53.4, 51.5, 47.1, 41.7, 28.0, 24.9, 24.6, 23.0, 22.7, 22.1.

tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-phenylalanyl-L-leucinate (7g)



White solid, 92% yield,  $R_f = 0.3$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (d, J = 7.5 Hz, 2H), 7.44 (t, J = 6.4 Hz, 2H), 7.30 (t, J = 7.4 Hz, 2H), 7.25 – 7.03 (m, 7H), 6.39 (s, 1H), 5.46 (s, 1H), 4.51 – 4.26 (m, 3H), 4.18 (t, J = 8.6 Hz, 1H), 4.12 – 4.02 (m, 1H), 2.99 (d, J = 6.5 Hz, 2H), 1.53 – 1.42 (m, 2H), 1.42 – 1.27 (m, 10H), 0.79 (t, J = 4.8 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.7, 170.5, 155.9, 143.8, 143.7, 141.3, 136.3, 129.5, 128.6, 127.7, 127.1, 127.0, 125.2, 125.1, 120.0, 82.0, 67.1, 55.9, 51.5, 47.1, 41.8, 38.6, 28.0, 24.8, 22.7, 22.1.

tert-butyl ((S)-2-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-(4-(tertbutoxy)phenyl)propanoyl)-L-leucinate (7h)



White solid, 91% yield,  $R_f = 0.75$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d, J = 7.5 Hz, 2H), 7.47 (dd, J = 7.4, 2.4 Hz, 2H), 7.31 (t, J = 7.4 Hz, 2H), 7.22 (t, J = 7.4 Hz, 2H), 7.00 (d, J = 8.0 Hz, 2H), 6.80 (d, J = 8.1 Hz, 2H), 6.26 (d, J = 8.0 Hz, 1H), 5.37 (d, J = 8.3 Hz, 1H), 4.46 – 4.28 (m, 3H), 4.24 (d, J = 7.4 Hz, 1H), 4.10 (t, J = 6.9 Hz, 1H), 2.96 (t, J = 6.6 Hz, 2H), 1.54 – 1.42 (m, 2H), 1.42 – 1.29 (m, 10H), 1.22 (s, 9H), 0.80 (dd, J = 6.0, 3.5 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.6, 170.4, 155.9, 154.4, 143.8, 143.7, 141.3, 131.0, 129.9, 127.7, 127.1, 125.1, 125.1, 124.2, 120.0, 82.0, 78.4, 67.1, 55.9, 51.5, 47.1, 41.7, 37.8, 28.8, 28.0, 24.8, 22.7, 22.1. HRMS m/z (ESI) calcd for C<sub>38</sub>H<sub>49</sub>N<sub>2</sub>O<sub>6</sub><sup>+</sup> [M+H]<sup>+</sup>: 629.3585, found: 629.3577.

tert-butyl 3-((S)-2-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-(((S)-1-(tertbutoxy)-4-methyl-1-oxopentan-2-yl)amino)-3-oxopropyl)-1H-indole-1carboxylate (7i) <sup>10</sup>



White solid, 93% yield,  $R_f = 0.75$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.04 (d, J = 8.1 Hz, 1H), 7.65 (d, J = 7.5 Hz, 2H), 7.57 (d, J = 7.6 Hz, 1H), 7.44 (t, J = 6.1 Hz, 3H), 7.28 (q, J = 5.7, 4.1 Hz, 2H), 7.24 – 7.09 (m, 4H), 6.25 (dd, J = 7.9, 4.3 Hz, 1H), 5.58 (d, J = 8.0 Hz, 1H), 4.50 (q, J = 7.2 Hz, 1H), 4.39 – 4.18 (m, 3H), 4.09 (t, J = 7.2 Hz, 1H), 3.28 – 2.97 (m, 2H), 1.54 (s, 9H), 1.48 – 1.38 (m, 2H), 1.39 – 1.24 (m, 10H), 0.77 (dd, J = 5.9, 2.7 Hz, 7H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.4, 170.5, 156.0, 149.5, 143.8, 143.7, 141.3, 135.5, 130.3, 127.7, 127.1, 125.2, 125.1, 124.7, 124.6, 122.8, 120.0, 119.1, 115.4, 115.3, 83.6, 81.9, 67.3, 54.7, 51.7, 47.1, 41.7, 28.4, 28.2, 28.0, 24.8, 22.7, 22.2.

tert-butyl 5-((S)-2-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-(((S)-1-(tertbutoxy)-4-methyl-1-oxopentan-2-yl)amino)-3-oxopropyl)-1H-imidazole-1carboxylate (7j) <sup>8</sup>



White solid, 96% yield,  $R_f = 0.4$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.96 (s, 1H), 7.68 (d, J = 7.5 Hz, 2H), 7.63 – 7.48 (m, 2H), 7.32 (t, J = 7.4 Hz, 2H), 7.23 (t, J = 7.5 Hz, 2H), 7.11 (d, J = 7.0 Hz, 2H), 6.62 (d, J = 7.5 Hz, 1H), 4.50 (d, J = 6.3 Hz, 1H), 4.40 – 4.22 (m, 3H), 4.19 (d, J = 7.4 Hz, 1H), 3.09 (dd, J = 14.8, 5.0 Hz, 1H), 2.89 (dd,

*J* = 14.9, 5.9 Hz, 1H), 1.51 (s, 9H), 1.44 – 1.22 (m, 12H), 0.77 (t, *J* = 6.0 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>) δ 171.6, 170.5, 156.2, 146.8, 143.9, 141.2, 139.1, 136.7, 127.7, 127.1, 125.2, 119.9, 114.9, 85.7, 81.7, 67.3, 54.7, 51.4, 47.1, 41.5, 30.3, 27.9, 27.8, 24.6, 22.8, 21.8.

tert-butyl N-(((9H-fluoren-9-yl)methoxy)carbonyl)-O-(tert-butyl)-L-seryl-Lleucinate (7k) <sup>11</sup>



White solid, 98% yield,  $R_f = 0.25$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.77 (d, J = 7.0 Hz, 2H), 7.69 – 7.53 (m, 2H), 7.40 (t, J = 6.8 Hz, 2H), 7.32 (t, J = 6.9 Hz, 2H), 7.28 – 7.19 (m, 1H), 5.79 (s, 1H), 4.60 – 4.33 (m, 3H), 4.25 (d, J = 5.9 Hz, 2H), 3.91 – 3.76 (m, 1H), 3.40 (t, J = 7.7 Hz, 1H), 1.70 – 1.57 (m, 2H), 1.54 (s, 1H), 1.46 (s, 9H), 1.22 (s, 9H), 0.95 (d, J = 4.8 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.6, 169.9, 156.0, 143.9, 143.8, 141.3, 127.7, 127.1, 125.1, 120.0, 81.7, 74.3, 67.1, 61.8, 54.3, 51.6, 47.1, 41.9, 28.0, 27.4, 24.9, 22.8, 22.1.

tert-butyl N-(((9H-fluoren-9-yl)methoxy)carbonyl)-O-(tert-butyl)-L-threonyl-Lleucinate (7l)



Yellow solid, 97% yield,  $R_f = 0.5$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d, J = 7.5 Hz, 2H), 7.53 (d, J = 7.5 Hz, 3H), 7.31 (t, J = 7.5 Hz, 2H), 7.23 (t, J = 7.4 Hz, 2H), 5.96 (d, J = 4.8 Hz, 1H), 4.49 – 4.23 (m, 3H), 4.22 – 3.98 (m, 3H), 1.66 – 1.50 (m, 2H), 1.51 – 1.32 (m, 10H), 1.22 (s, 9H), 1.05 (d, J = 6.2 Hz, 3H), 0.88 (t, J = 6.3 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.5, 169.2, 156.1, 144.0, 143.7, 141.3, 141.3, 127.7, 127.1, 125.2, 120.0, 120.0, 81.6, 75.6, 66.9, 66.8, 58.3, 51.9, 47.2, 41.4,  $S_{112}$ 

28.2, 28.0, 25.1, 22.8, 22.1, 16.5. HRMS m/z (ESI) calcd for  $C_{33}H_{47}N_2O_6^+$  [M+H]<sup>+</sup>: 567.3429, found: 567.3409.

tert-butyl N2-(((9H-fluoren-9-yl)methoxy)carbonyl)-N6-(tert-butoxycarbonyl)-Llysyl-L-leucinate (7m)<sup>12</sup>



White solid, 99% yield,  $R_f = 0.3$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.74 (d, J = 7.5 Hz, 2H), 7.58 (d, J = 7.5 Hz, 2H), 7.38 (t, J = 7.5 Hz, 2H), 7.30 (d, J = 7.5 Hz, 2H), 6.63 (d, J = 8.1 Hz, 1H), 5.68 (d, J = 8.3 Hz, 1H), 4.78 (d, J = 6.1 Hz, 1H), 4.53 – 4.41 (m, 1H), 4.36 (d, J = 7.2 Hz, 2H), 4.29 – 4.15 (m, 2H), 3.23 – 2.95 (m, 2H), 1.85 (p, J = 6.5 Hz, 1H), 1.74 – 1.57 (m, 3H), 1.43 (d, J = 6.3 Hz, 23H), 0.89 (d, J = 5.8 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.9, 171.5, 156.2, 143.9, 143.7, 141.3, 127.7, 127.1, 125.1, 120.0, 120.0, 81.9, 79.1, 67.1, 54.5, 51.5, 47.1, 41.4, 39.9, 32.4, 29.5, 28.4, 28.0, 24.9, 22.7, 22.3, 22.0.

tert-butyl N-(((9H-fluoren-9-yl)methoxy)carbonyl)-S-trityl-L-cysteinyl-Lleucinate (7n) <sup>13</sup>



Yellow solid, 96% yield,  $R_f = 0.5$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.08 (d, J = 7.7 Hz, 1H), 7.88 (d, J = 7.6 Hz, 2H), 7.78 – 7.60 (m, 3H), 7.50 – 7.10 (m, 19H), 4.42 – 4.10 (m, 4H), 4.05 (q, J = 7.7 Hz, 1H), 2.44 – 2.27 (m, 2H), 1.62 – 1.37 (m, 3H), 1.27 (s, 9H), 0.80 (dd, J = 17.4, 6.3 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.4,

169.6, 155.9, 144.4, 143.8, 143.7, 141.3, 129.6, 128.1, 127.8, 127.1, 126.9, 125.1, 120.0, 81.9, 67.3, 67.1, 54.0, 51.6, 47.1, 41.7, 33.9, 27.9, 24.9, 22.7, 22.2.

tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-methionyl-L-leucinate (70)



White solid, 97% yield,  $R_f = 0.4$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.75 (d, J = 7.5 Hz, 2H), 7.59 (d, J = 7.4 Hz, 2H), 7.39 (t, J = 7.4 Hz, 2H), 7.35 – 7.23 (m, 2H), 6.80 (d, J = 8.1 Hz, 1H), 5.87 (d, J = 8.3 Hz, 1H), 4.56 – 4.32 (m, 4H), 4.22 (d, J = 7.2 Hz, 1H), 2.61 (t, J = 7.3 Hz, 2H), 2.18 – 1.90 (m, 5H), 1.72 – 1.58 (m, 2H), 1.57 – 1.38 (m, 10H), 0.92 (d, J = 5.7 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.7, 170.9, 156.1, 143.8, 143.7, 141.3, 127.7, 127.1, 125.1, 125.1, 120.0, 120.0, 81.9, 67.1, 53.5, 51.6, 47.1, 41.5, 32.0, 29.9, 28.0, 24.9, 22.7, 22.1, 15.1. HRMS m/z (ESI) calcd for C<sub>30</sub>H<sub>41</sub>N<sub>2</sub>O<sub>5</sub>S<sup>+</sup> [M+H]<sup>+</sup>: 541.2731, found: 541.2715.

tert-butyl ((S)-2-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-5-(((Z)-amino ((2,2,4,6,7-pentamethyl-2,3-dihydrobenzofuran)-5-sulfonamido)methylene) amino)pentanoyl)-L-leucinate (7p)



Colorless oil, 80% yield,  $R_f = 0.4$  (PE/EA = 1:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>  $\delta$  7.72 (d, J = 7.5 Hz, 2H), 7.55 (d, J = 7.5 Hz, 2H), 7.35 (t, J = 7.3 Hz, 3H), 7.25 (d, J = 8.0 Hz, 3H), 6.38 (s, 2H), 6.09 (d, J = 8.2 Hz, 2H), 4.48 – 4.26 (m, 4H), 4.13 (t, J = 7.0 Hz, 1H), 3.24 (s, 2H), 2.89 (s, 2H), 2.59 (s, 3H), 2.51 (s, 3H), 2.06 (s, 3H), 2.00 – 1.89 (m,

1H), 1.80 - 1.68 (m, 1H), 1.68 - 1.49 (m, 5H), 1.41 (d, J = 2.7 Hz, 15H), 0.83 (d, J = 6.2 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.5, 172.1, 158.7, 156.4, 143.9, 143.7, 141.2, 141.2, 138.4, 132.9, 132.3, 127.7, 127.1, 125.2, 124.6, 119.9, 117.5, 86.4, 81.9, 67.1, 60.4, 53.9, 51.9, 47.0, 43.2, 40.5, 28.6, 28.0, 25.1, 24.8, 22.7, 21.8, 19.4, 18.0, 14.2, 12.5. HRMS m/z (ESI) calcd for C<sub>44</sub>H<sub>60</sub>N<sub>5</sub>O<sub>8</sub>S<sup>+</sup> [M+H]<sup>+</sup>: 818.4157, found: 818.4140.

tert-butyl ((S)-2-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-4-(tert-butoxy)-4oxobutanoyl)-L-leucinate (7q)



Colorless oil, 95% yield,  $R_f = 0.3$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 (d, J = 7.5 Hz, 2H), 7.50 (d, J = 7.4 Hz, 2H), 7.31 (t, J = 7.4 Hz, 2H), 7.22 (t, J = 7.4 Hz, 2H), 6.87 (d, J = 8.2 Hz, 1H), 5.97 (d, J = 8.4 Hz, 1H), 4.49 (q, J = 6.6 Hz, 1H), 4.44 – 4.26 (m, 3H), 4.14 (t, J = 7.1 Hz, 1H), 2.85 (dd, J = 17.3, 4.3 Hz, 1H), 2.54 (dd, J = 17.2, 6.8 Hz, 1H), 1.66 – 1.29 (m, 21H), 0.84 (d, J = 6.0 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.5, 171.4, 170.2, 156.0, 143.8, 143.7, 141.3, 127.8, 127.1, 125.1, 120.0, 81.9, 81.8, 67.3, 51.6, 51.0, 47.1, 41.6, 37.7, 28.0, 28.0, 24.8, 22.9, 22.0. HRMS m/z (ESI) calcd for C<sub>33</sub>H<sub>44</sub>N<sub>2</sub>NaO<sub>7</sub><sup>+</sup> [M+Na]<sup>+</sup>: 603.3041, found: 603.3027.

tert-butyl (S)-4-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-5-(((S)-1-(tertbutoxy)-4-methyl-1-oxopentan-2-yl)amino)-5-oxopentanoate (7r)



Yellow oil, 93% yield,  $R_f = 0.7$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 7.4 Hz, 2H), 7.59 (d, J = 7.3 Hz, 2H), 7.39 (t, J = 7.4 Hz, 2H), 7.30 (t, J = 7.4 Hz, 2H), 6.77 (d, J = 7.8 Hz, 1H), 5.79 (d, J = 7.6 Hz, 1H), 4.54 – 4.41 (m, 1H), 4.36 (d, J = 7.1 Hz, 2H), 4.32 – 4.25 (m, 1H), 4.21 (t, J = 7.1 Hz, 1H), 2.54 – 2.35 (m, 2H), 2.20 – 2.02 (m, 1H), 2.02 – 1.92 (m, 1H), 1.74 – 1.60 (m, 2H), 1.58 – 1.32 (m, 20H), 0.92 (d, J = 5.6 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  173.0, 171.6, 170.9, 156.1, 143.9, 143.7, 141.3, 141.3, 127.7, 127.1, 125.1, 120.0, 81.9, 81.0, 67.1, 53.9, 51.7, 47.1, 41.5, 31.6, 28.6, 28.1, 28.0, 24.9, 22.8, 22.0. HRMS m/z (ESI) calcd for C<sub>34</sub>H<sub>47</sub>N<sub>2</sub>O<sub>7</sub><sup>+</sup> [M+H]<sup>+</sup>: 595.3378, found:595.3362.

# tert-butyl N<sup>2</sup>-(((9H-fluoren-9-yl)methoxy)carbonyl)-N<sup>5</sup>-trityl-L-glutaminyl-L-leucinate (7s)



Yellow solid, 91% yield,  $R_f = 0.3$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (d, J = 7.5 Hz, 2H), 7.49 (d, J = 7.4 Hz, 2H), 7.29 (t, J = 7.5 Hz, 2H), 7.26 – 7.09 (m, 17H), 7.06 (s, 1H), 6.82 (d, J = 7.6 Hz, 1H), 5.79 (d, J = 7.2 Hz, 1H), 4.26 (d, J = 6.9 Hz, 3H), 4.18 – 4.07 (m, 1H), 4.08 – 3.96 (m, 1H), 2.44 (t, J = 6.5 Hz, 2H), 2.09 – 1.82 (m, 3H), 1.55 – 1.38 (m, 2H), 1.33 (s, 9H), 0.77 (d, J = 5.9 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.0, 171.9, 171.2, 156.1, 144.6, 143.9, 143.8, 141.3, 141.3, 128.7, 128.0, 127.7, 127.1, 127.0, 125.2, 120.0, 81.7, 70.7, 67.0, 53.5, 51.7, 47.2, 40.7, 33.4, 30.1, 28.0, 24.8, 22.8, 21.8. HRMS m/z (ESI) calcd for C<sub>49</sub>H<sub>54</sub>N<sub>3</sub>O<sub>6</sub><sup>+</sup> [M+H]<sup>+</sup>: 780.4007, found: 780.3986.

tert-butyl N<sup>2</sup>-(((9H-fluoren-9-yl)methoxy)carbonyl)-N<sup>4</sup>-trityl-L-asparaginyl-Lleucinate (7t)



White solid, 90% yield,  $R_f = 0.4$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.79 (d, J = 7.4 Hz, 2H), 7.60 (d, J = 7.4 Hz, 2H), 7.44 (d, J = 7.4 Hz, 2H), 7.38 – 7.10 (m, 19H), 7.03 (s, 1H), 6.49 (d, J = 7.4 Hz, 1H), 4.67 – 4.54 (m, 1H), 4.49 – 4.32 (m, 3H), 4.22 (t, J = 7.2 Hz, 1H), 3.10 (d, J = 15.2 Hz, 1H), 2.72 (dd, J = 15.6, 6.4 Hz, 1H), 1.71 – 1.54 (m, 2H), 1.53 – 1.36 (m, 10H), 0.91 (d, J = 4.9 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  170.9, 170.2, 155.8, 143.8, 143.3, 143.2, 140.8, 140.8, 128.2, 127.5, 127.2, 126.6, 124.7, 119.5, 81.1, 70.4, 66.8, 51.4, 50.6, 46.6, 40.5, 38.0, 27.5, 24.4, 22.2, 21.6. HRMS m/z (ESI) calcd for C<sub>48</sub>H<sub>52</sub>N<sub>3</sub>O<sub>6</sub><sup>+</sup> [M+H]<sup>+</sup>: 766.3851, found:766.3839.

tert-butyl ((benzyloxy)carbonyl)-L-alanyl-L-leucinate (7u)<sup>14</sup>

White solid, 95% yield,  $R_f = 0.2$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.43 – 7.27 (m, 5H), 6.70 (d, J = 6.1 Hz, 1H), 5.65 (d, J = 6.5 Hz, 1H), 5.08 (s, 2H), 4.52 – 4.40 (m, 1H), 4.39 – 4.21 (m, 1H), 1.68 – 1.53 (m, 2H), 1.50 – 1.40 (m, 10H), 1.36 (d, J = 6.9 Hz, 3H), 0.90 (d, J = 5.5 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.0, 171.8, 155.8, 136.2, 128.5, 128.1, 127.9, 81.8, 66.8, 51.4, 50.3, 41.5, 27.9, 24.8, 22.7, 22.0, 18.8.

tert-butyl (tert-butoxycarbonyl)-L-alanyl-L-leucinate (7v)<sup>15</sup>



White solid, 91% yield,  $R_f = 0.55$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.58 (d, J = 7.6 Hz, 1H), 5.15 (d, J = 7.2 Hz, 1H), 4.44 (q, J = 8.3 Hz, 1H), 4.17 (s, 1H), 1.69

- 1.50 (m, 3H), 1.42 (s, 9H), 1.41 (s, 9H), 1.32 (d, J = 7.0 Hz, 3H), 0.89 (d, J = 6.0 Hz, 6H).<sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>) δ 172.2, 171.9, 155.4, 81.7, 79.9, 51.3, 49.9, 41.7, 28.3, 27.9, 24.8, 22.8, 22.0, 18.3.

methyl N-((((9H-fluoren-9-yl)methoxy)carbonyl)-L-alanyl)-N-methyl-L-leucinate (7w)



White solid, 93% yield,  $R_f = 0.25$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 7.5 Hz, 2H), 7.60 (d, J = 7.5 Hz, 2H), 7.39 (t, J = 7.5 Hz, 2H), 7.31 (t, J = 7.4 Hz, 2H), 5.85 (d, J = 8.0 Hz, 1H), 5.39 – 5.24 (m, 1H), 4.72 (p, J = 7.1 Hz, 1H), 4.36 (d, J = 7.3 Hz, 2H), 4.21 (t, J = 7.3 Hz, 1H), 3.70 (s, 3H), 2.98 (s, 3H), 1.81 – 1.66 (m, 2H), 1.58 – 1.45 (m, 1H), 1.40 (d, J = 6.8 Hz, 3H), 0.94 (t, J = 6.8 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  173.4, 172.0, 155.6, 144.0, 143.8, 141.3, 127.7, 127.1, 125.2, 125.2, 120.0, 67.0, 54.7, 52.3, 47.3, 47.2, 36.9, 31.0, 24.9, 23.2, 21.4, 18.6. HRMS m/z (ESI) calcd for C<sub>26</sub>H<sub>33</sub>N<sub>2</sub>O<sub>5</sub><sup>+</sup> [M+H]<sup>+</sup>: 453.2384, found: 453.2371.

tert-butyl N-(((9H-fluoren-9-yl)methoxy)carbonyl)-N-methyl-L-alanyl-Lleucinate (7x)

Yellow oil, 94% yield,  $R_f = 0.65$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.77 (d, J = 7.5 Hz, 2H), 7.58 (d, J = 7.3 Hz, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.31 (t, J = 7.4 Hz, 2H), 6.38 (s, 1H), 4.81 (s, 1H), 4.56 – 4.39 (m, 3H), 4.26 (t, J = 6.8 Hz, 1H), 2.84 (s, 3H), 1.68 – 1.52 (m, 2H), 1.50 – 1.39 (m, 10H), 1.34 (d, J = 7.1 Hz, 3H), 0.90 (d, J = 5.4 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.8, 170.6, 157.0, 143.8, 141.3, 127.8,

127.1, 124.9, 120.0, 81.9, 67.9, 54.4, 51.3, 47.2, 41.6, 29.6, 28.0, 25.0, 22.8, 22.0, 13.7. HRMS m/z (ESI) calcd for C<sub>29</sub>H<sub>38</sub>N<sub>2</sub>NaO<sub>5</sub><sup>+</sup> [M+H]<sup>+</sup>: 517.2673, found:517.2665.

# methyl N-(N-(((9H-fluoren-9-yl)methoxy)carbonyl)-N-methyl-L-alanyl)-Nmethyl-L-leucinate (7y)



Yellow oil, 94% yield,  $R_f = 0.6$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.89 (d, J = 7.3 Hz, 2H), 7.63 (t, J = 6.8 Hz, 2H), 7.42 (t, J = 7.4 Hz, 2H), 7.38 – 7.29 (m, 2H), 5.09 – 4.48 (m, 3H), 4.47 – 4.09 (m, 2H), 3.61 (s, 3H), 2.87 – 2.54 (m, 3H), 2.51 – 2.34 (m, 2H), 1.87 – 1.51 (m, 2H), 1.35 – 1.21 (m, 1H), 1.11 (d, J = 6.7 Hz, 2H), 0.99 – 0.73 (m, 7H). <sup>13</sup>C {H} NMR (75 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  172.0, 171.8, 171.5, 170.5, 155.5, 144.2, 141.3, 128.1, 128.0, 127.5, 125.3, 125.2, 125.1, 120.6, 120.5, 66.9, 56.8, 55.0, 52.4, 51.5, 51.1, 47.4, 38.2, 37.1, 31.4, 29.7, 29.5, 29.1, 28.7, 24.9, 24.8, 23.6, 23.2, 21.8, 21.4, 14.9, 14.6. HRMS m/z (ESI) calcd for C<sub>27</sub>H<sub>35</sub>N<sub>2</sub>O<sub>5</sub><sup>+</sup> [M+H]<sup>+</sup>: 467.2540, found: 467.2532.

### methyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-alanyl-L-prolinate (7z)<sup>16</sup>



Colorless oil, 91% yield,  $R_f = 0.2$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.64 (d, J = 7.5 Hz, 2H), 7.49 (dd, J = 7.5, 2.9 Hz, 2H), 7.28 (t, J = 7.4 Hz, 2H), 7.19 (t, J = 7.5 Hz, 2H), 5.77 (d, J = 8.0 Hz, 1H), 4.54 – 4.37 (m, 2H), 4.31 – 4.16 (m, 2H), 4.10 (t, J = 7.2 Hz, 1H), 3.69 – 3.38 (m, 5H), 2.20 – 2.02 (m, 1H), 2.01 – 1.75 (m, 3H), 1.32 (d, J = 6.8 Hz, 3H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.3, 171.3, 155.7, 143.9, 143.8, 141.2, 127.7, 127.1, 125.2, 119.9, 67.0, 58.8, 52.3, 48.3, 47.1, 46.8, 28.9, 24.9, 18.3.



Colorless oil, 90% yield,  $R_f = 0.2$  (PE/EA = 1:1). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ 8.19 (d, *J* = 7.7 Hz, 1H), 7.89 (d, *J* = 7.5 Hz, 2H), 7.79 – 7.67 (m, 2H), 7.56 (d, *J* = 7.7 Hz, 1H), 7.42 (t, *J* = 7.4 Hz, 2H), 7.34 (d, *J* = 7.4 Hz, 2H), 5.09 (t, *J* = 5.5 Hz, 1H), 4.40 – 4.32 (m, 1H), 4.30 – 4.13 (m, 4H), 3.77 – 3.68 (m, 1H), 3.62 (s, 4H), 1.23 (d, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (75 {H} MHz, DMSO-*d*<sub>6</sub>)  $\delta$  172.9, 171.1, 155.7, 143.9, 143.8, 140.7, 127.7, 127.1, 125.3, 120.1, 65.6, 61.3, 54.6, 51.9, 49.7, 46.7, 18.3. HRMS m/z (ESI) calcd for C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>O<sub>6</sub><sup>+</sup> [M+H]<sup>+</sup>: 413.1707, found: 413.1717.

### tert-butyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-alanyl-L-glutaminate (7ab)<sup>8</sup>



White solid, 93% yield,  $R_f = 0.3$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.13 (d, J = 7.2 Hz, 1H), 7.82 (d, J = 7.4 Hz, 2H), 7.73 – 7.61 (m, 2H), 7.47 (d, J = 7.7 Hz, 1H), 7.35 (t, J = 7.3 Hz, 2H), 7.31 – 7.19 (m, 3H), 6.75 (s, 1H), 4.18 (d, J = 6.7 Hz, 3H), 4.10 – 3.93 (m, 2H), 2.10 (t, J = 7.5 Hz, 2H), 1.87 (dd, J = 14.9, 7.3 Hz, 1H), 1.73 (dt, J = 13.9, 8.0 Hz, 1H), 1.32 (s, 9H), 1.19 (d, J = 7.0 Hz, 3H). <sup>13</sup>C{H} NMR (75 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  173.8, 173.2, 171.4, 156.1, 144.4, 144.2, 141.2, 128.1, 127.5, 125.8, 120.5, 80.9, 66.1, 52.8, 50.1, 47.1, 31.6, 28.0, 27.1, 18.7.

### methyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-alanyl-L-tyrosinate (7ac)



White solid, 99% yield,  $R_f = 0.2$  (PE/EA = 1:1). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.93 (s, 1H), 8.45 – 8.23 (m, 1H), 7.88 (d, *J* = 7.5 Hz, 2H), 7.75 (t, *J* = 6.8 Hz, 2H), 7.63 – 7.48 (m, 2H), 7.50 – 7.26 (m, 5H), 7.30 – 7.16 (m, 1H), 7.09 (t, *J* = 7.5 Hz, 1H), 7.01 (t, *J* = 7.4 Hz, 1H), 4.59 (q, *J* = 7.1 Hz, 1H), 4.42 – 4.09 (m, 4H), 3.58 (s, 3H), 3.30 – 3.04 (m, 2H), 1.27 (d, *J* = 7.0 Hz, 3H). <sup>13</sup>C{H} NMR (75 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  173.0, 172.3, 156.3, 155.9, 144.2, 144.1, 141.0, 130.3, 127.9, 127.4, 127.3, 125.6, 120.4, 115.4, 65.9, 54.3, 52.1, 50.0, 47.0, 36.2, 18.5. HRMS m/z (ESI) calcd for C<sub>28</sub>H<sub>29</sub>N<sub>2</sub>O<sub>6</sub><sup>+</sup> [M+H]<sup>+</sup>: 489.2020, found: 489.2010.

### methyl (((9H-fluoren-9-yl)methoxy)carbonyl)-L-alanyl-L-tryptophanate (7ad)



White solid, 90% yield,  $R_f = 0.3$  (PE/EA = 2:1). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.93 (s, 1H), 8.46 – 8.22 (m, 1H), 7.88 (d, *J* = 7.5 Hz, 2H), 7.75 (t, *J* = 6.9 Hz, 2H), 7.54 (dd, *J* = 12.6, 7.9 Hz, 2H), 7.37 (dq, *J* = 15.7, 7.5 Hz, 5H), 7.23 (s, 0H), 7.09 (t, *J* = 7.5 Hz, 1H), 7.01 (t, *J* = 7.4 Hz, 1H), 4.59 (q, *J* = 7.1 Hz, 1H), 4.39 – 4.08 (m, 4H), 3.58 (s, 3H), 3.30 – 3.03 (m, 2H), 1.27 (d, *J* = 7.1 Hz, 3H). <sup>13</sup>C {H} NMR (75 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  173.2, 172.7, 156.1, 144.4, 144.2, 141.2, 136.5, 128.1, 127.5, 125.8, 124.2, 121.4, 120.5, 118.9, 118.4, 111.9, 109.7, 66.1, 53.6, 52.3, 50.2, 47.1, 27.5, 18.7. HRMS m/z (ESI) calcd for C<sub>30</sub>H<sub>30</sub>N<sub>3</sub>O<sub>5</sub><sup>+</sup> [M+H]<sup>+</sup>: 512.2180, found: 512.2173.

(9H-fluoren-9-yl)methyl ((S)-1-(((S)-1-amino-1-oxopropan-2-yl)amino)-1oxopropan-2-yl)carbamate (7ae)

White solid, 91% yield,  $R_f = 0.3$  (PE/EA = 4:1). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.08 (s, 1H), 7.90 (d, *J* = 6.2 Hz, 2H), 7.73 (s, 2H), 7.64 (s, 1H), 7.53 - 7.21 (m, 5H), 7.09 S27

(s, 1H), 4.39 – 4.11 (m, 4H), 4.05 (s, 1H), 1.31 – 1.10 (m, 6H); <sup>13</sup>C {H} NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 174.6, 172.6, 156.3, 144.2, 141.1, 128.1, 127.5, 125.7, 120.6, 66.1, 50.6, 48.4, 47.0, 18.7, 18.3. HRMS m/z (ESI) calcd for C<sub>21</sub>H<sub>23</sub>N<sub>3</sub>NaO<sub>4</sub><sup>+</sup> [M+H]<sup>+</sup>: 404.1581, found: 404.1574.

tert-butyl ((S)-2-((tert-butoxycarbonyl)amino)-3,3-dimethylbutanoyl)-L-leucinate (7af)



Yellow solid, 94% yield,  $R_f = 0.55$  (PE/EA = 5:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  6.01 (d, J = 8.3 Hz, 1H), 5.24 (d, J = 9.5 Hz, 1H), 4.56 – 4.40 (m, 1H), 3.81 (d, J = 9.5 Hz, 1H), 1.69 – 1.53 (m, 2H), 1.52 – 1.46 (m, 1H), 1.44 (s, 9H), 1.41 (s, 9H), 0.99 (s, 9H), 0.91 (d, J = 5.9 Hz, 6H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  171.7, 170.5, 155.8, 81.8, 79.6, 62.5, 51.4, 41.9, 34.5, 28.3, 27.9, 26.5, 24.8, 22.7, 22.1. HRMS m/z (ESI) calcd for C<sub>21</sub>H<sub>41</sub>N<sub>2</sub>O<sub>5</sub><sup>+</sup> [M+H]<sup>+</sup>: 401.3010, found:401.2999.

tert-butyl 2-(2-((((9H-fluoren-9-yl)methoxy)carbonyl)amino)-2methylpropanamido)-2-methylpropanoate (7ag)<sup>8</sup>

White solid, 91% yield,  $R_f = 0.4$  (PE/EA = 3:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 7.4 Hz, 2H), 7.62 (d, J = 7.3 Hz, 2H), 7.40 (t, J = 7.3 Hz, 2H), 7.32 (t, J = 7.4 Hz, 2H), 6.95 (s, 1H), 5.66 (s, 1H), 4.40 (d, J = 6.2 Hz, 2H), 4.22 (t, J = 6.7 Hz, 1H), 1.54 (s, 12H), 1.46 (s, 9H). <sup>13</sup>C{H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  173.8, 173.3, 155.0, 143.9, 141.3, 127.7, 127.1, 125.1, 120.0, 81.6, 66.5, 56.9, 56.8, 47.2, 27.8, 25.3, 24.2.

### Leuphasyl (L-Ala) (7ah)

A 10 mL round-bottomed flask was charged with **1ah** (Cbz-*L*-Tyr(O'Bu)-*L*-Ala-Gly-OH, 0.30 mmol), **TFPN** (0.36 mmol), DIPEA (0.36 mmol) and DMF (1 mL). The reaction solution was stirred at room temperature for 10 min. Then amine **5** (NH<sub>2</sub>-*L*-Phe-*L*-Leu-O'Bu, 0.36 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluoride intermediate was fully consumed. After these reactions were finished, water (20 mL) was added followed by EtOAc (3 x 10 mL), and the layers were separated. Combined organic layers were washed with brine (1 x 20 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford **7ah**.



White solid, 92% yield,  $R_f = 0.3$  (DCM/MeOH = 10:1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ 8.44 – 7.47 (m, 3H), 7.39 – 7.23 (m, 5H), 7.25 – 6.87 (m, 8H), 6.81 (d, *J* = 7.2 Hz, 2H), 6.64 (s, 1H), 5.31 (s, 1H), 5.13 (d, *J* = 12.3 Hz, 2H), 5.00 (d, *J* = 12.4 Hz, 1H), 4.83 (s, 1H), 4.52 (s, 2H), 3.90 (d, *J* = 16.6 Hz, 1H), 3.23 – 2.77 (m, 4H), 2.10 – 1.84 (m, 1H), 1.70 – 1.54 (m, 2H), 1.49 – 1.36 (m, 12H), 1.27 (s, 9H), 0.95 – 0.79 (m, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.8, 172.1, 171.4, 170.7, 168.5, 156.5, 154.2, 136.9, 136.6, 131.8, 130.0, 129.7, 128.4, 128.3, 128.0, 127.9, 126.7, 124.1, 81.8, 78.3, 66.8, 56.2, 53.8, 51.5, 48.6, 43.2, 42.0, 39.7, 28.9, 28.1, 25.0, 22.7, 22.6, 20.7. HRMS m/z (ESI) calcd for C<sub>45</sub>H<sub>62</sub>N<sub>5</sub>O<sub>9</sub><sup>+</sup> [M+H]<sup>+</sup>: 816.4542, found: 816.4536.

# The synthesis of protected Leu-enkephalin



### Boc-L-Phe-L-Leu-OMe (8) <sup>17</sup>:

A 10 mL round-bottomed flask was charged with **Boc-***L***-Phe-OH** (10 mmol, 2.65 g), **TFPN** (11 mmol), DIPEA (12 mmol) and DMF (15 mL). The reaction solution was stirred at room temperature for 10 min. Then amine **5** (NH<sub>2</sub>-*L*-Leu-OMe, 12 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluorides intermediates was fully consumed. After these reactions were finished, water (50 mL) was added followed by EtOAc (3 x 20 mL) and the layers were separated. Combined organic layers was washed with brine (1 x 30 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford **8** (93%, 3.65g).



<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.38 – 7.14 (m, 5H), 6.28 (s, 1H), 5.02 (s, 1H), 4.69 – 4.49 (m, 1H), 4.45 – 4.25 (m, 1H), 3.69 (s, 3H), 3.07 (d, *J* = 6.4 Hz, 2H), 1.64 – 1.45 (m, 3H), 1.42 (s, 9H), 0.90 (t, *J* = 5.0 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.8, 170.9, 155.4, 136.6, 129.4, 128.6, 126.9, 80.2, 55.6, 52.3, 50.7, 41.6, 38.0, 28.2, 24.6, 22.8, 21.9.

### Boc-Gly-L-Phe-L-Leu-OMe (9) 17:

**8** (7.2 mmol, 2.82 g) was dissolved in 10 mL of DCM and TFA (6 mL) was added dropwise at 0 °C. The mixture was allowed to warm to room temperature and stirred for 4 h. After the reaction was finished, the solvent was removed under reduced pressure. Then NaHCO<sub>3</sub> (30 mL, 2 M) was added and extracted with DCM (3 x 20 mL). The combined organic layers were washed with water (50 mL), brine (30 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was used for further reaction without purification.

A 10 mL round-bottomed flask was charged with **Boc-Gly-OH** (6.0 mmol, 1.05 g), **TFPN** (6.6 mmol), DIPEA (7.2 mmol), and DMF (15 mL). The reaction solution was stirred at room temperature for 10 min. Then the crude amine (6.6 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluoride intermediate was fully consumed. After these reactions were finished, water (40 mL) was added followed by EtOAc (3 x 20 mL), and the layers were separated. Combined organic layers were washed with brine (1 x 30 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford **9** (96%, 2.59 g).



<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.34 – 7.23 (m, 3H), 7.23 – 7.17 (m, 2H), 6.95 – 6.72 (m, 1H), 6.69 – 6.38 (m, 1H), 5.22 (s, 1H), 4.73 (d, *J* = 6.0 Hz, 1H), 4.62 – 4.44 (m, 1H), 3.77 (s, 2H), 3.69 (s, 3H), 3.22 – 2.91 (m, 2H), 1.66 – 1.47 (m, 4H), 1.43 (s, 9H), 0.88 (d, *J* = 4.2 Hz, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.8, 170.5, 169.5, 156.1, 136.4, 129.5, 128.7, 127.2, 80.5, 54.2, 52.4, 51.0, 44.5, 41.3, 38.2, 28.4, 24.8, 22.8, 22.0.

### Boc-Gly-Gly-L-Phe-L-Leu-OMe (10) 17:

**9** (3.6 mmol, 1.12 g) was dissolved in 5 mL of DCM and TFA (3 mL) was added dropwise at 0 °C. The mixture was allowed to warm to room temperature and stirred for 4 h. After the reaction was finished, the solvent was removed under reduced pressure. Then NaHCO<sub>3</sub> (20 mL, 2 M) was added and extracted with DCM (3 x 15 mL). The combined organic layers were washed with water (30 mL), brine (30 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was used for further reaction without purification.

A 10 mL round-bottomed flask was charged with **Boc-Gly-OH** (3.0 mmol, 0.53 g), **TFPN** (3.3 mmol), DIPEA (3.6 mmol), and DMF (15 mL). The reaction solution was stirred at room temperature for 10 min. Then the crude amine (3.6 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluoride intermediate was fully consumed. After these reactions were finished, water (30 mL) was added followed by EtOAc (3 x 20 mL), and the layers were separated. Combined organic layers were washed with brine (1 x 30 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford **10** (90%, 1.37 g).

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.01 – 7.54 (m, 3H), 7.30 – 7.06 (m, 5H), 5.90 (s, 1H), 5.04 (s, 1H), 4.59 (s, 1H), 4.18 – 3.84 (m, 4H), 3.71 (s, 3H), 3.24 – 3.05 (m, 1H), 2.96 (s, 1H), 1.71 – 1.56 (m, 3H), 1.45 (s, 9H), 0.90 (s, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  173.1, 171.4, 169.9, 168.6, 156.2, 136.5, 129.4, 128.3, 126.8, 79.8, 54.2, 52.2, 50.9, 43.7, 42.9, 40.9, 39.0, 28.4, 24.8, 22.7, 21.9.

### Fmoc-L-Tyr(O'Bu)-Gly-Gly-L-Phe-L-Leu-OMe (11):

**10** (0.48 mmol, 0.243 g) was dissolved in 1 mL of DCM, and TFA (1 mL) was added dropwise at 0 °C. The mixture was allowed to warm to room temperature and stirred for 4 h. After the reaction was finished, the solvent was removed under reduced pressure. Then NaHCO<sub>3</sub> (10 mL, 2 M) was added and extracted with DCM (3 x 10 mL). The combined organic layers were washed with water (10 mL), brine (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was used for further reaction without purification.

A 10 mL round-bottomed flask was charged with Fmoc-*L*-Tyr(O'Bu)-OH (0.4 mmol, 0.15 g), **TFPN** (0.44 mmol), DIPEA (0.48 mmol) and DMF (1 mL). The reaction solution was stirred at room temperature for 10 min. Then the crude amine (0.48 mmol) was added to the above solution and the reaction mixture was stirred at room temperature under air until the acyl fluoride intermediate was fully consumed. After these reactions were finished, water (20 mL) was added followed by EtOAc (3 x 10 mL) and the layers were separated. Combined organic layers were washed with brine (1 x 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated in vacuum. The crude product was purified by silica gel chromatography to afford **14** (92%, 0.31 g).



<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.67 – 7.92 (m, 3H), 7.69 (d, *J* = 7.8 Hz, 3H), 7.57 (d, *J* = 7.0 Hz, 1H), 7.47 – 7.14 (m, 5H), 7.13 – 6.87 (m, 8H), 6.75 (d, *J* = 7.7 Hz, 2H), 5.34 (s, 1H), 5.01 (s, 1H), 4.69 (s, 1H), 4.61 – 4.44 (m, 1H), 4.45 – 4.03 (m, 6H), 3.29 (s, 3H), 3.19 – 2.79 (m, 4H), 1.64 – 1.34 (m, 3H), 1.15 (s, 9H), 0.78 (s, 6H). <sup>13</sup>C {H} NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  172.9, 172.0, 170.6, 168.5, 168.1, 156.3, 154.1, 144.2, 143.8, 141.2, 136.4, 131.5, 130.0, 129.6, 128.1, 127.6, 127.0, 126.6, 125.6, 125.3, 123.9, 119.8, 78.1, 67.3, 55.5, 53.9, 51.8, 50.6, 46.9, 43.7, 43.3, 41.6, 39.9, 39.6, 28.7, 24.8, 22.6, 22.3. HRMS m/z (ESI) calcd for C<sub>48</sub>H<sub>58</sub>N<sub>5</sub>O<sub>9</sub><sup>+</sup> [M+H]<sup>+</sup>: 848.4229, found: 848.4224.

# HPLC studies for determining epimerization with Ser, Asn, Asp



### HPLC Studies for Determining Racemization of 7k

**HPLC condition:** Chiralpak®IC  $250 \times 4.6$  mm column; hexanes (solvent A): isopropanol (solvent B); isocratic 15% solvent B in 30 min; flow rate = 1.0 mL/min; detection wavelength = 254 nm.

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	21.877	49.64	615.1429	2250.94
2	23.537	50.36	624.0292	2095.08
Total:		100.00	1239.1721	4346.02

(1)	) Mixed	HPLC	data	of 7k	and	7k'
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(2) Crude HPLC data of reaction mixture for preparing compound 7k

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	23.643	100.00	236.0804	910.23
Total:		100.00	236.0804	910.23

(3) Pure HPLC data of 7k

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	23.537	100.00	538.4030	1875.91
Total:		100.00	538.4030	1875.91


HPLC Studies for Determining Racemization of 7q

**HPLC condition:** Chiralpak®IC  $250 \times 4.6$  mm column; hexanes (solvent A): isopropanol (solvent B); isocratic 15% solvent B in 35 min; flow rate = 1.0 mL/min; detection wavelength = 254 nm.

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	19.140	48.95	203.2103	365.79
2	21.363	51.05	211.9121	334.10
Total:		100.00	415.1224	699.89

(1) Mixed HPLC data of 7q and 7q'

(2) Crude HPLC data of reaction mixture for preparing compound 7q

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	19.553	0.40	5.4583	22.92
2	21.743	99.60	1367.4906	1778.45
Total:		100.00	1372.9489	1801.37

(3) Pure HPLC data of 7q

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	19.227	0.49	5.1248	11.54
2	21.010	99.51	1045.5387	1499.59
Total:		100.00	1050.6635	1511.13



HPLC Studies for Determining Racemization of 7t

**HPLC condition:** Chiralpak®IC  $250 \times 4.6$  mm column; hexanes (solvent A): isopropanol (solvent B); isocratic 15% solvent B in 35 min; flow rate = 1.0 mL/min; detection wavelength = 254 nm.

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	19.750	53.13	853.7111	1248.72
2	21.503	46.87	753.2121	844.08
Total:		100.00	1606.9232	2092.80

(1) Mixed HPLC data of 7t and 7t'

(2) Crude HPLC data of reaction mixture for preparing compound 7t

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	19.507	0.04	0.3512	0.85
2	21.953	99.96	937.0125	1078.06
Total:		100.00	937.3637	1078.92

(3) Pure HPLC data of 7t

Peak No	RT (min)	Area (%)	Area	Height (mAU)
1	20.633	0.02	0.2424	19.25
2	21.343	99.98	1172.9898	1341.83
Total:		100.00	1173.2322	1361.08

### **Procedures for the synthesis of peptides on resin (SPPS)**

## General procedure for incorporating the first amino acid on the solid support (2-CTC resin)

2-CTC resin (39.0 mg, 0.03 mmol) with a loading of 0.77 mmol/g was placed in a 5.0 mL fritted syringe, and the resin was swollen in DCM (2 mL) for 30 min. Then the syringe was drained, and Fmoc-Gly-OH (0.09 mmol 3.0 equiv.), DIPEA (18.2 mg, 6.0 equiv.), and 2 mL of a 1:1 DMF/DCM mixture (v/v) were added to the fritted syringe. The loading reaction was left to proceed for 2 h at room temperature before the syringe was drained. Then, the resin was washed with DMF (4 × 2 mL) and DCM (4 × 2 mL).

# General procedure for incorporating the first amino acid on the solid support (MBHA resin)

MBHA resin (0.03 mmol) with a loading of 0.79 mmol/g was placed in a 5.0 mL fritted syringe. The resin was successively swollen in DCM (2 mL) for 30 min, then drained and treated with a solution (2 mL) of piperidine: DMF (20:80, v/v, 20% piperidine in DMF) for 5 min before being drained. The solution was added again for 15 min, then drained and the resin was washed with DMF ( $4 \times 2$  mL) and DCM ( $4 \times 2$  mL). A 5 mL tube was charged with Fmoc-Xaa-OH (0.09 mmol), TFPN (0.09 mmol), DIPEA (0.11 mmol), and DMF (1 mL). The reaction solution was reacted at room temperature for 10 min. Then this solution was added to the fritted syringe. The loading reaction was left to proceed until the resin gave a negative color test. Then, the resin was washed with DMF ( $4 \times 2.0$  mL) and DCM ( $4 \times 2.0$  mL) before being drained.

#### General procedure for removing Fmoc-protection group

The solution of 20% piperidine in DMF (1 mL) was added to the fritted syringe to react for 5 min before being drained. A further 1 mL of 20% piperidine was added to react for 15 min. Then, the resin was again carefully washed with DMF ( $4 \times 2$  mL) and DCM ( $4 \times 2$  mL) before being drained.

#### General procedure for peptide elongation

A 5 mL tube was charged with Fmoc-Xaa-OH (0.09 mmol), TFPN (0.09 mmol), DIPEA (0.11 mmol), and DMF (1 mL). The reaction solution was reacted at room temperature for 10 min. Then this solution was added to the fritted syringe. The loading reaction was left to proceed until the resin gave a negative color test. Then, the resin was washed with DMF ( $4 \times 2.0$  mL) and DCM ( $4 \times 2.0$  mL) before being drained.

#### General procedure of resin cleavage

The resin was washed with DCM (4  $\times$  2 mL) and diethyl ether (4  $\times$  2 mL) immediately after synthesis was completed. Cleavage was then performed with 0.5 mL of a freshly prepared TFA/TIS/H<sub>2</sub>O (95:2.5:2.5) solution for 2 h. The resin was then filtered and washed with pure TFA (2  $\times$  0.5 mL). After combining the TFA solution, TFA was removed by Nitrogen, the residue was washed by ice-cold ether (4  $\times$  5 mL) and dissolved in MeCN/H<sub>2</sub>O (1:1, 3 mL). This solution was analyzed by analytical RP-HPLC subsequently.

**NH<sub>2</sub>-Ala-Ala-Val-Gly-Phe-OH** (12): HRMS m/z (ESI) calcd for  $C_{22}H_{34}N_5O_6^+$   $[M+H]^+$ : 464.2504, found: 464.2501.



HPLC condition: Jupiter 5 $\mu$ m C18 4.6 × 250 mm<sup>2</sup> column; 0.05% TFA (v/v) in H<sub>2</sub>O (solvent A), 0.05% TFA (v/v) in MeCN (solvent B); gradient 10%-100% (solvent B) in 30 min; flow rate = 1.0 mL/min; detection wavelength = 214 nm.



100.00

484.4316

Total

**NH<sub>2</sub>-Tyr-Ala-Gly-Phe-Leu-OH (13):** HRMS m/z (ESI) calcd for  $C_{29}H_{40}N_5O_7^+$  [M+H]<sup>+</sup>: 570.2922, found: 570.2916.



HPLC condition: Jupiter 5 $\mu$ m C18 4.6 × 250 mm<sup>2</sup> column; 0.05% TFA (v/v) in H<sub>2</sub>O (solvent A), 0.05% TFA (v/v) in MeCN (solvent B); gradient 10%-100% (solvent B) in 30 min; flow rate = 1.0 mL/min; detection wavelength = 214 nm.



**NH<sub>2</sub>-Tyr-Aib-Aib-Phe-Leu-NH<sub>2</sub> (14):** HRMS m/z (ESI) calcd for  $C_{32}H_{47}N_6O_6^+$  [M+H]<sup>+</sup>: 611.3552, found: 611.3549.



HPLC condition: Jupiter 5 $\mu$ m C18 4.6 × 250 mm<sup>2</sup> column; 0.05% TFA (v/v) in H<sub>2</sub>O (solvent A), 0.05% TFA (v/v) in MeCN (solvent B); gradient 10%-100% (solvent B) in 30 min; flow rate = 1.0 mL/min; detection wavelength = 214 nm.



NH<sub>2</sub>-Me-Ala-Tyr-Me-Ala-Gly-Phe-Leu-NH<sub>2</sub> (15): HRMS m/z (ESI) calcd for  $C_{34}H_{50}N_7O_7^+$  [M+H]<sup>+</sup>: 668.3766, found: 668.3757.



HPLC condition: Jupiter 5 $\mu$ m C18 4.6 × 250 mm<sup>2</sup> column; 0.05% TFA (v/v) in H<sub>2</sub>O (solvent A), 0.05% TFA (v/v) in MeCN (solvent B); gradient 10%-100% (solvent B) in 30 min; flow rate = 1.0 mL/min; detection wavelength = 214 nm.



NH2-Val-Gln-Ala-Ala-Ile-Asp-Tyr-Ile-Asn-Gly-OH (16): HRMS m/z (ESI) calcd for

 $C_{47}H_{75}N_{12}O_{16}^{+}$  [M+H]<sup>+</sup>: 1063.5419, found: 1063.5422.



HPLC condition: Jupiter 5 $\mu$ m C18 4.6 × 250 mm<sup>2</sup> column; 0.05% TFA (v/v) in H<sub>2</sub>O (solvent A), 0.05% TFA (v/v) in MeCN (solvent B); gradient 10%-100% (solvent B) in 30 min; flow rate = 1.0 mL/min; detection wavelength = 214 nm.



## NMR spectrum



Figure S1. <sup>1</sup>H NMR of compound **3a** in DMSO-*d*<sub>6</sub>, 400 MHz for <sup>1</sup>H NMR.



Figure S2. <sup>13</sup>C NMR of compound 3a in DMSO- $d_6$ , 100 MHz for <sup>13</sup>C NMR.



Figure S3. <sup>13</sup>F NMR of compound **3a** in DMSO- $d_6$ , 376 MHz for <sup>13</sup>F NMR.



Figure S4. <sup>1</sup>H NMR of compound 4a in DMSO-*d*<sub>6</sub>, 400 MHz for <sup>1</sup>H NMR.



Figure S5. <sup>13</sup>C NMR of compound 4a in DMSO- $d_6$ , 100 MHz for <sup>13</sup>C NMR.



Figure S6. <sup>1</sup>H NMR of compound 4b in DMSO-*d*<sub>6</sub>, 400 MHz for <sup>1</sup>H NMR.



Figure S7. <sup>13</sup>C NMR of compound 4b in DMSO- $d_6$ , 100 MHz for <sup>13</sup>C NMR.



Figure S8. <sup>1</sup>H NMR of compound 6a in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S9. <sup>13</sup>C NMR of compound 6a in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



**Figure S10.** <sup>1</sup>H NMR of compound **6b** in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S11. <sup>13</sup>C NMR of compound 6b in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S12. <sup>1</sup>H NMR of compound 6c in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S13. <sup>13</sup>C NMR of compound 6c in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S14. <sup>1</sup>H NMR of compound 6d in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S15. <sup>13</sup>C NMR of compound 6d in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S16. <sup>1</sup>H NMR of compound 6e in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S17. <sup>13</sup>C NMR of compound 6e in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S18. <sup>1</sup>H NMR of compound 6f in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S19. <sup>13</sup>C NMR of compound 6f in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S10. <sup>1</sup>H NMR of compound 6g in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S21. <sup>13</sup>C NMR of compound 6g in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S22. <sup>1</sup>H NMR of compound 6h in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S23. <sup>13</sup>C NMR of compound 6h in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S24. <sup>1</sup>H NMR of compound 6i in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S25. <sup>13</sup>C NMR of compound 6i in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S26. <sup>1</sup>H NMR of compound 6j in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S27. <sup>13</sup>C NMR of compound 6j in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S28. <sup>1</sup>H NMR of compound 7a in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S29. <sup>13</sup>C NMR of compound 7a in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S30. <sup>1</sup>H NMR of compound 7b in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S31. <sup>13</sup>C NMR of compound 7b in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S32. <sup>1</sup>H NMR of compound 7c in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S33. <sup>13</sup>C NMR of compound 7c in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S34. <sup>1</sup>H NMR of compound 7d in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S35. <sup>13</sup>C NMR of compound 7d in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S36. <sup>1</sup>H NMR of compound 7e in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S37. <sup>13</sup>C NMR of compound 7e in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S38. <sup>1</sup>H NMR of compound 7f in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S39. <sup>13</sup>C NMR of compound 7f in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S40. <sup>1</sup>H NMR of compound 7g in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S41. <sup>13</sup>C NMR of compound 7g in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S42. <sup>1</sup>H NMR of compound 7h in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S43. <sup>13</sup>C NMR of compound 7h in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S44. <sup>1</sup>H NMR of compound 7i in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S45. <sup>13</sup>C NMR of compound 7i in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S46. <sup>1</sup>H NMR of compound 7j in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S47. <sup>13</sup>C NMR of compound 7j in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S48. <sup>1</sup>H NMR of compound 7k in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S49. <sup>13</sup>C NMR of compound 7k in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S50. <sup>1</sup>H NMR of compound 71 in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S51. <sup>13</sup>C NMR of compound 7l in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S52. <sup>1</sup>H NMR of compound 7m in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S53. <sup>13</sup>C NMR of compound 7m in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.


Figure S54. <sup>1</sup>H NMR of compound 7n in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S55 <sup>13</sup>C NMR of compound 7n in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S56. <sup>1</sup>H NMR of compound 70 in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S57. <sup>13</sup>C NMR of compound 70 in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S58. <sup>1</sup>H NMR of compound 7p in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S59. <sup>13</sup>C NMR of compound 7p in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S60. <sup>1</sup>H NMR of compound 7q in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S61. <sup>13</sup>C NMR of compound 7q in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S62. <sup>1</sup>H NMR of compound 7r in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S63. <sup>13</sup>C NMR of compound 7r in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S64. <sup>1</sup>H NMR of compound 7s in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S65. <sup>13</sup>C NMR of compound 7s in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S66. <sup>1</sup>H NMR of compound 7t in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S67. <sup>13</sup>C NMR of compound 7t in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S68. <sup>1</sup>H NMR of compound 7u in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S69. <sup>13</sup>C NMR of compound 7u in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S70. <sup>1</sup>H NMR of compound 7v in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S71. <sup>13</sup>C NMR of compound 7v in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S72. <sup>1</sup>H NMR of compound 7w in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S73. <sup>13</sup>C NMR of compound 7w in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S74. <sup>1</sup>H NMR of compound 7x in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S75. <sup>13</sup>C NMR of compound 7x in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S76. <sup>1</sup>H NMR of compound 7y in DMSO-*d*<sub>6</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S77. <sup>13</sup>C NMR of compound 7y in DMSO-*d*<sub>6</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S78. <sup>1</sup>H NMR of compound 7z in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S79. <sup>13</sup>C NMR of compound 7z in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S80. <sup>1</sup>H NMR of compound 7aa in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S81. <sup>13</sup>C NMR of compound 7aa in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S82. <sup>1</sup>H NMR of compound 7ab in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S83. <sup>13</sup>C NMR of compound 7ab in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.





Figure S84. <sup>1</sup>H NMR of compound 7ac in DMSO-*d*<sub>6</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S85. <sup>13</sup>C NMR of compound 7ac in DMSO-*d*<sub>6</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S86. <sup>1</sup>H NMR of compound 7ad in DMSO-*d*<sub>6</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S87. <sup>13</sup>C NMR of compound 7ad in DMSO-*d*<sub>6</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S88. <sup>1</sup>H NMR of compound 7ae in DMSO-*d*<sub>6</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S89. <sup>13</sup>C NMR of compound 7ae in DMSO-*d*<sub>6</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S90. <sup>1</sup>H NMR of compound 7af in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S91. <sup>13</sup>C NMR of compound 7af in CDCl<sub>3</sub> 75 MHz for <sup>13</sup>C NMR.



Figure S92. <sup>1</sup>H NMR of compound 7ag in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S93. <sup>13</sup>C NMR of compound 7ag in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S94. <sup>1</sup>H NMR of compound 7ah in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S95. <sup>13</sup>C NMR of compound 7ah in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S96. <sup>1</sup>H NMR of compound 8 in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S97. <sup>13</sup>C NMR of compound 8 in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S98. <sup>1</sup>H NMR of compound 10 in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S99. <sup>13</sup>C NMR of compound 10 in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S100. <sup>1</sup>H NMR of compound 12 in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S101. <sup>13</sup>C NMR of compound 12 in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.



Figure S102. <sup>1</sup>H NMR of compound 14 in CDCl<sub>3</sub>, 300 MHz for <sup>1</sup>H NMR.



Figure S103. <sup>13</sup>C NMR of compound 14 in CDCl<sub>3</sub>, 75 MHz for <sup>13</sup>C NMR.

## **Reference:**

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