

Supporting information

Photocatalytic method for the generation of the 1,1,1,3,3,3-hexafluoroisopropyl radical

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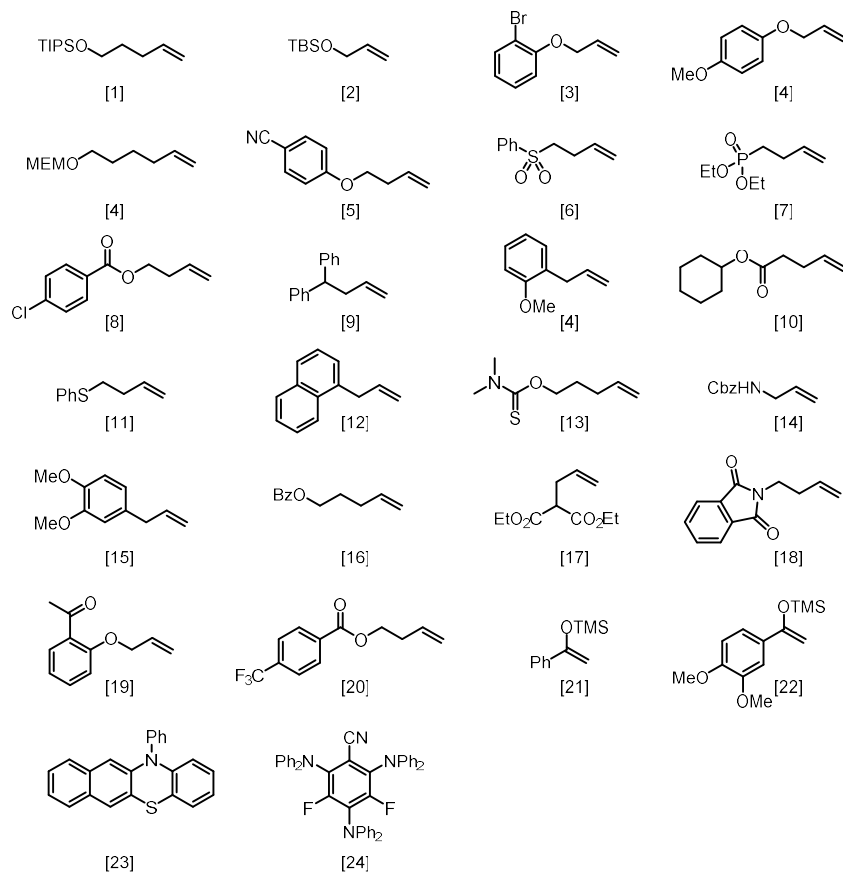
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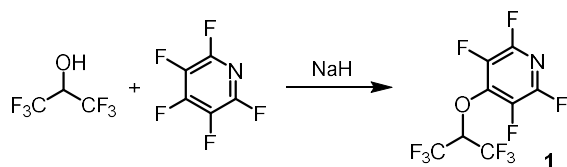
General methods

All reactions were performed in an atmosphere of dry argon. Column chromatography was carried out employing silica gel (230–400 mesh). Precoated silica gel plates F-254 were used for thin-layer analytical chromatography visualizing with UV and/or acidic aq. KMnO_4 solution. High resolution mass-spectra (HRMS) were measured using electrospray ionization (ESI) and a time-of-flight (TOF) mass analyzer (Bruker MicrOTOF II). The measurements were done in a positive-ion mode (interface capillary voltage -4500 V) or in a negative-ion mode (3200 V); the mass ranged from m/z 50 to m/z 3000. For NMR measurements, a Bruker AM300 spectrometer was used. Infrared spectra were recorded on Bruker Alpha-T spectrometer. Photo-induced reactions were performed in Duran culture tubes (Roth cat. no K248.1, outside diameter = 12 mm). The reaction tube was placed in a glass jacket covered with aluminium foil and cooled with water (18 °C). For irradiation with 400 nm light, COB LED matrix Hontiey (29-32 V, 3000 mA, 100W; the LED matrix was operated at 80W or 60W). For irradiation with 450 nm light, a strip of light emitting diodes (SMD 2835–120 LED 1 M Blue, 12 V, 24 W/m; 50 cm strip length; operated at 10W) was used. The distance between the reaction vessel and diodes was about 1 cm.

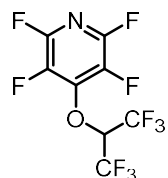
Starting materials

DMF, DMSO, and MeCN were distilled from CaH_2 and stored over $\text{MS } 4\text{\AA}$. Diethyl ether was distilled from lithium aluminum hydride. Following compounds were obtained according to literature procedures:



Reagent 1

In a round bottom flask in a stream of argon, sodium hydride (4.0 g, 60% in mineral oil, 100 mmol, 1.2 eq.) was washed with hexane (3×3 mL), and 50 mL of freshly distilled diethyl ether was added. The mixture was immersed into room temperature water bath. Hexafluoroisopropanol (HFIP) (16.8 g, 100 mmol, 1.2 eq.) was added dropwise via a syringe, and the mixture was stirred 10 minutes. The bath was replaced by ice/water, and the mixture was cooled to 0 °C, and then pentafluoropyridine (14.1 g, 83.3 mmol, 1 eq.) was rapidly added. The mixture was allowed to warm up to room temperature over 2 hours, while keeping the flask in the bath. The reaction was quenched with a solution of saturated ammonium chloride (10 mL). The organic layer was separated, the aqueous phase extracted with hexane (3×5 mL). The combined organic layers were dried over Na₂SO₄ and carefully fractionally distilled under stream of argon at ambient pressure. The fraction boiling at 142-143 °C was collected affording 25.3 g (80%). This material contains 5% impurity of the pentafluoropyridine disubstitution product. The resulting impurity does not interfere with the subsequent fluoroalkylation reaction. Its content can be reduced to 2% after additional fractional distillation through a column packed with Raschig rings.

2,3,5,6-Tetrafluoro-4-((1,1,1,3,3,3-hexafluoropropan-2-yl)oxy)pyridine (1)

Yield 25.3 g (80%), colorless liquid. Bp 142-143 °C.

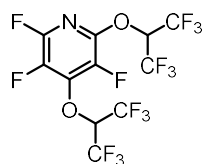
¹H NMR (300 MHz, Chloroform-d) δ 5.24 (hept, *J* = 5.2 Hz, 1H).

¹³C NMR (75 MHz, Chloroform-d) δ 144.53 (dtd, *J* = 245.2, 15.1, 2.9 Hz), 144.4 (tt, *J* = 10.0, 5.2 Hz), 138.0 – 133.7 (m), 120.5 (qq, *J* = 284.3, 2.4 Hz), 77.0 (hept. t, *J* = 35.0, 3.8 Hz).

¹⁹F NMR (282 MHz, Chloroform-d) δ -73.75 (dt, *J* = 5.1, 2.5 Hz, 6F), -86.65 – -87.02 (m, 2F), -155.85 – -156.18 (m, 2F).

MS (EI): 317 [M], 298 [M-F], 248 [M-CF₃], 69 [CF₃].

HRMS (ESI-TOF): calcd for C₈F₁₀NO, [M⁺] 316.9893; found 316.9882.

Impurity

Selected signals:

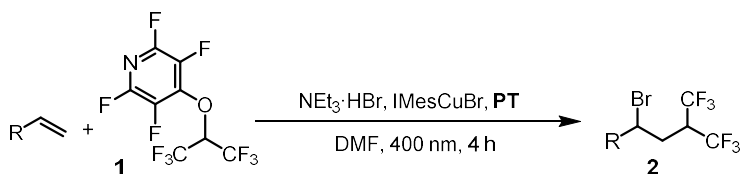
^1H NMR (300 MHz, Chloroform-*d*) δ 6.12 – 6.01 (m, 1H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 69.56 (hept, $J = 35.4$ Hz).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -73.36, -73.38, -88.28 (dd, $J = 26.7, 20.2$ Hz), -154.76 (d, $J = 26.7$ Hz), -159.07 (d, $J = 20.1$ Hz).

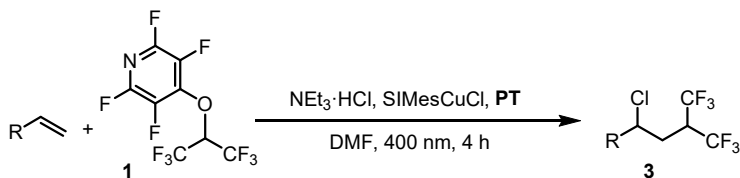
MS (EI): 465 [M], 446 [M-F], 396 [M-CF₃], 69 [CF₃].

Synthesis of bromides 2 (General Procedure A)



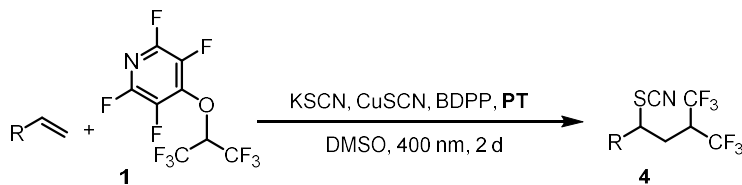
A screw test tube charged with $\text{NEt}_3 \cdot \text{HBr}$ (110 mg, 0.6 mmol 1.2 eq.), IMesCuBr (22 mg, 0.05 mmol 10 mol%), 12-phenyl-12H-benzo[*b*]phenothiazine (**PT**) (4 mg, 3 mol%) and a PTFE coated stirring bar was evacuated-back filled with argon three times, then dry DMF (1.0 mL) was added. Alkene (0.5 mmol, 1 eq.) and reagent **1** (238 mg, 0.75 mmol, 1.5 eq.) were added with a syringe. The tube was sealed and irradiated using 80W 400 nm LED for 4 hours. For the work-up, the reaction was quenched with water (4 mL) and the resulting mixture was extracted with MTBE (3×3 mL). The combined organic layers were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel.

Synthesis of chlorides 3 (General Procedure B)



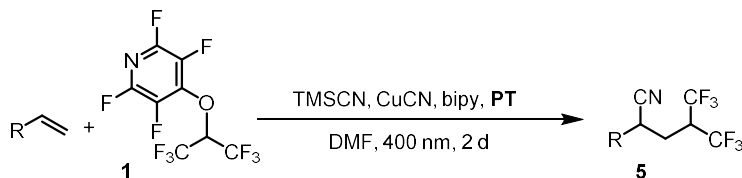
A screw test tube charged with $\text{NEt}_3 \cdot \text{HCl}$ (82 mg, 0.6 mmol 1.2 eq.), SIMesCuCl (20 mg, 0.05 mmol 10 mol%), 12-phenyl-12H-benzo[*b*]phenothiazine (**PT**) (4 mg, 3 mol%) and a PTFE coated stirring bar was evacuated-back filled with argon three times, then dry DMF (1.0 mL) was added. Alkene (0.5 mmol, 1 eq.) and reagent **1** (238 mg, 0.75 mmol, 1.5 eq.) were added with a syringe. The tube was sealed and irradiated using 80W 400 nm LED for 4 hours. For the work-up, the reaction was quenched with water (4 mL) and the resulting mixture was extracted with MTBE (3×3 mL). The combined organic layers were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel.

Synthesis of thiocyanates 4 (General Procedure C)



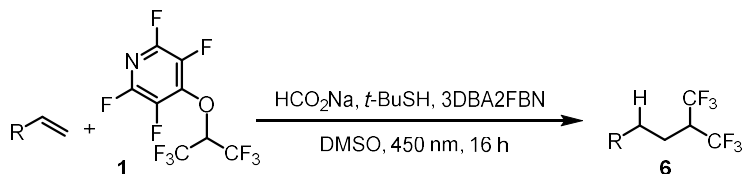
A screw test tube charged with CuSCN (6 mg, 0.05 mmol 10 mol%), KSCN (58 mg, 0.6 mol, 1.2 eq.), 2,6-bis(3,5-dimethyl-1H-pyrazol-1-yl)pyridine (BDPPENT, **L1**) (15 mg, 0.05 mmol 10 mol%), 12-phenyl-12H-benzo[b]phenothiazine (**PT**) (4 mg, 3 mol%) and a PTFE coated stirring bar was evacuated-back filled with argon three times, then dry DMSO (1.0 mL) was added. Alkene (0.5 mmol, 1 eq.) and reagent **1** (238 mg, 0.75 mmol, 1.5 eq.) were added with a syringe. The tube was sealed and irradiated using 60W 400 nm LED for 2 days. For the work-up, the reaction was quenched with water (4 mL) and the resulting mixture was extracted with MTBE (3×3 mL). The combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel.

Synthesis of nitriles 5 (General Procedure D)



A screw test tube charged with CuCN (5 mg, 0.05 mmol 10 mol%), 2,2'-bipyridine (bipy) (9 mg, 0.05 mmol 10 mol%), 12-phenyl-12H-benzo[b]phenothiazine (**PT**) (4 mg, 3 mol%) and a PTFE coated stirring bar was evacuated-back filled with argon three times, then dry DMF (1.0 mL) was added. Alkene (0.5 mmol, 1 eq.), trimethylsilyl cyanide (TMSCN) (59 mg, 75 μ l, 0.6 mmol, 1.2 eq.) and reagent **1** (238 mg, 0.75 mmol, 1.5 eq.) were added with a syringe. The tube was sealed and irradiated using 400 nm 60W LED for 2 days. For the work-up, the reaction was quenched with water (4 mL) and the resulting mixture was extracted with MTBE (3×3 mL). The combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel.

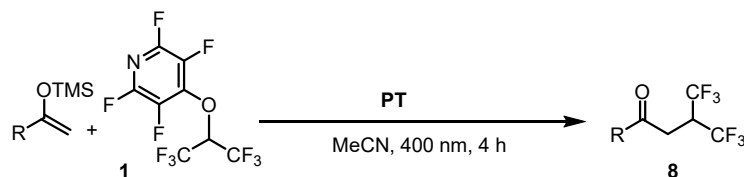
Synthesis of compounds 6 (General procedure E)



A test tube was evacuated and filled with argon. Then, DMSO (1 mL), reagent **1** (317 mg, 1 mmol, 2 eq.), alkene (0.5 mmol, 1 eq.), HCO₂Na (68 mg, 1 mmol, 2 eq.), 2-methylpropane-2-thiol (6 μ l,

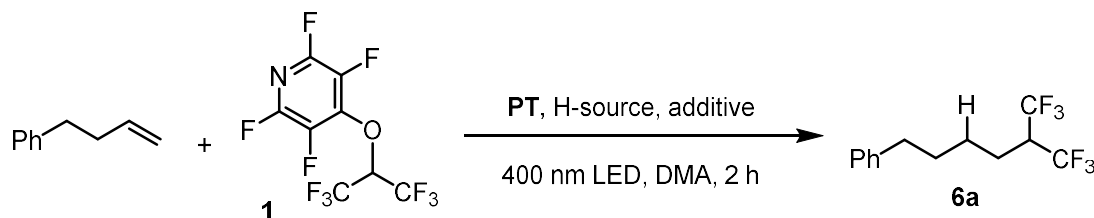
0.05 mmol, 10 mol%), 3DPA2FBN (1.6 mg, 2.5 μ mol, 0.5 mol%) were added. The tube was screw-capped and irradiated using 450 nm 10W LED strip for 16 hours. For the work-up, the reaction was quenched with water (5 mL) and extracted with hexane (3 \times 1.5 mL). The combined organic phases were filtered through a short pad of Na₂SO₄ and concentrated on a rotary evaporator. The residue was purified by column chromatography on silica gel.

Synthesis of ketones **8** (General procedure F)



A screw test tube charged with, 12-phenyl-12H-benzo[b]phenothiazine (**PT**) (4 mg, 3 mol%) and a PTFE coated stirring bar was evacuated-back filled with argon three times, then dry MeCN (1.0 mL) was added. Silyl enol ether (0.5 mmol, 1 eq.) and reagent **1** (238 mg, 0.75 mmol, 1.5 eq.) were added with a syringe. The tube was sealed and irradiated using and 400 nm 80W LED for 4 hours. For the work-up, reaction was quenched with water (4 mL) and the resulting mixture was extracted with MTBE (3 \times 3 mL). The combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel.

Variation of the source of hydrogen



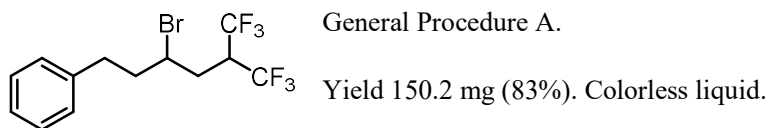
#	H-source	Additive	Conv. of alkene, %	Yield of 6a , % ^a
1	γ -terpinene	-	41	38
2	DIPEA	CySH	39	21
3	Ascorbic acid	NEt ₃	20	13
4	Hantzsch ester	-	trace	trace
5	<i>t</i> -Bu-triazinane ^b	-	26	14

^a Determined by ¹⁹F NMR with PhCF₃ as an internal standard.

^b 1,3,5-Tri-*tert*-butyl-1,3,5-triazinane.

Characterization of compounds

(3-Bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)benzene (2a)



Chromatography: hexanes/EtOAc, 20/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21 × 250 mm, 5 μm), flow rate 8 mL min⁻¹; mobile phase isocratic, acetonitrile/water, 15% water; t_R = 22.5 min

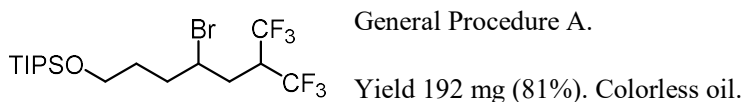
¹H NMR (300 MHz, Chloroform-*d*) δ 7.38 – 7.19 (m, 5H), 4.15 – 4.01 (m, 1H), 3.54 – 3.34 (m, 1H), 3.04 – 2.90 (m, 1H), 2.88 – 2.75 (m, 1H), 2.36 – 2.14 (m, 4H).

¹³C NMR (76 MHz, Chloroform-*d*) δ 140.2, 128.8, 128.6, 126.6, 123.9 (qq, *J* = 280.6, 2.4 Hz), 123.6 (qq, *J* = 280.0, 3.3 Hz), 52.5 (q, *J* = 2.1 Hz), 47.1 (hept, *J* = 28.1 Hz), 41.2, 33.7, 33.6 (t, *J* = 1.8 Hz).

¹⁹F NMR (282 MHz, Chloroform-*d*) δ -66.51 (qd, *J* = 9.7, 7.7 Hz, 3F), -67.80 (qd, *J* = 9.8, 8.2 Hz, 3F).

Calcd for C₁₃H₁₃BrF₆ (362.01): C 43.00%, H 3.63%; found: C 43.31%, H 3.78%.

((4-Bromo-7,7,7-trifluoro-6-(trifluoromethyl)heptyl)oxy)triisopropylsilane (2b)



Chromatography: hexane/EtOAc, 50/1.

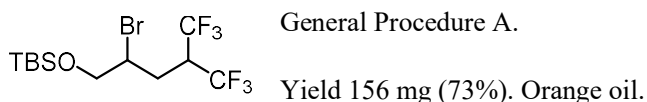
¹H NMR (300 MHz, Chloroform-*d*) δ 4.24 – 4.08 (m, 1H), 3.74 (t, *J* = 5.9 Hz, 2H), 3.53 – 3.32 (m, 1H), 2.39 – 2.14 (m, 2H), 2.14 – 1.60 (m, 4H), 1.15 – 0.93 (m, 21H).

¹³C NMR (75 MHz, Chloroform-*d*) δ 124.0 (qq, *J* = 280.7, 2.6 Hz), 123.7 (qq, *J* = 280.0, 3.3 Hz), 62.3, 53.3 (d, *J* = 2.1 Hz), 47.2 (hept, *J* = 28.1 Hz), 36.1, 33.8, 30.7, 18.1, 12.1.

¹⁹F NMR (282 MHz, Chloroform-*d*) δ -67.29 (pent, *J* = 9.6 Hz, 3F), -68.59 (pent, *J* = 9.5 Hz, 3F).

HRMS (ESI-TOF): calcd for C₁₇H₃₁[⁷⁹Br]F₆OSiNa [M+Na]: 495.1124; found 495.1117.

((2-Bromo-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)oxy)(*tert*-butyl)dimethylsilane (2c)



Chromatography: pentane/DCM, 5/1.

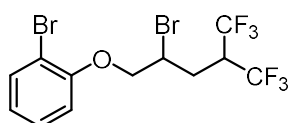
^1H NMR (300 MHz, Chloroform-*d*) δ 4.08 (ddt, $J = 10.6, 7.6, 3.9$ Hz, 1H), 3.97 (dd, $J = 10.7, 3.9$ Hz, 1H), 3.76 (dd, $J = 10.7, 7.7$ Hz, 1H), 3.52 – 3.30 (m, 2H), 2.59 (ddd, $J = 15.7, 9.2, 3.9$ Hz, 1H), 2.15 (ddd, $J = 15.7, 10.6, 1.3$ Hz, 1H), 0.91 (s, 9H), 0.10 (s, 3H), 0.09 (s, 3H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 124.0 (qq, $J = 280.3, 2.3$ Hz), 123.8 (qq, $J = 279.9, 3.2$ Hz), 67.5, 50.8 (d, $J = 2.2$ Hz), 46.8 (hept, $J = 28.1$ Hz), 30.2, 25.8, 18.3, -5.3, -5.4.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.49 (pent, $J = 9.5$ Hz, 3F), -68.64 (pent, $J = 9.5$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{12}\text{H}_{21}[^{79}\text{Br}]\text{F}_6\text{OSiNa}$ [$\text{M}+\text{Na}$]: 425.0341; found 425.0328.

1-Bromo-2-((2-bromo-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)oxy)benzene (2d)



General Procedure A.

Yield 171 mg (77%). Pale-yellow oil.

Chromatography: hexane/EtOAc, 50/1.

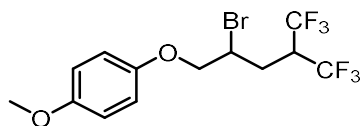
^1H NMR (300 MHz, Chloroform-*d*) δ 7.57 (dd, $J = 7.8, 1.5$ Hz, 1H), 7.28 (td, $J = 7.8, 1.5$ Hz, 1H), 6.97 – 6.85 (m, 2H), 4.54 – 4.34 (m, 2H), 4.28 – 4.11 (m, 1H), 3.66 – 3.42 (m, 1H), 2.82 (ddd, $J = 15.8, 9.1, 3.3$ Hz, 1H), 2.48 – 2.29 (m, 1H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 154.3, 133.8, 128.7, 123.9 (qq, $J = 280.3, 2.2$ Hz), 123.7 (qq, $J = 279.9, 3.2$ Hz), 123.3, 113.9, 112.6, 72.8, 46.8 (hept, $J = 28.2$ Hz), 46.5, 30.7.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.46 (q, $J = 9.6$ Hz, 3F), -68.55 (q, $J = 9.8$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{12}\text{H}_{10}[^{81}\text{Br}]\text{F}_6\text{ONa}$ [$\text{M}+\text{Na}$]: 468.8854; found 468.8857.

1-((2-Bromo-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)oxy)-4-methoxybenzene (2e)



General Procedure A.

Yield 146 mg (74%). Pale-yellow oil.

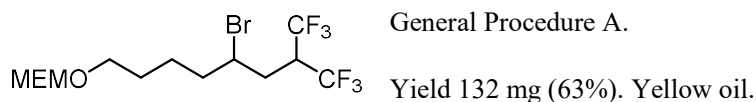
Chromatography: hexane/EtOAc, 35/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 6.86 (s, 4H), 4.42 – 4.25 (m, 2H), 4.12 (dd, $J = 9.7, 7.5$ Hz, 1H), 3.78 (s, 3H), 3.57 – 3.36 (m, 1H), 2.68 (ddd, $J = 15.9, 9.5, 3.1$ Hz, 1H), 2.29 (t, $J = 13.5$ Hz, 1H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 154.9, 152.1, 123.9 (qq, $J = 280.5, 2.2$ Hz), 123.6 (qq, $J = 279.4, 2.7, 2.2$ Hz), 116.2, 115.0, 72.9, 55.8, 47.4, 46.8 (hept, $J = 28.2$ Hz), 30.4.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.29 (pent, $J = 9.5$ Hz, 3F), -68.52 (pent, $J = 9.6$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{13}[^{79}\text{Br}]\text{F}_6\text{O}_2\text{Na}$ [$\text{M}+\text{Na}$]: 416.9895; found 416.9905.

4-Bromo-1,1,1-trifluoro-8-((2-methoxyethoxy)methoxy)-2-(trifluoromethyl)octane (2f)

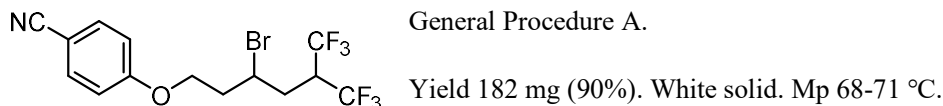
Chromatography: hexane/EtOAc, 4/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 4.70 (s, 2H), 4.17 – 4.01 (m, 1H), 3.69 (dd, $J = 6.1, 3.3$ Hz, 2H), 3.62 – 3.49 (m, 4H), 3.49 – 3.34 (m, 1H), 3.39 (s, 3H), 2.37 – 2.12 (m, 2H), 1.99 – 1.80 (m, 2H), 1.76 – 1.45 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 123.9 (qq, $J = 280.6, 2.4$ Hz), 123.6 (qq, $J = 280.0, 3.2$ Hz), 95.6, 71.9, 67.4, 66.9, 59.1, 53.0 (q, $J = 2.0$ Hz), 47.1 (hept, $J = 28.2$ Hz), 39.2, 33.5, 29.0, 24.4.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.33 (pent, $J = 9.6$ Hz, 3F), -68.67 (pent, $J = 9.4$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{21}[^{79}\text{Br}]\text{F}_6\text{O}_3\text{Na}$ [$\text{M}+\text{Na}$]: 441.0470; found 441.0475.

4-((3-Bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)oxy)benzonitrile (2g)

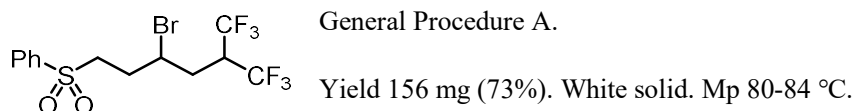
Chromatography: hexane/EtOAc, 8/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.59 (d, $J = 8.6$ Hz, 2H), 7.03 – 6.85 (m, 2H), 4.48 – 4.29 (m, 1H), 4.28 – 4.11 (m, 2H), 3.56 – 3.29 (m, 1H), 2.53 – 2.14 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 161.8, 134.2, 123.9 (qq, $J = 280.7, 2.3$ Hz), 123.5 (qq, $J = 280.3, 3.2$ Hz), 119.1, 115.4, 104.6, 65.6, 49.0, 47.0 (hept, $J = 28.2$ Hz), 38.6, 33.7.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.25 (pent, $J = 9.5$ Hz, 3F), -68.53 (pent, $J = 9.4$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{12}[^{81}\text{Br}]\text{F}_6\text{NONa}$ [$\text{M}+\text{Na}$]: 427.9879; found 427.9871.

((3-Bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)sulfonyl)benzene (2h)

Chromatography: hexane/EtOAc, 5/1.

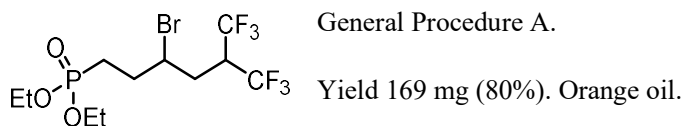
^1H NMR (300 MHz, Chloroform-*d*) δ 7.94 (d, $J = 7.5$ Hz, 2H), 7.72 (t, $J = 7.5$ Hz, 1H), 7.62 (t, $J = 7.5$ Hz, 2H), 4.15 (tt, $J = 10.3, 3.5$ Hz, 1H), 3.54 – 3.22 (m, 3H), 2.52 – 2.34 (m, 1H), 2.34 – 2.12 (m, 3H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 138.8, 134.23, 129.7, 128.1, 123.7 (qq, $J = 280.7, 2.2$ Hz), 123.4 (qq, $J = 280.2, 3.1$ Hz), 54.4, 50.1 (d, $J = 2.3$ Hz), 46.9 (hept, $J = 28.3$ Hz), 33.5, 32.3.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.26 (pent, $J = 9.5$ Hz, 3F), -68.58 (pent, $J = 9.8$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{13}[^{79}\text{Br}]\text{F}_6\text{O}_2\text{SNa}$ [$\text{M}+\text{Na}$]: 448.9616; found 448.9612.

Diethyl (3-bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)phosphonate (2i)



Chromatography: hexane/EtOAc, 1/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 4.19 – 3.95 (m, 5H), 3.49 – 3.23 (m, 1H), 2.34 – 1.74 (m, 6H), 1.29 (t, $J = 7.1$ Hz, 6H).

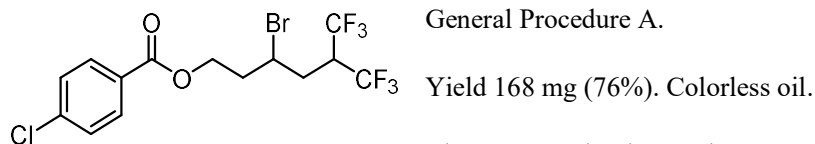
^{13}C NMR (75 MHz, Chloroform-*d*) δ 123.8 (qq, $J = 280.5, 2.4$ Hz), 123.5 (qq, $J = 279.6, 2.9$ Hz), 62.0 (dd, $J = 6.5, 4.6$ Hz), 52.8 (dt, $J = 17.8, 2.1$ Hz), 47.00 (hept, $J = 28.3$ Hz), 33.4, 32.6 (d, $J = 3.9$ Hz), 24.1 (d, $J = 143.2$ Hz), 16.5 (d, $J = 6.0$ Hz).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -66.56 (qd, $J = 9.7, 8.0$ Hz, 3F), -67.89 (qd, $J = 9.9, 8.1$ Hz, 3F).

^{31}P NMR (121 MHz, CDCl_3) δ 30.67.

HRMS (ESI-TOF): calcd for $\text{C}_{11}\text{H}_{18}[^{79}\text{Br}]\text{F}_6\text{O}_3\text{PNa}$ [$\text{M}+\text{Na}$]: 444.9973; found 444.9978.

3-Bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexyl 4-chlorobenzoate (2j)



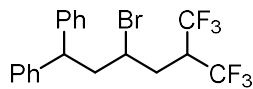
Chromatography: hexane/EtOAc, 20/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.95 (d, $J = 8.4$ Hz, 2H), 7.42 (d, $J = 8.7$ Hz, 2H), 4.68 – 4.43 (m, 2H), 4.27 (tt, $J = 9.0, 4.1$ Hz, 1H), 3.55 – 3.32 (m, 1H), 2.50 – 2.20 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 165.5, 139.9, 131.1, 129.0, 128.3, 123.9 (qq, $J = 280.7, 2.4$ Hz), 123.5 (qq, $J = 280.4, 3.1$ Hz), 62.6, 48.7, 47.0 (hept, $J = 28.6$ Hz), 38.3, 33.7.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.24 (pent, $J = 9.6$ Hz, 3F), -68.55 (pent, $J = 9.8$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{12}[^{79}\text{Br}]\text{F}_6\text{ClO}_2\text{Na}$ [$\text{M}+\text{Na}$]: 462.9506; found 462.9504.

(3-Bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexane-1,1-diyl)dibenzene (2k)

General Procedure A.

Yield 213 mg (97%). Pale-red oil.

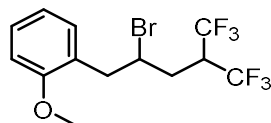
Chromatography: hexane/EtOAc, 20/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.40 – 7.14 (m, 10H), 4.35 (t, $J = 7.8$ Hz, 1H), 3.94 – 3.75 (m, 1H), 3.44 – 3.22 (m, 1H), 2.58 (dd, $J = 8.6, 5.9$ Hz, 2H), 2.41 – 2.18 (m, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 143.6, 142.3, 129.0, 128.9, 128.0, 127.7, 127.1, 126.8, 123.8 (qq, $J = 280.5, 2.1$ Hz), 123.6 (qq, $J = 280.0, 3.2$ Hz), 51.5, 49.1, 47.0 (hept, $J = 28.1$ Hz), 45.4, 33.8.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.53 (pent, $J = 9.6$ Hz), -68.64 (pent, $J = 9.7$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{19}\text{H}_{17}[^{79}\text{Br}]\text{F}_6\text{Na}$ [$\text{M}+\text{Na}$]: 461.0310; found 461.0322.

1-(2-Bromo-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)-2-methoxybenzene (2l)

General Procedure A.

Yield 135 mg (71%). Pale-yellow oil.

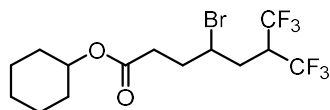
Chromatography: hexane/EtOAc, 50/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.23 (t, $J = 8.0$ Hz, 1H), 7.11 (d, $J = 7.4$ Hz, 1H), 6.93 – 6.78 (m, 2H), 4.44 (dtd, $J = 10.6, 6.8, 3.4$ Hz, 1H), 3.77 (s, 3H), 3.48 – 3.26 (m, 1H), 3.19 (d, $J = 7.0$ Hz, 2H), 2.34 – 2.07 (m, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 157.6, 131.4, 128.9, 125.7, 123.9 (qq, $J = 280.5, 2.3$ Hz), 123.7 (qq, $J = 280.0, 3.3$ Hz), 120.7, 110.6, 55.3, 51.8 (d, $J = 2.2$ Hz), 47.2 (hept, $J = 28.0$ Hz), 41.3, 33.0.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.42 (pent, $J = 9.5$ Hz 3F), -68.61 (pent, $J = 9.4$ Hz 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{13}[^{79}\text{Br}]\text{F}_6\text{ONa}$ [$\text{M}+\text{Na}$]: 400.9946; found 400.9945.

Cyclohexyl 4-bromo-7,7,7-trifluoro-6-(trifluoromethyl)heptanoate (2m)

General Procedure A.

Yield 153 mg (76%). Pale-red oil.

Chromatography: hexane/EtOAc, 25/1.

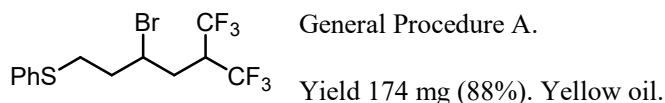
^1H NMR (300 MHz, Chloroform-*d*) δ 4.87 – 4.67 (m, 1H), 4.26 – 4.05 (m, 1H), 3.52 – 3.27 (m, 1H), 2.69 – 2.44 (m, 2H), 2.40 – 2.17 (m, 3H), 2.16 – 1.99 (m, 1H), 1.93 – 1.09 (m, 10H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 171.7, 123.9 (qq, $J = 280.7, 2.2$ Hz), 123.6 (qq, $J = 280.0, 3.3$ Hz), 73.3, 52.1, 47.1 (hept, $J = 28.0$ Hz), 34.5, 33.7, 32.6, 31.74, 31.72, 25.5, 23.9.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.36 (pent, $J = 9.4$ Hz, 3F), -68.65 (pent, $J = 9.6$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{19}[^{79}\text{Br}]\text{F}_6\text{O}_2\text{Na}$ [$\text{M}+\text{Na}$]: 435.0365; found 435.0357.

(3-Bromo-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)(phenyl)sulfane (2n)



Chromatography: hexane/EtOAc, 50/1.

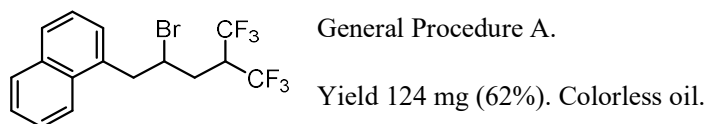
^1H NMR (300 MHz, Chloroform-*d*) δ 7.49 – 7.22 (m, 5H), 4.44 – 4.25 (m, 1H), 3.54 – 3.35 (m, 1H), 3.28 (ddd, $J = 13.2, 7.6, 4.9$ Hz, 1H), 3.09 (dt, $J = 13.2, 7.7$ Hz, 1H), 2.34 – 2.03 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 135.1, 130.4, 129.3, 126.9, 123.9 (qq, $J = 280.6, 2.2$ Hz), 123.6 (qq, $J = 280.1, 3.3$ Hz), 51.4 (d, $J = 2.0$ Hz), 47.0 (hept, $J = 28.3$ Hz), 38.6, 33.5, 31.9.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.27 (pent, $J = 9.4$ Hz, 3F), -68.59 (pent, $J = 9.4$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{13}[^{81}\text{Br}]\text{F}_6\text{S}[^{107}\text{Ag}]$ [$\text{M}+\text{Ag}$]: 502.8853; found 502.8855.

1-(2-Bromo-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)naphthalene (2o)



Chromatography: hexane/EtOAc, 100/1.

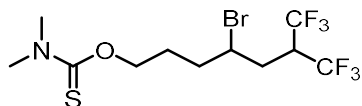
Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, 5 μm), flow rate 6 $\text{mL}\cdot\text{min}^{-1}$; mobile phase: isocratic, acetonitrile/water, 10% water; $t_{\text{R}} = 20.9$ min).

^1H NMR (300 MHz, Chloroform-*d*) δ 7.99 – 7.87 (m, 2H), 7.84 (d, $J = 8.2$ Hz, 1H), 7.65 – 7.31 (m, 4H), 4.67 – 4.41 (m, 1H), 3.78 (dd, $J = 14.6, 7.2$ Hz, 1H), 3.63 (dd, $J = 14.6, 7.0$ Hz, 1H), 3.55 – 3.33 (m, 1H), 2.52 – 2.21 (m, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 134.2, 133.1, 131.7, 129.3, 128.5, 127.9, 126.7, 126.0, 125.5, 123.8 (qq, $J = 280.5, 2.2$ Hz), 123.6 (qq, $J = 280.0, 3.1$ Hz), 122.9, 51.9, 47.3 (hept, $J = 28.0$ Hz), 43.3, 33.3.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.31 (pent, $J = 9.4$ Hz, 3F), -68.59 (pent, $J = 9.5$ Hz, 3F).

Anal. Calcd for $\text{C}_{16}\text{H}_{13}\text{BrF}_6$: C, 48.14; H, 3.28. Found: C, 48.27, H, 3.36.

O-(4-bromo-7,7,7-trifluoro-6-(trifluoromethyl)heptyl) dimethylcarbamothioate (2p)

General Procedure A.

Yield 156 mg (77%). Orange oil.

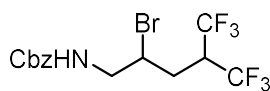
Chromatography: hexane/EtOAc, 12/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 4.59 – 4.39 (m, 2H), 4.20 – 4.01 (m, 1H), 3.49 – 3.27 (m, 1H), 3.35 (s, 3H), 3.10 (s, 3H), 2.39 – 2.13 (m, 2H), 2.12 – 1.78 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 188.2, 123.9 (qq, $J = 280.6, 2.3$ Hz), 123.5 (qq, $J = 280.1, 3.3$ Hz), 70.4, 52.6 (d, $J = 2.3$ Hz), 47.1 (hept, $J = 28.2$ Hz), 42.8, 37.8, 36.1, 33.6, 27.1.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.24 (pent, $J = 9.5$ Hz, 3F), -68.60 (pent, $J = 9.6$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{11}\text{H}_{16}[^{79}\text{Br}]\text{F}_6\text{NOSNa}$ [$\text{M}+\text{Na}$]: 425.9932; found 425.9922.

Benzyl (2-bromo-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)carbamate (2q)

General Procedure A.

Yield 188 mg (89%). Orange oil.

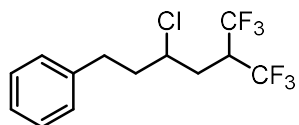
Chromatography: hexane/EtOAc, 5/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.44 – 7.30 (m, 5H), 5.25 (s, 1H), 5.14 (s, 2H), 4.33 – 4.15 (m, 1H), 3.76 – 3.49 (m, 2H), 3.49 – 3.29 (m, 1H), 2.36 (ddd, $J = 13.4, 9.0, 3.9$ Hz, 1H), 2.19 (t, $J = 13.5$ Hz, 1H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 156.5, 136.2, 128.8, 128.5, 128.3, 123.7 (qq, $J = 280.6, 2.2$ Hz), 123.5 (qq, $J = 280.2, 2.9$ Hz), 67.5, 51.7, 47.6, 46.8 (hept, $J = 28.4$ Hz), 30.4.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.32 (pent, $J = 9.6$ Hz, 3F), -68.34 (pent, $J = 9.5$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{14}[^{79}\text{Br}]\text{F}_6\text{NO}_2\text{Na}$ [$\text{M}+\text{Na}$]: 444.0004; found 443.9992.

(3-Chloro-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)benzene (3a)

General Procedure B.

Yield 127.2 mg (80%). Colorless liquid.

Chromatography: hexanes/EtOAc, 10/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, $5 \mu\text{m}$), flow rate 6 mL min^{-1} ; mobile phase isocratic, acetonitrile/water, 10% water; $t_R = 20.0$ min.

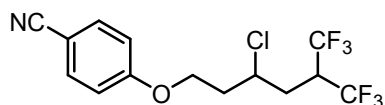
^1H NMR (300 MHz, Chloroform-*d*) δ 7.42 – 7.17 (m, 5H), 4.13 – 3.96 (m, 1H), 3.66 – 3.21 (m, 1H), 2.96 (dt, $J = 14.3, 7.2$ Hz, 1H), 2.81 (dt, $J = 13.8, 8.1$ Hz, 1H), 2.36 – 2.04 (m, 4H).

^{13}C NMR (76 MHz, Chloroform-*d*) δ 140.3, 128.8, 128.5, 126.53, 124.0 (qq, $J = 280.4, 2.3$ Hz), 123.7 (qq, $J = 280.1, 3.5$ Hz), 59.2 (q, $J = 1.9$ Hz), 45.9 (hept., $J = 28.2$ Hz), 40.6, 33.0, 32.7.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.40 (pent, $J = 9.6$ Hz, 3F), -68.65 (pent, $J = 9.5$ Hz, 3F).

Calcd for $\text{C}_{13}\text{H}_{13}\text{ClF}_6$ (318.06): C 49.00, H 4.11; found: C 48.92, H 4.31.

4-((3-Chloro-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)oxy)benzonitrile (3b)



General Procedure B.

Yield 158.0 mg (88%), colorless liquid.

Chromatography: hexanes/EtOAc, 10/1.

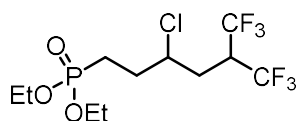
^1H NMR (300 MHz, Chloroform-*d*) δ 7.57 (d, $J = 8.8$ Hz, 2H), 6.95 (d, $J = 8.8$ Hz, 2H), 4.39 – 4.13 (m, 3H), 3.53 – 3.30 (m, 1H), 2.44 – 2.27 (m, 2H), 2.27 – 2.07 (m, 2H).

^{13}C NMR (76 MHz, Chloroform-*d*) δ 161.8, 134.10, 123.9 (qq, $J = 280.4, 2.4$ Hz), 123.5 (qq, $J = 280.0, 3.2$ Hz), 119.1, 115.3, 104.5, 64.6, 56.2 (d, $J = 2.0$ Hz), 45.7 (hept., $J = 28.3$ Hz), 37.9, 33.0.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -66.54 (pent, $J = 9.5$ Hz, 3F), -67.80 (qd, $J = 9.8, 8.0$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{12}[^{35}\text{Cl}]\text{F}_6\text{NONa}$, $[\text{M}+\text{Na}]$ 382.0404; found 382.0390

Diethyl (3-chloro-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)phosphonate (3c)



General Procedure B.

Yield 141.8 mg (75%). yellowish liquid.

Chromatography: hexanes/EtOAc, 1/1.

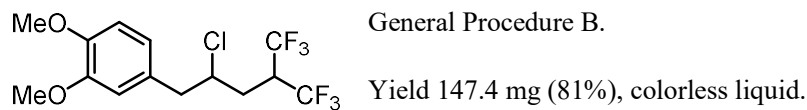
^1H NMR (300 MHz, Chloroform-*d*) δ 4.19 – 4.02 (m, 5H), 3.48 – 3.26 (m, 1H), 2.34 – 1.77 (m, 6H), 1.33 (t, $J = 7.1$ Hz, 6H).

^{13}C NMR (76 MHz, Chloroform-*d*) δ 123.8 (qq, $J = 280.5, 2.4$ Hz), 123.5 (qq, $J = 279.9, 3.2$ Hz), 61.9 (dd, $J = 6.5, 4.0$ Hz), 59.6 (dd, $J = 17.4, 2.1$ Hz), 45.7 (hept., $J = 28.3$ Hz), 32.7, 32.0 (d, $J = 4.0$ Hz), 22.84 (d, $J = 143.4$ Hz), 16.4 (d, $J = 5.9$ Hz).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -66.56 (pent, $J = 9.4$ Hz, 3F), -67.87 (qd, $J = 9.8, 8.1$ Hz, 3F).

^{31}P NMR (122 MHz, Chloroform-*d*) δ 29.98.

HRMS (ESI-TOF): calcd for $\text{C}_{11}\text{H}_{18}[^{35}\text{Cl}]\text{F}_6\text{O}_3\text{PNa}$, $[\text{M}+\text{Na}]$ 401.0478; found 401.0472.

4-(2-Chloro-5,5,5-trifluoro-4-(trifluoromethyl)pentyl)-1,2-dimethoxybenzene (3d)

Chromatography: hexanes/EtOAc, 10/1.

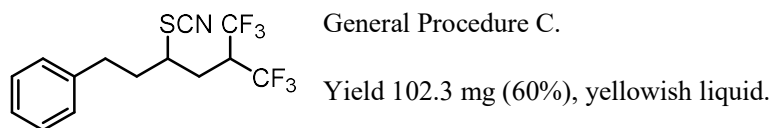
Final purification was performed by preparative HPLC (reversed-phase column C18, 21 × 250 mm, 5 μm), flow rate 8 mL min⁻¹; mobile phase isocratic, acetonitrile/water, 20% water; t_R = 15.1 min.

¹H NMR (300 MHz, Chloroform-d) δ 6.83 (d, *J* = 8.1 Hz, 1H), 6.79 – 6.67 (m, 2H), 4.32 – 4.14 (m, 1H), 3.88 (s, 6H), 3.53 – 3.30 (m, 1H), 3.12 (dd, *J* = 14.3, 7.1 Hz, 1H), 2.98 (dd, *J* = 14.3, 6.7 Hz, 1H), 2.37 – 2.18 (m, 1H), 2.04 (t, *J* = 13.5 Hz, 1H).

¹³C NMR (76 MHz, Chloroform-d) δ 149.17, 148.43, 128.90, 123.95 (qq, *J* = 280.6, 2.4 Hz), 123.60 (qq, *J* = 279.9, 3.3 Hz), 121.48, 112.35, 111.40, 59.94 (q, *J* = 1.9 Hz), 56.03, 55.99, 45.88 (hept, *J* = 28.2 Hz), 44.86, 32.16.

¹⁹F NMR (282 MHz, Chloroform-d) δ -66.46 (pent, *J* = 9.4 Hz, 3F), -67.84 (pent, *J* = 9.5 Hz, 3F).

HRMS (ESI-TOF): calcd for C₁₄H₁₅[³⁵Cl]F₆O₂Na, [M+Na] 387.0557; found 387.0567.

(6,6,6-Trifluoro-3-thiocyanato-5-(trifluoromethyl)hexyl)benzene (4a)

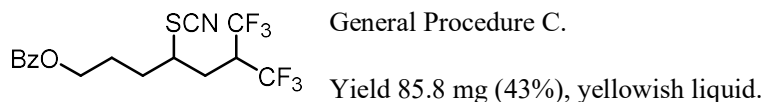
Chromatography: hexanes/EtOAc, 20/1.

¹H NMR (300 MHz, Chloroform-d) δ 7.40 – 7.18 (m, 5H), 3.43 – 3.17 (m, 1H), 3.12 – 2.93 (m, 2H), 2.89 – 2.75 (m, 1H), 2.42 – 2.14 (m, 4H).

¹³C NMR (76 MHz, Chloroform-d) δ 139.3, 128.9, 128.4, 126.9, 123.7 (qq, *J* = 280.6, 2.4 Hz), 123.3 (qq, *J* = 280.6, 3.0 Hz), 108.4, 47.3 (d, *J* = 1.7 Hz), 46.1 (hept., *J* = 28.3 Hz), 37.3, 33.0, 30.3 – 29.3 (m).

¹⁹F NMR (282 MHz, Chloroform-d) δ -66.41 (qd, *J* = 9.7, 7.6 Hz, 3F), -67.56 (qd, *J* = 9.8, 7.9 Hz, 3F).

HRMS (ESI-TOF): calcd for C₁₄H₁₃F₆NSNa, [M+Na] 364.0565; found 364.0578.

7,7,7-Trifluoro-4-thiocyanato-6-(trifluoromethyl)heptyl benzoate (4b)

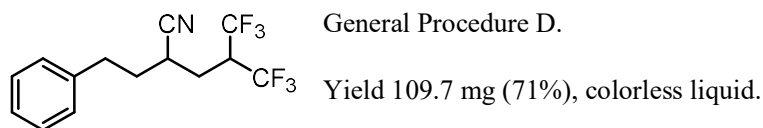
Chromatography: hexanes/EtOAc, 20/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 8.04 (d, $J = 7.3$ Hz, 2H), 7.58 (t, $J = 7.3$ Hz, 1H), 7.46 (t, $J = 7.3$ Hz, 2H), 4.49 – 4.28 (m, 2H), 4.05 – 3.88 (m, 1H), 3.27 – 3.08 (m, 1H), 2.14 – 1.79 (m, 6H).

^{13}C NMR (76 MHz, Chloroform-*d*) δ 166.62, 135.60, 133.30, 130.05, 129.72, 128.59, 123.75 (qq, $J = 280.7$, 2.3 Hz), 123.44 (qq, $J = 280.2$, 3.4 Hz), 63.84, 56.19 – 56.01 (m), 45.59 (hept, $J = 28.5$ Hz), 33.72, 30.40, 25.55.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.15 – -67.41 (m, 3F), -68.39 – -68.65 (m, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{16}\text{H}_{15}\text{F}_6\text{NO}_2\text{SNa}$, $[\text{M}+\text{Na}]$ 422.0620; found 422.0606.

5,5,5-Trifluoro-2-phenethyl-4-(trifluoromethyl)pentanenitrile (5a)

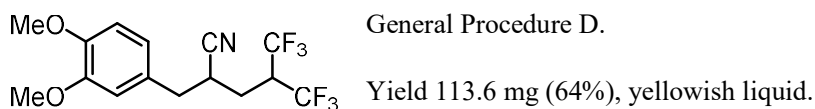
Chromatography: hexanes/EtOAc, 15/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.43 – 7.17 (m, 5H), 3.34 – 3.11 (m, 1H), 3.03 – 2.88 (m, 1H), 2.88 – 2.73 (m, 3H), 2.22 – 1.88 (m, 3H).

^{13}C NMR (76 MHz, Chloroform-*d*) δ 139.3, 128.9, 128.4, 126.9, 123.6 (qq, $J = 280.6$, 2.5 Hz), 123.3 (qq, $J = 280.4$, 3.2 Hz), 119.8, 46.5 (hept, $J = 28.5$ Hz), 34.3, 33.1, 29.4, 26.9 – 26.6 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -66.5 (qd, $J = 9.8$, 7.4 Hz, 3F), -67.8 (qd, $J = 9.9$, 7.7 Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{13}\text{F}_6\text{NNa}$, $[\text{M}+\text{Na}]$ 332.0844; found 332.0846.

2-(3,4-Dimethoxybenzyl)-5,5,5-trifluoro-4-(trifluoromethyl)pentanenitrile (5b)

Chromatography: hexanes/EtOAc, 3/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21 × 250 mm, 5 μm), flow rate 8 mL min^{-1} ; mobile phase isocratic, acetonitrile/water, 20% water; $t_R = 10.0$ min.

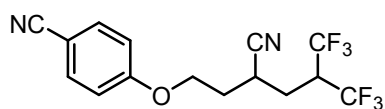
^1H NMR (300 MHz, Chloroform- d) δ 6.85 (d, $J = 7.9$ Hz, 1H), 6.81 – 6.73 (m, 2H), 3.88 (s, 6H), 3.31 – 3.12 (m, 1H), 3.11 – 2.97 (m, 1H) 2.92 (t, $J = 6.3$ Hz, 2H), 2.15 – 2.02 (m, 2H).

^{13}C NMR (76 MHz, Chloroform- d) δ 149.41, 148.81, 127.73, δ 123.61 (qq, $J = 280.8, 2.4$ Hz), 123.25 (qq, $J = 280.6$ Hz, 2.4Hz), 121.34, 119.81, 112.01, 111.64, 56.09, 56.05, 46.55 (hept, $J = 28.4$ Hz), 38.28, 32.13, 26.41.

^{19}F NMR (282 MHz, Chloroform- d) δ -66.3 (qd, $J = 9.8, 7.4$ Hz, 3F), -67.7 (qd, $J = 9.9, 7.8$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{15}\text{H}_{15}\text{F}_6\text{NO}_2\text{Na}$, $[\text{M}+\text{Na}]$ 378.0899; found 378.0901.

4-((3-Cyano-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)oxy)benzonitrile (5c)



General Procedure D.

Yield 106.8 mg (71%), colorless liquid.

Chromatography: hexanes/EtOAc, 3/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, $5 \mu\text{m}$), flow rate 6 mL min^{-1} ; mobile phase isocratic, acetonitrile/water, 15% water; $t_R = 12.0$ min.

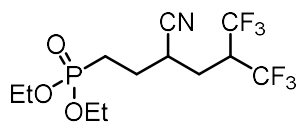
^1H NMR (300 MHz, Chloroform- d) δ 7.60 (d, $J = 8.9$ Hz, 2H), 6.96 (d, $J = 8.9$ Hz, 2H), 4.22 (t, $J = 5.7$ Hz, 2H), 3.34 – 3.10 (m, 2H), 2.24 – 2.11 (m, 4H).

^{13}C NMR (76 MHz, Chloroform- d) δ 161.3, 134.3, 123.6 (qq, $J = 280.8, 2.3$ Hz). 123.2 (qq, $J = 280.4, 3.2$ Hz), 119.2, 119.0, 115.3, 105.1, 64.7, 46.5 (hept., $J = 28.6$ Hz), 32.0, 27.2 (d, $J = 1.9$ Hz), 26.8.

^{19}F NMR (282 MHz, Chloroform- d) δ -66.4 (qd, $J = 9.9, 7.5$ Hz, 3F), -67.7 (qd, $J = 9.9, 7.7$ Hz, 3F).

HRMS (ESI-TOF): calcd for $\text{C}_{15}\text{H}_{12}\text{F}_6\text{N}_2\text{ONa}$, $[\text{M}+\text{Na}]$ 373.0746; found 373.0748.

Diethyl (3-cyano-6,6,6-trifluoro-5-(trifluoromethyl)hexyl)phosphonate (5d)



General Procedure D.

Yield 147.6 mg (80%), colorless liquid.

Chromatography: EtOAc.

^1H NMR (300 MHz, Chloroform- d) δ 4.21 – 4.01 (m, 4H), 3.31 – 3.08 (m, 1H), 3.08 – 2.90 (m, 1H), 2.14 – 1.81 (m, 6H), 1.32 (t, $J = 7.1$ Hz, 6H).

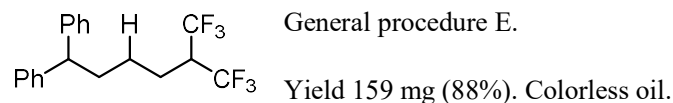
^{13}C NMR (76 MHz, Chloroform- d) δ 123.5 (qq, $J = 280.6, 2.5$ Hz), 123.1 (qq, $J = 280.3, 3.3$ Hz). 119.2, 62.1 (t, $J = 6.0$ Hz), 46.4 (hept., $J = 28.5$ Hz), 30.4 (d, $J = 15.4$ Hz), 26.6, 26.1 (d, $J = 4.3$ Hz), 23.24 (d, $J = 143.8$ Hz), 16.3 (d, $J = 6.0$ Hz).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -66.5 (pent, $J = 9.6$ Hz, 3F), -67.8 (pent, $J = 9.7$ Hz, 3F).

^{31}P NMR (122 MHz, Chloroform-*d*) δ 28.37.

HRMS (ESI-TOF): calcd for $\text{C}_{12}\text{H}_{19}\text{F}_6\text{NO}_3\text{P}$, $[\text{M}+\text{H}]$ 370.1001; found 370.1002.

(6,6,6-Trifluoro-5-(trifluoromethyl)hexane-1,1-diyl)dibenzene (6a)



Chromatography: hexane/EtOAc, 30/1.

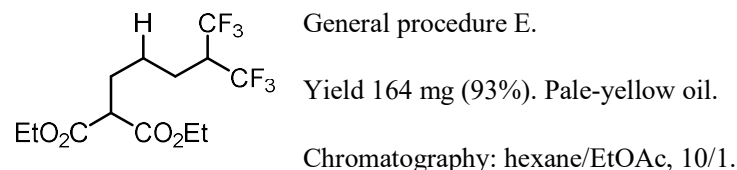
^1H NMR (300 MHz, Chloroform-*d*) δ 7.48 – 7.21 (m, 10H), 4.01 (t, $J = 7.8$ Hz, 1H), 3.01 – 2.73 (m, 1H), 2.19 (q, $J = 7.8$ Hz, 2H), 1.95 (q, $J = 7.0$ Hz, 2H), 1.77 – 1.49 (m, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 144.6, 128.7, 127.9, 126.5, 124.1 (qq, $J = 281.0, 3.6$ Hz), 51.1, 48.1 (hept, $J = 28.2, 27.7$ Hz), 35.4, 25.9, 23.9 – 23.6 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.94 (d, $J = 8.3$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{19}\text{H}_{19}\text{F}_6$ $[\text{M}+\text{H}]$: 361.1385; found 361.1381.

Diethyl 2-(5,5,5-trifluoro-4-(trifluoromethyl)pentyl)malonate (6b)

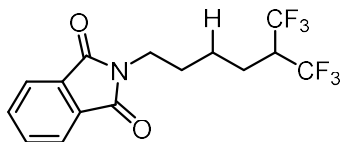


^1H NMR (300 MHz, Chloroform-*d*) δ 4.19 (q, $J = 7.1$ Hz, 4H), 3.32 (t, $J = 7.4$ Hz, 1H), 2.97 – 2.71 (m, 1H), 2.01 – 1.73 (m, 4H), 1.68 – 1.45 (m, 2H), 1.26 (t, $J = 7.1$ Hz, 7H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 169.2, 124.0 (qq, $J = 281.1, 3.9$ Hz), 61.6, 51.6, 47.9 (hept, $J = 28.1$ Hz), 28.4, 25.0, 23.6 – 23.4 (m), 14.1.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.96 (d, $J = 8.3$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{18}\text{F}_6\text{O}_4\text{Na}$ $[\text{M}+\text{Na}]$: 375.1001; found 375.0997.

2-(6,6,6-Trifluoro-5-(trifluoromethyl)hexyl)isoindoline-1,3-dione (6c)

General procedure E.

Yield 79 mg (45%). White solid (mp 63-66°C).

Chromatography: hexane/EtOAc, 6/1.

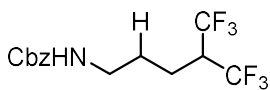
Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, 5 μm), flow rate 6 mL·min⁻¹; mobile phase: isocratic, acetonitrile/water, 20% water; tR = 15.7 min).

¹H NMR (300 MHz, Chloroform-*d*) δ 7.84 (t, *J* = 5.4, 3.2 Hz, 2H), 7.71 (dd, *J* = 5.4, 3.1 Hz, 2H), 3.70 (t, *J* = 7.1 Hz, 2H), 2.97 – 2.71 (m, 1H), 1.86 (dt, *J* = 7.3, 7.1 Hz, 2H), 1.73 (pent, *J* = 7.1 Hz, 2H), 1.65 – 1.49 (m, 2H).

¹³C NMR (75 MHz, Chloroform-*d*) δ 168.5, 134.1, 132.2, 124.0 (qq, *J* = 281.2, 3.9 Hz), 123.4, 48.1 (hept, *J* = 27.9 Hz), 37.4, 28.4, 24.6, 23.4 (pent, *J* = 1.9 Hz).

¹⁹F NMR (282 MHz, Chloroform-*d*) δ -67.90 (d, *J* = 8.3 Hz).

HRMS (ESI-TOF): calcd for C₁₅H₁₃F₆NO₂Na [M+Na]: 376.0743; found 376.0744.

Benzyl (5,5,5-trifluoro-4-(trifluoromethyl)pentyl)carbamate (6d)

General procedure E.

Yield 144 mg (84%). Colorless oil.

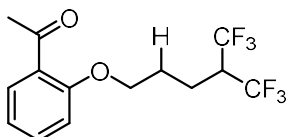
Chromatography: hexane/EtOAc, 5/1.

¹H NMR (300 MHz, Chloroform-*d*) δ 7.44 – 7.27 (m, 5H), 5.10 (s, 2H), 4.94 (s, 1H), 3.23 (q, *J* = 6.6 Hz, 2H), 3.04 – 2.74 (m, 1H), 1.97 – 1.54 (m, 4H).

¹³C NMR (75 MHz, Chloroform-*d*) δ 156.6, 136.6, 128.7, 128.3, 128.2, 124.0 (qq, *J* = 281.1, 3.5 Hz), 66.9, 47.8 (hept, *J* = 27.7, 27.2 Hz), 40.4, 27.7, 21.2 – 20.7 (m).

¹⁹F NMR (282 MHz, Chloroform-*d*) δ -67.75 (d, *J* = 8.1 Hz).

HRMS (ESI-TOF): calcd for C₁₄H₁₃F₆NO₂Na [M+Na]: 366.0899; found 366.0898.

1-(2-((5,5,5-Trifluoro-4-(trifluoromethyl)pentyl)oxy)phenyl)ethanone (6e)

General procedure E.

Yield 102 mg (62%). White solid (mp 56-58°C).

Chromatography: hexane/EtOAc, 8/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, 5 μm), flow rate 6 mL·min⁻¹; mobile phase: isocratic, acetonitrile/water, 20% water; tR = 18.3 min)

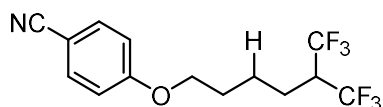
¹H NMR (300 MHz, Chloroform-*d*) δ 7.73 (dd, *J* = 7.5, 1.8 Hz, 1H), 7.45 (td, *J* = 8.4, 7.5, 1.8 Hz, 1H), 7.02 (t, *J* = 7.5 Hz, 1H), 6.93 (d, *J* = 8.4 Hz, 1H), 4.24 – 4.01 (m, 2H), 3.13 – 2.87 (m, 1H), 2.60 (s, 3H), 2.21 – 2.01 (m, 4H).

¹³C NMR (75 MHz, Chloroform-*d*) δ 199.6, 157.9, 133.7, 130.7, 128.7, 124.0 (qq, *J* = 281.1, 3.6 Hz), 121.1, 112.3, 67.6, 47.9 (hept, *J* = 27.7 Hz), 31.8, 27.1, 21.1 (pent, *J* = 2.0 Hz).

¹⁹F NMR (282 MHz, Chloroform-*d*) δ -67.77 (d, *J* = 8.0 Hz).

HRMS (ESI-TOF): calcd for C₁₄H₁₃F₆O₂ [M+H]: 329.0971; found 329.0981.

4-((6,6,6-Trifluoro-5-(trifluoromethyl)hexyl)oxy)benzonitrile (6f)



General procedure E.

Yield 128 mg (79%). Pale-yellow oil.

Chromatography: hexane/EtOAc, gradient elution from 8/1 to 6/1.

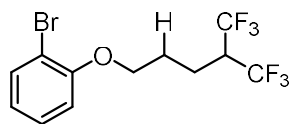
¹H NMR (300 MHz, Chloroform-*d*) δ 7.57 (d, *J* = 8.7 Hz, 2H), 6.92 (d, *J* = 8.7 Hz, 2H), 4.02 (t, *J* = 5.9 Hz, 2H), 3.05 – 2.74 (m, 1H), 2.05 – 1.79 (m, 4H), 1.78 – 1.54 (m, 2H).

¹³C NMR (75 MHz, Chloroform-*d*) δ 162.2, 134.1, 124.0 (qq, *J* = 281.5, 4.0 Hz), 119.3, 115.3, 104.1, 67.7, 48.1 (hept, *J* = 27.9 Hz), 28.8, 24.0, 23.7 – 23.4 (m).

¹⁹F NMR (282 MHz, Chloroform-*d*) δ -67.86 (d, *J* = 8.1 Hz).

HRMS (ESI-TOF): calcd for C₁₄H₁₃F₆NONa [M+Na]: 348.0794; found 348.0788.

1-Bromo-2-((5,5,5-trifluoro-4-(trifluoromethyl)pentyl)oxy)benzene (6g)



General procedure E.

Yield 84 mg (46%). Colorless oil.

Chromatography: hexane/EtOAc, 50/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, 5 μm), flow rate 6 mL·min⁻¹; mobile phase: isocratic, acetonitrile/water, 15% water; tR = 22.9 min).

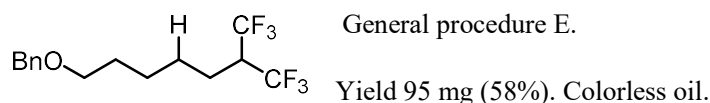
¹H NMR (300 MHz, Chloroform-*d*) δ 7.55 (dd, *J* = 7.7, 1.6 Hz, 1H), 7.26 (ddd, *J* = 8.3, 7.7, 1.6 Hz, 1H), 6.86 (t, *J* = 7.7 Hz, 2H), 4.07 (t, *J* = 5.3 Hz, 2H), 3.30 – 3.07 (m, 1H), 2.19 – 1.99 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 155.1, 133.6, 128.6, 124.2 (qq, $J = 281.5, 3.5$ Hz), 122.4, 113.3, 112.4, 68.4, 47.8 (hept, $J = 27.9$ Hz), 26.7, 21.2 – 21.0 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.79 (d, $J = 8.3$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{12}\text{H}_{11}[^{79}\text{Br}]\text{F}_6\text{ONa}$ [$\text{M}+\text{Na}$]: 386.9790; found 386.9785.

(((7,7,7-Trifluoro-6-(trifluoromethyl)heptyl)oxy)methyl)benzene (6h)



Chromatography: hexane/EtOAc, 30/1.

Final purification was performed by preparative HPLC (reversed-phase column C18, 21×250 mm, 5 μm), flow rate 6 mL·min⁻¹; mobile phase: isocratic, acetonitrile/water, 15% water; t_R = 27.1 min)

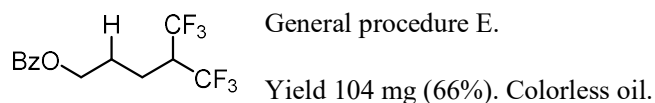
^1H NMR (300 MHz, Chloroform-*d*) δ 7.40 – 7.27 (m, 5H), 4.51 (s, 2H), 3.48 (t, $J = 6.3$ Hz, 2H), 2.93 – 2.72 (m, 1H), 1.93 – 1.73 (m, 2H), 1.71 – 1.35 (m, 6H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 138.7, 128.5, 127.8, 127.7, 124.2 (qq, $J = 281.0, 3.7$ Hz), 73.1, 70.1, 48.1 (hept, $J = 27.6$ Hz), 29.4, 27.2, 26.2, 23.8 (pent, $J = 1.9$ Hz).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.89 (d, $J = 8.3$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{15}\text{H}_{18}\text{F}_6\text{ONa}$ [$\text{M}+\text{Na}$]: 351.1154; found 351.1167.

5,5,5-Trifluoro-4-(trifluoromethyl)pentyl benzoate (6i)



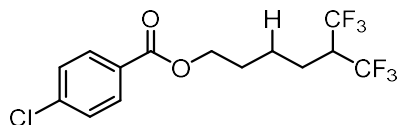
Chromatography: hexane/EtOAc, 20/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 8.03 (d, $J = 7.5$ Hz, 2H), 7.58 (tt, $J = 7.5, 1.5$ Hz, 1H), 7.45 (t, $J = 7.5$ Hz, 2H), 4.46 – 4.26 (m, 2H), 3.10 – 2.88 (m, 1H), 2.12 – 1.95 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 166.5, 133.2, 130.0, 129.6, 128.5, 124.0 (qq, $J = 281.1, 3.7$ Hz), 63.7, 47.8 (hept, $J = 27.8$ Hz), 26.3, 21.1 – 20.1 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.81 (d, $J = 8.0$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{12}\text{F}_6\text{O}_2\text{Na}$ [$\text{M}+\text{Na}$]: 337.0634; found 337.0636.

6,6,6-Trifluoro-5-(trifluoromethyl)hexyl 4-chlorobenzoate (6j)

General procedure E.

Yield 129 mg (71%). Colorless oil.

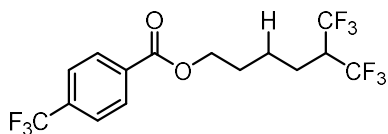
Chromatography: hexane/EtOAc, 15/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.95 (d, $J = 8.5$ Hz, 2H), 7.39 (d, $J = 8.5$ Hz, 2H), 4.34 (t, $J = 6.3$ Hz, 2H), 3.00 – 2.74 (m, 1H), 1.99 – 1.75 (m, 4H), 1.74 – 1.57 (m, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 165.8, 139.6, 131.0, 128.9, 124.1 (qq, $J = 281.2, 3.9$ Hz), 64.5, 48.1 (hept, $J = 27.9$ Hz), 28.6, 24.0, 23.7 – 23.4 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.89 (d, $J = 8.4$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{14}\text{H}_{13}[^{35}\text{Cl}]\text{F}_6\text{O}_2\text{Na}$ [$\text{M}+\text{Na}$]: 385.0400; found 385.0409.

6,6,6-Trifluoro-5-(trifluoromethyl)hexyl 4-(trifluoromethyl)benzoate (6k)

General procedure E.

Yield 149 mg (75%). Colorless oil.

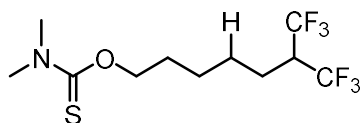
Chromatography: hexane/EtOAc, gradient elution from 25/1 to 15/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 8.14 (d, $J = 8.1$ Hz, 2H), 7.70 (d, $J = 8.1$ Hz, 2H), 4.38 (t, $J = 6.3$ Hz, 2H), 3.00 – 2.75 (m, 1H), 2.03 – 1.78 (m, 4H), 1.77 – 1.56 (m, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 165.5, 134.7 (q, $J = 32.7$ Hz), 133.6, 130.1, 125.6 (q, $J = 3.8$ Hz), 124.1 (qq, $J = 281.0, 3.6$ Hz), 123.81 (q, $J = 272.7$ Hz), 122.0, 64.8, 48.2 (hept, $J = 27.8$ Hz), 28.6, 24.0, 23.8 – 23.4 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -64.06 (s, 3F), -67.95 (d, $J = 8.0$ Hz, 6F).

HRMS (ESI-TOF): calcd for $\text{C}_{15}\text{H}_{13}\text{F}_9\text{O}_2\text{Na}$ [$\text{M}+\text{Na}$]: 419.0664; found 419.0664.

O-(7,7,7-Trifluoro-6-(trifluoromethyl)heptyl) dimethylcarbamothioate (6l)

General procedure E.

Yield 140 mg (86%). Colorless oil.

Chromatography: hexane/EtOAc, 12/1.

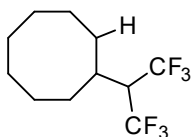
^1H NMR (300 MHz, Chloroform-*d*) δ 4.41 (t, $J = 6.5$ Hz, 2H), 3.32 (s, 3H), 3.07 (s, 3H), 2.93 – 2.71 (m, 1H), 1.93 – 1.63 (m, 4H), 1.62 – 1.32 (m, 4H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 188.3, 124.0 (qq, $J = 281.1, 3.9$ Hz), 71.2, 48.0 (hept, $J = 27.8$ Hz), 42.6, 37.6, 28.4, 26.9, 25.8, 23.78 – 23.45 (m).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -67.97 (d, $J = 8.4$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{11}\text{H}_{18}\text{F}_6\text{NOS}$ [$\text{M}+\text{H}$]: 326.1008; found 326.1006.

(1,1,1,3,3,3-Hexafluoropropan-2-yl)cyclooctane (6m)



General Procedure E.

Yield 104 mg (79%). Colorless liquid.

Chromatography: pentane.

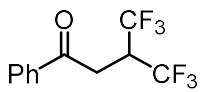
^1H NMR (300 MHz, Chloroform-*d*) δ 2.86 (heptd, $J = 9.0, 1.8$ Hz, 1H), 2.24 (t, $J = 9.8$ Hz, 1H), 1.54 (m, 14H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 124.4 (qq, $J = 282.6, 4.0$ Hz), 54.8 (hept, $J = 25.9$ Hz), 34.9, 31.2, 26.7, 26.2, 26.0.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -63.69 (d, $J = 9.3$ Hz).

MS (EI): 262 [M^+], 233, 111, 84, 69 [CF_3].

4,4,4-Trifluoro-1-phenyl-3-(trifluoromethyl)butan-1-one (8a)



General procedure F.

Yield 80 mg (59%). Colorless oil.

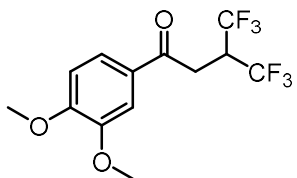
Chromatography: pentane/DCM, 5/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.99 (d, $J = 7.5$ Hz, 2H), 7.65 (t, $J = 7.5$ Hz, 1H), 7.52 (t, $J = 7.5$ Hz, 2H), 4.33 – 4.11 (m, 1H), 3.42 (d, $J = 5.4$ Hz, 2H).

^{13}C NMR (75 MHz, Chloroform-*d*) δ 192.83, 135.51, 134.32, 129.11, 128.38, 123.89 (qq, $J = 280.4, 3.7$ Hz), 43.09 (hept, $J = 29.3$ Hz), 32.54 (pent, $J = 1.7$ Hz), 29.86.

^{19}F NMR (282 MHz, Chloroform-*d*) δ -68.59 (d, $J = 8.4$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{11}\text{H}_8\text{F}_6\text{ONa}$ [$\text{M}+\text{Na}$]: 293.0372; found 293.0363.

1-(3,4-Dimethoxyphenyl)-4,4,4-trifluoro-3-(trifluoromethyl)butan-1-one (8b)

General procedure F.

Yield 142 mg (86%). Yellow oil.

Chromatography: Hexane/EtOAc, 5/1.

^1H NMR (300 MHz, Chloroform-*d*) δ 7.60 (dd, $J = 8.4, 2.1$ Hz, 1H), 7.53 (d, $J = 2.1$ Hz, 1H), 6.92 (d, $J = 8.4$ Hz, 1H), 4.27 – 4.09 (m, 1H), 3.95 (s, 3H), 3.93 (s, 3H), 3.38 (d, $J = 5.4$ Hz, 2H).

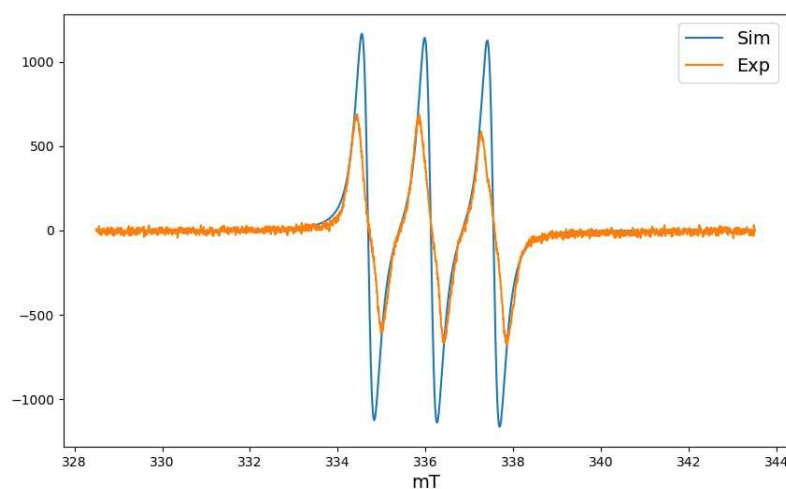
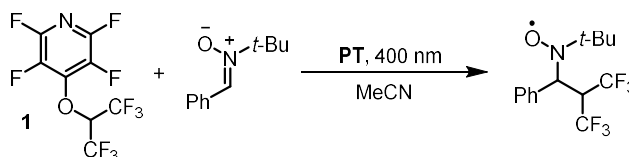
^{13}C NMR (75 MHz, Chloroform-*d*) δ 191.3, 154.4, 149.5, 128.7, 123.9 (qq, $J = 280.5, 3.7$ Hz), 123.1, 110.5, 110.3, 56.3, 56.2, 43.2 (hept, $J = 29.1$ Hz), 32.0 (s).

^{19}F NMR (282 MHz, Chloroform-*d*) δ -68.51 (d, $J = 8.5$ Hz).

HRMS (ESI-TOF): calcd for $\text{C}_{13}\text{H}_{12}\text{F}_6\text{O}_3\text{Na}$ [$\text{M}+\text{Na}$]: 353.0583; found 353.0579.

EPR study

A screw test tube was evacuated, refilled with argon, and charged with reagent **1** (32 mg, 0.1 mmol), *N*-*tert*-butyl-1-phenylmethanimine oxide (18 mg, 0.1 mmol) and 12-phenyl-12H-benzo[*b*]phenothiazine (**PT**) (1 mg, 0.01 mmol), followed by addition of MeCN (0.5 mL). From the resulting solution, an aliquot (ca. 10 μ l) was taken and placed into an EPR tube, which was then irradiated with 400 nm LED matrix (80W) for 180 seconds. The EPR spectrum was immediately recorded at 298 K on EPR spectrometer SPINSCAN X (ADANI).



The X-band EPR spectrum of the nitroxyl radical (orange line). Simulated EPR spectrum (blue line) based on hyperfine coupling constants of $a_N = 14.28$ G (g -factor = 2.0056).

Experiment parameters:

Center-Field: 336.0 mT

Width: 15 G

Points: 6000

Modulation Amplitude: 100 μ T

Modulation Frequency: 9.432985 GHz

Microwave Power: 31.6 mW

Time constant: 0.015 s

Cyclic Voltammetry

Voltammetric studies were carried out using potentiostat P30JM with a scan rates of $0.1 \text{ V}\cdot\text{s}^{-1}$ in a temperature-controlled ($25 \text{ }^\circ\text{C}$) glass cell ($V = 10 \text{ mL}$) under an argon atmosphere. Software *iR* compensation using ferrocene ($R = 1362 \Omega$) was used. A glassy carbon disk ($d = 2.9 \text{ mm}$) was used as the working electrode (carefully polished before each measurement). A saturated calomel electrode (SCE) separated from the solution being studied by a salt bridge filled with the supporting electrolyte ($0.1\text{M Et}_4\text{NClO}_4$ in DMSO) was used as the reference electrode. A platinum wire ($S = 3 \text{ cm}^2$) was used as the counter electrode. Experiment was performed with the concentration of compound **1** of 1 mM .

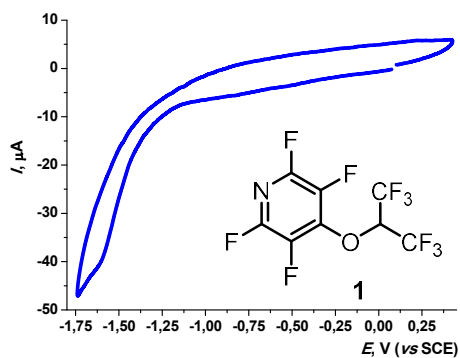


Figure S1. Compound **1** (initial cathodic scan).

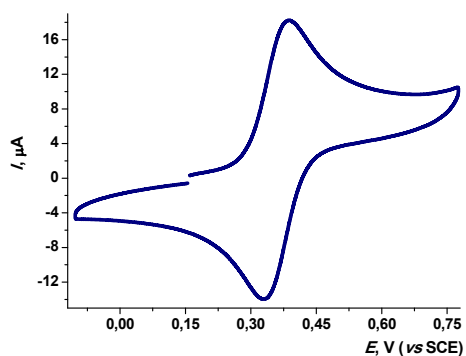
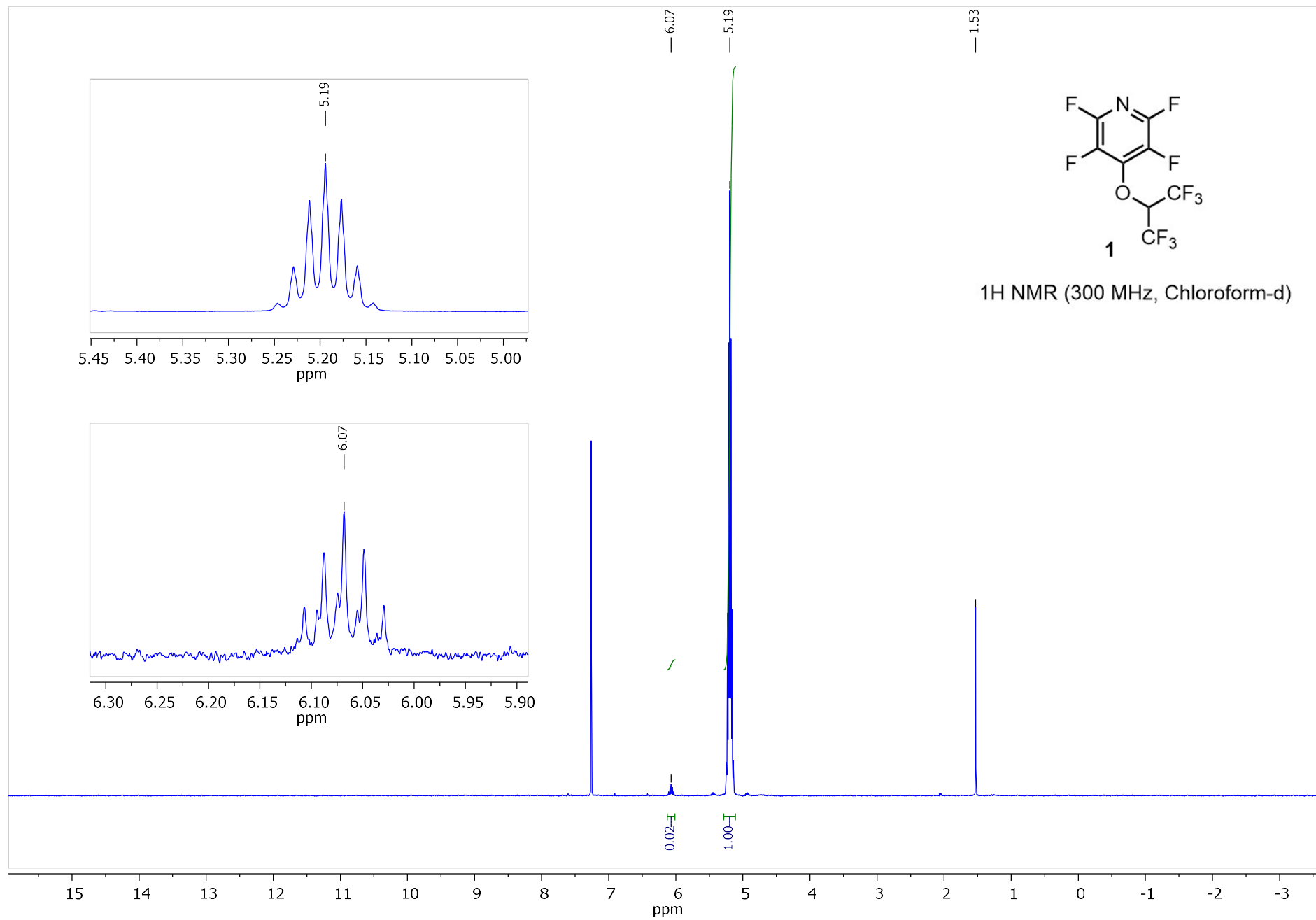


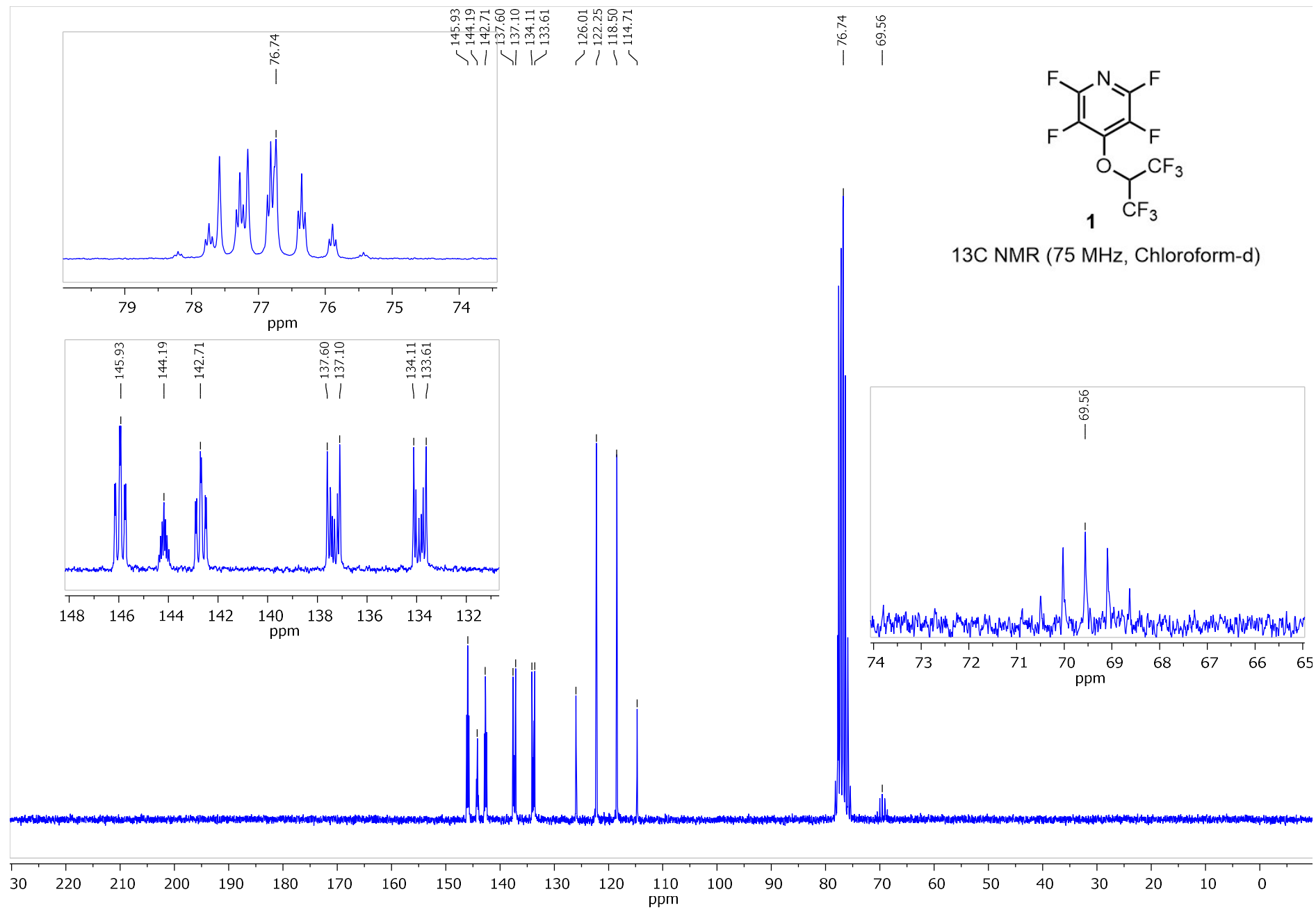
Figure S2. Ferrocene (initial anodic scan).

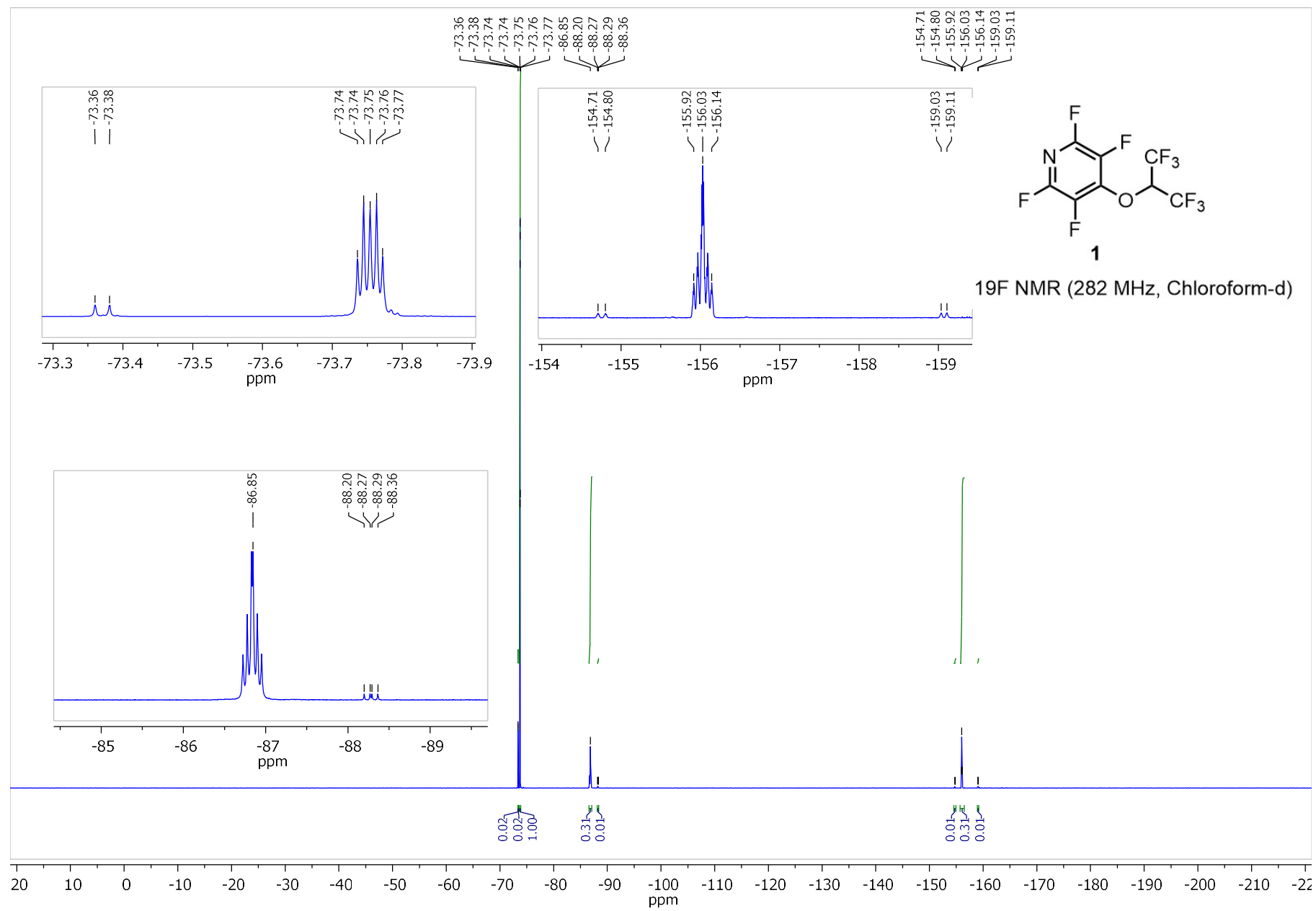
Compound **1** showed irreversible cathodic peak at $-1.65 \text{ V (vs. SCE)}$.

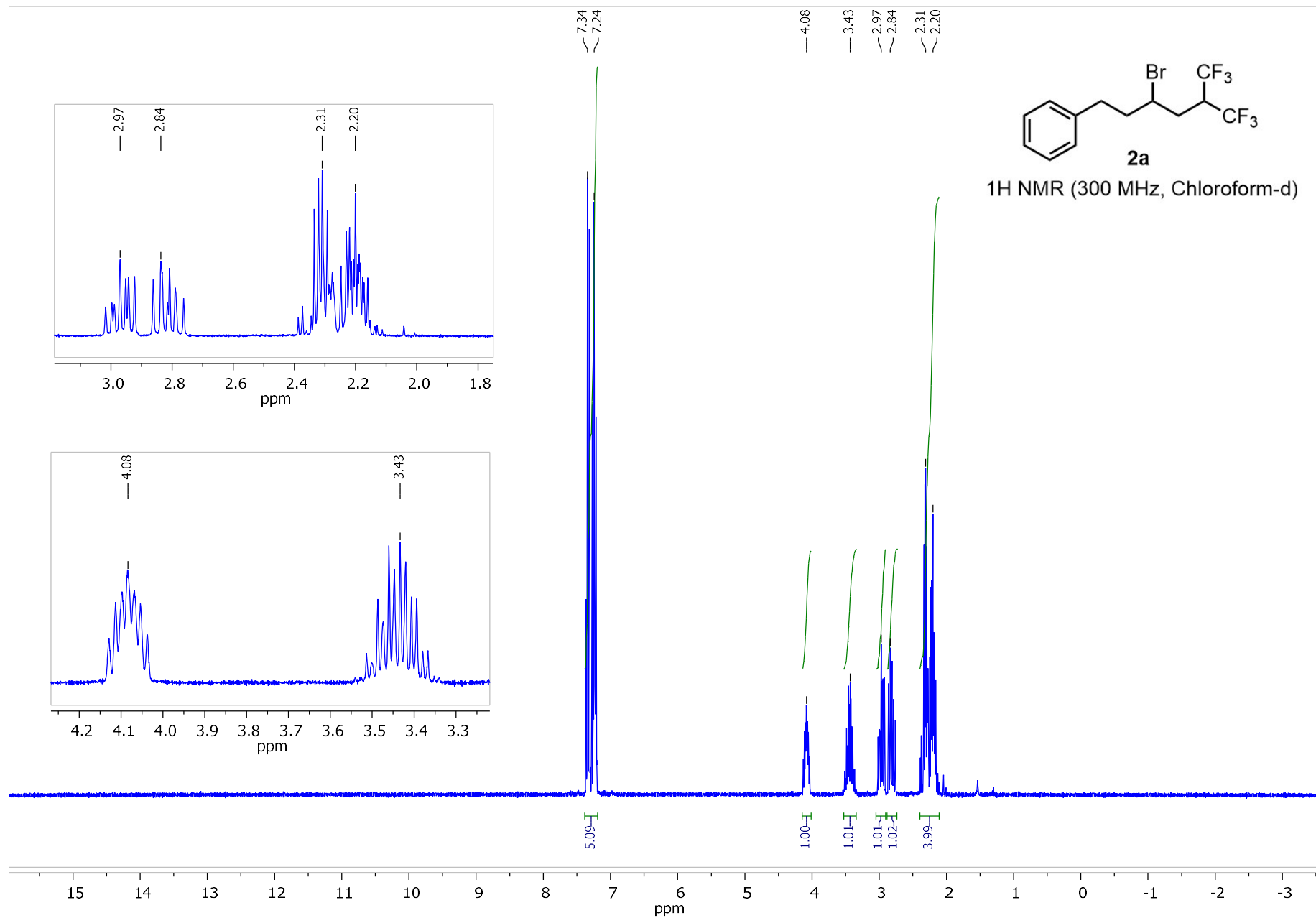
References

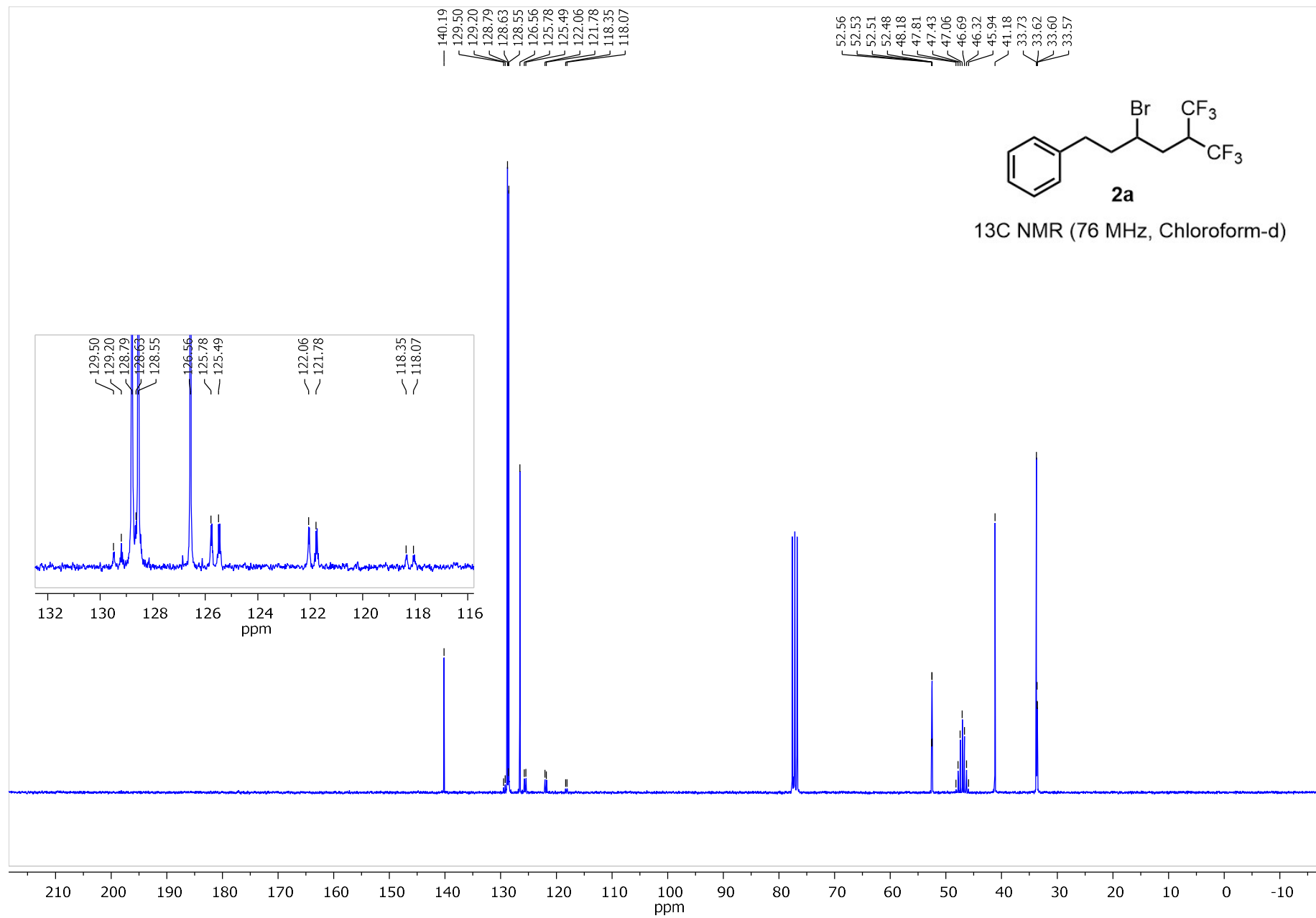
1. Yamasaki, Y.; Kumagai, T.; Kanno, S.; Kakiuchi, F.; Kochi, T., *J. Org. Chem.* **2018**, *83*, 9322–9333.
2. Scheller, M. E.; Frei, B., *Helv. Chim. Acta* **1986**, *69*, 44–52.
3. Anson, C. E.; Malkov, A. V.; Roe, C.; Sandoe, E. J.; Stephenson, G. R., *Eur. J. Org. Chem.* **2008**, *2008*, 196–213.
4. Yoshida, M.; Higuchi, M.; Shishido, K., *Org. Lett.* **2009**, *11*, 4752–4755.
5. Lipshutz, B. H.; Ghorai, S.; Leong, W. W. Y., *J. Org. Chem.* **2009**, *74*, 2854–2857.
6. Saha, M.; Li, X.; Collett, N. D.; Carter, R. G., *J. Org. Chem.* **2016**, *81*, 5963–5980.
7. Courtens, C.; Risseuw, M.; Caljon, G.; Maes, L.; Martin, A.; Van Calenbergh, S., *Bioorg. Med. Chem.* **2019**, *27*, 729–747.
8. He, D.; Guo, Y.; Chen, Q.-Y.; Yang, H.; Lv, T., *J. Fluorine Chem.* **2019**, *222–223*, 1–7.
9. Sharma, H.; Santra, S.; Debnath, J.; Antonio, T.; Reith, M.; Dutta, A., *Bioorg. Med. Chem.* **2014**, *22*, 311–324.
10. Bourgeois, M. J.; Montaudon, E.; Maillard, B., *Bull. Soc. Chim. Belg.* **1988**, *97*, 255–262.
11. Siu, J. C.; Parry, J. B.; Lin, S., *J. Am. Chem. Soc.* **2019**, *141*, 2825–2831.
12. Lin, S.; Song, C.-X.; Cai, G.-X.; Wang, W.-H.; Shi, Z.-J., *J. Am. Chem. Soc.* **2008**, *130*, 12901–12903.
13. Levin, V. V.; Dilman, A. D., *Chem. Commun.* **2021**, *57*, 749–752.
14. Warsitz, M.; Doye, S., *Chem. Eur. J.* **2020**, *26*, 15121–15125.
15. Coolen, H. K. A. C.; Meeuwis, J. A. M.; van Leeuwen, P. W. M. N.; Nolte, R. J. M., *J. Am. Chem. Soc.* **1995**, *117*, 11906–11913.
16. Voutyritsa, E.; Nikitas, N. F.; Apostolopoulou, M. K.; Gerogiannopoulou, A. D. D.; Kokotos, C. G., *Synthesis* **2018**, *50*, 3395–3401.
17. Edwards, G. A.; Culp, P. A.; Chalker, J. M., *Chem. Commun.* **2015**, *51*, 515–518.
18. Fan, B.-Z.; Hiasa, H.; Lv, W.; Brody, S.; Yang, Z.-Y.; Aldrich, C.; Cushman, M.; Liang, J.-H., *Eur. J. Med. Chem.* **2020**, *193*, 112222.
19. Cheung, W.-H.; Zheng, S.-L.; Yu, W.-Y.; Zhou, G.-C.; Che, C.-M., *Org. Lett.* **2003**, *5*, 2535–2538.
20. Matsumoto, A.; Maeda, N.; Maruoka, K., *J. Am. Chem. Soc.* **2023**, *145*, 20344–20354.
21. Fedorov, O. V.; Kosobokov, M. D.; Levin, V. V.; Struchkova, M. I.; Dilman, A. D., *J. Org. Chem.* **2015**, *80*, 5870–5876.
22. Talipov, M. R.; Navale, T. S.; Hossain, M. M.; Shukla, R.; Ivanov, M. V.; Rathore, R., *Angew. Chem. Int. Ed.* **2017**, *56*, 266–269.
23. Zemtsov, A. A.; Lunkov, S. S.; Levin, V. V.; Dilman, A. D., *Eur. J. Org. Chem.* **2021**, *2021*, 1007–1010.
24. Lux, M.; Klusmann, M., *Org. Lett.* **2020**, *22*, 3697–3701.

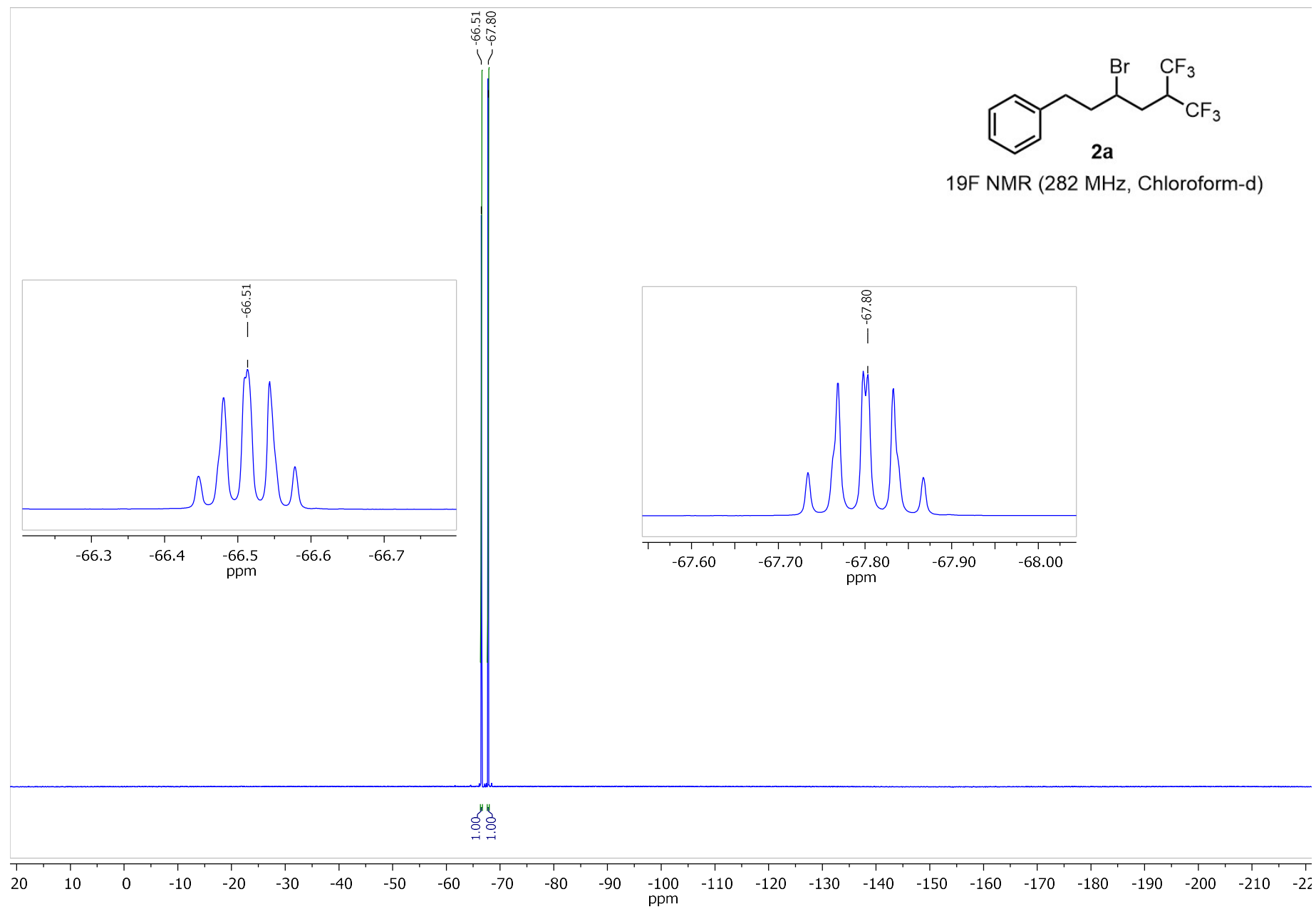


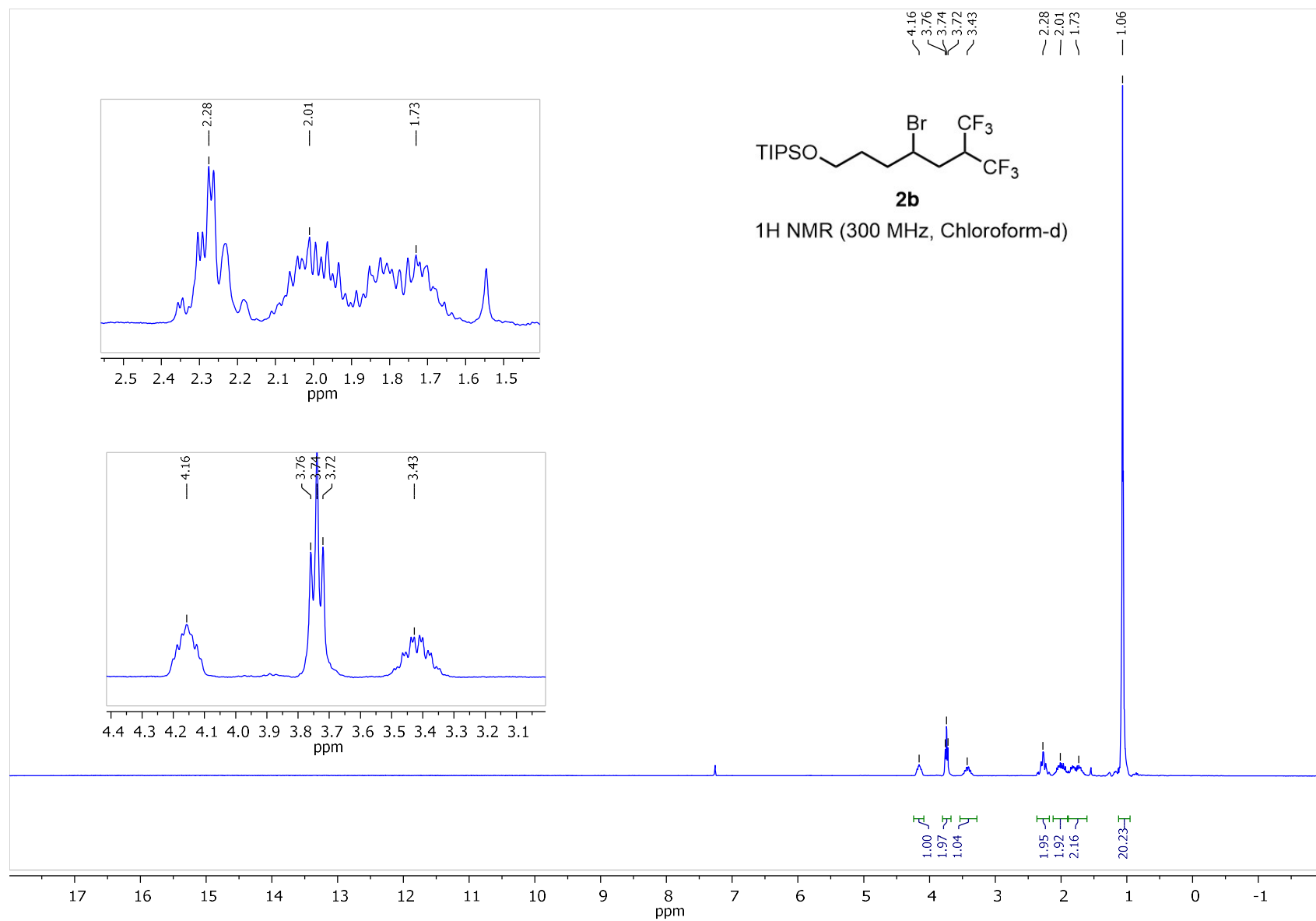


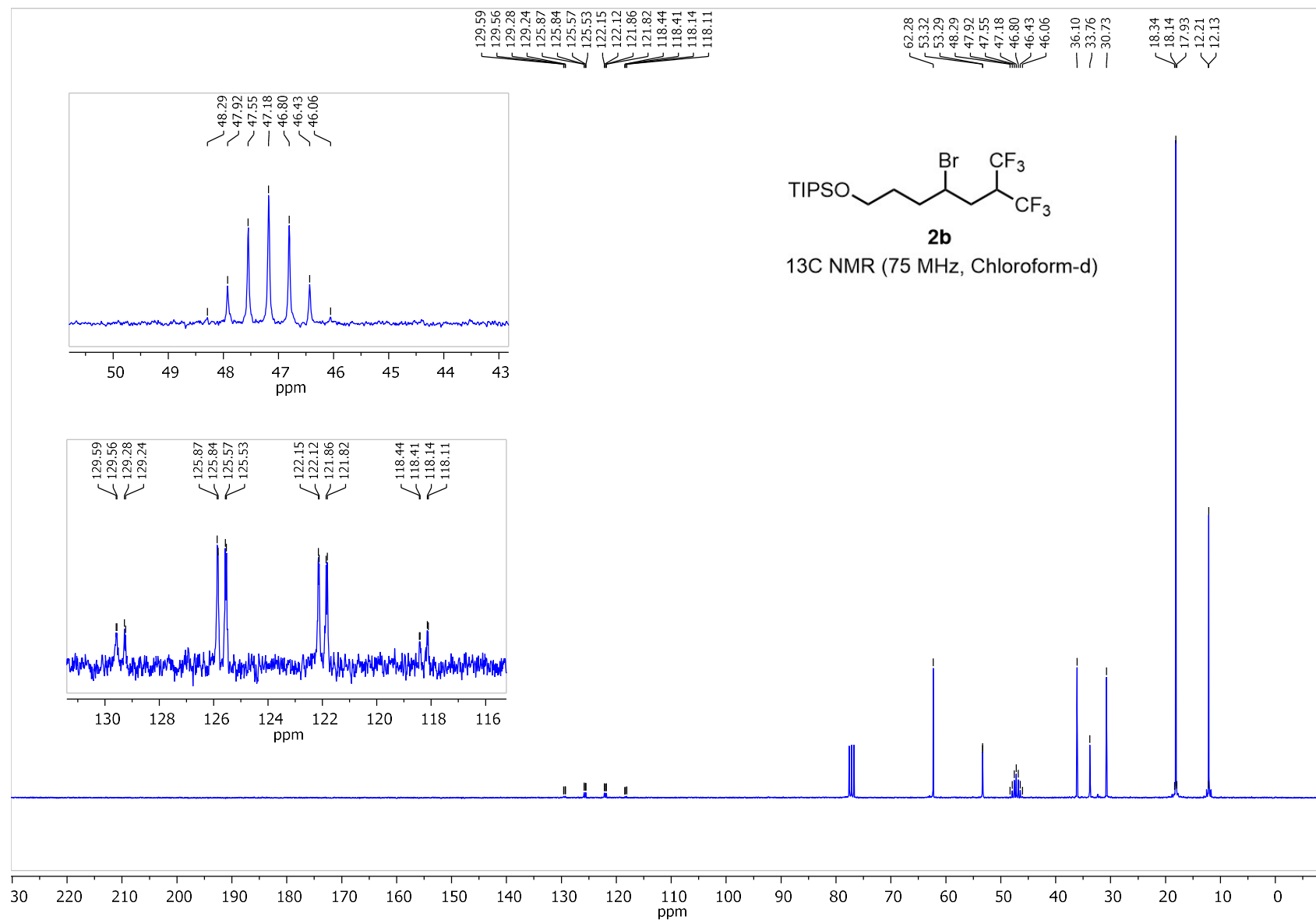


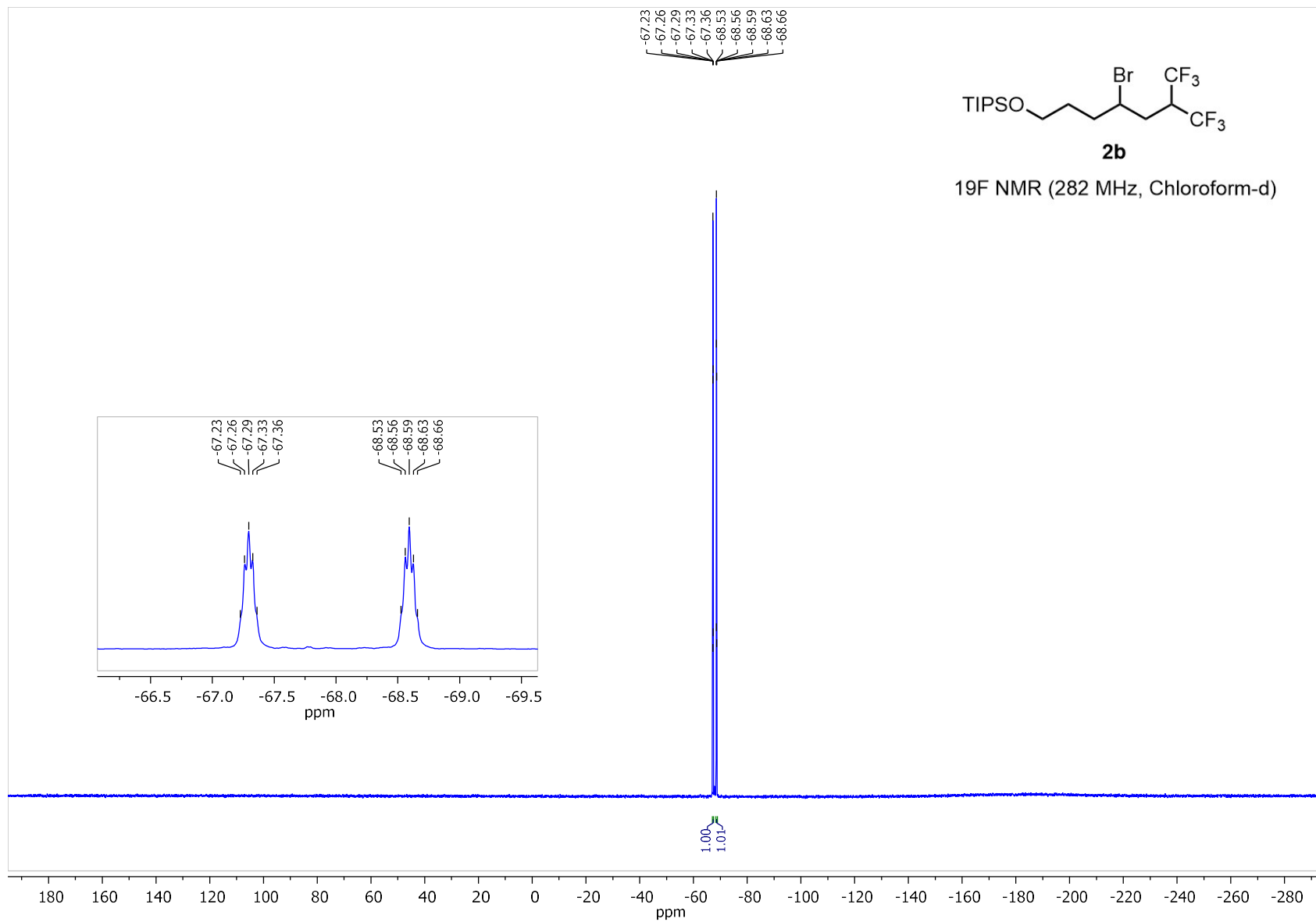


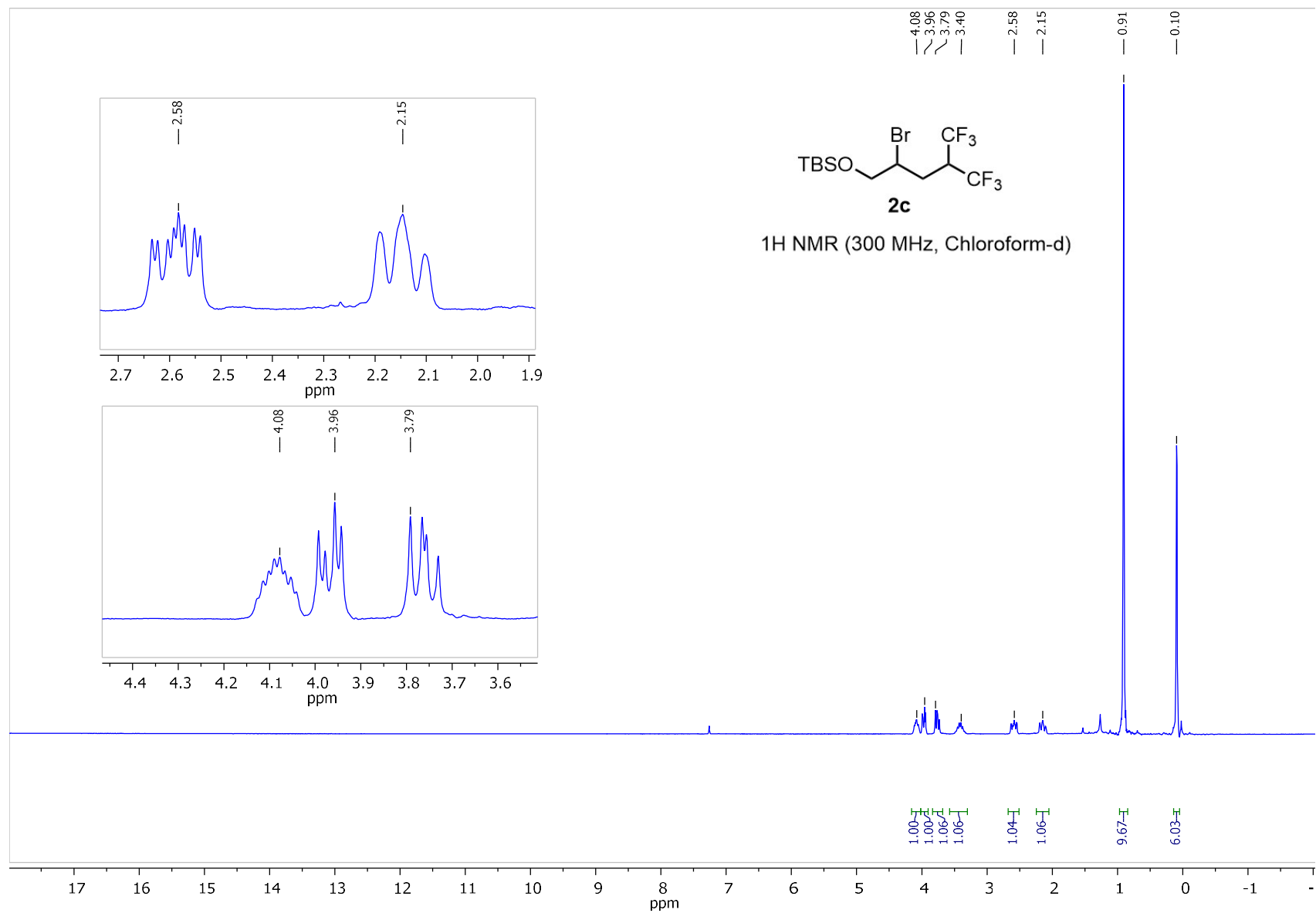


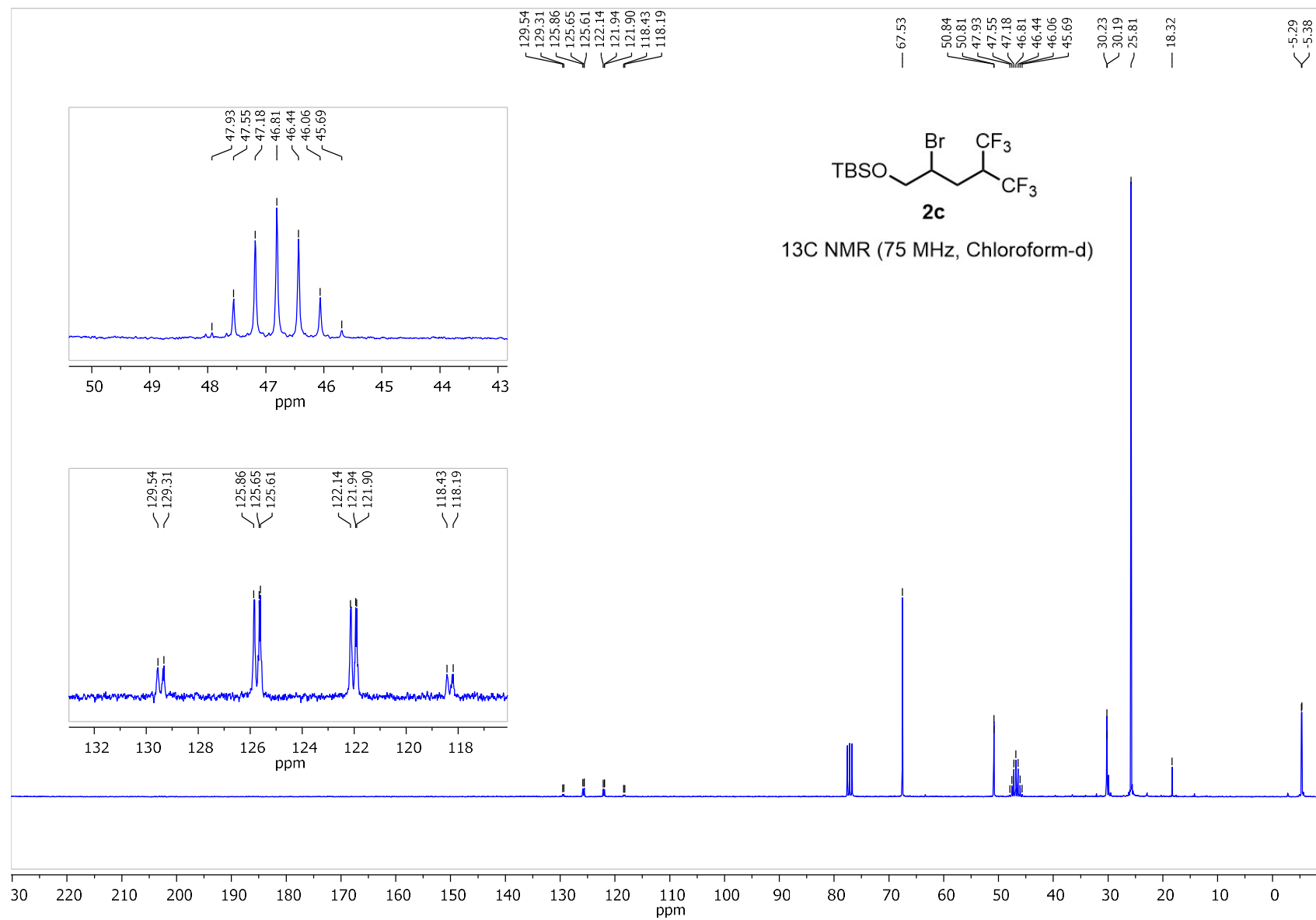


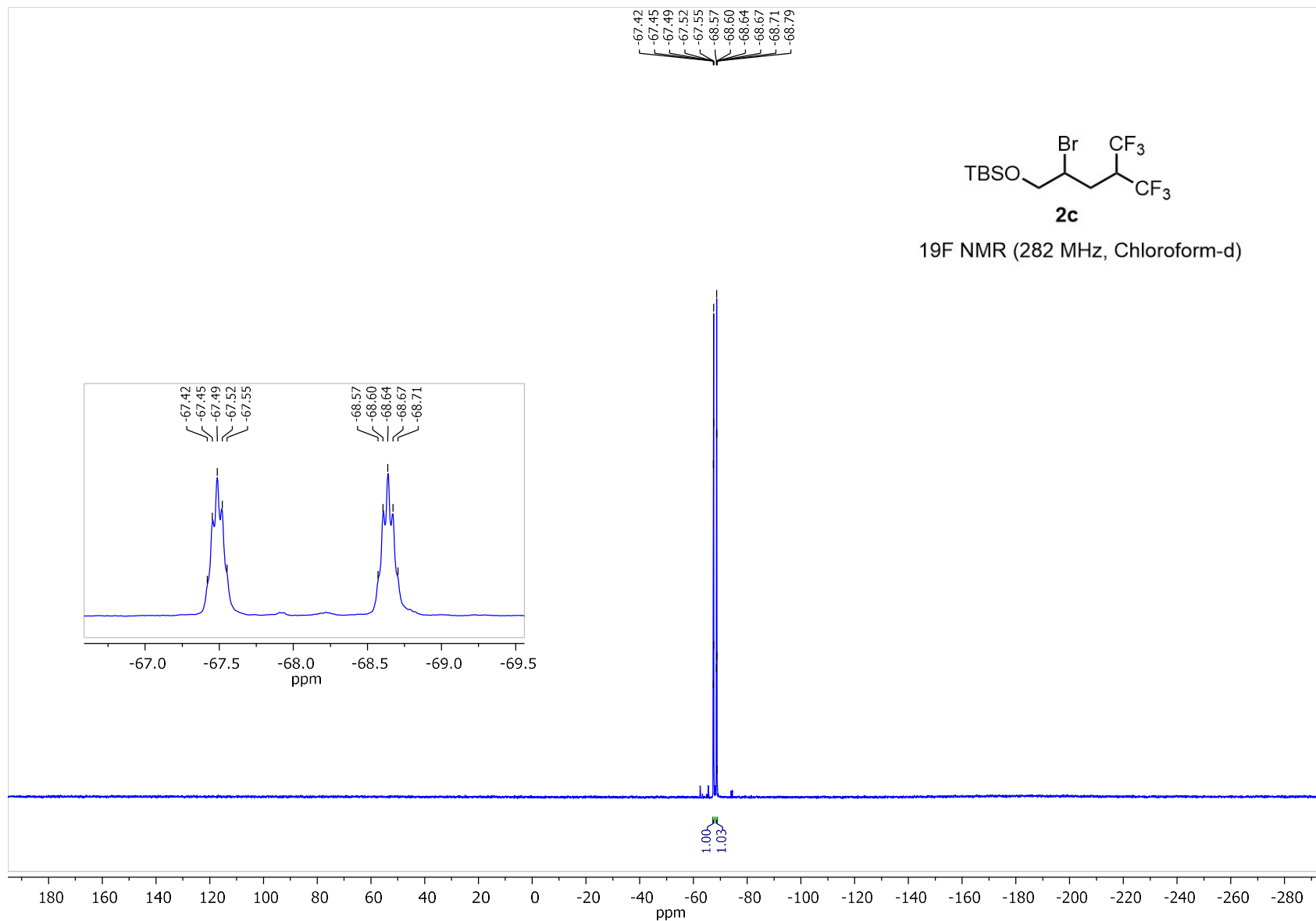


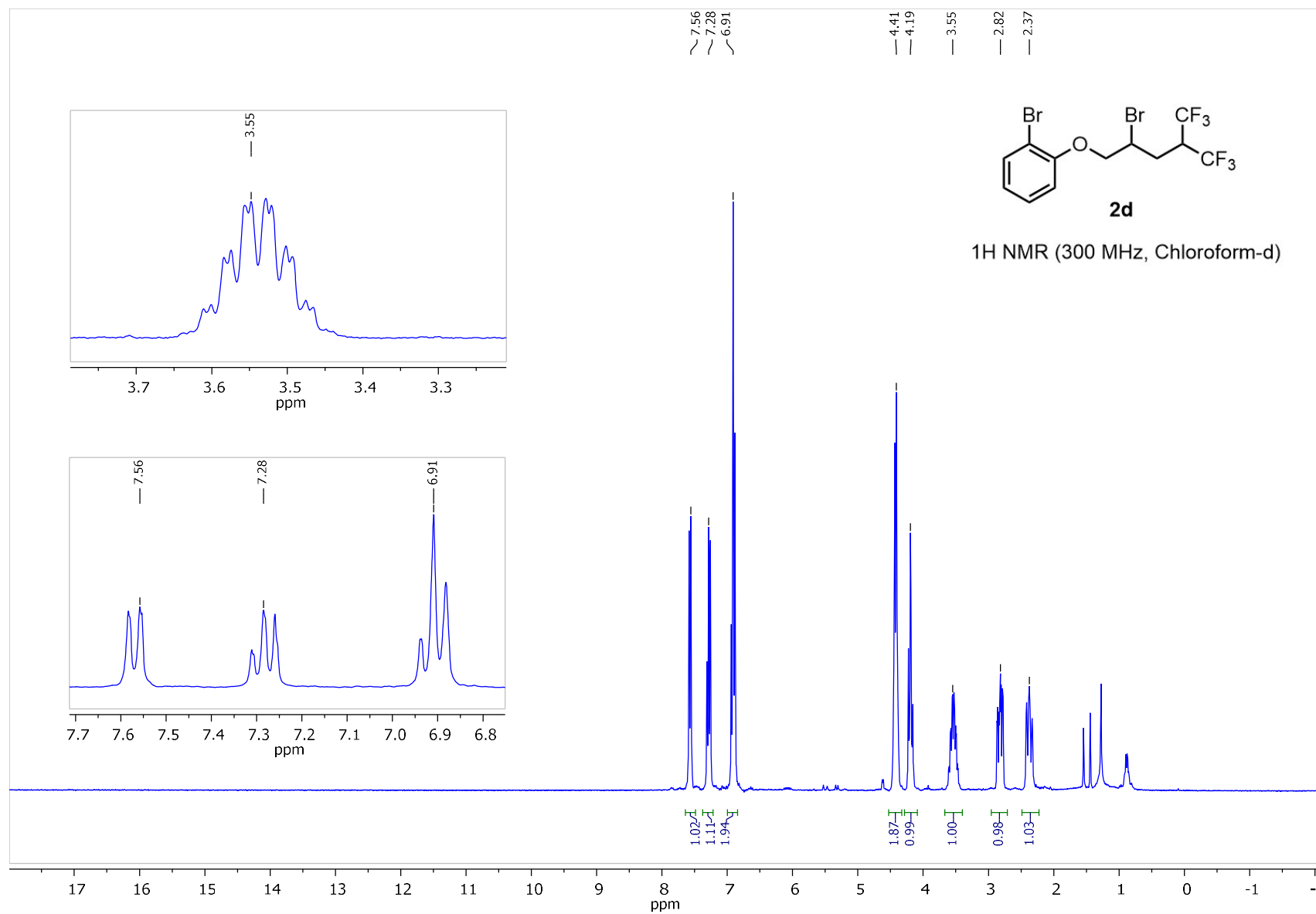


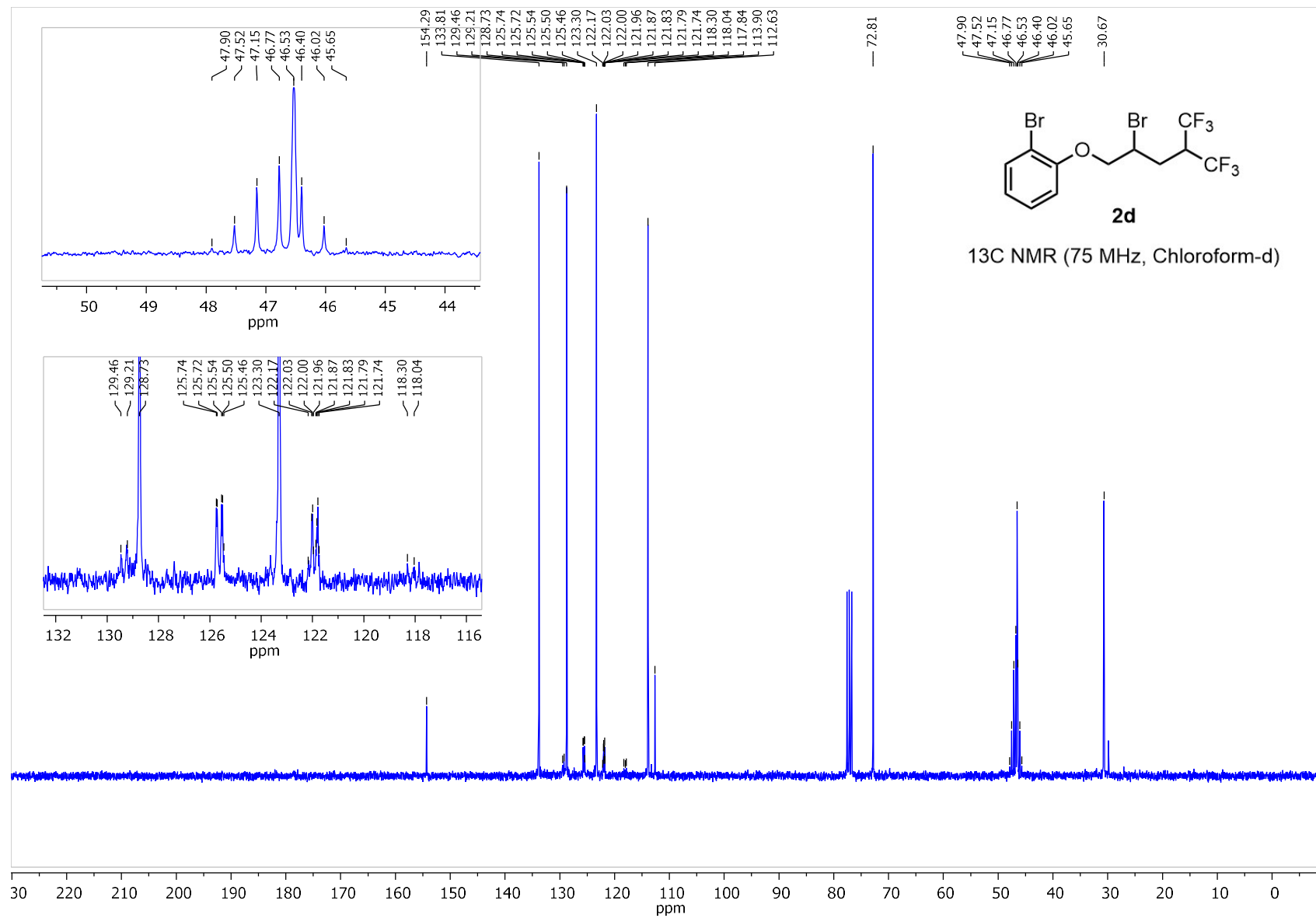


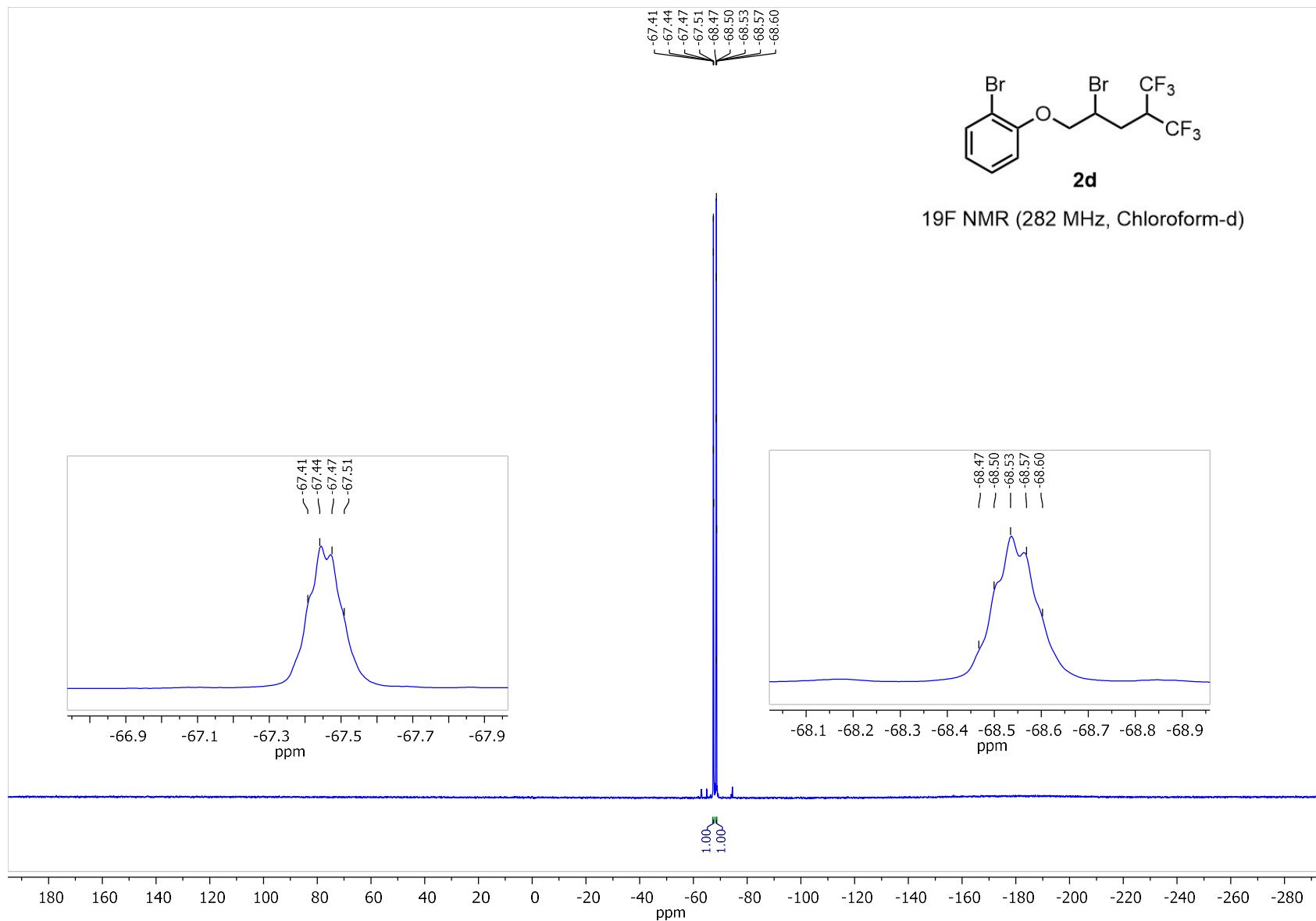


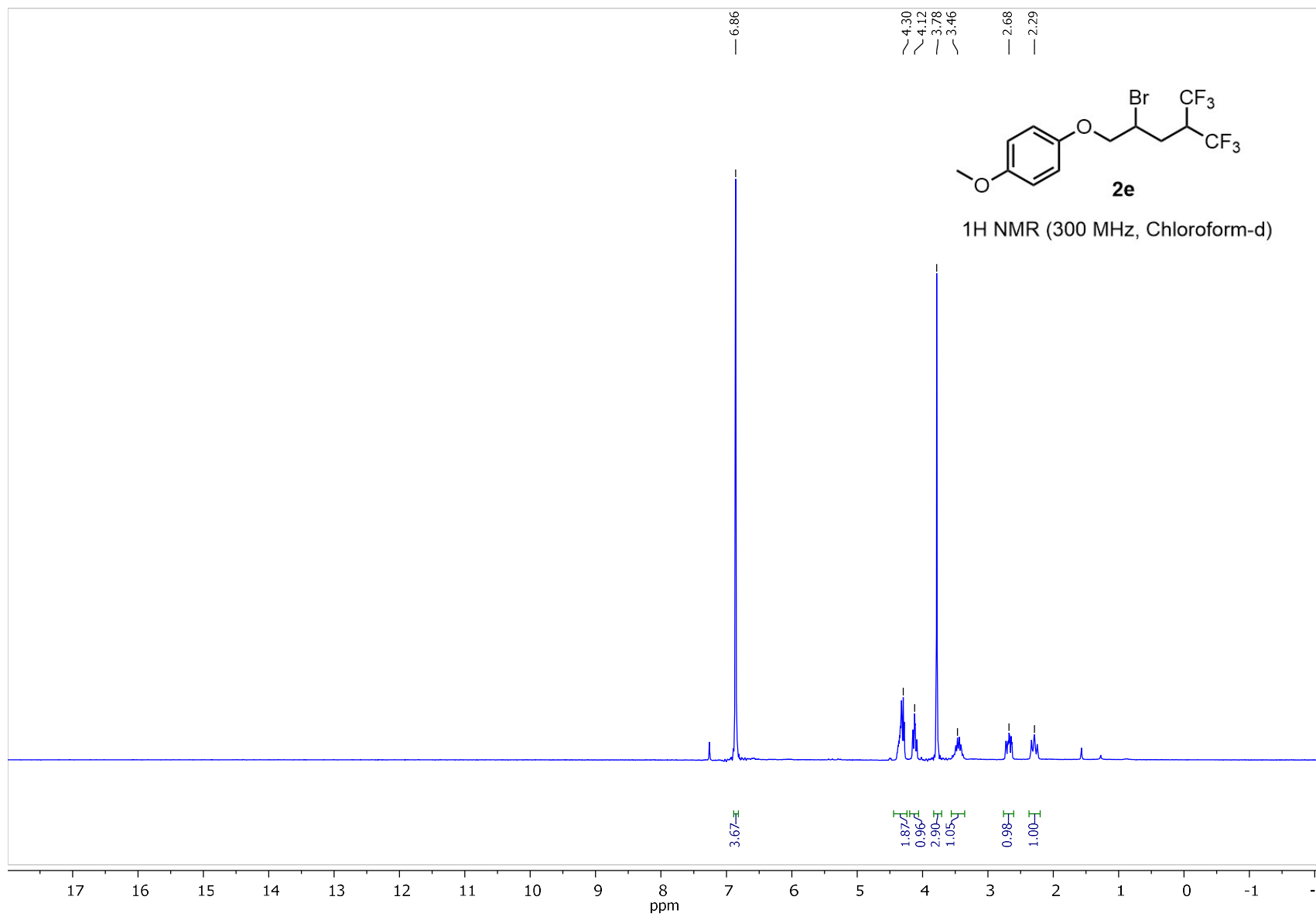


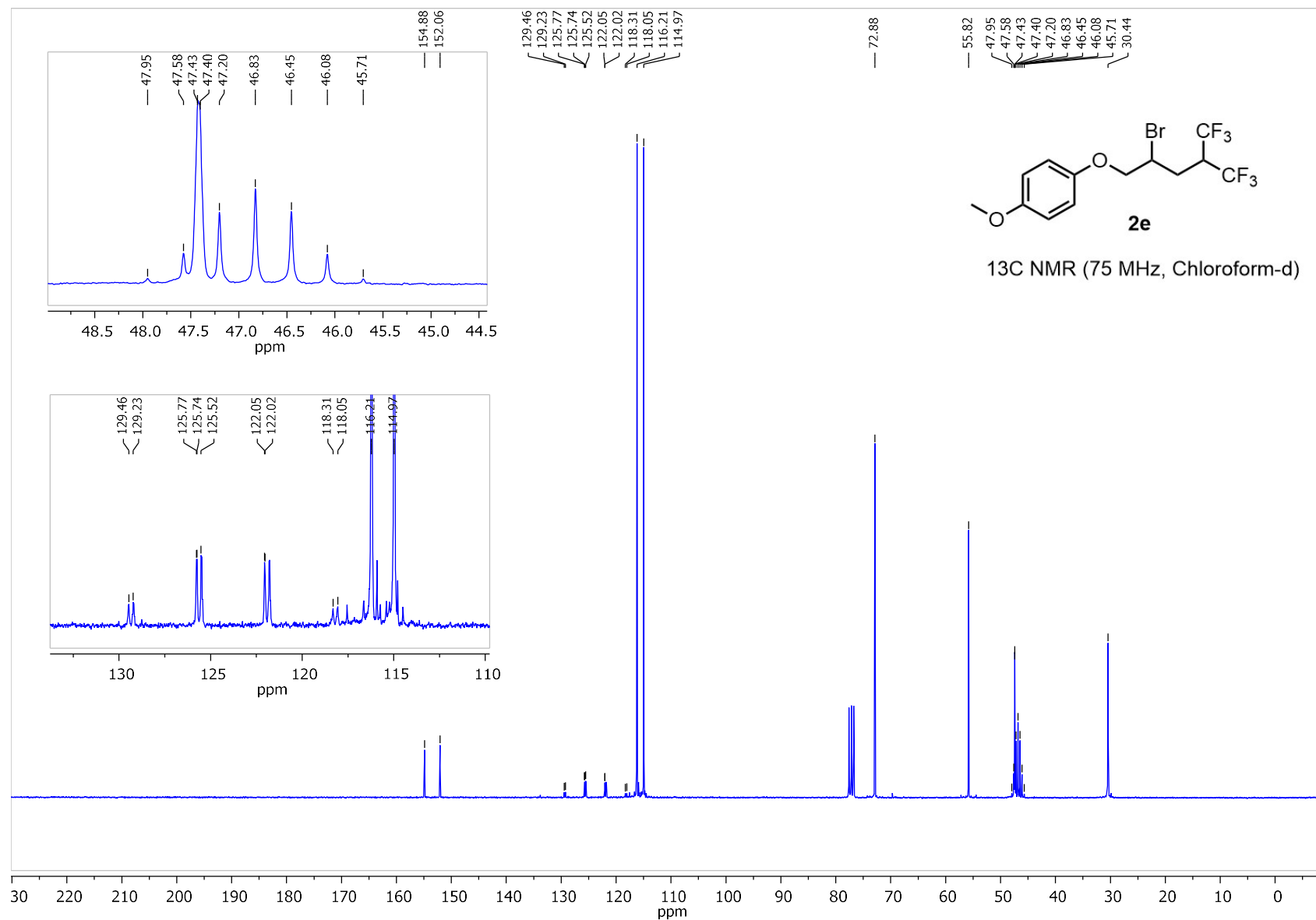


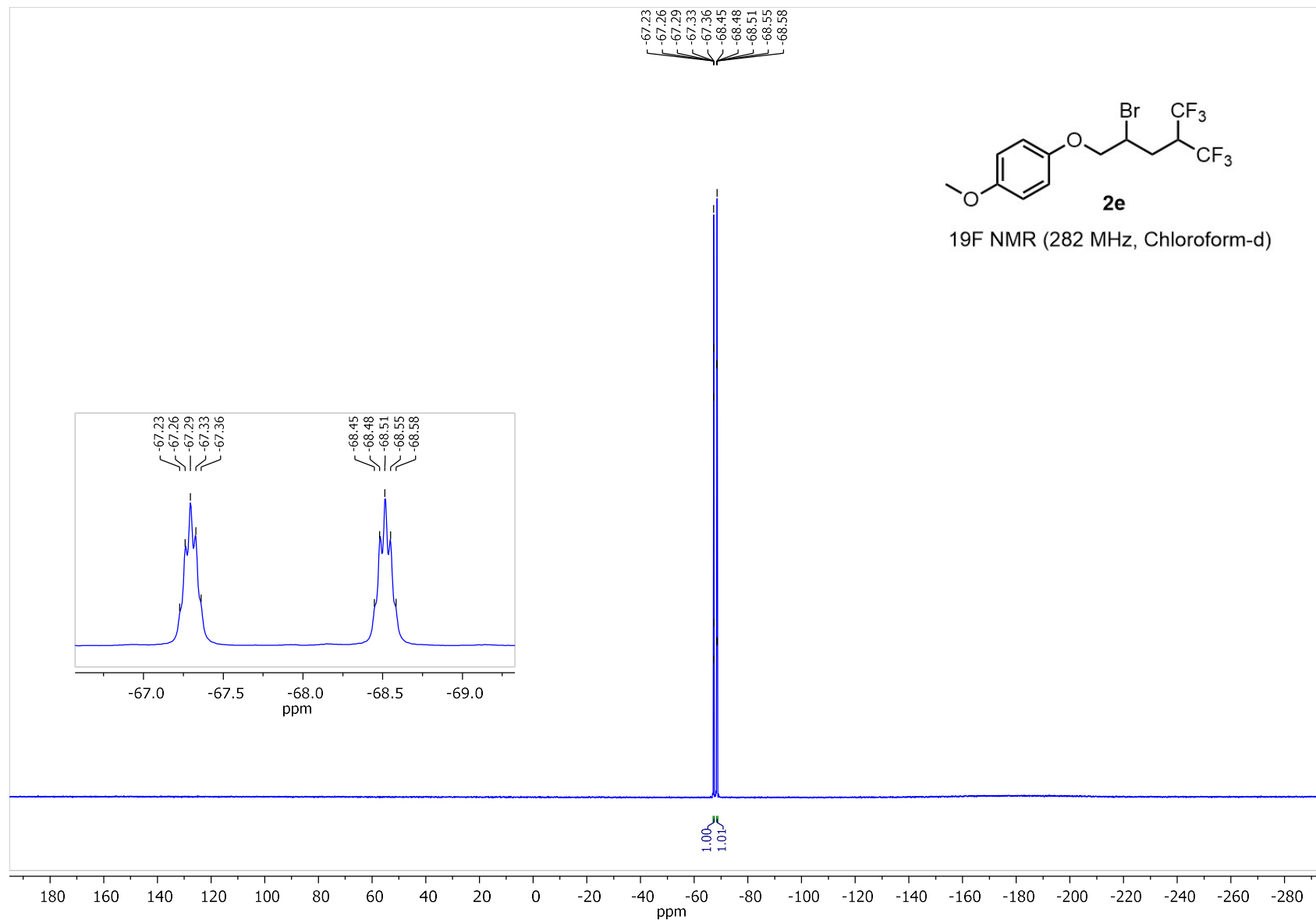


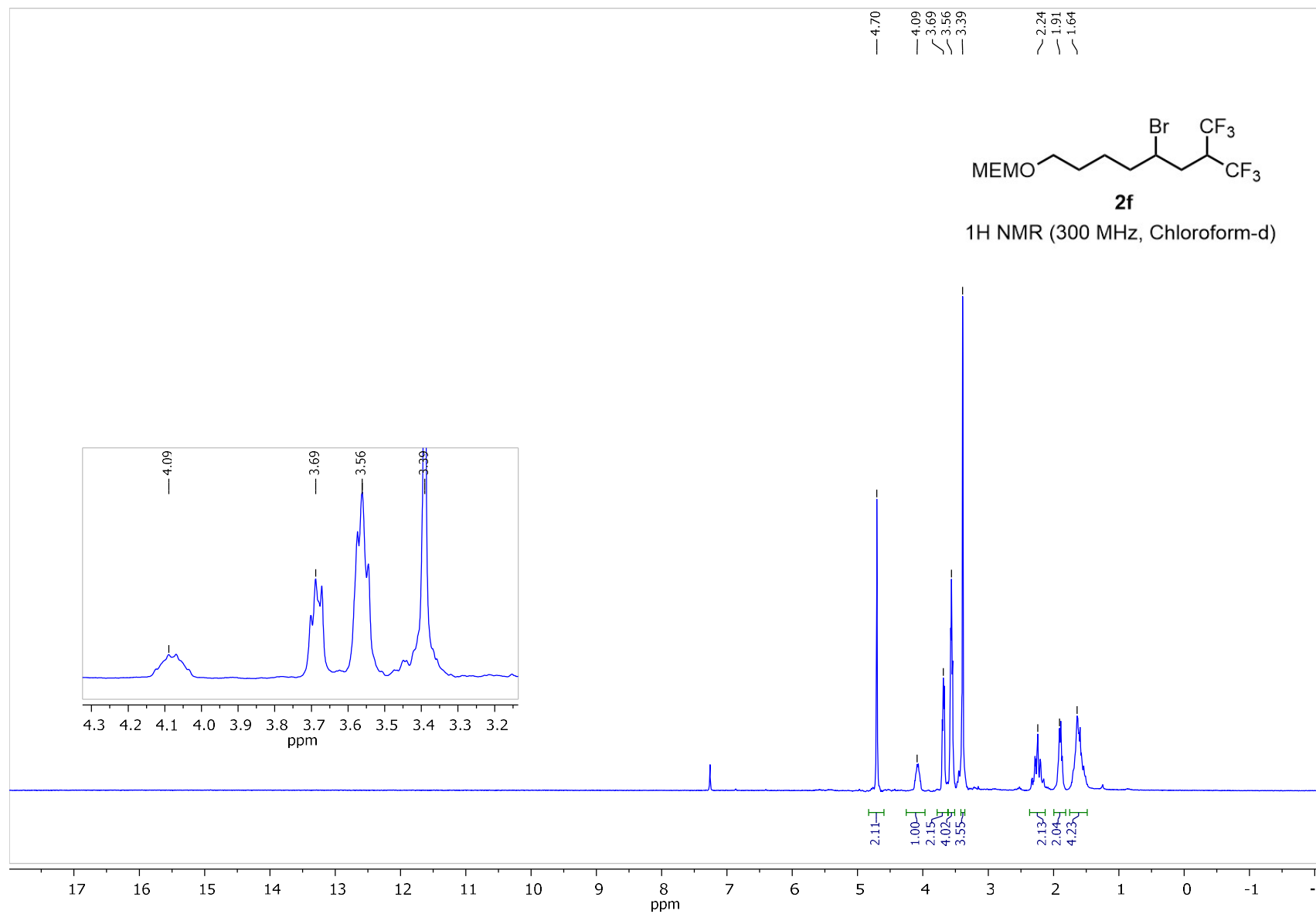


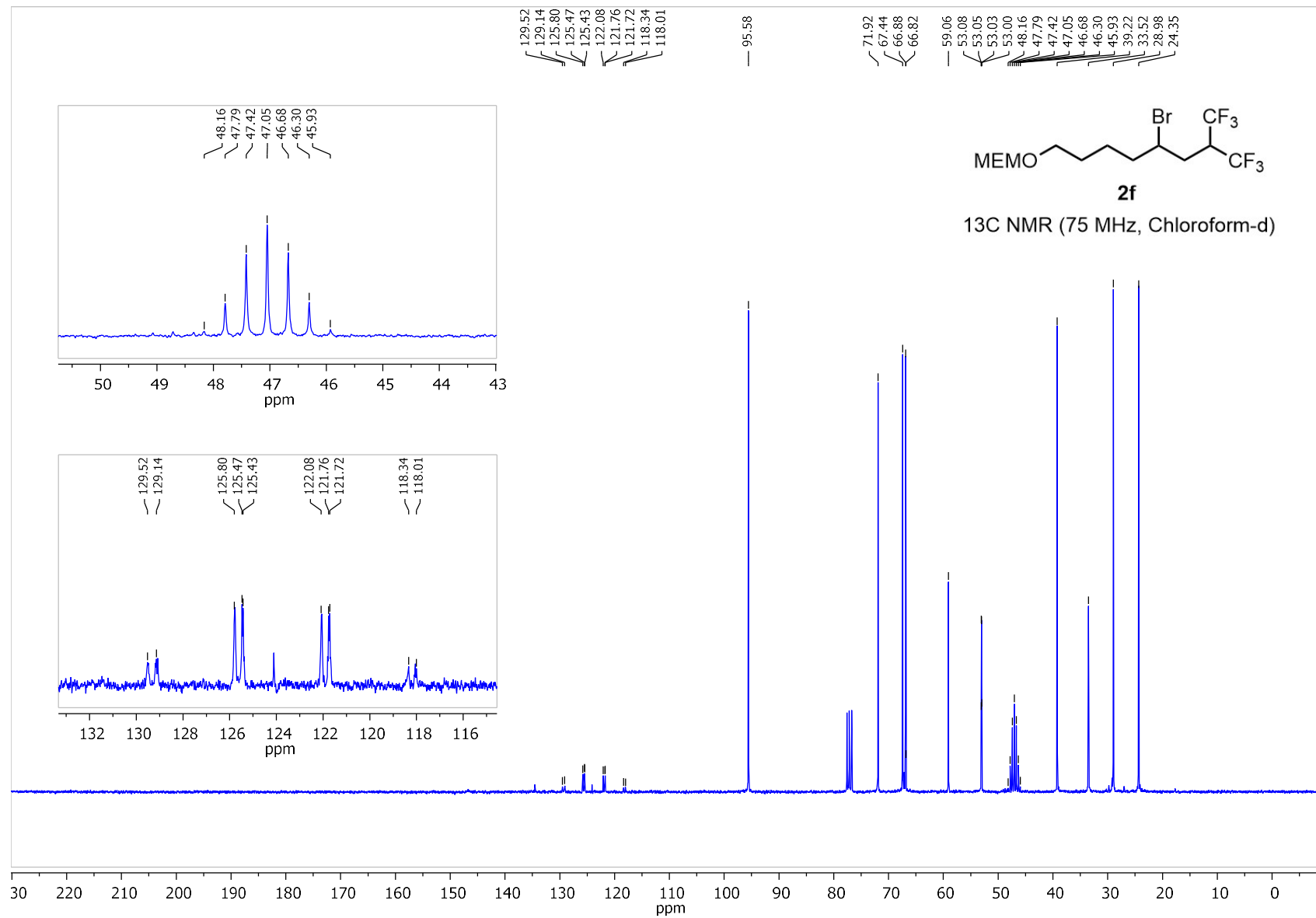


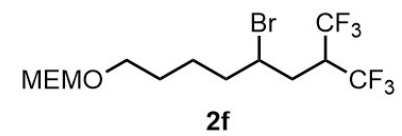




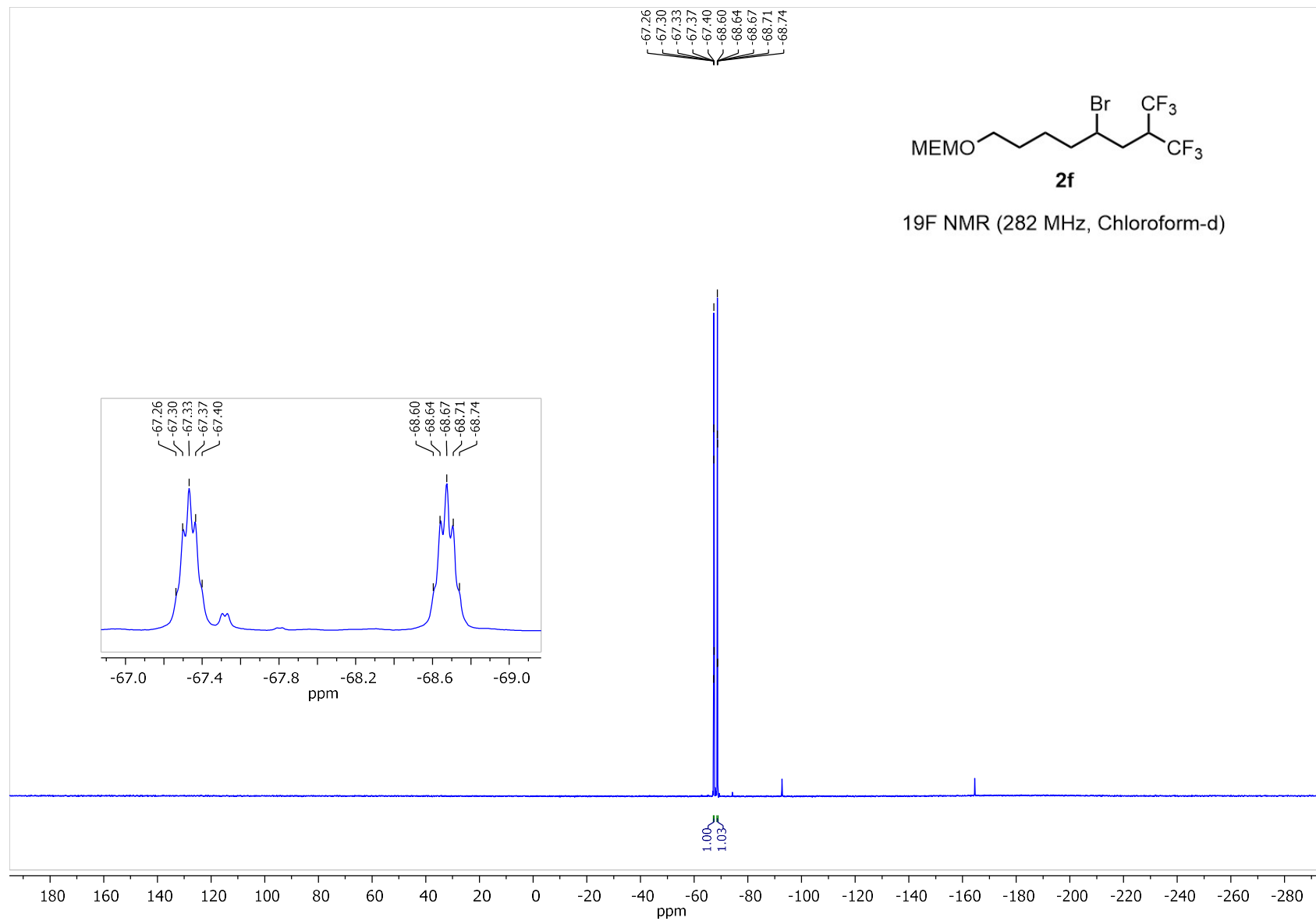


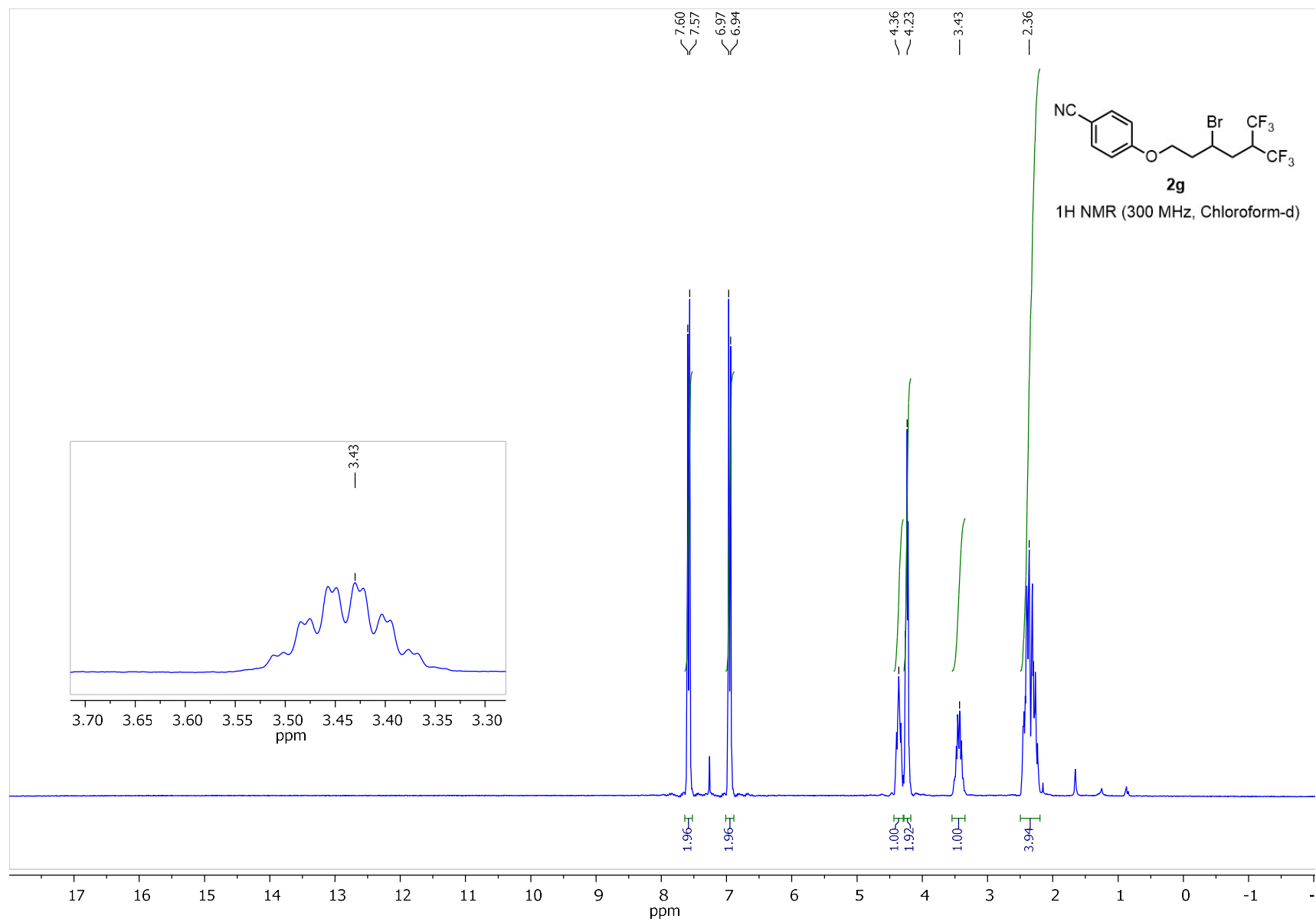


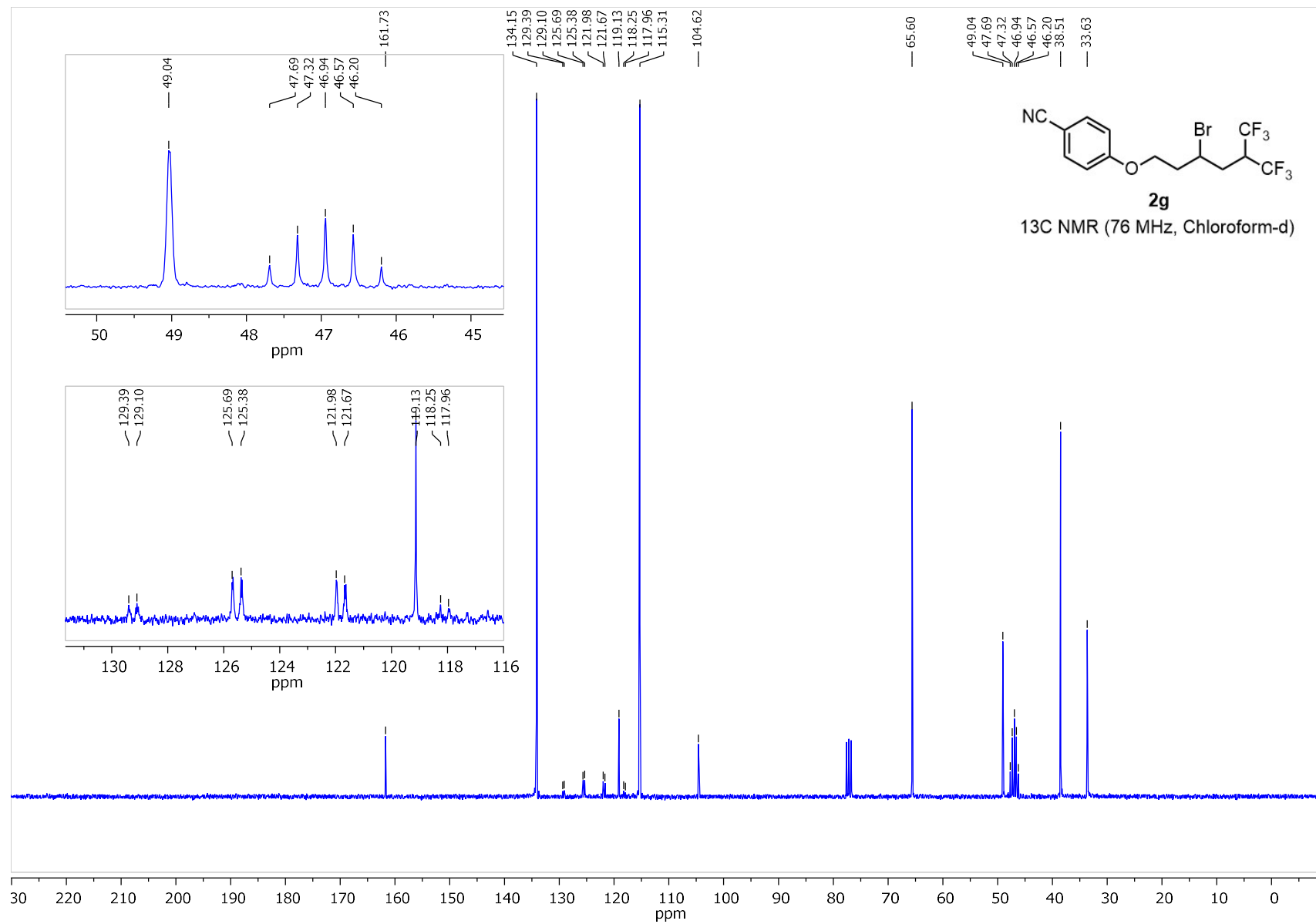


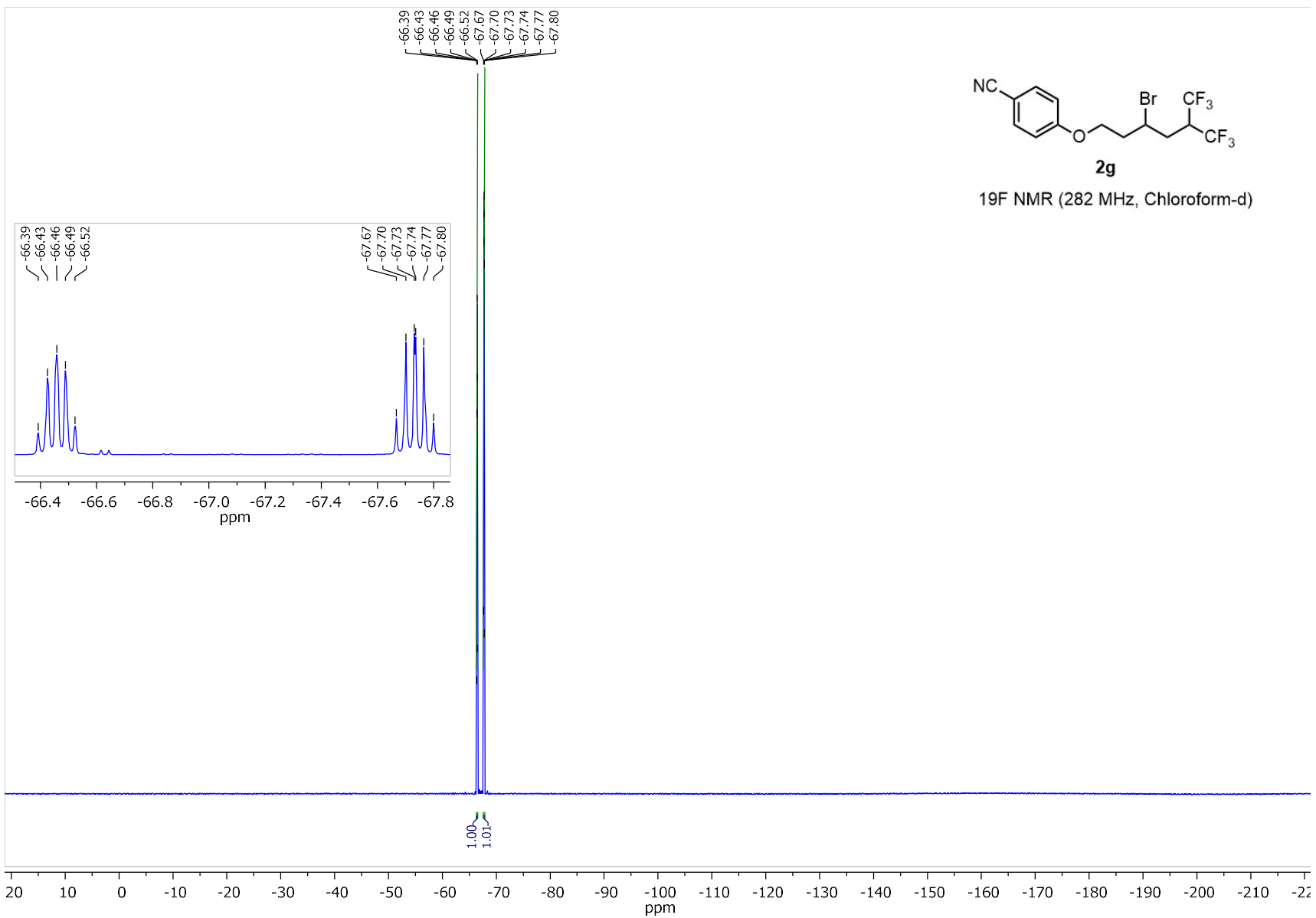


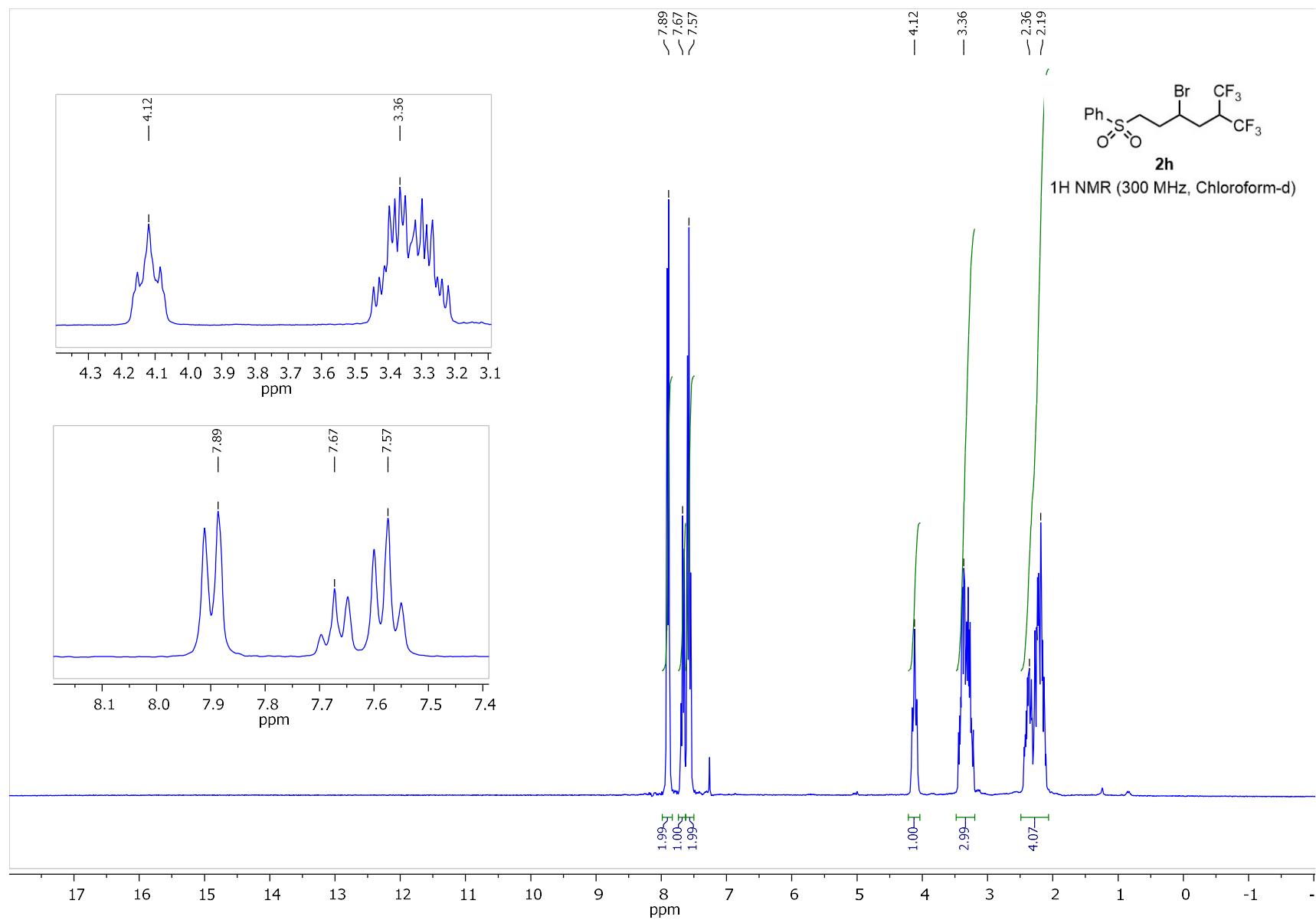
19F NMR (282 MHz, Chloroform-d)

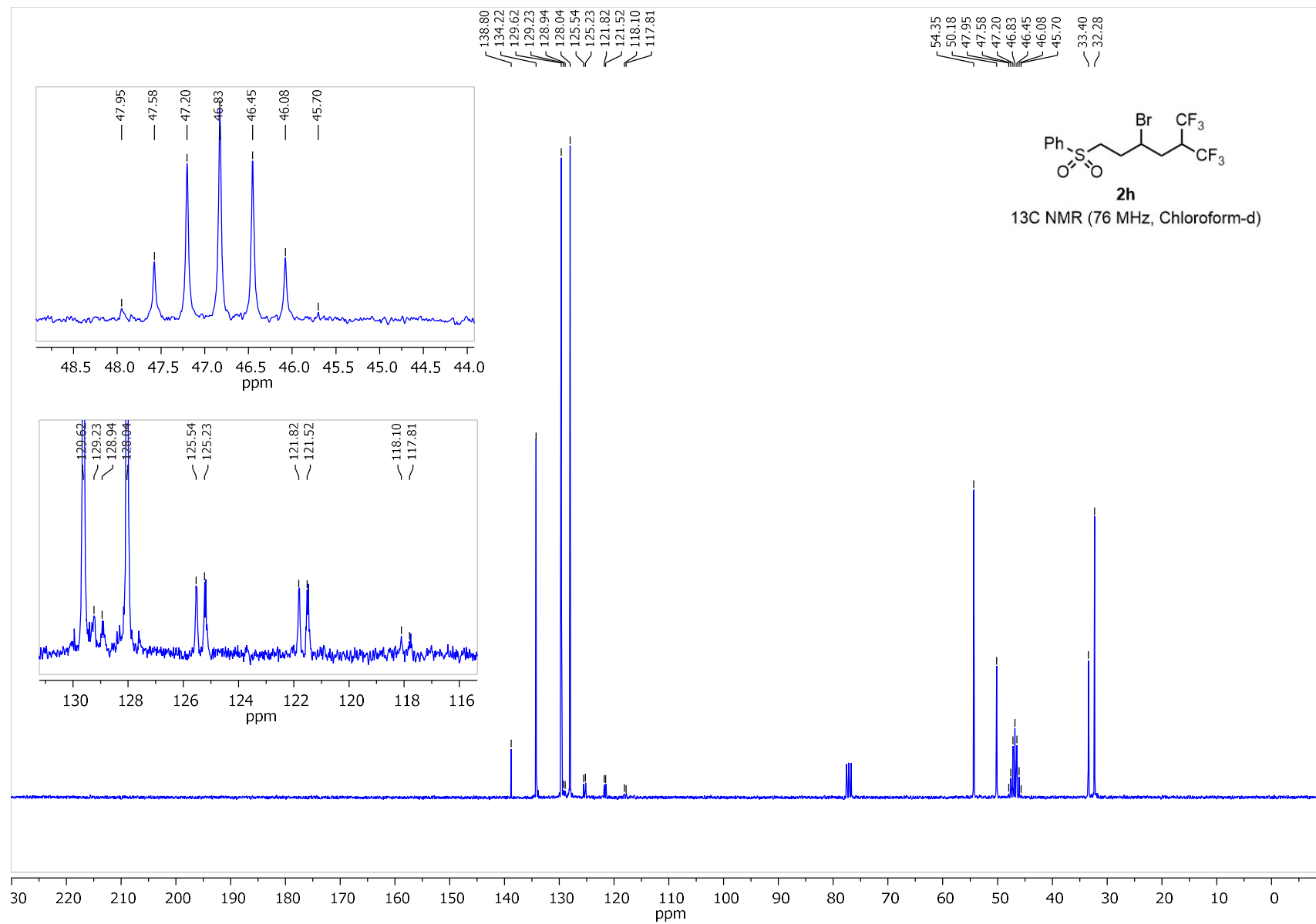


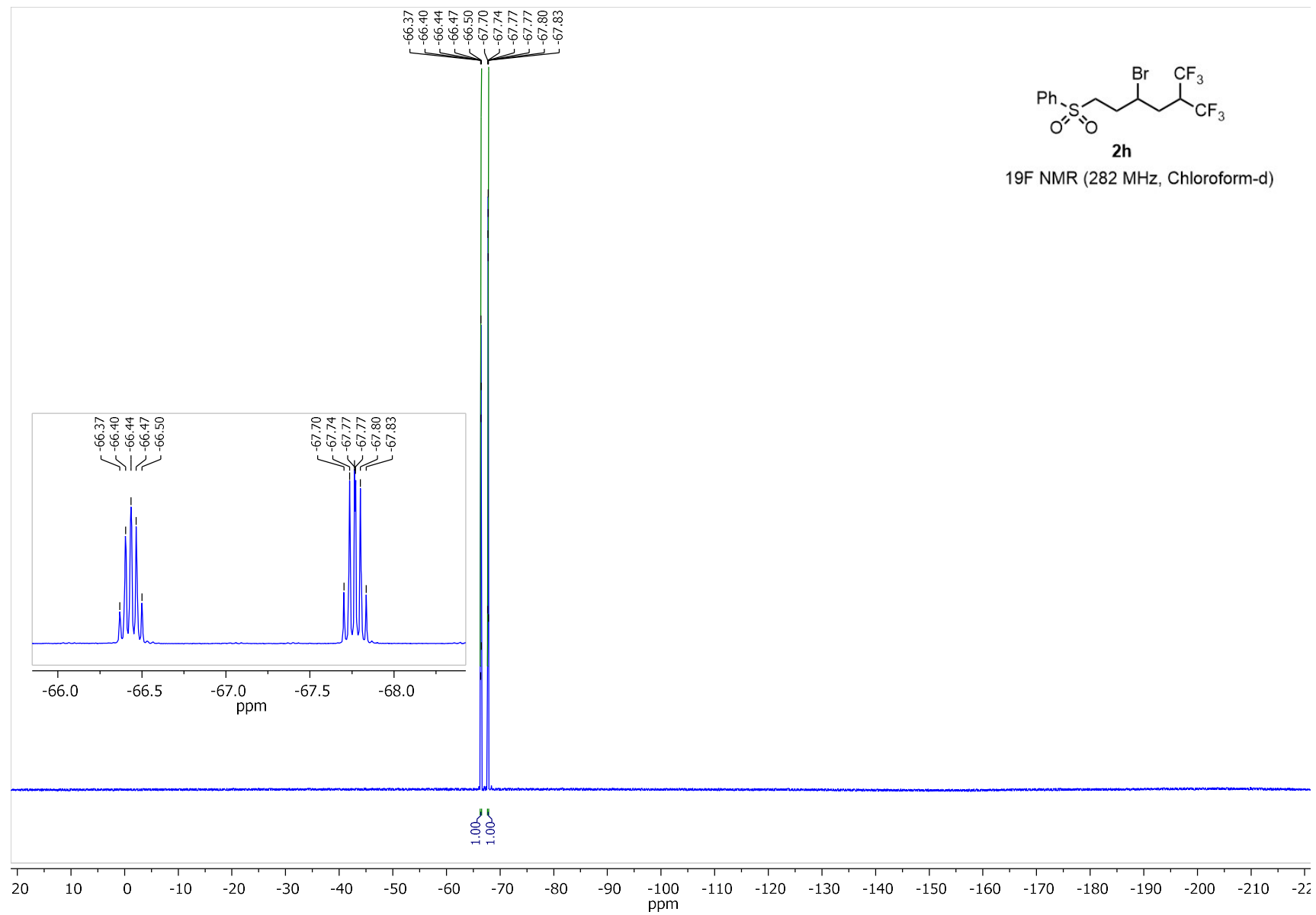


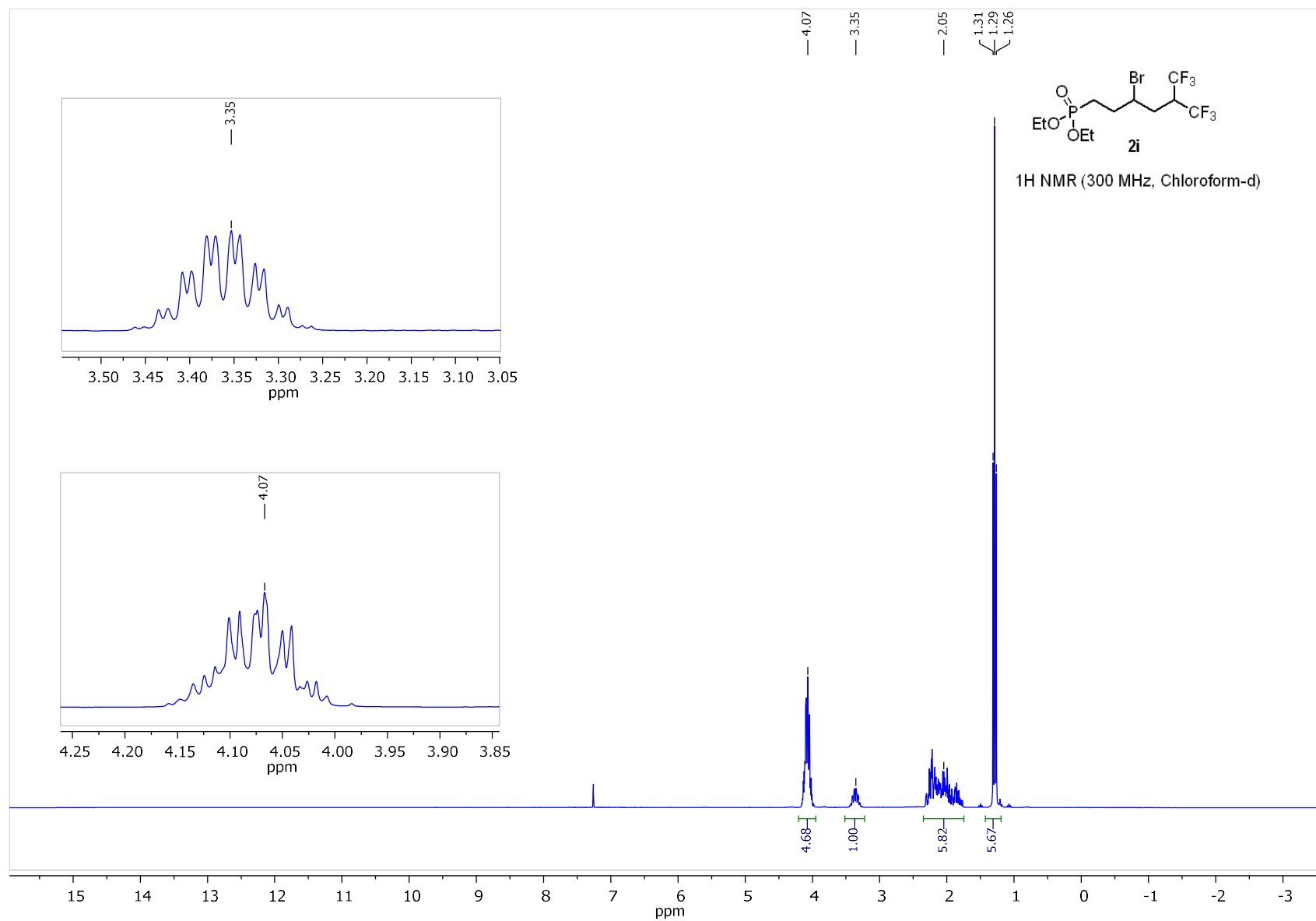


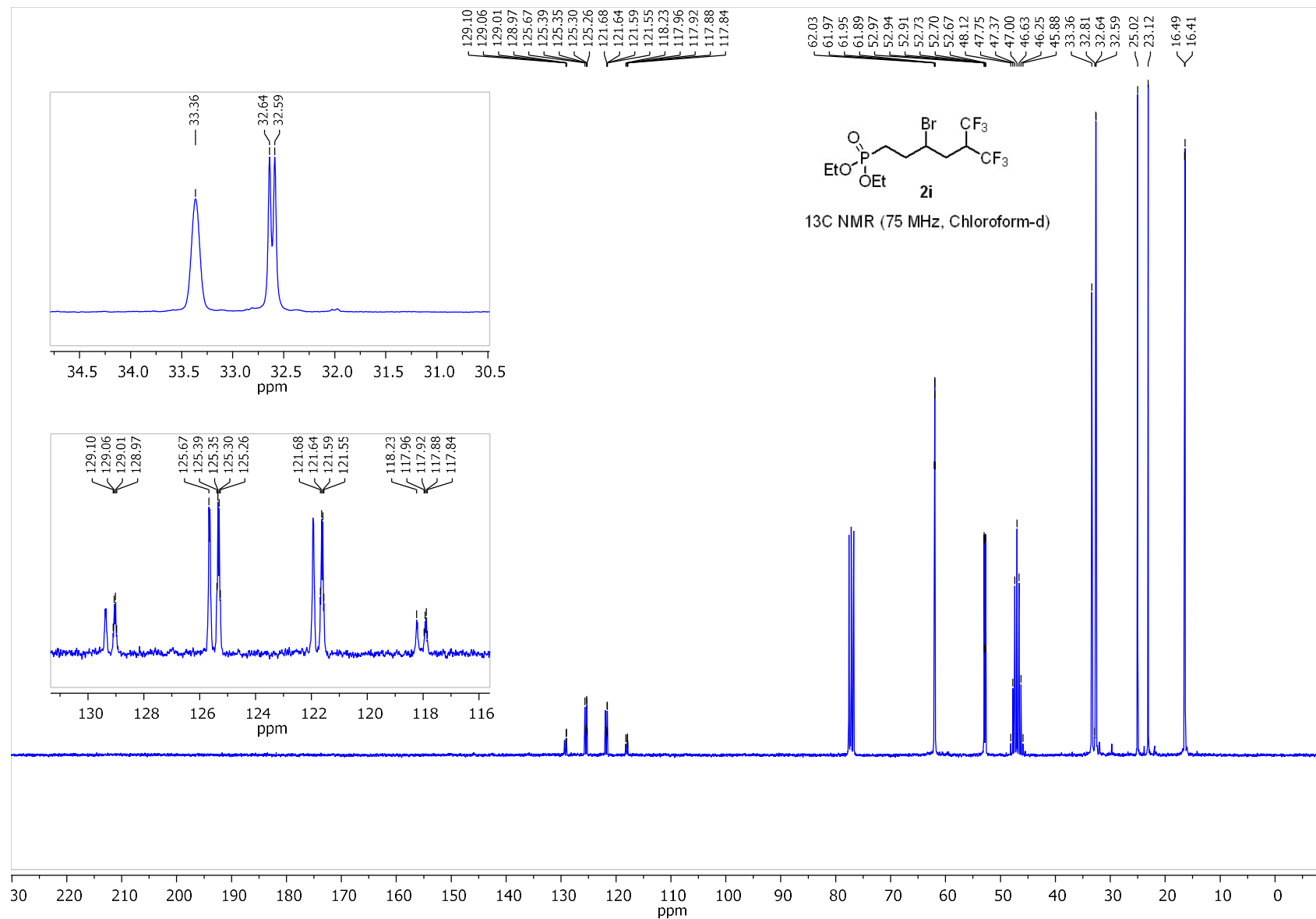


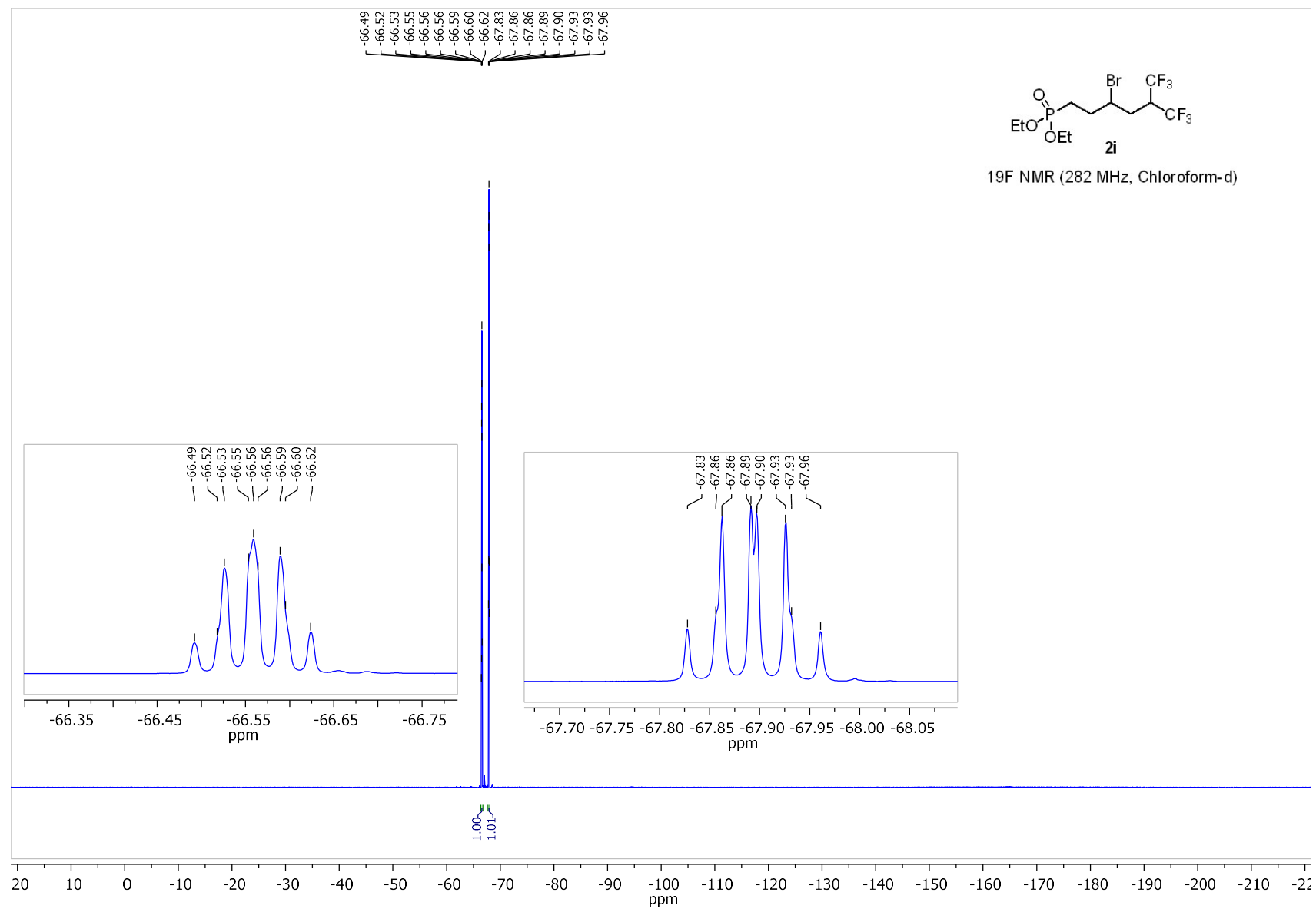


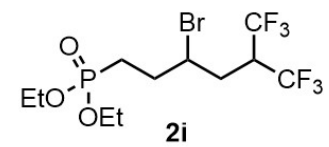




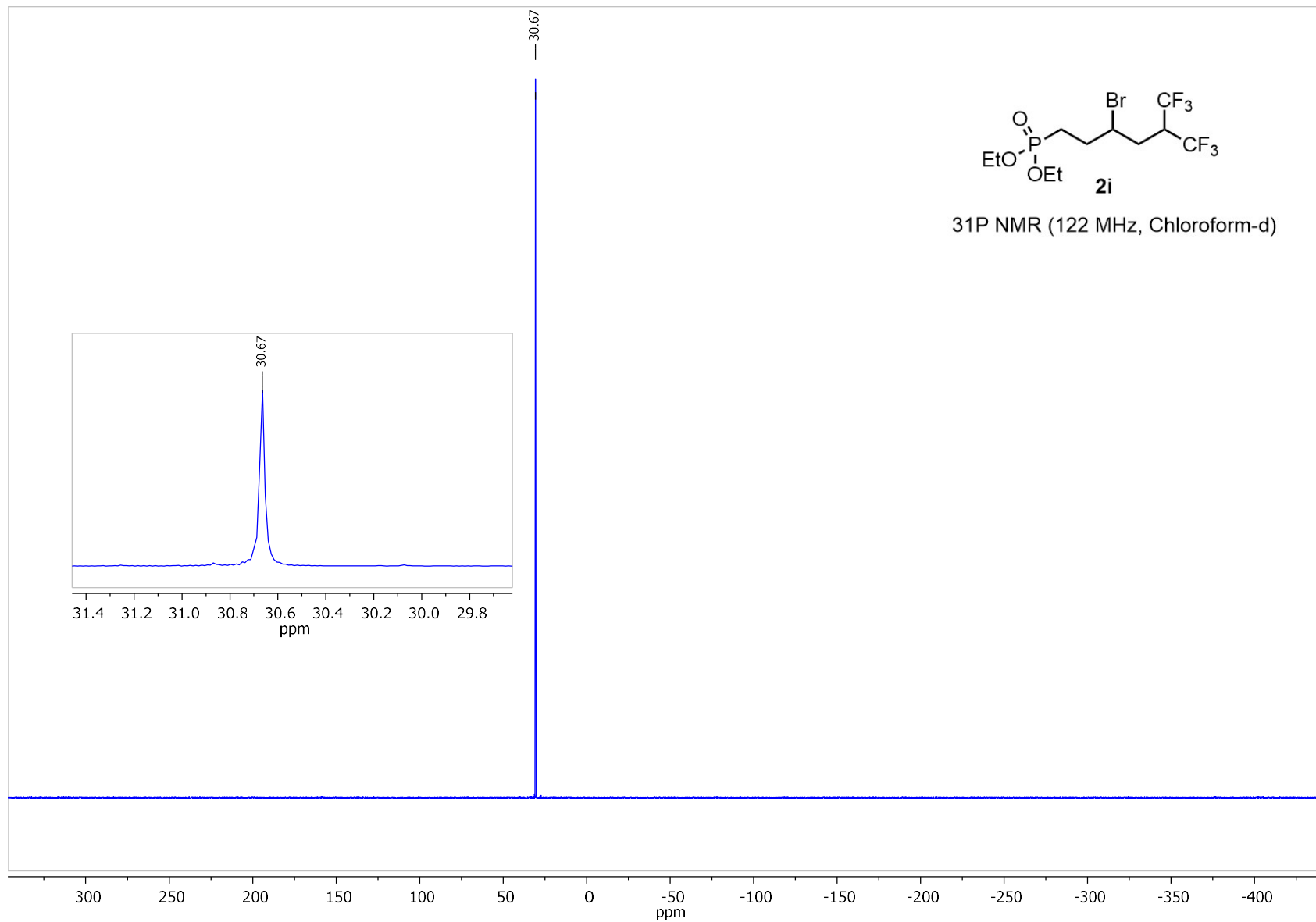


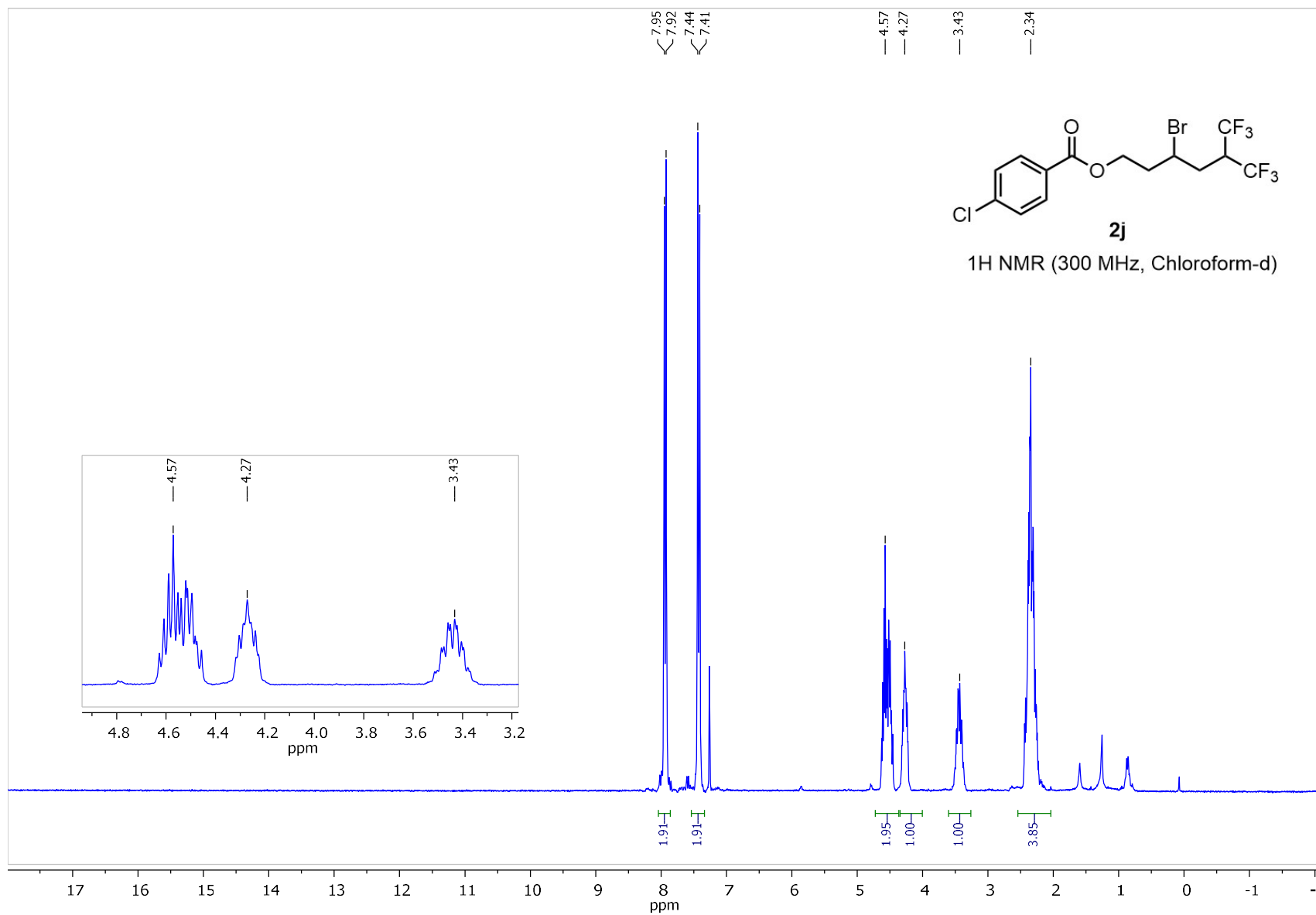


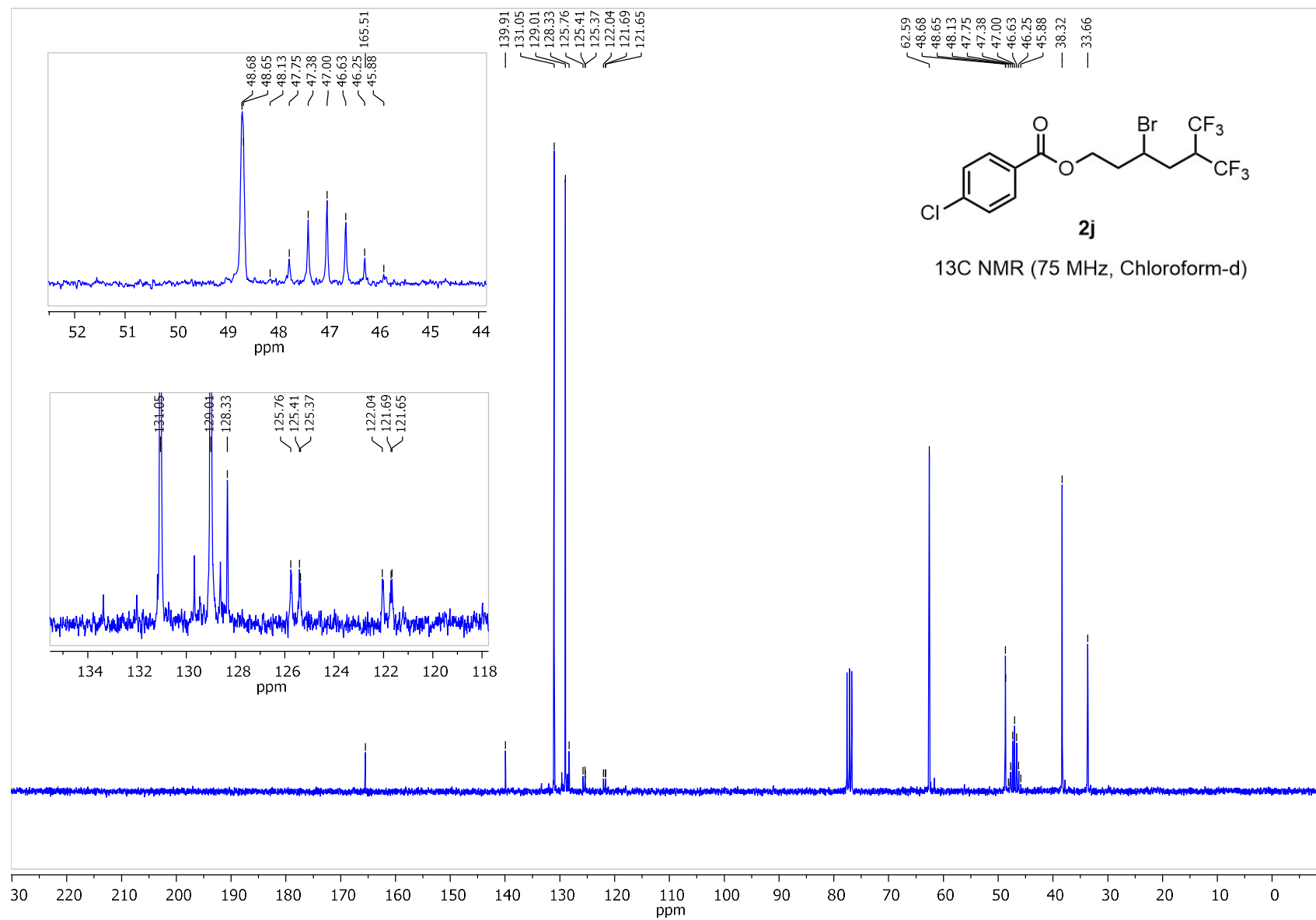


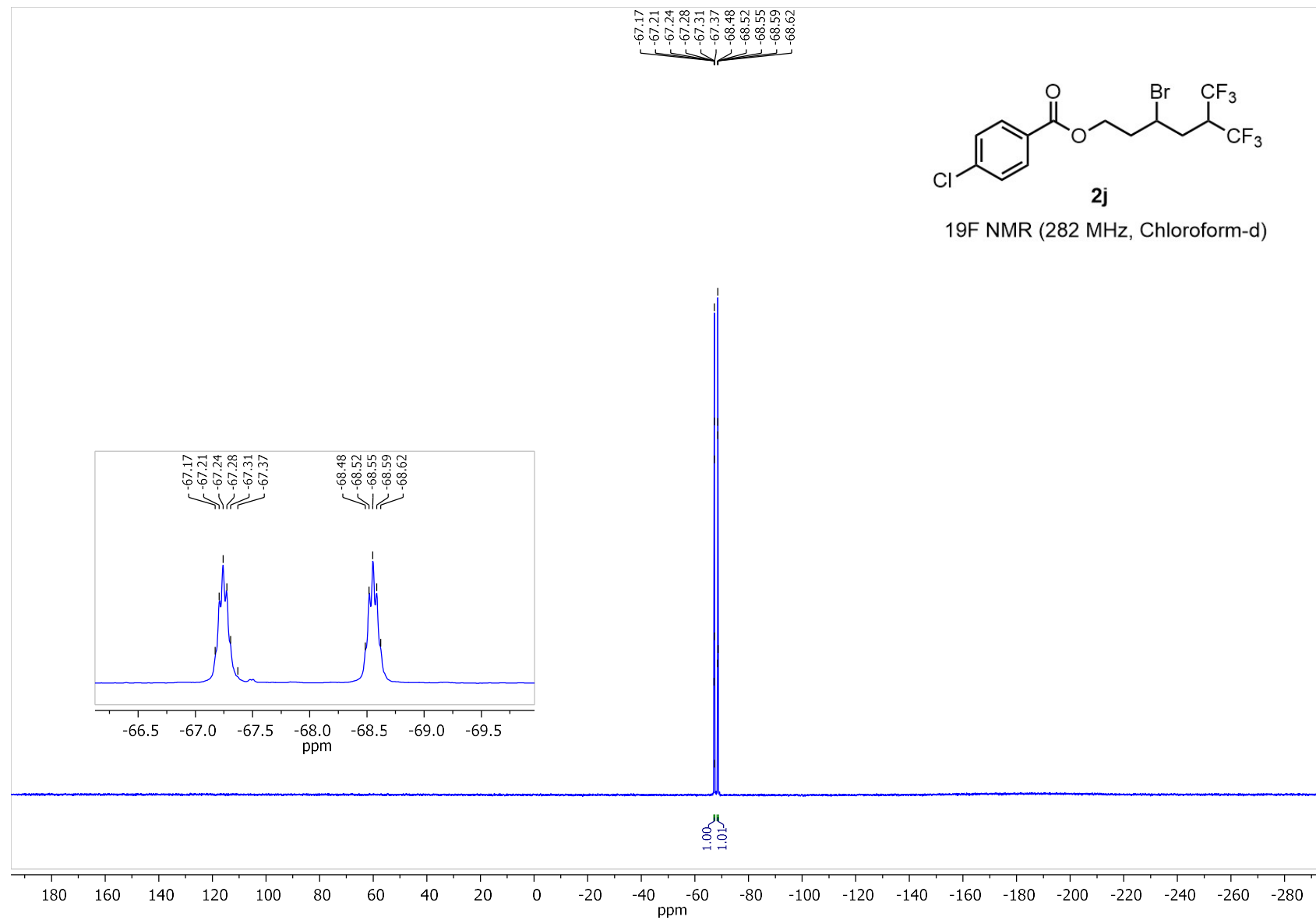


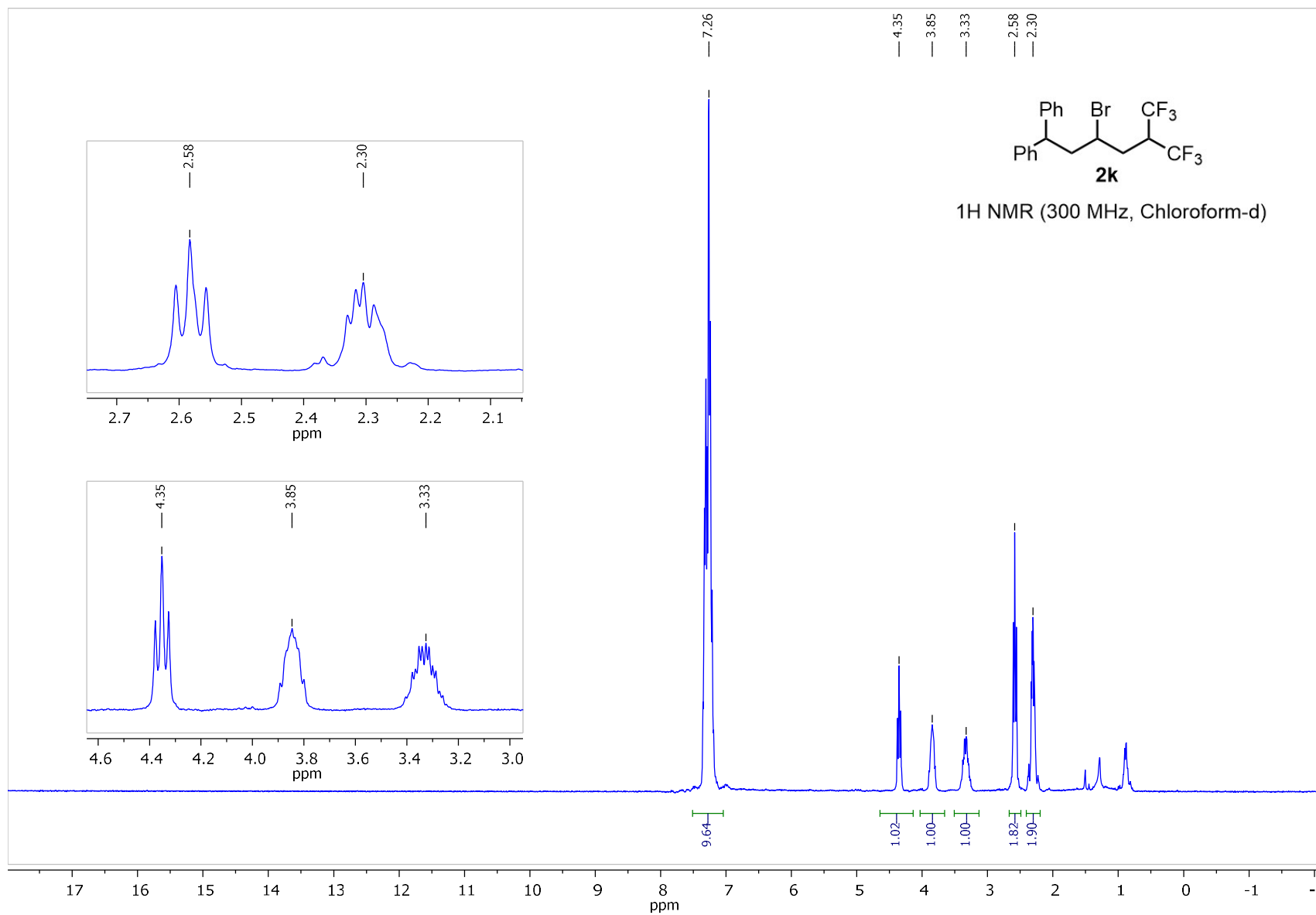
31P NMR (122 MHz, Chloroform-d)

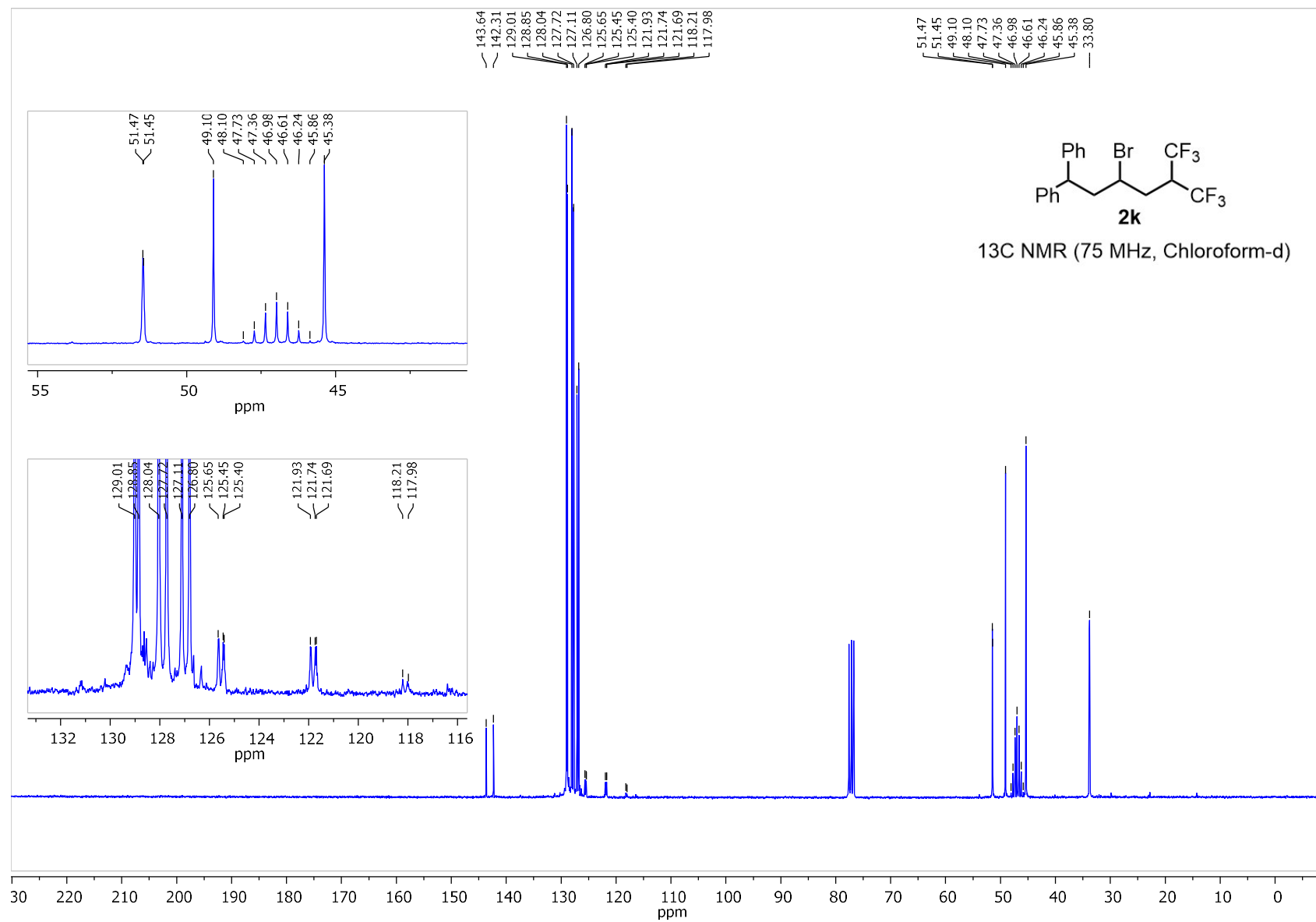


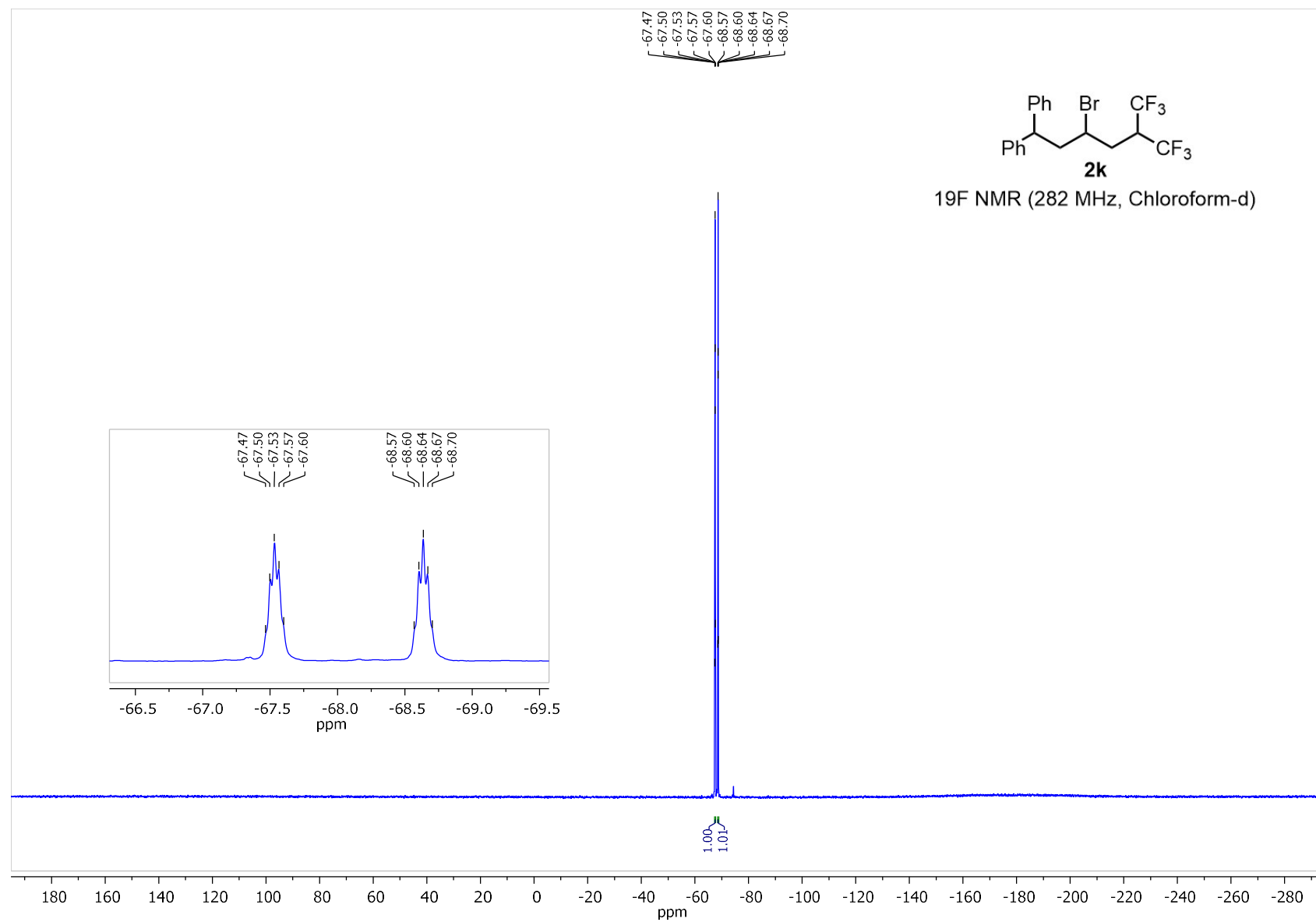


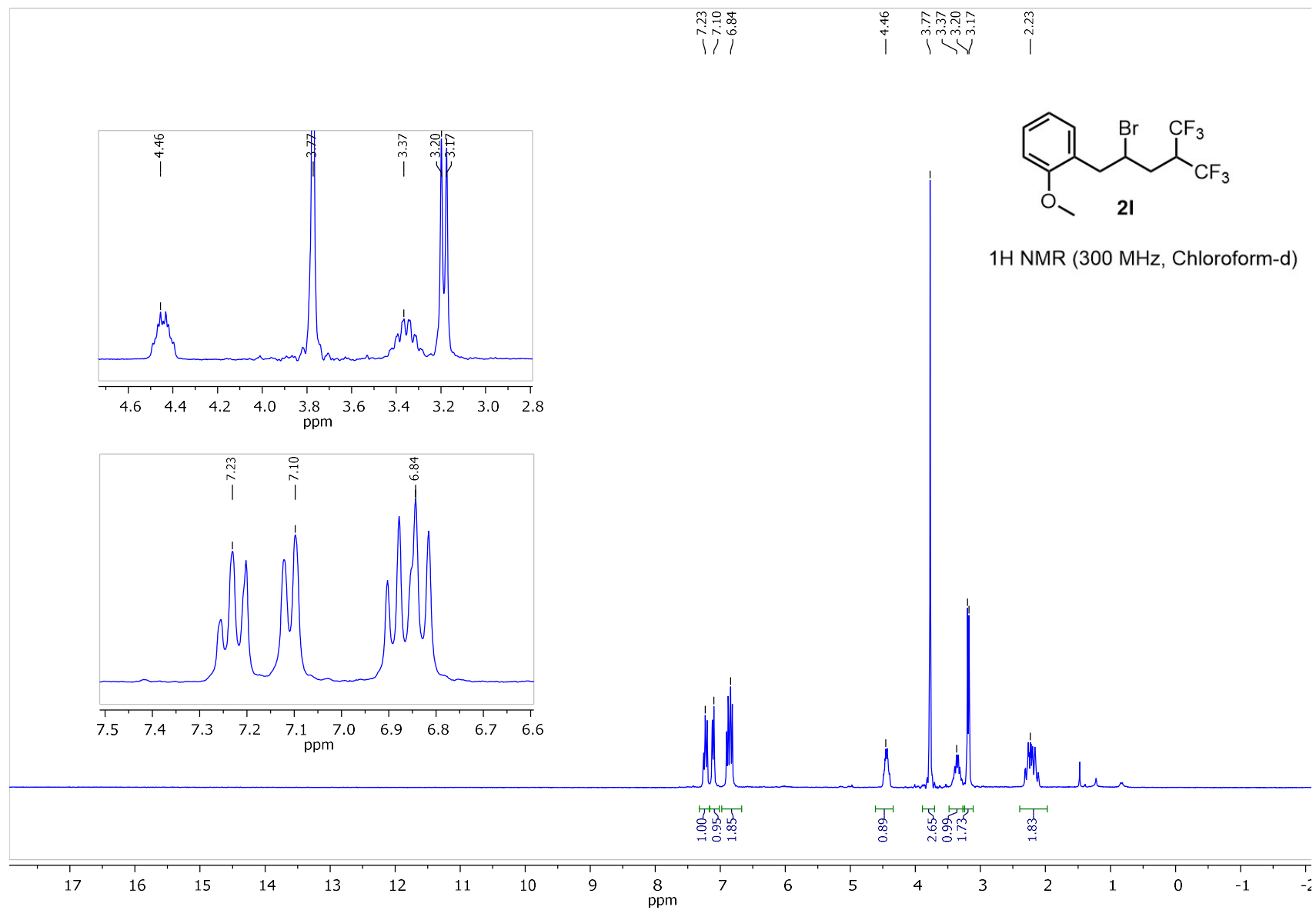


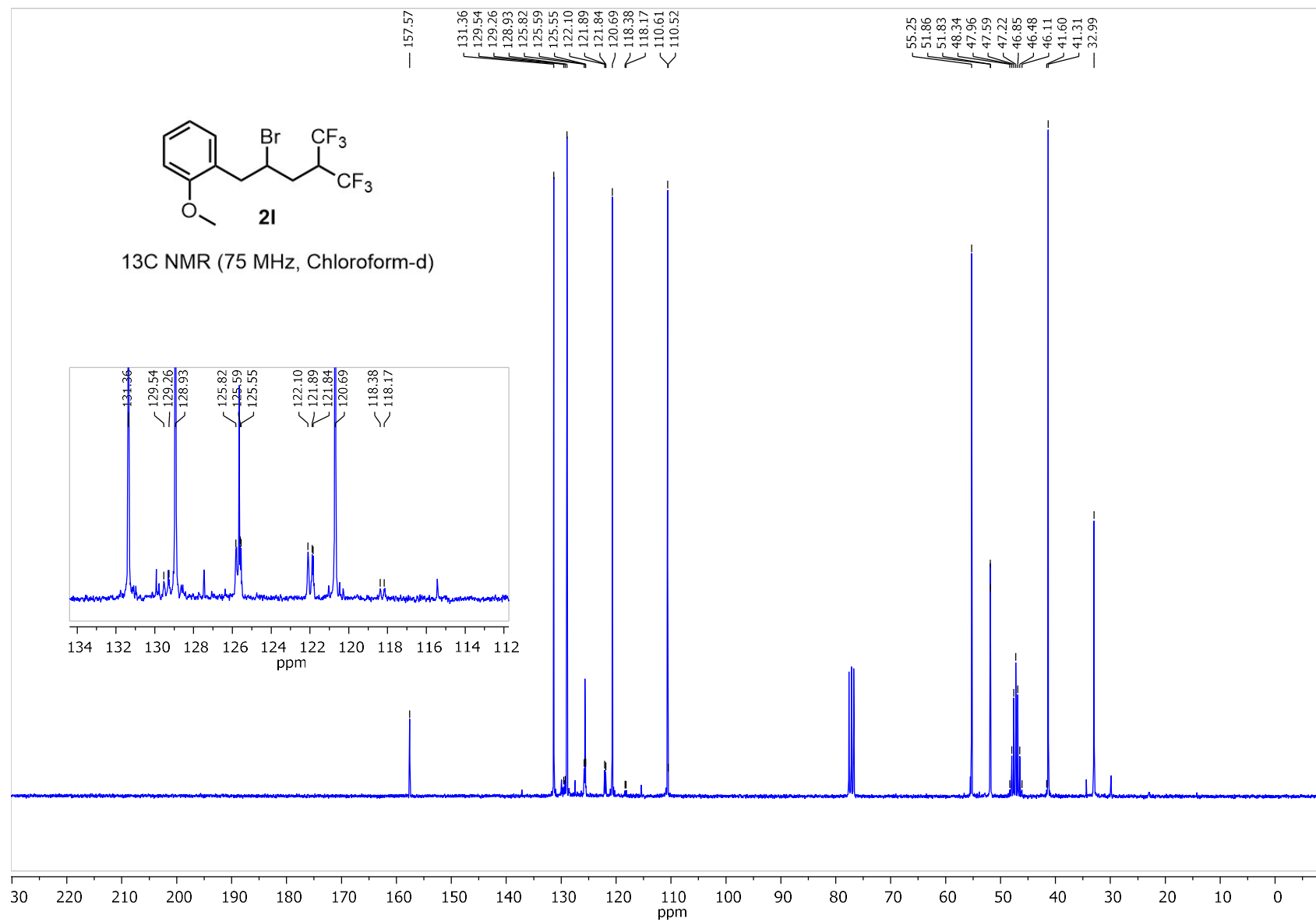


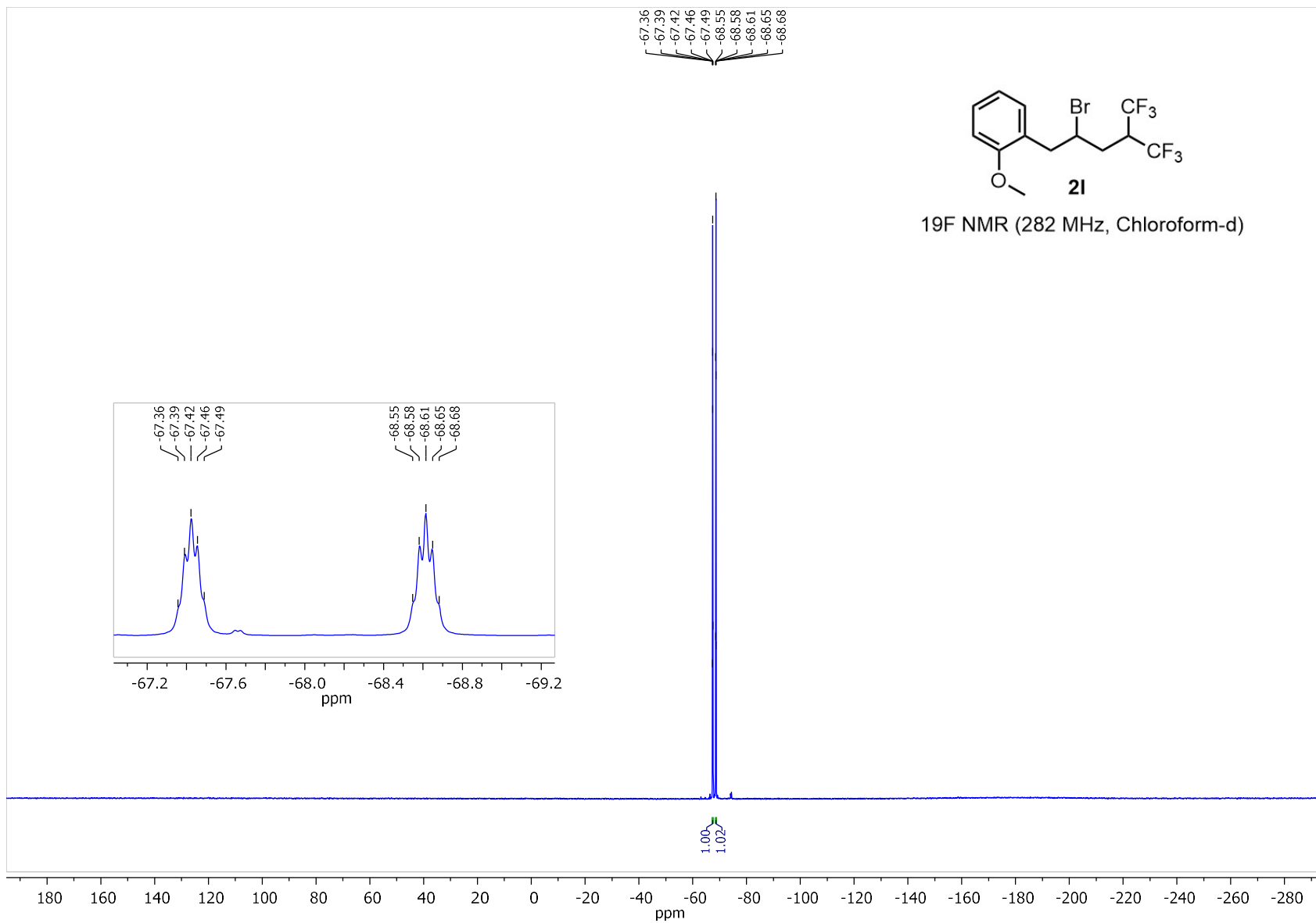


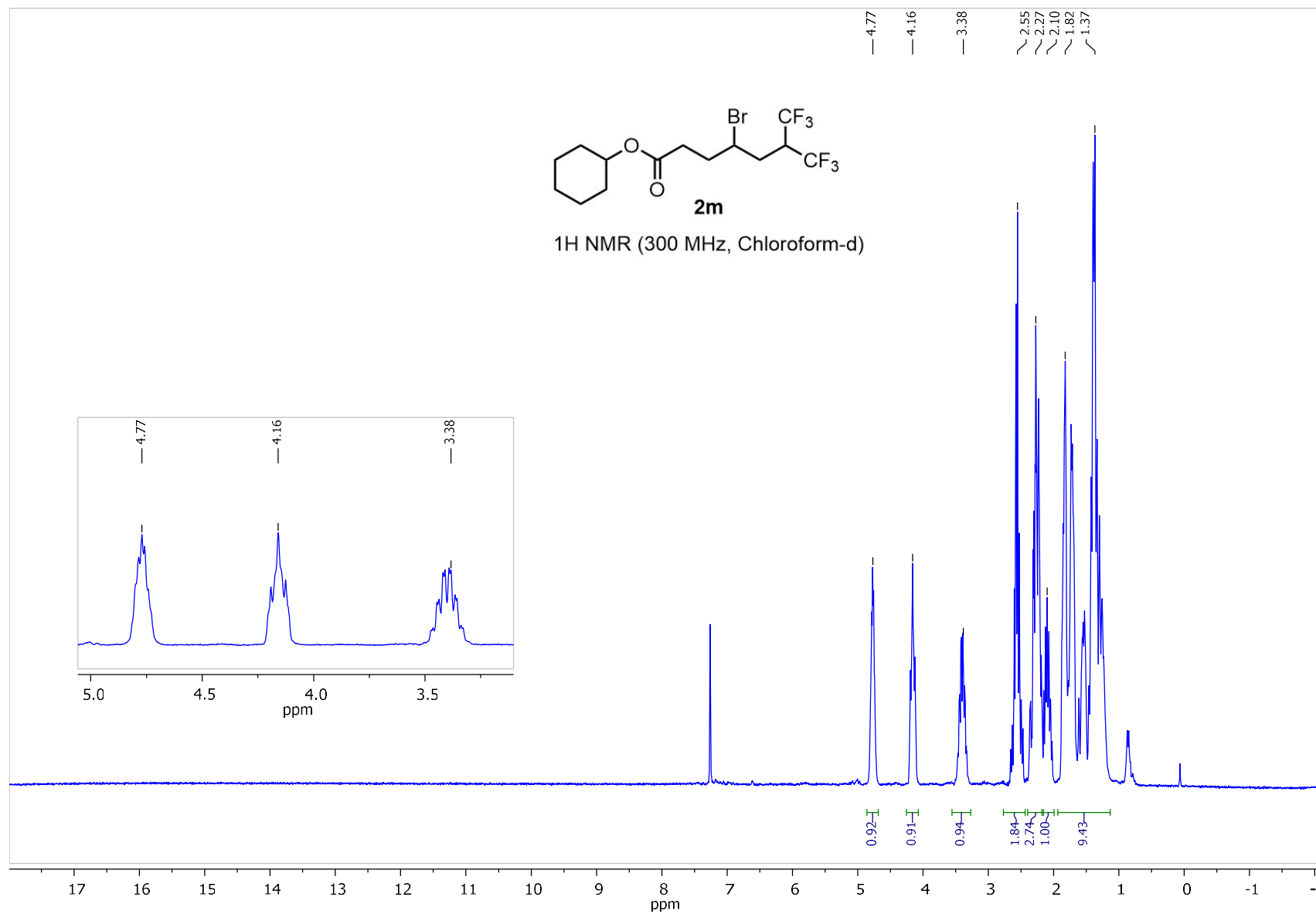


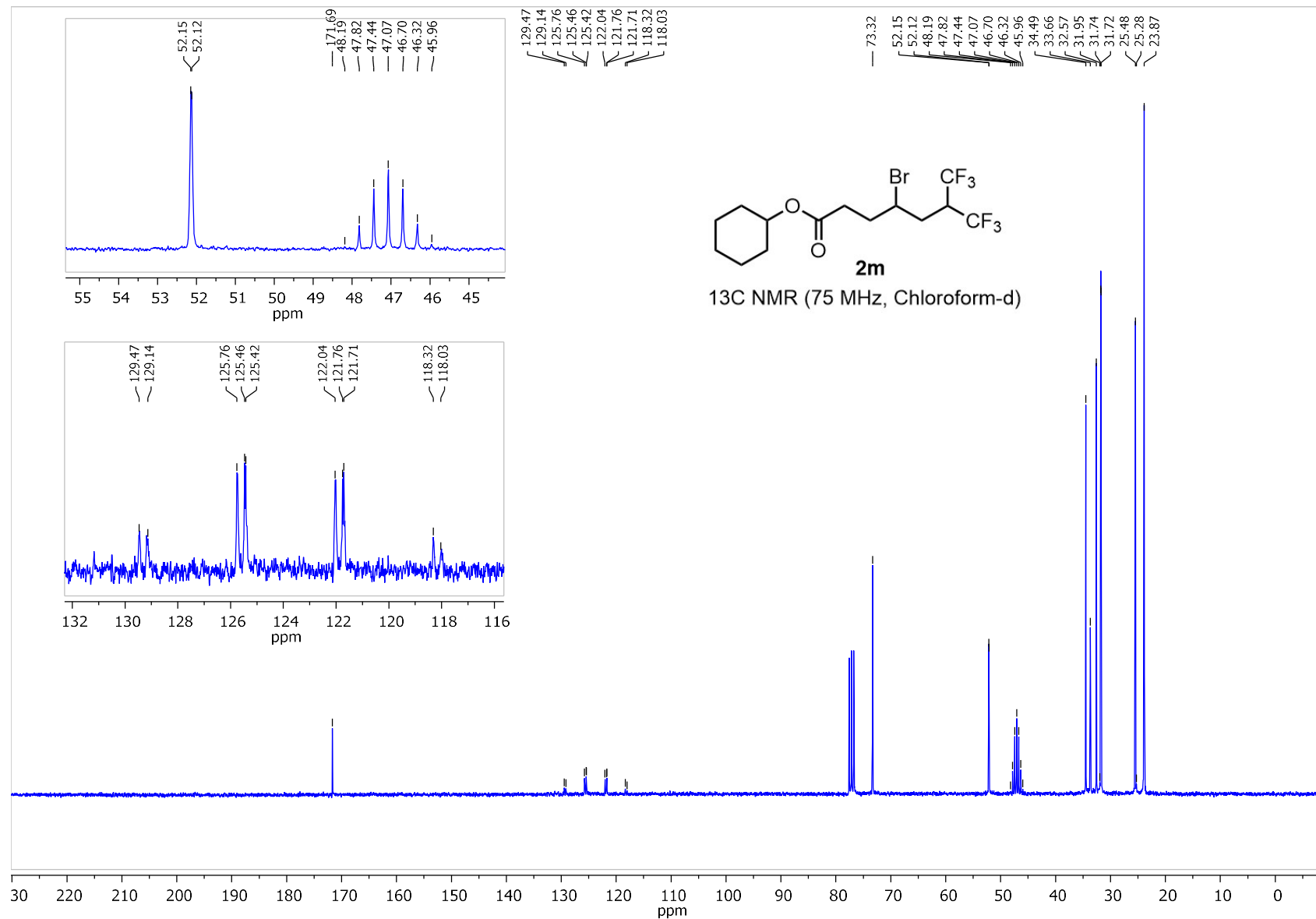


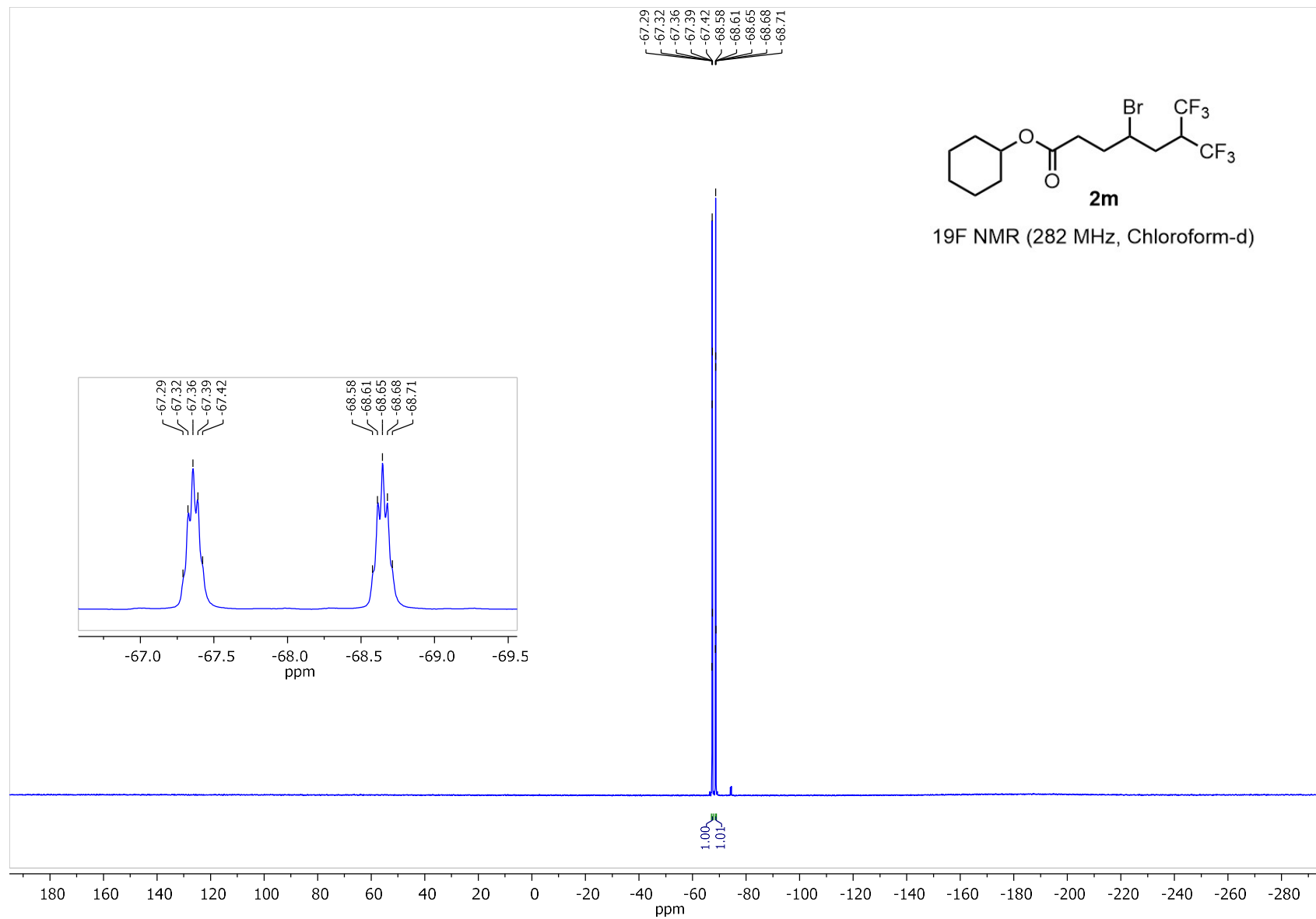


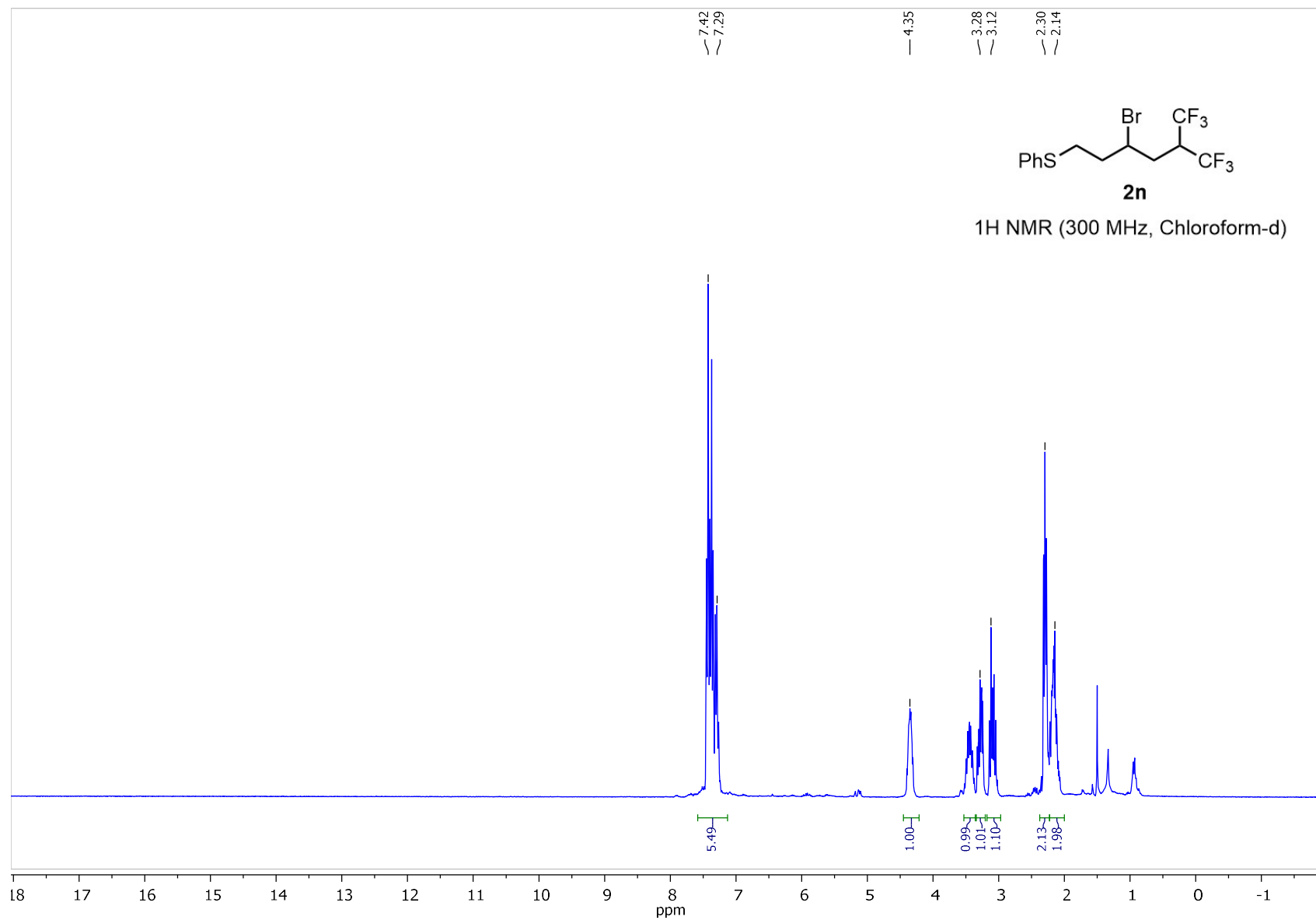


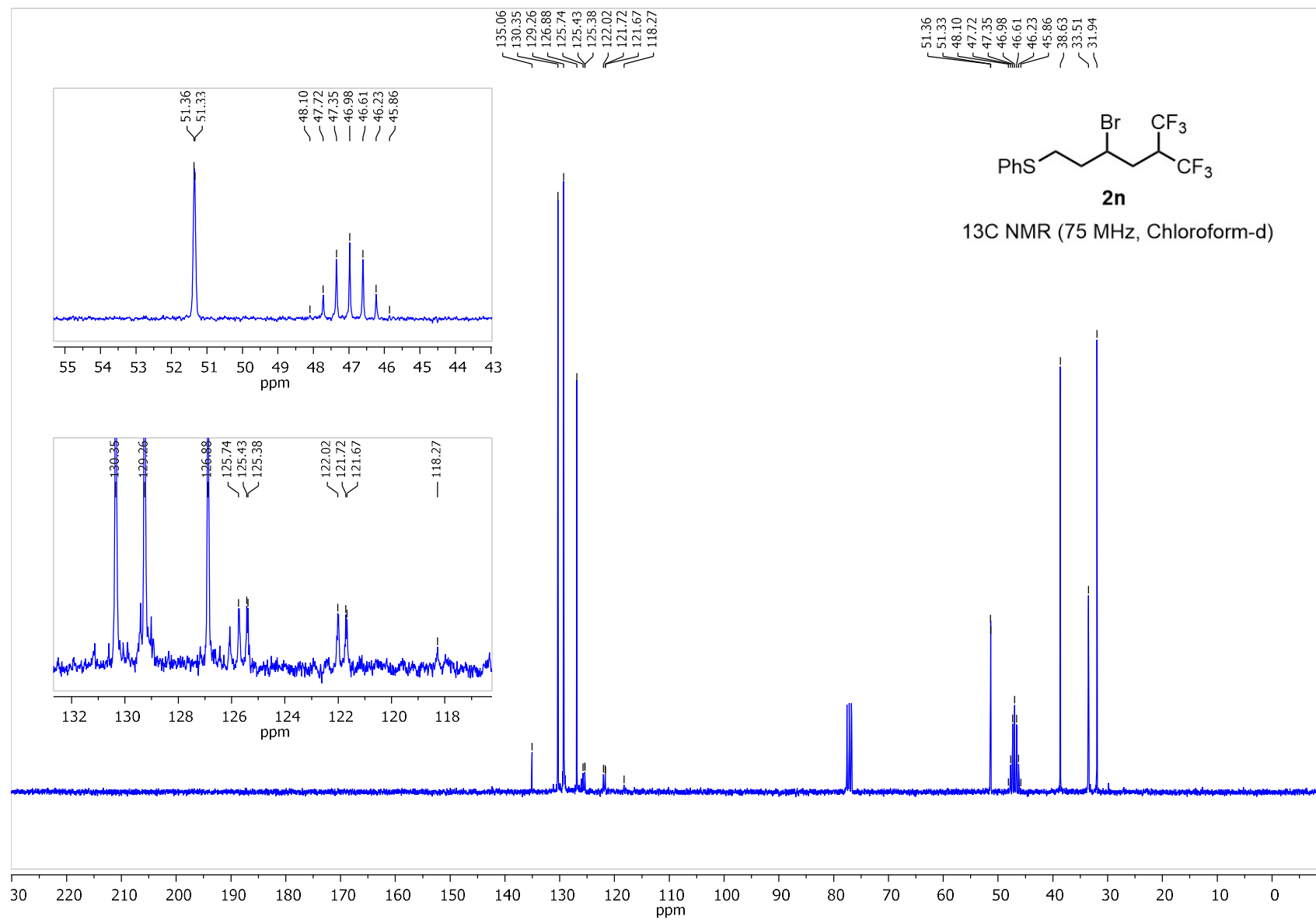


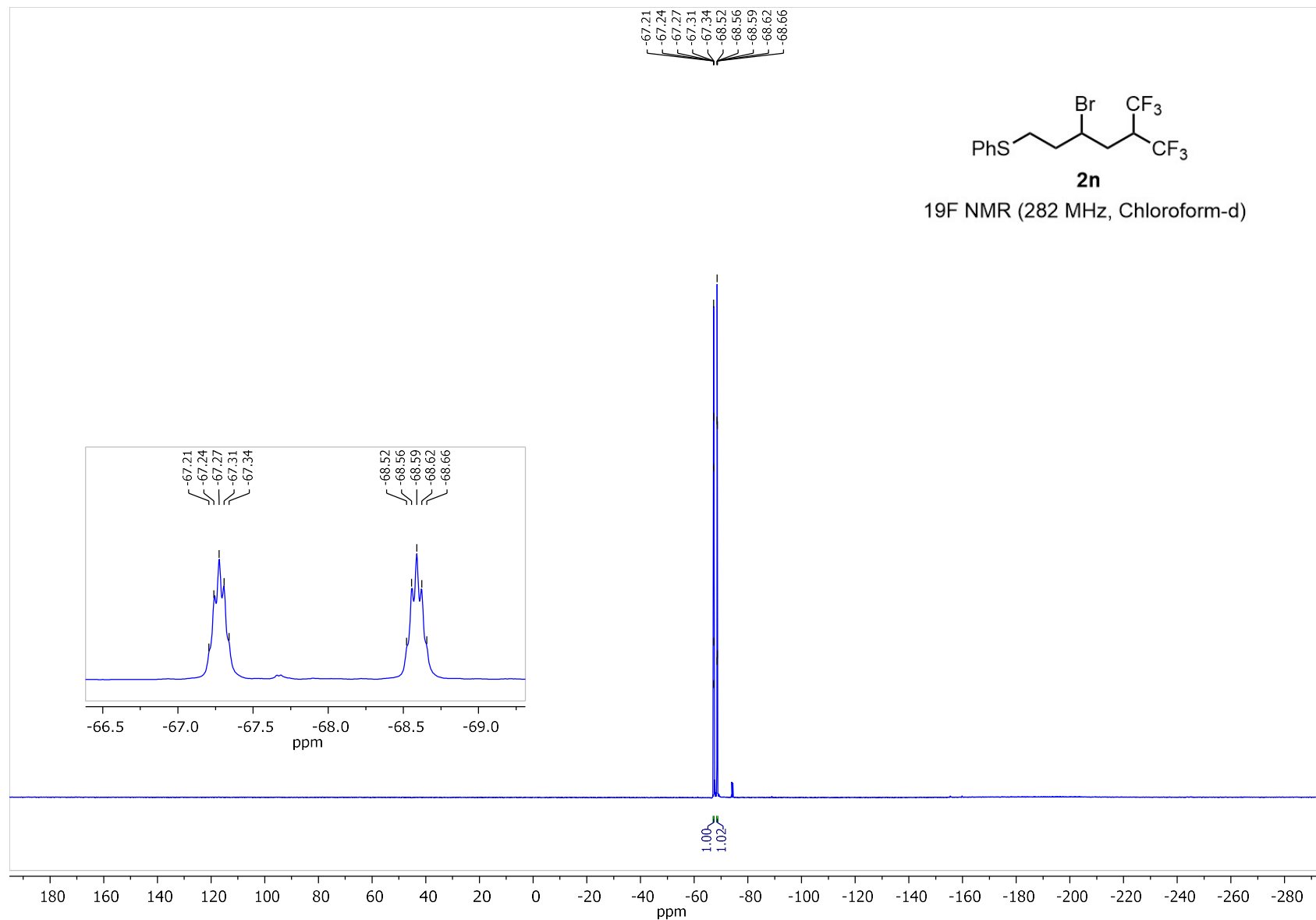
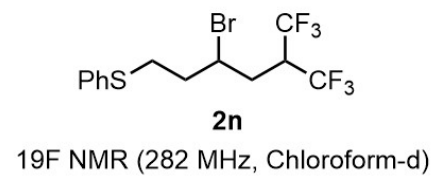


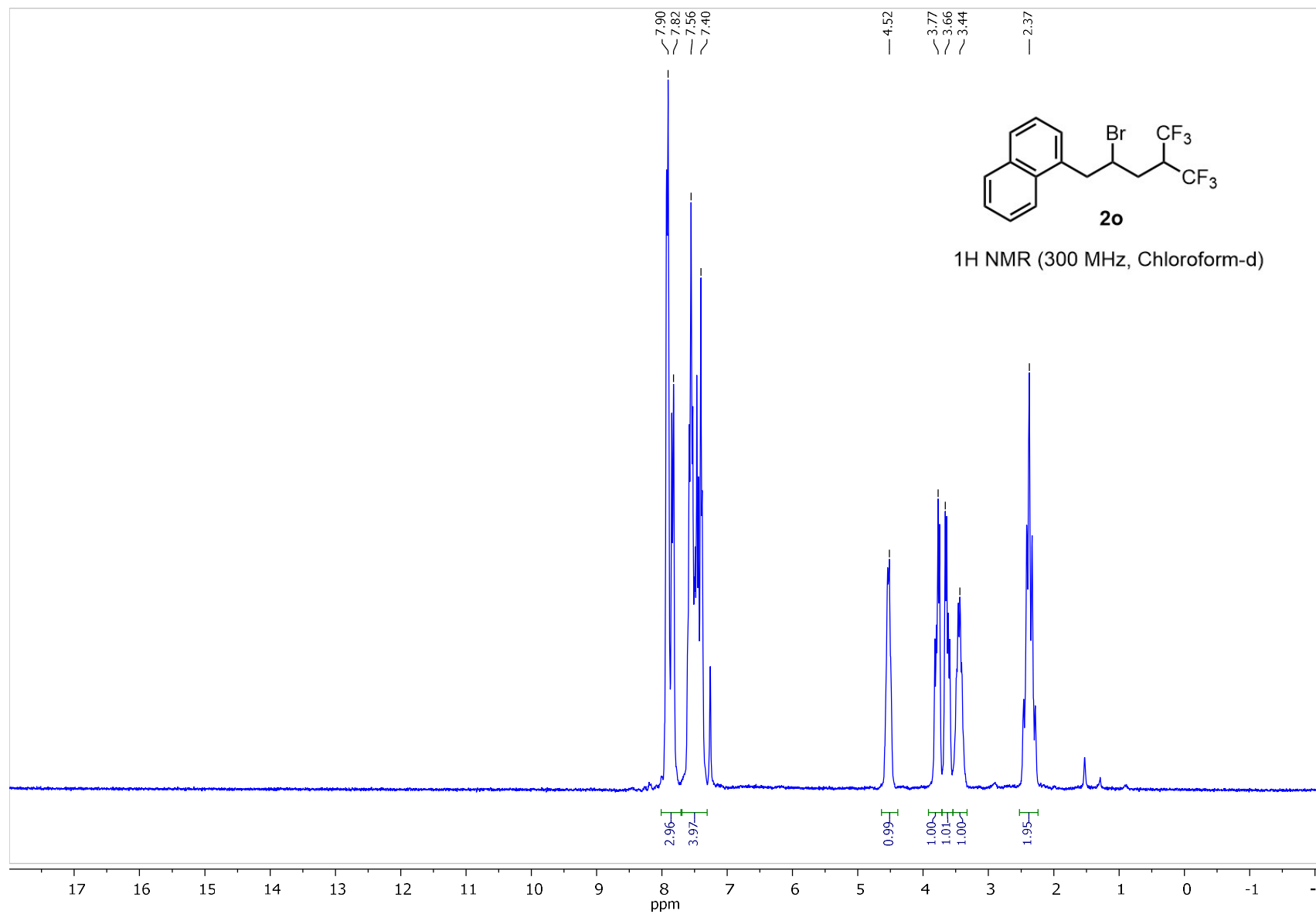


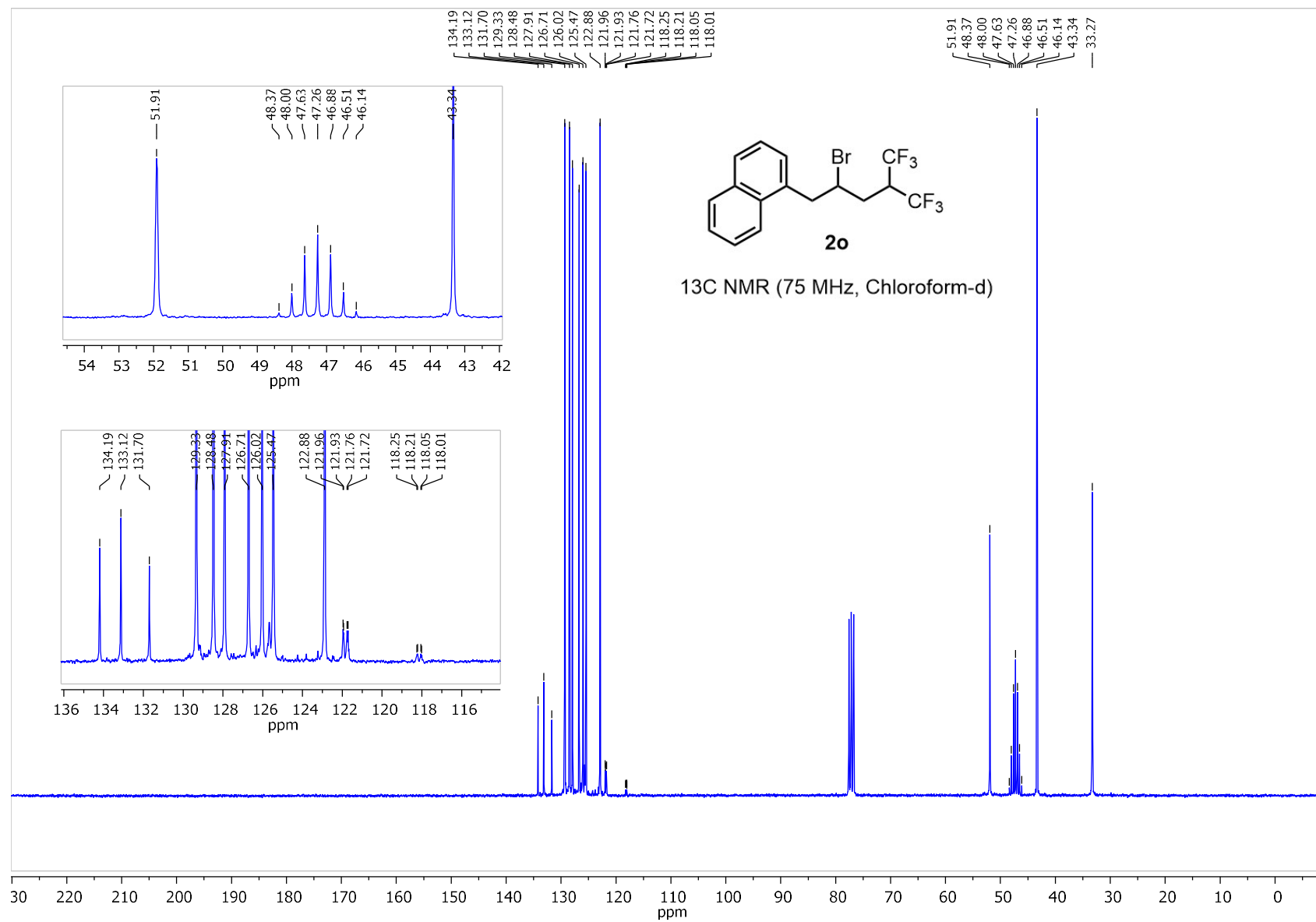


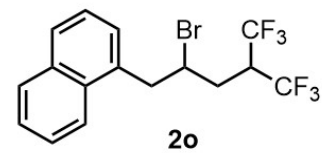




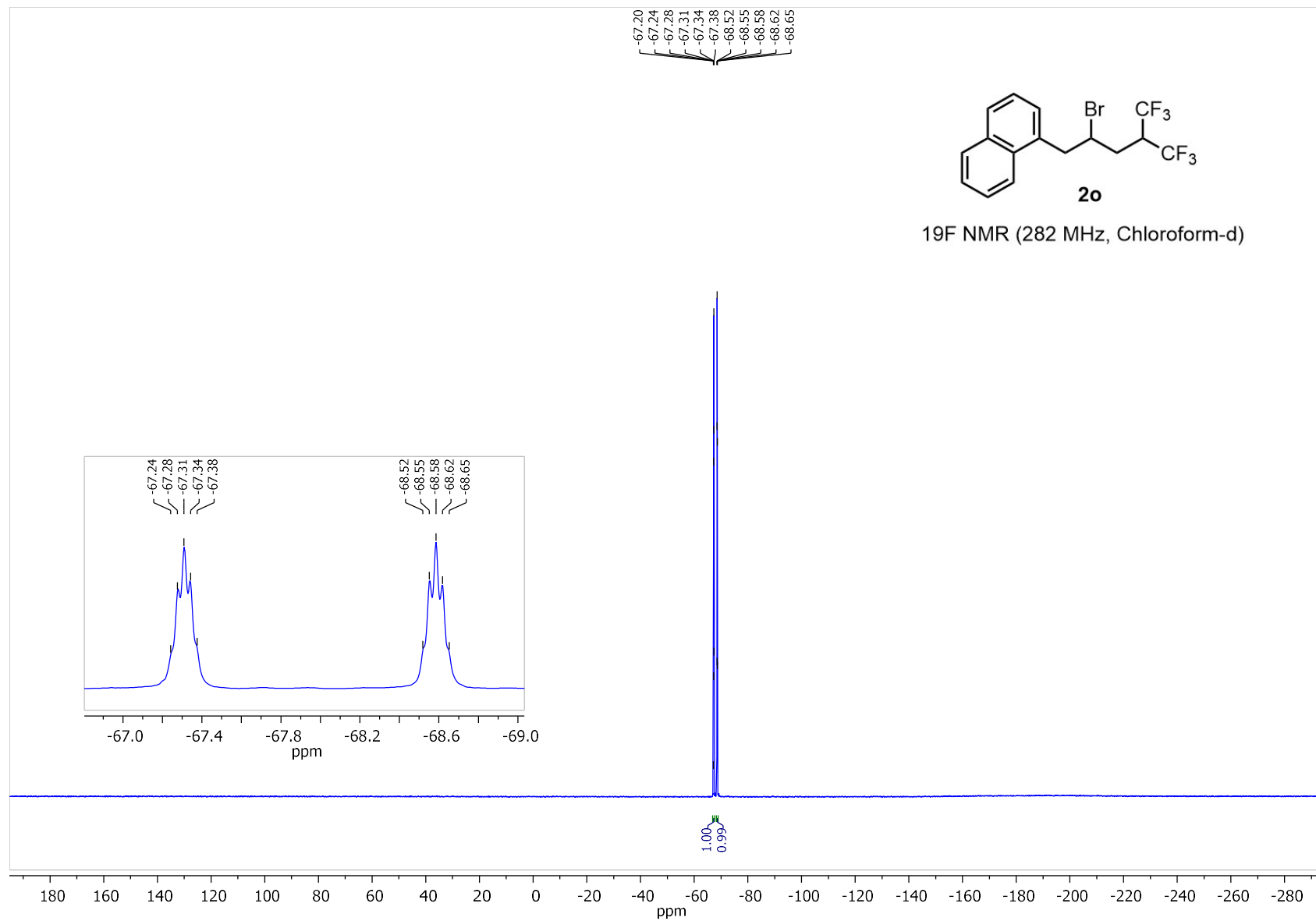


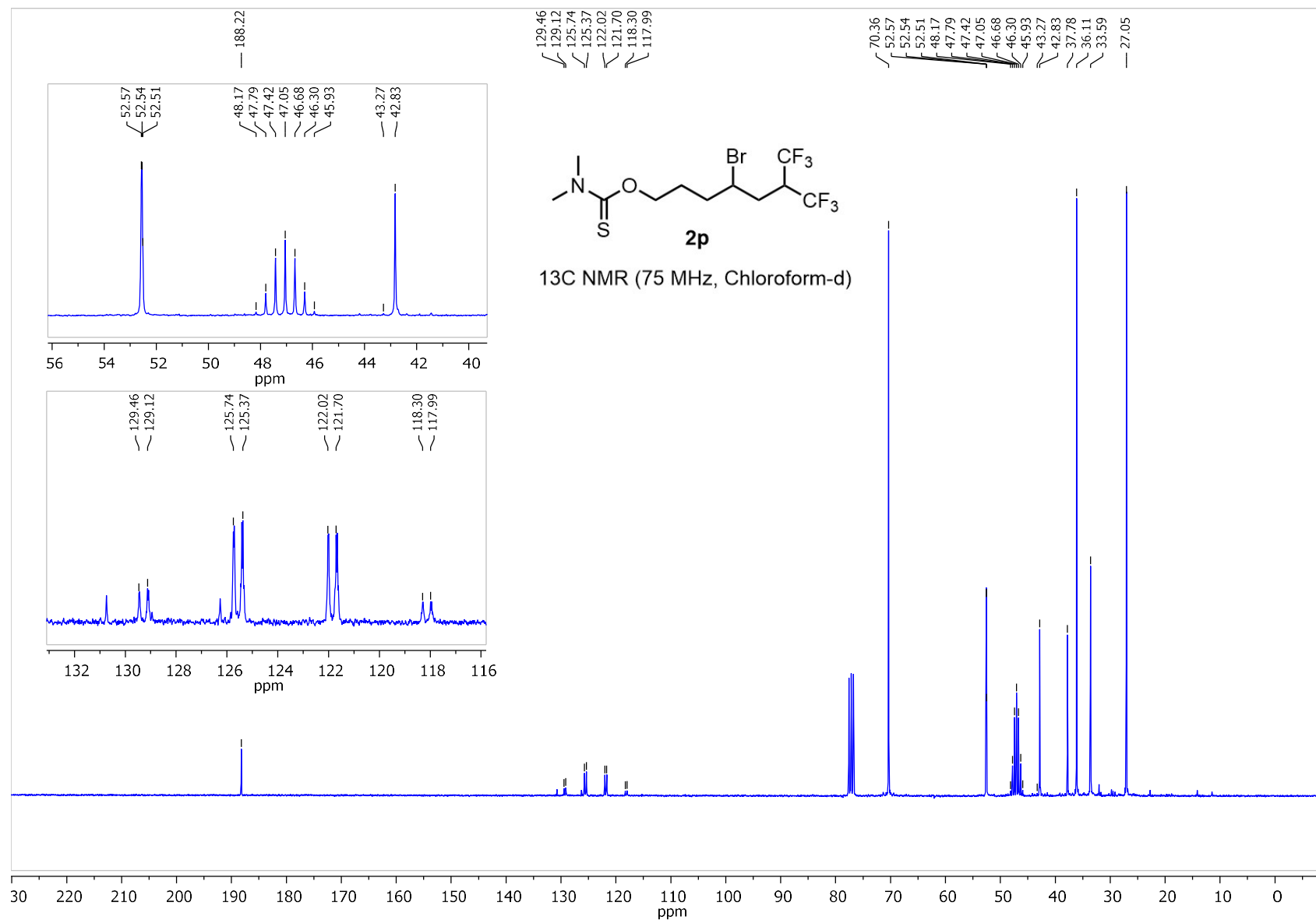


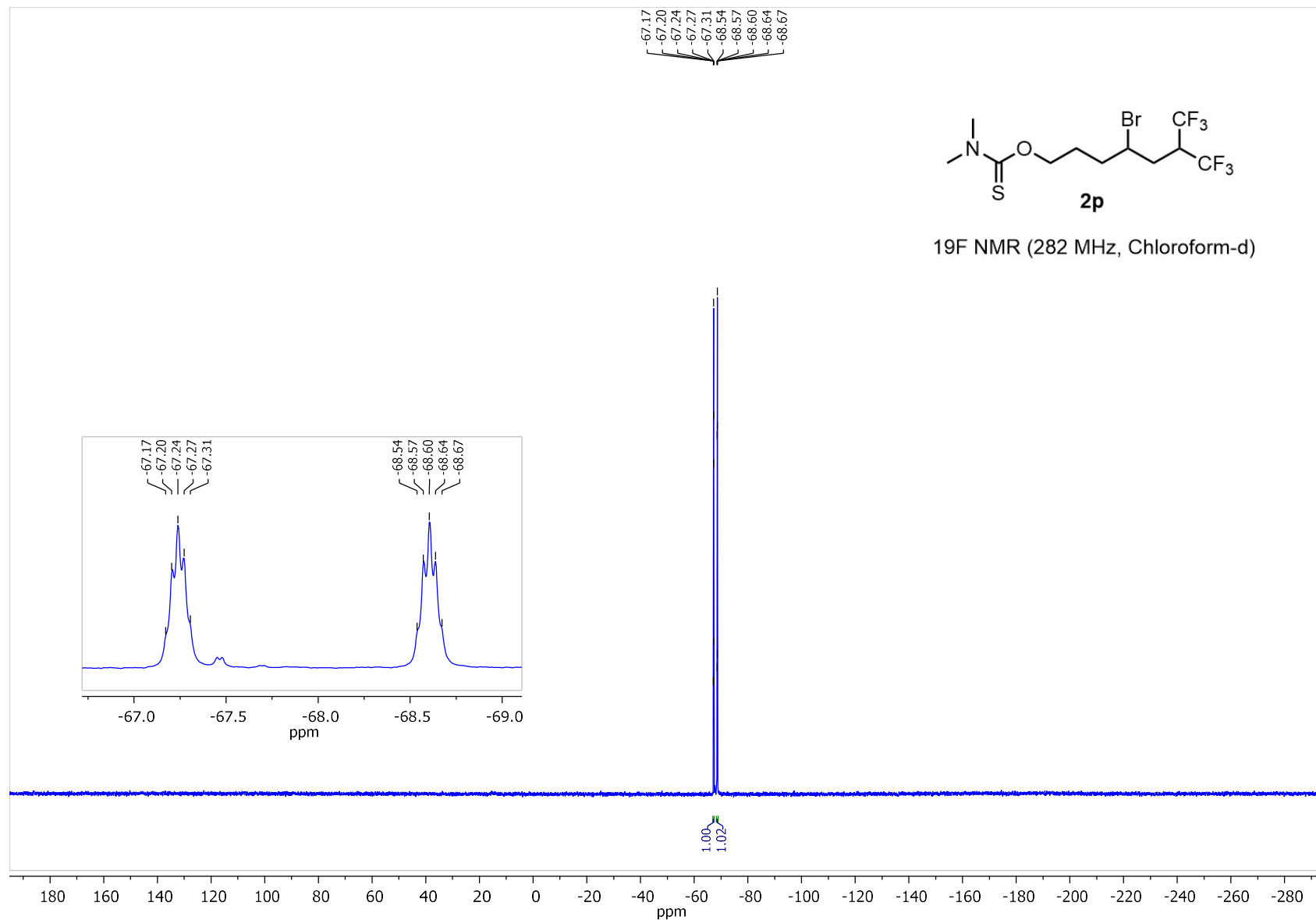


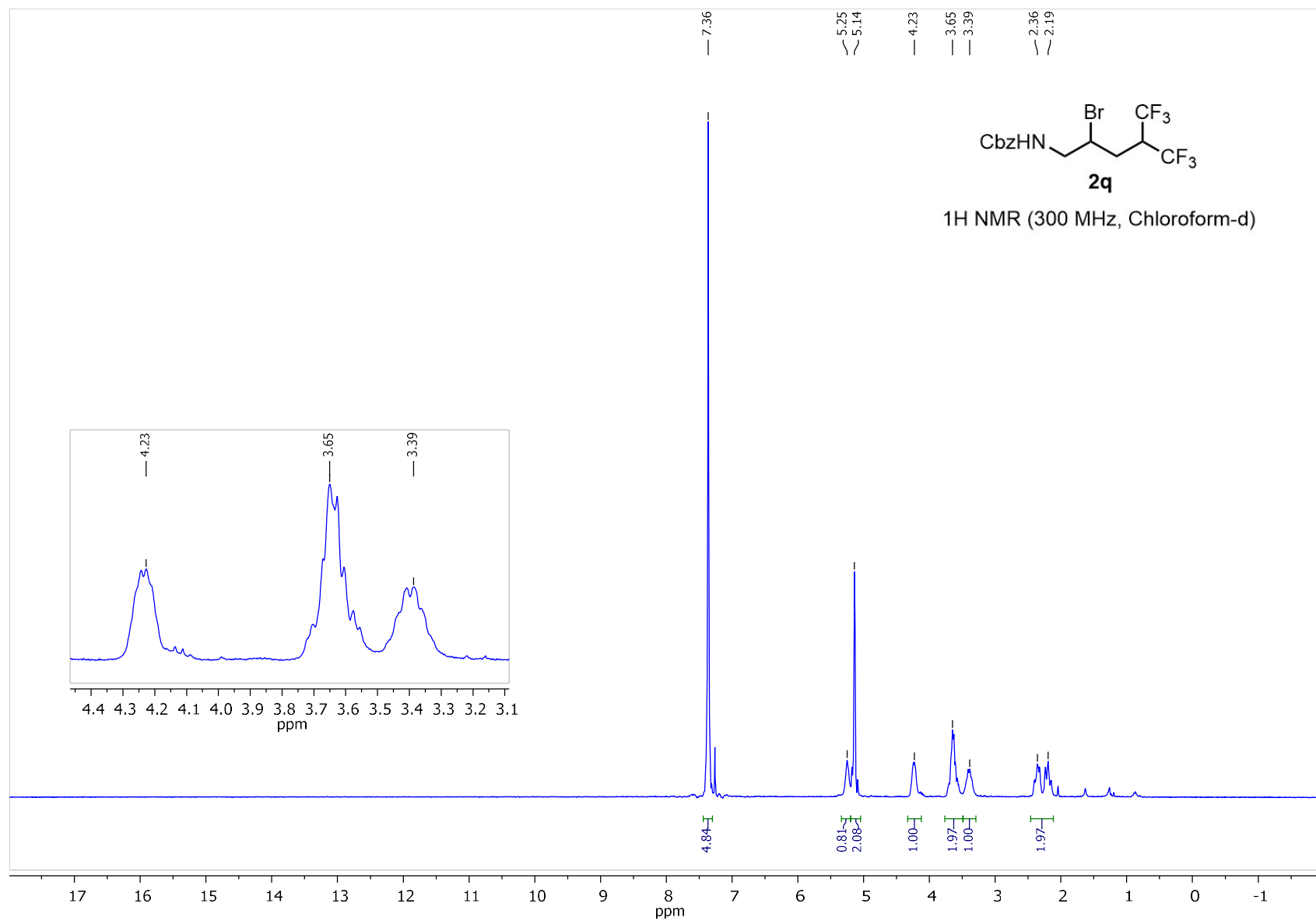


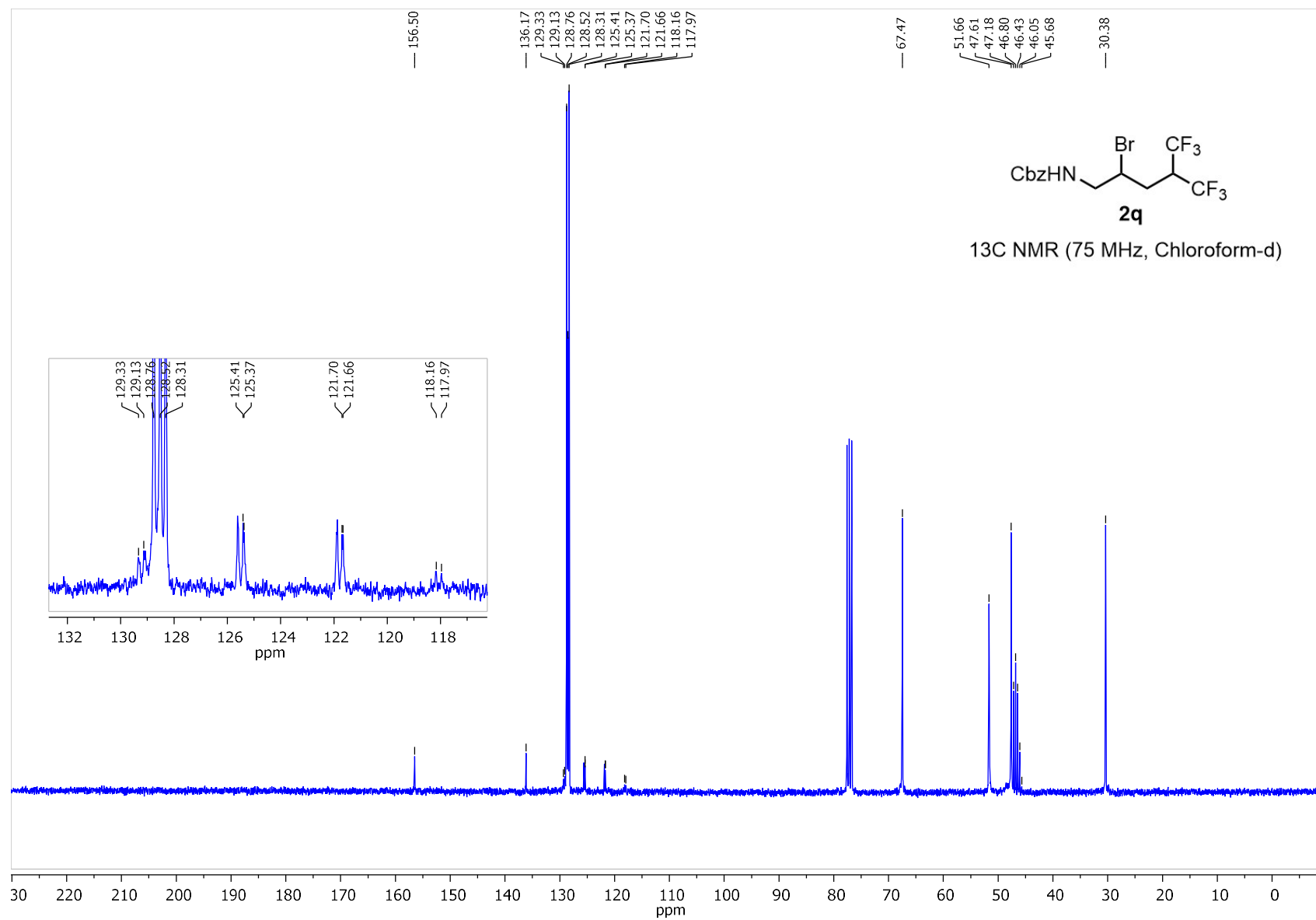
^{19}F NMR (282 MHz, Chloroform-d)

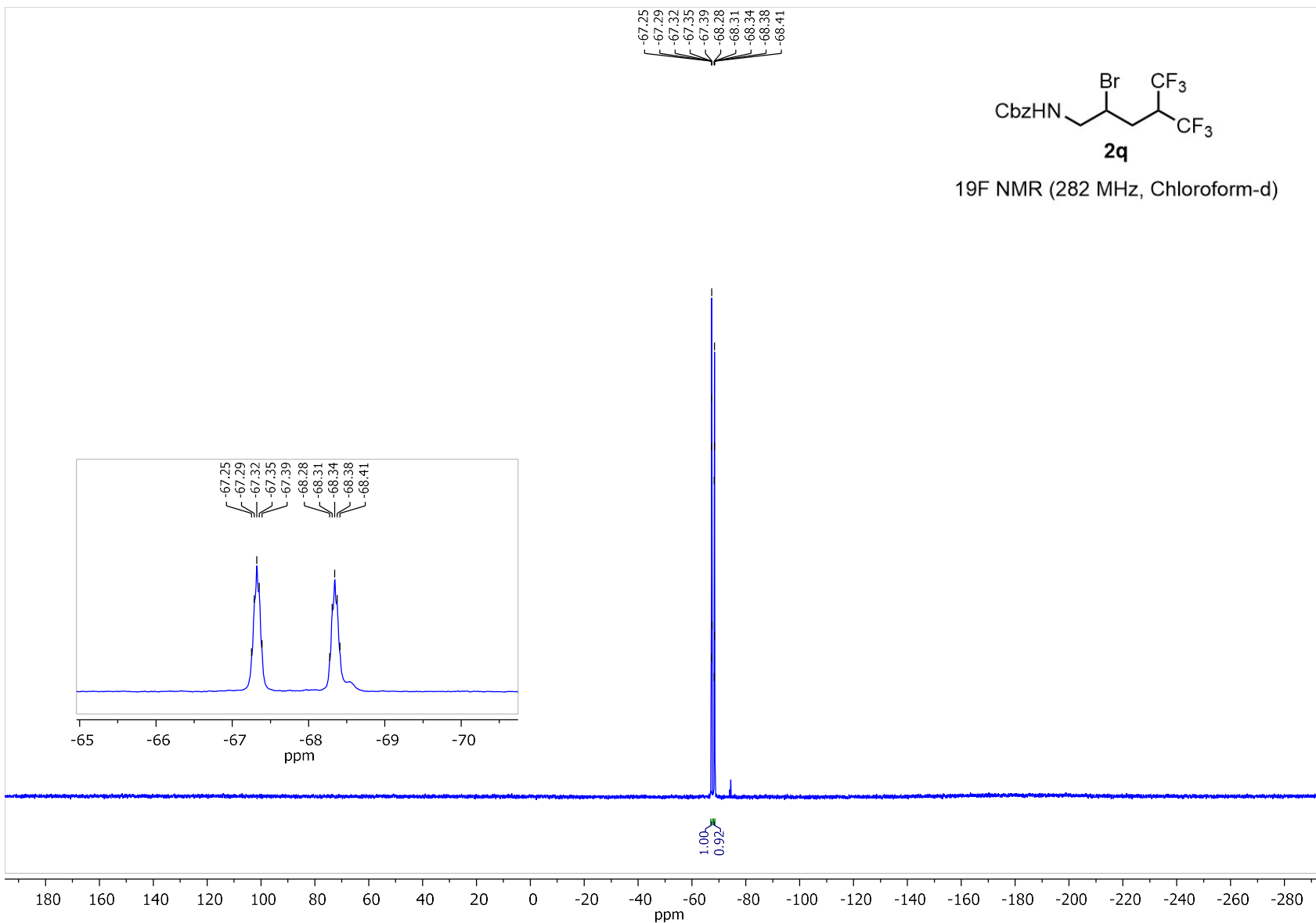


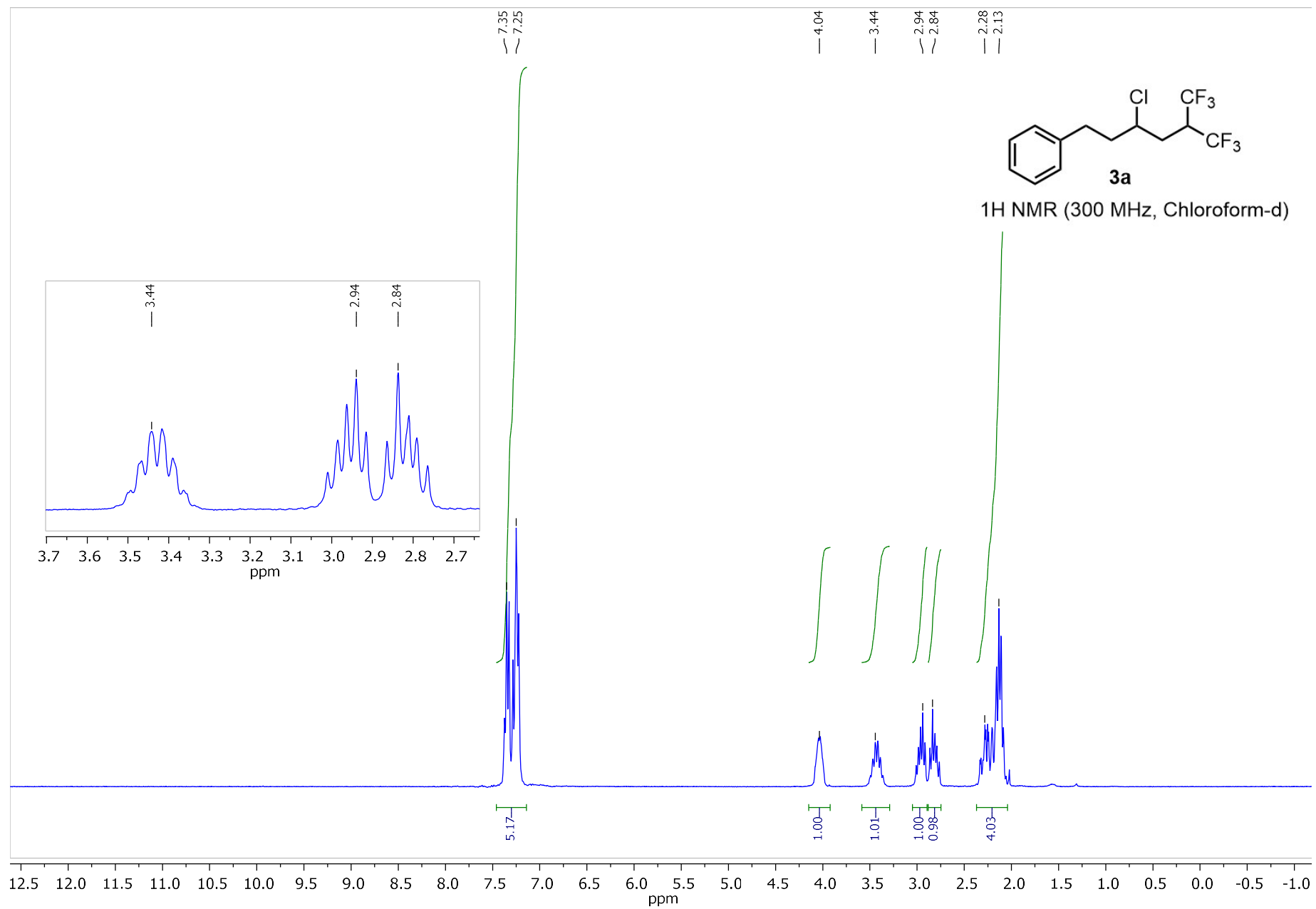


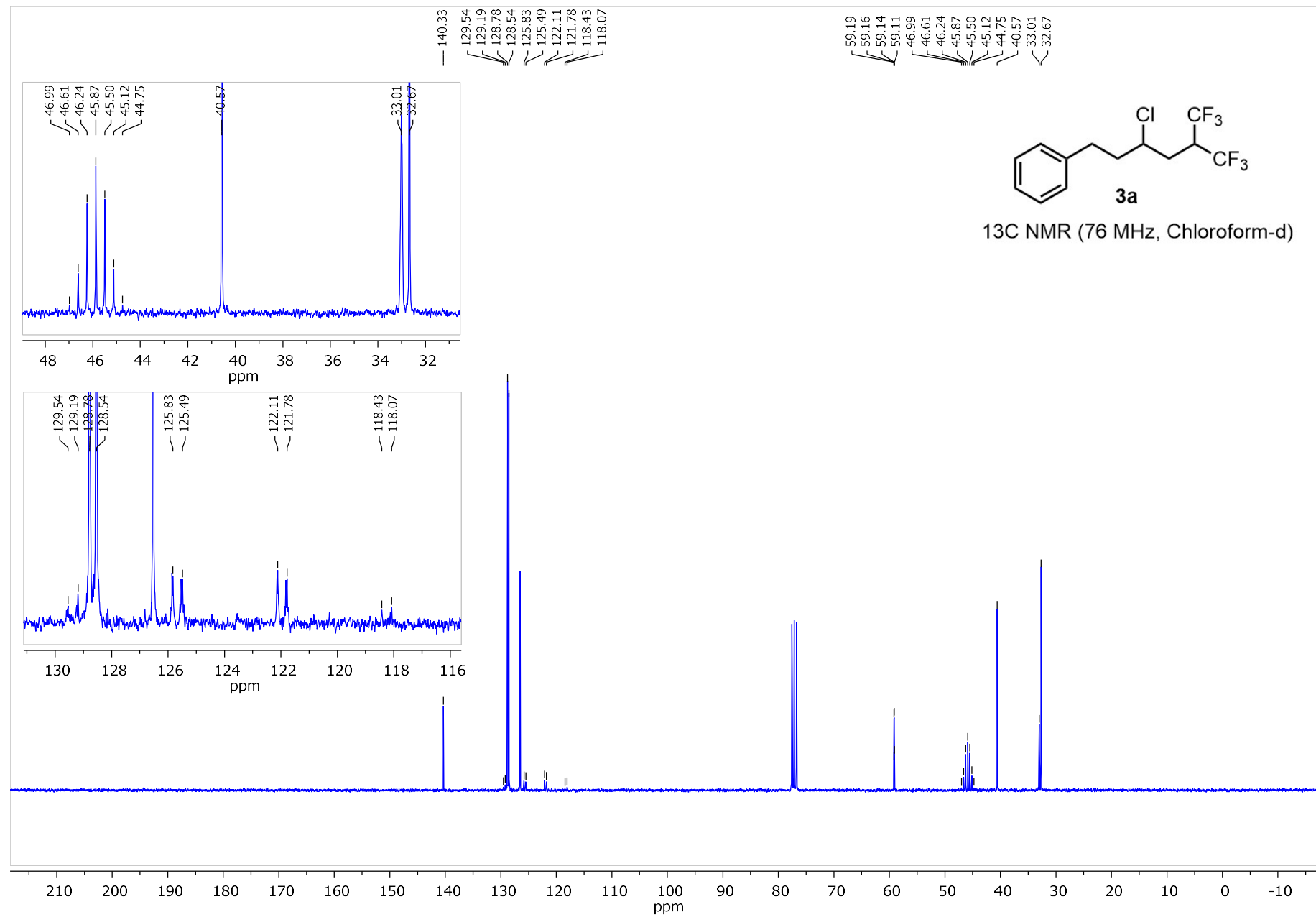


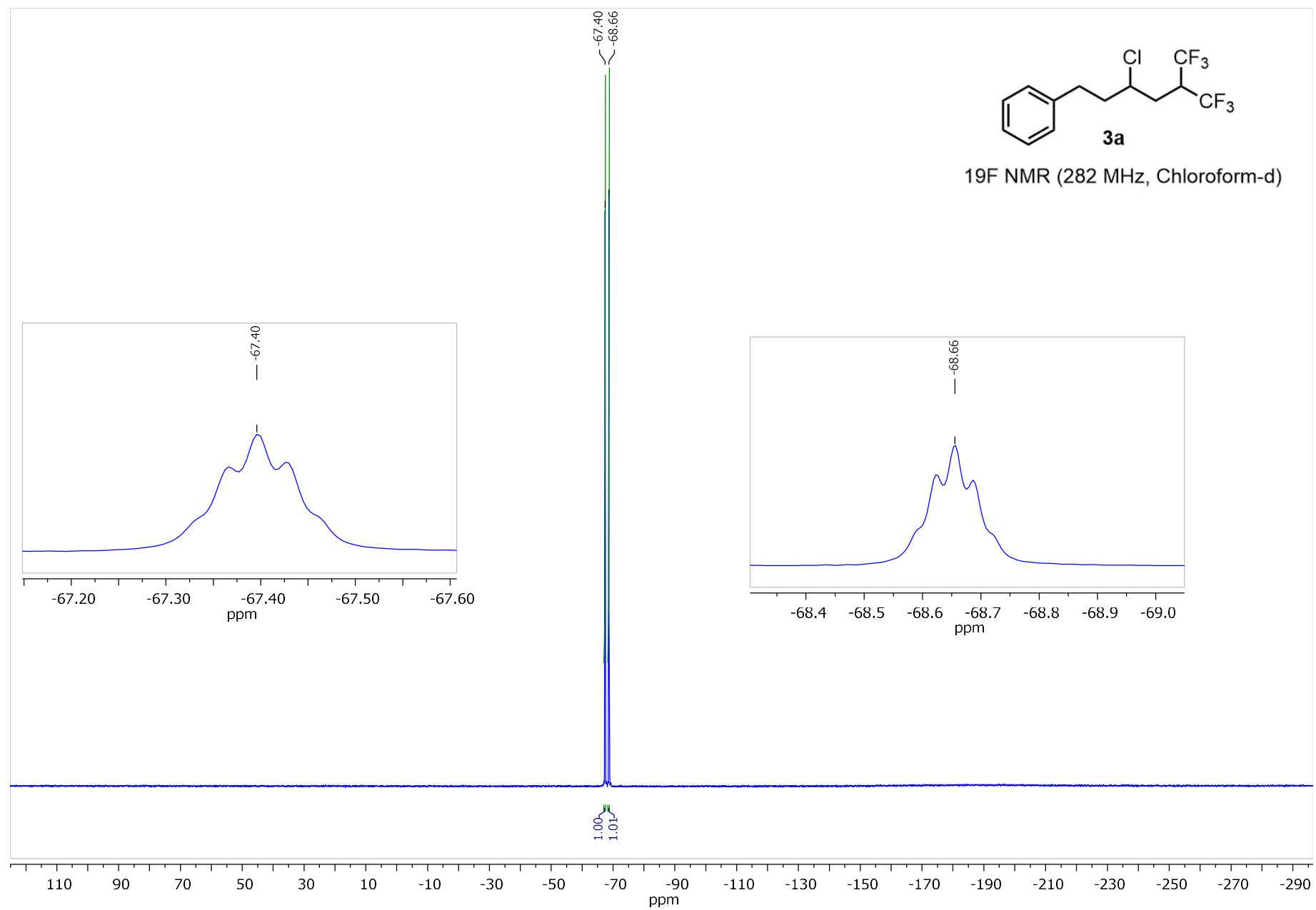


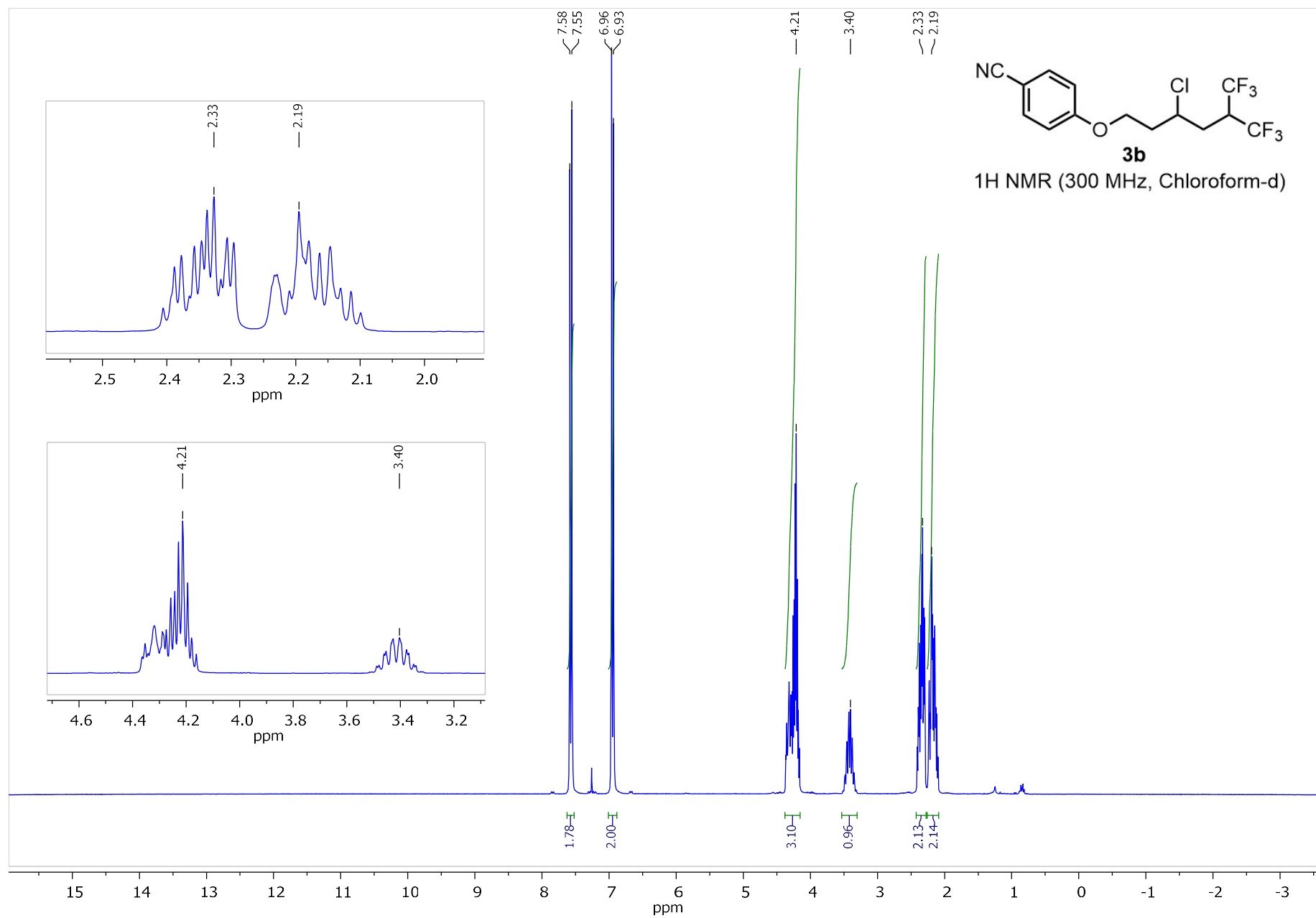


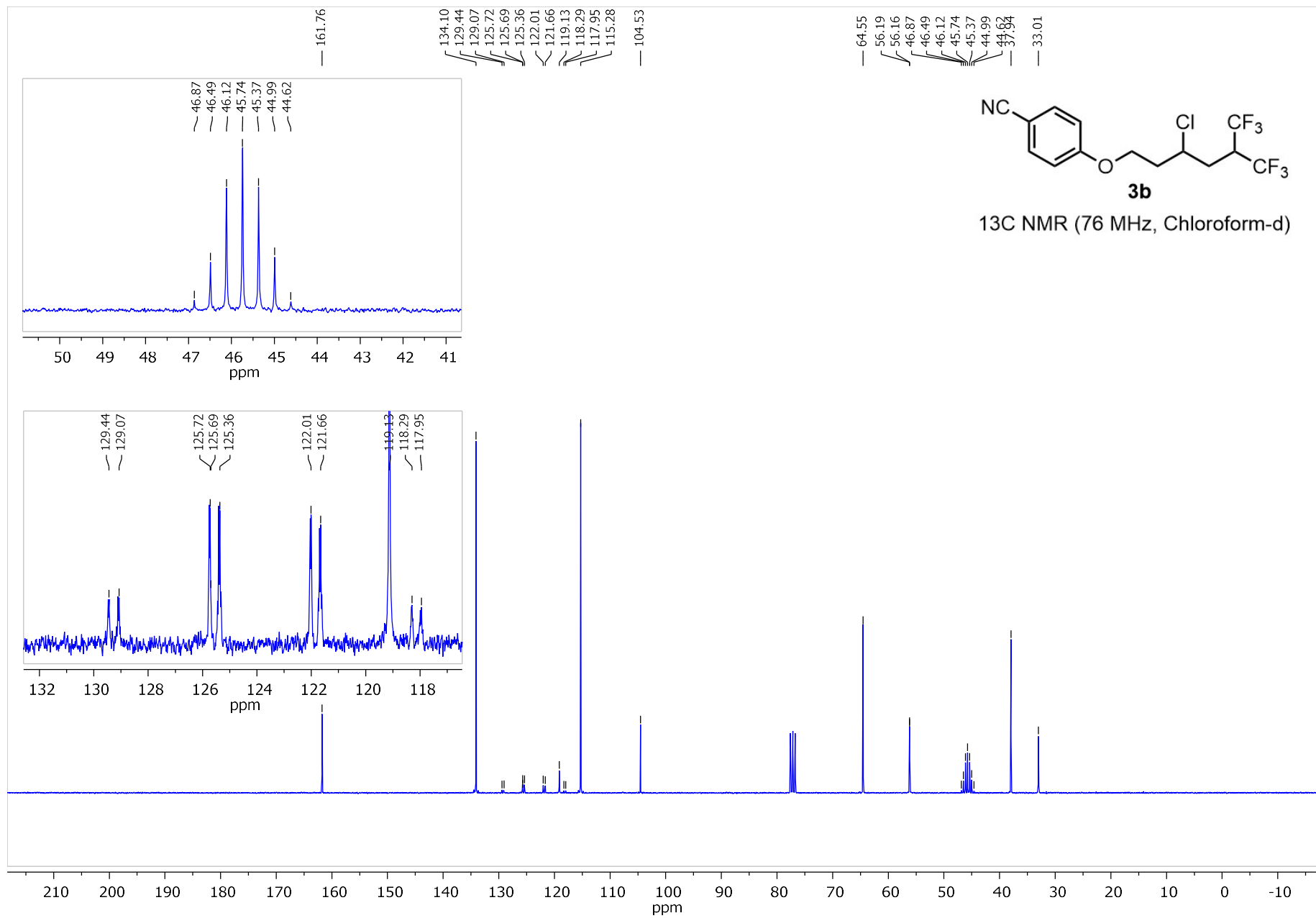


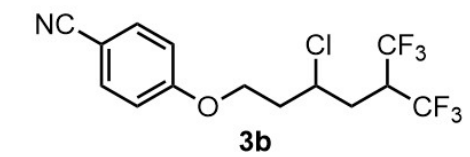




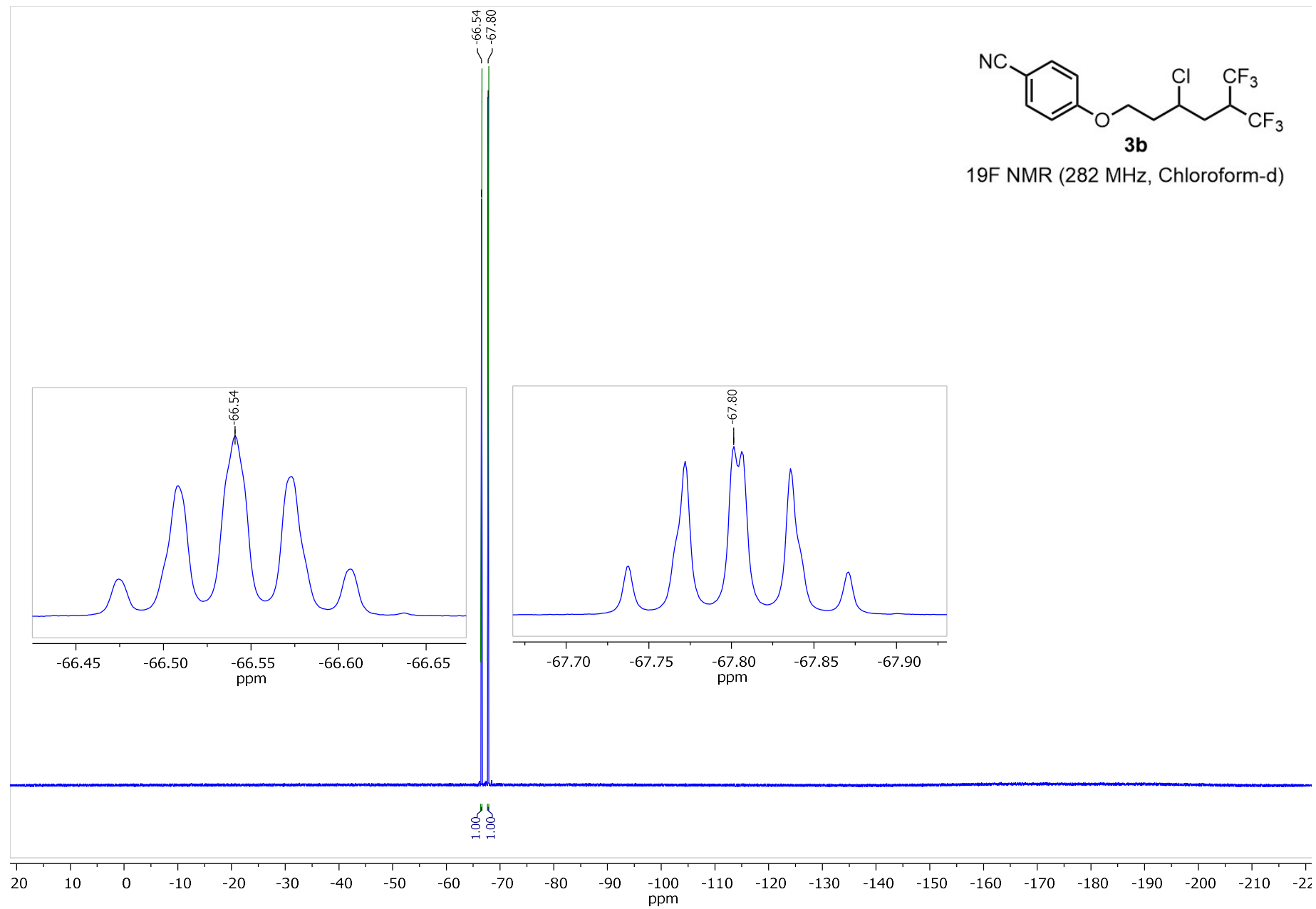


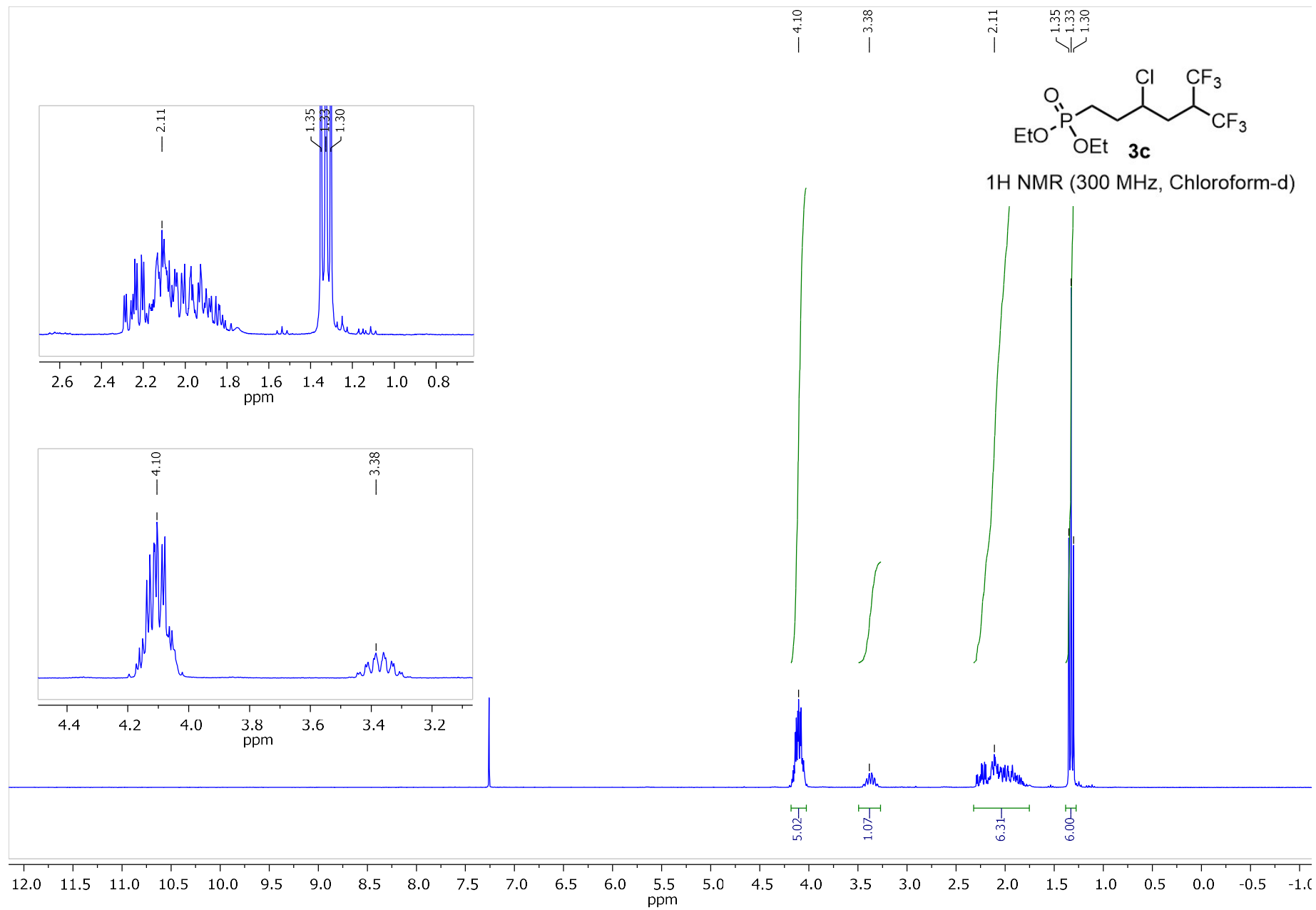


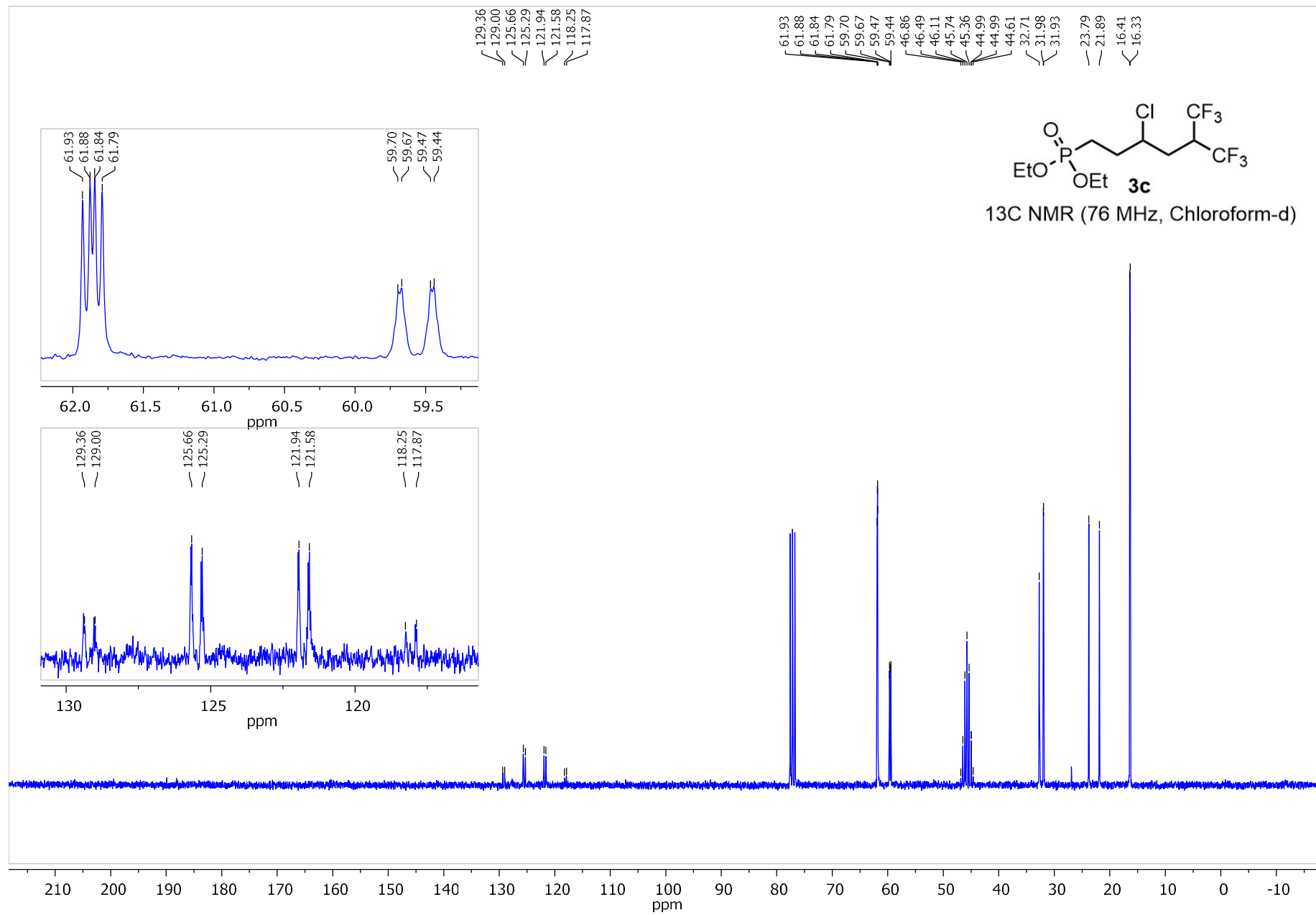


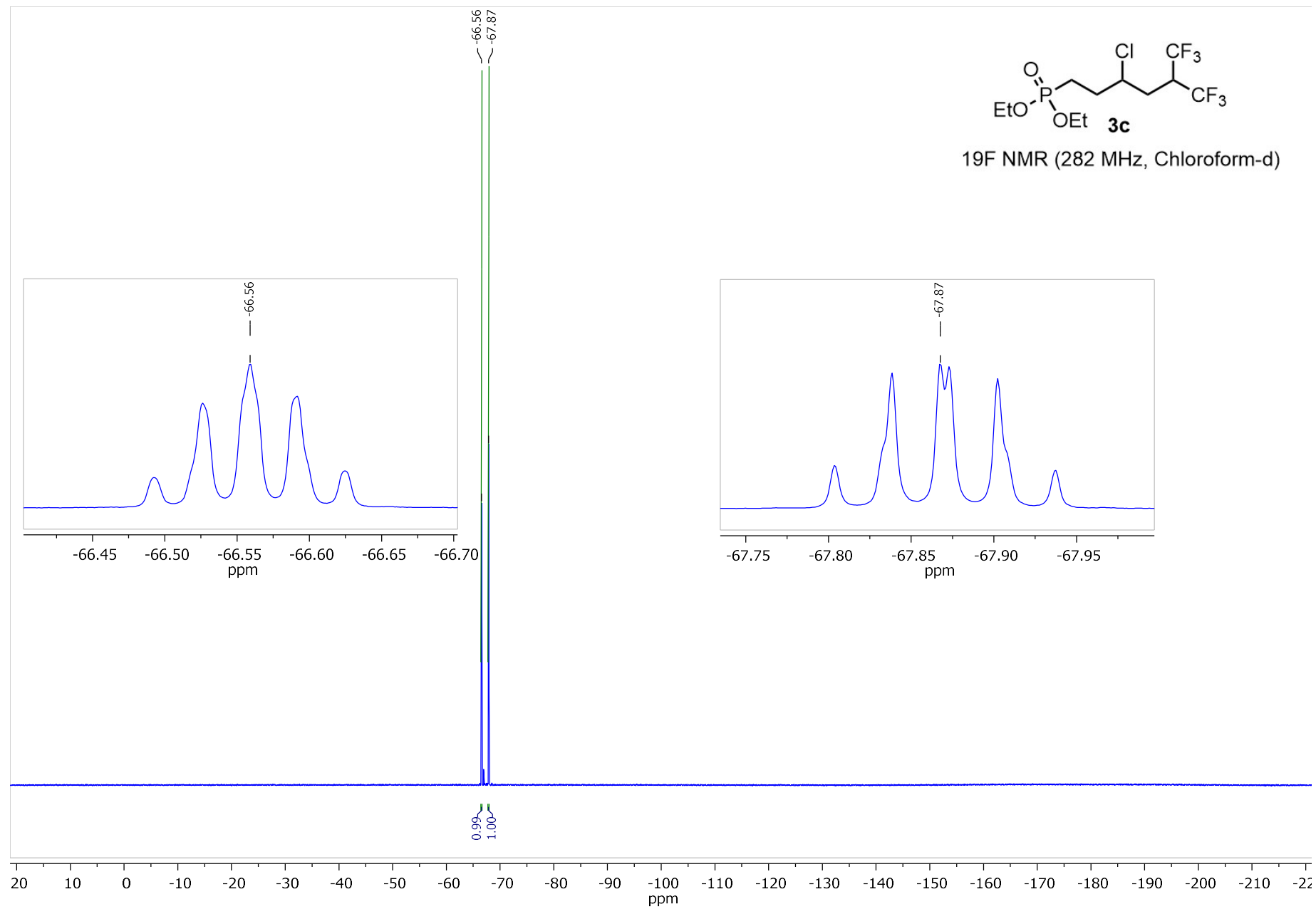


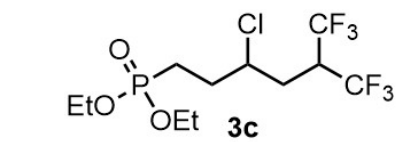
¹⁹F NMR (282 MHz, Chloroform-d)



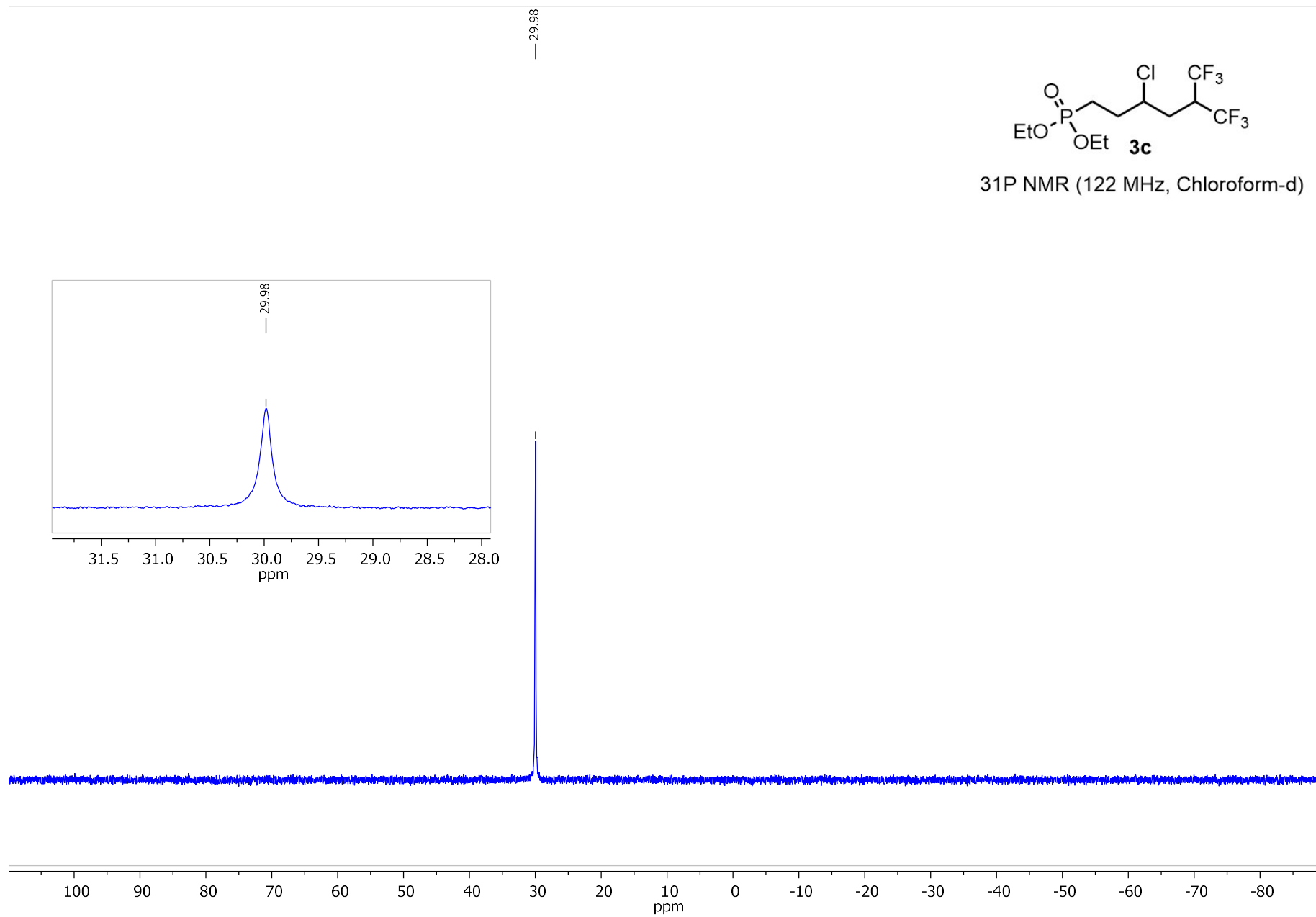


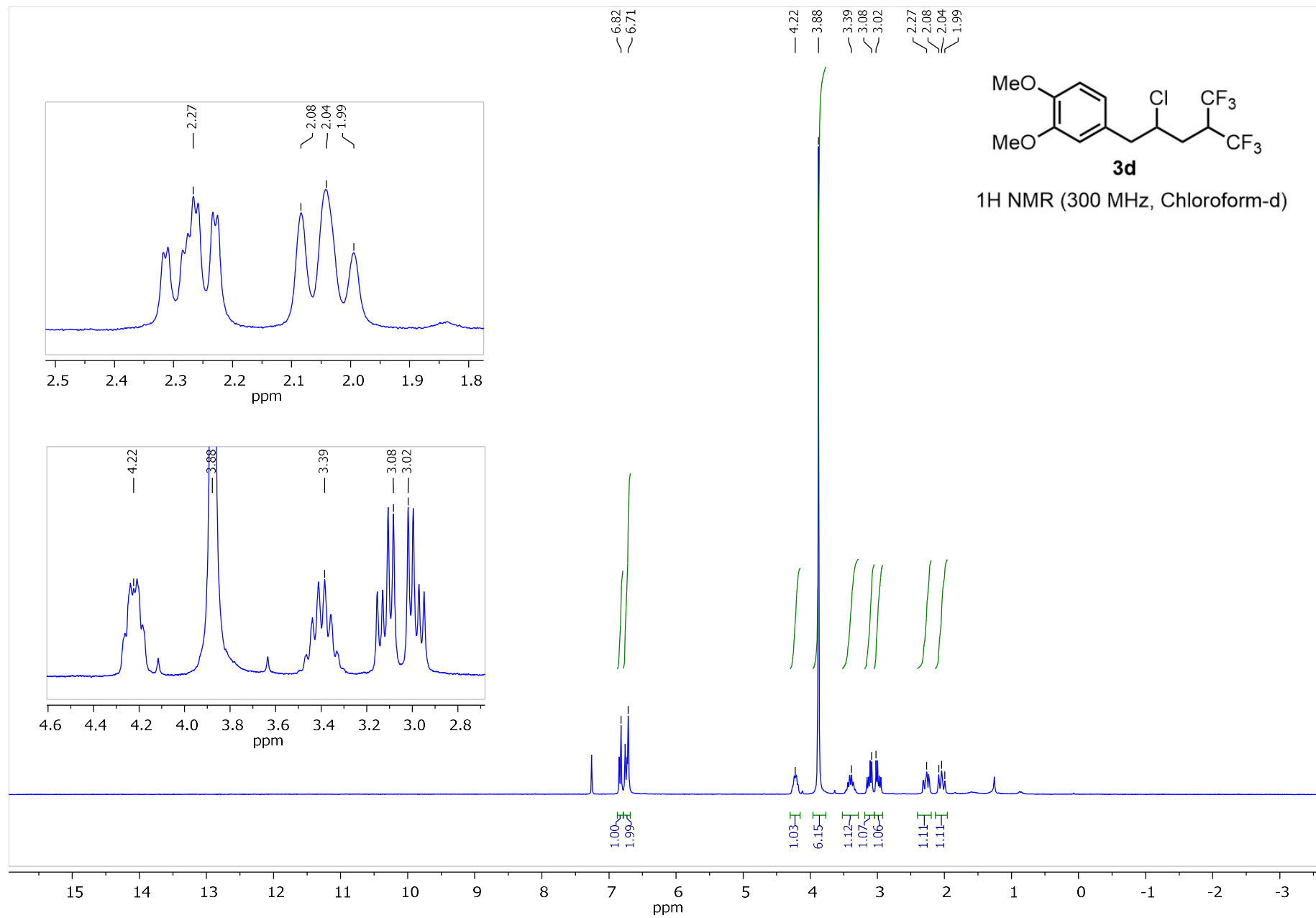


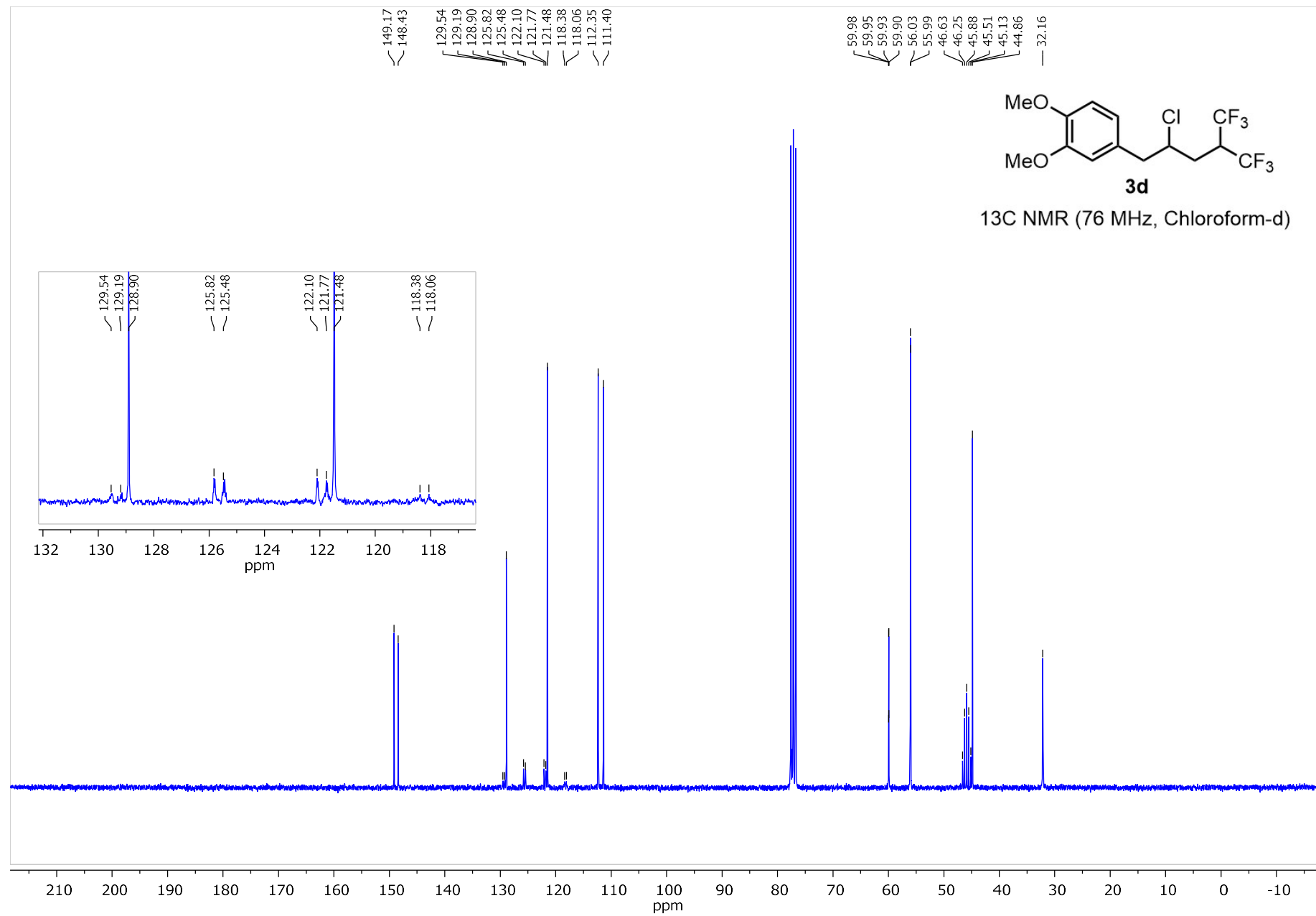


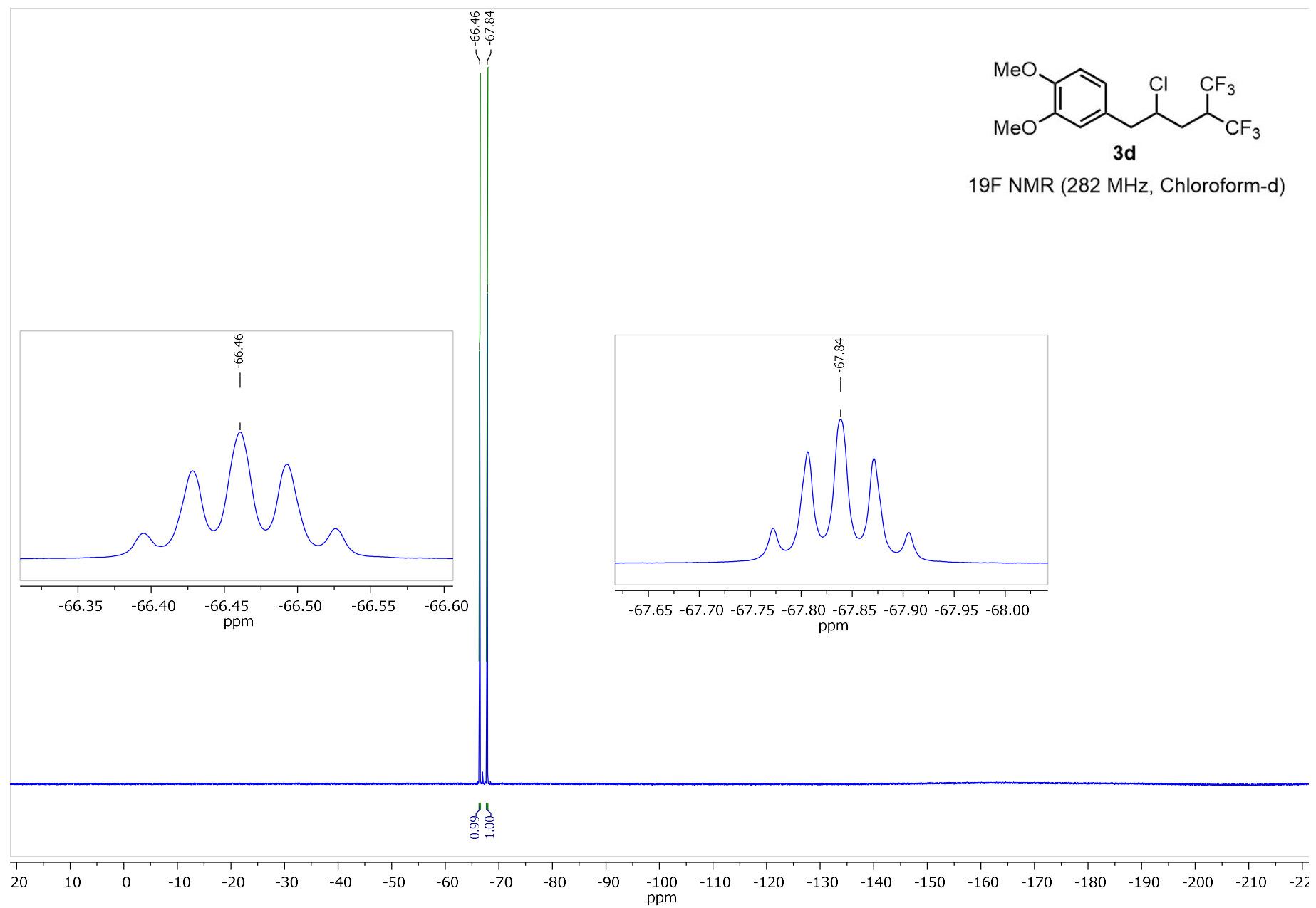


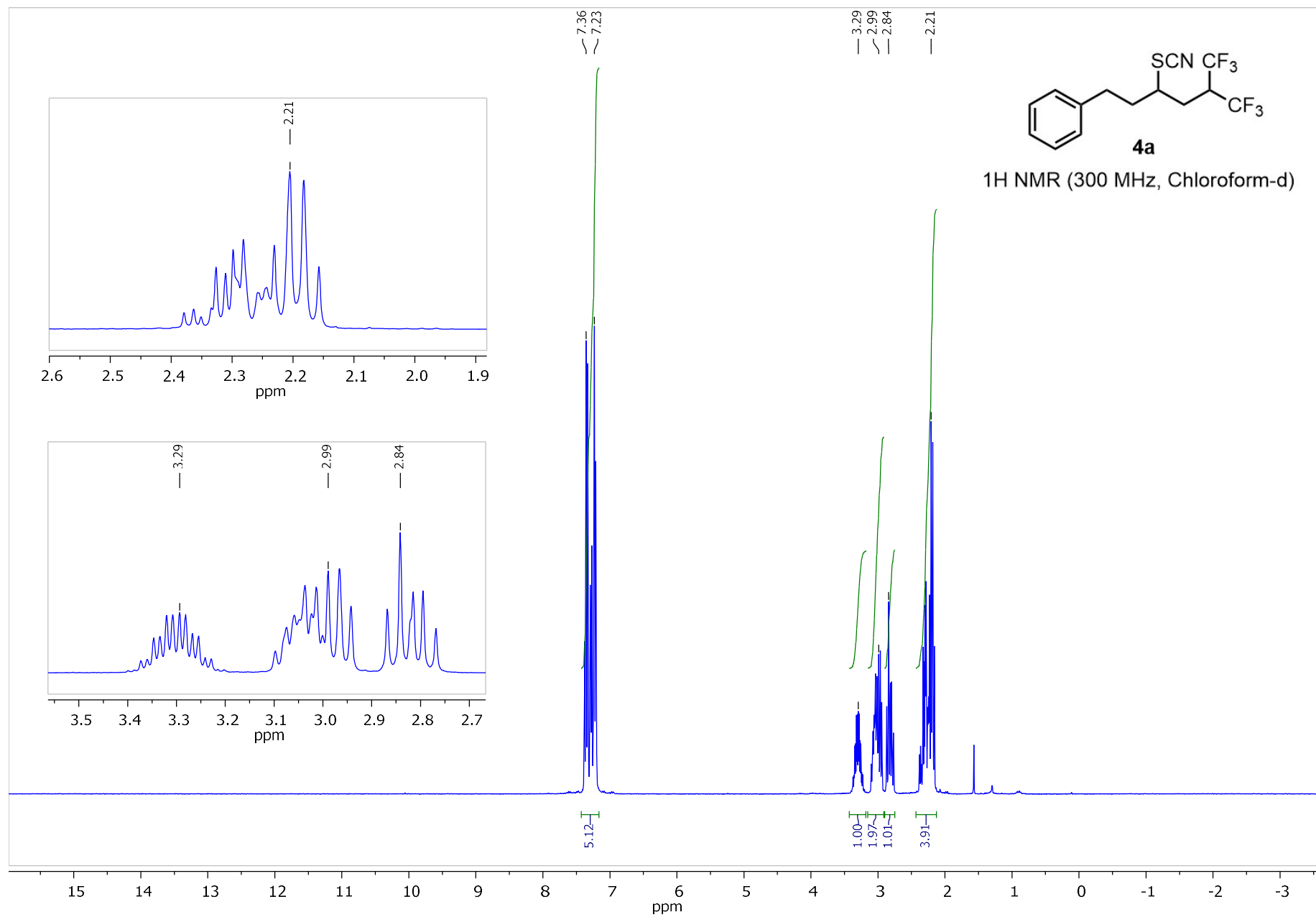
^{31}P NMR (122 MHz, Chloroform-d)

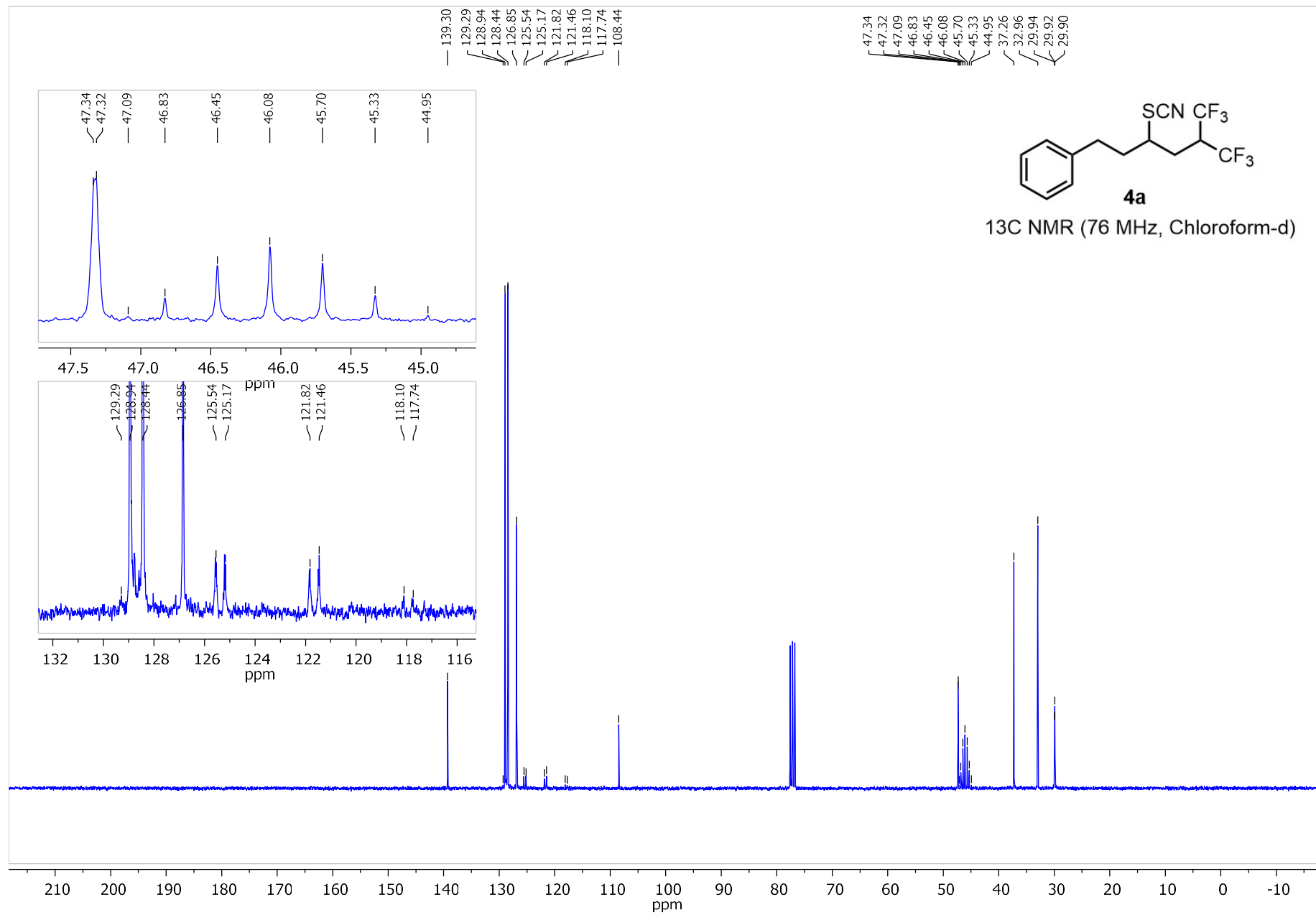


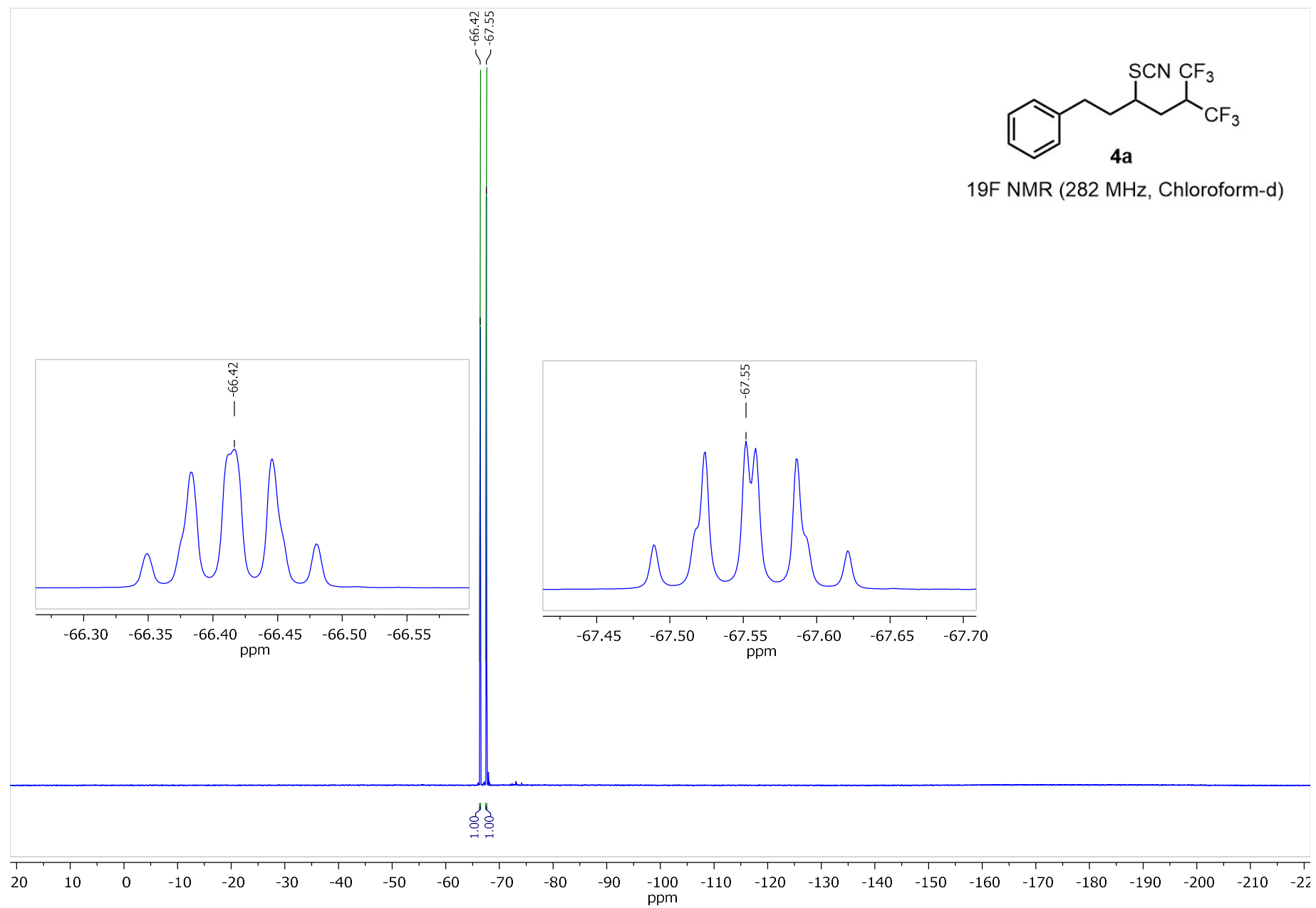


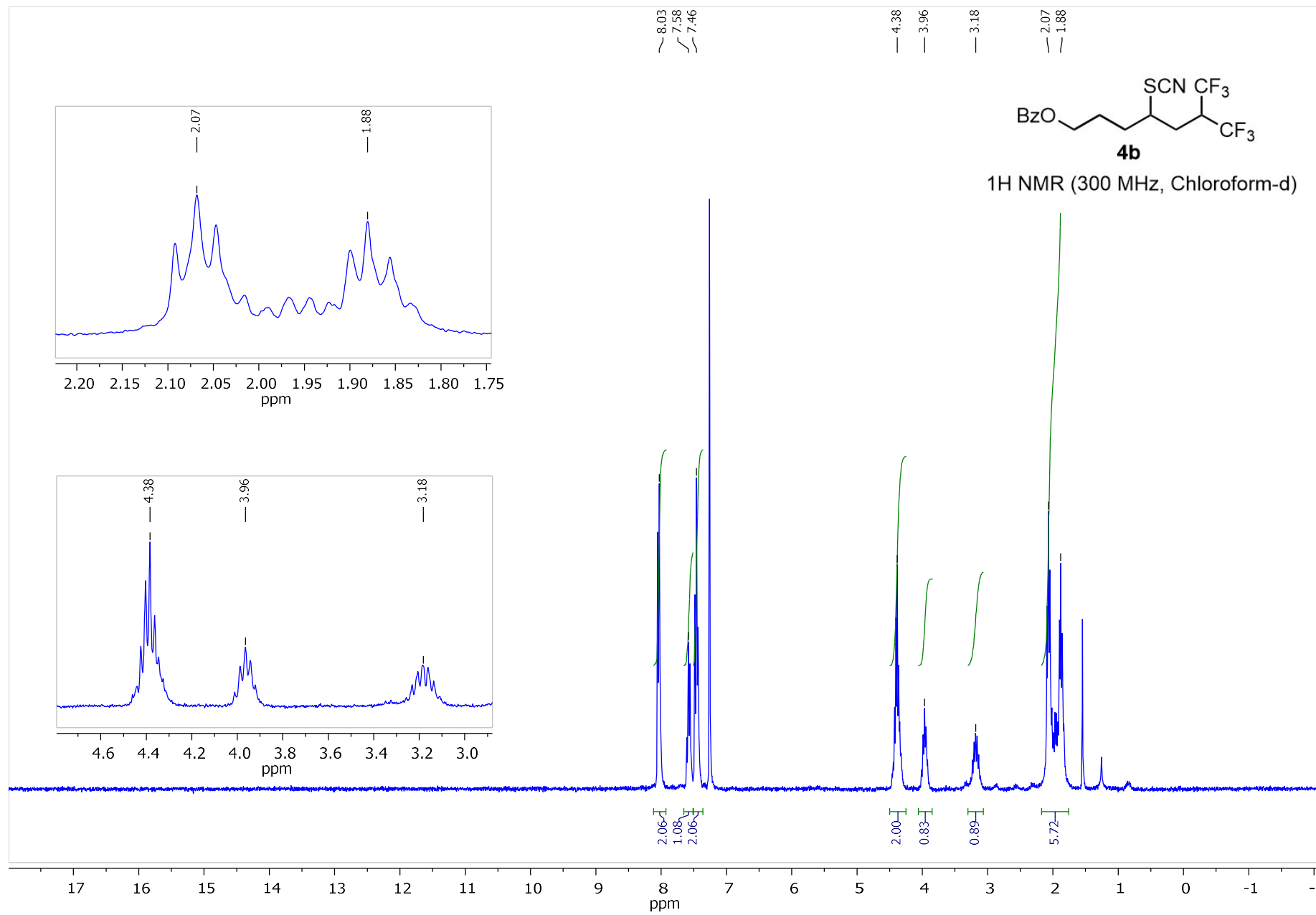


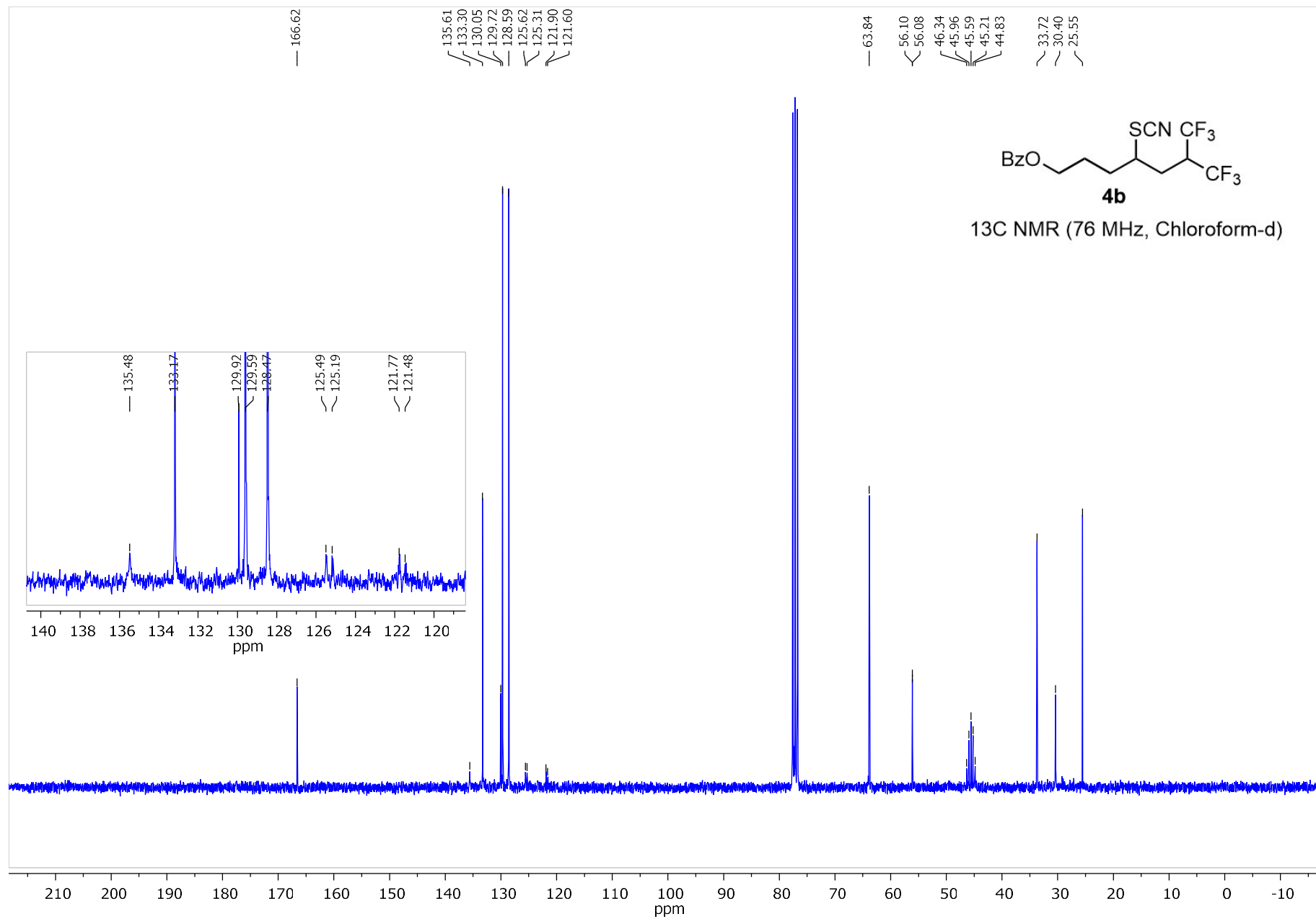


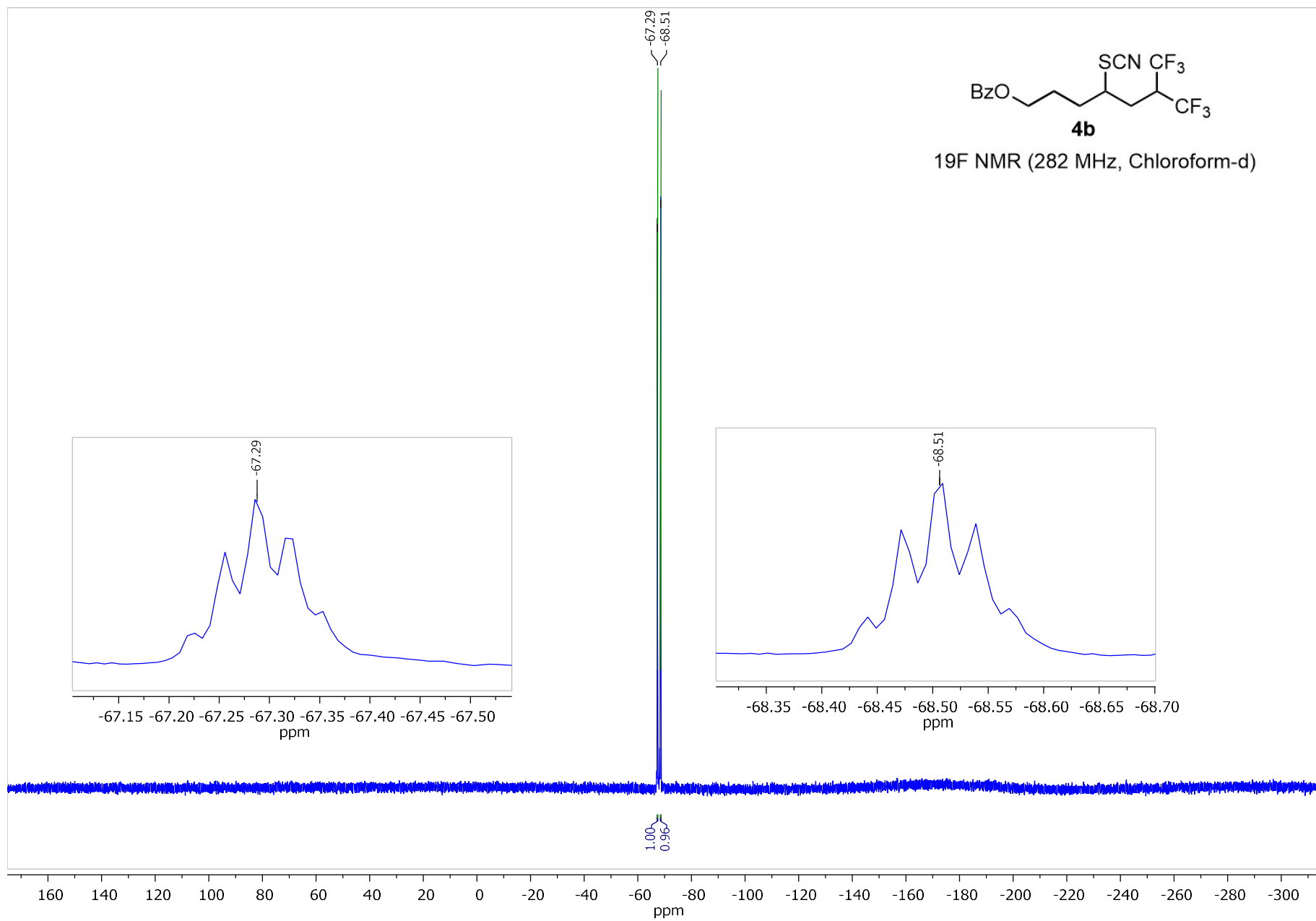


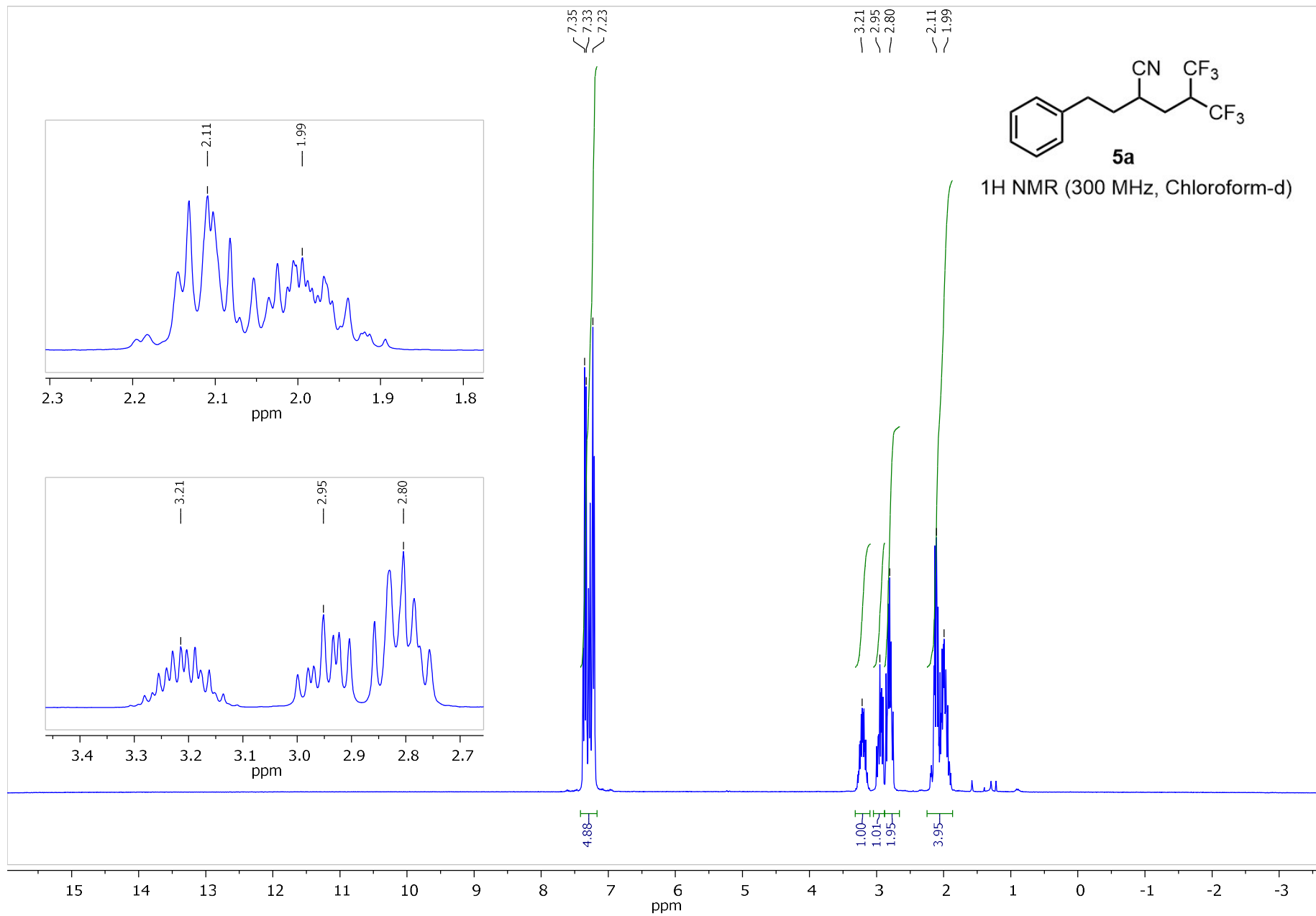


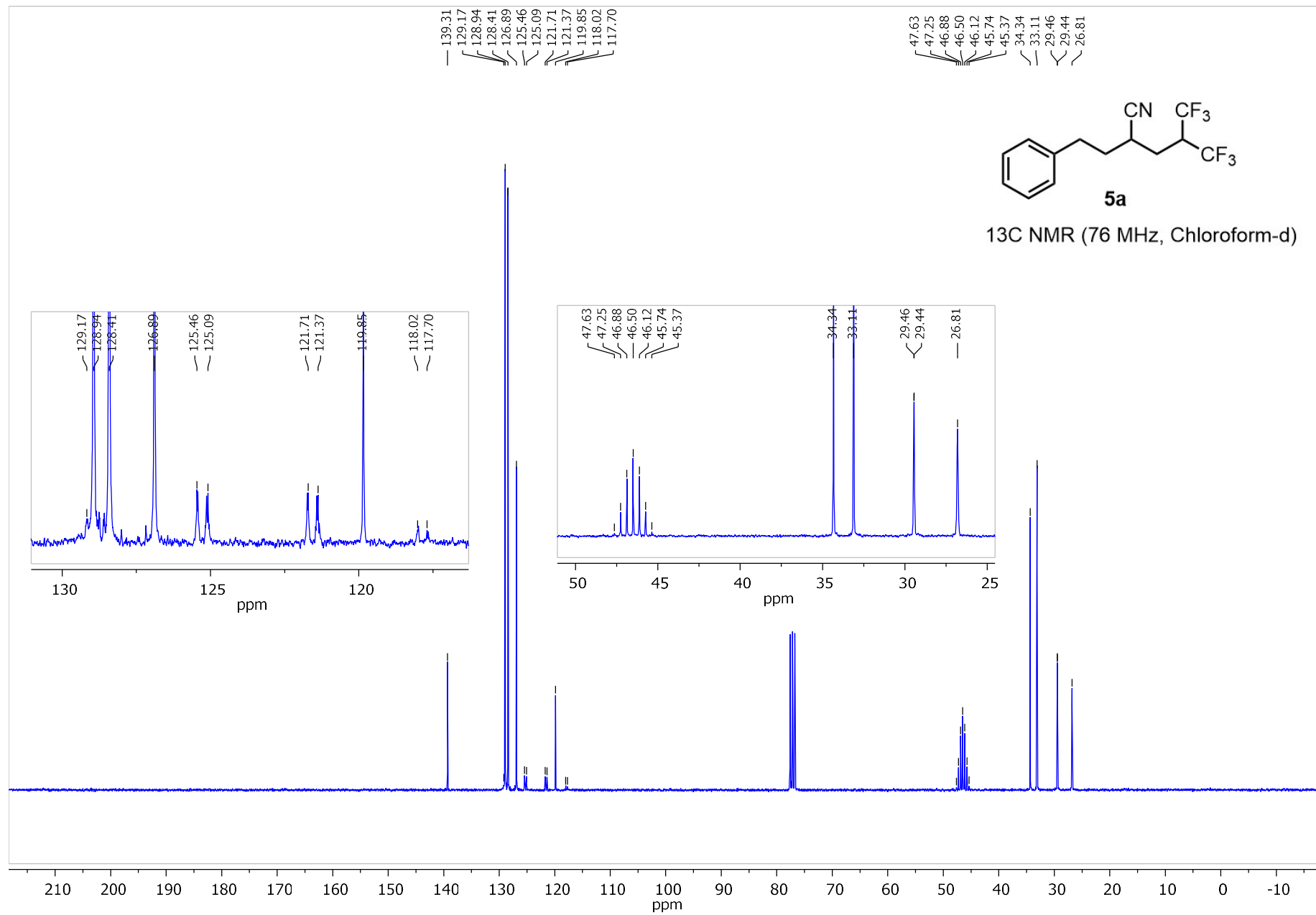


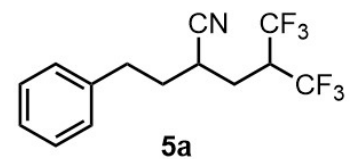




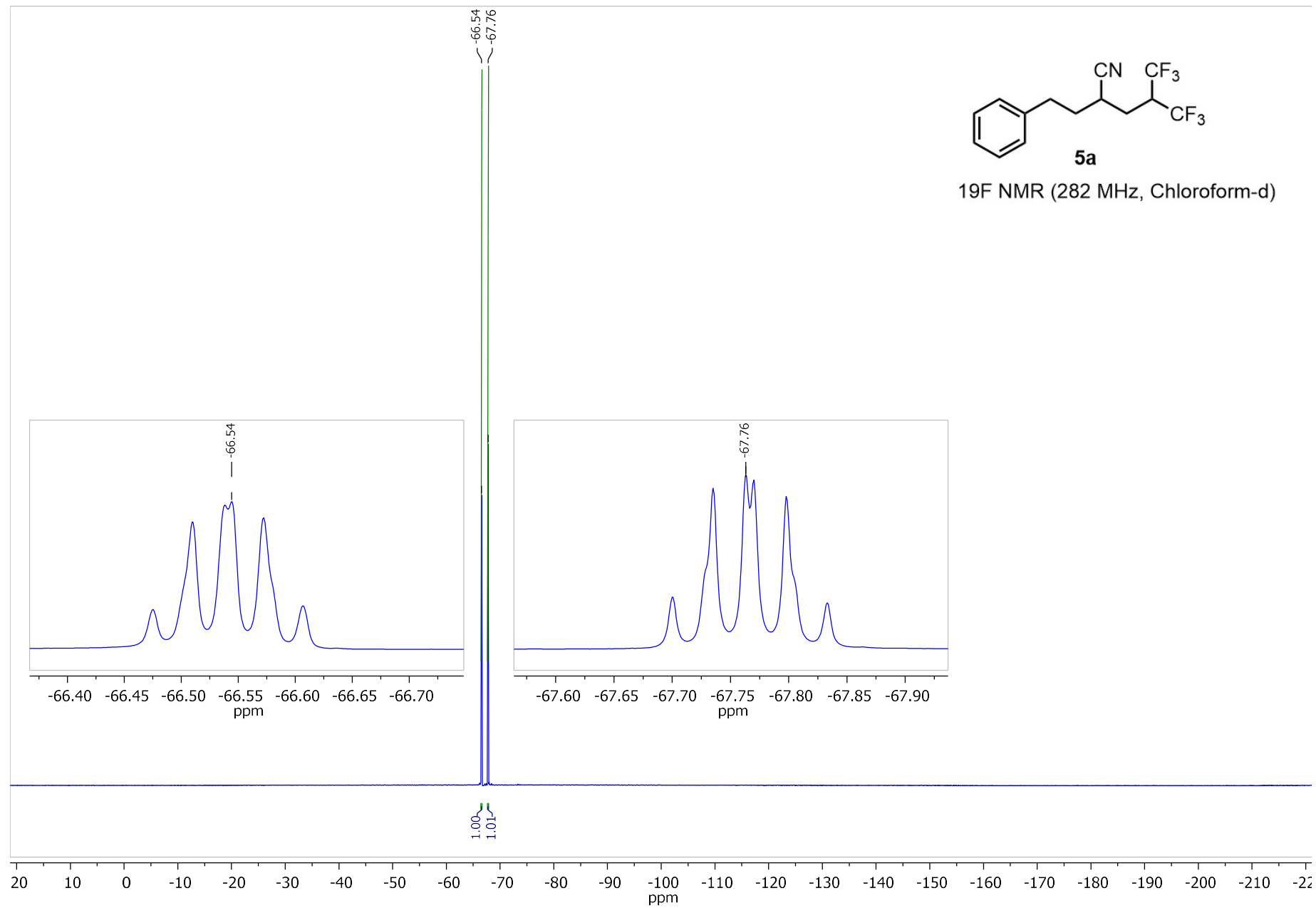


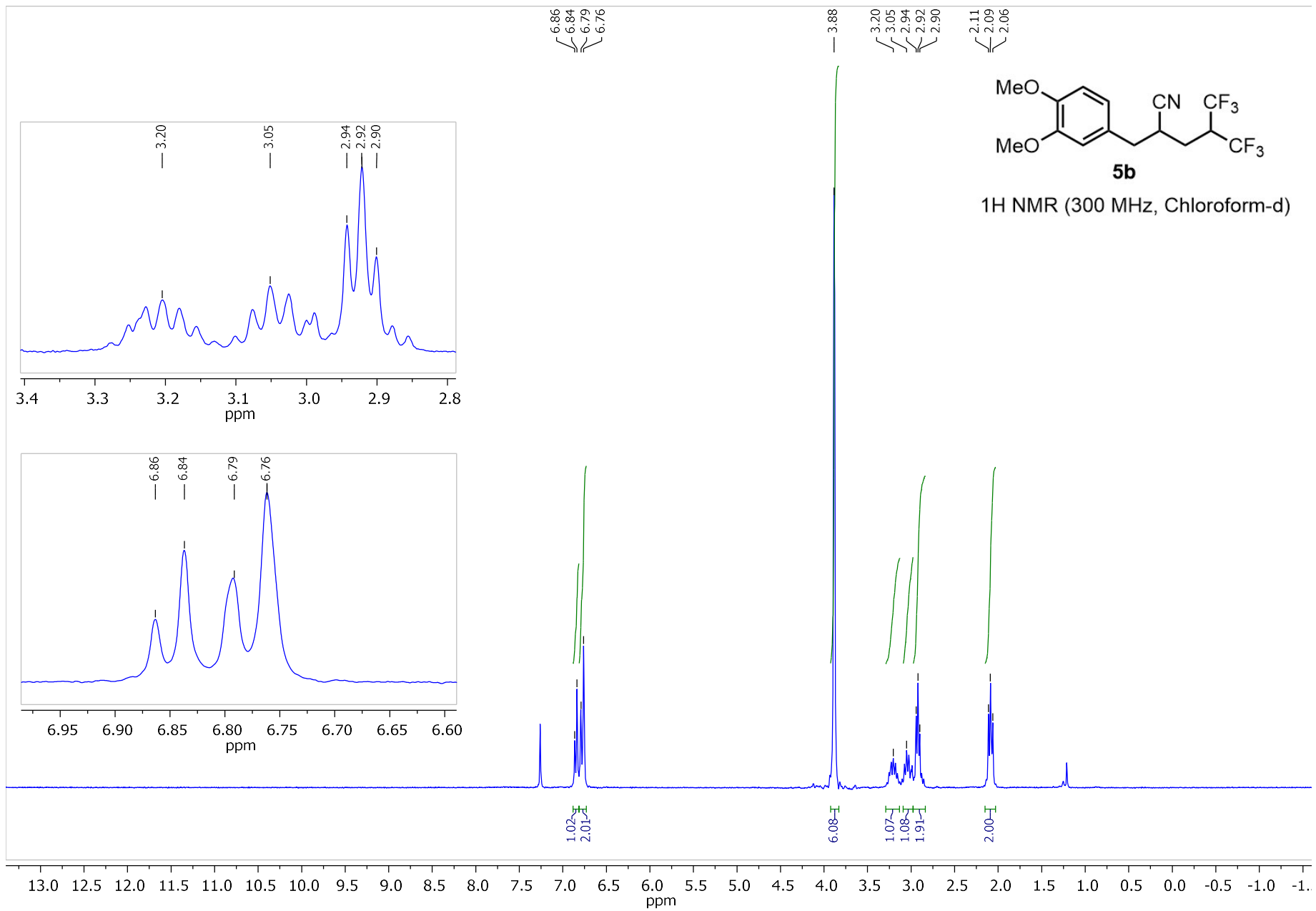


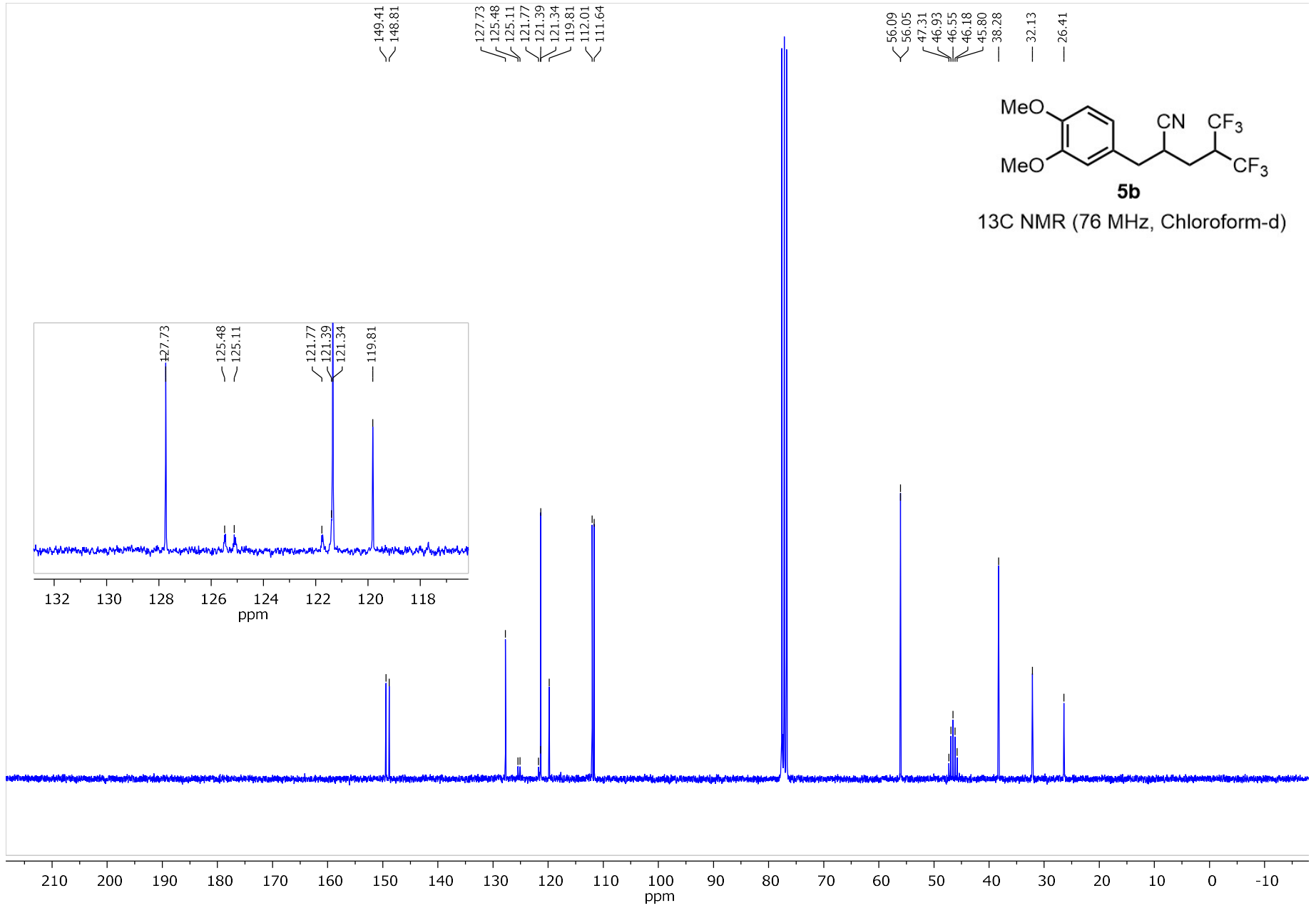


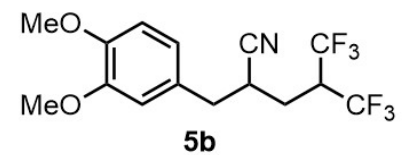


^{19}F NMR (282 MHz, Chloroform- d)

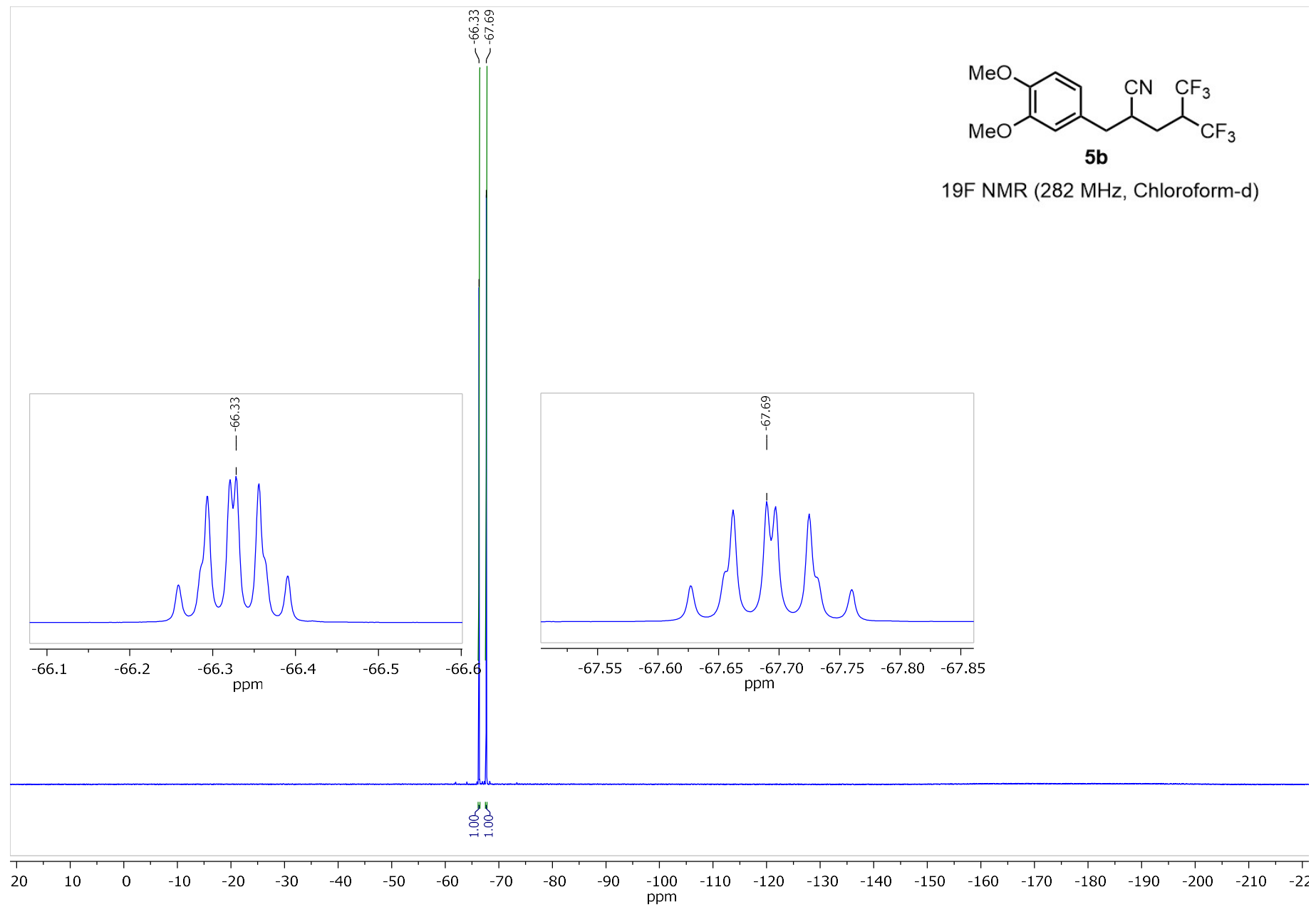


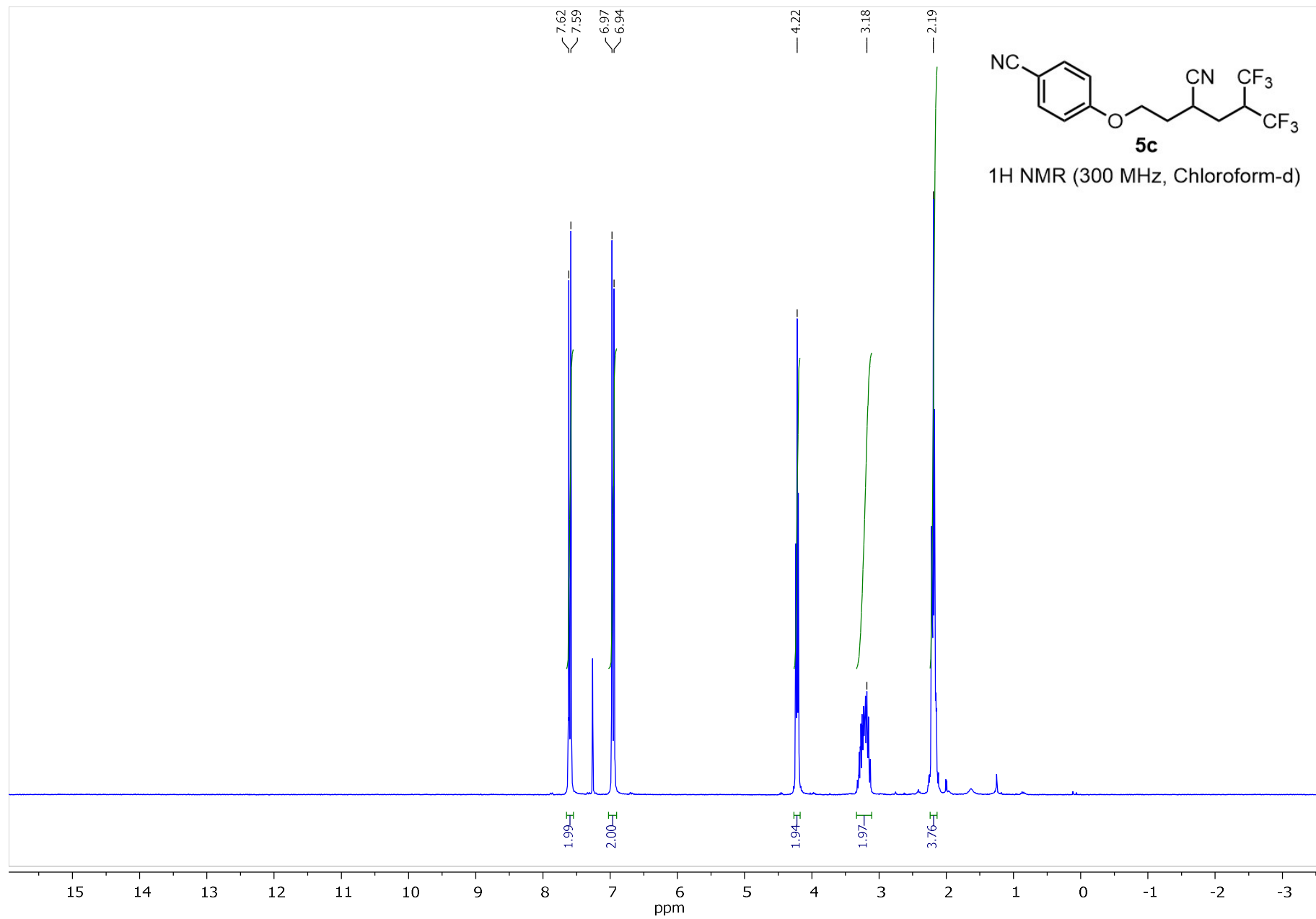


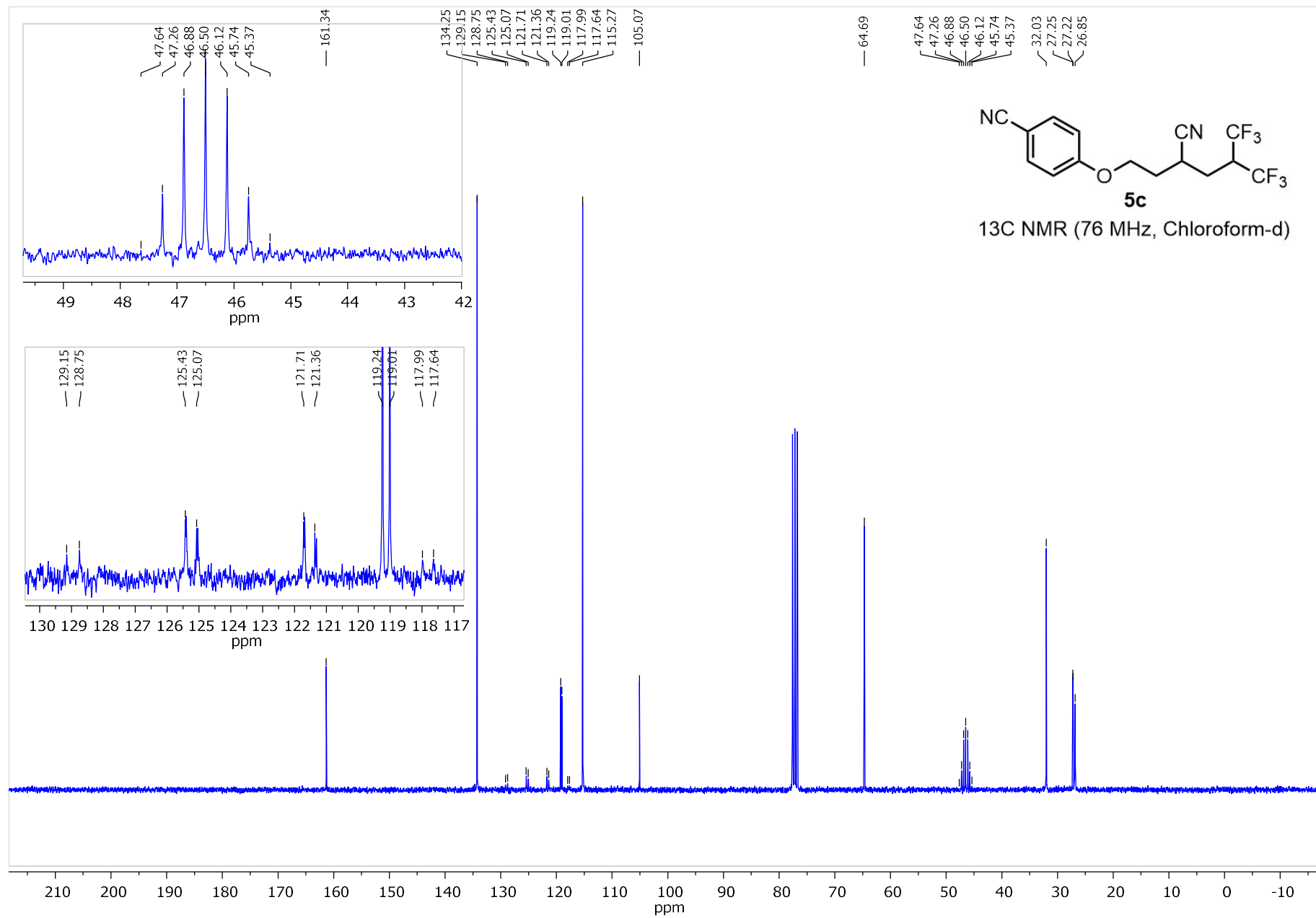


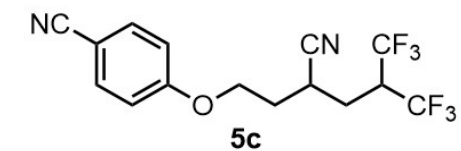


19F NMR (282 MHz, Chloroform-d)

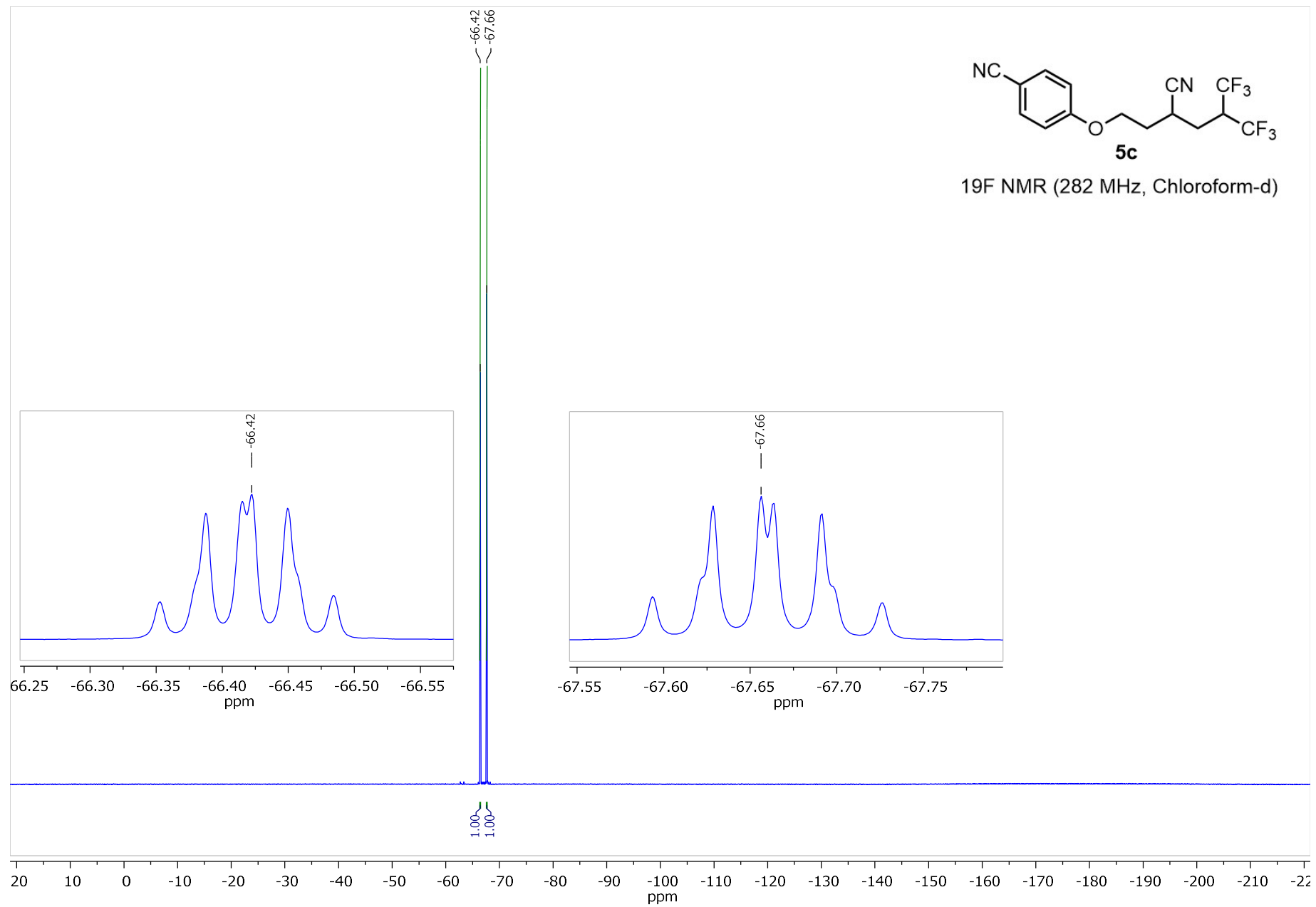


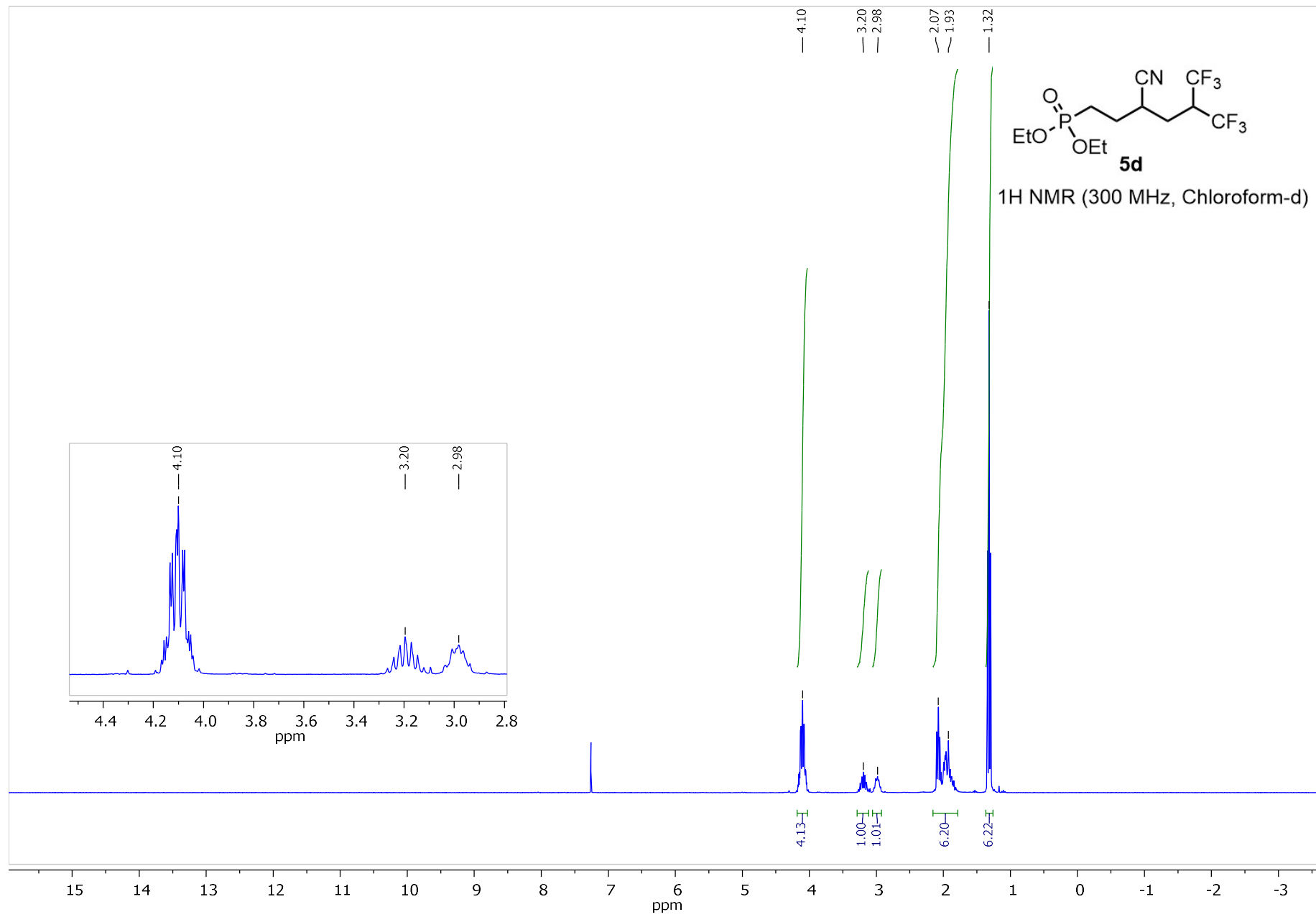


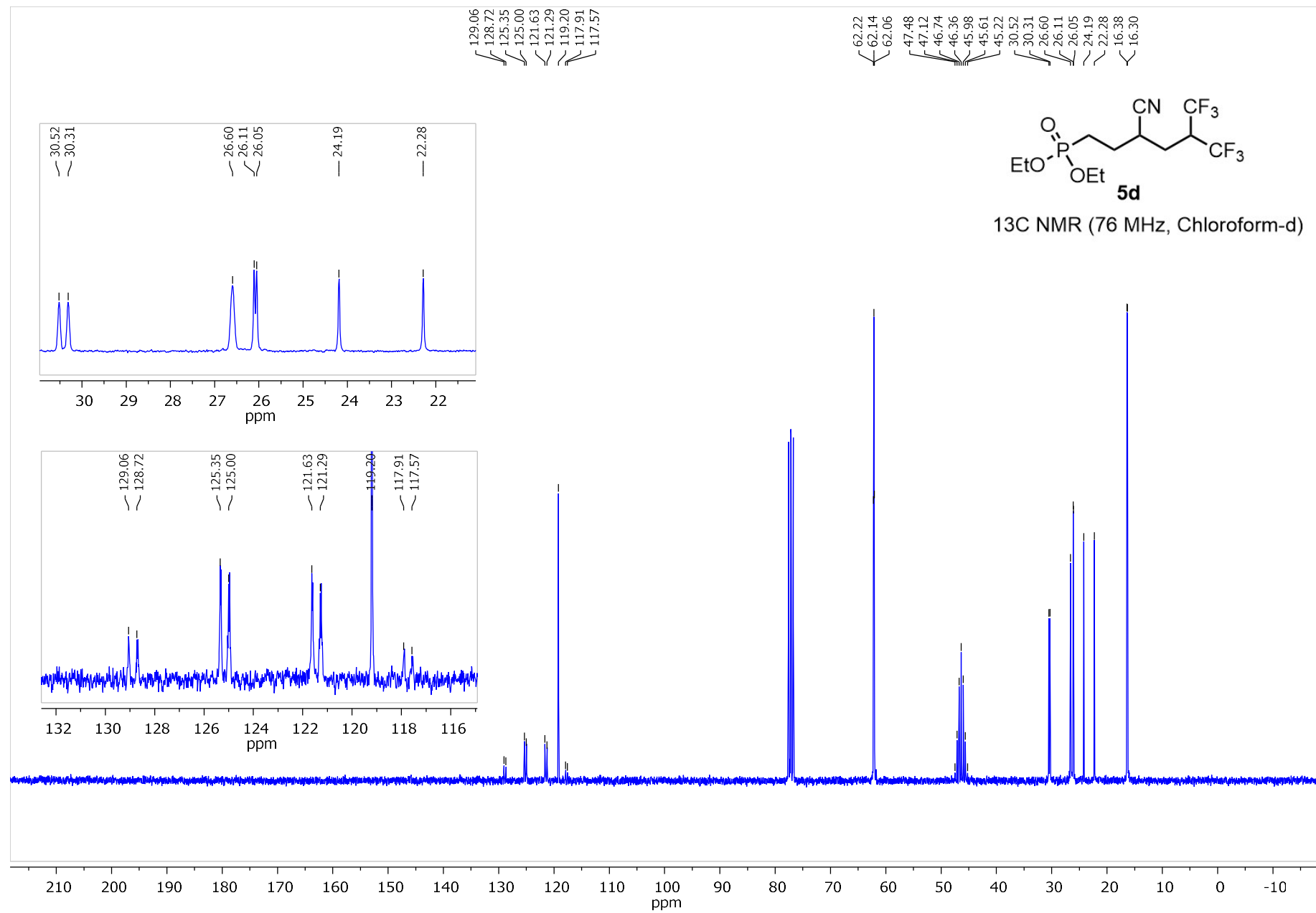


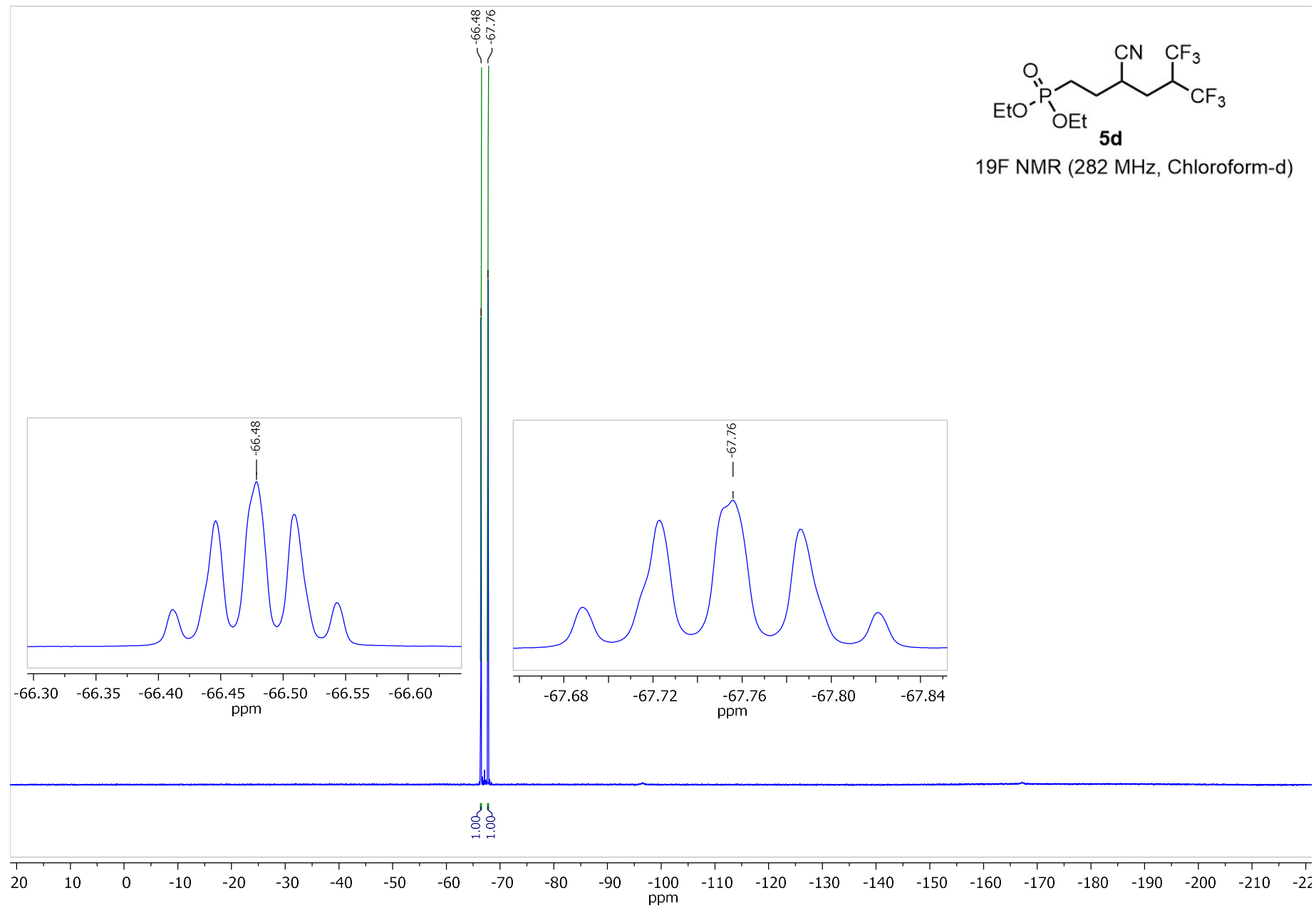


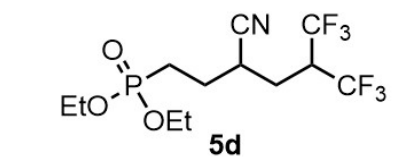
5c
19F NMR (282 MHz, Chloroform-d)



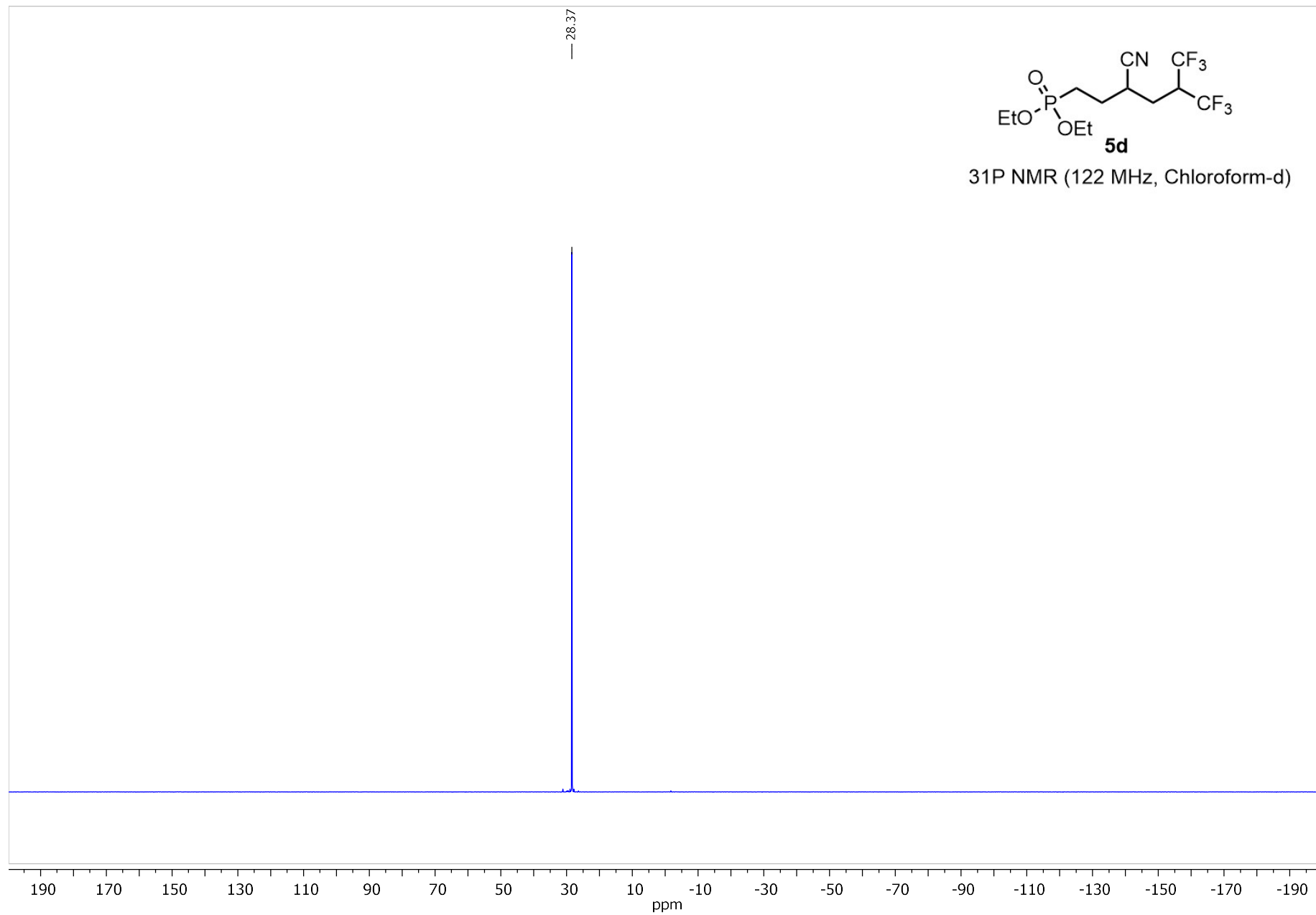


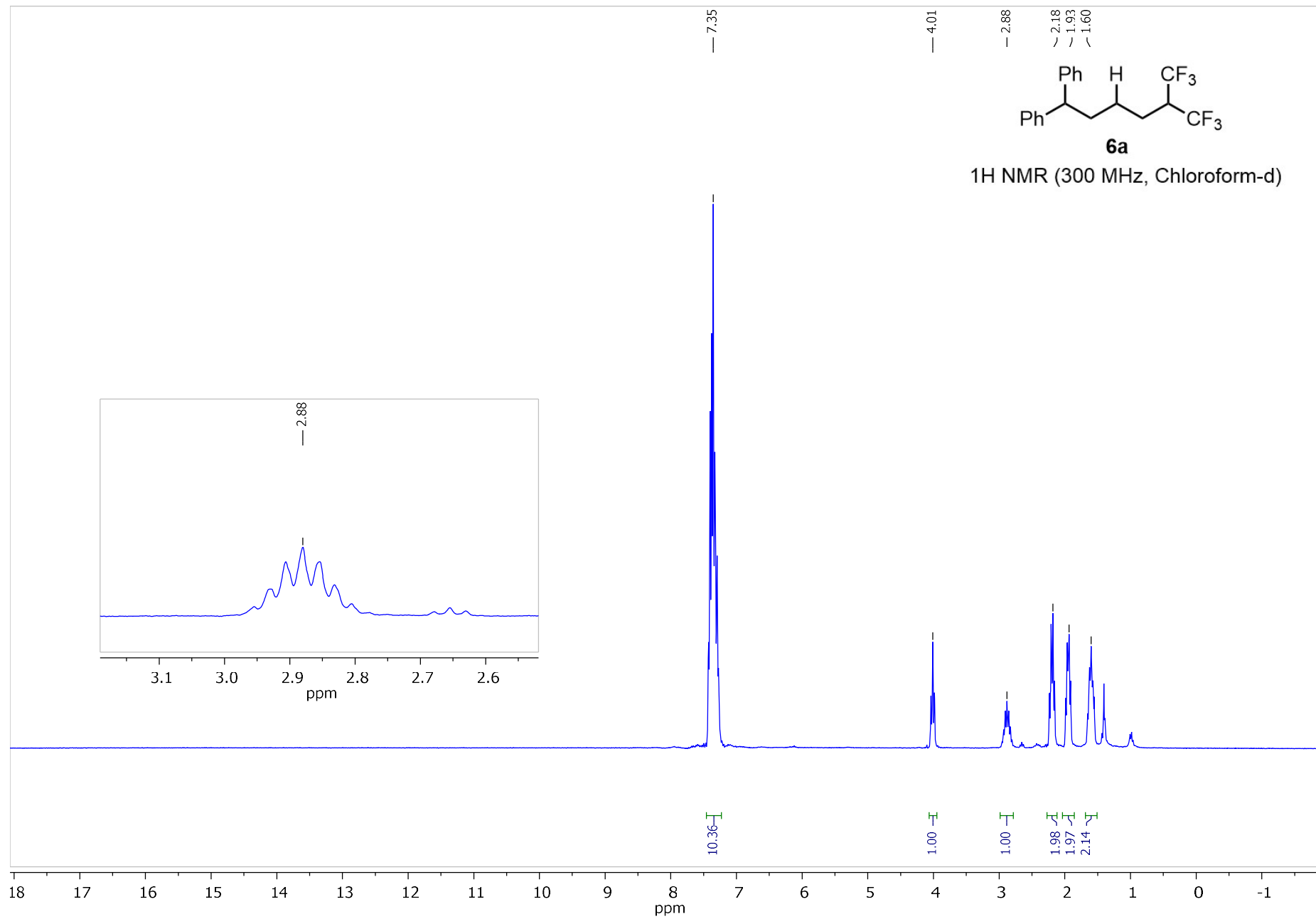


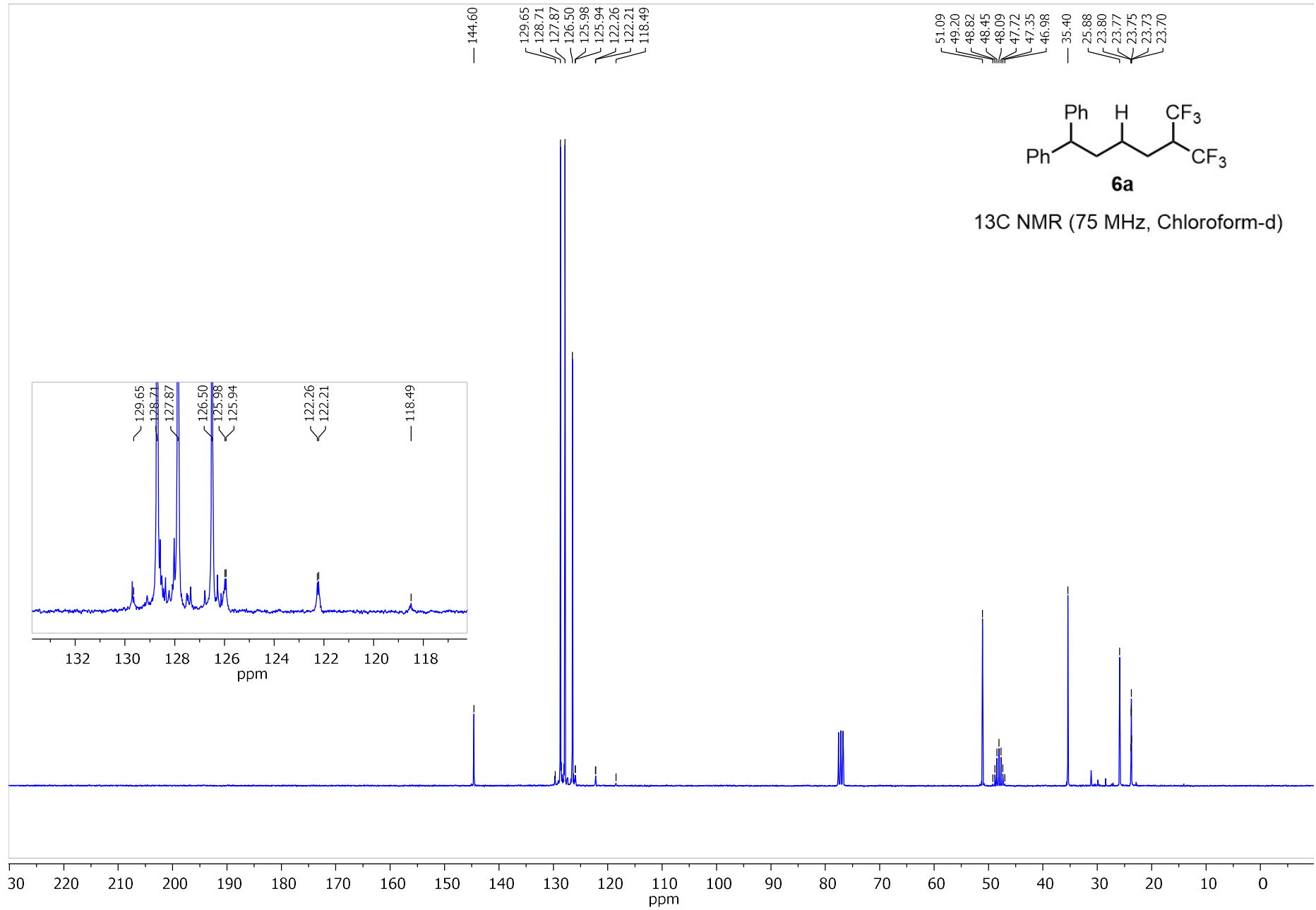


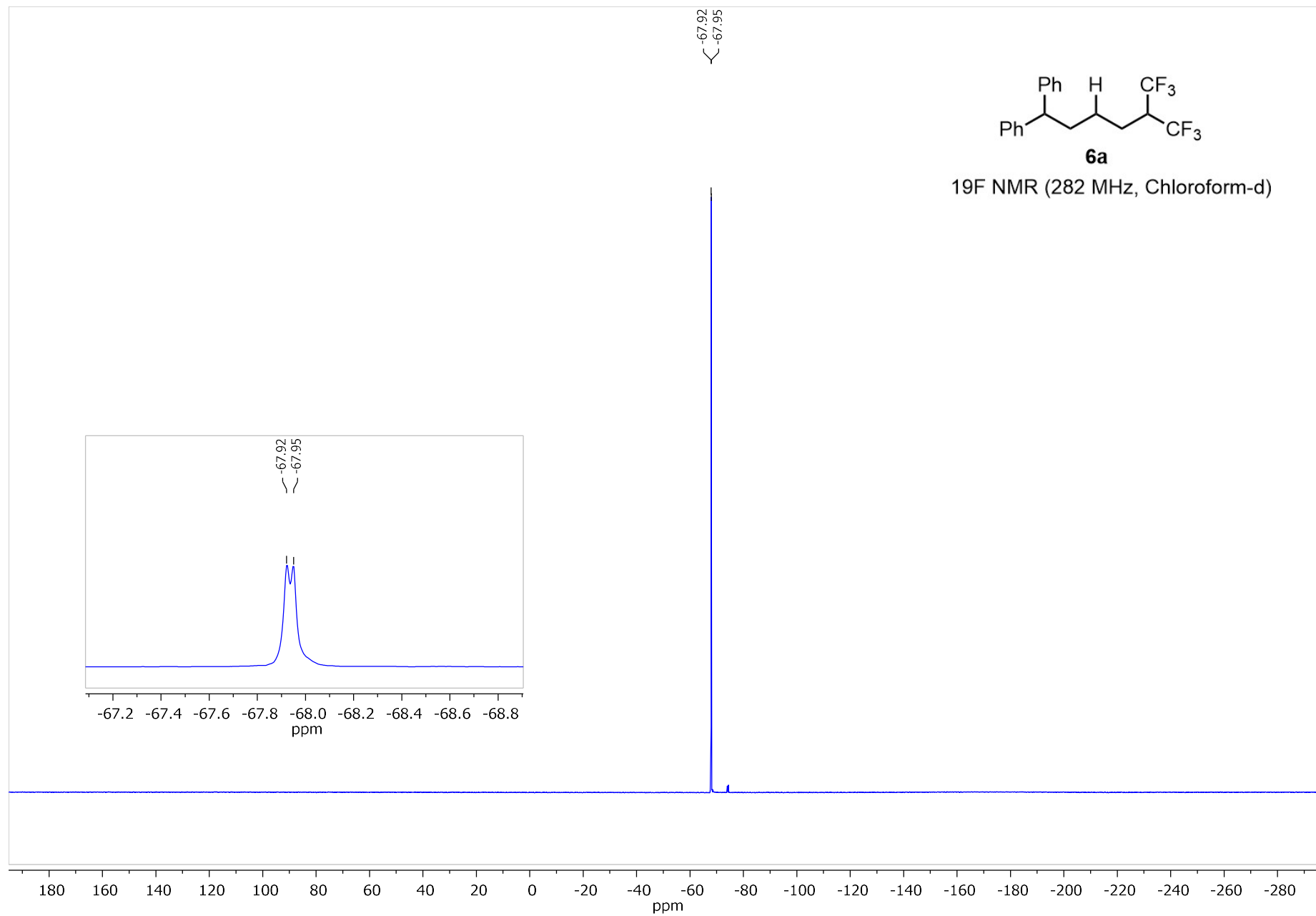
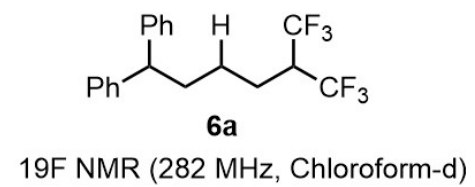


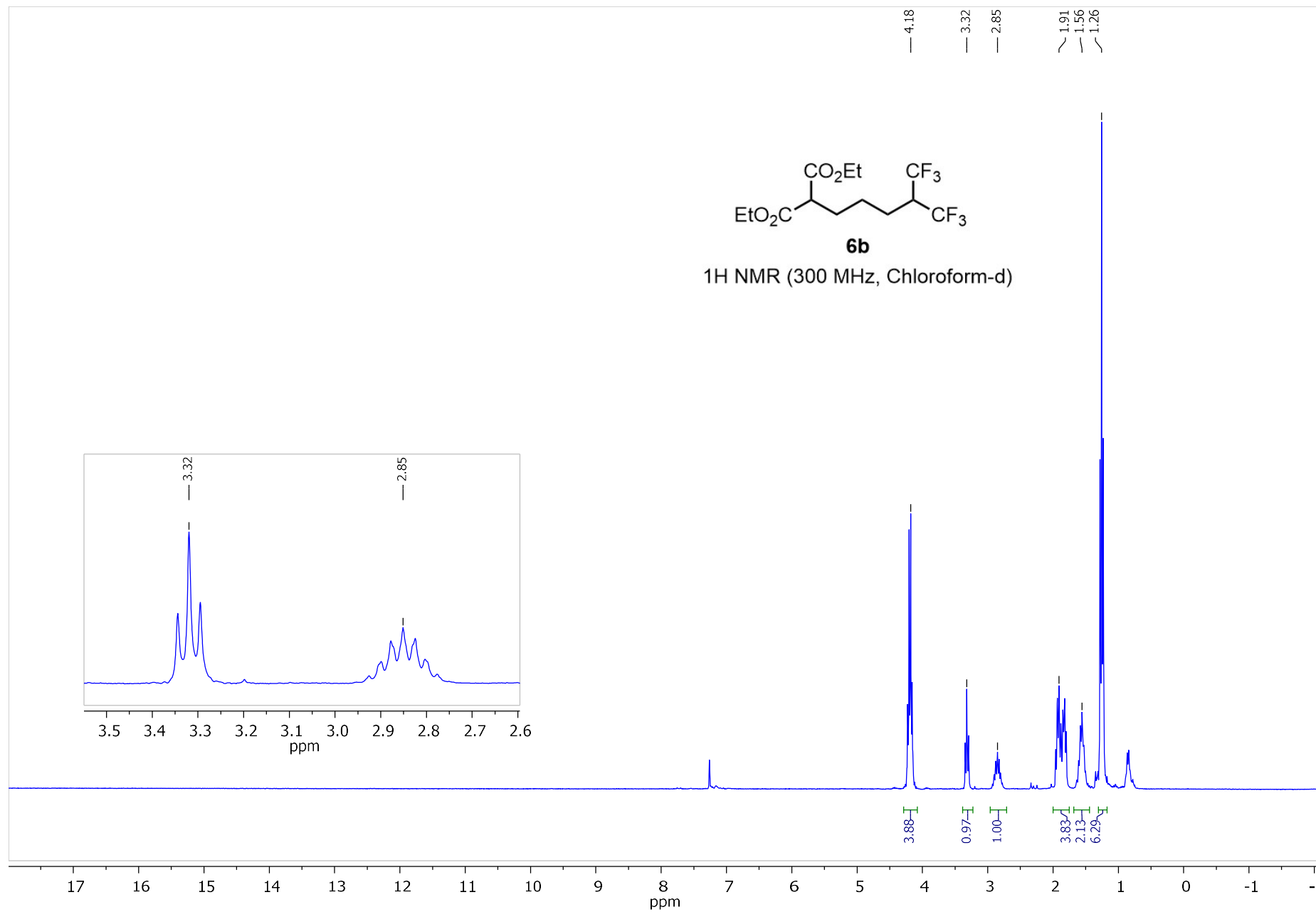
5d
31P NMR (122 MHz, Chloroform-d)

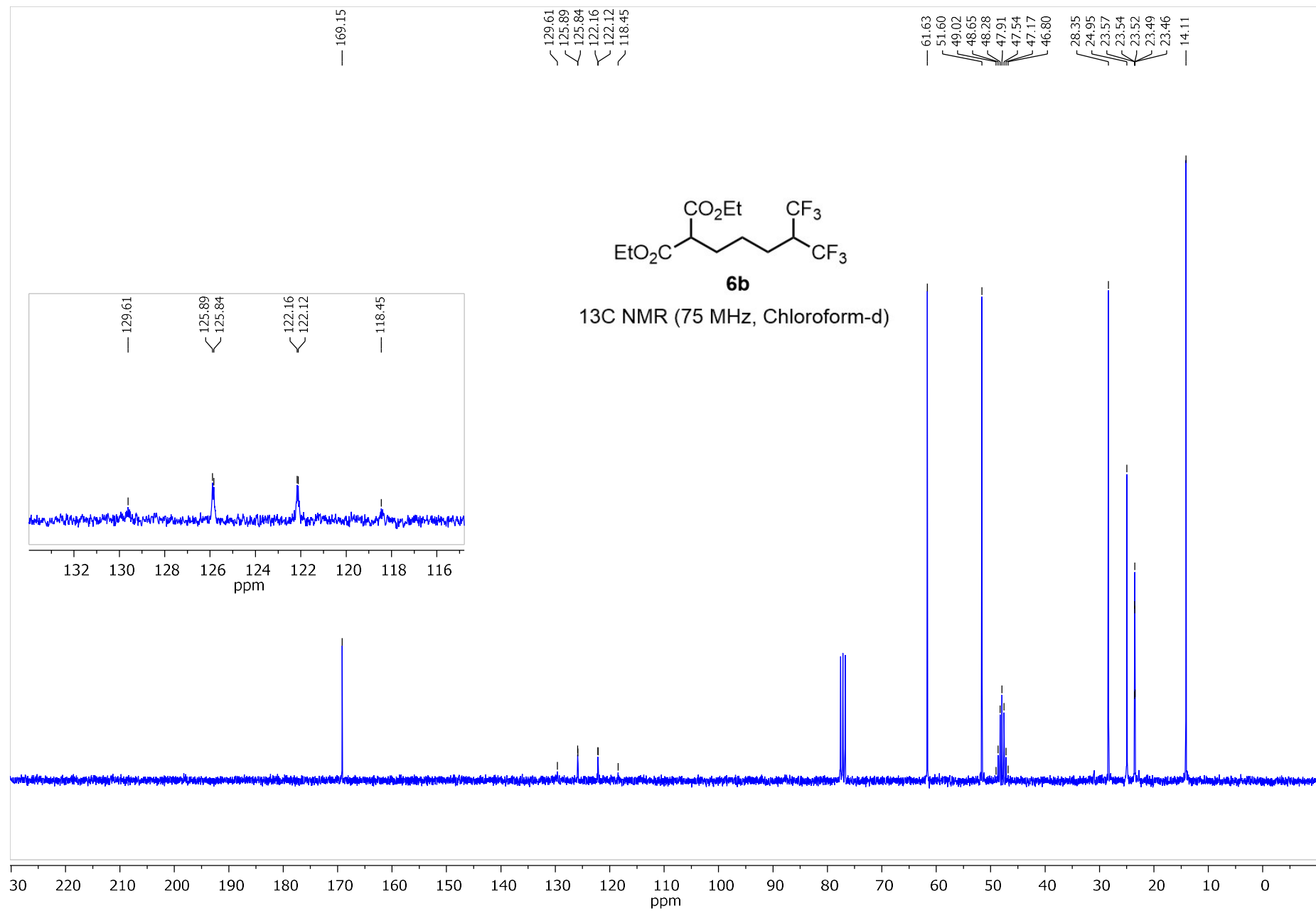


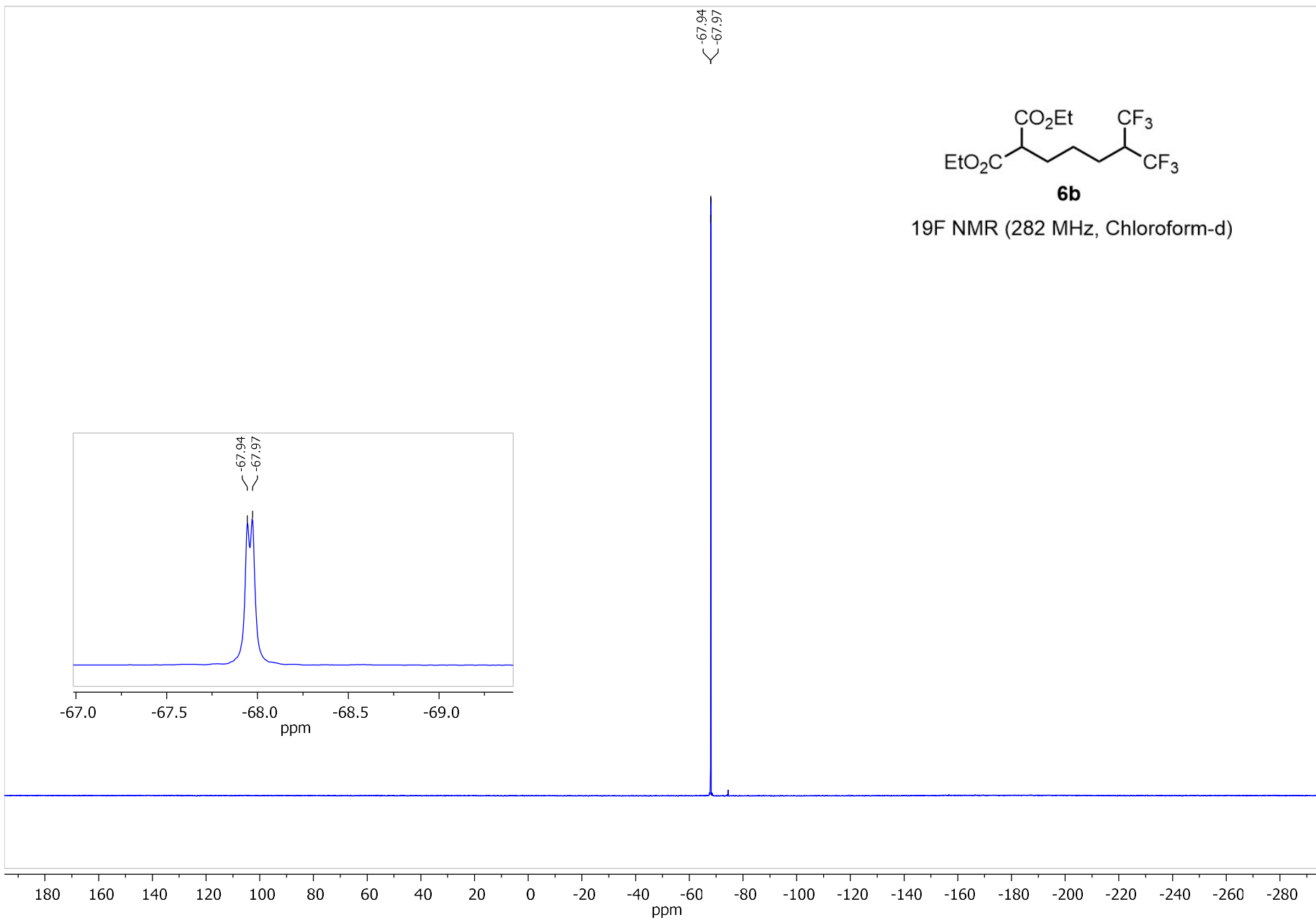


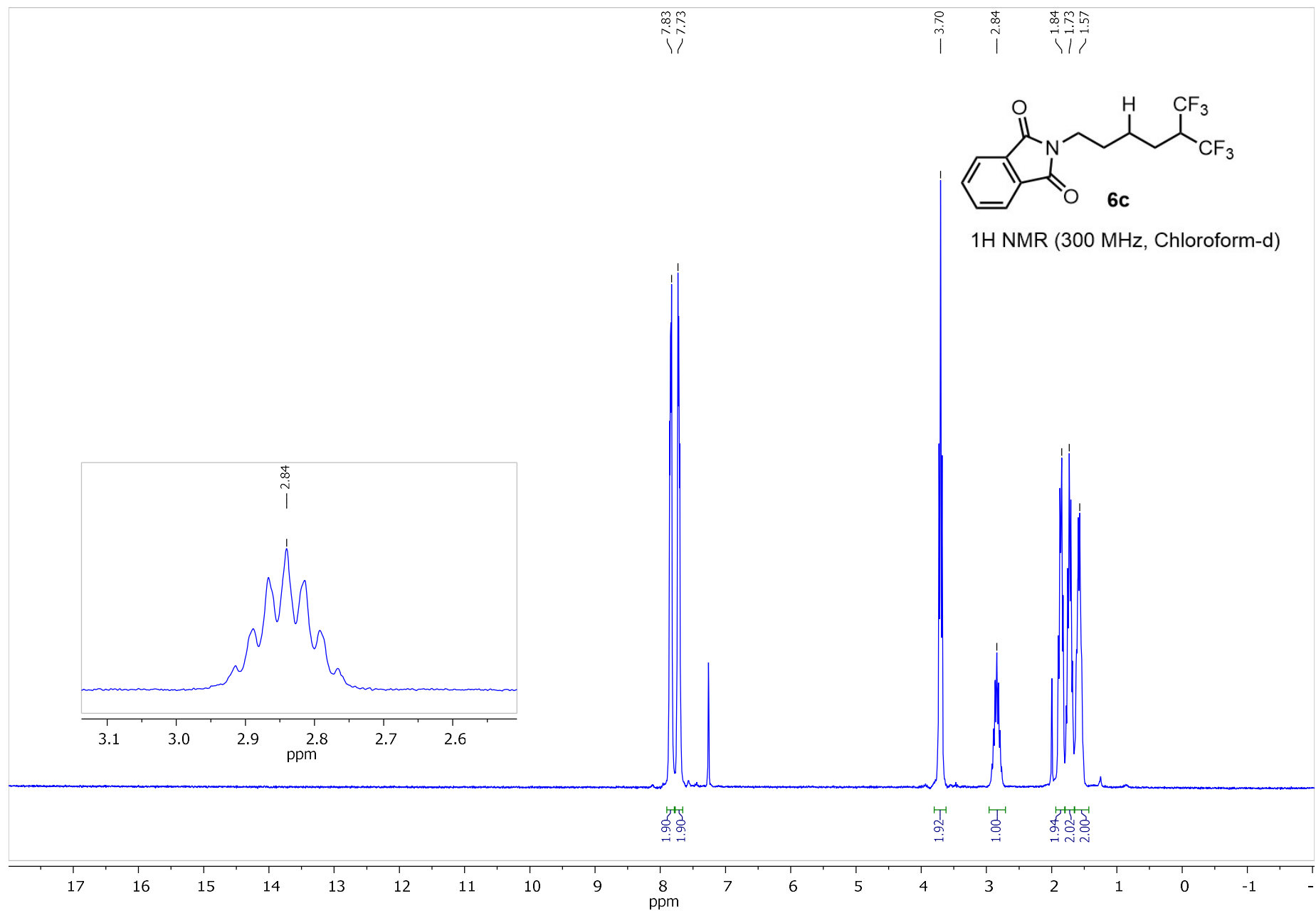


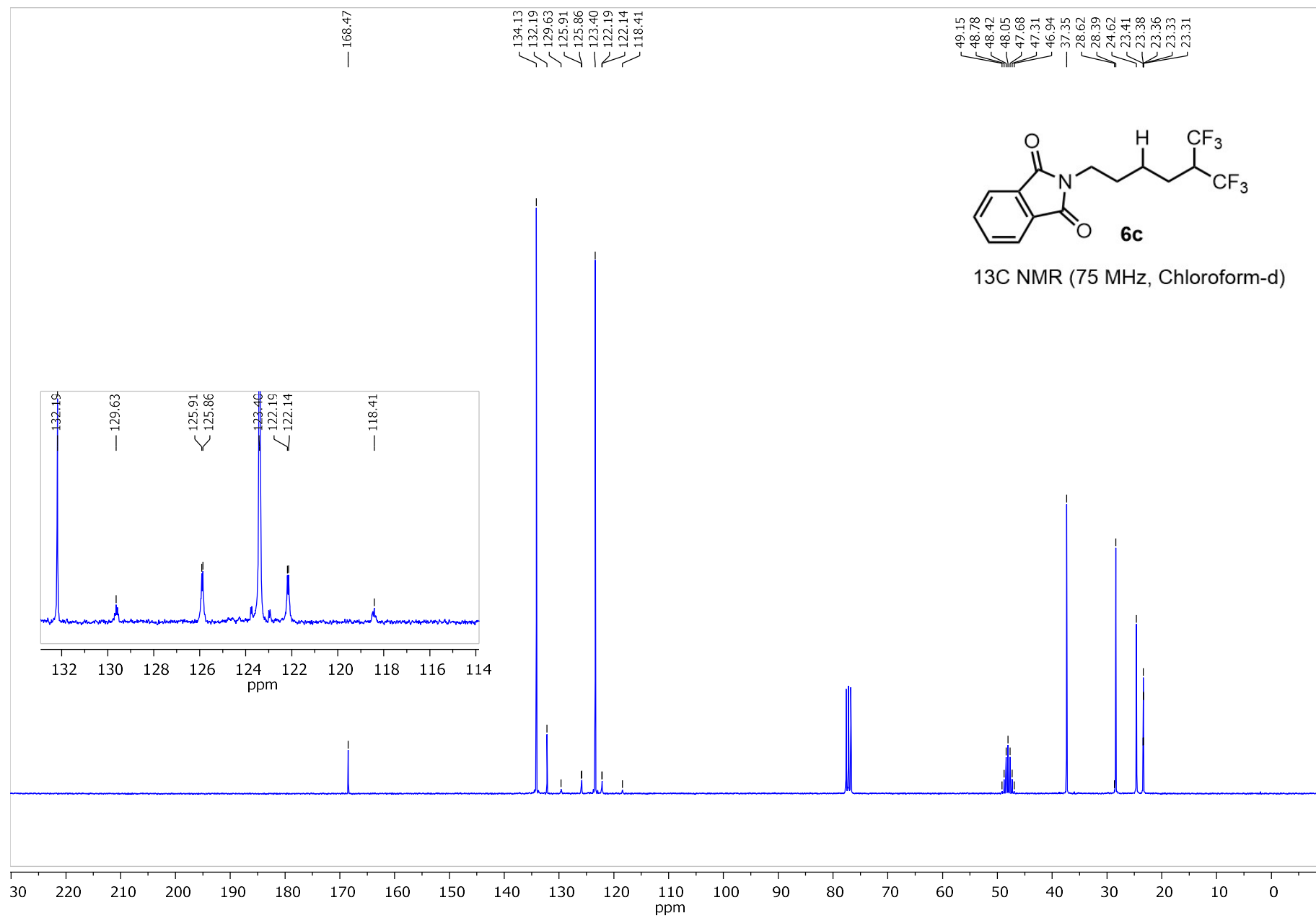


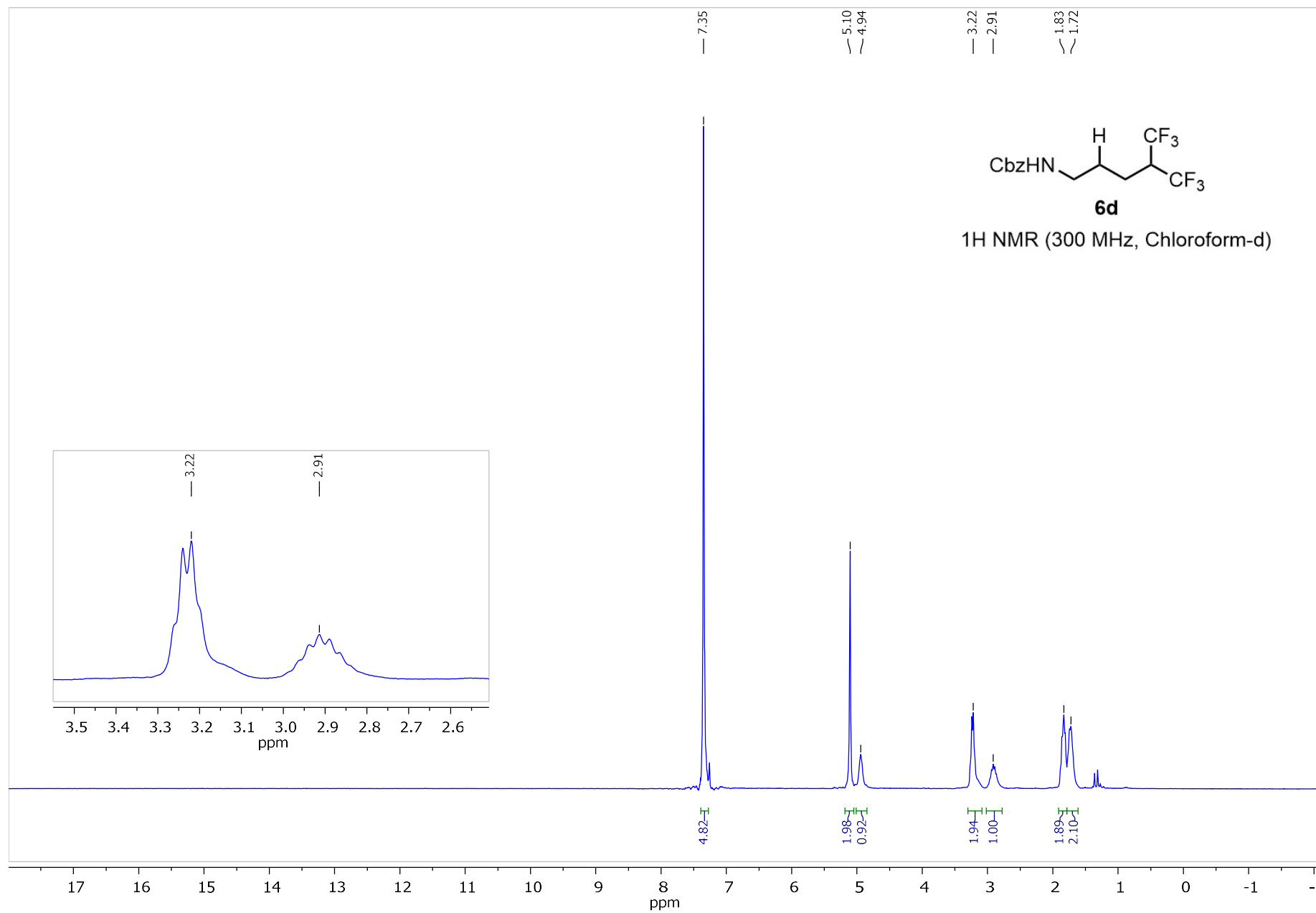


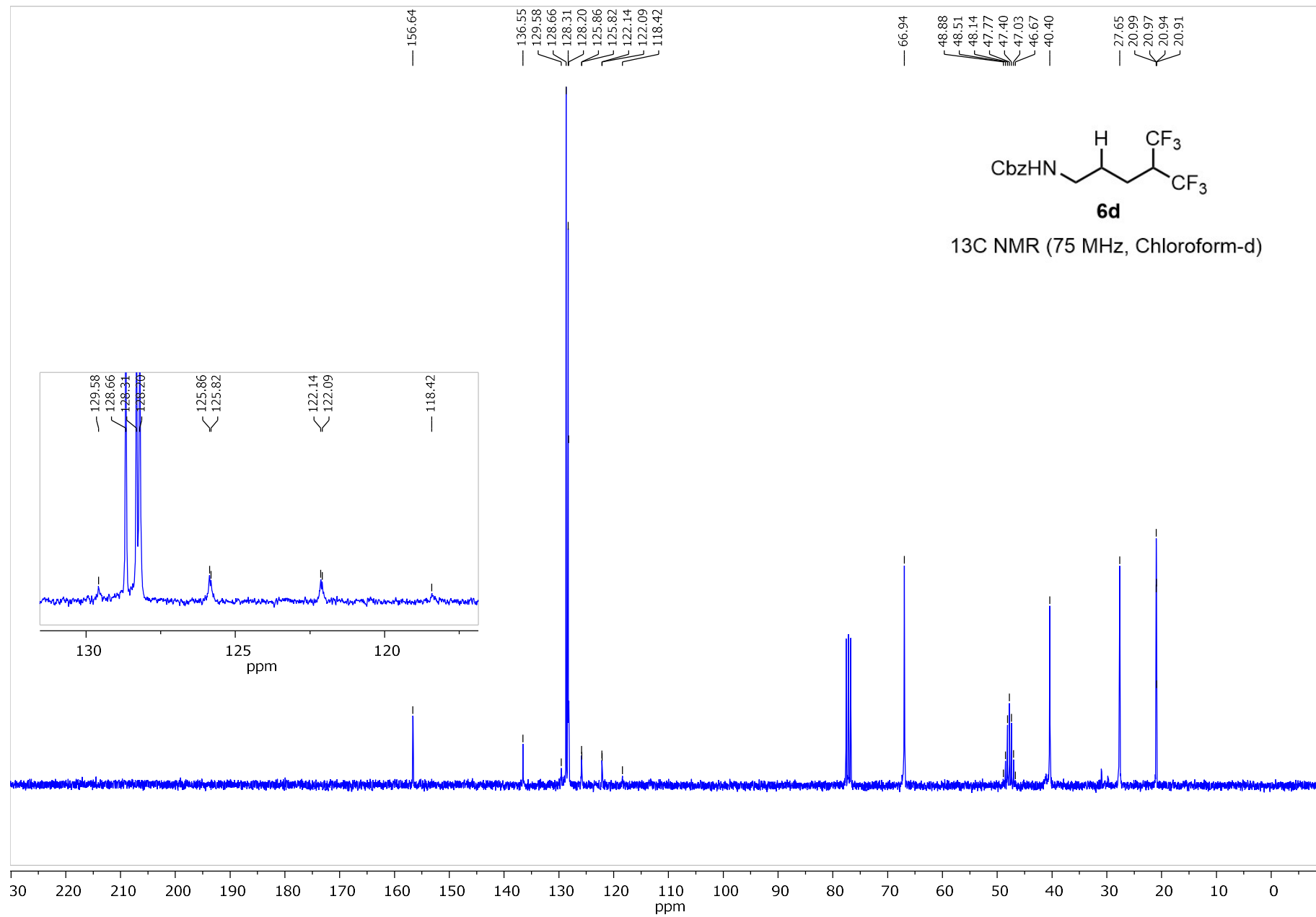


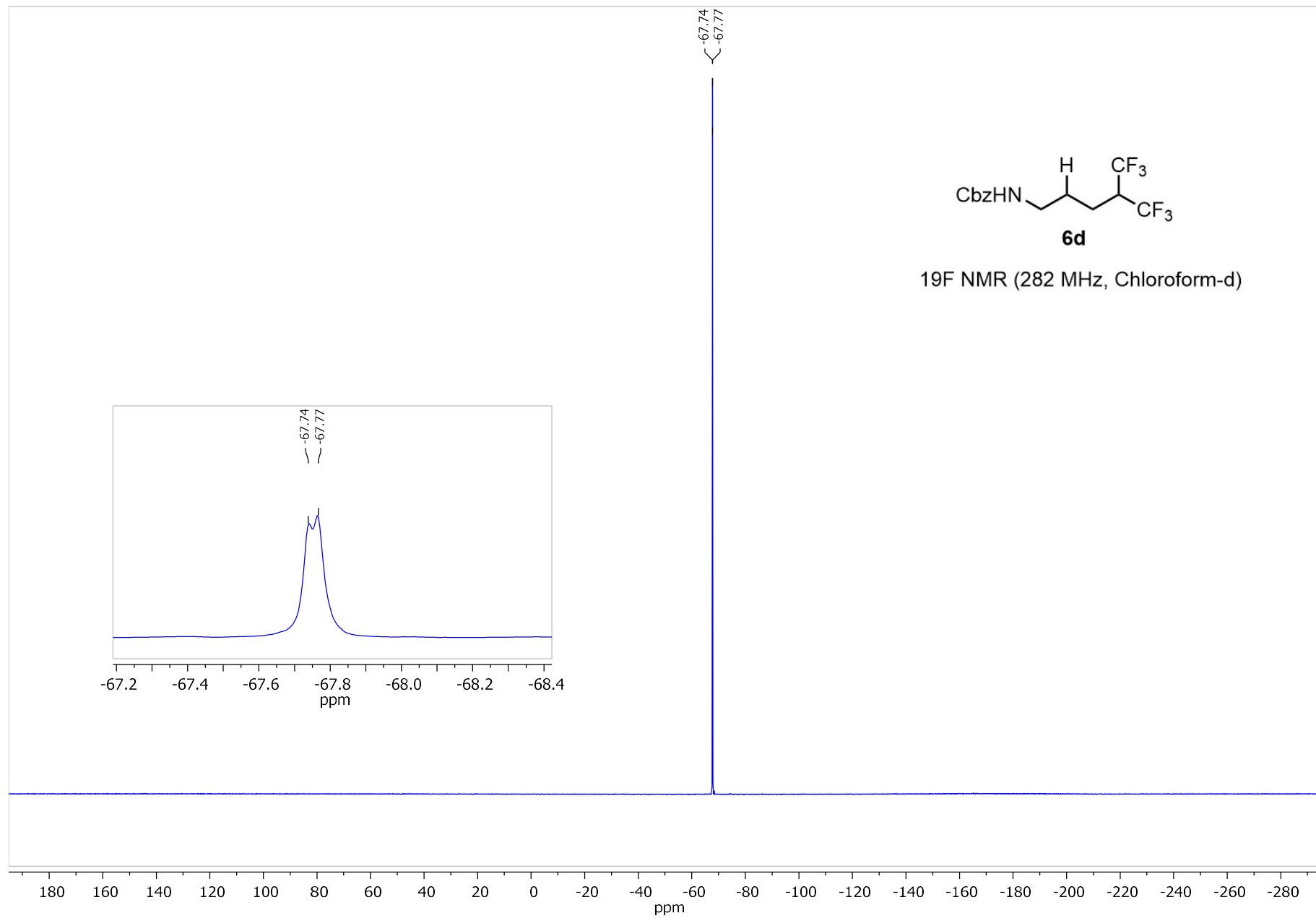
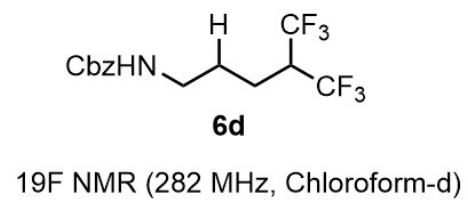


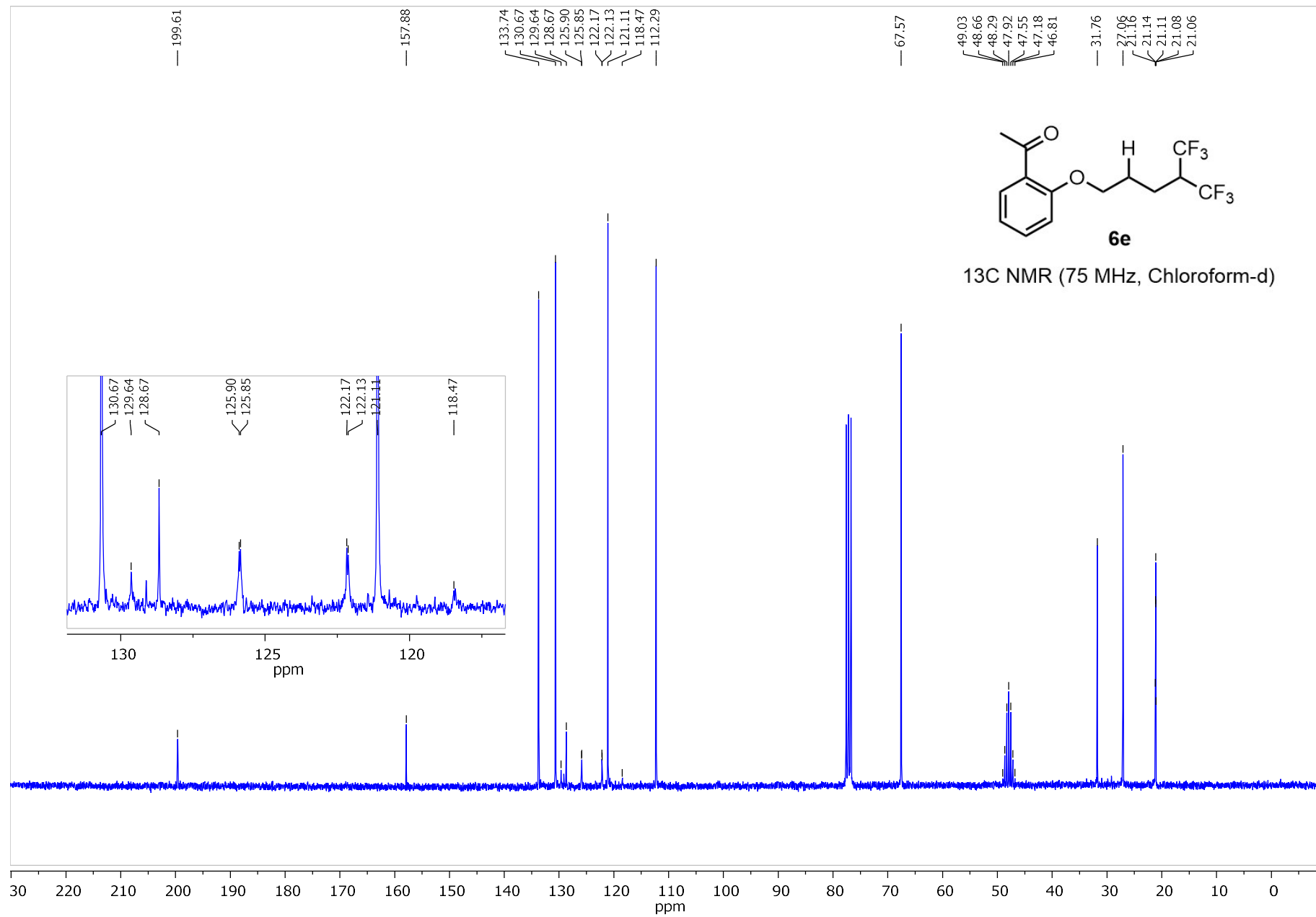


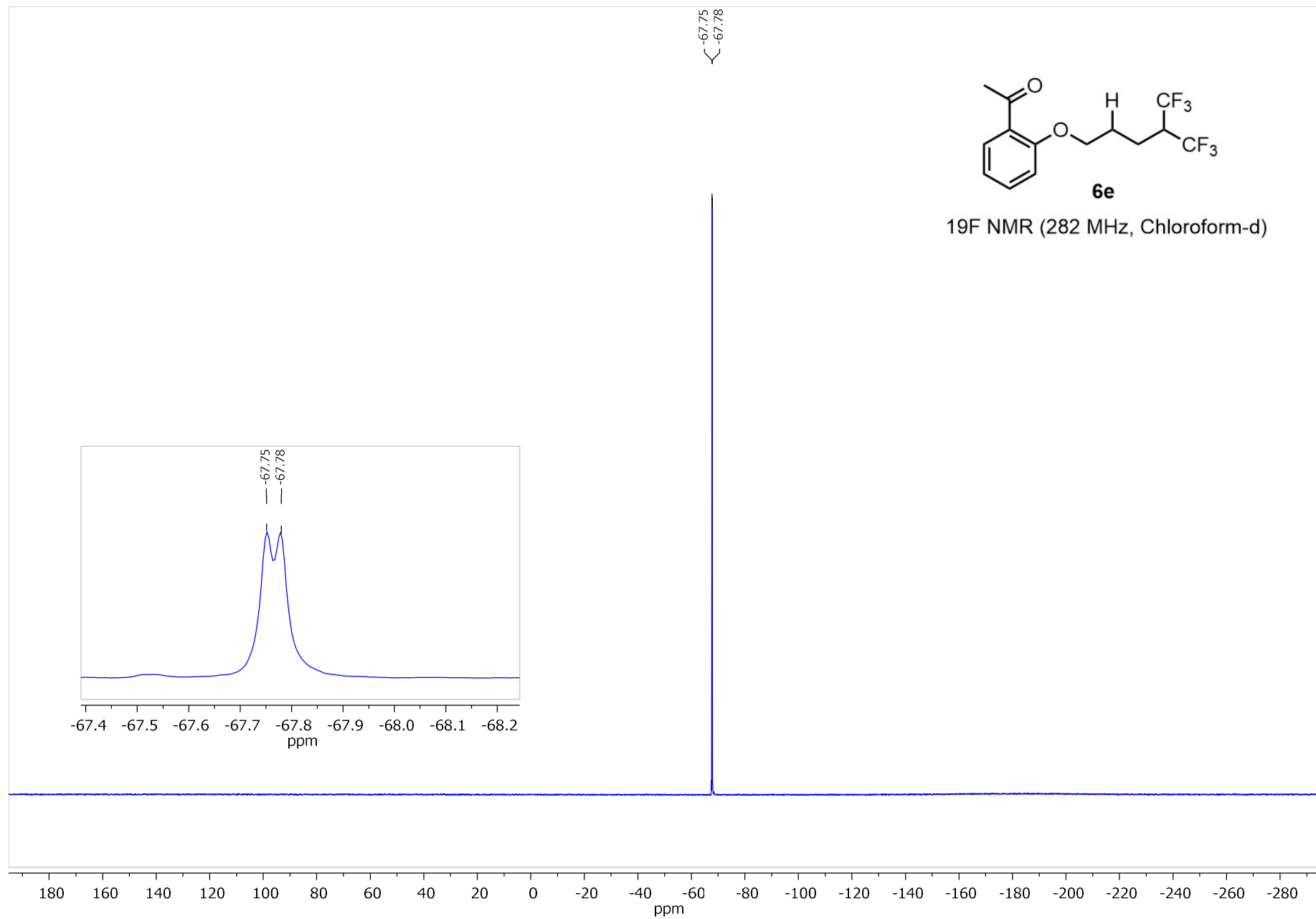


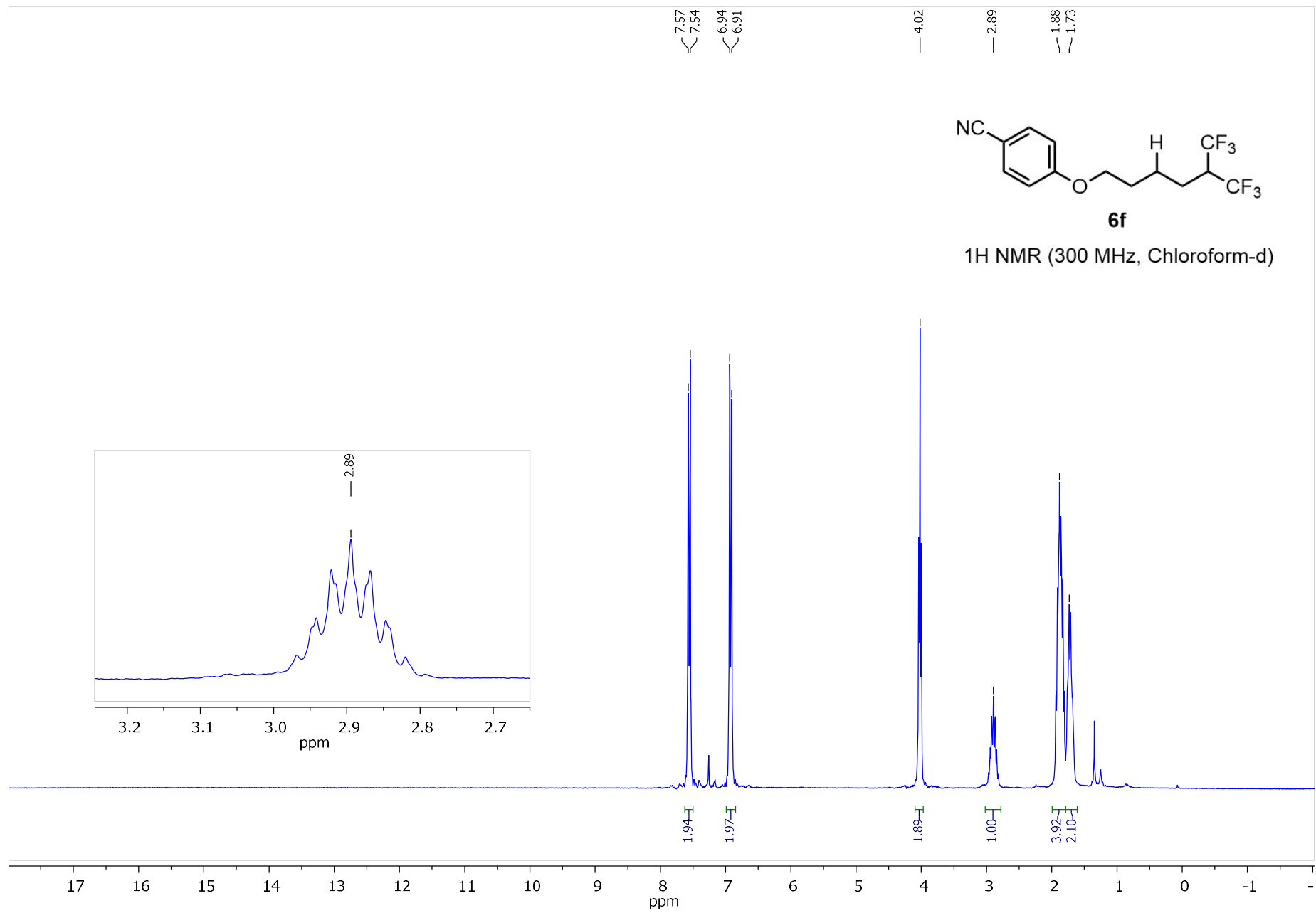


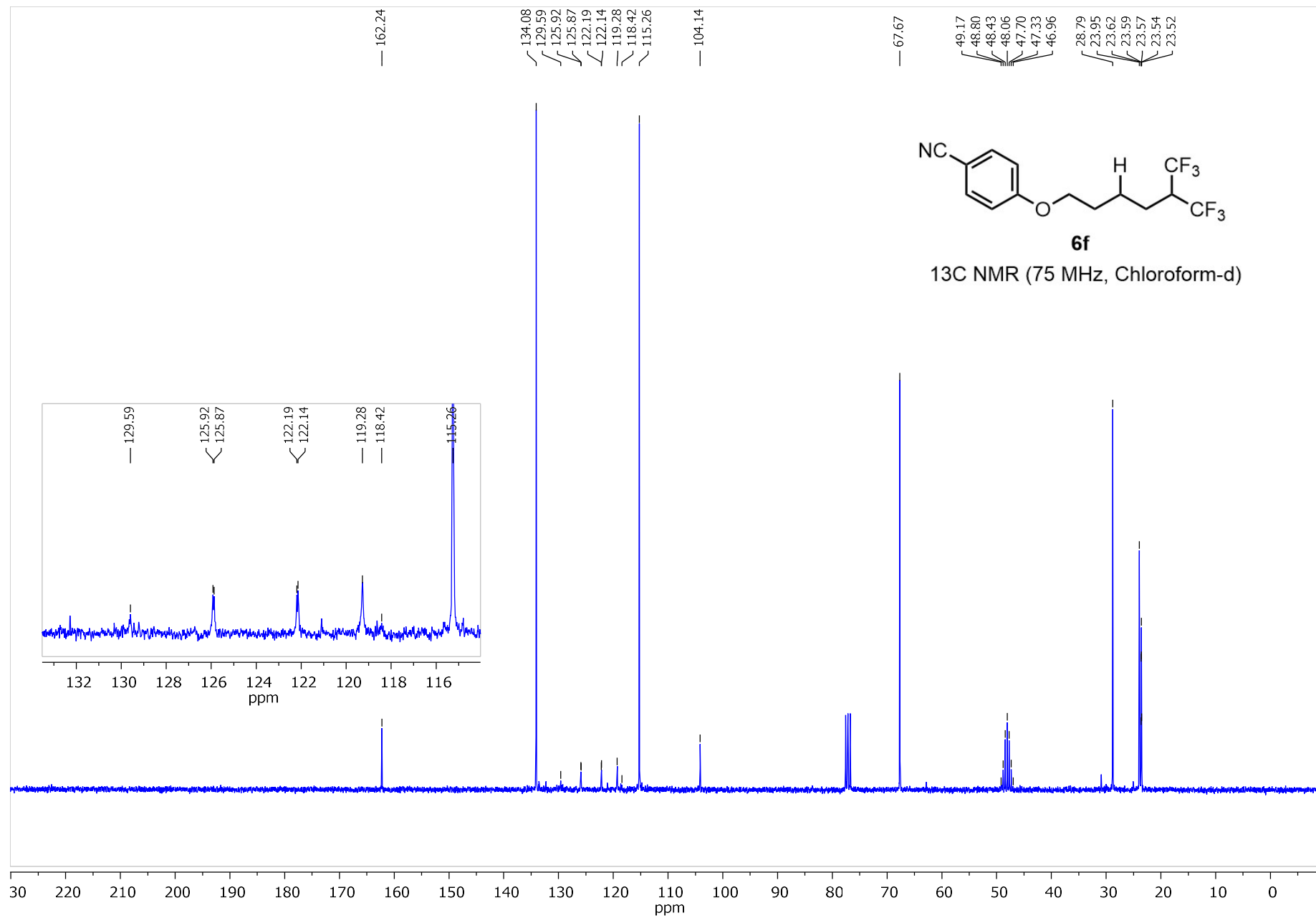


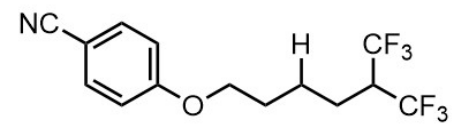




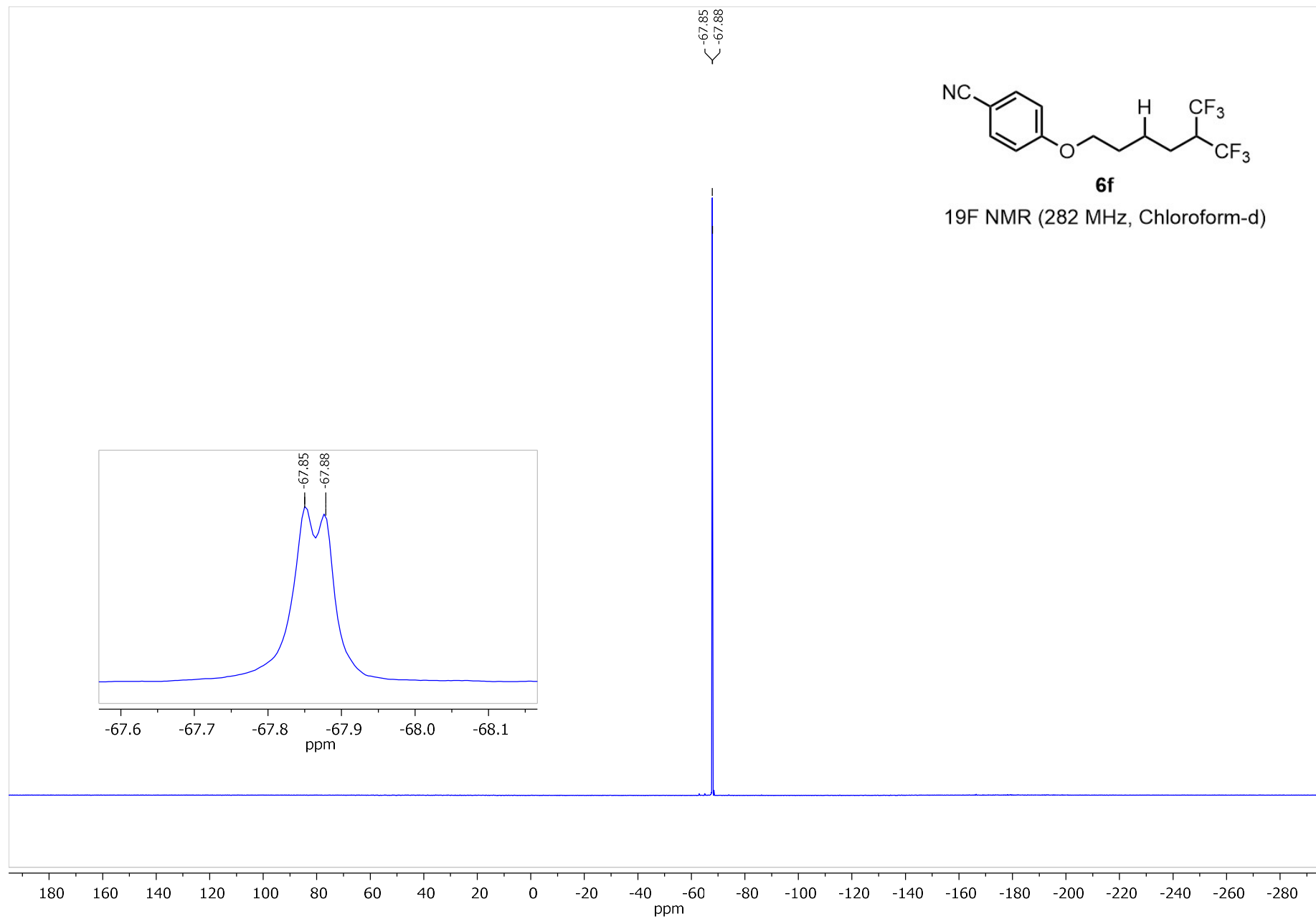


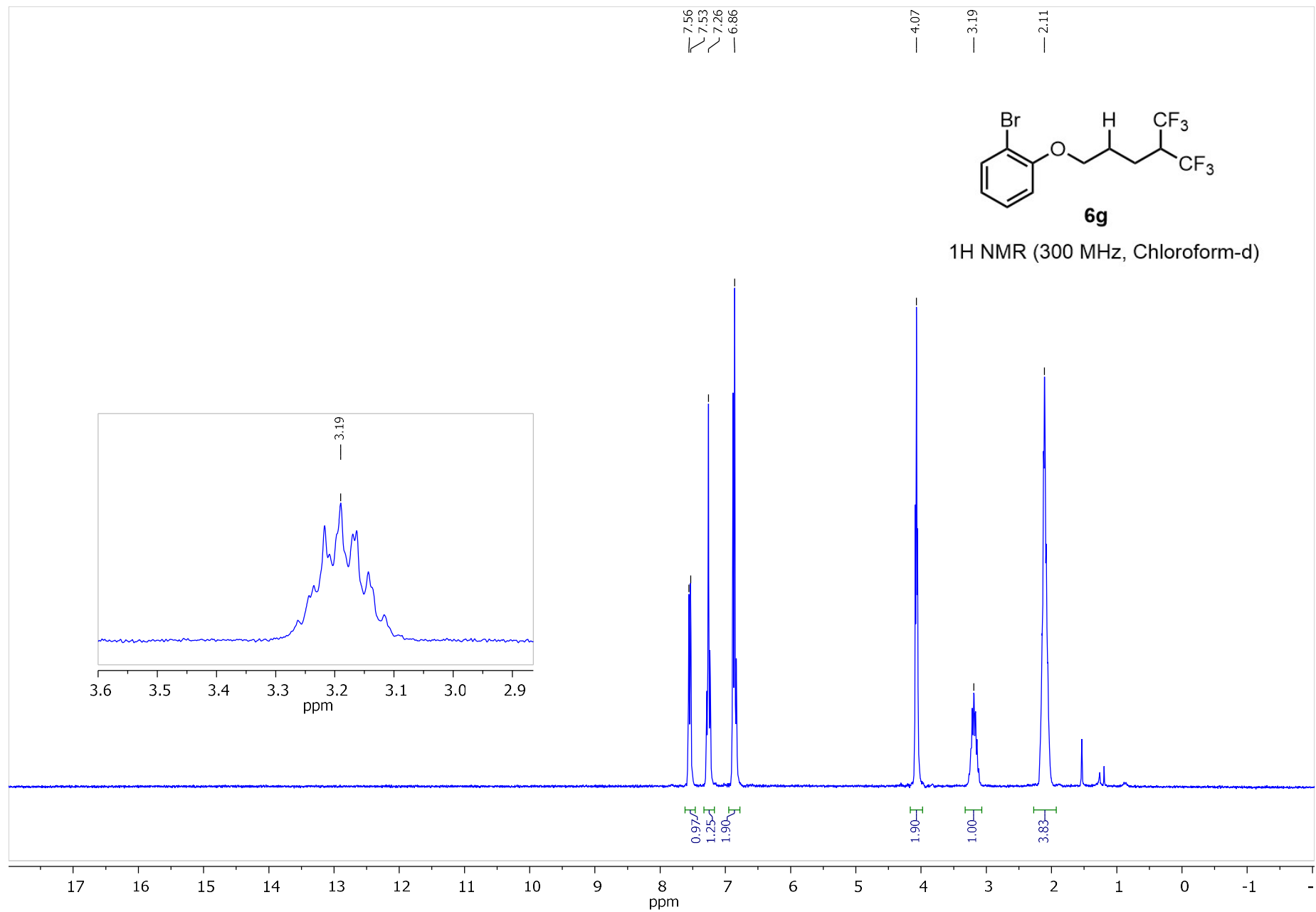


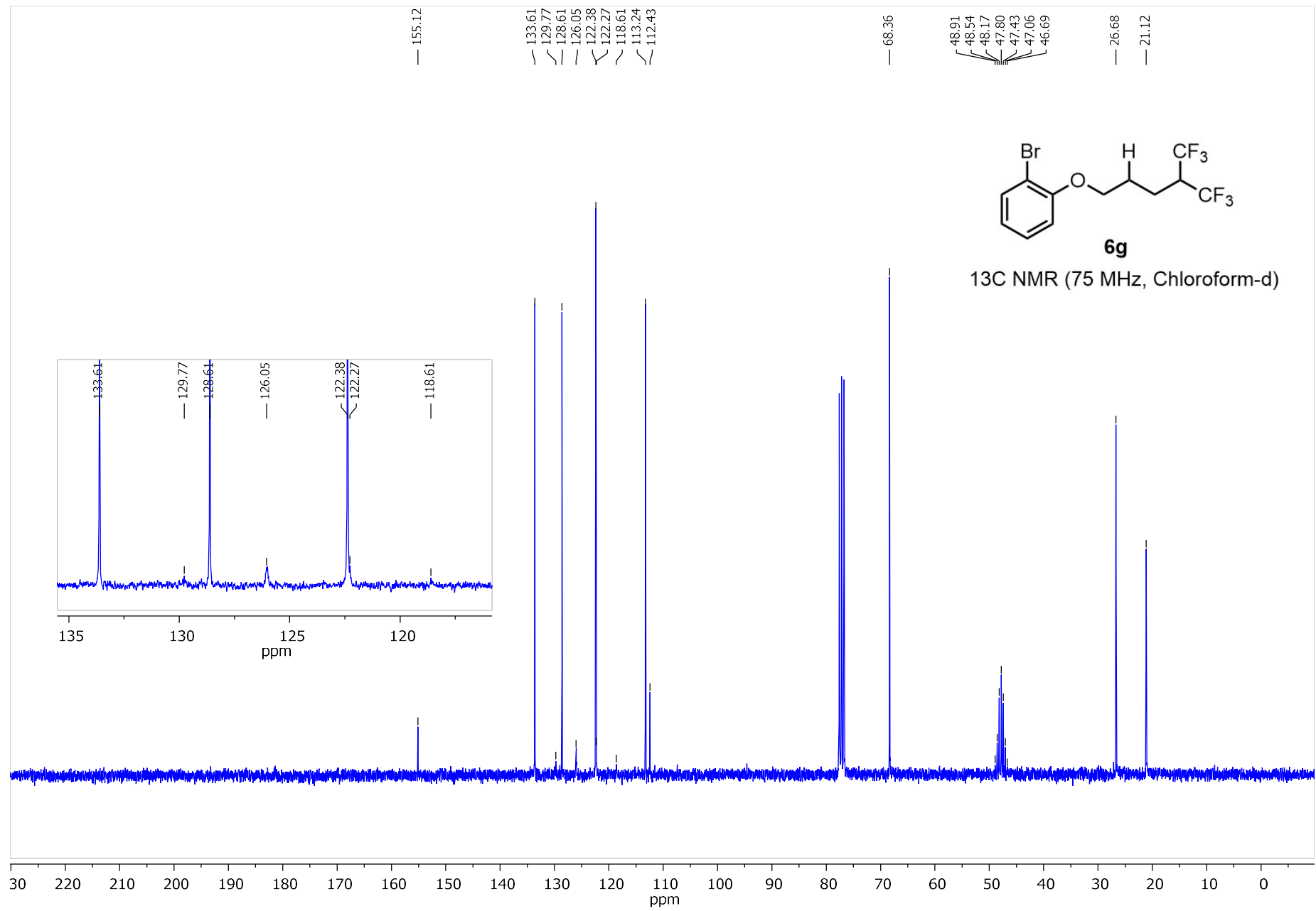


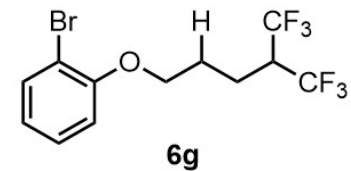
**6f**

19F NMR (282 MHz, Chloroform-d)

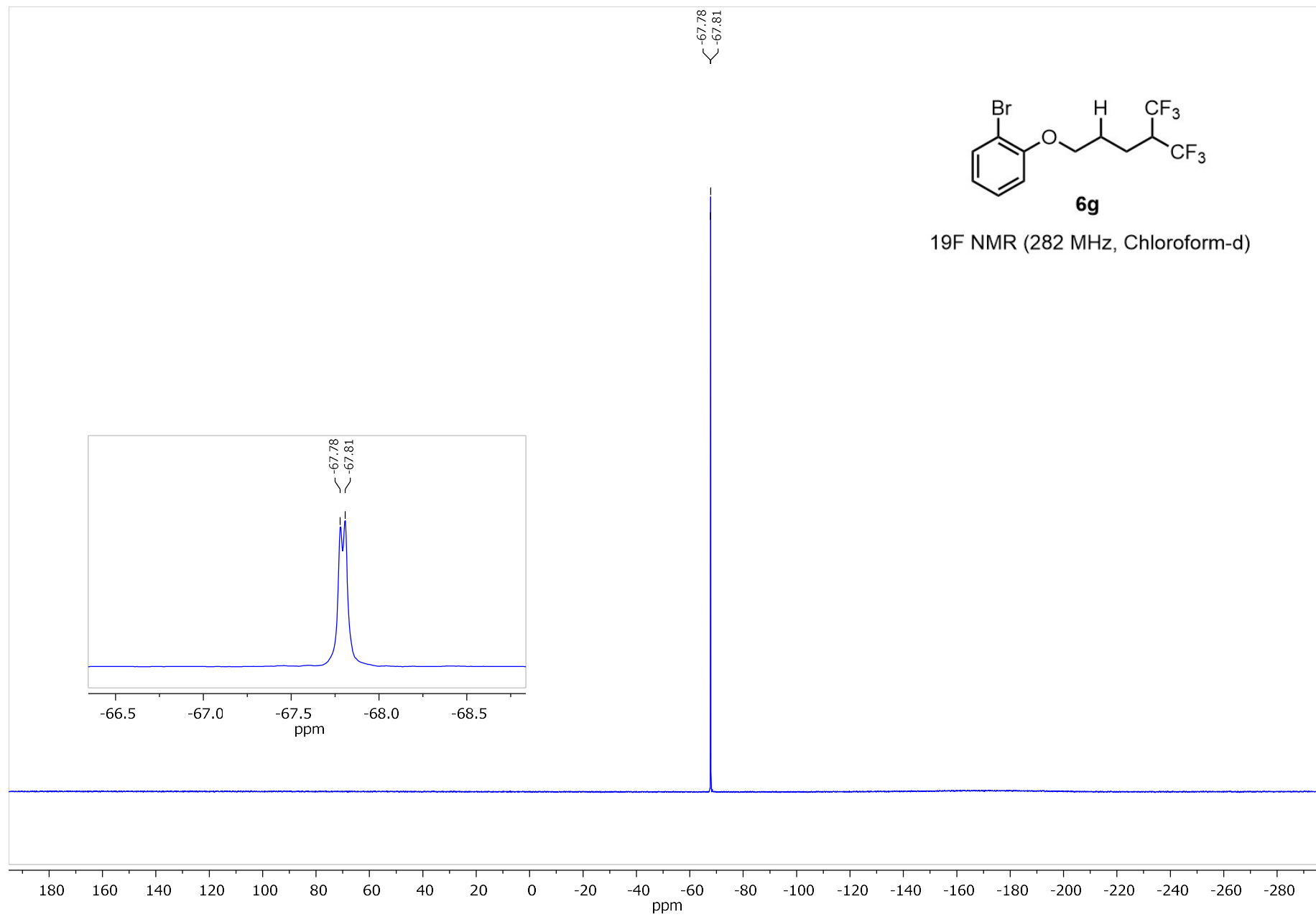


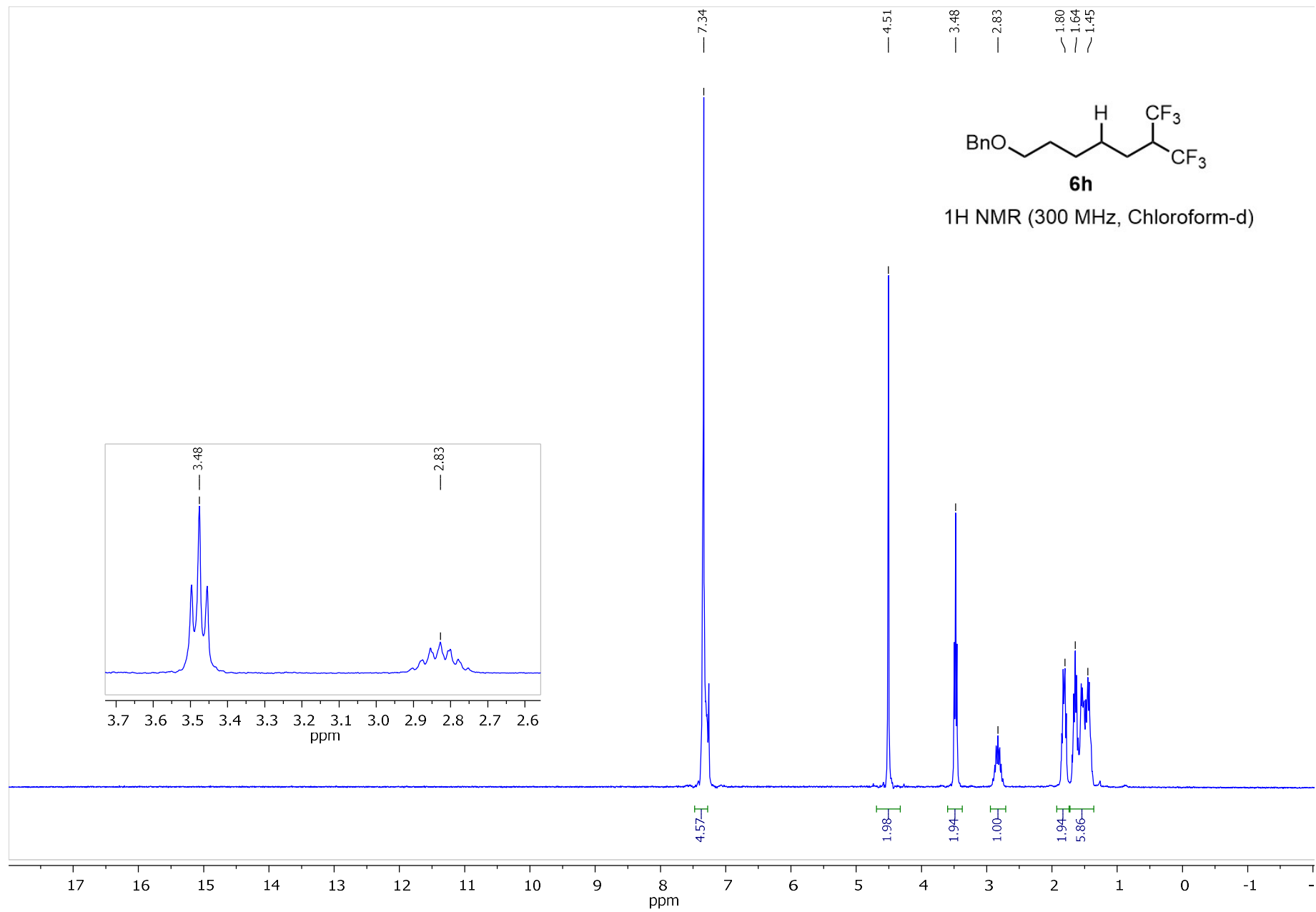


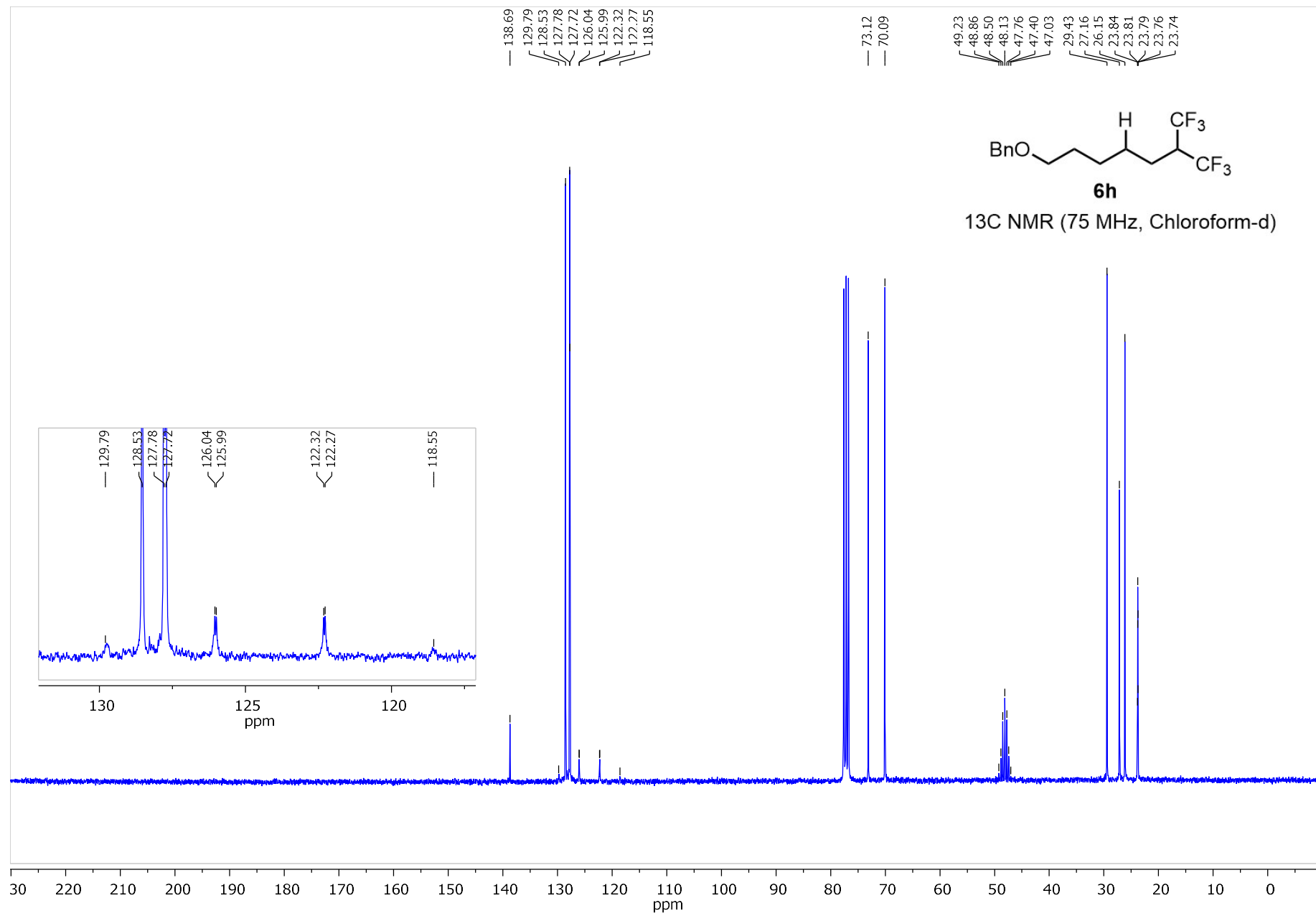


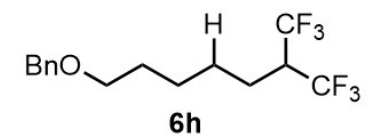


19F NMR (282 MHz, Chloroform-d)

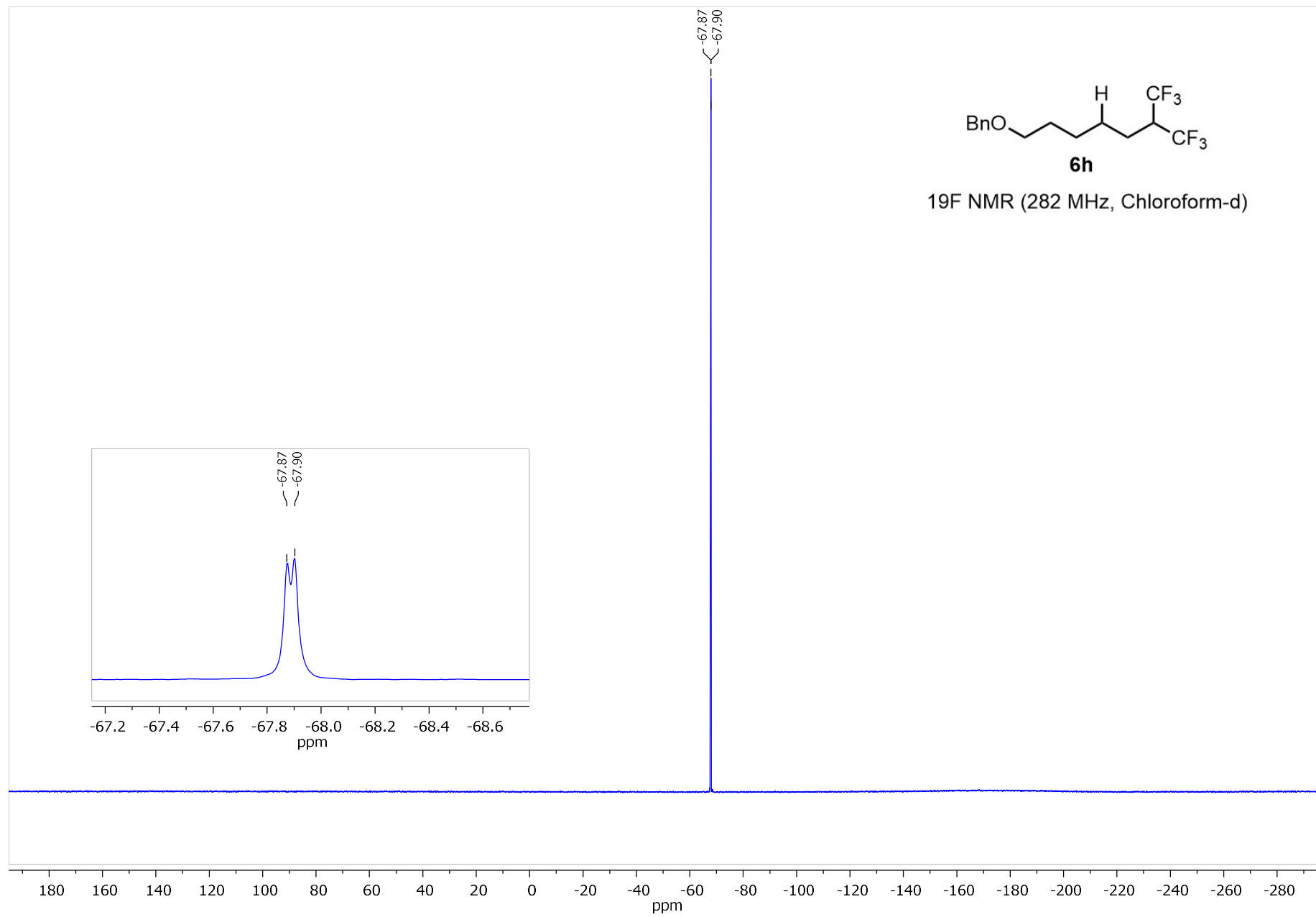


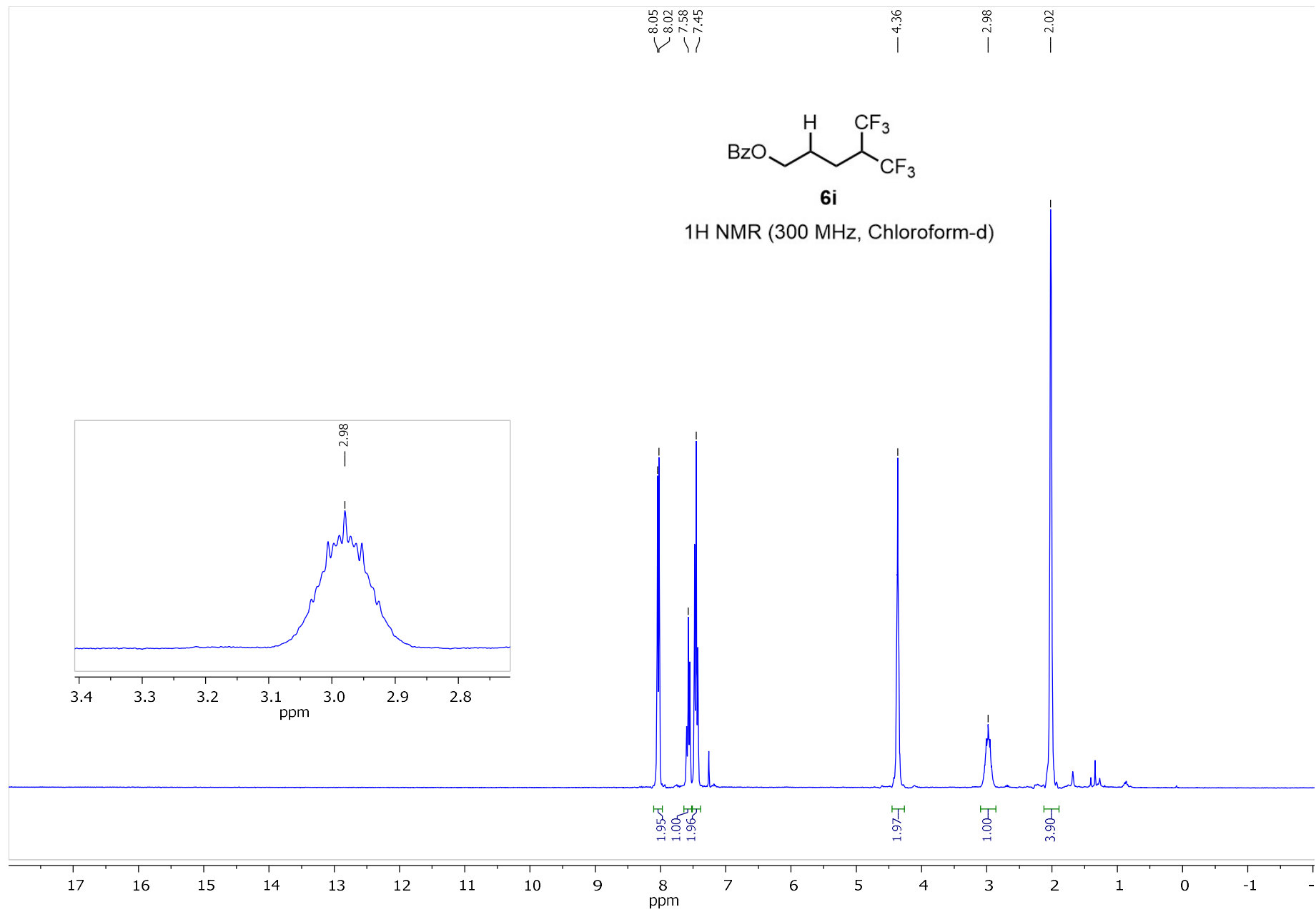


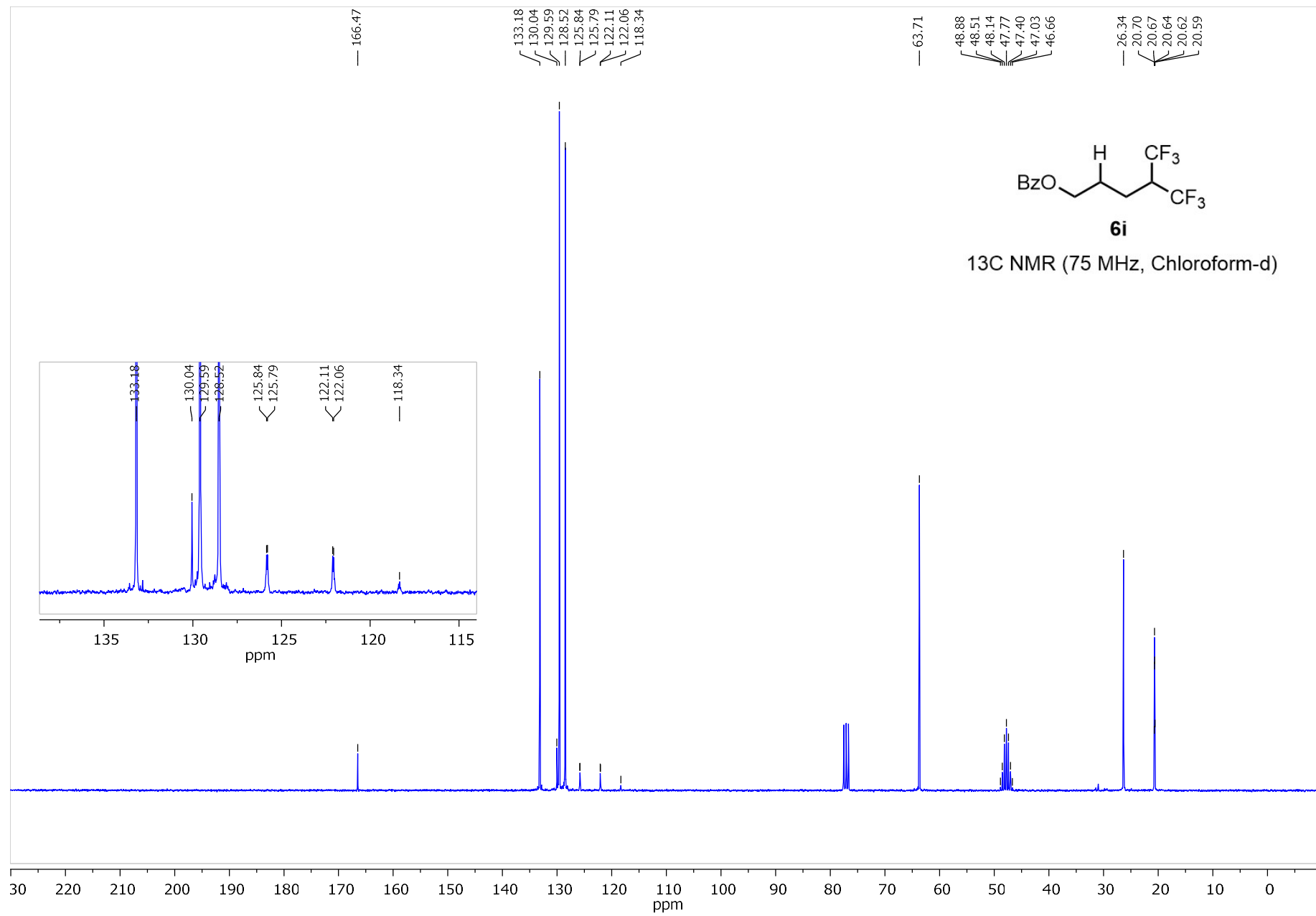


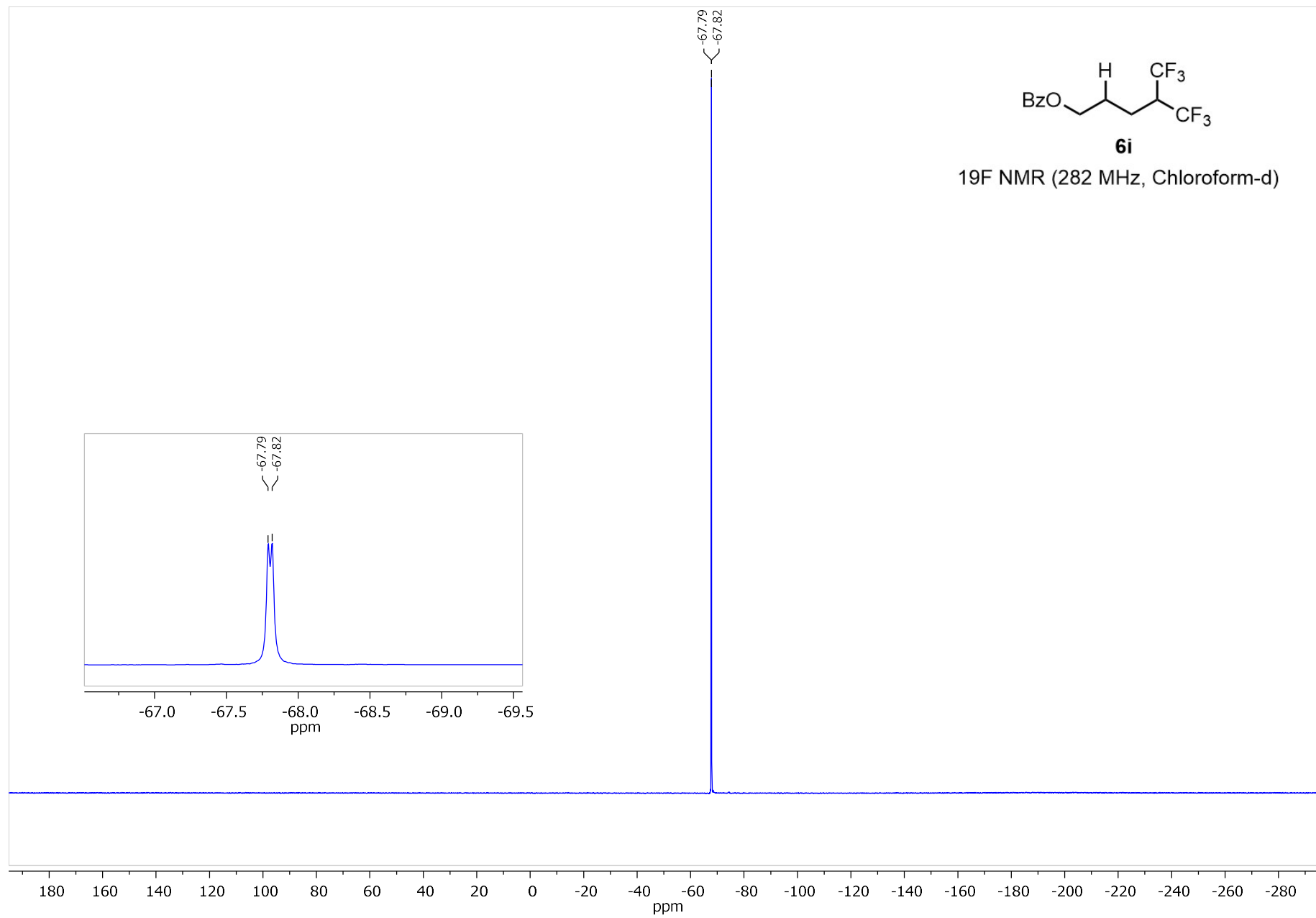
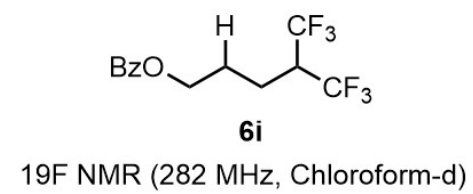


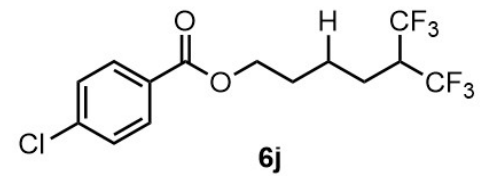
^{19}F NMR (282 MHz, Chloroform-d)



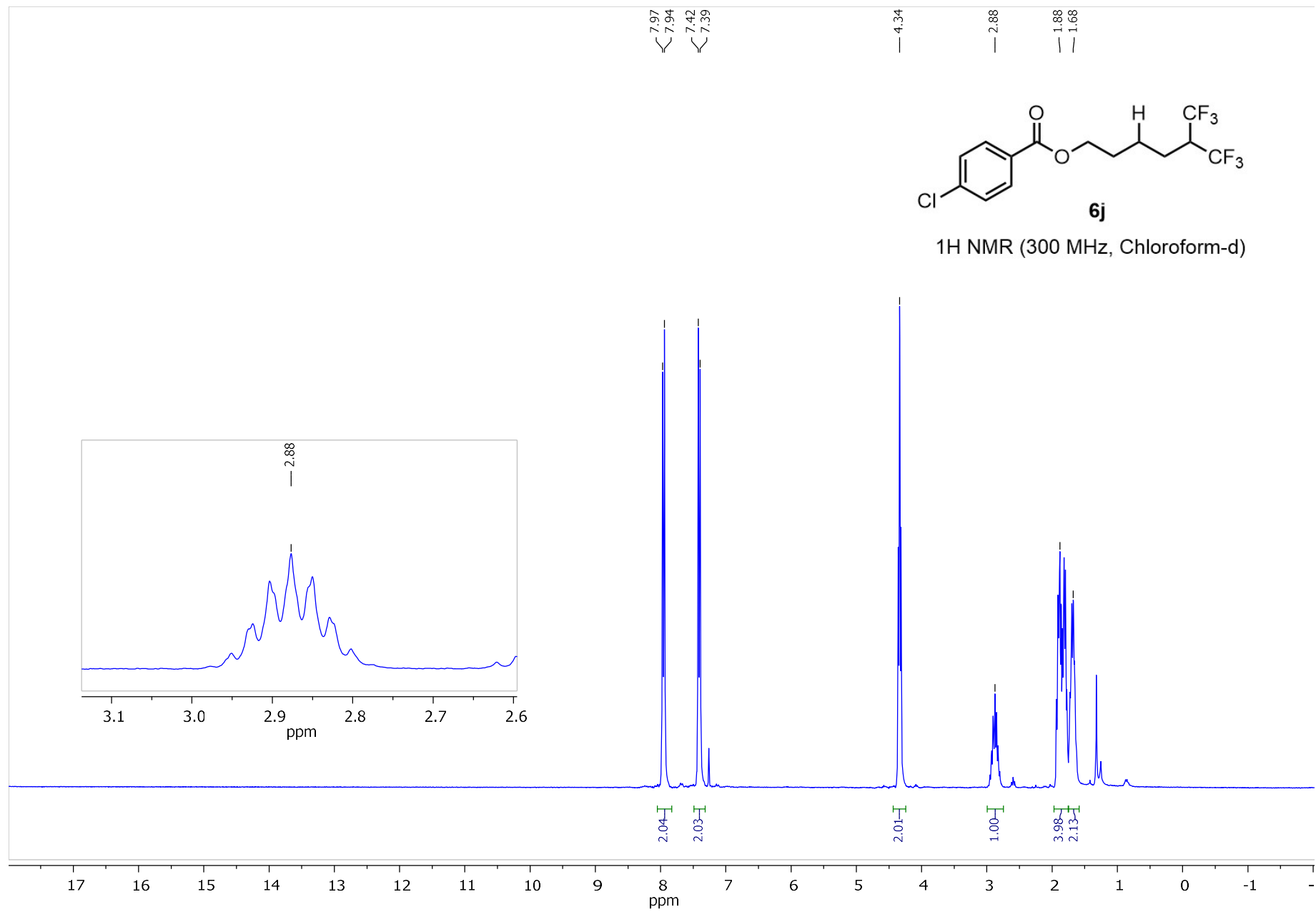


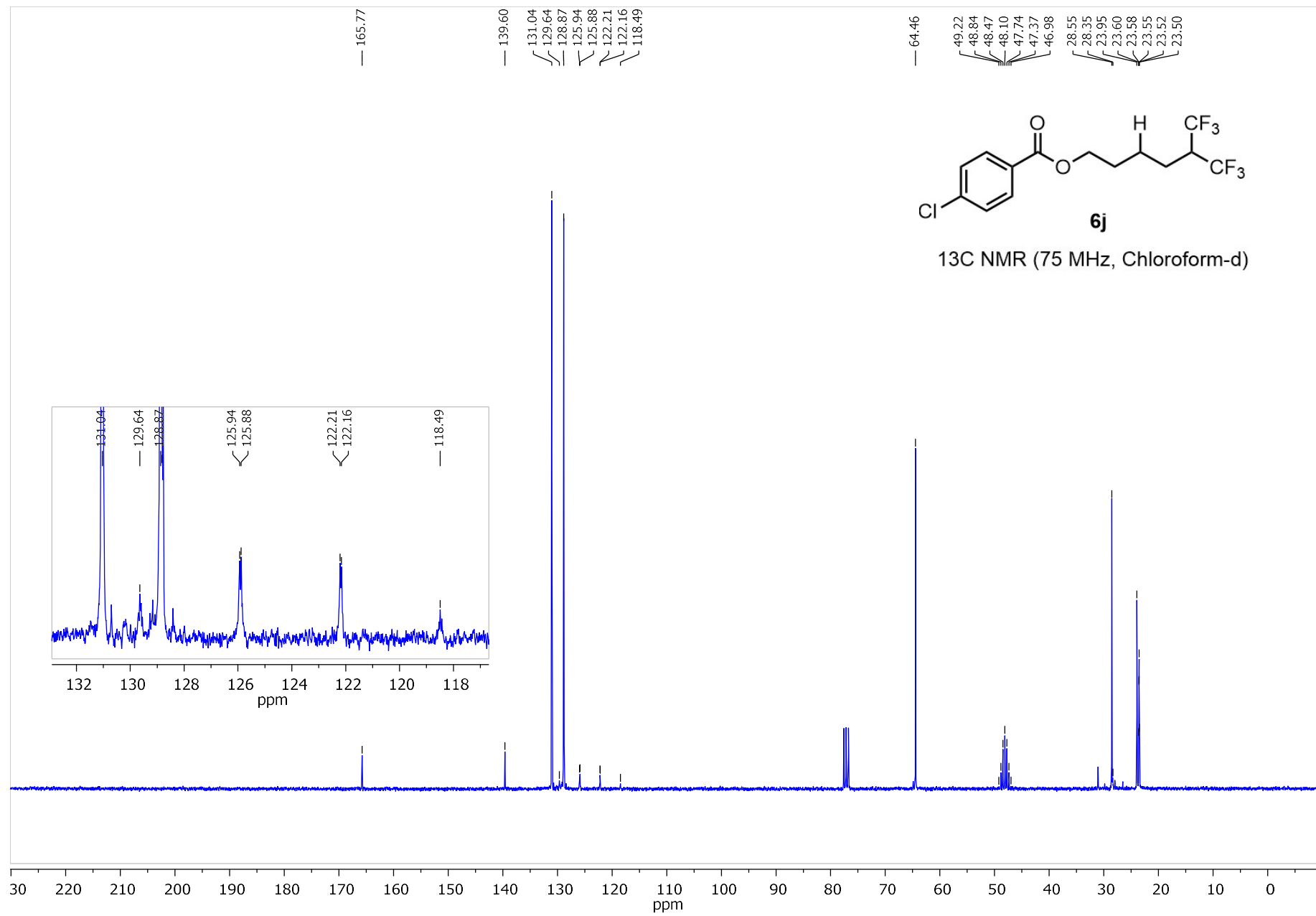


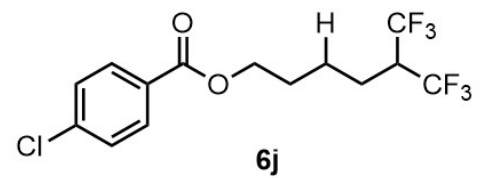




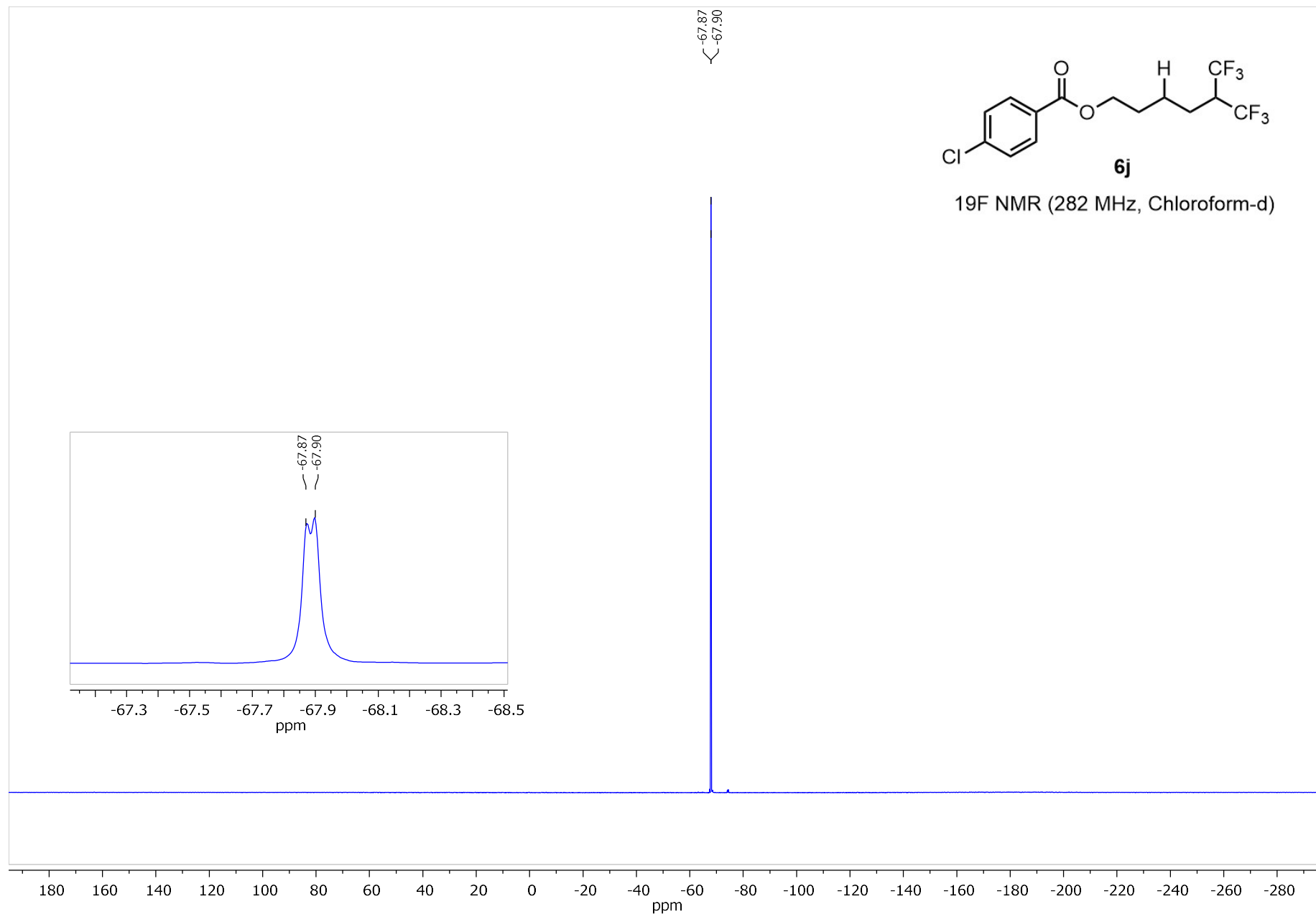
$^1\text{H NMR}$ (300 MHz, Chloroform- d)

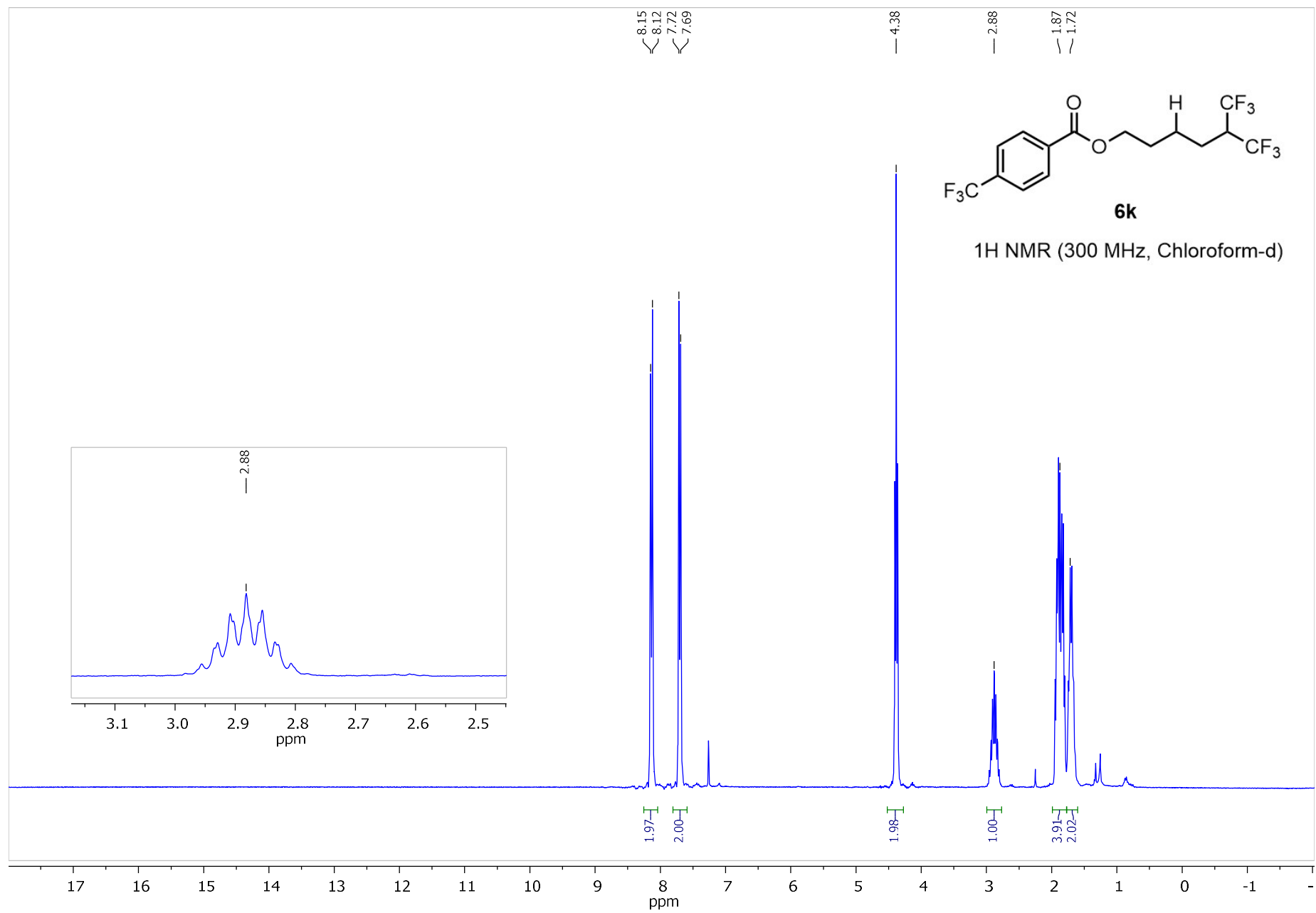


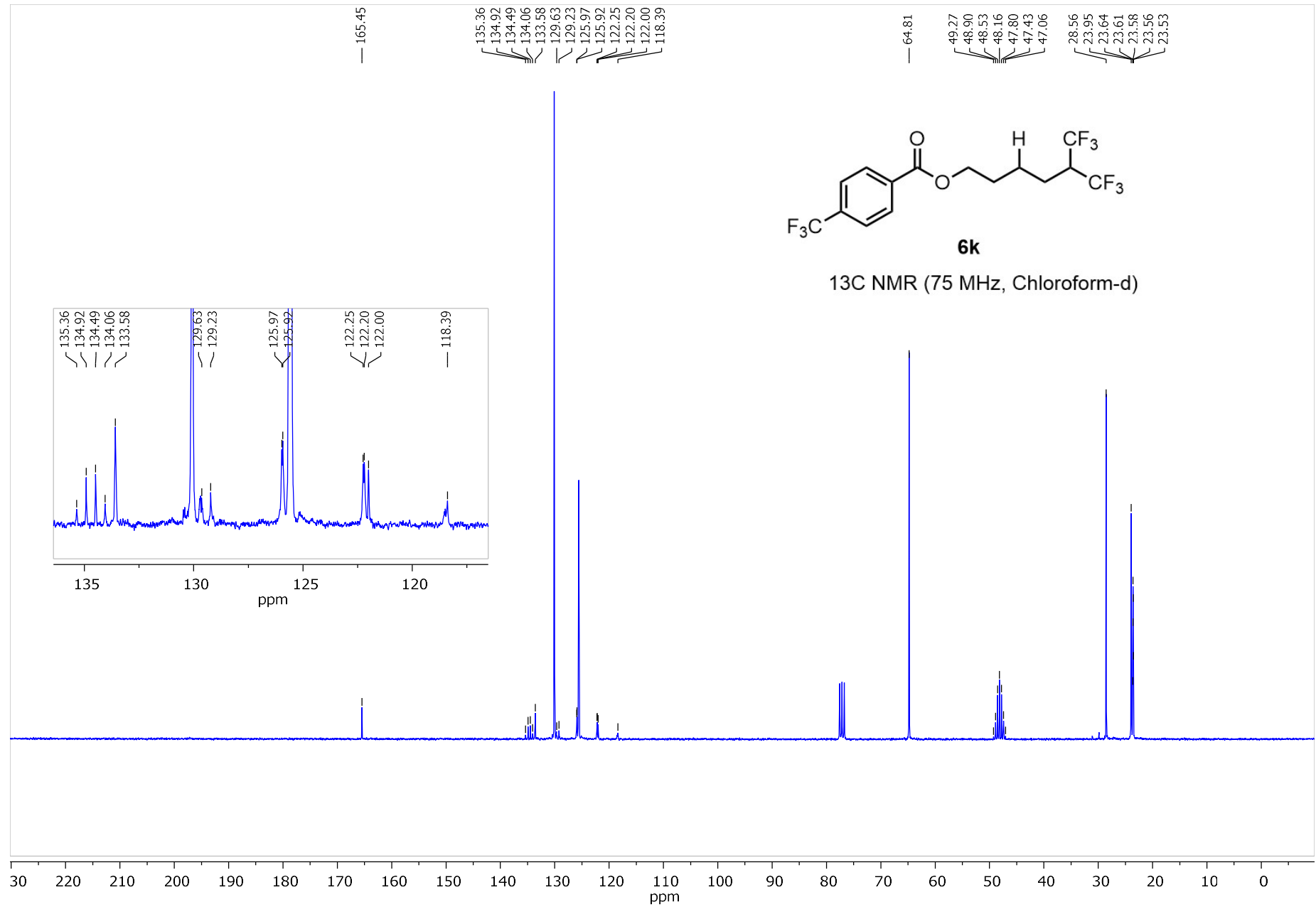


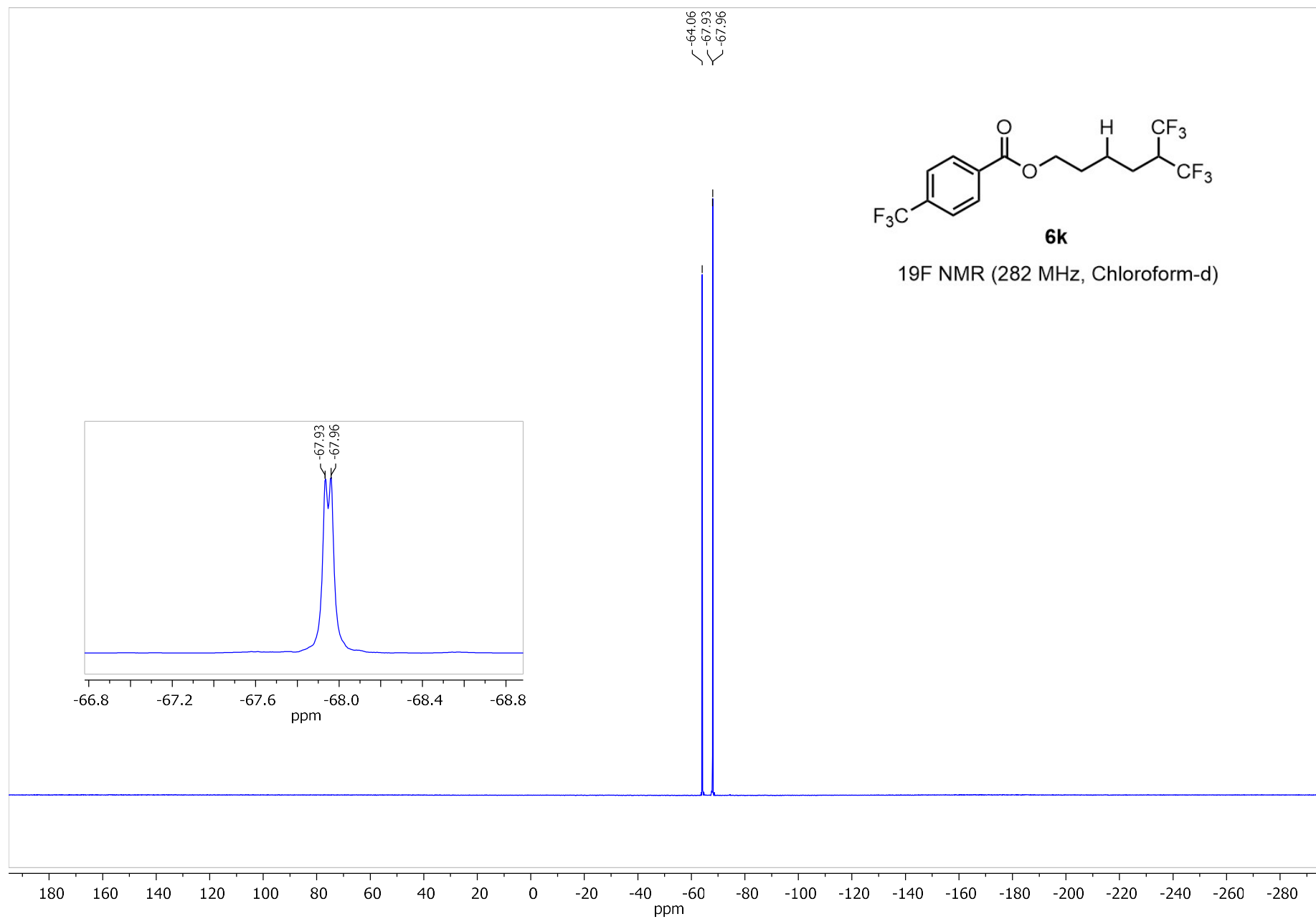


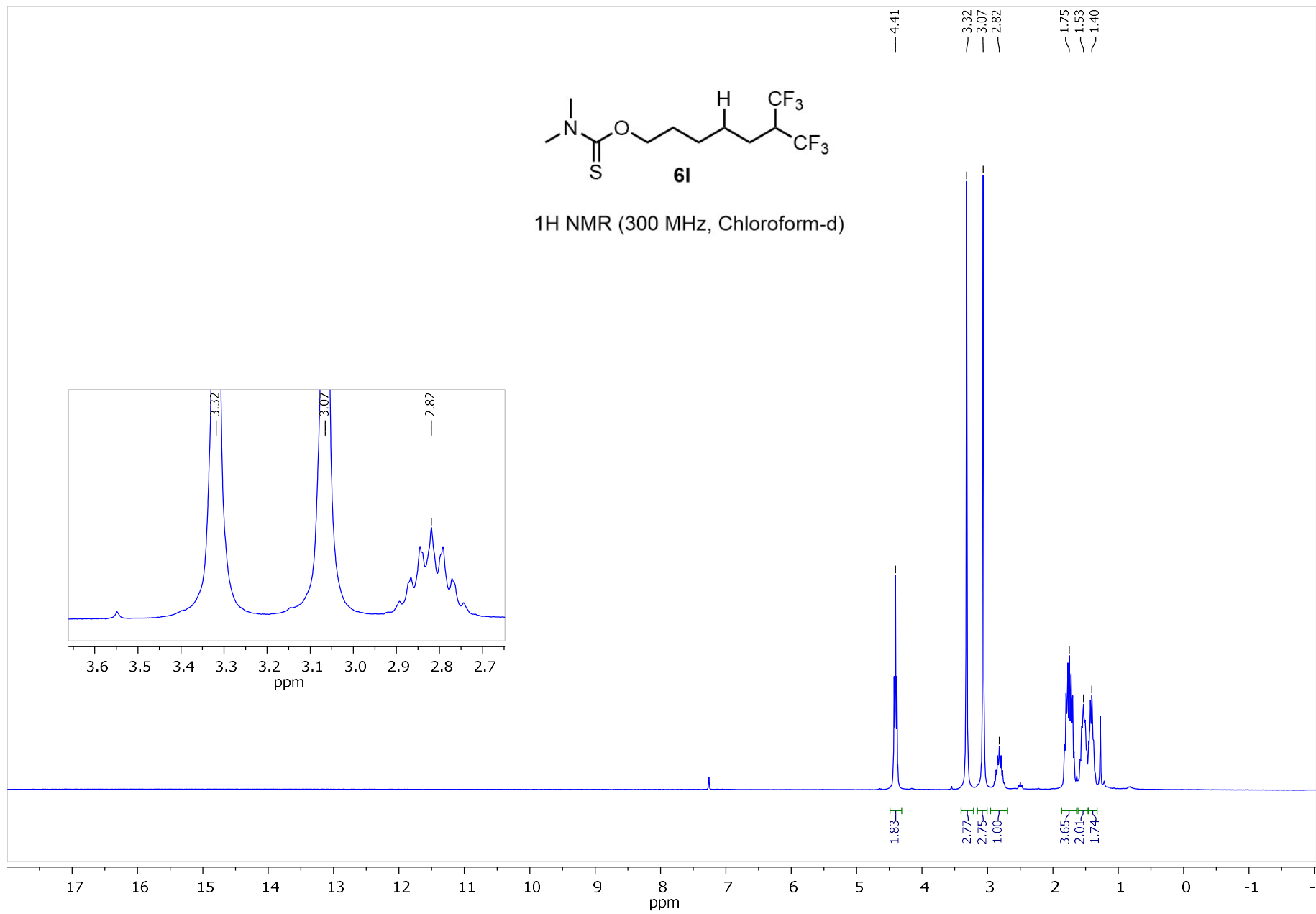
6j
19F NMR (282 MHz, Chloroform-d)

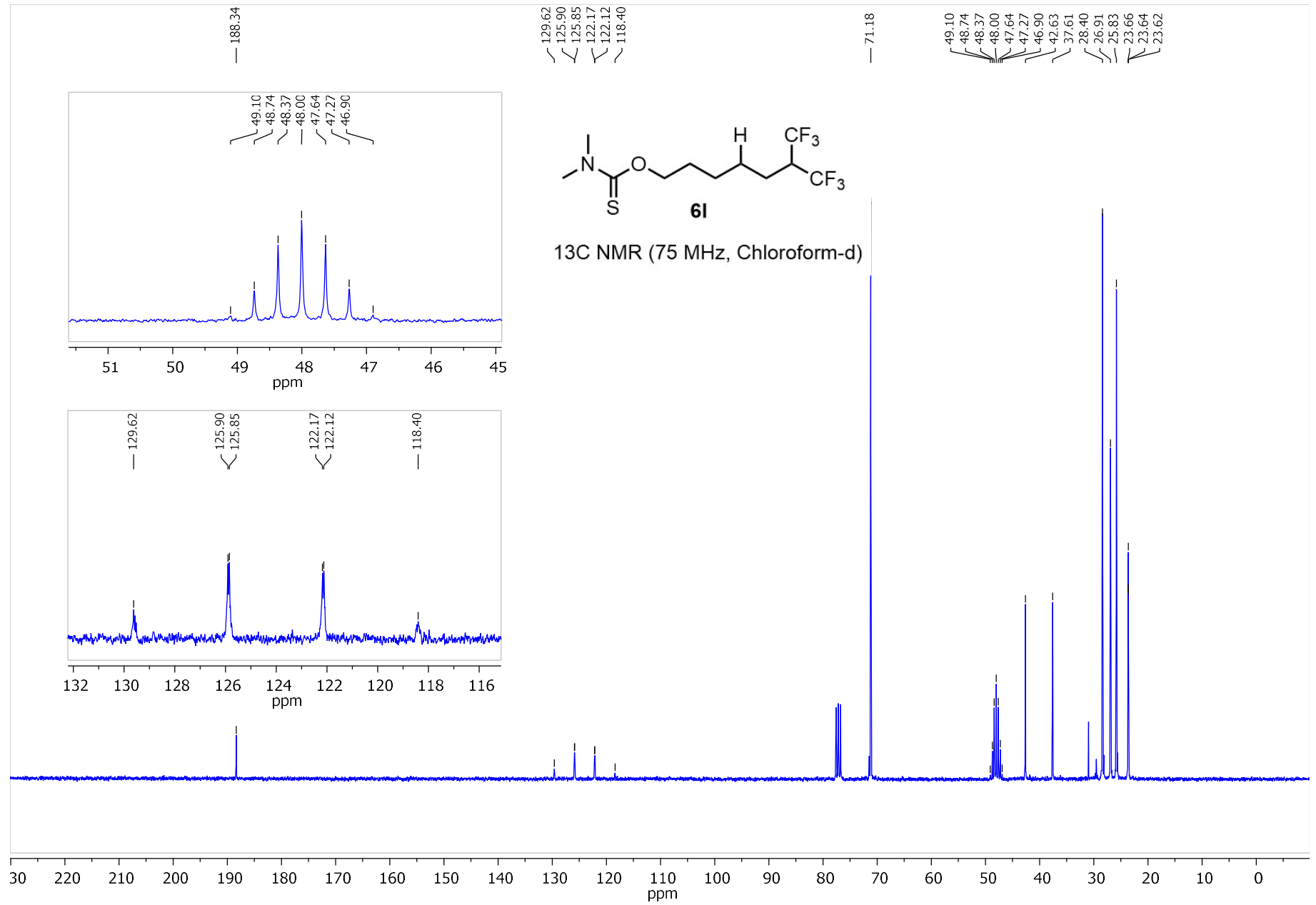


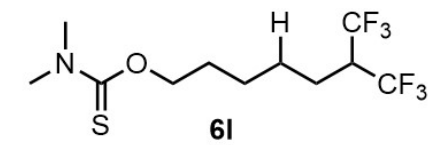




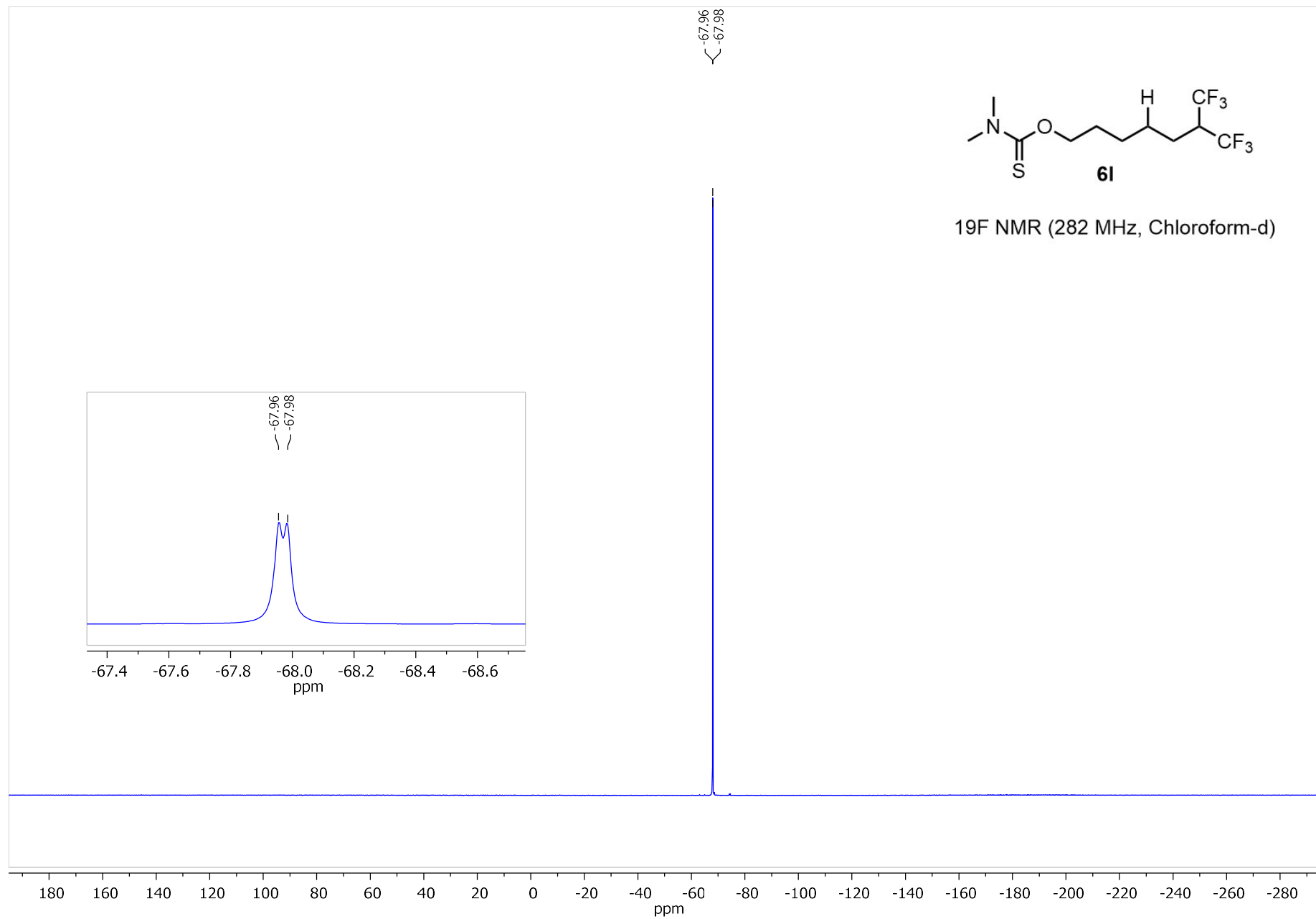


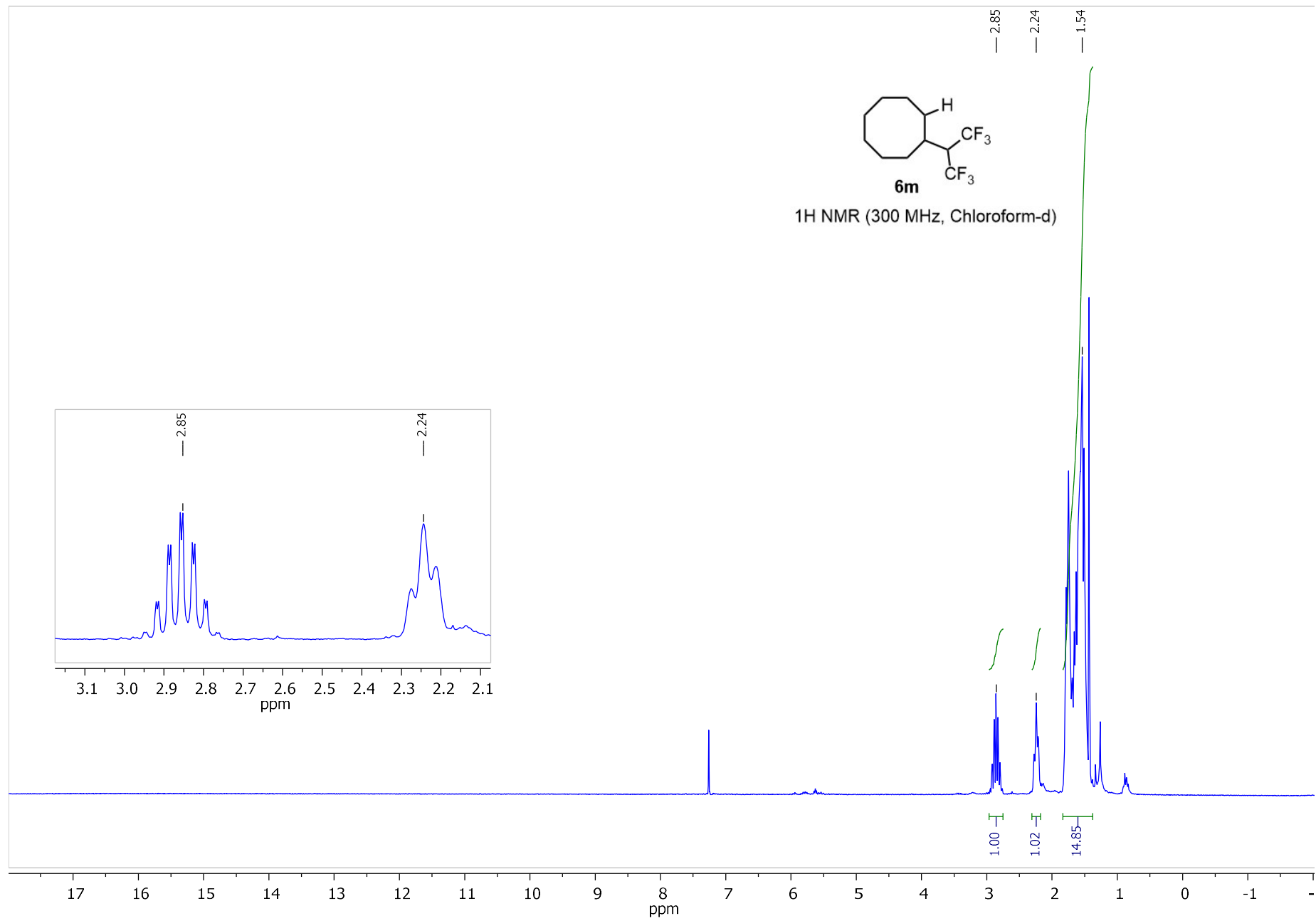


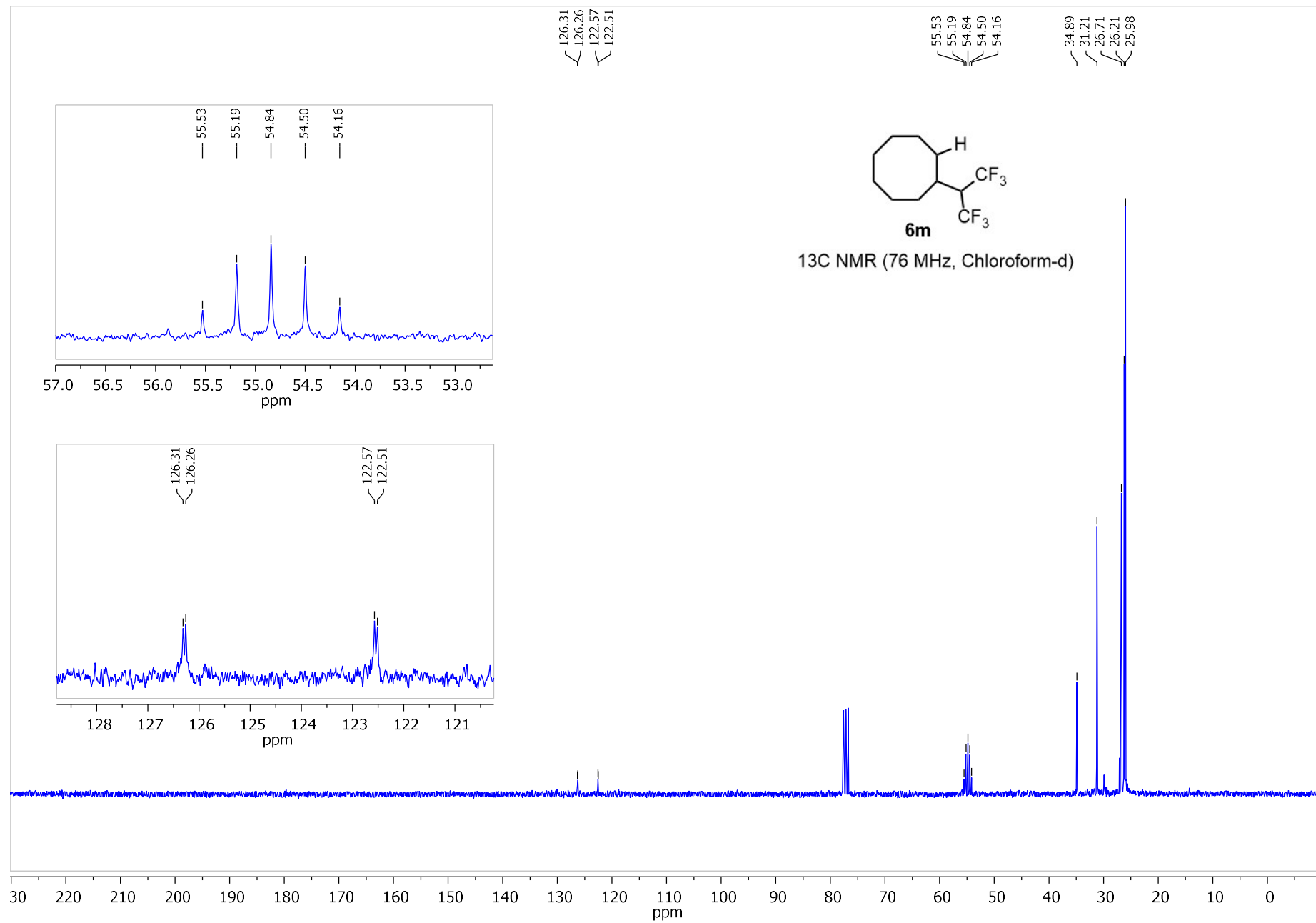


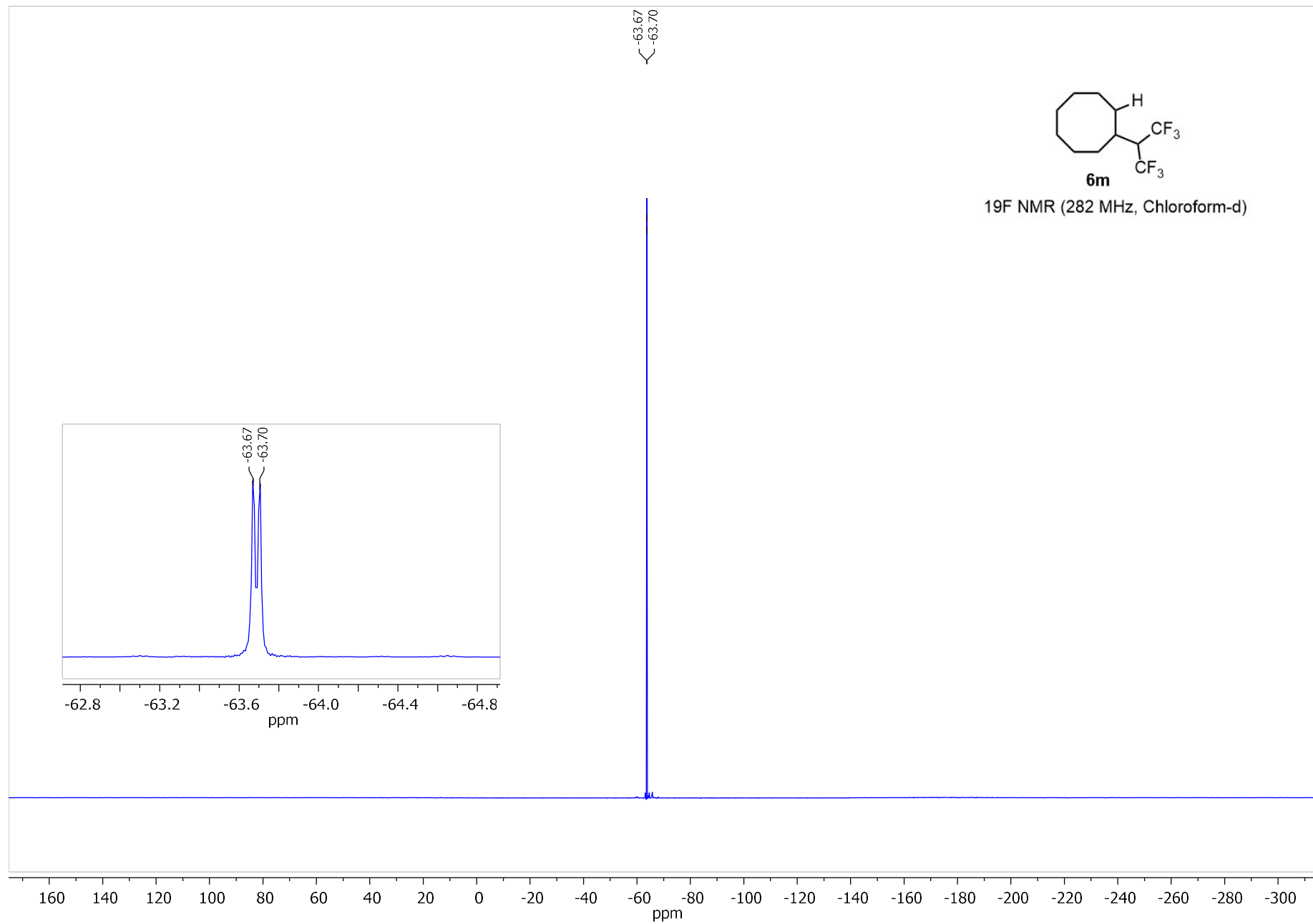


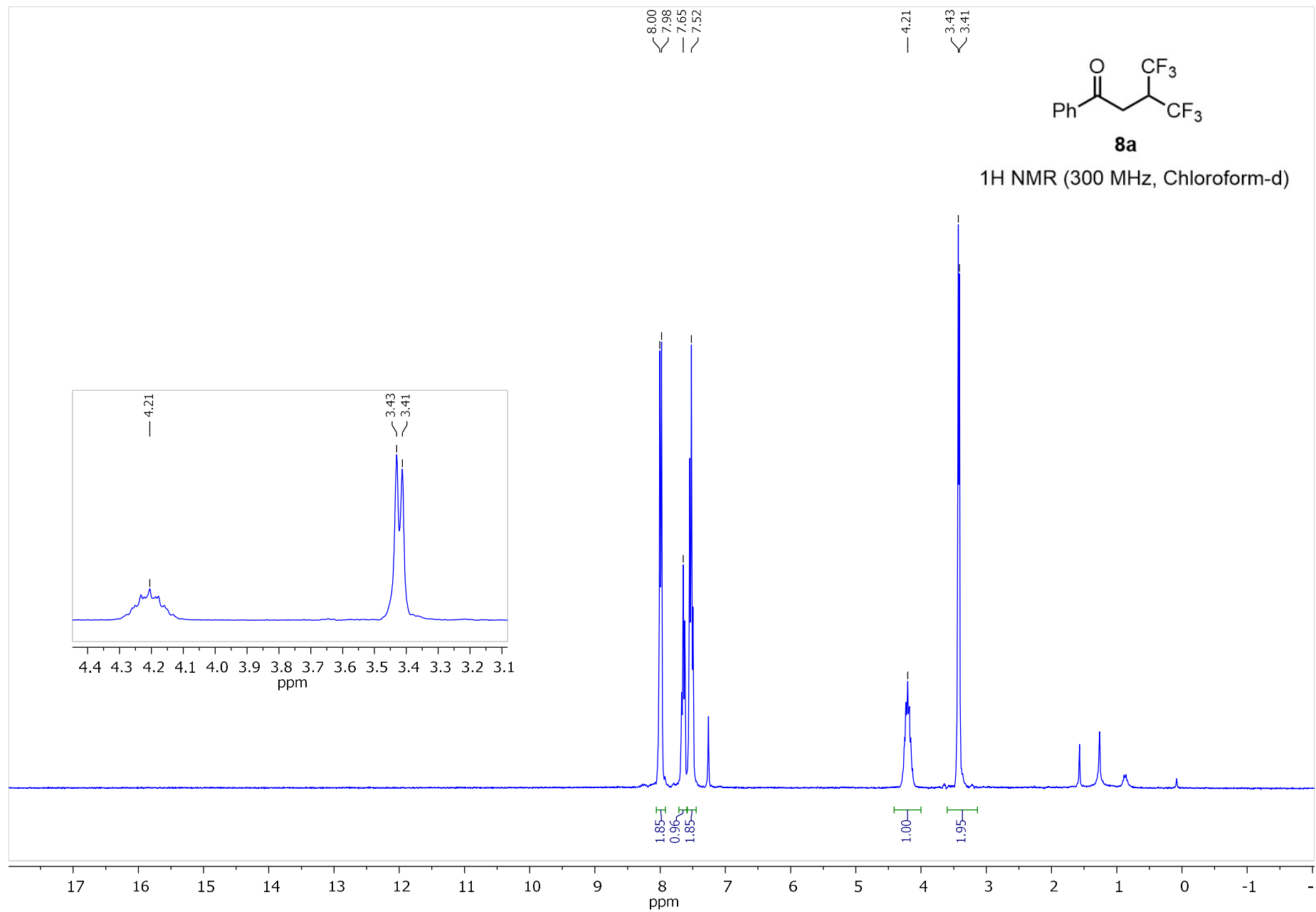
19F NMR (282 MHz, Chloroform-d)

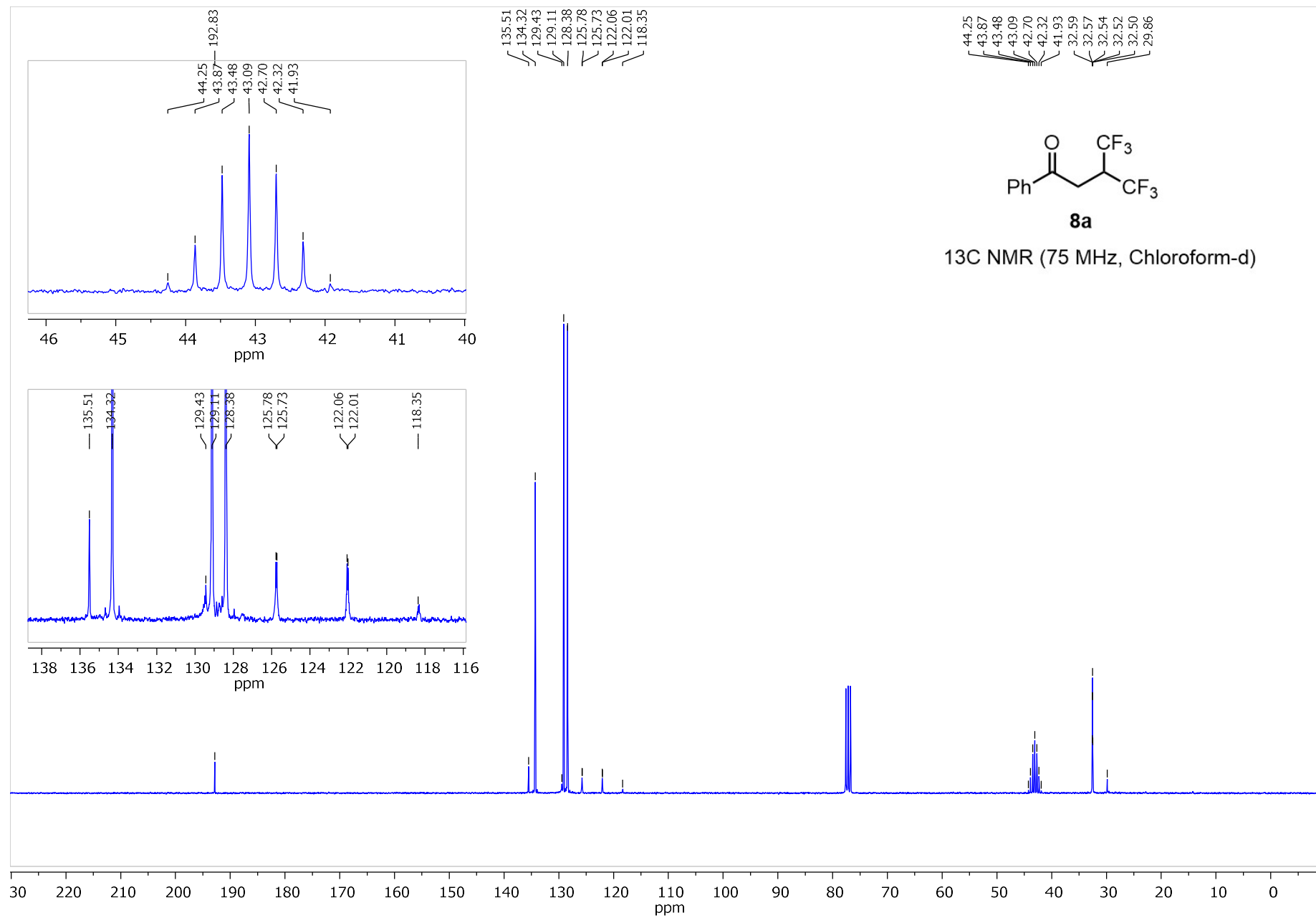


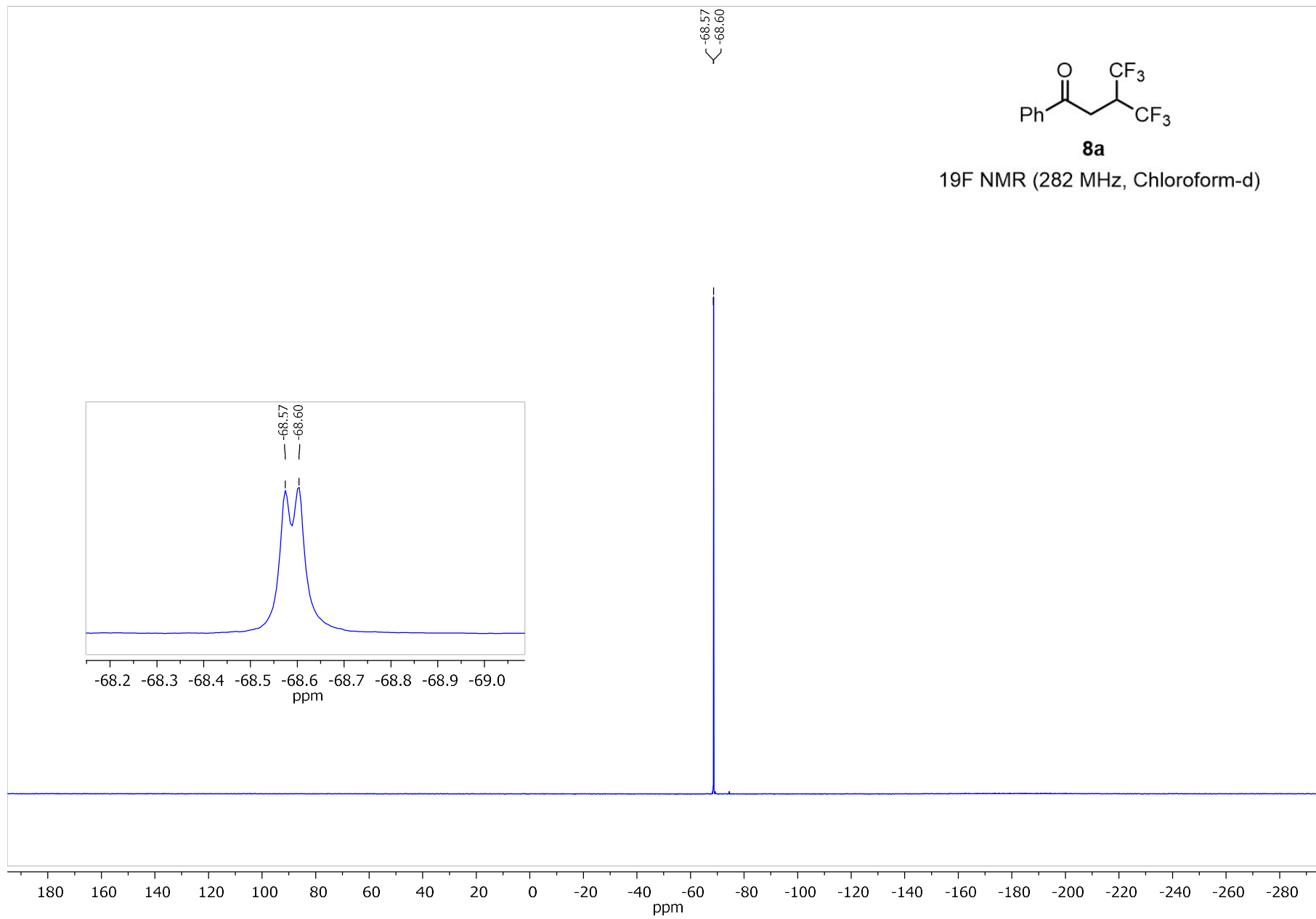
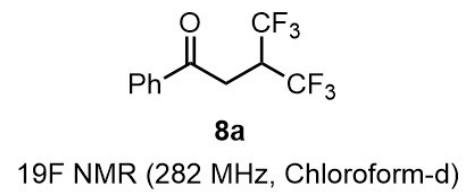


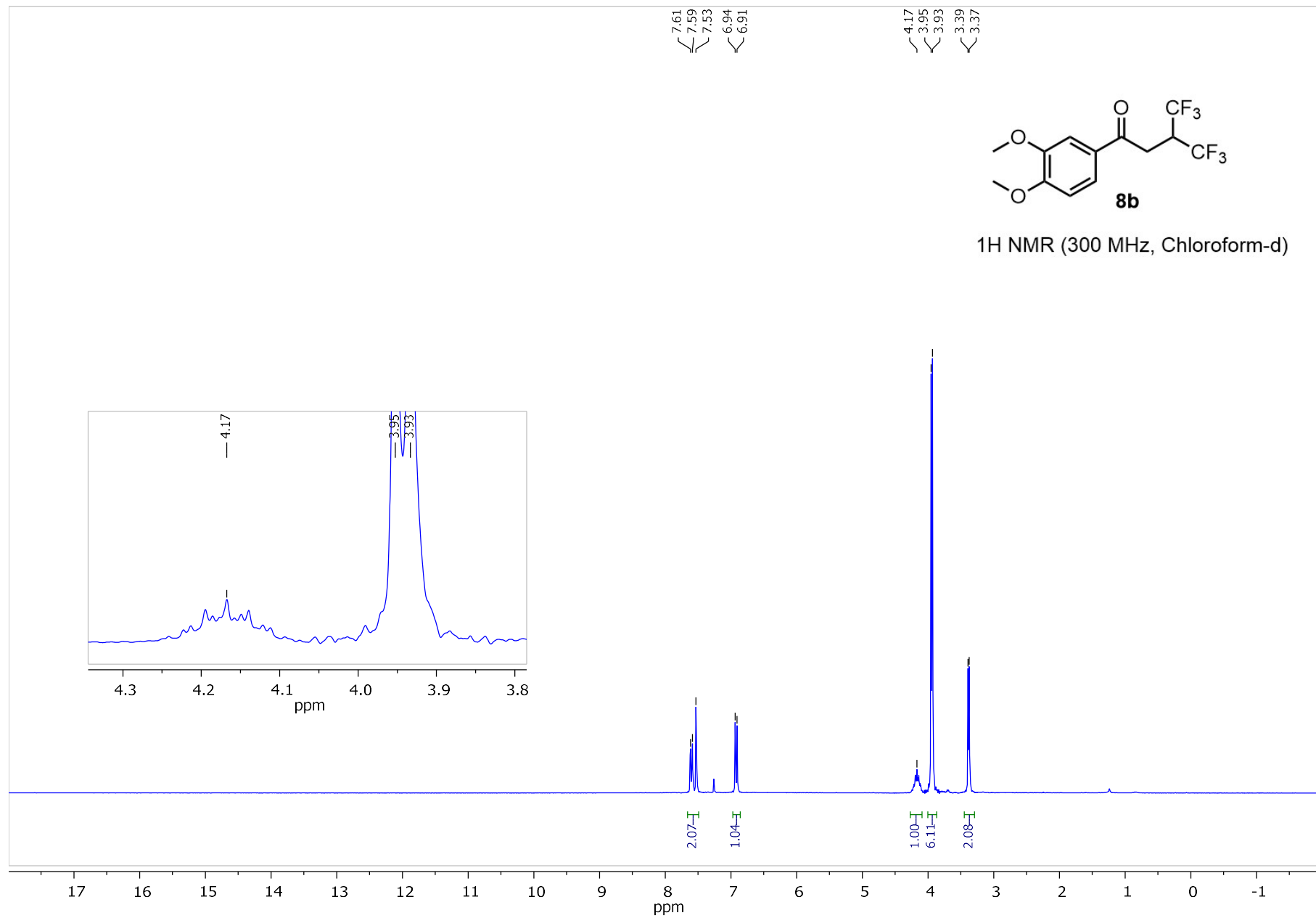


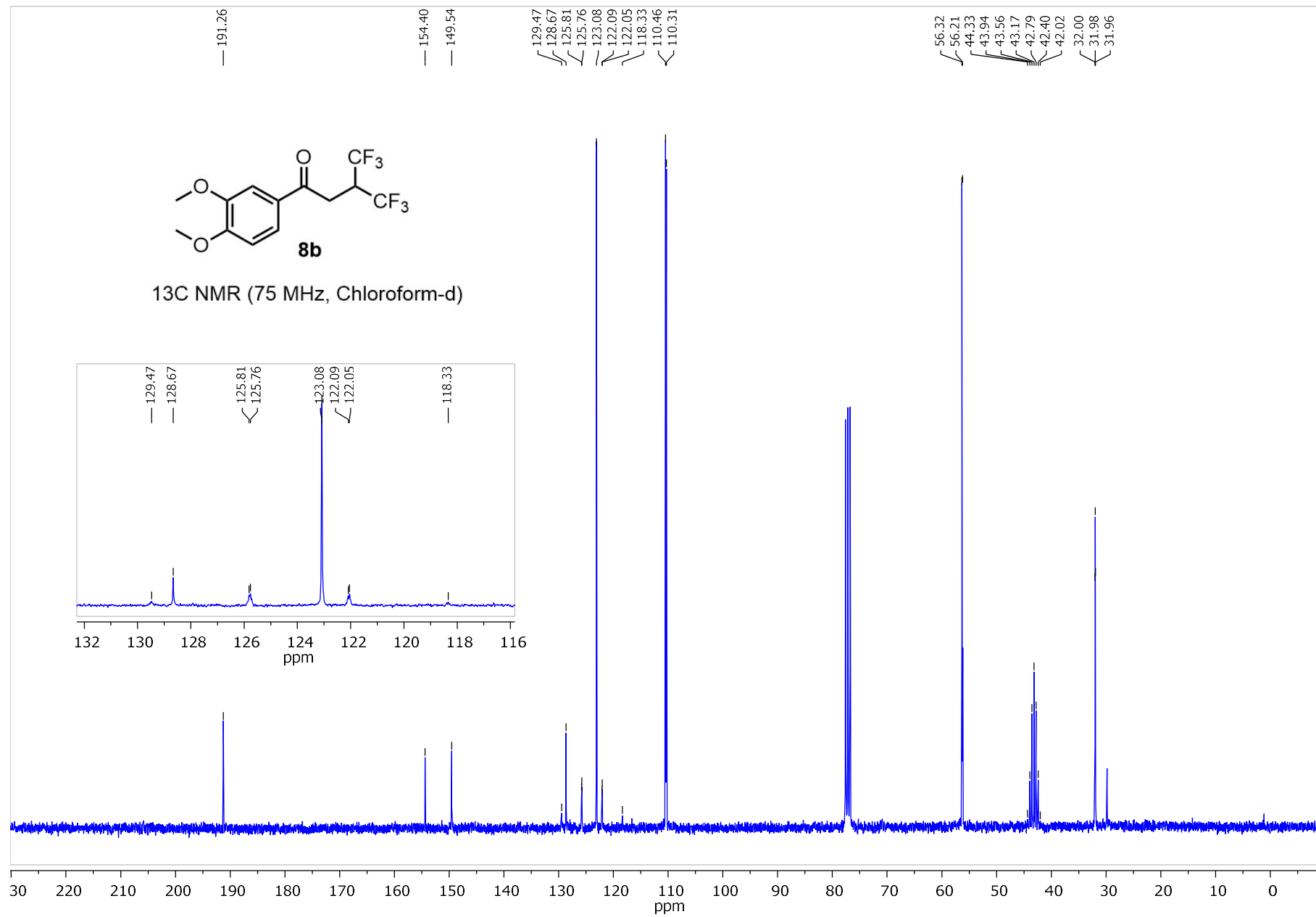


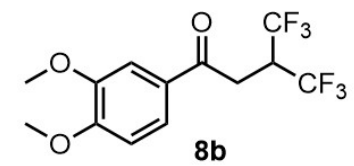












19F NMR (282 MHz, Chloroform-d)

